Model manipulation

Author(s): Vanja Vlasov, Systems Biochemistry Group, University of Luxembourg,

Thomas Pfau, Systems Biology Group, LSRU, University of Luxembourg.

Reviewer(s): Thomas Pfau, Systems Biology Group, LSRU, University of Luxembourg.

INTRODUCTION

In this tutorial, we will do a manipulation with the simple model of the first few reactions of the glycolisis metabolic pathway as created in the "Model Creation" tutorial.

Glycolysis is the metabolic pathway that occurs in most organisms in the cytosol of the cell. First, we will use the beginning of that pathway to create a simple constraint-based metabolic network (Figure 1).

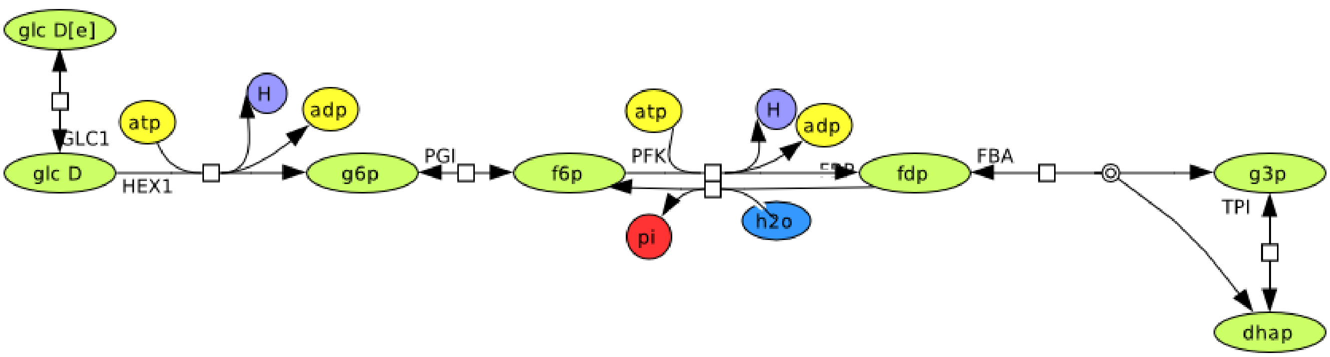

Figure 1: A small metabolic network consisting of the seven reactions in the glycolisis pathway.

At the beginning of the reconstruction, an integrity of the draft reconstruction must be initially assessed. The accuracy of the stochiometry, necessity of each reaction and metabolite, direction and reversibility of the reactions needed to be evaluated.

After creating or loading the model, the model can be modified to simulate different conditions, such as:

- Creating, adding and handling reactions;
- Adding Exchange, Sink and Demand reactions;
- Altering reaction bounds;
- Altering Reactions;
- Remove reactions and metabolites;
- Search for duplicates and comparison of two models;
- Changing the model objective;
- Changing the direction of reaction(s);

