

Standardizing-Marine-Biological-Data

Brett Johnson

2021-10-15

Contents

Preface	5
1 Introduction	7
1.1 Data Structures	7
1.2 Ontologies	7
1.3 Controlled Vocabularies	8
1.4 Technologies	10
1.5 Notes on Integrating OBIS, Darwin Core as it relates to OOS's .	10
1.6 Metadata	10
1.7 Data QC	10
2 Applications	13
2.1 Salmon Ocean Ecology Data	13
2.2 Example Two	18
3 Final Words	19
References	21

Preface

Biological data structures, definitions, measurements, and linkages are necessarily as diverse as the systems they represent. This presents a real challenge when integrating data across biological research domains such as ecology, oceanography, fisheries, and climate sciences.

Chapter 1

Introduction

The world of standardizing marine biological data can seem complex for the naive oceanographer, biologist, scientist, or programmer. Transforming and integrating data is about combining the right standards for your desired interoperability with other data types. For example, interoperating fish biology measurements with climate level variables. There are a few concepts necessary to make this possible such as standard data structures, controlled vocabularies and knowledge representations, along with metadata standards to facilitate data discovery. This will permit the inclusion of more data and broader access to better ecosystem based models. Many scientific domains data handling practices are currently being reshaped in light of recent advances in computing power, technology, and data science.

1.1 Data Structures

The OBIS-ENV Darwin Core Archive Data Structure.

OBIS manual

1.2 Ontologies

An ontology is a classification system for establishing a hierarchically related set of concepts. Concepts are often terms from controlled vocabularies.

From Marine Metadata: # TODO: add link

“Ontologies can include all of the following, but are not required to include them, depending on which perspective from above you adhere to:

Classes (general things, types of things) Instances (individual things) Relationships among things Properties of things Functions, processes, constraints, and rules relating to things”

TODO: Research Unified Modelling Language?

Environment Ontology (EnvO) EnvO is a community ontology for the concise, controlled description of environments.

1.3 Controlled Vocabularies

There are a number of controlled vocabularies that are used to describe parameters commonly used in specific research domains. This allows for greater interoperability of data sets within the domain, and ideally between domains. Here, we strive to document a number of relevant examples.

- Climate and Format (CF) Standard Names The purpose of the `standard_name` attribute is to provide a succinct and distinguishing description of a variable, in a way that encourages interoperability. These terms are typically for physical observations, however, there have been advancements in aligning biological taxa into the CF standard names (see here).
- NERC Vocabulary Server The NVS gives access to standardised and hierarchically-organized vocabularies.
 - Device categories using the SeaDataNet device categories
 - Device make/model using the SeaVoX Device Catalogue
 - Platform categories using SeaVoX Platform Categories
 - Platform instances using the ICES Platform Codes
 - Unit of measure
- GCMD Keywords (NASA)
- Geographic Domain/Features of Interest
- GeoLink base ontology was part of the EarthCube GeoLink Project

Note: To describe a measurement or fact of a biological specimen that conforms to Darwin Core standards, it’s necessary to use the ‘Biological entity described elsewhere’ method rather than taxon specific.

1.3.1 Taxonomy

- The World Registry of Marine Species (WoRMS) The aim of a World Register of Marine Species (WoRMS) is to provide an authoritative and comprehensive list of names of marine organisms, including information on synonymy. While the highest priority goes to valid names, other names in use are included so that this register can serve as a guide to interpret taxonomic literature.

1.3.2 Resources

1.3.3 Oceanography

Biological and Chemical Oceanography Data Management Office

Marine metadata interoperability vocab resources

1.3.4 Biology

BioPortal Ecosystem Ontology

1.3.5 NERC Search Interfaces

- SeaDataNet Common Vocab Search Interface:
- SeaDataNet Common Vocabularies:
- SeaDataNet Vocab Library
- Measurement Types in OBIS

1.3.6 Geosciences

UDUNITS are more common unit measurements in geosciences

1.3.7 Eco/EnvO

Environment Ontology including genomics.

1.3.8 Wild Cards

Question: Not sure use case for this.

