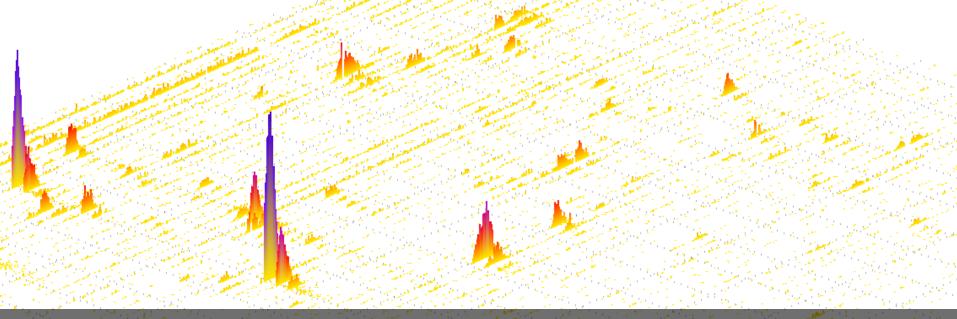
May Institute 2017

Computation and statistics for mass spectrometry and proteomics

Fundamentals of non-targeted proteomics and metabolomics





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Today's Schedule

Tuesday 5/2/2017	
8:00 AM	Bring your own data or Skyjam
9:00 AM	Lecture: Label-free quantitative proteomics.
10:30 AM	Refreshments
11:00 AM	Hands-on: Label-free quantification workflows
12:30 PM	Lunch Break
1:30 PM	Lecture: Introduction to non-targeted metabolomics.
2:30 PM	Hands-on: Metabolite profiling workflow.
3:00 PM	Refreshments
3:30 PM	Hands-on: Differential quantification of metabolites, visualization, report generation
5:00 PM	Questions and practice with own data
6:00 PM	Adjourn

Metabolome vs. Proteome

- Size and complexity of the metabolome still largely unknown
- Similar to protein sequence databases, there are also metabolite databases listing all known metabolites (usually contains tens of thousands of metabolites)
- Differences between proteome and metabolome
 - Metabolites belong to wider ranger of chemical compound classes (lipids, sugars, amino acids)
 - Proteins have a more homogenous chemistry (20 proteinogenic amino acids)
 - Metabolites can have complex structures that require a structural formula for a comprehensive description
 - Proteins have a simple, linear structure that can be represented by a sequence
 - Metabolites are light: average metabolite mass a 100-300 Da
 - Proteins are heavy: median protein length around 300-500 aa, about 40,000
 Da molecular weight

Metabolites

- Metabolites comprise a heterogeneous set of biomolecules: all small molecules in a system excepting salts and macromolecules (proteins, long peptides, RNA, DNA)
- Lipids and sugars are metabolites as well
- There are separate fields dealing with lipids and sugars (lipidomics, glycomics), techniques are very similar

Examples:

Metabolite	mol I ^{−1}	Metabolite	tabolite mol I ⁻¹		$ m mol~I^{-1}$	
Glutamate	9.6 × 10 ⁻²	UDP-glucuronate (51)	5.7×10^{-4}	N-Acetyl-ornithine (79)	4.3×10^{-5}	
Glutathione	1.7×10^{-2}	ADP	5.6×10^{-4}	Gluconate (80)	4.2×10^{-5}	
Fructose-1,6-bisphosphate	1.5×10^{-2}	Asparagine (52)	5.1×10^{-4}	Malonyl-CoA (81)	3.5×10^{-5}	
ATP	9.6×10^{-3}	α-Ketoglutarate	4.4×10^{-4}	Cyclic AMP (82)	3.5×10^{-5}	

Extracted from Bennett et al.: some of the most abundant small molecules in E. coli

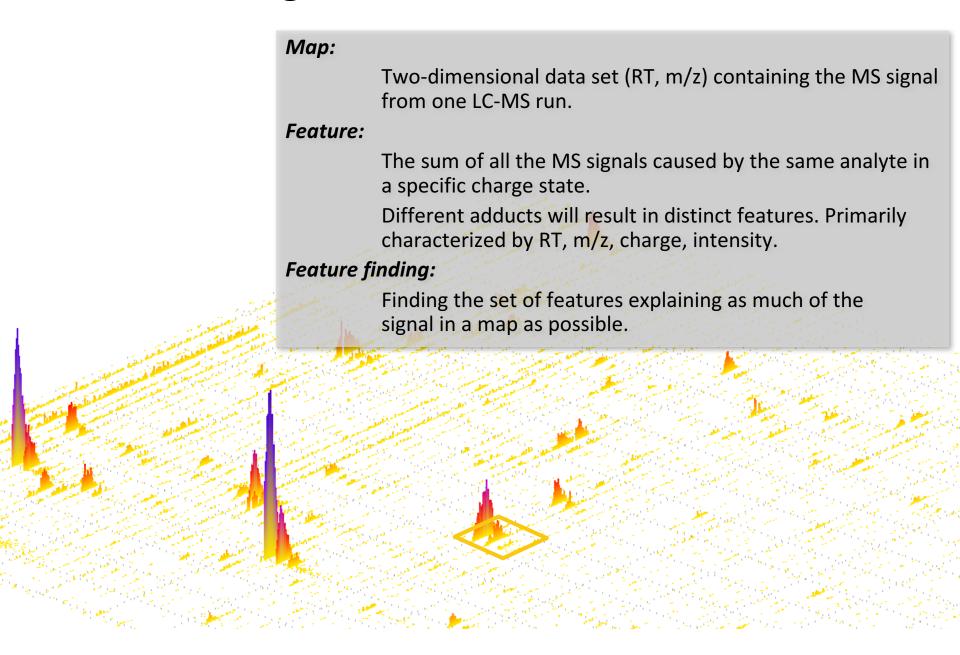
Metabolomics Techniques

- Fundamentally two types of approaches
 - Targeted metabolomics
 - Identify only a well-defined subset of metabolites, but those with higher accuracy (hundreds?)
 - All of these metabolites can then be identified
 - Non-targeted metabolomics (metabolic profiling)
 - Try to see as much of the metabolome as possible (thousands and more)
 - Majority of metabolites can be seen
 - Only a small fraction will be identified
- Similarly, there is also targeted and non-targeted proteomics
- In proteomics, the identification problem is less difficult, though, which is why this distinction is more relevant in metabolomics (where identification is much harder)

