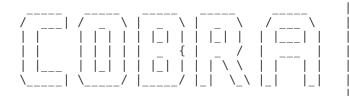
# Metabotools tutorial II - Integration of quantitative metabolomic data

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In this tutorial we ...

Clear workspace and initialize the COBRA Toolbox

```
clear
initCobraToolbox
```



COnstraint-Based Reconstruction and Analysis The COBRA Toolbox - 2017

Documentation:

http://opencobra.github.io/cobratoolbox

- > Checking if git is installed ... Done.
- > Checking if the repository is tracked using git ... Done.
- > Checking if curl is installed ... Done.
- > Checking if remote can be reached ... Done.
- > Initializing and updating submodules ... Done.
- > Adding all the files of The COBRA Toolbox ... Done.
- > Define CB map output... set to svg.
- > Retrieving models ... Done
- > TranslateSBML is installed and working properly.
- > Configuring solver environment variables ...
  - [----] ILOG\_CPLEX\_PATH: /opt/ibm/ILOG/CPLEX\_Studio1271/cplex/matlab/x86-64\_linux
  - [----] GUROBI PATH: /home/syarra/Dropbox/software/gurobi/gurobi652/linux64/matlab
  - [----] TOMLAB PATH: --> set this path manually after installing the solver ( see instructions )
  - [----] MOSEK\_PATH: /home/syarra/Dropbox/software/mosek/linux/8/

Done

- > Checking available solvers and solver interfaces ... Done.
- > Setting default solvers ... Done.
- > Saving the MATLAB path ... Done.
  - The MATLAB path was saved as ~/pathdef.m.
- > Summary of available solvers and solver interfaces

Support	LP MILP	QP	MIQP	NLP				
cplex_direct	full		0	0	0	0	-	
dqqMinos	full		1	-	-	-	-	
glpk	full		1	1	-	-	-	
gurobi	full		1	1	1	1	-	
ibm_cplex	full		1	1	1	-	-	
matlab	full		1	-	-	-	1	
mosek	full		1	1	1	-	-	
pdco	full		1	-	1	-	-	
quadMinos	full		1	-	-	-	1	
tomlab_cplex	full		0	0	0	0	-	
qpng	experiment	al	-	-	1	-	-	
tomlab_snopt	experiment	al	-	-	-	-	0	
gurobi_mex	legacy		0	0	0	0	-	
lindo_old	legacy		0	-	-	-	-	
lindo_legacy	legacy		0	-	-	-	-	
lp_solve	legacy		1	-	-	-	-	

```
opti
              legacy
Total
+ Legend: - = not applicable, 0 = solver not compatible or not installed, 1 = solver installed.
> You can solve LP problems using: 'dqqMinos' - 'glpk' - 'gurobi' - 'ibm_cplex' - 'matlab' - 'mosek' -
> You can solve MILP problems using: 'glpk' - 'gurobi' - 'ibm cplex' - 'mosek'
> You can solve QP problems using: 'gurobi' - 'ibm cplex' - 'mosek' - 'pdco' - 'qpng'
> You can solve MIQP problems using: 'gurobi'
> You can solve NLP problems using: 'matlab' - 'quadMinos'
> Checking for available updates ...
ssh: /usr/local/MATLAB/R2016a/bin/glnxa64/libcrypto.so.1.0.0: no version information available (required
ssh: /usr/local/MATLAB/R2016a/bin/glnxa64/libcrypto.so.1.0.0: no version information available (required
OpenSSL version mismatch. Built against 1000207f, you have 100010bf
fatal: Could not read from remote repository.
Please make sure you have the correct access rights
and the repository exists.
> You can update The COBRA Toolbox by running updateCobraToolbox() (from within MATLAB).
```

> The changes of The COBRA Toolbox could not be fetched. > There are 169 new commit(s) on <master> and

```
global CBTDIR
tol = 1e-6;
```

#### set and check solver

```
solver = 'gurobi'; % can be gurobi or tomlab cplex
solverQuant = 'ibm cplex';
outputPath = pwd; % output is saved to this location, can be the same as pathToCOBRA 'C: ...
```

