

## OptForce Tutorial

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### INTRODUCTION:

In this tutorial we will run optForce. For a detailed description of the procedure, please see [1]. Briefly, the problem is to find a set of interventions of size " $K$ " such that when these interventions are applied to a wild-type strain, the mutant created will produce a particular target of interest in a higher rate than the wild-type strain. The interventions could be knockouts (lead to zero the flux for a particular reaction), upregulations (increase the flux for a particular reaction) and downregulations (decrease the flux for a particular reaction).

For example, imagine that we would like to increase the production of succinate in *Escherichia coli*. Which are the interventions needed to increase the production of succinate? We will approach this problem in this tutorial and we will see how each of the steps of OptForce are solved.

### MATERIALS

#### EQUIPMENT

1. MATLAB
2. GAMS
3. A solver for Mixed Integer Linear Programming (MILP) problems. For example, Gurobi.

#### EQUIPMENT SETUP

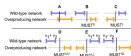
**GAMS:** Install the latest version of GAMS available. This is important as some GAMS functions are not supported in versions older than 24.7.1. During GAMS installation make sure to select the option "Add GAMS directory to PATH environment variable". In MATLAB, use functions `addpath()` and `savepath()` for adding and saving the directory where GAMS was installed. For a more detailed description of GAMS installation and setup, you can watch the official tutorial of GCOMPS at [https://www.youtube.com/watch?v=wPn\\_g9t8dE](https://www.youtube.com/watch?v=wPn_g9t8dE)

#### PROCEDURE

The procedure consists on the following steps:

- 1) Maximize specific growth rate and product formation.
- 2) Define constraints for both wild-type and mutant strain.
- 3) Perform flux variability analysis for both wild-type and mutant strain.
- 4) Find must sets, i.e. reactions that MUST increase or decrease their flux in order to achieve the phenotype in the mutant strain.

Figure 1.



3) Find the interventions needed that will ensure a increased production of the target of interest

Now, we will approach each step in detail.

#### STEP 1: Maximize specific growth rate and product formation

First, we load the model. This model comprises only 90 reactions, which describe the central metabolism of *E. coli* [2].

Then, we change the objective function to maximize biomass ("RTN"). We also change the lower bounds, so *E. coli* will be able to consume glucose, oxygen, sulfate, ammonium, citrate and glycerol.

```
global TUTORIAL_INIT_CB
if ~isempty(TUTORIAL_INIT_CB) && TUTORIAL_INIT_CB==0
    initializeData
    changeObjective('growth','all')
end

getTutorials = which('tutorial_optForce*.m');
getPath = fileparts(getTutorials);
cd(getPath)

modelName = 'bottom_wat';
modelDirectory = getcwd; % Get the full path for the distributed model.
modelPath = [modelDirectory filesep modelName]; % Get the full path. Necessary to be sure, that the right model is loaded
model = readModel(modelPath);

model.LC(1:ncsp(model.FMT,'RTN')) = 1;
model.L = changeFMO(model.L, 'RR_glc', -200, '1');
model.L = changeFMO(model.L, 'RR_o2', -200, '1');
model.L = changeFMO(model.L, 'RR_sul', -100, '1');
model.L = changeFMO(model.L, 'RR_cit', -100, '1');
model.L = changeFMO(model.L, 'RR_glyc', -100, '1');
model.L = changeFMO(model.L, 'RR_glyc', -200, '1');
```

Then, we calculate the maximum specific growth rate and the maximum production rate for succinate

```
growthRate = opt.is124CModel(model);
tryPrint("The maximum growth rate is %f", growthRate.f());
```

The maximum growth rate is 14.38

```
model = changeObjective(model, "R2_cac");
maxsucc = opt.is124CModel(model);
tryPrint("The maximum production rate of succinate is %f", maxsucc.f());
```

The maximum production rate of succinate is 235.56

**TP:** The biomass reaction is usually set to 1%-10% of maximum theoretical biomass yield when running the following steps, to prevent solutions with no biomass formation

1. maximizing product formation
2. finding MUST sets of second order
3. finding FORCE sets

## STEP 2: Define constraints for both wild-type and mutant strain

**TBWD:** This step should take a few days or weeks, depending on the information available for your species.

**CRITICAL STEP:** This is a manual task, so you should search for information in articles or even perform your own experiments. You can also make assumptions for describing the phenotypes of both strains which will make this task a little faster but make sure to have two strains different enough, because you should be able to find differences in reactions ranges.

