



National Institute of Environmental Health Sciences
Your Environment. Your Health.

Pathway Analysis

Biostatistics and Bioinformatics Short Course

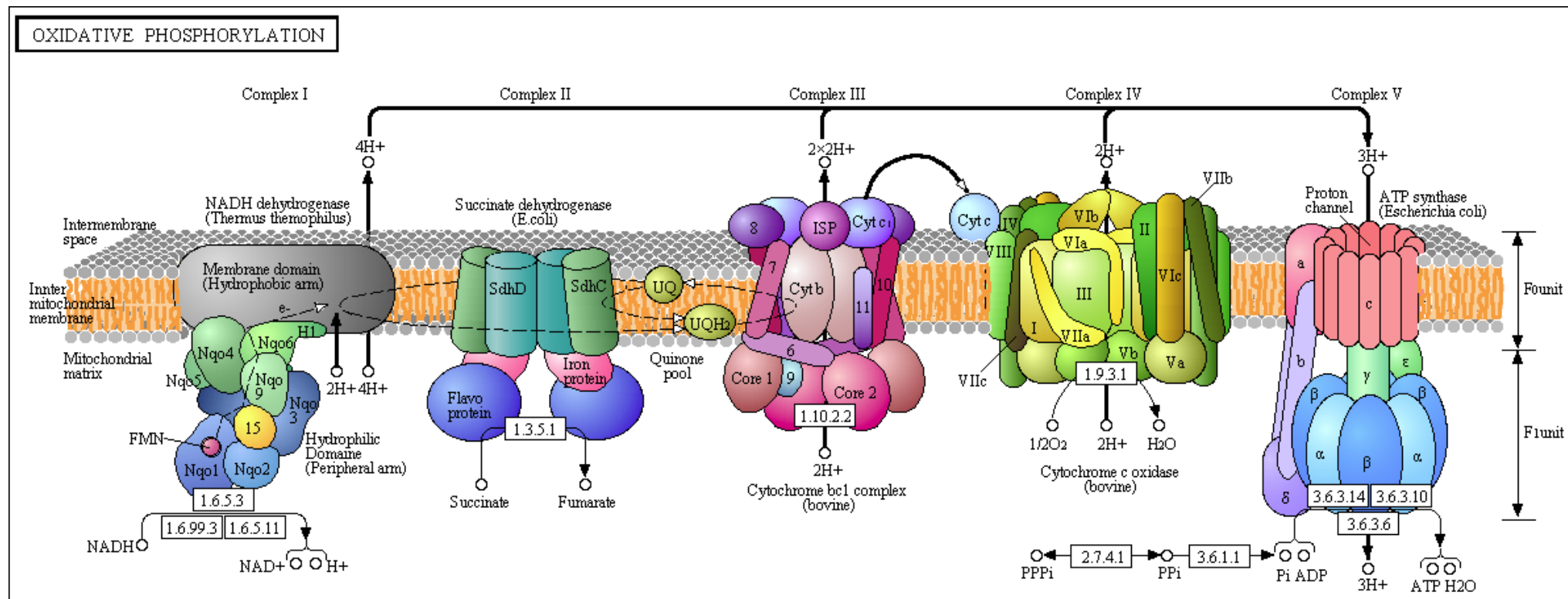
02/12/16

Brian Bennett, Ph.D.

(brian.bennett@nih.gov)

What is a Biological Pathway?

- A set of molecules in a cell that work together through a series of actions to achieve a particular outcome



What is Pathway Analysis?

- Identifying pathways whose genes are associated with a particular biological condition
- Examines the combined signal from multiple genes, as opposed to looking at individual genes separately
- Recently, this definition has been extended to include any set of genes that have some sort biological connection (gene sets)
 - Mechanistic and signaling cascades (KEGG, Biocarta)
 - Functional and biological processes (GO)
 - Associated with a disease or condition
 - Chromosomal proximity
 - Computationally derived

Advantages over Single-Gene Analysis

- Different samples from a common condition may have different key genes that are all driving changes in the same pathway



Advantages over Single-Gene Analysis

- Pathways may be perturbed by subtle changes in many genes

nature
genetics

ARTICLES

PGC-1 α -responsive genes involved in oxidative phosphorylation are coordinately downregulated in human diabetes

Vamsi K Mootha^{1,2,3,10}, Cecilia M Lindgren^{1,4,10}, Karl-Fredrik Eriksson⁴, Aravind Subramanian¹, Smita Sihag¹, Joseph Lehar¹, Pere Puigserver⁵, Emma Carlsson⁴, Martin Ridderstråle⁴, Esa Laurila⁴, Nicholas Houstis¹, Mark J Daly¹, Nick Patterson¹, Jill P Mesirov¹, Todd R Golub^{1,5}, Pablo Tamayo¹, Bruce Spiegelman⁵, Eric S Lander^{1,6}, Joel N Hirschhorn^{1,7,8}, David Altshuler^{1,2,7,9,11} & Leif C Groop^{4,11}

DNA microarrays can be used to identify gene expression changes characteristic of human disease. This is challenging, however, when relevant differences are subtle at the level of individual genes. We introduce an analytical strategy, Gene Set Enrichment Analysis, designed to detect modest but coordinate changes in the expression of groups of functionally related genes. Using this approach, we identify a set of genes involved in oxidative phosphorylation whose expression is coordinately decreased in human diabetic muscle. Expression of these genes is high at sites of insulin-mediated glucose disposal, activated by PGC-1 α and correlated with total-body aerobic capacity. Our results associate this gene set with clinically important variation in human metabolism and illustrate the value of pathway relationships in the analysis of genomic profiling experiments.

Type 2 diabetes mellitus (DM2) affects over 110 million people world-wide. One promising approach to increase power exploits the idea that

Advantages over Single-Gene Analysis

- A small list of enriched pathways may be easier to interpret than a large list of associated genes

Application of *a priori* established gene sets to discover biologically important differential expression in microarray data

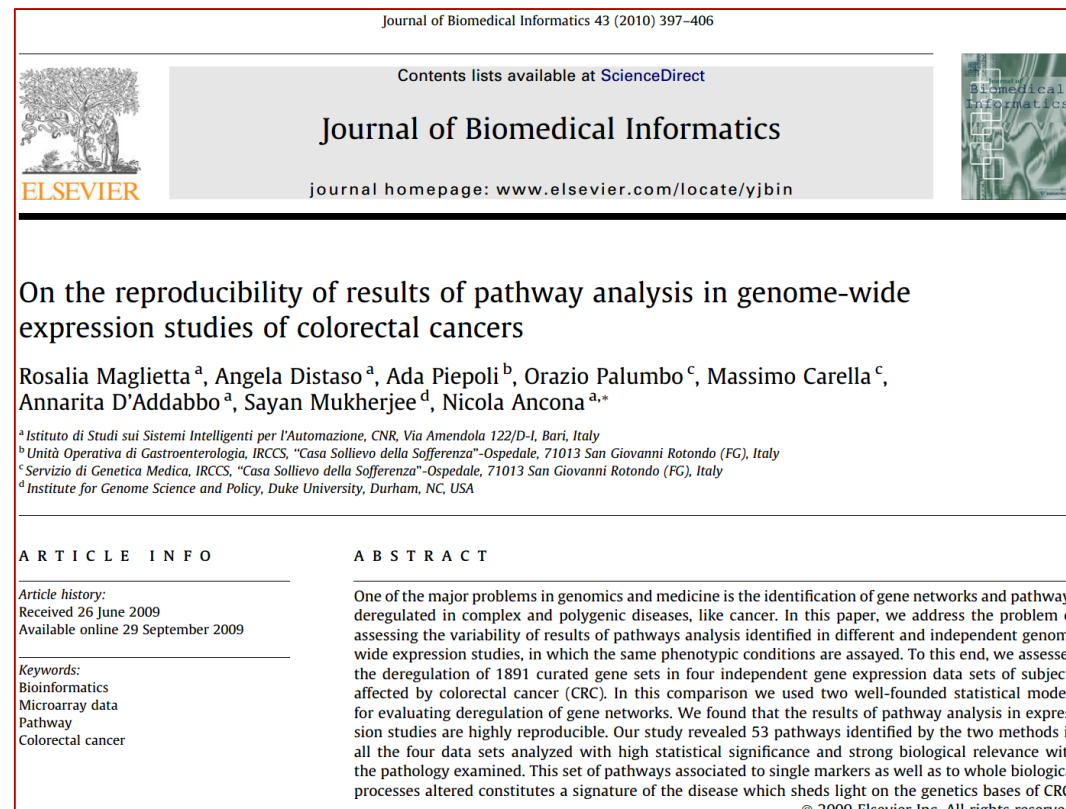
Andrea Bild*[†] and Phillip George Febbo*^{†§}

*Duke Institute for Genome Sciences and Policy and Departments of [†]Medicine and [‡]Molecular Genetics and Microbiology, Duke University Medical Center, Duke University, Durham, NC 27710

From inception, microarray analysis has facilitated discovery by associating gene expression with biological and/or clinical sample characteristics. However, cleaning biological data is a non-trivial task. The use of an established gene set (ES) that represents the difference between the observed rankings and that which would be expected assuming a random rank distribution (see figure 1 A and B in ref. 1). After establishing the ES for a given gene set, the location, Subramanian *et al.* successfully identify differential expression of genes located on the Y chromosome. In addition, a gene set containing genes known to escape X inactivation is significantly

Advantages over Single-Gene Analysis

- Pathways results from different studies may overlap better than single-gene results



Gene Expression Pathway Analysis

- Data: Gene expression data for samples in two groups (disease vs. control, treatment vs. no treatment, etc.)
- Biological Question: Which pathways are impacted by the condition or treatment?
- Statistical Question: Which pathways have more differentially expressed genes than expected by chance?

