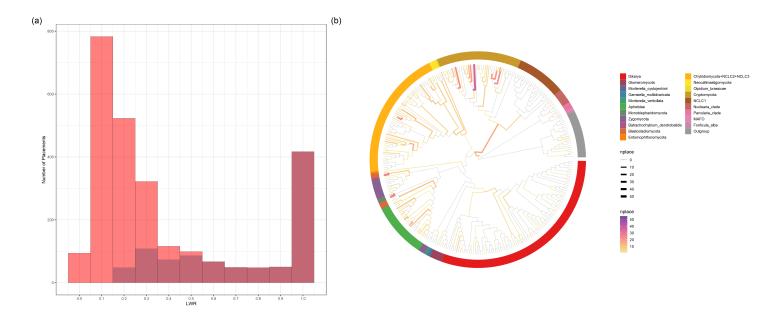
Phylogenetic placement visualization with treeio-ggtree method

0.1 Favorable placement filtration after parsing jplace file

```
# Required packages
library(treeio)
library(ggtree)
library(jsonlite)
library(dplyr)
library(ggtreeExtra)
library(RColorBrewer)
library(colorspace)
library(ggplot2)
library(aplot)
# Read jplace
jtree <- from JSON("../exampledata/Holomycota/HolomycotaV4_alignedtrim.jplace")
phylo <- treeio:::jplace_treetext_to_phylo(jtree$tree)</pre>
placements <- treeio:::extract.placement(jtree,phylo)</pre>
# placement filtering
filtered_placements <- group_by(placements, .data$name) %>%
    filter(.data$like_weight_ratio == max(.data$like_weight_ratio))
p1 <- ggplot(data = placements,aes(x=like_weight_ratio)) +</pre>
    geom_histogram(data=filtered_placements,aes(x=like_weight_ratio,..count..),fill="skyblue", color="white",s
    geom_histogram(data=placements,aes(x=like_weight_ratio,..count..),fill="red", color="black",size=0.1,binwi
    labs(y="Number of Placements",x="LWR") +
    scale_x_continuous(breaks = c(seq(0,1,0.1))) +
    theme_bw()
tree1 <- as.treedata(phylo)</pre>
td <- as_tibble(tree1)</pre>
tree1@data <- group_by(filtered_placements, .data$node) %>% summarize(nplace=n()) %>%
    full_join(td, by='node') %>%
    mutate(nplace = ifelse(is.na(.data$nplace), 0, .data$nplace))
p2 <- ggtree(tree1,layout = "circular",branch.length = "none",aes(color=nplace,size=nplace)) +
    scale_color_continuous_sequential(palette = "Sunset",limits =c(1, 55)) +
    scale_size_continuous(range=c(0.1, 2),limits =c(0, 55))
V4_group <- read.csv(file = "../exampledata/Holomycota/V4_group.csv")
getPalette <- colorRampPalette(brewer.pal(9, "Set1"))</pre>
cols <- getPalette(21)</pre>
levels <- rev(unique(V4_group$group))</pre>
p3 <- p2%<+% V4_group +
    geom_fruit(
        geom = geom_tile,
        mapping = aes(fill = factor(group,levels = levels)),
        width = 2,
        offset = 0.1
    ) +
    scale_fill_manual(
        name = "",
        values = cols,
```

