# CompMS2miner\_Workflow

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Matches MS1 features to MS2 spectra (.mzXML) files based on a mass-to-charge and retention time tolerance. Composite spectra and other data can subsequently be visualized during any stage of the CompMS2miner processing workflow. Composite spectra can be denoised, ion signals grouped and summed, substructure groups identified, common Phase II metabolites predicted, calculation of a correlation network and features matched to data bases monoisotopic mass data and insilico MS2 fragmentation data. The resulting data can then be readily curated by publishing as online shiny application on shinyapps.io, as a zip file which can be shared or by sending to a local or online couchDB database.

At any stage during the compMS2 workflow a compMS2 class object can be visualized with a Shiny application compMS2explorer. Full usage of the compMS2explorer application functionality requires an internet connection. The end result of following the workflow within this document using the example data provided can be visualized using the function, comments made in the metID comments tab can be saved on closing the application by assigning the compMS2explorer output to an object:

```
library(CompMS2miner)
# assign any metabolite identification comments to a new "CompMS2" object
compMS2example_commented <- compMS2explorer(compMS2example)</pre>
```

The following example illustrates the CompMS2miner workflow:

### 1. construct compMS2 class object.

From MS2 data (in the .mzXML file format) and an MS1feature table. The compMS2 object can also be constructed in as a parallel computation in the case of large peak tables and/ or larger numbers of MS2 data files.

```
# file path example MS1features in comma delimited csv file
# (see ?example_mzXML_MS1features for details).
MS1features_example <- system.file("extdata", "MS1features_example.csv",
                                    package = "CompMS2miner")
# mzXml file examples directory
mzXmlDir_example <- dirname(MS1features_example)</pre>
# observation MS1 feature table column names character vector for corrNetwork function
obsNames <- c(paste0(rep("ACN 80", 6), rep(LETTERS[1:3], each=2), rep(1:2, 3)),
              paste0(rep("MeOH_80_", 6), rep(LETTERS[1:3], each=2), rep(1:2, 3)))
# use parallel package to detect number of cores
nCores <- parallel::detectCores()</pre>
# read in example peakTable
peakTable <- read.csv(MS1features_example, header=TRUE, stringsAsFactors=FALSE)</pre>
# create compMS2 object
compMS2demo <- compMS2(MS1features = peakTable,</pre>
                       mzXMLdir = mzXmlDir_example, nCores=nCores,
                       mode = "pos", precursorPpm = 10, ret = 10,
                       TICfilter = 10000)
## creating compMS2 object in positive ionisation mode
```

```
## Loading required package: foreach
## Loading required package: Rcpp
## Loading required package: shiny
## 2 MS2 (.mzXML) files were detected within the directory...
## Starting SNOW cluster with 8 local sockets...
## matching MS1 peak table features to the following MS2 files:
## DDA_ACN_80.mzXML
## DDA_MeOH_80.mzXML
##
## "obsNames" in argsCorrNetwork argument missing. Not performing correlation network calculation.
# View summary of compMS2 class object at any time
compMS2demo
## A "CompMS2" class object derived from 2 MS2 files
## 163 MS1 features were matched to 1784 MS2 precursor scans
## containing 220161 ion features
## Average ppm match accuracy: 1.77
## with a ppm mass accuracy tolerance of (+/-) 10
##
## Average retention time match accuracy: 3.93 seconds
## with a retention time tolerance of (+/-) 10 seconds
## Memory usage: 4.57 MB
```

### 2. Dynamic noise filtration.

filter variable noise from the data using a dynamic noise filter.

```
# dynamic noise filter
compMS2demo <- deconvNoise(compMS2demo, "DNF")</pre>
## Applying dynamic noise filter to 257 composite spectra...
## Starting SNOW cluster with 8 local sockets...
## 256 composite spectra contained more than or equal to 5 peaks following dynamic noise filtration
# View summary of compMS2 class object at any time
compMS2demo
## A "CompMS2" class object derived from 2 MS2 files
## 163 MS1 features were matched to 1782 MS2 precursor scans
## containing 5030 ion features
##
## Average ppm match accuracy: 1.775
## with a ppm mass accuracy tolerance of (+/-) 10
## Average retention time match accuracy: 3.94 seconds
## with a retention time tolerance of (+/-) 10 seconds
## Memory usage: 1.33 MB
```

