

Mathematical Notes on Mutect

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I. SOMATIC LIKELIHOODS MODEL

We have a set of potential somatic alleles and read-allele likelihoods $\ell_{ra} \equiv P(\text{read } r | \text{allele } a)$. We don't know which alleles are real somatic alleles and so we must compute, for each subset \mathbb{A} of alleles, the likelihood that the reads come from \mathbb{A} . A simple model for this likelihood is as follows: each read r is associated with a latent indicator vector \mathbf{z}_r with one-hot encoding $z_{ra} = 1$ iff read r came from allele $a \in \mathbb{A}$. The conditional probability of the reads \mathbb{R} given their allele assignments is

$$P(\mathbb{R} | \mathbf{z}, \mathbb{A}) = \prod_{r \in \mathbb{R}} \prod_a \ell_{ra}^{z_{ra}}. \quad (1)$$

The alleles are not equally likely because there is a latent vector \mathbf{f} of allele fractions – f_a is the allele fraction of allele a . Since the components of \mathbf{f} sum to one it is a categorical distribution and can be given a Dirichlet prior,

$$P(\mathbf{f}) = \text{Dir}(\mathbf{f} | \boldsymbol{\alpha}). \quad (2)$$

Then f_a is the prior probability that a read comes from allele a and thus the conditional probability of the indicators \mathbf{z} given the allele fractions \mathbf{f} is

$$P(\mathbf{z} | \mathbf{f}) = \prod_r \prod_a f_a^{z_{ra}}. \quad (3)$$

The full-model likelihood is therefore

$$\mathbb{L}(\mathbb{A}) = P(\mathbb{R}, \mathbf{z}, \mathbf{f} | \mathbb{A}) = \text{Dir}(\mathbf{f} | \boldsymbol{\alpha}) \prod_a \prod_r (f_a \ell_{ra})^{z_{ra}}. \quad (4)$$

And the marginalized likelihood of \mathbb{A} , that is, the model evidence for allele subset \mathbb{A} , is

$$P(\mathbb{R} | \mathbb{A}) = \sum_{\mathbf{z}} \int d\mathbf{f} \text{Dir}(\mathbf{f} | \boldsymbol{\alpha}) \prod_a \prod_r (f_a \ell_{ra})^{z_{ra}}, \quad (5)$$

where the integral is over the probability simplex $\sum_a f_a = 1$.

The integral over \mathbf{f} is the normalization constant of a Dirichlet distribution and as such we can simply look up its formula. However, the sum over all values of \mathbf{z} for all reads has exponentially many terms. We will get around this difficulty by handling \mathbf{z} with a mean-field approximation in which we factorize the likelihood as $\mathbb{L} \approx q(\mathbf{z})q(\mathbf{f})$. This approximation is exact in two limits: first, if there are many reads, each allele is associated with many reads and therefore the Law of Large Numbers causes \mathbf{f} and \mathbf{z} to become uncorrelated. Second, if the allele assignments of reads are obvious \mathbf{z}_r is effectively not a random variable at all (there is no uncertainty as to which of component is non-zero) and also becomes uncorrelated with \mathbf{f} .

In the variational Bayesian mean-field formalism the value of \mathbf{f} that \mathbf{z} “sees” is the expectation of $\log \mathbb{L}$ with respect to $q(\mathbf{f})$ and vice versa. That is,

$$q(\mathbf{f}) \propto \text{Dir}(\mathbf{f} | \boldsymbol{\alpha}) \prod_a \prod_r f_a^{\bar{z}_{ra}} \propto \text{Dir}(\mathbf{f} | \boldsymbol{\alpha} + \sum_r \bar{\mathbf{z}}_r), \quad (6)$$

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where $\bar{z}_{ra} \equiv E_q[z_{ra}]$, and

$$q(\mathbf{z}_r) = \prod_a (\tilde{f}_a \ell_{ra})^{z_{ra}}, \tilde{f}_a = \exp E[\ln f_a] \quad (7)$$

