



Neuroinflammation in Alzheimer's disease

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Lancet Neurol 2015; 14: 388-405

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Increasing evidence suggests that Alzheimer's disease pathogenesis is not restricted to the neuronal compartment, but includes strong interactions with immunological mechanisms in the brain. Misfolded and aggregated proteins bind to pattern recognition receptors on microglia and astroglia, and trigger an innate immune response characterised by release of inflammatory mediators, which contribute to disease progression and severity. Genome-wide analysis suggests that several genes that increase the risk for sporadic Alzheimer's disease encode factors that regulate glial clearance of misfolded proteins and the inflammatory reaction. External factors, including systemic inflammation and obesity, are likely to interfere with immunological processes of the brain and further promote disease progression. Modulation of risk factors and targeting of these immune mechanisms could lead to future therapeutic or preventive strategies for Alzheimer's disease.

Introduction

At first glance, the specialties of immunology and neurobiology could not be more different. From a cellular perspective, the brain represents stasis, whereas the immune system represents motion. But these two perspectives have come together as efforts to understand the pathogenesis of neurodegenerative disease have borne fruit. Emerging evidence suggests inflammation has a causal role in disease pathogenesis, and understanding and control of interactions between the immune system and the nervous system might be key to the prevention or delay of most late-onset CNS diseases. In Alzheimer's disease, neuroinflammation is not a passive system activated by emerging senile plaques and neurofibrillar tangles, but instead contributes as much (or more) to pathogenesis as do plaques and tangles themselves.1 The important role of neuroinflammation is supported by findings that genes for immune receptors, including TREM22 and CD33,3,4 are associated with Alzheimer's disease. Analysis of clinical manifestations that precede the dementia stage of Alzheimer's disease, such as mild cognitive impairment, further support an early and substantial involvement of inflammation in disease pathogenesis. In this Review we provide an overview of the neuroinflammatory landscape during Alzheimer's disease, including associated cell types and mediators, methods used to visualise neuroinflammation, and its clinical presentation and potential treatments.

Cellular players

Microglia

Microglia, the resident phagocytes of the CNS, are ubiquitously distributed in the brain. Microglia constantly use highly motile processes to survey their assigned brain regions for the presence of pathogens and cellular debris, and simultaneously provide factors that support tissue maintenance (figure 1).⁵ At the same time, microglia are important players in the maintenance and

plasticity of neuronal circuits, contributing to the protection and remodelling of synapses. To some extent, this protective and remodelling action is mediated by release of trophic factors, including brain-derived neurotrophic factor, which contributes to memory formation.7 Once activated by pathological triggers, such as neuronal death or protein aggregates, microglia extend their processes to the site of injury, and migrate to the lesion, where they initiate an innate immune response (figure 2). Detection of pathological triggers is mediated by receptors that recognise danger-associated molecular patterns (DAMPs) or pathogen-associated molecular patterns (PAMPs). In Alzheimer's disease, microglia are able to bind to soluble amyloid β (A β) oligomers and A β fibrils via cell-surface receptors, including SCARA1, CD36, CD14, α6β1 integrin, CD47, and Toll-like receptors (TLR2, TLR4, TLR6, and TLR9),8-11 and this process is thought to be part of the inflammatory reaction in Alzheimer's disease. The AB peptide is derived from amyloid precursor protein (APP) by sequential cleavages by two membrane-bound proteases (figure 3).12,13 The 42-aminoacid form of Aβ has a particularly strong tendency to form soluble oligomers and fibrils. Binding of AB with CD36, TLR4, and TLR6 results in activation of microglia, which start to produce proinflammatory cytokines and chemokines (figure 4). 10,14 In turn, genetic deletion of CD36, TLR4, or TLR6 in vitro reduces Aβinduced cytokine production 10,14,15 and prevents intracellular amyloid accumulation and activation of multiprotein complexes known as inflammasomes.15

Microglial AB clearance mechanisms

In response to receptor ligation, microglia start to engulf $A\beta$ fibrils by phagocytosis. As a result, these fibrils enter the endolysosomal pathway. By contrast with fibrillar $A\beta$, which is mostly resistant to enzymatic degradation, soluble $A\beta$ can be degraded by various extracellular proteases. ¹⁶ In microglia, the proteases neprilysin and insulin-degrading enzyme (IDE) are of major importance.

In sporadic cases of Alzheimer's disease, inefficient clearance of AB has been identified as a major pathogenic pathway.¹⁷ Increased cytokine concentrations, by downregulation of expression of AB phagocytosis receptors, are suggested to be responsible for insufficient microglial phagocytic capacity.18 Further support for the hypothesis of compromised microglial function is provided by two studies^{2,3} identifying rare mutations that convey an increased risk of Alzheimer's disease. A rare mutation in the extracellular domain of TREM2 increases risk of Alzheimer's disease to a similar extent to apolipoprotein Ε (ApoE) ε4.2 TREM2 is highly expressed by microglia,19,20 and mediates phagocytic clearance of neuronal debris.21 Although a TREM2 ligand has not yet been discovered, TREM2 binding activity (putative TREM2 ligand expression) is detected on reactive astrocytes surrounding amyloid plaques and on damaged neurons and oligodendrocytes.²¹ Likewise, a single-nucleotide polymorphism (SNP) in the gene encoding the microglial surface receptor CD33 reduces AB phagocytosis by peripheral macrophages isolated from heterozygous and homozygous mutation carriers. Additionally, increased AB deposition, as shown by Pittsburgh compound B (PiB)-PET, was detected in the brains of carriers of the rs3865444 allele in the CD33 Alzheimer's disease susceptibility locus.³

Microglial diversity

Microglia activation is a complex process that results in several phenotypes. Outside the CNS, activated macrophages have been categorised as those with a classic, proinflammatory (M1) phenotype associated with expression of cytotoxic genes,22 and those with a noninflammatory, alternative activation (M2) phenotype, associated with induction of specific proteins, including ARG1, FIZZ1, YM1, and IGF1.23,24 Classic M1 activation is characterised by increased concentrations of proinflammatory cytokines, including TNFa, interleukin 1, interleukin 6, interleukin 12, and interleukin 18, and is accompanied by impaired phagocytic capacity.25 The M2 state is characterised by secretion of the antiinflammatory cytokines interleukin 4, interleukin 10, interleukin 13, and TGF-B, and increased phagocytic capacity without production of toxic nitric oxide.26-28 A third phenotype might be a deactivated one associated with corticosteroids or TGF-β.^{29,30} The M1 and M2 activation states represent the extremes of myeloid cell activation. Peripheral monocyte-derived macrophages exist in a diverse range of phenotypic states, particularly under conditions of chronic inflammation.31 Microglia are also likely to exist in a range of phenotypic states during chronic inflammation: these cells have a wide range of phenotypes that are indicative of their response to the local environment, including physical interaction with other cells and their physiological activity in the brain. Importantly, the ability to isolate or image subsets of unperturbed microglia to characterise their gene expression and mode of action as discriminated by physiological markers is restricted at present.

Microglia priming

In the ageing CNS of mice, rats, and primates, microglia show enhanced sensitivity to inflammatory stimuli,32 similar to that noted in microglia in brains with ongoing neurodegeneration. This phenomenon is termed priming. Priming might be caused by microglial senescence and might be associated with ageing. On the transcriptomic level, endogenous ligands downregulated during ageing, whereas factors for host defence and neuroprotection are upregulated. 20 To what extent age-related microglia priming results from cellautonomous cellular ageing, rather than prolonged exposure to the aged neural environment, is uncertain. In physiologically aged and senescence-accelerated mice, profound microglia priming was characterised by increased production of cytokines and reactive oxygen species, and enhanced phagocytic capacity. This model provided proof of principle that environmental effects, such as neuronal ageing, can drive microglia priming.

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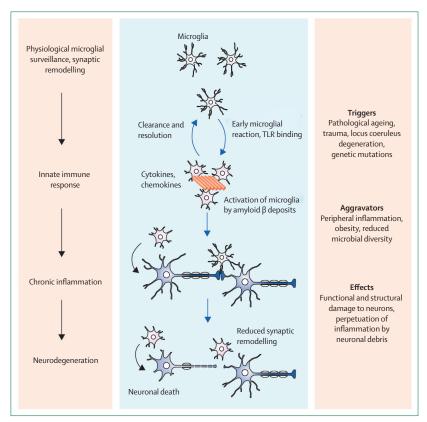


Figure 1: Pathomechanistic sequelae of microglia activation

Physiological functions of microglia, including tissue surveillance and synaptic remodelling, are compromised when microglia sense pathological amyloid β accumulations. Initially, the acute inflammatory response is thought to aid clearance and restore tissue homoeostasis. Triggers and aggravators promote sustained exposure and immune activation, which ultimately leads to chronic neuroinflammation. Perpetuation of microglia activation, persistent exposure to proinflammatory cytokines, and microglial process retraction cause functional and structural changes that result in neuronal degeneration. TLR=Toll-like receptor.

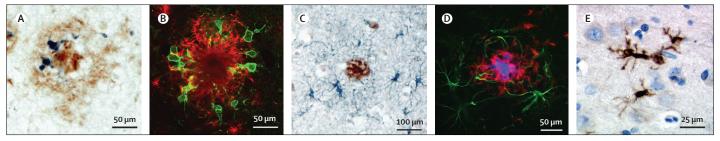


Figure 2: Changes in microglia and astroglia in Alzheimer's disease
Microglia and astroglia are key players in the inflammatory response: changes in microglia and astroglia are evident in the post-mortem brains of patients with Alzheimer's disease and in animal models of the disorder. (A) CD11b-positive microglia (blue) within an amyloid β (Aβ) deposit (brown) in the parietal cortex of a brain section from a patient with Alzheimer's disease. (B) Activated, IBA1-positive microglia (green) at an Aβ plaque site (red) in a brain section from an APP/PS1 transgenic mouse. (C) GFAP-positive astrocytes (blue) surround the site of Aβ deposition (brown) in the parietal cortex of a brain section from a patient with Alzheimer's disease. (D) GFAP-positive astrocytes (green) at an Aβ plaque site (red) in a brain section from an APP/PS1 transgenic mouse.

(E) Interleukin-1β-positive microglia (brown) in the frontal cortex of a brain section from a patient with Alzheimer's disease.

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Weighted gene correlation network analysis revealed a characteristic pattern of gene expression for microglia priming, featuring increased pattern recognition and expression of interferon signalling genes. A similar gene expression network was reported in mouse models of age-related neurodegeneration, including APP/PS1 transgenic mice.³³ Microglia might also be primed by systemic inflammation in response to peripheral immune reaction.

Modulation of microglia

The emerging role of microglia activation in Alzheimer's disease pathogenesis makes these cells a legitimate therapeutic target. However, depending on the circumstances, microglia activation can have both beneficial and detrimental effects. Thus, microglia might have different roles and effects depending on the particular disease stage and which brain region is affected in each model. After exposure to a DAMP or PAMP, the acute microglial reaction aims to remove the recognised abnormality or pathological change. In the case of Alzheimer's disease, this type of inflammatory reaction is sterile because it involves the same receptors but no living pathogens. Under normal circumstances, such a reaction quickly resolves pathological changes with immediate benefit to the nearby environment. However, in Alzheimer's disease, several mechanisms, including ongoing formation of AB and positivefeedback loops between inflammation and APP processing, compromise cessation of inflammation. Instead, further accumulation of AB, neuronal debris, and, most probably, further activating factors establish chronic, non-resolving inflammation. Sustained exposure to Aβ, chemokines, cytokines, and other inflammatory mediators seems to be responsible for the persistent functional impairment of microglial cells seen at plaque sites. 40,41 As an intracellular regulator of microglial function, expression of the autophagy protein Beclin 1 is reduced in the brains of patients with Alzheimer's disease. 42 Beclin 1 has a role in retromermediated sorting of cellular components, including

TREM2, APP, BACE1, and CD36, in the endolysosomal pathway. Reduction of Beclin 1 expression in vitro and in vivo interferes with efficient phagocytosis, resulting in decreased receptor recycling of CD36 and TREM2,⁴² but more receptors might be affected.

Plasticity of the microglial phenotype is of fundamental importance, since resolution of inflammation clearly involves conversion to an alternative (ie, similar to M2) activation state associated with tissue repair, phagocytosis, and anti-inflammatory actions. Conversion of microglia from detrimental to beneficial players might be achieved by modulation of proinflammatory signalling pathways such as the NLRP3 inflammasome. Successful modification of these pathways, however, necessitates that they are exclusively restricted to microglia and do not have crucial functions in other cell types. Pharmacologically, transition to an alternative activation state could be achieved through the heterodimeric type II nuclear receptors PPARγ/RXR, PPARδ/RXR, and LXR/RXR. Agonists of these receptors are robustly antiinflammatory and stimulate phagocytosis through induction of CD36, leading to increased microglial AB uptake.43 Another target is the RXR itself, which might have a positive effect on both LXR-controlled and PPARycontrolled genes. Agonism of RXR by bexarotene has been shown to cause rapid reduction of soluble AB, plaque load, and behavioural deficits by ApoE-dependent clearance of Aβ. 44 Nevertheless, results of this study could not be wholly reproduced by others. 45-48 Although aberrant and ineffective activation of microglia has been fairly well documented for prodromal Alzheimer's disease and moderate Alzheimer's disease, late-stage effects are less well understood. Some evidence exists of focal microglial senescence, especially surrounding neurofibrillary tangles.40

Blood-derived mononuclear cells

The precise contribution of blood-derived mononuclear cells infiltrating the CNS in Alzheimer's disease, such as innate immune responses of the brain, is so far unclear, and knowledge is restricted to animal studies. Results of

these animal studies have shown infiltration of peripheral mononuclear cells associated with amyloid plaques in mouse models.³⁴ Further, ablation of CD11bpositive cells in the APP/PS1 mouse model of peripheral Alzheimer's disease showed that mononuclear phagocytes have an important role to reduce the build-up of Aβ plaques.³⁴ Restriction of entry of blood-derived mononuclear cells into the brain, by deletion of the chemokine receptor CCR2 in the Tg2576 mouse model, led to increased plaque load,35 although the mononuclear cell type was not specified. However, most of these studies used bone marrow irradiation and subsequent transplantation with fluorescent, and therefore traceable, cells. Irradiation of whole animals is likely to cause damage to the blood-brain barrier. A further study in which the brain was shielded, thereby limiting irradiation to the rest of the body, did not report any cerebral infiltration by peripheral macrophages, but concluded that perivascular macrophages, protected by shielding of the brain, were able to modulate AB deposition depending on the presence of CCR2.36 Involvement of perivascular macrophages has also been shown for removal of $A\beta$ in a mouse model of cerebral amyloid angiopathy.³⁷ Nevertheless, recruitment of bonemarrow-derived cells is almost absent in parabiosis mouse models, even 12 months after initiation.³⁸ Notably, in this context, ablation of microglia in APP/PS1 mice by the HSV thymidine kinase/ganciclovir system did not change the amyloid pathology, although 95% of microglia were lost and blood-derived monocytes were spared by use of bone-marrow-chimeric mice.39 This result suggests that peripheral cells do not participate in phagocytosis of amyloid plaques, although the observation time was only 2-4 weeks. These results provide evidence against a substantial contribution of blood-derived monocytes, but support the idea that perivascular macrophages have some effect on removal of CNS Aβ depositions.