#### set and check solver

```
solverOK = changeCobraSolver(solverQuant, 'LP');
 > IBM ILOG CPLEX interface added to MATLAB path.
if solverOK == 1
    display('The solverQuant is set.');
else
    error('The solverQuant is not set.')
end
```

The solverQuant is set.

```
solverOK = changeCobraSolver(solver, 'LP');
```

> Gurobi interface added to MATLAB path.

```
if solverOK == 1
   display('The LP solver is set.');
else
   error('The LP solver is not set.')
```

```
end

The LP solver is set.

solverOK = changeCobraSolver(solver, 'QP');

> Gurobi interface added to MATLAB path.

if solverOK == 1
     display('The QP solver is set.');
else
    error('The QP solver is not set.')
end

The QP solver is set.
```

#### load and check tutorial input is loaded correctly

```
tutorialPath = [CBTDIR filesep 'tutorials' filesep 'metabotools' filesep 'tutorial_II'];
if exist([tutorialPath filesep 'starting_model.mat'], 'file') == 2 % 2 means it's a file.
        load([tutorialPath filesep 'starting_model.mat']);
        display('The model is loaded.');
else
        error('The ''starting_model'' could not be loaded.');
end
```

The model is loaded.

```
% Check output path and writing permission
if ~exist(outputPath) == 7
    error('Output directory in ''outputPath'' does not exist. Verify that you type it correct
end
% make and save dummy file to test writing to output directory
A = rand(1);
try
    save([outputPath filesep 'A']);
catch ME
    error('Files cannot be saved to the provided location: %s\nObtain rights to write into %s
end
```

## Section 1 - Define the model bounds using setMediumConstraints

```
set_inf = 2000;
current_inf = 1000;
medium_composition = {};
met_Conc_mM = [];
cellConc = [];
t = [];
cellWeight = [];

mediumCompounds = {'EX_h(e)', 'EX_h2o(e)', 'EX_hco3(e)', 'EX_nh4(e)', 'EX_o2(e)', 'EX_pi(e)',
```

#### Section 2 - Generate an individual exchange profiles for each sample

```
load([tutorialPath filesep 'tutorial_II_data.mat']);
model = modelMedium;
test_max = 500;
test_min = 0.0001;
variation = 20;

prepIntegrationQuant(model, metData, exchanges, samples, test_max, test_min, outputPath, tol,
clearvars -EXCEPT modelMedium samples tol solver outputPath tutorialPath solverQuant
```

#### Section 2B - Prepare table to check exchange profiles

```
nmets = 70;
[mapped_exchanges, minMax, mapped_uptake, mapped_secretion] = checkExchangeProfiles(samples, clearvars -EXCEPT modelMedium samples tol solver mapped_exchanges outputPath tutorialPath solver([outputPath filesep 'Result_checkExchangeProfiles']);
```

#### Section 3 - Generate contextualized models

```
changeCobraSolver(solverQuant, 'LP');
minGrowth = 0.008;
obj = 'biomass_reaction2';
no_secretion = {'EX_o2(e)'};
no_uptake = {'EX_o2s(e)', 'EX_h2o2(e)'};
medium = {};
tol = 1e-6;
model = modelMedium;
epsilon = 1e-4;
```

```
addExtraExch = {'EX_tdchola(e)', 'Ex_5hoxindoa[e]'};
addExtraExch_value = 1;
[ResultsAllCellLines, OverViewResults] = setQuantConstraints(model, samples, tol, minGrowth, on uptake, medium, addExtraExch, clearvars -EXCEPT modelMedium samples ResultsAllCellLines OverViewResults tol solver mapped_extraPython.
```

#### Section 4 - Analyze added exchanges

```
changeCobraSolver(solver, 'LP');

[Ex_added_all_unique] = statisticsAddedExchanges(ResultsAllCellLines, samples);
clearvars -EXCEPT modelMedium samples ResultsAllCellLines OverViewResults Ex_added_all_unique

[Added_all] = mkTableOfAddedExchanges(ResultsAllCellLines, samples, Ex_added_all_unique);
save([outputPath filesep 'statistics']);
clearvars -EXCEPT modelMedium samples ResultsAllCellLines OverViewResults tol solver mapped_exchanges(ResultsAllCellLines OverViewResults tol solver mapped_exchanges);
```