First, we load the model. This model comprises only 90 reactions, which describe the central metabolism of *E. coli* [2].

Then, we change the objective function to maximize biomass ("R75"). We also change the lower bounds, so *E. coli* will be able to consume glucose, oxygen, sulfate, ammonium, citrate and glycerol.

We define constraints for each strain

```
Constr_WT = struct("rowList", {{ "R75" }}, "rowValues", 10, "rowBoundType", "b");
Constr_RT = struct("rowList", {{ "R75", "R2_cac" }}, "rowValues", [0, 235.56], "rowBoundType", "lb");
```

## Step 3: Flux Variability Analysis

**TBWD:** This task should take from a few seconds to a few hours depending on the size of your reconstruction

We run the FVA analysis for both strains

```
[sInfFluxes_WT, sMaxFluxes_WT, sInfFluxes_RT, sMaxFluxes_RT, ~, ~] = FVaOptForce(model, Constr_WT, Constr_RT);
disp([sInfFluxes_WT, sMaxFluxes_WT, sInfFluxes_RT, sMaxFluxes_RT]);
```

-98.1251	97.1388	61.4313	108.8888
0	88.8788	61.4371	108.8888
0	88.8788	61.4371	108.8888
-98.1387	88.8788	-61.4388	11.1243
21.3831	103.1388	15.1588	111.1243
-3.8777	116.8888	15.1588	111.1243
0	115.1888	0	15.1823
0	187.1011	0	15.1887
0	108.1381	0	0.8537
-98.8888	182.9888	0	0.8537
18.8888	88.1754	0	0.8881
-98.8888	182.9888	0	0.8537
-88.8881	7.1888	-0.8881	0
-61.9981	2.1888	-0.8881	0
-61.9981	2.1888	-0.8881	0
-2.1888	11.9981	0	0.8881
0	88.8788	0	15.1823
0	88.8788	15.1588	15.1823
9.7828	116.8888	15.1588	15.1823
0	16.1881	15.1588	15.1871
16.8281	161.1888	115.1588	115.1881
16.8281	161.1888	115.1588	115.1881
6.8181	118.1128	115.1588	115.1881
-3.8736	123.1888	115.1588	115.1881
0	118.8576	0	0.8881
1.7888	123.1888	0	0.8881
-98.1188	123.1888	-15.1823	0.8881
0	115.1888	0	15.1823
0	115.1888	0	15.1823
0	116.1888	15.1588	117.1871
0	213.1881	0	122.1371
-7.1888	98.8888	0	0.8537
0	81.8887	88.8788	88.8888
0	81.8887	88.8788	88.8888
0	175.1881	188.1888	188.8888
0	175.1881	188.1888	188.8888
91.4138	187.1288	0	0
5.4188	8.4188	0	0
2.8188	2.8188	0	0
3.8188	3.8188	0	0
21.4128	16.8828	0	0
3.2888	3.2888	0	0
6.8128	6.8128	0	0
0	15.7188	0	0
-6.8888	8.8278	0	0
8.8788	16.1888	0	0
0	15.4188	0	0
3.2828	3.2828	0	0
6.1888	6.1888	0	0
6.1888	6.1888	0	0
7.2188	18.8888	0	0
2.8188	2.8188	0	0
3.6288	3.6288	0	0
3.9128	3.9128	0	0
3.9888	3.9888	0	0
2.4888	2.4888	0	0
1.8388	1.8388	0	0
8.7188	8.7188	0	0
1.2888	1.2888	0	0
2.8188	2.8188	0	0
1.2888	1.2888	0	0
79.7121	188.8888	188.9188	188.8888
0	118.8576	0	0.8881
-98.1881	113.8121	-12.1588	15.1888
0	213.1881	0	122.1371
88.8281	188.8888	98.9871	188.8888
11.8888	188.8888	98.9871	188.8888
-188.8888	81.8887	-188.8888	-98.9188
0	175.1881	188.1888	188.8888
0	181.8888	0	0.8537
116.9718	487.1271	111.1888	111.1187
62.1287	188.8888	98.9788	188.8888
97.8828	97.8828	0	0
3.2828	3.2828	0	0
16.8888	16.8888	0	0
0	175.1881	188.1888	188.8888
116.9718	487.1271	111.1888	111.1187
0	181.8888	0	0.8537
0	213.1881	0	122.1371
-188.8888	-18.8288	-188.8888	-98.9871
-188.8888	-15.8888	-188.8888	-98.9871
-188.8888	16.8887	-188.8888	-98.9188
-87.8828	-87.8828	0	0
-188.8888	-62.1287	-188.8888	-98.9788
-3.2828	-3.2828	0	0
0	181.8288	115.1588	115.1588
0	181.8288	115.1588	115.1588
11.8288	11.8288	0	0
3.8188	3.8188	0	0
3.9128	3.9128	0	0

Now, the run the second step of OptForce.