EQUIPMENT SETUP

Start CobraToolbox

```
initCobraToolbox;
```

```
                                                  |
  _____   _____   _____   _____     _____         |
 /  ___| /  _  \ |  _  \ |  _  \   / ___ \        |   COnstraint-Based Reconstruction and Analysis
 | |     | | | | | |_| | | |_| |  | |___| |       |   The COBRA Toolbox - 2017
 | |     | | | | |  _  { |  _  /  |  ___  |       |
```

```
    | |___  | |_| | | |_| | | | \ \  | |   | |   |   Documentation:
    \_____| \_____/ |_____/ |_|  \_\ |_|   |_|   |   http://opencobra.github.io/cobratoolbox
                                                |
```

```
> Checking if git is installed ...  Done.
> Checking if the repository is tracked using git ...  Done.
> Checking if curl is installed ...  Done.
> Checking if remote can be reached ...  Done.
> Initializing and updating submodules ... Done.
> Adding all the files of The COBRA Toolbox ...  Done.
> Define CB map output... set to svg.
> Retrieving models ...   Done.
> TranslateSBML is installed and working properly.
> Configuring solver environment variables ...
  - [----] ILOG_CPLEX_PATH :  --> set this path manually after installing the solver ( see instructions
  - [*---] GUROBI_PATH: /opt/gurobi702/linux64/matlab
  - [----] TOMLAB_PATH :  --> set this path manually after installing the solver ( see instructions )
  - [----] MOSEK_PATH :  --> set this path manually after installing the solver ( see instructions )
  Done.
> Checking available solvers and solver interfaces ... Done.
> Setting default solvers ... Done.
> Saving the MATLAB path ... Done.
  - The MATLAB path was saved as ~/pathdef.m.

> Summary of available solvers and solver interfaces

  Support     LP   MILP     QP   MIQP    NLP
----------------------------------------------------------------------
cplex_direct  full              0      0      0      0      -
dqqMinos      full              1      -      -      -      -
glpk          full              1      1      -      -      -
gurobi        full              1      1      1      1      -
ibm_cplex     full              0      0      0      -      -
matlab        full              1      -      -      -      1
mosek         full              0      0      0      -      -
pdco          full              1      -      1      -      -
quadMinos     full              1      -      -      -      1
tomlab_cplex  full              0      0      0      0      -
qpng          experimental      -      -      1      -      -
tomlab_snopt  experimental      -      -      -      -      0
gurobi_mex    legacy            0      0      0      0      -
lindo_old     legacy            0      -      -      -      -
lindo_legacy  legacy            0      -      -      -      -
lp_solve      legacy            1      -      -      -      -
opti          legacy            0      0      0      0      0
----------------------------------------------------------------------
Total         -                 7      2      3      1      2

+ Legend: - = not applicable, 0 = solver not compatible or not installed, 1 = solver installed.


> You can solve LP problems using: 'dqqMinos' - 'glpk' - 'gurobi' - 'matlab' - 'pdco' - 'quadMinos' - '
> You can solve MILP problems using: 'glpk' - 'gurobi'
> You can solve QP problems using: 'gurobi' - 'pdco' - 'qpng'
> You can solve MIQP problems using: 'gurobi'
> You can solve NLP problems using: 'matlab' - 'quadMinos'

> Checking for available updates ...
> The COBRA Toolbox is up-to-date.
```

PROCEDURE

Generate a network

A constraint-based metabolic model contains the stoichiometric matrix with reactions and metabolites [1].

S is stoichiometric representation of metabolic networks corresponding to the reactions in the biochemical pathway. In an each column of the *S* is a biochemical reaction and in each row is a precise metabolite. There is a stoichiometric coefficient of zero, which means that metabolite not participate in that distinct reaction. The coefficient also can be positive when the appropriate metabolite is produced, or negative for every metabolite depleted [1].

```
ReactionFormulas = {'glc_D[e]  -> glc_D[c]',...
    'glc_D[c] + atp[c]  -> h[c] + adp[c] + g6p[c]',...
    'g6p[c]  <=> f6p[c]',...
    'atp[c] + f6p[c]  -> h[c] + adp[c] + fdp[c]',...
    'fdp[c] + h2o[c]  -> f6p[c] + pi[c]',...
    'fdp[c]  -> g3p[c] + dhap[c]',...
    'dhap[c]  -> g3p[c]'};
ReactionNames = {'GLCt1r', 'HEX1', 'PGI', 'PFK', 'FBP', 'FBA', 'TPI'};
lowerbounds = [-20, 0, -20, 0, 0, -20, -20];
upperbounds = [20, 20, 20, 20, 20, 20, 20];
model = createModel(ReactionNames, ReactionNames, ReactionFormulas,...
                    'lowerBoundList', lowerbounds, 'upperBoundList', upperbounds);
```

```
Warning: Metabolite glc_D[e] not in model - added to the model
Warning: Metabolite glc_D[c] not in model - added to the model
GLCt1r glc_D[e]  <=> glc_D[c]
Warning: Metabolite atp[c] not in model - added to the model
Warning: Metabolite h[c] not in model - added to the model
Warning: Metabolite adp[c] not in model - added to the model
Warning: Metabolite g6p[c] not in model - added to the model
HEX1 glc_D[c] + atp[c]  -> h[c] + adp[c] + g6p[c]
Warning: Metabolite f6p[c] not in model - added to the model
PGI g6p[c]  <=> f6p[c]
Warning: Metabolite fdp[c] not in model - added to the model
PFK atp[c] + f6p[c]  -> h[c] + adp[c] + fdp[c]
Warning: Metabolite h2o[c] not in model - added to the model
Warning: Metabolite pi[c] not in model - added to the model
FBP fdp[c] + h2o[c]  -> f6p[c] + pi[c]
Warning: Metabolite g3p[c] not in model - added to the model
Warning: Metabolite dhap[c] not in model - added to the model
FBA fdp[c]  <=> g3p[c] + dhap[c]
TPI dhap[c]  <=> g3p[c]
```