Required Data

1. Gene expression data set
2. Collection of gene sets

Molecular Signatures Database (MSigDB)

- Free database (<http://www.broadinstitute.org/gsea/msigdb>)

H **hallmark gene sets** are coherently expressed signatures derived by aggregating many MSigDB gene sets to represent well-defined biological states or processes.

C1 **positional gene sets** for each human chromosome and cytogenetic band.

C2 **curated gene sets** from online pathway databases, publications in PubMed, and knowledge of domain experts.

C3 **motif gene sets** based on conserved cis-regulatory motifs from a comparative analysis of the human, mouse, rat, and dog genomes.

C4 **computational gene sets** defined by mining large collections of cancer-oriented microarray data.

C5 **GO gene sets** consist of genes annotated by the same GO terms.

C6 **oncogenic signatures** defined directly from microarray gene expression data from cancer gene perturbations.

C7 **immunologic signatures** defined directly from microarray gene expression data from immunologic studies.

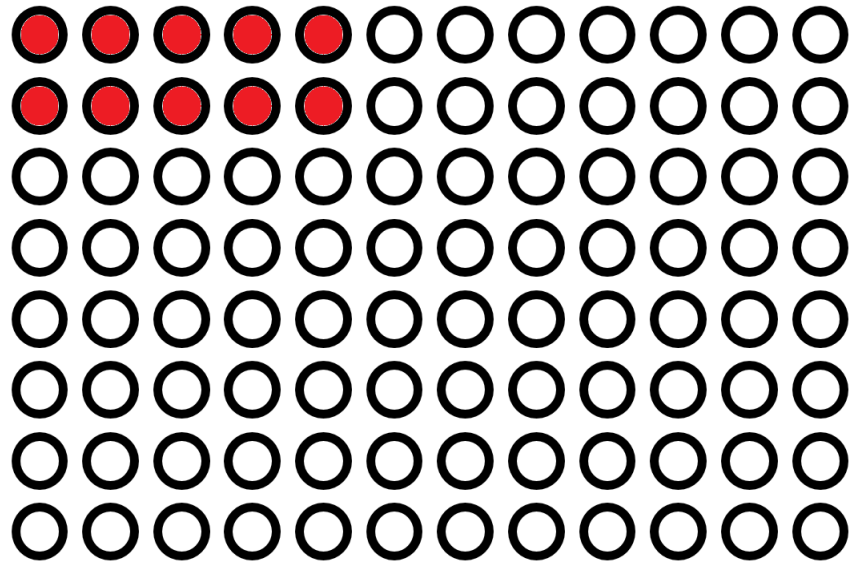
Molecular Signatures Database (MSigDB)

C2: curated gene sets (browse 4725 gene sets)	Gene sets collected from various sources such as online pathway databases, publications in PubMed, and knowledge of domain experts. The gene set page for each gene set lists its source. details	Download GMT Files original identifiers gene symbols entrez genes ids
CGP: chemical and genetic perturbations (browse 3395 gene sets)	Gene sets represent expression signatures of genetic and chemical perturbations. A number of these gene sets come in pairs: an xxx_UP (xxx_DN) gene set representing genes induced (repressed) by the perturbation. The gene set page for each gene set lists the PubMed citation on which it is based.	Download GMT Files original identifiers gene symbols entrez genes ids
CP: Canonical pathways (browse 1330 gene sets)	Gene sets from the pathway databases. Usually, these gene sets are canonical representations of a biological process compiled by domain experts. details	Download GMT Files original identifiers gene symbols entrez genes ids
CP:BIOCARTA: BioCarta gene sets (browse 217 gene sets)	Gene sets derived from the BioCarta pathway database (http://www.biocarta.com/genes/index.asp).	Download GMT Files original identifiers gene symbols entrez genes ids
CP:KEGG: KEGG gene sets (browse 186 gene sets)	Gene sets derived from the KEGG pathway database (http://www.genome.jp/kegg/pathway.html).	Download GMT Files original identifiers gene symbols entrez genes ids
CP:REACTOME: Reactome gene sets (browse 674 gene sets)	Gene sets derived from the Reactome pathway database (http://www.reactome.org/).	Download GMT Files original identifiers gene symbols entrez genes ids

Hypergeometric Test (Right-tailed)

- Parametric method
- Can also use Fisher's Exact (FE) test
- Required data:
 - List of DEGs (along with the number of total genes)
 - A gene set
- If the proportion of DEGs within the gene set is sufficiently higher than the proportion within the entire set, the gene set is considered significantly enriched

Hypergeometric Test (Right-tailed)

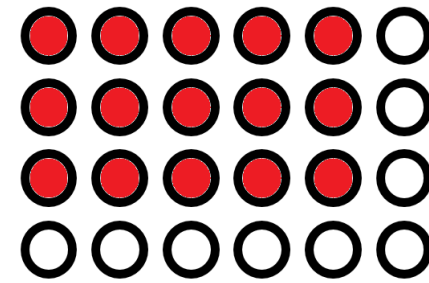
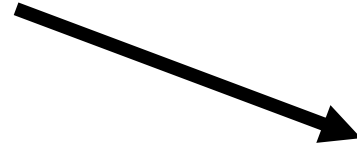


Full data

k = total DEGs = 1000

N = total overall genes = 10000

(10% are DEGs)



Gene set

x = total DEGs in gene set = 60

m = total genes in gene set = 100

(60% are DEGs)

Hypergeometric Test (Right-tailed)

- x = total DEGs in gene set = 60
- m = total genes in gene set = 100
- k = total DEGs = 1000
- N = total overall genes = 10000
- R code:
 - `phyper(x-1, m, N-m, k, lower.tail=FALSE)`
 - `fisher.test(matrix(c(x,k-x,m-x,N-m-k+x),2,2), alternative='greater')$p.value`
- P-value = 5.4×10^{-35}

Hypergeometric Test (Right-tailed)

- Used in Ingenuity Pathway Analysis (IPA)
 - Commercial software (<http://www.ingenuity.com/products/ipa>)
 - Pros:
 - Great source of clean, expertly curated gene sets
 - Cons:
 - Not free
 - Throws away information by only using DEG list

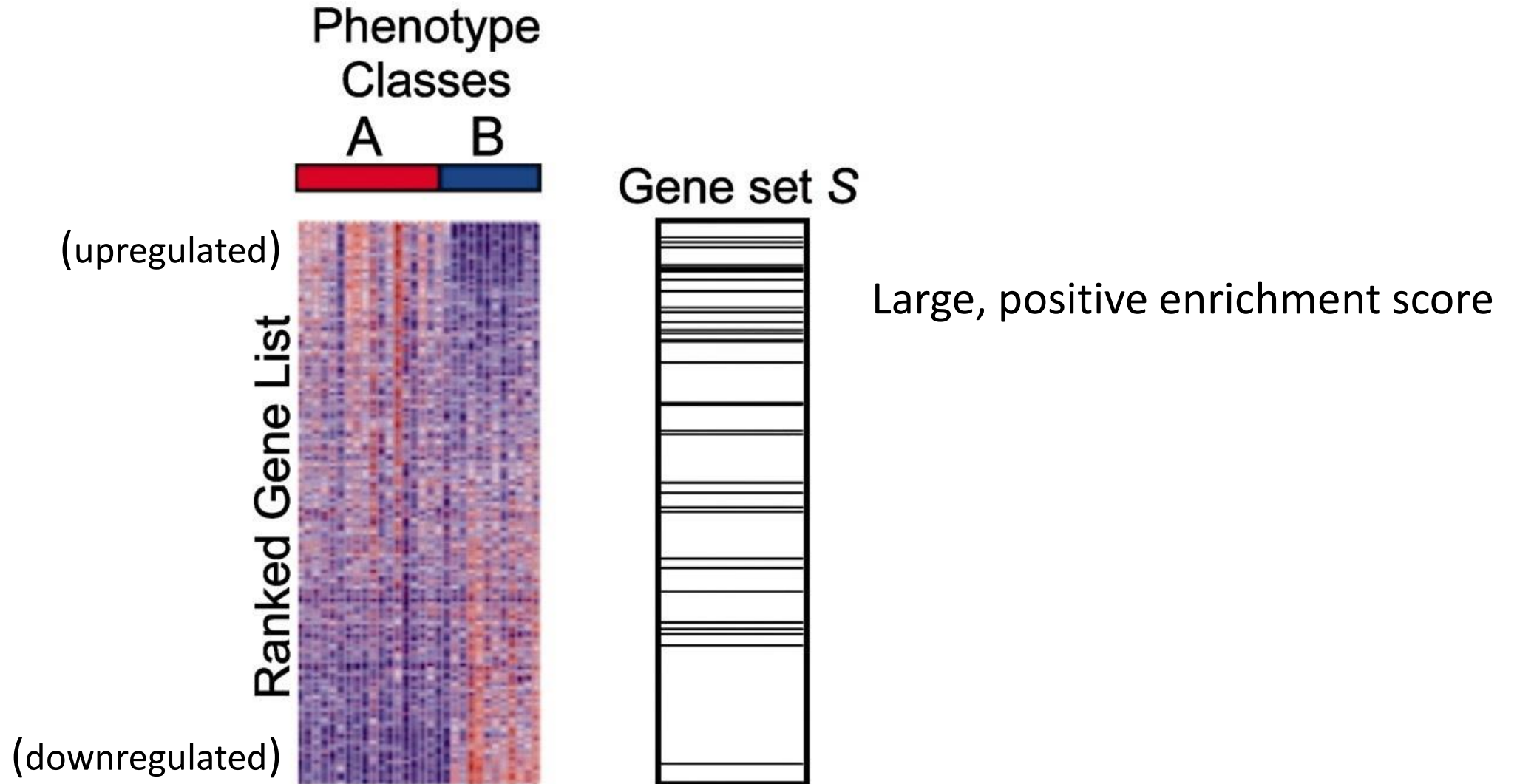
Hypergeometric Test (Right-tailed)