Because $q(\mathbf{z})$ is categorical and $q(\mathbf{f})$ is Dirichlet¹ the necessary mean fields are easily obtained and we have

$$\bar{z}_{ra} = \frac{\tilde{f}_a \ell_{ra}}{\sum_{a'} \tilde{f}_{a'} \ell_{ra'}} \quad (8)$$

and

$$\ln \tilde{f}_a = \psi(\alpha_a + \sum_r \bar{z}_{ra}) - \psi(\sum_{a'} \alpha_{a'} + N) \quad (9)$$

where ψ is the digamma function and N is the number of reads. To obtain $q(\mathbf{z})$ and $q(\mathbf{f})$ we iterate Equations 8 and 9 until convergence. A very reasonable initialization is to set $\bar{z}_{ra} = 1$ if a is the most likely allele for read r , 0 otherwise. Having obtained the mean field of \mathbf{z} , we would like to plug it into Eq 5. We can't do this directly, of course, because Eq 5 says nothing about our mean field factorization. Rather, we need the variational approximation (Bishop's Eq 10.3) to the model evidence, which is

$$\ln P(\mathbb{R}|\mathbb{A}) \approx \sum_{\mathbf{z}} \int d\mathbf{f} q(\mathbf{z}) q(\mathbf{f}) [\ln P(\mathbb{R}, \mathbf{z}, \mathbf{f}|\mathbb{A}) - \ln q(\mathbf{z}) - \ln q(\mathbf{f})] \quad (10)$$

$$= E_q [\ln P(\mathbb{R}, \mathbf{z}, \mathbf{f}|\mathbb{A})] - E_q [\ln q(\mathbf{z})] - E_q [\ln q(\mathbf{f})]. \quad (11)$$

Before we proceed, let's introduce some notation. First, from Eq 6 the posterior $q(\mathbf{f})$ is

$$q(\mathbf{f}) = \text{Dir}(\mathbf{f}|\boldsymbol{\beta}), \quad \boldsymbol{\beta} = \boldsymbol{\alpha} + \sum_r \bar{\mathbf{z}}_r. \quad (12)$$

Second, let's define the log normalization constant of a Dirichlet distribution as g so that

$$\ln \text{Dir}(\mathbf{f}|\boldsymbol{\omega}) = g(\boldsymbol{\omega}) + \sum_a (\omega_a - 1) \ln f_a, \quad g(\boldsymbol{\omega}) = \ln \Gamma(\sum_a \omega_a) - \sum_a \ln \Gamma(\omega_a). \quad (13)$$

Finally, define the Dirichlet mean log (aka "that digamma stuff") as h :

$$E_{\text{Dir}(\mathbf{f}|\boldsymbol{\omega})} [\ln f_a] = \psi(\omega_a) - \psi(\sum_{a'} \omega_{a'}) \equiv h_a(\boldsymbol{\omega}). \quad (14)$$

The log of Eq 4 is

$$\ln P(\mathbb{R}, \mathbf{z}, \mathbf{f}|\mathbb{A}) = g(\boldsymbol{\alpha}) + \sum_a (\alpha_a - 1) \ln f_a + \sum_{ra} z_{ra} (\ln f_a + \ln \ell_{ra}). \quad (15)$$

and thus the first term in Eq 11 is

$$E_q [\ln P(\mathbb{R}, \mathbf{z}, \mathbf{f}|\mathbb{A})] = g(\boldsymbol{\alpha}) + \sum_a (\alpha_a - 1) h_a(\boldsymbol{\beta}) + \sum_{ra} \bar{z}_{ra} (h_a(\boldsymbol{\beta}) + \ln \ell_{ra}) \quad (16)$$

$$= g(\boldsymbol{\alpha}) + \sum_a (\beta_a - 1) h_a(\boldsymbol{\beta}) + \sum_{ra} \bar{z}_{ra} \ln \ell_{ra}, \quad (17)$$

where we used the relationship $\boldsymbol{\beta} = \boldsymbol{\alpha} + \sum_r \bar{\mathbf{z}}_r$.

