iMKT using PopFly or PopHuman data

Brief intro about lots of data right now. Then, PopFly and PopHuman as great databases.

Therefore, iMKT package includes some functions which allow an easy retrieval and analysis of population genetics information stored in these genome browsers. Specifically, the functions permit the download of population genetics parameters computed for every gene annotation in several populations for both model species Drosophila and Homo sapiens.

Loading the row data

First step is to load this information into your working environment and how to access it in order to use it efficiently. To do so, use the loadPopFly() and loadPopHuman() functions.

Keep in mind that you are downloading the complete gene information of several populations (16 D. melanogaster and 26 Human), and this process could take a while.

Once the functions finish, two new objects are loaded into the workspace: PopFlyData and PopHumanData. These objects can be manually examined or used only when performing the main iMKT analysis.

```
library(iMKT)
#> Loading required package: ggplot2
loadPopFly()
#> Loading PopFly data into your workspace.
#> This process may take some seconds to complete, please be patient.
#loadPopHuman()
#knitr::kable(head(PopFlyData,4))
head(PopFlyData, 4)
     Pop
                Name
                        Start
                                   End Chr p0 pi d0
                                                     di
                                                                   alpha
#> 1 EA FBqn0000008 18024473 18060339
                                        2R 28 17 36
                                                     19 647 2449 -0.1504
#> 2 EA FBqn0000014 12632936 12655771
                                        3R
                                           5
                                              0 19
                                                      0 155
                                                             643
                                                                      NA
#> 3 EA FBqn0000015 12752932 12797958
                                        3R 6
                                              1 4
                                                                    -Inf
#> 4 EA FBqn0000017 16608966 16640982
                                       3L 10 5 83 134 867 3263
                                                                  0.6903
     fisher_pval
                     DoS
                           KaKs
                                               DAF0f
#> 1
          0.8348 -0.0323 0.5278 11;1;2;1;1;0;0;1;0;0 10;9;4;0;1;2;0;1;1;0
#> 2
          1.0000 0.0000 0.0000 0;0;0;0;0;0;0;0;0;0 0;1;2;0;0;0;0;2;0
#> 3
          1.0000 -0.1429 0.0000 0;1;0;0;0;0;0;0;0;0 0;2;1;0;2;0;0;0;1;0
          0.0530 0.2842 1.6145 0;3;1;1;0;0;0;0;0;0 0;6;1;1;0;0;0;0;2;0
#>
         cM_Mb
#> 1 2.1692837
#> 2 0.7616990
#> 3 0.4352566
#> 4 0.8999361
ls() ## new object created
#> [1] "PopFlyData"
```

Each row of the PopFlyData dataframe contains information regarding one gene annotation in one single population. In total, 16 populations and X genes are included. Metrics for each gene contain information about segrating, divergent and analyzed sites and the Derived Allele Frequency (DAF) for neutral (4fold) and putatively selected (0fold) sites; together with some neutrality tests statistics (Standard MKT, Direction of Selection, Ka/Ks).

Performing PopFly Analyses

The *PopFlyAnalisys()* function allows performing any MK test using a subset of PopFly data defined by custom genes and populations lists. It uses the previously loaded dataframe (PopFlyData). In addition to the genes and populations lists, the function also has the following parameters:

- recomb group genes according to recombination values (must specify number of bins). TRUE/FALSE. Recomb values (cM/Mb) from Comeron et al. 2012.
- bins number of recombination bins to compute (mandatory if recomb = TRUE)
- test which test to perform. Options include: standard (default), DGRP, FWW, asymptotic, iMK
- xlow lower limit for asymptotic alpha fit (default=0)
- xhigh higher limit for asymptotic alpha fit (default=1)

Hence, it allows deciding which test to perform, and whether to analyze genes groupped by recombination bins or not.

