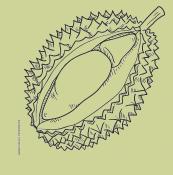
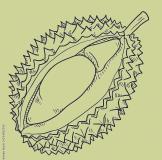
# The draft genome of tropical fruit durian (Durio zibethinus)





Genome Analysis 2023
Paper IV
Andreas Bergfeldt

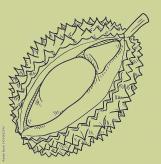
# What is a Durian and why do this assembly?

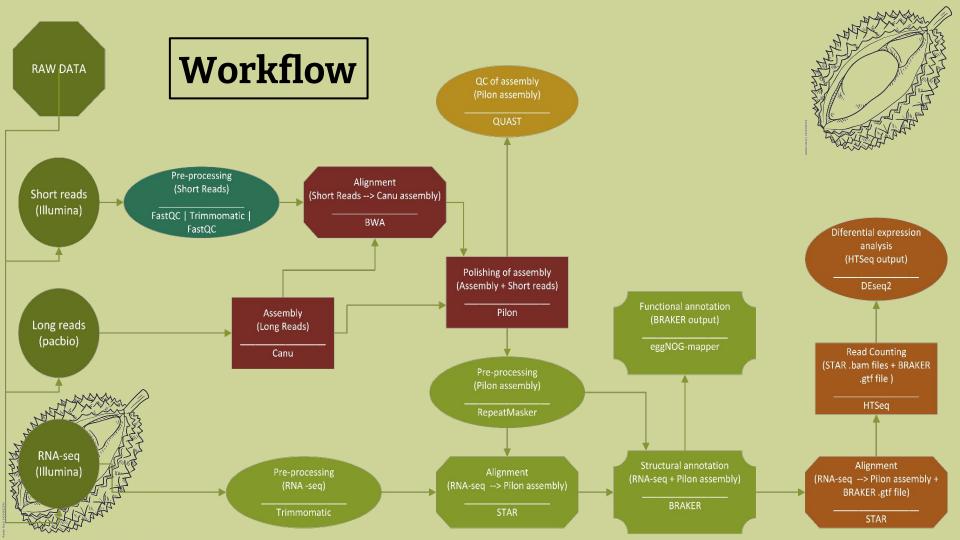
- What?
  - Fruit popular in southeast asia
  - Pungent odor, not allowed in some public spaces

- Why?
  - o 2016 China imported 600 mUSD
  - Almost non existent genetic research

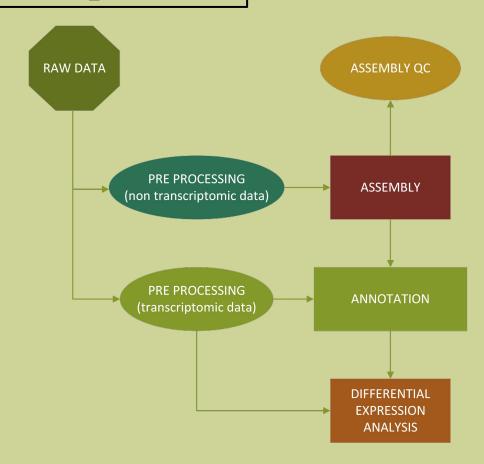
### My assembly

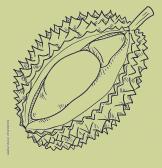
- One scaffold
  - Scaffold 10
- Too much to do whole genome
  - Computing power
  - o Time





# Workflow - simplified

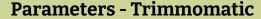




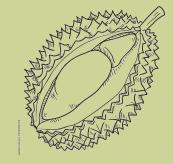
# Pre-processing - Trimmomatic

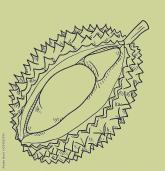
- Pre-trimmed data
  - Trimmed again to make sure it is good
- Trimming to remove adapters and low quality bases





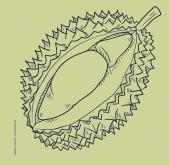
- ILLIUMINACLIP = TruSeq3-SE:2:30:10
- LEADING = 3
- TRAILING = 3
- SLIDINGWINDOW= 4:15
- MINLEN = 36



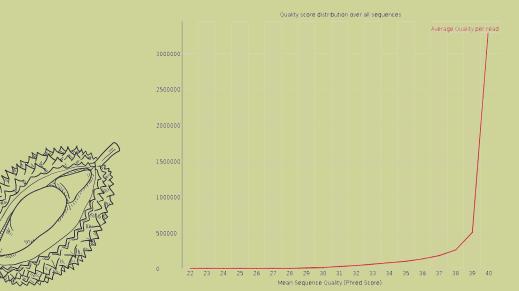


# Pre-processing - FastQC

Important to check data before proceeding!