The second term in Eq 11 is

$$-E_q [\ln q(\mathbf{z})] = -\sum_{ra} \bar{z}_{ra} \ln \bar{z}_{ra}. \quad (18)$$

¹ Note that we didn't *impose* this in any way. It simply falls out of the mean field equations.



FIG. 1: The probabilistic graphical model

The third term in Eq 11 is

$$-E_q[\ln q(\mathbf{f})] = -g(\boldsymbol{\beta}) - \sum_a (\beta_a - 1) E_q[\ln f_a] = -g(\boldsymbol{\beta}) - \sum_a (\beta_a - 1) h_a(\boldsymbol{\beta}). \quad (19)$$

Adding Eqs 17, 18, and 19 and noting the cancellation between parts of Eqs 17 and 19 we obtain

$$\ln P(\mathbb{R}|\mathbb{A}) \approx g(\boldsymbol{\alpha}) - g(\boldsymbol{\beta}) + \sum_{ra} \bar{z}_{ra} (\ln \ell_{ra} - \ln \bar{z}_{ra}). \quad (20)$$

We now have the model evidence for allele subset \mathbb{A} . This lets us choose which alleles are true somatic variants. It also lets us make calls on somatic loss of heterozygosity events. Furthermore, instead of reporting max-likelihood allele fractions as before, we may emit the parameters of the Dirichlet posterior $q(\mathbf{f})$, which encode both the maximum likelihood allele fractions and their uncertainty.

II. STRAND ARTIFACT MODEL

- \mathbf{z} is a latent random variable having a 1-of-K representation. For each variant locus, \mathbf{z} encodes the presence of strand artifact in forward reads ($[1, 0, 0]$), artifact in reverse reads ($[0, 1, 0]$), or no artifact ($[0, 0, 1]$)
- $f \sim \text{Unif}(0, 1)$ is a prior distribution over alt allele fraction f
- $\epsilon \sim \text{Beta}(\alpha, \beta)$ is a prior distribution over the error probability on a read on the artifact strand. For instance, if we have strand artifact on the reverse strand (i.e. $\mathbf{z} = [0, 1, 0]$), ϵ is the probability that the sequencer reads a ref allele on a reverse read as alt
- $x^+ | f, \epsilon, \mathbf{z}$ is the number of forward reads with the alt allele. It's a mixture of binomials, defined as follows:

$$x^+ | f, \epsilon, \mathbf{z} \sim \begin{cases} \text{Bin}(n^+, f + \epsilon(1 - f)) & \mathbf{z} = \text{Art}+ \\ \text{Bin}(n^+, f) & \mathbf{z} = \text{Art}- \\ \text{Bin}(n^+, f) & \mathbf{z} = \text{noArt} \end{cases} \quad (21)$$

We compute the conditional distributions of x^- analogously.

Having observed the read counts in the forward and reverse directions, we can compute the posterior probabilities of the latent variable \mathbf{z} . Below we derive the unnormalized posterior probability of strand artifact in forward reads ($\mathbf{z} = \text{art}+$), given that we observed x^+ forward alt reads and x^- reverse alt reads. We use a shorthand z_0 to denote $\mathbf{z} = \text{art}+$ for conciseness.