- Used in Database for Annotation, Visualization and Integrated Discovery (DAVID)
 - Free software (<https://david.ncifcrf.gov>)
 - Pros:
 - Easy to use (web-based)
 - Large collection of gene sets
 - Cons:
 - Gene sets are not as clean
 - Throws away information by only using DEG list

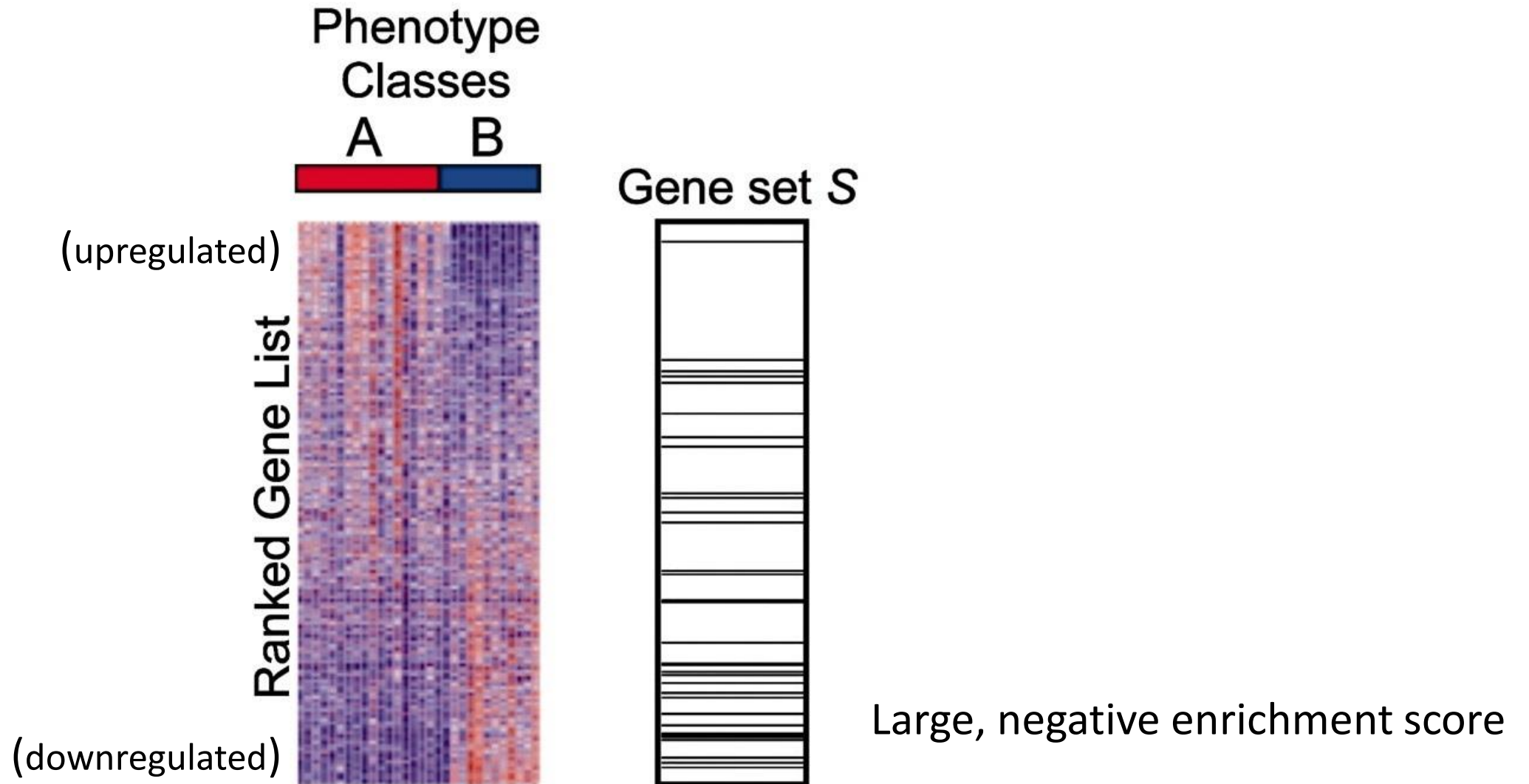
Kolmogorov–Smirnov (KS) Test

- Nonparametric method
- Required data:
 - Ranked list of genes sorted by differential expression (includes all genes)
 - A gene set
- If the genes in the gene set tend to fall near either end of the ranked list, the gene set is considered significantly enriched