Astroglia

Pathological responses of astrocytes include reactive astrogliosis, a complex, multistage and pathologyspecific reaction, whereas remodelling of astrocytes is generally aimed at neuroprotection and recovery of injured neural tissue. 49,50 Next to activated microglia, hypertrophic reactive astrocytes accumulate around senile plaques and are often seen in post-mortem human tissue from patients with Alzheimer's disease,51 and in animal models of the disorder.⁵² Glial cell activation might occur early in Alzheimer's disease, even before Aβ deposition.⁵³ Reactive astrocytes are characterised by increased expression of glial fibrillary acidic protein (GFAP) and signs of functional impairment;54 however, astrocytes do not seem to lose their domain organisation, and no evidence of scar formation exists (figure 2). In animal models of Alzheimer's disease, the early response is marked by astroglial atrophy, which might have far-reaching effects on synaptic connectivity, because astrocytes are central to the maintenance of synaptic transmission, thereby contributing to cognitive deficits.^{52,54–57} These signs of

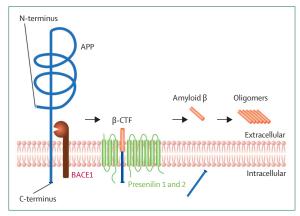


Figure 3: Amyloidogenic processing of amyloid precursor protein

Amyloid precursor protein (APP) is a type 1 transmembrane protein that is sequentially cleaved by two aspartate proteases. β -site APP cleaving enzyme 1 (the β -secretase BACE1) cleaves the protein to yield a C-terminal fragment (β -CTF) and secreted soluble peptide APP β . β -CTF is then processed by presenilin 1 and 2 (part of the γ -secretase complex) to release the amyloid β peptide. The process results in differentially truncated C-termini, ranging from aminoacid 37 to 42. The 42-aminoacid form ($A\beta_{1-\alpha}$) has a particularly strong tendency to form soluble oligomers and fibrils. These $A\beta$ aggregates bind to cell-surface receptors on microglia, inducing an inflammatory activation that results in the secretion of proinflammatory cytokines, including TNF α and interleukin 1 β . In this context, it has been shown that interleukin 1 β aggragvates plaque formation by modulation of APP expression. Additionally, expression of BACE1 is upregulated by some cytokines, resulting in increased production of $A\beta$ species.

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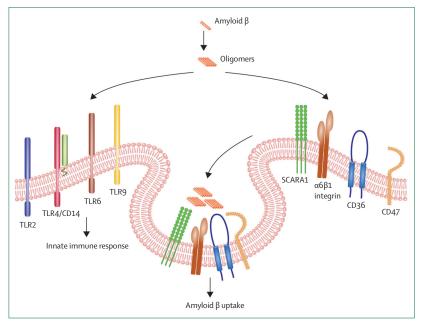


Figure 4: Activation of microglia by amyloid β Amyloid β (A β) aggregates (oligomers) act on several Toll-like receptors on the microglial surface, triggering reactions of the innate immune system, including production of proinflammatory cytokines and chemokines. A β oligomers are internalised by microglia, aided by SCARA1, α 6 β 1 integrins, CD36, and CD47.

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atrophy show clear spatiotemporal progression, appearing first in the entorhinal cortex, and affecting astrocytes located at a distance from senile plaques in the later stages of Alzheimer's disease. Like microglia, astrocytes release cytokines, interleukins, nitric oxide, and other potentially cytotoxic molecules after exposure to Aβ, thereby exacerbating the neuroinflammatory response. The importance of astroglial inflammation in Alzheimer's disease has been investigated by adenoassociated virus-driven suppression of the astrocytic reaction in APP/PS1 mice. Interference with the calcineurin/NFAT signalling pathway revealed improved cognition, reduced astrogliosis, and decreased AB concentrations.58 Additionally, astrocytes have a potential role in internalisation and degradation of Aβ in vivo.59 ApoE is needed for astrocyte-mediated clearance of Aβ,60 and astrocyte-dependent lipidation of ApoE increases the capability of microglia to clear Aβ. 61,62 Furthermore, adult astrocytes upregulate expression of extracellular Aβ-degrading proteases, 63 such as neprilysin, IDE, ECE2, and ACE, after exposure to native Aβ deposits. 64 These proteases, and impaired function and atrophy of astrocytes, might contribute to reduced proteolytic clearance of A\u03c3. In addition to these clearing pathways, astrocytes have a role in clearance of soluble Aß from the parenchyma by paravenous drainage. 65 This pathway depends on the astrocytic water channel aquaporin 4; deletion of this channel resulted in a substantial decrease in clearance via this pathway.

Mediators and modulators of neuroinflammation Cytokines

Microglia and astrocytes are arguably the major source of cytokines in Alzheimer's disease. Cytokines contribute to nearly every aspect of neuroinflammation, including proinflammatory and anti-inflammatory processes, bystander neuronal injury, chemoattraction, and response of microglia to AB deposits. Microglia activation is both characterised by and modulated by cytokines. Increases in Aβ concentration in ageing TgAPPsw and PSAPP transgenic mice are associated with increased concentrations of proinflammatory cytokines, including TNFα, interleukin 6, interleukin 1α, and GM-CSF.66 This observation suggests that pathological accumulation of AB is a key factor that drives neuroinflammatory responses in Alzheimer's disease. Additionally, exposure of microglia to preaggregated Aβ₁₋₄₂ increases production of proinflammatory cytokines (ie, pro-interleukin interleukin 6, and TNFα), MIP-1α, and M-CSF.67 Furthermore, M-CSF concentrations in the plasma and CNS of patients at the dementia stage of Alzheimer's disease are substantially increased compared with agematched healthy controls or patients with mild cognitive impairment. 67,68 Caspase 1 activation, which is needed for cleavage of interleukin 1β from its inactive proforms,69 is similarly elevated in the brains of patients with mild cognitive impairment and Alzheimer's disease dementia.70 As a result, high concentrations of the cardinal proinflammatory cytokine interleukin 1B are detected in microglial cells surrounding Aß plaques in brains of patients with Alzheimer's disease (figure 2) and in CSF of patients. In vitro, interleukin 1ß is released by activated microglia after stimulation with $A\beta$.⁷¹ Interleukin 1β can, at least under some circumstances, favour AB deposition by modulation of expression and proteolysis.72 Additionally, interleukin 12 and interleukin 23, which are known to be produced by leucocytes, are produced by microglia in mouse models of Alzheimer's disease,73 and inhibition of these cytokines reduces Alzheimer's disease-like pathology,73,74 although regulation of interleukin 12 in human CSF is debated.73,75

Evidence suggests that the proinflammatory environment present in the brains of patients with Alzheimer's disease and in transgenic mouse models of cerebral amyloidosis reaches damaging proportions. For example, risk for conversion from mild cognitive impairment to the dementia stage of Alzheimer's disease is increased in patients with elevated concentrations of the proinflammatory cytokine TNFa and decreased concentrations of anti-inflammatory TGF-β in the CSF.76 Interleukin 1β, TNFα, and other cytokines might impair neuronal function even before leading to structural changes, as shown by suppression of long-term potentiation (LTP) of synaptic transmission. Several interactions, and increased expression of additional cytokines, chemokines, and innate immune receptors, favour an M1-like activation state in Alzheimer's disease. For example, in neuron-microglia co-cultures, the synergistic action of A β with either interferon γ or CD40 ligand triggers TNFa secretion and production of neurotoxic reactive oxygen species.77-79 Additionally, the innate immune receptor TLR4 is responsible for increased concentrations of TNF α and MIP-1 α in mouse models of Alzheimer's disease.80

Conversely, stimulation of some proinflammatory signalling pathways seems to be beneficial in mouse models of Alzheimer's disease. Transgenic expression of interleukin 18 in APP/PS1 mice led to robust neuroinflammation and reduction of amyloid plaque pathology.^{81,82} These findings implicate interleukin 1B expression in activation of a beneficial form of neuroinflammation in APP/PS1 mice. In another study, AAV-mediated expression of interferon y in the brains of the TgCRND8 mouse model showed the ability of this proinflammatory cytokine to enhance clearance of amyloid plaques, with a widespread increase in astrogliosis and microgliosis.83 Additionally, these mice had decreased concentrations of soluble $A\beta$ and $A\beta$ plaque burden, without altered APP processing. Similar results were obtained using AAV-mediated expression of interleukin 6 and TNFa.84,85 Conversely, expression of the anti-inflammatory cytokine interleukin 4 resulted in exacerbation of A β deposition.⁸⁶ These results suggest that some beneficial forms of proinflammatory microglia activation potentially help to reduce Alzheimer's disease-like pathology in transgenic mouse models.

Chemokines

Chemokines have been suggested to regulate microglial migration to areas of neuroinflammation, thereby enhancing local inflammation in Alzheimer's disease.87 In Alzheimer's disease, upregulation of CCL2, CCR3, and CCR5 in reactive microglia has been reported, 88,89 whereas CCL4 has been detected in reactive astrocytes near AB plaques.88 In vitro, AB leads to generation of CXCL8 (also known as interleukin 8), CCL2, CCL3, and CCL4 in human macrophages and astrocytes,90 and microglia cultured from autopsies of patients with Alzheimer's disease revealed increased expression of CXCL8, CCL2, and CCL3 after experimental exposure to Aβ.91 In mouse models of Alzheimer's disease, modulation of neuronal survival,92 plaque load,93 and cognition94 by the CX3CR1/CX3CL1 system has been shown. Furthermore, the receptors CCR595 and CCR235,96,97 can modulate the course of disease through effects on microglial position and function.

Caspases

Caspases are a family of intracellular proteases that are key mediators of apoptosis and inflammation. Of the inflammatory caspases, the catalytic activity of caspase 1 is tightly regulated by signal-dependent autoactivation within inflammasomes, which mediate caspase 1 autocatalytic activation and subsequent cleavage of precursors of interleukin 18 and interleukin 18 into bioactive cytokines.98,99 Aβ fibrils can activate NRLP3 inflammasomes via lysosomal damage in mouse microglia.100 Increased concentrations of active caspase 1 are detected in the brains of patients with Alzheimer's disease and APP/PS1 mice. Additionally, APP/PS1 mice deficient in NLRP3 or caspase 1 are mostly protected from spatial memory impairment, loss of hippocampal synaptic plasticity, associated behavioural disturbances, and other effects associated with Alzheimer's disease. 70 Deficiency of NLRP3 or caspase 1 in APP/PS1 mice seemed to shift microglial cells from a proinflammatory M1-like phenotype to a more neuroprotective M2-like phenotype. 70 Further, stimulation of microglia with various proinflammatory mediators led to orderly activation of apoptotic caspase 8 and caspase 3/7. Activated caspase 3 modulates NF-κB activation via PKCδ and increases production of neurotoxic proinflammatory mediators, such as interleukin 1β , TNF α , and nitric oxide. Inhibition of these caspases hindered microglia activation and neurotoxicity.¹⁰¹ Incidentally, these caspases were activated in microglia in patients with Alzheimer's disease. 102 Pharmacological interventions with inhibitors of activated caspases have been reported to successfully exert neuroprotective effects in mouse models of Alzheimer's disease.103,104

Prostanoids and neuroprotectin D1

Prostanoids are derivatives of arachidonic acid synthesised by cyclooxygenase 1 and inducible cyclooxygenase 2, both of which are produced by microglia. In Alzheimer's disease, the suppressive effect of cyclooxygenase 1 inhibition on glia activation, amyloid deposition, and expression of inflammatory markersswitching microglia to an alternative phenotype—has been shown in a mouse model of Alzheimer's disease. 105 Additionally, concentrations of the proinflammatory prostaglandin PGE2, which binds to PTGER1-4 receptors, has proved to be elevated in patients with probable Alzheimer's disease. 106 PTGER1-3 receptors are expressed by microglia, 107 but are also expressed in other cells of the brain, particularly neurons. Microglial PTGER2 receptors inhibit Aβ phagocytosis and enhance neurotoxic activities of microglia in vitro. $^{\tiny 108}$ This effect is complemented by findings that deletion of PTGER2 or PTGER3 receptors in mouse models of Alzheimer's disease decreased oxidative stress, neuroinflammation, Aβ burden, and BACE1 expression. 109-111

Use of PTGER4 receptor agonist on microglia showed suppression of inflammation and increased uptake of synthetic A β , whereas deletion of PTGER4 receptor in the APP/PS1 mouse model of Alzheimer's disease increased plaque burden and production of proinflammatory cytokines such as interleukin 1 β and CCL3.¹¹² Notably, expression of PTGER4 receptor was decreased in the cortex of patients with mild cognitive impairment and Alzheimer's disease, ¹¹² suggesting that it might contribute to the inflammatory reaction in Alzheimer's disease. However, the role of PGE2 in neurodegeneration is probably complex owing to effects of PGE2 on other cell types such as neurons.