Quality deemed good enough



Pictures are from after trimming





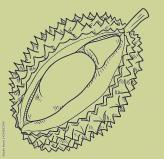
### Assembly - Canu

Assembly of the long reads



o corrMaxEvidanceErate parameter

• useGrid for running in UPPMAX



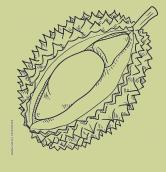
### Parameters - Canu

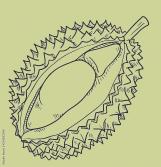
- useGrid = false
- genomeSize = 30m
- corMaxEvidenceErate = 0.15



### Assembly - BWA ( + samtools)

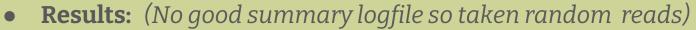
- BWA
- Mapping short reads to long reads
  - Necessary for polishing later
- Manipulating BWA output with samtools to get .bam file
  - $\circ$  .fasta  $\rightarrow$  .sai  $\rightarrow$  .sam  $\rightarrow$  .bam



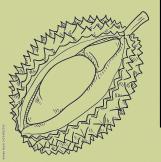


### Assembly - Pilon

- Pilon
- Polishing assembly with short read data
  - Higher accuracy to long scaffolds



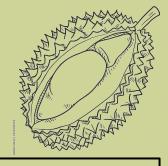
- o Total Reads: 18201
- Confirmed 177014 of 192342 bases (92.03%)
- Corrected 11 snps; 0 ambiguous bases
   corrected 96 small insertions totaling 102 bases
   41 small deletions totaling 42 bases



# Assembly QC - QUAST

• Was the assembly any good?



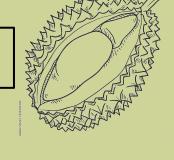


	Statistics without reference	durian_pilon
pilon	# contigs	608
	# contigs (>= 0 bp)	608
	# contigs (>= 1000 bp)	608
	# contigs (>= 5000 bp)	551
	# contigs (>= 10000 bp)	433
	# contigs (>= 25000 bp)	199
	# contigs (>= 50000 bp)	114
	Largest contig	826 217
	Total length	25 217 558
	Total length (>= 0 bp)	25 217 558
	Total length (>= 1000 bp)	25 217 558
	Total length (>= 5000 bp)	25 033 298
	Total length (>= 10000 bp)	24 133 282
	Total length (>= 25000 bp)	20 443 885
	Total length (>= 50000 bp)	17 656 612
	N50	120 097
	N90	14 884
	auN	198 308
	L50	48
	L90	318
	GC (%)	31.45
	Mismatches	
	# N's per 100 kbp	0
	# N's	0
ı		

### Annotation - Trimmomatic +

### repeat Maske Before annotation

• Trimmomatic → same as before

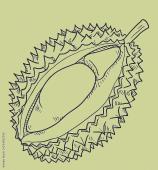


### RepeatMasker

- softmasking repeats for better annotation
- Important to specify softmasking

### Softmasking

- Identifies and "masks" repeats
- Changes bases in the fasta file from uppercase to lowercase
- Hardmasking removes info, softmasking does not



### Structural annotation -

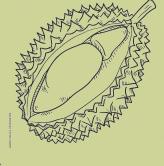
### BRAIKER consisting of Augustus and GeneMark

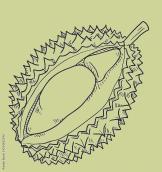
 Annotates based on reference genome (masked assembly), and transcriptomic data



### **BRAKER** results

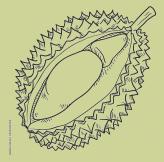
- Hardmasked scaffold → 96 genes identified
- Softmasked scaffold  $\rightarrow$  110 genes identified
- What are the genes?
  - o who knows...
    - will come later





# Functional annotation - eggNOG-mapper

Web based UI

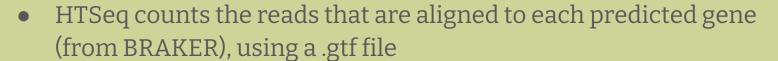


Loads of information about found hits

- Does not say a lot at this stage
  - Vital for differential expression analysis
    - Yes more patience is needed...

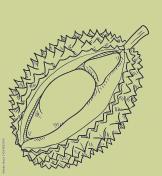
### Counting reads - STAR and HTSeq

- For transcriptomic reads to be counted they first need to be aligned
  - Done with STAR



### HTSeq results

- 8 files with reads of varying length
- Total 29 269 185 record pairs
- 835 record pairs with missing mate record
  - $\circ$  0.003% of total record pairs



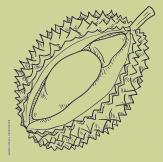


# Differential expression analysis

• R-module for expression analysis

Finally some tangible results!