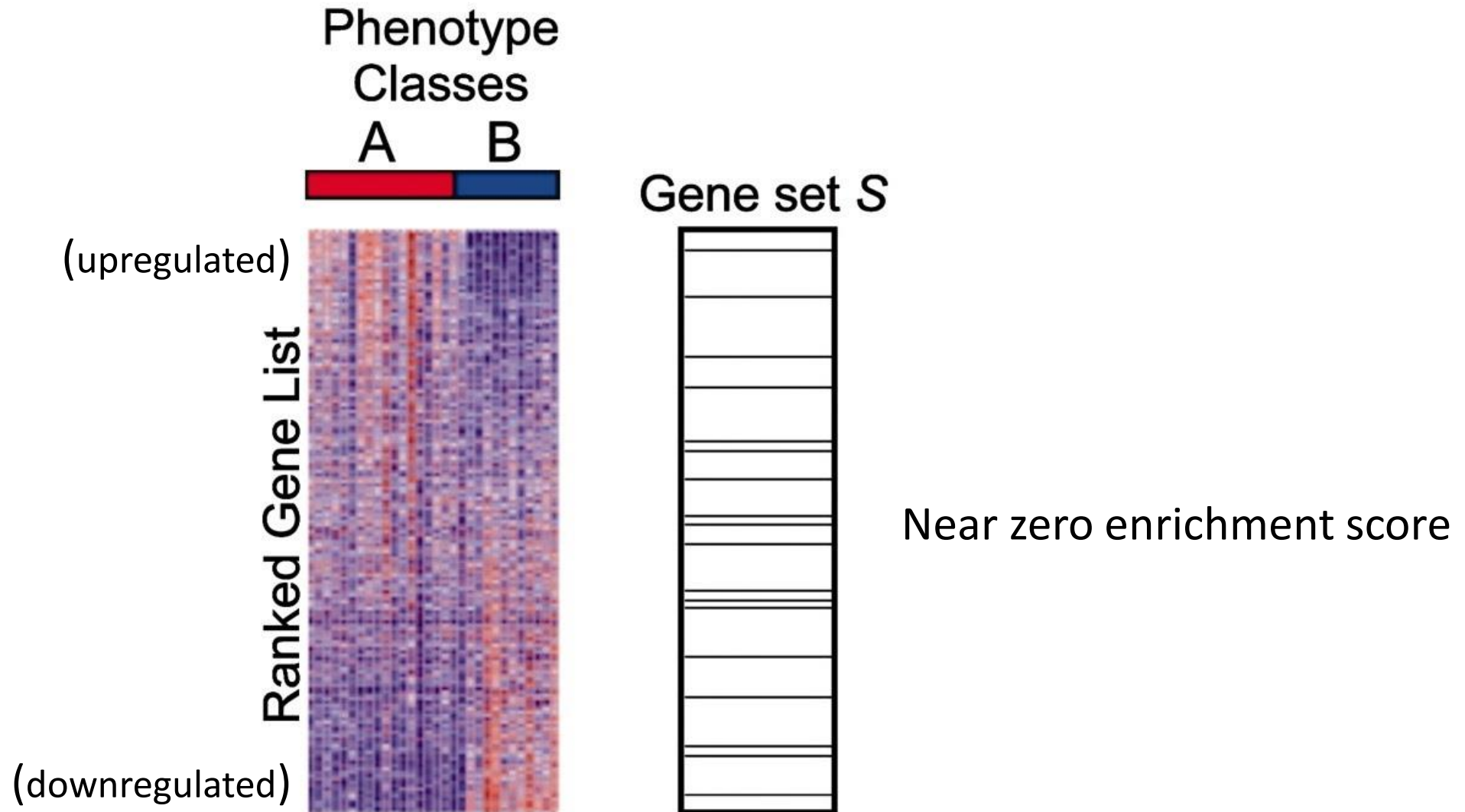
Kolmogorov–Smirnov (KS) Test



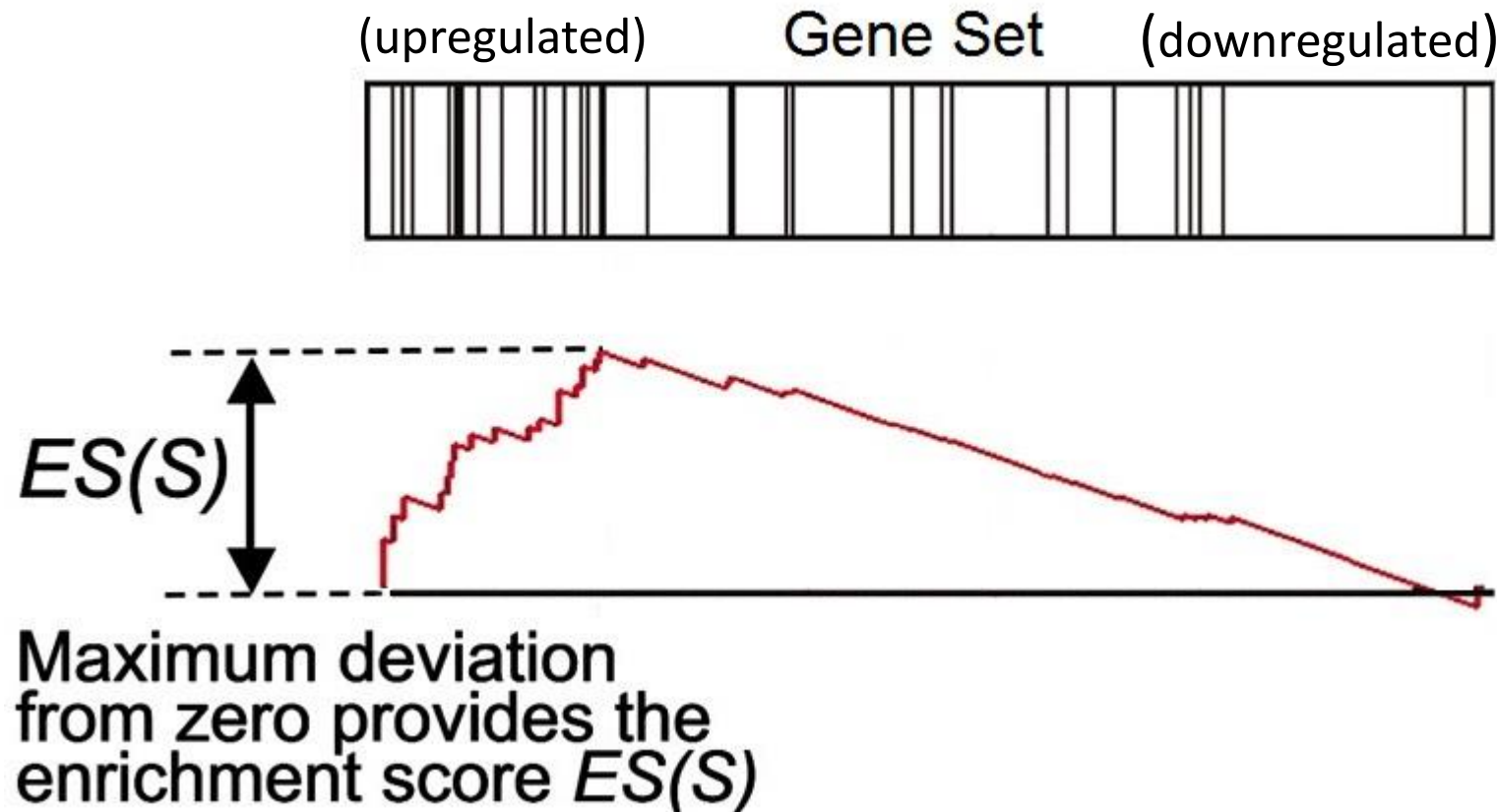
Kolmogorov–Smirnov (KS) Test



Kolmogorov–Smirnov (KS) Test

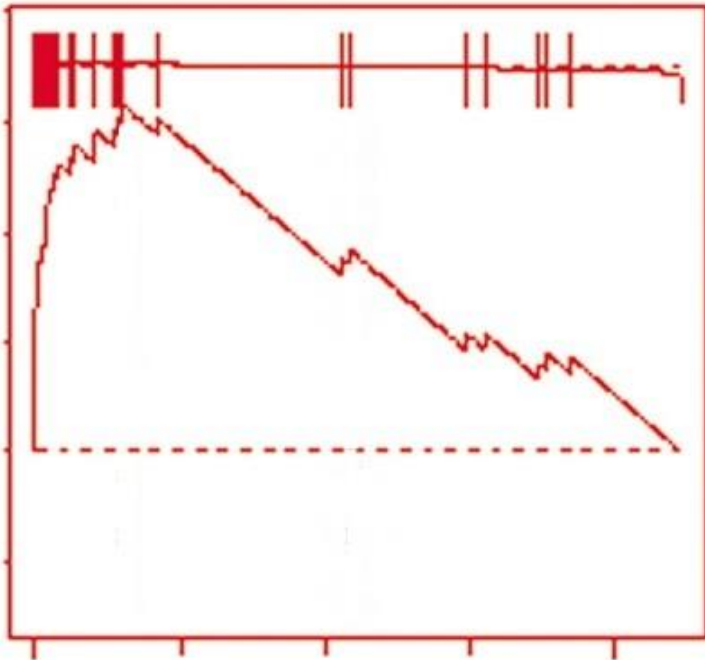


Kolmogorov–Smirnov (KS) Test



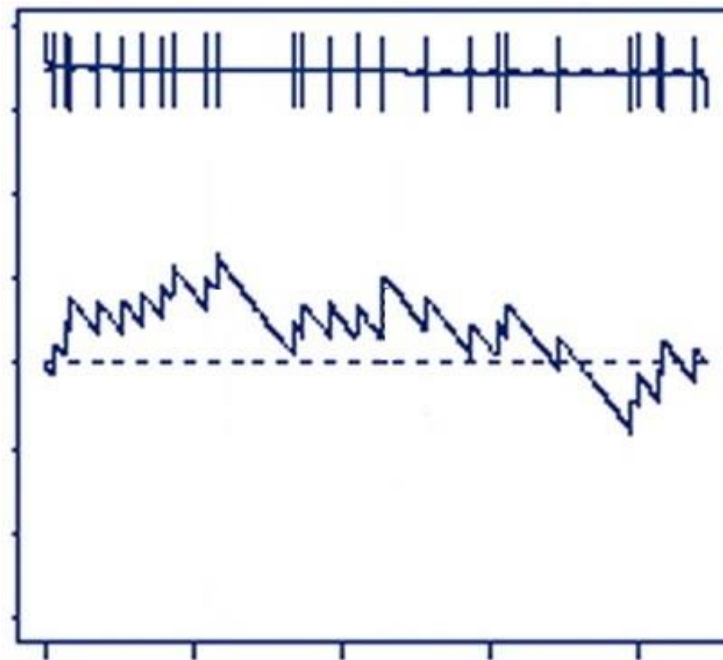
Kolmogorov–Smirnov (KS) Test

**Many genes
upregulated**



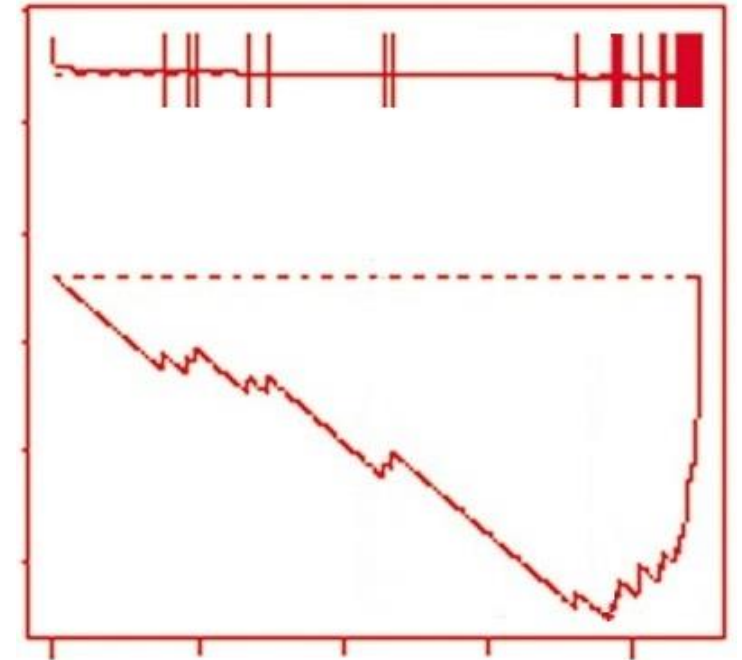
Large positive
enrichment score

**Genes
randomly distributed**



Near zero
enrichment score

**Many genes
downregulated**



Large negative
enrichment score

Kolmogorov–Smirnov (KS) Test

- Generate a null distribution of permuted ESs by shuffling the class labels
- Calculate normalized enrichment score (NES)
 - This adjusts for gene set size and correlation bias
 - Divide the original ES by the mean of permuted ESs with the same sign
- Calculate p-value
 - Compare the original ES to the distribution of permuted ESs (one-tailed)
 - Calculate the percentage of permuted ESs that are higher than the original ES

Kolmogorov–Smirnov (KS) Test

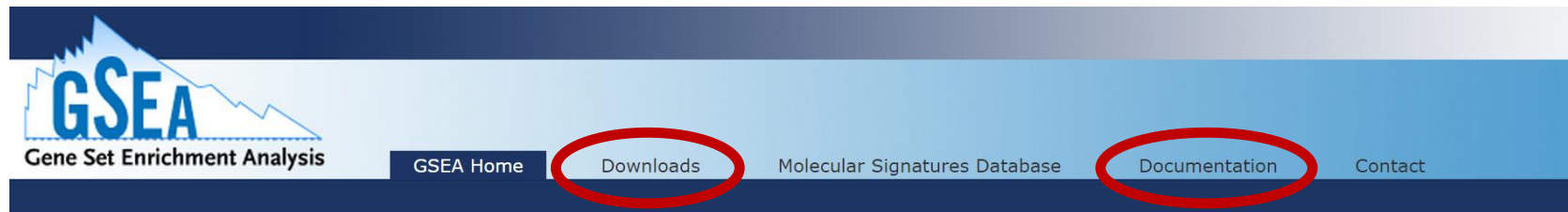
- Used in Gene Set Enrichment Analysis (GSEA)
 - Free software (<http://www.broadinstitute.org/gsea>)
 - Pros:
 - Large collection of gene sets
 - Uses more information than methods that only use DEG list
 - Enrichment plot improves interpretability
