

# Nonlinear LOB and LOD estimation

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Northeastern

# Outline

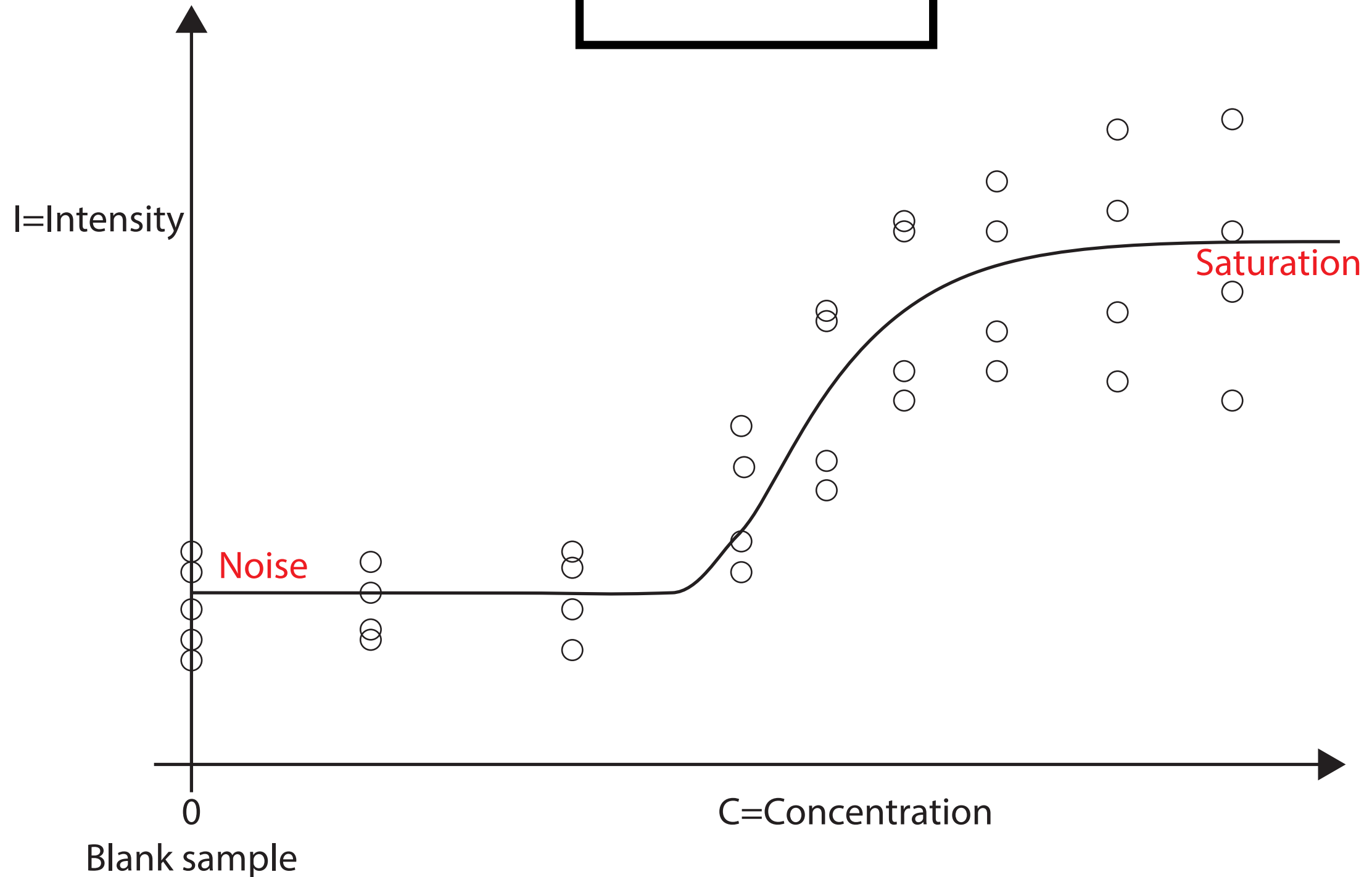
1. Basics of curve fit methods
2. An R package to calculate the LOB and LOD: MSstats

# Response curve

Input (=known)

Output

Protein concentration → LC-MS/MS → Signal Intensity



# LOB/LOD/LOQ definition

Will focus on curve fit methods

*Clinical Chemistry* 50:4  
732–740 (2004)

General Clinical  
Chemistry

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## Partly Nonparametric Approach for Determining the Limit of Detection

KRISTIAN LINNET<sup>1\*</sup> and MARINA KONDRATOVICH<sup>2</sup>

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A STATISTICAL OVERVIEW ON UNIVARIATE CALIBRATION,  
INVERSE REGRESSION, AND DETECTION LIMITS:  
APPLICATION TO GAS CHROMATOGRAPHY/MASS  
SPECTROMETRY TECHNIQUE

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**Irma Lavagnini and Franco Magno\***

*Dipartimento di Scienze Chimiche, Università di Padova,  
via Marzolo 1, 35131 Padova, Italy*

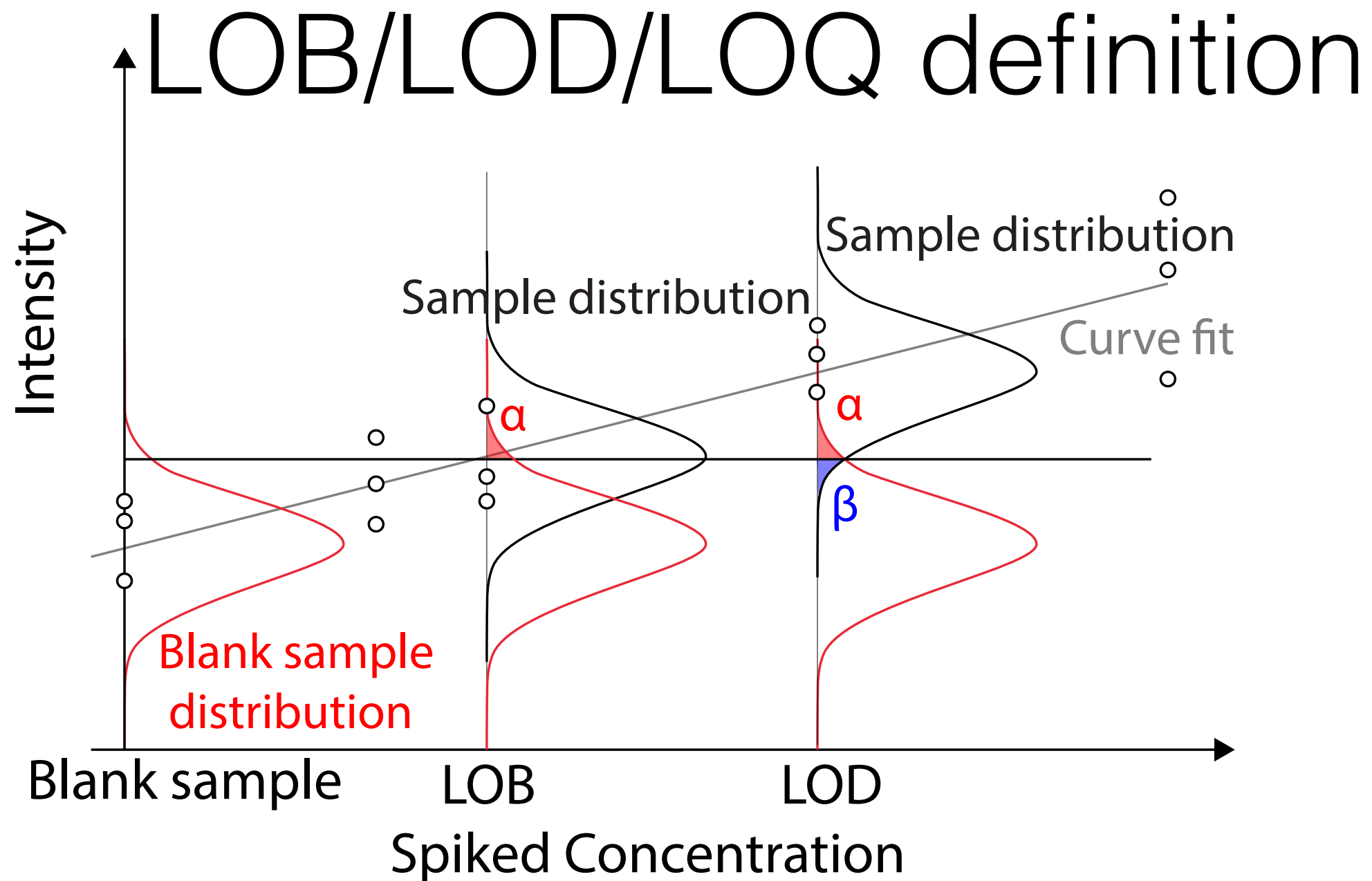
*Received 8 March 2006; accepted 21 April 2006*