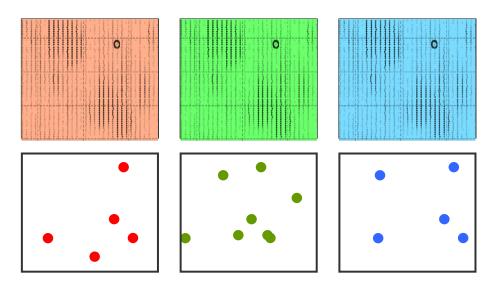
Metabolite Quantification

- Label-free proteomics is similar to non-targeted metabolomics
- Overall workflow is identical
 - Feature finding
 - Map alignment
 - Feature linking
- Feature-finding approaches are algorithmically similar to those used in proteomics
 - Mass traces usually at the heart of the algorithm
 - Assembly into features can be done similarly
- However, there are some differences
 - Isotopic patterns differ from proteomics (no averagine!)
 - Mass range and charge states are different

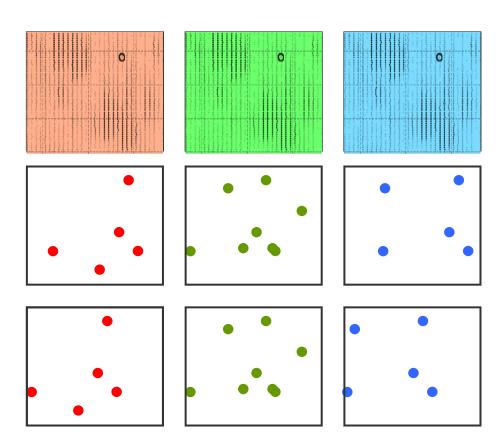
Feature Finding – Terms



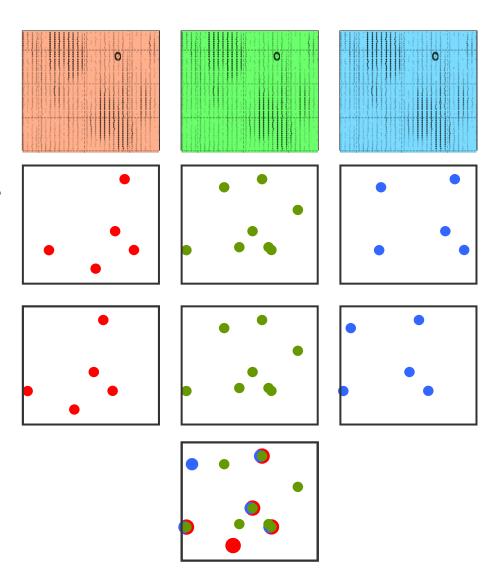
1. Find features in all maps



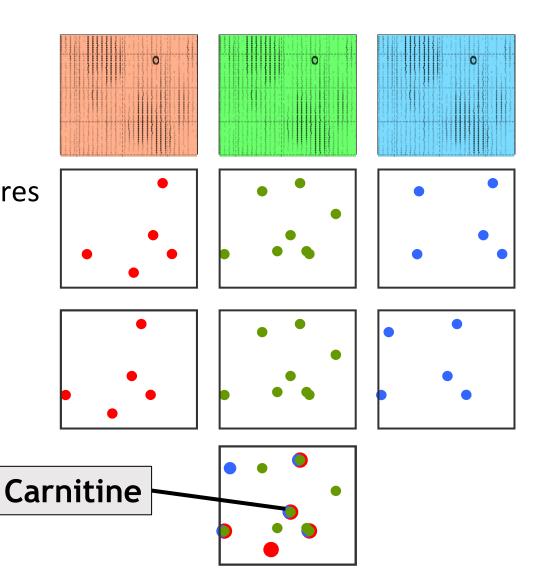
- 1. Find features in all maps
- 2. Align maps



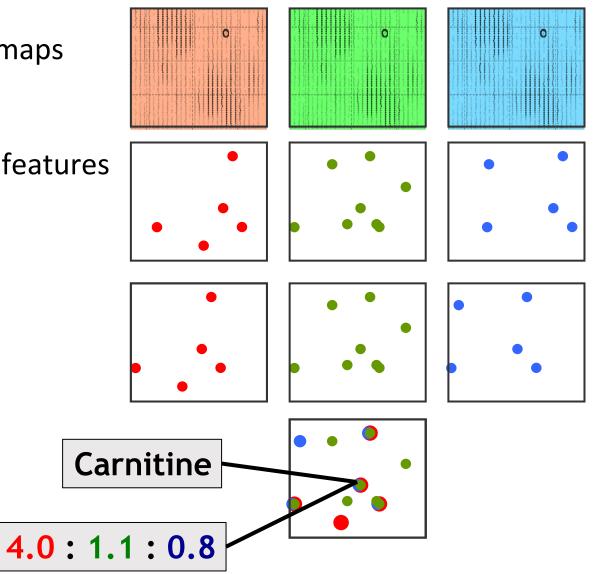
- 1. Find features in all maps
- 2. Align maps
- 3. Link corresponding features