We can now have a look at the different model fields created. The stoichiometry is stored in the S field of the model, which was described above. Since this is commonly a sparse matrix (i.e. it does contain a lot of zeros, to display it it is useful to use the full representation)

```
full(model.S)
```

```
ans =

    -1     0     0     0     0     0     0
     1    -1     0     0     0     0     0
     0    -1     0    -1     0     0     0
     0     1     0     1     0     0     0
     0     1     0     1     0     0     0
     0     1    -1     0     0     0     0
     0     0     1    -1     1     0     0
```

```
     0     0     0     1    -1    -1     0
     0     0     0     0    -1     0     0
     0     0     0     0     1     0     0
```

Some descriptive fields always present are model.mets and model.rxns which represent the metabolites and the reactions respectively.

```
model.mets
```

```
ans =
    'glc_D[e]'
    'glc_D[c]'
    'atp[c]'
    'h[c]'
    'adp[c]'
    'g6p[c]'
    'f6p[c]'
    'fdp[c]'
    'h2o[c]'
    'pi[c]'
    'g3p[c]'
    'dhap[c]'
```

```
model.rxns
```

```
ans =
    'GLCt1r'
    'HEX1'
    'PGI'
    'PFK'
    'FBP'
    'FBA'
    'TPI'
```

Fields in the COBRA model are commonly column vectors, which can be an important detail when writing functions manipulating these fields.

There are a few more fields present in each COBRA model:

model.lb, indicating the lower bounds of each reaction, and model.ub indicating the upper bound of a reaction.

```
% this displays an array with reaction names and flux bounds.
[{'Reaction ID', 'Lower Bound', 'Upper Bound'};...
 model.rxns, num2cell(model.lb), num2cell(model.ub)]
```

```
ans =
    'Reaction ID'    'Lower Bound'    'Upper Bound'
    'GLCt1r'         [        -20]    [         20]
    'HEX1'           [          0]    [         20]
    'PGI'            [        -20]    [         20]
    'PFK'            [          0]    [         20]
    'FBP'            [          0]    [         20]
    'FBA'            [        -20]    [         20]
    'TPI'            [        -20]    [         20]
```

```
% This is a convenience function which does pretty much the same as the line above
printFluxBounds(model);
```

```
Reaction ID    Lower Bound    Upper Bound
     GLCt1r        -20.000         20.000
       HEX1          0.000         20.000
        PGI        -20.000         20.000
        PFK          0.000         20.000
        FBP          0.000         20.000
        FBA        -20.000         20.000
        TPI        -20.000         20.000
```

Before we start to modify the model, it might be useful to store some of the current properties of the model

```
mets_length = length(model.mets);
rxns_length = length(model.rxns);
```

Creating, adding and handling reactions

If we want to add a reaction to the model or modify an existing reaction we are using function `addReaction`.