 - Cons:
 - Permutation-based p-values
 - Gene sets are not as clean

Additional Processing

- Apply a multiple comparison correction (FDR)
- Interpret p-values cautiously
- Filter out gene sets that are too small or too big
- Explore key drivers in significant pathways
- Experimentally validate or follow up on significant gene sets

GSEA Example

- <http://www.broadinstitute.org/gsea>

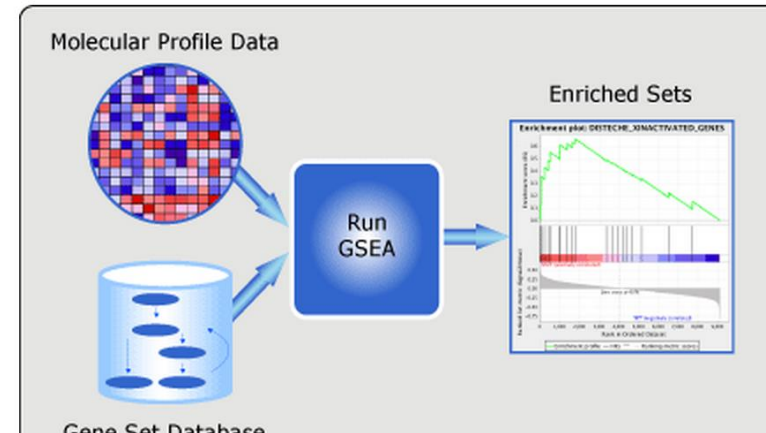


Overview

Gene Set Enrichment Analysis (GSEA) is a computational method that determines whether an a priori defined set of genes shows statistically significant, concordant differences between two biological states (e.g. phenotypes).

From this web site, you can:

- ▶ **Download** the GSEA software and additional resources to analyze, annotate and interpret enrichment results.
- ▶ **Explore the Molecular Signatures Database (MSigDB)**, a collection of annotated gene sets for use with GSEA software.
- ▶ **View documentation** describing GSEA and MSigDB.



GSEA Example

Downloads

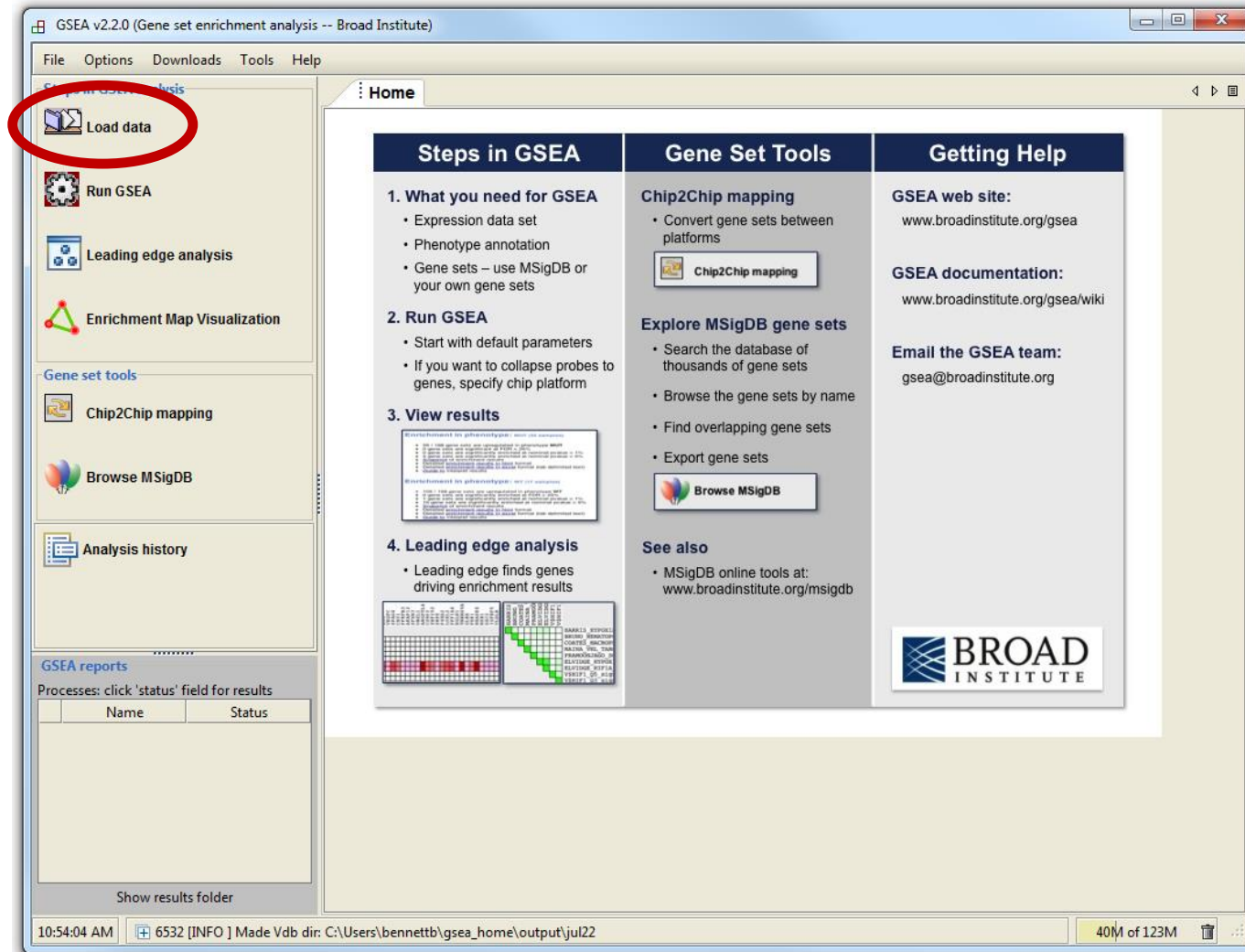
The GSEA software and source code and the Molecular Signatures Database (MSigDB) are freely available to individuals in both academia and industry for internal research purposes. Please see the [GSEA/MSigDB license](#) for more details.

