

# Package ‘iMKT’

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**Title** Integrative McDonald and Kreitman Test

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**Description** iMKT is an R package to compute the McDonald and Kreitman test (McDonald and Kreitman 1991 Nature) on polymorphism and divergence genomic data provided by the user or automatically downloaded from PopFly (Hervas et al. 2017 Bioinformatics) or PopHuman (Casillas et al. 2018 Nucleic Acids Res.). It includes five MK derived methodologies which allow inferring the rate of adaptive evolution ( $\alpha$ ) as well as the fraction of strongly deleterious (d), weakly deleterious (b), and neutral (f) sites.

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**License** GPL-3

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**LazyData** true

**RoxygenNote** 6.0.1

**Depends** R (>= 3.3), ggplot2

**Imports** knitr, utils, stats, cowplot, reshape2, nls2, MASS, ggthemes

**VignetteBuilder** knitr

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asymptoticMK	<i>Asymptotic MKT method</i>
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## Description

MKT calculation using asymptoticMK method (Messer and Petrov 2012 PNAS; Haller and Messer 2017 G3)

## Usage

```
asymptoticMK(daf, divergence, xlow, xhigh, seed)
```

## Arguments

daf	data frame containing DAF, Pi and P0 values
divergence	data frame containing divergent and analyzed sites for selected (i) and neutral (0) classes
xlow	lower limit for asymptotic alpha fit
xhigh	higher limit for asymptotic alpha fit
seed	seed value (optional). No seed by default

## Details

In the standard McDonald and Kreitman test, the estimate of adaptive evolution (alpha) can be easily biased by the segregation of slightly deleterious non-synonymous substitutions. Specifically, slightly deleterious mutations contribute more to polymorphism than they do to divergence, and thus, lead to an underestimation of alpha. Messer and Petrov proposed a simple asymptotic extension of the MK test that yields accurate estimates of alpha. Briefly, this method first estimates alpha for each DAF category using its specific Pi and P0 values and then fits an exponential function to this values, of the form:  $\alpha \text{ Fit}(x) = a + b \exp(-cx)$ . Although the exponential function is generally expected to provide the best fit, a linear function is also fit to the data, of the form:  $\alpha \text{ Fit}(x) = a + bx$ . Finally, the asymptotic alpha estimate is obtained by extrapolating the value of this function to  $x = 1$ :  $\alpha \text{ Asymptotic} = \alpha \text{ Fit}(x=1)$ . The exponential fit is always reported, except if the exponential fit fails to converge or if the linear fit is superior according to AIC. The code of this function is adapted from Haller and Messer 2017 G3 (<http://github.com/MesserLab/asymptoticMK>).

## Value

Estimation of asymptotic alpha and details about the model fit (function parameters, confidence intervals, etc.)

**Examples**

```
asymptoticMK(myDafData, myDivergenceData, xlow=0, xhigh=0.9)
```

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checkInput	<i>Check input data</i>
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**Description**

Check input data and return detailed errors when it is malformed

**Usage**

```
checkInput(daf, divergence, xlow, xhigh)
```

**Arguments**

daf	data frame containing DAF, Pi and P0 values
divergence	data frame containing divergent and analyzed sites for selected (i) and neutral (0) classes
xlow	lower limit for asymptotic alpha fit
xhigh	higher limit for asymptotic alpha fit

**Details**

Check input data used in most package's functions (arguments daf, divergence, xlow and xhigh) and return a brief description of the error(s) found. This function is called within each analysis function (standardMK, FWW, DGRP, asymptoticMK, iMK) and if data does not pass checkInput() without errors, the requested analysis is not performed.

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completeMK	<i>Complete MK methodologies</i>
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**Description**

MKT calculation using all methodologies included in the package: standardMK, FWW, DGRP, asymptoticMK, iMK.

**Usage**

```
completeMK(daf, divergence, xlow, xhigh, seed)
```

**Arguments**

daf	data frame containing DAF, Pi and P0 values
divergence	data frame containing divergent and analyzed sites for selected (i) and neutral (0) classes
xlow	lower limit for asymptotic alpha fit
xhigh	higher limit for asymptotic alpha fit
seed	seed value (optional). No seed by default

**Details**

Perform all MKT derived methodologies at once using the same input data and parameters.

**Value**

List with the diverse MKT results: standardMK, FWW, DGRP, asymptoticMK, iMK

**Examples**

```
completeMK(myDafData, myDivergenceData, xlow=0, xhigh=0.9)
```

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DGRP

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*DGRP correction method*


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**Description**

MKT calculation corrected using DGRP method (Mackay et al. 2012 Nature).