First we will derive the likelihood $p(x^+, x^-, f, \epsilon | z_0)$

$$p(x^+, x^- | z_0) = \iint_{f, \epsilon} p(x^+, x^-, f, \epsilon | z_0) df d\epsilon \quad (22)$$

$$= \iint_{f, \epsilon} p(f)p(\epsilon)p(x^+, x^- | z_0, f, \epsilon) df d\epsilon \quad (23)$$

$$= \iint_{f, \epsilon} p(f)p(\epsilon)p(x^+ | z_0, f, \epsilon)p(x^- | z_0, f, \epsilon) df d\epsilon \quad (24)$$

$$= \iint_{f, \epsilon} p(\epsilon)p(x^+ | z_0, f, \epsilon)p(x^- | z_0, f, \epsilon) df d\epsilon \quad (25)$$

$$= \iint_{f, \epsilon} \text{Beta}(\epsilon | \alpha, \beta) \text{Bin}(x^+ | f + \epsilon(1 - f), n^+) \text{Bin}(x^- | f, n^-) df d\epsilon \quad (26)$$

The posterior probability of strand artifact in forward reads is therefore

$$p(z_0 | x^+, x^-) \propto p(z_0)p(x^+, x^- | z_0) \quad (27)$$

$$= p(z_0) \iint_{f, \epsilon} \text{Beta}(\epsilon | \alpha, \beta) \text{Bin}(x^+ | f + \epsilon(1 - f), n^+) \text{Bin}(x^- | f, n^-) df d\epsilon \quad (28)$$

The derivation for the probability of strand artifact on reverse strand is analogous.

For the case of no strand artifact, the derivation of likelihoods is identical up to (25). Here we can simply the equation to a single integral over f because the conditional probabilities of x^+ and x^- do not depend on ϵ . We use a shorthand z_2 for $z = \text{noArt}$

$$\begin{aligned} p(x^+, x^- | z_2) &= \iint_{f, \epsilon} p(\epsilon)p(x^+ | z_2, f, \epsilon)p(x^- | z_2, f, \epsilon) df d\epsilon \\ &= \int_f p(x^+ | z_2, f)p(x^- | z_2, f) df \int_\epsilon p(\epsilon) d\epsilon \end{aligned} \quad (29)$$

$$= \int_f \text{Bin}(x^+ | f, n^+) \text{Bin}(x^- | f, n^-) df \quad (30)$$

And the posterior probability is

$$p(z_2 | x^+, x^-) \propto p(z_2)p(x^+, x^- | z_2) \quad (31)$$

$$= p(z_2) \int_f \text{Bin}(x^+ | f, n^+) \text{Bin}(x^- | f, n^-) df \quad (32)$$

III. GERMLINE FILTER

Suppose we have detected an allele such that its (somatic) likelihood in the tumor is ℓ_t and its (diploid) likelihood in the normal is ℓ_n . By convention, both of these are relative to a likelihood of 1 for the allele *not* to be found. If we have no matched normal, $\ell_n = 1$. Suppose we also have the population allele frequency f of this allele. Then the prior probability for the normal to be heterozygous or homozygous alt for the allele is $2f(1 - f) + f^2$ and the prior probability for the normal genotype not to contain the allele is $(1 - f)^2$. Finally, suppose that the prior for this allele to arise as a somatic variant is π .

We can determine the posterior probability that the variant exists in the normal genotype by calculating the unnormalized probabilities of four possibilities:

1. The variant exists in both the normal and the tumor samples. This has unnormalized probability $(2f(1 - f) + f^2) \ell_n \ell_t (1 - \pi)$.
2. The variant exists in the tumor but not the normal. This has unnormalized probability $(1 - f)^2 \ell_t \pi$.

3. The variant exists in neither the tumor nor the normal. This has unnormalized probability $(1 - f)^2(1 - \pi)$.
4. The variants exists in the normal but not the tumor. This is biologically very unlikely. Furthermore, if it *did* occur we wouldn't care about filtering the variant as a germline event because we wouldn't call it as a somatic event. Thus we neglect this possibility.

Normalizing, we obtain the following posterior probability that an allele is a germline variant:

$$P(\text{germline}) = \frac{(1)}{(1) + (2) + (3)} = \frac{(2f(1 - f) + f^2) \ell_n \ell_t (1 - \pi)}{(2f(1 - f) + f^2) \ell_n \ell_t (1 - \pi) + (1 - f)^2 \ell_t \pi + (1 - f)^2 (1 - \pi)}. \quad (33)$$

To filter, we set a threshold on this posterior probability.