- 1. Find features in all maps
- 2. Align maps
- 3. Link corresponding features
- 4. Identify features



- 1. Find features in all maps
- 2. Align maps
- 3. Link corresponding features
- 4. Identify features
- 5. Quantify



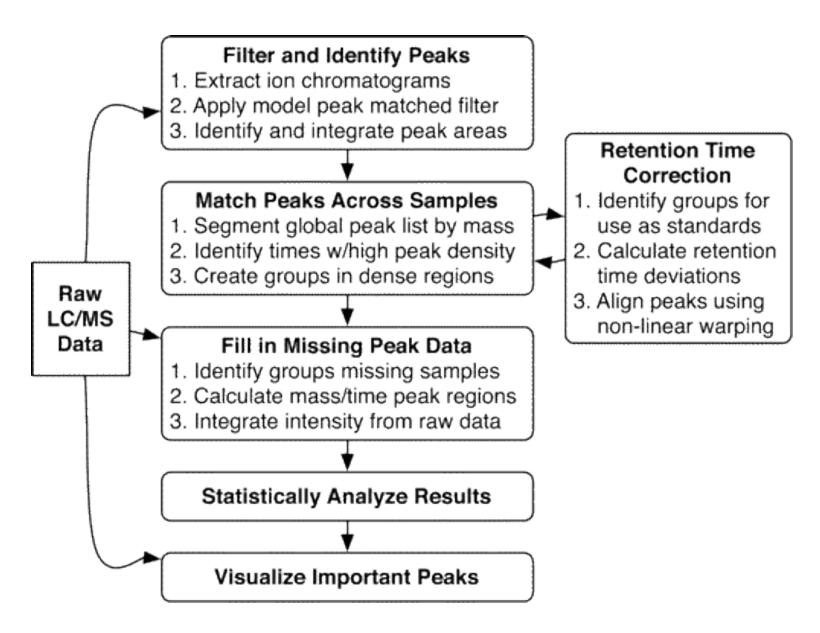
Feature Finding in MTX – Issues

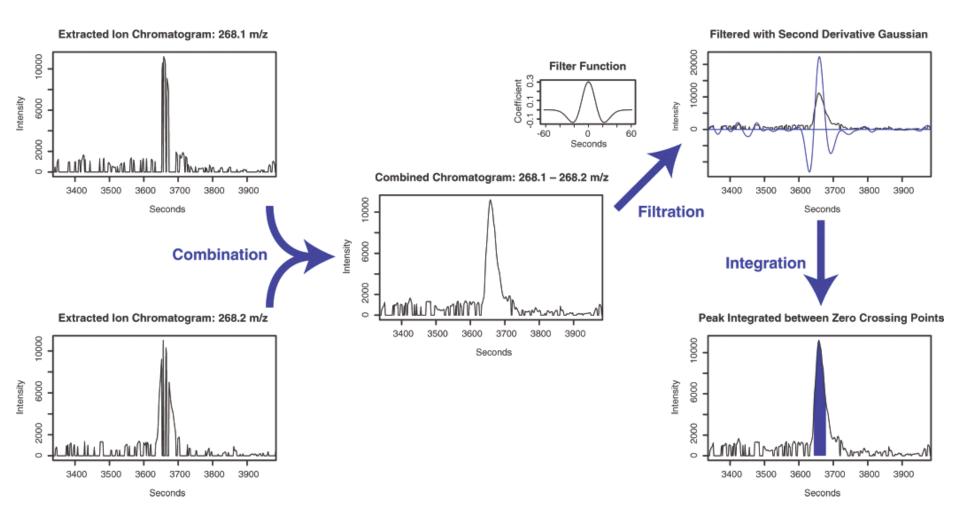
- Proteomics feature finding algorithms make extensive use of the averagine hypothesis: peptides have a well-defined average composition
- Metabolites are chemically much more diverse than peptides
- Feature finding algorithms are often very sensitive to the choice of parameters
- Tuning these parameters can be a challenge
- Sensitivity is often an issue in feature finding: distinguishing signal from noise can be a challenge
- Lack of sensitivity is often a problem for large-scale studies missing values

XCMS is a Bioconductor package, written in R

Key ideas

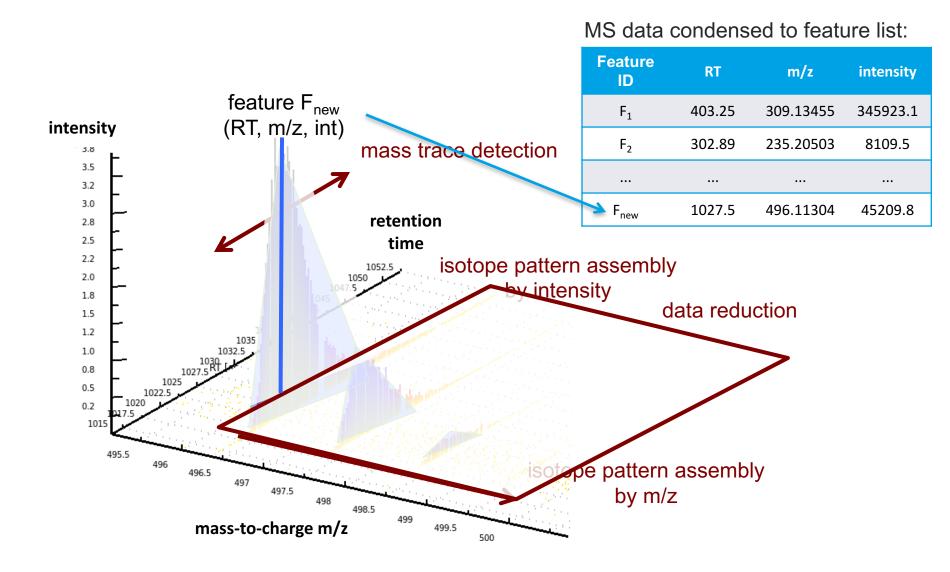
- Extract mass traces by binning peaks w.r.t. m/z
- Treat mass bins as distinct mass traces
- Detect peaks in these mass traces using standard methods from signal processing
- Align detected mass traces in the RT dimension across maps using nonlinear de-warping