P01 Biological Entity Parameter Code Builder

1.4 Technologies

1.4.1 ERDDAP

ERDDAP can be thought of as a data server. It provides ‘easier access to scientific data’ by providing a consistent interface that aggregates many disparate data sources. It does this by providing translation services between many common file types for gridded arrays (‘net CDF’ files) and tabular data (spreadsheets). Data access is also made easier because it unifies different types of data servers and access protocols. Here is a basic erddap installation that walks you through how to load a data set.

1.5 Notes on Integrating OBIS, Darwin Core as it relates to OOS’s

1.6 Metadata

OBIS uses the GBIF EML profile (version 1.1). In case data providers use ISO19115/ISO19139, there is a mapping available here: <http://rs.gbif.org/schema/eml-gbif-profile/1.1/eml2iso19139.xsl> This will be important for integrating OBIS datasets to other CIOOS and IOOS metadata profiles.

1.7 Data QC

There are a number of tools available to check the quality of data or check your data format against the expected standard.

OBIS Datatools shows some great R packages for this.

1.7.1 Compliance Checking

LifeWatch Belgium provides a number of tools to check your data against. Specifically you can test OBIS data format and see a map of your sample locations to check if they are on land. See <http://www.lifewatch.be/data-services/>

There’s also the GBIF data validator which allows anyone with a GBIF-relevant dataset to receive a report on the syntactical correctness and the validity of the content contained within the dataset.

1.7.2 Semantic Web and Darwin Core

Lessons learned from adapting the Darwin Core vocabulary standard for use in RDF

1.7.3 Resource Description Framework

Darwin Core Resource Description Framework Guide

Chapter 2

Applications

Some *significant* applications are demonstrated in this chapter.

2.1 Salmon Ocean Ecology Data

2.1.1 Intro

One of the goals of the Hakai Institute and the Canadian Integrated Ocean Observing System (CIOOS) is to facilitate Open Science and FAIR (findable, accessible, interoperable, reusable) ecological and oceanographic data. In a concerted effort to adopt or establish how best to do that, several Hakai and CIOOS staff attended an International Ocean Observing System (IOOS) Code Sprint in Ann Arbor, Michigan between October 7–11, 2019, to discuss how to implement FAIR data principles for biological data collected in the marine environment.

The Darwin Core is a highly structured data format that standardizes data table relations, vocabularies, and defines field names. The Darwin Core defines three table types: **event**, **occurrence**, and **measurementOrFact**. This intuitively captures the way most ecologists conduct their research. Typically, a survey (event) is conducted and measurements, counts, or observations (collectively measurementOrFacts) are made regarding a specific habitat or species (occurrence).

In the following script I demonstrate how I go about converting a subset of the data collected from the Hakai Institute Juvenile Salmon Program and discuss challenges, solutions, pros and cons, and when and what’s worthwhile to convert to Darwin Core.

The conversion of a dataset to Darwin Core is much easier if your data are already tidy (normalized) in which you represent your data in separate tables

that reflect the hierarchical and related nature of your observations. If your data are not already in a consistent and structured format, the conversion would likely be very arduous and not intuitive.

2.1.2 event

The first step is to consider what you will define as an event in your data set. I defined the capture of fish using a purse seine net as the **event**. Therefore, each row in the **event** table is one deployment of a seine net and is assigned a unique **eventID**.

My process for conversion was to make a new table called **event** and map the standard Darwin Core column names to pre-existing columns that serve the same purpose in my original **seine_data** table and populate the other required fields.

#TODO: Include abiotic measurements (YSI temp and salinity from 0 and 1 m) to hang off

```
event <- tibble(datasetName = "Hakai Institute Juvenile Salmon Program",
  eventID = survey_seines$seine_id,
  eventDate = date(survey_seines$survey_date),
  eventTime = paste0(survey_seines$set_time, "-0700"),
  eventRemarks = paste3(survey_seines$survey_comments, survey_seines$seine_id),
  decimalLatitude = survey_seines$lat,
  decimalLongitude = survey_seines$long,
  locationID = survey_seines$site_id,
  coordinatePrecision = 0.00001,
  coordinateUncertaintyInMeters = 10,
  country = "Canada",
  countryCode = "CA",
  stateProvince = "British Columbia",
  habitat = "Nearshore marine",
  geodeticDatum = "EPSG:4326 WGS84",
  minimumDepthInMeters = 0,
  maximumDepthInMeters = 9, # seine depth is 9 m
  samplingProtocol = "http://dx.doi.org/10.21966/1.566666", # This is the protocol
  language = "en",
  license = "http://creativecommons.org/licenses/by/4.0/legalcode",
  bibliographicCitation = "Johnson, B.T., J.C.L. Gan, S.C. Godwin, M. Krueger",
  references = "https://github.com/HakaiInstitute/jsp-data",
  institutionID = "https://www.gbif.org/publisher/55897143-3f69-42f1-810a-4b8b8b8b8b8b",
  institutionCode = "Hakai"
)
```