# 3. Intra-spectrum ion grouping and inter-MS2 file spectra grouping with signal summing.

group and sum ions from different scans and then combine summed ion composite spectra across multiple files. This create a single composite spectra for each MS1 EIC matched to MS2 precursor scans.

```
# intra-spectrum ion grouping and signal summing
compMS2demo <- combineMS2(compMS2demo, "Ions")</pre>
## Grouping ions in 256 composite spectra...
## Starting SNOW cluster with 8 local sockets...
## ...done
## 250 composite spectra contained more than or equal to 5 peaks following ion grouping
## The range of interfragment differences less than 0.1~\mathrm{m/z} in the composite spectra is min : 1~\mathrm{max} : 1~\mathrm{max}
## The average number of interfragment differences less than 0.1 m/z in the composite spectra is 5
## Warning in .local(object, ...): The average number of interfragment
## differences less than 0.1 m/z is greater than 2: please consider increasing
## the mzError parameter above 0.001
# View summary of compMS2 class object at any time
compMS2demo
## A "CompMS2" class object derived from 2 MS2 files
## 162 MS1 features were matched to 1738 MS2 precursor scans
## containing 2820 ion features
## Average ppm match accuracy: 1.789
## with a ppm mass accuracy tolerance of (+/-) 10
## Average retention time match accuracy: 3.93 seconds
## with a retention time tolerance of (+/-) 10 seconds
##
## Memory usage: 1.64 MB
#inter spectrum ion grouping and signal summing
compMS2demo <- combineMS2(compMS2demo, "Spectra")</pre>
## Combining 250 composite spectra by MS1 feature number...
## Starting SNOW cluster with 8 local sockets...
## ...done
## 162 composite spectra contained more than or equal to 5 peaks following ion grouping
## The range of interfragment differences less than 0.1 m/z in the composite spectra is min : 1 max : 1
## The average number of interfragment differences less than 0.1 m/z in the composite spectra is 6
## Warning in .local(object, ...): The average number of interfragment
## differences less than 0.1 m/z is greater than 2: please consider increasing
## the mzError parameter above 0.001
# View summary of compMS2 class object at any time
compMS2demo
## A "CompMS2" class object derived from 2 MS2 files
## 162 MS1 features were matched to 1738 MS2 precursor scans
## containing 2304 ion features
## Average ppm match accuracy: 1.817
## with a ppm mass accuracy tolerance of (+/-) 10
## Average retention time match accuracy: 3.77 seconds
## with a retention time tolerance of (+/-) 10 seconds
```

```
##
## Memory usage: 1.57 MB
```

### 4. Possible substructure identification.

Characteristic neutral losses/ fragments of electrospray adducts and metabolites from literature sources (?Substructure\_masses for details).

```
# annotate substructures
compMS2demo <- subStructure(compMS2demo, "Annotate")</pre>
## matching Precursor to fragment and interfragment neutral losses and fragments in 162 composite spect.
# identify most probable substructure annotation based on total relative intensity
# explained
compMS2demo <- subStructure(compMS2demo, "prob")</pre>
## Identifying likely substructure type...
# summary of most probable substructure annotation
mostProbSubStr <- subStructure(compMS2demo, "probSummary")</pre>
## Substructure annotation summary :
## 162 composite spectra
## 151 substructures identified above minimum sum relative intensity of 30
## SubStr.table
##
                phosphatidylcholine
                                                methionine side chain
##
                        polysiloxane
##
                                                             phthalate
##
## nitroaromatics, hydroxyaldehydes
                                                         nitrotoluenes
##
##
                           triazines
##
```

### 5. Metabolite identification methods.

A variety of metabolite identification methods are implemented through the function metID (see ?metID) for further details.

```
# annotate composite MS2 matched MS1 features to metabolomic databases (default
#is HMDB, also DrugBank, T3DB and ReSpect databases can also be queried).
#Warning: this may take 2-3 mins as large number of query masses
compMS2demo <- metID(compMS2demo, "dbAnnotate")
## matching 162 unknowns to 8,902,538 possible ESI artefacts and substructure mass shifts...
# select most probable annotations based on substructures detected
compMS2demo <- metID(compMS2demo, "dbProb")

# predict Phase II metabolites from SMILES codes
compMS2demo <- metID(compMS2demo, "predSMILES")

# metFrag insilico fragmentation.
compMS2demo <- metID(compMS2demo, "metFrag")</pre>
```

MetMSLine data-preprocessing and correlation network calculation.

```
### data pre-processing MS1features with MetMSLine package ###
if(!require(MetMSLine)){
 # if not installed then install from github
 devtools::install_github('WMBEdmands/MetMSLine')
 require(MetMSLine)
}
## Loading required package: MetMSLine
## Loading required package: tcltk2
## Loading required package: tcltk
##
## Attaching package: 'tcltk2'
## The following object is masked from 'package:shiny':
##
##
     tag
## Loading required package: zoo
## Attaching package: 'zoo'
## The following objects are masked from 'package:base':
##
##
     as.Date, as.Date.numeric
## Loading required package: fpc
## Loading required package: fastcluster
## Attaching package: 'fastcluster'
## The following object is masked from 'package:stats':
##
##
     hclust
# zero fill
peakTable <- zeroFill(peakTable, obsNames)</pre>
## zero filling with half the minimum non-zero value
# log transform
peakTable <- logTrans(peakTable, obsNames)</pre>
## log transforming to the base 2.718...
# add correlation network using pre-processed MS2 matched peak table
compMS2demo <- metID(compMS2demo, method='corrNetwork', peakTable, obsNames,</pre>
                 corrMethod='pearson', corrThresh=0.95, MTC='none', MS2only=3)
## Loading required package: igraph
## Attaching package: 'igraph'
## The following objects are masked from 'package:stats':
##
##
     decompose, spectrum
```

```
## The following object is masked from 'package:base':
##
## union
## Calculating correlation matrix for 162 features
## less than 300 nodes using Fruchterman-Reingold layout. see ?igraph::with_fr()
## 124 nodes with 446 edges identified at a correlation threshold >= 0.95 (pearson, p/q value <= 0.05, 1</pre>
```

## 6. Optional curate data in CouchDB/ publish results as a shiny App.

CompMS2 class objects can be sent to either a local or online couchDB database. Furthermore, the results of the CompMS2miner workflow can be published on the web as a shiny application to share metabolite identification information using the publishApp function.

```
# publish your app to shinyapps.io see ?publishApp for more details
# you may need to install the rsconnect and shinyapps packages and also sign up for a shinyapps.io acco
# quick guide here for setting up your account: http://shiny.rstudio.com/articles/shinyapps.html
publishApp(compMS2demo, appName='compMS2demo')
```