

DOCTORAL THESIS

Developments to established dose-finding methodologies for application in trials with complex and innovative designs

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Abstract

Developments to established dose-finding methodologies for application in trials with complex and innovative designs

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The results and decisions made in early phase clinical trials play an important role in the drug development process. Failure to make correct or accurate decisions may result in further unwarranted clinical research and may also cause potential future harm to patients.

This thesis explores methodologies used in dose-finding trials, specifically looking at the implementation of these methods and how they can be extended to answer different questions and assist with decision-making.

Firstly we detail our experiences implementing a novel methodology for a dose-finding trial where monotonic ordering is not possible. We then present an extension to a seamless adaptive phase I/II trial design which aims to conduct dose-finding and replace the need for a randomised phase II trial.

We also present two extensions to dose transition pathways, a visualisation tool to aid designing and decision making in early phase trials. The first looks at how they can be applied to more complex time-to-event methodologies and the second looks at how the same concept can be applied to single-arm phase II trials.

Throughout this thesis we include exemplar trials and showcase our methods being motivated or implemented into clinical trials conducted by the Cancer Research UK Clinical Trials Unit at the University of Birmingham.

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Chapter 1

Introduction

1.1 Aims of this thesis

Early phase clinical trials are essential in the drug development process as they provide key information about new interventions which can be used in later-phase testing. Therefore, it is important that the decisions made and conclusions reached in the early phase setting are correct or as accurate as possible. Failure to do so could lead to a waste of resources in pursuing unnecessary further research and could potentially negatively impact patients well-being. This thesis aims to explore methodologies used in early phase dose-finding trials. We look specifically at the implementation of novel methods as well as how they can be extended for use in complex and innovative designs.

In this chapter, we begin with a brief introduction of clinical trials and also provide a summary of the underlying methodology used within this thesis specific to early phase trials. A description of each chapter is then provided.

1.2 Clinical trials

Clinical trials are often a time-consuming and costly process [1]. It can take 10-20 years to get a new drug from inception to regulatory approval [2], [3].

Any new treatments that come to market must be thoroughly tested and examined, to make sure not only that they are safe but also effective and better than treatments currently in use.

The clinical trial process is split into multiple stages, commonly referred to as phases. Each phase has a different objective and builds upon knowledge and data collected in the previous phases [4]. Phase I trials aim to determine the safety of a treatment. This typically takes the form of a dose-finding study that aims to find a safe and tolerable dose that can be taken forward for further testing [5]. Phase II trials aim to determine if a treatment works or if there is some signal of efficacy. This is usually done with a single arm design i.e. a sample of patients is given the experimental treatment [6]. However, some Phase II trials can be RCTs (Randomised Controlled Trials), where the new treatment is compared with a treatment already in use or a placebo [7]. Finally, Phase III trials aim to establish the efficaciousness of an experimental treatment. Normally, this is done by comparing the new treatment against the standard of care. Results from Phase III clinical trials can then go on to influence clinical practice [8].

We usually consider Phase I and single arm Phase II trials to be early phase trials, then randomized Phase II and Phase III trials as late phase trials. However, there is no definitive boundary between early and late phase trials. Phases are determined by their intention and not by their design.

1.3 Introduction to early phase trials

For Phase I trials the main aim is to establish a dose, commonly referred to as the maximum tolerated dose (MTD), which can then be taken forward into later phase testing. Normally these may be considered first-in-human trials as Phase I is typically the point at which a drug would first be tested in healthy

human volunteers. However, in the oncology setting this is generally not the case. Often times, due to the nature of the treatments, such as chemotherapy or radiotherapy, they may be considered too toxic to give to healthy individuals so are rather tested in patients with the specific disease of interest [9].

Traditionally these trials adopted algorithm-based approaches. Here pre-determined rules were used during the trial to allocate patients to dose-levels and select the MTD. An example of this would be the 3+3 design [10], where patients were recruited in cohorts of three and dependent on the outcomes observed in each cohort a specific decision on the next investigative dose would be made. There were many criticisms of these approaches as they resulted in the sub-optimal treatment of patients, poor operating characteristics and a recommended dose or MTD that has limited interpretation as a dose yielding a specific target toxicity [11], [12].

This gave rise to the continual reassessment method (CRM) which was first introduced by O'Quigley et al. [13] in 1990. This methodology was developed as an approach to meet ethical requirements and use models to reasonably approximate the true probability of toxicity around the dose close to some pre-defined target toxicity. In their paper, O'Quigley et al. [13] demonstrate the CRM's superiority over various algorithm based designs via simulations. The main advantage of the CRM is that it is able to make use of all accumulated data whereas designs such as the 3+3 make decisions and recommendations based on data from the most recent cohort of patients. In the case of the 3+3 design, escalation continues until at least two patients in a cohort of three or six experience a DLT. More explicitly, the MTD is the dose level below the dose at which $\geq 33\%$ of patients experience a DLT [14].

With the CRM debuting over 30 years ago in the literature and multiple papers over the subsequent years confirming its advantages over rule-based designs you would expect model-based approaches for dose-finding trials to

become the norm however, this is not the case. A study by Rogatko et al. [15] published in 2007 looked into the translation of effective statistical designs into phase I trials for new anticancer therapies. Between 1991 and 2006 they searched for abstracts and categorised them as either clinical dose-finding trials or statistical methodology for dose-escalation trials. They found 1235 clinical trials and 90 methodological papers. Of those 1235 trials only 20 (1.6%) used statistical methodology, the remaining papers used various rule-based designs. A later review by Chiuzan et al. [16] looked at the number of phase I oncology articles published between 2008 and 2014. Out of the 1712 dose-finding trials 1591 (92.9%) used rule-based designs.

Based on these reviews we can see that the uptake of more efficient model-based designs such as the CRM has been slow and limited. There are probably a number of factors which cause this, such as lack of resources, access and understanding. There exists some guidance in the literature by Wages et al. [17] and Wheeler et al. [18] as well as practical examples from Mozgunov et al.[19]. The main issue is that implementation of these designs usually require the input of a statistician, more specifically one who is familiar with such approaches. They also need to be able to implement and conduct the trial with software available, for designs such as the CRM there are multiple options available such as the R packages dfcrm [20], escalation [21] and trialr [22]. However, for more complex and innovative designs, software may not be readily accessible and implementation may be difficult.

Phase II trials build upon the work of early Phase I trials. Here the focus shifts away from toxicity and looks more towards efficacy of these new treatments at the dose-levels previously determined in phase I trials [23]. The key purpose of phase II trials is to see if a new treatment or intervention works and establish if there is an efficacy signal. More specifically they aim to determine if there is a sufficient level of efficacy to warrant further research in for example

a Phase III setting [24].

An example of an early Phase II trial design would be a single arm trial. Eligible patients come into the trial and all of them will be allocated to the new treatment. Once they have completed their treatment period we would then assess the effectiveness of that treatment using some measure of success. Looking at the outcome of success in each patient, the success rate or proportion of success can then be determined. In the single arm setting, this success rate is then compared to some benchmark, which is determined from either historical data or clinical experience.

One approach to analysing a trial like this is Bayesian and utilises a Beta-Binomial conjugate analysis to estimate a response rate for a binary outcome. This is a fairly straight-forward analysis and can be found in most Bayesian text books, one example of which is by Lee [25]. Whilst this may be somewhat simplistic from a mathematical standpoint, clinicians may be less familiar with Bayesian approaches in general compared to frequentist methods. However, there is still value in using these methods to analyse trials as they can often be more efficient especially with smaller sizes [26], which is often the case in an early phase setting and they allow for greater flexibility in terms of practical considerations such as cohort sizes. Using a Bayesian approach allows for decisions to be made based on probabilities from a posterior distribution [27]. Which can be a more intuitive way to understand the treatment effect given observed data.

These types of analyses also better facilitate other complex and innovative designs such as platform/basket/umbrella trials where multiple biomarkers or treatments may be under investigation. The Bayesian approach will allow for information borrowing across any different arms in the trial, which is particularly useful when working with restricted resources [28].

Generally speaking, early phase trials work with less resources (i.e less patients, time, money), so it would be advantageous to use designs which are more efficient with the data collected, the majority of which would require a statistician familiar with these methods to implement. Clinicians may also push for designs that are easier to follow so either a algorithm-based approach in a Phase I or a frequentist approach for a Phase II trial. Clinicians may also not be familiar with these complex designs and how they work.

As developments occur in the medical field, e.g. introducing new interventions, trials may subsequently become more complex. This complexity arises because the assumptions made by current methodologies may no longer hold true. For example molecular targeted agents and certain biological therapies may not operate under the assumption that as dose increases efficacy increases. Additionally, the questions that trials aim to answer can also become more intricate due to various factors, including the disease setting under investigation and the type of treatment involved. Consequently, it is essential to address these challenges as they arise to ensure the optimal utilisation of our data and to make accurate decisions.

Whilst the development of new methodologies is necessary to facilitate better research, practical considerations also need to be made. The ease at which a new methodology can be implemented is important. Whilst a new method may be fantastic and solve a lot of issues, if researchers cannot easily implement it either through software or replicable code then the method will ultimately have little impact in the real world. Similarly, communication of new methods is also important. A summary of each chapter is provided in the next section, where we address different aspects of this further.

1.4 Chapters in this thesis

In Chapter 2, we detail our experiences implementing a novel methodology into the design of an early phase dose-finding trial in head and neck cancer. This trial, ADePT-DDR run by the University of Birmingham (UoB) Cancer Research Clinical UK Trials Unit (CRCTU), uses the partial ordering time-to-event continual reassessment method (PO-TITE-CRM) design [29]. The PO-TITE-CRM design was introduced in 2013 as an extension to the TITE-CRM design, itself an extension of the original continual reassessment method (CRM), a model-based approach to dose-finding trials. Despite the publication of this novel dose-escalation design its implementation appears to be rare, as highlighted in the reviews by Rogatko et al. [15] and Chiuzan et al. [16]. The CRM operates under the monotonicity assumption which assumes that as the dosage of drug increases so does the probability of toxicity. The PO-TITE-CRM was developed to address scenarios where the ability to fully order the doses based on increasing toxicity is missing. Multiple iterations of simulations were utilised to determine the optimal parameterisation of the design. Simulation results from the optimal parameterisation show the operating characteristics of the design perform well across a variety of scenarios. We present an overview of the design methodology and its application in this trial scenario.

The focus of Chapter 3 is on an extension to a seamless Phase I/II design. This design by Wages and Tait [30] uses adaptive randomisation to conduct its dose-finding. We aimed to extend this design to include a control comparator arm by leveraging the design's adaptive randomisation mechanism. Typically these types of seamless design utilise both toxicity and efficacy outcomes to select a dose for later phase testing. Our motivation was to take this one step further and develop a seamless Phase I/II design that would allow for a direct comparison of the selected dose to a control arm. We present the modification

we make to the design and detail how altering specific parameters impacts the allocation of patients to the control arm within the design. Simulations were then conducted to investigate the operating characteristics under certain specifications. Further simulation work is then used to compare our modified design with the original Wages and Tait design to ascertain if performance is impacted by our modification. Additionally, this control arm allows us to conduct power calculations comparing efficacy rates between patients allocated to the optimal dose and those in the control arm. These calculations give an insight into how our design would perform as a standard Phase II trial.

Decision-making is an important component of dose-finding trials. For non-statisticians, understanding the reason why certain decisions are being made from a model-based approach such as the CRM could be confusing or unintuitive compared to a 3+3 for example. In order to bridge that gap, Yap et al. developed a novel visualisation tool called dose transition pathways (DTPs) [31]. Our work in Chapter 4, explores the use of DTPs in a time-to-event (TITE) setting. TITE methodology, in a dose-finding context, allows decisions to be made at earlier time points based on partially observed data from patients currently in a trial. This leads to the number of outcomes being more complicated for a single cohort of patients in a dose-finding trial. This in turn further complicates DTPs which aim to effectively summarise the different dose-decisions that can be made based on all possible outcomes. Our work aims to reap the benefits of DTPs in a TITE setting. We detail what those benefits are and the challenges in presenting DTPs for TITE methodology. Using informative examples we demonstrate what these are and why they occur as well as potential solutions for how DTPs can still be incorporated for trials utilising this methodology.

In Chapter 5, we introduce efficacy transition pathways (ETPs) which are a new visualisation tool inspired by DTPs for use in Phase II trials. The idea

of visualising outcomes and decisions for dose-finding trials is a useful one, not only during the conduct of a trial but in its initial design stages as well. Through the development of three different trials at CRCTU it became apparent that a similar tool would be beneficial for Phase II trials. These trials, whilst not conducting dose-finding, aimed to assess efficacy and included multiple interim analyses at which different decisions could be made. We describe how ETPs are constructed and work, by providing examples from trials which inspired the idea behind ETPs. One of the success of DTPs was its ease of implementation through available software like the R package escalation by Brock [21]. In order to facilitate the same ease of use for ETPs we developed a R function along with a web based application to automatically generate these plots. In the chapter we detail how the application works to construct ETPs and additional features that we added so it could be used as an educational tool.

To end, Chapter 6, provides an overarching summary of the topics and ideas discussed in this thesis.

Chapter 2

Implementing the PO-TITE-CRM trial design into ADePT-DDR

This chapter contains work that has previously been published [32]. In this work I was responsible for the writing, analysis, conduct of simulations and generation of figures and tables used in this thesis.

2.1 Introduction

Worldwide there are approximately 600,000 new cases of Head and Neck Squamous Cell Carcinoma (HNSCC) each year [33]. Of which, 12,000 occur in the UK with the most common forms of treatment being surgery, radiotherapy and/or chemotherapy [34]. Radiotherapy is essential for the treatment of cancer. It has been estimated that more than 40% of patients will receive radiotherapy at some point in their treatment [35]. However, despite recent advancements in radiation techniques and the use of concomitant chemoradiotherapy, patients with solid tumours such as head and neck cancer have suboptimal cure rates [34], [36]. For those with advanced HNSCC primary radiotherapy with concurrent chemotherapy is often offered but, it has not been shown to improve survival in patients aged over 70 compared to radiotherapy alone

[37]. Therefore, any strategy to improve the efficacy of radiotherapy without increasing toxicity would have a significant impact on patient outcomes.

DNA damage repair (DDR) inhibition is a potential technique which could be utilised as it potentiates the therapeutic effects of ionising radiation in cancer cells [38]. Combining radiotherapy with DDR inhibition could improve clinical outcomes for these patients [39].

The ADePT-DDR trial ¹ is a platform trial which aims to evaluate the safety and efficacy of different DDR agents, or different immunotherapy agents and / or DDR and immunotherapy combinations, together with radiotherapy in patients with HNSCC. The initial component of this trial is a single-arm dose-finding trial investigating the ataxia telangiectasis and Rad3-related (ATR) inhibitor AZD6738 in combination with radiotherapy. ATR inhibitors not only stop DNA repair but impair the mechanism that allows for repairs to take place. Preclinical models have shown this double blocking to be effective in killing cancer cells [40].

In Chapter 1 we discussed model and algorithm-based dose-finding trial designs. Due to the historical use of rule-based designs [15], [16], the majority of the terminology used to describe them, and the ambiguity they raise, have been inherited by modern designs such as the CRM. The MTD in the context of a CRM is not the 'maximum' dose patients could tolerate but rather a dose in which there would be an acceptable target probability of a DLT occurring. For example, if the target is set at 25% the MTD would be the dose at which there is a 25% probability of experiencing a DLT. Rather than using the term MTD, the dose to be found will be referred to as the target dose (TD%, where the %'s are replaced by the target probability), i.e. TD25 would be the dose expected to be toxic in 25% of patients.

¹ Accelerating the Development and implementation of Personalised Treatments of DNA Damage Response agents and radiotherapy +/- immunotherapy for head and neck squamous cell cancer

The investigation of multiple-agent treatments, where the monotonicity assumption may not hold, is increasing in early phase trials. Finding the TD in combinations of treatments, compared to single-agents, presents methodological challenges. Each drug individually may obey the monotonicity assumption; we can refer to this as the doses being fully ordered. However, when multiple treatments are combined, the ordering of doses in terms of toxicity may not be fully apparent or may only be partially defined. An order may be identified for a subset of the doses which would result in a partial order. Without a full ordering it is uncertain which dose should be chosen in decisions of escalation and de-escalation and ultimately as the TD. This issue is not exclusively reserved for trials with multiple-agents. The monotonicity assumption may not hold for certain drugs in single-agent studies leading to partial orders of dose toxicity. For example, when dose and frequency of administration vary between dose levels. It requires that probability of toxicity always increases - staying the same is not enough. At high enough doses, this assumption is almost surely violated for all interventions when the event probability reaches its maximum. Thus, even when total ordering is possible, the monotonicity assumption could be violated [41]. Issues of partial ordering can occur in scenarios where multiple parameters of the treatment schedule are altered for each dose level. For example, either dose or treatment duration could be increased and even if patients receive an equal dose it would remain unclear as to if prolonged exposure to a lower dose is more toxic than short exposure to a higher dose, which implies a partial ordering of toxicity probabilities.

Further methodological challenges revolve around the issue of late-onset toxicities. Typically, early phase trials implement a short window to observe DLTs. This works well in situations where toxicities are likely to occur rapidly after treatment. However, this is not optimal for treatments that could cause late-onset toxicities such as radiotherapy. The aim with ADePT-DDR would be

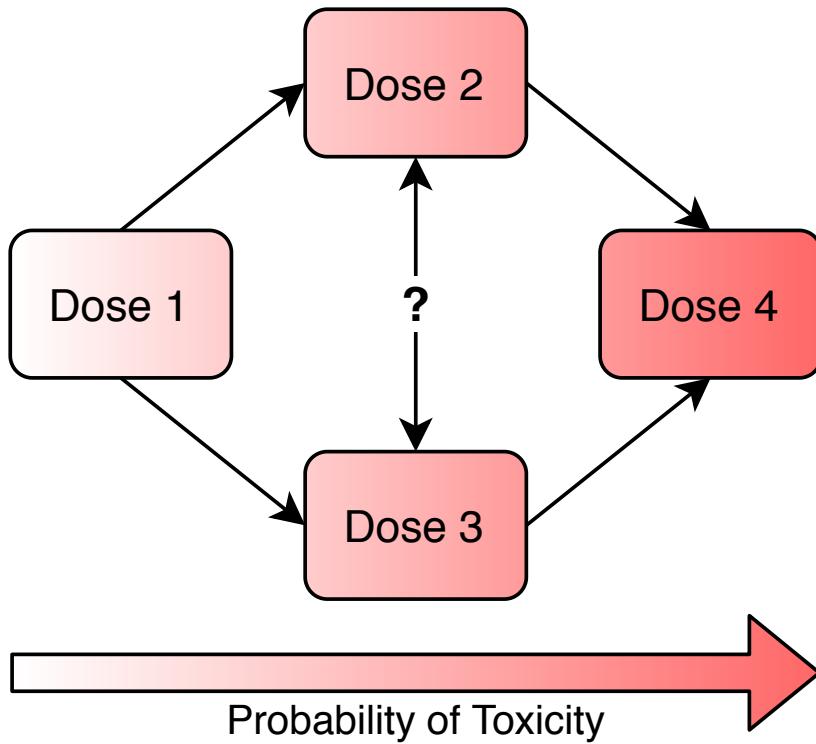
to incorporate a larger observation window to account for potential late-onset toxicities whilst also minimising the trial duration.

Cheung and Chappel [42] introduced an extension to the CRM to deal with the issues of treatments that may cause late-onset toxicity. This design referred to as the time-to-event CRM (TITE-CRM), uses a weighted dose-response model to incorporate the time it takes for a DLT to occur in a patient. There have also been published trial designs to deal with the issues that arise from investigating combinations of treatments. Thall et al. [43] proposed an adaptive two-stage Bayesian design which utilises a parametric model of toxicity as a function of two doses. Yin and Yuan [44] present a Bayesian design that uses a copula regression model to evaluate the joint toxicity probabilities of combined drugs. The continual reassessment method for partial orders (PO-CRM) developed by Wages et al. [45] extends the CRM design by relaxing the assumption of monotonicity and by modelling different potential orders. Figure 2.1 shows a simple example of partial ordering where the order of two out of the four dose levels are unknown.

Wages et al. [29], [45] further developed their work on the PO-CRM to deal with late-onset toxicities by implementing a TITE component. This trial design, referred to as the time-to-event continual reassessment method in the presence of partial orders (PO-TITE-CRM) by the authors, was chosen to be used in ADePT-DDR. A search of PubMed, conducted on the 25th of July 2020, found six articles that had cited the PO-TITE-CRM design by Wages et al. [29]. Of these six articles non actually implement the design into a trial. The following paragraphs provide more details.

Five of these papers were methodological in nature, two of which only include the PO-TITE-CRM design in a brief introduction to current methodology before going on to present new Bayesian trial designs [46], [47]. The other three

FIGURE 2.1: Example dose levels to illustrate partial ordering.



papers were authored by Wages. The first of which details practical considerations and specifications for the PO-CRM design, the TITE variant is only cited as the source of an example which is being used [48]. One paper presents an R package 'pocrm' [49], [50]. The package is only capable of analysing the PO-CRM design. The TITE variant is only referenced here as it illustrates the issue of partial ordering. The last methodological paper by Wages et al. [51] presents three different methods for phase I studies of drug combinations one of which is the PO-CRM however, PO-TITE-CRM is only mentioned as an extension to this design. A key message in this paper is the fact that novel methodologies are constantly emerging but are rarely implemented in practice.

The last paper is a protocol paper for a phase I/II study, OLA-TMZ-RTE-01 [52]. The phase I component of the study aims to determine the recommended phase II dose (RP2D) of olaparib combined with a standard schedule of radiotherapy and temozolomide (TMZ) as first line treatment for patients with

unresectable glioblastoma (GBM). The treatment schedule is divided into a radiotherapy and maintenance period. They propose to conduct two sequential dose-escalations of seven different olaparib dose-levels. Patients in the first escalation will be allocated to a dose level of olaparib for 10 weeks including radiotherapy for six weeks with TMZ given each day during radiotherapy and then for six cycles four weeks post radiotherapy during the maintenance period. They state the MTD1 will be determined using a TITE-CRM. Patients in the second escalation olaparib at the MTD1 during the radiotherapy period along with the same schedule of radiotherapy and TMZ. Those patients will then be allocated to one of the seven dose levels of olaparib during the maintenance period. Again, it is stated that the MTD2 will be determined using TITE-CRM modelling. The RP2D is the MTD1 and MTD2 during the radiotherapy and maintenance period respectively. Even though a combination of treatments is being investigated only olaparib is being escalated and doses for other treatments are fixed for all patients. Furthermore, the dose-levels for olaparib increase consistently in either amount or duration meaning there are no issues of partial ordering which would warrant the use of PO-TITE-CRM. The authors reference the TITE-CRM methodology with two papers. One of them being the paper detailing the PO-TITE-CRM design and the other being a paper by Huang and Kuan [53] which proposes an adaptive weight function that incorporates cyclical data of treatment into the TITE-CRM. It is unclear as to why the PO-TITE-CRM is cited as its methodology is not mentioned anywhere in methods.

An updated PubMed search conducted on the 20th July 2024 yielded an additional three papers. One of which was by van Werkhoven et al. [54] which details practical guidelines on running TITE-CRM trials providing some examples however no guidance is provided for PO-TITE-CRM trials specifically. Another paper published by Brown el at. [55] provides a road map to help

improve the design of phase I radiotherapy trials. Here PO-TITE-CRM is recommended as a methodology for dealing with dose-escalation of dual agents. The final paper [32] is a published form of the work in this chapter which documents our experience implementing this design.

We also conducted a search for papers that cited the original PO-CRM design paper by Wages et al. [45]. This was also conducted on the 20th July 2024 and yielded 77 results. The abstracts for all the papers were reviewed and we found four papers that discussed the implementation of PO-CRM methodology into trials (one of the papers is the published version of this chapter [32]). The remaining 73 papers were methodological in nature.

The first paper was by Walker et al. [56] and presents the results of an adaptive phase I trial. The trial aimed to identify if high-dose nitazoxanide is safe and well tolerated in healthy individuals also if it can reach and maintain antiviral concentrations predicted to be sufficient to prevent maturation of the SARS-CoV-2 spike protein. The paper primarily focused on presenting the results but does include details in the methods section and supplementary materials about the design of the trial and parameters used in the PO-CRM model. The trial included three different dosing schedules and there was uncertainty around the order with respect to toxicity which led to the use of PO-CRM.

Another of the three papers was by Mozgunov et al. [19]. This paper details their experience implementing the PO-CRM design. They provide a walk-through on how their design was implemented along with simulations and comparisons to other methods.

The final paper was also a results paper by Millard et al. [57] and it should be noted that Nolan Wages is also listed as a co-author for this paper. This trial was investigating a combination of entinostat and capecitabine in patients with HER-2 negative metastatic breast cancer. There were two doses of each drug under consideration leading to four different combinations/dose-levels. The

main focus of the paper were the results of the trial and there were limited details around the choice of the design and how it was parametrised.

It should be noted all of these papers were published after our work was conducted. So, we were unable to reference them or use them to help inform the choices made for our trial and design.

This is just a brief review of the current literature but it seems that the PO-TITE-CRM and PO-CRM have rarely been used or discussed since its inception.

This chapter provides novel insight into the methodology of PO-TITE-CRM through application in a real-world scenario. Section 2.2 will detail how the PO-TITE-CRM works. Section 2.3 discusses the justification for implementing the design into the ADePT-DDR trial and our experiences doing so. Section 2.4 explores other alternative designs which could have been implemented and assess how they perform in comparison to the PO-TITE-CRM. We provide some discussion in Section 2.5.

2.2 The PO-TITE-CRM design

Wages et al. [29] introduced the PO-TITE-CRM design which builds directly upon the PO-CRM design by incorporating a TITE component into the dose-toxicity model. The aim of which is to determine the target dose for combinations of drugs where the monotonicity assumption does not hold, in a setting where late-onset toxicities are possible.

TABLE 2.1: Example of drug combinations for a trial investigating two agents.

Agent	Drug combinations					
	d_1	d_2	d_3	d_4	d_5	d_6
A (mg/day)	0.25	0.5	1.0	0.25	0.5	1.0
B (mg/day)	1.0	1.0	1.0	1.5	1.5	1.5

To help understand partial ordering, consider an example of an early phase trial investigating the combination of two agents. Drug A which consists of three doses (0.25, 0.5, 1.0 mg/day) and drug B which consists of two doses (1.0, 1.5 mg/day), for a total of six drug combinations d_1, \dots, d_6 (Table 2.1). For each drug independently we assume they have a monotonic dose-toxicity curve however, the ordering of toxicity probabilities for some of the treatment combinations is unknown. Specifically, we can say d_1 is less toxic than d_2 as the dose of drug A increased whilst the dose of drug B stayed the same. This is also the case for d_2 and d_3 . So, d_1 can always be considered less toxic than d_2 which is always less toxic than d_3 . The same can be said for doses d_4, d_5 and d_6 , these three doses are can also all be considered more toxic than d_1 as well. The order between d_4 and d_5 in comparison to d_3 is not known because the dose of drug A decreases whilst the dose of drug B increases. Similarly the order between d_2 and d_4 is unknown. Also, we can say that d_6 is the always the most toxic dose. Assessing all these potential order toxicity relationships leaves five possible orderings.

1. $d_1 \rightarrow d_2 \rightarrow d_3 \rightarrow d_4 \rightarrow d_5 \rightarrow d_6$
2. $d_1 \rightarrow d_2 \rightarrow d_4 \rightarrow d_3 \rightarrow d_5 \rightarrow d_6$
3. $d_1 \rightarrow d_2 \rightarrow d_4 \rightarrow d_5 \rightarrow d_3 \rightarrow d_6$
4. $d_1 \rightarrow d_4 \rightarrow d_2 \rightarrow d_3 \rightarrow d_5 \rightarrow d_6$
5. $d_1 \rightarrow d_4 \rightarrow d_2 \rightarrow d_5 \rightarrow d_3 \rightarrow d_6$

Using the notation of Wages et al. [29], [45], let M denote the number of possible orders and Y be an indicator of a toxicity event. Then for a trial investigating k combinations, d_1, \dots, d_k , the dose for the j th patient, $X_j, j = 1, \dots, n$ can be thought of as random $x_j \in (d_1, \dots, d_k)$. For a specific ordering $m, m = 1, \dots, M$

the toxicity probability $R(d_i)$ is modelled by

$$R(d_i) = \phi_m(d_i, w, \beta) = w\psi_m(d_i, \beta) \quad i = 1, \dots, k; \quad m = 1, \dots, M \quad (2.1)$$

for a weighted dose-toxicity model $\phi_m(d_i, w, \beta)$ where $\beta \in (-\infty, \infty)$. The weight, w as defined by Cheung and Chappel [42], is a function of the time-to-event of each patient and is incorporated linearly with the dose-toxicity model ψ so that $0 \leq w \leq 1$. Each patient is followed for a fixed amount of time T . Let U_j represent the time-to-toxicity of patient j . Then for $u \leq T$,

$$P(U_j \leq u) = P(U_j \leq u | U_j \leq T)P(U_j \leq T) \equiv w(u; T)\psi_m(d_i, \beta). \quad (2.2)$$

For simplicity we will refer to the weight function $w(u; T)$ as w . The weight function will have to be decided upon by the trials team. Dependent on the scenario, a simple linear function or a more complex adaptive weights function could be utilised. There are also several working dose models which could be used for ψ . Wages et al. [29] present their design with the power parameter model given by

$$\psi_m(d_i, \beta) = \alpha_{mi}^{\exp(\beta)} \quad i = 1, \dots, k; \quad m = 1, \dots, M. \quad (2.3)$$

Here $0 < \alpha_{m1} < \dots < \alpha_{mk} < 1$ are the prior estimates of toxicity probabilities, or skeleton, for each potential ordering. Furthermore, prior probabilities are assigned to each order M to account for any prior information regarding the plausibility of each model such that, $p(m) = \{p(1), \dots, p(M)\}$, where $p(m) \geq 0$ and $\sum_m p(m) = 1$. When all orders are equally likely or there is no prior information available on possible orderings the prior is discretely uniform and would be $p(m) = 1/M$.

A Bayesian framework is used and a prior probability distribution $g(\beta)$ is

assigned to the parameter β . The ordering with the largest prior probability is selected as the starting ordering. In the scenario where all priors are equal an ordering is selected at random, subsequently a starting dose is also chosen. After j patients have been entered into the trial data is collected in the form of $\Omega_j = \{x_1, y_1, \dots, x_j, y_j\}$. A weighted likelihood for the parameter β is used to establish running probabilities of toxicity for each treatment combination. The weighted likelihood under ordering m , is given by

$$\tilde{L}_m(\beta|\Omega_j) = \prod_{l=1}^j \phi_m^{y_l}(x_l, w_l, \beta) \{1 - \phi_m(x_l, w_l, \beta)\}^{(1-y_l)} \quad (2.4)$$

which can be used to generate a summary value $\hat{\beta}_{mj}$ for each ordering. With the likelihood and the data Ω_j , the posterior density for β can be calculated using

$$\tilde{f}_m(\beta|\Omega_j) = \frac{\tilde{L}_m(\beta|\Omega_j)g(\beta)}{\int_{\beta} \tilde{L}_m(\beta|\Omega_j)g(\beta)d\beta} \quad (2.5)$$

This can then be used to establish posterior probabilities of the orderings given the data as

$$\tilde{\pi}(m|\Omega_j) = \frac{p(m) \int_{\beta} \tilde{L}_m(\beta|\Omega_j)g(\beta)d\beta}{\sum_{m=1}^M p(m) \int_{\beta} \tilde{L}_m(\beta|\Omega_j)g(\beta)d\beta}. \quad (2.6)$$

We select the single ordering, h , with the largest posterior probability along with its associated working model $\psi_h(d_i, \beta)$ and generate toxicity probabilities for each dose level. Once the j th patient has been included the posterior probability of DLT can be calculated for d_i so that

$$\hat{R}(d_i) = \psi_h(d_i, \hat{\beta}_{hj}); \hat{\beta}_h = \int_{\beta} \beta \tilde{f}_h(\beta|\Omega_j)d\beta. \quad (2.7)$$

In turn, the dose level $x_j \in \{d_1, \dots, d_k\}$ assigned to the $(j+1)$ th patient is the dose, d_i , which minimises

$$\triangle(\hat{R}(d_i), \theta) = |\hat{R}(d_i) - \theta|, \quad i = 1, \dots, k \quad (2.8)$$

where θ is the target toxicity rate. Similarly, once all patients have been recruited and observed and the trial ends, the target dose ($TD\theta$) is the dose, d_i , which minimises (2.8).

2.3 PO-TITE-CRM in ADePT-DDR

The decision to implement PO-TITE-CRM into ADePT-DDR was made by Piers Gaunt (PG) after discussions with other statisticians Kristian Brock (KB) and Daniel Slade (DS), as well as the chief investigator and other co-investigators. The design was chosen as the toxicity probabilities of the dose levels were not monotonically increasing which restricts the use of most early phase designs such as the CRM. Additionally, the design also handles late-onset toxicities which would be an issue in ADePT-DDR due to the treatment involving radiotherapy. The availability of software to conduct the trial was also a factor that was considered. The R package ‘pocrm’ [49] only provides a means for implementing the PO-CRM design but the easy accessibility to this code meant that it could be extended to include the TITE component.

The intended use of this design is for dose-finding in combinations of therapies, as this is the source of the partial ordering issue. ADePT-DDR however, is a unique implementation of the design as even though it involves a combination of therapies (radiotherapy and AZD6738) the dose of radiotherapy is fixed and dose-finding is only planned for AZD6738. PO-TITE-CRM is still applicable in this case as the design includes combinations of dose and duration

for AZD6738 which are partially ordered.

We detailed the design as presented by the authors. However, for our implementation we opted to utilise a two-stage likelihood based approach. Investigators wanted to start at a lower dose for safety reasons but still wanted to be able to escalate quickly in the scenario where no DLTs were observed. Implementing an initial design would allow us to do that. The ‘pocrm’ package also allowed for this option which helped when we developed our own code to implement this approach.

A two-stage PO-TITE-CRM will be used to find the TD25 of AZD6738. This will be determined by dose-limiting toxicities evaluated by Common Terminology Criteria for Adverse Events (CTCAE) v5.0 and Radiation Therapy Oncology Group (RTOG) late toxicity score. The binary DLT events are pre-defined by a variety of grade 3-4 adverse events notably, haematological, cardiovascular and gastrointestinal/hepatic toxicities as well as significant non-haematological events and specific treatment-related toxicities. DLTs will be monitored for the duration of treatment (seven weeks) and throughout the follow-up period. The total follow-up period post treatment is 52 weeks, so patients will spend a total of 59 weeks in the trial.

A maximum of 60 patients will be recruited for the dose-finding aspect of this trial and up to 20 patients as controls. Controls will be utilised to make comparisons for secondary outcomes such as survival and efficacy. Control patients will only be receiving radiotherapy, the dose of which is fixed at 70Gy/35F. Cohorts of three patients will be recruited and assigned to dose levels chosen by the PO-TITE-CRM. Controls will be recruited in the interim period between the recruitment of the third patient in a cohort and the completion of the minimum follow-up period.

2.3.1 Partial ordering in practice

Each patient entered into ADePT-DDR will receive fixed dose radiation, totalling 70 Gy in 35 fractions over seven weeks. For the dose-finding aspect we investigate six doses of AZD6738 detailed in Table 2.2. Treatment dose and duration to be selected for dose level 3 will be determined based on a combination of data observed, adverse events and compliance. The issue of partial ordering is illustrated in Figure 2.2 inspired from plots by Wages et al. [29]. The doses to be used in this trial are detailed in their appropriate box. It is clear that dose levels 2a and 2b would be considered more toxic than dose level 1 due to the increase in treatment duration and treatment dose respectively. When comparing 2a and 2b it is unknown whether the increase in dose or duration will be more toxic. Hence there are two possible orderings for ADePT-DDR.

$$1. \ d_{-1} \rightarrow d_0 \rightarrow d_1 \rightarrow d_{2a} \rightarrow d_{2b} \rightarrow d_3$$

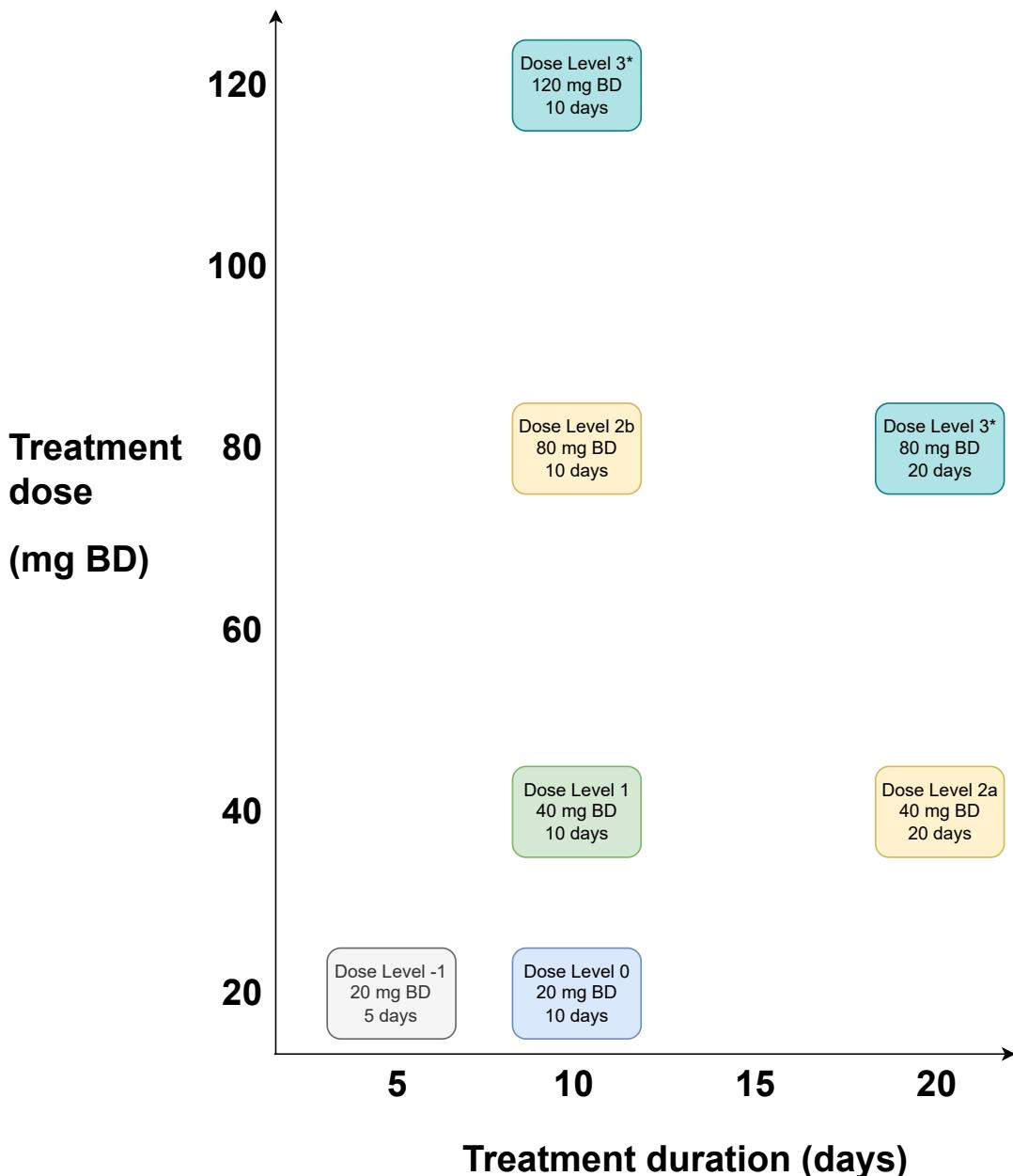
$$2. \ d_{-1} \rightarrow d_0 \rightarrow d_1 \rightarrow d_{2b} \rightarrow d_{2a} \rightarrow d_3$$

TABLE 2.2: ADePT-DDR dose-levels and duration of treatment for AZD6738.

Dose Level	AZD6738 Daily dose (mg BD)	Weeks	Duration (days)	Radiotherapy
-1	20	1	5	70 Gy / 35 F
0	20	1&4	10	70 Gy / 35 F
1	40	1&4	10	70 Gy / 35 F
2a	40	1,2,4&5	20	70 Gy / 35 F
2b	80	1&4	10	70 Gy / 35 F
3	120	1&4	10	70 Gy / 35 F
	80	1,2,4&5	20	70 Gy / 35 F

It should be noted that our definition of orderings is assuming that total dose is the key driver in terms of probability of toxicity. If we were to relax this assumption it could be possible that dose-level 3 at 120 mg for 10 days (1200 mg total) could be considered less toxic than 2a 40 mg for 20 days (800 mg

FIGURE 2.2: ADePT-DDR dose levels across dose and duration.



total). In this instance prolonged exposure to the treatment may be more toxic even though the total overall dose is lower. This would introduce a third ordering ($d_{-1} \rightarrow d_0 \rightarrow d_1 \rightarrow d_{2b} \rightarrow d_3 \rightarrow d_{2a}$). However, since we are operating under the assumption that more overall dose increases probability of toxicity we will only consider the two orderings listed above.

Traditionally, dose-finding trials for combinations would select dose levels

to form a 'path' through the dose combination space such that each subsequent dose level was logically more toxic. This avoids the issue of partial ordering but means doses of interest or effective dose combinations may be missed or not investigated. Specifically, for ADePT-DDR this allows two 'paths' from dose level 1 extending to 2a and 2b. In terms of dose level 3 only one of the doses in that tier will be investigated, it was unclear as to which dose level would be best due to a lack of historical data. Even though dose level 3 is not yet specified in terms of modelling and simulations it was treated as singular dose. This was done as clinicians thought that it would be unlikely that we would reach these doses and that the probability of toxicity between them would be similar.

Preliminary designs of the trial included only five dose levels and planned to use dose level 0 as the starting dose. During the trial design phase it was decided a new lower dose (dose level -1) would be introduced to allow for de-escalation if the initial dose was found to be too toxic. Dose escalation/de-escalation for subsequent cohorts would be determined from the two-stage PO-TITE-CRM. A two-stage design allows for escalation according to a pre-defined escalation scheme. The first stage dictates that if no DLT's are observed in the current cohort the dose allocated to the next cohort is the following dose in the escalation scheme. Dose levels continue to be incremented in this fashion until the first DLT is observed. In stage two, dose levels are determined by the PO-TITE-CRM.

Typically CRM designs begin by testing the first patient, or cohort, at the prior guess of TD or at a lower dose to be safe. However, clinicians may have safety concerns beginning the trial at higher dose levels as well as escalating to higher dose levels without testing lower ones. Investigators in ADePT-DDR expressed similar concerns as such a two-stage design was adopted. The escalation scheme used in stage one of ADePT-DDR will follow that of the first

ordering ($d_{-1} \rightarrow d_0 \rightarrow d_1 \rightarrow d_{2a} \rightarrow d_{2b} \rightarrow d_3$). If patients in the first cohort (assigned to dose level 0) do not experience a DLT the next cohort will be allocated to dose level 1 and then if no DLTs are observed again the third cohort will be allocated to dose level 2a and so on and so forth. The dose escalation scheme was determined based on the prior probabilities of toxicity generated for each dose level.

Information elicited from the investigators helped generate prior probabilities of toxicity for each dose level. They believed that dose level 2b would be the TD25 with 2a being less toxic. This was used in conjunction with the `getprior` function from the `dfcrm` R package [20] which yielded priors of 0.012, 0.036, 0.084, 0.157, 0.25 and 0.355 for dose levels -1, 0, 1, 2a, 2b and 3 respectively. The half-width of the indifference interval was set at 0.05. The indifference interval is an interval in which the toxicity probability of the selected dose will eventually fall. Prior probabilities are also required for the plausibility of each model and even though the clinicians think that 2b will be more toxic than 2a there is no clear evidence and still a lot of uncertainty. As such it is sensible to assume a plausibility probability of 0.5 for each ordering, implying both orders are equally likely to be the true ordering of these dose levels.

2.3.2 The TITE component

The observation window for this trial will be up to a year post-treatment as the combination of radiotherapy with AZD6738 is anticipated to cause late-onset toxicity. The Acute DLT observation period is 12 weeks (84 days) post radiotherapy end with a minimum of 8 weeks (56 days) for the last patient of each cohort. However, patients will continuously be monitored for occurrence of DLT for at least 12 weeks (84 days), i.e. at least 12 weeks (84 days) from

the end of radiotherapy. The full window will last for 52 weeks (365 days) post-treatment.

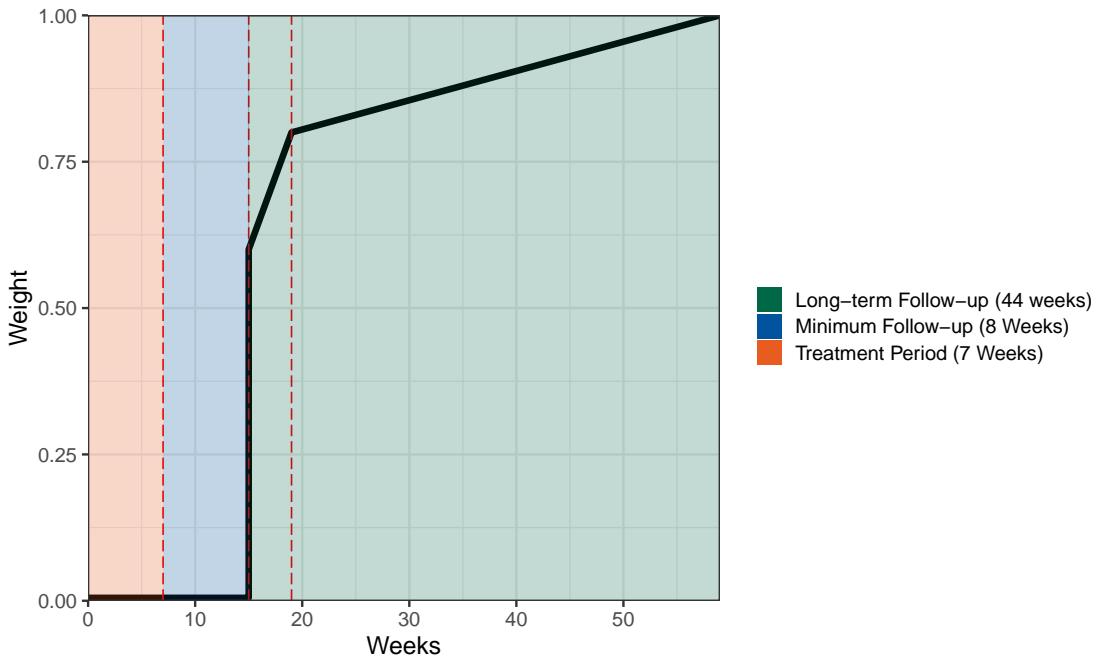
The TITE component incorporates a weighting contribution for each patient dependent on how long that patient has been evaluable in the study. This allows a patient to be evaluated once they have been observed for the minimum DLT period of 8 weeks (56 days). The weighting at this point is 60% rising to 80% at 12 weeks (84 days). A patient will not contribute fully to the model until they have completed 52 weeks (365 days) follow up (or have experienced a DLT at any stage in which case they will be weighted as a whole contribution). Linear weighting functions will be employed for any patient with a length of follow up between these three time points. One weight function to calculate weights between 8-12 weeks and another for weights between 12-52 weeks. For the weighting function $w(u; t_1, t_2, t_3)$ where u is the time-to-toxicity of patient j and t_1, t_2, t_3 is the time period with values 8, 12 and 52 respectively. Then for $t_1 \leq u \leq t_3$

$$w(u; t_1, t_2, t_3) = 0.6 + 0.2 \frac{\max(0, \min(u, t_2) - t_1)}{t_2 - t_1} + 0.2 \frac{\max(0, u - t_2)}{t_3 - t_2}. \quad (2.9)$$

All patients will have a minimum weight of 60% as that is the prescribed weighting to the minimum follow up period before dose escalation/de-escalation decisions can be made. For each additional week the patient is observed, without a DLT occurring, between weeks 8 and 12 their weighting increases by 5%. Similarly for each week between 12 and 52 weeks, without a DLT, weighting increases by 0.5%. Figure 2.3 illustrates the weight function and how the weight changes for patients dependent on how long they have been followed-up.

The TITE-CRM originally presented by Cheung and Chappel [42] did not

FIGURE 2.3: Weights of patients who have not experienced a DLT across the observation window.



incorporate a minimum follow-up period and their design allowed for the continual recruitment of patients whenever they became available. There are some practical considerations which make this infeasible in ADePT-DDR. The model would need to be run each time a new patient entered the study which requires statistical input hence the introduction of cohorts. Clinicians may also have safety concerns if we see rapid recruitment at the start of the trial and the model keeps escalating so we impose a minimum follow-up period. Initially this was set at 12 weeks (at 80% weighting) however, statisticians AP and PG pointed out that dose escalation/de-escalation decisions would have to take place 19 weeks (7 weeks treatment and 12 weeks follow-up) after recruitment of the third patient in the cohort. Dependent on the recruitment rates this could extend the duration of the trial and negates the benefits of using a TITE design. The investigators also agreed this was too long and settled on lowering this period to 8 weeks (at 60% weighting) whilst also including the original 12 week weighting of 80%.

2.3.3 Stopping rules

A practical modification was included to allow for early stopping of the trial if there is sufficient evidence that the TD has been reached. Sufficient evidence is achieved once 15 patients (five cohorts) have been treated at the same dose level and the model allocates that dose level again to a sixth cohort. This rule evolved from the original designs of the trial which involved 30 patients with a dose expansion cohort to ensure at least 15 patients were treated at the TD.

Initial simulations highlighted the inadequacy of these design parameters as operating characteristics for various scenarios were poor, specifically in terms of correct TD selection. Clinicians explained the inclusion of the dose expansion cohort was to ensure the dose-finding aspect of the trial did not take a large amount of time whilst also allowing safety to be assessed at the TD. In order to ensure that a reasonable amount of patients would be treated at the TD, the trial would not take longer than necessary and operating characteristics improved, the sample size was increased and this rule was introduced.

A rule was also implemented to allow for early termination of the trial in the case of excess toxicity at the lowest dose. If the probability of DLT at the lowest dose is higher than 0.35 with a probability of 80% and has been tested, the trials safety committee will be alerted and will recommend if the trial should be stopped. As the trial starts at dose level 0, which is not the lowest dose, it is possible for the trial to recommend terminating without ever allocating patients to the lowest dose level. As such it was decided early termination would only occur once at least 3 patients (1 cohort) have been allocated dose level -1.

An approximate estimate of the variance was calculated using methodology presented by O’Quigley and Shen [58]. The observed information matrix is obtained by taking the second derivative of the likelihood (eq. 2.4) which is

then used to calculate the variance $v(\hat{\beta}_j)$, for estimate β_j which becomes more accurate with larger sample sizes. After each cohort, we sample from a normal distribution with parameters based on the estimate of β_j and its variance. These samples are then plugged into our dose-toxicity model to ascertain the probability of toxicity at the lowest dose. The trial will be recommended to stop if it breaks the rule based on the criteria above.

2.3.4 Example trial run

To demonstrate how the trial will work in practice we present an example trial run. Patients will be recruited in cohorts of three. After each cohort a dose will be recommended based on our design and the subsequent cohort of patients will be given that dose. This process is then repeated until the stopping rules are triggered. In our example here we observe no DLTs to begin with allowing us to follow the initial escalation scheme up to dose-level 3. At this point we begin to observe DLTs and thus begin to use the model to recommend dose-levels until we stop for consensus.

Table 2.3 shows model outputs after each cohorts specifically, the estimated probability of toxicity or DLT rate at each dose and the estimated posterior probability of each ordering. Order 1 refers to the order assuming 2b is more toxic than 2a and order 2 assumes 2a is more toxic than 2b. For the first initial four cohorts we see that there are no model outputs. This is because the model was not fit in these instances as no DLTs were observed. You can see the initial escalation scheme was followed. Meaning after cohort 1 received dose-level 0 cohort 2 received dose-level 1, cohort 3 received dose-level 2a and cohort 4 received dose-level 2b.

For cohort 5 we see that one patient in this cohort observed a DLT. Based on the model outputs in this scenario the estimated probability of toxicity at

TABLE 2.3: Summary of model outputs for our example trial run.

Cohort	Dose	DLT	Estimated DLT rate					Order prob		Recommended Dose
			-1	0	1	2a	2b	3	1	
1	0	0								1
2	1	0								2a
3	2a	0								2b
4	2b	0								3
5	3	1	0	0	0.01	0.04	0.09	0.17	0.50	0.50
6	3	3	0.01	0.03	0.08	0.15	0.24	0.35	0.50	0.50
7	2b	2	0.02	0.06	0.12	0.21	0.31	0.42	0.66	0.34
8	2a	0	0.02	0.05	0.10	0.18	0.28	0.39	0.73	0.27
9	2b	0*	0.02	0.06	0.12	0.20	0.30	0.41	0.55	0.45
10	2a	0	0.02	0.04	0.10	0.18	0.27	0.38	0.62	0.38
11	2b	0	0.01	0.03	0.08	0.15	0.24	0.34	0.57	0.43
12	2b	0	0.01	0.03	0.07	0.13	0.22	0.32	0.51	0.49

* No DLTs in this cohort, but the previous cohort had a late onset DLT.

dose-level 3 is 0.17. As this is closer to our TD25 this becomes the dose that is recommended for cohort 6. We can also look at the estimates of probability for the orders, here both are at 0.5 indicating both orders are still equally likely. In cohort 6 a further three patients are recruited to dose-level 3 and we observe DLTs in all three patients. Overall, now there have been 4/6 DLTs at dose-level 3. Based on the model output here dose-level 2b with an estimated DLT rate of 0.24 is closest to the TD25 and is recommended for the next cohort. The posterior probability for the orders still indicate each order is equally likely.

