statTarget:

Statistical Analysis of Metabolite Profile

Hemi Luan

September 5, 2016

Contents

1 Background		kground	3
	1.1	System requirements	3
	1.2	Opening the GUI	3
2	GU	I overview	4
	2.1	What does statTarget offer statistically	4
	2.2	Primary functions of statTarget	4
3	Running Shift Correction from the GUI		6
4	Running Statistical Analysis from the GUI		7
5	Inve	estigating the results	9

6	References	9
7	Session Info	10

1 Background

Quality Control (QC) has been considered as an essential step in the metabolomics platform for high reproducibility and accuracy of data. The secondary use of the same QC samples is more and more acceptable for correcting the signal drift during the sequence of MS run order, especially benefiting to improve the quality of data in multi-block experiments of large-scale metabolomic study. statTarget is easy use tool to provide a graphical user interface for quality control based shift signal correction, integration of metabolomic data from multi-batch experiments, and the comprehensive statistic analysis in non-targeted or targeted metabolomics. This document is intended to guide the user to use statTarget-GUI to perform metabolomic data analysis. Note that this document will not describe the inner workings of statTarget algorithm.

1.1 System requirements

Dependent on R (>= 3.3.0)

1.2 Opening the GUI

Download it from https://www.xquartz.org.

Load the package with biocLite():
 source("https://bioconductor.org/biocLite.R")
 biocLite("statTarget")
 **For mac PC, the package "statTargetGUI" requires X11 support (XQuartz).

2 GUI overview

An easy to use tool provide a graphical user interface (Figure 1) for quality control based signal correction, integration of metabolomic data from multiple batches, and the comprehensive statistic analysis for non-targeted and targeted approaches. (URL: https://github.com/13479776/statTarget)

2.1 What does statTarget offer statistically

The main GUI of statTarget has two basic components. The first components is shift correction. It includes quality control-based robust LOESS signal correction (QC-RLSC) that is a widely accepted method for quality control based signal correction and integration of metabolomic data from multiple analytical batches (Dunn WB., et al. 2011; Luan H., et al. 2015). The second components is Statistical Analysis. It provides comprehensively computational and statistical methods that are commonly applied to analyze metabolomics data, and offers multiple results for biomarker discovery.

2.2 Primary functions of statTarget

Function 1 - Shift Correction provide QC-RLSC algorithm that fit the QC data, and each metabolites in the true sample will be normalized to the QC sample. To avoid overfitting of the observed data, LOESS based generalised cross-validation (GCV) would be automatically applied, when the QCspan was set at 0.

Function 2 - Statistical Analysis provide features including Data pre-

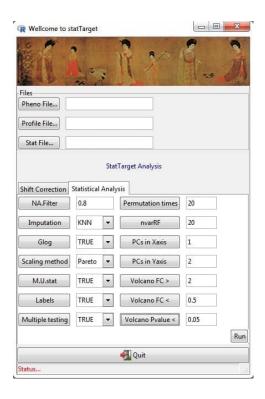


Figure 1: The main GUI of statTarget

processing, Data descriptions, Multivariate statistics analysis and Univariate analysis.

Data preprocessing: 80-precent rule, log transformation, KNN imputation, Median imputation and Minimum values imputation.

Data descriptions : Mean value, Median value, Sum, Quartile, Standard derivatives, etc.

Multivariate statistics analysis: PCA, PLSDA, VIP, Random forest.

 $\label{thm:constraint} \mbox{Univariate analysis}: \mbox{Student T-test}, \mbox{Shapiro-Wilk normality test and Mann-Whitney test}.$

Biomarkers analysis: ROC, Odd ratio.