IV. FINDING ACTIVE REGIONS

Mutect triages sites based on their pileup at a single base locus. If there is sufficient evidence of variation Mutect proceeds with local reassembly and realignment. As in the downstream parts of Mutect we seek a likelihood ratio between the existence and non-existence of an alt allele. Instead of obtaining read likelihoods via Pair-HMM, we assign each base a likelihood. For substitutions we can simply use the base quality. For indels we assign a heuristic effective quality that increases with length. Supposing we have an effective quality for each element in the read pileup we can now estimate the likelihoods of no variation and of a true alt allele with allele fraction f . Let \mathcal{R} and \mathcal{A} denote the sets of ref and alt reads. The likelihood of no variation is the likelihood that every alt read was in error. Letting ϵ_i be the error probability of pileup element i we have:

$$L(\text{no variation}) = \prod_{i \in \mathcal{R}} (1 - \epsilon_i) \prod_{j \in \mathcal{A}} \epsilon_j \quad (34)$$

$$L(f) = \prod_{i \in \mathcal{R}} [(1 - f)(1 - \epsilon_i) + f\epsilon_i] \prod_{j \in \mathcal{A}} [f(1 - \epsilon_j) + (1 - f)\epsilon_j] \quad (35)$$

The terms that signify observed ref reads that are actually alt reads with an error and vice versa are negligible² Then we get

$$L(f) \approx \prod_{i \in \mathcal{R}} [(1 - f)(1 - \epsilon_i)] \prod_{j \in \mathcal{A}} [f(1 - \epsilon_j)] \quad (36)$$

$$= (1 - f)^{|\mathcal{R}|} f^{|\mathcal{A}|} \prod_{i \in \mathcal{R}} (1 - \epsilon_i) \prod_{j \in \mathcal{A}} (1 - \epsilon_j) \quad (37)$$

We can integrate over the latent variable f from 0 to 1 with a flat prior analytically because the integral is the normalization constant of the beta distribution:

$$\int_0^1 (1 - f)^{|\mathcal{R}|} f^{|\mathcal{A}|} df = \frac{\Gamma(|\mathcal{R}| + 1) \Gamma(|\mathcal{A}| + 1)}{\Gamma(|\mathcal{R}| + |\mathcal{A}| + 2)} = \frac{|\mathcal{R}|! |\mathcal{A}|!}{(|\mathcal{R}| + |\mathcal{A}| + 1)!} \quad (38)$$

In the likelihood ratio the reference factors $\prod_{i \in \mathcal{R}} (1 - \epsilon_i)$ cancel, leaving a log-odds of

$$\text{LOD} \approx \sum_{j \in \mathcal{A}} [\log(1 - \epsilon_j) - \log \epsilon_j] + \log \frac{|\mathcal{R}|! |\mathcal{A}|!}{(|\mathcal{R}| + |\mathcal{A}| + 1)!} \quad (39)$$

$$\approx - \sum_{j \in \mathcal{A}} \log \epsilon_j + \log \frac{|\mathcal{R}|! |\mathcal{A}|!}{(|\mathcal{R}| + |\mathcal{A}| + 1)!}, \quad (40)$$

the first term of which is proportional to the sum of effective base qualities.

² We can set an upper bound on the error in the log likelihood by Taylor-expanding to first order. The error turns out to be quite small.

V. CALCULATING CONTAMINATION

Below, we present the GATK's fast, simple, and accurate method for calculating the contamination of a sample. This method does not require a matched normal, makes no assumptions about the number of contaminating samples, and remains accurate even when the sample has a lot of copy number variation.

The inputs to our tool are a bam file and a vcf of common variants – for example ExAC, gnomAD, or 1000 Genomes – with their allele frequencies. The basic idea is simply to count ref reads at hom alt sites and subtract the number of ref reads expected from sequencing error to obtain the number of ref reads contaminating these hom alt sites. Finally, we use the allele frequencies to account for the fact that some contaminating reads have the alt allele. The only subtlety is in distinguishing hom alt sites from loss of heterozygosity events, which we describe below.