*Anal. Chem.* **1997**, *69*, 3069–3075

## Weighted Least-Squares Approach To Calculating Limits of Detection and Quantification by Modeling Variability as a Function of Concentration

Michael E. Zorn,<sup>†</sup> Robert D. Gibbons,<sup>‡</sup> and William C. Sonzogni<sup>\*,†</sup>

*Water Chemistry Program, University of Wisconsin—Madison, Madison, Wisconsin 53706, and Department of Biostatistics, University of Illinois—Chicago, Chicago, Illinois 60612*



- LOB = Limit of Blank = Min. concentration above which false positive  $< \alpha$
- LOD = Limit of Detection = Min. concentration above which false negative  $< \alpha$  and false positive  $< \beta$

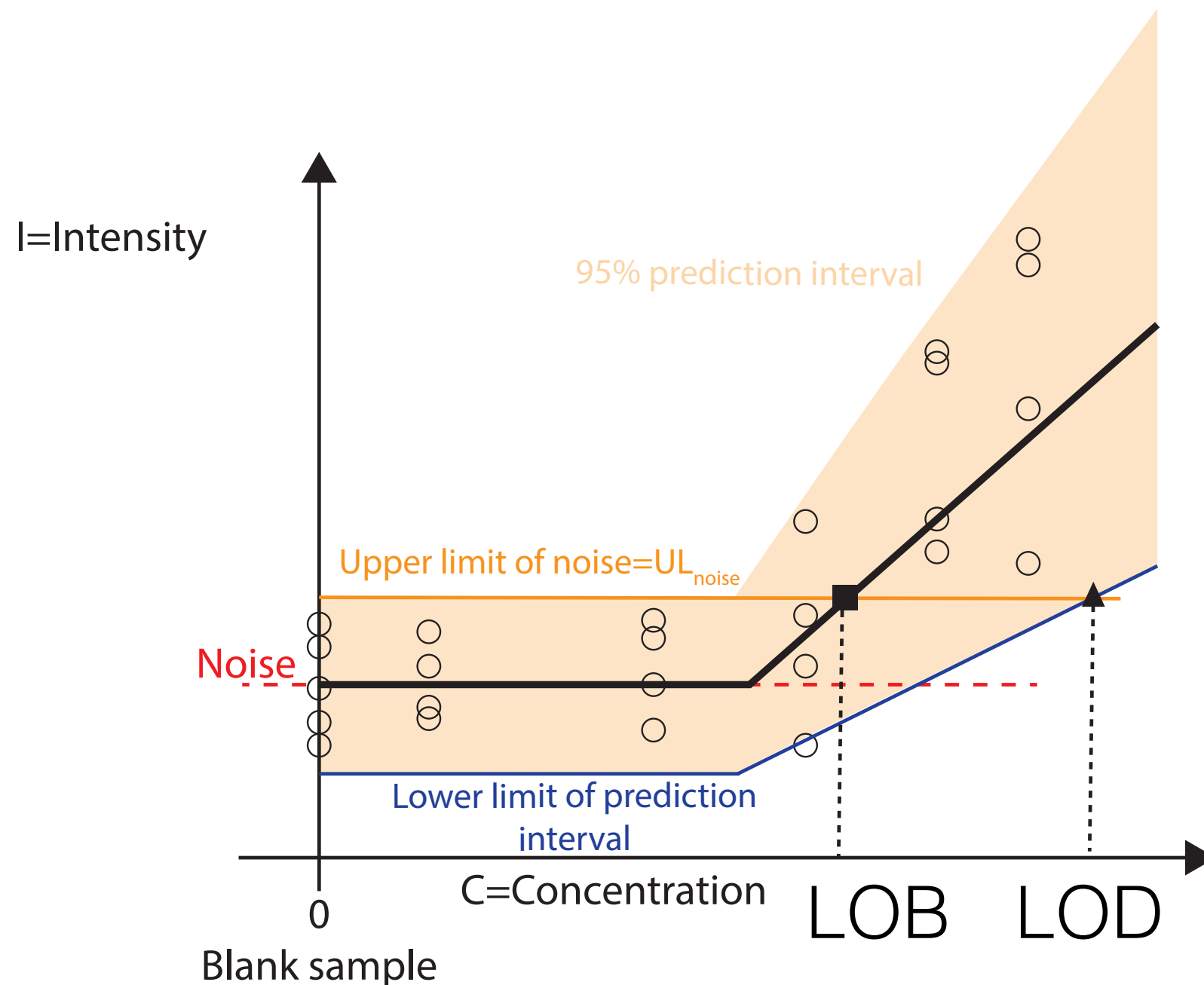
Usually  $\alpha = \beta = 5\%$

# LOB/LOD/LOQ definition

- LOQ = Limit of Quantification = Min. concentration above at which the assay is able to provide quantitative results of a stated analytical quality
- No “universal” definition exists for the LOQ
  - \* $CV = \text{mean}/sd < 0.2$
  - \*Recovery factor  $< 0.2$
  - \*Methods based on blank sample

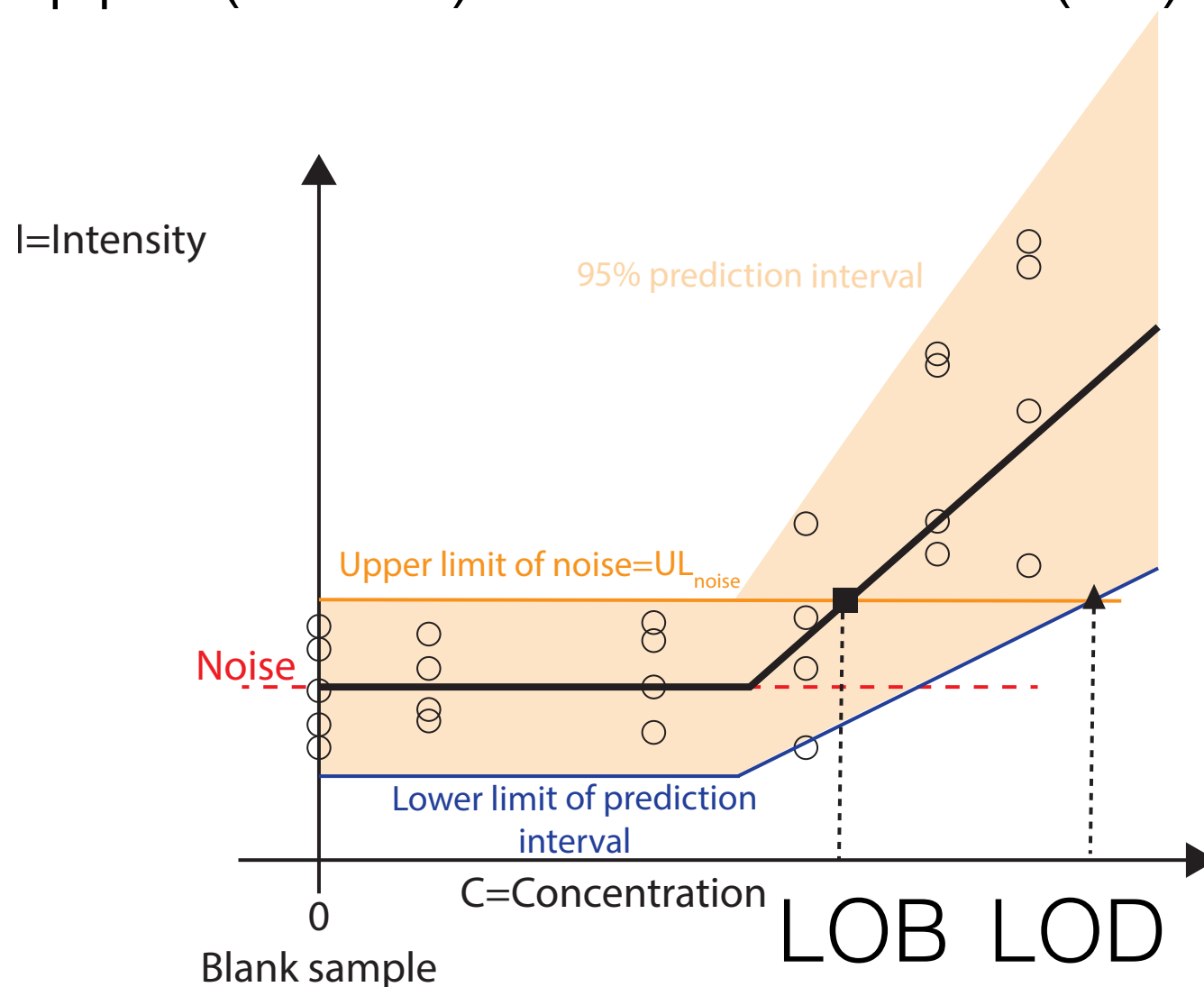
# LOD/LOQ definition

- LOB = Concentration when Intensity > Noise “on average”
- LOD = Concentration when Intensity > Noise “always”