2.1.3 occurrence

Next you'll want to determine what constitutes an occurrence for your data set. Because each event captures fish, I consider each fish to be an occurrence. Therefore, the unit of observation (each row) in the occurrence table is a fish. To link each occurrence to an event you need to include the `eventID` column for every occurrence so that you know what seine (event) each fish (occurrence) came from. You must also provide a globally unique identifier for each occurrence. I already have a locally unique identifier for each fish in the original `fish_data` table called `ufn`. To make it globally unique I pre-pend the organization and research program metadata to the `ufn` column.

Not every fish is actually collected and given a Universal Fish Number (UFN) in our fish data tables, so in our field data sheets we record the total number of fish captured and the total number retained. So to get an occurrence row for every fish captured I create a row for every fish caught (minus the number taken) and create a generic numeric id (ie hakai-jsp-1) in one table and then join that to the fish table that includes a row for every fish retained that already has a UFN.

```
## make table long first
seines_total_long <- survey_seines %>%
  select(seine_id, so_total, pi_total, cu_total, co_total, he_total, ck_total) %>%
  pivot_longer(-seine_id, names_to = "scientificName", values_to = "n")

seines_total_long$scientificName <- recode(seines_total_long$scientificName, so_total = "Oncorhynchus")

seines_taken_long <- survey_seines %>%
  select(seine_id, so_taken, pi_taken, cu_taken, co_taken, he_taken, ck_taken) %>%
  pivot_longer(-seine_id, names_to = "scientificName", values_to = "n_taken")

seines_taken_long$scientificName <- recode(seines_taken_long$scientificName, so_taken = "Oncorhynchus")

## remove records that have already been assigned an ID because they were actually retained
seines_long <- full_join(seines_total_long, seines_taken_long, by = c("seine_id", "scientificName"))
seines_long <- drop_na() %>%
  mutate(n_not_taken = n - n_taken) %>% #so_total includes the number taken so I subtract n_taken
  select(-n_taken, -n) %>%
  filter(n_not_taken > 0)

all_fish_not_retained <-
  seines_long[rep(seq.int(1, nrow(seines_long)), seines_long$n_not_taken), 1:3] %>%
  select(-n_not_taken) %>%
  mutate(prefix = "hakai-jsp-",
         suffix = 1:nrow(.),
         occurrenceID = paste0(prefix, suffix))
```

```

) %>%
  select(-prefix, -suffix)

#

# Change species names to full Scientific names
latin <- fct_recode(fish_data$species, "Oncorhynchus nerka" = "SO", "Oncorhynchus gorbuscha" = "GO",
  as.character())

fish_retained_data <- fish_data %>%
  mutate(scientificName = latin) %>%
  select(-species) %>%
  mutate(prefix = "hakai-jsp-",
    occurrenceID = paste0(prefix, ufn)) %>%
  select(seine_id, scientificName, occurrenceID)

occurrence <- bind_rows(all_fish_not_retained, fish_retained_data) %>%
  rename(eventID = seine_id) %>% # rename = dplyr::rename; vs plyr::rename
  mutate(`Life stage` = "juvenile")

unique_taxa <- unique(occurrence$scientificName)
worms_names <- wm_records_names(unique_taxa) # library(worms)
df_worms_names <- bind_rows(worms_names) %>%
  select(scientificName = scientificname,
    scientificNameAuthorship = authority,
    taxonRank = rank,
    scientificNameID = lsid
  )

#include bycatch species

unique_bycatch <- unique(bycatch$scientificName) %>% glimpse()

## chr [1:29] "Oncorhynchus nerka" "Oncorhynchus tshawytscha" ...

by_worms_names <- wm_records_names(unique_bycatch) %>%
  bind_rows() %>%
  select(scientificName = scientificname,
    scientificNameAuthorship = authority,
    taxonRank = rank,
    scientificNameID = lsid
  )

bycatch_occurrence <- bycatch %>%
  select(eventID = seine_id, occurrenceID, scientificName, `Life stage` = bm_ageclass)