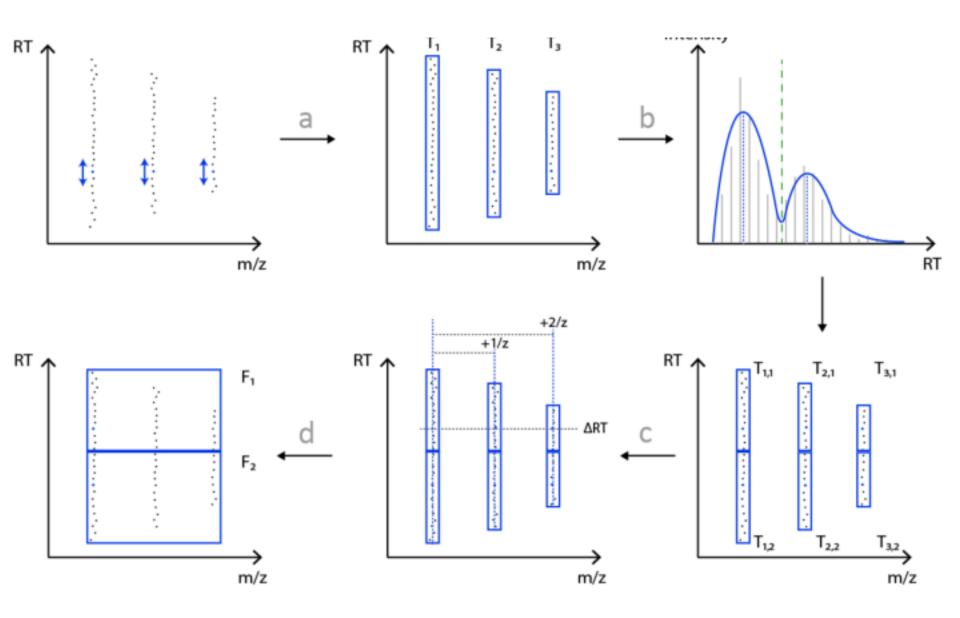


- XCMS has become the quasi standard for LC-MS metabolomics data analysis
- Recent versions include more advanced methods, including wavelet peak detection
- For many tasks (e.g., biomarker detection), the identification of differential mass traces is sufficient (lower complexity of metabolomics data sets)
- Other software packages also assemble mass traces back to features (e.g., OpenMS FeatureFinderMetabo)
- Advantages here:
 - Profit from additional information, increase specificity
 - Reduced number of signals (multiple mass traces per feature)

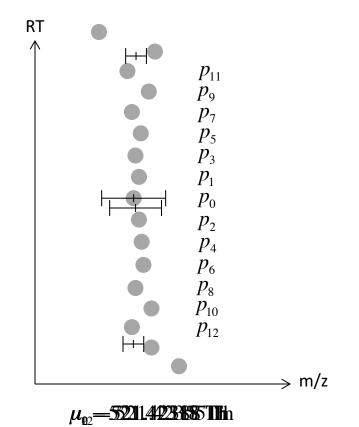
OpenMS - Metabolite Feature Finding



Algorithmic Overview



Mass Trace Detection



 $\sigma_{\rm pp}^2 \approx 0.0000037 \, \text{dr}$ param

A mass spectrometric peak p is given by

$$p = (t, m, i)$$

t: retention time, m: mass-to-charge ratio, i: intensity

A mass trace T is a list of peaks:

$$T = (p_1, p_2, ..., p_k, p_l, ..., p_n)$$
 $t_k < t_l \forall k < l$

- m/z error model is adaptive
- Online Gaussian density estimation

$$\mu_{n+1} = \frac{w_n \cdot \mu_n + i_{n+1} \cdot m_{n+1}}{w_n + i_{n+1}} \qquad \sigma_{n+1}^2 = \frac{w_n \cdot \sigma_n^2 + i_{n+1} \cdot (m_{n+1} - \mu_{n+1})^2}{w_n + i_{n+1}}$$
centroid m/z
$$m/z = \frac{v_n \cdot \mu_n + i_{n+1} \cdot m_{n+1}}{w_n + i_{n+1}}$$

$$m/z = \frac{v_n \cdot \sigma_n^2 + i_{n+1} \cdot (m_{n+1} - \mu_{n+1})^2}{w_n + i_{n+1}}$$