The neuroprotective docosahexaenoic acid derivative neuroprotectin D1 (also known as 10R,17S-DHA) is a major component of cell membranes, $^{\rm 113}$ and expression is decreased in early stages of Alzheimer's disease. $^{\rm 114}$ Neuroprotectin D1 is an autocrine/paracrine mediator of the resolution response during early stages of neuroinflammation, and downregulates amyloidogenic processing of APP, switches off proinflammatory gene expression, and promotes neural cell survival. Moreover, anti-amyloidogenic processing by neuroprotectin D1 targets α secretases and β secretases and PPAR γ receptor activation. sAPP α , a peptide with neurotrophin-like activity, is an agonist for neuroprotectin D1 synthesis and is part of a cycle that sustains generation of the lipid mediator. $^{\rm 115}$

Complement system

The complement system is a major constituent of the innate immune system, mainly involved in defence against pathogens. Activation of the proteolytic complement cascade results in opsonisation and, ultimately, in lysis of microorganisms. In the brain, the major cells that contribute to production of proteins of

the complement system are microglia and, to a lesser extent, astrocytes. ¹¹⁶ In Alzheimer's disease, activated factors of the complement system are associated with A β deposits. ¹¹⁷ Additionally, A β is able to activate the complement system in vitro via the so-called alternative pathway. The finding that variants of clusterin (apolipoprotein J), as a soluble inhibitor of the complement system, and the complement receptor CR1, involved in processing and clearance of opsonised immune complexes and a regulator of C3 convertase activity, are associated with Alzheimer's disease provides further evidence for the importance of the complement system in disease pathogenesis. ^{118,119}

Nitric oxide and reactive oxygen species

In addition to their direct actions via surface receptors, cytokines stimulate inducible nitric oxide synthase (iNOS) in microglia and astroglia, producing high concentrations of nitric oxide that can be toxic to neurons. iNOS is upregulated in brains of patients with Alzheimer's disease,120 and genetic knockout of iNOS is protective in mouse models of Alzheimer's disease.121 Likewise, NADPH oxidase (PHOX) is highly expressed by microglia, upregulated in Alzheimer's disease, and rapidly activated by inflammatory stimuli such as AB, resulting in production of hydrogen peroxide, which further promotes microglia activation. 222,123 Superoxide from PHOX reacts with iNOS-derived nitric oxide to form peroxynitrite.¹²⁴ Increased expression of iNOS in patients with Alzheimer's disease introduces posttranslational modifications caused by nitric oxide,125 which include nitration, S-nitrosylation, and dityrosine formation.¹²⁵ Nitration of the Aβ peptide at tyrosine 10 has been shown to increase the propensity of $A\beta$ to aggregate and has been identified in the core of amyloid plaques.¹²⁶ More compelling, this modified peptide was able to initiate plaque formation in APP/PS1 mice. suggesting that it has a central role during the early phase of Alzheimer's disease. Nitrated Aß suppressed hippocampal LTP more effectively than did non-nitrated Aβ, suggesting that this post-translational modification leads to functional and structural damage in the brains of patients with Alzheimer's disease. Evidence suggests that oxidative stress supports formation of this AB species. 127 Other nitric oxide-mediated modifications that might be relevant for Alzheimer's disease have already been reported, 128 and more are expected to follow.

Inflammatory changes of the neurovascular unit

Results of many epidemiological, clinical, and neuropathological studies have shown that vascular pathological change is an important risk factor for development of Alzheimer's disease. Moreover, Alzheimer's disease is associated with distinct inflammatory, functional, and morphological alterations of cerebral blood vessels and perivascular glia and neurons (the neurovascular unit). These early-onset and

progressive changes, which are induced by combined effects of soluble AB oligomers and vascular AB deposits, 129,130 ultimately lead to decreased cerebral blood flow and impaired functional hyperaemia (ie, the ability of local blood flow to increase in response to neuronal activation).¹³¹ Chronic cerebral hypoxia is further amplified by blood-borne factors such as platelets, which are chronically activated in models of, and patients with, disease,132 Alzheimer's ultimately resulting microinfarcts and neuronal injury. Moreover, the combination of mild hypoxia, inflammation of the neurovascular unit, and progressive AB accumulation in brain parenchyma, induces upregulation of AGER (also known as RAGE), which mediates Aβ transport into the brain across the blood-brain barrier.¹³³ Additionally, hypoxia directly induces amyloidogenic APP processing through several pathways involving β secretase, γ secretase, neprilysin, and others.134 Taken together, chronic hypoxia in Alzheimer's disease directly induces neuronal injury, but also amplifies neurodegeneration by induction of amyloidogenic pathways and reduction of brain clearance of Aβ.

Factors that drive neuroinflammation

A β deposition alone might be sufficient to induce an inflammatory reaction that subsequently contributes to cognitive decline and development of Alzheimer's disease. In view of the possibility that A β deposition precedes cognitive deficits or clinical manifestation by decades, one might speculate that exogenous or endogenous factors can modify the innate immune response mounted by A β -exposed microglia. Thus, environmentally modifiable Alzheimer's disease risk factors, including systemic inflammation, obesity, and traumatic brain injury, might affect risk through sustained neuroinflammatory drive.

Systemic inflammation

Development of sickness behaviour¹³⁵ after a peripheral inflammatory challenge, such as an infection or an aseptic injury,136 shows the communication between systemic inflammation and the brain. Sickness behaviour in response to an acute event is usually self-limited as a result of the presence of several regulatory mechanisms that dampen the central inflammatory response to peripheral challenge.¹³⁷ However, the inflammatory response to chronic, low-grade inflammation might be prolonged,138 possibly because no anti-inflammatory response occurs.¹³⁹ Neuroinflammation in Alzheimer's disease is such a chronic reaction, because microglia might already be primed and are therefore highly responsive to further activation, causing a rapid switch to a damaging M1 phenotype.^{23,140} This microglia priming is likely to result from various activators, such as chronic exposure to AB, neuronal debris, and chronic vascular changes, including cerebrovascular dysregulation and cerebral microinfarcts. This hypothesis is supported by results of animal studies showing an exaggerated inflammatory and oxidative stress response to peripheral stimuli in aged mice, 141 increased concentrations of interleukin 1 β in the CNS, and neuronal apoptosis in the ME7 prion mouse after peripheral challenge with lipopolysaccharide or polyinosinic-polycytidylic acid. $^{142-144}$ Other examples are the effects of osteoarthritis in APP/PS1/Col1-IL1 β XAT mice, resulting in accelerated neuroinflammation and A β pathology. 145 Additionally, results of clinical studies of Alzheimer's disease show increased cognitive decline and exacerbation of sickness behaviour after acute and chronic systemic inflammation. 146,147

Obesity

Obesity is defined as a medical disorder in which excess body fat has accumulated to the extent that it might have an adverse effect on health. Obesity increases a patient's propensity to acquire bacterial or viral infections, and thus directly increases the likelihood of systemic inflammation. 148,149 Moreover, white fat tissue has a high percentage of activated macrophages, which constantly secrete proinflammatory cytokines.¹⁵⁰ Notably, midlife obesity has been identified as a risk factor for Alzheimer's disease,151 which is related to the fact that other Alzheimer's disease risk factors, such as high-cholesterol diet, reduced physical activity, and sedentary lifestyle, are associated with obesity. 152 As a possible result of obesity, type 2 diabetes accelerates memory dysfunction and neuroinflammation in a mouse model of Alzheimer's disease.153 Obesity-associated reduced gut microbial diversity was associated with increased concentrations of proinflammatory markers in peripheral blood, and thus could be viewed as a factor contributing to risk for Alzheimer's disease.¹⁵⁴ Taken together, the evidence suggests that obesity increases the risk for Alzheimer's disease by the systemic and chronic presence of proinflammatory cytokines.

Traumatic brain injury

Several studies have established traumatic brain injury as a risk factor for development of Alzheimer's disease. 155 Experimentally, traumatic brain injury aggravates learning and memory deficits and deposition of AB in mouse models of Alzheimer's disease. 156,157 Results of animal and human studies have shown that microglia activation can persist for months or years after traumatic brain injury. 158,159 This inflammatory reaction might initially be important for phagocytic clearance of debris. However, sustained cerebral inflammation might either directly or indirectly promote development of Alzheimer's disease. Some cytokines implicated in traumatic brain injury can potentially increase BACE1 concentrations,160 thereby shifting APP processing to amyloidogenic generation of Aβ (figure 3). 161 Additionally, chronic release of cytokines might decrease the capability of microglia to phagocytose and degrade AB, or might directly affect neuronal functions.

Locus coeruleus degeneration

Noradrenaline, in addition to its role as a neurotransmitter, has potent anti-inflammatory, anti-oxidative, neurotrophic, and neuroprotective actions.¹⁶² The main source of noradrenaline in the brain is the locus coeruleus (LC), located at the dorsal part of the brain stem. The LC neurons project throughout the brain, although most terminals target the hippocampus and neocortex. Noradrenaline released from LC projections acts on adrenergic receptors expressed on neurons, microglia, and astrocytes. 162 The number of cells in the LC, and concentration of noradrenaline in the brain, decrease during normal ageing, 163 although more pronounced cell loss occurs in patients with Alzheimer's disease.¹⁶⁴ Experimental lesions of the LC in mouse models of Alzheimer's disease led to increased inflammation and neuronal damage, and an increase in A β plaque burden. 165,166 Thus, early degeneration of the LC and subsequent loss of noradrenaline-mediated innervation could substantially promote the inflammatory response to any stimulus, including Aβ. Experimental loss of noradrenaline compromised microglial migration and Aß phagocytosis in vivo, suggesting that a loss of noradrenaline tone increases not only inflammation, but also AB deposition. Selective noradrenaline reuptake inhibitors, α_2 -adrenoceptor antagonists, α_3 -adrenoceptor antagonists, and the noradrenaline precursor L-threo-3,4-dihydroxyphenylserine, 168 which increase endogenous noradrenaline concentrations, can reduce neuroinflammation and partly rescue microglial functions.

Analysis of immune activation

In-vivo laser-scanning microscopy

During the past few decades, analysis of innate immunity of the brain was restricted to cell culture experiments and immunohistochemical detection of microglia. Little was known about the functional state of these cells in vivo. Methodological advances such as generation of transgenic mice with enhanced-GFP-labelled microglia, cranial window implantation, and AB plaque labelling with the fluorescent dye methoxy-XO4 have enabled longitudinal and live monitoring of the functional state of microglia in mouse models of disease. This technique enables analysis of microglial phenotypes over time and in relation to deposition of AB. Studies have shown that, in control and plaque-bearing mice, microglia migrate within the brain parenchyma with an average speed of 5–9 mm/month. 169 After formation of Aβ plaques, microglia rearrange their processes, becoming polarised, and move their somata towards the plaque, temporarily leaving the formerly surveyed area.¹⁶⁹ Individual microglia migrate towards new or pre-existing plaques within 1-2 days, 169,170 similar to observations made after acute laser-induced brain injury. 171 In addition to A β deposits, microglia are attracted to neurons that have undergone Alzheimer's disease-associated elimination.92 Of note, microglial cells were recruited to neurons up to 7 days before their elimination, and neuronal loss could be

completely rescued by knockout of the receptor for chemokine fractalkine (CX3CR1).²²

Imaging of inflammation in animals

Various molecular imaging techniques are used in laboratory and clinical settings to study the temporal and spatial relation of inflammatory changes associated with neurodegeneration. Because microglia respond quickly to lesions, these cells are good candidates for diagnostic markers of disease progression.¹⁷² Therefore, several microglial cell-surface and mitochondrial receptors were used for development of in-vivo imaging ligands. One of these targets is the translocator protein TSPO,173 a protein of the outer mitochondrial membrane that is increasingly expressed under conditions of neuroinflammation. Binding of the radiolabelled ligands 11C-(R)-PK11195 and 18F-DPA-714 to TSPO can be visualised using PET and SPECT.174 In mouse models of Alzheimer's disease, progressive binding of PK11195 with age has been described. 175,176 This binding could be reliably detected by immunohistological means in APP/ PS1 mice only at a late stage, when microglia and astroglia activation is already high.177 Binding of both ³H-(R)-PK11195 and ³H-DPA-713 could be decreased by the PPARy agonists pioglitazone and ciglitazone in the TASTPM mouse model,176 and have anti-inflammatory and Aβ-lowering effects in mouse models of Alzheimer's disease.178 Development of new TSPO ligands and other probes with improved bioavailability, decreased nonspecific uptake, and increased specific binding 179,180 might enable detection of activated microglia at an even earlier disease stage.

Imaging of inflammation in human beings

Similar to results of studies in animals, increased microglia activation has been detected in patients with Alzheimer's disease by use of the TSPO ligand ¹¹C-PK11195. Binding potentials are reported to be increased up to 50% in association cortex.¹⁸¹ Uptake of another TSPO ligand, 11C-DAA1106, was reported to be increased by up to 33% in patients with Alzheimer's disease.182 Cortical distribution of 11C-PK11195 binding parallels that of amyloid deposition, as detected by the thioflavin analogue ¹¹C-PIB. ¹⁸³ Concentrations of cortical ¹¹C-PK11195 signals in patients with Alzheimer's disease are likewise associated with cognitive impairment rated with the Mini-Mental State Examination, 184,185 suggesting that cortical microglia activation is detrimental to cognitive function. Additionally, a relation of neuroinflammation and the severity of Alzheimer's disease has been reported for the TSPO marker 11C-PBR28.186 In patients with mild cognitive impairment, 11C-PK11195 PET detected inflammation in 40% of amnestic cases, 187 although another study with the same tracer failed to detect microglia activation. Seven patients with mild cognitive impairment showed a significant 27% mean increase in ¹¹C-DAA1106 uptake in the lateral temporal cortex,

compared with controls (p=0.008). Five of these seven patients with mild cognitive impairment, with $^{11}\text{C-DAA1106}$ uptake more than 0.5 SDs above the mean in controls, progressed to dementia during a 2-year follow-up period.