**Usage**

```
DGRP(daf, divergence, listCutoffs = c(0, 0.05, 0.1), plot = FALSE)
```

**Arguments**

daf	data frame containing DAF, Pi and P0 values
divergence	data frame containing divergent and analyzed sites for selected (i) and neutral (0) classes
listCutoffs	list of cutoffs to use (optional). Default cutoffs are: 0, 0.05, 0.1
plot	report plot (optional). Default is FALSE

Details

In the standard McDonald and Kreitman test, the estimate of adaptive evolution (alpha) can be easily biased by the segregation of slightly deleterious non-synonymous substitutions. Specifically, slightly deleterious mutations contribute more to polymorphism than they do to divergence, and thus, lead to an underestimation of alpha. Because adaptive mutations and weakly deleterious selection act in opposite directions on the MKT, alpha and the fraction of substitutions that are slightly deleterious, b, will be both underestimated when both selection regimes occur. To take adaptive and slightly deleterious mutations mutually into account, Pi, the count off segregating sites in class i, should be separated into the number of neutral variants and the number of weakly deleterious variants,  $P_i = P_{i\text{neutral}} + P_{i\text{weak del}}$ . Alpha is then estimated as  $1 - (P_{i\text{neutral}}/P_0)(D_0/D_i)$ . As weakly deleterious mutations tend to segregate at low frequencies, neutral and weakly deleterious fractions from Pi can be estimated based on any frequency cutoff established.

Value

MKT corrected by the DGRP method. List with alpha results, graph (optional), divergence metrics, MKT tables and negative selection fractions

Examples

```
## Using default cutoffs
DGRP(myDafData, myDivergenceData)
## Using custom cutoffs and rendering plot
DGRP(myDafData, myDivergenceData, c(0.05, 0.1, 0.15), plot=TRUE)
```

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FWW	<i>FWW correction method</i>
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Description

MKT calculation corrected using FWW method (Fay et al. 2001 Genetics).

Usage

```
FWW(daf, divergence, listCutoffs = c(0, 0.05, 0.1), plot = FALSE)
```

Arguments

daf	data frame containing DAF, Pi and P0 values
divergence	data frame containing divergent and analyzed sites for selected (i) and neutral (0) classes
listCutoffs	list of cutoffs to use (optional). Default cutoffs are: 0, 0.05, 0.1
plot	report plot (optional). Default is FALSE

Details

In the standard McDonald and Kreitman test, the estimate of adaptive evolution ( $\alpha$ ) can be easily biased by the segregation of slightly deleterious non-synonymous substitutions. Specifically, slightly deleterious mutations contribute more to polymorphism than they do to divergence, and thus, lead to an underestimation of  $\alpha$ . Because they tend to segregate at lower frequencies than do neutral mutations, they can be partially controlled by removing low frequency polymorphisms from the analysis. This is known as the FWW method.

Value

MKT corrected by the FWW method. List with  $\alpha$  results, graph (optional), divergence metrics, MKT tables and negative selection fractions

Examples

```
## Using default cutoffs
FWW(myDafData, myDivergenceData)
## Using custom cutoffs and rendering plot
FWW(myDafData, myDivergenceData, c(0.05, 0.1, 0.15), plot=TRUE)
```

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iMK	<i>integrative MKT method</i>
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Description

iMK: MKT using asymptoticMK method and estimation of negative selection fractions (d, b, f)

Usage

```
iMK(daf, divergence, xlow, xhigh, seed, plot = FALSE)
```

Arguments

daf	data frame containing DAF, Pi and P0 values
divergence	data frame containing divergent and analyzed sites for selected (i) and neutral (0) classes
xlow	lower limit for asymptotic alpha fit
xhigh	higher limit for asymptotic alpha fit
seed	seed value (optional). No seed by default
plot	report plots of daf, alpha and negative selection fractions (optional). Default is FALSE

## Details

The integrative MKT (iMKT) allows the estimation of the rate of adaptive evolution ( $\alpha$ ) and the diverse negative selection regimens. iMKT uses asymptotic MK method (Messer and Petrov 2012 PNAS; Haller and Messer 2017 G3) to estimate  $\alpha$  and the diverse negative selection fractions (d: strongly deleterious, b: weakly deleterious, f: neutral), based on the assumption that weakly deleterious mutations usually do not reach high allele frequencies and therefore, produce the underestimation of  $\alpha$  at low DAF categories. The fraction of strongly deleterious mutations is estimated as the difference between neutral (0) and selected (i) polymorphic sites relative to the number of analyzed sites:  $d = 1 - (P0/m0 / Pi/mi)$ . The fraction of weakly deleterious sites (b) corresponds to the relative proportion of selected polymorphic sites that cause the underestimation of  $\alpha$  at low DAF categories. Finally, the fraction of neutral sites (f) is estimated as:  $f = 1 - d - b$ . iMKT() only fits an exponential model for the computation of  $\alpha$ .