Suppose we have a set \mathbb{H} of SNPs at which our sample is homozygous for the alternate allele. Let N_{ref} be the total number of ref reads at these sites. We can decompose N_{ref} as follows:

$$N_{\text{ref}} = N_{\text{ref}}^{\text{error}} + N_{\text{ref}}^{\text{contamination}}, \quad (41)$$

where $N_{\text{ref}}^{\text{error}}$ and $N_{\text{ref}}^{\text{contamination}}$ are as the number of ref reads due to error and contamination, respectively. We can obtain N_{ref} by counting reads, and we estimate $N_{\text{ref}}^{\text{error}}$ as follows. Suppose, WLOG, that the ref allele is A and the alt is C. Then, assuming that all substitution errors are equally likely, $N_{\text{ref}}^{\text{error}}$ is approximately half the number of Gs and Ts. This is, of course, not a perfect assumption for any one site, but on average over all the sites in \mathbb{H} it is very good.

Next we take the expectation of both sides of Equation 41 to obtain

$$\langle N_{\text{ref}} - N_{\text{ref}}^{\text{error}} \rangle = \left\langle \sum_{s \in \mathbb{H}} \text{number of contaminant ref reads at } s \right\rangle \quad (42)$$

$$= \sum_{s \in \mathbb{H}} \langle \text{number of contaminant ref reads at } s \rangle \quad (43)$$

$$= \sum_{s \in \mathbb{H}} \langle \text{number of contaminant reads at } s \times \text{ref fraction of contaminant reads at } s \rangle \quad (44)$$

$$= \sum_{s \in \mathbb{H}} \langle \text{number of contaminant reads at } s \rangle \times \langle \text{ref fraction of contaminant reads at } s \rangle \quad (45)$$

where we have used the linearity of the expectation and the independence of the total number of contaminant reads with the fraction of contaminant reads that are ref. The expectation of the total number of contaminant reads is the depth d_s at site s times the contamination, which we denote by χ . The expected fraction of contaminant reads that are ref is one minus the alt allele frequency f_s . Crucially, this fact is independent of how many contaminating samples there are. Thus we have

$$\langle N_{\text{ref}} - N_{\text{ref}}^{\text{error}} \rangle = \chi \sum_{s \in \mathbb{H}} d_s (1 - f_s) \quad (46)$$

and obtain the estimate

$$\hat{\chi} \approx \frac{N_{\text{ref}} - N_{\text{ref}}^{\text{error}}}{\sum_{s \in \mathbb{H}} d_s (1 - f_s)} \quad (47)$$

Let us now roughly estimate the error bars on this result. Under normal conditions sequencing error is rare, so we will estimate the variance due to the sampling error of the number of contaminating ref reads. As discussed above, the probability of a read at site s being a contaminating ref is $\chi(1 - f_s)$, so the quantity in the numerator of Eq. 47 is distributed as $\sum_s \text{Binom}(d_s, \chi(1 - f_s))$ and therefore has variance

$$(\text{var})(N_{\text{ref}} - N_{\text{ref}}^{\text{error}}) = \sum_s d_s \chi (1 - f_s) (1 - \chi(1 - f_s)) < \chi \sum_s d_s (1 - f_s) \quad (48)$$

And thus the variance of Eq. 47 is bounded as

$$\text{var}(\hat{\chi}) < \frac{\chi \sum_s d_s (1 - f_s)}{(\sum_{s \in \mathbb{H}} d_s (1 - f_s))^2} = \frac{\chi}{\sum_{s \in \mathbb{H}} d_s (1 - f_s)}, \quad (49)$$

which gives the following bound for the standard deviation:

$$\text{std}(\hat{\chi}) < \left[\frac{\chi}{\sum_{s \in \mathbb{H}} d_s (1 - f_s)} \right]^{1/2} < \left[\frac{\chi}{\sum_{s \in \mathbb{H}} d_s} \right]^{1/2} \quad (50)$$

This standard deviation is quite small. For example, in an exome with 1000 hom alt sites (as is typical) and an average depth of 30 reads, a true contamination of 0.05 is estimated with an error of 0.0013.

