# Package 'iMKT'

December 30, 2017

Title McDonald and Kreitman Test and its extensions calculation

Version 0.1.1
Date 2017-12-01
<b>Description</b> McDonald and Kreitman Test and its extensions.
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License GPL-3
Encoding UTF-8
LazyData true
RoxygenNote 6.0.1
<b>Depends</b> R ( $>= 3.3$ ), ggplot2,
Imports knitr, utils, stats, cowplot, reshape2, nls2, MASS, ggthemes
VignetteBuilder knitr
NeedsCompilation no
R topics documented:
i topics documented.
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#### **Description**

MKT calculation using asymptoticMK method (Messer and Petrov 2012 PNAS; Haller and Messer 2017 G3)

## Usage

```
asymptoticMK(daf, divergence, xlow, xhigh, seed)
```

## **Arguments**

daf data frame containing DAF, Pi and I	P0 values
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divergence data frame containing divergent and analyzed sites for selected (i) and neutral

(0) classes

xlow lower limit for asymptotic alpha fit xhigh higher limit for asymptotic alpha fit

seed value (optional). No seed by default

#### Details

In the standard McDonald and Kreitman test, the estimate of adaptive evolution (alpha) can be easily biased by the segregation of slightly deleterious non-synonymous substitutions. Specifically, slightly deleterious mutations contribute to polymorphism but not to divergence, and thus, lead to an underestimation of alpha. Messer and Petrov proposed a simple asymptotic extension of the MK test that yields accurate estimates of alpha. Briefly, this method first estimates alpha for each DAF category using its specific Pi and P0 values and then fits an exponential function to this values, of the form: alphaFit(x) = x + x

## Value

Estimation of asymptotic alpha and details about the model fit (function parameters, confidence intervals, etc.)

#### **Examples**

```
asymptoticMK(myDafData, myDivergenceData, xlow=0, xhigh=0.9)
```

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heckInput	Check input data

## **Description**

Check input data and return detailed errors when it is malformed.

## Usage

```
checkInput(daf, divergence, xlow, xhigh)
```

## **Arguments**

daf data frame containing DAF, Pi and P0 values

divergence data frame containing divergent and analyzed sites for selected (i) and neutral

(0) classes

xlow lower limit for asymptotic alpha fit xhigh higher limit for asymptotic alpha fit

#### **Details**

Check input data used in most package's functions (arguments daf, divergence, xlow and xhigh) and return a brief description of the error(s) found. This function is called within each analysis function (standardMK, FWW, DGRP, asymptoticMK, iMK) and if data does not pass checkInput() without errors, the requested analysis is not performed.

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#### **Description**

MKT calculation using all methodologies included in the package: standardMK, FWW, DGRP, asymptoticMK, iMK.

#### Usage

```
completeMK(daf, divergence, xlow, xhigh, seed)
```

## **Arguments**

daf	data frame containing DAF, Pi and P0 values
divergence	data frame containing divergent and analyzed sites for selected (i) and neutral (0) classes
xlow	lower limit for asymptotic alpha fit
xhigh	higher limit for asymptotic alpha fit
seed	seed value (optional). No seed by default

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#### **Details**

Perform all MKT derived methodologies at the same time using the same input parameters.

#### Value

List with all MKT results: standardMK, FWW, DGRP, asymptoticMK, iMK

#### **Examples**

```
completeMK(myDafData, myDivergenceData, 0, 0.9)
```

DGRP

DGRP correction method

#### **Description**

MKT calculation corrected using DGRP method (Mackay et al. 2012 Nature).