We will add some more reactions from glycolysis.

- The formula approach

```
model = addReaction(model, 'GAPDH',...
       'reactionFormula', 'g3p[c] + nad[c] + 2 pi[c] -> nadh[c] + h[c] + 13bpg[c]');
```

```
Warning: Metabolite nad[c] not in model - added to the model
Warning: Metabolite nadh[c] not in model - added to the model
Warning: Metabolite 13bpg[c] not in model - added to the model
GAPDH 2 pi[c] + g3p[c] + nad[c]  -> h[c] + nadh[c] + 13bpg[c]
```

```
model = addReaction(model, 'PGK',...
       'reactionFormula', '13bpg[c] + adp[c] -> atp[c] + 3pg[c]');
```

```
Warning: Metabolite 3pg[c] not in model - added to the model
PGK adp[c] + 13bpg[c]  -> atp[c] + 3pg[c]
```

```
model = addReaction(model, 'PGM', 'reactionFormula', '3pg[c] <=> 2pg[c]' );
```

```
Warning: Metabolite 2pg[c] not in model - added to the model
PGM 3pg[c]  <=> 2pg[c]
```

Display of stoichiometric matrix after adding reactions. Note the enlarge link when you move your mouse over the output to display the full matrix:

```
full(model.S)
```

```
ans =

    -1     0     0     0     0     0     0     0     0     0
```

```
     1    -1     0     0     0     0     0     0     0     0
     0    -1     0    -1     0     0     0     0     1     0
     0     1     0     1     0     0     0     1     0     0
     0     1     0     1     0     0     0     0    -1     0
     0     1    -1     0     0     0     0     0     0     0
     0     0     1    -1     1     0     0     0     0     0
     0     0     0     1    -1    -1     0     0     0     0
     0     0     0     0    -1     0     0     0     0     0
     0     0     0     0     1     0     0    -2     0     0
```

```
% one extra column is added(for added reaction) and 5 new
% rows(for nadh, nad, 13bpg, 2pg and 3pg metabolites)
```

The following functions are used when we want to search reactions sequence in the model and change the order of the selected reaction.

```
rxnID = findRxnIDs(model, model.rxns);
model = moveRxn(model, 8, 1);
```

While the latter does not modify the structure as such it can help in keeping a model tidy.

- The list approach

The `addReaction` function has ability to recognize duplicate reactions when an order of metabolites and an abbreviation of the reaction are different.

```
model = addReaction(model, 'GAPDH2',...
    'metaboliteList', {'g3p[c]', 'nad[c]', 'pi[c]', '13bpg[c]', 'nadh[c]', 'h[c]' },...
    'stoichCoeffList', [-1; -1; -2; 1; 1; 1], 'reversible', false);
```

```
Warning: Model already has the same reaction you tried to add: GAPDH
```

Since the second call should not have added anything we will check if the number of the reaction increased by the three reactions we added (and not by the one duplicated) and the number of metabolites was incremented by five (13bpg, nad, nadh, 23bpg and 2pg).

```
assert(length(model.rxns) == rxns_length + 3);
assert(length(model.mets) == mets_length + 5);
```

Adding Exchange, Sink and Demand reactions

Specific type of reactions in the constraint-based models are reactions that are using and recycling accumulated metabolites, or producing required metabolites in the model.

1. *Exchange reactions* - Reactions added to the model to move metabolites across the created *in silico* compartments. Those compartments represent intra- and intercellular membranes.
2. *Sink reactions* - The metabolites, produced in reactions that are outside of an ambit of the system or in unknown reactions, are supplied to the network with reversible sink reactions.
3. *Demand reactions* - Irreversible reactions added to the model to consume metabolites that are deposited in the system.