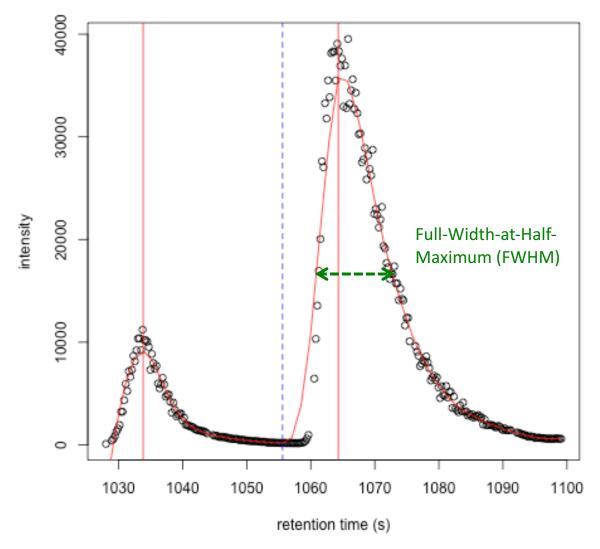
$$w_n = \sum_{k=1}^{n} i_k$$

$$\mu_n - 3 \cdot \sigma_n \le m_{n+1} \le \mu_n + 3 \cdot \sigma_n$$
weight
$$m/z \text{ constraint}$$

 $T = (p_0)p_1, p_2, p_3, p_4, p_2, p_0, p_1, p_2, \dots, p_{11})$

Peak Separation

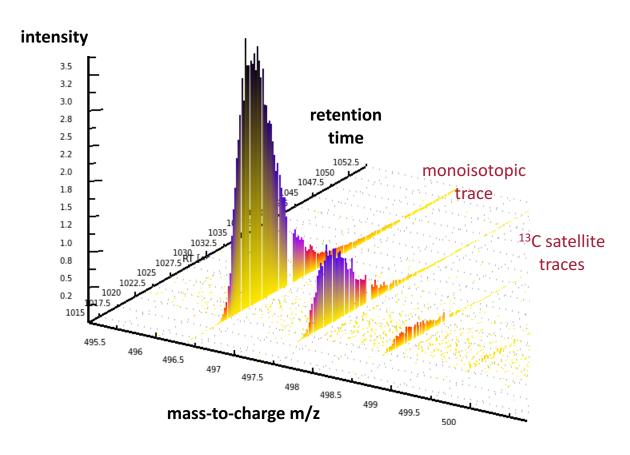
Split chromatographic peaks overlapping in retention time



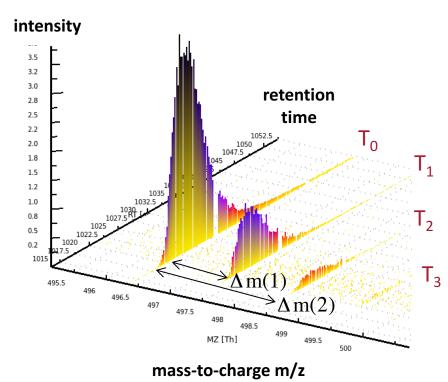
Kenar et al., Mol. Cell. Prot., 2014, 13(1):348-59. doi: 10.1074/mcp.M113.031278

Feature Assembly

- Identify mass traces belonging to the same feature
- Multiple explanations are possible
- Create all potential hypotheses and score them



Feature Scoring - m/z



• m/z distances T_0 and T_j :

$$\Delta m(j) = \left| \overline{m}_0 - \overline{m}_j \right|$$

Theoretical m/z distances:

$$\mu(j) = 1.0033 \, \text{Da} \cdot \frac{j}{z}$$

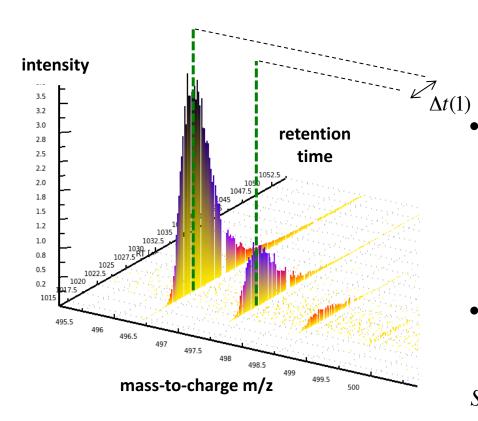
• Mass errors for T_0 and T_j :

$$\sigma^2(j) = \sigma_0^2 + \sigma_j^2$$

Pairwise scoring function:

$$S_{\Delta m}(j) = \begin{cases} e^{-\frac{(\Delta m(j) - \mu(j))^2}{2\sigma^2(j)}}, & \text{if } \mu(j) - 3 \cdot \sigma(j) \le \Delta m(j) \le \mu(j) + 3 \cdot \sigma(j) \\ 0 & \text{else.} \end{cases}$$

Feature Scoring – RT



RT shifts between T₀ and T_j:

$$\Delta t(j) = \left| \overline{t_0} - \overline{t_j} \right|$$

Gaussian error model with

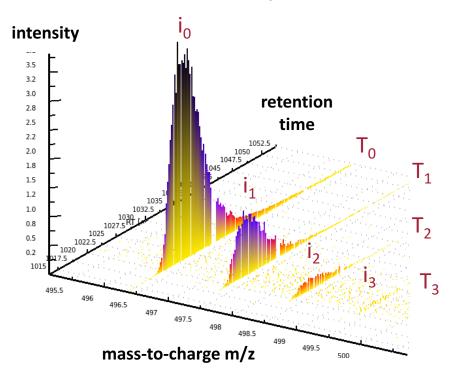
$$\mu_{\Delta RT} = 0 \qquad \sigma_{\Delta RT}^2 = \left(\frac{\Delta t_{0.5}}{2\sqrt{2\ln 2}}\right)^2$$