It remains to describe how we determine which sites are hom alt. Our simple heuristic is to consider only sites with sufficient depth and fraction of alt reads, say 80 percent. (If this measure fails to find enough hom alt sites, we can conclude that contamination is greater than 20 percent, which is so large as to make the sample worthless for somatic variant calling). We must additionally reject apparent hom alt sites that are actually het sites with deletion of the ref allele. We do this by looking for copy number variation and loss of heterozygosity in the vicinity of this site. To find sites with anomalous copy number, we compute the average depth smoothed over a scale of one megabase and compare it to the average depth of the sample as a whole and reject sites that show evidence of a deletion. To find loss of heterozygosity, we obtain ref and alt read counts of common SNPs within a megabase and compare the number of hets found to that expected based on the known allele frequencies. If too few hets are found, we reject the site. These two heuristics suffice to screen out false hom alts.

VI. PROPOSED TUMOR IN NORMAL ESTIMATION TOOL

Similar to the spirit of CalculateContamination, the fraction of tumor reads in the normal bam is a single number with a large amount of evidence and is probably well-estimated by simple descriptive statistics rather than a full-fledged probabilistic model. It shouldn't be much more complicated than finding somatic variants and comparing their signal in the normal sample to that in the tumor.

We propose the following steps to obtain our input of confident somatic SNVs:

- Run Mutect in tumor-only mode to obtain a preliminary list of somatic SNVs. For the sake of speed, we could implement a pileup-based mode in which we skip reassembly and equate read likelihoods with base qualities. This would allow us to obtain variant annotations using the existing architecture of Mutect and therefore to filter calls with no new code. It would probably make sense at this stage to filter more stringently than usual based on population allele frequencies in gnomAD.
- Run FilterMutectCalls. With default settings this eliminates the great majority of sequencing artifacts. To eliminate even more we could increase the log odds threshold slightly, essentially requiring a slightly larger alt allele count. Normally we don't do this because it sacrifices some sensitivity, but for our purposes here a 10 or 20 percent loss of sensitivity is perfectly acceptable as long as we are left with enough SNVs for our estimate.
- Remove all SNVs that have enough read counts in the normal that we conclude they are germline variants, as opposed to tumor in normal contamination. We could also accomplish this by using tumor-normal mode in the first step, but we would need to modify our active region determination which currently would mark a region as inactive based only on a small number of tumor-in-normal alt reads. We would also have to turn off the normal artifact filter.

The above steps are all very reliable, so at this point we can assume we have a collection of confident biallelic somatic SNVs that are hom ref in the germline. Similar to CalculateContamination, we can now estimate the number of alt reads in the normal at these sites:

$$\text{alt in normal} \approx \sum_{\text{sites}} (\text{depth in normal}) \times (\text{alt fraction in tumor}) \times (\text{tumor in normal fraction}) \quad (51)$$

Hence we estimate

$$\text{tumor in normal fraction} \approx \frac{\text{total number of alt reads in normal at somatic SNV sites}}{\sum_{\text{somatic SNV sites}} (\text{depth in normal}) \times (\text{alt fraction in tumor})} \quad (52)$$

VII. MUTECT FILTERS

Mutect2 emits candidate variants with a set of annotations. After that FilterMutectCalls produces filtered calls by subjecting these variants to a series of hard filters that reject sites if some annotation is out of an allowable range. Here are the command line arguments that control these filters, along with the annotations they relate to.