Now at cohort 7, patients are allocated to dose-level 2b. Here two DLTs are observed. The model recommends dose-level 2a. We can see the estimated probability of the orders suggests order 1 is more likely, which is indicative after having observed multiple toxicities at higher doses. Cohort 8 at dose-level 2a observes no toxicities which leads to the recommendation of going back to dose-level 2b. Here the estimated DLT rate here is 0.28 which is closer to the TD25 compared to any of the other doses. For the order probabilities we see again order 1 is still more likely.

Cohort 9 recruits patients to dose-level 2b and observes no DLTs however,

one patient from the previous cohort (cohort 8 dose-level 2a) has now experienced a late-onset DLT after the minimum follow-up period. Incorporating this new cohort data as well as the new DLT data for the previous cohort gives a model recommendation of going back to dose-level 2a. The exact estimated DLT rates for 2a and 2b were 0.2036 and 0.3037 respectively, 2a was narrowly the closest to dose-level 2b hence why it was recommended. The DLT at 2a also impacted the probabilities for each ordering. These still indicated 1 was more likely but they are much closer now.

Going forward no more DLTs were observed and we can see the model keeps recommending dose-level 2b. Cohort 12 is the fifth cohort to treat patients at dose-level 2b, after observing no DLTs the model once again recommends dose-level 2b. This then triggers our consensus stopping rule as 15 patients have been treated at the same dose which is then recommended again. In this example dose-level 2b, with an estimated DLT rate of 0.22, is our TD25.

Probabilities for the orders are now 0.51 and 0.49 for 1 and 2 respectively, meaning order 1 is slightly more likely. Overall dose-level 2a observed DLTs in 1/9 patients and dose-level 2b observed DLTs in 2/15 patients. In the data we observed there was no observable difference overall between the two doses. Earlier on in the trial the probabilities of the order fluctuated as more DLTs were observed. With potentially more data again this would change and keep updating.

An additional point to consider is that at each dose decision or when the model is fit patients will have different weights based on the time spent in the trial without a DLT. For this example we assigned patients weights based on the weight function and the minimum follow-up period required. Figure 2.4 visualises the data that was used for the model output after each cohort. The shape of the points represent whether or not a patient had a DLT, the dose is represented by colour and the weight each patient had is represented by the

transparency of the point. Later patients in a cohort tend to have less weight as they would have been recruited later, this is represented by the fainter points on the plot. For each subsequent cohort these points then become darker as patients gain more weight in the model after having been observed for a longer period of time. Patients with DLTs are always at full weight.

Figure 2.5 shows an overall summary of the DLTs in each patient at each dose-level and shows how the doses change over time. One patient at dose-level 2a who had their DLT after the initial monitoring period which did not affect the recommendation for the next dose-level but for the cohorts thereafter.

FIGURE 2.4: Plot of data included for each dose decision at each cohort.

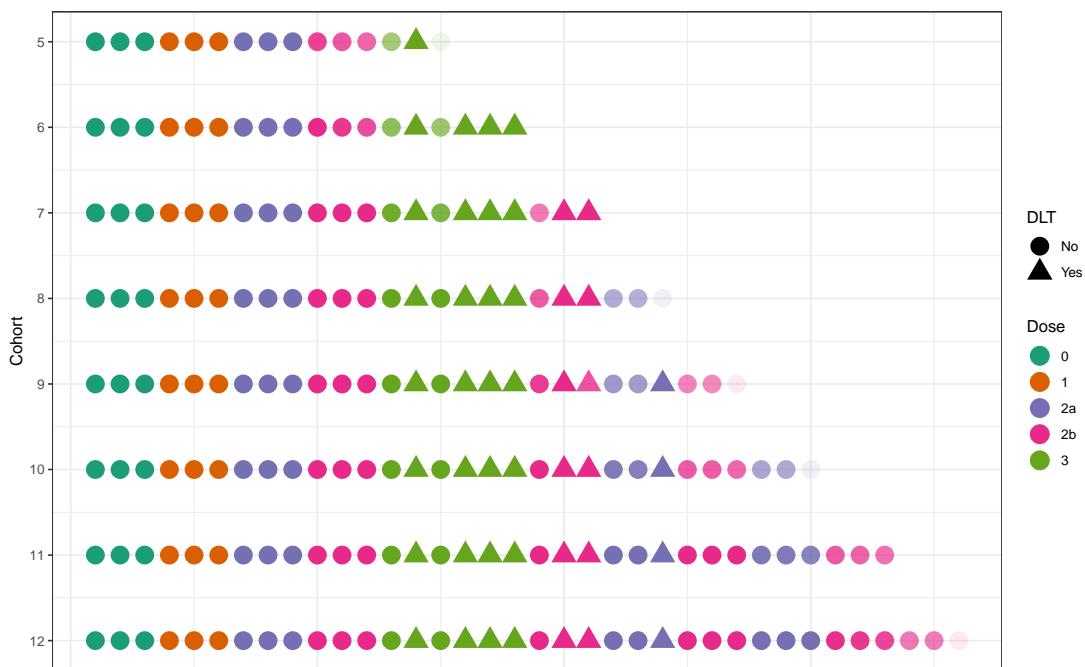
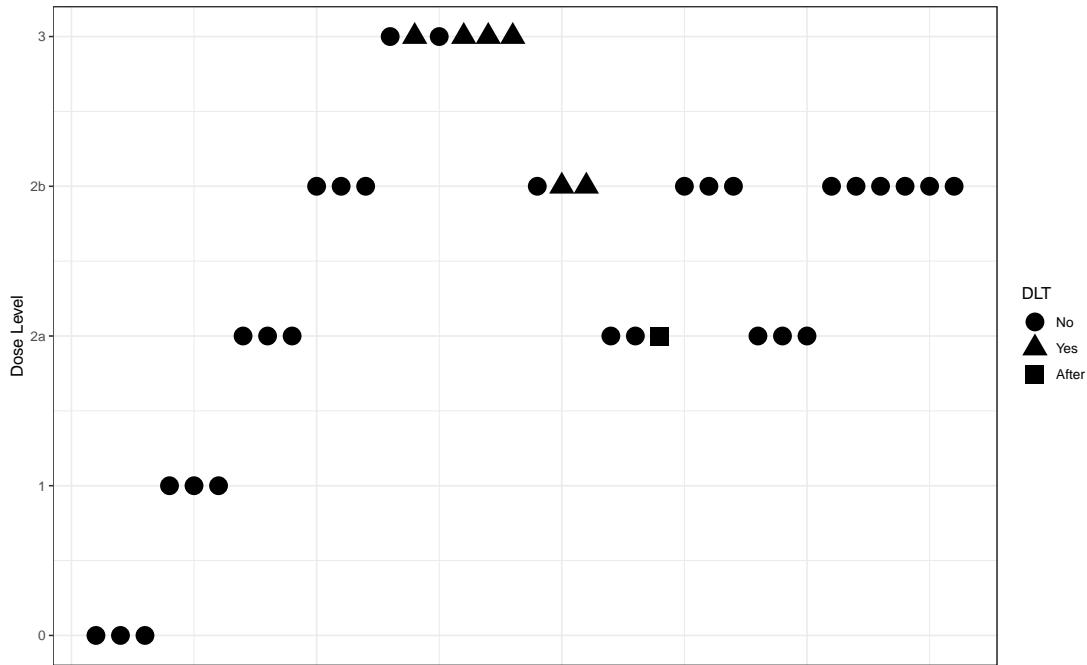


FIGURE 2.5: Plot of dose decisions throughout the example trial.



2.3.5 Operating characteristics

Simulations were continually utilised during the design process of the trial to assess how various changes impact the overall performance. These changes to design features such as the sample size, weight function and stopping rules helped inform decisions which led to the design specified in the previous section.

Functions from pocrm package in R [49], [50] were modified in order to perform simulations and conduct the trial. The majority of work involved integrating the TITE component and the stopping rules into the code. In standard CRM designs a binary outcome for toxicity is generated for each patient based on a pre-specified true DLT rates for the dose they are assigned. Adding the TITE component means the time the toxicity occurs also has to be generated, the simulation must also track this time and incorporate this information into the PO-TITE-CRM model when it needs to make dose allocation decisions for the next cohort. We defined multiple scenarios to reflect various real life

possibilities in order to assess the design's performance.

Standard scenarios to run involve adjusting the true DLT rates to reflect each dose being the TD25. For each of these we calculate the probability of selecting each dose as the TD25. It would be expected the dose with the highest probability of being selected has its true DLT rate set at 25% to match the target rate. A high probability of selection for the correct dose implies the design works well in the specified scenario. Additional characteristics such as the average number of patients at each dose level are also investigated. This can be used to look at how many patients may potentially be allocated to a toxic dose. It is also necessary to consider performance when all doses are too toxic, here we would want the design to recommend stopping early. Usually the true DLT rates used to define these scenarios abide by the monotonicity assumption. Due to the partial ordering we consider scenarios in which the true DLT rates follow both orders. However, as ADePT-DDR only has two orders we explored all scenarios for each ordering.

We simulated 10000 trials for each scenario using the finalised design detailed in section 2.3. Simulations were based on the assumption that the trial would recruit one patient per month. The occurrence of DLT's were randomly generated for patients in each cohort using a Bernoulli distribution with the probability set at the true DLT rate for the cohorts assigned dose level in the specific scenario. For patients who had a DLT occur, the time at which the DLT occurred was randomly generated using a uniform distribution which spanned the start of treatment to the end of follow-up. The simulations presented in Tables 2.4 and 2.5 took approximately 5 hours and 53 minutes to run. It is recommended by Morris et al. [59] to detail the Monte Carlo standard error in order to quantify the simulations uncertainty. In the case of a 50% selection probability the Monte Carlo standard error estimated by 10000 simulations is $\sqrt{0.5 \times 0.5 / 10000} = 0.5\%$. This implies that differences of 1% can be deemed

as significant.

Table 2.4 details simulations for eight scenarios to test the performance of the PO-TITE-CRM design using true DLT rates which reflect the first ordering. We analyse scenarios where each dose is the TD25 (scenarios 1-6) and when all doses are too toxic (scenario 8). Additionally, we also investigate performance under conditions where the probability of DLT is fairly similar between doses (scenario 7). This is a notoriously difficult circumstance for CRM designs to deal with as the limited number of patients and events at each dose make it hard to accurately estimate toxicity probabilities if they are similar. Simulation results for ordering 2 are shown in Table 2.5 where dose level 2a is considered more toxic than 2b. This is achieved by altering the true DLT rates so 2b has a lower probability of DLT compared to 2a.

Ideally we want the probability of selection for the dose allocated at TD25 to be as high as possible and greater than other dose levels. For scenarios 1-7 the TD25 is highlighted in bold along with results from the simulations. However, for scenario 8 where all doses are too toxic we expect the trial to terminate early, here 'stop' should be selected and its associated probability of stopping is shown in bold.

In scenarios 1 - 6 (Table 2.4), this design correctly selects the TD25 with probabilities between 43% and 78%, under the assumption 2b is more toxic than 2a. Likewise, for the ordering where 2a is more toxic than 2b, scenarios 9-14 (Table 2.5) have probabilities between 43% and 78% of correctly selecting the TD25. Correct selection probabilities are generally higher when the TD25 is at the first and last dose levels compared to dose levels 2a and 2b. However, these dose levels are still chosen with the highest probability as the TD25 in their given scenarios. For scenarios 7 and 15, the probabilities of toxicity are equally spaced, approximately 5% apart. This is a relatively difficult scenario for dose-finding studies to handle. The probability of selecting the TD25 is 28%

TABLE 2.4: Operating Characteristics of the two-stage PO-TITE-CRM (with true DLT rates that imply 2b is more toxic than 2a) based on 10000 simulated trials. Definitions: DLT: Dose-limiting toxicity. P(select): Probability of selecting a dose as the TD25.

Scenario	Prior DLT	Dose Levels						Stop
		-1	0	1	2a	2b	3	
1: TD25 @-1	Prior DLT	0.01	0.04	0.08	0.16	0.25	0.35	
	True DLT rate	0.25	0.4	0.45	0.5	0.55	0.6	
	P(select)	0.68	0.18	0.05	0.01	0	0	0.08
	% of patients	39	32	20	6	3	0	
2: TD25 @0	Mean number of patients	10.17	8.46	5.33	1.67	0.69	0.07	
	True DLT rate	0.12	0.25	0.4	0.45	0.5	0.55	
	P(select)	0.23	0.51	0.2	0.03	0.02	0	0.01
	% of patients	17	35	29	11	6	1	
3: TD25 @1	Mean number of patients	5.24	10.48	8.75	3.4	1.83	0.26	
	True DLT rate	0.09	0.12	0.25	0.4	0.45	0.5	
	P(select)	0.02	0.2	0.55	0.14	0.09	0.01	<0.01
	% of patients	4	20	34	23	16	3	
4: TD25 @2a	Mean number of patients	1.22	6.41	10.97	7.23	5.14	1.02	
	True DLT rate	0.06	0.09	0.12	0.25	0.4	0.45	
	P(select)	0	0.02	0.22	0.48	0.23	0.05	<0.01
	% of patients	1	12	20	31	25	11	
5: TD25 @2b	Mean number of patients	0.47	3.88	6.74	10.43	8.2	3.5	
	True DLT rate	0.03	0.06	0.09	0.12	0.25	0.4	
	P(select)	0	0	0.02	0.3	0.43	0.25	0
	% of patients	1	10	12	24	28	25	
6: TD25 @3	Mean number of patients	0.25	3.36	4.15	8.17	9.33	8.33	
	True DLT rate	0.01	0.03	0.06	0.09	0.12	0.25	
	P(select)	0	0	0	0.09	0.13	0.78	0
	% of patients	0	10	11	18	18	42	
7: Equal steps in DLT rate	Mean number of patients	0.1	3.13	3.49	5.46	5.6	13.14	
	True DLT rate	0.05	0.1	0.15	0.2	0.25	0.3	
	P(select)	0	0.03	0.12	0.31	0.28	0.26	<0.01
	% of patients	2	13	18	26	23	19	
8: All toxic	Mean number of patients	0.55	4.03	5.72	8.32	7.15	5.96	
	True DLT rate	0.5	0.6	0.65	0.7	0.75	0.8	
	P(select)	0.26	0	0	0	0	0	0.74
	% of patients	56	26	15	2	0	0	
	Mean number of patients	9.05	4.27	2.4	0.37	0.04	0	

and 32% for orderings 1 and 2 respectively and even if the performance is poor the correct dose is still likely to be selected. In scenarios 8 and 16, where all the doses are too toxic, the design very seldom allocates patients higher than the first three doses and there is a high chance (74% and 73% respectively) that the trial will recommend early stopping.

Additionally, we assess designs based on how doses are allocated to patients. Designs may correctly select the TD however, this could be undesirable

TABLE 2.5: Operating Characteristics of the two-stage PO-TITE-CRM (with true DLT rates that imply 2a is more toxic than 2b) based on 10000 simulated trials. Definitions: DLT: Dose-limiting toxicity. P(select): Probability of selecting a dose as the TD25.

Scenario	Prior DLT	Dose Levels						Stop
		-1	0	1	2a	2b	3	
9: TD25 @-1	Prior DLT	0.01	0.04	0.08	0.16	0.25	0.35	
	True DLT rate	0.25	0.4	0.45	0.55	0.5	0.6	
	P(select)	0.67	0.19	0.05	0	0.01	0	0.08
	% of patients	39	32	20	6	3	0	
10: TD25 @0	Mean number of patients	10.19	8.43	5.27	1.6	0.68	0.07	
	True DLT rate	0.12	0.25	0.4	0.5	0.45	0.55	
	P(select)	0.23	0.52	0.2	0.02	0.02	0	0.01
	% of patients	18	36	29	11	6	1	
11: TD25 @1	Mean number of patients	5.24	10.64	8.82	3.16	1.85	0.24	
	True DLT rate	0.09	0.12	0.25	0.45	0.4	0.5	
	P(select)	0.02	0.2	0.55	0.09	0.14	0.01	<0.01
	% of patients	4	20	34	21	17	3	
12: TD25 @2a	Mean number of patients	1.16	6.43	11.07	6.83	5.6	1.07	
	True DLT rate	0.06	0.09	0.12	0.25	0.15	0.45	
	P(select)	0	0.01	0.08	0.44	0.33	0.14	<0.01
	% of patients	1	11	16	30	24	18	
13: TD25 @2b	Mean number of patients	0.48	3.78	5.24	10.1	7.9	6.07	
	True DLT rate	0.03	0.06	0.09	0.35	0.25	0.4	
	P(select)	0	0	0.15	0.31	0.43	0.11	0
	% of patients	1	11	18	30	28	14	
14: TD25 @3	Mean number of patients	0.25	3.5	5.9	9.82	9.14	4.54	
	True DLT rate	0.01	0.03	0.06	0.12	0.09	0.25	
	P(select)	0	0	0	0.13	0.09	0.78	0
	% of patients	0	10	11	19	16	43	
15: Equal steps in DLT rate	Mean number of patients	0.1	3.13	3.51	5.88	5.06	13.13	
	True DLT rate	0.05	0.1	0.15	0.25	0.2	0.3	
	P(select)	0	0.02	0.12	0.32	0.27	0.26	<0.01
	% of patients	2	13	19	27	22	18	
16: All toxic	Mean number of patients	0.54	4.02	5.93	8.56	6.89	5.75	
	True DLT rate	0.5	0.6	0.65	0.75	0.7	0.8	
	P(select)	0.27	0	0	0	0	0	0.73
	% of patients	56	27	15	2	0	0	
	Mean number of patients	9.01	4.28	2.39	0.38	0.05	0	

and unethical if the majority of patients are over-dosed at the more toxic dose levels. The average number and the percentage of patients at each dose level, for each scenario, is recorded in Tables 2.4 and 2.5.

The percentage of patients treated at the TD25 ranges between 23% and 43% for each scenario under both orderings. The design also allocates the most patients on average to the TD25 apart from in scenario 7. In this case more patients were allocated to the next lowest dose, we have already discussed the

difficulties of this scenario so this characteristic is not too concerning. The mean number of patients recruited for scenarios 1-6 is 26, 30, 32, 33, 34 and 31 respectively. Similarly for scenarios 9-14 its 26, 30, 32, 34, 33 and 31. Even though we allow for up to 60 patients the majority of trials terminate early based on the pre-defined rules for selecting the TD25. This information is presented in Table 2.6 which also shows how often the max sample size is reached from the 10000 trials for each scenario.

TABLE 2.6: Summary of simulated patient numbers for each scenario.

Scenario	Max no. of patients	% max reached	Mean no. of patients
1: TD25 @-1	60	0.21	26.38
2: TD25 @0	60	0.08	29.97
3: TD25 @1	60	0.05	32.01
4: TD25 @2a	60	0.12	33.22
5: TD25 @2b	60	0.06	33.60
6: TD25 @3	60	0.02	30.92
7: Equal steps	60	0.01	31.74
8: All toxic	54	0.01	16.14
9: TD25 @-1	60	0.17	26.24
10: TD25 @0	60	0.11	29.95
11: TD25 @1	60	0.06	32.15
12: TD25 @2a	60	0.07	33.56
13: TD25 @2b	60	0.03	33.16
14: TD25 @3	60	0.08	30.81
15: Equal steps	60	0.02	31.69
16: All toxic	51	0.01	16.11

Overall, the simulation results show the specification of this design performs relatively well in a number of scenarios. We have shown there is a high probability of the trial stopping early if all dose-levels are too toxic. We have also shown the design behaves in an appropriate manner when there is a lack of disparity between dose-levels in terms of toxicity. Finally, we have demonstrated that regardless of the ordering we observe the PO-TITE-CRM has a

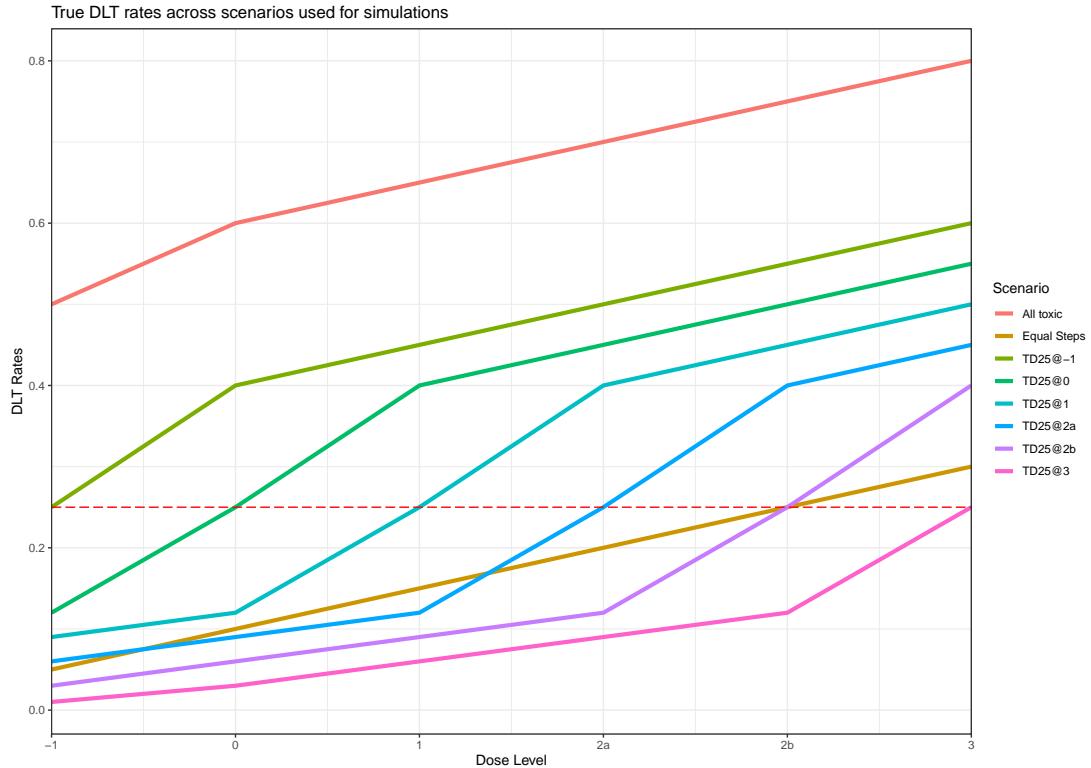
high probability of selecting the correct dose. There are a number of limitations to the operating characteristics presented here which are due to the specification of the simulations and trial design. Section 2.4 explores and discusses these limitations in more detail.

2.4 Exploring other designs

The operating characteristics presented in section 2.3.5 provide an insight into how the trial design operates and its effectiveness at selecting the TD25. However, several factors impact the results seen here. These factors can be grouped into two main categories, limitations with the simulations performed or the trial design.

To simulate various scenarios the true DLT rates are adjusted to reflect the TD25 being at different dose levels. There is no formal process to select these values as such their selection is fairly arbitrary. We set one dose level as the TD25 with lower and higher dose levels set at lower and higher DLT rates respectively. Figure 2.6 illustrates the dose levels for scenarios in Table 2.4 where dose level 2b is more toxic than 2a. The DLT rates cover some possible scenarios and account for a range of plausible values. However, these true DLT rates may not accurately reflect what we observe once the trial begins. Also, the relationship between the rates and the dose levels may not be similar to what we use in the simulations. Multiple other scenarios could be investigated but it would still be impossible to account for all possible variations which may occur. Hence when evaluating the performance of a design it is important to note the scenario in which it is being evaluated and whether or not the design performs as expected and to an adequate level. For ADePT-DDR, the design produces reasonable operating characteristics under each scenario.

FIGURE 2.6: True DLT rates used for each of the scenarios where dose level 2b is more toxic than 2a. The dotted red line represents the target dlt rate of 25% (TD25).



The original methodological papers by Wages et al. [29], [45] only provide simulations for their examples using true DLT rates that are monotonically increasing which represented one of their possible orderings. Initially, in ADePT-DDR we followed suit and only produced simulations under a monotonically increasing DLT rate (order where 2b is more toxic than 2a, Table 2.4). However, as we are unclear on the ordering of 2a and 2b there is a possibility that 2b is less toxic. So those initial simulations would not provide an accurate assessment of the design in that circumstance. This was the main motivation for running scenarios in Table 2.5, in which we see the design performs at a similar level regardless of the partial ordering. ADePT-DDR is a simple case of partial ordering as there are only two possible orderings and six dose-levels. For trials with higher numbers of orderings or dose-levels the number of scenarios that

would have to be evaluated would increase which may be infeasible. Here it may be more beneficial to choose a handful of scenarios from multiple different orderings to cover a wider range of possible outcomes for the trial to assess the design.

There are various features in this trial design that impact how it performs. The partial ordering caused by dose levels 2a and 2b adds complexity to design. If one of these dose-levels were to be removed or a normal ordering was assumed a standard TITE-CRM design could be used instead. However, this would take away from what the trial is trying to discover. This trial also has a long follow-up period due to potential late-onset toxicities and in turn, will have a long duration. The TITE component will allow for the duration to be a lot shorter than it would be otherwise. TITE-CRM designs allow for patients to be recruited sequentially and allocated a dose based on available information from patients already in the trial. The design for ADePT-DDR uses cohorts of three and a minimum follow-up period. The dose-escalation decisions will only be made every third patient after a specific amount of time. This is done for safety and practicality reasons but means that some patients may not be able to enter the trial and it also loses some of the benefits of the TITE-CRM. We also have a sample size of 60 patients but include a stopping rule for when a consensus is reached which means we often do not recruit the maximum sample size. Further simulations were produced to investigate how these features affected the trial. Tables 2.7 and 2.8 compares selection probabilities as well as duration measured in months from the ADePT-DDR trial design with five alternative designs based on the two different orderings. Figures 2.7 and 2.8 visualise the result from each of these tables respectively. 10000 trials were simulated for each scenario which took 62 hours and 22 minutes to complete.

1. TITE-CRM design. This design assumes that partial ordering does not

exist and that dose-level 2b is more toxic than 2a. A TITE-CRM is used instead of a PO-TITE-CRM. All other stopping rules and details remain the same.

2. PO (Partial Ordering design). This design removes the TITE component and uses a PO-CRM as detailed by [45]. This requires the removal of the minimum follow-up period so all dose allocation decisions are made once all 3 patients in a cohort have been observed for the full follow-up period of one year. All other stopping rules and details remain the same.
3. $N = 30$. This design uses a fixed sample size of 30 patients and removes the stopping rule for reaching consensus. The analysis is still conducted using the PO-TITE-CRM. All other stopping rules and details remain the same.
4. $N = 60$. This design uses a fixed sample size of 60 patients and removes the stopping rule for reaching consensus. The analysis is still conducted using the PO-TITE-CRM. All other stopping rules and details remain the same.
5. $CS = 1$. This design uses a cohort size (CS) of one. All other stopping rules remain the same.

The TITE-CRM performs comparably to our original design for certain scenarios, specifically where 2a is assumed less toxic than 2b (Table 2.4). Compared to the PO-TITE design we see increases in probability selection for scenarios 4 and 5 where the target dose is at 2a and 2b respectively. This increase in performance can be attributed to the fact that the partial ordering no longer exists as we have assumed an ordering. The lower selection probabilities for the PO-TITE-CRM can be seen as the price to pay for the uncertainty of not knowing the order of 2a and 2b. However, the TITE-CRM underperforms in

TABLE 2.7: Selection probabilities of the TD25 and expected trial duration (in months) for the PO-TITE, TITE and PO-CRM designs as well as modified PO-TITE-CRM designs for scenarios 1-8 (where 2b is considered more toxic than 2a) based on 10000 simulated trials.

Scenario	CRM details	Prior DLT	Dose Levels						Stop	Duration	Mean N
			-1	0	1	2a	2b	3			
1: TD25 @-1	PO-TITE TITE PO N = 30 N = 60 CS = 1	True DLT rate	0.25	0.4	0.45	0.5	0.55	0.6			
		P(select)	0.68	0.18	0.05	0.01	0	0	0.08	57.61	26.38
		P(select)	0.7	0.21	0.05	0.01	0	0	0.03	59.55	27.46
		P(select)	0.59	0.18	0.04	0.01	0	0	0.19	132.19	22.11
		P(select)	0.67	0.19	0.05	0.01	0	0	0.08	60.02	27.72
		P(select)	0.78	0.12	0.01	0	0	0	0.09	106.61	53.62
		P(select)	0.63	0.17	0.04	0.01	0.01	0	0.15	87.62	22.44
2: TD25 @0	PO-TITE TITE PO N = 30 N = 60 CS = 1	True DLT rate	0.12	0.25	0.4	0.45	0.5	0.55			
		P(select)	0.23	0.51	0.2	0.03	0.02	0	0.01	64.06	29.97
		P(select)	0.22	0.54	0.2	0.04	0.01	0		63.5	29.65
		P(select)	0.2	0.54	0.21	0.02	0.01	0	0.02	163.53	27.79
		P(select)	0.23	0.5	0.21	0.03	0.02	0	0.01	63.26	29.52
		P(select)	0.15	0.69	0.14	0.01	0	0	0.01	116.29	59
		P(select)	0.25	0.46	0.19	0.03	0.02	0.01	0.04	105.41	27.59
3: TD25 @1	PO-TITE TITE PO N = 30 N = 60 CS = 1	True DLT rate	0.09	0.12	0.25	0.4	0.45	0.5			
		P(select)	0.02	0.2	0.55	0.14	0.09	0.01		67.74	32.01
		P(select)	0.01	0.2	0.54	0.2	0.04	0		64.44	30.18
		P(select)	0.02	0.16	0.59	0.14	0.07	0.01		178.12	30.43
		P(select)	0.01	0.19	0.52	0.15	0.12	0.01		64.04	29.96
		P(select)	0	0.14	0.7	0.1	0.06	0		117.89	59.89
		P(select)	0.05	0.18	0.51	0.14	0.09	0.02	0.01	114.18	30.13
4: TD25 @2a	PO-TITE TITE PO N = 30 N = 60 CS = 1	True DLT rate	0.06	0.09	0.12	0.25	0.4	0.45			
		P(select)	0	0.02	0.22	0.48	0.23	0.05		69.91	33.22
		P(select)	0	0.02	0.21	0.56	0.18	0.03		66.99	31.59
		P(select)	0	0.02	0.22	0.49	0.22	0.05		189.69	32.52
		P(select)	0	0.01	0.22	0.45	0.27	0.05		64.1	29.99
		P(select)	0	0	0.18	0.64	0.18	0.01		118.01	59.96
		P(select)	0.03	0.02	0.22	0.45	0.2	0.08		113.77	30.01
5: TD25 @2b	PO-TITE TITE PO N = 30 N = 60 CS = 1	True DLT rate	0.03	0.06	0.09	0.12	0.25	0.4			
		P(select)	0	0	0.02	0.3	0.43	0.25		70.6	33.6
		P(select)	0	0	0.02	0.22	0.56	0.19		68.34	32.34
		P(select)	0	0	0.03	0.26	0.44	0.27		194.41	33.38
		P(select)	0	0	0.03	0.3	0.43	0.24		64.11	29.99
		P(select)	0	0	0	0.24	0.61	0.14		118.08	59.99
		P(select)	0.01	0	0.02	0.26	0.4	0.3		110.26	29
6: TD25 @3	PO-TITE TITE PO N = 30 N = 60 CS = 1	True DLT rate	0.01	0.03	0.06	0.09	0.12	0.25			
		P(select)	0	0	0	0.09	0.13	0.78		65.78	30.92
		P(select)	0	0	0	0.03	0.23	0.73		65.6	30.82
		P(select)	0	0	0	0.08	0.12	0.8		183.48	31.4
		P(select)	0	0	0	0.1	0.14	0.75		64.12	30
		P(select)	0	0	0	0.06	0.1	0.85		118.09	60
		P(select)	0.01	0	0	0.06	0.1	0.83		92.92	23.98
7: Equal steps	PO-TITE TITE PO N = 30 N = 60 CS = 1	True DLT rate	0.05	0.1	0.15	0.2	0.25	0.3			
		P(select)	0	0.03	0.12	0.31	0.28	0.26		67.25	31.74
		P(select)	0	0.03	0.14	0.3	0.32	0.21		65.22	30.61
		P(select)	0.01	0.03	0.14	0.3	0.27	0.25		186.6	31.96
		P(select)	0	0.02	0.12	0.3	0.3	0.25		64.07	29.97
		P(select)	0	0	0.07	0.33	0.35	0.25		118.04	59.97
		P(select)	0.03	0.02	0.09	0.24	0.23	0.39		101.81	26.55
8: All toxic	PO-TITE TITE PO N = 30 N = 60 CS = 1	True DLT rate	0.5	0.6	0.65	0.7	0.75	0.8			
		P(select)	0.26	0	0	0	0	0	0.74	39.19	16.14
		P(select)	0.37	0	0	0	0	0	0.63	44.65	19.17
		P(select)	0.13	0	0	0	0	0	0.86	70.38	10.92
		P(select)	0.23	0	0	0	0	0	0.77	41.35	17.34
		P(select)	0.13	0	0	0	0	0	0.87	47.35	20.68
		P(select)	0.33	0	0	0	0	0	0.67	46.43	10.51

TABLE 2.8: Selection probabilities of the TD25 and expected trial duration (in months) for the PO-TITE, TITE and PO-CRM designs as well as modified PO-TITE-CRM designs for scenarios 9-16 (where 2a is considered more toxic than 2b) based on 10000 simulated trials.

Scenario	CRM details	Prior DLT	Dose Levels						Stop	Duration	Mean N
			-1	0	1	2a	2b	3			
9: TD25 @-1	PO-TITE	True DLT rate	0.25	0.4	0.45	0.55	0.5	0.6			
		P(select)	0.67	0.19	0.05	0	0.01	0	0.08	57.35	26.23
		TITE	0.7	0.21	0.05	0	0	0	0.03	59.34	27.34
		PO	0.58	0.17	0.04	0	0	0	0.2	131.46	21.98
		N = 30	0.68	0.18	0.04	0	0.01	0	0.09	59.85	27.62
		N = 60	0.78	0.13	0.01	0	0	0	0.09	106.5	53.56
		CS = 1	0.62	0.16	0.05	0	0.01	0	0.15	87.6	22.43
10: TD25 @0	PO-TITE	True DLT rate	0.12	0.25	0.4	0.5	0.45	0.55			
		P(select)	0.23	0.52	0.2	0.02	0.02	0	0.01	64.02	29.95
		TITE	0.21	0.56	0.19	0.02	0.01	0		63.57	29.69
		PO	0.2	0.54	0.2	0.01	0.02	0	0.02	163.06	27.7
		N = 30	0.24	0.51	0.2	0.02	0.03	0	0.01	63.35	29.57
		N = 60	0.15	0.7	0.13	0	0.01	0	0.01	116.22	58.96
		CS = 1	0.25	0.47	0.19	0.02	0.03	0.01	0.04	105.04	27.49
11: TD25 @1	PO-TITE	True DLT rate	0.09	0.12	0.25	0.45	0.4	0.5			
		P(select)	0.02	0.2	0.55	0.09	0.14	0.01		68	32.15
		TITE	0.01	0.22	0.58	0.14	0.04	0		64.86	30.41
		PO	0.03	0.17	0.59	0.09	0.12	0.01		177.68	30.35
		N = 30	0.02	0.2	0.51	0.1	0.16	0.01		64.03	29.95
		N = 60	0	0.14	0.71	0.05	0.1	0		117.88	59.88
		CS = 1	0.05	0.17	0.53	0.09	0.13	0.02	0.01	113.98	30.07
12: TD25 @2a	PO-TITE	True DLT rate	0.06	0.09	0.12	0.25	0.15	0.45			
		P(select)	0	0.01	0.08	0.44	0.33	0.14		70.52	33.56
		TITE	0	0.02	0.14	0.26	0.45	0.14		67.23	31.73
		PO	0.01	0.02	0.09	0.45	0.28	0.15		194.42	33.38
		N = 30	0	0.01	0.08	0.42	0.35	0.14		64.09	29.98
		N = 60	0	0	0.03	0.57	0.33	0.07		117.98	59.94
		CS = 1	0.02	0.01	0.07	0.4	0.32	0.18		113.02	29.8
13: TD25 @2b	PO-TITE	True DLT rate	0.03	0.06	0.09	0.35	0.25	0.4			
		P(select)	0	0	0.15	0.31	0.43	0.11		69.8	33.16
		TITE	0	0.01	0.26	0.36	0.28	0.1		65.95	31.02
		PO	0	0.01	0.14	0.34	0.43	0.1		190.68	32.7
		N = 30	0	0	0.15	0.3	0.44	0.11		64.12	30
		N = 60	0	0	0.1	0.29	0.57	0.04		118.07	59.99
		CS = 1	0.02	0	0.14	0.27	0.4	0.17		112.27	29.58
14: TD25 @3	PO-TITE	True DLT rate	0.01	0.03	0.06	0.12	0.09	0.25			
		P(select)	0	0	0	0.13	0.09	0.78		65.58	30.81
		TITE	0	0	0	0.04	0.21	0.74		65.66	30.86
		PO	0	0	0	0.13	0.07	0.8		183.9	31.47
		N = 30	0	0	0	0.13	0.11	0.75		64.12	30
		N = 60	0	0	0	0.09	0.06	0.85		118.09	60
		CS = 1	0.01	0	0	0.1	0.07	0.82		93.1	24.03
15: Equal steps	PO-TITE	True DLT rate	0.05	0.1	0.15	0.25	0.2	0.3			
		P(select)	0	0.02	0.12	0.32	0.27	0.26		67.17	31.69
		TITE	0	0.03	0.19	0.27	0.28	0.24		65.01	30.49
		PO	0	0.03	0.16	0.32	0.26	0.23		186.35	31.92
		N = 30	0	0.02	0.13	0.3	0.29	0.25		64.1	29.99
		N = 60	0	0	0.07	0.36	0.31	0.25		117.97	59.94
		CS = 1	0.03	0.02	0.09	0.23	0.23	0.4		102.22	26.67
16: All toxic	PO-TITE	True DLT rate	0.5	0.6	0.65	0.75	0.7	0.8			
		P(select)	0.27	0	0	0	0	0	0.73	39.14	16.11
		TITE	0.37	0	0	0	0	0	0.63	44.23	18.94
		PO	0.13	0	0	0	0	0	0.87	69.79	10.81
		N = 30	0.23	0	0	0	0	0	0.77	41.18	17.24
		N = 60	0.13	0	0	0	0	0	0.87	47.29	20.64
		CS = 1	0.33	0	0	0	0	0	0.67	45.76	10.31

FIGURE 2.7: Plot of the simulation results presented in Table 2.7 detailing selection probabilities for multiple designs across scenarios 1-8 (where 2b is considered more toxic than 2a).

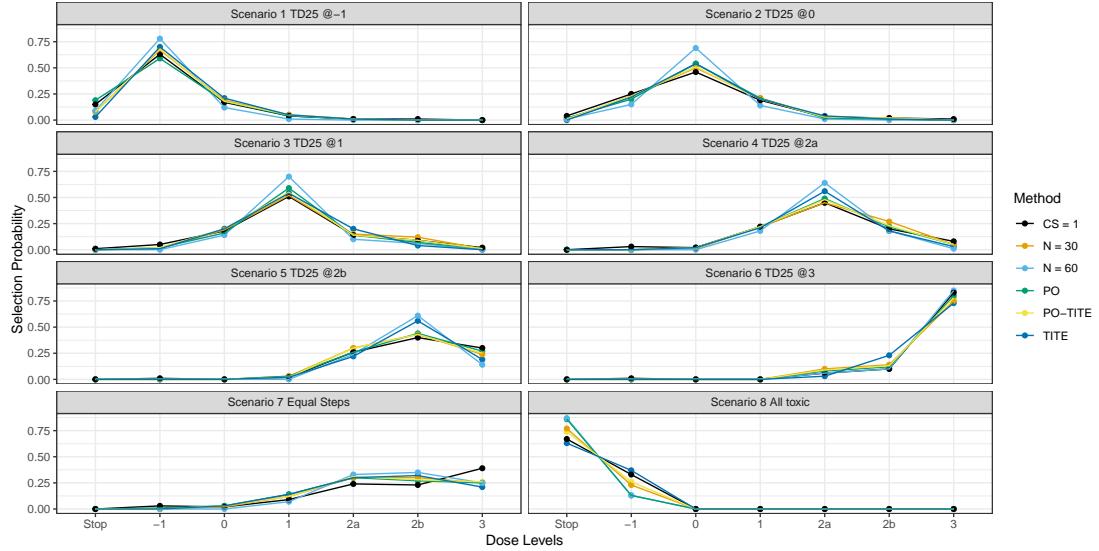
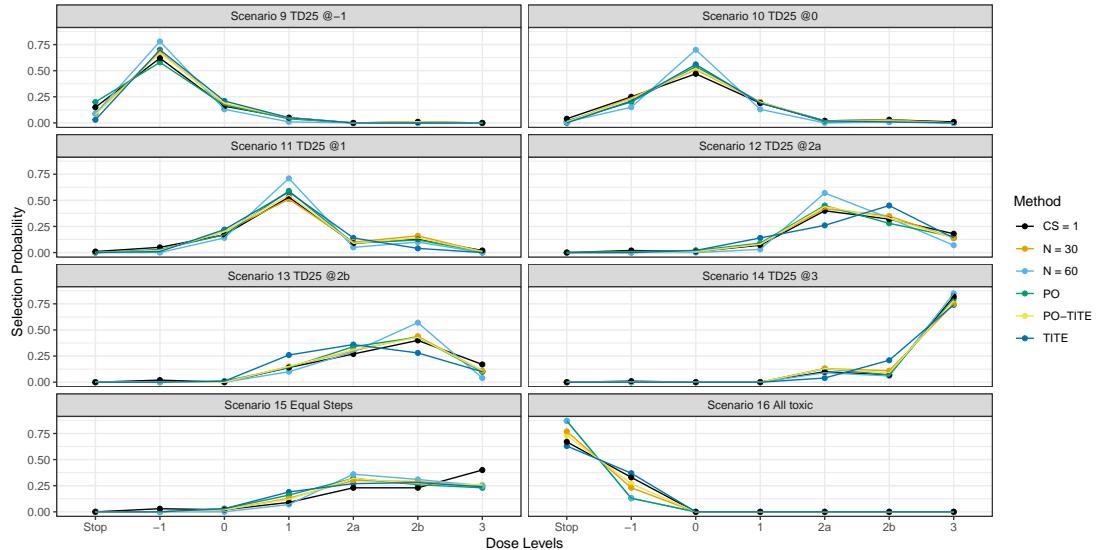


FIGURE 2.8: Plot of the simulation results presented in Table 2.8 detailing selection probabilities for multiple designs across scenarios 9-16 (where 2a is considered more toxic than 2b).



scenarios 9-16 where 2a is assumed more toxic than 2b. Specifically, scenarios 12 and 13 where it fails to identify the TD25 the majority of the time. This is also explored by Abbas et al. [60].

The PO-CRM design without a TITE component also performs similarly

except for scenarios 1, 8, 9 and 16 where the trial stops more regularly for excess toxicity at the lowest dose. This would be because patients complete the full follow-up window before the next dose allocation decision is made. In a TITE setting a new cohort could be recruited before patients in previous cohorts experience a DLT. The main difference between these designs is the trial duration. Without the TITE component the trial duration is significantly longer, with the average length ranging from 70 to 195 months compared to 39 to 71 months for PO-TITE-CRM. In scenario 1 TITE-CRM duration is longer than the PO-TITE-CRM design this can be attributed to the lower chance of stopping early. If the trial is stopping early less it has more chance of going on for longer thus increasing the duration.

The design with a fixed sample size of 30 is comparable to our design with the sample size of 60 and the consensus stopping rule. With a sample size of 30 selection probabilities are only 2-5% lower. For the design with 60 patients, we see much improved operating characteristics with selection probabilities ranging from 31% to 85% for the various scenarios. Even though our original design specifies a sample size of 60 we rarely ever reach it as we often stop for consensus hence why this design performs better. The trade-off here is trial duration. Recruitment and follow-up under the constraints of these simulations will take much longer compared to our specification which is not ideal for an early-phase trial. Originally our design had a fixed sample size of 30 but as the clinicians wanted a dose expansion cohort we opted to use the consensus rule to ensure a minimum number of patients would be treated at the TD25.

For the design with cohort size of one, we see somewhat comparable performance to that of the PO-TITE-CRM design. The design performs similarly for scenarios where the TD25 is at the lowest or highest dose level but underperforms for the more complex scenarios in terms of selection probabilities.

This discrepancy in performance may be related to how the simulations recruit patients into the trial and the large DLT follow-up period, meaning more frequent dose allocation decisions are being made each with less available information. This also leads to the no cohorts design having a longer duration. Patients entered into the trial in cohorts of three will not have to wait the full minimum follow-up period between patients within the cohort.

2.5 Discussion

The PO-CRM and PO-TITE-CRM designs offer solutions to the issue of partial ordering where the order of the treatments is only partially known. The original methodology details that this issue commonly arises in trials of multiple agents, where each drug individually may follow the monotonicity assumption but when combined at certain dose levels this may not hold. This issue is typically dealt with either by fixing the dose of one of the agents and escalating the other or escalating both agents simultaneously. This means certain drug combinations that are clinically relevant may not be investigated or even considered.

Here we have shown that these issues can also arise in other situations. Even though the ADePT-DDR trial uses multiple agents, the issue of partial ordering occurs in this case due to the varying treatment dose and schedule for one of its agents AZD6738. Implementing the PO-TITE-CRM design allowed us to deal with this issue effectively. There may be other factors or variables in single-agent dose-finding trials that would lead to the issue of partial ordering and would warrant the use of either PO-CRM or PO-TITE-CRM. The issue of dose scheduling and partial ordering is further explored by Wages et al. [61] where they propose further methods for dealing with this issue.

The limited literature review that was conducted highlighted that this may be the first instance of the PO-TITE-CRM design being applied. It is important to note that although this methodology takes into account all the various orderings, the main aim is to identify the TD and does not attempt to identify the order that is more correct.

Compared to other CRM-based designs only a few additional pieces of information are required to implement the PO-CRM design. More important is the number of toxicity orderings and prior probabilities for the orders. Dependent on how many dose combinations are available it may not be feasible to investigate all combinations and all orderings. Careful thought and consideration should be given to the combinations and orderings selected which would require input from all relevant investigators. In terms of priors for orderings if no prior information is available, all orders should be treated as equally likely to occur. Extending this design to the PO-TITE-CRM requires a fit-for-purpose weight function and is applied similarly to the TITE-CRM methodology. There is an R package available with functions that can be used to run and simulate a PO-CRM trial. These functions were extended to included weighted dose-toxicity models as described in this chapter to implement PO-TITE-CRM into ADePT-DDR. The lack of available software for PO-TITE-CRM specifically may be one of the reasons for its lack of use.

In terms of ADePT-DDR, dose combinations were decided upon by the clinical investigators. The issue of partial ordering was due to the dose-levels 2a and 2b, as such this methodology was employed to deal with that scenario. This is a very simple example of partial ordering as we only have two possible orderings and six dose levels. The necessity of implementing this methodology was discussed and whether or not adopting an easier solution by simply altering the dose levels would have been better. Ultimately, the dose levels selected by the clinicians were deemed the most relevant with the TD25 likely to

be one of these doses.

Simulations and operating characteristics were the main tools used to assess the design's performance as well as help understand the impact of sample size and stopping rules. This was an iterative process that involved running multiple iterations of simulations under various scenarios until the design was finalised. A key point is that scenarios from simulations should account for each of the possible orderings. ADePT-DDR only has two orderings, we ran scenarios for both. For a trial with a greater number of orderings, this may be unfeasible but at least some scenarios should be assessed to ensure the design is behaving as expected. Overall, the design operating characteristics performed reasonably well even in difficult scenarios.

One limitation of the simulations is how the time-to-event data is generated. The time of DLTs is sampled from a uniform distribution $U(0, 413)$, where the time of the DLT can occur at any time between the patient beginning treatment and the end of follow-up (413 days). Using this uniform distribution implies that a DLT has an equal probability of occurring at any time-point in the observation window. This may not be an accurate representation of what happens in the actual trial. Similar comments can be made about the accrual rate used in the simulations. Here we specified the recruitment of one patient per month which is in no way guaranteed for the actual trial. Wages et al. [29], when presenting this methodology investigated four different applications of the PO-TITE-CRM which used different models to enroll patients and allocate DLTs. Results across these four applications were comparable.

The simulations are also able to instantaneously determine dose-levels for incoming cohorts with all available information. This does not fully reflect the process in which dose-escalation decisions would be made during the actual running of the trial. The analysis would require a data snapshot and time would have to be spent cleaning the data and determining the next dose-level.

This would mean any data from the point of the snapshot would not be included in any dose escalation/de-escalation decisions.

Similarly, there may also be limitations with some of the design choices made concerning cohort size and sample size. These were investigated alongside a variety of other trial designs that could have been implemented. This was done to validate the choices we made with the design and highlight the differences in operating characteristics due to the varying assumptions and components in the designs. The standard PO-CRM had a much longer average duration due to the lack of TITE component whereas a standard TITE-CRM overall performs better but assumes the ordering of toxicity is known.

Here we have detailed the issue of partial ordering with a time-to-event component. We discuss how we implemented the trial design, in what we believe is the first real-world application of this specific design. A large amount of simulation work is required to assess the performance of the design. This is often an iterative process to refine decisions that were made and often requires input from both clinical and statistical investigators. We recommend running several varied scenarios for each potential ordering that will be investigated. Finally, we also compared the implementation of PO-TITE-CRM to various other designs and showed it performs relatively well given all of the methodological and practical challenges.

Chapter 3

Extension to a Phase I/II trial design

3.1 Introduction

In Chapters 1 and 2 we discuss the main aim of a Phase I is to establish the maximum tolerated dose (MTD) for a treatment under investigation. Model-based designs such as the continual reassessment method (CRM) [13] operate with dose selection and escalation decisions being determined by the occurrence of toxicities. These designs also operate under the cytotoxic assumption which assumes the most toxic dose is also the most efficacious. Subsequent Phase II trials aim to assess the efficacy of the treatment at the recommended dose (MTD). Usually, these two phases are conducted independently of each other and as such, the ability to share information across the phases is somewhat lost.

For treatments like chemotherapy which kills all cells including cancer cells [62], the cytotoxic assumption is valid. However, the emergence of modern treatments such as immunotherapy and molecular targeted agents challenges this paradigm. Immunotherapy is a form of treatment that utilises the body's immune system to fight cancer [63]. Molecular targeted agents (MTA) work by interfering with specific molecules responsible for the growth, spread, and progression of cancer [64]. The monotonic assumption of dose-efficacy may

not hold for these new types of treatments. Furthermore, these treatments, in general, are less toxic than traditional cytotoxic agents such as chemotherapy therefore it is possible the most efficacious dose may occur at a dose-level below the MTD [65]. This produces some methodological challenges for dose-finding trials. Instead of trying to identify the MTD, the goal would be to determine the optimal biological dose (OBD). Depending on the aims of the trial and the design implemented the definition of the OBD may vary. The OBD could be a dose that provides the maximum probability of efficacy with the probability of toxicity being less than a pre-defined target value, or the dose that has a beneficial trade-off between toxicity and efficacy. To determine an optimal dose both toxicity and efficacy outcomes need to be considered and this leads to a need for joint phase I/II trial designs.

Here we will briefly explore some of these designs. This is not an extensive review of the literature but just an illustration of these types of trial design.

Braun [66] proposed the bivariate continual reassessment method (bCRM), an extension to the CRM which incorporates competing outcomes for both toxicity and disease progression. The design models the probabilities of toxicity and progression independently, it is suggested that either empiric, logistic, or hyperbolic tangent functions are used dependent on their biological plausibility. Both outcomes are then combined into a joint distribution which is used to estimate posterior means based on priors and observed data.

Thall & Cook [67] developed EffTox, a Bayesian adaptive dose-finding trial based on trade-offs between the probabilities of toxicity and efficacy. Marginal probabilities of efficacy and toxicity at each dose are modelled and used with utility contours to determine the desirability of each dose based on posterior probabilities of efficacy and toxicity [68].

Zhou et al. [69] introduced a Utility-based Bayesian Optimal Interval (U-BOIN) phase I/II design to identify the OBD. This design is an extension of

the Bayesian optimal interval (BOIN) design for phase I trials developed by Liu and Yuan [46]. U-BOIN jointly models toxicity and efficacy with a multinomial-Dirichlet model and uses a utility function to measure the dose risk-benefit trade-off. The design consists of two seamless stages. Firstly, in stage I the BOIN design is used to explore the dose levels and determine a set of admissible doses and collect preliminary efficacy data. In stage II, posterior estimates of utility for each dose are continuously updated after each cohort using toxicity and efficacy data from both stages.

Zhang et al. [70] introduced the trivariate CRM (TriCRM) design. The design considers patients to have one of three possible outcomes: no efficacy and toxicity, efficacy without toxicity, and toxicity. These outcomes are then modelled using a continuation-ratio model. A Bayesian approach and dose-finding algorithm are then used to identify the OBD similar to the CRM.

Anathakrishnan et al. [71] produced extensions to the modified Toxicity Probability Interval (mTPI) design by Ji & Wang [72] and Toxicity Equivalent Range (TEQR) design by Blanchard & Longmate [73] to include efficacy outcomes. In both designs, isotonic regression is applied to the observed DLT rates at the end of the trial. Dependent on the shapes of the dose-response curves and the underlying response rates, isotonic regression is applied to the observed response rates or the differences in observed response rates to determine the optimal dose.