# General LOB/LOD calculation procedure

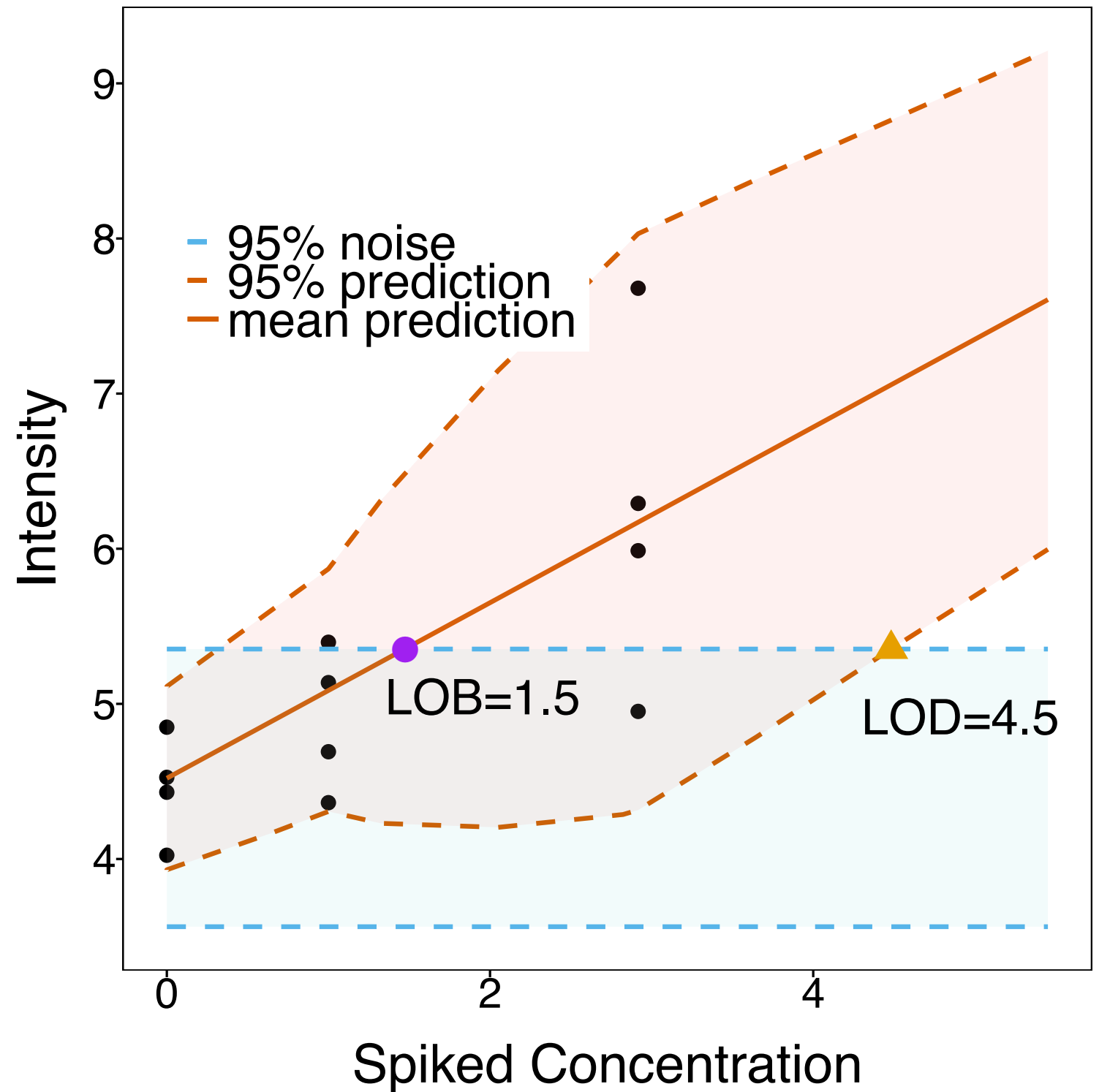
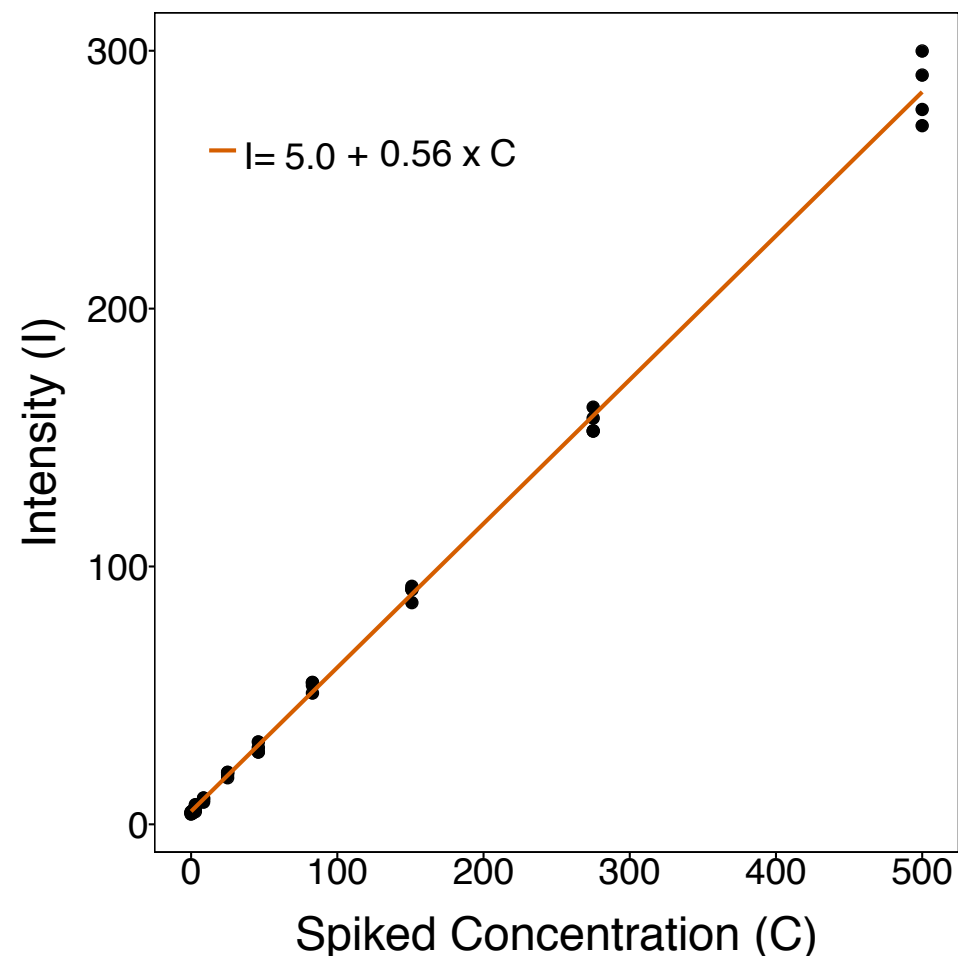
1. Normalize data (e.g. with heavy peptides)
2. Do a curve fit of Intensity  $\sim$  Concentration
3. Calculate prediction interval of fit
4. Estimate noise of blank samples
5.  $LOB = \text{Fit} \cap \text{Upper(Noise)}$      $LOD = \text{Lower(Fit)} \cap \text{Upper(Noise)}$