There are two ways to implement that kind of reactions:

1. **Use `addReaction` with the documented function call:**

```
model = addReaction(model, 'EX_glc_D[e]', 'metaboliteList', {'glc_D[e]'} ,...
                    'stoichCoeffList', [-1]);
```

```
EX_glc_D[e] glc_D[e]  <=>
```

In the bigger networks we can find our exchange reactions with the following functions:

```
% determines whether a reaction is a general exchange reaction and
% whether its an uptake.
[selExc, selUpt] = findExcRxns(model, 0, 1)
```

```
selExc =
   0
   0
   0
   0
   0
   0
   0
   0
   0
   0
   1

selUpt =
   0
   0
   0
   0
   0
   0
   0
   0
   0
   0
   0
```

2. Use a utility function to create a particular reaction type: `addExchangeRxn`, `addSinkReactions`, `addDemandReaction`.

```
model = addExchangeRxn(model, {'glc_D[e]', 'glc_D[c]'})
```

```
Warning: Reaction with the same name already exists in the model, updating the reaction
EX_glc_D[e] glc_D[e]  <=>
EX_glc_D[c] glc_D[c]  <=>
```

```
model =
          rxns: {12×1 cell}
             S: [17×12 double]
            lb: [12×1 double]
            ub: [12×1 double]
             c: [12×1 double]
          mets: {17×1 cell}
             b: [17×1 double]
         rules: {12×1 cell}
```

```
         genes: {0×1 cell}
        osense: -1
        csense: [17×1 char]
    rxnGeneMat: [12×0 double]
      rxnNames: {12×1 cell}
    subSystems: {12×1 cell}
      metNames: {17×1 cell}
       grRules: {12×1 cell}
```

```
model = addSinkReactions(model, {'13bpg[c]', 'nad[c]'})
```

```
sink_13bpg[c] 13bpg[c]  <=>
sink_nad[c] nad[c]  <=>
```

```
model =
          rxns: {14×1 cell}
             S: [17×14 double]
            lb: [14×1 double]
            ub: [14×1 double]
             c: [14×1 double]
          mets: {17×1 cell}
             b: [17×1 double]
         rules: {14×1 cell}
         genes: {0×1 cell}
        osense: -1
        csense: [17×1 char]
    rxnGeneMat: [14×0 double]
      rxnNames: {14×1 cell}
    subSystems: {14×1 cell}
      metNames: {17×1 cell}
       grRules: {14×1 cell}
```

```
model = addDemandReaction(model, {'dhap[c]', 'g3p[c]'})
```

```
DM_dhap[c] dhap[c]  ->
DM_g3p[c] g3p[c]  ->
```

```
model =
          rxns: {16×1 cell}
             S: [17×16 double]
            lb: [16×1 double]
            ub: [16×1 double]
             c: [16×1 double]
          mets: {17×1 cell}
             b: [17×1 double]
         rules: {16×1 cell}
         genes: {0×1 cell}
        osense: -1
        csense: [17×1 char]
    rxnGeneMat: [16×0 double]
      rxnNames: {16×1 cell}
    subSystems: {16×1 cell}
      metNames: {17×1 cell}
       grRules: {16×1 cell}
```

Setting ratio between the reactions and changing reactions boundary

It is important to emphasize that previous knowledge base informations should be taken into account. Most of them could disrupt future analysis of the model.

For instance, if it is familiar that flux through one reaction is *X* times the flux through another reaction, it is recommended to specify that in your model.