```



```

  filter(scientificName != "unknown")

  bycatch_occurrence$`Life stage`[bycatch_occurrence$`Life stage` == "J"] <- "juvenile"
  bycatch_occurrence$`Life stage`[bycatch_occurrence$`Life stage` == "A"] <- "adult"
  bycatch_occurrence$`Life stage`[bycatch_occurrence$`Life stage` == "Y"] <- "Young of year"

  combined_worms_names <- bind_rows(by_worms_names, df_worms_names) %>%
    distinct(scientificName, .keep_all = TRUE)

  occurrence <- bind_rows(bycatch_occurrence, occurrence)

  occurrence <- left_join(occurrence, combined_worms_names) %>%
    mutate(basisOfRecord = "HumanObservation",
           occurrenceStatus = "present")

  write_csv(occurrence, "../datasets/hakai_salmon_data/raw_data/occurrence.csv") # here::here("../",

# This removes events that didn't result in any occurrences
  event <- dplyr::semi_join(event, occurrence, by = 'eventID') %>%
    mutate(coordinateUncertaintyInMeters = ifelse(is.na(decimalLatitude), 1852, coordinateUncertain

  simple_sites <- sites %>%
    select(site_id, ocgy_std_lat, ocgy_std_lon)

  event <- dplyr::left_join(event, simple_sites, by = c("locationID" = "site_id")) %>%
    mutate(decimalLatitude = coalesce(decimalLatitude, ocgy_std_lat),
           decimalLongitude = coalesce(decimalLongitude, ocgy_std_lon)) %>%
    select(-c(ocgy_std_lat, ocgy_std_lon))

  write_csv(event, "../datasets/hakai_salmon_data/raw_data/event.csv") # here::here("../", "datasets/

```

2.1.4 measurementOrFact

To convert all your measurements or facts from your normal format to Darwin Core you essentially need to put all your measurements into one column called `measurementType` and a corresponding column called `MeasurementValue`. This standardizes the column names are in the `measurementOrFact` table. There are a number of predefined `measurementTypes` listed on the NERC database that should be used where possible. I found it difficult to navigate this page to find the correct `measurementType`.

Here I convert length, and weight measurements that relate to an event and an occurrence and call those `measurementTypes` as `length` and `weight`.

```

mof_types <- read_csv("https://raw.githubusercontent.com/HakaiInstitute/jsp-data/master/

fish_data$weight <- coalesce(fish_data$weight, fish_data$weight_field)
fish_data$fork_length <- coalesce(fish_data$fork_length, fish_data$fork_length_field)
fish_data$`Life stage` <- "juvenile"

measurementOrFact <- fish_data %>%
  mutate(occurrenceID = paste0("hakai-jsp-", ufn)) %>%
  select(occurrenceID, eventID = seine_id, "Length (fork length)" = fork_length,
        "Standard length" = standard_length, "Weight" = weight, `Life stage`) %>%
  pivot_longer(`Length (fork length)`:`Life stage`,
               names_to = "measurementType",
               values_to = "measurementValue",
               values_transform = list(measurementValue = as.character)) %>%
  filter(measurementValue != "NA") %>%
  left_join(mof_types, by = c("measurementType")) %>%
  mutate(measurementValueID = case_when(measurementValue == "juvenile" ~ "http://vocab
        measurementID = paste(eventID, measurementType, occurrenceID, sep = "-"))

write_csv(measurementOrFact, "../datasets/hakai_salmon_data/raw_data/extendedMeasurement

#Check that every eventID in Occurrence occurs in event table
no_keys <- dm(event, occurrence, measurementOrFact)
only_pk <- no_keys %>%
  dm_add_pk(event, eventID) %>%
  dm_add_pk(occurrence, occurrenceID) %>%
  dm_add_pk(measurementOrFact, measurementID)
dm_examine_constraints(only_pk)

model <- only_pk %>%
  dm_add_fk(occurrence, eventID, event) %>%
  dm_add_fk(measurementOrFact, occurrenceID, occurrence)
dm_examine_constraints(model)

#TODO: Fix bookdown issues so that dm_draw shows data model html output. Perhaps add
# dm_draw(model, view_type = "all")

```

2.2 Example Two

Add another example here, perhaps zooplankton?