Riviere et al. [74] developed a Bayesian dose-finding design for MTA. The design works on the premise that for MTA efficacy initially increases with dose then eventually plateaus. They use a logistic model with a plateau parameter to capture the dose-level at which plateaus begin in the dose-efficacy relationship. A weighted likelihood approach is also used to accommodate for any potential late-onset toxicities. This methodology incorporates adaptive randomisation to allocate patients to the dose-level closest to the likely plateau

point.

Mozgunov et al. [75] discuss randomised dose-escalation designs for drug combination trials with immunotherapy. Here they investigate the use of a 4 parameter EMAX model with a randomisation scheme. The use of a four parameter model allows for toxicity risk at the standard of care to be modelled independently, modelling a plateau at a particular level and a steep increase in toxicity.

This chapter revolves around the seamless phase I/II dose-finding adaptive design by Wages and Tait [30], which we will refer to as the WT design. This design models toxicity and efficacy independently. To model the probability of efficacy, a set of possible efficacy skeletons are considered which would correspond to plausible dose-efficacy relationships. For the class of dose-efficacy models, a single parameter model is used similar to the empiric model of the CRM. The authors recommend that $(2n - 1)$ efficacy skeletons are specified where n is the number of doses being investigated. Toxicity is modelled using a CRM approach with an empiric model that requires a skeleton for toxicity. The dose-finding operates in two stages the adaptive randomisation (AR) phase and the maximisation phase. In the AR phase patients are adaptively randomised amongst a set of tolerable doses (determined by the CRM toxicity model), where probabilities of randomisation to each dose are proportional to their posterior probabilities of efficacy. A pre-defined number of patients enter the AR phase and once recruitment has been completed we move to the maximisation phase. In this phase, patients are allocated to the dose in the tolerable set which maximises the probability of efficacy.

The incorporation of an AR phase early on into the trial is beneficial since there may be a lack of data to rely on decisions made by the maximisation of efficacy probabilities. Also, there may be doses that have not been tested and randomisation allows for information to be collected from these. It also helps

avoid getting stuck repeatedly recruiting to the same dose and allows for a more broad understanding of the dose-efficacy and toxicity relationships. One extension that we propose is the inclusion of randomisation to a control arm in the design. This would provide a set of patients who receive standard of care to act as controls and allow for comparisons to be made with outcomes from patients receiving the OBD. There is also the added benefit of being able to include standard of care into the models to get a better understanding of the dose-efficacy and toxicity relationships.

Section 3.2, details the statistical aspects of the WT design and how it works. We introduce our extension to the design to include randomisation to control in Section 3.3. Section 3.4 evaluates the performance of the new design with a simulation study. Section 3.5 explores how efficient our design would be at performing an efficacy test. Finally, we finish with a discussion in Section 3.6.

3.2 The Wages and Tait design

In this section, we detail the Wages and Tait design. Here we have used slightly different notation than first introduced in Chapter 2, this was to ensure that the design we present here is as specified in the original paper [30] and also to make it simpler when cross referencing with available code. A set of I doses under investigation can be denoted as $\mathcal{D} = \{d_1, \dots, d_i\}$. For each patient j entered into the trial they are allocated to a dose-level and joint outcomes for toxicity and efficacy are measured. The dose for the j th patient, $X_j, j = 1, \dots, n$ can be thought of as random, taking values $x_j \in \mathcal{D}$. Let Y_j and Z_j be the random variables for binary toxicity and efficacy events respectively. For an individual patient j , toxicity and efficacy outcomes can take values $y_j, z_j \in \{0, 1\}$ where 0 indicates an event did not happen and 1 indicates that it did.

Wages and Tait [30] utilise the CRM approach of O’Quigley et al. [13] to model toxicity. A univariate Bayesian method is used which begins by assuming a monotonically increasing dose-toxicity curve. The DLT probabilities, $\pi_T(d_i)$, are modelled at each dose level i where $i = 1, \dots, I$. The power model is specifically used by Wages and Tait in this design given by

$$F(d_i, \beta) = p_i^{\exp(\beta)}. \quad (3.1)$$

A working model or skeleton containing the prior beliefs of toxicity at each dose-level is required in the form $0 < p_1 < \dots < p_I < 1$. For the single parameter in the power model β we assume it has a prior distribution $g(\beta)$. After the inclusion of j subjects into the trial, we have data in the form of $\Omega_j = \{(x_1, y_1, z_1), \dots, (x_j, y_j, z_j)\}$. The toxicity data can be used with Equation 3.1 to give the likelihood for β

$$L(\beta|\Omega_j) = \prod_{l=1}^j \{F(x_l, \beta)\}^{y_l} \{1 - F(x_l, \beta)\}^{1-y_l}, \quad (3.2)$$

the posterior density for β can be calculated using

$$P(\beta|\Omega_j) = \frac{L(\beta|\Omega_j)g(\beta)}{\int_{-\infty}^{\infty} L(\beta|\Omega_j)g(\beta)d\beta}. \quad (3.3)$$

This can then be used to establish the posterior mean of β

$$\hat{\beta}_j = \int_{-\infty}^{\infty} \beta P(\beta|\Omega_j)d\beta. \quad (3.4)$$

Using $\hat{\beta}_j$ estimates of DLT probabilities at each dose level can be obtained via

$$\hat{\pi}_T(d_i) = F(d_i, \hat{\beta}_j) = p_i^{\exp(\hat{\beta}_j)}. \quad (3.5)$$

For a specific maximum acceptable toxicity rate, ϕ_T a set of acceptable or admissible doses can be declared as follows

$$\mathcal{A}_j = \{d_i : \hat{\pi}_T(d_i) \leq \phi_T; i = 1, \dots, I\}. \quad (3.6)$$

To model efficacy, a Bayesian approach is taken similar to how toxicity was modelled but rather than using a singular working model a class of working models is considered. They use a class of skeletons that correspond to various dose-efficacy relationships. These relationships might be monotonically increasing (as the dose-levels increase efficacy increases), unimodal (initially increasing then decreasing) or plateau (initially increase then level off). As a guide, it is suggested that $(2I - 1)$ working models should be specified. The probability of an efficacious response at dose d_i is denoted as $\pi_E(d_i)$. The primary aim of the trial is to identify the optimal dose from doses considered safe $d_v \in \mathcal{D}$ which is defined such that

$$\pi_E(d_1) \leq \dots \leq \pi_E(d_v) \geq \dots \geq \pi_E(d_I). \quad (3.7)$$

Let K denote the number of efficacy skeletons being used. Then for each skeleton k we have $0 < q_{1k} < \dots < q_{Ik} < 1$ and for a particular skeleton $k; k = 1, \dots, K$ the true probability of efficacious response $\pi_E(d_i)$ at d_i is modelled by

$$\pi_E(d_i) = Pr(Z_j = 1|d_i) \approx G_k(d_i, \theta) = q_{ik}^{exp(\theta)}. \quad (3.8)$$

As with the modelling of toxicity the power model is used again. Similarly as with β a prior distribution $h(\theta)$ is assumed for θ . For both the toxicity and efficacy models a Normal prior is used as first suggested by O'Quigley and Shen [58] such that $\beta, \theta \sim N(0, 1.34)$. Additionally for the modelling of efficacy prior information regarding the plausibility of each model is taken into account

using a weight function $v(k) = \{v(1), \dots, v(K)\}$, where $v(k) \geq 0$ and where $\sum_k v(k) = 1$. If no information is available a discrete uniform distribution can be specified for $v(k)$. After j patients have been included and observed in the study we have efficacy data from Ω_j and the likelihood model under k is given by

$$L(\theta|\Omega_j) = \prod_{l=1}^j \{G_k(x_l, \theta)\}^{z_l} \{1 - G_k(x_l, \theta)\}^{1-z_l}, \quad (3.9)$$

the posterior density is

$$P(\theta|\Omega_j) = \frac{L(\theta|\Omega_j)h(\theta)}{\int_{-\infty}^{\infty} L(\theta|\Omega_j)h(\theta)d\theta}, \quad (3.10)$$

and under skeleton k the posterior mean is given by

$$\hat{\theta}_{jk} = \int_{-\infty}^{\infty} \theta P(\theta|\Omega_j)d\theta. \quad (3.11)$$

This information can be used to establish posterior model probabilities

$$w(k|\Omega_j) = \frac{v(k) \int_{-\infty}^{\infty} L_k(\theta|\Omega_j)h(\theta)d\theta}{\sum_{k=1}^K v(k) \int_{-\infty}^{\infty} L_k(\theta|\Omega_j)h(\theta)d\theta}. \quad (3.12)$$

The posterior model probabilities are then used to determine which skeleton will be selected to model the dose-efficacy relationship. Each time a new patient is to be entered into the study and a dose-escalation decision needs to be made, the skeleton k^* with the largest posterior probability is selected such that

$$k^* = \arg \max_k w(k|\Omega_j). \quad (3.13)$$

After determining the best skeleton and calculating the posterior mean of θ estimates of efficacy probabilities are then generated for each dose.

$$\hat{\pi}_E(d_i) = G_{k^*}(d_i, \hat{\theta}_{jk^*}) \quad (3.14)$$

Dose-finding is conducted in two stages. The first stage begins with the adaptive randomisation (AR) phase. Here the next dose is randomly selected from the set of admissible doses determined by the CRM toxicity model. Randomisation probabilities for each dose are proportional to $\hat{\pi}_E(d_i)$ so that doses with higher estimated efficacy are more likely to be assigned to patients. For doses in \mathcal{A}_j their adaptive randomisation probability R_i is

$$R_i = \frac{\hat{\pi}_E(d_i)}{\sum_{d_i \in \mathcal{A}_j} \hat{\pi}_E(d_i)}. \quad (3.15)$$

The AR phase lasts for a subset of j_R patients such that $j_R \leq J$, where J is the total number of patients to be entered into the trial. Wages and Tait suggest as a general rule of thumb to allocate 50% of patients to both stages. It was shown that this approach works well in a variety of scenarios. However, this can be easily be adapted to suit individual trials.

Once the AR phase has been completed the design switches to the second stage called the maximisation phase. Here the next dose is the dose from the admissible set with the highest estimated probability of efficacy. For a dose-escalation decision that needs to be made in the maximisation phase for the $(j + 1)$ th patient the dose x_{j+1} is selected from the admissible set of doses \mathcal{A}_j with the highest estimated efficacy probability $\hat{\pi}_E(d_i)$ i.e.

$$x_{j+1} = \arg \max_{d_i \in \mathcal{A}_j} \hat{\pi}_E(d_i) \quad (3.16)$$

The design also incorporates stopping rules for safety and futility. The safety rule stops the trial if too much toxicity is observed at the lowest dose

level. This rule is applied throughout the trial for each dose-escalation decision. Exact binomial 95% confidence intervals are calculated for the lowest dose. The lower bound of the interval is then compared to the acceptable toxicity rate ϕ_T . If the lower bound interval is greater than the acceptable rate it can be said that the treatment is too toxic to warrant further investigation. Patients need to have been observed at the lowest dose for this rule to trigger, if there is no data available at the lowest dose the binomial confidence interval is effectively 0.

The futility rule stops the trial if there are too few observed efficacy events. This rule only comes into play during the maximisation phase. This rule uses a similar method to the stopping rule by utilising binomial 95% confidence intervals. During the maximisation phase, the dose with the highest probability of efficacy is selected. At this point, the 95% binomial confidence interval is calculated for the current dose and if the upper bound is less than the futility threshold ϕ_E the trial is stopped as the treatment is ineffectual at all doses. Although 95% confidence intervals are used by Wages and Tait these can be altered accordingly.

3.3 RtC-WT: An extension to the Wages and Tait design

In this section, we introduce our proposed extension to the Wages and Tait (WT) design named Randomisation to Control Wages and Tait (RtC-WT). As the name states, the design will allow investigators to utilise the WT design with the ability to recruit patients to a control arm/dose-level. This idea was initially conceived by Kristian Brock (KB) whilst working on the design of a new dose-finding trial.

3.3.1 The Rationale for Incorporating Randomisation to Control

Typically, seamless phase I/II trial designs perform the tasks of phase I and phase II trials. However, they do not replace the need for randomised phase II trials entirely where preliminary efficacy data is collected on an experimental treatment versus control to determine the need for a larger phase III study [76]. This is our main motivation for introducing RtC-WT. By introducing the ability to randomise to control in the Wages and Tait method we can achieve similar objectives to randomised phase II studies.

An example of where this design may be beneficial is in the investigation of a standard of care treatment in combination with an experimental treatment. The standard of care treatment could be included as the control dose and should have a well-understood toxicity and efficacy profile which could be incorporated into the toxicity and efficacy skeletons. Further dose levels would also receive standard of care along with increasing levels of the experimental treatment, here the interaction between the two treatments in terms of toxicity and efficacy could be investigated and an OBD could be found using the RtC-WT design.

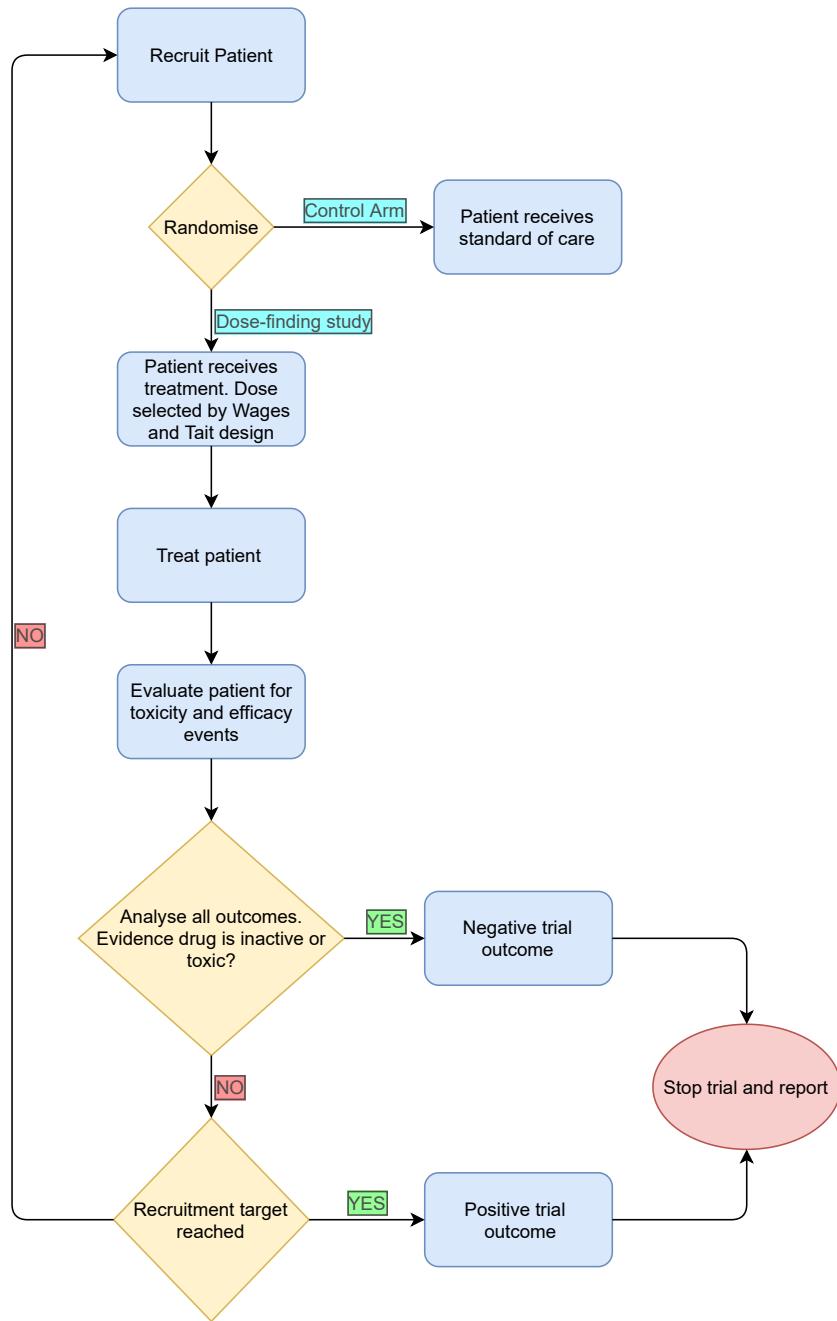
As a seamless phase I/II design WT is relatively simple and effective. The familiarity of using a CRM design to model toxicity and naturally extending that methodology to model efficacy with multiple working models means the design is not particularly difficult to implement. The mathematics behind the design is also not too intense so extensive computation will not be necessary. Given some effort, this design could be implemented in a variety of programming languages although Brock offers easy implementation of this design in his R package escalation [21]. Considering all these factors, extensions to this design can easily be executed without too many obstacles.

The WT design can be considered fairly unique due to its use of adaptive randomisation. Whilst adaptive randomisation is not the core focus of the design it is still a distinguishing factor that could be leveraged to help investigators answer questions other designs are unable to. Specifically, the randomisation allows for more dose-levels to be explored and perhaps obtain a better understanding of the dose-toxicity and efficacy relationships.

Conceptually the WT design could include a control arm without any modification to the design. All that this would require is the addition of a new dose-level at which patients receive control treatment/standard of care. This would need to be implemented as the lowest dose-level as dose-levels still need to obey the monotonicity assumption for toxicity. The issue with taking this approach is that the design is unlikely to allocate patients to the control dose-level. Even though adaptive randomisation is being used the randomisation probabilities are based on estimates of efficacy probabilities and control patients may be unexpected to have an efficacious event. This is a desirable characteristic when investigating treatments as we do not want to allocate too many patients to inefficacious doses. However, if the aim is to establish a cohort of patients as controls to facilitate comparisons to the OBD this is not an optimal characteristic.

There is another approach that could also be used to include a control arm rather than our proposed design RtC-WT. A two-arm randomised design could be used where patients are allocated to either a control arm or a dose-finding arm. Those patients allocated in the dose-finding arm will then be a part of the WT design (see Figure 3.1). This approach maintains many of the traditional qualities of a two-arm randomised trial. The number of patients in each arm can be specified this way and we guarantee a minimum number of patients in our control arm. Also, the characteristics of patients in both arms are likely to be similar which would be beneficial when making comparisons

FIGURE 3.1: Flowchart of how a two arm randomised dose-finding design would operate using the Wages and Tait design.



between the two arms. A downside of this method is that the data for control patients are no longer included in the modelling process. Whilst control patients may still be observed for efficacious and toxic events these will not be included in the modelling. As such the ability to make inferences on the

dose-toxicity and efficacy relationships in reference to a control/ standard of care dose is lost.

Both of these approaches have their merit but also have flaws as well. RtC-WT is somewhat of a middle ground that aims to recruit patients to a control dose and include the control patients' data in the modelling process all whilst maintaining reasonable operating characteristics. We detail RtC-WT in Section 3.3.2 and explore the operating characteristics of this design in Section 3.4.

3.3.2 Design of the Proposed Extension RtC-WT

With this extension, much of the Wages and Tait design stays the same. The modification only impacts the adaptive randomisation (AR) phase and requires some additional specifications at the start of the trial. Firstly, we set the lowest dose-level d_1 to be the control dose-level. This dose-level should be included in the working models for efficacy and toxicity and should be treated like any other dose-level. Even if toxicity and efficacy events are expected to be non-existent for control, their corresponding skeleton values must be non-zero. Investigators also need to consider a randomisation probability ϕ_R for the control dose. During the AR phase, ϕ_R represents the probability of selecting the control dose as the next dose level. The probability of randomisation R_i for other dose in \mathcal{A}_j is scaled accordingly such that the $\sum_{d_i \in \mathcal{A}_j} R_i = 1$. The adaptive randomisation probabilities can now be expressed as

$$R_1 = \phi_R, \quad (3.17)$$

$$R_i = (1 - \phi_R) \frac{\hat{\pi}_E(d_i)}{\sum_{d_i \in \mathcal{A}_j} \hat{\pi}_E(d_i)}, \quad i = 2, \dots, I. \quad (3.18)$$

Compared to Equation 3.15 the adaptive randomisation probability is fixed to ϕ_R at the lowest dose (the control dose) and for all other dose levels in the admissible set \mathcal{A}_j a scaled randomisation probability is calculated. By fixing the probability for the control dose we guarantee a greater chance of patients being allocated to this dose-level. Although estimates of efficacy at the control dose-level $\hat{\pi}_E(d_1)$ do not directly impact its associated randomisation probability, the efficacy data that generated the estimate is still included in the efficacy modelling and impacts probabilities for the remaining dose-levels. Also, by scaling the remaining probabilities of dose-levels in the admissible set we ensure that those doses with high estimates of efficacy maintain their proportional advantage of selection over the other non-control doses.

Some adjustments were made to the stopping rule for safety. The WT design assesses the lower bound of the 95% binomial confidence interval of DLT rate for the lowest dose to determine whether or not the trial should be stopped. However, with the RtC-WT design since the lowest dose is the control, it makes little sense to surmise treatment is toxic here since none of the patients on control would have received the experimental treatment. It is also likely the trial would never recommend stopping even if the treatment is toxic since patients on control are unlikely to experience excess toxic events. The RtC-WT design stops for safety by checking for excess toxicity at the second-lowest dose-level (the first treatment dose-level).

Once the AR phase ends, dose-levels are no longer selected by adaptive randomisation. At this point, it will be difficult for patients to be recruited to the control dose since recommended doses will be based on those with the greatest estimates of efficacy. As such it is important to consider the values set for both your randomisation probability for control ϕ_R and the size of the AR phase j_R . Wages and Tait simply suggest a 50:50 split between both the AR phase and the maximisation phase and show relatively good performance

at this level. However, for RtC-WT the AR phase is the main component and more thought should be given here. In the next section, we explore multiple combinations to better understand how these choices impact the operating characteristics of the design. We also compare RtC-WT to the two alternative designs mentioned in section 3.3.1 via simulations and the inspection of operating characteristics specifically, the selection probabilities of the OBD and patient allocation numbers at each dose-level.

3.4 Evaluation and exploration of the extension via simulations

In this section, we evaluate the performance of RtC-WT in comparison to the two alternative designs mentioned in Section 3.3.1. We also explore the impact of changing the probability of randomisation to control and the number of patients included in the adaptive randomisation phase. These will both be assessed via simulation and inspection of their operating characteristics. To facilitate simulations, a generic trial example will be utilised along with a variety of scenarios. Code for the original Wages and Tait design was provided to us by Nolan Wages. We then altered this code to work for our new modified design RtC-WT.

3.4.1 Design Specification

Here we detail the design specifications for RtC-WT that we will be using throughout this section. We assume five dose-levels, where the lowest dose is considered to be the control dose-level. The maximum sample size of the trial is set at 60 with patients recruited in cohorts of three and the first cohort starting at dose-level two (the first treatment dose-level). The pre-specified toxicity

upper bound and efficacy lower bound are set at $\phi_T = 0.35$ and $\phi_E = 0.50$ respectively. Toxicity and efficacy skeletons, p_i and q_i respectively, are presented in Table 3.1. In terms of efficacy relationships monotonic, unimodal and plateau skeletons were all used. We assume that each of the seven efficacy skeletons is equally likely and set $v(k) = \frac{1}{7}$.

TABLE 3.1: Toxicity and efficacy skeletons for RtC-WT in the example trial.

Skeleton	Dose-levels				
	1	2	3	4	5
p_i	0.1	0.15	0.25	0.35	0.45
q_{i1}	0.3	0.7	0.6	0.5	0.4
q_{i2}	0.3	0.6	0.7	0.6	0.5
q_{i3}	0.3	0.5	0.6	0.7	0.6
q_{i4}	0.3	0.4	0.5	0.6	0.7
q_{i5}	0.3	0.5	0.6	0.7	0.7
q_{i6}	0.3	0.6	0.7	0.7	0.7
q_{i7}	0.3	0.7	0.7	0.7	0.7

For the control dose, we have set our prior beliefs for the probability of toxicity at 10% and the probability of efficacy at 30%. These values can be altered if there is reason to believe that the control dose may have higher or lower probabilities of toxicity and efficacy.

Wages and Tait recommend $(2I - 1)$ efficacy skeletons be used which in this example would be nine, however, we have only considered seven. As there are only four active doses and we are assuming we understand the control dose in terms of toxicity and probability then seven different skeletons fits in the Wages and Tait's recommendation. As we are investigating control plus some experimental treatment it is implausible to have scenarios where efficacy falls as dose increases. So, we have removed the two efficacy skeletons with dose-efficacy relationships that suggest the lowest dose would be the most efficacious. For completeness, the first extra skeleton would be unimodal with the highest efficacy occurring at dose-level one (i.e. 0.7, 0.6, 0.5, 0.4, 0.3 for

dose-levels 1-5) and the second skeleton would be a plateau relationship with the plateau beginning at dose-level one (i.e. 0.7, 0.7, 0.7, 0.7, 0.7 for dose-levels 1-5).

We also include the same stopping rules for safety and futility with the safety rule assessing toxicity at dose-level two. A rule will also be implemented to prevent the skipping of untried doses when escalating. This rule does not apply when de-escalating.

The two parameters we have left to specify are the fixed adaptive randomisation probability for control ϕ_R and the number of patients included in the adaptive randomisation phase j_R . In Section 3.3.2 we briefly discussed the importance of giving thought when setting these values. This is due to the fact they are the main factors driving how RtC-WT works compared to the standard WT design. For example, one could set the AR phase to last for the whole trial and keep a relatively low probability to randomise to control. Alternatively, the AR phase can be set for half the patients in the trial and double the probability of randomisation could be used. These two approaches could allocate the same number of patients in the control arm but have different operating characteristics. It could be hypothesised that by setting the AR phase for the whole trial you miss out on the maximisation phase where patients are allocated to the estimated most efficacious dose which could yield slightly worse operating characteristics. We explore different combinations of these parameters in the next section.

3.4.2 Impact of AR phase size and probability of randomisation to control on RtC-WT

The effect of adjusting the probability of randomising to control ϕ_R is fairly intuitive, as the probability increases the percentage chance that patients are

allocated to the control dose-level also increases. However, this is only in isolation without considering the size of the AR phase. Increasing the AR phase would also mean more patients are likely allocated to the control dose-level since the randomisation only occurs in the AR phase. The interest lies within the interaction of both of these components and their impacts on operating characteristics. To gain a better understanding of this impact on RtC-WT we consider multiple combinations.

We look at two different probabilities for randomisation to control, $\phi_R = 0.2$ and $\phi_R = 0.33$ i.e. 20% and 33% probability of patients being allocated to the control dose-level during the AR phase of the trial. We also consider varying AR phase sizes, specifically $j_R = 0, 15, 30, 45, 60$ essentially looking at when the AR phase lasts 0%, 25%, 50%, 75% and 100% of the trial. The inclusion of setting the AR phase as 0 is somewhat counter-intuitive since the trial will just be run using the maximisation phase where the most efficacious doses are allocated. As such it is unlikely that the control dose-level would ever be the most efficacious specifically in our scenario here. However, its inclusion will serve as a benchmark as the design most likely to achieve optimal performance in terms of locating the OBD since there will be no randomisation and the estimated most efficacious dose will always be the one being tested. Similarly, by setting the AR phase at 60 we limit some of the designs features by never entering the maximisation phase to select dose levels based on efficacy. Also, the stopping rule for futility does not come into play. Although, many more combinations could be explored this set provide a good basis for us to gain a better understanding of how RtC-WT works. It also helps us understand how best to optimise RtC-WT for comparisons with alternative designs later on.

To compare these different combinations we use simulations covering a wide range of scenarios. For each scenario, we simulate 10000 trials each consisting of 60 patients recruited in cohorts of three. Patient outcomes for toxicity

and efficacy are randomly sampled using true toxicity and true efficacy probabilities, these are assumed to be independent of each other. Dose-allocation decisions are made after each cohort of patients and then the subsequent cohort is allocated the recommended dose. The trial could also be stopped if the recruitment target is reached or if any of the stopping rules are triggered. The rest of the design specification is as defined in Section 3.4.1.

The true toxicity and efficacy probabilities are manipulated to produce each scenario for the simulations. Table 3.2 shows a summary of the scenarios that will be used. We look at a combination of five different efficacy curves with three toxicity curves giving 15 scenarios altogether. The five dose-efficacy relationships we consider are; monotonically increasing, unimodal (at dose level 3), plateau (starting at dose level 3), monotonically decreasing and finally no efficacy. For toxicity we look at scenarios where all doses are tolerable, all doses are toxic and a scenario where only higher doses (dose-levels 4 and 5) are toxic. We also list which doses would be considered the OBD under the designs specification along with which doses would be good for each of the scenarios. The OBD in this context would be the dose which maximises efficacy whilst not breaching the toxicity limit. Good doses are those which are considered safe (probability of toxicity $\leq 35\%$) and efficacious (probability of efficacy $\geq 50\%$). For scenarios that are too toxic/lack efficacy, we would expect the trial to stop early, here we have labelled the OBD as being the probability of stopping and good doses as the probability of stopping and selecting dose-level 1, the control dose. Whilst allocating patients to dose-level 1 in these scenarios is not necessarily a bad thing it would likely mean more patients are exposed to the toxic/inefficacious doses which is not optimal, hence the distinction.

Operating characteristics for the scenarios under investigation are given in Table 3.3. The table provides the following operating characteristics:

- $P(OBD)$ - Probability of selecting the OBD
- $P(Good)$ - Probability of selecting a good dose
- $N(OBD)$ - Mean number of patients treated at the OBD
- $N(Good)$ - Mean number of patients treated at good doses
- $N(Control)$ - Mean number of patients treated at the control dose (dose-level 1)

These are provided for each scenario under the 10 different parameterisations of ϕ_R and j_R . For certain scenarios, where the ideal outcome would be to stop early, $N(OBD)$ is left blank as patients are not allocated to a specific dose. Also, for these scenarios $N(Good)$ and $N(Control)$ are the same as the only good dose patients can be allocated to is the control. Then for scenarios where there is only one good dose-level that would be the OBD as well so $P(OBD)$ and $P(Good)$ would be the same as would $N(OBD)$ and $N(Good)$.

TABLE 3.2: Summary of the efficacy and toxicity curves used in each scenario.

Scenario	1	2	3	4	5	Description	OBD	Good Dose	
1	tox	0.1	0.2	0.25	0.3	0.35	All doses tolerable	5	3-5
	eff	0.3	0.4	0.5	0.6	0.7	Monotone increasing		
2	tox	0.1	0.45	0.5	0.55	0.6	Too toxic	Stop	Stop/Control
	eff	0.3	0.4	0.5	0.6	0.7	Monotone increasing		
3	tox	0.1	0.25	0.35	0.45	0.55	High doses toxic	3	3
	eff	0.3	0.4	0.5	0.6	0.7	Monotone increasing		
4	tox	0.1	0.2	0.25	0.3	0.35	All doses tolerable	3	3-4
	eff	0.3	0.4	0.7	0.5	0.4	Unimodal		
5	tox	0.1	0.45	0.5	0.55	0.6	Too toxic	Stop	Stop/Control
	eff	0.3	0.4	0.7	0.5	0.4	Unimodal		
6	tox	0.1	0.25	0.35	0.45	0.55	High doses toxic	3	3
	eff	0.3	0.4	0.7	0.5	0.4	Unimodal		
7	tox	0.1	0.2	0.25	0.3	0.35	All doses tolerable	3	3-5
	eff	0.3	0.4	0.6	0.6	0.6	Plateau		
8	tox	0.1	0.45	0.5	0.55	0.6	Too toxic	Stop	Stop/Control
	eff	0.3	0.4	0.6	0.6	0.6	Plateau		
9	tox	0.1	0.25	0.35	0.45	0.55	High doses toxic	3	3
	eff	0.3	0.4	0.6	0.6	0.6	Plateau		
10	tox	0.1	0.2	0.25	0.3	0.35	All doses tolerable	2	2-4
	eff	0.3	0.7	0.6	0.5	0.4	Monotone decreasing		
11	tox	0.1	0.45	0.5	0.55	0.6	Too toxic	Stop	Stop/Control
	eff	0.3	0.7	0.6	0.5	0.4	Monotone decreasing		
12	tox	0.1	0.25	0.35	0.45	0.55	High doses toxic	2	2-3
	eff	0.3	0.7	0.6	0.5	0.4	Monotone decreasing		
13	tox	0.1	0.2	0.25	0.3	0.35	All doses tolerable	Stop	Stop/Control
	eff	0.3	0.3	0.3	0.3	0.3	No Efficacy		
14	tox	0.1	0.45	0.5	0.55	0.6	Too toxic	Stop	Stop/Control
	eff	0.3	0.3	0.3	0.3	0.3	No Efficacy		
15	tox	0.1	0.25	0.35	0.45	0.55	High doses toxic	Stop	Stop/Control
	eff	0.3	0.3	0.3	0.3	0.3	No Efficacy		

TABLE 3.3: Operating characteristics for multiple combinations of AR phase size and probabilities for randomisation to control. Probability of selecting the OBD or good dose levels, mean number of patients treated at those dose levels and at the control dose after 10000 simulations.

Scenario	ϕ_R	j_R	OBD	Good Doses	P(OBD)	P(Good)	N(OBD)	N(Good)	N(Control)
1	0.2	0	5	3-5	0.05	0.60	2.8	32.1	1.2
		15	5	3-5	0.05	0.65	2.7	32	3.6
		30	5	3-5	0.05	0.71	2.2	31.1	6.4
		45	5	3-5	0.05	0.76	1.5	27.5	9.4
	0.33	60	5	3-5	0.04	0.80	1.1	22.9	12.4
		0	5	3-5	0.05	0.60	2.8	32.1	1.2
		15	5	3-5	0.07	0.67	3.1	32.6	4.9
		30	5	3-5	0.06	0.75	2.2	30.4	9.8
		45	5	3-5	0.05	0.79	1.3	25.5	14.7
		60	5	3-5	0.03	0.82	0.8	19.4	19.6
2	0.2	0	stop	stop/1	0.81	0.85	-	10.1	10.1
		15	stop	stop/1	0.80	0.85	-	11	11
		30	stop	stop/1	0.77	0.81	-	13.2	13.2
		45	stop	stop/1	0.72	0.75	-	15.6	15.6
	0.33	60	stop	stop/1	0.41	0.50	-	17.9	17.9
		0	stop	stop/1	0.81	0.85	-	10.1	10.1
		15	stop	stop/1	0.77	0.83	-	11.4	11.4
		30	stop	stop/1	0.75	0.79	-	14	14
		45	stop	stop/1	0.67	0.70	-	17.6	17.6
		60	stop	stop/1	0.38	0.42	-	20.7	20.7
3	0.2	0	3	3	0.25	0.25	15.5	15.5	2.6
		15	3	3	0.31	0.31	16.8	16.8	4.7
		30	3	3	0.35	0.35	16.8	16.8	7.5
		45	3	3	0.42	0.42	15.5	15.5	10.4
	0.33	60	3	3	0.48	0.48	12.7	12.7	13.3
		0	3	3	0.25	0.25	15.5	15.5	2.6
		15	3	3	0.32	0.32	17.2	17.2	5.8
		30	3	3	0.39	0.39	17.3	17.3	10.4
		45	3	3	0.46	0.46	15.2	15.2	15.2
		60	3	3	0.52	0.52	11.5	11.5	20.1

TABLE 3.3: Operating characteristics (continued)

Scenario	ϕ_R	j_R	OBD	Good Doses	P(OBD)	P(Good)	N(OBD)	N(Good)	N(Control)
4	0.2	0	3	3-4	0.69	0.75	32.3	37.2	1.3
		15	3	3-4	0.69	0.76	30.3	35.5	3.5
		30	3	3-4	0.71	0.81	26.4	32.3	6.5
	0.33	45	3	3-4	0.73	0.83	21.6	27.4	9.5
		60	3	3-4	0.71	0.83	16	21.7	12.4
		0	3	3-4	0.69	0.75	32.3	37.2	1.3
	0.33	15	3	3-4	0.69	0.77	29.5	35.1	5
		30	3	3-4	0.70	0.81	24.8	31	9.9
		45	3	3-4	0.70	0.84	19.7	25.4	14.7
	5	60	3	3-4	0.68	0.83	13.7	18.5	19.5
		0	stop	stop/1	0.81	0.86	-	10.1	10.1
		15	stop	stop/1	0.79	0.84	-	11.1	11.1
		30	stop	stop/1	0.77	0.82	-	13.1	13.1
		45	stop	stop/1	0.71	0.75	-	15.6	15.6
		60	stop	stop/1	0.41	0.49	-	17.9	17.9
		0	stop	stop/1	0.81	0.86	-	10.1	10.1
		15	stop	stop/1	0.78	0.83	-	11.3	11.3
		30	stop	stop/1	0.75	0.78	-	14.1	14.1
	6	45	stop	stop/1	0.67	0.70	-	17.6	17.6
		60	stop	stop/1	0.38	0.43	-	20.8	20.8
		0	3	3	0.37	0.37	21.2	21.2	2.6
		15	3	3	0.41	0.41	21.3	21.3	4.5
		30	3	3	0.45	0.45	19.6	19.6	7.5
		45	3	3	0.50	0.50	17.1	17.1	10.4
		60	3	3	0.54	0.54	12.8	12.8	13.4
		0	3	3	0.37	0.37	21.2	21.2	2.6
		15	3	3	0.42	0.42	21.4	21.4	5.8
	0.33	30	3	3	0.50	0.50	20.3	20.3	10.4
		45	3	3	0.55	0.55	16.6	16.6	15.1
		60	3	3	0.59	0.59	11.5	11.5	20.1

TABLE 3.3: Operating characteristics (continued)

Scenario	ϕ_R	j_R	OBD	Good Doses	P(OBD)	P(Good)	N(OBD)	N(Good)	N(Control)
7	0.2	0	3	3-5	0.48	0.70	23.9	35.6	1.3
		15	3	3-5	0.50	0.73	23.7	35	3.5
	0.33	30	3	3-5	0.52	0.78	21.8	32.4	6.4
		45	3	3-5	0.56	0.81	19.3	27.9	9.4
	0.33	60	3	3-5	0.57	0.82	15.8	22.5	12.5
		0	3	3-5	0.48	0.70	23.9	35.6	1.3
	0.33	15	3	3-5	0.49	0.75	22.7	35.1	5
		30	3	3-5	0.51	0.80	20.8	31.6	9.7
	0.33	45	3	3-5	0.55	0.84	17.7	26	14.7
		60	3	3-5	0.56	0.84	13.7	19.6	19.5
8	0.2	0	stop	stop/1	0.81	0.85	-	10.2	10.2
		15	stop	stop/1	0.79	0.84	-	11.1	11.1
	0.33	30	stop	stop/1	0.77	0.81	-	13.2	13.2
		45	stop	stop/1	0.71	0.76	-	15.6	15.6
	0.33	60	stop	stop/1	0.41	0.50	-	17.9	17.9
		0	stop	stop/1	0.81	0.85	-	10.2	10.2
	0.33	15	stop	stop/1	0.78	0.82	-	11.3	11.3
		30	stop	stop/1	0.75	0.79	-	14	14
	0.33	45	stop	stop/1	0.69	0.71	-	17.4	17.4
		60	stop	stop/1	0.38	0.43	-	20.8	20.8
9	0.2	0	3	3	0.34	0.34	18.6	18.6	2.6
		15	3	3	0.36	0.36	19.2	19.2	4.6
	0.33	30	3	3	0.42	0.42	18.3	18.3	7.5
		45	3	3	0.45	0.45	16.3	16.3	10.3
	0.33	60	3	3	0.50	0.50	12.9	12.9	13.3
		0	3	3	0.34	0.34	18.6	18.6	2.6
	0.33	15	3	3	0.38	0.38	19.7	19.7	5.9
		30	3	3	0.45	0.45	18.6	18.6	10.4
	0.33	45	3	3	0.52	0.52	15.9	15.9	15.2
		60	3	3	0.56	0.56	11.5	11.5	20.1

TABLE 3.3: Operating characteristics (continued)

Scenario	ϕ_R	j_R	OBD	Good Doses	P(OBD)	P(Good)	N(OBD)	N(Good)	N(Control)
10	0.2	0	2	2-4	0.77	0.97	45.1	57.1	1.3
		15	2	2-4	0.79	0.97	41.8	55.3	3.5
		30	2	2-4	0.79	0.98	37.2	52.5	6.5
	0.33	45	2	2-4	0.79	0.99	32.2	49.6	9.4
		60	2	2-4	0.80	0.99	26.4	46.4	12.3
		0	2	2-4	0.77	0.97	45.1	57.1	1.3
	0.33	15	2	2-4	0.77	0.97	40.1	53.6	5
		30	2	2-4	0.76	0.98	34.6	49.1	9.8
		45	2	2-4	0.76	0.99	28.4	44.3	14.7
	11	60	2	2-4	0.78	0.99	22.3	39.2	19.6
		0	stop	stop/1	0.65	0.74	-	11.7	11.7
		15	stop	stop/1	0.65	0.72	-	12.3	12.3
		30	stop	stop/1	0.62	0.69	-	13.8	13.8
		45	stop	stop/1	0.57	0.63	-	15.7	15.7
		60	stop	stop/1	0.41	0.49	-	17.9	17.9
		0	stop	stop/1	0.65	0.74	-	11.7	11.7
		15	stop	stop/1	0.64	0.72	-	12.5	12.5
		30	stop	stop/1	0.60	0.67	-	14.7	14.7
		45	stop	stop/1	0.52	0.56	-	17.7	17.7
		60	stop	stop/1	0.38	0.42	-	20.9	20.9
12	0.2	0	2	2-3	0.83	0.92	47.3	53.7	2.7
		15	2	2-3	0.83	0.93	43.6	51.8	4.7
		30	2	2-3	0.82	0.95	39.7	49.3	7.4
	0.33	45	2	2-3	0.84	0.96	35.9	46.4	10.4
		60	2	2-3	0.85	0.97	31.7	43.5	13.2
		0	2	2-3	0.83	0.92	47.3	53.7	2.7
	0.33	15	2	2-3	0.81	0.94	41.9	50.7	5.9
		30	2	2-3	0.80	0.95	36.6	46.3	10.4
		45	2	2-3	0.81	0.96	31.7	41.8	15.3
	0.33	60	2	2-3	0.82	0.97	26	36.8	20

TABLE 3.3: Operating characteristics (continued)

Scenario	ϕ_R	j_R	OBD	Good Doses	P(OBD)	P(Good)	N(OBD)	N(Good)	N(Control)
13	0.2	0	stop	stop/1	0.82	0.82	-	1.2	1.2
		15	stop	stop/1	0.78	0.78	-	3.5	3.5
		30	stop	stop/1	0.70	0.70	-	6.5	6.5
	0.33	45	stop	stop/1	0.57	0.57	-	9.5	9.5
		60	stop	stop/1	0.01	0.01	-	12.4	12.4
		0	stop	stop/1	0.82	0.82	-	1.2	1.2
	0.33	15	stop	stop/1	0.76	0.76	-	4.9	4.9
		30	stop	stop/1	0.68	0.68	-	9.7	9.7
		45	stop	stop/1	0.54	0.54	-	14.7	14.7
	14	60	stop	stop/1	0.01	0.01	-	19.6	19.6
	0.2	0	stop	stop/1	0.96	0.97	-	8.3	8.3
		15	stop	stop/1	0.95	0.96	-	9.6	9.6
		30	stop	stop/1	0.94	0.95	-	12.4	12.4
	0.33	45	stop	stop/1	0.92	0.93	-	15.5	15.5
		60	stop	stop/1	0.42	0.50	-	17.7	17.7
		0	stop	stop/1	0.96	0.97	-	8.3	8.3
	0.33	15	stop	stop/1	0.95	0.96	-	9.9	9.9
		30	stop	stop/1	0.93	0.94	-	13.5	13.5
		45	stop	stop/1	0.89	0.90	-	17.3	17.3
	15	60	stop	stop/1	0.38	0.43	-	20.8	20.8
	0.2	0	stop	stop/1	0.89	0.89	-	2.3	2.3
		15	stop	stop/1	0.87	0.87	-	4.5	4.5
		30	stop	stop/1	0.82	0.82	-	7.3	7.3
	0.33	45	stop	stop/1	0.73	0.73	-	10.4	10.4
		60	stop	stop/1	0.02	0.03	-	13.4	13.4
		0	stop	stop/1	0.89	0.89	-	2.3	2.3
	0.33	15	stop	stop/1	0.86	0.86	-	5.7	5.7
		30	stop	stop/1	0.80	0.80	-	10.3	10.3
		45	stop	stop/1	0.69	0.70	-	15.2	15.2
	16	60	stop	stop/1	0.02	0.03	-	20.1	20.1

For scenario 1 it is relatively simple to select an admissible dose since all doses are tolerable and efficacy increases monotonically. The difficulty is locating the OBD. All of these combinations fail to identify the OBD (dose-level 5) more than 7% of the time. Whereas the probability of selecting a good dose is between 60 and 82%. As the size of the AR phase increases from 0 to 60 so do the selection probabilities for a good dose, going from 60% to 80% and 60% to 82% for randomisation probabilities of 0.2 and 0.33 respectively. It should be noted that the two designs with no AR phase are identical since the randomisation probabilities are never used. In terms of the number of patients treated we see more patients in the control arm as AR phase size and randomisation probability increase. This is expected since if you increase the amount of time available for cohorts to be randomised or the probability in which that is done, more patients will be recruited to control. By increasing the probability of randomising to control we can also see that fewer patients are being treated at good doses at higher AR sizes. Also, the ϕ_R figure does not guarantee that exact percentage of patients in the control dose will be allocated to control but for this scenario, it appears to be somewhat accurate.

Scenario 2 has no OBD as all treatment doses are considered toxic. For most of the combinations, stopping occurs 67-81% of the time and including allocation to the control arm we see this increase to 70-85%. Slightly concerning is the case where the AR phase lasts the whole trial. Here stopping is less frequent at 50% and 42% for probabilities of randomisation to control of $\phi_R = 0.2$ and $\phi_R = 0.33$ respectively. To understand why this was occurring we investigated the failure mechanism in individual simulation runs. We found that the design was stopping appropriately for excess toxicity. However, due to setting the AR phase at 60 the maximisation phase never starts and thus the stopping rule for futility never triggers. This is why this parametrisation performs worse comparatively. Even though this scenario is to check for excess

toxicity the true efficacy rates used in this scenario could also potentially trigger the futility rule as they are set at 40% and 50% for dose-levels 2 and 3. To confirm this we ran some of the parametrisations where the AR size is less than 60 using a design without the futility rule and observed similar stopping rates of 40% in this scenario. This indicates that overall our design is not that great at stopping for potential toxicities. This could be improved by utilising different stopping criteria. We also observe this difference in scenarios 5, 8, 11 and 14 where we should also be stopping for excess toxicity. The results for $j_R = 60$ in these scenarios could be interpreted as our baseline for stopping for excess toxicity and the increase stopping in other parametrisations represent how often the futility rule is triggered.

In scenario 3, the treatment is toxic at high doses and ineffective at lower doses meaning only one dose can be considered good or the OBD. This is a difficult scenario since only one of the five dose-levels is suitable to allocate patients to. The selection probabilities range from 25% to 52%. We see as j_R increases selection probabilities also increase. As those designs with smaller AR phases go into the maximisation phase they would be selecting dose-levels based on those in the admissible set with higher efficacy. Since the doses with higher efficacy also have a high toxicity rate there is a chance early on in the trial this is not detected resulting in toxicities at the higher dose-levels causing the trial to stop early. This could be a reason why the designs with the larger AR phase perform better as doses would be randomly selected. This means there is more chance for the lower dose-levels to be chosen and since those are not toxic and the futility rule does not kick in until the maximisation phase there is a higher chance that the OBD can be found.

Scenarios 4-6 look at a unimodal dose-efficacy relationship where efficacy peaks at dose-level 3, with the three same dose-toxicity relationships as before: tolerable, toxic and toxic at high doses for each scenario respectively.

Firstly, in scenario 4 we see good performance from all the parametrisations with the probability of selecting the OBD ranging between 68% and 73% and the probability of selecting a good dose ranging from 75% to 84%. We also see an appropriate amount of patients in the control arm for each of our parameter combinations. An important characteristic of this design to note is the ratio of patients allocated to the control arm compared to the OBD or the good doses. For the higher randomisation probability and maximum AR phase, we can see close to a 1:1 (18.5 at good dose levels to 19.5 at control) allocation between patients treated at control and the best doses. For $\phi_R = 0.2$, and $j_R = 45$ we can see close to a 2:1 allocation between those treated at the OBD (21.6) and those treated at the control (9.5).

Scenario 5 is where treatment is too toxic. Like scenario 2 we see high probabilities of stopping 67%-81% and even higher probabilities for stopping and including patients in the control arm ($P(\text{Good})$) 70%-86%. Similarly, the design with an AR phase of 60 performs poorly here and the number of patients allocated to the control is also comparable.

Scenario 6 only has one good dose to select, making it similar to scenario 3 except with a unimodal efficacy curve. Here, selection probabilities are slightly better ranging from 37%-59%. In this scenario, we also observe that the larger the AR phase the greater the selection probability of the OBD. The unimodal efficacy curve means that the dose we want to select is also the dose with the highest efficacy making it easier for the model to pick out.

A plateau relationship where dose-efficacy stops increasing after dose 3 is looked at in scenarios 7-9 with the three different toxicity curves. In scenario 7 the probability of selecting the OBD ranges from 48%-56%, with the probability of selecting a good dose ranging from 70-84%. In this instance, we see quite a bit of discrepancy going from selecting the OBD to selecting the good doses in terms of the selection probabilities being much higher for good doses. Here we

have three doses with the same efficacy level, two of which only have a slight increase in toxicity, which is still below the pre-specified target level. This can also be seen in the number of patients being treated at the good dose versus at the OBD. Scenario 8 exhibits the same behaviour as the other toxic scenarios 2 and 5. Then with scenario 9, the designs also behave similarly to scenarios 3 and 6 where higher dose-levels are too toxic. Selection probabilities range from 34% to 56% with the designs with higher AR phases performing better.

For scenarios 10-12 we look at a monotone decreasing efficacy curve where dose-level 2 is the most efficacious for each of the toxicity curves. In scenario 10 we see very high probabilities of selecting the OBD (76%-80%) with the probability of selecting a good dose being 97% or higher. We can also see that in terms of the numbers treated at the OBD and the good doses as these values are relatively high compared to other scenarios. As the OBD is one of the lower dose-levels it makes it relatively easy for the model to select since it is more likely that patients will be allocated there early on into recruitment. Once the maximisation phase starts there would be a lot of efficacy data for that dose and it would be favoured by the model. Even in the cases with larger AR phases, the adaptive randomisation probabilities are still scaled based on efficacy, so it would be more likely they would be allocated to dose-level 2 as well.

In terms of stopping early if the dose-levels are too toxic for this efficacy curve (scenario 11), performance appears to be worse compared to other too-toxic scenarios (scenarios 2,5,8 and 14). Even so, we still see the same patterns where when the AR phase is 60, the same size as the trial stopping is relatively even worse. Ignoring those designs the stopping probability ranges from 52%-65%, adding in the percentage of patients allocated to the control arm this increases to 56%-74%. One reason why this may be worse is due to the very high efficacy rates early on and the toxicity rate only being slightly above our target

rate by 5%. Early on into the trial, that 5% would be difficult for the model to detect but the high efficacy rate is likely to lead to more events so the trial would be less likely to stop until it went to higher dose-levels. Additionally, as the efficacy rates are so high as well for early doses the futility rule is less likely to be triggered meaning less stopping overall compared to other toxic scenarios.

Typically, the scenarios where the higher doses are more toxic have been the most difficult for the design to deal with. However for scenario 12, with the monotone decreasing efficacy curve we see probabilities of selecting the OBD range from 80%-85%. This is even higher than the selection probabilities in scenario 10. However, when we look at the probabilities of selecting good doses (92%-97%), whilst still very high it is still slightly less than in scenario 10 where there was one more dose that could be considered good.

Finally, the last efficacy curve we look at is one where no efficacy is apparent, so efficacy stays at the same level as the control dose. Ideally, in all these scenarios (13-15) we would stop for lack of efficacy. One thing to point out is that the rule for stopping for futility only triggers in the maximisation phase. So, for the designs where $j_R = 60$ stopping for lack of efficacy will not occur. For scenario 13 we have doses that are all tolerable and can see stopping probabilities ranging from 54% to 82%. The only reason to stop in this scenario is for lack of efficacy and so those designs with larger AR phases will not be able to do this until later on into the trial meaning they are less likely to stop as reflected in the stopping probabilities. In terms of selecting the good dose, this is identical to selecting the OBD. So, in this scenario, the control dose is seldom selected as the OBD, even though patients are still being allocated to that dose-level. As stated for an AR phase size of 60 the trial can not be stopped for futility, so the probability of stopping is due to toxicity. In scenario 14, all

the dose-levels are toxic, here we have very high stopping rates between 89%-96% except in the case when the AR phase is 60. Scenario 15, where only the higher doses are too toxic stops most of the time as well given a reasonable AR phase between 69% and 89% of the time. Again, very rarely is the control dose selected as the OBD.

Table 3.4 provides a summary of the selection probabilities of the OBD and good doses respectively for all 15 scenarios and 10 parameter combinations. The mean provides a rudimentary glance across the 15 scenarios of how well it selects the OBD / good dose-levels. The standard deviation is a representation of the variability of performance across the different designs. The lower the standard deviation the more homogenous the performance. These statistics have been calculated for all scenarios and then for 'Non-Stopping' scenarios, which are just the scenarios where stopping early for toxicity/futility is not the ideal outcome i.e. scenarios 1,3,4,6,7,9,10 and 12.