#### Usage

```
DGRP(daf, divergence, list_cutoffs = c(0, 0.05, 0.1), plot = FALSE)
```

## **Arguments**

daf data frame containing DAF, Pi and P0 values

divergence data frame containing divergent and analyzed sites for selected (i) and neutral

(0) classes

list\_cutoffs list of cutoffs to use (optional). Default cutoffs are: 0, 0.05, 0.1

plot report plot (optional). Default is FALSE

#### **Details**

In the standard McDonald and Kreitman test, the estimate of adaptive evolution (alpha) can be easily biased by the segregation of slightly deleterious non-synonymous substitutions. Specifically, slightly deleterious mutations contribute to polymorphism but not to divergence, and thus, lead to an underestimation of alpha. Because adaptive mutations and weakly deleterious selection act in opposite directions on the MKT, alpha and the fraction of substitutions that are slighlty deleterious, b, will be both underestimated when both selection regimes occur. To take adaptive and slighlty deleterious mutations mutually into account, Pi, the count off segregatning sites in class i, should be separated into the number of neutral variants and the number of weakly deleterious variants, Pi = Pineutral + Pi weak del. Alpha is then estimated as 1-(Pineutral/P0)(D0/Di). As weakly deleterious mutations tend to segregate at low frequencies, neutral and weakly deleterious fractions from Pi can be estimated based on any frequency cutoff established.

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#### Value

MKT corrected by the DGRP method. List with alpha results, graph (optional), divergence metrics, MKT tables and negative selection fractions

## **Examples**

```
## Using default cutoffs
DGRP(myDafData, myDivergenceData)
## Using custom cutoffs and rendering plot
DGRP(myDafData, myDivergenceData, c(0.05, 0.1, 0.15), plot=TRUE)
```

FWW

FWW correction method

#### **Description**

MKT calculation corrected using FWW method (Fay et al. 2001 Genetics).

#### Usage

```
FWW(daf, divergence, list_cutoffs = c(0, 0.05, 0.1), plot = FALSE)
```

## **Arguments**

daf data frame containing DAF, Pi and P0 values

divergence data frame containing divergent and analyzed sites for selected (i) and neutral

(0) classes

list\_cutoffs list of cutoffs to use (optional). Default cutoffs are: 0, 0.05, 0.1

plot report plot (optional). Default is FALSE

## **Details**

In the standard McDonald and Kreitman test, the estimate of adaptive evolution (alpha) can be easily biased by the segregation of slightly deleterious non-synonymous substitutions. Specifically, slightly deleterious mutations contribute to polymorphism but not to divergence, and thus, lead to an underestimation of alpha. Because they tend to segregate at lower frequencies than do neutral mutations, they can be partially controlled by removing low frequency polymorphisms from the analysis. This is known as the FWW method.

## Value

MKT corrected by the FWW method. List with alpha results, graph (optional), divergence metrics, MKT tables and negative selection fractions

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## **Examples**

```
## Using default cutoffs
FWW(myDafData, myDivergenceData)
## Using custom cutoffs and rendering plot
FWW(myDafData, myDivergenceData, c(0.05, 0.1, 0.15), plot=TRUE)
```

iMK

integrative MKT method

### **Description**

iMK: MKT using asymptoticMK method and estimation of negative selection fractions (d, b, f)

#### Usage

```
iMK(daf, divergence, xlow, xhigh, seed, plot = FALSE)
```

## **Arguments**

daf	data frame containing DAF, Pi and P0 values
divergence	data frame containing divergent and analyzed sites for selected (i) and neutral (0) classes
xlow	lower limit for asymptotic alpha fit
xhigh	higher limit for asymptotic alpha fit
seed	seed value (optional). No seed by default
plot	report plots of daf, alpha and negative selection fractions (optional). Default is

## FALSE

## **Details**

The integrative MKT (iMKT) allows the estimation of the rate of adaptive evolution (alpha) and the diverse negative selection regimens. iMKT uses asymptotic MK method (Messer and Petrov 2012 PNAS; Haller and Messer 2017 G3) to estimate alpha and the diverse negative selection fractions (d: strongly deleterious, b: weakly deleterious, f: neutral), based on the assumption that weakly deleterious mutations usually do not reach high allele frequencies and therefore, produce the underestimation of alpha at low DAF categories. The fraction of strongly deleterious mutations is estimated as the difference between neutral (0) and selected (i) polymorphic sites relative to the number of analyzed sites: d = 1 - (P0/m0 / Pi/mi). The fraction of weakly deleterious sites (b) corresponds to the relative proportion of selected polymorphic sites that cause the underestimation of alpha at low DAF categories. Finally, the fraction of neutral sites (f) is estimated as: f = 1 - d - b.