Chapter 3

Final Words

We have finished a nice book.

References

- Allaire, JJ, Yihui Xie, Jonathan McPherson, Javier Luraschi, Kevin Ushey, Aron Atkins, Hadley Wickham, Joe Cheng, Winston Chang, and Richard Iannone. 2021. *Rmarkdown: Dynamic Documents for r*. <https://CRAN.R-project.org/package=rmarkdown>.
- Barbier, Edward B. 2017. “Marine Ecosystem Services.” *Current Biology*. Cell Press. <https://doi.org/10.1016/j.cub.2017.03.020>.
- Benson, Abigail, Cassandra M. Brooks, Gabrielle Canonico, Emmett Duffy, Frank Muller-Karger, Heidi M. Sosik, Patricia Miloslavich, and Eduardo Klein. 2018. “Integrated Observations and Informatics Improve Understanding of Changing Marine Ecosystems.” *Frontiers in Marine Science*. Frontiers Media S.A. <https://doi.org/10.3389/fmars.2018.00428>.
- Benson, Abigail, Tylar Murray, Gabrielle Canonico, Enrique Montes, Frank Muller-Karger, Maria T. Kavanaugh, Joaquin Trinanés, and Lynn M. deWitt. 2021. “Data Management and Interactive Visualizations for the Evolving Marine Biodiversity Observation Network.” *Oceanography*. <https://doi.org/10.5670/oceanog.2021.220>.
- Borer, Elizabeth T., Eric W. Seabloom, Matthew B. Jones, and Mark Schilbauer. 2009. “Some Simple Guidelines for Effective Data Management.” *Bulletin of the Ecological Society of America* 90 (April). <https://doi.org/10.1890/0012-9623-90.2.205>.
- Canonico, Gabrielle, Pier Luigi Buttigieg, Enrique Montes, Frank E. Muller-Karger, Carol Stepien, Dawn Wright, Abigail Benson, et al. 2019. “Global Observational Needs and Resources for Marine Biodiversity.” *Frontiers in Marine Science*. Frontiers Media S.A. <https://doi.org/10.3389/fmars.2019.00367>.
- Crystal-Ornelas, Robert, Charuleka Varadharajan, Ben Bond-Lamberty, Kristin Boye, Madison Burrus, Shreyas Cholia, Michael Crow, et al. 2021. “A Guide to Using GitHub for Developing and Versioning Data Standards and Reporting Formats.” *Earth and Space Science*, July, e2021EA001797. <https://doi.org/10.1029/2021EA001797>.
- Davies, Neil, John Deck, Eric C Kansa, Sarah Whitcher Kansa, John Kunze, Christopher Meyer, Thomas Orrell, et al. 2021. “Internet of Samples (iSamples): Toward an Interdisciplinary Cyberinfrastructure for Material Samples.” *GigaScience* 10: 1–5. <https://doi.org/10.1093/gigascience/giab028>.