# Linear LOD/LOQ calculation method

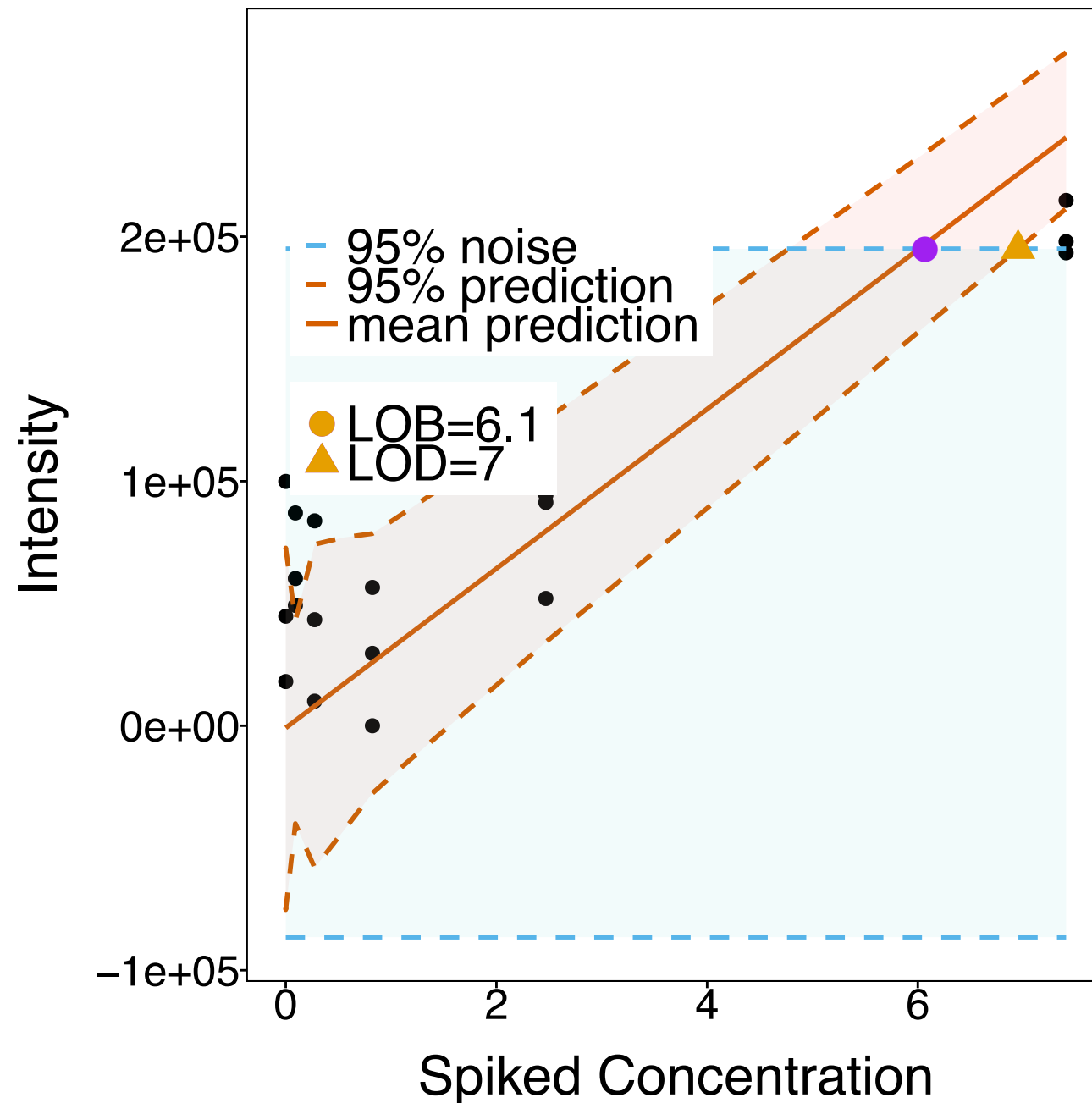
- Based on a weighted linear fit
- Prediction interval estimated by resampling data



CPTAC 7, Peptide ESDTSYVSLK, Site 86, Study IIIc

# Linear LOB/LOD calculation method

- Does not capture threshold (=leveling off at low concentrations)
- Curve fit, LOB and LOD are clearly not accurate



# Nonlinear LOB/LOD calculation methods

- Multiple methods developed to remedy problems with linear fits


Drug Development and Industrial Pharmacy, 26(6), 661–669 (2000)

## Statistical Method for Determining and Comparing Limits of Detection of Bioassays

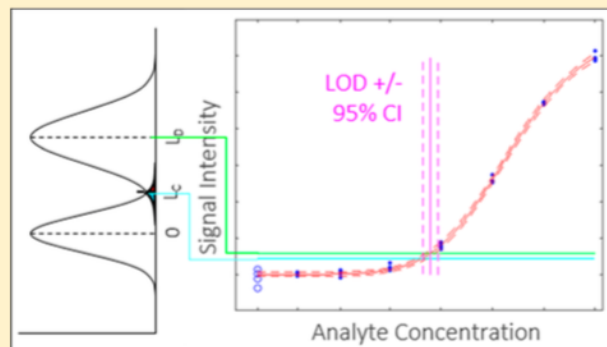
Carly A. Holstein,<sup>\*,†</sup> Maryclare Griffin,<sup>‡</sup> Jing Hong,<sup>‡</sup> and Paul D. Sampson<sup>‡</sup>

<sup>†</sup>Department of Bioengineering, University of Washington, Box 355061, Seattle, Washington 98195, United States

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 Supporting Information

**ABSTRACT:** The current bioassay development literature lacks the use of statistically robust methods for calculating the limit of detection of a given assay. Instead, researchers often employ simple methods that provide a rough estimate of the limit of detection, often without a measure of the confidence in the estimate. This scarcity of robust methods is likely due to a realistic preference for simple and accessible methods and to a lack of such methods that have reduced the concepts of limit of detection theory to practice for the specific application of bioassays. Here, we have developed a method for determining limits of detection for bioassays that is statistically robust and reduced to practice in a clear and accessible manner geared at researchers, not statisticians. This method utilizes a four-



RESEARCH PAPER

## Calibration and LOD/LOQ Estimation of a Chemiluminescent Hybridization Assay for Residual DNA in Recombinant Protein Drugs Expressed in *E. coli* Using a Four-Parameter Logistic Model

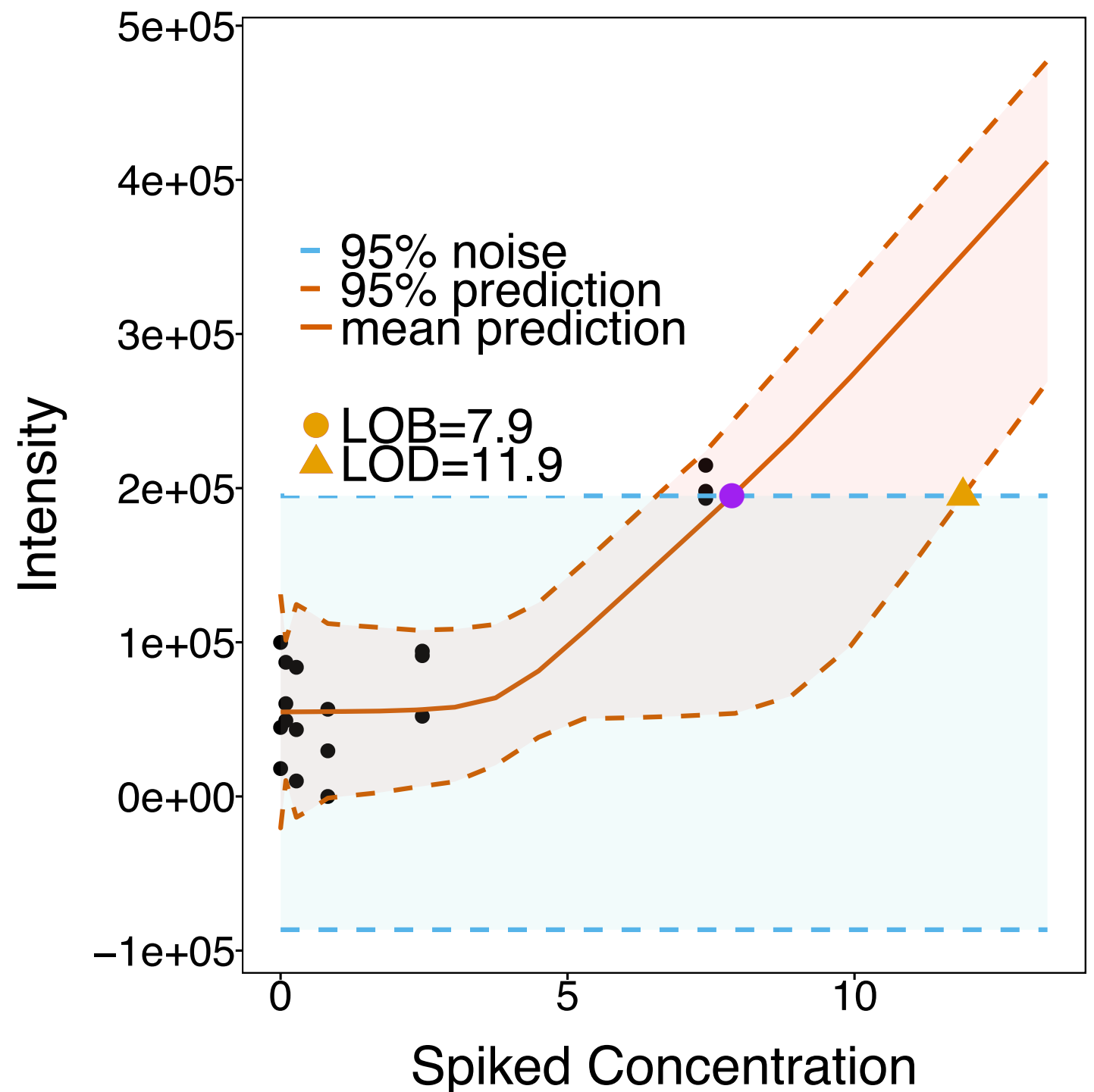
Kwan R. Lee,<sup>1,\*</sup> Byron Dipaolo,<sup>2</sup> and Xiaoying Ji<sup>2</sup>