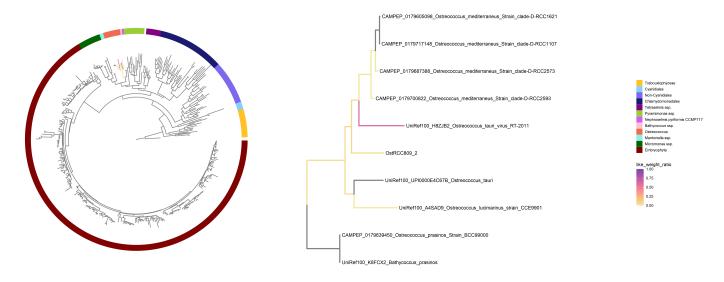


0.2 Utility to explore the placement uncertainty

```
# Required packages
library(ggplot2)
library(treeio)
library(ggtree)
library(jsonlite)
library(dplyr)
library(colorspace)
library(ggtreeExtra)
library(aplot)
library(patchwork)
# Read jplace
jtree <- fromJSON("../exampledata/subtree/pplacer_Amt_subtree.jplace")</pre>
phylo <- treeio:::jplace_treetext_to_phylo(jtree$tree)</pre>
placements <- treeio:::extract.placement(jtree,phylo)</pre>
seq_name <- "saltern1"</pre>
sgplacement <- group_by(placements, .data$name) %>%
    filter(.data$name == seq_name)
tree1 <- as.treedata(phylo)</pre>
tree2 <- left_join(tree1,sgplacement,by="node")</pre>
p1 <- ggtree(tree2,layout = "circular",aes(color=like_weight_ratio)) +</pre>
    scale_color_continuous_sequential(palette = "Sunset",limits =c(0, 1)) +
    ggtitle(seq_name)
```

```
Amt_tiplabel <- read.csv(file = "../exampledata/subtree/Amt_tiplabel.csv",</pre>
                         header = TRUE)
head(Amt_tiplabel)
##
     node
               tiplabel
                                    Group
## 1
        1 Asterochloris Trebouxiophyceae
## 2
        2 Asterochloris Trebouxiophyceae
## 3
              Coccomyxa Trebouxiophyceae
## 4
        4 Stichococcus Trebouxiophyceae
## 5
        5
              Galdieria
                              Cyanidiales
## 6
              Galdieria
                              Cyanidiales
taxon_groups <- c("Trebouxiophyceae", "Cyanidiales", "Non-Cyanidiales",
                  "Chlamydomonadales", "Tetraselmis ssp.", "Pyramimonas ssp.",
                  "Nephroselmis pyriformis CCMP717", "Bathycoccus ssp.",
                  "Ostreococcus", "Mantoniella ssp.", "Micromonas ssp.",
                  "Embryophyta")
Amt_tiplabel$Group <- factor(Amt_tiplabel$Group, levels=taxon_groups)</pre>
cols <- c("#FFC125","#87CEFA","#7B68EE","#191970","#800080",
          "#9ACD32","#D15FEE","#FFCOCB","#EE6A50","#8DEEEE",
          "#006400", "#800000", "#B0171F")
p2 <- p1%<+% Amt_tiplabel +
    geom_fruit(
        geom = geom_tile,
        mapping = aes(fill = Group),
        width = 0.2,
        offset = 0.01
    ) +
    scale_fill_manual(
        name = "",
        values = cols,
        guide = guide_legend(
            keywidth = 1,
            keyheight = 1,
            order = 3
        ),
        na.translate = FALSE
    )
# viewClade(p1,659) # viewClade
tree3 <- tree_subset(tree2, node=657, levels_back=0)</pre>
p3 <- ggtree(tree3,layout = "rect",aes(color=like_weight_ratio),size=1) +
    scale_color_continuous_sequential(palette = "Sunset",limits =c(0, 1)) +
    geom_tiplab(size=4,color="black",offset = 0.01) + xlim(0,2)
plot_list(p2, p3, tag_levels = list(c('(a)', '(b)'), '1'), guides = 'collect',
          nrow = 1, tag_size = 20)
```

(a) saltern1 (b)