Software

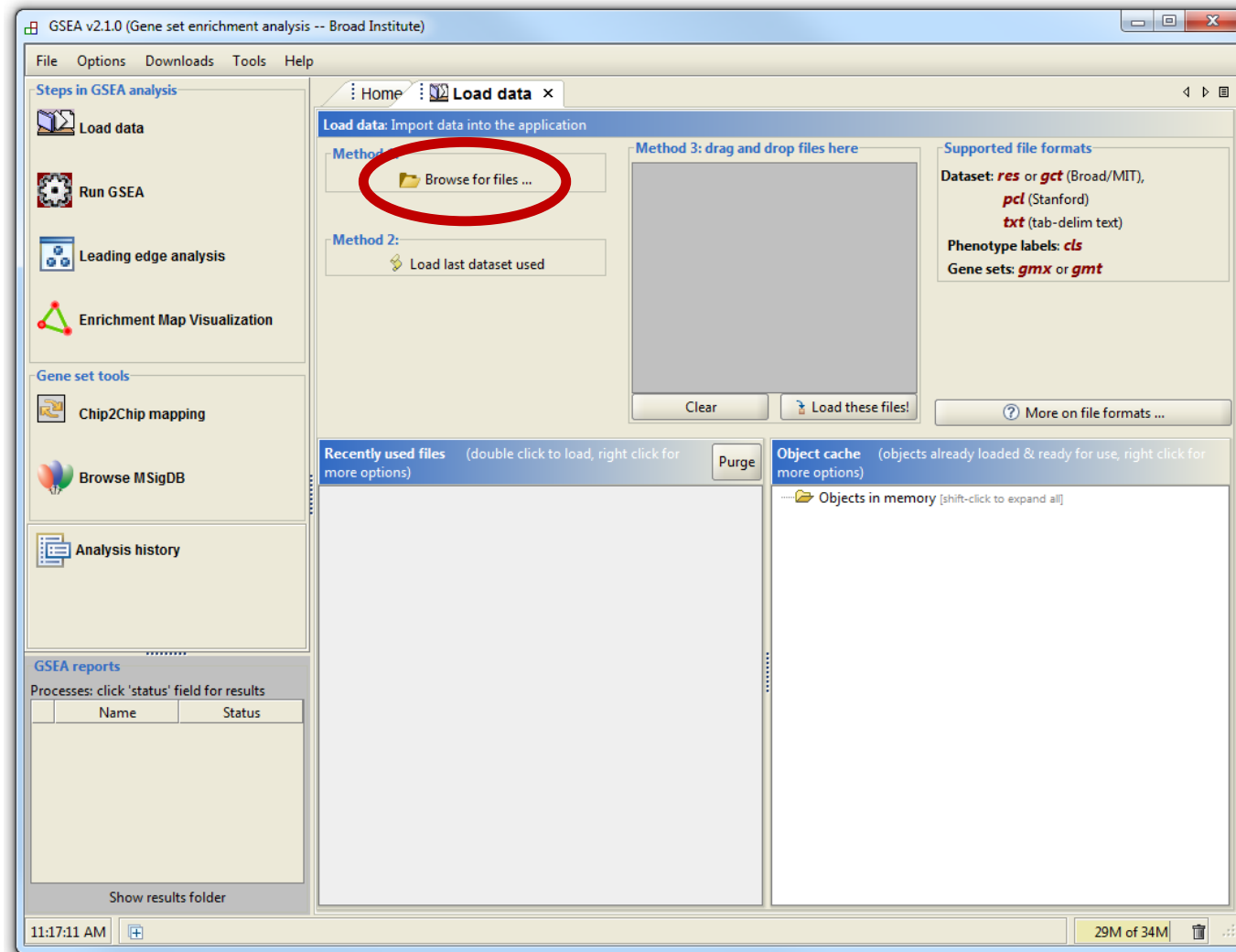
There are several options for GSEA software. All options implement exactly the same algorithm. Usage recommendations and installation instructions are listed below. Current Java implementations of GSEA require Java 6 or 7.

javaGSEA Desktop Application	<ul style="list-style-type: none">▶ Easy-to-use graphical user interface▶ Runs on any desktop computer (Windows, Mac OS X, Linux etc.) that supports Java 6 or 7▶ Produces richly annotated reports of enrichment results▶ Integrated gene sets browser to view gene set annotations, search for gene sets and map gene sets between platforms	Launch with 1GB (for 32 or 64-bit Java) ▾ memory: 
javaGSEA Java Jar file	<ul style="list-style-type: none">▶ Command line usage▶ Runs on any platform that supports Java 6 or 7▶ We recommend using the 'Launch' buttons above instead of this mode for most users	
R-GSEA	<ul style="list-style-type: none">▶ Usage from within the R programming environment	