<sup>1</sup>Statistical Sciences Department and <sup>2</sup>Bioanalytical Sciences Department, SmithKline Beecham Pharmaceuticals R&D, King of Prussia, PA 19406

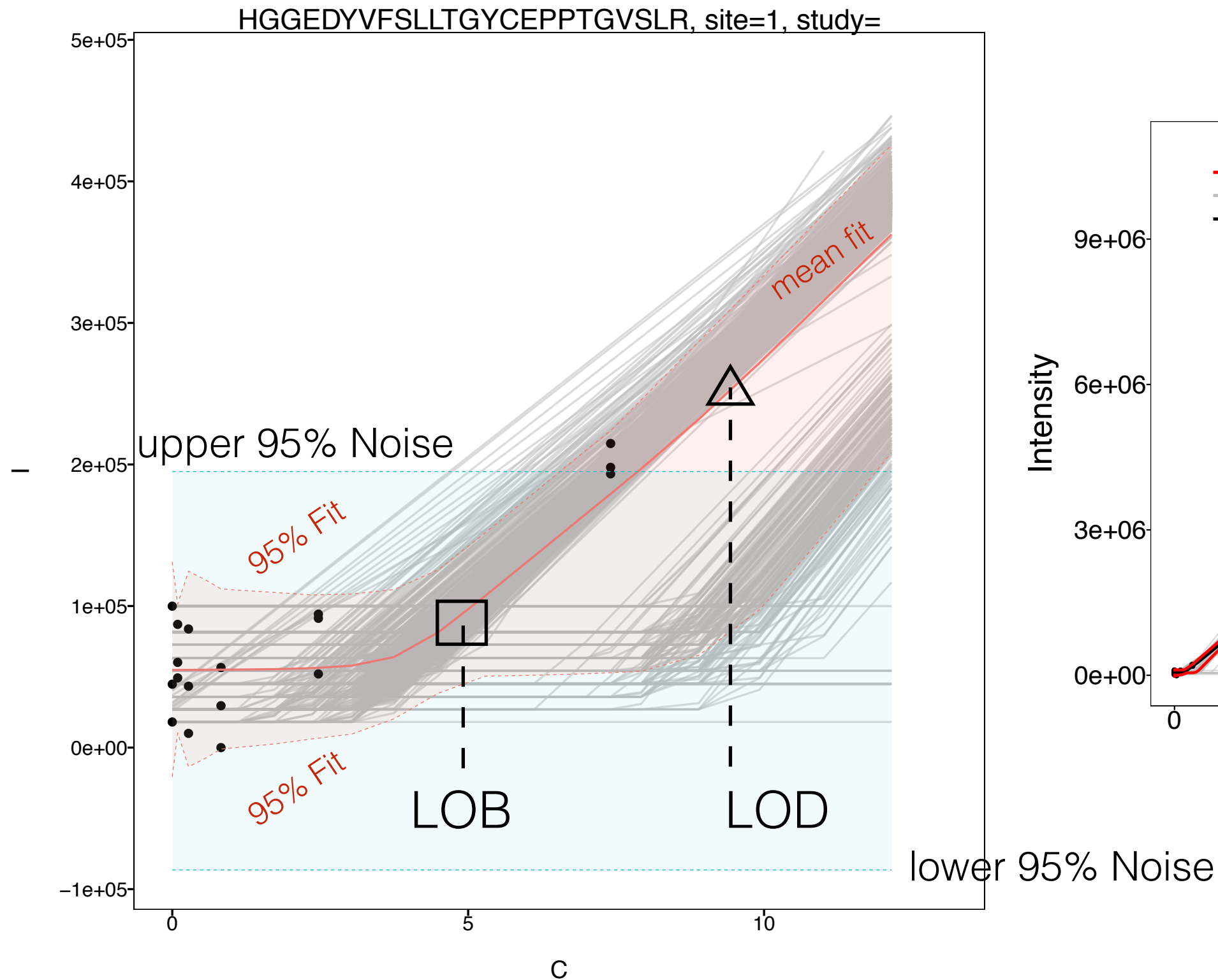
- Do not however take canonical shape of response into account
  - \*Overfitting
  - \*Numerical issues (convergence etc.)

# A new Nonlinear LOB/LOD calculation method

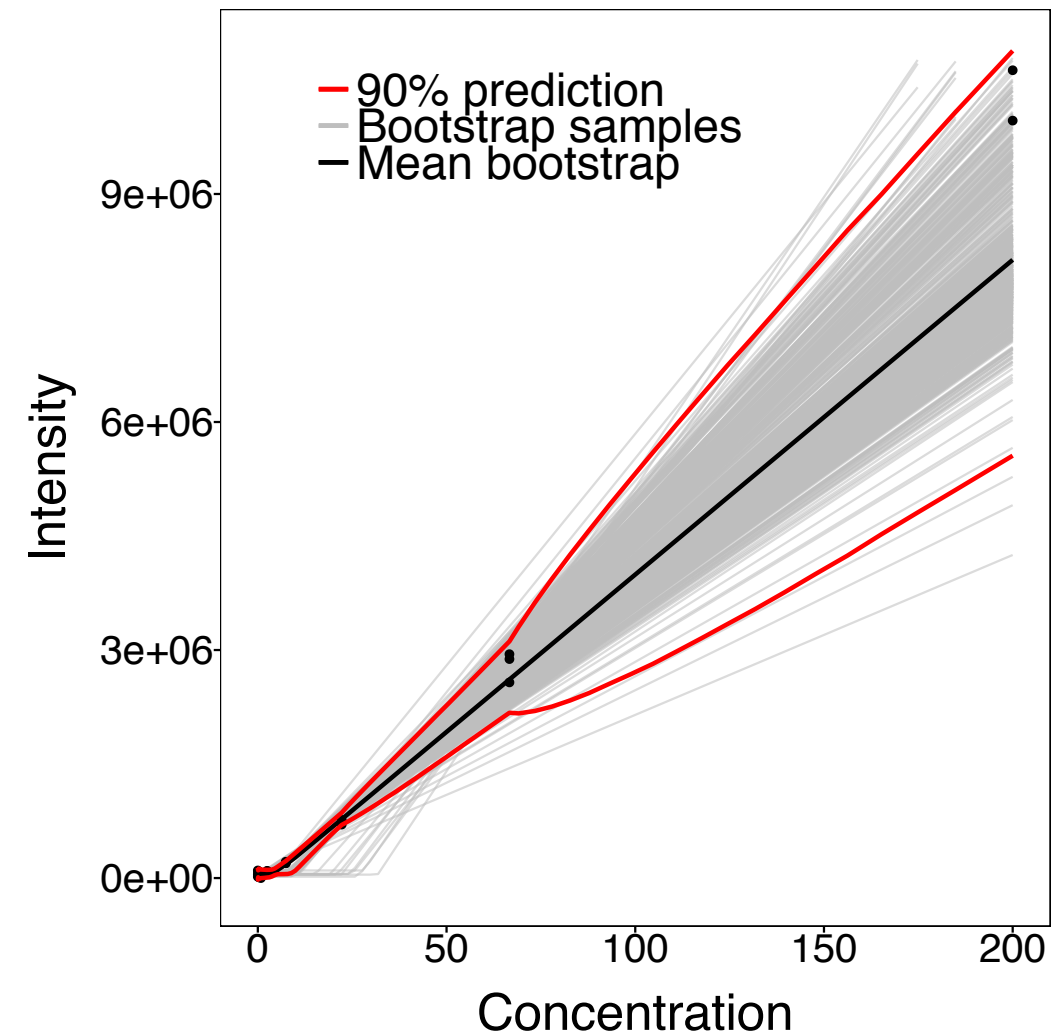
- More advanced method recently developed by the Vitek Lab
- Captures threshold at low concentrations but linear when no threshold is present
- Avoids problems with logistic regression



# A new Nonlinear LOB/LOD calculation method



Blank sample



# A new Nonlinear LOB/LOD calculation method: Results

The screenshot shows a web browser window with the address bar displaying <https://panoramaweb.org/labkey/project/CPTAC%20Assay%20Portal/begin.view?>. The browser's address bar also shows a "Secure" lock icon and a search bar labeled "Search PanoramaWeb".