Example 1

In this first example, the analysis is focused in two genes and 3 populations, without considering gene's recombination context, and using DGRP methodology.

```
PopFlyAnalysis(genes=c("FBgn0000055","FBgn0003016"), pops=c("RAL","ZI","FR"), recomb=F, test="DGRP")
\# [1] "Population = FR"
#> Warning in check_input(daf, divergence, 0, 1): Input daf file contains PO values = 0.
#> This can bias the function fitting and the estimation of alpha.
#> [1] "Population = RAL"
#> Warning in check_input(daf, divergence, 0, 1): Input daf file contains PO values = 0.
#> This can bias the function fitting and the estimation of alpha.
#> [1] "Population = ZI"
#> $`Population = FR`
#> $`Population = FR`$Results
#>
                  alpha.symbol Fishers exact test P-value
\# Cutoff = 0
                    -0.3675214
                                                0.5274392
#> Cutoff = 0.05
                    -0.3675214
                                                0.5274392
#> Cutoff = 0.2
                                                0.5274392
                    -0.3675214
#>
#> $`Population = FR`$`Divergence metrics`
#> $`Population = FR`$`Divergence metrics`$`Global metrics`
#>
             Ka
                         Ks
                                 omega
#> 1 0.003767024 0.05807201 0.06486815
#>
#> $`Population = FR`$`Divergence metrics`$`Estimates by cutoff`
#>
                  omegaA.symbol omegaD.symbol
\# Cutoff = 0
                    -0.02384043
                                  0.08870859
#> Cutoff = 0.05
                    -0.02384043
                                   0.08870859
\# Cutoff = 0.2
                    -0.02384043
                                   0.08870859
#>
#>
#> $`Population = FR`$`MKT tables`
#> $`Population = FR`$`MKT tables`$`Number of segregating sites by DAF category - Cutoff = 0`
#>
#>
#> Table: cutoff
#>
#>
                     DAF.below.cutoff
                                       DAF. above. cutoff
```

```
0
#> Neutral class
#> Selected class
#>
#> $`Population = FR`$`MKT tables`$`Number of segregating sites by DAF category - Cutoff = 0.05`
#>
#>
#> Table: cutoff
#>
#>
                DAF.below.cutoff DAF.above.cutoff
#> -----
                    27
                                             18
#> Neutral class
                             10
#> Selected class
#> $`Population = FR`$`MKT tables`$`Number of segregating sites by DAF category - Cutoff = 0.2`
#>
#>
#> Table: cutoff
#>
                {\it DAF.below.cutoff} {\it DAF.above.cutoff}
#>
#> -----
#> Neutral class
                            27
#> Selected class
                             10
                                              6
#> $`Population = FR`$`MKT tables`$`MKT standard table`
#>
#>
                Polymorphism Divergence
                      45
16
#> Neutral class
                                 13
#> Selected class
#>
#> $`Population = FR`$Fractions
#> 0 0.05
#> d 0.91129141 0.909073698 0.909073698
#> f 0.08870859 0.088708587 0.088708587
#> b 0.00000000 0.002217715 0.002217715
#>
#>
#> $`Population = RAL`
#> $`Population = RAL`$Results
              alpha.symbol Fishers exact test P-value
\# Cutoff = 0 -1.19780220 0.03712979
#> Cutoff = 0.05 -0.04395604
                                       1.00000000
#> Cutoff = 0.2 -0.37362637
                                       0.44760661
#>
#> $`Population = RAL`$`Divergence metrics`
#> $`Population = RAL`$`Divergence metrics`$`Global metrics`
#> Ka Ks omega
#> 1 0.003767024 0.05226481 0.07207573
#> $`Population = RAL`$`Divergence metrics`$`Estimates by cutoff`
              omegaA.symbol omegaD.symbol
```

```
\#> Cutoff = 0   -0.086332464   0.15840819   \#> Cutoff = 0.05   -0.003168164   0.07524389
#> Cutoff = 0.2 -0.026929392 0.09900512
#>
#> $`Population = RAL`$`MKT tables`
#> $`Population = RAL`$`MKT tables`$`Number of segregating sites by DAF category - Cutoff = 0`
#>
#>
#> Table: cutoff
#>
                 {\it DAF.below.cutoff} {\it DAF.above.cutoff}
#> -----
#> Neutral class
                              0
#> Selected class
                                                40
#>
\# $`Population = RAL`$`MKT tables`$`Number of segregating sites by DAF category - Cutoff = 0.05`
#>
#>
#> Table: cutoff
#>
          {\it DAF.below.cutoff} {\it DAF.above.cutoff}
#> ------
#> Neutral class
                              20
                                                43
#> Selected class
                              34
                                                 6
#> $`Population = RAL`$`MKT tables`$`Number of segregating sites by DAF category - Cutoff = 0.2`
#>
#>
#> Table: cutoff
#>
#>
                {\it DAF.below.cutoff} {\it DAF.above.cutoff}
#> ------
#> Neutral class
                              30
                                               33
#> Selected class
                               34
                                                 6
#> $`Population = RAL`$`MKT tables`$`MKT standard table`
#>
#>
                 Polymorphism Divergence
#> -----
                         63
#> Neutral class
                                     45
#> Selected class
                          40
#>
#>
#> $`Population = RAL`$Fractions
#> 0 0.05 0.2
#> d 0.8415918 0.84039746 0.84178039
#> f 0.1584082 0.07524389 0.09900512
#> b 0.0000000 0.08435865 0.05921449
#>
#>
#> $`Population = ZI`
#> $`Population = ZI`$Results
```

```
#> alpha.symbol Fishers exact test P-value 
#> Cutoff = 0 -0.5279503 0.2545795
#> Cutoff = 0.05 -0.1180124
                                          0.8623250
#> Cutoff = 0.2 -0.2670807
                                           0.6114433
#> $`Population = ZI`$`Divergence metrics`
#> $`Population = ZI`$`Divergence metrics`$`Global metrics`
#> Ka Ks omega
#> 1 0.003765933 0.04524362 0.08323677
#>
#> $`Population = ZI`$`Divergence metrics`$`Estimates by cutoff`
               omegaA.symbol\ omegaD.symbol
#> Cutoff = 0 -0.043944879 0.12718165
#> Cutoff = 0.05 -0.009822973 0.09305974
#> Cutoff = 0.2 -0.022230939 0.10546771
#>
#>
#> $`Population = ZI`$`MKT tables`
#> $`Population = ZI`$`MKT tables`$`Number of segregating sites by DAF category - Cutoff = 0`
#>
#> Table: cutoff
#>
                  DAF.below.cutoff DAF.above.cutoff
#> ------
#> Neutral class
                               0
                                               161
#> Selected class
                                0
                                                82
#> $`Population = ZI`$`MKT tables`$`Number of segregating sites by DAF category - Cutoff = 0.05`
#>
#> Table: cutoff
#>
#>
                 DAF.below.cutoff DAF.above.cutoff
#> -----
                             108
#> Neutral class
                                                53
#> Selected class
                               77
#>
\# $`Population = ZI`$`MKT tables`$`Number of segregating sites by DAF category - Cutoff = 0.2`
#>
#>
#> Table: cutoff
#>
                 DAF.below.cutoff DAF.above.cutoff
#> Neutral class
                             125
                                                 36
                               78
#> Selected class
                                                  4
#>
#> $`Population = ZI`$`MKT tables`$`MKT standard table`
#>
#>
                 Polymorphism Divergence
#> Neutral class
                           161
                                       39
```

```
#> Selected class 82 13
#>
#>
#>
#> $`Population = ZI`$Fractions
#> 0 0.05 0.2
#> d 0.8728183 0.87282798 0.87229814
#> f 0.1271817 0.09305974 0.10546771
#> b 0.0000000 0.03411227 0.0223415
```