Characterisation and monitoring of neuroinflammation in Alzheimer's disease

Although emerging evidence suggests that inflammation has a causal role in Alzheimer's disease pathogenesis, detection of inflammatory markers has not yet been established as a valuable method for diagnosis or monitoring of Alzheimer's disease. Nevertheless, novel data from a gene-expression analysis of post-mortem brains from patients with late-onset Alzheimer's disease highlighted an immune and microglia network dominated by genes implicated in phagocytosis. These data, together with analysis of inflammation-related biomarkers in the CSF, peripheral blood, or directly in the brain by imaging, will be the focus of future studies. An important aspect of this search will be discovery of inflammatory biomarkers that can be used to identify prodromal stages of Alzheimer's disease (panel).

Detection of neuroinflammatory markers in CSF

During the past 25 years, several studies have investigated concentrations of proinflammatory and anti-inflammatory cytokines in the CSF of patients with mild cognitive impairment and Alzheimer's disease. Results of these studies are often debated;189 however, the timepoint of sampling—that is, the stage of the disease—seems to be a crucial factor for investigation. Some studies highlight increased concentrations of cytokines in the CSF as risk factors for conversion of mild cognitive impairment to the dementia stage of Alzheimer's disease or as markers of the speed of cognitive decline and disease progression. 76,190 As with imaging consortia, to overcome inter-individual differences and to obtain a definite description of cytokine regulation and function in Alzheimer's disease, a high degree of method harmonisation and patient characterisation, together with longitudinal sampling over years, seems to be essential to progress beyond crosssectional descriptions.

Systemic biomarkers

Results of a growing number of studies suggest that a sophisticated interaction occurs between the systemic environment and the brain. Thus, systemic immune cells and secreted signalling proteins communicate with the brain, and have been associated not only with neuroinflammation, but also with neurodegenerative processes in general.¹⁹¹ Although some of these interactions might involve cells entering the nervous tissue, many more are likely to be mediated by soluble signalling molecules, such as blood-borne factors, present in the systemic environment. These factors can inhibit or promote adult neurogenesis in an age-

dependent manner or restore regeneration of the ageing brain in mice. 192,193 To identify such factors, scientists have tried to discover molecular or cellular changes in blood associated with neurodegenerative diseases. 194 Various proteomic methods have been used to identify blood-based biomarkers. Cytokines and trophic factors such as BDNF are typical biomarkers. 195-198 Using multiplex ELISA in plasma from controls and patients with mild dementia, mild cognitive impairment or Alzheimer's disease, protein signatures were described that might be specific to prodromal stages of the disease, 199 or that characterise patients who progress from a prodromal stage to the dementia stage of Alzheimer's disease.200 Other signatures seem to correlate with ApoE²⁰¹ or with pathological changes such as AB and tau protein concentrations in CSF of patients with Alzheimer's disease. 202 About 200 communicome proteins were measured in plasma samples from patients participating in the Alzheimer's disease neuroimaging initiative, yielding protein signatures associated with patients who converted from mild cognitive impairment to the dementia stage of Alzheimer's disease.203

Clinical trials and epidemiological findings

Anti-inflammatory drugs

Non-steroidal anti-inflammatory drug (NSAID) epidemiology and clinical trial results (table)204-219 have produced some healthy scepticism about apparent stagedependent outcomes, but the disappointing results of these studies are perhaps not surprising when one considers that normal physiological cytokine regulation of glia activation and microglial phenotypes is highly context-dependent and stage-dependent.5 Contextual factors modulating glial-activated phenotypes include immune-modulatory APOE genotype and newly identified Alzheimer's disease genes. Normal ageing is likewise associated with chronic activation of glia²²⁰ and focal stage-dependent injury-induced factors. Contextdependent responses should be expected for NSAIDs that act as cyclooxygenase inhibitors to reduce concentrations of prostaglandin products—notably PGE2-which act through PTGER1-4 receptors to produce very different outcomes. For example, PTGER2 activation predominantly engages proinflammatory neurotoxic pathways and downregulates AB phagocytosis, whereas PTGER4 ligands can produce antiinflammatory and neuroprotective effects.^{221,222} Most importantly, these receptors have a role in promotion of resolution of chronic neuroinflammation.²²³ Thus, conventional cyclooxygenase-inhibiting NSAIDs could block incipient inflammation-driven Alzheimer's disease pathogenesis at early stages. Additionally, these NSAIDS can have adverse effects in advanced disease, potentially by restriction of resolution and interference with phagocytic clearance of AB and extracellular tau aggregates.

Panel: Recommended steps to advance and harness understanding of neuroinflammation in Alzheimer's disease

Develop new animal models

New models should recapitulate multiple facets of Alzheimer's disease and should not be restricted to transgenic expression of human mutations of familial Alzheimer's disease. In addition to amyloid β and tau pathology, models should include aspects of multiple neurotransmitter loss, disease spreading, and late onset of disease. Ideally, new models would show Alzheimer's disease-like vascular pathology, synaptic destruction, and neuronal loss. Future experiments with animal models should take into account disease modifiers such as systemic inflammation, insulin resistance, brain trauma, nutritional states, physical inactivity, and obesity.

Identify biomarkers of the inflammatory component

New biomarkers could be developed for disease diagnosis, and for monitoring of preventive and therapeutic strategies. Such biomarkers could be blood-based, CSF-based, or imaging-based, and enable discrimination of acute from chronic neuroinflammation.

Define pathologies and periods of neuroinflammation

Understanding is needed of the contributions of microglia, macrophages, astrocytes, neurons, and endothelial cells during the course of Alzheimer's disease. These insights could help to identify which inflammatory processes are protective, which are harmful, and which are relevant for disease pathogenesis at different stages of Alzheimer's disease.

Exploit effects of mutations, epigenetics, and the microbiome on neuroinflammation Discoveries that suggest a direct immune-related modification of the onset, progress, and phenotype of Alzheimer's disease, including single-nucleotide polymorphisms (SNPs) in immune-associated genes, epigenetic immune regulation, and the effect of the microbiome on innate immunity, will be important to consider.

Design observational studies and preventive clinical trials to target immune response Observational studies should closely monitor the clinical course of patients with SNPs in immune-related genes that have been associated with increased risk of Alzheimer's disease. Clinical trials could include preventive trials of strategies to inhibit detrimental aspects of neuroinflammation before any cognitive decline or to foster beneficial immunity, relevant to non-steroidal anti-inflammatory drug epidemiology.

Stage-dependent efficacy has likewise been suggested for anti-Aβ immunotherapy that stimulates microglial phagocytosis of AB, and potential benefits might be seen only with early intervention. A plausible argument is that ageing or Aβ-induced or injury-induced inflammation initiates tauopathy to drive neurodegeneration and downstream clinical decline. Thus, possible explanations for failure of immunotherapy or anti-inflammatory therapy to treat established dementia include an inability to halt the spread of fully established and seeded tauopathy and to rescue deficits driven by neuron loss. Whatever the explanations for past NSAID trial failures are, on the basis of compelling new genetic evidence for a causal role for innate immunity in Alzheimer's disease risk, new trials with both longer and earlier interventions and alternative approaches to favourably modulate neuroinflammation are warranted.

Interventional anti-inflammatory trials

Randomised trials with NSAIDs in patients with Alzheimer's disease show some evidence of success

	Drug	Participants	Treatment duration*	Primary endpoint	Finding
Rogers et al (1993) ²⁰⁴	Indometacin 100-150 mg daily versus placebo	28 patients with AD dementia randomised 1:1	6 months	Cognitive trajectory on a battery of psychometric tests	Positive effects (after 36% attrition; p<0.003)
De Jong et al (2008) ²⁰⁵	Indometacin 100 mg daily with omeprazole versus placebo	51 patients with mild-to- moderate AD randomised 1:1	1 year	Change in score on ADAS-Cog	Neutral-to-positive effects (after 25% attrition; not significant)
Aisen et al (2000) ²⁰⁶	Prednisone (20 mg once daily tapered to 10 mg) versus placebo	138 patients with AD randomised 1:1	1 year	Change in score on ADAS-Cog	Neutral-to-negative effects (worsening of secondary endpoint behavioural measures; not significant)
Aisen et al (2003) ²⁰⁷	Naproxen sodium 220 mg twice daily or rofecoxib 25 mg once daily versus placebo	351 patients with mild-to- moderate AD (MMSE score 13–26)	1 year	Change in score on ADAS-Cog	Neutral-to-negative effects greater decline in rofecoxib group (p=0·09 after adjustment for multiple comparisons)
Aisen et al (2002) ²⁰⁸	Nimesulide 100 mg twice daily versus placebo	40 patients with AD randomised 1:1	3 months	Composite of cognitive, behavioural, and functional outcomes	No apparent effect
Reines et al (2004) ²⁰⁹	Rofecoxib 25 mg once daily versus placebo	692 patients with mild-to- moderate AD randomised 1:1	1 year	ADAS-Cog, CIBIC+	Trend towards negative effects (after 30% attrition)
Thal et al (2005) ²¹⁰	Rofecoxib 25 mg once daily versus placebo	1457 patients with MCI randomised 1:1	3·5 years	Change in status from MCI to AD dementia	Increased progression to AD dementia in rofecoxib grou (p=0·011); no effects on secondary outcomes
Van Gool et al (2001) ²¹¹	Hydroxychloroquine (200–400 mg once daily by body weight) versus placebo	168 patients with mild AD randomised 1:1	18 months	Functional status questionnaire, ADAS- Cog, behavioural symptoms	No apparent effect
ADAPT Research Group (2007 and 2008) ^{212,213}	Celecoxib 100 mg twice daily or naproxen sodium 220 mg twice daily versus placebo	2528 healthy individuals with family history of AD randomised 1:1:1.5	1–3 years	Onset of AD	Trend towards negative effects
ADAPT Research Group (2007 and 2008) ^{212,213}	Celecoxib 100 mg twice daily or naproxen sodium 220 mg twice daily versus placebo	2528 healthy individuals with family history of AD randomised 1:1:1.5	1–3 years	Cognitive decline on battery of neuropsychological tests	Trend towards negative effects
Simons et al (2002) ²¹⁴	Simvastatin up to 80 mg per day as tolerated versus placebo	44 patients with AD randomised 1:1	26 weeks	CSF biomarkers $A\beta_{_{1\text{-}40}}$ and $A\beta_{_{1\text{-}42}}$	No apparent effect
Sparks et al (2005) ²¹⁵	Atorvastatin 80 mg once daily versus placebo	67 patients with mild AD randomised 1:1	1 year	ADAS-Cog, CGI (co-primaries), LOCF analysis	Trend towards positive effects
Feldman et al (2010) ²¹⁶	Atorvastatin 80 mg once daily versus placebo	640 patients with mild-to- moderate AD (MMSE 13-25) randomised 1:1	72 weeks	ADAS-Cog, CGI (co-primaries)	No apparent effect
Harrington et al (2011) ²¹⁷	Rosiglitazone 2 mg or 8 mg daily	2981 patients with mild-to- moderate AD randomised 1:1	48 weeks	ADAS-Cog, CDR sum of boxes	No apparent effect
Breitner et al (2011) ²¹⁸	Celecoxib 100 mg twice daily or naproxen sodium 220 mg twice daily versus placebo	Follow-up of 2071 participants randomised in ADAPT	Follow-up 2–4 years after termination of treatments	Onset of AD, CSF tau, plasma tau, and CSF $A\beta_{_{1\!-\!1\!2}}$	No apparent effect for celecoxib, possible positive effects for naproxen (including reduced ratio of CSF tau to $A\beta_{i\rightarrow e}$)
ADAPT Research Group (2013) ²¹⁹	Celecoxib 100 mg twice daily or naproxen sodium 220 mg twice daily versus placebo	Follow-up of 1537 participants randomised in ADAPT	Follow-up 5-7 years after termination of treatments	Onset of AD	No apparent effect

AD=Alzheimer's disease. ADAS-Cog=Alzheimer Disease Assessment Scale-cognitive portion. MMSE=Mini-Mental State Examination. CIBIC+=Clinician Interview-Based Impression of Change plus Caregiver Input. MCI=mild cognitive impairment. ADAPT= Alzheimer's Disease Anti-inflammatory Prevention Trial. Aβ=amyloid β. CGI=Clinician Global Impression. LOCF-last observation carried forward. CDR=Clinical Dementia Rating. "Duration of follow-up is given for participants followed after randomisation in the ADAPT trial. "Duration of follow-up is given for participants followed after randomisation in the ADAPT trial." We searched PubMed for publications up until 2014 using the search terms "anti-inflammatory", "Alzheimer" and "trial" for randomised controlled trials published in English. The table is not an exhaustive list of studies, but provides a list of trial results that have, in our view, had a notable effect on the direction of subsequent research. Priority was given to trials with sufficient power to give a meaningful result, definition of clinical outcomes, and specification of design method to enable firm conclusions to be drawn (including inference of uncertainty). Two trials^{204,205} were included because of their effect on later work, despite the fact that they failed to meet the aforementioned criteria.