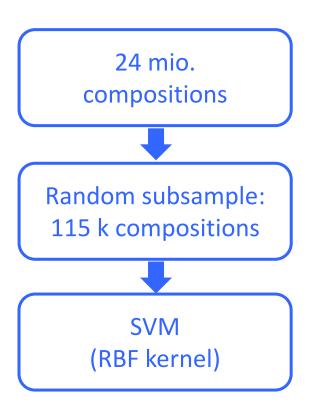
• Pairwise scoring function:

$$S_{\Delta RT}(j) = \begin{cases} e^{-\frac{(\Delta t(j))^2}{2\sigma_{\Delta t}^2}}, & \text{if } -3 \cdot \sigma_{\Delta t} \leq \Delta t(j) \leq 3 \cdot \sigma_{\Delta t} \\ 0 & \text{else.} \end{cases}$$

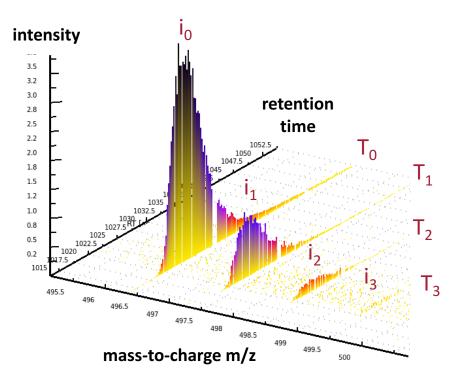
Feature Scoring – Intensity

- Problem: There is no 'averagine' for metabolites
- Idea
 - Enumerate metabolite compositions and learn intensities
 - 'Golden rules' describe likely chemistry (Kind & Fiehn, BMC Bioinfo, 2007)
 - Generate all compositions, remove unlikely ones based on heuristics





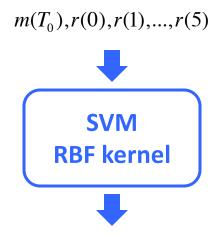
Feature Scoring – Intensity



Intensity ratio of T₀ and T_i:

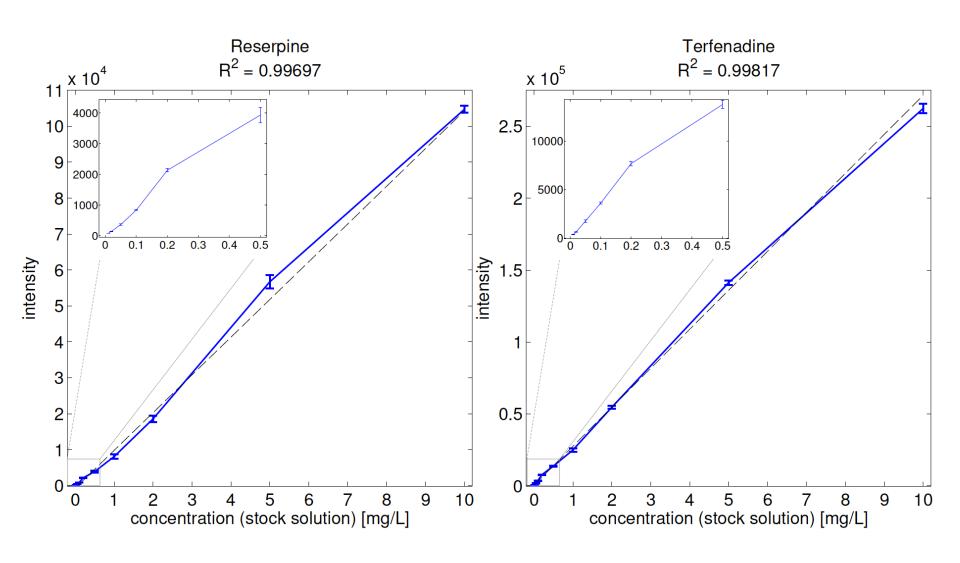
$$r(j) = \frac{i_j}{i_0}$$

Assess if valid isotope ratios:

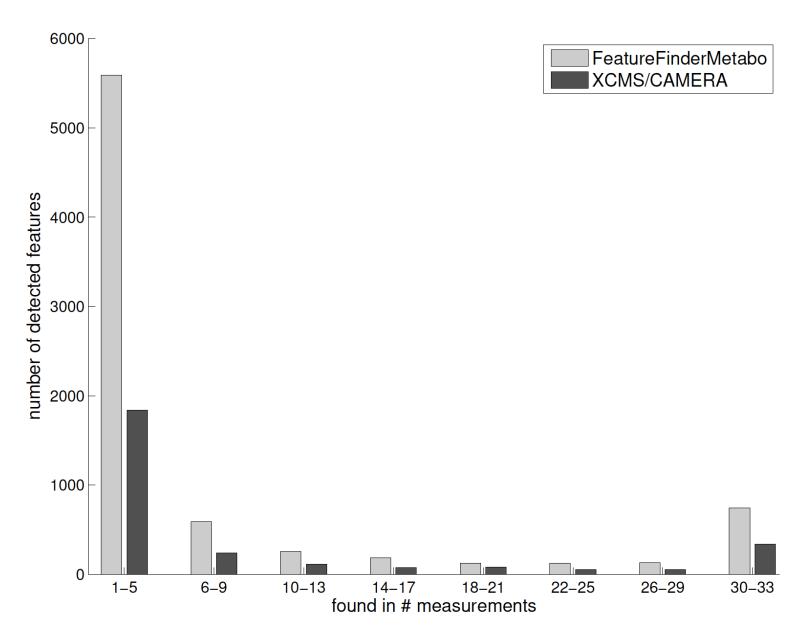


Yes, it is a legal isotope pattern, **keep it**Or
No, it is not a legal isotope pattern, **discard it**