- Djurhuus, Anni, Collin J. Closek, Ryan P. Kelly, Kathleen J. Pitz, Reiko P. Michisaki, Hilary A. Starks, Kristine R. Walz, et al. 2020. “Environmental DNA Reveals Seasonal Shifts and Potential Interactions in a Marine Community.” *Nature Communications* 11 (December): 1–9. <https://doi.org/10.1038/s41467-019-14105-1>.
- Duffy, J. Emmett, Linda A. Amaral-Zettler, Daphne G. Fautin, Gustav Paulay, Tatiana A. Rynearson, Heidi M. Sosik, and John J. Stachowicz. 2013. “Envisioning a Marine Biodiversity Observation Network.” *BioScience* 63 (May): 350–61. <https://doi.org/10.1525/bio.2013.63.5.8>.
- Fornwall, M, R Gisiner, S E Simmons, H Moustahfid, G Canonico, P Halpin, P Goldstein, et al. 2012. “Expanding Biological Data Standards Development Processes for US IOOS: Visual Line Transect Observing Community for Mammal, Bird, and Turtle Data.” IOOS. <https://www.researchgate.net/publication/255681522>.
- Hardisty, Alex R., William K. Michener, Donat Agosti, Enrique Alonso García, Lucy Bastin, Lee Belbin, Anne Bowser, et al. 2019. “The Bari Manifesto: An Interoperability Framework for Essential Biodiversity Variables.” *Ecological Informatics* 49 (January): 22–31. <https://doi.org/10.1016/j.ecoinf.2018.11.003>.
- Heberling, J Mason, Joseph T Miller, Daniel Noesgaard, Scott B Weingart C, Dmitry Schigel, and Douglas E Soltis. 2021. “Data Integration Enables Global Biodiversity Synthesis.” *Proceedings of the National Academy of Sciences of the United States of America*. <https://doi.org/10.1073/pnas.2018093118/-/DCSupplemental>.
- Jonathan A. Hare, Mark W. Nelson, Wendy E. Morrison. n.d. “A Vulnerability Assessment of Fish and Invertebrates to Climate Change on the Northeast u.s. Continental Shelf.” *PLoS ONE* 11 (2): e0146756. <https://doi.org/10.1371/journal.pone.0146756>.
- Jones, Matthew B., Mark P. Schildhauer, O. J. Reichman, and Shawn Bowers. 2006. “The New Bioinformatics: Integrating Ecological Data from the Gene to the Biosphere.” *Annual Review of Ecology, Evolution, and Systematics*. <https://doi.org/10.1146/annurev.ecolsys.37.091305.110031>.
- Kavanaugh, Maria T., Matthew J. Oliver, Francisco P. Chavez, Ricardo M. Letelier, Frank E. Muller-Karger, and Scott C. Doney. 2016. “Seascapes as a New Vernacular for Pelagic Ocean Monitoring, Management and Conservation.” *ICES Journal of Marine Science* 73 (July): 1839–50. <https://doi.org/10.1093/icesjms/fsw086>.
- Kot, Connie Y., Ei Fujioka, Lucie J. Hazen, Benjamin D. Best, Andrew J. Read, and Patrick N. Halpin. 2010. “Spatio-Temporal Gap Analysis of OBIS-SEAMAP Project Data: Assessment and Way Forward.” *PLoS ONE* 5 (September): 12990. <https://doi.org/10.1371/journal.pone.0012990>.
- Lab, Malin Pinsky. n.d. “OceanAdapt.” *GitHub*. <https://oceanadapt.rutgers.edu/>.
- Lamprecht, Anna-Lena, Leyla Garcia, Mateusz Kuzak, Carlos Martinez, Ricardo Arcila, Eva Martin Del Pico, Victoria Dominguez Del Angel, et al. 2019. “Towards FAIR Principles for Research Software.” Edited by Paul