The means for selecting a good dose are about 10% higher and appear to be slightly less variable. This is to be expected as when selecting the good dose-levels we are allowing for a wider range of doses to be included. There appear to be limited differences between the various parameterisations, except for in the case of when $j_R = 60$ where its performance is poor in scenarios that require stopping. There are only 1-2 percentage points difference in the mean selection of the OBD and good dose-levels for the various designs.

In general, these scenarios show us that there are some issues with certain specifications of ϕ_R and j_R in some of the scenarios presented. Specifically, in the case of stopping for toxicity, having the AR phase being the same as the sample size causes some issues. However, as we investigated this is largely due to the stopping criteria we applied and as such the design may perform better under a different stopping rule. For the randomisation probabilities, performance was mostly similar between the two values we chose.

TABLE 3.4: Probabilities of selecting the OBD and good dose levels for multiple combinations of AR phase size and probabilities for randomisation to control, plus summary statistics.

ϕ_R	j_R	Selection probabilities: Scenarios 1-15															All scenarios		Non Stopping	
		1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	Mean	StDev	Mean	StDev
Selection probabilities for the OBD																				
	0	0.05	0.81	0.25	0.69	0.81	0.37	0.48	0.81	0.34	0.77	0.65	0.83	0.82	0.96	0.89	0.63	0.27	0.47	0.27
	15	0.05	0.80	0.31	0.69	0.79	0.41	0.50	0.79	0.36	0.79	0.65	0.83	0.78	0.95	0.87	0.64	0.25	0.49	0.26
0.20	30	0.05	0.77	0.35	0.71	0.77	0.45	0.52	0.77	0.42	0.79	0.62	0.82	0.70	0.94	0.82	0.63	0.23	0.51	0.26
	45	0.05	0.72	0.42	0.73	0.71	0.50	0.56	0.71	0.45	0.79	0.57	0.84	0.57	0.92	0.73	0.62	0.21	0.54	0.26
	60	0.04	0.41	0.48	0.71	0.41	0.54	0.57	0.41	0.50	0.80	0.41	0.85	0.01	0.42	0.02	0.44	0.26	0.56	0.25
	0	0.05	0.81	0.25	0.69	0.81	0.37	0.48	0.81	0.34	0.77	0.65	0.83	0.82	0.96	0.89	0.63	0.27	0.47	0.27
	15	0.07	0.77	0.32	0.69	0.78	0.42	0.49	0.78	0.38	0.77	0.64	0.81	0.76	0.95	0.86	0.63	0.24	0.49	0.25
0.33	30	0.06	0.75	0.39	0.70	0.75	0.50	0.51	0.75	0.45	0.76	0.60	0.80	0.68	0.93	0.80	0.63	0.22	0.52	0.24
	45	0.05	0.67	0.46	0.70	0.67	0.55	0.55	0.69	0.52	0.76	0.52	0.81	0.54	0.89	0.69	0.60	0.20	0.55	0.24
	60	0.03	0.38	0.52	0.68	0.38	0.59	0.56	0.38	0.56	0.78	0.38	0.82	0.01	0.38	0.02	0.43	0.26	0.57	0.24
Selection probabilities for good dose-levels																				
	0	0.60	0.85	0.25	0.75	0.86	0.37	0.70	0.85	0.34	0.97	0.74	0.92	0.82	0.97	0.89	0.73	0.23	0.61	0.27
	15	0.65	0.85	0.31	0.76	0.84	0.41	0.73	0.84	0.36	0.97	0.72	0.93	0.78	0.96	0.87	0.73	0.21	0.64	0.26
0.20	30	0.71	0.81	0.35	0.81	0.82	0.45	0.78	0.81	0.42	0.98	0.69	0.95	0.70	0.95	0.82	0.74	0.19	0.68	0.25
	45	0.76	0.75	0.42	0.83	0.75	0.50	0.81	0.76	0.45	0.99	0.63	0.96	0.57	0.93	0.73	0.72	0.18	0.71	0.23
	60	0.80	0.50	0.48	0.83	0.49	0.54	0.82	0.50	0.50	0.99	0.49	0.97	0.01	0.50	0.03	0.56	0.29	0.74	0.21
	0	0.60	0.85	0.25	0.75	0.86	0.37	0.70	0.85	0.34	0.97	0.74	0.92	0.82	0.97	0.89	0.73	0.23	0.61	0.27
	15	0.67	0.83	0.32	0.77	0.83	0.42	0.75	0.82	0.38	0.97	0.72	0.94	0.76	0.96	0.86	0.73	0.21	0.65	0.25
0.33	30	0.75	0.79	0.39	0.81	0.78	0.50	0.80	0.79	0.45	0.98	0.67	0.95	0.68	0.94	0.80	0.74	0.18	0.70	0.23
	45	0.79	0.70	0.46	0.84	0.70	0.55	0.84	0.71	0.52	0.99	0.56	0.96	0.54	0.90	0.70	0.72	0.17	0.74	0.21
	60	0.82	0.42	0.52	0.83	0.43	0.59	0.84	0.43	0.56	0.99	0.42	0.97	0.01	0.43	0.03	0.55	0.30	0.76	0.18

Many of the discrepancies in the scenarios were due to the size of the AR phase. Seemingly, performance was generally unaffected by the percentage of patients being randomised to control but rather the amount of time spent being randomised. In terms of patient numbers at the control dose, we see on average a similar number to what would be expected i.e. for $\phi_R = 0.2$ and $j_R = 45$ you would expect 9 control patients (20% of 45) our simulations yielded values ranging from 9.4 to 15.6 across the 15 scenarios.

When we look at the mean of the non-stopping scenarios there appears to be a monotonic increase in the selection of both the OBD and good doses. As we increase the size of the AR phase we are more likely to make a correct decision. There is also a similar pattern when we increase the probability of randomising to control. So, accuracy appears to increase as more adaptive randomisation is allowed to take place in the design. This may not ethically be the best as we would want to prioritise giving patients the most efficacious and tolerable dose. However, by allowing for more adaptive randomisation we increase the probability of spreading out patients across the doses and gaining

more information about all of the dose-levels which appears to make the final selection more accurate. The uncertainty around the probability of efficacy at each dose is reflected in our randomisation probabilities. Which is a positive from an ethical perspective since patients are more likely to be allocated to the more efficacious treatment and at worse would be given control. The main caveat to this is that when we consider all scenarios the same relationship is not observed and this is mainly due to the futility stopping rule.

Based on these simulations it would be best to use an AR phase sized between 25% and 75% of the total sample size and a randomisation to control probability that will produce the desired number of control patients. This can be determined by dividing the desired number of controls by the size of the AR phase. For example, say our AR size is 30 patients and we want 15 controls the probability of randomising to control ϕ_R should be set to 0.5 (15/30). However, it may be beneficial to investigate various values of ϕ_R as there does appear to be some trade-off in terms of performance and the number of patients recruited to the control dose.

3.4.3 Example trial run

To further demonstrate how the design works we provide a illustrative example of a single trial run. Here we will use the same design as specified in the previous section. Specifically we will be using the design with an AR phase size of 30 and an adaptive randomisation probability of 0.33. All other parameters and priors are as defined previously. In practice the trial will recruit patients in cohorts of three, they will then be given treatment and observed. After the observation period has been collected we will update the model with the data that has been observed and determine the dose to be recommended for the next cohort.

Toxicity and efficacy events are treated as binary variables, so for each patient there are four different potential outcomes. We will use a specific notation to denote each of these outcomes. A patient can experience a toxicity event (T), a efficacy event (E), both a toxicity and efficacy event (B) or neither a toxicity or efficacy event (N). To reference the outcomes for a specific cohort we label them with the dose-level of that cohort. For example a cohort at dose-level 4, where 1 patient had a toxicity, 1 patient experienced a toxicity and efficacy event and 1 patient had neither would be presented as 4TBN. It should be noted the order of the outcomes does not matter either, 4TBN is equivalent to 4BTN, 4BNT, etc.

For our example trial run we start at dose-level 2. Overall we will recruit 60 patients, unless the model recommends stopping early. The adaptive randomisation phase will last for 30 patients (10 cohorts). The results for our observed outcomes are shown in Table 3.5. Here for each cohort we show the estimated probabilities from the model for toxicity and efficacy at each dose-level, we also present the randomisation probabilities as well which indicate the probability of randomly allocating the next cohort to the next dose-level. The table also shows what the recommended and optimal dose is after each cohort.

For the first cohort we see 2EEN, so two patients had efficacy events and one patient has neither. Based on this data the model shows that all doses are considered safe as the probability of toxicity is less than 0.35 for all five doses. Looking at the estimated probability of efficacy we can see that dose-level 2 has the highest probability at 0.68. This implies that the optimal dose would be dose-level 2 as it has the highest estimated efficacy probability and is within the admissible safe set of doses. However, the recommended dose is dose-level 3. This is because the recommended dose in the AR phase is selected randomly based on the randomisation probabilities. As dose-level 2 has the highest efficacy probability we can see its randomisation probability is also the highest

TABLE 3.5: Summary of model outputs during the adaptive randomisation phase for our example trial run.

Cohort	Dose	Outcome	Estimated probabilities					Recommended	Optimal	
			1	2	3	4	5			
1	2	EEN	tox	0.06	0.09	0.14	0.19	0.26	3	2
			eff	0.34	0.68	0.59	0.5	0.42		
			rand.	0.33	0.21	0.18	0.15	0.13		
2	3	EEE	tox	0.02	0.04	0.07	0.11	0.17	3	3
			eff	0.51	0.74	0.8	0.74	0.67		
			rand.	0.33	0.17	0.18	0.17	0.15		
3	3	EEE	tox	0.02	0.04	0.07	0.11	0.17	4	3
			eff	0.51	0.74	0.8	0.74	0.67		
			rand.	0.33	0.17	0.18	0.17	0.15		
4	4	ENN	tox	0.01	0.01	0.03	0.05	0.09	1	3
			eff	0.44	0.69	0.77	0.69	0.61		
			rand.	0.33	0.17	0.19	0.17	0.15		
5	1	NNN	tox	0.01	0.01	0.03	0.05	0.09	2	3
			eff	0.32	0.6	0.7	0.6	0.5		
			rand.	0.33	0.17	0.19	0.17	0.14		
6	2	ENN	tox	0	0.01	0.02	0.05	0.08	4	3
			eff	0.27	0.56	0.66	0.56	0.46		
			rand.	0.33	0.17	0.2	0.17	0.14		
7	4	ENT	tox	0.01	0.03	0.06	0.12	0.19	1	3
			eff	0.23	0.53	0.64	0.53	0.42		
			rand.	0.33	0.17	0.2	0.17	0.13		
8	1	ENN	tox	0.01	0.03	0.06	0.11	0.18	5	3
			eff	0.25	0.54	0.65	0.54	0.44		
			rand.	0.33	0.17	0.2	0.17	0.14		
9	5	ETT	tox	0.03	0.05	0.1	0.17	0.26	1	3
			eff	0.24	0.53	0.64	0.53	0.43		
			rand.	0.33	0.17	0.2	0.17	0.13		
10	1	ENN	tox	0.03	0.05	0.1	0.17	0.25	3	3
			eff	0.25	0.55	0.65	0.55	0.44		
			rand.	0.33	0.17	0.2	0.17	0.14		

apart from dose-level 1 which is fixed at 0.33. Here there is still a chance to recommend the other doses as there randomisation probabilities are non zero. It is important to note if we were to run the model again with the exact same data, the estimated toxicity, efficacy and randomisation probabilities and optimal dose would all be the same but the recommended dose could be any of the five doses.

Continuing this example as the model recommended dose-level 3 after the first cohort the second cohort is recruited and assigned dose-level 3. In this example all 3 patients experience an efficacy event. When this is put into the model, dose-level 3 is determined to be the optimal dose. The recommended dose in this case is also dose-level 3. We see it has the highest estimated efficacy and therefore also has the highest randomisation probability outside of the control dose. As the efficacy probabilities are similar for the dose-levels 2-5 the randomisation probabilities are also similar (0.15-0.18). So, the model could have easily recommended any of the other dose-levels.

This process is repeated for the whole AR phase. After observing each cohort we can see from cohort 2 onwards the optimal dose is considered to be dose-level 3. This is due to the dose having the highest estimated efficacy probability whilst remaining in the admissible set of doses in terms of safety based on the estimated toxicity probability. Even though the optimal dose is considered to be dose-level 3 we see after most of the cohorts the recommended dose is usually a different dose. This is because the recommended dose is being randomly allocated and as there have been limited toxicities in this example all of the doses are considered safe and have a chance of being allocated to. In this example we see that all of the dose-levels have been allocated patients at some point during this AR phase.

As 10 cohorts of three patients have now been recruited we have reached the 30 patient limit for the AR phase. Moving into the maximisation phase

TABLE 3.6: Summary of model outputs during the maximisation phase for our example trial run.

Cohort	Dose	Outcome	Estimated probabilities					Recommended/Optimal	
			1	2	3	4	5		
11	3	EEN	tox eff	0.02 0.25	0.04 0.55	0.09 0.65	0.15 0.55	0.24 0.44	3
12	3	BBE	tox eff	0.04 0.28	0.07 0.57	0.14 0.68	0.23 0.57	0.32 0.47	3
13	3	BEE	tox eff	0.05 0.3	0.08 0.59	0.16 0.69	0.25 0.59	0.34 0.49	3
14	3	EET	tox eff	0.06 0.3	0.09 0.59	0.17 0.69	0.26 0.59	0.36 0.49	3
15	3	ENT	tox eff	0.06 0.27	0.1 0.57	0.18 0.67	0.28 0.57	0.37 0.47	3
16	3	BEE	tox eff	0.07 0.29	0.11 0.59	0.19 0.69	0.29 0.59	0.39 0.49	3
17	3	BET	tox eff	0.09 0.29	0.13 0.59	0.22 0.69	0.32 0.59	0.42 0.49	3
18	3	BNN	tox eff	0.09 0.27	0.14 0.57	0.23 0.67	0.33 0.57	0.43 0.47	3
19	3	EEN	tox eff	0.08 0.27	0.13 0.57	0.22 0.67	0.31 0.57	0.41 0.47	3
20	3	ENN	tox eff	0.08 0.25	0.12 0.55	0.21 0.66	0.3 0.55	0.4 0.45	3

doses are no longer randomly allocated and instead the optimal dose is now the dose that is recommended. From cohort 10 in Table 3.5 we can see the optimal dose is dose-level 3 and there would be the dose recommended for cohort 11. Table 3.6 provides a summary of the model outputs for the next 10 cohorts based on specific outcomes. The adaptive randomisation probabilities are no longer presented as they are not required to allocate patients and as the optimal dose is now always the recommended dose these can be considered the same.

In maximisation phase we can see after each cohort the decision remains unchanged an the recommended/optimal dose is always dose-level 3. This is because that dose has the highest estimated probability of efficacy. Whilst doses 4 and 5 are still within the admissible set of doses in terms of probability

of toxicity the estimated efficacy is not as high compared to dose-level 3. If perhaps we were to observe fewer efficacy events in some of the cohorts this may change and the model may consider moving to a higher or lower dose. Also in terms of toxicity whilst some patients do start to experience toxicity events there is not enough for the model to consider dose-level 3 no longer safe and move to a lower dose.

For example we see in cohorts 12 and 13 outcomes of BBE and BEE respectively so all six of those patients had an efficacy event and three also had an efficacy event. This causes the estimated probability of toxicity to go from 0.09 after evaluating outcomes for cohort 11 to 0.16 after cohort 13. Similarly, the estimated efficacy goes from 0.65 to 0.68. For future cohorts there would need to be a difference in the outcomes observed in order for the model to recommend a different dose as up to this point there has been data collected on 39 patients, with 15 who have been treated at dose-level 3.

After the final cohort has been observed the model still recommends dose-level 3 so that would be declared as the OBD. Table 3.7 shows a summary of the number of patients treated at each dose-level along with the number of efficacy and toxicity events and the percentage of patients in each dose-level who experienced an event. The majority of patients end up being allocated to dose-level 3 as this was continually defined as the optimal dose during the maximisation phase.

Overall, our design seems to work as intended. During the AR phase multiple different doses were allocated to cohorts of patients along with the control dose (dose-level 1). It is possible that just by chance during the AR phase you may end up selecting the same dose over and over again and not allocate patients to other doses. So we could include additional rules to mitigate against this. Ultimately the impact on performance of the design may be limited as a range of doses could still be explored during the maximisation phase. Here

TABLE 3.7: Number and percentage of events for our example trial run by dose-level.

Dose	No. of patients	N (%)	
		Toxicity	Efficacy
1	9	0 (0%)	2 (22.22%)
2	6	0(0%)	3(50%)
3	36	9 (25%)	26 (72.22%)
4	6	1(16.67%)	2(33.33%)
5	3	2 (66.67%)	1 (33.33%)

the outcomes observed could cause the estimated probabilities of efficacy and toxicity to change and thus give a different dose recommendations.

3.4.4 Comparison of RtC-WT against Alternative Designs

The simulations presented in Section 3.4.2 were about exploring the impact of varying the parameters controlling the randomisation in RtC-WT. In this section we investigate two alternative trial approaches which could be used to achieve the same aims as RtC-WT, that is to conduct a dose-finding study locating the ODB whilst recruiting patients to a control arm. Simulations will be conducted for these two alternatives across a variety of scenarios and operating characteristics will be compared against those for RtC-WT.

The first alternative approach would be just to use a standard Wages and Tait design and include the lowest dose-level as control. We will refer to this approach as the standard Wages and Tait (WT). Technically this design does not aim to recruit control patients but by including it as a dose-level there is still a probability during the AR phase that this occurs. Either way, this will be a good comparator for RtC-WT as we will be able to directly compare how our extension impacts performance compared to a traditional Wages and Tait design. Theoretically, since the standard Wages and Tait design will not be forced to allocate patients to the control dose-level you would expect more patients

to be allocated at the experimental treatment dose-levels leading to more data on efficacy and toxicity relationships making it easier to locate the ODB. The differences in performance between these two designs could be considered as the cost of including a control dose.

The second design uses a two-arm randomised approach. Patients once recruited are randomised to either a control arm or a dose-finding study arm. The dose-finding study arm will use the method of Wages and Tait to identify an OBD. We will refer to this method as the two-arm approach. One of the benefits of this approach is that it is fairly simplistic. This could be considered a straightforward way of including a control arm into complex designs without having to deal with any mathematics. For example, the EffTox design could be used for the dose-finding arm, and due to the two-arm approach, we now have a cohort of control patients without building that methodology directly into the design. For the dose-finding study, any methodology could be used here to locate an OBD, however, we selected WT to provide more comparisons for RtC-WT. Since the randomisation occurs upfront a guaranteed number of patients can be expected in the control arm, which may be a desirable characteristic of this design. It is important to note in our simulations here that we will not consider any data from the patients in the control arm to have an impact on the dose-finding study arm. Both arms can be considered independent for our simulations. In terms of comparisons to RtC-WT, this design will allow us to see if it is worth including the control patients directly in the dose-finding aspect of the design and if there is any benefit in terms of operating characteristics.

For RtC-WT we will be using the same design specification as detailed in Section 3.4.1. In terms of parameters for the number of patients in the AR phase and the probability of randomising to control these will be set at $j_R = 30$ and $\phi_R = 0.33$ respectively. These values were selected based on the work done

in the previous section. This combination of parameters seemed to perform consistently across all the scenarios explored. The standard Wages and Tait approach will also be using the same specification except for the fixed probability of randomisation to control.

For the two-arm approach, patients are being randomised first. Looking at the RtC-WT design we have specified a sample size of 60, an AR phase size of 30 and a probability of randomising to control at 33%. Here we would expect roughly ten patients to be allocated to the control arm (33% of 30), looking at Table 3.3 for this combination we see on average we achieve around 9-10 patients at the control dose-level in non-toxic scenarios. To mimic this behaviour for the two-arm approach we would need to specify a randomisation ratio upfront. Based on the parameters set for RtC-WT this can be done generally using the formula:

$$1 : \frac{J}{\phi_R j_r} - 1, \quad (3.19)$$

where J is the maximum sample size. Alternatively, if the number of control patients desired is known the denominator can be replaced by that number. In this scenario, using the equation above we obtain a randomisation ratio of approximately 1:5. This would lead to 10 patients in the control arm and 50 in the dose-finding arm. As we are using cohorts of 3 it would be preferable to have the sample size of the dose-finding study be divisible by 3. Therefore we set the desired number of control patients as 9, leaving 51 for the dose-finding study.

The specifications for the dose-finding study in the two-arm approach will be somewhat similar as well. Here there will only be four dose levels (no control dose-level) i.e doses 2-5 in Section 3.4.1. We also adjust the toxicity and efficacy skeletons accordingly in Table 3.1 as well by removing the values from

dose-level 1. Since there is no control dose-level in the design we will just be using approximately 50% of the patients in the AR phase, as recommended by Wages and Tait. As there will be 51 patients in the dose-finding study we set the AR phase to 24, slightly less than 50% as we are using cohorts of three and 51 patients can not be evenly split up between the two phases. Each design has an effective total sample size of 60 patients, however the allocation of patients to the specific dose-finding portion of the Two-Arm design differs slightly. All other design specifications remain the same such as the stopping rules and the pre-specified toxicity upper bound and lower bound. We summarise the three designs being compared in Table 3.8.

TABLE 3.8: Summary of the three designs being compared.

Design	Specification	Benefits	Flaws	Assumptions
RtC-WT	5 Doses N = 60 Cohort size = 3 AR size (j_R) = 30 $q_R = 0.33$	Guaranteed patients recruited to the control dose. Able to compare control to experimental doses.	Performance may suffer. Extra complexity in the design.	The lowest dose level is a control dose.
WT	5 Doses N = 60 Cohort size = 3 AR size (j_R) = 30	Simple to implement. Able to compare control to experimental doses.	Patients not guaranteed at the control dose.	Control dose treated in the same was as an experimental dose.
Two-Arm	4 Doses N = 51 9 control patients Cohort size = 3 AR size (j_R) = 24	Simpler to implement. Exact number of control patients is known.	Cannot include the control dose when modelling toxicity and efficacy	Recruit dose-finding and control patients separately. Allocation ratio is based on the number of control patients desired.

To compare these different approaches simulations will be used covering the same 15 scenarios as in section 3.4.2. For each scenario, we simulate 10000 trials using the designs mentioned above. Table 3.9 shows the operating characteristics comparing the three designs. Table 3.10 provides summary statistics of the operating characteristics. The mean and standard deviation of the selection probabilities are calculated for all scenarios and also all scenarios excluding those where stopping is the best option (non stopping).

TABLE 3.9: Operating characteristics for alternative designs. Probability of selecting the best or good dose levels as the OBD, mean number of patients treated at those dose levels and at the control dose after 10000 simulations.

Scenario	Design	OBD	Good Doses	P(OBD)	P(Good)	N(OBD)	N(Good)	N(Control)
1	RtC-WT	5	3-5	0.06	0.75	2.2	30.4	9.8
	WT	5	3-5	0.05	0.72	2.1	31.8	6.6
	Two-Arm	5	3-5	0.04	0.61	1.8	25.5	9
2	RtC-WT	stop	stop/1	0.75	0.79	-	14	14
	WT	stop	stop/1	0.76	0.80	-	13.9	13.9
	Two-Arm	stop	stop	0.75	0.75	-	-	9
3	RtC-WT	3	3	0.39	0.39	17.3	17.3	10.4
	WT	3	3	0.37	0.37	17.7	17.7	8.4
	Two-Arm	3	3	0.26	0.26	12.6	12.6	9
4	RtC-WT	3	3-4	0.70	0.81	24.8	31	9.9
	WT	3	3-4	0.73	0.81	27.6	33.5	6.6
	Two-Arm	3	3-4	0.72	0.76	24	28.1	9
5	RtC-WT	stop	stop/1	0.75	0.78	-	14.1	14.1
	WT	stop	stop/1	0.76	0.80	-	13.9	13.9
	Two-Arm	stop	stop	0.75	0.75	-	-	9
6	RtC-WT	3	3	0.50	0.50	20.3	20.3	10.4
	WT	3	3	0.48	0.48	21.2	21.2	8.4
	Two-Arm	3	3	0.38	0.38	15.9	15.9	9
7	RtC-WT	3	3-5	0.51	0.80	20.8	31.6	9.7
	WT	3	3-5	0.54	0.79	22.8	33.4	6.8
	Two-Arm	3	3-5	0.52	0.71	19.5	27.8	9
8	RtC-WT	stop	stop/1	0.75	0.79	-	14	14
	WT	stop	stop/1	0.75	0.79	-	13.9	13.9
	Two-Arm	stop	stop	0.76	0.76	-	-	9
9	RtC-WT	3	3	0.45	0.45	18.6	18.6	10.4
	WT	3	3	0.43	0.43	19.4	19.4	8.4
	Two-Arm	3	3	0.34	0.34	14.2	14.2	9

TABLE 3.9: Operating characteristics (continued)

Scenario	Design	OBD	Good Doses	P(OBD)	P(Good)	N(OBD)	N(Good)	N(Control)
10	RtC-WT	2	2-4	0.76	0.98	34.6	49.1	9.8
	WT	2	2-4	0.78	0.98	36.4	52.2	6.7
	Two-Arm	2	2-4	0.83	0.99	37.1	50.2	9
11	RtC-WT	stop	stop/1	0.60	0.67	-	14.7	14.7
	WT	stop	stop/1	0.60	0.67	-	14.3	14.3
	Two-Arm	stop	stop	0.53	0.53	-	-	9
12	RtC-WT	2	2-3	0.80	0.95	36.6	46.3	10.4
	WT	2	2-3	0.82	0.95	38.5	48.5	8.3
	Two-Arm	2	2-3	0.90	0.97	41.2	48.6	9
13	RtC-WT	stop	stop/1	0.68	0.68	-	9.7	9.7
	WT	stop	stop/1	0.71	0.71	-	6.3	6.3
	Two-Arm	stop	stop	0.68	0.68	-	-	9
14	RtC-WT	stop	stop/1	0.93	0.94	-	13.5	13.5
	WT	stop	stop/1	0.94	0.95	-	13.2	13.2
	Two-Arm	stop	stop	0.95	0.95	-	-	9
15	RtC-WT	stop	stop/1	0.80	0.80	-	10.3	10.3
	WT	stop	stop/1	0.82	0.82	-	8.1	8.1
	Two-Arm	stop	stop	0.81	0.81	-	-	9

TABLE 3.10: Selection probabilities and summary statistics for multiple designs.

Design	Selection probabilities: Scenarios 1-15															All scenarios		Non Stopping	
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	Mean	StDev	Mean	StDev
Selection probabilities for the OBD																			
RtC-WT	0.06	0.75	0.39	0.70	0.75	0.50	0.51	0.75	0.45	0.76	0.60	0.80	0.68	0.93	0.80	0.63	0.22	0.52	0.24
WT	0.05	0.76	0.37	0.73	0.76	0.48	0.54	0.75	0.43	0.78	0.60	0.82	0.71	0.94	0.82	0.64	0.23	0.53	0.26
Two-Arm	0.04	0.75	0.26	0.72	0.75	0.38	0.52	0.76	0.34	0.83	0.53	0.90	0.68	0.95	0.81	0.62	0.26	0.50	0.30
Selection probabilities for good dose-levels																			
RtC-WT	0.75	0.79	0.39	0.81	0.78	0.50	0.80	0.79	0.45	0.98	0.67	0.95	0.68	0.94	0.80	0.74	0.18	0.70	0.23
WT	0.72	0.80	0.37	0.81	0.80	0.48	0.79	0.79	0.43	0.98	0.67	0.95	0.71	0.95	0.82	0.74	0.19	0.69	0.24
Two-Arm	0.61	0.75	0.26	0.76	0.75	0.38	0.71	0.76	0.34	0.99	0.53	0.97	0.68	0.95	0.81	0.68	0.22	0.63	0.28

To compare the Two-Arm design with the others, we still consider there to be five dose-levels, with the first dose-level being a separate arm and dose-levels 2-5 comprising the dose-finding arm. Also, since the two arms can be

considered independent in scenarios where stopping is preferred, this would only stop the dose-finding arm so patients at dose-level 1, the control arm, can still be recruited.

Across most scenarios RtC-WT and WT designs outperform the Two-Arm approach in terms of selection of the OBD if only by a few percentage points. Since the Two-Arm approach is essentially the same as the standard WT design but with fewer patients, it is expected to be slightly inferior as with most dose-finding studies a higher number of patients yield better operating characteristics. However, with the RtC-WT design actively recruiting to the control dose there should be an equal number of patients receiving control and treatment dose-levels to equate to similar operating characteristics. There appears to be some added benefit in terms of modelling for including control as a dose-level rather than using the Two-Arm approach when it comes to selecting the OBD. It should be mentioned that the differences in selection of the OBD are only slight between 1% and 8%. In terms of MC error, which was discussed in Chapter 2, Section 2.3.5, a slight difference of 1% can still be interpreted as significant. Also, the Two-Arm approach manages to allocate more patients to the OBD in certain scenarios (10 and 12) which is the monotone decreasing efficacy curve. In these scenarios having fewer dose-levels is an advantage as the earlier dose-levels are the most optimal making selection easier. It should be mentioned that an alternative specification of the Two-Arm approach may lead to better performance overall. In scenarios where stopping early would be ideal all three designs seem to behave similarly and have relatively comparable operating characteristics.

The performance of the RtC-WT and WT design is quite similar across all the scenarios. The key differences between RtC-WT and WT can be seen in the number of patients recruited to the control dose-level. The WT design recruits fewer patients here as expected since the randomisation probability is

not fixed. This usually results in more patients being allocated to the OBD. RtC-WT would experience situations where it would allocate a cohort to the control dose-level but the standard WT in the same situation would be able to randomise between the efficacious doses. This advantage however does not consistently yield better selection probabilities. Generally, we can say the WT design is not ideal if a trial aims to recruit control patients whilst conducting a dose-finding study. However, in the case where the control dose has high enough efficacy, the WT design can successfully still randomise patients there as can be seen in these simulations. It is possible under a different parametrisation, perhaps a design with a larger adaptive randomisation phase those numbers of control patients would be higher, although this may negatively impact performance. Overall, we can see the trade-off between allowing for this recruitment to control and overall performance. Based on these simulations and scenarios it seems RtC-WT performs relatively well.

3.5 Efficiency of an efficacy test

Typically seamless Phase I/II adaptive trial designs, such as the Wages and Tait design, allow us to conduct dose-finding whilst considering both toxicity and efficacy outcomes. However, they cannot often make comparisons between experimental treatment and placebo/control which would typically occur in a randomised Phase II setting. Our design, RtC-WT, takes these seamless Phase I/II designs one step further by incorporating a control arm which allows us to make these comparisons.

Trials that use an efficacy outcome investigating multiple doses along with a control dose will usually include a test of differences between the selected dose and control. If RtC-WT would be implemented in an actual trial it would be plausible to conduct an efficacy test between the control arm and eventual

OBD. A two proportions test [77] could be utilised which would simply compare the proportion of efficacy events in the control arm with the proportion of efficacy events at the OBD. However, dependent on the exact circumstance of that trial and the data observed the efficiency of the efficacy test may vary. To evaluate RtC-WT and the efficiency of conducting this test we can use our previous simulation results and calculate the power that each specific trial would generate.

We will utilise simulation results from the design in Section 3.4.4. For each relevant scenario and individual trial run, we will take the number of patients treated at the control dose and the number of patients treated at the OBD to conduct power calculations. These results will then be aggregated and summarised.

Power calculations will be done for a two-sided hypothesis test of the difference between two independent proportions using the effect size. These methods are detailed by Cohen [78]. The proportions are compared by looking at their difference which is calculated from transformed values of the proportions in both groups.

Let P_1 and P_2 represent the proportions for the two arms then the effect size is represented by the difference h :

$$h = \psi_1 - \psi_2 \quad (3.20)$$

where

$$\psi_e = 2\arcsine(\sqrt{P_e}), e = 1, 2. \quad (3.21)$$

To calculate the power we need to specify the effect size. Cohen [78] suggests using 0.2, 0.5 and 0.8 as effect sizes and indicated these could be interpreted as small, medium and large effects respectively. Our example design has an efficacy rate specified in the control arm of 30% and an efficacy lower bound of 50% which informs the futility stopping rule. From this, we could say the proportion of patients we expect to have an efficacy event in the control arm is 0.3 and a minimum of 0.5 in the OBD. Plugging these values into the equations above gives an effect size of 0.41 which we also investigate. For our calculations we considered multiple null hypotheses of $H_0 : h = 0.2, 0.41, 0.5, 0.8$ and the alternative hypothesis of $H_1 : h \neq 0.2, 0.41, 0.5, 0.8$ respectively. The type I error rate was set at 10%.

Table 3.11 lists the effect sizes used for the power calculations. For each effect size, the power (1 - the type II error probability) was calculated. It should be noted that the value of the effect size does not directly match values in the differences in the proportions. For example proportions of 0.21 and 0.1 will give an effect size of 0.3 but so will proportions of 0.55 and 0.44. In our simulations, we have set the efficacy rate in the control arm at 30% so here it is reasonable to assume that on average the proportion in the control arm will be similar. We can then calculate, using our effect size, what we would expect the proportion in the OBD arm to be.

TABLE 3.11: List of effect sizes used in power calculations.

Effect Size	P_{OBD}	$P_{Control}$	$P_{OBD} - P_{Control}$
0.2	0.395	0.3	0.095
0.41	0.5	0.3	0.2
0.5	0.544	0.3	0.244
0.8	0.689	0.3	0.389

For scenarios where stopping was the optimal outcome, we did not calculate power. In these scenarios, we would not want to reach this stage as either the treatment was found to be ineffective or too toxic. Similarly, power was

also not calculated in individual trial runs where stopping occurred in other scenarios. There were a small number of runs in which either no patients were recruited to the control dose or the OBD was declared as a dose that had not treated any patients. In both of these instances, power could not be calculated as the number of patients for either the control or OBD arm is 0.

Table 3.12 details the power calculations which are based on the results of 10000 simulations for each scenario. For each effect size, the average power is presented. In some instances, this average is not across the 10000 simulated trials but a smaller number due to either stopping rules or insufficient patient numbers. For reference, we also present the total number of trials in which power is calculated for each scenario.

TABLE 3.12: Mean power achieved across multiple scenarios for different assumptions of effect size.

Scenario	Effect Sizes - Mean Power (sd)				
	0.2	0.41	0.5	0.8	N
1	0.15 (0.02)	0.28 (0.07)	0.36 (0.09)	0.64 (0.14)	8445
3	0.15 (0.02)	0.3 (0.07)	0.38 (0.09)	0.67 (0.14)	7096
4	0.15 (0.02)	0.29 (0.07)	0.37 (0.09)	0.66 (0.14)	8771
6	0.15 (0.02)	0.3 (0.07)	0.38 (0.09)	0.68 (0.14)	7490
7	0.15 (0.02)	0.28 (0.06)	0.36 (0.09)	0.64 (0.14)	8777
9	0.15 (0.02)	0.3 (0.07)	0.38 (0.09)	0.67 (0.14)	7358
10	0.15 (0.02)	0.3 (0.07)	0.39 (0.09)	0.68 (0.14)	9556
12	0.15 (0.02)	0.31 (0.07)	0.4 (0.1)	0.7 (0.14)	9391

We can see for the effect sizes 0.2, 0.41 and 0.5 power is relatively low ranging from 0.15 to 0.4 depending on the scenario. For a large effect size, the average power is a bit higher ranging from 0.64 to 0.7. The higher the power the lower the probability of committing a type II error wherein the hypothesis test fails to reject a false null hypothesis. Our design appears to have somewhat reasonable power to detect larger differences in the proportions of efficacy events in the control and OBD arm. Altering the parameters of our design

could lead to results that gives us a higher power but we would also need to evaluate how those parameters impacted the dose-finding aspect of the design.

The main way to increase the power would be to increase the sample size specifically by having more patients in the control and OBD arm. Increasing the total sample size of the trial would not always achieve this goal as the extra patients could potentially be recruited into other dose-levels. One way around this would be to utilise an idea like a dose expansion cohort. Once the OBD has been established recruitment could continue and patients could be randomised to either the control arm or the OBD to better power any efficacy comparisons. Alternatively, a two-stage approach could be used where the first stage consists of dose-finding which is then built upon in a subsequent stage to compare efficacy more efficiently.

3.6 Discussion

In this chapter, we proposed RtC-WT, an extension to the seamless Phase I/II Wages & Tait trial design to allow for the ability to randomise patients to a control dose. The main motivation was to add a control arm as a dose level to achieve similar objectives as a randomised phase II study whilst maintaining efficiency in the dose-finding process. We examined the impact of various combinations of the AR phase size and the probability of randomising to control. We then compared RtC-WT to alternative approaches. We also assessed the impact of different efficacy skeletons and assumptions we made about the control dose on the operating characteristics of the design. In summary, we found that RtC-WT maintains reasonable operating characteristics when randomising patients to a control dose, although this depends on the parametrisation of the design.

When examining the various combinations of AR phase sizes and randomisation to control probabilities we consistently saw several issues with the extreme values of AR phase sizes. For instance, when the AR phase lasts the whole trial the design had some issues regarding stopping for excess toxicity. There were also problems generally with the design when attempting to stop for futility. It may be necessary to employ alternative stopping rules to achieve acceptable probabilities of stopping in certain scenarios. Also, when there was no AR phase the design had issues locating the OBD. This led to the recommendation of using 25-75% of the trial size as the AR phase as these were the values we investigated.

In terms of randomisation probabilities, we looked at two different possibilities in conjunction with the various AR phase sizes. There are practically countless combinations that could have been investigated and different combinations may have led to alternative conclusions. What can be said however is that in a practical clinical trial setting, thought should be given as to how many control patients the investigators want to recruit and then use that to determine the optimal sample size and AR phase size. For example, if 20 control patients are desired from a maximum sample size of 60 a randomisation to control probability of 0.66 for an AR phase size of 30 could be used or a 0.5 randomisation provability with an AR size of 40. Also, here it may be unfeasible to look at every combination but a sample should be investigated to see how it impacts the design.

Furthermore, the scenarios we presented in our simulations are again only a small handful of all possible scenarios that could be investigated. The scenarios we chose represent a variety of dose-efficacy curves which may be plausible in this trial setting. Another limitation of the simulations is how toxicity and efficacy data were generated. We have assumed that both of these events are independent but in a practical scenario, this may not be the case. A patient

may withdraw from the trial after having a toxicity, meaning no efficacy data can be observed for that individual. Further work could be done to look at the impact of sampling dependent toxicity and efficacy data.

Additionally, the model does not aim to balance toxicity against efficacy or consider a trade-off between the two like some designs do for example EffTox. The Wages and Tait design and the RtC-WT select doses from a subset of tolerable doses based on the probability of efficacy. This guarantees that the dose chosen will be safe but there may be a slightly less effective dose which is a lot less toxic which may be more appropriate or could be considered optimal. Additional scenarios could be investigated that look into how this balance works and those results could be contrasted against designs like EffTox that are designed with those relationships in mind.

To our knowledge, there are no other phase I/II designs that share the same aims as RtC-WT as such it is difficult to make comparisons with alternative approaches. We opted to compare our design against an easy-to-implement two-arm method. Although our design outperformed the two-arm approach in the scenarios presented we did not optimise the design of the dose-finding arm. Usually specifying a design for a dose-finding study is an iterative process in which certain parameters are tweaked to produce optimal operating characteristics. As such a more optimised two-arm approach could yield better performance than RtC-WT. The same could be said for the RtC-WT design and there could exist a better combination of parameters for the example trial we presented.

For all our simulations and when comparing designs, we used the same exemplar trial. This involved a control dose that had a known toxicity and efficacy profile. The probability of observing an efficacious event at this dose was specified at 30%. Even though this is relatively high when comparing our design directly to a standard Wages and Tait design we saw an increase in the

number of patients at the control dose and no real loss in terms of performance. This shows our design achieves what was intended as well as the fact that the standard Wages and Tait design can perform a similar job if the efficacy rates of the control dose are high enough. One caveat to this however is that we have just looked at one example trial and these conclusions may vary for a trial in which the control dose is not quite as efficacious. Further work could be done comparing these designs using different trials with different underlying assumptions.

If choosing between these two designs to be implemented into an actual trial there may be multiple things to consider. Whilst the RtC-WT and WT perform similarly in these simulations, in practice this may not be the case. The RtC-WT may be a more flexible option. For example, in the case where it is assumed that the control dose would have a relatively high probability of efficacy, simulations would show several patients being recruited to that dose; however, if in the actual trial the efficacy rate at the control dose is much lower than anticipated you would end up with fewer patients at that dose than the initial simulations would suggest. With the RtC-Wt this would not be the case since the probability to randomise to the control dose is fixed up front and would remain constant throughout a variety of scenarios as we have demonstrated.

There may also be many practical considerations to accommodate when considering implementing RtC-WT into an actual trial. For example, the adaptive randomisation component may require validation from multiple statisticians every time the probabilities are updated. In the case of RtC-WT this would need to be done for each cohort. As statisticians are a rare resource in most trial units this may be a limitation to using the design. It could be said however in the case of Wages and Tait and other dose-finding trials which use adaptive randomisation that patients are not being adaptively randomised but

rather the dose-levels are. This distinction may circumvent the need to have the randomisation probabilities validated. Typically, in normal dose-finding studies, patients who enter the trial later on are more likely to receive the higher dose-levels dependent on whatever escalation occurred so far. With RtC-WT and Wages and Tait method in general patients will be able to receive any of the admissible doses with probabilities scaled to the most efficacious doses.

It should be noted that the inclusion of a control dose depends on the definition of efficacy and toxicity events. It should be possible to measure the efficacy and toxicity events in the control arm. For example, the standard of care may aim to treat the underlying symptoms of a condition whereas the treatment aims to reduce some biological measure that the standard of care does not impact. This is more of a practical concern to determine if RtC-WT is a suitable design for a trial since during the modelling phase if no toxic or efficacy events are observed for the control this should not be an issue.

We also explored how efficient our design would be when conducting efficacy tests. The power for detecting large differences in the effect size ranged from 64-70%. Most Phase II trials would strive to achieve higher power and may be able to detect smaller differences. Further work could be done by building upon this design to achieve this same goal. One other limitation is that whilst we have used some form of randomisation it is not entirely similar to randomisation that may occur in a Phase II setting. So, it may be possible when using RtC-WT there is an imbalance in key patient characteristics when comparing the control and OBD doses.

A recent development by Yan et al. [79] suggested three alternative strategies of adaptive randomisation for the Wages and Tait design. They showed that their final recommended strategy achieved better accuracy when selecting the OBD as well as allocating more patients to that dose-level as well. The

strategy works by gradually excluding efficacy skeletons as data is collected. RtC-WT could be improved further by the incorporation of this alternative adaptive randomisation method as we have already shown its effectiveness in comparison to the traditional Wages and Tait design.

In the paper by Mozgunov et al. [75] they compare the use of a 4 parameter EMAX model with a CRM one parameter model, a two parameter logistic model with and without randomisation. It was found that the four parameter model with the inclusion of randomisation out performed the alternative approaches in terms of identification of the optimal dose combination. It was also shown that the two parameter logistic models performed relatively well and out performed the one parameter model. For the RtC-WT design one limitation is we only investigated use of one parameter models for toxicity and efficacy. Based on the work by Mozgunov et al. [75] the use of a two parameter model may yield better performance from our design as it would be able to model the probability of toxicity at the control dose and the experimental doses separately. Based on the randomisation in our design patients would be allocated to doses around two points the control dose and OBD. Thus, more flexible models may be more appropriate. Considering our aims were to investigate the adaptive randomisation component of the WT design to see if it could be leveraged to include a control arm optimisation of this design could be apart of further research specifically investigating the benefit of using different models.

Whilst relatively simplistic, our modification allowed for the inclusion of a control dose-level to the Wages and Tait method without compromising operating characteristics. We presented results from multiple simulations showing good performance, especially in non-monotonic efficacy scenarios. Further

work could be done to consider a wider range of design specifications and scenarios. We utilised a relatively simple trial scenario and made a lot of assumptions that may not necessarily be applicable in a real-world trial setting.

Chapter 4

Extending Dose Transition Pathways for use in TITE-CRMs

4.1 Introduction

Dose transition pathways (DTPs), were developed as a tool by Yap et al. [31] in order to address some of the issues around understanding and implementing complex and innovative designs. DTPs can be considered a tool primarily for dose-finding trials where the primary objective is to determine an MTD (or TD% at a specific target toxicity level) which is determined by the occurrence of DLTs recorded as a binary variable. The purpose of DTPs is to aid the design and analysis of these types of trials. This is done by projecting in advance the dose-escalation decisions for future cohorts based on the different potential data that may accrue. It can also be used as a calibration tool during the design stage of a trial to ascertain how the model behaves under certain outcomes and modify its specifications if necessary. These projections can then be visualised and help illustrate how the model operates and communicate possible future decisions that may be made.

In the paper by Yap et al. [31], DTPs are introduced through their illustrative use in a trial with a CRM design. They discuss that the idea of DTPs can

be extended to other model-based designs such as the TITE-CRM and phase I/II designs that consider efficacy and toxicity. DTPs are produced based on the different potential outcome data that will be collected and for both of these possible extensions, the outcome data is more complex which makes producing DTPs for these designs slightly more challenging. Specifically, for the TITE-CRM additional data is needed depending on if the patient is to be fully or partially weighted. So, if the patient has not experienced a DLT they will be partially weighted in the model based on the time they have spent in the trial. This makes mapping out dose-escalation decisions difficult since we are no longer dealing with a simple binary variable of DLT/no DLT.

In this chapter, we aim to explore the potential of extending DTPs for use in TITE-CRMs. We start by providing an example of a DTP for a simple CRM design to better understand what they are and how they can be used. We then look at some of the issues with extending DTPs to TITE-CRMs and present possible solutions for how they may be overcome.

4.2 Dose Transition Pathways

To explore how DTPs can be extended for use in TITE-CRMs we first look at how they would be used for a simple CRM trial. This will involve looking at how CRM designs are implemented and analysed and how DTPs can be incorporated into these processes.

When implementing a CRM design multiple parameters need to be considered and specified. These are the number of doses, target toxicity level, dose-toxicity model, dose-toxicity skeleton, method of inference, decision rules, sample size, cohort size, safety modifications, and stopping rules [18]. This usually requires input from multiple stakeholders such as the statistician, clinician

and trial management team. Typically, once these parameters have been specified, simulations can be run for various clinically relevant scenarios to obtain operating characteristics for the specified CRM design. At this point, these results can be reassessed and the specifications of the CRM can be updated. This whole process can be repeated iteratively until an acceptable design is reached.

Even after multiple rounds of simulations, there is still the risk that a statistically optimal design may not be optimal in practice. This could be due to the scenarios used in simulations not being representative of what is observed during the running of the trial. This may also occur when model recommendations are deemed clinically unacceptable resulting in dose-escalation decisions recommended by the model being ignored or overruled. Whilst dose recommendations from the model should be used as guidance, if its recommendations are constantly being ignored it undermines the model and brings into question its specification. DTPs can be used to avoid this from occurring and can help calibrate the model. They provide insight during the design stages of the trial into what recommendations the model will make under the range of potential outcomes. This will help clinicians as well as statisticians better understand how the model works and calibrate it accordingly so clinically reasonable decisions are being made.

DTPs can also be utilised during the analysis of a trial. In a 3+3 trial just by knowing the rules of the design we already know what dose-escalation decision will be made for all possible outcomes of one cohort. If zero out of three patients (0/3) experience a DLT we escalate to the next highest dose; if one out of three patients (1/3) experiences a DLT we recruit a further 3 patients to the same dose-level; and if two out of three (2/3) patients experience a DLT we de-escalate to a lower dose or stop the trial if already at the lowest dose. This can be done and computed without a statistician. On the other hand, we

have designs such as the CRM where this is not as simple since the next recommended dose will be based on the accrued data. Here DTPs can be utilised to present this information by analysing all possible outcomes and summarising the dose recommendations these lead to.

The number of pathways can be calculated using the number of cohorts (x) and the number of patients in each cohort (y). Here, the total sample size is xy and the number of pathways is $(y + 1)^x$. So, for a trial recruiting 10 cohorts of 3 patients the number of pathways would be over 1 million ($[3 + 1]^{10} = 1,048,576$). There may be fewer pathways based on any stopping rules which causes the trial to stop early. Presenting this many pathways is difficult and may also be unintuitive. Yap et al. [31] suggest using the first group of cohorts to help facilitate discussion with investigators during the design stage of the trial. DTPs can also be updated during dose-escalation phases as well by incorporating the accrued data and then projecting pathways for subsequent cohorts. In the next section, section 4.2.1 we provide a generic trial example to show how DTPs can be implemented.

4.2.1 Example trial to illustrate DTPs

Consider an early phase trial aiming to determine the TD25 of a single agent; Table 4.1 details all the parameters we need to set up the CRM design. First, we specify the clinical parameters; there will be 5 dose levels (d_1, \dots, d_5), the trial will start at dose-level 2 (d_2), the target sample size will be 30 patients, patients will be recruited in cohorts of 3 (10 cohorts overall) and the target DLT rate will be 25% (TD25). A single-stage CRM will be used with a power model to model the dose-toxicity curve, Bayesian estimation methods will also be used. Prior estimates for the DLT rates are specified as 0.04, 0.08, 0.16, 0.25 and 0.35 for dose-levels 1-5 respectively, this assumes dose-level 4 (d_4) will be the TD25.

TABLE 4.1: Specification of parameters for an example CRM trial.

Parameter	Value
Number of dose-levels	5
Starting dose-level	2
Sample size	30
Cohort size	3
Target DLT rate	25%
Dose-toxicity model	Power model
Dose-toxicity skeleton	0.04, 0.08, 0.16, 0.25, 0.35
Method of inference	Bayesian

We assume the prior distribution for the slope parameter in the power model will be normally distributed with a mean of 0 and variance of 1.34. These values are based on work by O’Quigley and Shen [58] that proposed a suitable prior distribution would be a normal distribution with a mean of 0 and a sufficiently large variance. They go on to use a variance of 1.34 which was adopted by others. As this example is for only illustrative purposes and given its simplicity we feel these are adequate choices. Wheeler et al [18] also discuss choices of prior parameters when designing a dose-finding study. Obviously, in a real trial scenario more thought should be given to the selection of all parameters.

Most of the parameter specifications made here are for simplicity. In practice, these specifications should have either a clinical or statistical rationale behind them based on the context of the trial. Once an initial set of these parameters have been selected, simulations are conducted to assess the operating characteristics of the design under various scenarios. At this point, DTPs can also be generated.

For reference we have conducted a small simulation study to evaluate our example CRM. 10000 trials were simulated under six different scenarios, 5 where each of the dose-levels is the TD25 and a sixth where all doses are too toxic. The results for the simulations are presented in Table 4.2 and they show

the selection probabilities for each dose across each scenario.

TABLE 4.2: Selection probabilities from 10000 simulated trials under various scenarios for the example CRM.