Table: Selected clinical trials of anti-inflammatory drugs in patients with Alzheimer's disease

(table). Early trials with indometacin that suggested reduced cognitive decline²⁰⁴ were not replicated, ²⁰⁵ and large-scale trials with other NSAIDs seemed to be unsuccessful.207,209 Randomised trials with other antiinflammatory drugs, including prednisone,206 hydroxychloroquine, 211 simvastatin, 214 atorvastatin, 215,216 aspirin, 224 and rosiglitazone,217 likewise showed no clinically significant changes in primary cognitive outcomes in patients with prodromal symptoms or Alzheimer's disease dementia. However, although a large randomised study of the NSAIDs naproxen and celecoxib initially reported a detrimental effect for both,212 a longer-term follow-up of these patients suggested that timing and choice of specific NSAID might be key.²¹⁸ Thus, the early detrimental effects were mostly in a small group of patients with early cognitive impairment and, in keeping with epidemiological studies, naproxen seemed thereafter to be protective in patients who had been asymptomatic at baseline.²¹⁸

Immunisation in patients and mouse models of Alzheimer's disease

In some (but not all) studies of effects of immunisation, preponderance of M1 and M2 phenotypes has been reported in response to specific conditions and cytokine exposures. With ageing, Aβ-depositing mice increase expression of alternative activation state genes and deactivation state genes at the expense of classic activation state genes. 225 However, Jimenez and colleagues²²⁶ show age-associated increased expression of mRNA for TNFα, interleukin 1 and iNOS, but also interleukin 4, interleukin 10 and TGF-B, which they interpret as a shift from an alternative to classical activation state. When these mouse models are treated with antibodies, a shift in the activation state occurs over the course of treatment. Initial studies²²⁷ noted reciprocal changes in markers such as kinases, with MAP kinase p38 declining and MAP kinase p44/42 increasing during antibody treatment. Subsequent work using mRNA markers to distinguish between the M1 and M2 phenotypes identified a transition from an M2 phenotype before treatment to an M1 phenotype after antibody treatment.225 Similar shifts in microglia activation states seem to be associated with Alzheimer's disease in vaccination studies. Compared with tissues from non-vaccinated patients with Alzheimer's disease, tissue from vaccinated patients has reduced staining for several microglial markers, including the scavenger receptor SCARA1 and Fcy receptor, and deposited AB. 228 Cases coming to autopsy within 2 years of the vaccination also showed increased levels microhaemorrhage and vascular AB deposits, plus appearance of phagocytic microglia.²²⁹ Similar outcomes were previously seen in aged mice treated with antibodies.230 In the phase 2 and phase 3 trials with the antibody bapineuzumab, events referred to as Aβrelated imaging abnormalities were seen on MRI scans.231 To some extent, these abnormalities in mouse models could be diminished by reduction of antibody affinity for Fcy receptors via deglycosylation, ²³² suggesting that the microglia activation caused by the antibody–antigen interaction might have a role in these vascular responses to immunotherapy.

Conclusions and future directions

Evidence exists that neuroinflammation might drive the pathogenic process in Alzheimer's disease. The brain can no longer be viewed as an immune-privileged organ, and advances in immunology need to be integrated into the known pathogenic pathways of diverse neurodegenerative disorders. The ligandreceptor interactions in the CNS microenvironment that keep microglia under tight control in the healthy brain are perturbed in chronic neurodegenerative disease, but when and how this occurs in Alzheimer's disease is unclear. Although the simple idea of activated microglia has been a useful one, it has no doubt hindered understanding and recognition of the diversity of microglial phenotypes and the extraordinary plasticity of these cells. An important goal of future studies will be to better understand the individual contributions of microglia and other cell types to the neuroinflammatory response during the course of Alzheimer's disease (panel). Other priorities include development of animal models that recapitulate several facets of the disease. The scarcity of methods to assay the different states of microglia activation in vivo adds to the difficulty of understanding the role of neuroinflammation in the human CNS at present. Improved ligands to target microglial activation for PET or other imaging modalities will be key to progress.

The innate immune cells of the brain respond rapidly to systemic events, and these responses are exaggerated in the ageing and diseased brain. In future studies, the effect of systemic comorbidities of Alzheimer's disease (such as diabetes and hypertension), associated systemic inflammation, and ageing as a major risk factor for Alzheimer's disease, should be considered in efforts to understand and exploit the immunological processes associated with the disease (panel). Recognition that modification of the immune system contributes to pathogenesis of chronic neurodegenerative diseases opens many potential routes to delay their onset and progression.

Search strategy and selection criteria

We searched PubMed for journal articles published in English between Jan 1, 2009, and Oct 31, 2014, for the terms "neuroinflammation" or "inflammation" and "Alzheimer", and included those papers judged to be most relevant to the focus of this Review. Additionally, we identified and included older papers with ground-breaking findings that led to recent research, using PubMed and by searches of the authors' own files and the reference lists of selected papers.

Contributors

All authors provided sections of text covering their area of expertise and participated in the proofreading and discussion. MTH and MPK wrote the manuscript and drafted the figures.

Declaration of interests

MTH and MPK have a patent pending on nitration of amyloid β peptides (WO 2011006871 A1). GEL has a patent pending on an RXR agonist in Alzheimer's disease (WO 2011006157 A2). AH reports grants from Boehringer Ingelheim Pharma during the preparation of the Review. SAF reports grants from Veterans Affairs and the US National Institutes of Health (NIH) during the preparation of the Review, and a patent for curcumin formulation (application no. 60,779,817 [WO 2007103435 A3]) with royalties paid. RMR reports grants from the UK National MS Society during the preparation of the Review. CV reports grants from Pfizer, outside the submitted work. KY reports grants from MEXT, Japan, during the preparation of the Review. JK reports grants from Baxter, and personal fees from Medeia Therapeutics, outside the submitted work. CH reports grants from Pfizer, outside the submitted work. GMC reports grants from Veterans Affairs and NIH during the preparation of the Review. All other authors declare no competing interests.

Acknowledgments

The authors of this Review were participants of the 3rd Venusberg Meeting on Neuroinflammation on Feb 28–March 2, 2013, in Bonn, Germany.

References

- Zhang B, Gaiteri C, Bodea L-G, et al. Integrated systems approach identifies genetic nodes and networks in late-onset Alzheimer's disease. Cell 2013; 153: 707–20.
- 2 Guerreiro R, Wojtas A, Bras J, et al, and the Alzheimer Genetic Analysis Group. TREM2 variants in Alzheimer's disease. N Engl J Med 2013; 368: 117–27.
- 3 Bradshaw EM, Chibnik LB, Keenan BT, et al, and the Alzheimer Disease Neuroimaging Initiative. CD33 Alzheimer's disease locus: altered monocyte function and amyloid biology. *Nat Neurosci* 2013; 16: 848–50.
- 4 Griciuc A, Serrano-Pozo A, Parrado AR, et al. Alzheimer's disease risk gene CD33 inhibits microglial uptake of amyloid beta. *Neuron* 2013; 78: 631–43.
- Kettenmann H, Hanisch U-K, Noda M, Verkhratsky A. Physiology of microglia. Physiol Rev 2011; 91: 461–553.
- 6 Ji K, Akgul G, Wollmuth LP, Tsirka SE. Microglia actively regulate the number of functional synapses. PLoS One 2013; 8: e56293.
- 7 Parkhurst CN, Yang G, Ninan I, et al. Microglia promote learningdependent synapse formation through brain-derived neurotrophic factor. *Cell* 2013; 155: 1596–609.
- 8 Bamberger ME, Harris ME, McDonald DR, Husemann J, Landreth GE. A cell surface receptor complex for fibrillar beta-amyloid mediates microglial activation. J Neurosci 2003; 23: 2665–74.
- 9 Paresce DM, Ghosh RN, Maxfield FR. Microglial cells internalize aggregates of the Alzheimer's disease amyloid beta-protein via a scavenger receptor. *Neuron* 1996; 17: 553–65.
- Stewart CR, Stuart LM, Wilkinson K, et al. CD36 ligands promote sterile inflammation through assembly of a Toll-like receptor 4 and 6 heterodimer. Nat Immunol 2010; 11: 155–61.
- Liu Y, Walter S, Stagi M, et al. LPS receptor (CD14): a receptor for phagocytosis of Alzheimer's amyloid peptide. *Brain* 2005; 128: 1778–89.
- 12 Querfurth HW, LaFerla FM. Alzheimer's disease. N Engl J Med 2010: 362: 329–44.
- 13 Kummer MP, Heneka MT. Truncated and modified amyloid-beta species. Alzheimers Res Ther 2014; 6: 28.
- 14 El Khoury JB, Moore KJ, Means TK, et al. CD36 mediates the innate host response to beta-amyloid. J Exp Med 2003; 197: 1657–66.
- 15 Sheedy FJ, Grebe A, Rayner KJ, et al. CD36 coordinates NLRP3 inflammasome activation by facilitating intracellular nucleation of soluble ligands into particulate ligands in sterile inflammation. Nat Immunol 2013; 14: 812–20.
- 16 Lee CYD, Landreth GE. The role of microglia in amyloid clearance from the AD brain. J Neural Transm 2010; 117: 949–60.
- 17 Mawuenyega KG, Sigurdson W, Ovod V, et al. Decreased clearance of CNS β-amyloid in Alzheimer's disease. Science 2010; 330: 1774.

- 18 Hickman SE, Allison EK, El Khoury J. Microglial dysfunction and defective beta-amyloid clearance pathways in aging Alzheimer's disease mice. J Neurosci 2008; 28: 8354–60.
- Frank S, Burbach GJ, Bonin M, et al. TREM2 is upregulated in amyloid plaque-associated microglia in aged APP23 transgenic mice. Glia 2008; 56: 1438–47.
- 20 Hickman SE, Kingery ND, Ohsumi TK, et al. The microglial sensome revealed by direct RNA sequencing. *Nat Neurosci* 2013; 16: 1896–905.
- 21 Hsieh CL, Koike M, Spusta SC, et al. A role for TREM2 ligands in the phagocytosis of apoptotic neuronal cells by microglia. J Neurochem 2009; 109: 1144–56.
- 22 Sierra-Filardi E, Puig-Kröger A, Blanco FJ, et al. Activin A skews macrophage polarization by promoting a proinflammatory phenotype and inhibiting the acquisition of anti-inflammatory macrophage markers. *Blood* 2011; 117: 5092–101.
- 23 Colton CA, Mott RT, Sharpe H, Xu Q, Van Nostrand WE, Vitek MP. Expression profiles for macrophage alternative activation genes in AD and in mouse models of AD. J Neuroinflammation 2006; 3: 27.
- 24 Mantovani A, Sica A, Sozzani S, Allavena P, Vecchi A, Locati M. The chemokine system in diverse forms of macrophage activation and polarization. *Trends Immunol* 2004: 25: 677–86.
- 25 Mantovani A, Sozzani S, Locati M, Allavena P, Sica A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol* 2002; 23: 549–55.
- 26 Koenigsknecht-Talboo J, Landreth GE. Microglial phagocytosis induced by fibrillar beta-amyloid and IgGs are differentially regulated by proinflammatory cytokines. J Neurosci 2005; 25: 8240–49.
- 27 Goerdt S, Orfanos CE. Other functions, other genes: alternative activation of antigen-presenting cells. *Immunity* 1999; 10: 137–42.
- Zelcer N, Khanlou N, Clare R, et al. Attenuation of neuroinflammation and Alzheimer's disease pathology by liver x receptors. Proc Natl Acad Sci USA 2007; 104: 10601–06.
- Colton C, Wilcock DM. Assessing activation states in microglia. CNS Neurol Disord Drug Targets 2010; 9: 174–91.
- 30 Town T, Laouar Y, Pittenger C, et al. Blocking TGF-beta-Smad2/3 innate immune signaling mitigates Alzheimer-like pathology. Nat Med 2008; 14: 681–87.
- 31 Xue J, Schmidt SV, Sander J, et al. Transcriptome-based network analysis reveals a spectrum model of human macrophage activation. *Immunity* 2014; 40: 274–88.
- 32 Perry VH, Teeling J. Microglia and macrophages of the central nervous system: the contribution of microglia priming and systemic inflammation to chronic neurodegeneration. *Semin Immunopathol* 2013; 35: 601–12.
- 33 Orre M, Kamphuis W, Dooves S, et al. Reactive glia show increased immunoproteasome activity in Alzheimer's disease. *Brain* 2013; 136: 1415–31.
- 34 Simard AR, Soulet D, Gowing G, Julien J-P, Rivest S. Bone marrow-derived microglia play a critical role in restricting senile plaque formation in Alzheimer's disease. *Neuron* 2006; 49: 489–502.
- 35 El Khoury J, Toft M, Hickman SE, et al. Ccr2 deficiency impairs microglial accumulation and accelerates progression of Alzheimer-like disease. *Nat Med* 2007; 13: 432–38.
- 36 Mildner A, Schlevogt B, Kierdorf K, et al. Distinct and nonredundant roles of microglia and myeloid subsets in mouse models of Alzheimer's disease. J Neurosci 2011; 31: 11159–71.
- 37 Hawkes CA, McLaurin J. Selective targeting of perivascular macrophages for clearance of beta-amyloid in cerebral amyloid angiopathy. Proc Natl Acad Sci USA 2009; 106: 1261–66.
- 38 Ginhoux F, Greter M, Leboeuf M, et al. Fate mapping analysis reveals that adult microglia derive from primitive macrophages. *Science* 2010; 330: 841–45.
- 39 Grathwohl SA, Kälin RE, Bolmont T, et al. Formation and maintenance of Alzheimer's disease beta-amyloid plaques in the absence of microglia. Nat Neurosci 2009; 12: 1361–63.
- 40 Streit WJ, Braak H, Xue Q-S, Bechmann I. Dystrophic (senescent) rather than activated microglial cells are associated with tau pathology and likely precede neurodegeneration in Alzheimer's disease. Acta Neuropathol 2009; 118: 475–85.