Step 4: Find Must Sets

**TIMING:** This task should take from a few seconds to a few hours depending on the size of your reconstruction

First, we define an ID for this run. Each time you run the functions associated to the optForce procedure, some folders can be generated to store inputs used in that run. Outputs are stored as well. These folders will be located inside the folder defined by your run ID. Thus, if your runID is "TestOptForce", the structure of the folders will be the following:

```
└─ CurrentFolder
  │ └─ TestOptForce
  │   │
  │   └─ Inputs
  │       │
  │       └─ Outputs
```

To avoid the generation of inputs and outputs folders, set keepInputs = 0, printExcel = 0, printText = 0 and keepGameOutputs = 0

Also, a report of the run is generated each time you run the functions associated to the optForce procedure. So, the idea is to give a different runID each time you run the functions, so you will be able to see the report (inputs used, outputs generated, errors in the run) for each run.

We define then our runID

```
runID = "TestOptForce";
```

For now, only functions to find first and second order must sets are supported. As depicted in **Figure 1**, the first order must sets are MUSTU and MUSTL, and second order must sets are MUSTLU, MUSTLL and MUSTUL.

#### A) Finding first order must sets

We define constraints

```
constraintOpt = struct('runID', {( 'EX_gluco', 'XTS', 'EX_suc' )}, 'values', [-100, 0, 100.5]);
```

We then run the functions findMustWithGAMS.m and findMustWithGAMS.m that will find mustU and mustL sets, respectively.

Important: To run these functions you will need a solver able to solve Mixed Integer Linear Programming (MILP or MIP) problems. Some popular options are: cplex and gurobi. You can see which games solvers are available in your systems to solve MIP problems by running checkGAMSsolvers('MIP')

```
saSolve = checkGAMSsolvers('MIP'); disp(saSolve);
```

Columns 1 through 12

'AMPL' 'BARON' 'BIRLP' 'BIRSP' 'CBC' 'CONVERT' 'CPLEX' 'EAGLE' 'EAGLE2' 'GUROBI' 'KISTREL' 'LINGO'

Columns 13 through 22

'LINGOGLOBAL' 'LINGO2' 'LOCALSOLVER' 'MOSEK' 'OPTICOMP' 'OptiCplex' 'OptiGurobi' 'OptiMosek' 'OptiMip' 'XPRESS'

Columns 23 through 26

'SCIP' 'SA' 'XPRESS'

We then run findMustWithGAMS.m and findMustWithGAMS.m.

#### i) MustL Set:

```
[mustLSet, pos_mustL] = findMustWithGAMS(model, solveMip_WT, solveMip_WT, 'constraintOpt', constraintOpt, ...  
'solveMip_WT', 'cplex', 'runID', runID, 'outputFolder', 'outputsFindMustL', 'outputFilebase', 'MustL', ...  
'printExcel', 1, 'printText', 1, 'printReport', 1, 'keepInputs', 1, 'keepGameOutputs', 1, 'verbose', 0);
```

Note that the folder "TestOptForce" was created. Inside this folder, two additional folders were created: "InputsMustL" and "OutputsFindMustL". In the inputs folder you will find all the inputs required to run the GAMS function "findMustL.gms". Additionally, in the outputs folder you will find the mustL set found, which were saved in two files (.xls and .txt), as well as other files generated automatically by GAMS. Furthermore, a report which summarize all the inputs and outputs used during your running was generated. The name of the report will be in this format "report-Day-Month-Year-Hour-Minutes". So, you can maintain a chronological order of your experiments

We display the reactions that belongs to the mustL set

```
disp(mustLSet)
```

```

'K11'
'K26'
'K37'
'K38'
'K39'
'K48'
'K51'
'K62'
'K63'
'K64'
'K65'
'K66'
'K67'
'K68'
'K69'
'K70'
'K71'
'K72'
'P0000pvc_1'
'P0000pvc_1'
'P0000pvc_1'

```

#### j) MustU set

```

[mustUset, gsk_outU] = findFUNCTIONS(model, slistFuncs_WT, slistFuncs_WT, 'constUopt', constUopt, ...
    'solveName', 'cplex', 'runID', 'outputFolder', 'outputFolderU', 'outputFolderU', 'MustU', ...
    'printBase', 1, 'printText', 1, 'printReport', 1, 'keepInputs', 1, 'keepOutputs', 1, 'verbose', 0);

```

Note that the folders "InputMustU" and "OutputFindMustU" were created. These folders contain the inputs and outputs of findMustUWithGAMS.m, respectively.