## Value

iMKT method. List with asymptotic MK table and values, fractions of sites and graphs of DAF, asymptotic  $\alpha$  model and negative selection fractions (optional).

## Examples

```
## Without plot
iMK(myDafData, myDivergenceData, xlow=0, xhigh=0.9)
## With plot
iMK(myDafData, myDivergenceData, xlow=0, xhigh=0.9, plot=TRUE)
```

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loadPopFly

*Load PopFly dataset*


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## Description

Load PopFly dataset with information regarding protein coding gene annotations

## Usage

```
loadPopFly()
```

## Details

This function loads PopFly data (Hervas et al. 2017 Bioinformatics, <http://popfly.uab.cat/>) into the current workspace. Data is stored in a dataframe named PopFlyData, which includes population genetics estimates (nucleotide diversity, divergence, basic tests of neutrality, recombination rates, etc.) regarding each protein coding gene for 16 worldwide wild-derived *Drosophila melanogaster* populations from the *Drosophila* Genome Nexus project (Lack et al. 2015 Genetics, Lack et al. 2016 MBE).

**Value**

PopFlyData object loaded in the workspace

**Examples**

```
## Load PopFly data if necessary. This process may take several seconds to complete.
# loadPopFly()
```

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loadPopHuman	<i>Load PopHuman dataset</i>
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**Description**

Load PopHuman dataset with information regarding protein coding gene annotations

**Usage**

```
loadPopHuman()
```

**Details**

This function loads PopHuman data (Mulet et al. 2017 NAR, <http://pophuman.uab.cat/>) into the current workspace. Data is stored in a dataframe named PopHumanData, which includes population genetics estimates (nucleotide diversity, divergence, basic tests of neutrality, recombination rates, etc.) regarding each protein coding gene for 26 worldwide Homo sapiens populations from the 1000 Genomes Project (The 1000 Genomes Project Consortium 2012 Nature, The 1000 Genomes Project Consortium 2015 Nature).

**Value**

PopHumanData object loaded in the workspace

**Examples**

```
## Load PopHuman data if necessary. This process may take several seconds to complete.
# loadPopHuman()
```



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`myDafData`*Sample DAF data frame*

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**Description**

Data frame containing polymorphism sample data

- daf. derived allele frequency (DAF) categories
- Pi. number of selected (i) polymorphic sites for each daf category
- P0. number of neutral (0) polymorphic sites for each daf category

**Usage**

```
myDafData
```

**Format**

A data frame containing polymorphic sites for selected (i) and neutral (0) classes at different DAF categories

---

`myDivergenceData`*Sample Divergence data frame*

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**Description**

Data frame containing divergence sample data

- mi. number of selected (i) analyzed sites
- Di. number of selected divergent sites
- m0. number of neutral (0) analyzed sites
- D0. number of neutral divergent sites

**Usage**

```
myDivergenceData
```

**Format**

A data frame containing divergent and analyzed sites for selected (i) and neutral (0) classes

---

PopFlyAnalysis

*iMKT using PopFly data*


---

## Description

Perform any MKT method using a subset of PopFly data defined by custom genes and populations lists

## Usage

```
PopFlyAnalysis(genes = c("gene1", "gene2", "..."), pops = c("pop1", "pop2",
  "..."), recomb = TRUE/FALSE, bins = 0, test = c("standardMK", "DGRP",
  "FWW", "asymptoticMK", "iMK"), xlow = 0, xhigh = 1, plot = FALSE)
```

## Arguments

genes	list of genes to analyze
pops	list of populations to analyze
recomb	group genes according to recombination values (TRUE/FALSE)
bins	number of recombination bins to compute (mandatory if recomb=TRUE)
test	which test to perform. Options include: standardMK (default), DGRP, FWW, asymptoticMK, iMK
xlow	lower limit for asymptotic alpha fit (default=0)
xhigh	higher limit for asymptotic alpha fit (default=1)
plot	report plot (optional). Default is FALSE

## Details

Execute any MKT method (standardMK, FWW, DGRP, asymptoticMK, iMKT) using a subset of PopFly data defined by custom genes and populations lists. It uses the dataframe PopFlyData, which can be already loaded in the workspace (using loadPopFly()) or is directly loaded when executing this function. It also allows deciding whether to analyze genes grouped by recombination bins or not, using recombination rate estimates from Comeron et al. 2012 Plos Genetics.