- 41 Krabbe G, Halle A, Matyash V, et al. Functional impairment of microglia coincides with beta-amyloid deposition in mice with Alzheimer-like pathology. PLoS One 2013; 8: e60921.
- 42 Lucin KM, O'Brien CE, Bieri G, et al. Microglial beclin 1 regulates retromer trafficking and phagocytosis and is impaired in Alzheimer's disease. Neuron 2013; 79: 873–86.
- 43 Yamanaka M, Ishikawa T, Griep A, Axt D, Kummer MP, Heneka MT. PPARγ/RXRα-induced and CD36-mediated microglial amyloid-β phagocytosis results in cognitive improvement in amyloid precursor protein/presenilin 1 mice. J Neurosci 2012; 32: 17321–31.
- 44 Cramer PE, Cirrito JR, Wesson DW, et al. ApoE-directed therapeutics rapidly clear β-amyloid and reverse deficits in AD mouse models. *Science* 2012; 335: 1503–06.
- 45 Fitz NF, Cronican AA, Lefterov I, Koldamova R. Comment on "ApoE-directed therapeutics rapidly clear β-amyloid and reverse deficits in AD mouse models". Science 2013; 340: 924–c.
- 46 Price AR, Xu G, Siemienski ZB, et al. Comment on "ApoE-directed therapeutics rapidly clear β-amyloid and reverse deficits in AD mouse models". Science 2013; 340: 924–d.
- 47 Tesseur I, Lo AC, Roberfroid A, et al. Comment on "ApoE-directed therapeutics rapidly clear β-amyloid and reverse deficits in AD mouse 'models". Science 2013; 340: 924–e.
- 48 Veeraraghavalu K, Zhang C, Miller S, et al. Comment on "ApoE-directed therapeutics rapidly clear β-amyloid and reverse deficits in AD mouse models". Science 2013; 340: 924–f.
- 49 Sofroniew MV. Molecular dissection of reactive astrogliosis and glial scar formation. *Trends Neurosci* 2009; 32: 638–47.
- 50 Sofroniew MV, Vinters HV. Astrocytes: biology and pathology. Acta Neuropathol 2010; 119: 7–35.
- 51 Medeiros R, LaFerla FM. Astrocytes: conductors of the Alzheimer disease neuroinflammatory symphony. Exp Neurol 2013; 239: 133–38.
- Olabarria M, Noristani HN, Verkhratsky A, Rodríguez JJ. Concomitant astroglial atrophy and astrogliosis in a triple transgenic animal model of Alzheimer's disease. Glia 2010; 58: 831–38.
- 53 Kummer MP, Hammerschmidt T, Martinez A, et al. Ear2 deletion causes early memory and learning deficits in APP/PS1 mice. J Neurosci 2014; 34: 8845–54.
- Olabarria M, Noristani HN, Verkhratsky A, Rodríguez JJ. Age-dependent decrease in glutamine synthetase expression in the hippocampal astroglia of the triple transgenic Alzheimer's disease mouse model: mechanism for deficient glutamatergic transmission? Mol Neurodegener 2011; 6: 55.
- 55 Yeh C-Y, Vadhwana B, Verkhratsky A, Rodríguez JJ. Early astrocytic atrophy in the entorhinal cortex of a triple transgenic animal model of Alzheimer's disease. ASN Neuro 2011; 3: 271–79.
- 56 Kulijewicz-Nawrot M, Verkhratsky A, Chvátal A, Syková E, Rodríguez JJ. Astrocytic cytoskeletal atrophy in the medial prefrontal cortex of a triple transgenic mouse model of Alzheimer's disease. J Anat 2012; 221: 252–62.
- 57 Beauquis J, Pavía P, Pomilio C, et al. Environmental enrichment prevents astroglial pathological changes in the hippocampus of APP transgenic mice, model of Alzheimer's disease. Exp Neurol 2013; 239: 28–37.
- 58 Furman JL, Sama DM, Gant JC, et al. Targeting astrocytes ameliorates neurologic changes in a mouse model of Alzheimer's disease. J Neurosci 2012; 32: 16129–40.
- 59 Wyss-Coray T, Loike JD, Brionne TC, et al. Adult mouse astrocytes degrade amyloid-beta in vitro and in situ. Nat Med 2003; 9: 453–57.
- 60 Koistinaho M, Lin S, Wu X, et al. Apolipoprotein E promotes astrocyte colocalization and degradation of deposited amyloid-beta peptides. *Nat Med* 2004; 10: 719–26.
- 61 Jiang Q, Lee CYD, Mandrekar S, et al. ApoE promotes the proteolytic degradation of Aβ. Neuron 2008; 58: 681–93.
- 62 Terwel D, Steffensen KR, Verghese PB, et al. Critical role of astroglial apolipoprotein E and liver X receptor-α expression for microglial Aβ phagocytosis. J Neurosci 2011; 31: 7049–59.
- 63 Saido T, Leissring MA. Proteolytic degradation of amyloid β-protein. Cold Spring Harb Perspect Med 2012; 2: a006379.
- 64 Pihlaja R, Koistinaho J, Kauppinen R, Sandholm J, Tanila H, Koistinaho M. Multiple cellular and molecular mechanisms are involved in human Aβ clearance by transplanted adult astrocytes. *Glia* 2011; 59: 1643–57.

- 65 Iliff JJ, Wang M, Liao Y, et al. A paravascular pathway facilitates CSF flow through the brain parenchyma and the clearance of interstitial solutes, including amyloid β. Sci Transl Med 2012; 4: 147ra111.
- 66 Patel NS, Paris D, Mathura V, Quadros AN, Crawford FC, Mullan MJ. Inflammatory cytokine levels correlate with amyloid load in transgenic mouse models of Alzheimer's disease. J Neuroinflammation 2005; 2: 9.
- 67 Lue LF, Rydel R, Brigham EF, et al. Inflammatory repertoire of Alzheimer's disease and nondemented elderly microglia in vitro. Glia 2001; 35: 72–79.
- 68 Laske C, Stransky E, Hoffmann N, et al. Macrophage colonystimulating factor (M-CSF) in plasma and CSF of patients with mild cognitive impairment and Alzheimer's disease. Curr Alzheimer Res 2010; 7: 409–14.
- 69 Heneka MT, Kummer MP, Latz E, et al. Innate immune activation in neurodegenerative disease. Nat Rev Immunol 2014; 14: 463–77.
- 70 Heneka MT, Kummer MP, Stutz A, et al. NLRP3 is activated in Alzheimer's disease and contributes to pathology in APP/PS1 mice. Nature 2013; 493: 674–78.
- 71 Akama KT, Van Eldik LJ. β-amyloid stimulation of inducible nitric-oxide synthase in astrocytes is interleukin-1β- and tumor necrosis factor-α (TNFα)-dependent, and involves a TNFα receptor-associated factor- and NFκB-inducing kinase-dependent signaling mechanism. J Biol Chem 2000; 275: 7918–24.
- 72 Mrak RE, Sheng JG, Griffin WS. Glial cytokines in Alzheimer's disease: review and pathogenic implications. *Hum Pathol* 1995; 26: 816–23
- 73 Vom Berg J, Prokop S, Miller KR, et al. Inhibition of IL-12/IL-23 signaling reduces Alzheimer's disease-like pathology and cognitive decline. Nat Med 2012; 18: 1812–19.
- 74 Tan M-S, Yu J-T, Jiang T, Zhu X-C, Guan H-S, Tan L. IL12/23 p40 inhibition ameliorates Alzheimer's disease-associated neuropathology and spatial memory in SAMP8 mice. J Alzheimers Dis 2014; 38: 633–46.
- 75 Rentzos M, Paraskevas GP, Kapaki E, et al. Interleukin-12 is reduced in cerebrospinal fluid of patients with Alzheimer's disease and frontotemporal dementia. J Neurol Sci 2006; 249: 110–14.
- 76 Tarkowski E, Andreasen N, Tarkowski A, Blennow K. Intrathecal inflammation precedes development of Alzheimer's disease. J Neurol Neurosurg Psychiatry 2003; 74: 1200–05.
- 77 Meda L, Cassatella MA, Szendrei GI, et al. Activation of microglial cells by β -amyloid protein and interferon- γ . Nature 1995; 374: 647–50.
- 78 Tan J, Town T, Paris D, et al. Microglial activation resulting from CD40–CD40L interaction after beta-amyloid stimulation. *Science* 1999; 286: 2352–55.
- 79 Tan J, Town T, Crawford F, et al. Role of CD40 ligand in amyloidosis in transgenic Alzheimer's mice. *Nat Neurosci* 2002; 5: 1288–93.
- 80 Jin J-J, Kim H-D, Maxwell JA, Li L, Fukuchi K. Toll-like receptor 4-dependent upregulation of cytokines in a transgenic mouse model of Alzheimer's disease. *J Neuroinflammation* 2008; 5: 23.
- 81 Shaftel SS, Carlson TJ, Olschowka JA, Kyrkanides S, Matousek SB, O'Banion MK. Chronic interleukin-1β expression in mouse brain leads to leukocyte infiltration and neutrophil-independent blood brain barrier permeability without overt neurodegeneration. J Neurosci 2007; 27: 9301–09.
- 82 Ghosh S, Wu MD, Shaftel SS, et al. Sustained interleukin-1β overexpression exacerbates tau pathology despite reduced amyloid burden in an Alzheimer's mouse model. *J Neurosci* 2013; 33: 5053–64.
- 83 Chakrabarty P, Ceballos-Diaz C, Beccard A, et al. IFN-gamma promotes complement expression and attenuates amyloid plaque deposition in amyloid beta precursor protein transgenic mice. *J Immunol* 2010; 184: 5333–43.
- 84 Chakrabarty P, Jansen-West K, Beccard A, et al. Massive gliosis induced by interleukin-6 suppresses Aβ deposition in vivo: evidence against inflammation as a driving force for amyloid deposition. FASEB I 2010: 24: 548–59.
- 85 Chakrabarty P, Herring A, Ceballos-Diaz C, Das P, Golde TE. Hippocampal expression of murine TNFα results in attenuation of amyloid deposition in vivo. Mol Neurodegener 2011; 6: 16.

- 86 Chakrabarty P, Tianbai L, Herring A, Ceballos-Diaz C, Das P, Golde TE. Hippocampal expression of murine IL-4 results in exacerbation of amyloid deposition. Mol Neurodegener 2012; 7: 36.
- 87 Savarin-Vuaillat C, Ransohoff RM. Chemokines and chemokine receptors in neurological disease: raise, retain, or reduce? Neurotherapeutics 2007; 4: 590–601.
- 88 Xia MQ, Qin SX, Wu LJ, Mackay CR, Hyman BT. Immunohistochemical study of the beta-chemokine receptors CCR3 and CCR5 and their ligands in normal and Alzheimer's disease brains. Am J Pathol 1998; 153: 31–37.
- 89 Ishizuka K, Kimura T, Igata-yi R, Katsuragi S, Takamatsu J, Miyakawa T. Identification of monocyte chemoattractant protein-1 in senile plaques and reactive microglia of Alzheimer's disease. Psychiatry Clin Neurosci 1997; 51: 135–38.
- 90 Smits HA, Rijsmus A, van Loon JH, et al. Amyloid-beta-induced chemokine production in primary human macrophages and astrocytes. J Neuroimmunol 2002; 127: 160–68.
- 91 Lue LF, Walker DG, Rogers J. Modeling microglial activation in Alzheimer's disease with human postmortem microglial cultures. Neurobiol Aging 2001; 22: 945–56.
- 92 Fuhrmann M, Bittner T, Jung CKE, et al. Microglial Cx3cr1 knockout prevents neuron loss in a mouse model of Alzheimer's disease. Nat Neurosci 2010: 13: 411–13.
- 93 Lee S, Varvel NH, Konerth ME, et al. CX3CR1 deficiency alters microglial activation and reduces beta-amyloid deposition in two Alzheimer's disease mouse models. Am J Pathol 2010; 177: 2549–62.
- 94 Cho S-H, Sun B, Zhou Y, et al. CX3CR1 protein signaling modulates microglial activation and protects against plaqueindependent cognitive deficits in a mouse model of Alzheimer disease. *I Biol Chem* 2011: 286: 32713–22.
- 95 Lee YK, Kwak DH, Oh KW, et al. CCR5 deficiency induces astrocyte activation, Aβ deposit and impaired memory function. Neurobiol Learn Mem 2009; 92: 356–63.
- 96 Kiyota T, Yamamoto M, Xiong H, et al. CCL2 accelerates microgliamediated Aβ oligomer formation and progression of neurocognitive dvsfunction. PLoS One 2009: 4: e6197.
- Semple BD, Frugier T, Morganti-Kossmann MC. CCL2 modulates cytokine production in cultured mouse astrocytes. J Neuroinflammation 2010; 7: 67.
- 98 Schroder K, Tschopp J. The inflammasomes. Cell 2010; 140: 821–32.
- 99 van de Veerdonk FL, Netea MG, Dinarello CA, Joosten LAB. Inflammasome activation and IL-1β and IL-18 processing during infection. *Trends Immunol* 2011; 32: 110–16.
- 100 Halle A, Hornung V, Petzold GC, et al. The NALP3 inflammasome is involved in the innate immune response to amyloid-β. Nat Immunol 2008; 9: 857–65.
- 101 Fricker M, Vilalta A, Tolkovsky AM, Brown GC. Caspase inhibitors protect neurons by enabling selective necroptosis of inflamed microglia. J Biol Chem 2013; 288: 9145–52.
- 102 Burguillos MA, Deierborg T, Kavanagh E, et al. Caspase signalling controls microglia activation and neurotoxicity. *Nature* 2011; 472: 319–24.
- 103 Rohn TT, Kokoulina P, Eaton CR, Poon WW. Caspase activation in transgenic mice with Alzheimer-like pathology: results from a pilot study utilizing the caspase inhibitor, Q-VD-OPh. Int J Clin Exp Med 2009; 2: 300–08.
- 104 Biscaro B, Lindvall O, Tesco G, Ekdahl CT, Nitsch RM. Inhibition of microglial activation protects hippocampal neurogenesis and improves cognitive deficits in a transgenic mouse model for Alzheimer's disease. Neurodegener Dis 2012; 9: 187–98.
- 105 Choi S-H, Aid S, Caracciolo L, et al. Cyclooxygenase-1 inhibition reduces amyloid pathology and improves memory deficits in a mouse model of Alzheimer's disease. J Neurochem 2013; 124: 59–68.
- 106 Montine TJ, Sidell KR, Crews BC, et al. Elevated CSF prostaglandin E2 levels in patients with probable AD. *Neurology* 1999; 53: 1495–98.
- 107 Slawik H, Volk B, Fiebich B, Hüll M. Microglial expression of prostaglandin EP3 receptor in excitotoxic lesions in the rat striatum. Neurochem Int 2004; 45: 653–60.
- 108 Shie F-S, Montine KS, Breyer RM, Montine TJ. Microglial EP2 as a new target to increase amyloid beta phagocytosis and decrease amyloid beta-induced damage to neurons. *Brain Pathol* 2005; 15: 134–38.