#### Section 5 - Analyze the sets of essential genes

```
cutoff = 0.05;
[genes, ResultsAllCellLines, OverViewResults] = analyzeSingleGeneDeletion(ResultsAllCellLines,
clearvars -EXCEPT modelMedium samples ResultsAllCellLines OverViewResults Ex_added_all_unique
```

## Section 6 - Check which individual gene-associated reaction makes the model infeasible

```
samples_to_test = samples;
fill = 'NAN';
genes_to_test = {'55293.1'};

[FBA_Rxns_K0, ListResults] = checkEffectRxnK0(samples_to_test, fill, genes_to_test, samples, Fill)
clearvars -EXCEPT modelMedium samples ResultsAllCellLines OverViewResults Ex_added_all_unique
```

#### Section 7 - Make intersect and union model

```
mk_union = 1;
mk_intersect = 1;
mk_reactionDiff = 1;
load([tutorialPath filesep 'starting_model.mat']);
model = starting_model;

[unionModel, intersectModel, diffRxns, diffExRxns] = makeSummaryModels(ResultsAllCellLines, sacclearvars -EXCEPT modelMedium samples ResultsAllCellLines OverViewResults Ex_added_all_unique save([outputPath filesep 'summary']);
```

### **Section 8A ATP production**

#### **Section 8B NADH production**

```
met2test = {'nadh[c]', 'nadh[m]', 'nadh[n]', 'nadh[x]', 'nadh[r]'};

transportRxns = {'NADHtpu'; 'NADHtru'; 'NADtpu'};

[BMall, ResultsAllCellLines, metRsall, maximum_contributing_rxn, maximum_contributing_flux_NADHtru'; 'nadh[x]', 'nadh
```

#### **Section 8C FADH2 production**

```
transportRxns = {'FADH2tru'; 'FADH2tx'};

met2test = {'fadh2[c]', 'fadh2[m]', 'fadh2[n]', 'fadh2[x]', 'fadh2[r]'};
 [BMall, ResultsAllCellLines, metRsall, maximum_contributing_rxn, maximum_contributing_flux_FAI
    clear transportRxns met2test

PHs = [PHs maximum_contributing_rxn(:, 1)];
```

## **Section 8D NADPH production**

```
transportRxns = {'NADPHtru'; 'NADPHtxu'};
```

```
met2test = {'nadph[c]', 'nadph[m]', 'nadph[n]', 'nadph[x]', 'nadph[r]'};
[BMall, ResultsAllCellLines, metRsall, maximum_contributing_rxn, maximum_contributing_flux_NAC
clear transportRxns met2test

PHs = [PHs maximum_contributing_rxn(:, 1)];
save([outputPath filesep 'fluxSplits']);
```

#### Section 8E illustrate the phenotypes (PHs) on 3Dplot

```
diff_view = 1;
fonts = 18;
make3Dplot(PHs, maximum_contributing_flux_ATP, fonts, outputPath, diff_view);
```

#### **Section 9 Perform phase Plane Analysis**

```
mets = {'EX_glc(e)', 'EX_o2(e)'; 'EX_gln_L(e)', 'EX_o2(e)'; 'EX_lac_L(e)', 'EX_o2(e)'};
step_size = [40, 40; 20, 40; 40, 40];
step_num = [28, 26; 21, 26; 42, 26];
direct = [-1, -1; -1, -1; 1, -1];
[ResultsAllCellLines] = performPPP(ResultsAllCellLines, mets, step_size, samples, step_num, disave([outputPath filesep 'PPP']);
```

#### Section 9b illustrate phase plane analysis results

```
label = {'Glucose uptake (fmol/cell/hr)'; 'Oxygen uptake (fmol/cell/hr)'; 'Growth rate (hr-1)
mets = {'EX_glc(e)'; 'EX_o2(e)'};
fonts = 12;
samples = {'IGROV1'};
illustrate_ppp(ResultsAllCellLines, mets, outputPath, samples, label, fonts, tol);
```