		Dose Levels					
		1	2	3	4	5	Stop
Scenario	Prior DLT	0.04	0.08	0.16	0.25	0.35	
1:TD25 @1	True DLT rate	0.25	0.35	0.45	0.55	0.65	
	P(Select)	0.68	0.27	0.05	0	0	0
2:TD25 @2	True DLT rate	0.15	0.25	0.35	0.45	0.55	
	P(Select)	0.22	0.48	0.26	0.04	0	0
3:TD25 @3	True DLT rate	0.1	0.15	0.25	0.35	0.45	
	P(Select)	0.02	0.21	0.48	0.24	0.04	0
4:TD25 @4	True DLT rate	0.05	0.1	0.15	0.25	0.35	
	P(Select)	0	0.03	0.25	0.47	0.25	0
5:TD25 @5	True DLT rate	0.01	0.05	0.1	0.15	0.25	
	P(Select)	0	0	0.04	0.26	0.71	0
6:All toxic	True DLT rate	0.5	0.55	0.65	0.75	0.85	
	P(Select)	1	0	0	0	0	0

4.2.2 Using DTPs to calibrate the CRM

Since we have specified a sample size of 30 and cohort size of three that means we will have 10 cohorts and therefore 1,048,576 pathways ($[3+1]^{10} = 1,048,576$). Patients in each cohort are considered to either experience a toxic event (T) or have no toxic event (N). For the first cohort of three patients, there are four possible outcomes: all patients in the cohort experience no toxicity (NNN), one

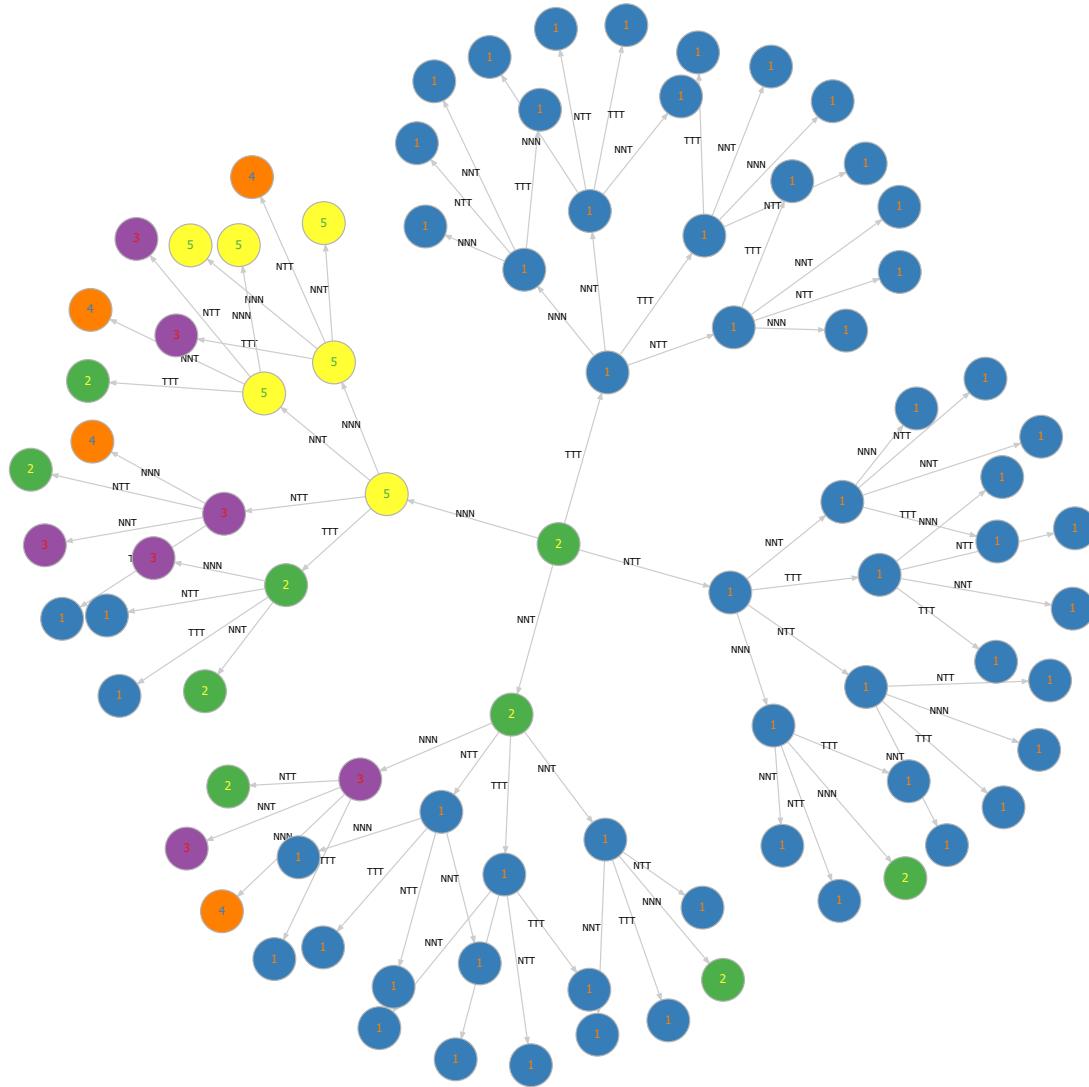
patient experiences a toxic event (NNT), two patients experience a toxic event (NTT) or all three patients experience a toxic event (TTT). For the subsequent cohort, the same set of four outcomes can be observed but in combination with the previous cohort, this creates 16 different outcomes for the first two cohorts (six patients). This process is then continued for each cohort creating exponentially more pathways.

Given the impracticalities of presenting and summarising all these pathways, we can instead present the pathways of the first three cohorts. In this case, there are only 64 potential pathways ($[3 + 1]^3 = 64$). Table 4.3 lists all the pathways for the first three cohorts of patients. Similarly, we can also represent these pathways visually using either a node plot (Figure 4.1, generated using the R package escalation [21]) or a flow plot which was originally used to visualise the pathways when DTPs were first introduced. These have also been produced (Figure 4.2, generated using the R package dtpcrm [80]) but due to limitations with the software they only show the pathways for the first two cohorts.

TABLE 4.3: Initial DTP for the first three cohorts of our example CRM.

Pathway	Cohort 1		Cohort 2		Cohort 3		Cohort 4
	Dose	Outcomes	Dose	Outcomes	Dose	Outcomes	Dose
1	2	NNN	5	NNN	5	NNN	5
2	2	NNN	5	NNN	5	NNT	5
3	2	NNN	5	NNN	5	NTT	4
4	2	NNN	5	NNN	5	TTT	3
5	2	NNN	5	NNT	5	NNN	5
6	2	NNN	5	NNT	5	NNT	4
7	2	NNN	5	NNT	5	NTT	3
8	2	NNN	5	NNT	5	TTT	2
9	2	NNN	5	NTT	3	NNN	4
10	2	NNN	5	NTT	3	NNT	3
11	2	NNN	5	NTT	3	NTT	2
12	2	NNN	5	NTT	3	TTT	1
13	2	NNN	5	TTT	2	NNN	3
14	2	NNN	5	TTT	2	NNT	2
15	2	NNN	5	TTT	2	NTT	1
16	2	NNN	5	TTT	2	TTT	1
17	2	NNT	2	NNN	3	NNN	4
18	2	NNT	2	NNN	3	NNT	3
19	2	NNT	2	NNN	3	NTT	2
20	2	NNT	2	NNN	3	TTT	1
21	2	NNT	2	NNT	1	NNN	2
22	2	NNT	2	NNT	1	NNT	1
23	2	NNT	2	NNT	1	NTT	1
24	2	NNT	2	NNT	1	TTT	1
25	2	NNT	2	NTT	1	NNN	1
26	2	NNT	2	NTT	1	NNT	1
27	2	NNT	2	NTT	1	NTT	1
28	2	NNT	2	NTT	1	TTT	1
29	2	NNT	2	TTT	1	NNN	1
30	2	NNT	2	TTT	1	NNT	1
31	2	NNT	2	TTT	1	NTT	1
32	2	NNT	2	TTT	1	TTT	1
33	2	NTT	1	NNN	1	NNN	2
34	2	NTT	1	NNN	1	NNT	1
35	2	NTT	1	NNN	1	NTT	1
36	2	NTT	1	NNN	1	TTT	1
37	2	NTT	1	NNT	1	NNN	1
38	2	NTT	1	NNT	1	NNT	1
39	2	NTT	1	NNT	1	NTT	1
40	2	NTT	1	NNT	1	TTT	1
41	2	NTT	1	NTT	1	NNN	1
42	2	NTT	1	NTT	1	NNT	1
43	2	NTT	1	NTT	1	NTT	1
44	2	NTT	1	NTT	1	TTT	1
45	2	NTT	1	TTT	1	NNN	1
46	2	NTT	1	TTT	1	NNT	1
47	2	NTT	1	TTT	1	NTT	1
48	2	NTT	1	TTT	1	TTT	1
49	2	TTT	1	NNN	1	NNN	1
50	2	TTT	1	NNN	1	NNT	1
51	2	TTT	1	NNN	1	NTT	1
52	2	TTT	1	NNN	1	TTT	1
53	2	TTT	1	NNT	1	NNN	1
54	2	TTT	1	NNT	1	NNT	1
55	2	TTT	1	NNT	1	NTT	1
56	2	TTT	1	NNT	1	TTT	1
57	2	TTT	1	NTT	1	NNN	1
58	2	TTT	1	NTT	1	NNT	1
59	2	TTT	1	NTT	1	NTT	1
60	2	TTT	1	NTT	1	TTT	1
61	2	TTT	1	TTT	1	NNN	1
62	2	TTT	1	TTT	1	NNT	1
63	2	TTT	1	TTT	1	NTT	1
64	2	TTT	1	TTT	1	TTT	1

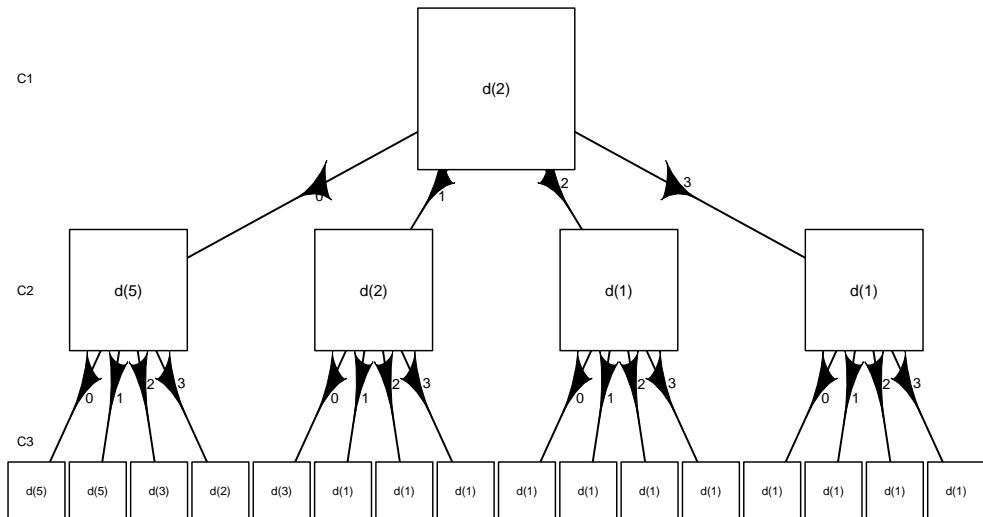
FIGURE 4.1: Initial DTP node plot for the first three cohorts of our example CRM.



From Table 4.3, looking at pathways 33-64, we can see that if there are two or more toxicities in the first cohort the CRM will always de-escalate the dose and if there are one or more toxicities in the next two cohorts it will stay at dose-level 1. We can also see from pathways 17-32 that if we observe a toxicity event in the first cohort we will stay at the same dose-level for the next cohort. If no toxicities occur we escalate straight to the highest dose.

Figure 4.1 also shows the same information. The central node represents the starting dose and first cohort, from here we have 4 branches showing the

FIGURE 4.2: Flow plot of the initial DTP for the first two cohorts of our example CRM.



various outcomes and which dose-level is allocated to the next cohort. In the case where each patient in the first cohort experience a DLT (TTT), we see that subsequent cohorts are all allocated to dose-level 1 regardless of their outcomes. Similarly, when two patients from the first cohort experience DLTs (NTT) all resulting branches show that dose-level 1 would be selected except in one case where no further DLTs occur and the CRM would escalate back to the starting dose. When one DLT occurs in the first cohort (NNT), we remain at the same dose-level. Looking at these branches if one or more DLTs are experienced in the next cohorts the dose-level is de-escalated, there is only potential for escalation in the scenario where no further DLTs occur. For the case when no DLTs occur (NNN) we see the dose for the second cohort escalated to dose-level 5. At this point, if the second cohort experiences 3 DLTs the CRM will de-escalate to dose-level 2. If there are only 2 DLTs the CRM goes to dose-level

3 and one or fewer DLTs and the next cohort will remain at dose-level 5. The flow plot, Figure 4.2, only shows outcomes up to the third cohort but can be interpreted similarly to the node plot.

In combination with operating characteristics from simulations, DTPs can be used to facilitate discussions to see if the CRM can be better calibrated and how it allocates doses. In our example here there may be a few things that would concern clinicians, the first being that we skip doses when escalating and the second that in the cases where lots of toxicity occurs recruitment continues. To remedy this we can include a rule to not skip untried doses and add a safety rule to stop the trial if too many toxicities occur at the lowest dose.

In a Bayesian setting, an appropriate method to stop early would be to test the posterior distribution for the probability of toxicity. For our example here we will stop if there is at least a 90% probability that the toxicity rate is 10% greater than the target level at the lowest dose. This can be expressed as $P(\text{true DLT rate at } d_1 > 0.25 + 0.1 \mid \text{observed data and prior information}) > 0.9$. With the addition of these two rules we now have a modified CRM design. Again, for reference, updated simulations are provided in Table 4.4. The same scenarios as presented in Table 4.2 were used to evaluate this new modified CRM.

TABLE 4.4: Updated selection probabilities from 10000 simulated trials under various scenarios for the example CRM with additional rules.

		Dose Levels					
		1	2	3	4	5	Stop
Scenario	Prior DLT	0.04	0.08	0.16	0.25	0.35	
1:TD25 @1	True DLT rate	0.25	0.35	0.45	0.55	0.65	
	P(Select)	0.66	0.26	0.05	0	0	0.02
2:TD25 @2	True DLT rate	0.15	0.25	0.35	0.45	0.55	
	P(Select)	0.23	0.47	0.25	0.04	0	0
3:TD25 @3	True DLT rate	0.1	0.15	0.25	0.35	0.45	
	P(Select)	0.03	0.21	0.48	0.24	0.04	0
4:TD25 @4	True DLT rate	0.05	0.1	0.15	0.25	0.35	
	P(Select)	0	0.03	0.25	0.46	0.26	0
5:TD25 @5	True DLT rate	0.01	0.05	0.1	0.15	0.25	
	P(Select)	0	0	0.04	0.26	0.71	0
6:All toxic	True DLT rate	0.5	0.55	0.65	0.75	0.85	
	P(Select)	0.34	0	0	0	0	0.66

So, now along with our new simulation results DTPs can also be updated. Table 4.5 shows the pathways for the first three cohorts. The node and flow plots were also updated, Figures 4.3 and 4.4 respectively. Since we included a rule to stop in the case of excess toxicity we see several pathways terminate early so overall there are fewer pathways compared to the initial set that was produced. Here we see six different branches where it recommended that the trial stop early (pathways 32, 44, 45, 53, 54, 55 Table 4.5). This can also be seen

in Figure 4.3, we can also see three of these nodes recommend stopping before recruiting a third cohort. Using the flow plot, in Figure 4.4, we can see that stopping is suggested when five out of the first six patients experience a DLT. Also, escalation of doses no longer skips dose-levels. With these new rules, we observe that if there are no DLTs in the first cohort the dose for the next cohort is dose-level 3 and not 5. We still observe that one toxicity in the first cohort leads to recruiting the next cohort at that same dose-level and with two or more toxicities de-escalation occurs.

FIGURE 4.3: Updated DTP node plot for the first three cohorts of our example CRM with additional rules.

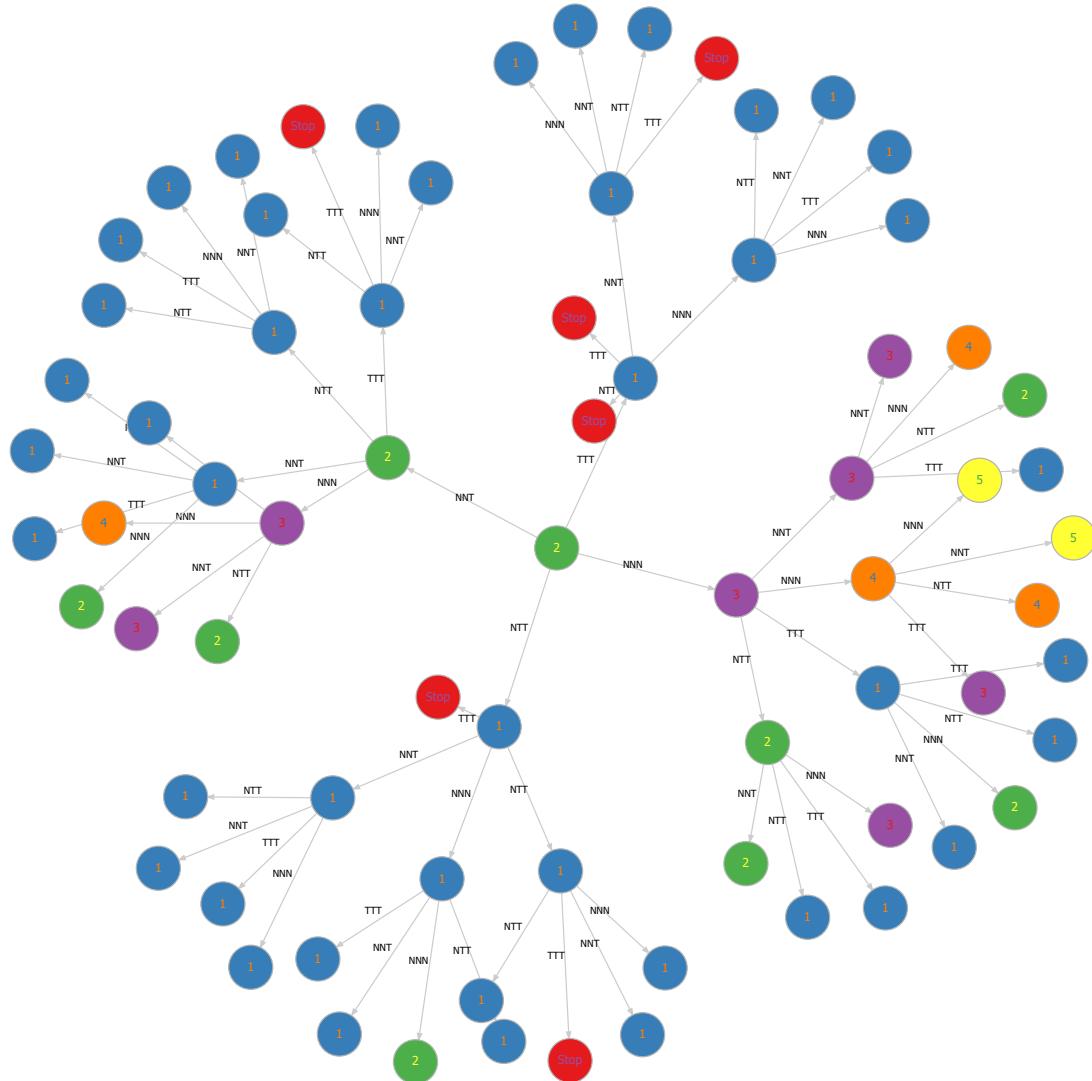


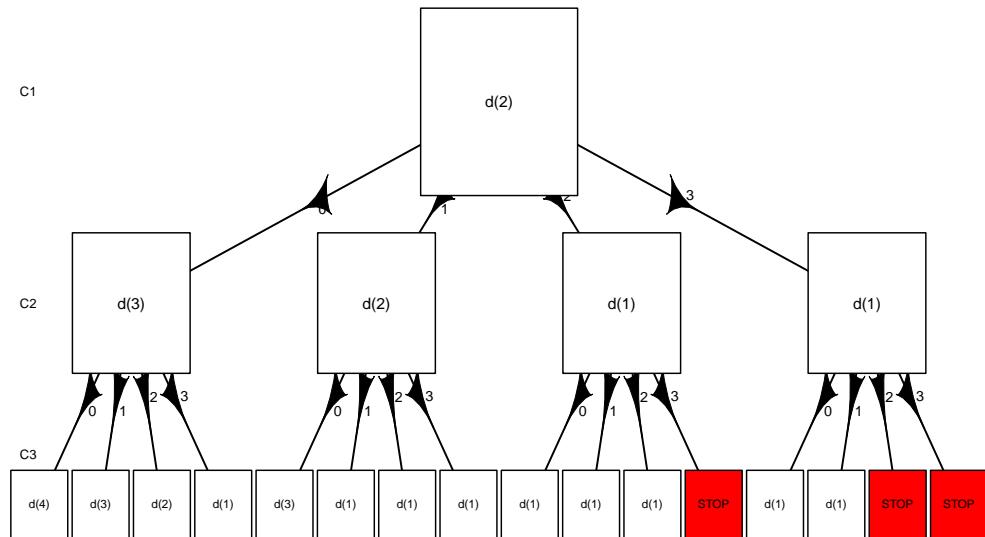
TABLE 4.5: Updated DTPs for the first three cohorts of our example CRM with additional rules.

Pathway	Cohort 1		Cohort 2		Cohort 3		Cohort 4
	Dose	Outcomes	Dose	Outcomes	Dose	Outcomes	Dose
1	2	NNN	3	NNN	4	NNN	5
2	2	NNN	3	NNN	4	NNT	5
3	2	NNN	3	NNN	4	NTT	4
4	2	NNN	3	NNN	4	TTT	3
5	2	NNN	3	NNT	3	NNN	4
6	2	NNN	3	NNT	3	NNT	3
7	2	NNN	3	NNT	3	NTT	2
8	2	NNN	3	NNT	3	TTT	1
9	2	NNN	3	NTT	2	NNN	3
10	2	NNN	3	NTT	2	NNT	2
11	2	NNN	3	NTT	2	NTT	1
12	2	NNN	3	NTT	2	TTT	1
13	2	NNN	3	TTT	1	NNN	2
14	2	NNN	3	TTT	1	NNT	1
15	2	NNN	3	TTT	1	NTT	1
16	2	NNN	3	TTT	1	TTT	1
17	2	NNT	2	NNN	3	NNN	4
18	2	NNT	2	NNN	3	NNT	3
19	2	NNT	2	NNN	3	NTT	2
20	2	NNT	2	NNN	3	TTT	1
21	2	NNT	2	NNT	1	NNN	2
22	2	NNT	2	NNT	1	NNT	1
23	2	NNT	2	NNT	1	NTT	1
24	2	NNT	2	NNT	1	TTT	1
25	2	NNT	2	NTT	1	NNN	1
26	2	NNT	2	NTT	1	NNT	1
27	2	NNT	2	NTT	1	NTT	1
28	2	NNT	2	NTT	1	TTT	1
29	2	NNT	2	TTT	1	NNN	1
30	2	NNT	2	TTT	1	NNT	1
31	2	NNT	2	TTT	1	NTT	1
32	2	NNT	2	TTT	1	TTT	STOP
33	2	NTT	1	NNN	1	NNN	2
34	2	NTT	1	NNN	1	NNT	1
35	2	NTT	1	NNN	1	NTT	1
36	2	NTT	1	NNN	1	TTT	1
37	2	NTT	1	NNT	1	NNN	1
38	2	NTT	1	NNT	1	NNT	1
39	2	NTT	1	NNT	1	NTT	1
40	2	NTT	1	NNT	1	TTT	1
41	2	NTT	1	NTT	1	NNN	1
42	2	NTT	1	NTT	1	NNT	1
43	2	NTT	1	NTT	1	NTT	1
44	2	NTT	1	NTT	1	TTT	STOP
45	2	NTT	1	TTT	STOP	NA	STOP
46	2	TTT	1	NNN	1	NNN	1
47	2	TTT	1	NNN	1	NNT	1
48	2	TTT	1	NNN	1	NTT	1
49	2	TTT	1	NNN	1	TTT	1
50	2	TTT	1	NNT	1	NNN	1
51	2	TTT	1	NNT	1	NNT	1
52	2	TTT	1	NNT	1	NTT	1
53	2	TTT	1	NNT	1	TTT	STOP
54	2	TTT	1	NTT	STOP	NA	STOP
55	2	TTT	1	TTT	STOP	NA	STOP

At this stage, further discussions could be held about the updated DTPs and simulations. Here there may be more subtle points to discuss such as the parameters of the stopping rule. Dependent on the clinical rationale investigators may be inclined to impose either looser or stricter stopping rules. In our example, this can be done by altering the threshold values in our test of the posterior distribution of the probability of toxicity at the lowest dose.

We also see in pathway 2 that an escalation occurs after observing a toxicity

FIGURE 4.4: Flow plot of the updated DTP for the first two cohorts of our example CRM with additional rules.



event in the previous cohort. This shows our design to be incoherent. A CRM design is considered coherent if escalation only occurs when the previous cohort experiences no DLTs and de-escalation only occurs when a DLT has been observed in the previous cohort. This property limits the risk of unnecessarily exposing patients to toxic doses whilst also ensuring patients get treated at a reasonable dose within the safety limit [81]. This became an issue due to the rule we enforced not to skip doses in escalation, the previous design without this rule was coherent. Further rules could be added to ensure the design remains coherent such that escalation will only take place if the previous cohort experience no DLTs likewise, de-escalation will only occur if the previous cohort did experience DLTs.

We have mentioned how DTPs can be used in conjunction with operating characteristics from simulations to assist with the design of a trial. There is

a further added benefit that probabilistic inference can be used with DTPs to ascertain exact operating characteristics of a design. This work was developed by Brock [21] as part of the escalation package which he referred to as crystallised dose-paths. As we do with simulations for a specific scenario, if we specify an assumed truth about the toxicity probabilities at each dose we can calculate the likelihood of each dose-path according to that truth. From this we can then determine the probability of recommending each dose or stopping the trial. The benefits of calculating operating characteristics this way is that it can potentially be quicker than simulations. This is specifically true for smaller trials where if we were to run 10000 iterations for a trial with five cohorts of three patients that would require 50000 model fits. Whereas with crystallised dose-paths models only have to be fit once at each node which in this example corresponds to 1365 model fits. The first cohort has four outcomes NNN, NNT, NTT and TTT which requires four model fits. These four nodes each then have four outcomes representing outcomes for the second cohort which leads to 16 additional model fits. So, for each subsequent cohort the number of model fits increases exponentially based on the number of outcomes that can be observed. Even in instances where it may not be more efficient to produce these crystallised dose-paths they have the added benefit of not suffering from Monte Carlo error. There is no uncertainty associated with these exact probabilities but rather about which path will be taken based on outcomes observed. There are some inherent advantages that mean in practice it may be a better option to use dose-paths over simulations.

Here we have highlighted how DTPs can be utilised during the initial stages of setting up a trial. Due to our example, some changes could be implemented into our suggested design to improve it. However, this was to illustrate what the pathways look like and how they can be used to facilitate discussions with the relevant clinicians and the trials team. Although we just looked at DTPs

any changes being made to the design should also take into account operating characteristics from simulations or from crystallised dose-paths. CRM designs may not be intuitively understood by clinicians but DTPs should help make them more accessible. Next, we will look at how DTPs can be used during a trial.

4.2.3 Using DTPs during a trial

Dependent on the size of the dose-finding trial it will often be infeasible to present all the different pathways. In our example with 10 cohorts of 3 patients there are approximately 1 million different pathways. As the trial progresses we observe outcomes for each patient and thus the number of pathways is reduced. This makes it possible to present DTPs for future cohorts of patients once we have accrued the outcome data of previous cohorts.

DTP's main use in the design stage is allowing you to see how the model behaves with certain data generated by the earlier cohorts and we can see if escalation and stopping are occurring as expected. We can then also communicate more effectively with clinicians and investigators about what our design is doing. So, once a dose-finding trial has been designed, we can continue using DTPs whilst we are accruing data to project in advance dose decisions that may occur. This has the potential to alleviate some of the burden place on a statistician during a trial. for example clinicians may be interested in understanding what future dose decisions may be if a certain number of DLTs were to be observed and rather than having the statistician run the models DTPs generated ahead of time could be used. Additionally, the time between the recruitment of cohorts could also be reduced if the next recommended dose is the same dose regardless of the outcomes observed in the current cohort. It also allows the statistician to check that the model is still escalating and stopping as

expected. Although it may be more difficult to make changes to the design of the trial once it is underway and typically this isn't usually done unless there are extenuating circumstances.

To see how this would work in practice we will use the same example as specified in Section 4.2.1 along with the stopping rule we introduced in Section 4.2.2. Essentially, the same design that was used to produce the DTPs in Table 4.5 and Figure 4.3. Consider a trial where we observe outcomes for the first three cohorts of patients that match pathway 6 in Table 4.5. That implies the first cohort of patients is recruited to dose-level 2 and no toxicities are observed, cohort 2 is allocated to dose-level 3 where one toxicity is observed, then cohort 3 is allocated to dose-level 3 where again only one toxicity is observed and that leads the model recommending dose-level 3 for cohort 4. To refer to previous cohorts' outcomes we will use the nomenclature introduced by Brock [68]. Outcomes for patients, either toxicity (T) or no toxicity (N) are strung behind a numeric dose-level. For instance, 2TTN denotes a cohort of three patients that were allocated to dose-level 2, two of whom experienced toxicity and one who did not. In our example, using pathway 6 from Table 4.5, these outcomes can be denoted as 2NNN 3NNT 3NNT. In this scenario we are unsure about the toxicity of dose-level 3 as we have seen toxicities in two separate cohorts, however, this is mainly due to only having recruited three cohorts. If we consider this our new starting point we can produce new DTPs based on the observed data at that time.

Table 4.6 shows the new set of DTPs following previous outcomes (2NNN 3NNT 3NNT), these are also visualised in Figure 4.5. For each pathway, cohort 4 patients start at dose-level 3 as this is the model recommendation based on the previously observed outcomes. We have 64 pathways again which indicates that regardless of how many toxicities are observed the trial does not recommend stopping. In pathway 64 three patients have a toxicity event at

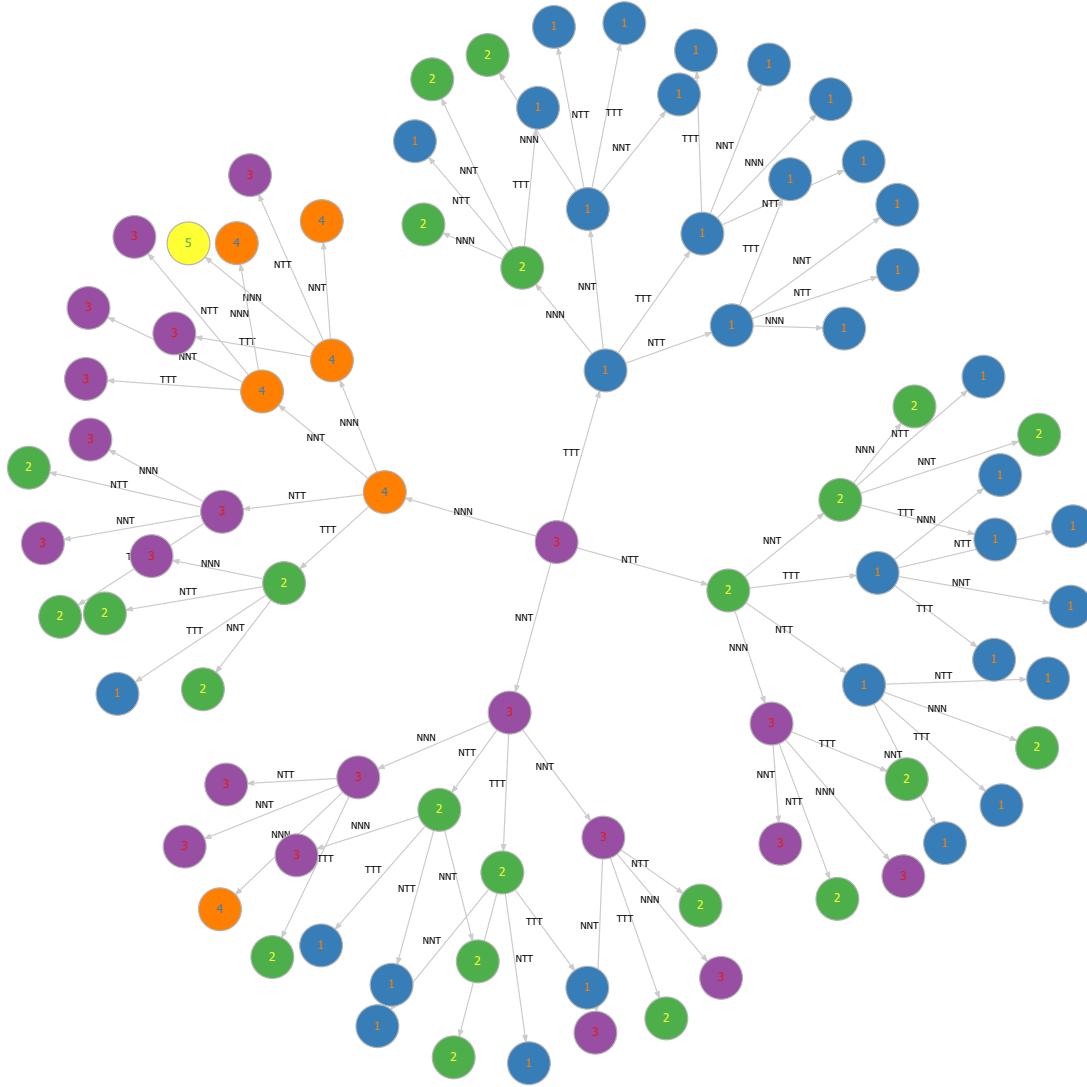
dose-level 3 and then six have toxicities at dose-level 1. This does not necessarily mean that the stopping rule is not working as intended rather due to the non-toxicities observed in the first three cohorts there would need to be more toxicity events before the rule we specified is triggered. This could be investigated further by looking at the DTPs following the outcomes of pathway 64 (i.e 2NNN 3NNT 3NNT 3TTT 1TTT 1TTT).

It can also be seen if there are two in cohort 4 (pathways 33-48) we de-escalate to dose-level 2 and if there are three (pathways 49-64) we de-escalate to dose-level 1. If one toxicity occurs we stay at dose-level 3 (pathways 17-32). If no toxicities are observed we escalate to dose-level 4 (pathways 1-16). There are also only four pathways where we end up at a higher dose in cohort 7 (pathways 1, 2, 5 and 17). Given the data we have already observed and if another toxicity occurs in cohort 4 there would need to be two cohorts of no toxicities before escalation can take place (pathway 17).

TABLE 4.6: DTPs for three additional cohorts after observing outcomes for the first three cohorts.

Pathway	Cohort 4		Cohort 5		Cohort 6		Cohort 7
	Dose	Outcomes	Dose	Outcomes	Dose	Outcomes	Dose
1	3	NNN	4	NNN	4	NNN	5
2	3	NNN	4	NNN	4	NNT	4
3	3	NNN	4	NNN	4	NTT	3
4	3	NNN	4	NNN	4	TTT	3
5	3	NNN	4	NNT	4	NNN	4
6	3	NNN	4	NNT	4	NNT	3
7	3	NNN	4	NNT	4	NTT	3
8	3	NNN	4	NNT	4	TTT	3
9	3	NNN	4	NTT	3	NNN	3
10	3	NNN	4	NTT	3	NNT	3
11	3	NNN	4	NTT	3	NTT	2
12	3	NNN	4	NTT	3	TTT	2
13	3	NNN	4	TTT	2	NNN	3
14	3	NNN	4	TTT	2	NNT	2
15	3	NNN	4	TTT	2	NTT	2
16	3	NNN	4	TTT	2	TTT	1
17	3	NNT	3	NNN	3	NNN	4
18	3	NNT	3	NNN	3	NNT	3
19	3	NNT	3	NNN	3	NTT	3
20	3	NNT	3	NNN	3	TTT	2
21	3	NNT	3	NNT	3	NNN	3
22	3	NNT	3	NNT	3	NNT	3
23	3	NNT	3	NNT	3	NTT	2
24	3	NNT	3	NNT	3	TTT	2
25	3	NNT	3	NTT	2	NNN	3
26	3	NNT	3	NTT	2	NNT	2
27	3	NNT	3	NTT	2	NTT	1
28	3	NNT	3	NTT	2	TTT	1
29	3	NNT	3	TTT	2	NNN	2
30	3	NNT	3	TTT	2	NNT	1
31	3	NNT	3	TTT	2	NTT	1
32	3	NNT	3	TTT	2	TTT	1
33	3	NTT	2	NNN	3	NNN	3
34	3	NTT	2	NNN	3	NNT	3
35	3	NTT	2	NNN	3	NTT	2
36	3	NTT	2	NNN	3	TTT	2
37	3	NTT	2	NNT	2	NNN	2
38	3	NTT	2	NNT	2	NNT	2
39	3	NTT	2	NNT	2	NTT	1
40	3	NTT	2	NNT	2	TTT	1
41	3	NTT	2	NTT	1	NNN	2
42	3	NTT	2	NTT	1	NNT	1
43	3	NTT	2	NTT	1	NTT	1
44	3	NTT	2	NTT	1	TTT	1
45	3	NTT	2	TTT	1	NNN	1
46	3	NTT	2	TTT	1	NNT	1
47	3	NTT	2	TTT	1	NTT	1
48	3	NTT	2	TTT	1	TTT	1
49	3	TTT	1	NNN	2	NNN	2
50	3	TTT	1	NNN	2	NNT	2
51	3	TTT	1	NNN	2	NTT	1
52	3	TTT	1	NNN	2	TTT	1
53	3	TTT	1	NNT	1	NNN	2
54	3	TTT	1	NNT	1	NNT	1
55	3	TTT	1	NNT	1	NTT	1
56	3	TTT	1	NNT	1	TTT	1
57	3	TTT	1	NTT	1	NNN	1
58	3	TTT	1	NTT	1	NNT	1
59	3	TTT	1	NTT	1	NTT	1
60	3	TTT	1	NTT	1	TTT	1
61	3	TTT	1	TTT	1	NNN	1
62	3	TTT	1	TTT	1	NNT	1
63	3	TTT	1	TTT	1	NTT	1
64	3	TTT	1	TTT	1	TTT	1

FIGURE 4.5: Node plot of three additional cohorts after observing outcomes for the first three cohorts.



Once cohort 7 is reached in the trial the DTPs can be updated again using the observed outcomes of whichever pathway occurred. These will show the potential pathways up to the 10th cohort, which is the maximum sample size in our example, and will also detail the final dose recommendation. Throughout the trial the DTPs allow us to map out what doses are recommended until the final decision.

DTPs can also work with non-uniform cohorts. So far our examples have

been fairly simple however, in practice there may be complications with running a dose-finding trial. For instance, there may be issues with recruitment leading to long time periods between the evaluation of cohorts and dose decisions. This may be due to a number of factors such as a lack of recruiting sites, underestimation of the prevalence of disease in the patient population, or a global pandemic. One solution to this may be to reduce the cohort size and make dose decisions earlier. Model-based designs are fairly flexible at dealing with issues like these as the model could just be updated after fewer patients instead. As these considerations may not have been made in the design stages of the trial, DTPs could be used as a way to evaluate any changes to cohort sizes that are made during the trial.

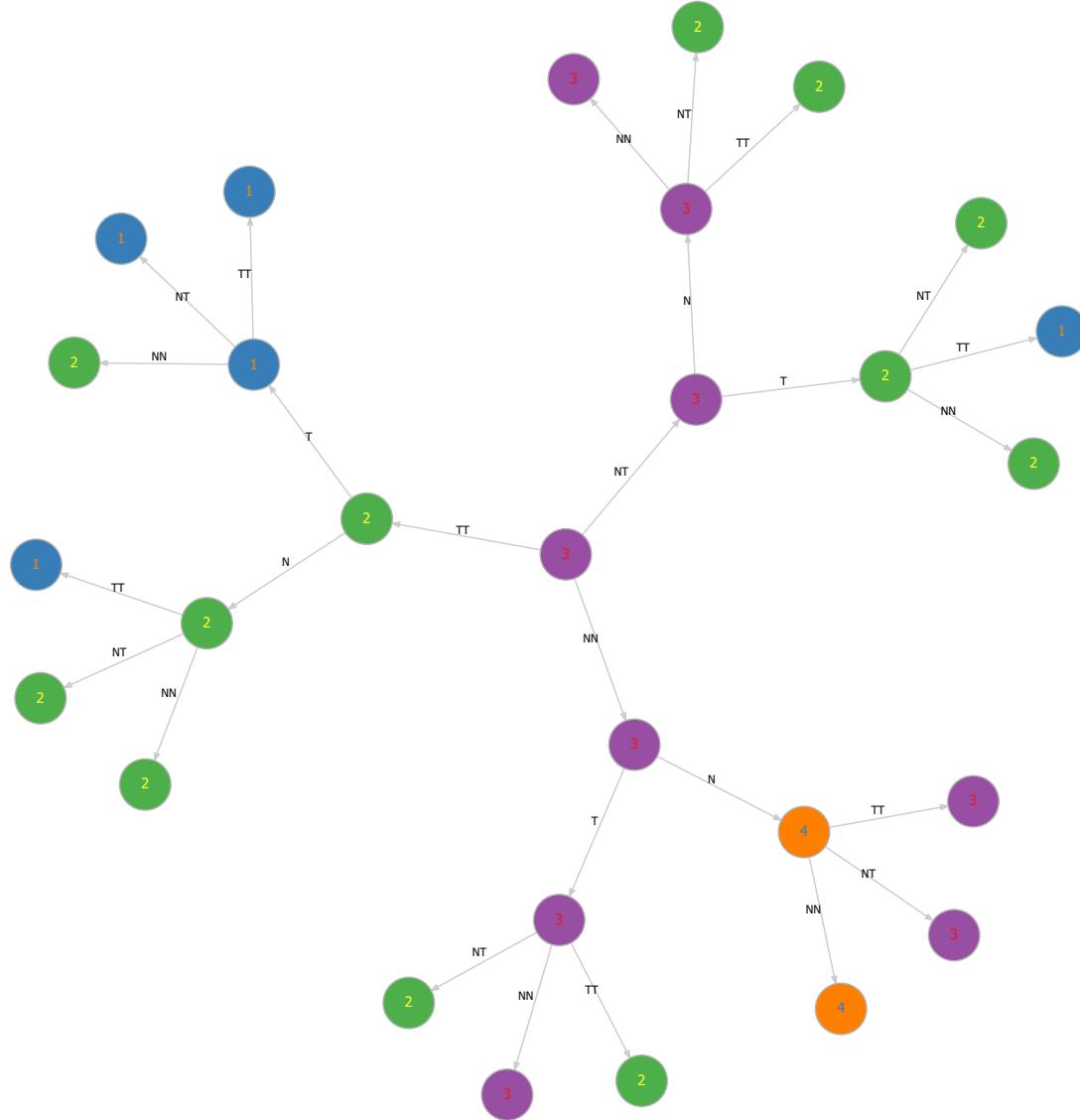
We will recreate the DTPs presented earlier in this section except we will now use varying cohort sizes. These DTPs will use the same previously specified outcomes of 2NNN 3NNT 3NNT. DTPs will be calculated assuming the next three cohorts (cohorts 4,5 and 6) will only be able to recruit 2, 1 and 2 patients respectively. Table 4.7 lists the different pathways and they are also visualised in Figure 4.6.

TABLE 4.7: DTPs for three additional cohorts with varying cohort sizes after observing outcomes for the first three cohorts.

Pathway	Cohort 4		Cohort 5		Cohort 6		Cohort 7
	Dose	Outcomes	Dose	Outcomes	Dose	Outcomes	Dose
1	3	NN	3	N	4	NN	4
2	3	NN	3	N	4	NN	4
3	3	NN	3	N	4	NT	3
4	3	NN	3	N	4	NT	3
5	3	NN	3	N	4	TT	3
6	3	NN	3	N	4	TT	3
7	3	NN	3	T	3	NN	3
8	3	NN	3	T	3	NN	3
9	3	NN	3	T	3	NT	2
10	3	NN	3	T	3	NT	2
11	3	NN	3	T	3	TT	2
12	3	NN	3	T	3	TT	2
13	3	NT	3	N	3	NN	3
14	3	NT	3	N	3	NN	3
15	3	NT	3	N	3	NT	2
16	3	NT	3	N	3	NT	2
17	3	NT	3	N	3	TT	2
18	3	NT	3	N	3	TT	2
19	3	NT	3	T	2	NN	2
20	3	NT	3	T	2	NN	2
21	3	NT	3	T	2	NT	2
22	3	NT	3	T	2	NT	2
23	3	NT	3	T	2	TT	1
24	3	NT	3	T	2	TT	1
25	3	TT	2	N	2	NN	2
26	3	TT	2	N	2	NN	2
27	3	TT	2	N	2	NT	2
28	3	TT	2	N	2	NT	2
29	3	TT	2	N	2	TT	1
30	3	TT	2	N	2	TT	1
31	3	TT	2	T	1	NN	2
32	3	TT	2	T	1	NN	2
33	3	TT	2	T	1	NT	1
34	3	TT	2	T	1	NT	1
35	3	TT	2	T	1	TT	1
36	3	TT	2	T	1	TT	1

These DTPs can be interpreted in the same way as before but now just correspond to a different number of patients. Where the recommended dose is set at dose-level 3 for cohort 5 we can consider these pathways equivalent to those presented earlier in Table 4.6 for cohort 4. As in both of these scenarios 3 patients would have been treated at dose-level 3. For instance pathways 1-6 and their recommended dose for cohort 6 in Table 4.7 are equivalent to pathways 1-16 and its recommended dose for cohort 5 in Table 4.6. This is because in both instances 3 patients have been treated at the same dose-level and all

FIGURE 4.6: Node plot of three additional cohorts with varying cohort sizes after observing outcomes for the first three cohorts.



have the same outcome. Of course, this does not always hold as a dose decision made on less data may lead to a different dose decision and hence create a different pathway. In the scenarios where two patients experience a DLT the model de-escalates and since there is not a third patient at that dose-level 3 the following pathways will all differ from those presented previously.

We have established how DTPs can be used during the design and calibration of a trial but also whilst it is running. They have the ability to communicate dose decisions effectively. They also help alleviate some of the potential mystery behind model-based designs where clinicians and non-statisticians involved in trials may not appreciate how dose decisions are being made by a model such as the CRM. Additionally, they can also be used to assess any modifications that need to be implemented due to practical or logistical issues. It is clear DTPs are a valuable tool to incorporate in any dose-finding trial and with the escalation package by Brock [21] they are very easy to implement, only requiring a few lines of code.

Yap et al. [31] briefly discuss the idea of implementing DTPs in TITE-CRMs. However, limited advice or guidance was provided regarding the problems statisticians may face when trying to do this and there were no examples to refer to. In the next section, we explore the possibility of extending DTPs to work in TITE-CRMs using an illustrative example. We also discuss potential limitations and issues faced when attempting this.

4.3 TITE-DTPs

One reason why DTPs are effective is due to the simplicity of the outcomes being presented. With a CRM these outcomes are either toxicity (T) or no toxicity (N). Similarly, with a design like Wages and Tait or EffTox, outcomes are either toxicity (T), efficacy (E), both toxicity and efficacy (B) or neither (N). Whilst the number of outcomes contributes to the number of potential pathways, other aspects of the trial also go into determining this such as cohort size and the number of cohorts.

However, when we move to the TITE setting the problem becomes more complex. TITE methodology works by using the idea of a partial tolerance

event. At the time of analysis for a dose decision, patients without a toxicity event who have not completed their evaluation period can be considered as having a partial tolerance. These patients can then be weighted according to how much of the evaluation period they have completed and be included in the model. If the patient completes the evaluation period without having a toxicity, they have tolerated treatment and are fully weighted. This means they can be analysed and included in the model as is normally done in a standard CRM design. If a patient experiences a toxicity at any time point they are also fully weighted.

With TITE-DTPs we still maintain the issues from normal DTPs. The number of cohorts and the cohort sizes determines the number of pathways that are produced, but now the outcome is also more complex. More specifically, just considering DTPs for a standard CRM design, patients would have either a toxicity or no toxicity whereas now in the TITE setting we also have to account for the time each patient has spent in the trial. This problem could also be extended depending on how much precision is used in the measurement of time.

To explore the problem further we will use an illustrative example. Consider the example trial we presented in Section 4.2.1, except now we include a TITE component with a 35-day evaluation period. The TITE-CRM can use all the same parameters and specifications we chose for the CRM trial. When setting up a time-to-event trial we also need to specify a weight function. This function determines how patients with partial tolerances will be weighted in the TITE-CRM model. For this example, we will use a linear weight function where patients are weighted as a proportion of the time they have completed in the evaluation period. So, a patient who has completed 20 days would have a weight of 0.571 (i.e. $20 \div 35$). The original CRM design used cohorts of three

however, due to the complexity of TITE-DTPs we will begin by looking at cohorts of one and two patients. For similar reasoning, we will also only detail pathways for the first cohort.

4.3.1 Cohort of one patient

First we will just look at a cohort of one patient, i.e the first patient recruited into the trial. There are 36 possible outcomes for this patient. The simplest pathway to consider is what happens when this patient has a toxicity, as the timing of the toxicity does not impact how the patient is weighted. In this scenario, the recommended dose for the next patient according to the TITE-CRM model would be dose-level 1. So, if we see toxicity at any time point we de-escalate.

Now in the scenario where no toxicity is observed, there are 35 different possible outcomes. Either it is day one and the patient has no toxicity, or it is day two, or day three, all the way up to no toxicity at day 35. Here we can fit the TITE-CRM model for each different outcome and see what the model will recommend as the next dose. Where the patient has between 1-19 days of follow-up the model recommends dose-level 4, anything greater than 19 and the model recommends dose-level 5.

After calculating all possible outcomes we can produce a TITE-DTP for this cohort (Table 4.8). As previously specified the first patient starts at dose-level 2, and as there are 36 possible outcomes there are 36 pathways to the next cohort.

We also extend the nomenclature of Brock et al. [68] to express the different outcomes; here the number in brackets represents the amount of follow-up or observation period the patient has completed. So, N(14) indicates at 14 days the patient has had a partial tolerance event and the corresponding recommended dose for that pathway is dose-level 4. To summarise a large group of

these outcomes we may include inequalities in the notation as well; $N(\leq 14)$ would refer to all the outcomes where the patient had 14 days of follow-up or less. Similarly, $N(\geq 14)$ would indicate 14 days or more.

TABLE 4.8: TITE-DTP for a cohort of one.

Pathway	Dose	Cohort 1	Cohort 2
		Outcomes	Dose
1	2	T	1
2	2	N(1)	4
3	2	N(2)	4
4	2	N(3)	4
5	2	N(4)	4
6	2	N(5)	4
7	2	N(6)	4
8	2	N(7)	4
9	2	N(8)	4
10	2	N(9)	4
11	2	N(10)	4
12	2	N(11)	4
13	2	N(12)	4
14	2	N(13)	4
15	2	N(14)	4
16	2	N(15)	4
17	2	N(16)	4
18	2	N(17)	4
19	2	N(18)	4
20	2	N(19)	4
21	2	N(20)	5
22	2	N(21)	5
23	2	N(22)	5
24	2	N(23)	5
25	2	N(24)	5
26	2	N(25)	5
27	2	N(26)	5
28	2	N(27)	5
29	2	N(28)	5
30	2	N(29)	5
31	2	N(30)	5
32	2	N(31)	5
33	2	N(32)	5
34	2	N(33)	5
35	2	N(34)	5
36	2	N	5

DTPs presented earlier in this chapter have been able to convey a lot of information. Previously with 64 pathways, we were able to specify all the possible outcomes and dose recommendations up to the 4th cohort of three patients. Whereas in the TITE setting, we have 36 just for one cohort of one patient. One way to improve on this would be to more succinctly summarise the TITE-DTP by aggregating the pathways which lead to the same recommendations (Table 4.9).

TABLE 4.9: Summary of TITE-DTP for a cohort of one.

No. of Pathways	Cohort 1			Cohort 2
	Dose	Outcome	Follow-up	Dose
1	2	T		1
19			1-19	4
16	2	N	20-35	5

Whilst we can still apply the concept of DTPs to TITE-CRMs we can see even with just one patient and one cohort there are a lot of possible outcomes we have to look at. Also, we have used a small observation period of only 5 weeks. If a larger observation period were to be used the number of pathways would increase exponentially. In the next section, we explore how adding in an extra patient affects these TITE-DTPs.

4.3.2 Cohort of two patients

Now we consider a cohort of two patients who start at dose-level 2. As before we will consider this the first cohort of patients and only calculate pathways for the next cohort. Here there are a greater number of potential outcomes. There are three potential scenarios either both patients could have a toxicity, one of the patients could have a toxicity and one could not and finally both

patients could have no toxicity. Within the two options where patients could potentially have no toxicity, there are multiple outcomes depending on how much follow-up time is observed. The different scenarios and the associated number of outcomes are:

- TT - 1 outcome
- NT - 35 outcomes
- NN - 630 outcomes

So, for a cohort of two patients with a follow-up period of 35 days, there are 666 possible pathways. The simplest of these is if both patients have a toxicity. Here both patients are fully weighted and when put into the model the dose recommendation is dose-level 1.

If one patient has a toxicity and the other one does not there will be 35 different outcomes. One patient has a toxicity and the other has a partial tolerance event on day one, day two, day three, all the way till day 35 where they have fully tolerated the dose (i.e. N(1)T, N(2)T, N(3)T, ..., N(34)T, NT). These outcomes are just an extension of the pathways in Section 4.3.1 for one cohort of one patient, except now when we model these we include an extra patient in the model who experienced a toxicity. Table 4.10 presents the pathways for this scenario. Here we can see, regardless of how much follow-up time the patient with no toxicity has, the model will always recommend de-escalating.

TABLE 4.10: TITE-DTP for a cohort of two for scenario 2NT.

Pathway	Cohort 1		Cohort 2
	Dose	Outcomes	Dose
1	2	N(1)T	1
2	2	N(2)T	1
3	2	N(3)T	1
4	2	N(4)T	1
5	2	N(5)T	1
6	2	N(6)T	1
7	2	N(7)T	1
8	2	N(8)T	1
9	2	N(9)T	1
10	2	N(10)T	1
11	2	N(11)T	1
12	2	N(12)T	1
13	2	N(13)T	1
14	2	N(14)T	1
15	2	N(15)T	1
16	2	N(16)T	1
17	2	N(17)T	1
18	2	N(18)T	1
19	2	N(19)T	1
20	2	N(20)T	1
21	2	N(21)T	1
22	2	N(22)T	1
23	2	N(23)T	1
24	2	N(24)T	1
25	2	N(25)T	1
26	2	N(26)T	1
27	2	N(27)T	1
28	2	N(28)T	1
29	2	N(29)T	1
30	2	N(30)T	1
31	2	N(31)T	1
32	2	N(32)T	1
33	2	N(33)T	1
34	2	N(34)T	1
35	2	NT	1

The most complicated scenario is when both patients have no toxicity. The number of outcomes for this scenario can be calculated using the combinations

with replacement formula where n represents the number of follow-up days and r represents the number of patients:

$$\mathbb{C}(n + r - 1, r) = \frac{(n + r - 1)!}{r!(n - 1)!} \quad (4.1)$$

Here we have to consider every combination of follow-up days both patients could have completed. We only consider unique combinations of days for example, N(21)N(34) would indicate one patient had been observed for 21 days and the other for 34 days, this would be the same as N(34)N(21) and so would only require one pathway.

For our example with two patients and an observation window of 35 days, we get 630 different combinations, hence the 630 pathways. Trying to show all these pathways in a table as we did before would be infeasible and hard to interpret so instead we just present the aggregate dose recommendations in Table 4.11.