GSEA Example



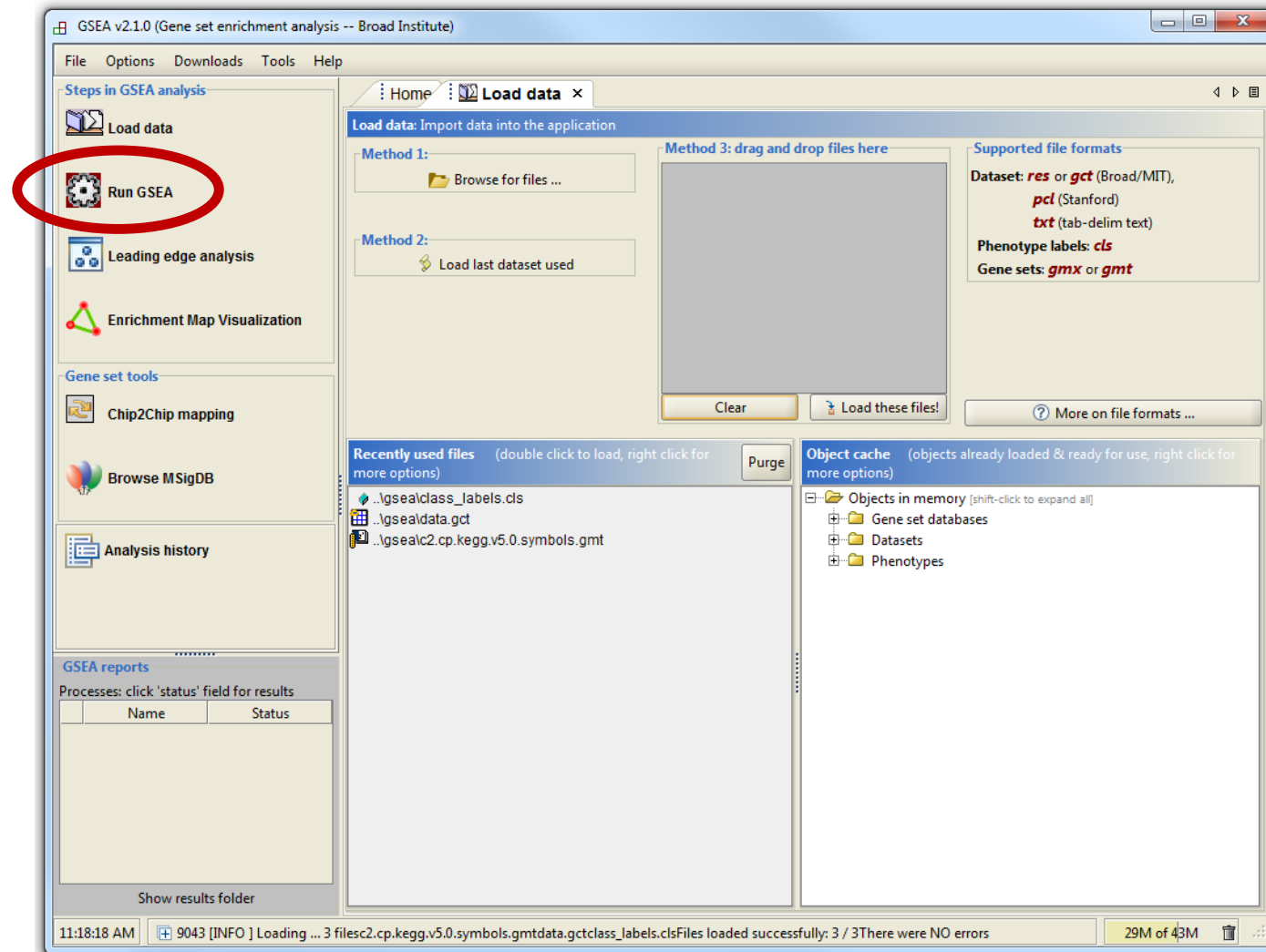
GSEA Example



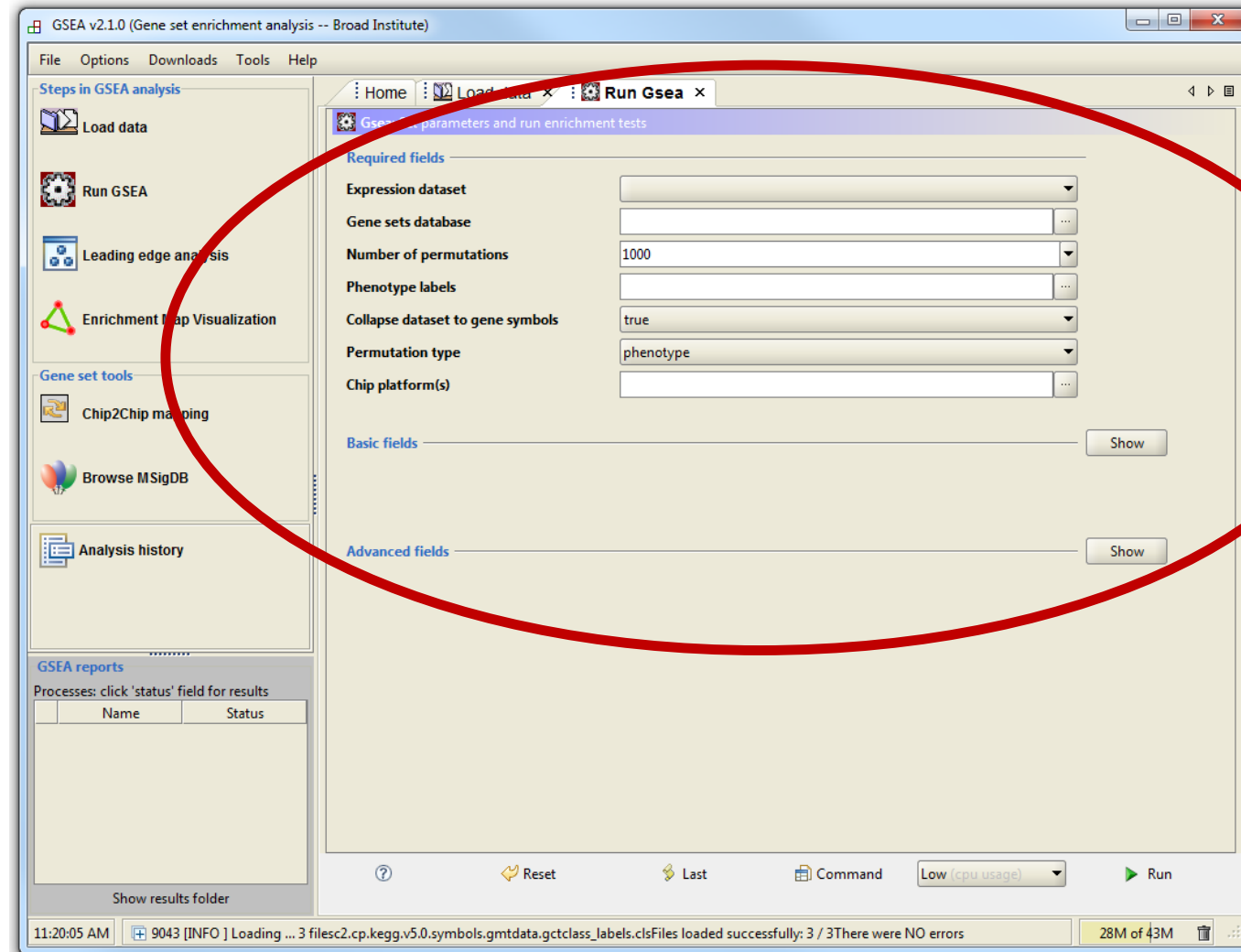
GSEA Example

- Supported file types:
 - http://www.broadinstitute.org/cancer/software/gsea/wiki/index.php/Data_formats
- Required:
 - Expression data file
 - Class label file
- Optional:
 - Gene set file