The main content area of the browser displays the "CPTAC Assay Portal" page. The page has a blue header with the "Panorama" logo and "PanoramaWeb" text. A navigation menu on the left lists various assay types, including "Broad\_Carr", "Broad\_Carr\_FHCRC\_Paulovich", "FHCRC\_Paulovich", "FHCRC\_Paulovich\_2", "FHCRC\_Paulovich\_3", "JHU\_DChan\_HZhang\_ZZhang", "SNU\_KIST\_Kim\_FHCRC\_Paulovich", "UVicPC\_Borchers", and "WUSTL\_Townsend".

The main content area contains a welcome message and a description of the CPTAC Assay Portal. It states: "Welcome to the CPTAC Assay Portal on panoramaweb.org. The CPTAC Assay Portal is a web-based resource for the Clinical Proteomic Tumor Analysis Consortium (CPTAC) of the National Cancer Institute. The purpose of the CPTAC Assay Portal is to provide access to proteomic assays to the scientific community, via access to SOPs, reagents, and assay validation data. The portal is designed to support hypothesis-driven questions of the proteome, with analytical chemists equipped to perform mass spectrometry-based assays."

At the bottom of the page, there is a search section with three tabs: "Protein Search", "Peptide Search", and "Modification Search". The "Protein Search" tab is selected. Below the tabs, there is a search input field labeled "Protein name \*?", a "SEARCH" button, and two checkboxes: "Search in subfolders?" (unchecked) and "Exact matches only?" (checked).

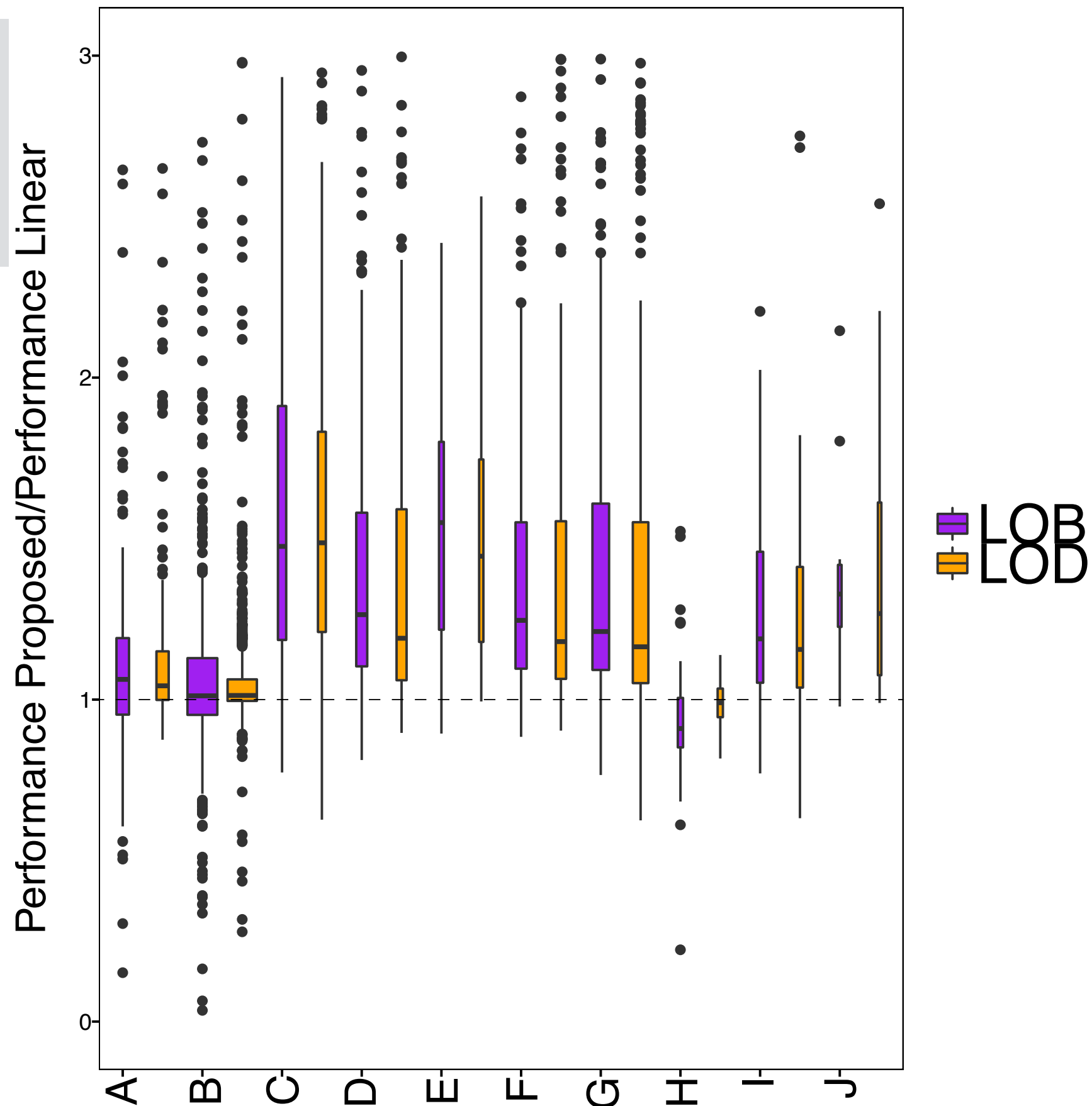
On the right side of the page, there is a "Panorama Dashboard" section with a list of links: "Panorama: Repository Software for Targeted Proteomics Assays from Skyline", "Panorama Partners Program", "Documentation", "Tutorials", "Webinars", "Publications", "Dashboard", "Example Data", "Related", and "University of Washington". Below this list, there are links for "NEXT", "EXPAND ALL", and "COLLAPSE ALL".



# A new Nonlinear LOB/LOD calculation method: Results

10 datasets from  
CPTAC assay portal

Nonlinear method  
corrects  
underestimation of  
LOD and LOB with  
linear method



# A nonlinear LOB/LOD calculation method: R package

Part of MSstats R package: <http://msstats.org>



## MSstats

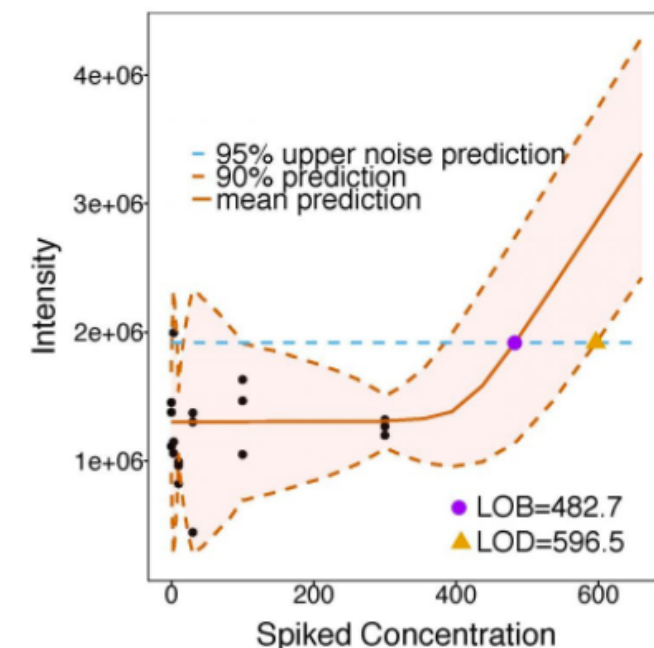
Statistical Tool For Quantitative Mass Spectrometry-Based Proteomics

[HOME](#) [MSSTATS](#) [LOB/LOD](#) [MSSTATSQC](#) [DATASETS](#) [TRAINING](#) [CONTACT](#)

## LOB/LOD ANALYSIS

### Assay characterization : estimation of limit of blanc (LoB) and limit of detection (LoD)

The need for assay characterization is ubiquitous in quantitative mass spectrometry-based proteomics. Although many assay characteristics exist, the limit of blank (LOB) and limit of detection (LOD) are particularly useful figures of merit. LOB and LOD are determined by repeatedly measuring the peak intensities of peptides in samples with known peptide concentrations, and deriving an intensity versus concentration response curve. Most commonly, a weighted linear regression is fit to the intensity-concentration response, and LOB and LOD are estimated from the fit. Linear methods, however, inaccurately characterize assays containing a noise threshold at low concentrations, which is a very common situation. We propose a new approach based on non-linear regression that correctly captures the noise threshold. In absence of a noise threshold, the estimates of LOB/LOD obtained with non-linear statistical modeling are identical to those of weighted linear regression. However, in presence of a noise threshold the non-linear model changed the estimates of LOB/LOD by up to 20-40%. It improved the accuracy of the results, and avoided the unduly optimistic estimation of these figures of merit. We implemented the non-linear regression approach in the open-source R-based software MSstats, and advocate its general use for mass spectrometric protein assay characterization.