Quantification Linearity – Spike-In



Sensitivity – Human Plasma



Specificity – Synthetic Data

Benchmarking feature detection algorithms is HARD

- Multiple metrics are required: linearity, sensitivity, specificity
- Sensitivity needs to be balanced with specificity
- Experimental data does not come with a well-defined ground truth

Idea

- Simulated LC-MS data with known composition
- Take a well-defined experimental dataset (identification lists from a metabolomics study, plant metabolites)
- OpenMS LC-MS simulator was expanded to generate metabolite data

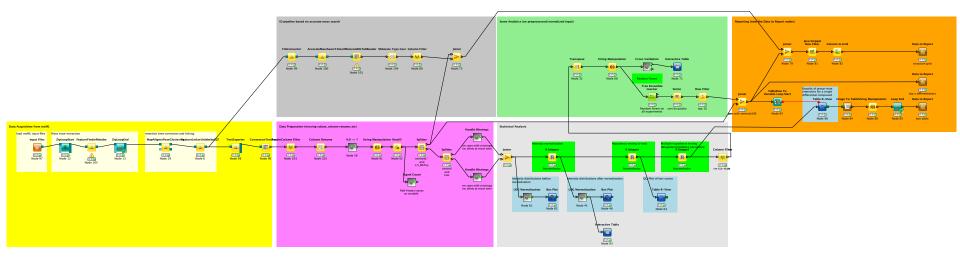
Method	Recall	Precision	F-score
OpenMS	96%	97 %	0.97
XCMS/Camera	88%	37%	0.52

NON-TARGETD METABOLOMICS WITH OPENMS

- Workflows for non-targeted metabolomics
- Metabolomics workflows with OpenMS in KNIME
- Integration into Compound Discoverer

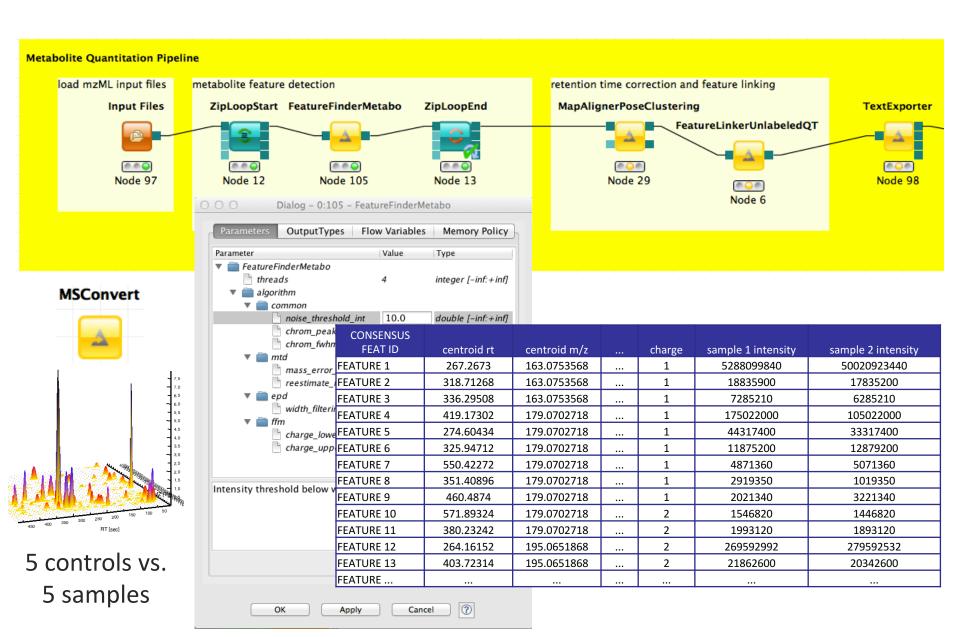


Metabolomics – Biomarker ID

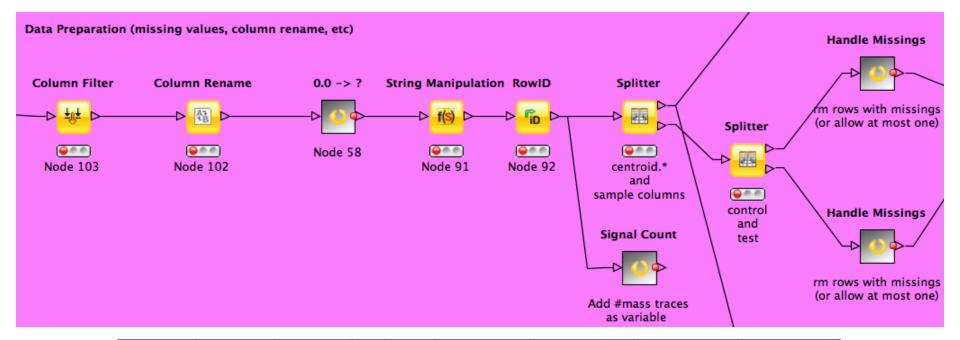


- Complex workflow analyzing a diabetes-related metabolomics biomarker study
 - Data preprocessing (yellow)
 - Quantification (purple)
 - Identification based on accurate mass/HMDB (gray)
 - Detection of distinctive features, statistics (green/gray)
 - Reporting of differential features and their structures (orange)