#### Value

iMKT method. List with asymptotic MK table and values, fractions of sites and graphs of DAF, asymptotic alpha model and negative selection fractions (optional).

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### **Examples**

```
## Without plot
iMK(myDafData, myDivergenceData, xlow=0, xhigh=0.9)
## With plot
iMK(myDafData, myDivergenceData, xlow=0, xhigh=0.9, plot=TRUE)
```

loadPopFly

Load PopFly dataset

#### **Description**

Load PopFly dataset with information regarding protein coding gene annotations.

## Usage

loadPopFly()

## **Details**

This function loads PopFly data (Hervas et al. 2017 Bioinformatics, http://popfly.uab.cat/) into the current workspace. Data is stored in a dataframe named PopFlyData, which includes population genetics estimates (nucleotide diversity, divergence, basic tests of neutrality, recombination rates, etc.) regarding each protein coding gene for 16 worldwide wild-derived Drosophila melanogaster populations from the Drosophila Genome Nexus project (Lack et al. 2015 Genetics, Lack et al. 2016 MBE).

## Value

PopFlyData object loaded in the workspace

#### **Examples**

```
\mbox{\tt \#\#} Load PopFly data if necessary. This process may take several seconds to complete. \mbox{\tt \#} loadPopFly()
```

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loadPopHuman

Load PopHuman dataset

#### Description

Load PopHuman dataset with information regarding protein coding gene annotations.

#### Usage

loadPopHuman()

#### **Details**

This function loads PopHuman data (Mulet et al. 2017 NAR, http://pophuman.uab.cat/) into the current workspace. Data is stored in a dataframe named PopHumanData, which includes population genetics estimates (nucleotide diversity, divergence, basic tests of neutrality, recombination rates, etc.) regarding each protein coding gene for 26 worldwide Homo sapiens populations from the 1000 Genomes Project (The 1000 Genomes Project Consortium 2012 Nature, The 1000 Genomes Project Consortium 2015 Nature).

#### **Examples**

## Load PopHuman data if necessary. This process may take several seconds to complete.
# loadPopHuman()

myDafData

Example data frames

## Description

Data frame containing polymorphism sample data

- daf. derived allele frequency (DAF) categories
- Pi. number of selected (i) polymorphic sites for each daf category
- P0. number of neutral (0) polymorphic sites for each daf category

## Usage

myDafData

#### **Format**

A data frame containing polymorphic sites for selected (i) and neutral (0) classes at different DAF categories

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myDivergenceData	Example data frames	

#### **Description**

Data frame containing divergence sample data

- mi. number of selected (i) analyzed sites
- Di. number of selected divergent sites
- m0. number of neutral (0) analyzed sites
- D0. number of neutral divergent sites

## Usage

myDivergenceData

#### **Format**

A data frame containing divergent and analyzed sites for selected (i) and neutral (0) classes

PopFlyAnalysis	PopFlyAnalysis		
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## **Description**

Perform any MK test using a subset of PopFly data defined by custom genes and populations lists

## Usage

```
PopFlyAnalysis(genes = c("gene1", "gene2", "..."), pops = c("pop1", "pop2",
    "..."), recomb = TRUE/FALSE, bins = 0, test = c("standardMK", "DGRP",
    "FWW", "asymptoticMK", "iMK"), xlow = 0, xhigh = 1)
```

## Arguments

genes	list of genes
pops	list of populations
recomb	group genes according to recombination values (must specify number of bins). TRUE/FALSE. Recomb values (cM/Mb) from Comeron et al. 2012.
bins	number of recombination bins to compute (mandatory if recomb = TRUE)
test	which test to perform. Options include: standardMK (default), DGRP, FWW, asymptoticMK, iMK
xlow	lower limit for asymptotic alpha fit (default=0)
xhigh	higher limit for asymptotic alpha fit (default=1)

#### **Details**

Recombination values (recomb=T) from Comeron et al. 2012 (reference!)

