wsl.biodiv

November 9, 2022

```
Type Package
Title A toolbox to efficiently handle Species Distribution Modelling (SDMs) in collaborative projects
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Depends R (>= 3.1.0), snow, mgcv, dismo, MASS, gbm, randomForest, ROCR, cluster, NMOF, spat-
      stat, raster, foreach, doParallel, usdm, glmnet, neuralnet, rgbif, Coordinate-
      Cleaner, rgdal, lme4, MuMIn
Description Package aiming at varied and flexible model fitting and tools for SDMs.
      Meta information should be stored (author, date, taxon,...) efficiently.
      Model evaluation and prediction should flexibly handle various fitted model objects.
      Evaluation metrics for pres-only and pres-abs models should be mutually implemented.
      Relationships between saved objects should be preserved.
      Filtering and testing tools should help the user to find adequate predictors.
      Poisson Point Process models (PPPM) implementation should remain user-friendly.
      Regularization and variable selection should be implemented in this framework.
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 $Boyce\ index\ model\ evaluation\ sub-function\ II$

Description

boycei

Not to be called directly by the user

Usage

```
boycei(interval, obs, fit)
```

Author(s)

'boycei' from the 'ecospat' package

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Do the actual model evaluations

Description

Not to be called directly by the user

Usage

```
ceval(f, pa, tesdat, crit, tre = numeric())
```

Author(s)

Philipp Brun

copy_PseuAbs

Copy pseudoabsences from wsl.pseudoabsences object for new species

Description

Copy pseudoabsences from existing wsl.pseudoabsences object to wsl.pseudoabsences object for new species

Usage

```
copy_PseuAbs(env.stack, pres = SpatialPoints(), taxon = character(), x)
```

Arguments

env.stack	the same layers as in the wsl.pseudoabsences object.
pres	SpatialPoints object. Location of presence points. Necessary for 'geographic' sampling strategy and the best adding point if the downstream functions will be used.
taxon	Character; name of taxon of interest to keep track of meta information.

x A wsl.pseudoabsences object

Details

if the desired pseudoabsence sampling strategy is not species-specific it may be more efficient to copy on the sampled pseudoabsences from another object.

Value

an object of class 'wsl.pseudoabsences' that can be plotted and passed on to wsl.flex

Author(s)

Philipp

copy_PseuAbs 5

Examples

```
### Data preparation
# Predictors
bio=raster::getData('worldclim', var='bio', lon=16, lat=48, res=.5)
bio=bio[[c(1,4,12)]]
# install.packages("rgbif")
library(rgbif)
# extract species
spn1='Boletus aestivalis'
xt=as.vector(extent(bio))
baest <- occ_search(scientificName=spn1,</pre>
              hasCoordinate=TRUE,
              decimalLongitude=paste0(xt[1],",",xt[3]),
              decimalLatitude=paste0(xt[2],",",xt[4]))
pbaest=baest$data[,c('decimalLongitude','decimalLatitude')]
baest_spp=SpatialPoints(pbaest,proj4string = crs(bio))
spn2='Boletus edulis'
bedu <- occ_search(scientificName=spn2,</pre>
              hasCoordinate=TRUE,
              decimalLongitude=paste0(xt[1],",",xt[3]),
              decimalLatitude=paste0(xt[2],",",xt[4]))
pbedu=bedu$data[,c('decimalLongitude','decimalLatitude')]
bedu_spp=SpatialPoints(pbedu,proj4string = crs(bio))
### Sample pseudoabsences
# Geo.start
pseu.abs1=wsl.samplePseuAbs(type="geo.strat",
                     n=5000,
                     env.stack=bio,
                     pres=baest_spp,
                     geores_fact=3,
                     add.strat=0,
                     taxon=spn1)
plot(pseu.abs1)
pseu.abs2=copy_PseuAbs(env.stack=bio,
                 pres=bedu_spp,
                 taxon=spn2,
                 x=pseu.abs1)
plot(pseu.abs2)
```

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create_envstrat	Create environmental strata for environmentally stratified pseudoabsence sampling.

Description

Create environmental strata for environmentally stratified pseudoabsence sampling.

Usage

```
create_envstrat(
  env.stk,
  rAll = TRUE,
  save_it = TRUE,
  strat_dir = NA,
  poolsiz = 5 * 10^6,
  sampsiz = 4e+05,
  type
)
```

Arguments

env.stk	raster stack with environmental layers of interest			
rAll	should raster be read entirely into memory? This is faster but may blow the RAMs of your machine.			
save_it	Should a stratified sample of c. 400000 points be saved			
strat_dir	Directory to save the stratified sample in, Not to be called directly by the user			

Author(s)

Philipp

df_or_	rast
--------	------

Predict to data.frame or raster

Description

Not to be called directly by the user

Usage

```
df_or_rast(mod, nwdat, clust, ...)
```

Author(s)

Philipp Brun

eco.adj.D2.glm

Function originally written by A. Guisan (UNIL-ECOSPAT,Switzerland) and modified by C. Randin (UNIL-ECOSPAT,Switzerland) S-Plus Function for calculating an adjusted D2 (see Weisberg, 1980) Takes any GLM object as argument

Description

Not to be called directly by the user

Usage

```
eco.adj.D2.glm(glm.obj)
```

Author(s)

'ecospat.adj.D2.glm' from the 'ecospat' package

ecoBoyce

Boyce index model evaluation sub-function I

Description

Not to be called directly by the user

Usage

```
ecoBoyce(fit, obs, nclass = 0, window.w = "default", res = 100, PEplot = TRUE)
```

Author(s)

'ecospat.boyce' from the 'ecospat' package

fitdoc

Fit presence-absence models

Description

Flexibly fit various types of functions but let framework take care of resampling, meta-info storage, and file saving. wsl.flex basically allows supplying any possible model, however, there may be problems with prediction/evaluation for exotic functions. Available model algorithms for fitting are: Generalized Linear Models (GLM), Generalized additive models (GAM), Maximum Entropy (MaxEnt), Artificial Neural Networks (ANN), Generalized Boosted regression Models/Boosted Regression Trees (GBM) and Random Forest (RF)

Usage

```
wsl.glm(
  x = numeric(),
  pa = numeric(),
  env_vars = data.frame(),
  taxon = character(),
  replicatetype = character(),
  reps,
  strata = NA,
  save = FALSE,
  project = NA,
  path = NA,
  step = FALSE,
  mod_tag = "",
  xy = NULL,
)
wsl.gam(
  x = numeric(),
  pa = numeric(),
  env_vars = data.frame(),
  taxon = character(),
  replicatetype = character(),
  reps,
  strata = NA,
  save = FALSE,
  project = NA,
  path = NA,
  step = FALSE,
  mod_tag = "",
  xy = NULL,
  . . .
wsl.maxent(
  x = numeric(),
  pa = numeric(),
  env_vars = data.frame(),
  taxon = character(),
  replicatetype = character(),
  reps,
  strata = NA,
  save = FALSE,
  project = NA,
  path = NA,
  mod_tag = "",
  xy = NULL,
wsl.gbm(
```

```
x = numeric(),
  pa = numeric(),
  env_vars = data.frame(),
  taxon = character(),
  replicatetype = character(),
  reps,
  strata = NA,
  save = FALSE,
  project = NA,
  path = NA,
  mod_tag = "",
  xy = NULL,
)
wsl.ann(
  x = numeric(),
  pa = numeric(),
  env_vars = data.frame(),
  taxon = character(),
  replicatetype = character(),
  reps,
  strata = NA,
  save = FALSE,
  project = NA,
  path = NA,
  mod_tag = "",
  xy = NULL,
)
wsl.flex(
  x = numeric(),
  pa = numeric(),
  env_vars = data.frame(),
  taxon = character(),
  replicatetype = character(),
  reps,
  strata = NA,
  save = FALSE,
  project = NA,
  path = NA,
  mod_args = list(),
  xy = NULL
)
```

Arguments

x Optional. Object of class 'wsl.pseudoabsences'. If used 'pa', 'env_vars' and 'taxon' will be replaced by new set of values set in the object.

pa Object of class 'vector' with presence/absence values

env_vars Object of class 'data.frame' with environmental predictors

taxon Name of the taxon for which models are fitted (How) should replicates be generated? May be 'none', 'splitsample', 'cv' or replicatetype 'block-cv' Number of replicates reps A numeric vector of the same length as observations with integers separating strata cross validation replicates. Only used when replicatetype='block-cv' save Should the model be saved in a structured way? (not implemented yet) Character indicating the name of the project within which the models are run project (later used to define saving directories) path Where to save? (not implemented yet) For glms and gams only. Should the models be updated with the step function? step Not in wsl.flex. Descriptive label for current model mod_tag Optional. XY coordinates of observations. Used for post evaluation with presenceху only metric when pres_abs=TRUE in wsl.evaluate.pres(). Must be a data.frame()

Value

mod_args

Object of class wsl.fit including slots for meta info, testing data for evaluation, and model objects

List with elements of class 'multi.input' which specify models to be fitted in

Author(s)

Philipp Brun, Yohann Chauvier

or matrix().

wsl.flex

Examples

```
# Take anguilla data set from dismo package
data("Anguilla_train")
vrs=c("SegSumT","USRainDays","USSlope")
env=Anguilla_train[,vrs]
### Check out wsl.glm
form.glm=as.formula(paste("Presence",paste(paste("poly(",vrs,",2)"),collapse="+")))\\
modi1=wsl.glm(pa=Anguilla_train$Angaus,
              env_vars = env,
              taxon="Angaus",
              replicatetype="cv",
              reps=5,
              project="prototest",
              mod_tag="test-glm",
              formula=form.glm,
              family="binomial",
              step=TRUE)
# Try out custom summary function
summary(modi1)
# Access glm object of first replicate
```

```
summary(modi1@fits$replicate_01$`test-glm`)
# Evaluate the model
eval1=wsl.evaluate.pa(modi1)
# Get evaluation summary
summary(eval1)
### Check out wsl.gam
form.gam=as.formula(paste("Presence~",paste(paste0("s(",vrs,")"),collapse="+")))
# Try out wsl.glm funcion
modi2=wsl.gam(pa=Anguilla_train$Angaus,
              env_vars = env,
              taxon="Angaus",
              replicatetype="splitsample",
              reps=3,
              project="prototest",
              mod_tag="test-gam",
              formula=form.gam,
              family="binomial",
              step=FALSE)
# Try out custom summary function
summary(modi2)
# Access glm object of first replicate
summary(modi2@fits$replicate_01$`test-gam`)
# Evaluate the model
eval2=wsl.evaluate.pa(modi2,crit="maxTSS")
# Get evaluation summary
summary(eval2)
### Check out wsl.gbm
form.gbm=as.formula(Presence ~ .)
# Try out wsl.glm funcion
modi3=wsl.gbm(pa=Anguilla_train$Angaus,
              env_vars = env,
              taxon="Angaus",
              replicatetype="none",
              reps=1,
              project="prototest",
              mod_tag="test-brt",
              formula= form.gbm,
              distribution = "bernoulli",
              interaction.depth = 1,
              shrinkage=.01,
              n.trees = 3500)
# Try out custom summary function
summary(modi3)
# Access glm object of first replicate
summary(modi3@fits$replicate_01$`test-brt`)
```

```
# Prepare external testing data
tste=data.frame(Presence=Anguilla_train$Angaus,env)
# Evaluate the model
eval3=wsl.evaluate.pa(modi3,crit="maxTSS",tester=tste)
# Get evaluation summary
summary(eval3)
### Check out wsl.maxent
feat=c("linear=true","quadratic=true","hinge=true","product=true","threshold=false")
# Try out wsl.glm funcion
modi4=wsl.maxent(pa=Anguilla_train$Angaus,
                env_vars = env,
                taxon="Angaus",
                replicatetype="block-cv",
                reps=3.
                strata=sample(1:3,nrow(env),replace=TRUE),
                project="prototest",
                mod_tag="test-mxe",
                args=feat)
# Try out custom summary function
summary(modi4)
# Access glm object of first replicate
summary(modi4@fits$replicate_01$`test-mxe`)
# Define external threshold
thmxe=c(`test-mxe`=0.5)
# Evaluate the model
eval4=wsl.evaluate.pa(modi4,crit="external",thres=thmxe)
# Get evaluation summary
summary(eval4)
# Get thresholds
thr.4=get_thres(eval4)
### Check out wsl.flex
form.glm.2=as.formula(paste("Presence~",paste(vrs,collapse="+")))
multi("gbm",list(formula=form.gbm,
                distribution = "bernoulli",
                interaction.depth = 1,
                shrinkage=.01,
                n.trees = 3500), "gbm-simple"),
multi("gam",list(formula=form.gam,family="binomial"),"gam-simple",step=FALSE,weight=TRUE)),
multi("maxent", list(args=feat), "mxe-simple"),
multi("randomForest",list(formula=form.gbm,ntree=500,maxnodes=NULL),"waud1"),
multi("glm",list(formula=form.glm.2,family="binomial"),"glm-lin",step=TRUE,weight=TRUE)))
# Try out wsl.glm funcion
```