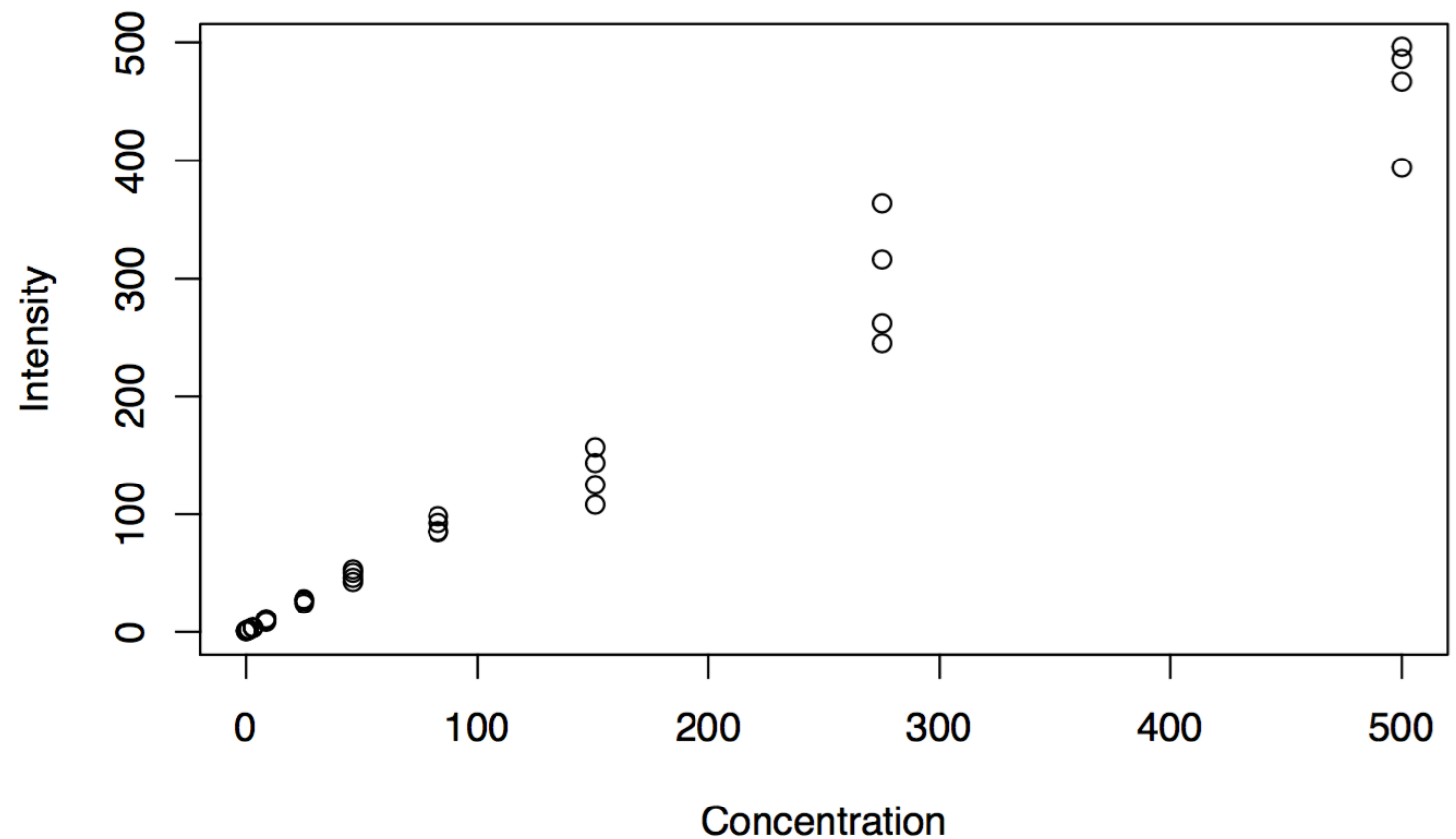
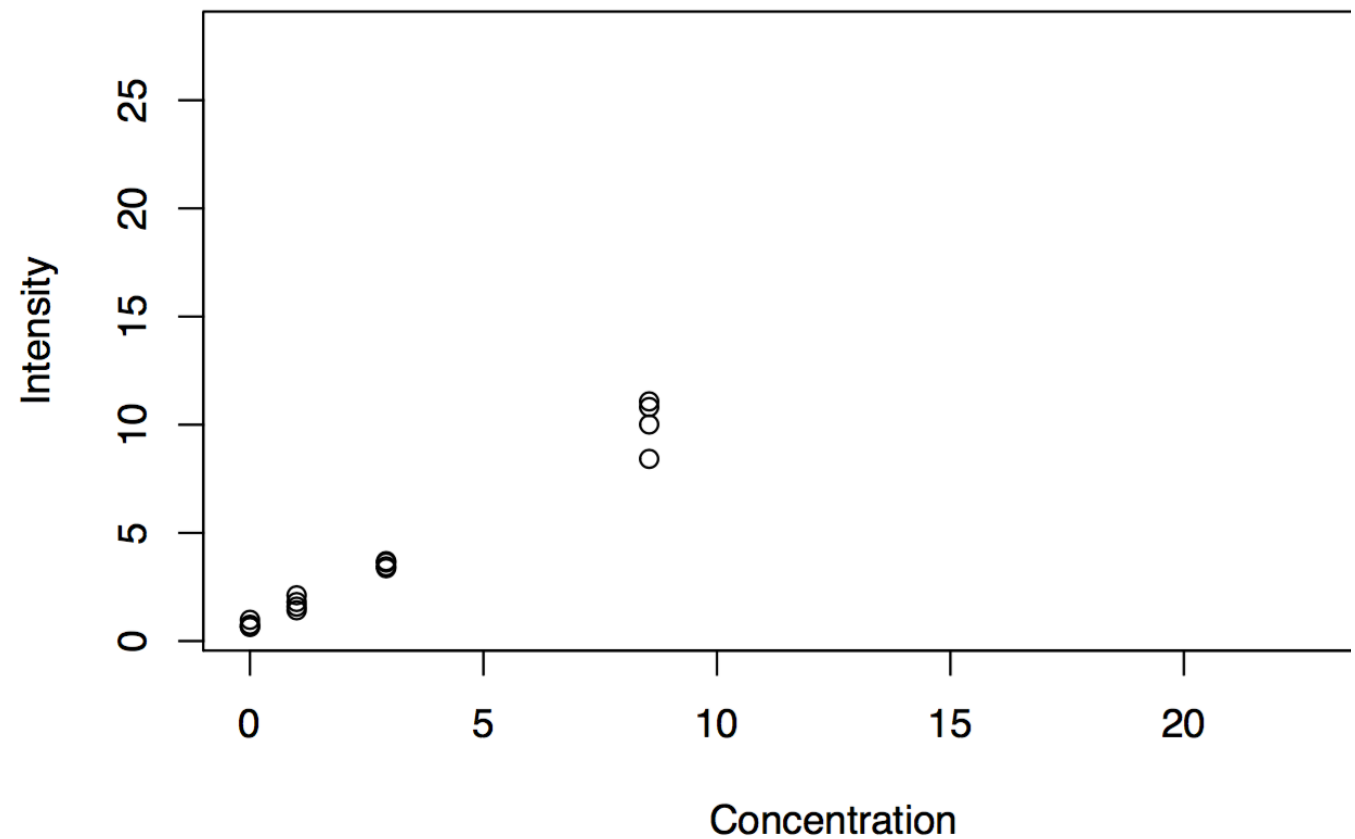