- 109 Liang X, Wang Q, Hand T, et al. Deletion of the prostaglandin E2 EP2 receptor reduces oxidative damage and amyloid burden in a model of Alzheimer's disease. J Neurosci 2005; 25: 10180–87.
- 110 Shi J, Wang Q, Johansson JU, et al. Inflammatory prostaglandin E2 signaling in a mouse model of Alzheimer disease. Ann Neurol 2012; 72: 788–98.
- 111 Xiang Z, Ho L, Yemul S, et al. Cyclooxygenase-2 promotes amyloid plaque deposition in a mouse model of Alzheimer's disease neuropathology. Gene Expr 2002; 10: 271–78.
- 112 Woodling NS, Wang Q, Priyam PG, et al. Suppression of Alzheimer-associated inflammation by microglial prostaglandin-E2 EP4 receptor signaling. J Neurosci 2014; 34: 5882–94.
- 113 Bazan NG, Molina MF, Gordon WC. Docosahexaenoic acid signalolipidomics in nutrition: significance in aging, neuroinflammation, macular degeneration, Alzheimer's, and other neurodegenerative diseases. Annu Rev Nutr 2011; 31: 321–51.
- 114 Lukiw WJ, Cui J-G, Marcheselli VL, et al. A role for docosahexaenoic acid-derived neuroprotectin D1 in neural cell survival and Alzheimer disease. J Clin Invest 2005; 115: 2774–83.
- 115 Zhao Y, Calon F, Julien C, et al. Docosahexaenoic acid-derived neuroprotectin D1 induces neuronal survival via secretase- and PPARγ-mediated mechanisms in Alzheimer's disease models. PLoS One 2011; 6: e15816.
- 116 Veerhuis R, Nielsen HM, Tenner AJ. Complement in the brain. Mol Immunol 2011; 48: 1592–603.
- 117 Strohmeyer R, Ramirez M, Cole GJ, Mueller K, Rogers J. Association of factor H of the alternative pathway of complement with agrin and complement receptor 3 in the Alzheimer's disease brain. J Neuroimmunol 2002; 131: 135–46.
- 118 Lambert J-C, Heath S, Even G, et al, and the European Alzheimer's Disease Initiative Investigators. Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. Nat Genet 2009; 41: 1094–99.
- 119 Harold D, Abraham R, Hollingworth P, et al. Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. Nat Genet 2009; 41: 1088–93.
- 120 Vodovotz Y, Lucia MS, Flanders KC, et al. Inducible nitric oxide synthase in tangle-bearing neurons of patients with Alzheimer's disease. J Exp Med 1996; 184: 1425–33.
- 121 Nathan C, Calingasan N, Nezezon J, et al. Protection from Alzheimer's-like disease in the mouse by genetic ablation of inducible nitric oxide synthase. *J Exp Med* 2005; 202: 1163–69.
- 122 Jekabsone A, Mander PK, Tickler A, Sharpe M, Brown GC. Fibrillar β-amyloid peptide Aβ1–40 activates microglial proliferation via stimulating TNF-α release and H2O2 derived from NADPH oxidase: a cell culture study. J Neuroinflammation 2006; 3: 24.
- 123 Choi S-H, Aid S, Kim H-W, Jackson SH, Bosetti F. Inhibition of NADPH oxidase promotes alternative and anti-inflammatory microglial activation during neuroinflammation. J Neurochem 2012; 120: 292–301.
- 124 Mander P, Brown GC. Activation of microglial NADPH oxidase is synergistic with glial iNOS expression in inducing neuronal death: a dual-key mechanism of inflammatory neurodegeneration. J Neuroinflammation 2005; 2: 20.
- 125 Butterfield DA, Reed TT, Perluigi M, et al. Elevated levels of 3-nitrotyrosine in brain from subjects with amnestic mild cognitive impairment: implications for the role of nitration in the progression of Alzheimer's disease. *Brain Res* 2007; 1148: 243–48.
- 126 Kummer MP, Hermes M, Delekarte A, et al. Nitration of tyrosine 10 critically enhances amyloid β aggregation and plaque formation. Neuron 2011; 71: 833–44.
- 127 Thiabaud G, Pizzocaro S, Garcia-Serres R, Latour J-M, Monzani E, Casella L. Heme binding induces dimerization and nitration of truncated β-amyloid peptide Aβ16 under oxidative stress.

 Angew Chem Int Ed Engl 2013; 52: 8041–44..
- 128 Cho D-H, Nakamura T, Fang J, et al. S-nitrosylation of Drp1 mediates β-amyloid-related mitochondrial fission and neuronal injury. Science 2009; 324: 102–05.
- 129 Iadecola C. Neurovascular regulation in the normal brain and in Alzheimer's disease. Nat Rev Neurosci 2004; 5: 347–60.
- 130 Shin HK, Jones PB, Garcia-Alloza M, et al. Age-dependent cerebrovascular dysfunction in a transgenic mouse model of cerebral amyloid angiopathy. *Brain* 2007; 130: 2310–19.

- 131 Petzold GC, Murthy VN. Role of astrocytes in neurovascular coupling. Neuron 2011; 71: 782–97.
- 132 Jarre A, Gowert NS, Donner L, et al. Pre-activated blood platelets and a pro-thrombotic phenotype in APP23 mice modeling Alzheimer's disease. Cell Signal 2014; 26: 2040–50.
- 133 Deane R, Du Yan S, Submamaryan RK, et al. RAGE mediates amyloid-β peptide transport across the blood-brain barrier and accumulation in brain. Nat Med 2003; 9: 907–13.
- 134 Zlokovic BV. Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat Rev Neurosci* 2011; 12: 723–38.
- 135 Hart BL. Biological basis of the behavior of sick animals. Neurosci Biobehav Rev 1988; 12: 123–37.
- 136 Yang H, Ochani M, Li J, et al. Reversing established sepsis with antagonists of endogenous high-mobility group box 1. Proc Natl Acad Sci USA 2004; 101: 296–301.
- 137 Rivest S. Regulation of innate immune responses in the brain. Nat Rev Immunol 2009: 9: 429–39.
- 138 Undén A-L, Andréasson A, Elofsson S, et al. Inflammatory cytokines, behaviour and age as determinants of self-rated health in women. Clin Sci (Lond) 2007; 112: 363–73.
- 139 Maitra U, Deng H, Glaros T, et al. Molecular mechanisms responsible for the selective and low-grade induction of proinflammatory mediators in murine macrophages by lipopolysaccharide. *J Immunol* 2012; **189**: 1014–23.
- 140 Pace JL, Russell SW, Torres BA, Johnson HM, Gray PW. Recombinant mouse gamma interferon induces the priming step in macrophage activation for tumor cell killing. J Immunol 1983; 130: 2011–13.
- 141 Godbout JP, Johnson RW. Age and neuroinflammation: a lifetime of psychoneuroimmune consequences. *Immunol Allergy Clin North Am* 2009; 29: 321–37.
- 142 Cunningham C, Wilcockson DC, Campion S, Lunnon K, Perry VH. Central and systemic endotoxin challenges exacerbate the local inflammatory response and increase neuronal death during chronic neurodegeneration. J Neurosci 2005; 25: 9275–84.
- 143 Cunningham C, Campion S, Lunnon K, et al. Systemic inflammation induces acute behavioral and cognitive changes and accelerates neurodegenerative disease. *Biol Psychiatry* 2009; 65: 304–12.
- 144 Field R, Campion S, Warren C, Murray C, Cunningham C. Systemic challenge with the TLR3 agonist poly I:C induces amplified IFNalpha/beta and IL-1beta responses in the diseased brain and exacerbates chronic neurodegeneration. Brain Behav Immun 2010; 24: 996–1007.
- 145 Kyrkanides S, Tallents RH, Miller JN, et al. Osteoarthritis accelerates and exacerbates Alzheimer's disease pathology in mice. J Neuroinflammation 2011; 8: 112.
- 146 Holmes C, Cunningham C, Zotova E, et al. Systemic inflammation and disease progression in Alzheimer disease. *Neurology* 2009; 73: 768–74.
- 147 Holmes C, Cunningham C, Zotova E, Culliford D, Perry VH. Proinflammatory cytokines, sickness behavior, and Alzheimer disease. *Neurology* 2011; 77: 212–18.
- 148 Almond MH, Edwards MR, Barclay WS, Johnston SL. Obesity and susceptibility to severe outcomes following respiratory viral infection. *Thorax* 2013; 68: 684–86.
- 149 Chidiac C. Pneumococcal infections and adult with risk factors. Med Mal Infect 2012; 42: 517–24.
- 150 Bastard J-P, Maachi M, Lagathu C, et al. Recent advances in the relationship between obesity, inflammation, and insulin resistance. Eur Cytokine Netw 2006; 17: 4–12.
- 151 Whitmer RA, Gustafson DR, Barrett-Connor E, Haan MN, Gunderson EP, Yaffe K. Central obesity and increased risk of dementia more than three decades later. *Neurology* 2008; 71: 1057–64.
- 152 Brown BM, Peiffer JJ, Martins RN. Multiple effects of physical activity on molecular and cognitive signs of brain aging: can exercise slow neurodegeneration and delay Alzheimer's disease? Mol Psychiatry 2013; 18: 864–74.
- 153 Takeda S, Sato N, Uchio-Yamada K, et al. Diabetes-accelerated memory dysfunction via cerebrovascular inflammation and Aβ deposition in an Alzheimer mouse model with diabetes. Proc Natl Acad Sci USA 2010; 107: 7036–41.
- 154 Le Chatelier E, Nielsen T, Qin J, et al, and the MetaHIT consortium. Richness of human gut microbiome correlates with metabolic markers. Nature 2013: 500: 541–46.

- 155 Sivanandam TM, Thakur MK. Traumatic brain injury: a risk factor for Alzheimer's disease. Neurosci Biobehav Rev 2012; 36: 1376–81.
- 156 Brody DL, Holtzman DM. Morris water maze search strategy analysis in PDAPP mice before and after experimental traumatic brain injury. Exp Neurol 2006; 197: 330–40.
- 157 Tajiri N, Kellogg SL, Shimizu T, Arendash GW, Borlongan CV. Traumatic brain injury precipitates cognitive impairment and extracellular Aβ aggregation in Alzheimer's disease transgenic mice. PLoS One 2013; 8: e78851.
- 158 Koshinaga M, Katayama Y, Fukushima M, Oshima H, Suma T, Takahata T. Rapid and widespread microglial activation induced by traumatic brain injury in rat brain slices. J Neurotrauma 2000; 17: 185–92.
- 159 Ramlackhansingh AF, Brooks DJ, Greenwood RJ, et al. Inflammation after trauma: microglial activation and traumatic brain injury. Ann Neurol 2011; 70: 374–83.
- 160 Sastre M, Dewachter I, Landreth GE, et al. Nonsteroidal anti-inflammatory drugs and peroxisome proliferator-activated receptor-gamma agonists modulate immunostimulated processing of amyloid precursor protein through regulation of beta-secretase. J Neurosci 2003; 23: 9796–804.
- 161 Corrigan F, Pham CLL, Vink R, et al. The neuroprotective domains of the amyloid precursor protein, in traumatic brain injury, are located in the two growth factor domains. *Brain Res* 2011; 1378: 137–43.
- 162 O'Donnell J, Zeppenfeld D, McConnell E, Pena S, Nedergaard M. Norepinephrine: a neuromodulator that boosts the function of multiple cell types to optimize CNS performance. *Neurochem Res* 2012; 37: 2496–512.
- 163 Marien MR, Colpaert FC, Rosenquist AC. Noradrenergic mechanisms in neurodegenerative diseases: a theory. Brain Res Brain Res Rev 2004; 45: 38–78.
- 164 Zarow C, Lyness SA, Mortimer JA, Chui HC. Neuronal loss is greater in the locus coeruleus than nucleus basalis and substantia nigra in Alzheimer and Parkinson diseases. Arch Neurol 2003; 60: 337–41.
- 165 Kalinin S, Gavrilyuk V, Polak PE, et al. Noradrenaline deficiency in brain increases beta-amyloid plaque burden in an animal model of Alzheimer's disease. Neurobiol Aging 2007; 28: 1206–14.
- 166 Heneka MT, Ramanathan M, Jacobs AH, et al. Locus ceruleus degeneration promotes Alzheimer pathogenesis in amyloid precursor protein 23 transgenic mice. J Neurosci 2006; 26: 1343–54.
- 167 O'Sullivan JB, Ryan KM, Harkin A, Connor TJ. Noradrenaline reuptake inhibitors inhibit expression of chemokines IP-10 and RANTES and cell adhesion molecules VCAM-1 and ICAM-1 in the CNS following a systemic inflammatory challenge. J Neuroimmunol 2010; 220: 34–42.
- 168 Kalinin S, Polak PE, Lin SX, Sakharkar AJ, Pandey SC, Feinstein DL. The noradrenaline precursor L-DOPS reduces pathology in a mouse model of Alzheimer's disease. *Neurobiol Aging* 2012; 33: 1651–63.
- 169 Bolmont T, Haiss F, Eicke D, et al. Dynamics of the microglial/ amyloid interaction indicate a role in plaque maintenance. J Neurosci 2008; 28: 4283–92.
- 170 Meyer-Luehmann M, Coomaraswamy J, Bolmont T, et al. Exogenous induction of cerebral beta-amyloidogenesis is governed by agent and host. *Science* 2006; 313: 1781–84.
- 171 Kim JV, Dustin ML. Innate response to focal necrotic injury inside the blood-brain barrier. J Immunol 2006; 177: 5269–77.
- 172 Perry VH, Nicoll JAR, Holmes C. Microglia in neurodegenerative disease. Nat Rev Neurol 2010; 6: 193–201.
- 173 Rupprecht R, Papadopoulos V, Rammes G, et al. Translocator protein (18 kDa) (TSPO) as a therapeutic target for neurological and psychiatric disorders. Nat Rev Drug Discov 2010; 9: 971–88.
- 174 Jacobs AH, Tavitian B, and the INMiND consortium. Noninvasive molecular imaging of neuroinflammation. J Cereb Blood Flow Metab 2012: 32: 1393–415.
- 175 Venneti S, Lopresti BJ, Wang G, et al. PK11195 labels activated microglia in Alzheimer's disease and in vivo in a mouse model using PET. Neurobiol Aging 2009; 30: 1217–26.
- 176 Roberts JC, Friel SL, Roman S, et al. Autoradiographical imaging of PPARgamma agonist effects on PBR/TSPO binding in TASTPM mice. Exp Neurol 2009; 216: 459–70.
- 177 Rapic S, Backes H, Viel T, et al. Imaging microglial activation and glucose consumption in a mouse model of Alzheimer's disease. Neurobiol Aging 2013; 34: 351–54.