```
modi5=wsl.flex(pa=Anguilla_train$Angaus,
               env_vars = env,
               taxon="Angaus"
               replicatetype="block-cv",
               reps=3,
               strata=sample(1:3,nrow(env),replace=TRUE),
               project="multitest",
               mod_args=modinp)
# Try out custom summary function
summary(modi5)
# Access glm object of first replicate
summary(modi5@fits$replicate_01$`glm-simple`)
# Evaluate the model
eval5<-wsl.evaluate.pa(modi5,crit="pp=op")</pre>
# Get evaluation summary
summary(eval5)
# Get thresholds
thr.5=get_thres(eval5)
```

fitppm

Fit Poisson Point Process Models (PPPM)

Description

PPPM (Poisson Point Process Models) is a modelling approach genrally design to use observation-only data aka point occurences. It allows the user to model species observations intensity per unit area (i.e. the density of presence points over a spatial grid) as a log linear function of predictors. Quadrature points (or background points) are necessary in the model, and may be generated with the 'wsl.quadrature' function. Those points apply a spatial scaling proportional to the study area and estimate the maximised model log likelihood (see Renner 2013, Renner et al. 2015). Two version of the function are available. 'wsl.ppmO' applies a simple point process model using the 'ppm' function from the package "spatstat" i.e. without any further validation possible (only for calibration and prediction use). 'wsl.ppmGlasso' applies a similar point process model but by implementing choices of regularisation and variable selection following the package "glmnet".

Usage

```
wsl.ppmGlasso(
  pres = data.frame(),
  env_vars = matrix(),
  quadPoints = wsl.quads(),
  asurface = numeric(),
  taxon = character(),
  replicatetype = character(),
  reps,
  strata = NA,
  save = FALSE,
```

```
project = NA,
  path = NA,
  mod\_tag = ""
  poly = TRUE,
  which_poly = NULL,
  lasso = TRUE,
  penalty.glmnet = NULL,
)
wsl.ppmO(
  pres = data.frame(),
  quadPoints = ppp(),
  env_vars = list(),
  window = owin(),
  taxon = character(),
  replicatetype = character(),
  reps,
  strata = NA,
  save = FALSE,
  project = NA,
  path = NA,
  formula = NULL,
)
```

Arguments

pres Object of class 'data.frame' or 'matrix'. Coordinates xy of Species observations.

env_vars If 'wsl.ppmGlasso' used, must be an object of class 'matrix' or 'data.frame' with

environmental predictor values. If 'wsl.ppmO' used, must be an object of class 'list' generated with the wsl.ppm.env' function. Note that categorical predictor values must be of class factor for both 'env_vars' and 'quadPoints@Qenv'

quadPoints If 'wsl.ppmGlasso' used, must a 'wsl.quads' object generated with 'wsl.quadrature'

function and 'lasso=TRUE'. If 'wsl.ppmO' used, must be a 'ppp' object generated with the 'wsl.quadrature' function with 'lasso=FALSE'; here, may be one same object for each 'reps' or a list of objects whose length equal to 'reps'

asurface Only when 'wsl.ppmGlasso' used. The surface of the study area in square kilo-

meters

taxon Name of the taxon for which models are fitted

replicatetype (How) should replicates be generated? may be 'none', 'splitsample', 'cv' or

'block-cv'

reps Number of replicates

strata A numeric vector of the same length as observations + quadrature points with in-

tegers assigning cross validation replicates. Only used when replicatetype='block-cv'. Note: the vector must first integrate CV information for observation points.

save Should the model be saved in a structured way? (not implemented yet)

project Character indicating the name of the project within which the models are run

(later used to define saving directories)

path Where to save? (not implemented yet)

mod_tag Descriptive label for current model

poly If TRUE, PPPM fits a second order polynomial regression

which_poly Which predictors should be using polynomial terms? Use a binary vector that

specify which variables/predictors. Length of vector must be equal to the number of input variables. '1' stands for poly=TRUE whereas '0' stands for poly=FALSE.

Default is polynomial for all if poly=TRUE.

lasso If FALSE no regularisation is applied

penalty.glmnet If 'lasso=TRUE', a binary vector that specify which variables/predictors used

to model should be shrinked. Length of vector must be equal to the number of input variables. '1' stands for shrinkage whereas '0' stands for no shrinkage, i.e.

the variable will always be included in the model.

... If 'wsl.ppmGlasso' used with lasso = TRUE, arguments passed on to the cv.glmnet()

function (package 'glmnet') use to apply a Lasso, Ridge or Elastic Net regularisation. To notice that the package's argument 'penalty.factor' is not needed here. If used, the parameter 'penalty.glmnet' must instead be filled for each 'env_vars'. If lasso = FALSE, arguments passed on to the glm("poisson") function. If 'wsl.ppmO' used, arguments passed on to the ppm() function (package

'spatstat')

window Only when 'wsl.ppmO' used. An object of class 'owin' generated with the

'wsl.ppm.window' function

formula Equation of the desired fit

Value

Object of class wsl.fit including slots for meta info, testing data for evaluation, and model objects

Author(s)

Yohann Chauvier, Philipp Brun

Examples

```
# Grid regular
quadG1 = wsl.quadrature(mask = maskR,
                        area.win = wind,
                        random = FALSE,
                        lasso = TRUE,
                        env_vars = rst)
### Define quadrature points for 'wsl.ppm0'
   # Grid regular
quadO1 = wsl.quadrature(mask = maskR,
                        area.win = wind,
                        random = FALSE,
                        lasso = FALSE,
                        env_vars = NULL)
### Define your environments
   # For 'wsl.ppmGlasso' (observations focus)
envG = raster::extract(rst,mypoints)
   # For 'wsl.ppm0' (study area focus)
env0 = wsl.ppm.env(rst,maskR)
### Modelling
   # 'wsl.ppmGlasso' (alpha = 0.5 => Elastic net, see package 'glmnet')
       # Complex PPPM lasso (poly = TRUE & lasso=TRUE)
lasso1 = wsl.ppmGlasso(pres = mypoints,
                       quadPoints = quadG1$Edata,
                       asurface = raster::area(shp.lonlat)/1000,
                       env_vars = envG,
                       taxon = "species_eg1",
                       replicatetype = "cv",
                       reps = 5,
                       strata = NA,
                       save=FALSE,
                       project = "lasso_eg1",
                       path = NA,
                       poly = TRUE,
                       lasso = TRUE,
                       alpha = 0.5,
                       type.measure = "mse",
                       standardize = TRUE,
                       nfolds = 5,
                       nlambda = 100)
       # Simple PPPM non lasso (poly = FALSE & lasso=FALSE)
lasso2 = wsl.ppmGlasso(pres = mypoints,
                       quadPoints = quadG1$Edata,
                       asurface = raster::area(shp.lonlat)/1000,
                       env_vars = envG,
                       taxon = "species_eg2",
                       replicatetype = "cv",
                       reps = 5,
```

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```
strata = NA,
                       save = FALSE,
                       project = "lasso_eg2",
                       path = NA,
                       poly = FALSE,
                       lasso = FALSE)
   # 'wsl.ppm0'
       # Simple PPPM non lasso (same as above with poly = FALSE & lasso = FALSE)
form.Sppm = as.formula(paste("~",paste(names(env0),collapse="+")))
lasso3 = wsl.ppmO(pres = mypoints,
                  quadPoints = quadO1,
                  env_vars = env0,
                  window = wind,
                  taxon = "species_eg3",
                  replicatetype = "cv",
                  reps = 5,
                  strata = NA,
                  save = FALSE,
                  project = "lasso_eg3",
                  path = NA,
                  formula = form.Sppm)
       # Complex PPPM non lasso
form.Cppm = as.formula(paste("~",paste(paste0("poly(",names(env0),",2)"),collapse="+")))
lasso4 = wsl.ppmO(pres = mypoints,
                  quadPoints = quadO1,
                  env_vars = env0,
                  window = wind,
                  taxon = "species_eg3",
                  replicatetype = "cv",
                  reps = 5,
                  strata = NA,
                  save = FALSE,
                  project = "lasso_eg3",
                  path = NA,
                  formula = form.Cppm)
```

get_thres

Get threshold

Description

Extracts thresholds from wsl.evaluation objects and names them so they can be fed to the wsl.predict function. At the moment only averages over replicates can be obtained.

Usage

```
get_thres(x, mean)
```

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Arguments

x An object of class wsl.evaluation

mean Logical. If TRUE, a mean is applied to all thresholds

Value

```
numeric() or 'vector' with thresholds
```

Author(s)

Philipp Brun, Yohann Chauvier

Examples

```
# Take anguilla data set from dismo package
data("Anguilla_train")
vrs=c("SegSumT","USRainDays","USSlope")
env=Anguilla_train[,vrs]
### Check out wsl.glm
form.glm=as.formula(paste("Presence",paste(paste("poly(",vrs,",2)"),collapse="+")))\\
modi1=wsl.glm(pa=Anguilla_train$Angaus,
              env_vars = env,
              taxon="Angaus",
              replicatetype="cv",
              reps=5,
              project="prototest",
              mod_tag="test-glm",
              formula=form.glm,
              family="binomial",
              step=TRUE)
# Evaluate the model
eval1=wsl.evaluate.pa(modi1)
# Get thresholds
get_thres(eval1, mean = FALSE)
get_thres(eval1, mean = TRUE)
```

hde

Hide annoying prints

Description

Not to be called directly by the user

Usage

hde(x)

make_blocks 19

Author(s)

Philipp Brun

make_blocks

Block-wise split data into training and testing

Description

Creates a stratum vector based on a data.frame with n columns. If the data.frame has one column strata are created based on clusters separated by quantiles. If the data.frame has two or more columns, strata ere created based on k-medoid clusters (function 'pam' from package cluster). Instead of a data.frame also the argument 'npoints' can be provided, then groups are created by random sampling. An opitimization algorithm (function 'gridSearch' from package NMOF) optimizes for equal stratum sizes.

Usage

```
make_blocks(
  nstrat = 4,
  df = data.frame(),
  nclusters = nstrat * 5,
  npoints = NA,
  pres = numeric()
)
```

Arguments

nstrat	Number of approximately equal-sized classes to separate groups in block-cross validation
df	Object of class 'data.frame' with n columns containing critera for cluster building. Not necessary if argument npoints is supplied
nclusters	Number of clusters based on which strata should be built. Minimum is the same number as 'nstrat'. Maximum is nrow(df)/10.
npoints	Optional argument if 'df' is not supplied. For how many points should random sampling be made?
pres	Optional argument if 'df' is not supplied. For how many points should random sampling be made?

Value

Object of class 'vector' of length nrow(df) or 'npoints', with integers defining different strata

Author(s)

Philipp Brun

20 make_tiles

Examples

```
### Test out block generation function
data("Anguilla_train")
vrs=c("SegSumT","USRainDays","USSlope")
env=Anguilla_train[,vrs]
# No layers supplied
strt.1=make_blocks(npoints=1000)
table(strt.1)
# Stratified by 1d layer a
strt.2=make_blocks(df=env[,1,drop=F],nstrat=5,nclusters=5)
table(strt.2)
# Stratified by 1d layer b
strt.3=make_blocks(df=env[,1,drop=F],nstrat=5,nclusters=15)
table(strt.3)
# Stratified by 2d layer a
strt.4 = make\_blocks(df = env[,c(1,3)], nstrat = 3, nclusters = 3)
table(strt.4)
# Stratified by 2d layer b
strt.5=make_blocks(df=env[,c(1,3)],nstrat=5,nclusters=50)
table(strt.5)
# Stratified by 3d layer
strt.6=make_blocks(df=env[,1:3],nstrat=5,nclusters=50)
table(strt.6)
par(mfrow=c(3,2))
plot(env[,c(1,3)],col=strt.1)
plot(env[,c(1,3)],col=strt.2)
plot(env[,c(1,3)],col=strt.3)
plot(env[,c(1,3)],col=strt.4)
plot(env[,c(1,3)],col=strt.5)
plot(env[,c(1,3)],col=strt.6)
```

make_tiles

Create a specific number of tiles based on a raster extent

Description

Based on a specific extent, one or several tiles are generated. Tiles can be smaller raster extents or geometry arguments POLYGON(). The original extent is therefore either converted into a POLYGON() argument, or divided into Ntiles of regular fragments which are converted into POLYGON() arguments and smaller SpatExtent.

Usage

```
make_tiles(geo, Ntiles, sext = TRUE)
```

multi 21

Arguments

geo	Object of class 'Extent', 'SpatExtent', 'SpatialPolygon', 'SpatialPolygonDataframe',
	10 17 1 7770004 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1

or 'SpaVector' (WGS84 or planar) to define the study's area extent. Default is

NULL i.e. the whole globe.

Ntiles Numeric. In how many tiles/fragments should geo be divided approximately?

sext Logical. Should a list of SpatExtent also be returned for each generated POLY-

GON()?