TABLE 4.11: Summary of pathways for a cohort of two for scenario 2NN.

Recommend Dose	No. of Pathways	Combined no. of Follow-up Days	
		Minimum	Maximum
4	102	2	21
5	528	20	70

We can see out of the 630 pathways, 102 recommend dose 4 for the next cohort and 528 recommend dose 5. Dose-level 4 is recommended when the combined follow-up between patients is between 2 and 21 days and dose-level 5 is recommended when the combined number of follow-up days is between 20 and 70. This presents another challenge with TITE-DTPs, as there is some overlap in dose recommendations depending on how much combined follow-up patients have. So, if the combined follow-up between the two patients in

the cohort is 20 or 21 days they could potentially be allocated to either dose-level 4 or 5 and the way that decision is made is dependent on the split in follow-up between the two patients. This problem is also visualised in Figure 4.7. The red lines indicate the combined follow-up time between both patients of 19 and 22 days respectively. Anything greater than 22 days combined follow-up and the model recommends dose-level 5, anything less than 19 and the recommendation is dose-level 4. In between those two time points there is some discrepancy with the model selecting to escalate higher, to dose-level 5, with less data i.e. combined follow-up time of 20 days. Table 4.12 provides a breakdown of these specific combinations.

FIGURE 4.7: Plot illustrating combined-follow up and overlap of dose recommendations for a cohort of two patients for scenario NN.

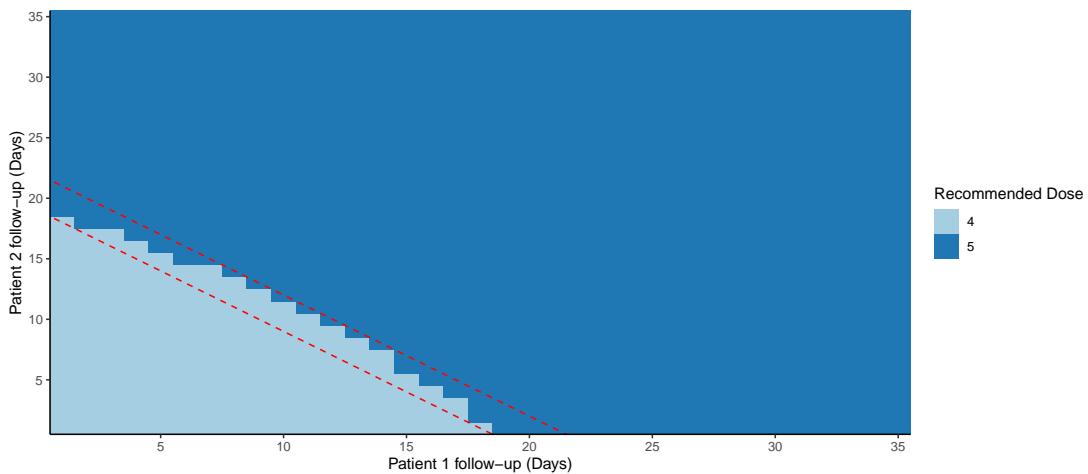


TABLE 4.12: Different dose recommendations with overlapping combined follow-up times.

Follow-up			Posterior Estimates		
Patient 1	Patient 2	Combined	Dose Recommendation	β	Variance
1	20	21	5	0.1606	1.2024
2	19	21	5	0.1579	1.2063
3	18	21	5	0.1555	1.2097
4	17	21	5	0.1534	1.2127
5	16	21	5	0.1516	1.2153
6	15	21	5	0.1501	1.2174
7	14	21	4	0.1489	1.2191
8	13	21	4	0.1480	1.2204
9	12	21	4	0.1474	1.2212
10	11	21	4	0.1471	1.2216
1	19	20	5	0.1520	1.2110
2	18	20	5	0.1495	1.2146
3	17	20	4	0.1473	1.2178
4	16	20	4	0.1453	1.2205
5	15	20	4	0.1437	1.2228
6	14	20	4	0.1424	1.2247
7	13	20	4	0.1414	1.2261
8	12	20	4	0.1406	1.2272
9	11	20	4	0.1402	1.2278
10	10	20	4	0.1400	1.2280

In the case where one patient has 14 days or less of follow-up, the model recommends dose-level 4, similarly, if one patient has at least 18 days of follow up the model recommends dose-level 5. Then when one patient has follow-up times of 15, 16 or 17 the model requires the second patient to have enough

follow-up to make the combined total of 21 days in order to recommend dose-level 5. So, there exists some threshold whereby the model is happy to escalate further if a single patient has enough follow-up (in this case 18 days) and some critical range where the model will only escalate further if a minimum combined follow-up threshold is met (here this is between 15-17 days for one patient with the threshold being 21 days).

This could perhaps be similar to an incoherent CRM design, which escalates after observing a toxicity, except here we escalate after an inadequate amount of follow-up. It should also be noted that in practice rules may be employed to stop the trial skipping untried doses or skipping multiple doses which is what is being recommended in this case. However, that does not mean that this issue would still not occur even when selecting between two more appropriate dose-levels.

This issue was examined further by assessing the posterior estimates of our model parameter β and the variance, from the power model used within our TITE-CRM, for each of these different follow-up combinations. Table 4.12 shows these values. What can be seen is that there is a specific value of β at which point the dose-decision changes from dose-level 4 to 5. The skeleton is used with the power model and the estimate of β to generate posterior probabilities of toxicity for each dose. The dose-level then closest to our target of 25% is then selected as the recommended dose. So, there must exist some value of β at which dose-level 5 now becomes the dose closest to our target. From Table 4.12 we can see that a β value less than or equal to 0.1489 leads to a dose-recommendation of 4 and a value of 0.1495 or higher leads to a dose recommendation of 5. More specifically, we evaluated values between 0 and 1 up to four decimal places to see exactly where this boundary occurs and found that a β value of 0.1492 or lower leads to dose-level 4 and 0.1493 and higher leads to dose-level 5.

In Figure 4.8 for each possible combination of follow-up from 2 to 70 days we plotted the estimated value of β as a single point. Specific combinations of combined follow-up times are represented by only one point on the plot as there is only once combination which leads to that combined follow-up time. For example combined follow-up time of two days only occurs in the scenario where both patients have one day of follow-up. For other combinations there are multiple ways to achieve that so we can see multiple points in these instances. For example, 35 days can be achieved with one patient having 20 and the other 15, or 13 and 22, or 31 and 4 so for these combinations we can see multiple points on the plot. The red line indicates that critical value of 0.1492 and we can see that any combination of follow-up on or before that line recommends dose-level 4 and any above it recommends dose-level 5.

Figure 4.9 focuses specifically on a combined follow-up time of 20 and 21 days and shows how these two combinations of days are the only ones which cross the boundary where the dose decision changes. Each point on the plot is labelled with the individual days of follow-up for both patients. This is a zoomed in snapshot of Figure 4.8 highlighting the combined follow-up times of 20 and 21 days.

Intuitively, since we are using a linear weight function, we would expect each day of follow-up to be weighted the same across each patient. That is to say if we observe 20 days of follow-up with no toxicity the model should make the same recommendation regardless of if that is 20 patients with only one day of follow-up each or only one patient with 20 days of follow-up. Clearly, that is not the case here. When we look back at dose transition pathways for a cohort of one (Table 4.8) for N(20) and N(21) the model recommends dose-level 5. So, when two patients have a combined (i.e total) follow-up time of 20 or 21 days the dose recommendation could potentially be lower than if just one patient had the same amount of follow-up.

FIGURE 4.8: Plot illustrating how the posterior estimate of β and dose recommendation change based on combined follow-up for two patients.

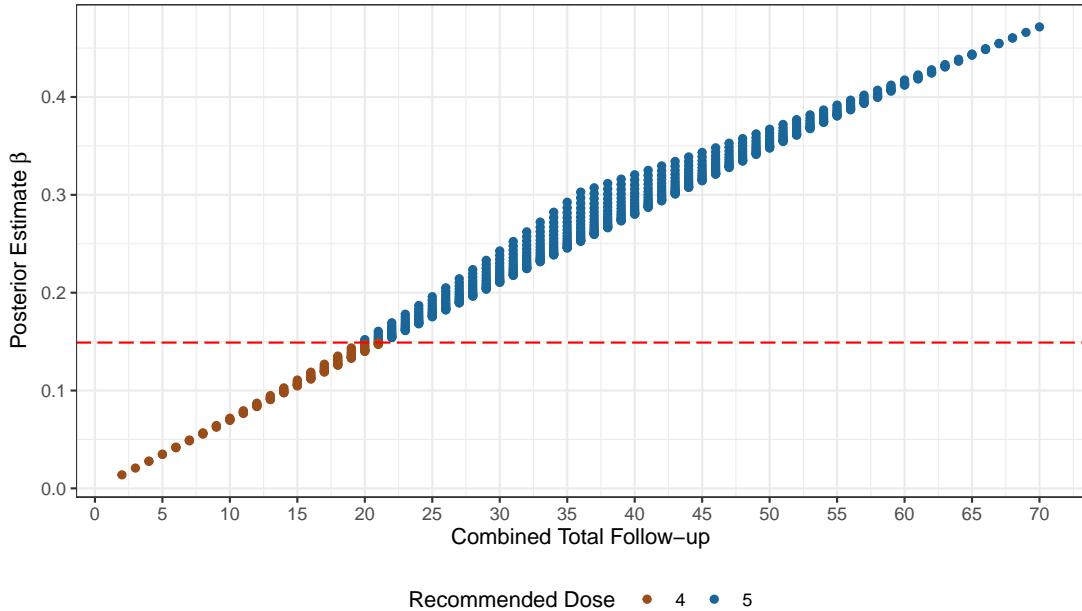
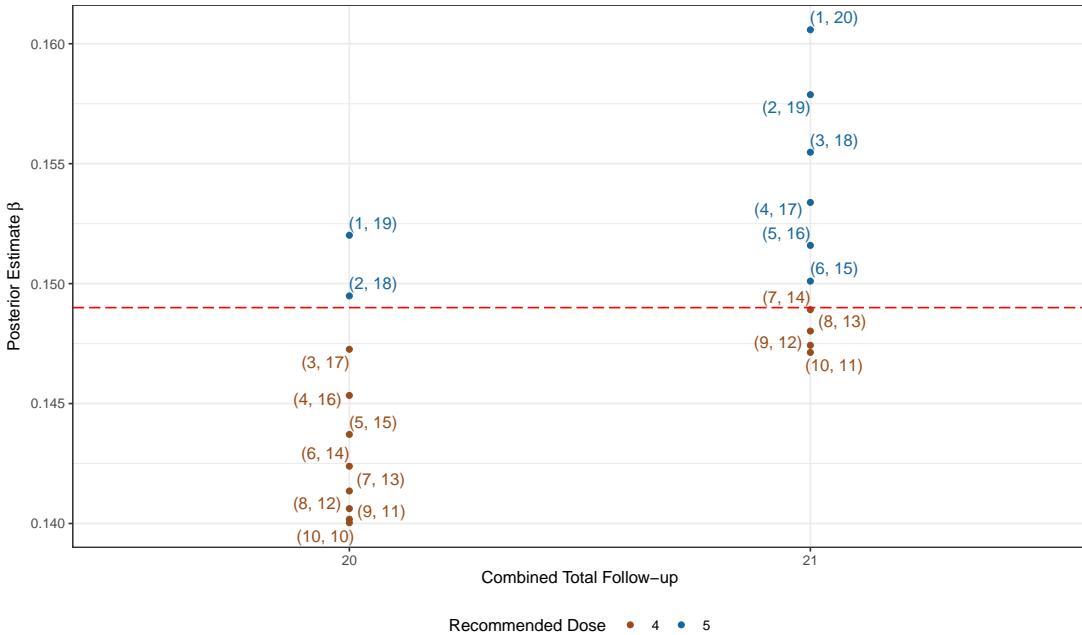


FIGURE 4.9: Plot illustrating how the posterior estimate of β and dose recommendation change based on combined follow-up of 20 and 21 days for two patients.



In order to explore why this discrepancy exists we look into how the TITE-CRM works. For reference the TITE-CRM was originally introduced by Cheung and Chappel [42]. A further detailed description of the TITE-CRM is also

provided by Cheung [81]. We also detail the TITE-CRM design in the presence of partial orders in Chapter 2 (Section 2.2). The TITE-CRM makes use of a weighted likelihood for the model parameter β , this is given by the equation below:

$$L_n(\beta|w) = \prod_{i=1}^n \{w_i F(x_i, \beta)\}^{Y_i} \{1 - w_i F(x_i, \beta)\}^{1-Y_i} \quad (4.2)$$

where $F(x_i, \beta)$ is our dose toxicity model, Y_i is a toxicity indicator for each patient and w_i is the weight associated for the observation of each patient. When there are no toxicities i.e. $Y_i = 0$ the first part of the likelihood formula ($w_i F(x_i, \beta)^{Y_i}$) is reduced to 1 and we are left with $1 - w_i F(x_i, \beta)^{1-Y_i}$. At each dose-level $F(x_i, \beta)$ will remain fixed and is only affected by the weight w_i . This value is different for each patient depending on how long they have been observed in the trial. Then the likelihood is calculated by multiplying these terms for each patient together. This leads us to the root of the issue. As each w_i is independent for each patient and each of these terms is then multiplied together there is no linear relationship between the combined follow-up times for patients at the same dose-level. So, we will get different likelihood estimates which leads to different estimates of the β parameter which ultimately leads to different dose decisions as illustrated in our example.

This was shown in Figure 4.8, where all combinations of follow-up times yielded different values of β . For example, a combined total follow-up of 20 days where one patient has 2 days and the other has 18 equates to weights of $\frac{2}{35}$ and $\frac{18}{35}$ for each patient respectively (this is based on our linear weight function and observation window of 35 days). For the same combined total follow-up of 20 days but split where one patient has 3 days and the other has 17 leads to weights of $\frac{3}{35}$ and $\frac{17}{35}$. These two different combinations of weights when entered into the likelihood formula would yield different results. The

plot also shows this occurs at every combination. However, in most instances the different values of β for the same overall combined follow-up time would either be all above or below the critical value we discovered so as such would not change the dose decision.

When using a linear weight function we cannot say that each day in each patient at the same dose-level is worth the same amount. Essentially, the sum of follow-up time between patients at the same dose-level cannot be thought of as the same. 10 days of follow-up in one patient and 10 days in another is not the same as 20 days in one patient or 19 days in one patient and one in another. This is mainly due to how the likelihood is calculated. We can still say that weighting is linear in individual patients, due to the linear weight function we use, this just does not apply across patients.

So far we have split up the presentation of the pathways dependent on the scenario as there are too many to tabulate and present at once. Table 4.13 attempts to summarise all the different pathways for a cohort of two patients. However, due to the issue with the NN scenario, it is difficult to adequately summarise the combined follow-up required for the different dose decisions. Here we have opted to show the exact minimum and maximum combined follow-up that leads to different dose-recommendations. We then add in extra rows to provide a specific breakdown of how many days individual patients need.

From these values you can still determine the minimum and maximum that guarantee a specific dose as well. For a combined follow-up of 20 days where one patient has 17 days or less of follow-up the recommendation is dose-level 4 and for the same combination but with one patient with 18 days or more follow-up the recommendation is dose-level 5. This may be confusing at first as if you assume a combined follow-up of 20 days with one patient only having two days you would say that is less than 17 so the dose-recommendation is 4.

However, if one patient has two days and the combination is 20 that implies the other patient has 18 days of follow-up so we should use the recommendation from the $N(\geq 18)$ row which is dose-level 5.

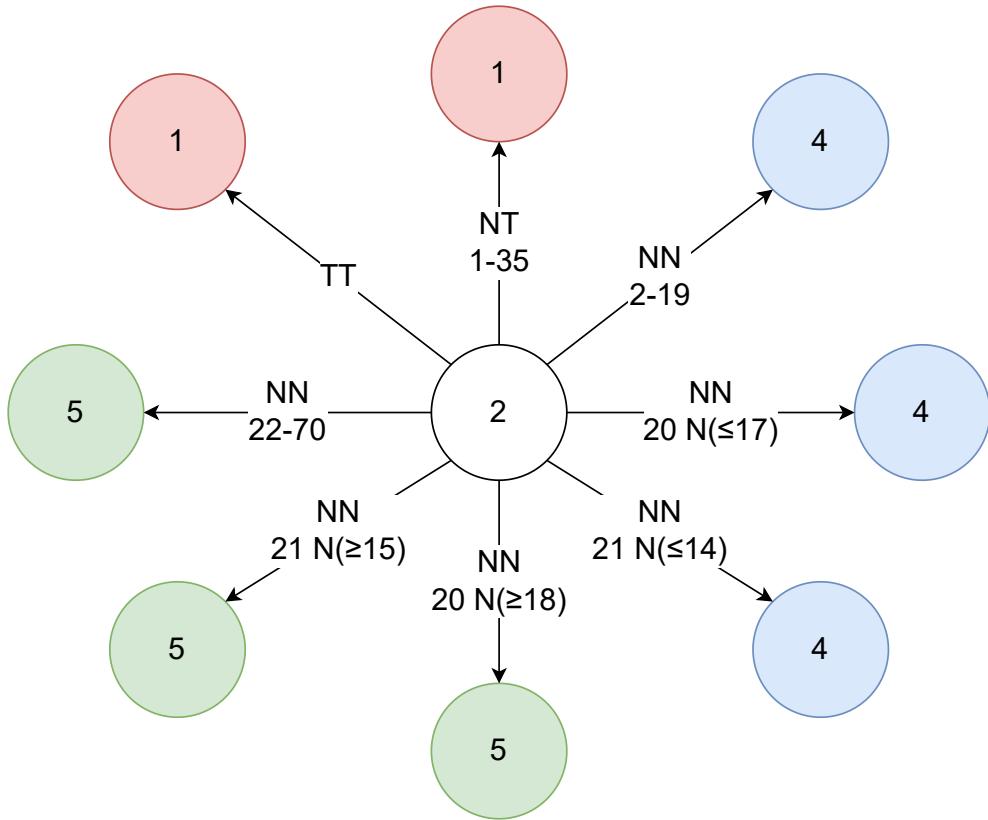
This table could be simplified just by including the overlap in the follow-up column but this would lose out on the granular detail. So, for the outcome NN we could say a follow-up of 2-21 leads to dose-level 4 and a follow-up of 20-70 leads to dose-level 5. An asterisk or some text could accompany the table to perhaps explain the overlap or add more details.

TABLE 4.13: Summary of TITE-DTP for a cohort of two.

No. of Pathways	Cohort 1		Cohort 2	
	Dose	Outcome	Follow-up	Dose
1	2	TT		1
35	2	NT	1-35	1
90			2-19	4
8			20 $N(\leq 17)$	4
4	2	NN	21 $N(\leq 14)$	4
2			20 $N(\geq 18)$	5
6			21 $N(\geq 15)$	5
520			22-70	5

As with classic DTPs we can also visually show these pathways. Figure 4.10 shows a summary of the pathways as a node plot. Rather than present each possible pathway as we do with normal DTPs here we just present the same summary that is shown in Table 4.13. Inside each node is the dose-level. The arrows show each possible outcome and its associated dose-decision. Each arrow is labelled with the specific outcome and the combined amount of follow-up time which leads to the specific recommendation.

FIGURE 4.10: TITE-DTP Node plot for a cohort of two patients.



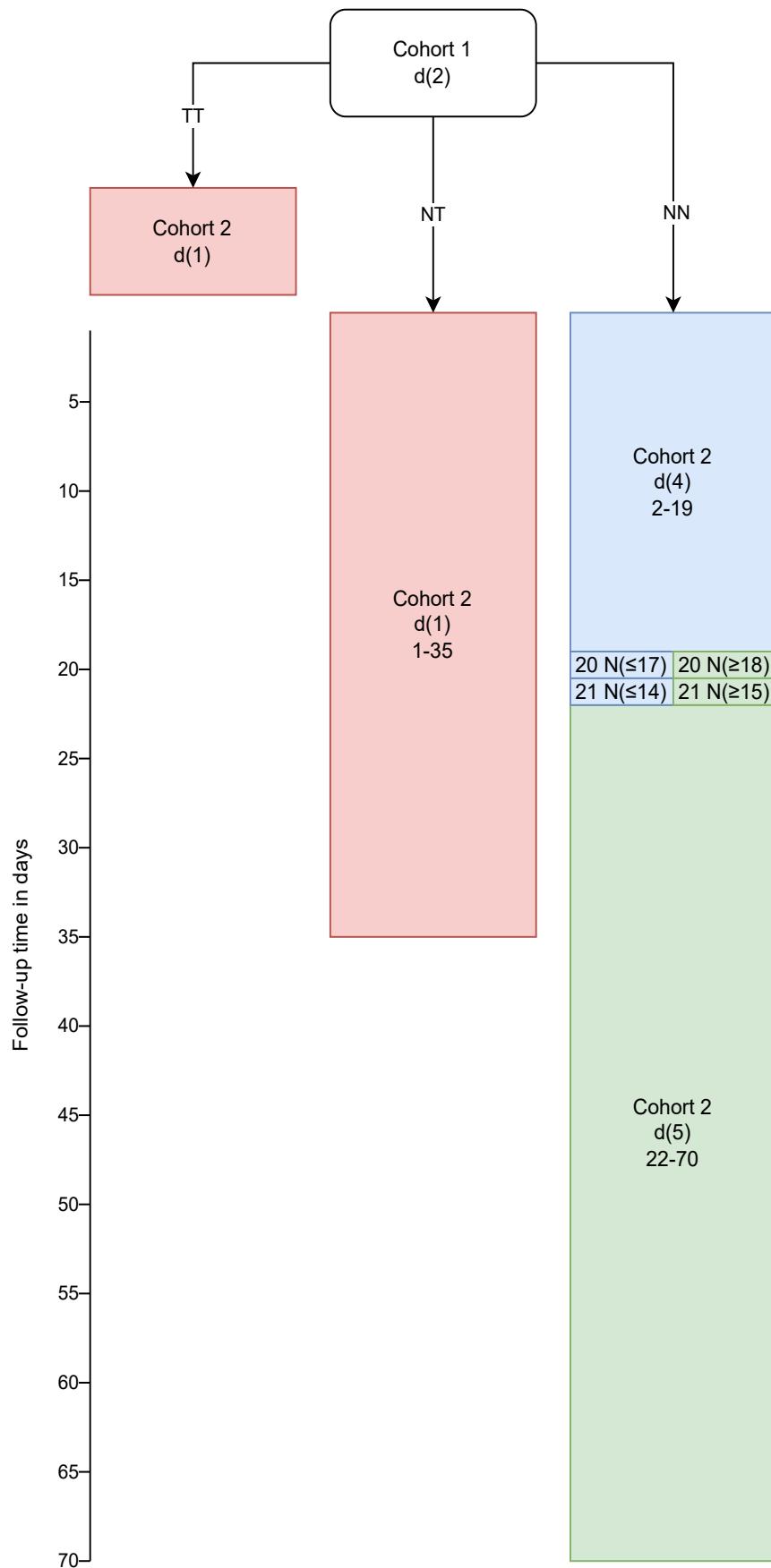
An alternative presentation of the pathways is provided in Figure 4.11. This plot shows a summary of the pathways in a similar manner to the flow plots of DTPs. It shows how the dose decision changes with the combined amount of follow-up time. Each box is labelled with the specific cohort it is referring to. The number in brackets corresponds to the dose-level and the combined follow-up times required to reach that decision are also listed below the dose-level. Additionally, each box is coloured based on the dose-level that is being recommended for the subsequent cohort. In the case where the boxes are too small we have just contained some relevant details about the specific outcome that leads to that recommendation.

For this plot it is important to note that follow-up time is only relevant to outcomes of no toxicity (N). So, for TT the outcome is not dependent on follow-up time. However, for NT the outcomes could change based on the amount of

follow-up time the patient who experienced no toxicity (N) has. This is why TT, NT and NN have different sized boxes as the total combined follow-up for each is different. Neither of these plots capture how many pathways these outcomes represent but this information could easily be added as required. This should be done with caution as we said before this could be slightly misleading as the number of pathways does not indicate that one outcome is more likely than another.

Just by adding an extra patient to a cohort of one the number of pathways we have has increased almost 20-fold. We have also discovered when looking at specific combinations of partial tolerance events there are some inconsistencies with the way the TITE-CRM is recommending dose-levels. Finally, for completeness, we attempt producing TITE-DTPs for a cohort of 3 patients.

FIGURE 4.11: TITE-DTP Flow plot for a cohort of two patients.



4.3.3 Cohort of three patients

Consider instead we have a cohort of 3 patients starting at dose-level 2. Here we will explore all the possible pathways for the first cohort. With 3 patients there are four possible scenarios, the number of possible outcomes relating to each scenario is listed below:

- TTT - 1 outcome
- NTT - 35 outcomes
- NNT - 630 outcomes
- NNN - 7770 outcomes

In total for just the first cohort of three patients, there are 8436 pathways. The first three scenarios listed here are just extensions of what has been previously presented. When all 3 patients have a toxicity, these can just be entered into the model as fully weighted patients and the model recommends de-escalating to dose-level 1.

The NTT scenario is the same as the cohorts of two NT scenario except now there is an extra patient in the cohort who also has a toxicity. Table 4.14 shows the pathways for this scenario. Regardless of the number of follow-up days the patient with no toxicity has the model will always recommend dose-level 1 if the other two patients in the cohort have toxicities.

TABLE 4.14: TITE-DTP for a cohort of three for scenario 2NTT.

Pathway	Cohort 1		Cohort 2
	Dose	Outcomes	Dose
1	2	N(1)TT	1
2	2	N(2)TT	1
3	2	N(3)TT	1
4	2	N(4)TT	1
5	2	N(5)TT	1
6	2	N(6)TT	1
7	2	N(7)TT	1
8	2	N(8)TT	1
9	2	N(9)TT	1
10	2	N(10)TT	1
11	2	N(11)TT	1
12	2	N(12)TT	1
13	2	N(13)TT	1
14	2	N(14)TT	1
15	2	N(15)TT	1
16	2	N(16)TT	1
17	2	N(17)TT	1
18	2	N(18)TT	1
19	2	N(19)TT	1
20	2	N(20)TT	1
21	2	N(21)TT	1
22	2	N(22)TT	1
23	2	N(23)TT	1
24	2	N(24)TT	1
25	2	N(25)TT	1
26	2	N(26)TT	1
27	2	N(27)TT	1
28	2	N(28)TT	1
29	2	N(29)TT	1
30	2	N(30)TT	1
31	2	N(31)TT	1
32	2	N(32)TT	1
33	2	N(33)TT	1
34	2	N(34)TT	1
35	2	NTT	1

Similarly, the NNT scenario is an extension of the NN scenario for a cohort of two patients. We add an extra patient who experiences a toxicity and fit

the same models and observe the outcomes. Table 4.15 shows a summary of the 630 possible pathways. We can see that the majority of the time the model recommends de-escalating. However, there are six pathways where the model recommends staying at the same dose, dose-level 2. This is when the combined follow-up time from the two patients who do not have a toxicity is between 67 and 70 days. We also no longer have that inconsistency issue that we saw before where different dose decisions were being made on the same amount of follow-up time dependent on the split of days between patients. Here the TITE DTP is more clear and a combined follow-up time of 66 days or less leads to de-escalation otherwise the next dose should be recruited at the same dose-level.

TABLE 4.15: Summary of pathways for a cohort of three for scenario 2NNT.

Recommend Dose	No. of Pathways	Combined no. of Follow-up Days	
		Minimum	Maximum
1	624	2	66
2	6	67	70

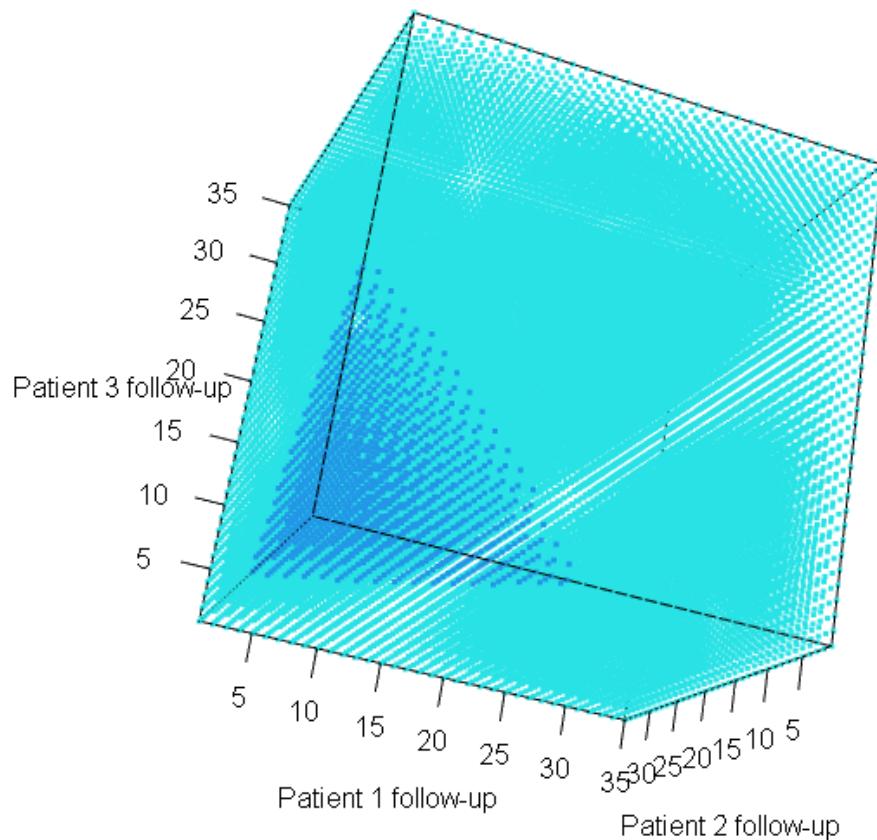
When an additional patient is added to the cohort, the most complicated scenario is when all patients in the cohort experience no toxicity. With 3 patients and 35 days follow-up, 7770 different possible combinations of follow-up days can be observed. The results from fitting all these models are presented in Table 4.16. In this scenario, if the combined number of follow-up between the three patients is between 3 and 23 days the model will recommend dose-level 4 and if it is above 22 dose-level 5 will be recommended. Again we see the same problem as before with a cohort of two patients for the NN scenario. There appears to be some overlap in dose decisions for certain combinations of follow-up days between the three patients. Anything between 20 and 21 days

may result in either a recommendation of dose-level 4 or 5 depending on the split of follow-up time between the three patients. Figure 4.12 also provides a 3D illustration of these pathways with each dot representing a different decision, and the colour corresponding to the dose recommended. A dark blue dot indicates dose-level 4 and light blue is dose-level 5. This is just for illustration purposes and due to its complexity it would not be suitable way to communicate these results.

TABLE 4.16: Summary of pathways for a cohort of three for scenario 2NNN.

Recommend Dose	No. of Pathways	Combined no. of Follow-up Days	
		Minimum	Maximum
4	265	3	21
5	7505	20	105

FIGURE 4.12: Dose recommendations for a cohort of three patients for scenario 2NNN.



In this example, there are 70 different pathways where the combined follow-up between the three patients is 20-21 days. By examining these pathways specifically we can define the different thresholds that are needed to make different decisions. Out of the 70 pathways dose-level 5 is recommended in only 9 of those instances (Table 4.17). Of those only one is for a combined total follow-up of 20 days and the other 8 have a combined total follow-up of 21 days. So, if one of the three patients has a minimum of 18 days of follow-up then the model will recommend dose-level 5. This is why the minimum combined days of follow-up is 20 to recommend dose-level 5 in Table 4.16 as one patient will have 18 days and the other two will have one day each. In the case where the combined follow-up is 21 days, the model will only recommend

dose-level 5 if one of the three patients has a minimum of 15 days and other has a minimum of 4.

TABLE 4.17: Follow-up combinations totalling 20 or 21 days leading to dose-level 5.

Follow-up					Posterior Estimates	
Patient 1	Patient 2	Patient 3	Combined	Dose Recommendation	β	Variance
1	1	19	21	5	0.1578	1.2064
1	2	18	21	5	0.1553	1.2100
1	3	17	21	5	0.1531	1.2131
1	4	16	21	5	0.1512	1.2158
1	5	15	21	5	0.1496	1.2181
2	2	17	21	5	0.1530	1.2133
2	3	16	21	5	0.1510	1.2161
2	4	15	21	5	0.1493	1.2185
1	1	18	20	5	0.1494	1.2147

We have not shown all 70 pathways but we observe the β estimate in each of these cases leading to a dose recommendation of 5 is above 0.1493. This is the same threshold we saw in the scenario of 2 patients with NN outcomes. Similarly, all of the combinations with β estimate of 0.1492 or less resulted in a dose-recommendation of dose-level 4. As before we can also visualise this issue with Figures 4.13 and 4.14. These plots show the β estimates for each possible total follow-up combination and more specifically the combinations for a total of 20 and 21 days. Figure 4.14 labels each point to show the individual follow-up of each of the patients. Here we see the same pattern before as with the 2NN scenario. A minimum number of days is needed in a specific patient in order to obtain an estimate of β which is high enough to warrant escalation to the highest dose. We can see when the follow-up time is split more evenly between the patients the estimates of β are lower. This also supports our findings earlier where we surmised that weightings between patients at the same

dose-level are not entirely equivalent due to how the likelihood is calculated.

FIGURE 4.13: Plot illustrating how the posterior estimate of β and dose recommendation change based on combined follow-up for three patients.

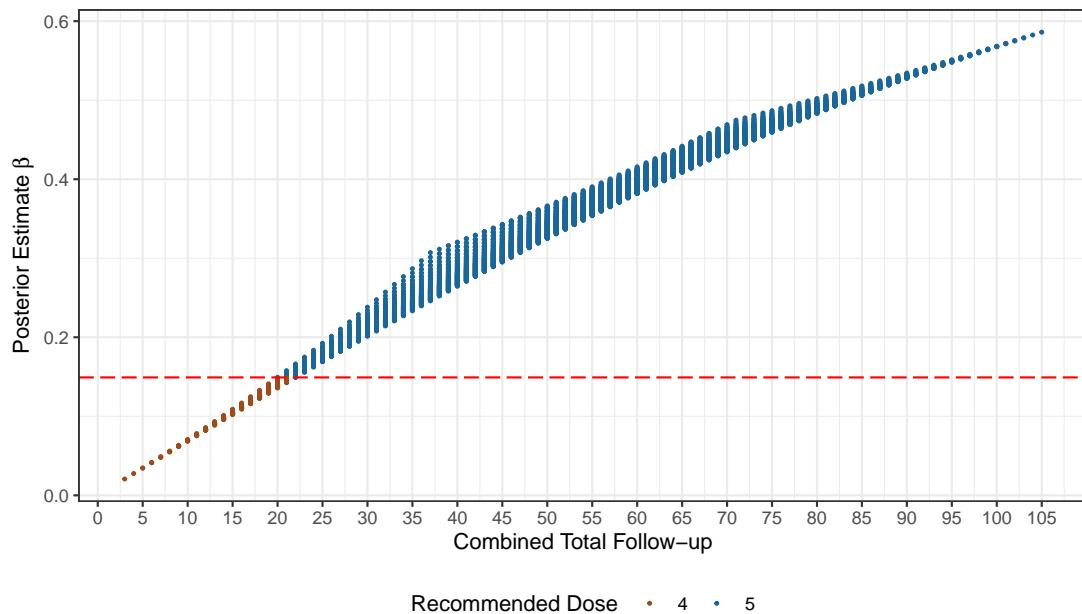
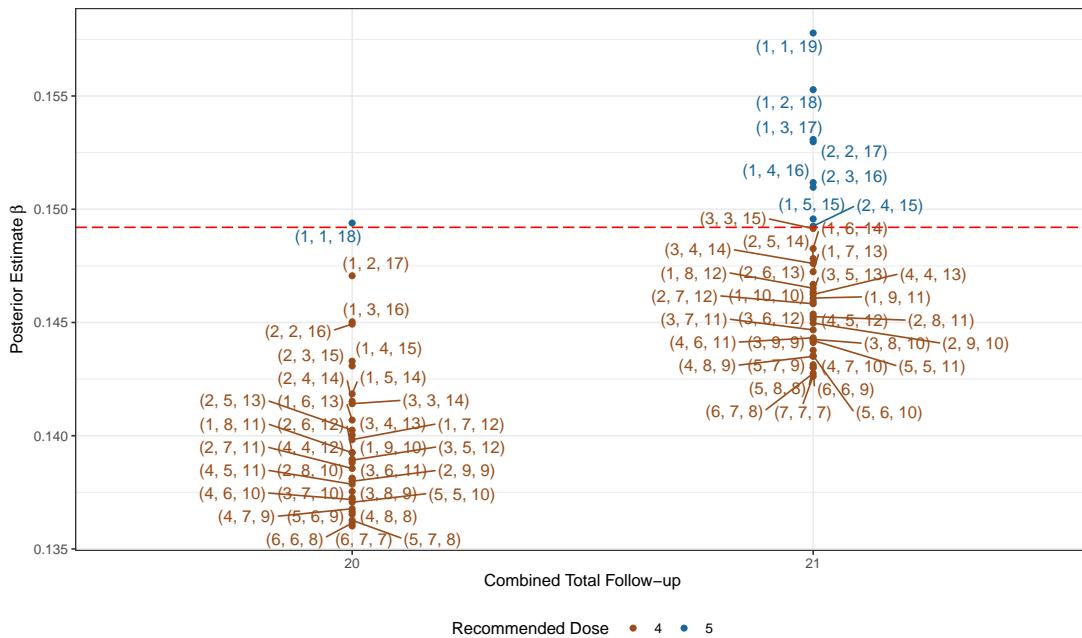


FIGURE 4.14: Plot illustrating how the posterior estimate of β and dose recommendation change based on combined follow-up of 20 and 21 days for three patients.



It is important to note that the threshold of the β estimate which causes the change in dose-recommendation should remain constant throughout the trial. This threshold can be thought of as the the values of β required to make a dose-recommendation picking between doses 4 and 5. This is what we have observed in our two examples of 2NN and 2NNN outcomes respectively. This is due to the way in which we determine the recommended dose for the next patient. The next dose is recommended based on minimising the following equation:

$$|F(d_k, \beta) - \theta| \quad k = i, \dots, 5 \quad (4.3)$$

Here d_k represents each dose-level 1 to 5 and θ is our TD25. The dose with the probability of toxicity closest to our target of 25% will be selected as our next recommended dose. This probability is determined by our estimate of β . What we see in our example is that when β is less than 0.1492 dose-level 4 is the closest in probability to our target level and when it is 0.1493 or higher it is dose-level 5. We did not observe any values of β between 0.1492 and 0.1493 but there must exist some asymptotic value at which the decision changes. This boundary cannot be explicitly defined as its based on the absolute minimum difference which could theoretically be infinitesimally small.

You would also expect there to be similar boundaries/thresholds when choosing between any two other adjoining dose-levels. There should exist a value of β at which dose-level 1 would be recommended over dose-level 2 and vice versa. Similarly, for dose-levels 2 and 3 and then 3 and 4. Whilst we may be able to identify these boundary values of β we cannot know what combination of patients, weights and dose-levels will lead us there. In our example for the cohort of two (2NN) and three (2NNN) in both instances we see that at a combined follow-up of 20 and 21 days our dose-decision changes. However,

if we were to change all of the patients or just one of the patients dose-levels the combined follow-up of where the decision changes may be different. This goes back to equation 4.2. At a different dose-level the value of $F(x_i, \beta)$ will differ leading to a different likelihood and estimate of the β parameter. At lower dose-levels we would expect the combination of follow-up days to recommend the same dose to be higher and lower for higher dose-levels. So the amount of follow-up time required to recommend dose-level 5 would increase as you decrease dose-level.

Table 4.18 combines all the different scenarios and creates a summary table of the TITE-DTPs for a cohort of three. As before we have used a similar notation to express each possible pathway. There is some additional complexity due to there being three patients instead of two and this can be seen in the pathways for the NNN outcome where the combined follow-up time is 21 days.

These can be interpreted as follows. If there are no toxicities in the three patients and their combined follow-up time is 21 days the recommended dose will be dose level-4, if one of those patients has a follow-up time of less than or equal to 14 days ($21 \cap (\leq 14)$), or if one of the patients has a follow-up time of less than or equal to 15 days and another patient has less than or equal to 3 days ($21 \cap (\leq 15) \cap (\leq 3)$). This constitutes 29 different pathways. The scenario with $N(\leq 15)N(\leq 3)$ is included as the cut-off in this scenario is not dependent on a specific amount of follow-up in one patient but in two. In the scenario where patients have 15, 3 and 3 days of follow-up respectively the dose recommendation is 4 but a combination of (15, 4, 2) or (15, 5, 1) leads to a dose recommendation of 5. This can also be seen in the next row of the table where if the combined follow-up is 21 days a dose recommendation of 5 is made if one patient has at least 16 days of follow-up or one patient has at least 15 with another having at least 4. Looking back at Figure 4.14 we can see this exact

change by looking at the points around the line representing the threshold value of β .

Looking at the number of pathways can be quite misleading as from this table it would seem that a large number of them would end up recommending dose-level 5 however, this scenario might not be the most likely depending on what the underlying toxicity is of the dose-level. A higher number of pathways does not correlate to that outcome being more likely it just indicates that it is more complex. Therefore, extra care should be taken when interpreting TITE-DTPs.

Figures 4.15 and 4.16 visualise the TITE-DTP as we did for a cohort of two patients using a node plot and flow plot respectively. Due to the additional patient and the potential for more outcomes we can see these figures are somewhat more complicated.

TABLE 4.18: Summary of TITE-DTP for a cohort of three.

No. of Pathways	Cohort 1			Cohort 2
	Dose	Outcome	Follow-up	Dose
1	2	TTT		1
35	2	NTT	1-35	1
624			2-66	1
6	2	NNT	67-70	2
204			3-19	4
32			20 N(≤ 17)	4
1			20 N(≥ 18)	5
29	2	NNN	21 N(≤ 14) N(≤ 15)N(≤ 3)	4
8			21 N(≥ 16) N(≥ 15)N(≥ 4)	5
7496			22-105	5

FIGURE 4.15: TITE-DTP Node plot for a cohort of two patients.

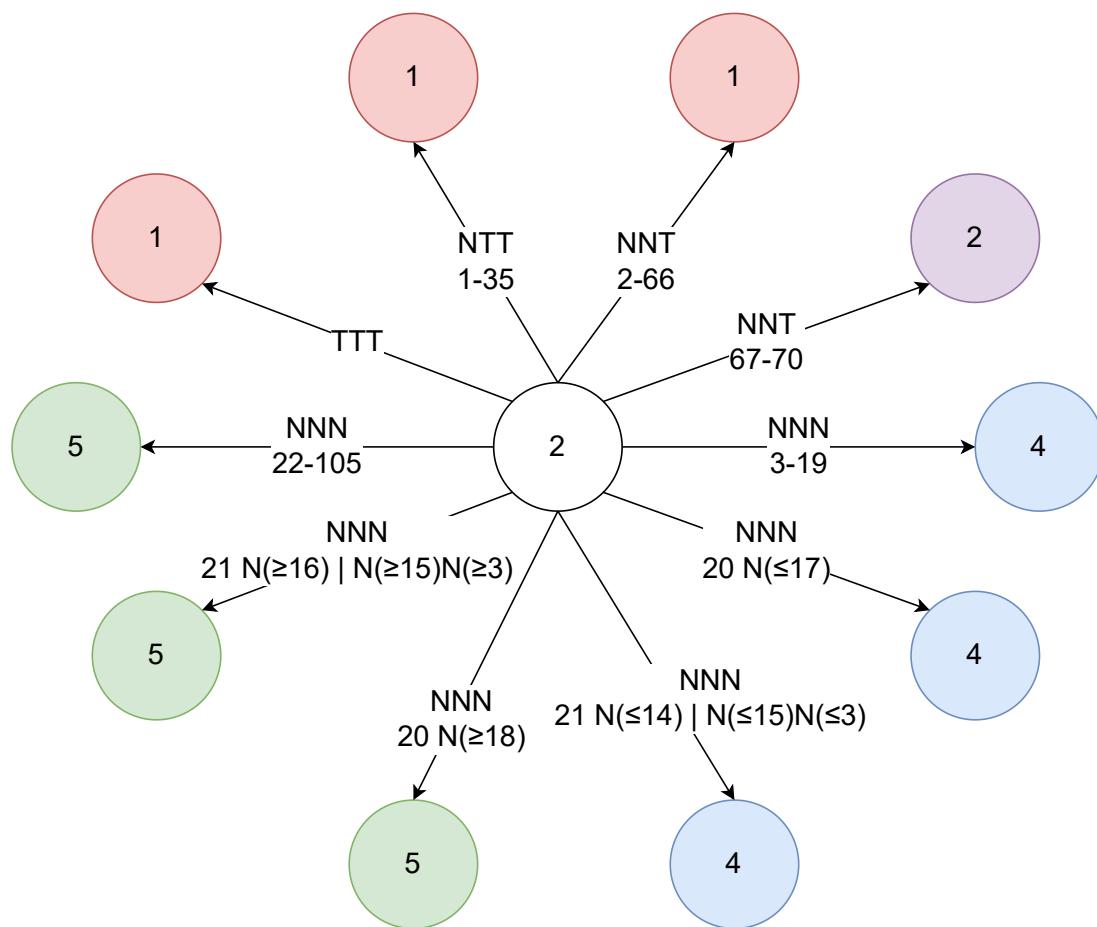
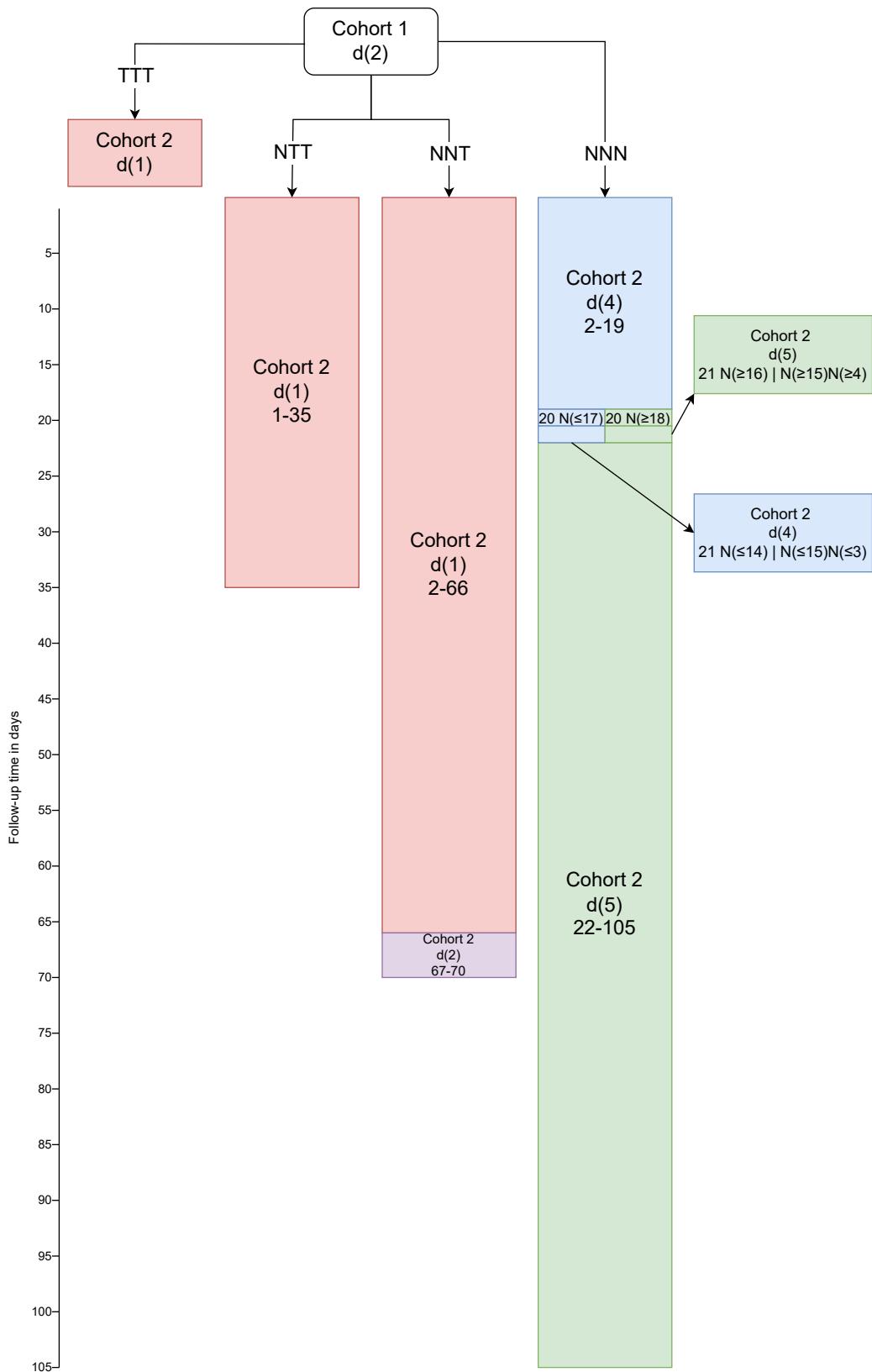


FIGURE 4.16: TITE-DTP Flow plot for a cohort of two patients.



In section 4.2.2, we introduced a simple trial example and produced DTPs (Table 4.3). The pathways for cohort 1 in that table are the full information equivalent of the TITE-DTPs in Table 4.18. As the same trial example was used to produce both of these pathways the only difference is for one of them we have allowed the ability to use partial information in the form of the TITE-CRM and introduce a follow-up period of 35 days. In Table 4.3 pathways 1-16 indicate NNN which is equivalent to Table 4.18 outcome of NNN when the follow-up is max for each patient i.e. 105 days of combined follow-up. In both sets of pathways we can see the recommended dose for cohort 2 is dose-level 5. The outcome where all three patients have a toxicity is exactly the same for both the DTP and TITE-DTP. For the outcome of NTT we can see the recommended dose for cohort 2 is the same in both pathways, this implies allowing for partial information does not change the recommendation for this cohort. When we come to compare NNT we can see that allowing for partial information does have a slightly different impact on dose recommendations. If the cohort is evaluated when there are two partial tolerances with the number of combined follow-up days being 66 or lower the model recommends dose-level 1. Contrast this to when we have full information (each patient has 35 days of follow-up with no toxicity or just the CRM version of the DTP, Table 4.3) and the recommended dose is 2. TITE-DTPs also allow you to compare your design to one with full information i.e the CRM equivalent of a TITE-CRM and allow you to evaluate the length of the follow-up period and see how the dose decisions change as you move through it.

4.4 Alternative weight functions

In this section we explore how TITE-DTPs are impacted by the use of different weight functions. Previously, in Section 4.3, we used a TITE-CRM with a linear

weight function. We will use the same design as specified previously with the only change being the use of different weight functions. Here we investigate two different weight functions which we will refer to as Weight 1 and Weight 2 respectively. The formula for Weight 1 is given in equation 4.4 and the formula for Weight 2 is given in equation 4.5. Here, x represents the number of days patients would have spent in the trial without observing a DLT.

$$f(x) = 1 - e^{-(\frac{x}{7})^2} \quad (4.4)$$

$$f(x) = 1 - e^{-(\frac{x}{22})^6} \quad (4.5)$$

These weight functions represent two different curves. Weight 1 is a curve which increases from 0 to 1 rather quickly. This means a patient would gain more weight in a short amount of time during the start of the observation window. Weight 2 is a curve that increases more slowly over the start of the window and then increases rapidly. This means that patients would have to be observed for a large majority of the window before they are weighted more highly in the model. These two curves are presented in Figure 4.17 along with the linear weight function for reference.

Weight 1 may be used in scenarios where we expect DLTs to occur very early on into a window. So here if patients pass that initial period we may want to weight them more highly in the model. Weight 2 is more conservative to begin with and may be used in scenarios where we want to wait for a large part of the observation window to pass before we start assigning more weight to patients. To give more context Table 4.19 provides a summary of what the specific weights would be for every 5 days without DLT for the three different weight functions.

FIGURE 4.17: Plot of weight values for different weight functions.

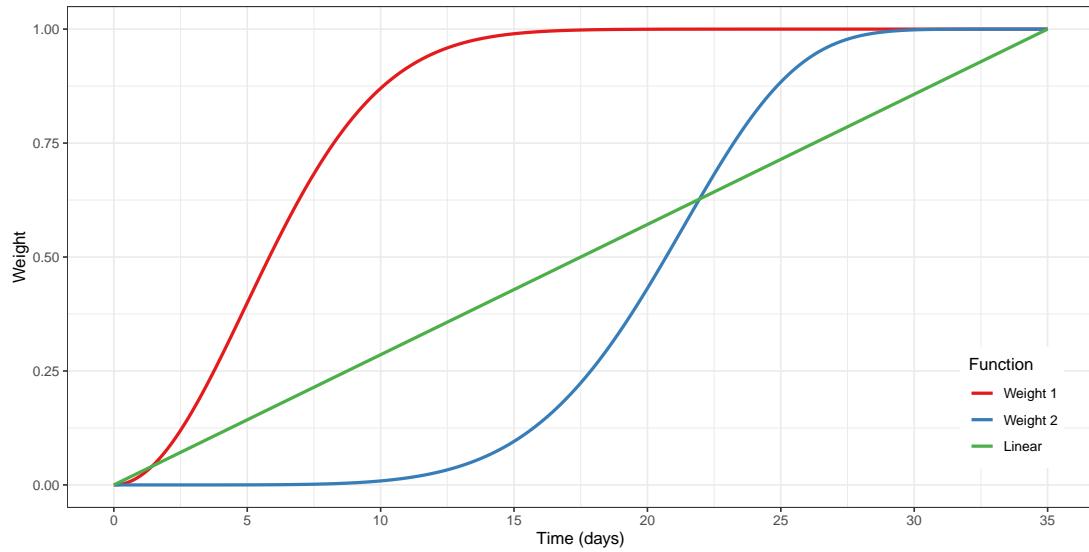


TABLE 4.19: Summary of weight values for different weight functions.