E.g. $1\, v\, \mathrm{EX\_glc\_}D[c] = 2\, v\, \mathrm{EX\_glc\_}D[e]$

```
model = addRatioReaction (model, {'EX_glc_D[c]', 'EX_glc_D[e]'}, [1; 2]);
```

Altering Reaction bounds

In order to respect the transport and exchange potential of a particular metabolite, or to resemble the different conditions in the model, we frequently need to set appropriate limits of the reactions.

```
model = changeRxnBounds(model, 'EX_glc_D[e]', -18.5, 'l');
```

Modifiying Reactions

The `addReaction` function also is a good choice when modifying reactions. By supplying a new stoichiometry, the old will be overwritten. For example further up, we added awrong stoichiometry for the GAP-Dehydrogenase with a phosphate coefficient of 2. (easily visiple by printing the reaction)

```
printRxnFormula(model, 'rxnAbbrList', 'GAPDH');
```

```
GAPDH 2 pi[c] + g3p[c] + nad[c]  -> h[c] + nadh[c] + 13bpg[c]
```

We can correct this by simply calling ddReaction again with the corrected stoichiometry. In essence parts which are not provided are taken from the old reaction, and only the new ones overwrite the existing data

```
model = addReaction(model, 'GAPDH',...
    'metaboliteList', {'g3p[c]', 'nad[c]', 'pi[c]', '13bpg[c]', 'nadh[c]','h[c]' },...
    'stoichCoeffList', [-1; -1; -1; 1; 1; 1]);
```

```
Warning: Reaction with the same name already exists in the model, updating the reaction
GAPDH pi[c] + g3p[c] + nad[c]  -> h[c] + nadh[c] + 13bpg[c]
```

We might also want to add a gene rule to the reaction. This can either be done using

```
model = changeGeneAssociation(model, 'GAPDH', 'G1 and G2');
```

```
New gene G1 added to model
New gene G2 added to model
```

```
printRxnFormula(model, 'rxnAbbrList', {'GAPDH'}, 'gprFlag', true);
```

```
GAPDH pi[c] + g3p[c] + nad[c]  -> h[c] + nadh[c] + 13bpg[c]  G1 and G2
```

The other option to achieve this is to use `addReaction` and the `geneRule` parameter

```
model = addReaction(model, 'PGK', 'geneRule', 'G2 or G3', 'printLevel', 0);
```

```
Warning: Reaction with the same name already exists in the model, updating the reaction
New gene G3 added to model
```

```
printRxnFormula(model, 'gprFlag', true);
```

```
GAPDH pi[c] + g3p[c] + nad[c]  -> h[c] + nadh[c] + 13bpg[c]  G1 and G2
GLCt1r glc_D[e]  <=> glc_D[c]
HEX1 glc_D[c] + atp[c]  -> h[c] + adp[c] + g6p[c]
PGI g6p[c]  <=> f6p[c]
PFK atp[c] + f6p[c]  -> h[c] + adp[c] + fdp[c]
FBP fdp[c] + h2o[c]  -> f6p[c] + pi[c]
FBA fdp[c]  <=> g3p[c] + dhap[c]
TPI dhap[c]  <=> g3p[c]
PGK adp[c] + 13bpg[c]  -> atp[c] + 3pg[c]  G2 or G3
PGM 3pg[c]  <=> 2pg[c]
EX_glc_D[e] glc_D[e]  <=> 2 Ratio_EX_glc_D[c]_EX_glc_D[e]
EX_glc_D[c] glc_D[c] + Ratio_EX_glc_D[c]_EX_glc_D[e]  <=>
sink_13bpg[c] 13bpg[c]  <=>
sink_nad[c] nad[c]  <=>
DM_dhap[c] dhap[c]  ->
DM_g3p[c] g3p[c]  ->
```

Remove reactions and metabolites

In order to detach reactions from the model, the following function has been used:

```
model = removeRxns(model, {'EX_glc_D[c]', 'EX_glc_D[e]', 'sink_13bpg[c]', ...
                           'sink_nad[c]', 'DM_dhap[c]', 'DM_g3p[c]'});

assert(rxns_length + 3 == length(model.rxns));
% The reaction length has been reevaluated
```

Remove metabolites

```
model = removeMetabolites(model, {'3pg[c]', '2pg[c]'}, false);
```

For instance, in previous code the many metabolites from 'GAPDH' were deleted, but the reaction is still present in the model (since there are more metabolites left). The false indicates, that empty reactions should not be removed.