Value

A list of geometry arguments POLYGON() of length Ntiles (and of SpatExtent if sext=TRUE)

References

Chauvier, Y., Thuiller, W., Brun, P., Lavergne, S., Descombes, P., Karger, D. N., ... & Zimmermann, N. E. (2021). Influence of climate, soil, and land cover on plant species distribution in the European Alps. Ecological monographs, 91(2), e01433. 10.1002/ecm.1433

Examples

```
# Load the European Alps Extent
data(geo_dat)

# Apply the function to divide the extent in ~20 fragments
mt = make_tiles(geo=shp.lonlat,Ntiles=20,sext=TRUE); mt
```

multi

Define settings for function that should be supplied to wsl.flex

Description

Create a multi.input object that efficiently stores model specifications

Usage

```
multi(mod, args, tag = "", step = FALSE, weight = FALSE)
```

Arguments

mod	A character with the name of the function to be called. E.g. "gam"
args	A list with arguments to be passed to the function specified in 'mod'
tag	Character with name for model set-up
step	Should step function be applied to update model

Value

Object of class 'multi'

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Author(s)

Philipp Brun

Examples

```
### Preliminary
data("Anguilla_train")
vrs=c("SegSumT","USRainDays","USSlope")
form.glm=as.formula(paste("Presence~",paste(paste0("poly(",vrs,",2)"),collapse="+")))
form.gbm=as.formula(Presence ~ .)
form.glm.2=as.formula(paste("Presence~",paste(vrs,collapse="+")))
feat=c("linear=true","quadratic=true","hinge=true","product=true","threshold=false")
form.gam=as.formula(paste("Presence~",paste(paste0("s(",vrs,")"),collapse="+")))
### Multi examples
multi("glm",list(formula=form.glm,family="binomial"),"glm-simple",step=TRUE)
multi("gbm",list(formula=form.gbm,
                 distribution = "bernoulli",
                 interaction.depth = 1,
                 shrinkage=.01,
                 n.trees = 3500), "gbm-simple")
multi("gam",list(formula=form.gam,family="binomial"),"gam-simple",step=FALSE)
multi("maxent",list(args=feat),"mxe-simple")
multi("randomForest",list(formula=form.gbm,ntree=500,maxnodes=NULL),"waud1")
multi("glm",list(formula=form.glm.2,family="binomial"),"glm-lin",step=TRUE)
```

multi.input-class

An S4 class to store evaluation data

Description

An S4 class to store evaluation data

Slots

```
mod Name of model algorithm to be called (character) e.g., 'glm' args A list of arguments to be supplied to the model fitting alrogrithm tag The name of the model setup (character) step A logical indicating whether step function should be ran (for glm and gam) weight Should observations be weighted based on their prevalence?
```

Author(s)

Philipp Brun

optme 23

optme	Optimization 'make blocks	9	to	create	equal-sized	strata	in	the
	mane_brocks	junction						

Description

Not to be called directly by the user

Usage

```
optme(x, nms, grps, tot)
```

Author(s)

Philipp Brun

prd.pa

Correctly feed the predict functions depending on model type (glm, gbm, maxent...)

Description

Not to be called directly by the user

Usage

```
prd.pa(mod, tst, clust = FALSE)
```

Author(s)

Philipp Brun

prd.pres

Correctly feed the predict functions depending on model type (ppm, glm, gbm, maxent...)

Description

Not to be called directly by the user

24 preva.meta

Usage

```
prd.pres(
  mod,
  env_vars,
  window,
  polly,
  meta,
  coefs,
  id.fact = NULL,
  valid.pres = NULL,
  env_samp = NULL
)
```

Author(s)

Yohann Chauvier

preps

Check input data, collect meta information, take care of data subsetting. Called by model fitting functions.

Description

Not to be called directly by the user

Usage

```
preps(env = parent.frame(), call)
```

Author(s)

Philipp Brun, Yohann Chauvier

preva.meta

Generate meta information for prediction and evaluation

Description

Not to be called directly by the user

Usage

```
preva.meta(env = parent.frame(), type = character())
```

Author(s)

Philipp

prop.sampling 25

p.sam		

Sample pseudo-absences proportional to target group distribution

Description

Uses the 'density' function from the package spatstat to create a density surface of the supplied point pattern and samples pseudo-absences from this density-distribution.

Usage

```
prop.sampling(points, nsamples = 1000, res = 1, ...)
```

Arguments

points matrix or data.frame with column names 'x' and 'y' assumed to be on the same

scale and metric distances (m, km,...)

nsamples number of pseudoabsences to be generated

res resolution of the denstiy grid from which pseudo-absences are drawn. default

(1) corresponds to 1000 cells on the x-axis

... arguments passed on to the 'density' function from package spatstat. Note in

particluar the argument 'adjust' which controls the kernel size in the density

interpolation.

Value

nsamples x 2 matrix with drawn psuedo-absences

Author(s)

Philipp Brun

Examples

```
### Load points

data(mypts)

### Test proportional sampling

pseu.1=prop.sampling(points=my.pts,nsamples=10000,adjust=1,res=1)
pseu.2=prop.sampling(points=my.pts,nsamples=10000,adjust=.5,res=1)
pseu.3=prop.sampling(points=my.pts,nsamples=10000,adjust=.2,res=1)
pseu.4=prop.sampling(points=my.pts,nsamples=10000,adjust=.1,res=1)

par(mfrow=c(2,2))

plot(pseu.1,pch=16,col="red",cex=.5)
points(my.pts)
plot(pseu.2,pch=16,col="red",cex=.5)
points(my.pts)
plot(pseu.3,pch=16,col="red",cex=.5)
```

26 pseu.targr

```
points(my.pts)
plot(pseu.4,pch=16,col="red",cex=.5)
points(my.pts)
```

pseu.targr

Sample pseudo-absences proportional to target group distribution 2.0

Description

This is essentially an improved version of prop.sampling. It uses the 'density' function from the package 'spatstat' to create a density surface of the supplied point pattern and samples pseudo-absences from this density-distribution. It additionally requires a raster layer (env.layer) as input, typically containing an environmental variable used for modelling to extract information about the extent of the study area and the resolution of the environmental information. Furthermore, a mask of class Spatial can be supplied to constrain the area within which pseudoabsences should be sampled. If no mask is supplied, pseudoabsences are sampled in the non-NA cells of the env.layer.

Usage

```
pseu.targr(
  points,
  nsamples = 1000,
  p.avoid = NULL,
  env.layer,
  mask = NULL,
  ...
)
```

Arguments

points	Matrix or data.frame with column names 'x' and 'y' assumed to be on the same scale and metric distances (m, km,)
nsamples	Number of pseudoabsences to be generated
p.avoid	Same type and conditions as "points" argument; avoid sampling pseudo-absences in cells where input coordinates fall
env.layer	A raster layer of desired resolution and extent (and area of interest -> non_NA cells)
mask	Optional. An object of class Spatial to mask for sampling area of interest
	Additional arugements to the density.ppp function (package 'spatstat')

Value

nsamples x 2 matrix with drawn psuedo-absences

Author(s)

Philipp Brun, Yohann Chauvier

stratify 27

Examples

```
# Load files
data(AlpineConvention_lonlat)
data(exrst)
data(xy_ppm)
mypoints=xy.ppm[,c("x","y")]
id.avoid=sample(1:nrow(mypoints),500,replace=TRUE)
xy.avoid=mypoints[id.avoid,]
xy.process=mypoints[-id.avoid,]
# Run pseu.targ()
dens1=pseu.targr(points=xy.process,nsamples=5000,p.avoid=xy.avoid,
                 env.layer=rst[[1]],mask=shp.lonlat,adjust=0.1)
dens2=pseu.targr(points=xy.process,nsamples=5000,p.avoid=xy.avoid,
                 env.layer=rst[[1]],mask=shp.lonlat,adjust=0.5)
dens3=pseu.targr(points=xy.process,nsamples=1000,p.avoid=NULL,
                 env.layer=rst[[1]],mask=NULL,adjust=0.1)
dens4=pseu.targr(points=xy.process,nsamples=1000,p.avoid=NULL,
                 env.layer=rst[[1]],mask=NULL,adjust=0.5)
# Plot results
par(mfrow=c(2,2))
plot(rst[[1]])
points(dens1,pch=20,cex=0.3)
plot(rst[[1]])
points(dens2,pch=20,cex=0.3)
plot(rst[[1]])
points(dens3,pch=20,cex=0.3)
plot(rst[[1]])
points(dens4,pch=20,cex=0.3)
```

stratify

Subsample based on two stratification critera

Description

Subsample based on two stratification critera

Usage

```
stratify(spp, type, sampsiz)
```

Arguments

spp Spatial data.frame

type either 'env.strat' or 'env.semi.strat'

sampsiz desired size of the subsample Not to be called directly by the user

28 upsample_strategic

Author(s)

Philipp

thin_them

Thin a spatial points object by number of points or minimum distance

Description

Thin a spatial points object by number of points or minimum distance

Usage

```
thin_them(spdf, lim_dist = NA, lim_n = NA)
```

Arguments

spdf SpatialPoints or SpatialPointsDataFrame object lim_dist Minimum tolerated distance between the points

lim_n Number of points to be kept

Details

iteratively the closest point pairs are compared and the point with the lower overall distinctiveness is removed

Value

thinned SpatialPoints or SpatialPointsDataFrame object

Author(s)

Philipp

upsample_strategic

Sample a supsample from a large set of points with a minimum distance constraint

Description

Thin a spatial points object by number of points or minimum distance

Usage

```
upsample_strategic(spdf, lim_dist, n_tot, warnig = TRUE)
```

Arguments

spdf	SpatialPoints or SpatialPointsDataFrame object
lim_dist	Minimum tolerated distance between the points
n_tot	Minimum tolerated distance between the points

upsample_thin 29

Details

iteratively samples points and rejects them if min thist to all existing points i the sample are not respected

Value

SpatialPoints or SpatialPointsDataFrame with at least lim_dist between all points

Author(s)

Philipp

upsample_thin Sam

Sample a supsample from a large set of points with a minimum distance constraint

Description

Thin a spatial points object by number of points or minimum distance

Usage

```
upsample_thin(spdf, lim_dist, n_tot)
```

Arguments

spdf SpatialPoints or SpatialPointsDataFrame object
lim_dist Minimum tolerated distance between the points
n_tot Minimum tolerated distance between the points

Details

iteratively samples points and rejects them if min thist to all existing points i the sample are not respected

Value

SpatialPoints or SpatialPointsDataFrame with at least lim_dist between all points

Author(s)

Philipp

30 wsl.ebc

wsl.ebc	Correct environmental bias of species observations via environmental clustering

Description

Our wsl.ebc function corrects environmental bias from an observational dataset based on environmental stratification / clustering. A map of n clusters is generated based on input raster layers (in the article, climate predictors used in PPMs are employed as inputs). Following random equal-stratified sampling design (EBCe; Hirzel & Guisan, 2002), for each cluster, the number of observations per species (relative to all others) is artificially rescaled to the total number of observations found in the cluster presenting the highest observation density. Proportional-stratified sampling design (EBCp; Hirzel & Guisan, 2002) adds a second step: for each cluster, the number of observations per species may additionally be multiplied by the cluster's area (i.e. proportion of pixels in percentage relative to the study area), or by its logarithm (default; consensus between EBCe and EBCp). Resulting output indicates a new number of observations per cluster and species, that the function automatically sub-samples with replacement over the original observational dataset. This function may be used for presences and absences distinctively.

Usage

```
wsl.ebc(
  obs = NULL,
  ras = NULL,
  pportional = TRUE,
  plog = TRUE,
  nclust = 50,
  sp.specific = TRUE,
  sp.cor = 0.5,
  keep.bias = TRUE,
  filter = FALSE,
  path = NULL,
  ...
)
```

Arguments

obs	Object of class matrix or data frame with three columns named sp.id (character), x (numeric) and y (numeric). More than one observation per species must be referenced.
ras	Object of class RasterBrick, RasterStack, list of RasterLayer of desired resolution and extent. Used to generate the map of clusters needed to summarize the environmental space of the study area.
pportional	Logical. Should environmental stratification of observations be proportional to the clusters' areas? If TRUE, EBCp applies.
plog	Logical. Should EBCp apply with a logarithm? If TRUE, a stratification consensus between EBCp and EBCe applies.
nclust	Number of chosen clusters. Default is 50.

wsl.evaluate.pa 31

sp.specific	Logical. Should EBC apply only for species whose environmental bias follows the overall one (i.e. the number of original species observations per cluster is correlated with that of the full dataset)?
sp.cor	If sp.specific = TRUE, spearman's correlation tests are by default set to 0.5; i.e. species with $r < 0.5$ are excluded from the function outputs.
keep.bias	Default is TRUE. Strongly recommended to use when sp.cor = TRUE. Per species, should the number of observations of the cluster in which the species was originally sampled most often, be preserved? Said differently, for each species, should the cluster with the most original species observations be as representative as the cluster with the most corrected observations? If TRUE, after EBC applies, the number of observations per species in their densest original cluster is set to that of the densest corrected cluster.
filter	Logical. Should the observations be filtered according to 'ras' resolution?
path	Path folder where the new species observation files should be saved.
	Additional arguments passed on to the 'clara' function (package 'cluster')

Value

The function returns in 'path', one text file of corrected observations (presences or absences) per species. If the number of new EBC observations per species is too large, sampling those randomly without replacement before model calibrations is advised.