## Value

List of lists with the default test output for each selected population (and recombination bin when defined)

## Examples

```
## List of genes
mygenes <- c("FBgn0053196", "FBgn0086906", "FBgn0261836", "FBgn0031617",
  "FBgn0260965", "FBgn0028899", "FBgn0052580", "FBgn0036181",
  "FBgn0263077", "FBgn0013733", "FBgn0031857", "FBgn0037836")
## Perform analyses
```

```
PopFlyAnalysis(genes=mygenes, pops="RAL", recomb=FALSE, test="iMK", xlow=0, xhigh=0.9, plot=TRUE)
PopFlyAnalysis(genes=mygenes, pops=c("RAL","ZI"), recomb=TRUE, bins=3, test="DGRP", plot=FALSE)
```

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PopHumanAnalysis	<i>iMKT using PopHuman data</i>
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## Description

Perform any MKT method using a subset of PopHuman data defined by custom genes and populations lists

## Usage

```
PopHumanAnalysis(genes = c("gene1", "gene2", "..."), pops = c("pop1",
  "pop2", "..."), recomb = TRUE/FALSE, bins = 0, test = c("standardMK",
  "DGRP", "FWW", "asymptoticMK", "iMK"), xlow = 0, xhigh = 1,
  plot = FALSE)
```

## Arguments

genes	list of genes to analyze
pops	list of populations to analyze
recomb	group genes according to recombination values (TRUE/FALSE)
bins	number of recombination bins to compute (mandatory if recomb=TRUE)
test	which test to perform. Options include: standardMK (default), DGRP, FWW, asymptoticMK, iMK
xlow	lower limit for asymptotic alpha fit (default=0)
xhigh	higher limit for asymptotic alpha fit (default=1)
plot	report plot (optional). Default is FALSE

## Details

Execute any MKT method (standardMK, FWW, DGRP, asymptoticMK, iMKT) using a subset of PopHuman data defined by custom genes and populations lists. It uses the dataframe PopHuman-Data, which can be already loaded in the workspace (using loadPopHuman()) or is directly loaded when executing this function. It also allows deciding whether to analyze genes grouped by recombination bins or not, using recombination rate values corresponding to the sex average estimates from Bh  rer et al. 2017 Nature Commun.

## Value

List of lists with the default test output for each selected population (and recombination bin when defined)

Examples

```
## List of genes
mygenes <- c("ENSG0000011021.21_3","ENSG00000091483.6_3","ENSG00000116191.17_3",
             "ENSG00000116337.15_4","ENSG00000116584.17_3","ENSG00000116745.6_3",
             "ENSG00000116852.14_3","ENSG00000116898.11_3","ENSG00000117010.15_3",
             "ENSG00000117090.14_3","ENSG00000117222.13_3","ENSG00000117394.20_3")

## Perform analyses
PopHumanAnalysis(genes=mygenes , pops=c("CEU","YRI"), recomb=FALSE, test="standardMK")
PopHumanAnalysis(genes=mygenes , pops=c("CEU"), recomb=TRUE, bins=3, test="DGRP")
```

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standardMK	<i>Standard MKT</i>
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---

Description

Standard MKT calculation (McDonald and Kreitman 1991 Nature).

Usage

```
standardMK(daf, divergence)
```

Arguments

daf	data frame containing DAF, Pi and P0 values
divergence	data frame containing divergent and analyzed sites for selected (i) and neutral (0) classes

Details

The standard McDonald and Kreitman test (MKT) is used to detect the signature of selection at the molecular level. The MKT compares the amount of variation within a species (polymorphism, P) to the divergence (D) between species at two types of sites, one of which is putatively netral and used as the reference to detect selection at the other type of site. In the standard MKT, these sites are synonymous (putatively neutral, 0) and non-synonymous sites (selected sites, i) in a coding region. Under strict neutrality, the ratio of the number of selected and neutral polymorphic sites ( $P_i/P_0$ ) is equal to the ratio of the number of selected and neutral divergence sites ( $D_i/D_0$ ). The null hypothesis of neutrality is rejected in a MKT when  $D_i/D_0 > P_i/P_0$ . The excess of divergence relative to polymorphism for class i, is interpreted as adaptive selection for a subset of sites i. The fraction of adaptive fixations ( $\alpha$ ) is estimated from  $1-(P_i/P_0)(D_s/D_n)$ . The significance of the test can be assesed with a Fisher exact test.

Value

Standard MKT. List with  $\alpha$  estimate, Fisher’s exact test p-value, MKT table and divergence metrics.

**Examples**

```
standardMK(myDafData, myDivergenceData)
```

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themePublication	<i>ggplot Theme for publication ready plots</i>
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**Description**

Theme with the configuration and parameters necessary to generate publication ready plots using ggplot

**Usage**

```
themePublication(base_size = 14, base_family = "sans")
```

**Arguments**

base_size	base size required from themePublication
base_family	font to load in themePublication

**Details**

Theme used for plot images developed by Koundinya Desiraju (04/07/2015). Code adapted from <http://rpubs.com/Koundy/71792>.

**Value**

plot theme

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