3 Running Shift Correction from the GUI

Pheno File

Meta information includes the Sample name, class, batch and order. Do not change the name of each column. (a) Class: The QC should be labeled as NA. (b) Order: Injection sequence. (c) Batch: The analysis blocks or batches with ordinal number, e.g., 1,2,3,.... (d) Sample name should be consistent in Pheno file and Profile file. (See the example data)

Profile File

Expression data includes the sample name and expression data. (See the example data)

NA.Filter

NA.Filter: Removing peaks with more than 80 percent of missing values (NA or 0) in each group. (Default: 0.8)

QCspan

The smoothing parameter which controls the bias-variance tradeoff. The common range of QCspan value is from 0.2 to 0.75. If you choose a span that is too small then there will be a large variance. If the span is too large, a large bias will be produced. The default value of QCspan is set at '0', the generalised cross-validation will be performed for choosing a good value, avoiding overfitting the observed data. (Default: 0)

degree

Lets you specify local constant regression (i.e., the Nadaraya-Watson estimator, degree=0), local linear regression (degree=1), or local polynomial fits (degree=2). (Default: 2)

Imputation

Imputation: The parameter for imputation method. (i.e., nearest neighbor averaging, "KNN"; minimum values for imputed variables, "min"; median values for imputed variables (Group dependent) "median". (Default: KNN)

4 Running Statistical Analysis from the GUI

Stat File

Expression data includes the sample name, group, and expression data.

NA.Filter

Removing peaks with more than 80 percent of missing values (NA or 0) in each group. (Default: 0.8)

Imputation

The parameter for imputation method. (i.e., nearest neighbor averaging, "KNN"; minimum values for imputed variables, "min"; median values for imputed variables (Group dependent) "median". (Default: KNN)

Glog

Generalised logarithm (glog) transformation for Variance stabilization (Default: TRUE)

Scaling Method

Scaling method before statistic analysis (PCA or PLS). Pareto can be used for specifying the Pareto scaling. Auto can be used for specifying the Auto scaling (or unit variance scaling). Vast can be used for specifying the vast scaling. Range can be used for specifying the Range scaling. (Default: Pareto)

M.U.Stat

Multiple statistical analysis and univariate analysis (Default: TRUE)

Permutation times

The number of random permutation times for PLS-DA model (Default: 20)

PCs

PCs in the Xaxis or Yaxis: Principal components in PCA-PLS model for the x or y-axis (Default: 1 and 2)

nvarRF

The number of variables in Gini plot of Randomforest model (=< 100).

(Default: 20)

Labels

To show the name of sample in the Score plot. (Default: TRUE)

Multiple testing

This multiple testing correction via false discovery rate (FDR) estimation with Benjamini-Hochberg method. The false discovery rate for conceptualizing the rate of type I errors in null hypothesis testing when conducting multiple comparisons. (Default: TRUE)

Volcano FC

The up or down -regulated metabolites using Fold Changes cut off values in the Volcano plot. (Default: > 2 or < 1.5)

Volcano Pvalue

The significance level for metabolites in the Volcano plot. (Default: 0.05)

5 Investigating the results

Once data files have been analysed it is time to investigate them. Please get this info. through the GitHub page. (URL: https://github.com/13479776/statTarget)

6 References

Dunn, W.B., et al., Procedures for large-scale metabolic profiling of serum and plasma using gas chromatography and liquid chromatography coupled to mass

spectrometry. Nature Protocols 2011, 6, 1060.

Luan H., LC-MS-Based Urinary Metabolite Signatures in Idiopathic Parkinson's Disease. J Proteome Res., 2015, 14,467.

Luan H., Non-targeted metabolomics and lipidomics LC-MS data from maternal plasma of 180 healthy pregnant women. GigaScience 2015 4:16

7 Session Info

- R version 3.3.0 (2016-05-03), x86_64-apple-darwin13.4.0
- Locale:
 en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
- Base packages: base, datasets, graphics, grDevices, methods, stats, utils
- Other packages: devtools 1.11.1, digest 0.6.9, gWidgets2 1.0-7,
 gWidgets2RGtk2 1.0-5, memoise 1.0.0, RGtk2 2.20.31, statTarget 1.2.6
- Loaded via a namespace (and not attached): BiocInstaller 1.22.3, cluster 2.0.4, DEoptimR 1.0-4, grid 3.3.0, impute 1.46.0, lattice 0.20-33, magrittr 1.5, mvtnorm 1.0-5, pcaPP 1.9-60, pdist 1.2, pls 2.5-0, plyr 1.8.3, pROC 1.8, randomForest 4.6-12, Rcpp 0.12.4, robustbase 0.92-6, roxygen2 5.0.1, rrcov 1.3-11, stats4 3.3.0, stringi 1.0-1, stringr 1.0.0, tools 3.3.0, withr 1.0.1