Author(s)

Yohann Chauvier

wsl.evaluate.pa Evaluate presence-absence models

Description

Assess several model skill metrics for all models in a wsl.fit object. Currently AUC, RMSE, TSS, PPV, Accuracy, and Cohen's Kappa are evaluated. Furthermore, the threshold applied is returned.

Usage

```
wsl.evaluate.pa(
    x,
    tester = data.frame(),
    window = NULL,
    thres = numeric(),
    crit = "pp=op",
    prevalence_correction = FALSE,
    pres_only = FALSE,
    log_trans = TRUE,
    bias_cov = NULL
)
```

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Arguments

x A wsl.fit object

tester Optional. A data frame with testing data. Only mandatory if replicatetype='none'

was chosen when models were fitted. Otherwise, used when evaluation against external dataset is needed. Must be a data.frame with as columns in order: "Presence" ('0' and '1'), "CV" (numeric: chosen cv-folds; if replicatetype='none' -> only '1') and associated environmental values (same ones as for fitted models; i.e. same columns order and names). Note that categorical predictor values must be of class factor. NB: Here, model evaluation will only be initiated for the

new testing data.

window Only when 'wsl.ppmO' used'. Same object of class 'owin' used for models (in

developments)

thres Vector of the same length as the number of reps in model fit object. For wsl.flex

model outputs, thresholds have to be labelled with the same names provided to

models.

crit Which threshold criterion should be considered? Currently 'pp=op' (predicted

prevalence = observed prevalence), 'maxTSS' (threshold yielding maximum

TSS), and 'external' (thresholds manually supplied) are possible

prevalence_correction

logical. Should imbalanced presence/absence data be upsampled to prevalence

0.5 for model evaluation.

pres_only Logical. If TRUE, evaluation metrics of presence-absence models is applied to

a wsl.fit object of presence-only models i.e. generated with wsl.ppmGlasso()

arithm before evaluation? Prevent model evaluation errors.

bias_cov A numerical vector whose length equal the number of environmental layers/columns.

Only used when a bias covariate is implemented in the model calibration i.e. to fit species obs. with a potential spatial observer bias. Default is 1 for each variable, whereas designated bias covariate(s) (i.e. 0) will be reset everywhere to

zero in order to evaluate corrected predictions

Value

An object of class 'wsl.evaluation'

Author(s)

Philipp Brun, Yohann Chauvier

Examples

```
# Take anguilla data set from dismo package
data("Anguilla_train")
vrs=c("SegSumT","USRainDays","USSlope")
env=Anguilla_train[,vrs]

### Check out wsl.gam
form.gam=as.formula(paste("Presence~",paste(paste0("s(",vrs,")"),collapse="+")))
# Try out wsl.gam funcion
```

```
modi2=wsl.gam(pa=Anguilla_train$Angaus,
              env_vars = env,
              taxon="Angaus"
              replicatetype="splitsample",
              reps=3,
              project="prototest",
              mod_tag="test-gam",
              formula=form.gam,
              family="binomial",
              step=FALSE)
# Try out custom summary function
summary(modi2)
# Access gam object of first replicate
summary(modi2@fits$replicate_01$`test-gam`)
# Evaluate the model
eval2=wsl.evaluate.pa(modi2,crit="maxTSS")
# Get evaluation summary
summary(eval2)
```

wsl.evaluate.pres

Evaluate presence-only models

Description

Assess model skill metrics for presence-only models in a wsl.fit object. Currently Boyce index is evaluated. Furthermore, the threshold applied is returned.

Usage

```
wsl.evaluate.pres(
    X,
    tester = data.frame(),
    env_vars,
    window = NULL,
    thres = numeric(),
    pres_abs = FALSE,
    log_trans = TRUE,
    speedup = FALSE,
    env_samp = 5000,
    bias_cov = NULL,
    ...
)
```

Arguments

Х

A wsl.ppm fit object

tester	Optional. A data.frame with testing data. Only mandatory if replicatetype='none' was chosen when models were fitted. Otherwise, used when evaluation against external dataset is needed. Must be a data.frame with as columns in order: "x", "y", "Presence" ('0' and '1'), "CV" (numeric: chosen cv-folds; if replicate-type='none' -> only '1') and associated environmental values (same ones as for fitted models; i.e. same columns order and names). Note that categorical predictor values must be of class factor. NB: Here, model evaluation will only be initiated for the new testing data.
env_vars	Same spatial layers used in the fitted model or an object of class 'data.frame' or 'matrix' defining a sample of the target layers by keeping same order for columns. If spatial layers are used, object of class 'RasterStack' or RasterBrick'.
window	Only when 'wsl.ppmO' used'. Same object of class 'owin' used for models (in developments)
thres	Vector of the same length as the number of reps in model fit object
pres_abs	Logical. If TRUE, evaluation metrics of presence-only models is applied to a wsl.fit object of presence-absence models
log_trans	Logical. Use only if pres_abs=FALSE. Should predictions be converted to logarithm before evaluation? Prevent model evaluation errors.
speedup	If env_vars is a 'RasterStack' or RasterBrick', should the boyce evaluation be faster? If TRUE, the algorithm uses a sample of the environmental layers
env_samp	If speedup=TRUE, how many environmental cells should be sampled with replacement? Default is 50'000 samples. The sample may be smaller than requested because of NAs
bias_cov	A numerical vector whose length equal the number of environmental layers/columns. Only used when a bias covariate is implemented in the PPM calibration i.e. to fit species observations with a potential spatial observer bias. Default is 1 for each variable, whereas designated bias covariate(s) (i.e. 0) will be reset everywhere to zero in order to evaluate corrected predictions
	Additional arguments supplied to ecospat.boyce function (package 'ecospat')

Value

an object of class 'wsl.evaluation'. If in slot "performance" NA as thresholds are found, it indicates a lack of convergence in the model tested, and so, a biased/invalid threshold

Author(s)

Yohann Chauvier, Philipp Brun

Examples

```
### Load

data(AlpineConvention_lonlat)
data(exrst)
rst = rst[[1:6]]
data(xy_ppm)
mypoints = xy.ppm[,c("x","y")]
```

```
### Define mask
maskR = mask(rst[[1]],shp.lonlat)
### Run 'wsl.ppm.window' function
wind = wsl.ppm.window(mask = maskR,
                      val = 1,
                      owin = TRUE)
### Define quadrature points for 'wsl.ppmGlasso'
   # Grid regular
quadG1 = wsl.quadrature(mask = maskR,
                        area.win = wind,
                        random = FALSE,
                        lasso = TRUE,
                        env_vars = rst)
### Define quadrature points for 'wsl.ppm0'
   # Grid regular
quadO1 = wsl.quadrature(mask = maskR,
                        area.win = wind,
                        random = FALSE,
                        lasso = FALSE,
                        env_vars = NULL)
### Define your environments
   # For 'wsl.ppmGlasso' (observations focus)
envG = raster::extract(rst,mypoints)
   # For 'wsl.ppm0' (study area focus)
env0 = wsl.ppm.env(rst,maskR)
### Modelling
   # 'wsl.ppmGlasso' (alpha = 0.5 => Elastic net, see package 'glmnet')
       # Complex PPPM lasso (poly = TRUE & lasso=TRUE)
lasso1 = wsl.ppmGlasso(pres = mypoints,
                       quadPoints = quadG1$Edata,
                       asurface = raster::area(shp.lonlat)/1000,
                       env_vars = envG,
                       taxon = "species_eg1",
                       replicatetype = "cv",
                       reps = 5,
                       strata = NA,
                       save=FALSE,
                       project = "lasso_eg1",
                       path = NA,
                       type = "binomial",
                       poly = TRUE,
                       lasso = TRUE,
                       alpha = 0.5,
                       type.measure = "mse",
```

```
standardize = TRUE,
                       nfolds = 5,
                       nlambda = 100)
       # Simple PPPM non lasso (poly = FALSE & lasso=FALSE)
lasso2 = wsl.ppmGlasso(pres = mypoints,
                       quadPoints = quadG1$Edata,
                       asurface = raster::area(shp.lonlat)/1000,
                       env_vars = envG,
                       taxon = "species_eg2",
                       replicatetype = "cv",
                       reps = 5,
                       strata = NA,
                       save = FALSE,
                       project = "lasso_eg2",
                       path = NA,
                       type = "binomial",
                       mask = maskR,
                       poly = FALSE,
                       lasso = FALSE)
   # 'wsl.ppm0'
       # Simple PPPM non lasso (same as above with poly = FALSE & lasso = FALSE)
form.Sppm = as.formula(paste("~",paste(names(env0),collapse="+")))
lasso3 = wsl.ppmO(pres = mypoints,
                  quadPoints = quadO1,
                  env_vars = env0,
                  window = wind,
                  taxon = "species_eg3",
                  replicatetype = "cv",
                  reps = 5,
                  strata = NA,
                  save = FALSE,
                  project = "lasso_eg3",
                  path = NA,
                  formula = form.Sppm)
       # Complex PPPM non lasso
form.Cppm = as.formula(paste("~",paste(paste0("poly(",names(env0),",2)"),collapse="+")))
lasso4 = wsl.ppmO(pres = mypoints,
                  quadPoints = quadO1,
                  env_vars = env0,
                  window = wind,
                  taxon = "species_eg3",
                  replicatetype = "cv",
                  reps = 5,
                  strata = NA,
                  save = FALSE,
                  project = "lasso_eg3",
                  path = NA,
                  formula = form.Cppm)
```

Evaluation

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```
# Example for 'wsl.ppmGlasso'
eval1 = wsl.evaluate.pres(x = lasso1,
                          tester = NULL,
                          env_vars = rst,
                          mask = maskR,
                          window = NULL,
                          thres = NULL)
eval2 = wsl.evaluate.pres(x = lasso2,
                          tester = NULL,
                          env_vars = rst,
                          mask = maskR,
                          window = NULL,
                          thres = 0.001)
   # Example for 'wsl.ppm0'
eval3 = wsl.evaluate.pres(x = lasso3,
                          tester = NULL,
                          env_vars = env0,
                          mask = maskR,
                          window = wind,
                          thres = NULL)
eval4 = wsl.evaluate.pres(x = lasso4,
                          tester = NULL,
                          env_vars = env0,
                          mask = maskR,
                          window = wind,
                          thres = NULL)
```

wsl.evaluation-class An S4 class to store evaluation data

Description

An S4 class to store evaluation data

Slots

meta A list with meta information

thres A vector with externally supplied thresholds

performance A list with model performance estimates

Author(s)

Philipp Brun

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wsl.fit-class

An S4 class to store fitted objects

Description

An S4 class to store fitted objects

Slots

```
meta A list with meta information
tesdat A list with held out data to be used for testing
performance fitted model objects
call the function call
```

Author(s)

Philipp Brun, Yohann Chauvier

wsl.ppm.env

Poisson Point Process Models (PPPM) 'env'

Description

Convert envrionmental data for 'wsl.ppmO' fitting.

Usage

```
wsl.ppm.env(env_vars, mask)
```

Arguments

env_vars Object of class 'RasterBrick' or 'RasterStack'. Same extent and resolution as

'mask'

mask Object of class 'RasterLayer' or 'RasterBrick' or 'RasterStack'. Mask is defined

according to non NAs values and the resolution of the study

Value

List of objects of class 'im'

Author(s)

Yohann Chauvier

wsl.ppm.qscheme 39

Examples

```
### Load

data(exrst)
data(xy_ppm)
mypoints = xy.ppm[,c("x","y")]

### Define mask

maskR = mask(rst[[1]],shp.lonlat)

### Define your environments

    # For 'wsl.ppmGlasso' (observations focus)
envG = raster::extract(rst,mypoints)

    # For 'wsl.ppmO' (study area focus)
envO = wsl.ppm.env(rst,maskR)
```

wsl.ppm.qscheme

To prepare PPM general quadratic environment with input species points

Description

Not to be called directly by the user

Usage

```
wsl.ppm.qscheme(data, area.win, quads)
```

Author(s)

Yohann Chauvier

wsl.ppm.window

Poisson Point Process Models (PPPM) 'window'

Description

Set up a study window of class 'owin' or 'im' necessary to run 'wsl.ppmO' fit function.

Usage

```
wsl.ppm.window(mask, val = 1, owin)
```

Arguments

mask Object of class 'RasterLayer' or 'RasterBrick' or 'RasterStack'. Mask is de-

fined according to non NAs values. Defined study area in pixels. Same as in

'wsl.quadrature'

val Values that are assigned to the 'owin' or 'im' mask

owin TRUE or FALSE

Value

Object of class 'owin' or 'im'

Author(s)

Yohann Chauvier

Examples

wsl.predict.pa

Make predictions

Description

Make predictions with all models from a wsl.fit object. If thresholds are supplied binary predictions are made if convert=TRUE, otherwise continuous predictions and separate description of thresholds are returned

Usage

```
wsl.predict.pa(
   x,
   predat = data.frame(),
   thres = numeric(),
   bias_cov = NULL,
   clust = FALSE
)
```

Arguments

An object of class wsl.fit predat Data.frame or raster for which predictions should be made thres Optional. Object of the same length as the number of replicates, or model types if a mean is applied accross model types. Obtained with 'get_thres' bias_cov A numerical vector whose length equal the number of environmental layers/columns. Only used when a bias covariate is implemented in calibrations i.e. to fit species observations with a potential spatial observer bias. Default is 1 for each variable,

whereas designated bias covariate(s) (i.e. 0) will be reset everywhere to zero in

order to evaluate corrected predictions

Logical. If raster predictions are made, should the operation be run in parallel? clust

Value

Object of class wsl.prediction with slots for meta info, and model predictions

Author(s)

Philipp Brun, Yohann Chauvier

Examples