We display the reactions that belongs to the mustU set

```

disp([mustUset])

'K21'
'K22'
'K23'
'K24'
'K33'
'K34'
'K35'
'K36'
'K6'
'EX_glu'
'EX_ohf'
'EX_oxf'
'EX_oxf'
'EXC1'

```

#### k) Finding second order must sets

First, we define the reactions that will be excluded from the analysis. It is suggested to eliminate reactions found in the previous step as well as exchange reactions

```

constUopt = strcat('runID', {'EX_glu', 'K75', 'EX_oxf'}), 'values', [-100, 0, 100.5]);
exchangeFunc = model.runIDwithFunctionID, strcat(model.runID, 'EX_') == 0);
excludedReac = unique([mustUset; mustUset; exchangeFunc]);
mustUPL1Order = unique([mustUset; mustUset]);

```

Now, we run the functions for finding second order must sets

#### j) MustU2:

```

[mustU2, gsk_outU2, mustU2_linear, gsk_outU2_linear] = findFUNCTIONS(model, slistFuncs_WT, slistFuncs_WT, ...
    'constUopt', constUopt, 'excludeReac', excludedReac, 'mustUPL1Order', mustUPL1Order, 'solveName', 'cplex', ...
    'runID', runID, 'outputFolder', 'outputFolderU2', 'outputFolderU2', 'MustU2', 'printBase', 1, 'printText', 1, ...
    'printReport', 1, 'keepInputs', 1, 'keepOutputs', 1, 'verbose', 0);

```

Note that the folders "InputMustU2" and "OutputFindMustU2" were created. These folders contain the inputs and outputs of findMustU2WithGAMS.m, respectively.

We display the reactions that belongs to the mustU2 set

```

disp([mustU2])

'K36' 'K63'
'K63' 'K11'

```

#### k) MustL:

```
[musTLL, get_musTLL, musTLL_linear, get_musTLL_linear] = findMusTLLWithGAMS(m, s, sspNames_MT, sspNames_MT, ...
    'constraint', constraint, 'excludedReactions', excludedReactions, 'musTLLFirstOrder', 'musTLLFirstOrder', 'solverName', 'cplex', ...
    'runID', runID, 'outputFolder', 'outputFolderMusTLL', 'outputFileName', 'MusTLL', 'printHeader', 1, 'printTest', 1, ...
    'printReport', 1, 'keepInputs', 1, 'keepAllOutputs', 1, 'verbose', 0);
```

Note that the folders "InputMusTLL" and "OutputFindMusTLL" were created. These folders contain the inputs and outputs of findMusTLLWithGAMS.m, respectively. We display the reactions that belongs to the musTLL set. In this case, MusTLL is an empty array because no reaction was found in the musTLL set.

```
disp(musTLL);
```

**ii) MusTLL:**

```
[musTLL, get_musTLL, musTLL_linear, get_musTLL_linear] = findMusTLLWithGAMS(m, s, sspNames_MT, sspNames_MT, ...
    'constraint', constraint, 'excludedReactions', excludedReactions, 'musTLLFirstOrder', 'musTLLFirstOrder', 'solverName', 'cplex', ...
    'runID', runID, 'outputFolder', 'outputFolderMusTLL', 'outputFileName', 'MusTLL', 'printHeader', 1, 'printTest', 1, ...
    'printReport', 1, 'keepInputs', 1, 'keepAllOutputs', 1, 'verbose', 0);
```

Note that the folders "InputMusTLL" and "OutputFindMusTLL" were created. These folders contain the inputs and outputs of findMusTLLWithGAMS.m, respectively. We display the reactions that belongs to the musTLL set. In this case, MusTLL is an empty array because no reaction was found in the musTLL set.

```
disp(musTLL);
```

**TROUBLESHOOTING 1:** "I didn't find any reaction in my must sets"

**TROUBLESHOOTING 2:** "I got an error when running the findMusTLLWithGAMS.m functions (X = L or U or LL or UL or UU depending on the case)"

**Step 5: OptForce**

**TIPING:** This task should take from a few seconds to a few hours depending on the size of your reconstruction

We define constraints and we define "K" the number of interventions allowed, "idlex" the maximum number of sets to find, and "targetReac" the reaction producing the metabolite of interest (in this case, succinate).