Metabolite Quantitation

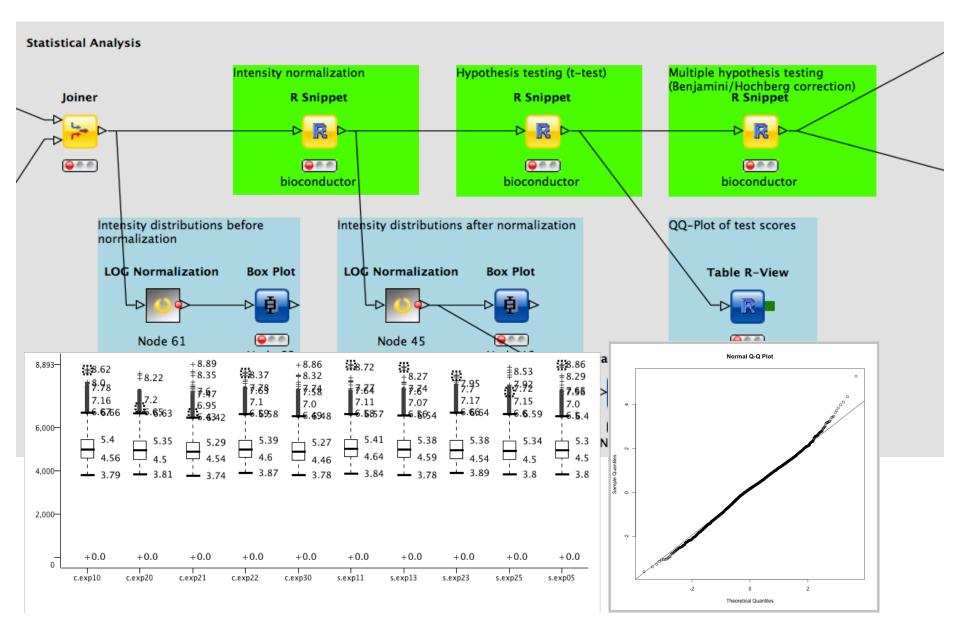


Data Table Magic

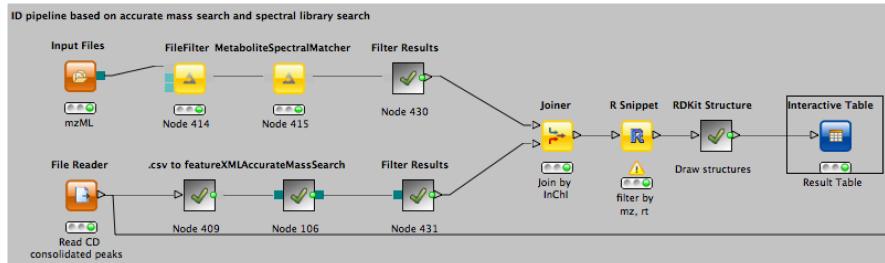


0 10				0 1 14	0		
Row ID	centroid.rt	centroid.mz	 charge	Control_1	Control_2	Sample_2	Sample_2
FEATURE 1	267.2673	163.0753568	 1	5288099840	50020923440	5288099840	50020923440
FEATURE 2	318.71268	163.0753568	 1	18835900	17835200	18835900	17835200
FEATURE 3	336.29508	163.0753568	 1	7285210	6285210	7285210	6285210
FEATURE 4	419.17302	179.0702718	 1	175022000	105022000	175022000	105022000
FEATURE 5	274.60434	179.0702718	 1	44317400	33317400	44317400	33317400
FEATURE 6	325.94712	179.0702718	 1	11875200	12879200	11875200	12879200
FEATURE 7	550.42272	179.0702718	 1	4871360	5071360	4871360	5071360
FEATURE 8	351.40896	179.0702718	 1	2919350	1019350	2919350	1019350
FEATURE 9	460.4874	179.0702718	 1	2021340	3221340	2021340	3221340
FEATURE 10	571.89324	179.0702718	 2	1546820	1446820	1546820	1446820
FEATURE 11	380.23242	179.0702718	 2	1993120	1893120	1993120	1893120
FEATURE 12	264.16152	195.0651868	 2	269592992	279592532	269592992	279592532
FEATURE 13	403.72314	195.0651868	 2	21862600	20342600	21862600	20342600
FEATURE			 				

Multiple Hypothesis Testing



Metabolite ID

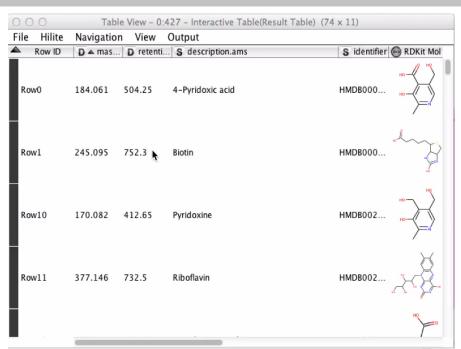


Multiple ID strategies

- Accurate mass
- Retention time database
- Retention time prediction
- Spectral matching

KNIME provides

- Online access to structure databases
- Structure visualization
- Cheminformatics
 - Metabolization
 - Substructure search



References

XCMS

• C.A. Smith, E.J. Want, G.C. Tong, R. Abagyan, and G. Siuzdak. XCMS: Processing Mass Spectrometry Data for Metabolite Profiling Using Nonlinear Peak Alignment, Matching, and Identification. Anal. Chem., 2006,

FeatureFinderMetabo

Kenar, E, Franken, H, Forcisi, S, Wörmann, K, Häring, H, Lehmann, R, Schmitt-Kopplin, P, Zell, A, and Kohlbacher, O (2014). Automated Label-Free Quantification of Metabolites from LC-MS Data. Mol. Cell. Prot., 13(1):348-59. http://dx.doi.org/10.1074/mcp.M113.031278