```
# Take anguilla data set from dismo package
data("Anguilla_train")
vrs=c("SegSumT","USRainDays","USSlope")
env=Anguilla_train[,vrs]
### Check out wsl.glm
form.glm=as.formula(paste("Presence~",paste(paste0("poly(",vrs,",2)"),collapse="+")))
modi1=wsl.glm(pa=Anguilla_train$Angaus,
              env_vars = env,
              taxon="Angaus",
              replicatetype="cv",
              reps=5,
              project="prototest",
              mod_tag="test-glm",
              formula=form.glm,
              family="binomial",
              step=TRUE)
# Try out custom summary function
summary(modi1)
# Access glm object of first replicate
summary(modi1@fits$replicate_01$`test-glm`)
# Evaluate the model
eval1=wsl.evaluate.pa(modi1)
# Get evaluation summary
summary(eval1)
```

```
### Check out wsl.gam
form.gam=as.formula(paste("Presence~",paste(paste0("s(",vrs,")"),collapse="+")))
# Try out wsl.glm funcion
modi2=wsl.gam(pa=Anguilla_train$Angaus,
              env_vars = env,
              taxon="Angaus",
              replicatetype="splitsample",
              reps=3,
              project="prototest",
              mod_tag="test-gam",
              formula=form.gam,
              family="binomial",
              step=FALSE)
# Try out custom summary function
summary(modi2)
# Access glm object of first replicate
summary(modi2@fits$replicate_01$`test-gam`)
# Evaluate the model
eval2=wsl.evaluate.pa(modi2,crit="maxTSS")
# Get evaluation summary
summary(eval2)
### Check out wsl.gbm
form.gbm=as.formula(Presence ~ .)
# Try out wsl.glm funcion
modi3=wsl.gbm(pa=Anguilla_train$Angaus,
              env_vars = env,
              taxon="Angaus",
              replicatetype="none",
              reps=1,
              project="prototest",
              mod_tag="test-brt",
              formula= form.gbm,
              distribution = "bernoulli",
              interaction.depth = 1,
              shrinkage=.01,
              n.trees = 3500)
# Try out custom summary function
summary(modi3)
# Access glm object of first replicate
summary(modi3@fits$replicate_01$`test-brt`)
# Prepare external testing data
tste=data.frame(Presence=Anguilla_train$Angaus,env)
# Evaluate the model
eval3=wsl.evaluate.pa(modi3,crit="maxTSS",tester=tste)
# Get evaluation summary
```

```
summary(eval3)
### Check out wsl.maxent
feat=c("linear=true","quadratic=true","hinge=true","product=true","threshold=false")
# Try out wsl.glm funcion
modi4=wsl.maxent(pa=Anguilla_train$Angaus,
                 env_vars = env,
                 taxon="Angaus",
                 replicatetype="block-cv",
                 strata=sample(1:3,nrow(env),replace=TRUE),
                 project="prototest",
                 mod_tag="test-mxe",
                 args=feat)
# Try out custom summary function
summary(modi4)
# Access glm object of first replicate
summary(modi4@fits$replicate_01$`test-mxe`)
# Define external threshold
thmxe=c('test-mxe'=0.5)
# Evaluate the model
eval4=wsl.evaluate.pa(modi4,crit="external",thres=thmxe)
# Get evaluation summary
summary(eval4)
# Get thresholds
thr.4=get_thres(eval4)
### Check out wsl.flex
form.glm.2=as.formula(paste("Presence~",paste(vrs,collapse="+")))
\verb|modinp=list(multi("glm", list(formula=form.glm, family="binomial"), "glm-simple", step=TRUE), \\
multi("gbm",list(formula=form.gbm,
                 distribution = "bernoulli",
                 interaction.depth = 1,
                 shrinkage=.01,
                 n.trees = 3500), "gbm-simple"),
multi("gam",list(formula=form.gam,family="binomial"),"gam-simple",step=FALSE),
multi("maxent", list(args=feat), "mxe-simple"),
multi("randomForest",list(formula=form.gbm,ntree=500,maxnodes=NULL),"waud1"),
multi("glm",list(formula=form.glm.2,family="binomial"),"glm-lin",step=TRUE))
# Try out wsl.glm funcion
modi5=wsl.flex(pa=Anguilla_train$Angaus,
               env_vars = env,
               taxon="Angaus",
               replicatetype="block-cv",
               strata=sample(1:3,nrow(env),replace=TRUE),
               project="multitest",
               mod_args=modinp)
```

```
# Try out custom summary function
summary(modi5)

# Access glm object of first replicate
summary(modi5@fits$replicate_01$`glm-simple`)

# Evaluate the model
eval5<-wsl.evaluate.pa(modi5,crit="pp=op")

# Get evaluation summary
summary(eval5)

# Get thresholds
thr.5=get_thres(eval5)

### Make some predictions
pred4=wsl.predict.pa(modi4,predat=env)
pred5=wsl.predict.pa(modi5,predat=env,thres=thr.5)</pre>
```

wsl.predict.pres

Make predictions

Description

Make predictions with all models from a wsl.fit object. If thresholds are supplied binary predictions are made, otherwise continuous predictions are returned.

Usage

```
wsl.predict.pres(
    x,
    thres = numeric(),
    predat = list(),
    window = NULL,
    log_trans = TRUE,
    raster = FALSE,
    bias_cov = NULL
)
```

Arguments

X	An object of class	wsl.fit
X	An object of class	WSI.I

thres Optional. Object of the same length as the number of replicates, or model types

if a mean is applied accross model types. Obtained with 'get_thres'

predat Same spatial layers used in the fitted model or an object of class 'data.frame'

or 'matrix' defining a sample of the target layers by keeping same order for columns. If spatial layers, when wsl.ppmGlasso' used, object of class 'Raster-Stack' or RasterBrick'. When 'wsl.ppmO' used, a list of 'im' object. 'wsl.ppmO'

must be used with bias_cov = NULL (in developments)

window Only when 'wsl.ppmO' used'. Same object of class 'owin' used for models

binary? Should be TRUE if log.trans was TRUE when using wsl.evaluate

raster Logical. Should the output be a list of rasters or matrix?

bias_cov A numerical vector whose length equal the number of environmental layers/columns.

Only used when a bias covariate is implemented in the PPM calibration i.e. to fit species observations with a potential spatial observer bias. Default is 1 for each variable, whereas designated bias covariate(s) (i.e. 0) will be reset everywhere

to zero in order to evaluate corrected predictions

Value

Object of class wsl.prediction with slots for meta info, and model predictions

Author(s)

Yohann Chauvier, Philipp Brun

Examples

```
### Load
data(AlpineConvention_lonlat)
data(exrst)
rst = rst[[1:6]]
data(xy_ppm)
mypoints = xy.ppm[,c("x","y")]
### Define mask
maskR = mask(rst[[1]],shp.lonlat)
### Run 'wsl.ppm.window' function
wind = wsl.ppm.window(mask = maskR,
                      val = 1,
                      owin = TRUE)
### Define quadrature points for 'wsl.ppmGlasso'
   # Grid regular
quadG1 = wsl.quadrature(mask = maskR,
                        area.win = wind,
                        random = FALSE,
                        lasso = TRUE,
                        env_vars = rst)
### Define quadrature points for 'wsl.ppm0'
   # Grid regular
quad01 = wsl.quadrature(mask = maskR,
                        area.win = wind,
                        random = FALSE,
                        lasso = FALSE,
                        env_vars = NULL)
```

```
### Define your environments
   # For 'wsl.ppmGlasso' (observations focus)
envG = raster::extract(rst,mypoints)
   # For 'wsl.ppm0' (study area focus)
env0 = wsl.ppm.env(rst,maskR)
### Modelling
   # 'wsl.ppmGlasso' (alpha = 0.5 => Elastic net, see package 'glmnet')
       # Complex PPPM lasso (poly = TRUE & lasso=TRUE)
lasso1 = wsl.ppmGlasso(pres = mypoints,
                       quadPoints = quadG1$Edata,
                       asurface = raster::area(shp.lonlat)/1000,
                       env_vars = envG,
                       taxon = "species_eg1",
                       replicatetype = "cv",
                       reps = 5,
                       strata = NA,
                       save=FALSE,
                       project = "lasso_eg1",
                       path = NA,
                       type = "binomial",
                       mask = maskR,
                       poly = TRUE,
                       lasso = TRUE,
                       alpha = 0.5,
                       type.measure = "mse",
                       standardize = TRUE,
                       nfolds = 5,
                       nlambda = 100)
       # Simple PPPM non lasso (poly = FALSE & lasso=FALSE)
lasso2 = wsl.ppmGlasso(pres = mypoints,
                       quadPoints = quadG1$Edata,
                       asurface = raster::area(shp.lonlat)/1000,
                       env_vars = envG,
                       taxon = "species_eg2",
                       replicatetype = "cv",
                       reps = 5,
                       strata = NA,
                       save = FALSE,
                       project = "lasso_eg2",
                       path = NA,
                       type = "binomial",
                       mask = maskR,
                       poly = FALSE,
                       lasso = FALSE)
   # 'wsl.ppm0'
       # Simple PPPM non lasso (same as above with poly = FALSE & lasso = FALSE)
form.Sppm = as.formula(paste("~",paste(names(env0),collapse="+")))
lasso3 = wsl.ppmO(pres = mypoints,
```

```
quadPoints = quadO1,
                  env_vars = env0,
                  window = wind,
                  taxon = "species_eg3",
                  replicatetype = "cv",
                  reps = 5,
                  strata = NA,
                  save = FALSE,
                  project = "lasso_eg3",
                  path = NA,
                  formula = form.Sppm)
       # Complex PPPM non lasso
form.Cppm = as.formula(paste("~",paste(paste0("poly(",names(env0),",2)"),collapse="+")))
lasso4 = wsl.ppmO(pres = mypoints,
                  quadPoints = quadO1,
                  env_vars = env0,
                  window = wind,
                  taxon = "species_eg3",
                  replicatetype = "cv",
                  reps = 5,
                  strata = NA,
                  save = FALSE,
                  project = "lasso_eg3",
                  path = NA,
                  formula = form.Cppm)
### Evaluation
   # Example for 'wsl.ppmGlasso'
eval1 = wsl.evaluate.pres(x = lasso1,
                          tester = NULL,
                          env_vars = rst,
                          mask = maskR,
                          window = NULL,
                          thres = NULL)
eval2 = wsl.evaluate.pres(x = lasso2,
                          tester = NULL,
                          env_vars = rst,
                          mask = maskR,
                          window = NULL,
                          thres = 0.001)
   # Example for 'wsl.ppmO'
eval3 = wsl.evaluate.pres(x = lasso3,
                          tester = NULL,
                          env_vars = env0,
                          mask = maskR,
                          window = wind,
                          thres = NULL)
eval4 = wsl.evaluate.pres(x = lasso4,
                          tester = NULL,
                          env_vars = env0,
```

```
mask = maskR,
                          window = wind,
                          thres = NULL)
### Thresholds
get_thres(eval1, mean = FALSE)
get_thres(eval1, mean = TRUE)
get_thres(eval2, mean = FALSE)
get_thres(eval2, mean = TRUE)
get_thres(eval3, mean = FALSE)
get_thres(eval3, mean = TRUE)
get_thres(eval4, mean = FALSE)
get_thres(eval4, mean = TRUE)
### Predictions
   # 'wsl.ppmGlasso'
pred1 = wsl.predict.pres(x = lasso1,
                         predat = rst,
                         window = NULL,
                         mask = maskR,
                         thres = get_thres(eval1, mean=FALSE),
                         raster = TRUE)
par(mfrow=c(2,3))
sapply(1:5,function(x) plot(pred1@predictions[[x]][[1]]))
pred2 = wsl.predict.pres(x = lasso2,
                         predat = rst,
                         window = NULL,
                         mask = maskR,
                         thres = NULL,
                         raster = TRUE)
par(mfrow=c(2,3))
sapply(1:5,function(x) plot(pred2@predictions[[x]][[1]]))
pred3 = wsl.predict.pres(x = lasso1,
                         predat = rst,
                         window = NULL,
                         mask = maskR,
                         thres = get_thres(eval1, mean=TRUE),
                         raster = TRUE)
par(mfrow=c(2,3))
sapply(1:5,function(x) plot(pred3@predictions[[x]][[1]]))
pred4 = wsl.predict.pres(x = lasso2,
                         predat = rst,
                         window = NULL,
                         mask = maskR,
                         thres = NULL,
                         raster = FALSE)
```

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```
'wsl.ppm0'
pred5 = wsl.predict.pres(x = lasso3,
                         predat = env0,
                         window = wind,
                         mask = NULL,
                         thres = get_thres(eval3,mean=FALSE),
                         raster = TRUE)
par(mfrow=c(2,3))
sapply(1:5,function(x) plot(pred5@predictions[[x]][[1]]))
pred6 = wsl.predict.pres(x = lasso4,
                         predat = env0,
                         window = wind,
                         mask = NULL,
                         thres = get_thres(eval4,mean=FALSE),
                         raster = TRUE)
par(mfrow=c(2,3))
sapply(1:5,function(x) plot(pred6@predictions[[x]][[1]]))
```

wsl.prediction-class An S4 class to store prediction data

Description

An S4 class to store prediction data

Slots

meta A list with meta information
thres A vector with externally supplied thresholds
predictions A list with model predictions