0.3 Merge placement information to the placement tree

```
# Required packages
library(treeio)
library(ggtree)
library(ggplot2)
library(ggtreeExtra)
library(colorspace)
library(dplyr)
library(patchwork)
tree1 <- read.jplace(file = "../exampledata/Mitsi/rsbl20190182supp2.jplace")</pre>
tree2 <- read.tree(file = "../exampledata/Mitsi/rsbl20190182supp7.tre")</pre>
tree2 <- as.treedata(tree2)</pre>
p1 <- ggtree(tree1,layout = "circular",aes(color=nplace,size=nplace)) +
    scale_color_viridis_c(option="viridis",limit=c(0.1,120)) +
    scale_size_continuous(range=c(0, 2))
# Extract placement informations
pla <- get.placements(tree1)</pre>
# merge placement data onto best-hit tree
\#pla$label \leftarrow unlist(lapply(pla$name,function(x){paste("QUERY_",x,sep = "_")}))
pla$label <- pla$name</pre>
pla2 <- select(pla,c("name","label", "likelihood","like_weight_ratio"))</pre>
tree3 <- as_tibble(tree2)</pre>
tree4 <- left_join(tree3,pla2,by="label",)</pre>
tree4$group <- gsub("_1","",tree4$label)</pre>
tree4$group <- gsub("_2","",tree4$group)</pre>
tree4$group <- lapply(tree4$group,function(x){unlist(strsplit(x,"_"))[c(2)]})</pre>
tree4$group <- gsub("Rhabditophora", "Bothrioplana_semperi", tree4$group)</pre>
s_group <- c("Catenulida", "Polycladida", "Macrostomorpha", "Prorhynchidae",</pre>
              "Proseriata", "Rhabdocoela", "Bothrioplana_semperi", "Fecampiidae",
              "Tricladida", "Prolecithophora", "Monogenea", "Trematoda", "Cestoda")
```

```
otu_group <- unlist(list(tree4[grep("^OTU.",tree4$group),"group"]))</pre>
all_group <- c(s_group,otu_group)</pre>
tree4[!(tree4$group %in% all_group), "group"] <- "Outgroup"</pre>
tree4[seq(193,201),"group"] <- "Prorhynchidae"</pre>
tree4[seq(249,252),"group"] <- "Fecampiidae"</pre>
tree4[seq(169,171), "group"] <- "Gnosonesimidae"
tree4[seq(119,121),"group"] <- "Clade 1"</pre>
tree4[seq(174,192),"group"] <- "Clade 2"</pre>
s_group2 <- c("Trematoda", "Cestoda", "Monogenea",</pre>
              "Prolecithophora", "Tricladida", "Fecampiidae",
              "Bothrioplana_semperi", "Rhabdocoela", "Proseriata", "Prorhynchidae",
              "Clade 2", "Gnosonesimidae", "Macrostomorpha", "Polycladida",
              "Clade 1", "Catenulida", "Outgroup")
tree4[is.na(tree4$label), "group"] <- NA</pre>
tree4$group <- factor(tree4$group, levels=s_group2)</pre>
tree4[is.na(tree4$like_weight_ratio),"like_weight_ratio"] <- 0</pre>
tree5 <- as.treedata(tree4)</pre>
# tree5 <- root(tree5, node=572)</pre>
p2<- ggtree(tree5,
              layout = "circular",
              # branch.length = "none",
) + geom_tree(
    aes(
        color=like_weight_ratio,
        size=like_weight_ratio
    )
) +
    # geom_tiplab(size=2,offset=0.02,align=TRUE) +
    scale color continuous sequential(palette = "Sunset", limits=c(0.000001, 0.2)) +
    scale_size_continuous(range=c(0, 1.5)) +
    geom_tiplab2(data=td_filter(like_weight_ratio > 0),align=TRUE,size=3,offset=0.1)
p3 <- scaleClade(p2,
            node=c(MRCA(tree5, "SA_OTU14871", "SA_OTU158753")),
            scale=5)
p4 <- scaleClade(p3,
                  node=c(MRCA(tree5, "SA_OTU22294", "SA_OTU9482")),
                  scale=5)
# Annotate Group Labels
#ggtree(tree5) + geom_text2(aes(x=branch, label=node))
p5 < - p4 +
    geom_fruit(geom = geom_tile,
                mapping = aes(fill = group),
                width = 0.1,
                offset = 0.01)
p5
```