- 178 Heneka MT, Reyes-Irisarri E, Hüll M, Kummer MP. Impact and Therapeutic Potential of PPARs in Alzheimer's Disease. Curr Neuropharmacol 2011; 9: 643–50.
- 179 Chauveau F, Boutin H, Van Camp N, et al. In vivo imaging of neuroinflammation in the rodent brain with [11C]SSR180575, a novel indoleacetamide radioligand of the translocator protein (18 kDa). Eur J Nucl Med Mol Imaging 2011; 38: 509–14.
- 180 Chauveau F, Van Camp N, Dollé F, et al. Comparative evaluation of the translocator protein radioligands 11C-DPA-713, 18F-DPA-714, and 11C-PK11195 in a rat model of acute neuroinflammation. J Nucl Med 2009; 50: 468–76.
- 181 Schuitemaker A, Kropholler MA, Boellaard R, et al. Microglial activation in Alzheimer's disease: an (R)-[11C]PK11195 positron emission tomography study. Neurobiol Aging 2013; 34: 128–36.
- 182 Yasuno F, Kosaka J, Ota M, et al. Increased binding of peripheral benzodiazepine receptor in mild cognitive impairment-dementia converters measured by positron emission tomography with [11C] DAA1106. Psychiatry Res 2012; 203: 67–74.
- 183 Edison P, Archer H, Hinz R, et al. Relationship between the distribution of microglial activation and amyloid deposition in Alzheimer's disease: An 11C-PK11195 and 11C-PIB PET study. J Neurol Neurosurg Psychiatry 2007; 78: 219.
- 184 Yokokura M, Mori N, Yagi S, et al. In vivo changes in microglial activation and amyloid deposits in brain regions with hypometabolism in Alzheimer's disease. Eur J Nucl Med Mol Imaging 2011; 38: 343–51.
- 185 Edison P, Archer HA, Hinz R, et al. Amyloid, hypometabolism, and cognition in Alzheimer disease: an [11C]PIB and [18F]FDG PET study. Neurology 2007; 68: 501–08.
- 186 Kreisl WC, Lyoo CH, McGwier M, et al, and the Biomarkers Consortium PET Radioligand Project Team. In vivo radioligand binding to translocator protein correlates with severity of Alzheimer's disease. *Brain* 2013; 136: 2228–38.
- 187 Okello A, Edison P, Archer HA, et al. Microglial activation and amyloid deposition in mild cognitive impairment: a PET study. Neurology 2009; 72: 56–62.
- 188 Shi H, Belbin O, Medway C, et al. Genetic variants influencing human aging from late-onset Alzheimer's disease (LOAD) genome-wide association studies (GWAS). Neurobiol Aging 2012; 33: 1849 e5–18.
- 189 Brosseron F, Krauthausen M, Kummer M, Heneka MT. Body fluid cytokine levels in mild cognitive impairment and Alzheimer's disease: a comparative overview. Mol Neurobiol 2014; 50: 534–44.
- 190 Galimberti D, Fenoglio C, Scarpini E. Inflammation in neurodegenerative disorders: friend or foe? *Curr Aging Sci* 2008; 1: 30–41.
- 191 Czirr E, Wyss-Coray T. The immunology of neurodegeneration. J Clin Invest 2012; 122: 1156–63.
- 192 Ruckh JM, Zhao J-W, Shadrach JL, et al. Rejuvenation of regeneration in the aging central nervous system. *Cell Stem Cell* 2012; 10: 96–103.
- 193 Villeda S, Wyss-Coray T. Microglia—a wrench in the running wheel? Neuron 2008; 59: 527–29.
- 194 Lista S, Faltraco F, Hampel H. Biological and methodical challenges of blood-based proteomics in the field of neurological research. Prog Neurobiol 2013; 101–102: 18–34.
- 195 Green MJ, Matheson SL, Shepherd A, Weickert CS, Carr VJ. Brain-derived neurotrophic factor levels in schizophrenia: a systematic review with meta-analysis. *Mol Psychiatry* 2011; 16: 960–72.
- 196 Kurita M, Nishino S, Kato M, Numata Y, Sato T. Plasma brainderived neurotrophic factor levels predict the clinical outcome of depression treatment in a naturalistic study. PLoS One 2012; 7: e39212.
- 197 Lee B-H, Kim H, Park S-H, Kim Y-K. Decreased plasma BDNF level in depressive patients. J Affect Disord 2007; 101: 239–44.
- 198 Martinotti G, Di Iorio G, Marini S, Ricci V, De Berardis D, Di Giannantonio M. Nerve growth factor and brain-derived neurotrophic factor concentrations in schizophrenia: a review. J Biol Regul Homeost Agents 2012; 26: 347–56.
- 199 Hu WT, Holtzman DM, Fagan AM, et al, and the Alzheimer's Disease Neuroimaging Initiative. Plasma multianalyte profiling in mild cognitive impairment and Alzheimer disease. *Neurology* 2012; 79: 897–905.

- 200 Ray S, Britschgi M, Herbert C, et al. Classification and prediction of clinical Alzheimer's diagnosis based on plasma signaling proteins. *Nat Med* 2007; 13: 1359–62.
- 201 Soares HD, Potter WZ, Pickering E, et al, and the Biomarkers Consortium Alzheimer's Disease Plasma Proteomics Project. Plasma biomarkers associated with the apolipoprotein E genotype and Alzheimer disease. Arch Neurol 2012; 69: 1310–17.
- 202 Britschgi M, Rufibach K, Huang SLB, et al. Modeling of pathological traits in Alzheimer's disease based on systemic extracellular signaling proteome. Mol Cell Proteomics 2011; 10: M111.008862.
- 203 Johnstone D, Milward EA, Berretta R, Moscato P, and the Alzheimer's Disease Neuroimaging Initiative. Multivariate protein signatures of pre-clinical Alzheimer's disease in the Alzheimer's disease neuroimaging initiative (ADNI) plasma proteome dataset. PLoS One 2012; 7: e34341.
- 204 Rogers J, Kirby LC, Hempelman SR, et al. Clinical trial of indomethacin in Alzheimer's disease. *Neurology* 1993; 43: 1609–11.
- 205 de Jong D, Jansen R, Hoefnagels W, et al. No effect of one-year treatment with indomethacin on Alzheimer's disease progression: a randomized controlled trial. PLoS One 2008; 3: e1475.
- 206 Aisen PS, Davis KL, Berg JD, et al. A randomized controlled trial of prednisone in Alzheimer's disease. Alzheimer's Disease Cooperative Study. *Neurology* 2000; 54: 588–93.
- 207 Aisen PS, Schafer KA, Grundman M, et al, and the Alzheimer's Disease Cooperative Study. Effects of rofecoxib or naproxen vs placebo on Alzheimer disease progression: a randomized controlled trial. JAMA 2003; 289: 2819–26.
- 208 Aisen PS, Schmeidler J, Pasinetti GM. Randomized pilot study of nimesulide treatment in Alzheimer's disease. *Neurology* 2002; 58: 1050–54.
- 209 Reines SA, Block GA, Morris JC, et al, and the Rofecoxib Protocol 091 Study Group. Rofecoxib: no effect on Alzheimer's disease in a 1-year, randomized, blinded, controlled study. *Neurology* 2004; 62: 66–71.
- 210 Thal LJ, Ferris SH, Kirby L, et al, and the Rofecoxib Protocol 078 study group. A randomized, double-blind, study of rofecoxib in patients with mild cognitive impairment. *Neuropsychopharmacology* 2005; 30: 1204–15.
- 211 Van Gool WA, Weinstein HC, Scheltens P, Walstra GJ. Effect of hydroxychloroquine on progression of dementia in early Alzheimer's disease: an 18-month randomised, double-blind, placebo-controlled study. *Lancet* 2001; 358: 455–60.
- 212 Martin BK, Szekely C, Brandt J, et al, and the ADAPT Research Group. Cognitive function over time in the Alzheimer's Disease Anti-inflammatory Prevention Trial (ADAPT): results of a randomized, controlled trial of naproxen and celecoxib. Arch Neurol 2008: 65: 896–905.
- 213 Lyketsos CG, Breitner JC, Green RC, et al, and the ADAPT Research Group. Naproxen and celecoxib do not prevent AD in early results from a randomized controlled trial. *Neurology* 2007; 68: 1800-08
- 214 Simons M, Schwärzler F, Lütjohann D, et al. Treatment with simvastatin in normocholesterolemic patients with Alzheimer's disease: a 26-week randomized, placebo-controlled, double-blind trial. Ann Neurol 2002; 52: 346–50.
- 215 Sparks DL, Sabbagh MN, Connor DJ, et al. Atorvastatin for the treatment of mild to moderate Alzheimer disease: preliminary results. Arch Neurol 2005; 62: 753–57.
- 216 Feldman HH, Doody RS, Kivipelto M, et al, and the LEADe Investigators. Randomized controlled trial of atorvastatin in mild to moderate Alzheimer disease: LEADe. *Neurology* 2010; 74: 956–64.
- 217 Harrington C, Sawchak S, Chiang C, et al. Rosiglitazone does not improve cognition or global function when used as adjunctive therapy to AChE inhibitors in mild-to-moderate Alzheimer's disease: two phase 3 studies. Curr Alzheimer Res 2011; 8: 592–606.
- 218 Breitner JC, Baker LD, Montine TJ, et al, and the ADAPT Research Group. Extended results of the Alzheimer's disease antiinflammatory prevention trial. Alzheimers Dement 2011; 7: 402–11.
- 219 Alzheimer's Disease Anti-inflammatory Prevention Trial Research Group. Results of a follow-up study to the randomized Alzheimer's Disease Anti-inflammatory Prevention Trial (ADAPT). Alzheimers Dement 2013; 9: 714–23.

- 220 Cribbs DH, Berchtold NC, Perreau V, et al. Extensive innate immune gene activation accompanies brain aging, increasing vulnerability to cognitive decline and neurodegeneration: a microarray study. *J Neuroinflammation* 2012; 9: 179.
- 221 Shi J, Johansson J, Woodling NS, Wang Q, Montine TJ, Andreasson K. The prostaglandin E2 E-prostanoid 4 receptor exerts anti-inflammatory effects in brain innate immunity. *J Immunol* 2010; 184: 7207–18.
- 222 Tang EHC, Libby P, Vanhoutte PM, Xu A. Anti-inflammation therapy by activation of prostaglandin EP4 receptor in cardiovascular and other inflammatory diseases. *J Cardiovasc Pharmacol* 2012; 59: 116–23.
- 223 Brenneis C, Coste O, Altenrath K, et al. Anti-inflammatory role of microsomal prostaglandin E synthase-1 in a model of neuroinflammation. *J Biol Chem* 2011; 286: 2331–42.
- 224 Bentham P, Gray R, Sellwood E, Hills R, Crome P, Raftery J, and the AD2000 Collaborative Group. Aspirin in Alzheimer's disease (AD2000): a randomised open-label trial. *Lancet Neurol* 2008; 7: 41–49.
- 225 Wilcock DM, Zhao Q, Morgan D, et al. Diverse inflammatory responses in transgenic mouse models of Alzheimer's disease and the effect of immunotherapy on these responses. ASN Neuro 2011; 3: 249–58.
- 226 Jimenez S, Baglietto-Vargas D, Caballero C, et al. Inflammatory response in the hippocampus of PS1M146L/APP751SL mouse model of Alzheimer's disease: age-dependent switch in the microglial phenotype from alternative to classic. J Neurosci 2008; 28: 11650–61.

- 227 Morgan D, Gordon MN, Tan J, Wilcock D, Rojiani AM. Dynamic complexity of the microglial activation response in transgenic models of amyloid deposition: implications for Alzheimer therapeutics. J Neuropathol Exp Neurol 2005; 64: 743–53.
- 228 Zotova E, Bharambe V, Cheaveau M, et al. Inflammatory components in human Alzheimer's disease and after active amyloid-β42 immunization. Brain 2013; 136: 2677–96.
- 229 Boche D, Zotova E, Weller RO, et al. Consequence of Aβ immunization on the vasculature of human Alzheimer's disease brain. *Brain* 2008; 131: 3299–310.
- 230 Wilcock DM, Rojiani A, Rosenthal A, et al. Passive immunotherapy against Aβ in aged APP-transgenic mice reverses cognitive deficits and depletes parenchymal amyloid deposits in spite of increased vascular amyloid and microhemorrhage. J Neuroinflammation 2004; 1: 24.
- 231 Sperling R, Salloway S, Brooks DJ, et al. Amyloid-related imaging abnormalities in patients with Alzheimer's disease treated with bapineuzumab: a retrospective analysis. *Lancet Neurol* 2012; 11: 241–49.
- 232 Wilcock DM, Alamed J, Gottschall PE, et al. Deglycosylated antiamyloid-β antibodies eliminate cognitive deficits and reduce parenchymal amyloid with minimal vascular consequences in aged amyloid precursor protein transgenic mice. J Neurosci 2006; 26: 5340–46.