Author(s)

Philipp Brun

```
wsl.pseudoabsences-class
```

An S4 class to store pseudoabsence data

Description

An S4 class to store pseudoabsence data

50 wsl.quadrature

Slots

```
meta A list with meta information
type The type of pseudoabsence sampling
pa a vector with presences and pseudo absences
env_vars data.frame with environmental predictors
xy a matrix with coordinates of the points
```

Author(s)

Philipp

wsl.quadrature

Poisson Point Process Models (PPPM) 'quadrature'

Description

Set up quadrature points (or "background points") necessary to run 'wsl.ppmO' and 'wsl.ppmGlasso' fit functions. Those points apply a spatial scaling proportional to the study area and estimate the maximised model log likelihood (see Renner 2013, Renner et al. 2015)

Usage

```
wsl.quadrature(
  mask,
  area.win,
  random = FALSE,
  nQ = 1e+05,
  lasso = TRUE,
  env_vars = NULL
)
```

Arguments

mask	Object of class 'RasterLayer' or 'RasterBrick' or 'RasterStack'. Mask is defined according to non NAs values. Defines study area in pixels used to grid sample the quadrature points. Sampling is done over raster centroids so the resolution of the mask defines the desired sampling.
area.win	Object of class 'owin'. 'owin' output of 'wsl.ppm.window'
random	Logical. Should quadrature points be generated randomly or according to a regular mask?
nQ	To choose the number of random quadrature points in case 'random=TRUE'

lasso Logical. Form of the output. If TRUE, 'wsl.ppmGlasso' is used

env_vars Only when 'lasso=TRUE'. Object of class 'RasterBrick' or 'RasterStack'. Set

of predictors the user wants to use to fit the model

Value

Object of class 'ppp' or 'wsl.quads'. Points associated to NAs env. values are removed

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Author(s)

Yohann Chauvier

Examples

```
#### Load
data(AlpineConvention_lonlat)
data(exrst)
### Define mask
maskR = mask(rst[[1]],shp.lonlat)
### Run 'wsl.ppm.window' function
wind = wsl.ppm.window(mask = maskR,
                      val = 1,
                      owin = TRUE)
### Define quadrature points for 'wsl.ppmGlasso'
   # Grid regular
quadG1 = wsl.quadrature(mask = maskR,
                        area.win = wind,
                        random = FALSE,
                        lasso = TRUE,
                        env_vars = rst)
   # Randomly
quadG2 = wsl.quadrature(mask = maskR,
                        area.win = wind,
                        random = TRUE,
                        nQ = 100000,
                        lasso = TRUE,
                        env_vars = rst)
### Define quadrature points for 'wsl.ppm0'
   # Grid regular
quadO1 = wsl.quadrature(mask = maskR,
                        area.win = wind,
                        random = FALSE,
                        lasso = FALSE,
                        env_vars = NULL)
   # Randomly
quadO2 = wsl.quadrature(mask = maskR,
                        area.win = wind,
                        random = TRUE,
                        nQ = 100000,
                        lasso = FALSE,
                        env_vars = NULL)
```

wsl.quads-class

An S4 class to store quadrature objects

Description

An S4 class to store quadrature objects

Slots

```
coordinates informations on the generated quadratures predictors values extracted for each quadrature
```

Author(s)

Yohann Chauvier, Philipp Brun

wsl.samplePseuAbs

Sample pseudoabsences using various strategies

Description

Flexlible function to sample pseudoabsences with various strategies and store the results in a 'wsl.pseudoabsences' object that can be passed on in the 'wsl.biodiv' pipeline.

Usage

```
wsl.samplePseuAbs(
  n = 10000,
  env.stack,
  type = "geographic",
  add.strat = 0,
  pres = numeric(),
  taxon = character(),
  geodist_fact = 1,
  geores_fact = 20,
  template_dir = NA,
  geo\_nrep = 7,
  target.group_dir = NA,
  env.strat_path = NA,
  rAll = TRUE,
  force_spat_thin = "no",
  limdist = NA,
  set_max_npres_to_nabs = TRUE
)
```

Arguments

n number of pseudoabsence points desired. Default is 10000.

env. stack RasterStack with environmental layers for sampling and extraction

type Desired sampling strategy. Options are 'geographic', 'density', 'random', 'tar-

get.group', 'geo.strat', 'env.strat' and 'env.semi.strat' (see details). Default is

'geographic'

add.strat Fraction between 0 and 1; should strategy be complemented by a fraction of

environmental strata.

pres SpatialPoints object. Location of presence points. Necessary for 'geographic'

sampling strategy and the best adding point if the downstream functions will be

used.

taxon Character; name of taxon of interest to keep track of meta information.

geodist_fact Factor to adjust spatial autocorrelation lengths: for 'geographic' pseudoabsence

point patterns, values below 1 increase autocorrelation length; values above 1

decrease it; for 'density' sampling it is the other way around.

geores_fact Aggregation factor 'geographic' template. Larger values save calculation time,

but decrease resolution of sampling points.

template_dir Directory where template raster for 'geographic' sampling should be saved in/loaded

from. If NA, nothing will be saved; if provided, template will be saved in/loaded

from directory depending on whether a file already exists.

geo_nrep number of replicates of geographic models. More will create a smoother pattern

but increase computation time.

target.group_dir

Directory where xy files of target group taxa are stored. Must be supplied if sampling stragegy is 'target.group', must contain a column names 'x' and 'y'

with coordinates in the same projection as other spatial data

env.strat_path Directory where sample of environmental strata for 'env.strat' sampling should

be saved in/loaded from. If NA, nothing will be saved; if provided, environmental strata will be saved in/loaded from directory depending on whether a file

already exists.

rAll should all data be read into memory for computation of environmental strata?

this is faster but you may run into memory issues for large rasters.

force_spat_thin

Should minimum distance be enforced between points? Options are 'no', 'presences', 'absences', and 'both'. By default thinning is defined for pseudoab-

sences from 'geographic', 'density', 'random', and 'geo.strat' methods with minimum distance according to the resolution of the template raster. 'presences' takes the minimum distance criterion from the template raster over to the 'presence' points; 'absences' takes the criterion over to 'env.strat', 'env.semi.strat',

and 'target.group'; 'both' does it for both.

limdist The minimum distance accepted for spatial thinning. Units should be km if the

spatial data is projected, otherwise the units of the coordinate reference system used. If no value is supplied, the maximum distance between two cells of the

template raster will be taken.

set_max_npres_to_nabs

logical. Should the maximum number of presences be equal to the number of

pseudoabsences defined.

Details

'geographic' samples pseudoabsences with a sampling probability. inversely proportional to the geographic distance to presence observations. 'density' samples pseudoabsences proportional to the density of presence observations. 'random' samples pseudoabsences randomly with a sampling probability proportional the area of the cells. 'target.group' samples pseudoabsences from the presences of the taxa of the target group, attempting to correct for sampling bias. It depends on a directory with taxa defined by the user as target group. 'geo.strat' samples pseudoabsences geographically stratified either on a plane, or on a sphere depending on the projection of the supplied env.stack. 'env.strat' samples pseudoabsences environmentally stratified. Points are sampled form all realized combinations of environmental conditions occuring in the environmental stack that have a minimal occurrence frequency. By default environmental strata are calculated based on all raster layers supplied. If a directory is supplied as 'env.strat_path', a large sample of stratified points will be saved to speed up computations for follow-up species. If environmental strata based on different predictors than supplied are preferred 'env.strat_path'can be an .RData file from a previous sampling of strata form different environmental predictors. 'env.semi.strat' is similar to 'env.strat' but samples environmental strata proportional to the logarithm of the area they cover.

Value

an object of class 'wsl.pseudoabsences' that can be plotted and passed on to wsl.flex

Author(s)

Philipp

Examples

```
### Data preparation
# Predictors
bio=getData('worldclim', var='bio', lon=16, lat=48, res=.5)
bio=bio[[c(1,4,12)]]
# install.packages("rgbif")
library(rgbif)
# extract species
spn='Boletus aestivalis'
xt=as.vector(extent(bio))
baest <- occ_search(scientificName=spn,</pre>
                hasCoordinate=TRUE,
                 decimalLongitude=paste0(xt[1],",",xt[3]),
                 decimalLatitude=paste0(xt[2],",",xt[4]))
pbaest=baest$data[,c('decimalLongitude','decimalLatitude')]
baest_spp=SpatialPoints(pbaest,proj4string = crs(bio))
# extract target group
targr <- occ_search(familyKey = 8789,</pre>
                 hasCoordinate=TRUE,
                 limit = 10000,
                 decimalLongitude=paste0(xt[1],",",xt[3]),
                 decimalLatitude=paste0(xt[2],",",xt[4]))
```

```
ptargr=as.matrix(targr$data[,c('decimalLongitude','decimalLatitude')])
colnames(ptargr)=c("x","y")
# create temporary directory for target.group info
tdir=paste0(tempdir(),"/trgr")
dir.create(tdir)
write.table(ptargr,file=paste0(tdir,"/targetxy.txt"),row.names = F)
# create temporary directory for template raster and env strata
strdir=paste0(tempdir(),"/str")
dir.create(strdir)
# Note that for these should not be temporary files for a real analysis.
### Sample pseudoabsences
# Geograppic method with 20% env strata
pseu.abs1=wsl.samplePseuAbs(type="geographic",
                       n=5000,
                       env.stack=bio,
                       pres=baest_spp,
                       add.strat=0.2,
                       template_dir=strdir,
                       env.strat_path=strdir,
                       geodist_fact=1,
                       geores_fact=3,
                       geo_nrep=7,
                       taxon=spn)
plot(pseu.abs1)
# Only geographic with longer autocorrelation length
pseu.abs2=wsl.samplePseuAbs(type="geographic",
                        n=5000,
                        env.stack=bio,
                        pres=baest_spp,
                        add.strat=0,
                        template_dir=strdir,
                        env.strat_path=strdir,
                        geodist_fact=.5,
                        geores_fact=3,
                        geo_nrep=7,
                        taxon=spn)
plot(pseu.abs2)
# Random and thin presences
pseu.abs3=wsl.samplePseuAbs(type="random",
                        n=5000,
                        env.stack=bio,
                        template_dir=strdir,
                        pres=baest_spp,
                        geores_fact=3,
                        add.strat=0,
                        taxon=spn,
```

```
force_spat_thin="presences")
plot(pseu.abs3)
# Geo.start
pseu.abs4=wsl.samplePseuAbs(type="geo.strat",
                           n=5000,
                           env.stack=bio,
                           template_dir=strdir,
                           pres=baest_spp,
                           geores_fact=3,
                           add.strat=0,
                           taxon=spn)
plot(pseu.abs4)
# Target group with 20% env strat
pseu.abs5=wsl.samplePseuAbs(type="target.group",
                         n=5000.
                         env.stack=bio,
                         template_dir=strdir,
                         target.group_dir=tdir,
                         env.strat_path=strdir,
                         geores_fact=3,
                         pres=baest_spp,
                         add.strat=0.2,
                         taxon=spn,
                         force_spat_thin="both")
plot(pseu.abs5)
# Environmental semi-stratified
pseu_abs6=wsl.samplePseuAbs(n = 5000,
                           env.stack=bio,
                           type = "env.semi.strat",
                           add.strat = 0,
                           pres = baest_spp,
                           taxon = spn,
                           template_dir=strdir,
                           env.strat_path=strdir)
plot(pseu_abs6)
# Environmental semi-stratified with min dist
pseu_abs7=wsl.samplePseuAbs(n = 5000,
                           env.stack=bio,
                           type = "env.semi.strat",
                           add.strat = 0,
                           geores_fact=3,
                           pres = baest_spp,
                           taxon = spn,
                           template_dir=strdir,
                           env.strat_path=strdir)
plot(pseu_abs7)
# Density dependent
```

```
pseu_abs8=wsl.samplePseuAbs(n = 5000,
                     env.stack=bio,
                     type = "density",
                     add.strat = 0,
                     pres = baest_spp,
                     taxon = spn,
                     geores_fact=3,
                     template_dir=strdir,
                     env.strat_path=strdir)
plot(pseu_abs8)
### Fit SDMs
# Define model settings
vrs=names(bio)
form. glm=as. formula (paste("Presence", paste(paste0("poly(", vrs, ", 2)"), collapse="+"))) \\
form. gam=as. formula (paste("Presence", paste(paste0("s(", vrs, ")"), collapse="+"))) \\
form.tree=as.formula(Presence ~ .)
modinp=list(multi("glm",list(formula=form.glm,family="binomial"),"glm-simple",step=FALSE),
        multi("gbm",list(formula=form.tree,
            distribution = "bernoulli",
            interaction.depth = 1,
            shrinkage=.01,
            n.trees = 3500), "gbm-simple"),
       multi("gam",list(formula=form.gam,family="binomial"),"gam-simple",step=FALSE),
       multi("randomForest",list(formula=form.tree,ntree=500,maxnodes=NULL),"waud1"))
# Fit models using wsl.flex function
library(gam)
library(gbm)
library(randomForest)
modi5=wsl.flex(x=pseu.abs5,
           replicatetype="block-cv",
           reps=3,
           strata=sample(1:3,nrow(pseu.abs5@env_vars),replace=TRUE),
           project="multitest",
           mod_args=modinp)
### Evaluate
# Evaluate the models with wsl.evaluate function
eval5<-wsl.evaluate(modi5,crit="maxTSS",prevalence_correction = TRUE)</pre>
summary(eval5)
### Predict
modi_pred=wsl.flex(x=pseu.abs5,
              replicatetype="none",
              reps=1,
```

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wsl.test.quads

Poisson Point Process Models (PPPM) 'quadrature' for presenceabsence evaluation metrics

Description

Sample independent test quadrature points over the study area used for evaluating presence-only models with presence-absence metrics (i.e. AUC, TSS, Kappa, PPV...). Presence-only models can be evaluated with other metrics than Boyce index by using random sampling points as absences. But these absences validation data need to be generated independently over the study area i.e. by avoiding resampling quadrature points already used in the models.