#### Value

None

#### **Examples**

```
## List of genes
# mygenes <- c("FBgn0053196", "FBgn0086906", "FBgn0261836", "FBgn0031617",
# "FBgn0260965", "FBgn0028899", "FBgn0052580", "FBgn0036181",
# "FBgn0263077", "FBgn0013733", "FBgn0031857", "FBgn0037836")
## Perform analyses
# PopFlyAnalysis(genes=mygenes , pops=c("RAL","ZI"), recomb=F, test="iMK", xlow=0, xhigh=0.9)
# PopFlyAnalysis(genes=mygenes , pops=c("RAL","ZI"), recomb=T, bins=3, test="DGRP")</pre>
```

PopHumanAnalysis

**PopHumanAnalysis** 

## **Description**

Perform any MK test using a subset of PopHuman data defined by custom genes and populations lists

## Usage

```
PopHumanAnalysis(genes = c("gene1", "gene2", "..."), pops = c("pop1",
   "pop2", "..."), recomb = TRUE/FALSE, bins = 0, test = c("standardMK",
   "DGRP", "FWW", "asymptoticMK", "iMK"), xlow = 0, xhigh = 1)
```

## Arguments

genes	list of genes
pops	list of populations
recomb	group genes according to recombination values (must specify number of bins). TRUE/FALSE. Recomb values (cM/Mb) from Bheller et al.
bins	number of recombination bins to compute (mandatory if recomb = TRUE)
test	which test to perform. Options include: standardMK (default), DGRP, FWW, asymptoticMK, iMK
xlow	lower limit for asymptotic alpha fit (default=0)
xhigh	higher limit for asymptotic alpha fit (default=1)

#### **Details**

Recombination values (recomb=T) from Bheller et al. (reference!)

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#### Value

None

#### **Examples**

```
## List of genes
# mygenes <- c("AHNAK2","MUC5B","MUC4","TTN","MUC16","PLIN4",
# "OBSCN","PLEC","MUC12","PKD1","LAMA5","HELZ2")
## Perform analyses
# PopHumanAnalysis(genes=mygenes , pops=c("CEU","YRI"), recomb=F, test="standardMK")
# PopHumanAnalysis(genes=mygenes , pops=c("CEU"), recomb=T, bins=3, test="DGRP")</pre>
```

standardMK

Standard MKT

#### **Description**

Standard MKT calculation (McDonald and Kreitman 1991 Nature).

#### Usage

```
standardMK(daf, divergence)
```

#### Arguments

daf data frame containing DAF, Pi and P0 values

divergence data frame containing divergent and analyzed sites for selected (i) and neutral

(0) classes

#### **Details**

The standard McDonald and Kreitman test (MKT) is used to detect the signature of selection at the molecular level. The MKT compares the amount of variation within a species (polymorphism, P) to the divergence (D) between species at two types of sites, one of which is putatively netral and used as the reference to detect selection at the other type of site. In the standard MKT, these sites are synonymous (putatively neutral, 0) and non-synonymous sites (selected sites, i) in a coding region. Under strict neutrality, the ratio of the number of selected and neutral polymorphic sites (Pi/P0) is equal to the ratio of the number of selected and neutral divergence sites (Di/D0). The null hypothesis of neutrality is rejected in a MKT when Di/D0 > Pi/P0. The excess of divergence relative to polymorphism for class i, is interpreted as adaptive selection for a subset of sites i. The fraction of adaptive fixations (alpha) is estimated from 1-(Pi/P0)(Ds/Dn). The significance of the test can be assessed with a Fisher exact test.

#### Value

Standard MKT. List with alpha estimate, Fisher's exact test p-value, MKT table and divergence metrics.

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## **Examples**

standardMK(myDafData, myDivergenceData)

 $the {\tt mePublication}$ 

ggplot Theme for publication ready plots

## Description

Theme with the configuration and parameters necessary to generate publication ready plots using ggplot

## Usage

```
themePublication(base_size = 14, base_family = "sans")
```

## Arguments

base\_size base size required from themePublication

base\_family font to load in themePublication

## **Details**

Theme used for plot images developed by Koundinya Desiraju (04/07/2015). Code adapted from http://rpubs.com/Koundy/71792.

#### Value

plot theme

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