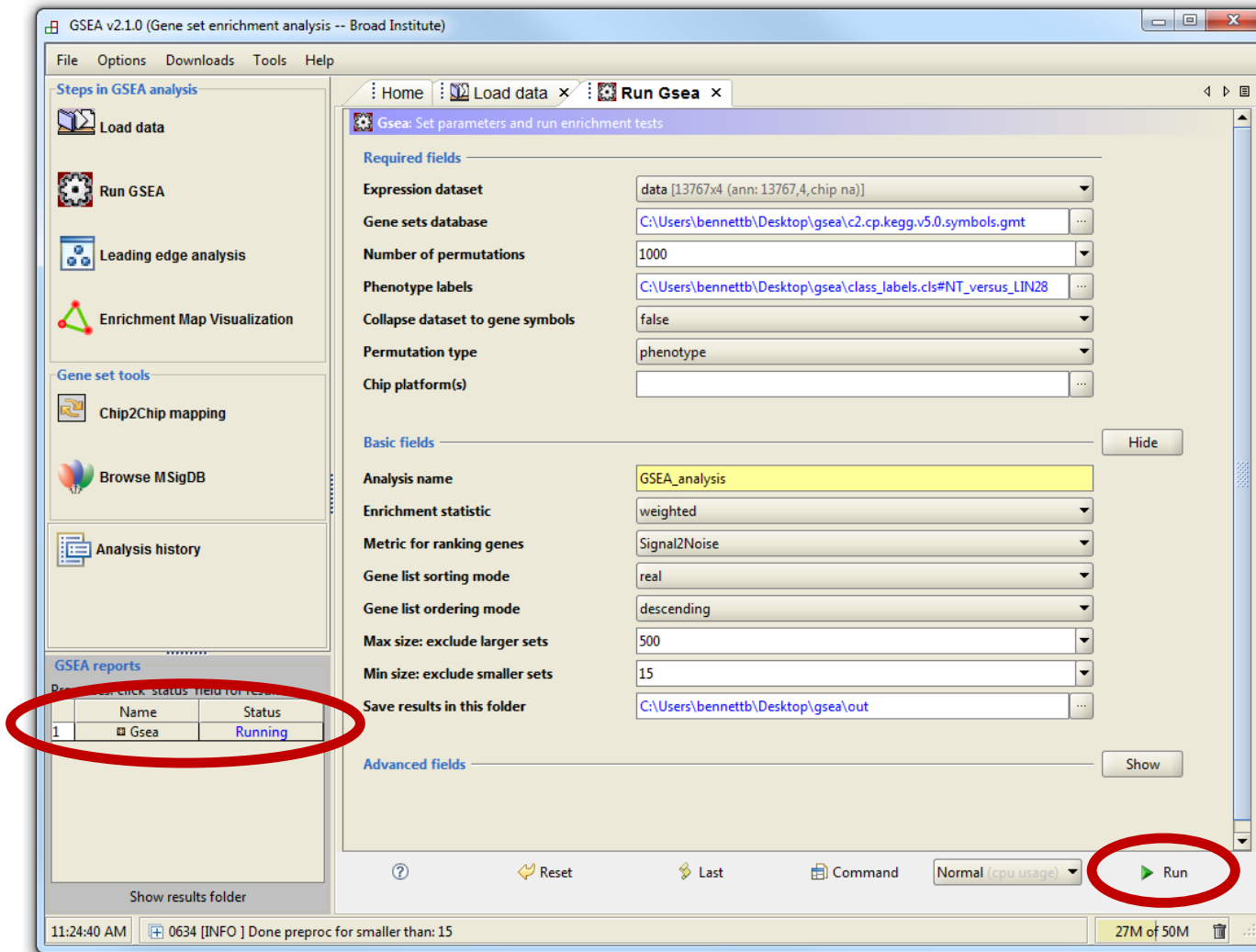
GSEA Example



GSEA Example

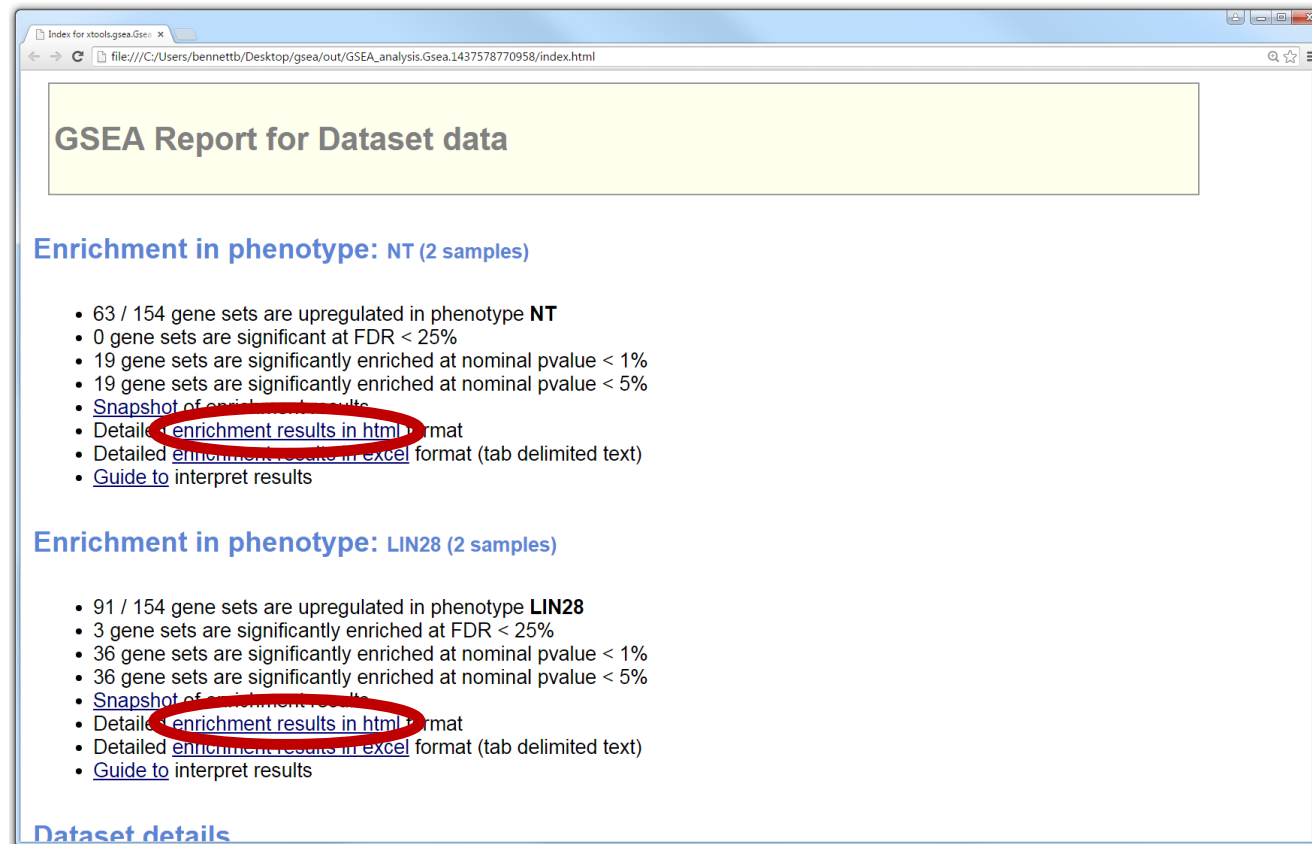


GSEA Example



GSEA Example

- Within output directory: index.html



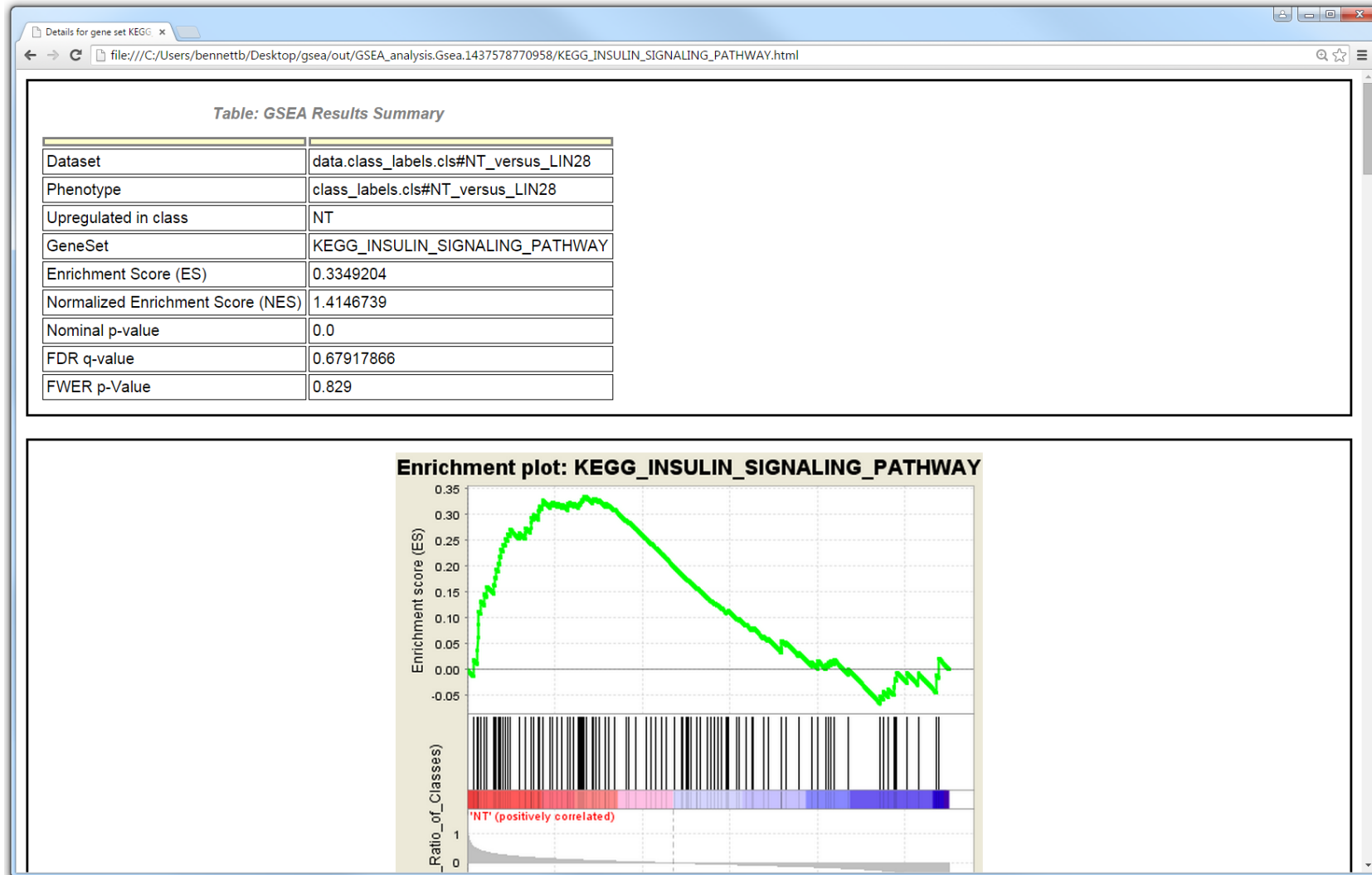
GSEA Example

Report for NT 143757877/ x
file:///C:/Users/bennettb/Desktop/gsea/out/GSEA_analysis.Gsea.1437578770958/gsea_report_for_NT_1437578770958.html

Table: Gene sets enriched in phenotype NT (2 samples) [plain text format]

	GS follow link to MSigDB	GS DETAILS	SIZE	ES	NES	NOM p-val	FDR q-val	FW p-v
1	KEGG REGULATION OF AUTOPHAGY	Details ...	1	0.46	1.60	0.000	0.527	0.17
2	KEGG VASOPRESSIN REGULATED WATER REABSORPTION	Details ...	36	0.55	1.56	0.000	0.488	0.66
3	KEGG MTOR SIGNALING PATHWAY	Details ...	43	0.30	1.44	0.000	0.743	0.82
4	KEGG INSULIN SIGNALING PATHWAY	Details ...	110	0.33	1.41	0.000	0.679	0.82
5	KEGG VASCULAR SMOOTH MUSCLE CONTRACTION	Details ...	66	0.34	1.34	0.000	1.000	0.82
6	KEGG LONG TERM POTENTIATION	Details ...	46	0.35	1.33	0.000	1.000	0.82
7	KEGG N GLYCAN BIOSYNTHESIS	Details ...	45	0.36	1.29	0.000	1.000	0.82

GSEA Example



Other Types of Pathway Analysis

- Other data types (genotype, copy number, etc.)
- Other types of gene lists (genes from proteomics, ChIP targets, etc.)
- Custom gene sets (genes of interest, DEGs from another analysis, etc.)

Resources

- Pathway analysis tools
 - Gene Set Enrichment Analysis (GSEA)
 - <http://www.broadinstitute.org/gsea>
 - Ingenuity Pathway Analysis (IPA)
 - <http://www.ingenuity.com/products/ipa>
 - Database for Annotation, Visualization and Integrated Discovery (DAVID)
 - <https://david.ncifcrf.gov>
 - Gene Ontology Enrichment Analysis Software Toolkit (GOEAST)
 - <http://omicslab.genetics.ac.cn/GOEAST>

Resources

- Gene set and pathway databases
 - Molecular Signatures Database (MSigDB)
 - <http://www.broadinstitute.org/gsea/msigdb>
 - Kyoto Encyclopedia of Genes and Genomes (KEGG)
 - <http://www.genome.jp/kegg>
 - BioCarta
 - <http://www.biocarta.com>
 - Gene Ontology (GO)
 - <http://geneontology.org>
 - PANTHER GO-slim
 - <http://www.pantherdb.org/panther/ontologies.jsp>