To delete metabolites and reactions with zero rows and columns,the following function can be used:

```
model = removeTrivialStoichiometry(model)
```

```
model =
          rxns: {9×1 cell}
             S: [15×9 double]
            lb: [9×1 double]
            ub: [9×1 double]
             c: [9×1 double]
          mets: {15×1 cell}
             b: [15×1 double]
         rules: {9×1 cell}
         genes: {3×1 cell}
       osense: -1
       csense: [15×1 char]
```

```
    rxnGeneMat: [9×3 double]
      rxnNames: {9×1 cell}
    subSystems: {9×1 cell}
      metNames: {15×1 cell}
       grRules: {9×1 cell}
          note: 'EX_glc_D[c] andEX_glc_D[e]are set to have a ratio of1:2.'
```

```
model = removeRxns(model,{'GAPDH', 'PGK'});
```

Search for duplicates and comparison of two models

Since genome-scale metabolic models are expanding every day [2], the need for comparison and merge of them is also spreading.

The elementary functions for the model manipulation, besides main actions, simultaneously perform the structural analysis and comparison (e.g. `addReaction`). Likewise, there are additional functions that are only dealing with analysing similarities and differences within and between the models.

- Checking for reaction duplicates by reaction abbreviation, by using method 'S' that will not detect reverse reactions, and method 'FR' that will neglect reactions direction:

```
[model, removedRxn, rxnRelationship] = checkDuplicateRxn(model, 'S', 1, 1);
```

```
Checking for reaction duplicates by stoichiometry ...
 no duplicates found.
```

Adding duplicate reaction to the model:

```
model = addReaction(model, 'GLCt1r_duplicate_reverse',...
                    'metaboliteList', {'glc_D[e]', 'glc_D[c]'},...
                    'stoichCoeffList', [1 -1], 'lowerBound', 0, ...
                    'upperBound', 20, 'checkDuplicate', 0);
```

```
GLCt1r_duplicate_reverse glc_D[c]  -> glc_D[e]
```

```
fprintf('>> Detecting duplicates using S method\n');
```

```
>> Detecting duplicates using S method
```

```
method = 'S';
% will not be removed as does not detect reverse reaction
[model,removedRxn, rxnRelationship] = checkDuplicateRxn(model, method, 1, 1);
```

```
Checking for reaction duplicates by stoichiometry ...
 no duplicates found.
```

```
assert(rxns_length + 1 == length(model.rxns));
% The reaction length has been reevaluated

fprintf('>> Detecting duplicates with using FR method\n');
```

```
>> Detecting duplicates with using FR method
```

```
method = 'FR';
% will be removed as detects reverse reaction
```

```
[model, removedRxn, rxnRelationship] = checkDuplicateRxn(model, method, 1, 1);
```

```
Checking for reaction duplicates by stoichiometry (up to orientation) ...
     Keep:  GLCt1r glc_D[e]  <=> glc_D[c]
Duplicate:  GLCt1r_duplicate_reverse glc_D[c]  -> glc_D[e]
```

```
assert(rxns_length == length(model.rxns));
```

- Function `checkCobraModelUnique` marks the reactions and metabolites that are not unique in the model.

```
model = checkCobraModelUnique(model, false)
```

```
model =
          rxns: {7×1 cell}
             S: [12×7 double]
            lb: [7×1 double]
            ub: [7×1 double]
             c: [7×1 double]
          mets: {12×1 cell}
             b: [12×1 double]
         rules: {7×1 cell}
         genes: {3×1 cell}
        osense: -1
        csense: [12×1 char]
    rxnGeneMat: [7×3 double]
      rxnNames: {7×1 cell}
    subSystems: {7×1 cell}
      metNames: {12×1 cell}
       grRules: {7×1 cell}
          note: 'EX_glc_D[c] andEX_glc_D[e]are set to have a ratio of1:2.'
```