# MSstats R package: linear fit for LOB/LOD calculation

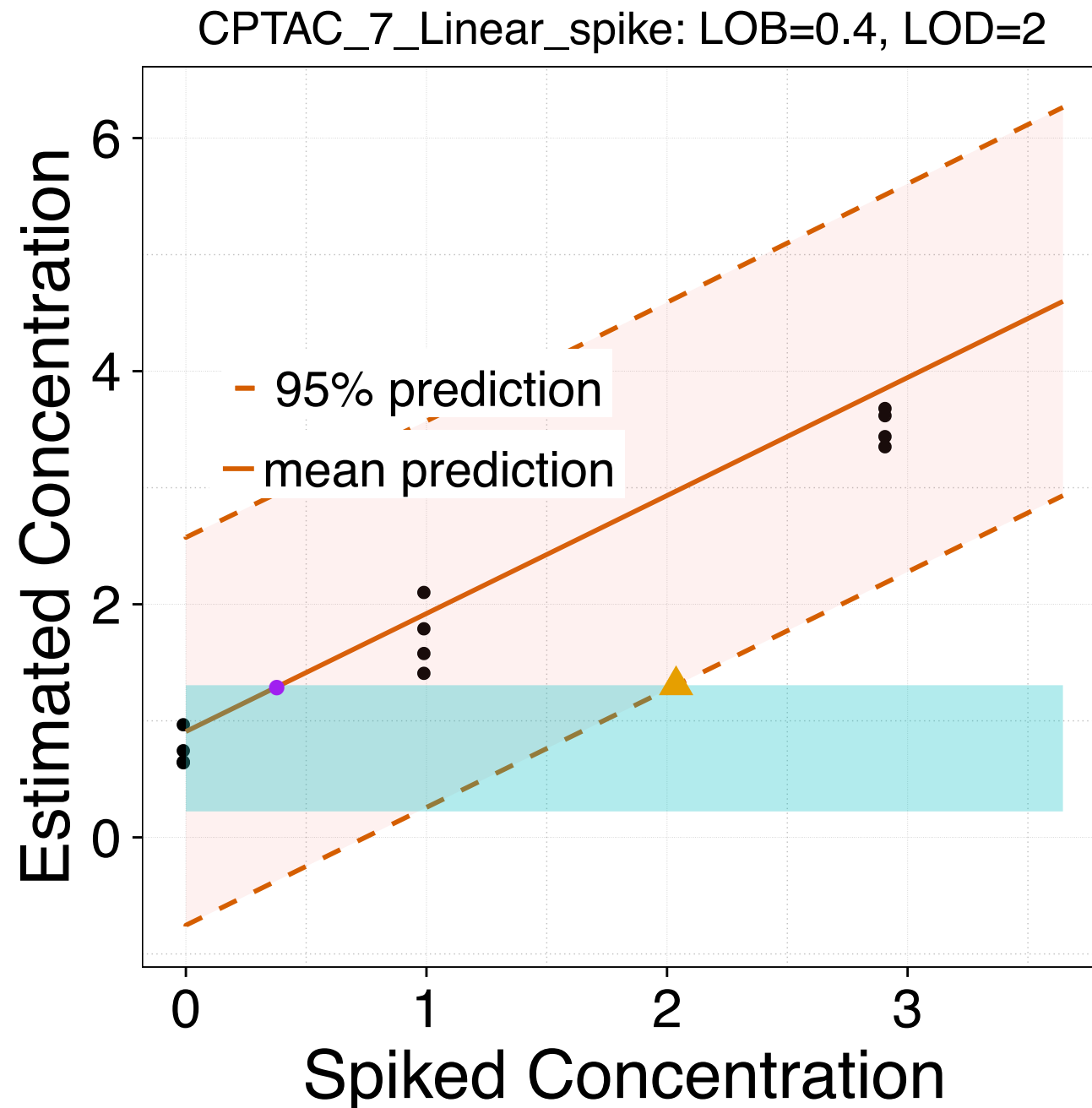
Input data set:

	INTENSITY	CONCENTRATION	NAME	REPLICATE
1	2.629110e+04	0.0576	LPPGLLANFTLLR	1
2	2.448406e+05	0.2880	LPPGLLANFTLLR	1
3	0.000000e+00	0.0000	LPPGLLANFTLLR	1
4	7.742742e+05	0.0000	LPPGLLANFTLLR	1
5	4.820082e+05	0.0000	LPPGLLANFTLLR	1
6	7.807922e+05	0.0000	LPPGLLANFTLLR	1
7	1.377281e+06	1.4400	LPPGLLANFTLLR	1
8	9.861708e+06	7.2000	LPPGLLANFTLLR	1
9	6.757440e+07	36.0000	LPPGLLANFTLLR	1
10	2.875355e+08	180.0000	LPPGLLANFTLLR	1
11	2.427150e+09	900.0000	LPPGLLANFTLLR	1
12	2.546919e+04	0.0576	LPPGLLANFTLLR	2
13	1.886122e+05	0.2880	LPPGLLANFTLLR	2
14	0.000000e+00	0.0000	LPPGLLANFTLLR	2
15	1.415216e+05	0.0000	LPPGLLANFTLLR	2
16	1.931595e+05	0.0000	LPPGLLANFTLLR	2

Showing 1 to 17 of 30 entries



# MSstats R package: Linear fit for LOB/LOD calculation: linear\_quantlim()



```
linear_quantlim(datain, alpha = 0.05, Npoints = 100, Nbootstrap = 500)
```

## Arguments

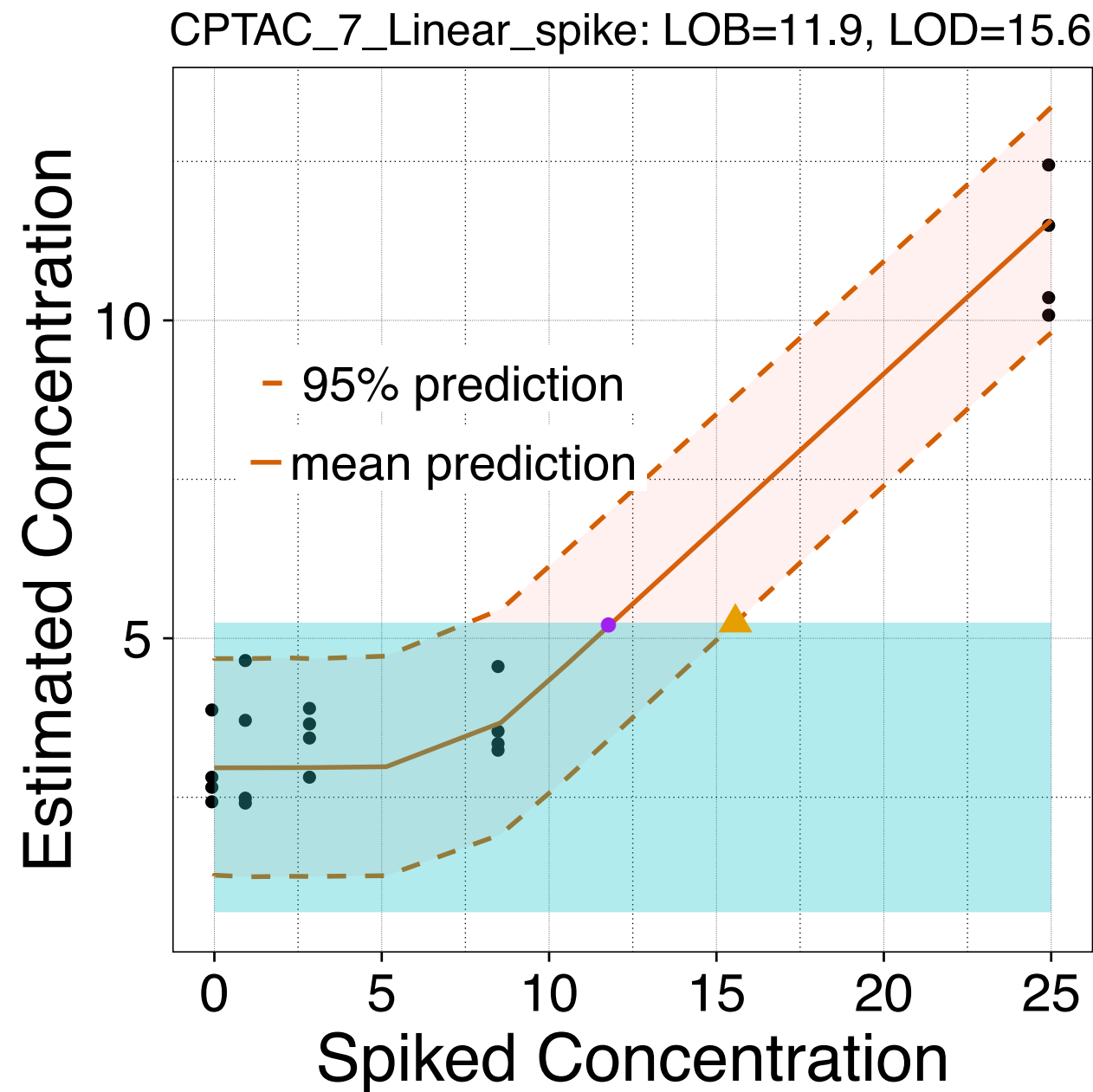
`datain`

Data frame that contains the input data. The input data frame has to contain the following columns : CONCENTRATION, INTENSITY (both of which are measurements from the spiked in experiment) and NAME which designates the name of the assay (e.g. the name of the peptide or protein)

`alpha`

Probability level to estimate the LOB/LOD

# MSstats R package: Nonlinear fit for LOB/LOD calculation: `nonlinear_quantlim()`



## Usage

```
nonlinear_quantlim(datain, alpha = 0.05, Npoints = 100, Nbootstrap = 500)
```

## Arguments

- datain** Data frame that contains the input data. The input data frame has to contain the following columns : CONCENTRATION, INTENSITY (both of which are measurements from the spiked in experiment) and NAME which designates the name of the assay (e.g. the name of the peptide or protein)
- alpha** Probability level to estimate the LOB/LOD

# MSstats R package: Nonlinear vs Linear fit for LOB/LOD calculation