- **tumor_lod** is the minimum likelihood of an allele as determined by the somatic likelihoods model required to pass.
- **maxEventsInHaplotype** is the maximum allowable number of called variants co-occurring in a single assembly region. If the number of called variants exceeds this they will all be filtered. Note that this filter is misnamed because it counts the total number of events over all haplotypes in an assembly region.
- **uniqueAltReadCount** is the minimum number of unique (start position, fragment length) pairs required to make a call. This count is a proxy for the number of unique molecules (as opposed to PCR duplicates) supporting an allele. Normally PCR duplicates are marked and filtered by the GATK engine, but in UMI-aware calling this may not be the case, hence the need for this filter.
- **maxAltAllelesThreshold** is the maximum allowable number of alt alleles at a site. By default only biallelic variants pass the filter.
- **max_germline_posterior** is the maximum posterior probability, as determined by the above germline probability model, that a variant is a germline event.
- **normal_artifact_lod** is the maximum acceptable likelihood of an allele in the normal *by the somatic likelihoods model*. This is different from the normal likelihood that goes into the germline model, which makes a diploid assumption. Here we compute the normal likelihood as if it were a tumor in order to detect artifacts.
- **strandArtifactPosteriorProbability** is the posterior probability of a strand artifact, as determined by the model described above, required to apply the strand artifact filter. This is necessary but not sufficient – we also require the estimated max a posteriori allele fraction to be less than **strandArtifactAlleleFraction**. The second condition prevents filtering real variants that also have significant strand bias, i.e. a true variant that *also* has some artifactual reads.
- **minMedianBaseQuality** is the minimum median base quality of bases supporting a SNV.
- **minMedianMappingQuality** is the minimum median mapping quality of reads supporting an allele.
- **maxMedianFragmentLengthDifference** is the maximum difference between the median fragment lengths reads supporting alt and reference alleles. Note that fragment length is based on where paired reads are mapped, not the actual physical fragment length.
- **minMedianReadPosition** is the minimum median length of bases supporting an allele from the closest end of the read. Indels positions are measured by the end farthest from the end of the read.
- If **FilterMutectCalls** is passed a **contaminationTable** from **CalculateContamination** it will filter alleles with allele fraction less than the whole-bam contamination in the table³.

Additionally, there are two unadjustable filters. the panel of normals filter removes all alleles at a site belonging to the panel of normals, which is a vcf of blacklisted artifact sites. It can be disabled by not passing a panel of normals to **Mutect2**. There is also an STR contraction filter which removes variants that are the deletion of a single repeat unit of an STR when this repeat unit contains more than one base. This filter can be disabled with the argument **-XA TandemRepeat** which turns off the **TandemRepeat** annotation.

Here for convenience is a table of **Mutect2** filters with their corresponding annotations specified by the **-A** argument⁴, vcf keys for these annotations, and command line arguments controlling filtering thresholds.

³ This should be made more sophisticated by integrating the possibility of contamination into the germline model.

⁴ Most of these are default annotations and do not need to be invoked explicitly.

Filter	Annotation	Key	Argument
t_lod	-	TLOD	tumor_lod
clustered_events	-	ECNT	maxEventsInHaplotype
duplicate_evidence	UniqueAltReadCount	UNIQ_ALT_READ_COUNT	uniqueAltReadCount
multiallelic	-	-	maxAltAllelesThreshold
germline_risk	-	P_GERMLINE	max_germline_posterior
artifact_in_normal	-	N_ART_LOD	normal_artifact_lod
strand_artifact	StrandArtifact	SA_POST_PROB, SA_MAX_AF	strandArtifactPosteriorProbability
base_quality	BaseQuality	MBQ	minMedianBaseQuality
mapping_quality	MappingQuality	MMQ	minMedianMappingQuality
fragment_length	FragmentLength	MFRL	maxMedianFragmentLengthDifference
read_position	ReadPosition	MPOS	minMedianReadPosition
panel_of_normals	-	IN_PON	normal_panel
contamination	-	-	contaminationTable
str_contraction	TandemRepeat	RU, RPA	-