Changing the model's objective

Simulating different conditions in the model is often necessery in favor of performing calculations that investigate a specific objective. One of the elementary objectives is optimal growth [3]. Model can be modified to get different conditions with changing the model objective:

```
modelNew = changeObjective(model, 'GLCt1r', 0.5);

% multiple rxns, default coefficient (1)
modelNew = changeObjective(model, {'PGI'; 'PFK'; 'FBP'});
```

The direction of reactions

For some purposes, it is important to only have irreversible reactions in a model, i.e. only allowing positive flux in all reactions. This can be important if e.g. absolute flux values are of interest and negative flux would reduce an objective while it should actually increase it. The COBRA toolbox offers functionality to change a model to an irreversible format, by splitting all reversible reactions and adjusting the respective lower and upper bounds, such that the model capacities stay the same.

Lets see, how the glycolysis model currently looks:

```
printRxnFormula(model);
```

```
GLCt1r glc_D[e]  <=> glc_D[c]
HEX1 glc_D[c] + atp[c]  -> h[c] + adp[c] + g6p[c]
PGI g6p[c]  <=> f6p[c]
PFK atp[c] + f6p[c]  -> h[c] + adp[c] + fdp[c]
FBP fdp[c] + h2o[c]  -> f6p[c] + pi[c]
FBA fdp[c]  <=> g3p[c] + dhap[c]
TPI dhap[c]  <=> g3p[c]
```

To convert a model to an irreversible model use the following command:

```
[modelIrrev, matchRev, rev2irrev, irrev2rev] = convertToIrreversible(model);
```

Lets compare the irreversible model with the original model:

```
printRxnFormula(modelIrrev);
```

```
GLCt1r_f glc_D[e]  -> glc_D[c]
HEX1 glc_D[c] + atp[c]  -> h[c] + adp[c] + g6p[c]
PGI_f g6p[c]  -> f6p[c]
PFK atp[c] + f6p[c]  -> h[c] + adp[c] + fdp[c]
FBP fdp[c] + h2o[c]  -> f6p[c] + pi[c]
FBA_f fdp[c]  -> g3p[c] + dhap[c]
TPI_f dhap[c]  -> g3p[c]
GLCt1r_b glc_D[c]  -> glc_D[e]
PGI_b f6p[c]  -> g6p[c]
FBA_b g3p[c] + dhap[c]  -> fdp[c]
TPI_b g3p[c]  -> dhap[c]
```

You will notice, that there are more reactions in this model, and that all reactions which have a lower bound < 0 are split in two.

There is also a function to convert an irreversible model to a reversible model:

```
modelRev = convertToReversible(modelIrrev);
```

If we now compare the reactions of this model with those from the original model, they should look the same.

```
printRxnFormula(modelRev);
```

```
GLCt1r glc_D[e]  <=> glc_D[c]
HEX1 glc_D[c] + atp[c]  -> h[c] + adp[c] + g6p[c]
PGI g6p[c]  <=> f6p[c]
PFK atp[c] + f6p[c]  -> h[c] + adp[c] + fdp[c]
FBP fdp[c] + h2o[c]  -> f6p[c] + pi[c]
FBA fdp[c]  <=> g3p[c] + dhap[c]
TPI dhap[c]  <=> g3p[c]
```

REFERENCES

[1] Orth, J. D., Thiele I., and Palsson, B. Ø. (2010). What is flux balance analysis? *Nat. Biotechnol., 28*(3), 245–248.

[2] Feist, A. M., Palsson, B. (2008). The growing scope of applications of genome-scale metabolic reconstructions: the case of *E. coli*. *Nature Biotechnology, 26*(6), 659–667.

[3] Feist, A. M., Palsson, B. O. (2010). The Biomass Objective Function. *Current Opinion in Microbiology, 13*(3), 344–349.