Example 2

#> 3

b 0.000000

In this second example, genes from RAL population are groupped in 2 recombination bins, using recombination values from Comeron et al (ref). The test used is iMK, with xlow and xhigh values set to 0 and 0.9, respectively.

```
geneList <- c("FBgn0053196", "FBgn0086906", "FBgn0261836", "FBgn0031617", "FBgn0260965",
              "FBgn0028899", "FBgn0052580", "FBgn0036181", "FBgn0263077", "FBgn0013733",
              "FBgn0031857", "FBgn0037836")
PopFlyAnalysis(genes=geneList , pops=c("RAL"), recomb=T, bins=2, test="iMK", xlow=0, xhigh=0.9)
#> [1] "Population = RAL"
#> [1] "Recombination bin = 1"
#> [1] "Recombination bin = 2"
#> $`Population = RAL`
#> $`Population = RAL`$`Recombination bin = 1`
#> $`Population = RAL`$`Recombination bin = 1`$`Asymptotic MK table`
#>
          model a
                            b c alpha_asymptotic CI_low CI_high
#> 1 exponential 0.4626 -0.8511 14.8205
                                                 0.4626 -0.9731 32.4937
     alpha_original
#> 1
            0.2489
#>
#> $`Population = RAL`$`Recombination bin = 1`$`Fractions of sites`
     Type Fraction
       d 0.6515607
#> 1
#> 2
        f 0.3484393
        ь 0.0000000
#> 3
#>
#> $`Population = RAL`$`Recombination bin = 1`$`Recombination bin Summary`
    numGenes minRecomb medianRecomb meanRecomb maxRecomb
#> 1
            6 0.6230327
                            1.216535
                                        1.20733 1.759126
#>
#>
#> $`Population = RAL`$`Recombination bin = 2`
#> $`Population = RAL`$`Recombination bin = 2`$`Asymptotic MK table`
                                    c alpha_asymptotic CI_low CI_high
                     \boldsymbol{a}
                             b
                                                 0.6628 -0.2964
#> 1 exponential 0.6632 -0.5395 7.3407
#>
     alpha original
#> 1
             0.426
\# $`Population = RAL`$`Recombination bin = 2`$`Fractions of sites`
#>
     Type Fraction
#> 1
       d 0.727223
#> 2
       f 0.272777
```

#>

#> \$`Population = RAL`\$`Recombination bin = 2`\$`Recombination bin Summary`

#> numGenes minRecomb medianRecomb meanRecomb maxRecomb

#> 1 6 2.277748 3.518252 3.398377 4.104627