Single cell RNA sequencing data analysis, 27-29 January 2020

Åsa Björklund asa.bjorklund@scilifelab.se





Thursday – Bring your own data

- Bring your own data or a publically available dataset from a tissue of interest.
- We will help you import the data into appropriate pipelines.
- During the day we will be around to help you with interpretation of the data.
- At the end we will wrap up and compare experiences.





Raw data: scRNA-seq analysis overview fastq files Mapping & Data normalization Gene expression estimate Gene set selection Batch effect removal Removal of other Data: QC: confounders **Expression profiles** Remove low Q cells Remove contaminants Clustering methods Visualization / Trajectory Defining cell types/lineages **Dimensionality reduction** assignment Gene signatures Verification experiments

Some take-home messages





- Data analysis is very seldom a straight line one pipeline fits all.
 - Often requires several iterations of filtering data, exploring data, refiltering, exploring again, discovering technical artifacts, normalization, exploring again, etc. etc.





- Get to know your data what types of variation do you have?
 - PCA is a good tool for exploring data
- Apply appropriate methods to control for problems that you see.





- Always check for:
 - Batch effects think of all possible batches.
 - Cell cycle effects if appropriate
 - Separation due to nUMI / nGene
- Both at the start of a project and at the end for your final clustering.





- Clustering try out a few different approaches
 - Consensus of different methods gives confidence
 - If they do not agree figure out why!





- Use your biological knowledge to evaluate the results
- Warning! Do not overfit your data to fit your initial hypotheses. Keep an open mind;-)



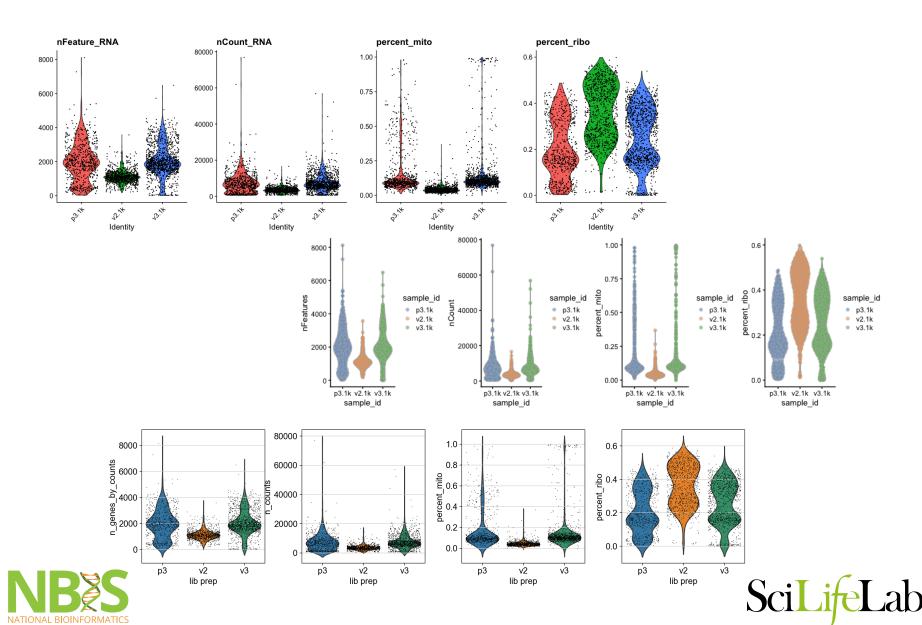


- scRNAseq analysis is a fast evolving field with new methods being published all the time.
 - Try to keep up with development
 - BUT! You cannot test every new method out there!