- Groth. *Data Science*, 1–23. <https://doi.org/10.3233/DS-190026>.
- Maria Dornelas, Brian McGill, Nicholas J. Gotelli. 2014. “Assemblage Time Series Reveal Biodiversity Change but Not Systematic Loss.” *Science* 344 (6181): 296–99. <https://doi.org/10.1126/science.1248484>.
- McKenna, Megan F., Simone Baumann-Pickering, Annebelle C. M. Kok, William K. Oestreich, Jeffrey D. Adams, Jack Barkowski, Kurt M. Fristrup, et al. 2021. “Advancing the Interpretation of Shallow Water Marine Soundscapes.” *Frontiers in Marine Science* 0 (September): 1426. <https://doi.org/10.3389/FMARS.2021.719258>.
- Miloslavich, Patricia, Nicholas J. Bax, Samantha E. Simmons, Eduardo Klein, Ward Appeltans, Octavio Aburto-Oropeza, Melissa Andersen Garcia, et al. 2018. “Essential Ocean Variables for Global Sustained Observations of Biodiversity and Ecosystem Changes.” *Global Change Biology* 24 (June): 2416–33. <https://doi.org/10.1111/gcb.14108>.
- Montes, Enrique, Anni Djurhuus, Frank E. Muller-Karger, Daniel Otis, Christopher R. Kelble, and Maria T. Kavanaugh. 2020. “Dynamic Satellite Seascapes as a Biogeographic Framework for Understanding Phytoplankton Assemblages in the Florida Keys National Marine Sanctuary, United States.” *Frontiers in Marine Science* 7 (July): 575. <https://doi.org/10.3389/fmars.2020.00575>.
- Moustahfid, Hassan, and Philip Goldstein. 2014. “IOOS Biological Data Services Enrollment Procedures.”
- Moustahfid, Hassan, Jim Potemra, Philip Goldstein, Roy Mendelssohn, and Annette Desrochers. 2011. “Making United States Integrated Ocean Observing System (u.s. IOOS) Inclusive of Marine Biological Resources.” <https://www.researchgate.net/publication/254013004>.
- Muller-Karger, Frank E., Erin Hestir, Christiana Ade, Kevin Turpie, Dar A. Roberts, David Siegel, Robert J. Miller, et al. 2018. “Satellite Sensor Requirements for Monitoring Essential Biodiversity Variables of Coastal Ecosystems.” *Ecological Applications* 28 (April): 749–60. <https://doi.org/10.1002/eap.1682>.
- Muller-Karger, Frank E., Patricia Miloslavich, Nicholas J. Bax, Samantha Simmons, Mark J. Costello, Isabel Sousa Pinto, Gabrielle Canonico, et al. 2018. “Advancing Marine Biological Observations and Data Requirements of the Complementary Essential Ocean Variables (EOVs) and Essential Biodiversity Variables (EBVs) Frameworks.” *Frontiers in Marine Science*. Frontiers Media S.A. <https://doi.org/10.3389/fmars.2018.00211>.
- Neeley, Aimee, Stace E. Beaulieu, Chris Proctor, Ivona Cetinić, Joe Futrelle, Inia Soto Ramos, Heidi M. Sosik, et al. 2021. “Standards and Practices for Reporting Plankton and Other Particle Observations from Images.” <https://doi.org/10.1575/1912/27377>.
- O’Brien, Margaret, Colin A. Smith, Eric R. Sokol, Corinna Gries, Nina Lany, Sydne Record, and Max C. N. Castorani. 2021. “ecocomDP: A Flexible Data Design Pattern for Ecological Community Survey Data.” *Ecological Informatics* 64 (September): 101374. <https://doi.org/10.1016/J.ECOINF.2021.101374>.