Usage

```
wsl.test.quads(mask, quadrature, env_vars, nQ = 10000, replace = TRUE)
```

Arguments

mask	Object of class 'RasterLayer' or 'RasterBrick' or 'RasterStack' of. Mask is defined according to non NAs values. Defines study area in pixels used to grid sample the quadrature points. Sampling is done over raster centroids so the resolution of the mask defines the desired sampling. Same as in 'wsl.quadrature'
quadrature	Object of class 'wsl.quads' or 'ppp'. May be one object or a list of object. Same as in wsl.ppm.fit
env_vars	An object of class 'RasterStack' or 'RasterBrick'. Same spatial layers used in the models. Use to extract env values from test quadrature points
nQ	Mumber of random quadrature points to sample
replace	Logical. If TRUE, quadrature sampling is done with replacements

Value

Object of class 'wsl.quads' or 'list'. Points associated to NAs env. values are removed

Author(s)

Yohann Chauvier

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Examples

```
#### Load
data(AlpineConvention_lonlat)
data(exrst)
### Define mask
maskR = mask(rst[[1]],shp.lonlat)
### Define quadrature points for 'wsl.ppmGlasso'
quadG2 = wsl.quadrature(mask = maskR,
                        area.win = wind,
                        random = TRUE,
                        nQ = 10000,
                        lasso = TRUE,
                        env_vars = rst)
### Define quadrature points for 'wsl.ppm0'
quadO2 = wsl.quadrature(mask = maskR,
                        area.win = wind,
                        random = TRUE,
                        nQ = 10000,
                        lasso = FALSE,
                        env_vars = NULL)
### Define quadrature test points for 'wsl.ppmGlasso'
test.quadG2=wsl.quads.test(mask=maskR,
                           quadrature=quadG2,
                           nQ=1000,
                           replace=TRUE)
### Define quadrature test points for 'wsl.ppm0'
test.quadG2=wsl.quads.test(mask=maskR,
                           quadrature=quad02,
                           nQ=1000,
                           replace=TRUE)
```

wsl.varKeep

Pre-select set of predictors

Description

Rank spatial predictors "ras" in descending order following their mean predictive power found with wsl.varPPower(), and apply descending successive correlations tests to obtain a pre-selection of predictors. Additionally a Variance Inflation Factor Test (VIF) may also be applied to predictors which succeeded the descending tests to identify the ones that show high mutlicollinearity.

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Usage

```
wsl.varKeep(
    PPower.object,
    ras,
    corTEST = 0.7,
    vifTEST = FALSE,
    corVIF = 0.7,
    ...
)
```

Arguments

PPower.object	'wsl.varPPower' object
ras	Object of class 'RasterLayer' or 'list' of 'RasterLayer'. Same as in wsl.varPPower(). Layers must be of same resolution i.e. excatly the same number of cells
corTEST	Numeric. Threshold of accepted correlation applied in the descending tests
vifTEST	Logical. If TRUE, a VIF test is applied to the set of predictors that succeeded descending correlation tests
corVIF	Numeric. Threshold of accepted correlation within VIF if "vifTEST=TRUE"
	Arguments passed on to the cor() function used for "corTEST"

Value

Object of class list with three elements: 1) ranking of non correlated predictors, 2) ranking of leftovers correlated predictors, 3) vif test results

Author(s)

Yohann Chauvier

Examples

```
### Data Preparation
\mbox{\#}\mbox{Load} environmental rasters and assign random raster Class
data(exrst)
rCLASS = c(rep("Pollution",4),rep("Climate",4),rep("LandCover",4))
# Create a list of rasters out of the rasterBRICK
rasterL = unstack(rst)
# Load my binary observations species data
data(var_select_XYtest.RData)
### wsl.varPPower(): example with a data.frame & rasters not in classes
PPower.DF = wsl.varPPower(points=sp.DF[,c("x","y")],
                          val = sp.DF$myPA,
                          species = sp.DF$spCODES,
                          ras = rasterL,
                          rasCLASS = NULL,
                          mlinear = FALSE,
                           glmMODE = "binomial",
```

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```
weight = sp.DF$myWEIGHT,
                          poly = TRUE,
                          polyBRUT = TRUE,
                          parallel = FALSE,
                          cores = NULL)
### wsl.varPPower(): example with SpatialPointsDataFrame & rasters in classes
PPower.DF = wsl.varPPower(points=sp.DF[,c("x","y")],
                          val = sp.DF$myPA,
                          species = sp.DF$spCODES,
                          ras = rasterL,
                          rasCLASS = NULL,
                          mlinear = FALSE,
                          glmMODE = "binomial",
                          weight = sp.DF$myWEIGHT,
                          poly = TRUE,
                          polyBRUT = TRUE,
                          parallel = FALSE,
                          cores = NULL)
### wsl.varPPower(): example with SpatialPointsDataFrame & rasters in classes
# Checking length of every species input
c(length(mySP),length(myPA),length(spCODES),length(myWEIGHT))
# Running the function
PPower.SPDF = wsl.varPPower(points = mySP,
                            val = myPA,
                            species = spCODES,
                            ras = rasterL,
                            rasCLASS = rCLASS,
                            mlinear = FALSE,
                            glmMODE = "binomial",
                            weight = myWEIGHT,
                            poly = TRUE,
                            polyBRUT = TRUE,
                            parallel = FALSE,
                            cores = NULL)
 ### wsl.varKeep() with a descending correlation of 0.7 with a VIF test at 0.7
# Example with no raster classes
KeepVar = wsl.varKeep(PPower.object = PPower.DF,
                      ras = rasterL,
                      corTEST = 0.7,
                      vifTEST = TRUE,
                      corVIF = 0.7,
                      use="complete.obs")
#Example with raster classes
KeepVar = wsl.varKeep(PPower.object = PPower.SPDF,
                      ras = rasterL,
                      corTEST = 0.7,
                      vifTEST = TRUE,
```

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```
corVIF = 0.7,
use = "complete.obs")
```

wsl.varPPower

Predictive power assessments of predictors

Description

Evaluate the predictive power of single spatial predictors for fitting spatial observations or continous values. Predictors may be classify in classes to separate the ouputs, and data to explain (Y input) may be binary (e.g. presences/absences), discrete (e.g. diversity count) or continuous. Fix and random effects may be used with generalized linear model (GLM) and simple linear model (LM). Therefore, the function currently implement D2.adj::glm(), R2.adj::lm(), pseudo-R2::glmer() and pseudo-R2::lmer(). D2 is calculated via the ecospat package and pseudo-R2 with the MuMIn package for mixed models. Quadratic terms may also be used for more flexible fits, and a parallel argument is available if processing the data is too time consuming.

Usage

```
wsl.varPPower(
  points,
  val,
  species = NULL,
  ras,
  rasCLASS = NULL,
  mlinear = FALSE,
  mixEffect = FALSE,
  mixCat = NULL,
  glmMODE = "binomial",
  weight = 1,
  poly = FALSE,
  polyBRUT = FALSE,
  parallel = FALSE,
  parINFOS = NULL,
  cores = detectCores()/2,
)
```

Arguments

points	Numeric. Object of class 'matrix' or 'data frame' with two columns name	1
	1 10 10 1 11 1 10 1	

and "y", or 'SpatialPoints', or 'list' of 'SpatialPoints'. "points" should be of

same length as "val", "species" and "weight"

Numeric. Object of class 'vector' if "points" is of class 'matrix' or 'data.frame',

otherwise object of class 'list'. Binary observations if "glmMODE="binomial", discrete if "glmMODE="poisson", continious if "mlinear=TRUE". Must be of

same length as "points", "species" and "weight"

species Character. Object of class 'vector' specifying the ID of your observations. No mandatory use except when "points" is a 'list' of 'SpatialPoints'. If used, must

be of same length as "points", "val" and "weight"

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ras	Object of class 'RasterLayer' or 'list' of 'RasterLayer'. Layers must be of exact same resolution and extent i.e. exactly the same number of cells
rasCLASS	Object of class 'vector' to associate IDs to the raster(s) if "ras" is a 'list' of 'RasterLayer'
mlinear	'TRUE' run lm(), 'FALSE' run glm()
mixEffect	Default is FALSE. If 'TRUE' run mLinear with random effect (i.e. mix LM or GLM)
mixCat	Character. Object of class 'vector' if "points" is of class 'matrix' or 'data.frame', otherwise object of class 'list'. Use to assign categories to your observations. Those are used for potential random effects. Must be of same length as "points", "val" and "species"
glmMODE	glm() link function if "mlinear=FALSE" (defaut::"binomial" or "poisson")
weight	Numeric. Object of class 'vector' if "points" is of class 'matrix' or 'data.frame', otherwise object of class 'list'. Used for weighting your observations when running glm() or lm(). Must be of same length as "points", "val" and "species"
poly	If TRUE, glm() or lm() uses a polynomial quadratic transformation on "ras"
polyBRUT	If TRUE, force "poly" parameter in cases NAs are found in "ras"
parallel	If TRUE, parallelisation is active
parINFOS	Path. Create a txt file to write parallelisation infos if "parallel=TRUE".
cores	Number of cores if "parallel=TRUE"
	Arguments passed on to the lm() or glm() function

Value

'wsl.varPPower' object with n slots corresponding to n "rasCLASS". In each slots: matrix of nrow::c("species" + mean + standard deviation) & ncol::"ras". Where NAs occur, models could not correctly converged.

Author(s)

Yohann Chauvier

Examples

```
### Data Preparation

rm(list = setdiff(ls(), lsf.str()))

# Load environmental rasters and assign random raster Class
data(exrst)
rCLASS = c(rep("Pollution",4),rep("Climate",4),rep("LandCover",4))

# Create a list of rasters out of the rasterBRICK
rasterL = unstack(rst)

# Load my binary observations species data
data(var_select_XYtest)

# Create a category vector
mixV = sample(LETTERS[1:3], 35743, replace=TRUE)
```

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```
### wsl.varPPower(): example with a data.frame & rasters not in classes
PPower.DF = wsl.varPPower(points=sp.DF[,c("x","y")],
                      val = sp.DF$myPA,
                      species = sp.DF$spCODES,
                      ras = rasterL,
                      rasCLASS = NULL,
                      mlinear = FALSE.
                      glmMODE = "binomial",
                      weight = sp.DF$myWEIGHT,
                      poly = TRUE,
                      polyBRUT = TRUE,
                      parallel = FALSE,
                      cores = NULL)
### wsl.varPPower(): Same example including random effect
PPower.DF = wsl.varPPower(points = sp.DF[,c("x","y")],
                      val = sp.DF$myPA,
                      species = sp.DF$spCODES,
                      ras = rasterL,
                      rasCLASS = NULL,
                      mlinear = FALSE,
                      mixEffect = TRUE,
                      mixCat = mixV,
                      glmMODE = "binomial",
                      weight = sp.DF$myWEIGHT,
                      poly = TRUE,
                      polyBRUT = FALSE,
                      parallel = FALSE,
                      cores = NULL)
### wsl.varPPower(): example with a data.frame & rasters in classes
PPower.DF = wsl.varPPower(points=sp.DF[,c("x","y")],
                        val = sp.DF$myPA,
                        species = sp.DF$spCODES,
                        ras = rasterL,
                        rasCLASS = rCLASS,
                        mlinear = FALSE,
                        glmMODE = "binomial",
                        weight = sp.DF$myWEIGHT,
                        poly = TRUE,
                        polyBRUT = TRUE,
                        parallel = FALSE,
                        cores = NULL)
### wsl.varPPower(): Same example including random effect
PPower.DF = wsl.varPPower(points=sp.DF[,c("x","y")],
                        val = sp.DF$myPA,
                        species = sp.DF$spCODES,
                        ras = rasterL,
                        rasCLASS = rCLASS,
                        mlinear = FALSE,
                        mixEffect = TRUE,
                        mixCat = mixV,
                        glmMODE = "binomial",
                        weight = sp.DF$myWEIGHT,
                        poly = TRUE,
```

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```
polyBRUT = TRUE,
                        parallel = FALSE,
                        cores = NULL)
### wsl.varPPower(): example with SpatialPointsDataFrame & rasters in classes
# Checking length of every species input
c(length(mySP),length(myPA),length(spCODES),length(myWEIGHT))
# Generate new random effect classes
mixL = lapply(sapply(mySP,length),function(x) sample(LETTERS[1:3],x, replace=TRUE))
# Running the function
PPower.SPDF = wsl.varPPower(points = mySP[[1]],
                          val = myPA[[1]],
                          species = NULL,
                          ras = rasterL,
                          rasCLASS = rCLASS,
                          mlinear = FALSE,
                          mixEffect=TRUE,
                          mixCat=mixL[[1]],
                          glmMODE = "binomial"
                          weight = myWEIGHT[[1]],
                          poly = TRUE,
                          polyBRUT = TRUE,
                          parallel = FALSE,
                          cores = NULL)
```

wsl_doi

Get a custom DOI for a GBIF filtered dataset

Description

A small user friendly wrapper of the derived_dataset() function of the rgbif R package, compatible with one or several wsl_gbif() outputs.