Time (Days)	Weight functions		
	Linear	Weight 1	Weight 2
0	0.00	0.00	0.00
5	0.14	0.40	0.00
10	0.29	0.87	0.01
15	0.43	0.99	0.10
20	0.57	1.00	0.43
25	0.71	1.00	0.88
30	0.86	1.00	1.00
35	1.00	1.00	1.00

To evaluate TITE-DTPs with these new weight functions we will look specifically at a cohort of two patients with 2NN outcomes (two patients at dose-level 2 who don't experience a DLT). In Section 4.3.3, where we explored TITE-DTPs for cohorts of three, we saw this was just a more complicated version

of the cohorts of two TITE-DTPs. For simplicity, we will just investigate the different weight functions for cohorts of two. We will only consider the NN outcomes as the NT and TT outcomes are unaffected by changing the weight function.

In the TT scenario, both patients are given full weighting regardless of how much time they have spent in the trial as they have both experienced a DLT. This would lead to the model recommending to de-escalate to dose-level 1. Neither of the new weight functions would have an impact in this scenario.

The NT scenario, is where one patient has a DLT and the other does not. Here, there are 35 different outcomes, these were all presented in Table 4.10. The results here imply regardless of what weight the patient without the DLT has the model will always recommend dose-level 1. We could apply our new weight functions here but the results would still be the same. It should be noted this only applies to our specific example due to how we have parametrised the TITE-CRM design. There may exist a design which has different priors, target toxicity rates, number of doses etc. which would make a different dose decision based on different weights. However, this is not the case for our design so we will only investigate the NN scenario. Sections 4.4.1 and 4.4.2 investigate this scenario using Weight 1 and Weight 2 respectively.

4.4.1 Weight function 1 in the NN scenario

In the NN scenario there are 630 possible pathways. We consider every unique possible combination of follow-up days both patients could have completed. A summary of the dose recommendations are presented in Table 4.20. Here we can see the majority of pathways recommend dose 5. However, the same issue occurs where there is some overlap with the recommendation when the combined follow-up time is 8 days.

TABLE 4.20: Summary of pathways for scenario 2NN with Weight 1.

Recommend Dose	No. of Pathways	Combined no. of Follow-up Days	
		Minimum	Maximum
4	14	2	8
5	616	8	70

Table 4.21 provides a breakdown of the specific instances where the combined follow-up time is 8 days. We can see in the case where one patient has a follow-up time of 6 days or more the dose recommendation is dose-level 5. As before we also see that these dose decisions match the boundary values we discovered before where all β estimates above 0.1493 lead to dose-level 5 and estimates below 0.1492 lead to dose-level 4.

TABLE 4.21: Follow-up combinations totalling 8 days for scenario NN using Weight 1.

Follow-up			Dose Recommendation	Posterior Estimates	
Patient 1	Patient 2	Combined		Beta	Variance
1	7	8	5	0.1775	1.1845
2	6	8	5	0.1557	1.2092
3	5	8	4	0.1415	1.2255
4	4	8	4	0.1365	1.2311

To illustrate this further we plot the estimated value of β for each possible combination in figure 4.18. Here we see only for the combination of 8 days do the points cross the boundary resulting in the different dose recommendations. Figure 4.19 shows the same but only for combinations of 7, 8 and 9 days.

FIGURE 4.18: Plot illustrating how the posterior estimate of β and dose recommendation change based on combined follow-up for two patients with Weight 1.

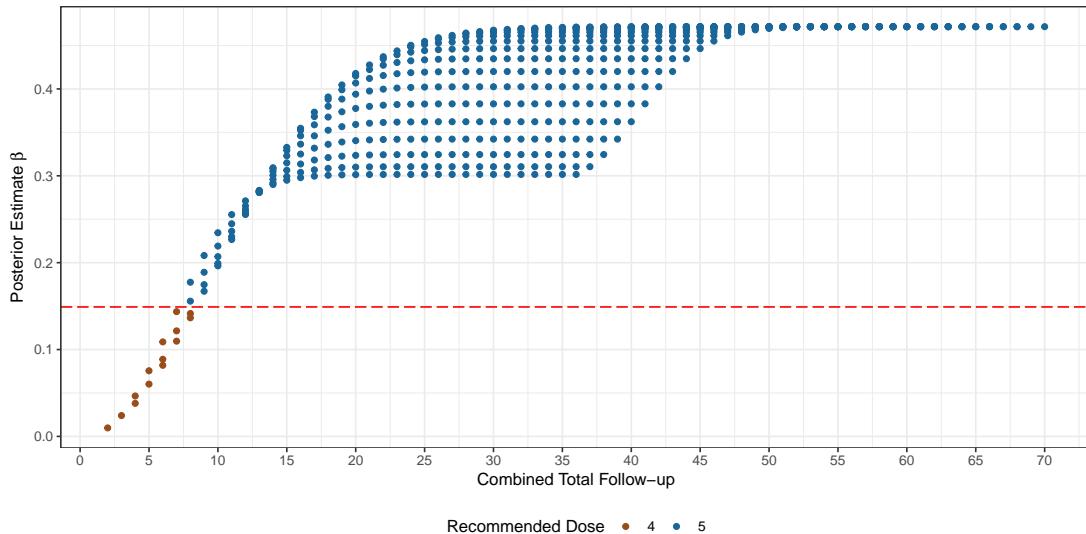
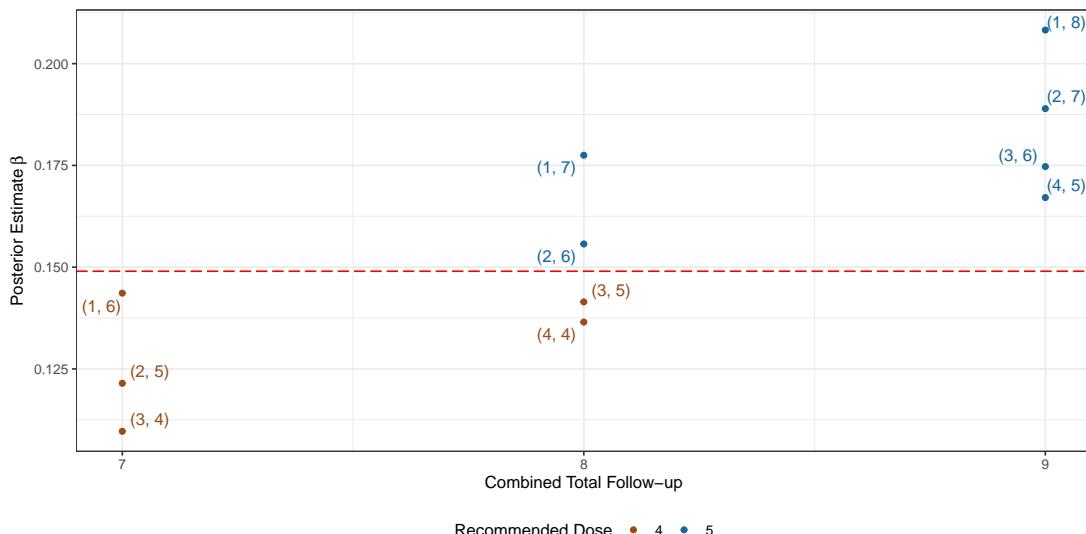


FIGURE 4.19: Plot illustrating how the posterior estimate of β and dose recommendation change based on combined follow-up of 20 and 21 days for two patients with Weight 1.



Compared to the scenario with the linear weight function we still observe that for specific combinations of combined follow-up time the dose recommendation could change based on how much follow-up time an individual patient has. The effect of this specific weight function has made it so we observe this at

a lower amount of combined follow-up time. Using the linear weight function this occurred at 20 and 21 days. This makes sense since with Weight 1 we are allocating more weight to patients earlier on compared to the linear function. The point at which the model recommends dose 5 is now based on a lower amount of follow-up time than under the linear function.

For this scenario under Weight 1 we are still able to construct a TITE-DTP, which is presented in Table 4.22. Now we can clearly see that if the combined follow-up time is greater than 9 the model would always recommend dose 5. Here it is only under the specific scenario where the combined follow-up time is exactly 8 where different doses can be recommended.

TABLE 4.22: Summary of TITE-DTP for scenario NN under Weight 1.

No. of Pathways	Cohort 1			Cohort 2	
	Dose 1	Dose 2	Outcome TT	Follow-up	Dose 1
35		2	NT	1-35	1
12				2-7	4
2		2	NN	8 N(≤ 5)	4
2				8 N(≥ 6)	5
614				9-70	5

4.4.2 Weight function 2 in the NN scenario

Now we will consider the scenario NN using Weight 2. As before there are 630 possible pathways and a summary of the dose recommendations is given in Table 4.23. There are 217 pathways that recommend dose 4 and these have combined follow-up time ranging from 2 to 37. Then there are 413 pathways that recommend dose 5 with combined follow-up times ranging from 23-70.

With this weight function when the combined follow-up time between the two patients is 23 and 37 the dose recommendation will depend on the specific combination that has been observed.

TABLE 4.23: Summary of pathways for scenario 2NN with Weight 2.

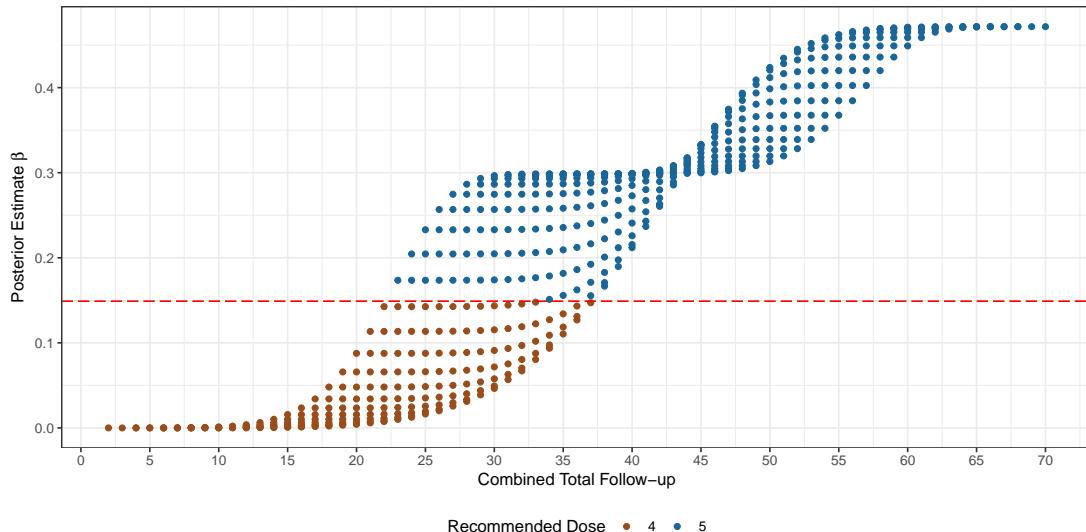
Recommend Dose	No. of Pathways	Combined no. of Follow-up Days	
		Minimum	Maximum
4	217	2	37
5	413	23	70

There are 220 pathways where the combined follow-up time is between 23 and 37 days. Of the 220 pathways 124 lead to recommending dose level 4 and 96 to recommending dose level 5. We investigate the β estimates for all of these combinations and present them in Figure 4.20. The same boundary as before is observed where β estimates above 0.1493 lead to dose-level 5 and estimates below 0.1492 lead to dose-level 4. Compared to the linear weight function this only occurred in 20 pathways where the combined follow-up time was 20 or 21 days.

As this weight function allocates less weight, compared to the linear weight function, to patients earlier on in the trial we see a larger amount of combined follow-up time needed to change the dose recommendation. This is why we see a wide range of combined follow-up times where the dose recommendation could be either 4 or 5. Under this specific weight function there may be one patient who has sufficiently high follow-up time such that the model would recommend the higher dose compared to if two patients had a similar amount of follow-up time but over a shorter time period.

Specifically, we have one pathway where the combined follow-up time is 23 days with one patient having 22 days and the other having just one day

FIGURE 4.20: Plot illustrating how the posterior estimate of β and dose recommendation change based on combined follow-up for two patients with Weight 2.



which leads to a dose recommendation of 5. In this instance the weight for the patient with 22 days of follow-up is 0.63 and the weight for the patient with 1 day is 8.8×10^{-9} . Another pathway is when the combined follow-up time is 37 days with 18 days observed for one patient and 19 days for the other. This corresponds to weights of 0.26 and 0.34 respectively. Even though there are 14 more days of follow-up without DLT the dose recommended is lower. This is not incorrect mathematically and the TITE-CRM model is behaving as expected. This is just a consequence of the weight function we have specified.

With this weight function it is difficult to specify the TITE-DTP with the same amount of granularity as we have done before. This is mainly because we have 220 pathways and there is no way to succinctly summarise them in a meaningful manner. Previously where the dose decision was different for the same amount of combined follow-up time we were able to present two statements to indicate how the recommendation changes based on specific follow-up in a specific patient (See Table 4.22 where 8 N(≤ 5) leads to dose 4 and 8

$N(\geq 6)$ leads to dose 5). With this weight function there are 15 different combinations where this issue occurs and so we would have to specify 30 statements to show specifically when the dose recommendation changes based on follow-up times for one of the patients in the cohort.

Table 4.24 presents the TITE-DTP under this weight function. Here we have had to simplify what is presented when the combined follow-up time can lead to differing recommendations. Overall, we can still see if the combined follow-up time is 22 days or less the recommendations is always dose 4 and likewise if it is 38 days and above the recommended dose is 5.

TABLE 4.24: Summary of TITE-DTP for scenario NN under Weight 2.

No. of Pathways	Cohort 1			Cohort 2
	Dose	Outcome	Follow-up	Dose
1	2	TT		1
35	2	NT	1-35	1
121			2-22	4
220	2	NN	23-37	4/5*
289			38-70	5

* Depends on the specific amount of follow-up time for each patient.

4.5 Discussion

Dose transition pathways as a tool were developed to improve communication and understanding of model-based designs. Often clinicians may not feel comfortable with having a model select doses when compared to the standard approach of a traditional and easy to follow rule-based design [82]. DTPs try to bridge this gap and make these model-based designs more approachable.

They do this by summarising model recommendations based on possible outcomes into simple pathways of dose decisions. They also can be used by the statistician to help calibrate the model and any design specifications. In particular, this helps with implementing stopping rules and investigating how escalation occurs. There is also a potential operational upside where DTPs can aid the running of a trial. By looking ahead there may be instances where regardless of any outcomes observed on the trial the dose-level will not change. In scenarios like this the need for a statistician may be lessened.

DTPs can also be implemented as a visualisation tool to help visualise all the pathways in advance. This may have benefits in terms of raising any imminent safety concerns if certain pathways are followed. They can also be used throughout a trial's life cycle at safety committee meetings to discuss potential doses for future cohorts of patients. They are also very adaptive and are able to handle many challenging circumstances such as a change in cohort size or a patient receiving an incorrect dose. If these circumstances were to occur new DTPs could simply be calculated to account for any trial deviations. Although this is not an exclusive feature of DTPs, they are only capable of handling these scenarios because they can be accounted for in model-based designs like the CRM.

A lot of this chapter focused on providing examples of how DTPs could be implemented specifically for a CRM design. However, the concept can easily be applied to many other model-based dose-finding trial designs such as BOIN and EffTox and even the 3+3. Implementation of these DTPs is relatively simple as well with the escalation package by Brock [21]. It should be noted, as mentioned above, some of the flexibility of DTPs is due to the underlying designs that are used to make them. So, some challenges may be found when producing DTPs for certain types of trial designs.

It is clear that the inclusion and use of DTPs is a net positive for dose-finding trials, not just in their design but also during the running of the trial. This is also reflected in guidance that is being published relating to the conduct and reporting of trials. Statistical Analysis Plan (SAP) guidelines produced by Gamble et al. [83] in 2011 were extended by Homer et al. [84] in 2022 to provide guidance for early phase trials. The authors here advocate for the use of DTPs where appropriate when producing SAPs for dose-finding trials. The DEFINE study [85] aimed to extend the SPIRIT (Standard Protocol Items: Recommendations for Interventional Trials) 2013 statement [86] and the CONSORT (CONsolidated Standards Of Reporting Trials) 2010 statement [87] for use in early phase dose-finding trials. This led to the development of SPIRIT-DEFINE [88] and CONSORT-DEFINE [89]. Whilst SPIRIT-DEFINE explicitly include DTPs in their checklist they are not included in CONSORT-DEFINE. This makes sense as DTPs are more of a tool during the design and conduct of a trial and therefore more relevant to include in protocols. However, in scenarios where a dose-finding trial is stopped early, for issues other than safety, the reporting of these trials could also include DTPs to report what the potential results of the trial may have been.

In the discussion section of the Yap et al. [31] paper which first introduces the idea, there is some mention of applying DTPs to TITE-CRMs. They mention the problems with patients having either partial or full tolerance and how DTPs may differ depending on how much follow-up time they achieve. One recommendation they gave was to produce the CRM equivalent DTPs. This would be useful during design stages to assess whether dose decisions change with full or partial information.

Our work agrees with what Yap et al. [31] originally theorised. Extending DTPs to a TITE-CRM is problematic. Firstly, due to the idea of partial tolerances, trying to account for every possibility and time point a patient has not

had a toxicity is an exponentially increasing problem. The complexity of DTPs is intrinsically linked to patients with partial tolerances as to fully build out the TITE-DTP you need to calculate every possible time point at which that patient could be observed. Then also for cohorts of more than one patient, considering the outcome where both patients experience a partial tolerance exponentially increases the number of pathways. We demonstrated that by showing the different potential DTPs for cohorts of patients from sizes one to two to three and how the number of pathways kept increasing with each iteration.

We also found the TITE-CRM to have some issues when recommending doses. In some instances, different dose decisions were being made based on the split of follow-up time between patients for the same overall combined follow-up time. So, in our examples and TITE-DTPs for a cohort of two or three patients we saw different recommendations when the combined amount of follow-up time between the three patients was 20 and 21 days dependent on how those were split between the patients. We have shown this is dependent on the model parameter and that different combinations of follow-up times produce different estimates of the parameter due to the likelihood function. The model parameter β can take any real value. As such there are specific boundaries or thresholds that exist at which the dose-decision will change dependent on the value of β . However, trying to strictly define these as a specific value is difficult as the way a dose is selected is based on the smallest difference between the estimated probability of toxicity at that dose and the target toxicity level. This difference can be infinitesimally small and as that tends to zero the value of β which leads to that small difference tends to a specific boundary/threshold value. However, we can define a two values for β at each adjoining dose-level to say a value above will result in the higher dose being recommended and a value below the other value will mean the lower dose is recommended.

Understanding this exact boundary is not paramount as in a practical setting due to the observation window or the number of patients you have you may not get close to that specific value of β where the dose decision changes. As with our examples where values of β below 0.1492 resulted in a dose-recommendation of dose-level 4 and any value above 0.1493 resulted in dose-level 5 being recommended. We saw this corresponded with combined follow-up times between 20 and 21 days. Understanding the exact combination of follow-up days for patients that lead to a dose decision is important to help us construct the TITE-DTPs. This is another advantage of DTPs in a TITE setting as they allow us to see exactly what amount of data we need to observe for specific decisions to be made. This may be helpful in practice when it comes to timing analyses for dose-decisions as it may be beneficial to observe patients for a few extra days in order to escalate to a higher doses. Whilst this may be a rare occurrence in an actual trial it will still be beneficial to be aware that this problem could arise and develop a plan ahead of time to deal with it.

It is also important to note that this inconsistency may change how we intuitively interpret patients weighting in a TITE-CRM, specifically with a linear weight function. Through our examples we have shown that follow-up time between patients at the same dose-levels is not equally weighted. We saw that for the same combination of or total follow-up time different dose recommendations were made based on the split of follow-up time between the patients. Even in instances where different dose-recommendations were not made we still saw different estimates of β for different combinations of the same total. Even with the linear weights it is the non-linearity of the weights that causes this problem. An extensive review of the literature has not been conducted however we do not think that this issue with the TITE-CRM has been highlighted previously, this may also extend to other methodology which implements a TITE component.

We also investigated the use of alternative weight functions. Here we still observed under certain combinations of follow-up times between patients different dose recommendations could be made. Compared to a linear weight function having a weight function that gives more weight early resulted in fewer pathways being impacted by this. by attributing more weight to the patients early on they contribute more to the model which may result in a different dose decision. For a weight function which allocates more weight later into the observation window there are a lot more pathways for a wider range of combined follow-up times where different decisions could be made. In our example one pathway escalated to a higher dose even though there were 14 less total days of combined follow-up compared to another pathway. Whilst this is not necessarily incorrect it helps us better understand the impact of weight functions in TITE-CRMs. For example there could be a medical context where it is more meaningful to have a single patient go 4 weeks without a DLT compared to two patients going 2 weeks without one. In this scenario you may want to allocate more weight to the patient at 4 weeks. TITE-DTPs are possible with different weight functions but may be more difficult to construct. They could still be a useful tool in helping understand weight functions and when a specific dose decision would be made for a specific amount of time/weight.

Further work may be done to explore this issue and the idea of TITE-DTPs. Here we have only looked at a relatively simple example using a TITE-CRM with a power model. We could also investigate more weight functions such as the adaptive weight function presented by Cheung and Chappel [42], Huang and Kuan [53], or Braun [90]. Our example design was not a modified TITE-CRM so we did not make use of any stopping rules or rules about skipping doses. Changes to any of the specifications or the inclusion of these rules would alter the results we produced. It is also possible that this issue may

be less apparent in the early stages of a modified TITE-CRM example. If a rule was included to avoid skipping doses the initial set of doses the TITE-CRM could explore would be restricted. However at a certain point once multiple dose-levels had been tested the issue may reappear.

Additionally, the observation window of 35 days that we selected was also fairly arbitrary and as discussed previously any significant increase in this value will drastically increase the number of pathways that need to be calculated. We only looked at patients at the start of the trial all of whom received the same dose-level. An additional approach would be to look at the pathways halfway through the trial assuming previous patients had been fully observed so we would not have to consider multiple pathways from that data. Looking into a more complex example and multiple scenarios to see if TITE-DTPs still behave as we have described would be beneficial to validate the work we have already done as well as highlight any different features that may emerge such as the inconsistency with weighting that we found.

The examples of TITE-DTPs we provided were for only one cohort as well. Obviously, this becomes more and more difficult to deal with as we add in extra patients and cohorts. It is also not as trivial as just presenting a summary table as we did for the first cohort as well. Any additional cohort will have to take into account not only partial tolerance events from the new cohort but would have to consider every remaining possible partial tolerance event from a previous cohort. Consider one pathway from a cohort of three, N(31)N(23)N(9), all patients here experienced a partial tolerance and as their combined follow-up time adds up to 63 days we can see from the TITE-DTP in Table 4.18 the dose recommendation would be dose-level 5. If we were to then recruit cohort 2 to dose-level 5 and attempt to produce more pathways we would need to check combinations for each possible amount of remaining follow-up time for the patients in the previous cohort as well as the full 35 days

for each of the three new patients. So, this can essentially be thought of as a cohort of six, where some patients already have some data available. Equation 4.1 can then be used to give us a rough estimate of how many combinations need to be considered. For a cohort of 6 patients with 35 days of follow-up, the number of combinations is 1×10^{41} . Now since we already have some data a few of these combinations are redundant but that is still an astronomical amount of pathways. For context the universe is approximated to be 13.7 billion years old [91] which roughly converts to 4.3×10^{17} seconds i.e. there are more possible combinations than there are seconds that have passed since the beginning of time.

The TITE-CRM was originally designed to work without the use of cohorts. The idea was that patients could be recruited into the trial continually and they would be allocated a dose-level based on data from patients already in the trial. As those patients already in the trial may not have completed their DLT observation window they would be partially weighted using the idea of them having achieved a partial tolerance. As recruitment to a clinical trial may not be consistent or predictable this made it possible to not pause recruitment in a dose-finding trial which would typically be done whilst a cohort was being observed for DLTs. However, in a practical setting and with guidance from regulators it may not always be possible to run trials in this manner. In order to make a dose-decision data may have to be source verified and cleaned. A report and analysis will have to be produced and used to make a dose-decision which is then discussed with an independent committee. This process may take weeks and recruitment may have to be paused to allow for this to occur before the next patient is recruited. Werkhoven et al. [54] provide a full discussion on the practicalities of running a TITE-CRM and the challenges it raises. However, the TITE-CRM still remains useful as we can have long observation windows to monitor late-onset toxicities and still make dose-decisions whilst

observing patients (Like we have done in Chapter 2 for the ADePT-DDR trial). As such TITE-DTPs can still be a useful tool.

One way in which TITE-DTPs could work with multiple cohorts would be if previous cohorts were completed and had full information. That data could go directly into the model and you would not need to consider any previous partial tolerances. When a dose decision for cohort 2 is made on partial information from cohort 1, TITE-DTPs could be created showing outcomes for cohort 3 that assume cohort 1 has full information. In practice, this may very much be the case as well depending on factors such as recruitment time and the follow-up period.

The way DTPs are calculated the outcomes are specified and then entered into the model and then the recommended dose based on those is extracted. Those values are then used to construct the DTPs. That means for each outcome we have to specify an individual model. So, earlier in the chapter when there were 64 pathways 64 models were fitted and in the example of TITE-DTPs for a cohort of 3 we had 8436. As you increase the cohort size and the number of patients or the number of follow-up days in the trial, the number of pathways increases hence the number of models required to compute the DTP increase and the more models required the more computing time is needed. One way around this may be to stop computing once a dose-decision threshold is reached. This specifically relates to any outcomes where a partial tolerance occurs. In the TITE-DTP example for the NNN outcome, we see anything after 27 days of combined follow-up recommends dose-level 5, We could incorporate a rule into our code that checks after each combination of follow-up days if the recommended dose changes and remains the same across every permutation of follow-up time across the patients DTPs would stop being calculated and you can assume the recommended dose will be the same. That is to say in this scenario for every combination of the three patients' follow-up

time that adds up to 27 the model recommends dose-level 5 so we can assume that for any additional follow-up time the model will make the same recommendation as that is the maximum dose. This could cut down computing time on thousands of additional pathways depending on the scenario and context.

An alternative method of producing TITE-DTPs may be to use patients' weight as a reference instead of their follow-up time. Even though weight is a function of follow-up time it might make presenting the TITE-DTPs simpler. A set of weights could be specified and pathways for those could be calculated instead. So rather than calculating pathways for N(1), N(2), ..., N(34), N you just calculate pathways when a patient is at 25%, 50%, 75% and 100% weighting. The issue with this is in order to make it interpretable you would have to back-transform the weighting. So in our example with a 35 day follow-up period and a linear weight function weightings of 25%, 50%, 75% and 100% would correspond to 8.75, 17.5, 26.25, and 35 days respectively. This might not be intuitive but it is one way to reduce the number of potential pathways. For cohorts of two or more where multiple patients are experiencing partial tolerance rather than looking at every possible combination you could just look at specific weightings. For example what is the pathway when both patients are at 50% weight, or perhaps when one is at 75% and the other at 25%.

As Yap et al. [31] suggested perhaps the easiest approach is just to assume that full tolerance will be achieved and calculate DTPs from that viewpoint. So treat the trial like a CRM and give full weighting to all the patients. This could be used as an alternative and a way to compare dose decisions made on partial information versus full information. This comparison was also made in our example and we saw how some decisions may change with only partial information. During the design stage of a trial, this may be useful in helping determine the length of any potential observation window and how dose decisions may change during it.

Overall it is possible to produce TITE-DTPs but it very much depends on the number of days that patients can have a partial tolerance. It also depends on the size of the cohort you are evaluating. There are also some practical suggestions for how TITE-DTPs could be used during a trials design as well as it is running. Based on all these factors it appears problematic to produce TITE-DTPs for more than one cohort at a time and this will only be feasible during a trial if you can assume complete information on previous cohorts. Ultimately TITE-DTPs are still able to achieve the same aims as DTP except its ability to look ahead is a lot less.

Statisticians should attempt to produce them wherever possible. They can be used to project the pathways for the first cohort of patients. This will help provide insight into the specific amount of follow-up that is required for certain dose decisions to be made. They can also be used to help calibrate the weight function as well. It could also be beneficial to produce DTPs for the CRM in this instance as a starting point and then work on implementing TITE-DTPs. However, if the scenario or trial parameters mean the TITE-DTPs is too complicated they may be more of a hindrance than a benefit.

Chapter 5

Efficacy Transition Pathways

5.1 Introduction

In Phase II trials we are often attempting to determine whether or not a new treatment or intervention works and establish if there is an efficacy signal. More specifically we aim to determine if there is sufficient level of evidence to warrant further research in a Phase III setting [92]. In addition to assessing efficacy there is also opportunity to further explore the toxicity profile of the treatment. Compared to Phase I trials, Phase II trials are typically conducted using a larger sample size [93]. Generally speaking Phase II trials should be efficient and quick such that we can progress to Phase III as quickly as possible or drop any ineffective treatments.

The output from a Phase II trial should be either a 'GO' or 'No GO' decision i.e should we or should we not proceed to later phase testing based on the data observed in this trial. One of the more important aspects of these trials is that we do not want to make any incorrect decisions and if there is an effective treatment that is being investigated we want to make sure that it is taken forward into Phase III. As such it is important that we try to make correct decisions in Phase II trials. Failure to do so could result in potentially beneficial treatments being rejected or bad treatments being investigated further, which

could negatively impact patients and waste a lot of time and money [94].

However, Phase II trials still face some issues which may make this challenging. Whilst it is the case that Phase II trials are typically larger than Phase I trials, there are still some instances in which we would be dealing with small sample sizes, such as, in a rare disease setting. In these instances we may employ single arm Phase II trial designs along with Bayesian methods to make better use of the data we are able to collect.

Furthermore, we may also be interested in taking a look at some safety data and check that there is sufficient signal of efficacy to warrant continuing the trial. This may take the form of interim analyses and can be thought of in a similar manner to dose decisions in dose-finding trials. Rather than assessing the data and selecting a dose, we are assessing if the trial should continue or not, stopping for either futility in terms of efficacy or safety reasons.

Whilst these designs may be simple to implement they also suffer from similar drawbacks as certain dose-finding methodologies, that have previously been discussed. There may be some issues parametrising these designs e.g. selecting adequate decision rules. Any Bayesian approaches may be less familiar than traditional frequentist approaches that are more commonly used [95], such as the Simon's two-stage design [96]. Clinicians and non-statisticians may struggle to understand why certain decisions are being made during interim and final analyses.

In order to solve some of these issues Lucinda Billingham (LB) developed the idea of Efficacy Transition Pathways (ETPs), a novel visualisation tool to aid the design and interpretation of these types of designs. ETPs are an extension on the concept of Dose Transition Pathways (DTPs). Here they help map out and visualise the different decisions that can be made at interim and final analyses based on different observed outcomes in a similar manner as how DTPs present different dose-decisions that can be made for each cohort.

In this chapter we will detail how ETPs are constructed as well as how they may be used in practice. In addition we present motivating examples where ETPs have been actively implemented and used in clinical trials. The primary aim of this chapter is to develop software to facilitate the implementation of ETPs in order to make them as accessible and as DTPs.

5.2 Efficacy Transition Pathways

ETPs were primarily designed for use in single arm Phase II trials using Bayesian methods where the data is being looked at and assessed frequently. Typically a trial like this, at least in an oncology setting, will have a short-term binary outcome either success or failure, response or no response as its primary outcome.

One approach for these sorts of trials is to use a Beta-Binomial conjugate analysis to estimate a response rate for the binary outcome. Posterior probabilities can be used to inform decision-making and predictive probabilities can be used during interim analyses in a similar manner. Any decisions made will be done using pre-specified decision rules.

In order to demonstrate how ETPs are constructed and used we will look at the design process for a trial using a Beta-Binomial conjugate analysis. When implementing a design like this we need to consider a number of factors such as: the total sample size, the number and timing of any interim analyses, and decision criteria for interim and final analyses. Then, much like with a dose-finding trial, simulations can be conducted to obtain operating characteristics of the design. In addition we can also calculate the number of responses required at each analysis to continue the trial. This is also an iterative process so the design and decision rules can then be tweaked until an acceptable design is reached.

ETPs can be utilised during this process as they are able to present the decisions that will be made based on the current design specification and the potential outcomes that can be observed for each analysis time point. As the design process for a trial involves multiple stakeholders it may not be easy to understand why or when certain decisions are being made. By mapping this all out in a single plot it may be easier to visualise.

For example, based on whatever the current specification is, it may be the case that in the first interim analysis 2/5 patients require a response to continue the trial. Once the clinicians see this they may feel its too strict and would only think about stopping if there are no responses or alternatively they may want to remove this interim analysis and decide to only look at the data once 10 patients have been recruited. This is far more intuitive then trying to explain these concepts and using different boundaries for predictive probabilities. This could then be used to facilitate further discussions about the design specification and if decision rules need to be more or less lenient.

ETPs much like DTPs could also be used throughout the trial as well. So, based on how many responses have already been observed you can easily figure out how many responses would be required for the next cohort of patients to obtain a 'GO' decision. Once the ETPs are produced all future decisions based on the number of responses can also be seen. So, this has the benefit of not requiring a statistician to run an analysis to calculate the specific number of responses that are required for a 'GO' or 'No GO' decision. Whilst, ETPs should not replace the need for a statistician and they should still be involved with all the analyses they act as a tool that can help monitor the progress of a trial and can be referred to throughout the trials life cycle. In the next sections we provide an example trial to show how ETPs are constructed and utilised.

5.2.1 Illustrative example to showcase a Beta-Binomial design

Consider a Phase II trial where the objective is to evaluate the efficacy of some new treatment. This will be evaluated using an outcome measure of response. Patients can either be considered a responder or a non-responder. Consider this is in a rare disease setting so patient numbers are limited and as such we will be using a single-arm design. The treatment effect will be the response rate which will be estimated using a Beta-Binomial conjugate model. We will require a sufficient level of evidence that there is adequate treatment effect to warrant further research in a Phase III trial. Here, we will be utilising a decision space with two outcomes: GO or No GO. However, this approach can be adapted for more complex scenarios, such as the Lalonde framework [97] which implements a Go/Pause/Stop decision structure.

The Beta-Binomial conjugate model is described by Lee [25]. Consider a parameter of interest θ that represents some treatment effect. More specifically, for binomial data in a single arm Phase II clinical trial, let's say the parameter θ is the probability of response in a number of patients following a some new treatment. Each patient can experience either a response or no response, with the same probability of response and each patient being independent from each other. For a fixed sample size with n patients and number of responses (y) we have:

$$Y \sim \text{Binomial}(n, \theta) \quad (5.1)$$

So, y is from a binomial distribution which produces the following likelihood:

$$L(\theta) = P(y|\theta) = \binom{n}{y} \theta^y (1-\theta)^{n-y} \quad (y = 0, 1, \dots, n) \quad (5.2)$$

If the prior for θ is from a Beta distribution such that

$$P(\theta) = \text{Beta}(a, b) \quad (5.3)$$

then the posterior distribution is also from a Beta distribution and can be expressed as

$$P(\theta|y) = \text{Beta}(a + y, b + n - y) \quad (5.4)$$

To avoid confusion with the prior we will let $\alpha = a + y$ and $\beta = b + n - y$ which gives

$$P(\theta|y) = \text{Beta}(\alpha, \beta) \quad (5.5)$$

Bayesian inference can then be used for estimation and decision making. Features such as the mean and variance of the treatment effect θ can be estimated from the posterior distribution given in equation 5.5. For the posterior distribution of $\theta \sim \text{Beta}(\alpha, \beta)$ the mean, variance and mode are.

$$E[\theta] = \frac{\alpha}{\alpha + \beta} \quad (5.6)$$

$$\text{Var}[\theta] = \frac{\alpha\beta}{(\alpha + \beta)^2(\alpha + \beta + 1)} \quad (5.7)$$

$$\text{mode}[\theta] = \frac{\alpha - 1}{\alpha + \beta - 2} \quad (5.8)$$

The proofs for these formula are relatively simple and can be found in [98]. We can also establish credible intervals or make other probability statements in a similar manner.

We can also establish credible intervals or make other probability statements in a similar manner. Then pre-specified rules based on these direct probabilities from the posterior can be used for decision making purposes. For example, if

$$P(\theta > c|y) \geq q \quad \text{then GO else No GO} \quad (5.9)$$

where c is some target level of treatment effect and q is some threshold of sufficient evidence.

For our example we specify a minimally informative Beta(1,1) prior. This represents a 50% response rate from a group of two patients. Data will also need to be collected from patients recording if they had a response or not. This in turn is combined with the prior distribution to generate a posterior distribution for the treatment effect.

We will also specify the following decision rule will also be used $P(\theta \geq 30\%) \geq 0.9$. So, if there is a greater than 90% chance that the true response rate is at least 30% this will be considered sufficient evidence to warrant a 'GO' decision. This criteria is just for illustrative purposes, in practice both clinical and statistical considerations should be made for specifying these parameters. In a rare disease setting there may not be many effective treatments so targeting a low level of treatment effect could still be considered an improvement for patients. Depending on the scenario it may also be appropriate to require a high level of evidence that the treatment is effective. There may be a heavy treatment burden on patients with multiple hospital visits or negative side effects and so clinicians may want a higher amount of certainty that the targeted level of treatment is reached to trigger a 'GO' decision. For this example trial we will set our sample size at 30 patients.

We will also include interim analyses after every five patients to evaluate

whether or not the trial should be stopped for futility. To do this we will use the predictive probability of success (PPoS).

Consider for a fixed sample of size n patients an interim analysis that will be conducted after n_{int} patients, such that $n_{int} < n$. Again, the parameter of interest, treatment effect or response rate will be represented by θ . Let the number of responses x among the n_{int} patients follow a binomial distribution:

$$X \sim \text{Binomial}(n_{int}, \theta) \quad (5.10)$$

Let Z be the number of responses in the remaining m patients, where $m = n - n_{int}$. When $Z = i$, where $i = 1, \dots, m$, the posterior distribution can be expressed as

$$P(\theta|x, Z = i) = \text{Beta}(a + x + z, b + n - x - z) \quad (5.11)$$

Where a and b are parameters from the Beta prior distribution, see equation 5.3.

Suppose we have a decision rule where a GO decision is made if the posterior probability of θ exceeds some pre-specified target treatment effect/ response rate c with a probability greater than some threshold q (as defined in equation 5.9).

The predictive probability of success (PPoS) can be calculated as follows. Let $B_i = P(\theta > c|x, Z = i)$ and $I_i = I(B_i \geq q)$ be an indicator variable taking a value of 1, if the criteria $B_i \geq q$ is satisfied or 0 otherwise. Then we have

$$\begin{aligned}
\text{PPoS} &= E\{I[P(\theta > c|x, Z) \geq q]|x\} \\
&= \int I[P(\theta > c|x, Z) \geq q]dP(Z|x) \\
&= \sum_{i=0}^m P(Z = i|x)I[P(\theta > c|x, Z = i) \geq q] \\
&= \sum_{i=0}^m P(Z = i|x)I(B_i \geq q) \\
&= \sum_{i=0}^m P(Z = i|x)I_i
\end{aligned} \tag{5.12}$$

Where $P(Z = i|x)$ is the probability of observing Z responses in m patients based on a beta probability distribution with parameters $a + x$ and $b + n_{int} - x$. This can be calculated from the probability mass function

$$\begin{aligned}
f(Z = i|m, a + x, b + n_{int} - x) &= \binom{m}{z} \frac{\text{Beta}(a + x + z, b + n_{int} + m - x - i)}{\text{Beta}(a + x, b + n_{int} - x)} \\
&= \binom{m}{z} \frac{\text{Beta}(a + x + z, b + n - x - i)}{\text{Beta}(a + x, b + n_{int} - x)}
\end{aligned} \tag{5.13}$$

The Beta functions can be further simplified or alternatively expressed using the gamma function. Note $\Gamma(k) = (k - 1)!$

$$\begin{aligned}
f(Z = i) &= \frac{m!}{z!(m-z)!} \frac{\frac{\Gamma(a + x + z)\Gamma(b + n - x - z)}{\Gamma(a + b + n)}}{\frac{\Gamma(a + x)\Gamma(b + n_{int} - x)}{\Gamma(a + b + n)}} \\
&= \frac{\Gamma(m+1)}{\Gamma(z+1)\Gamma(m-z+1)} \frac{\Gamma(a + x + z)\Gamma(b + n - x - z)}{\Gamma(a + b + n)} \frac{a + b + n_{int}}{\Gamma(a + x)\Gamma(b + n_{int} - x)}
\end{aligned} \tag{5.14}$$

The process to calculate the PPoS starts with the quantity B_i which represents the probability that the response rate is greater than some target c given x responses in n_{int} patients and assuming i future responses in the remaining

m patients. The quantity B_i is then compared to our probability threshold q which provides a value for the indicator variable I_i and informs us if the trial would result in a GO decision at the end of the trial dependent on the data observed and the value of $Z = i$. The indicators I_i are then weighted by the probability $P(Z = i)$ and summed to give the PPoS.

PPoS is then interpreted and used to make decisions at interim analyses. Low values of PPoS suggest there is a low probability of achieving a GO decision at the final analysis based on the data accrued so far. Similarly, a high PPoS suggests the opposite and that the trial would likely be a success based on the current data. An interim decision rule can then be implemented around values of PPoS to recommend stopping the trial early for either efficacy or futility. To stop for futility a the following rule could be imposed

$$\text{PPoS} < t \quad \text{then STOP for futility} \quad (5.15)$$

Where t is some PPoS acceptable probability threshold. Values of t range from 0 to 1 but would typically be small. For example a value of 0.2 may be used to indicate if there is a less than 20% chance that the response rate at the end of the trial will be greater than c with some probability q then the trial should stop early.

For our example trial we will specify a PPoS acceptable probability threshold where if $\text{PPoS} < 0.05$ then we should stop the trial for futility. This implies we would stop the trial if there is a less than 5% chance that the response rate at the end of the trial will be greater than 30% with a probability of 0.9 i.e a less than 5% chance that the trial would reach a GO decision or be considered a success.

Table 5.1 details the minimum number of responses required under this design at each analysis time point. For the final analysis, once 30 patients

have been recruited, a minimum of 13 responses must be observed in order to have a GO decision. We can also see what the minimum number of responses required are for each interim analysis.

TABLE 5.1: Specification of parameters for the example Beta-Binomial trial.

Analysis	Minimum No. Responses for GO Decision
N = 5	1
N = 10	2
N = 15	3
N = 20	7
N = 25	9
N = 30	13

Most of the specifications we have made here along with our decision rules are fairly arbitrary. In practice decision rules should be decided before the trial starts. This is typically done via the evaluation of simulations and looking at the minimum number of responses required for GO decisions. Multiple scenarios corresponding to different true response rates can be investigated. The probability of making a correct decision can then be calculated. This represents the power of our design. For scenarios with low true response rates (relative to the target response rate) we want the probability of making a No GO decision to be high. Similarly, for scenarios with high true response rates we want the probability of making a GO decision to be high. The decision rule parameters, the target response rate and probability threshold can then be adjusted to ensure the design is making appropriate decisions in these scenarios.

Simulations for this example trial are shown in Table 5.2. There are five different scenarios where we vary the true response rate. For each scenario we simulate 10000 trials. for each individual trial run we sample responses from patients using the underlying true response rate and determine if the criteria are met to pass the interim analyses and final analysis decision criteria. The table records the probability in each scenario of there being a 'GO' decision. In

scenarios where the true response rate is low (10% and 20%) we have almost no chance of a GO decision. As our decision criteria targets a response rate of 30% if the actual response rate was also 30% a 'GO' decision would be the desired outcome. However, the simulations show with the current design this does not often occur. For higher true response rates the design does perform slightly better in reaching 'GO' decisions with probabilities of 0.414 and 0.796 for 40% and 50% respectively.

TABLE 5.2: Simulations for example Beta-Binomial design.

True response rate	Probability of 'GO' decision
10%	0.000
20%	0.003
30%	0.078
40%	0.414
50%	0.796

Simulations are always an important aspect of the design process however, they may not convey what is most meaningful for other collaborators when assessing the merits of each iterative design specification. This is where ETPs can be used to try and bridge that gap by offering a way to visually represent key information and decisions that are made with these types of designs. In the next section we detail how an ETP is constructed using the example trial design specified here.

5.2.2 Constructing Efficacy Transition Pathways

At each interim analysis PPoS is calculated based on the number of responses observed thus far and evaluated to see if it meets the decision criteria. Therefore there will be a minimum number of responses that have to be observed in

order to continue recruitment. This is similar to how we can calculate the number of responses required at the end of the trial to warrant a GO decision. The minimum number of responses required at each interim and the final analysis depends on the decision criteria that is specified.

Intuitively, it is easier to understand that three responses have to be observed from 15 patients rather than a $\text{PPoS} \leq 0.05$ is needed. Through discussions with clinicians we can then calibrate our decision criteria based on these interpretations. We may want to be more strict or lenient at our interim. If the clinicians would be happy to continue recruitment after seeing only two responses we could lower the PPoS threshold, likewise if they wanted to be confident and only continue if six responses were observed we would increase the PPoS threshold. Similarly, this can also be done for the final analysis decision criteria. The acceptable probability level or target response rate could be adjusted so a specific minimum number of responses observed achieves a GO decision. Any changes made should be assessed by simulations as drastically changing the decision criteria could have a negative impact on performance.

As more interim analyses are added to a design, it may be more complicated to keep track of the specific number of responses required for 'GO' decisions at each interim. A solution for this is ETPs. In our example trial we have a maximum sample size of 30 patients with interim analyses planned after every five patients. This results in a total of six analyses, one final analysis and five interims. The decision criteria at the end of the trial requires the treatment effect to be greater than 30% with a probability of 0.9 and to pass the interim analyses we require the PPoS to be greater than 0.05.

To construct an ETP we produce individual cells which contain key information about a specific outcome i.e. a certain amount of responses. If we consider our first interim at five patients, at that point there are six different possible outcomes that can be observed. Either one, two, three, four or all five

of the patients had a response or none of them did. For each possible outcome we can then calculate the PPoS as well as the Bayesian estimate of the response rate and an associated credible interval. Figure 5.1 shows what this cell would look like.

FIGURE 5.1: ETP cell plot for 0 responses in 5 patients.

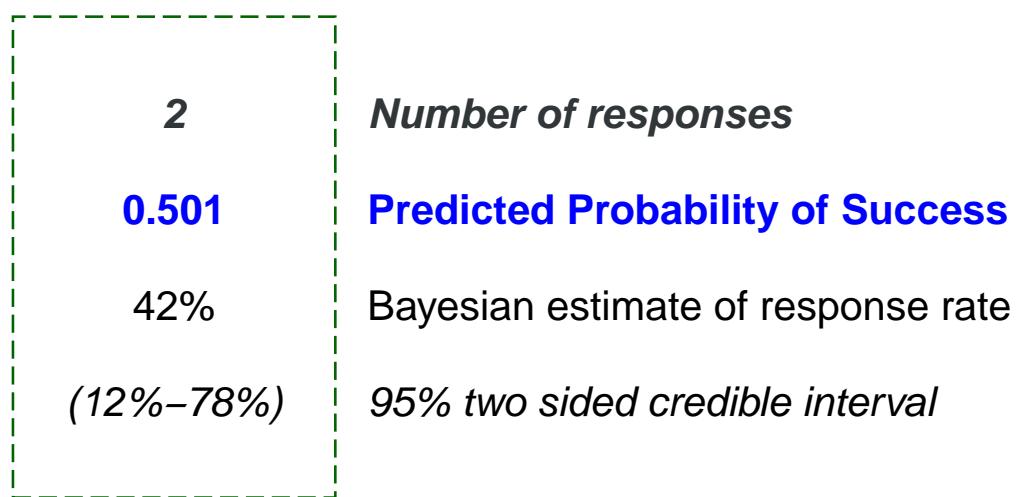
0	Number of responses
0.025	Predicted Probability of Success
11%	Bayesian estimate of response rate
(0%–46%)	<i>95% two sided credible interval</i>

The number at the top indicates the outcome for the cell, in this case the number of responses which is 0. The second row shows the PPoS in this scenario, the third row showing the Bayesian posterior estimate of response rate and the last row shows the 95% credible interval. For 0 responses the PPoS is 0.025 which is less than our threshold so the decision here would be to stop. This is represented by the red dashed line. From this one cell we are able to see what the decision would be at the interim analysis time point if this is the outcome that is observed. We are also able to see specifically what the PPoS and estimated response rate would be as well. The choice was made to present probabilities with decimals and any estimates of response rates with percentages. This was so the two could easily be differentiated.

Cells are generated for each possible outcome at each interim time point. Figure 5.2 shows the cell for two responses in five patients. Here we can see

the PPoS is 0.501 which is greater than our threshold so the decision would be made to continue recruitment. This is indicated by the green dashed line. As we put each cell together we can then clearly see the minimum number of responses required to continue recruiting.

FIGURE 5.2: ETP cell plot for 2 responses in 5 patients.



This process is then repeated for each interim analysis. So, the next analysis would be at 10 patients. Here we would generate 11 cells for all the different possible outcomes (no response, one response, two responses, ..., 10 responses). The same would then be done for the analysis at 15, 20 and 25 patients.

For the final analysis the presentation of the cells is slightly different. Here we are no longer interested in PPoS as no more patients will be recruited and rather we can just evaluate if the trial has met the decision criteria. So, in each cell rather than present PPoS, the posterior probability that the response rate is greater than our target rate is presented instead. Figures 5.3 and 5.4 show the cells for 10 responses and 14 responses out of 30 patients respectively. In this example our q is set at 0.9 so if the posterior probability is greater than that we have a GO decision.

We opted to present the PPoS and posterior probability as decimals. This was done to avoid these values being confused with estimated treatment effect. It may be easy for a non-statistician to see a posterior probability of 0.976 and interpret it as the treatment having a 97.6% response rate. These are also presented to three significant figures. This was done as in some instances the posterior probability was shown to be the same value for adjacent cells which could also cause some confusion. When constructing ETPs the presentation of these values and the cells in general can be altered to suit the needs of the users.

FIGURE 5.3: ETP cell plot for 10 responses in 30 patients.

10 0.688 34% (19%–51%)	Number of responses Posterior Probability Bayesian estimate of response rate <i>95% two sided credible interval</i>
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The efficacy transition pathway is then constructed by grouping each cell for each interim analysis and then stacking those group of cells together. For our example trial the ETP is shown in Figure 5.5. Each row of cells in the ETP represents each interim analysis with the final row representing the final analysis. One adaptation made with the cells is that the confidence interval is presented across the bottom two rows in each cell just to make the figure easier to read and more scalable.

FIGURE 5.4: ETP cell plot for 14 responses in 30 patients.

14 0.976 47% (30%–64%)	Number of responses Posterior Probability Bayesian estimate of response rate <i>95% two sided credible interval</i>
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From this figure what can clearly be seen is when we do or do not have a GO decision. At each interim of 5, 10, 15, 20, 25 patients we can see that the minimum number of responses for a GO decision is 1, 2, 4, 7 and 9 respectively. Also, for the final analysis a minimum of 13 responses is required. We can also read down the figure to gauge an idea of how many additional responses would be required in future analyses to warrant a GO decision. For instance if you observed five responses in your first cohort of five patients, that is enough observed data to continue passing the interim decision criteria until the fourth interim analysis in which case you would need an additional two responses. This can easily be seen by reformatting the ETP to be aligned to the left so the same number of responses are stacked on top of each other for each cohort. This is illustrated in Figure 5.6.

In addition to the easy visualisation of the number of responses required to achieve a GO decision, we can also see the estimates of the treatment effect for each potential outcome and each analysis time point. This is useful for interpreting the results for the final analysis (the bottom row of the ETP plot). Right away from interpreting the cell for the minimum number of responses

(13/30) for a GO decision we can see the estimated response rate would be 44% with a probability of 0.95 that the true response rate is between 27% and 61%. It is important to remember the original decision rule for the final analysis which for any GO decision implies there is a greater than 0.9 probability that the response rate is greater than 30%. This can then be used to facilitate discussions with the clinicians to deem if this is an acceptable level of evidence to warrant further research or potentially bring into practice, based on previous studies, current treatments or their experience. We can then adjust our design and decision rules accordingly, which we discuss in the next section.

FIGURE 5.5: Example of a constructed ETP.