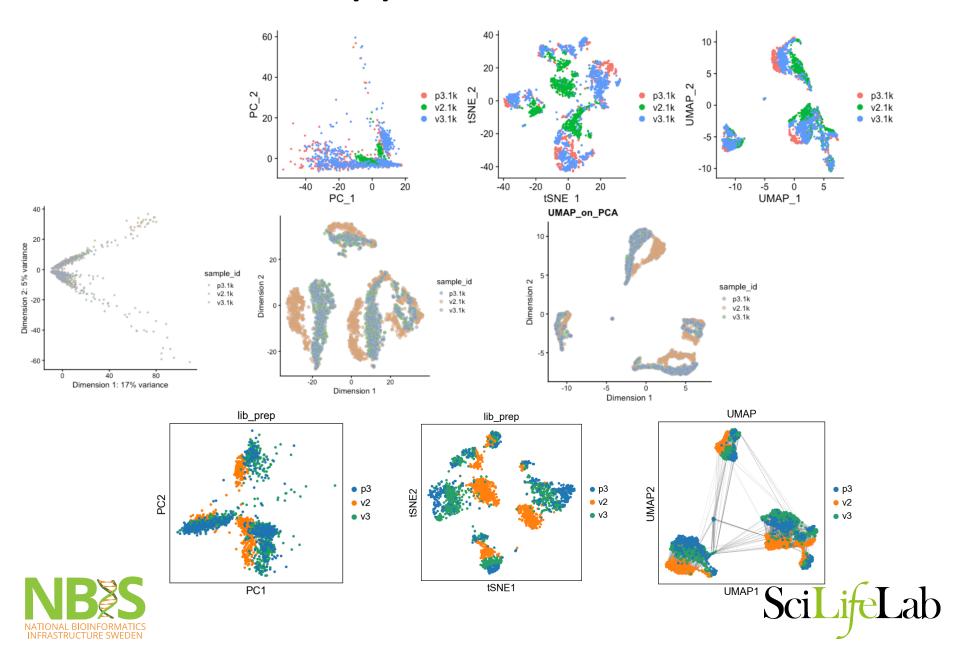




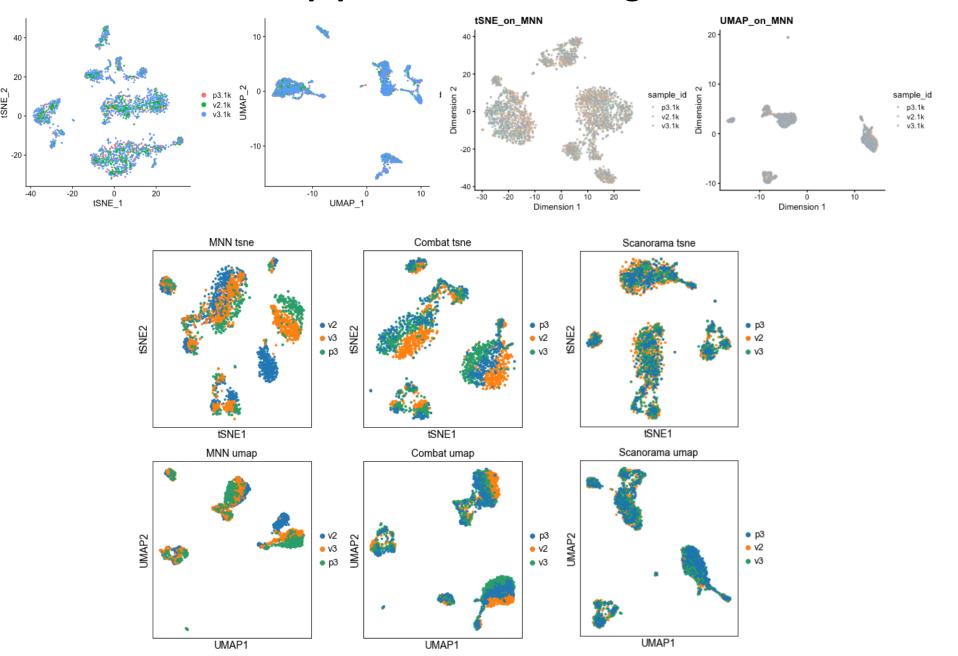
Different pipelines - QC



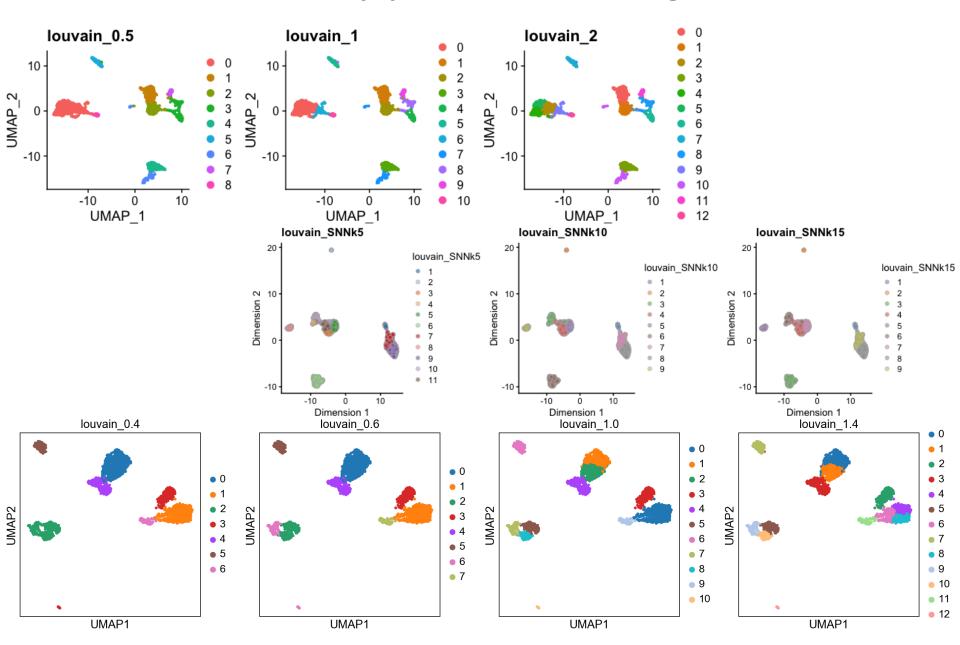
Different pipelines – Dim Reduction



Data pipelines – Data integration

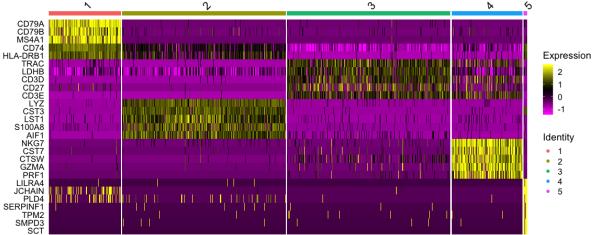


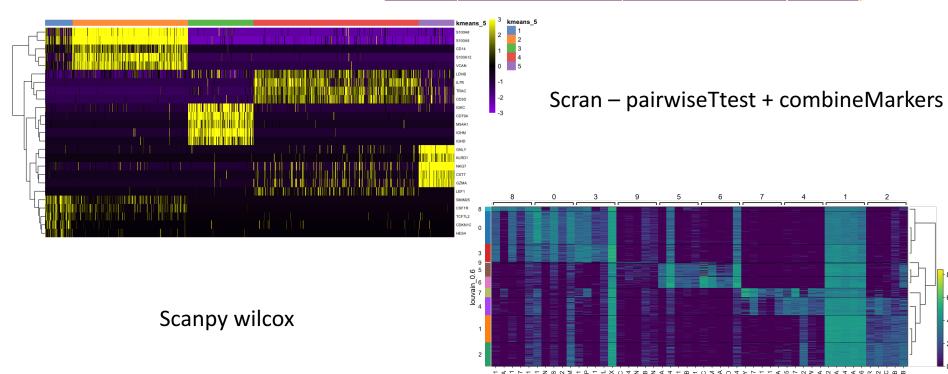
Data pipelines - Clustering



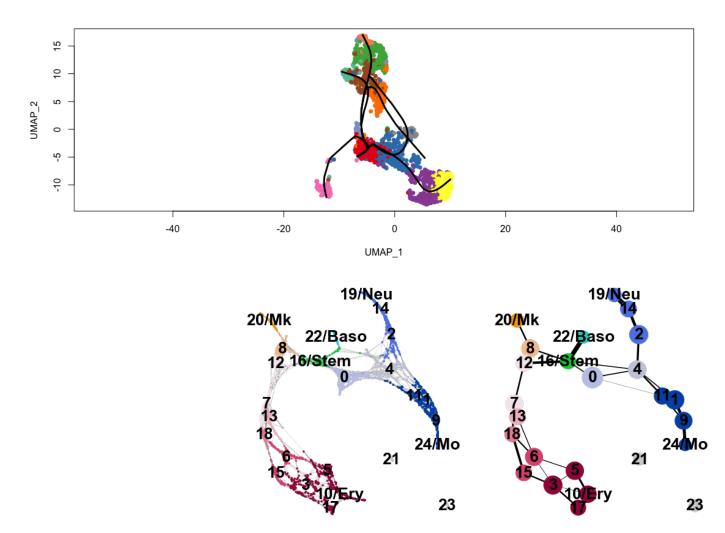
Data pipelines - DE

Seurat wilcox





Trajectory – PAGA and Slingshot







Additional comments to the exercises?





Need help?

- NBIS Long term support (aka WABI) application rounds 3 times a year – 500h for free
- NBIS Short term support fee for service support.
 Apply any time.
- Drop-in sessions weekly/bi-weekly at several sites across Sweden
 - SciLifeLab Stockholm Gamma 2, lunch room, Tuesdays 10.30
 - SciLifeLab Uppsala Navet floor 3, Thursdays 10.00
 - Umeå, Linköping, Stockholm University, Lund and Götebort as well.
- More info at: http://nbis.se/





Reproducible research in R

- R / Rstudio in Docker containers
 - https://www.andrewheiss.com/blog/2017/04/27/super-basicpractical-guide-to-docker-and-rstudio/
 - https://github.com/rocker-org/rocker
- OBS! On Uppmax only Singularity containers are allowed. Most Docker images can be converted.
- Learn more on containers etc:
 - http://nbis-reproducible-research.readthedocs.io/en/latest/
- Rstudio package management Packrat
 - https://rstudio.github.io/packrat/
- Conda installations of packages can use conda on both bianca and rackham – module load conda





Please fill in the Evaluation Form

Your feedback is important so that we can help improve the course.

Good luck with your analyses!