Additionally, we define the mustL set as the union of the reactions that must be up-regulated in both first and second order must sets, and must\_U set as the union of the reactions that must be down-regulated in both first and second order must sets

```
musLU = setUnion(musTLLset, musTUU);
musUL = setUnion(musTLLset, musTUL);
targetReac = 'SU_C';
K = 1;
nReacts = 1;
constraint = struct('ruleList', {{('RX_gluc', 'KX')}, 'values', [-200, 0]});

[gytReactions, gpyOptReactions, typeOptGytReactions] = gytForceWithGAMS(m, targetReac, musLU, musUL, sspNames_MT, ...
    sspNames_MT, sspNames_MT, sspNames_MT, 'K', K, 'sets', sets, 'constraint', constraint, ...
    'runID', runID, 'outputFolder', 'outputFolderOptForce', 'outputFileName', 'OptForce', 'solverName', 'cplex', 'printHeader', 1, 'printTest', 1, ...
    'printReport', 1, 'keepInputs', 1, 'keepAllOutputs', 1, 'verbose', 0);
```

Note that the folders "InputOptForce" and "OutputOptForce" were created. These folders contain the inputs and outputs of gytForceWithGAMS.m, respectively.

We display the reactions found by OptForce

```
disp(gytReactions)
```

```
'SU_C'
```

The reaction found was "SU\_C", i.e. a transporter for succinate (a very intuitive solution).

Next, we will increase "K" and we will exclude "SU\_C" from up-regulations to find non-intuitive solutions.

**TIP:** Sometimes the product is at the end of a long linear pathway. In that case, the recommendation is to also exclude most reactions on the linear pathway. Essential reactions and reactions not associated with any gene should also be excluded.

We will only search for the 20 best solutions, but you can try with a higher number.

We will change the runID to save both results (K = 1 and K = 2) in different folders

```
K = 2;
nReacts = 20;
runID = 'TestOptForce2';
excludedReac = struct('ruleList', {{('SU_C')}, 'typeOpt', 'U'});
[gytReactions, gpyOptReactions, typeOptGytReactions] = gytForceWithGAMS(m, targetReac, musLU, musUL, sspNames_MT, ...
    sspNames_MT, sspNames_MT, sspNames_MT, 'K', K, 'sets', sets, 'constraint', constraint, 'excludedReactions', excludedReac, ...
    'runID', runID, 'outputFolder', 'outputFolderOptForce', 'outputFileName', 'OptForce', 'solverName', 'cplex', 'printHeader', 1, 'printTest', 1, ...
    'printReport', 1, 'keepInputs', 1, 'keepAllOutputs', 1, 'verbose', 0);
```

Note that the folders "InputOptForce" and "OutputOptForce" were created inside TestOptForce2. These folders contain the inputs and outputs of gytForceWithGAMS.m, respectively.



## Acknowledgments

I would like to thank the research group of Costas D. Maranas who provided the GAMS functions to solve this example. In particular I would like to thank to Chiam Yu Ng who kindly provided examples for using GAMS and support for writing GAMS functions.

## References

- [1] Ranganathan S, Suthes PF, Maranas CD (2010) CydForce: An Optimization Procedure for Identifying All Genetic Manipulations Leading to Targeted Overproductions. *PLoS Computational Biology* 6(4): e1000764. <https://doi.org/10.1371/journal.pcbi.1000764>.
- [2] Madek R, Antoniewicz, David F. Kraynie, Lisa A. Laffend, Joanna-Gordziej-Legler, Joanne K. Keliher, Gregory Stephanopoulos, Metabolic flux analysis in a nonstationary system: Fed-batch fermentation of a high-yielding strain of *E. coli* producing 1,3-propanediol, *Metabolic Engineering*, Volume 9, Issue 3, May 2007, Pages 277-292, ISSN 1086-7178, <https://doi.org/10.1016/j.jmbs.2007.01.003>.