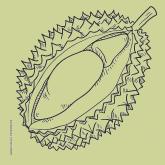


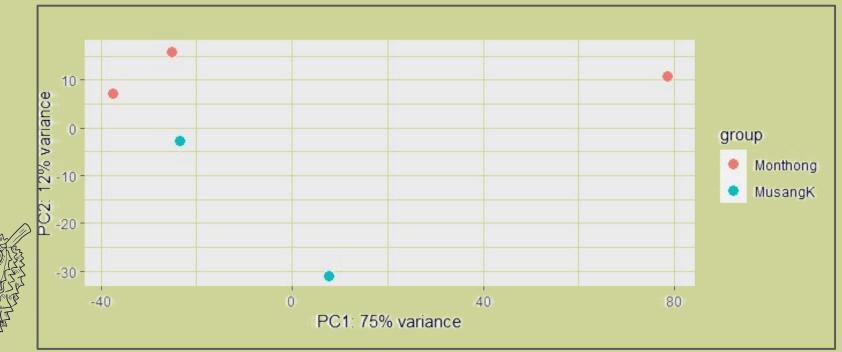
### The analysis in short

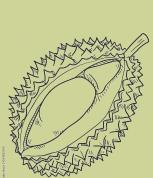
(In depth on future slides)

- 2 different species
  - Musang King and Mothong
- 2 different type of analysis
  - Between species (Musang king, Mothong)
  - Within species (Musang King)
    - Different plant organs (aril, stem, leaf, root)
- Types of visualization
  - PCA
  - Heatmap

### DE - Between species - PCA



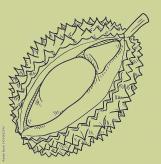




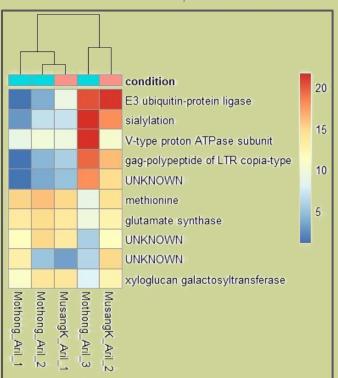
### DE - Between species - Heatmap

- Overall hard to separate the two species
- On PCA the grouping is not good
  - o Groups Musang with Mothong
  - O Does not group all of same species

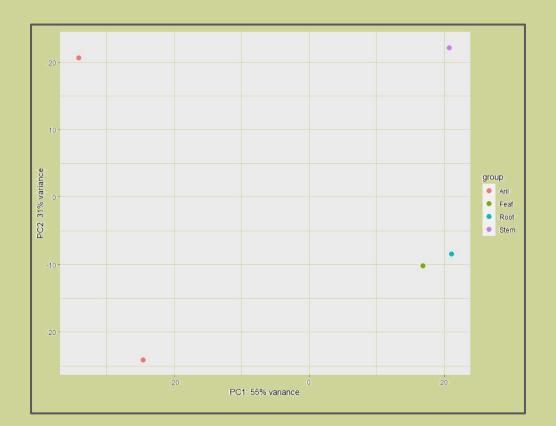
 The heatmap does not show any patterns in which the species can be separated

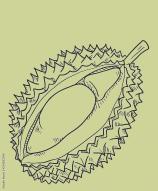






# DE - Within species - PCA



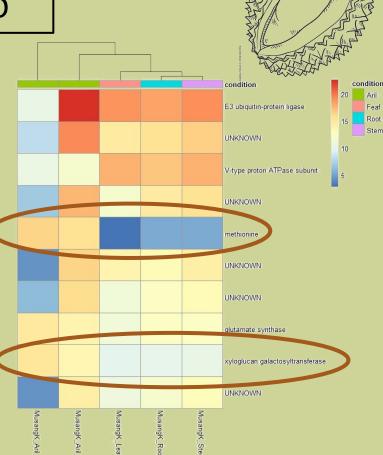




### DE - Within species - Heatmap

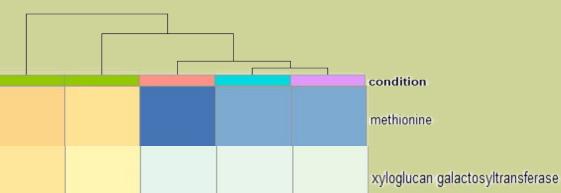
- Better separation between plant organs compared to plant species
  - PCA did okay
  - Possible separations at least

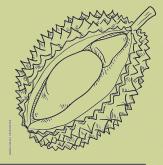
- Some patterns can be seen that separates arils from the rest of the plant organs
  - Eg. methionine and xyloglucan



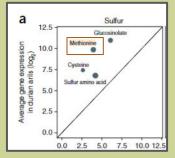
### DE - Biological conclusions

- Methionine converted to methanethiol
   with the enzyme methionine γ-lyase
  - o Methanethiol has key role in odor of durian
- Xyloglucan building block of cell wall 2
  - "interlace cellulose microfibrils in most flowering plants."
  - Genes with association to cell wall 2 upregulated in arils

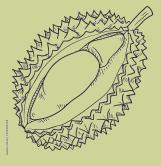




### Figures from paper IV







# Thank you for your attention!

For more in depth information about programs etc. please look at my github wiki for this project: <a href="https://github.com/A-Bergfeldt">https://github.com/A-Bergfeldt</a>