- Pooter, Daphnis De, Ward Appeltans, Nicolas Bailly, Sky Bristol, Klaas Deneudt, Menashè Eliezer, Ei Fujioka, et al. 2017. “Toward a New Data Standard for Combined Marine Biological and Environmental Datasets - Expanding OBIS Beyond Species Occurrences.” *Biodiversity Data Journal* 5 (January): 10989. <https://doi.org/10.3897/BDJ.5.e10989>.
- Provoost, Pieter. n.d. “Iobis/Ebsa.” *GitHub*. <https://github.com/iobis/ebsa>.
- R Core Team. 2021. *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foundation for Statistical Computing. <https://www.R-project.org/>.
- Rücknagel, J., P. Vierkant, R. Ulrich, G. Kloska, E. Schnepf, D. Fichtmüller, E. Reuter, et al. 2015. “Metadata Schema for the Description of Research Data Repositories. Version 3.0.” <https://doi.org/10.2312/re3.008>.
- Rule, Adam, Amanda Birmingham, Cristal Zuniga, Ilkay Altintas, Shih-Cheng Huang, Rob Knight, Niema Moshiri, et al. 2019. “Ten Simple Rules for Writing and Sharing Computational Analyses in Jupyter Notebooks.” *PLOS Computational Biology* 15 (July): e1007007. <https://doi.org/10.1371/JOURNAL.PCBI.1007007>.
- Santora, Jarrod A., Elliott L. Hazen, Isaac D. Schroeder, Steven J. Bograd, Keith M. Sakuma, and John C. Field. 2017. “Impacts of Ocean Climate Variability on Biodiversity of Pelagic Forage Species in an Upwelling Ecosystem.” *Marine Ecology Progress Series* 580 (September): 205–20. <https://doi.org/10.3354/meps12278>.
- Schmid, Moritz S, Dominic Daprano, Kyler M Jacobson, Christopher Sullivan, Christian Briseño-Avena, Jessica Y Luo, and Robert K Cowen. 2021. *A Convolutional Neural Network Based High-Throughput Image Classification Pipeline - Code and Documentation to Process Plankton Underwater Imagery Using Local HPC Infrastructure and NSF’s XSEDE*. Zenodo. <https://doi.org/10.5281/ZENODO.4641158>.
- Taylor, Gordon T., Frank E. Muller-Karger, Robert C. Thunell, Mary I. Scranton, Yrene Astor, Ramon Varela, Luis Troccoli Ghinaglia, et al. 2012. “Ecosystem Responses in the Southern Caribbean Sea to Global Climate Change.” *Proceedings of the National Academy of Sciences of the United States of America* 109 (November): 19315–20. <https://doi.org/10.1073/pnas.1207514109>.
- Tittensor, Derek P, Camilo Mora, Walter Jetz, Heike K Lotze, Daniel Ricard, Edward Vanden Berghe, and Boris Worm. 2010. “Global Patterns and Predictors of Marine Biodiversity Across Taxa.” *Nature* 466 (7310): 1098.
- Warren, R., J. Price, E. Graham, N. Forstenhaeusler, and J. VanDerWal. 2018. “The Projected Effect on Insects, Vertebrates, and Plants of Limiting Global Warming to 1.5°C Rather Than 2°C.” *Science* 360 (May): 791–95. <https://doi.org/10.1126/science.aar3646>.
- Wieczorek, John, David Bloom, Robert Guralnick, Stan Blum, Markus Döring, Renato Giovanni, Tim Robertson, and David Vieglais. 2012. “Darwin Core: An Evolving Community-Developed Biodiversity Data Standard.” *PLoS ONE* 7 (January): 29715. <https://doi.org/10.1371/journal.pone.0029715>.
- Wilkinson, Mark D., Michel Dumontier, IJsbrand Jan Aalbersberg, Gabrielle

- Appleton, Myles Axton, Arie Baak, Niklas Blomberg, et al. 2016. “The FAIR Guiding Principles for Scientific Data Management and Stewardship.” *Scientific Data* 3 (March): 1–9. <https://doi.org/10.1038/sdata.2016.18>.
- Xie, Yihui. 2014. “Knitr: A Comprehensive Tool for Reproducible Research in R.” In *Implementing Reproducible Computational Research*, edited by Victoria Stodden, Friedrich Leisch, and Roger D. Peng. Chapman; Hall/CRC. <http://www.crcpress.com/product/isbn/9781466561595>.
- . 2015b. *Dynamic Documents with R and Knitr*. 2nd ed. Boca Raton, Florida: Chapman; Hall/CRC. <http://yihui.name/knitr/>.
- . 2015a. *Dynamic Documents with R and Knitr*. 2nd ed. Boca Raton, Florida: Chapman; Hall/CRC. <https://yihui.org/knitr/>.
- . 2016. *Bookdown: Authoring Books and Technical Documents with R Markdown*. Boca Raton, Florida: Chapman; Hall/CRC. <https://bookdown.org/yihui/bookdown>.
- . 2021a. *Bookdown: Authoring Books and Technical Documents with r Markdown*. <https://CRAN.R-project.org/package=bookdown>.
- . 2021b. *Knitr: A General-Purpose Package for Dynamic Report Generation in r*. <https://yihui.org/knitr/>.
- Xie, Yihui, J. J. Allaire, and Garrett Grolemund. 2018. *R Markdown: The Definitive Guide*. Boca Raton, Florida: Chapman; Hall/CRC. <https://bookdown.org/yihui/rmarkdown>.
- Xie, Yihui, Christophe Dervieux, and Emily Riederer. 2020. *R Markdown Cookbook*. Boca Raton, Florida: Chapman; Hall/CRC. <https://bookdown.org/yihui/rmarkdown-cookbook>.