Usage

```
wsl_doi(
  wsl_gbif = NULL,
  title = NULL,
  description = NULL,
  source_url = "https://example.com/",
  user = "",
  pwd = "",
  ...
)
```

Arguments

wsl_gbif data.frame or list. One wsl_gbif() output or a list of several.title The title for your derived dataset.

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source_url A link to where the dataset is stored.

user Your GBIF username. pwd Your GBIF password.

. . . Additional parameters for derived_dataset() in rgbif.

description A description of the dataset.

Details

see derived_dataset() function from the rgbif R package

Value

One citable DOI and its information.

References

Chamberlain, S., Oldoni, D., & Waller, J. (2022). rgbif: interface to the global biodiversity information facility API. 10.5281/zenodo.6023735

See Also

The rgbif package for additional and more general approaches to get GBIF DOI

Examples

wsl_gbif Massively download and filter GBIF observations for sound spatial analyses

Description

Implement an user-friendly workflow to download and clean gbif taxa observations. The function uses the rgbif R package but (1) implements the same search result found if www.gbif.org is employed i.e., based on the input taxa name, all species records related to its accepted name and synonyms are extracted. The function also (2) bypasses rgbif hard limit on the number of records (100'000 max). For this purpose, a dynamic moving window is created and used across the geographic extent defined by the user. This window automatically fragments the specified study area in succesive tiles of different sizes, until all tiles include < 100'000 observations. The function

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also (3) automatically applies a post-filtering of observations based on the chosen resolution of the study/analysis and by partly employing the CoordinateCleaner R package. Filtering options may be chosen and involve several choices: study's extent, removal of duplicates, removal of absences, basis of records selection, removal of invalid/uncertain xy coordinates (WGS84), time period selection and removal of raster centroids. By default, the argument hasGeospatialIssue in occ_search() (implemented rgbif function) is set to FALSE.

Usage

```
wsl_gbif(
  sp_name = NULL,
  conf_match = 90,
  geo = NULL,
  grain = 1000,
  duplicates = FALSE,
  absences = FALSE,
  no_xy = FALSE,
  basis = c("OBSERVATION", "HUMAN_OBSERVATION", "MACHINE_OBSERVATION",
  "MATERIAL_SAMPLE", "PRESERVED_SPECIMEN", "FOSSIL_SPECIMEN", "LIVING_SPECIMEN",
    "LITERATURE", "UNKNOWN"),
  add_infos = NULL,
  time_period = c(1000, 3000),
  identic_xy = FALSE,
  wConverted_xy = FALSE,
  centroids = FALSE,
  ntries = 10.
  error.skip = TRUE,
)
```

Arguments

Summer		
	sp_name	Character. Scientific name to run an online search (i.e. with GBIF-API) for species observations. Works also for genus and higher taxa levels.
	conf_match	Numeric from 0 to 100. Determine the confidence threshold of match of 'sp_name' with the GBIF backbone taxonomy. Default is 90.
	geo	Object of class 'Extent', 'SpatExtent', 'SpatialPolygon', 'SpatialPolygonDataframe', or 'SpaVector' (WGS84) to define the study's area extent. Default is NULL i.e. the whole globe.
	grain	Numeric. Specify in meters the study resolution. Used to filter gbif records (x2) according to their uncertainties and number of coordinate decimals. Records with no information on coordinate uncertainties (column 'coordinateUncertaintyInMeters') are be kept by default. See details.
	duplicates	Logical. Should duplicated records be kept?
	absences	Logical. Should absence records be kept?
	no_xy	Logical. Default is FALSE i.e. only records with coordinates are downloaded. If TRUE, only records with no coordinates are downloaded.
	basis	Character. Which basis of records should be selected? Default is all i.e. c("OBSERVATION", "HUMAN_OBSERVATION", "MACHINE_OBSERVATION", "MATERIAL_SAMPLE", "PRESERVED_SPECIMEN", "FOSSIL_SPECIMEN", "LIVING_SPECIMEN",

data-use/en/basis-of-record.html

"LITERATURE", "UNKNOWN"). Description may be found here: https://docs.gbif.org/course-

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add_infos Character. Infos that may be added to the default output information. List of IDs may be found at: https://www.gbif.org/developer/occurrence. Default IDs contain 'taxonKey', 'scientificName', 'acceptedTaxonKey', 'acceptedScientificName', 'individualCount', 'decimalLatitude', 'decimalLongitude', 'basisOfRecord', 'coordinateUncertaintyInMeters', 'country', 'year', 'datasetKey', 'institutionCode', 'publishingOrgKey', 'taxonomicStatus' and 'taxonRank'. time_period Numerical vector. Observations will be downloaded according to the chosen year range. Default is c(1000,3000). Observations with year = NA are kept by default. Logical. Should records with identical xy be kept? identic_xy Logical. Should incorrectly lon/lat converted xy be kept? Uses 'cd_ddmm' from wConverted_xy 'CoordinateCleaner' R package. centroids Logical. Should species records from raster centroids be kept? Uses 'cd_round' from 'CoordinateCleaner' R package. Numeric. In case of failure from GBIF server or within the rgbif package, how ntries many download attempts should the function request? Default is '10' with a 2 seconds interval between tries. If attempts failed, an empty data.frame is return by default. Logical. Should the search process continues if ntries failed? error.skip Additional parameters for the function cd_round() of CoordinateCleaner.

Details

Argument 'grain' used for two distinct gbif records filtering. (1) Records filtering according to gbif 'coordinateUncertaintyInMeters'; every records uncertainty > grain/2 are removed. Note: Records with no information on coordinate uncertainties are kept by default. (2) Records filtering according to the number of longitude/latitude decimals; if 110km < grain <= 11km, lon/lat with >= 1 decimal are kept, if 11km < grain <= 1100m, lon/lat with >= 2 decimals kept; if 1100m < grain <= 110m, lon/lat with >= 4 decimals are kept; if 11m < grain <= 1.1m, lon/lat with >= 5 decimals are kept etc...

Value

Object of class data.frame with requested GBIF information. Although the function works accurately, error outputs might still occur depending on the 'sp_name' used. Therefore, default information detailed in 'add_infos' is stored so that sanity checks may still be applied afterwards. Although crucial preliminary checks of species records are done by the function, addtional post exploration with the CoordinateCleaner R package is still highly recommended.

References

Chauvier, Y., Thuiller, W., Brun, P., Lavergne, S., Descombes, P., Karger, D. N., ... & Zimmermann, N. E. (2021). Influence of climate, soil, and land cover on plant species distribution in the European Alps. Ecological monographs, 91(2), e01433. 10.1002/ecm.1433

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Zizka, A., Silvestro, D., Andermann, T., Azevedo, J., Duarte Ritter, C., Edler, D., ... & Antonelli, A. (2019). CoordinateCleaner: Standardized cleaning of occurrence records from biological collection databases. Methods in Ecology and Evolution, 10(5), 744-751. 10.1111/2041-210X.13152

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See Also

The (1) rgbif and (2) CoordinateCelaner packages for additional and more general approaches on (1) downloading GBIF observations and (2) post-filetering those.

Examples

```
# Load maptools for the map world
library(maptools)
data(wrld_simpl)
# Load the Alps Extend
data(geo_dat)
# Downloading worldwide the observations of Panthera tigris
test1 = wsl_gbif("Panthera tigris",basis=c("OBSERVATION","HUMAN_OBSERVATION"))
plot(wrld_simpl)
points(test1[,c("decimalLongitude","decimalLatitude")],pch=20,col="#238b4550",cex=4)
# Downloading in the Alps the observations of Cypripedium calceolus (with a 100m grain and
# by adding the 'issues' column)
test2 = wsl_gbif("Cypripedium calceolus", geo = shp.lonlat, grain = 100, add_infos = c("issue"))
plot(shp.lonlat)
points(test2[,c("decimalLongitude","decimalLatitude")],pch=20,col="#238b4550",cex=1)
# Downloading worlwide the observations of Ailuropoda melanoleuca (with a 100km grain, after 1990
# and by keeping duplicates and by adding the name of the person who collected the panda records)
test3 = wsl_gbif("Ailuropoda melanoleuca", grain = 100000 , duplicates = TRUE,
    time\_period = c(1990,3000), add\_infos = c("recordedBy","issue"))
plot(wrld_simpl)
points(test3[,c("decimalLongitude","decimalLatitude")],pch=20,col="#238b4550",cex=4)
# Downloading worlwide the observations of Phascolarctos cinereus (with a 1km grain, after 1980,
# and keeping raster centroids)
test4 = wsl_gbif("Phascolarctos cinereus", grain = 1000,
    time\_period = c(1990,3000), centroids = TRUE)
```

wsl_obs_filter

Filter a set of GBIF observations according to a defined grain

Description

Whereas the 'grain' parameter in wsl_gbif() allows GBIF observations to be filtered according to a certain spatial precision, wsl_obs_filter() accepts as input a wsl_gbif() output (one or several species) and filter the observations according to a specific given grid resolution (one observation per pixel grid kept). This function allows the user to refine the density of GBIF observations according to a defined analysis/study's resolution.

Usage

```
wsl_obs_filter(wsl_gbif, grid)
```

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Arguments

wsl_gbif one wsl_gbif() output including one or several species. Note that if GBIF ab-

sences are kept in the output(s), the function should be used distinctively for

observations and absences.

grid Object of class 'SpatRaster', 'RasterLayer', 'RasterBrick' or 'RasterStack' of

desired resolution and extent (WGS84)

Value

a data frame with two columns named 'x' and 'y' comprising the new set of observations filtered at grid resolution.

Examples

```
# Load the European Alps extent and a raster of a random resolution
data(geo_dat)
data(exrst)
# Downloading in the European Alps the observations of two plant species
obs.arcto = wsl_gbif("Arctostaphylos alpinus",geo=shp.lonlat)
obs.saxi = wsl_gbif("Saxifraga cernua",geo=shp.lonlat)
plot(vect(shp.lonlat))
points (obs.arcto[,c("decimalLongitude","decimalLatitude")], pch=20,col="#238b4550",cex=1)\\
points(obs.saxi[,c("decimalLongitude","decimalLatitude")],pch=20,col="#99000d50",cex=1)
# rbind both datasets
both.sp = rbind(obs.arcto,obs.saxi)
# Run function
obs.filt = wsl_obs_filter(both.sp,rst)
# Check new points
x11();plot(vect(shp.lonlat))
points(obs.filt[obs.filt$Species%in%"Arctostaphylos alpinus",c("x","y")],pch=20,col="#238b4550",cex=1)
points(obs.filt[obs.filt$Species%in%"Saxifraga cernua",c("x","y")],pch=20,col="#99000d50",cex=1)
```

wsl_taxonomy

Retrieve from GBIF all scientific names of a specific Taxa

Description

Generates, based on a given species name, a list of all its scientific names (accepted, synonyms) found in the GBIF backbone taxonomy to download the data. Children and related doubtful names not use to download the data may also be extracted.

Usage

```
wsl_taxonomy(sp_name = NULL, conf_match = 90, all = FALSE)
```

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Arguments

sp_name	Character. Species name from which the user wants to retrieve all existing GBIF names
conf_match	Numeric. From 0 to 100. Determine the confidence threshold of match of 'sp_name' with the GBIF backbone taxonomy. Default is 90.
all	Logical. Default is FALSE. Should all species names be retrieved or only the accepted name and its synonyms?

Value

A data.frame with two columns: (1) Names and (2) Backbone Taxonomy Status

References

Chamberlain, S., Oldoni, D., & Waller, J. (2022). rgbif: interface to the global biodiversity information facility API. 10.5281/zenodo.6023735

See Also

The rgbif package for additional and more general approaches on how to retrieve scientific names from the GBIF backbone taxonomy.

Examples

```
wsl_taxonomy("Cypripedium calceolus",all=FALSE)
wsl_taxonomy("Cypripedium calceolus",all=TRUE)
```

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