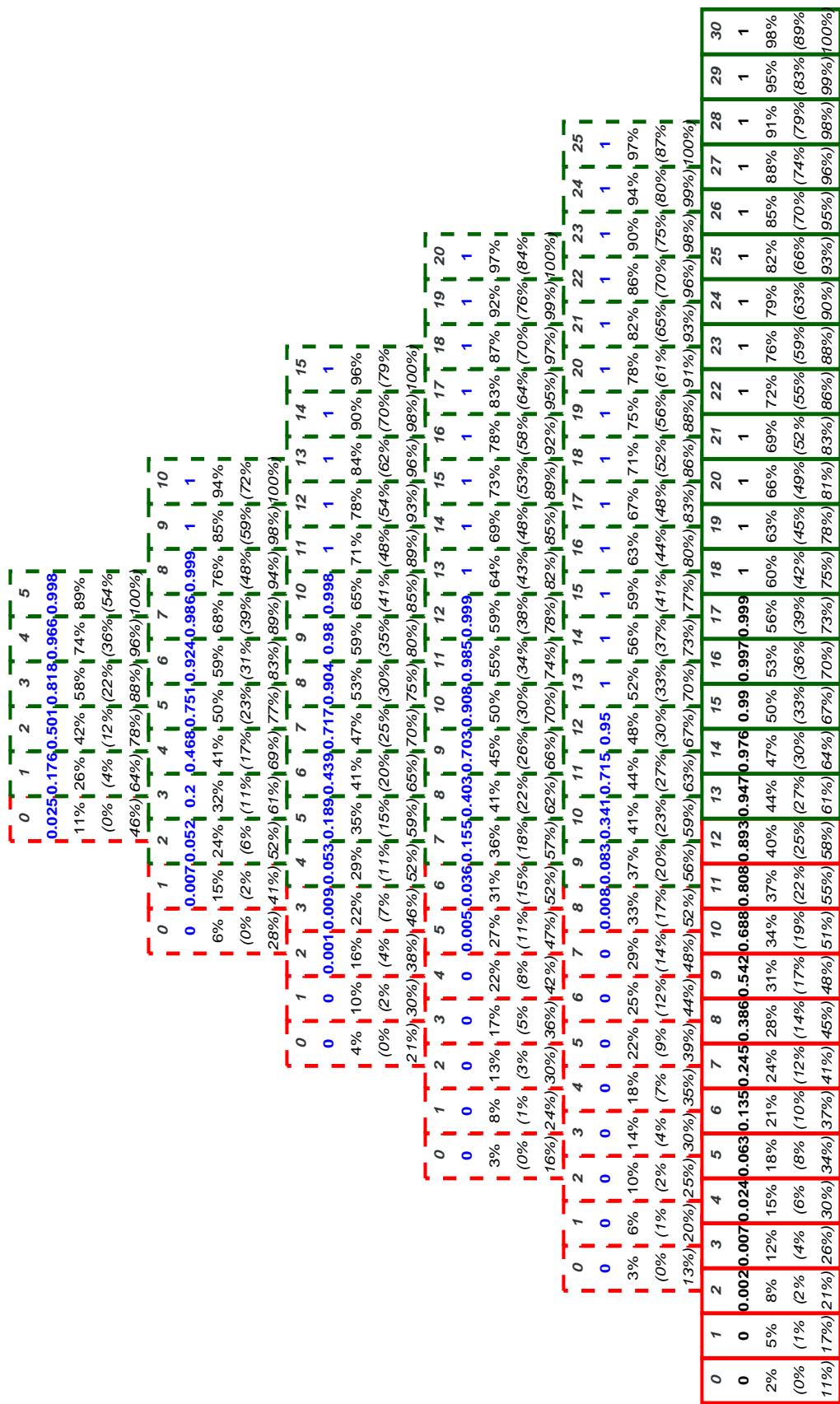
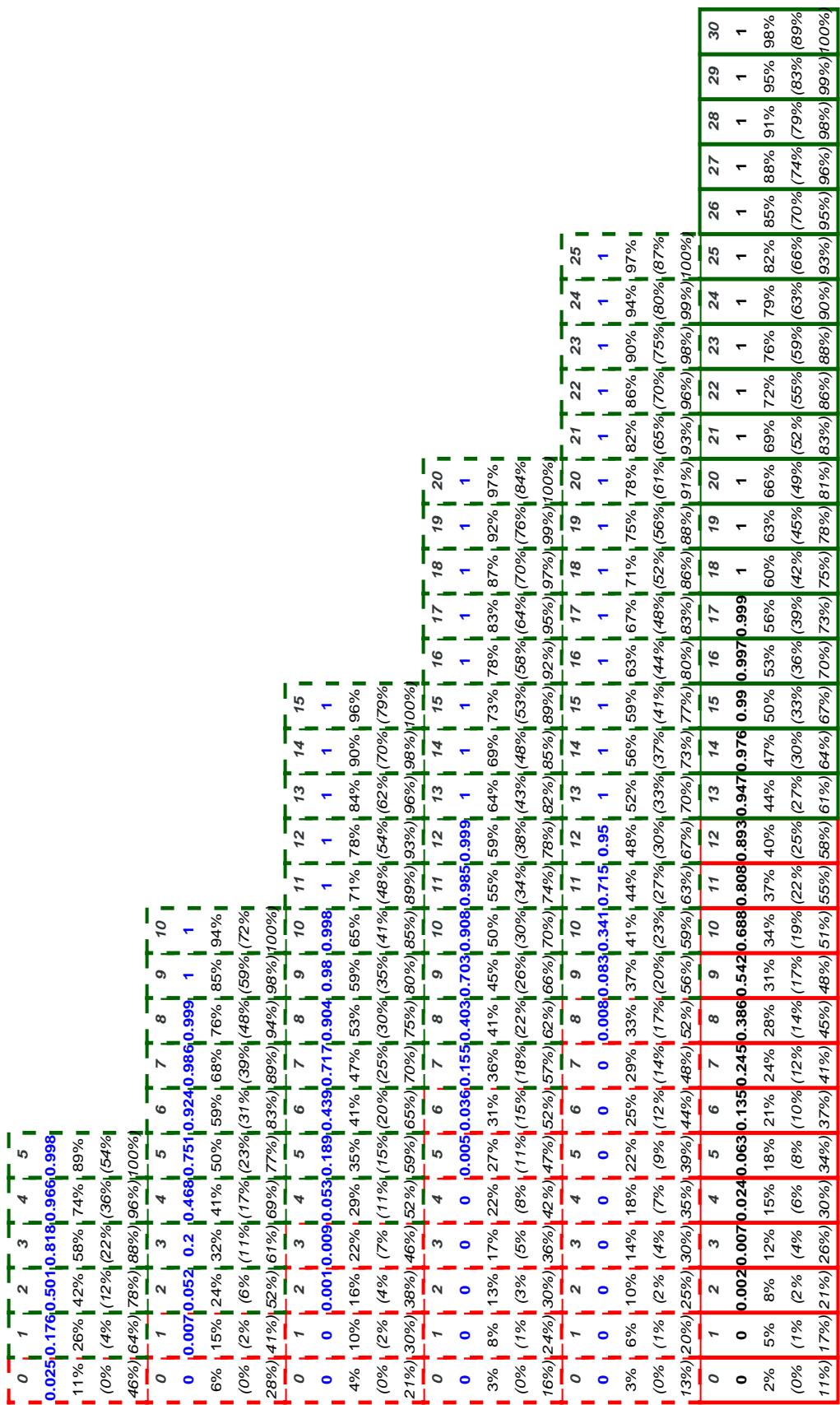


FIGURE 5.6: A left aligned version of the constructed ETP.



5.2.3 Updating decision criteria based on Efficacy Transition Pathways

Having seen the ETP we created for our example trial, suppose we want to modify our decision criteria such that a GO decision at the final analysis requires only nine patients to achieve a response. Compared to the initial decision criteria, which required 13 responses, we may want to consider lowering this due to the treatment options available for these patients may not being very effective. Obviously there may be many reasons why we would want to increase or decrease this requirement and this would depend on the context and background of each individual trial.

By looking directly at the ETP Figures 5.5 or 5.6 we can see that nine responses out of 30 patients has a posterior probability of 0.542. So, based of the initial decision rule of $P(\theta \geq 30\%) \geq 0.9$, this implies that the probability that the response rate is greater than or equal to 30% is 0.542. Therefore, if we wanted to make this a GO decision we would change our decision criteria such that our acceptable level of probability was something smaller than that posterior probability. For example, a new decision rule could be $P(\theta \geq 30\%) \geq 0.5$. This would mean that nine responses out of 30 patients would now be a GO decision. This is another benefit of ETPs, we can quickly ascertain how we would need to change our decision criteria to be in order for a specific minimum number of responses to be a GO decision.

Given the ETPs observed in the previous section and the simulations shown in Table 5.2 we will implement this new decision rule to see how it impacts our design. We provide updated simulations in Table 5.3. Here we can see that there is now an increase in probability of making a 'GO' decision if the true response rates are low (10% and 20%) however these are still relatively small.

Compared to before there is now higher probabilities of making a correct decision when the true response rates are above 30%.

TABLE 5.3: Simulations for example Beta-Binomial design with new decision rule.

True response rate	Probability of GO decision
10%	0.001
20%	0.126
30%	0.558
40%	0.892
50%	0.988

We can then also update our ETPs. In Figure 5.7 we can see for the final analysis (the bottom row of the plot) GO decisions start from nine responses. It is important to note the content of these cells have not changed. The estimates of response rates and credible intervals are still the same. This is because we fundamentally have not made any changes to the design, just the criteria for which we are making decisions. Changes in these estimates would only be triggered if we were to change the total sample size or prior. Additionally the posterior probabilities are also consistent with the previous ETP, this is because we have not altered the targeted response rate in our decision criteria. This value is still showing the probability that the treatment has a response rate greater than 30%. These values would only differ if our target response rate was set as something else. For the rest of the cells showing previous cohorts and the interim analyses we can see the minimum number of responses required for a GO decision is now 0, 1, 3, 4 and 6 for each interim analysis respectively. Also, note how the PPoS is different compared to the last ETP. This

is because PPoS takes into account the final decision rule we are using. Any change to that rule will impact PPoS calculations.

By altering our decision rule like this we ensure that if we observe the desired number of responses, in this case nine, the trial would be a success. We did this by just changing q , the acceptable probability level. However, it should be noted that this can also be achieved by changing both c (the target response rate) and q . These values can both be manipulated so we still maintain the same practical decision making. This is important to note as whilst clinicians may find it more intuitive to specify decision rules based on a minimum number of responses there are different ways in which that can be achieved.

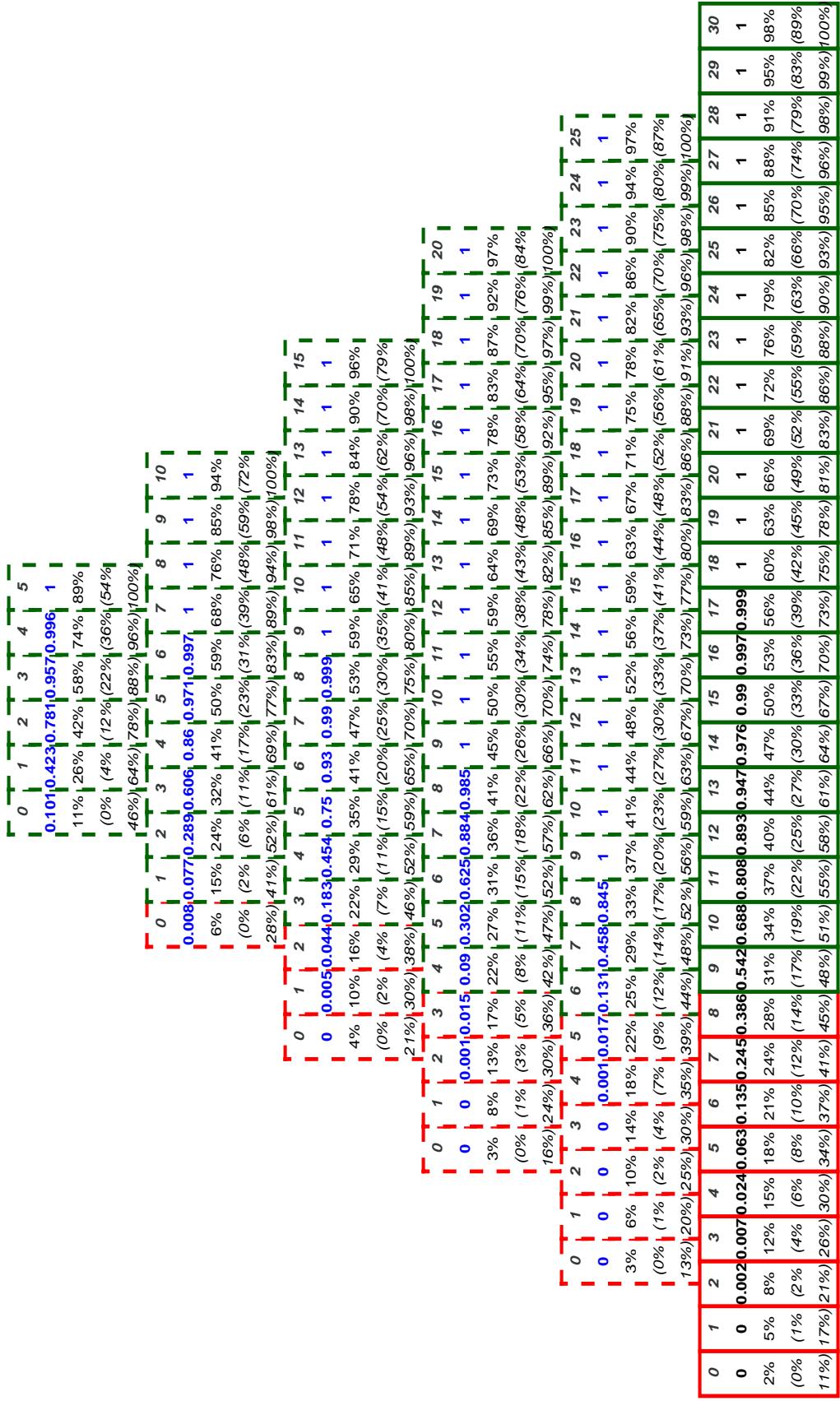
Table 5.4 details the posterior probability that the estimated response rate is greater than four different target response rates given we observe nine responses out of 30 patients. If our target response rate is 10% and we observe nine responses the posterior probability is 0.999 so then our decision rule can be specified as if $P(\theta \geq 10\%) \geq 0.95$. Here our value of q just has to take a value lower than the posterior probability but we should be careful to make sure it is not low enough such that observing 8 responses also becomes a GO decision. Similar statements can also be made about the other target response rates presented in the table. The associated decision criteria for those response rates are also presented.

TABLE 5.4: Examples of different decision criteria.

Posterior Probability for 9 Responses	c	q
$P(\theta \geq 10\%) = 0.999$	10%	0.95
$P(\theta \geq 20\%) = 0.926$	20%	0.90
$P(\theta \geq 30\%) = 0.542$	30%	0.50
$P(\theta \geq 40\%) = 0.143$	40%	0.10

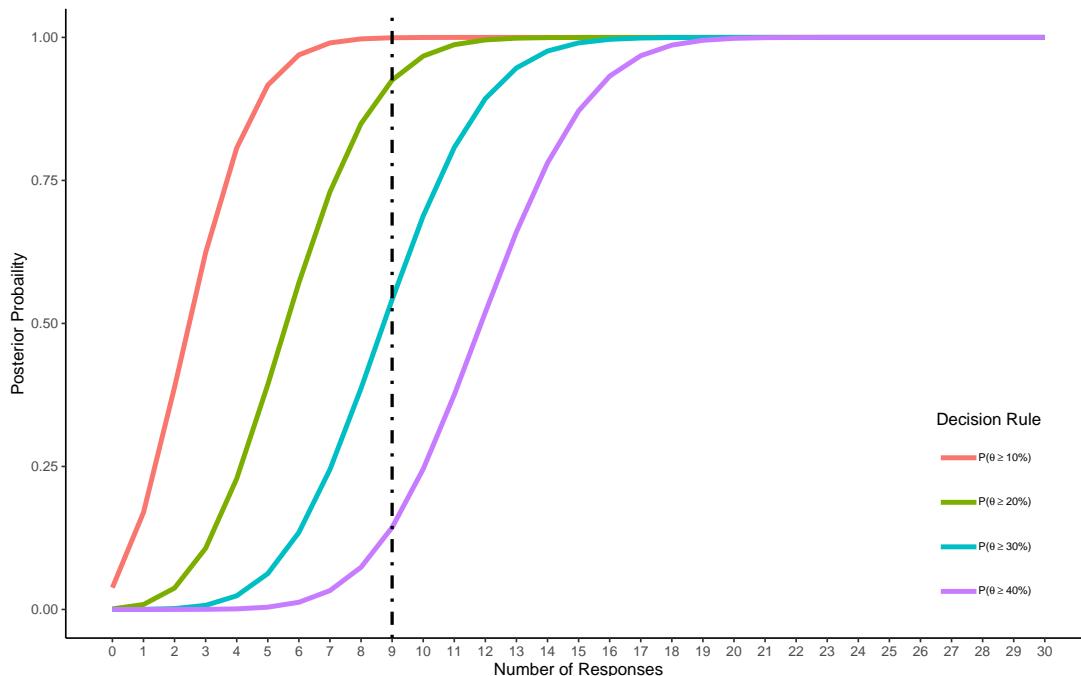
This is also shown in Figure 5.8. Here there are four curves each representing the posterior probability that the estimated response rate is greater than the target response rates of 10%, 20%, 30% and 40% for all possible number

FIGURE 5.7: ETP with updated final decision rule.



of responses. The dashed vertical line represents the minimum number of responses we would require for a GO decision and where this crosses with each curve is the posterior probability that the treatment effect is greater than those target response rates and should be used to inform our values of q . By adding in more curves for different target response rates or moving the dashed line for a different number of minimum responses we can determine appropriate values for our decision criteria.

FIGURE 5.8: Changes in posterior probability for decision criteria.



Here we have shown how ETPs are constructed and how they change and react to modifications in our decision rules. There are several other factors which can impact an ETP such as the prior used in the Beta-Binomial conjugate analysis as well as the timing of each interim analysis and the overall sample size of the trial. Changes to the prior will have an impact on all of the calculations as it is used to generate the posterior distribution on which all of the other calculations are based upon. Adding more interim analyses would

add more rows to the plot and changing the sample size of the trial alters the number of cells in each row.

Overall ETPs can be a useful tool during the design stages of a trial as we can experiment with different decision rules and see what practical effect it has on the trial in terms of the number of responses that need to be observed for a GO decision. They can be used to facilitate discussions with non-statistical experts involved in the design of the trial. Much like dose transition pathways in a dose-finding trial they can also provide some transparency as to what decisions will be made and when they would be made.

A single ETP also provides us with the ability to see how multiple different decision rules may change the outcome for a trial. If just the acceptable probability levels for the PPoS and final analysis, t and q , are changing in the decision rule the impact of those changes should be apparent just by comparing the PPoS and posterior probability without the need of generating a new ETP like we have in our example.

Whilst the calculations needed to produce these plots can be simple, constructing the plots can be challenging. To overcome this issue and make ETPs easily accessible and producible, we developed a web based application to generate ETPs. All the ETPs shown in this chapter were generated using this application.

In the next sections we detail trials that have been designed using ETPs. These served as further motivation for development of the application to produce ETPs. We then go on to detail how the app was developed and how it works.

5.3 Implementation of ETPs

With the development of any new methodology or novel tool such as ETPs there will be some barriers that will impede its use. One of those barriers will be the difficulty of implementing the methodology. If the intention of some newly developed methodology is for it be applied in a practical setting, when presented it should be accompanied by appropriate software or code such that the target audience are able to implement it with minimal effort. Otherwise the new methodology may remain purely theoretical and would rely on others to come up with a solution for its implementation.

To overcome this barrier for ETPs we developed a R function to produce these plots given the input of key details such as the decision criteria, sample size and cohort size. We then used this function and built a web application around it. Rather than just offering code to implement ETPs a web app makes implementation even easier as it does not require knowledge or experience with a specific piece of stats software such as R, STATA or SAS.

Another barrier that limits the rate at which new methodology is implemented is a lack of awareness or familiarity with the methodology. Specifically, with clinical trials that often involve a multidisciplinary team it is unrealistic to expect clinicians or trial management to be up to date with the latest statistical innovations. Some statisticians may not be familiar with the latest methodology if it is not primarily an area they specialise in. As a result, newer methods may be overlooked even if it would be beneficial to implement. Even if statisticians are aware of new methodology, the struggle may then become explaining the methodology to non-statisticians and convincing them it will be beneficial to use.

Inherently ETPs as a tool were created to help better explain the analysis that is done in these phase II trials as well as the decisions that are made. They

exist more as a tool to help promote the underlying Bayesian methodology. Therefore ETPs may be simple to implement and explain to non-statisticians. Instead the issue may come from a lack of understanding of a Beta-Binomial conjugate analysis, predictive probabilities or the background methodology behind the ETPs.

To address this within our web application we included detailed information about ETPs as well as a breakdown of the methodology behind them. This was done through text and images in combination with custom built interactive tools that illustrates how these trials are run in a practical setting. These additional explanations and features were built in to make understanding ETPs easier, especially for those with a non stats background. These sections can also be used as a teaching tool to help educate those who are working on trials but not familiar with this methodology.

Lucinda Billingham (LB) as the visionary behind ETPs had began implementing them in a number of different trial designs despite these barriers. Some of these trials were being designed as umbrella, basket or platform trials and involved multiple arms. Here analyses and decisions were being made in each arm independently so ETPs were employed to help design these trials.

This leads to more issues where during the design stages of a trial multiple ETPs may need to be generated. If changes were made to specific decision criteria the ETP would need to be updated so you could communicate how those changes would affect the outcome of the trial. This is then further compounded with the complex trial designs where there may different criteria dependent on the arms in the trial.

Prior to the development of the app ETPs were being constructed by hand which was a time consuming endeavour. In Appendix A we detail three trials that were designed by the Cancer Research UK Clinical Trials Unit (CRCTU) at the University of Birmingham and show how ETPs have previously been

implemented.

5.4 Development of a web application for ETPs

R shiny [99] is a tool available in R to facilitate the production of interactive web applications (apps). It allows R users to create useful and accessible tools that would otherwise be out of reach of non R users without requiring any programming or statistical knowledge. In the context of medical statistics the usefulness of these types of apps is also acknowledged by funding bodies such as the NIHR (National Institute for Health and Care Research). They have awarded grants which in part helped the development of apps relating to network meta-analysis [100] and meta-analyses of diagnostic test accuracy studies [101], [102].

Even without funding there have been apps that have been developed to help address issues around the implementation of early phase and adaptive designs. For example, there is a whole suite of software and shiny applications made available by the MD Anderson Cancer Centre [103] to help implement methodology they have developed. Wheeler et al. [104] produced the A plus B app to produce operating characteristics for A+B trial designs [105]. There is also the MoDEsT app created by Pallmann et al. [106] for designing and conducting single-agent dose-finding trials. Grayling and Wason [107] developed an app to aid the implementation of multi-arm clinical trial designs. These are just a few examples of shiny apps already available, all of which aim to make certain methodologies more accessible and is something we also wanted to do for ETPs.

Initially we produced a R function that was able to create ETPs. Then using R shiny an application was built which utilised this function to produce ETPs. Whilst the R function makes implementing ETPs a lot easier, the R shiny

app offers an even better solution. Firstly, it does not require any previous knowledge of a specific statistical package so any statistician should be able to use the application with ease. This also applies for non-statisticians and an application would make ETPs more accessible.

R shiny also has the ability to make graphs interactive. Whilst ETPs produce a lot of information they are static in nature. As such a change in parameters or design characteristics means that a separate ETP would need to be produced. With R shiny a interface allows for these changes to be easily made and the plots automatically be updated. Additionally, elements of interactivity can also be included in an application. In the application by clicking on a specific cell of the ETP we produce additional plots which give the user more details than they would otherwise get with just a single ETP. The ETPs presented in Sections 5.2.2 and 5.2.3 were all produced in a few clicks using this app.

In our app we have a specific page which is labelled "plot builder" that will generate ETPs. This page is split into two sections. The section on the left can be considered as the input and the section on the right is the output. There are three tabs in the input section and each one deals with a separate set of input parameters. The prior parameters tab allows the user to specify priors that are being used in the beta-binomial conjugate analysis and also produces a plot of the corresponding beta distribution. The app can be accessed at the following link - <https://amit-patel.shinyapps.io/beta-binomialapp/>. A video demonstrating the key features of the app is also available on YouTube through this link - <https://www.youtube.com/watch?v=vfVzwBDp9-E>.

The design parameters tab allows the user to specify the details of their trial and all the relevant information required to produce an ETP. This includes the number of cohorts and the size of each cohort, this assumes that an interim analysis will be performed once each cohort has been recruited. Details also

need to be inputted regarding the decision criteria at both the interim analysis and final analysis stage. As these design parameters are input the output in the right hand side section named 'Decision Rules' also updates. Here the decision criteria based on the inputs are presented to the user in a mathematical format. There is also text to explain what each of the rules mean so the users can ensure they have inputted the correct details. The interpretation will also be useful for non statisticians and provide some understanding of the decision criteria and what it means.

The final input tab is for parameters corresponding to the visualisation of the plot. These correspond to arguments within the R function which allow the user to adjust how the ETPs look. There is the ability to change the alignment of the ETP such that the cells are either centred or left aligned. There is also an option to change the size of the text in the ETP. As ETPs can become more and more complicated depending on the design of the trial the text may become over crowded so would need to be adjusted, which is what this option allows for. There is also the option to turn the legend on and off.

In the output section the tab labelled "Efficacy Transition Pathway" contains the ETP which is generated based on all the inputs from the input section (prior, design and plot parameters). The user can clearly see what the ETP for their given design would look like. From here the user also has the ability to download the ETP in multiple formats. Any changes made to the input parameters will result in the ETP being updated. This allows for the user to easily and quickly make tweaks to the trial design and see how the decisions made would change. They can also easily save/download all versions of the ETPs they make to compare across the different designs.

There is also an added layer of interactivity that was incorporated into the app. By clicking on an individual cell in the ETP additional information and plots will be generated. Once a cell has been clicked on a line of text will appear

below the ETP which provides specific details about what each of the numbers in the cell represent. Additionally, there will also be a plot of the individual cell with added text to explain what each number in the cell represents. This will allow users to investigate and further explore individual cells. This is specifically useful in scenarios where the ETP may be large and contain many cells and some may not be clear on the ETP. Additionally, a posterior distribution plot is also generated when a user clicks on a specific cell. This will allow the user to see at the specific analysis time point what the posterior distribution of the treatment effect looks like given a specific number of responses. The plot also includes key statistics as well as the specified credible intervals. This plot is produced for all cells, even those at the interim analysis, whilst these may not be the final posterior distribution it provides a visualisation of the treatment effect at that interim analysis which can than either be compared to other time points or changes in the number of responses.

The final tab here presents the data that is used to generate the ETP plot. Each cells data is included in a row and contains results from the PPoS and posterior probability calculations. There is also some functionality within the app that so that the data can be ordered by specific columns or searched. Additionally, this dataset can also be downloaded. This allows users to take the key data and calculations for use outside of the app. Users can also take the data and use it to create their own ETP as well. Whilst the primary objective of the app is to produce the ETP it also serves as a quick calculator for PPoS. Which is a useful feature in its own right as users could specify there design and download the data and use the PPoS values in their SAPs, protocols, grant applications etc.

The way in which the app works is that all the parameters for the Beta-Binomial trial design and inputs for the ETP are stored. They are then fed into the R function which outputs the data table. Based on the data the function

will produce the required number of cells which are then plotted on a cartesian plane. Then each of the values which make up the individual cells are plotted at fixed y coordinates. The interactive element works by registering the coordinates of where the user clicks on the ETP plot. Then it finds the coordinates of the centre of the nearest cell in a specific margin and extracts the data for that cell. That extracted data is then used to create the additional elements like the text explaining that cell, the enhanced individual cell plot and the specific posterior distribution plot.

5.4.1 Additional Features of the App

We have shown how the app allows for easy creation of ETPs so anyone familiar with the methodology and an internet connection is just a few clicks away from being able to produce these plots. The other issue we mentioned earlier with the introduction of new methodology is lack of awareness or familiarity. In order to address this we built additional pages in the app that act as an educational resource which provide all the prerequisite knowledge required to understand ETPs. These pages cover all the material covered in this chapter. Starting from the basics of Phase II trials, Bayesian statistics, Beta-Binomial conjugate analyses and PPoS. The traditional way to explain new concepts would usually be through some combination of text and images and whilst we employ these to introduce more of the simpler elements of ETPs, R shiny gives us the ability to incorporate some level of interactivity within the material.

The navigation bar on the right hand side of the app can be used to navigate through the various pages. The "Introduction" tab contains two pages.

The first is an introduction to clinical trials which contains background information on the phases of clinical trials and then goes into more detail specifically about Phase II single-arm trials. This also serves to provide some set-up and context about when a beta-binomial conjugate analysis might be used. The second page is about Bayesian analysis, which is used to introduce Bayes' theorem and concepts like priors and likelihood. Whilst both of these pages are not fully detailed or insightful renditions of the topics, they do provide the basic information required to use the app and create ETPs. Additionally, the content on these pages should be accessible for almost anybody regardless of their experience with clinical trials or statistics.

The next tab in the navigation bar is labelled "Beta-Binomial Designs" which is also split into two further pages. The first page is about the basics of a Beta-Binomial conjugate analysis. Here we have a brief introduction to conjugate models and more specifically a Beta-Binomial analysis. To illustrate how it works we use some interactive elements. We start by splitting the conjugate model into its three main components the prior probability distribution, the likelihood and the posterior probability distribution. In each of these sections we detail what these components are how they are presented mathematically and how they can be interpreted. Additionally, in each section we also include a visual representation of each component along with the ability to modify the plots. All of these plots between each of the sections interact with each other. So, for the prior section we default by showing a Beta(1,1) prior distribution. The likelihood plot shows the likelihood function which is default set to 8 responses from 15 patients. Finally, the posterior section shows the posterior probability distribution based on the prior and likelihood sections, the default here is a Beta(9, 8) based on a Beta(1,1) prior and having 8 responses for 15 patients. Additionally, we also introduce the idea of decision criteria in the posterior probability section and on the plot we visualise the cut-off for a GO/No

GO decision. The default decision criteria specified here is $P(\theta \geq 40\%) \geq 0.8$ which based on the other defaults results in a GO decision. As well as all of these plots interacting with each other we have specified controls such that the user can change any of the parameters, data or decision rules used to generate these plots. Changes to any of these inputs results in all the corresponding plots being updated. As such the user can experiment and investigate the affect of changing any of the default specifications. For clarity, we include text statements on the interpretation that can be made from the posterior which also update automatically based on the details specified.

The second page in this tab allows the user to run a practice trial using a Beta-Binomial conjugate analysis. The previous page shows the mechanics of how the design works but that is based on knowing the final number of patients and responses. The top left box has multiple tabs containing the instructions, decision criteria and controls that are being used. For consistency we use the same decision criteria and priors as before. We also specify a true response rate, defaulted at 50% which is the response rate we sample patients from. Again, All of these specifications can be changed by the user. Using the "Add patient" tab the user has the option to add or remove patients from the trial. A slider between 1-10 allows the user to select how many patients they want to add and then by clicking the add patients button they can add that many patients, this can then be repeated many times. Once patients are added you will see a plot in the top right box that shows a circle for each patient, coloured green or red to indicate if they had a response or no response respectively. Patients responses are determined based on the true response rate specified earlier. The bottom left box will also produce a plot showing the posterior distribution based on the number of patients added and the number of responses. The plot has a checkbox option to display the decision criteria which will show whether or not based on the data generated and decision rule

if there would be a GO decision. Then the bottom right box gives statements on the interpretation of the posterior probability distribution and decision rules in the "Analysis" tab. The "Summary Estimates" tab then provides summary estimates from the posterior with the option of including these in the plot.

This page was developed as more of a demonstration tool which can be used to illustrate how decisions we make may end up being incorrect based on data we observe or the timing of the decision. Users have the ability to add multiple sets of patients in the form of cohorts and can see what the decision or results from the trial would be based on the data they generate. As it is based on a true response rate of 50% they may get lucky and get enough responses from 10 patients to make a GO decision but if they were to re-run this they may get a different number of responses and hence a different result. Users then can see the affect of adding more patients or changing the decision rules or the true response rate.

Finally, we have an "App Details" tab which contains two pages. The first one includes a list of references with external links to more material on topics covered by the app. We also, reference the R packages that were used to make the app and link to their respective CRAN pages. We also have a page for version history which details what was added, changed or updated for each version of the app. A link to the code used to create the app is also hosted on GitHub and linked for users to see.

All of these additional features allow us to utilise this app as an effective teaching tool as well. There is some evidence that suggests the use of apps helps students learn more effectively [108]. R shiny allows us to develop these custom features without any relevant knowledge of languages such as Java or HTML.

5.5 Discussion

In this chapter we introduced the idea of Efficacy Transition Pathways. These were initially thought of as an extension to Dose Transition Pathways which are used as a visualisation tool to communicate decisions made in dose-finding trials. ETPs act in a similar manner and serve as a tool to better illustrate and communicate decisions made in single arm Phase II trials that utilise a Beta-Binomial conjugate analysis. We detail the basics of these analyses and how posterior probability of success can be used to make decisions during a trial at the interim analysis stage. Just like in dose-finding, depending on the number of outcomes we observe, i.e. number of responses, we can know ahead of time the decisions that will be made. Depending on the complexity of the design and decision rules the number of different scenarios can be difficult to comprehend. So, ETPs help visualise these scenarios. These can be beneficial for not only statisticians but other investigators involved with the design and conduct of the study.

ETPs consist of an array of cells for each cohort in a trial, with each cell containing key information pertinent to a specific number of responses being observed in the total cohort size. Constructing ETPs requires numerous calculations to be run including determining PPoS as well as Bayesian estimates and credible intervals from the posterior distribution. These then need to be evaluated against the decision criteria to determine what the outcome of the trial would be based on the data that cell represents. All this then needs to be constructed into a larger image with multiple cells. A benefit of this approach is that the cells can be tailored to include information or other details that is felt relevant to portray. However, this whole process can be time consuming especially during the design stages of a trial where specifications and decision rules may constantly be changed and iterated upon. This was further illustrated in

the three examples we gave of trials where ETPs had been implemented and used. Each of these trials could be considered as using complex and innovative designs which involve multiple treatment arms with various time-points for analyses and different decision rules. Throughout the design of these trials multiple different ETPs were having to be produced in order to facilitate discussions and showcase the design of the trials.

This is one of the issues with ETPs that motivated us to create an app and some software that would automatically produce these plots. The other motivation came from issues surrounding the introduction of new methodologies. Often times when a new methodology or tool is introduced it takes a long amount of time before it gets picked up and used by those other than the original creators of the methodology. This is often due to multiple factors such as lack of awareness about the new methodology or lack of useable code or relevant materials. From the perspective of statisticians working on and applying methodology, keeping up-to-date with all the latest innovations is often unfeasible. Similarly, if you do come across a new methodology where there is access to code, trying to implement the methods become a time-consuming task so we may default to standard or typical practice even if it is sub-optimal. Additionally, they may also need to take on the role of explaining the new methodology to non-statisticians involved in the oversight and management of the trial. This burden should fall on those developing the methodology if they want it to be used more frequently. In order to facilitate this for ETPs we created a simple R function along with a shiny app to make ETPs more accessible and easy to explain.

The primary feature of the app is to produce ETPs, we achieve this by having a simple to use interface which allows users to produce ETPs just by clicking a few buttons. What should be stated here is that currently there is limited flexibility with adjusting the ETPs. For example, the current app and function

only allows for fixed cohort sizes and one set of decision rules. If you are working in a rare disease setting, recruitment may be fragmented so your trial may employ flexible cohort sizes and as such ETPs do not allow for that but could easily be modified to have a different number of cells on each row for each cohort. Similarly, you may want to consider multiple decision rules or have more complicated rules during your interim analyses. For example, if your PPoS is between a specific range say 5% and 15% you may want to consider stopping but if it is definitely less than that you would want to stop and if its more you would want to continue. Whilst not technically difficult to account for that in our code or function, these are more niche scenarios that may not be too common. As such the basic functionality of the app can still be used to investigate these things. Future versions of the code and app will be updated to allow for more of these options.

The app also serves the secondary feature of acting as an educational tool. We include materials and features which will show people what ETPs are and how they are used as well as the very basic concepts and ideas in a beta-binomial conjugate analysis. This makes uses of R shiny interactivity features and is a different method to introduce people to a new methodology compared to something just like a publication. There exists resources such as the PANDA toolkit [109] which aim to educate trialists on adaptive and novel designs however as of yet they do not cover early phase trials. As such our app could be considered an introduction to some of these ideas and could also be expanded in the future to cover other adaptive designs in early phase trials.

The material and content included, in our app, has been produced and reviewed by statisticians and we feel it should be widely accessible. As such we may pilot using the app for teaching purposes and show the contents to people of a non-statistical background to determine if it is appropriate. Some of the features such as the page that lets you run a practice clinical trial may be best

utilised by a statistician trying to explain decision-making concepts in these trials. By having control over the true objective response rate you can show how by having small numbers you are prone to making the wrong decisions or if you have strict decisions rules you may need a lot of patients to obtain GO decision.

Away from ETPs, additions could be made to the app to add additional features such as options to run simulations. The 'run a clinical trial' page is essentially manually running one iteration of a simulation. This would also be an additional draw to use the app. Whilst simulations for a Beta-Binomial conjugate analysis are not difficult to run by having an app or a tool that does these for you could be beneficial. R shiny could be utilised to automatically produce graphics and summary tables then the design parameters that you specify could also feed in to produce ETPs, this could all then be summarised on the web page and printed off as a pdf. This may make it so more users are inclined to use the app and thus ETPs.

Our implementation of ETPs can also be extended to cover a larger outcome space. Currently we only consider GO or No GO outcomes, but it is possible to have an indeterminate outcome or a result in-between good and bad. In this instance there may not be enough evidence to declare the treatment a success but also not enough evidence to definitively say the treatment does not work. In order to account for these different outcomes we would need to provide additional decisions rules. The Lalonde framework [97], is an example of this which employs three different rules for each of their Go/Pause/Stop criteria. The flexibility of ETPs means that they could still be used in these more complex scenarios with multiple decision rules. This could be achieved just by using more colours to represent more outcomes.

Overall, we believe ETPs to be an effective tool in detailing decision-making for these types of trials. We have shown that with ETPs they can be used to

help iterate on the design of the trial and communicate the decision making with non-statisticians. To avoid many of the pitfalls of new methodology that never gets used, we have created code and an app which is publicly available that will easily produce these plots and act as an educational tool and thereby benefit the clinical trial research community.

Chapter 6

Summary and Conclusions

Throughout this thesis, we have explored various methods relating to early phase trials and dose-finding methodologies and investigated how they could be applied in practical contexts through illustrative examples and real clinical trials. In this concluding chapter, we summarise our key points.

We began in Chapter 2 by detailing the innovative PO-TITE-CRM design [29], [45] and our experiences implementing the design, which we believe is the first application of its kind. Partial ordering was originally hypothesised for use in investigations of escalation in combination of multiple-agents. Depending on the doses selected, it is possible in these scenarios the monotonicity assumption no longer holds. However our implementation, whilst also in a trial investigating combination treatment, observed partial ordering due to the dosing schedule of one of the treatments. This highlights the fact that this issue can also arise in multiple settings and more generally, methodology can still be implemented in scenarios outside of its original purview.

We showcased the performance of the design via simulations and then contrasted that with potential alternative designs. Even with the additional complexity of the design the operating characteristics were comparable across the variety of alternative options. The caveat was that some of those alternatives would have to make concessions and assumptions about the dose-levels we

were investigating. Without this methodology answering the relevant clinical question of the trial would become difficult. The doses selected for investigation were deemed clinically relevant and if we could not resolve the issue around the uncertainty of the order of the doses different doses would have had to been selected.

Whilst for the development of a new drug typically the maximum tolerated dose (MTD) is of interest, for repurposed treatments or combination treatments there may be additional interest in also understanding the optimal dosing schedule. Whilst this was not one of the main aims of the ADePT-DDR trial, it was of interest. An understanding of the best dosing schedule would then feed into the decision for what the final dose (dose-level 3) would eventually become based on the aggregate data.

If biologically plausible that the same overall dose administered across a shorter duration is more or less toxic when administered over a longer duration then time window for administration may be more important to consider than overall dose. For example, if it is understood that 400mg of a specific drug over five days is more toxic than 400mg over 20 days, it does not always follow then that by increasing the overall dose we may be increasing toxicity. So, 401mg over 20 days may still be less toxic than 400mg over five. Obviously this is an extreme example but it could still be clinically relevant in some areas and maybe either warrant further research. The partial ordering approach is one way to solve this issue but their could potentially be more efficient methods.

As the problem and issue of partial ordering is fairly unique there are not many approaches to dealing with it or accounts of practical applications. This may not be surprising considering it was only introduced in 2013 and it still shows there is somewhat of a lag between methodology being developed and implemented. One barrier we experienced was the limited availability of software. Along with the original methodology Nolan et al. [49], [50] developed

R functions for the implementation of the PO-CRM but not its time-to-event counterpart. However, having this as a basis to work from was crucial in our success at implementing the design. I further developed the existing code to incorporate the TITE element which still took a bit of time given the available code. From an academic perspective finding the resources in terms of time and statistical expertise may also be another barrier especially during initial design stages of a trial where there is limited funding.

Our work also has the added benefit of validating the original PO-TITE-CRM design and confirming the results found in that paper. Whilst we did not directly replicate the exact work presented there we have reached similar conclusions. Hopefully, our account and experiences implementing this methodology will help others to do the same. We have shared this work through a poster presentation at ICTMC 2022 [110] and also with a publication currently in pre-print. The ADePT-DDR trial opened in August 2021 and is currently recruiting to its third cohort of patients. Due to practical and logistical issues some aspects of the design have been altered however, the underlying methodology used in the trial still remains the same.

In Chapter 3 we explored the development of our own methodology which is specifically an extension we made to the Wages and Tait design [30]. This design is considered an adaptive or seamless phase I/II design as it uses both toxicity and efficacy outcomes to obtain an optimal biological dose (OBD). However, just by including efficacy outcomes does not mean we fully eradicate the need to conduct a randomised phase II study. We sought, through our design modification, to address this and have a trial design capable of conducting dose-finding and making direct comparisons to a control arm.

By leveraging the adaptive randomisation mechanism we were able to force the design to allocate patients to a control arm. We demonstrated through simulations that the design worked as intended. A reasonable rate of selection for

the OBD or "good" dose-levels was achieved along with allocating a sufficient number of patients to a control arm. By contrasting our design to potential alternatives we also showed there is minimal loss in terms of efficiency when using this modification. However, when it came to comparing the control arm to the OBD our results showed that we would be extremely underpowered unless there was a particularly large effect size. In order to improve this we would need to increase the overall sample size of the study or add expansion cohorts for both the OBD dose and control dose.

We identify that the most common situation in which it would be appropriate to implement this design is when investigating a new treatment in combination with some form of standard of care. Here there should be some understanding on the efficacy and toxicity profile of the standard of care treatment and this design would allow you to assess if there are any benefits to adding a new intervention to that standard of care. An additional limitation of this work is that we only explored this design modification using one exemplar trial. Therefore, our results and overall conclusions of the design may have slight variations under a different example.

There may also be issues with this design that we have not considered and would only become apparent when implementing the design. Simulations showed adequate allocation of patients to the control arm however, in practice we only run the trial once not 10,000 times so perhaps by chance we could end up with very few or limited patients in the control arm. So, we may want to consider changing our fixed rate of randomisation to control to also be adaptive dependent on the number of patients already in the control arm. Similarly, more work could be done to look at different randomisation mechanisms to allocate dose-levels.

Overall it may still be more efficient to set up a trial in this manner rather than having an independent phase I trial into an independent phase II trial.

This may be the future of trial designs where they are fully seamless and adaptive going through all the phases aiming to answer multiple questions with multiple decision points. You could imagine using our RtC-WT design to determine an OBD then potentially expand recruitment to confirm if there is enough of an efficacy signal to prompt a 'GO' decision for a fully randomised phase III trial. If this was all incorporated into one trial you could save resources in terms of time and money. There may also be efficiency gains in directly borrowing information from patients within the same trial from previous phases.

This work was motivated by a few potential trials being designed at the University of Birmingham's Cancer Research UK Clinical Trials Unit. Unfortunately, none of those trials came to fruition due to other circumstances, so the methodology itself is yet to be implemented.

Chapter 4 looked at applying dose transition pathways (DTPs) to trials using time-to-event dose-finding methodology. The original paper by Yap et al. [31] touches on the issue of applying DTPs to designs like the TITE-CRM. The authors briefly discussed that projected DTPs can differ dependent on how information was available for each dose decision and that it would be difficult to map out decisions in advance. Our work looked into this issue deeper to detail the exact issues and to see if there were any potential solutions.

This work was also partly motivated by the ADePT-DDR trial as well. Due to the PO-TITE-CRM methodology and the long observation periods there was interest in mapping out decisions ahead of time to facilitate faster decision making and better communicate the decision that would be made throughout the trial. It quickly became apparent due to the length of the observation window (52 weeks post treatment) this would be unfeasible.

Here we present the idea of introducing categories of combined follow-up

time for patients in the same cohort at the same dose-level. By grouping together the follow-up times of each patient in a cohort we can then apply the TITE-CRM model for each combination of time in order to figure out what the dose decisions would be for each. Based on this we could then see how much information or combined follow-up was required to make specific dose decisions. During this process we made an interesting observation that depending on how the combined follow-up time was split between patients a different dose-decision could be made. We clarified that this was due to the way in which the likelihood was calculated and that, at least for a linear weight function, patients weights are linear at the individual level but not at the cohort level. This could be a potential area for further research by investigating how the likelihood calculation is impacted by different weight functions and what impact that has on the dose recommendations being made.

The other main issue in presenting TITE-DTPs comes from the number of possible outcomes that need to be mapped out. Even in our simple example of just a five-week observation window, mapping out all the potential variations of outcomes for six patients results in more outcomes then seconds since the universe began. Our suggested solution to this was to limit the pathways that were presented just to be for the next cohort assuming you had all the available data for all previous cohorts. This still does not help for trials with much longer observation windows. In these scenarios it would be suggested that outcomes be limited to plausible scenarios. For instance it is unlikely that you would recruit a cohort on one day and need to immediately make a dose-decision for a new patient on the next day. So, the number of outcomes could be simplified to just look at combined follow-up times which were appropriate or more realistic. Though, if the observation window is long enough this still may not be sufficient in reducing the number of outcomes.

Anecdotally speaking DTPs appear to be one of best tools for communicating methodology in early phase trials. Especially when it comes to presenting the different decisions that can be made during a trial. I have received compliments from presenting DTPs whether it be patient representatives or independent clinicians sitting on data monitoring committees or representatives from pharma partners when setting up a trial. So, it would seem beneficial to try and extend DTPs or the concept of them for presenting decisions as far as possible in trials.

Staying with that idea, Chapter 5 introduces efficacy transition pathways (ETPs), a visualisation tool inspired by DTPs but for use in some types of more complex phase II trials. Much like DTPs they aim to better illustrate and communicate decisions made in these types of trials. We detail how ETPs can be constructed and how they can be used to assist during the running of a trial and the development of the trials design and stopping rules.

Through three motivating illustrative example trials (see Appendix A) we detailed how they have already begun to be implemented and used in practice. All three trials present ETPs in a similar manner however they illustrate the potential applications in different settings. What became apparent from these implementations is that whilst theoretically simple to produce, ETPs were often time consuming to put together. The calculations required for presentation in the ETPs could be automated by constructing the plot could become laborious if other elements of the trial design were altered. This motivated us to develop software in order to allow for automatic generation of ETPs.

We began by developing a simple function in R that could generate these plots. Around that we built a web based application using shiny. The app makes producing these plots even easier as it does not require the user to be familiar with any particular software package. All the inputs required for the function are presented in the app through various widgets. From the app there

is the ability to download the ETP as an image. Additionally the user can also download a copy of the data used to generate the ETP. Other benefits of the app also allow us to include some interactivity with the ETP. By clicking on the plot the user can see additional information for the specific cell they clicked on. A larger version of the cell is produced, which is useful in cases where the ETP is rather large with lots of cells. A posterior distribution is also plotted based on the data in that cell showing other key information like credible intervals and the decision criteria.

We also developed additional pages on the app to introduce some of the fundamentals required to understand ETPs. This is done mainly through text and images. However, we include interactive elements to explain the basics of beta-binomial conjugate analyses. Users can interact and specify different priors, likelihoods and decision rules and see how interpretations from the posterior distribution change accordingly. There is also the facility to simulate a practice trial so users can get an idea of how a trial designed in this way could work. All this additional content was incorporated in a way that is hopefully approachable by people with limited knowledge on these topics. We still envisage that these pages could be used by statisticians as well to also help explain these concepts to others.

Even though the tools such as DTPs and ETPs are beneficial in and of themselves, having the ability to automatically produce them by clicking a few buttons could further enhance how they are used. During discussions with clinicians you could easily demonstrate how changes to the design or decision criteria would affect the decisions made through the ETPs. The added layer of interactivity through the app could also assist the statistician in better communicating the changes that are being suggested. By developing all these materials and making them easily accessible we hope it makes ETP simple to implement where appropriate.

Further research for work presented in this thesis would revolve around extending the methodologies presented in each of the chapters and investigating more practical applications. For the ADePT-DDR trial we designed work could be done to investigate, develop or extend alternative methodologies and compare how they perform. For our extension to the Wages and Tait design future work should look into the use of models with multiple parameters and different adaptive randomisation methods as these have been shown to yield better performance. TITE-DTPs could be explored further by investigating the use of different weight functions in more detail. Additionally, it would be beneficial to gain experience into how TITE-DTPs work in practice by implementing them into trials that are in development or ongoing. These case studies would provide information on how we could further improve TITE-DTPs, For ETPs further work can be done to improve functionality within the app to allow for more features such as running simulations. ETPs and the app could also be developed to work with other designs rather than just the Beta-Binomial design.

All the work presented in this thesis was motivated by the development of clinical trials at the University of Birmingham's Cancer Research UK Clinical Trials Unit. Novel methods are generally speaking very useful and valuable. In the context of dose-finding methodologies they can often times provide solutions to complex clinical questions without compromising efficiency or in some cases improve on it. Furthermore, the development of appropriate software and tools to facilitate decision making are equally important in advancing the implementation of these novel methods.

Appendix A

Examples implementing ETPs

The following sections contain examples of clinical trials that have implemented Efficacy Transition Pathways (ETPs). All of these trials were designed by statisticians at the Cancer Research UK Clinical Trials Unit (CRCTU) at the University of Birmingham.

A.1 MonoGerm

This trial was designed by Lucinda Billingham (LB) and Laura Kirton (LK). What is presented here is the design that was used as part of a grant application which has since been accepted. The trial is currently in the process of being set-up so may be subject to some design changes before it is open.

MonoGerm¹ is a Phase II trial investigating, in parallel, two single-agent chemotherapies (carboplatin or vinblastine) as monotherapy prior to standard of care radiotherapy in patients with intracranial germinoma. This trial utilises a Bayesian approach for analysis and decision making. A 'flip-flop' approach

¹A phase II trial of carboplatin or vinblastine monotherapy induction prior to radiotherapy for patients with intracranial germinoma

is used for recruitment [111]. Essentially, recruitment will begin to the carboplatin arm then once three patients have been recruited and considered evaluable for an interim analysis recruitment flips to the vinblastine arm. This process is then repeated by switching recruitment back and forth between the two arms till recruitment is complete.

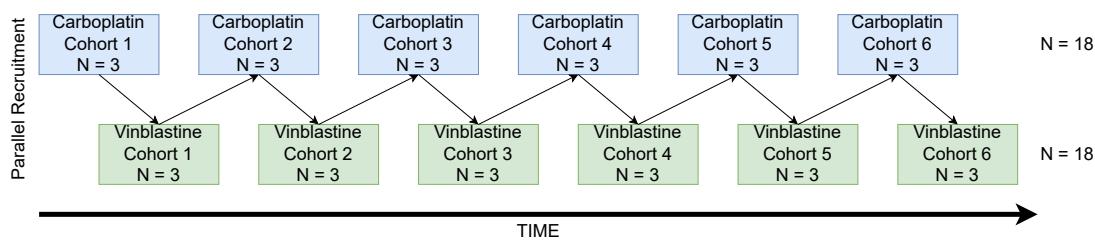
Typically intracranial germinoma is chemosensitive but it requires radiotherapy for a cure. This mainly affects a paediatric population. For patients with localised disease, standard of care involves three-drug chemotherapy consisting of ifosfamide, carboplatin and etoposide followed by subsequent radiotherapy. Treatment involving multiple chemotherapies allows for reduction in radiotherapy doses and fields. However, there is still an increased burden of treatment on patients which can cause both short and long-term harm. This can lead to patients experiencing multiple toxicities such as diabetes insipidus, myelosuppression, vomiting/diarrhoea, electrolyte disturbances, renal impairment, and elevation of liver enzymes.

The aim of the trial is to evaluate whether a single-agent chemotherapy (carboplatin or vinblastine) is non-inferior to the standard of care multi-drug chemotherapy for inducing complete response (CR), and is associated with reduced harm and improved quality of life. The primary outcome measure will be CR based on MRI scans at 6 and 12 weeks.

The trial will consist of two arms, one arm for carboplatin and one for vinblastine. There will be 36 patients recruited in total, 18 per arm. Patients will be enrolled in cohorts of three to each treatment-arm by the flip-flop design. This is illustrated in Figure A.1. Recruitment begins in the carboplatin arm and then once three patients have been recruited recruitment is paused and then begins in the vinblastine arm. Whilst recruitment is paused in the carboplatin arm an interim analysis will be performed for that first cohort. Here primary outcome data and key safety data will be assessed. Once recruitment to the first cohort

of the vinblastine arm is complete and the interim analysis for the first cohort of carboplatin patients is done, recruitment can begin for the second cohort in the carboplatin arm and the interim analysis for the first cohort of vinblastine patients can be conducted. This process is then repeated for the subsequent cohorts. This design allows for continuous enrolment and monitoring of the primary outcome.

FIGURE A.1: Flip-flop recruitment design in the MonoGerm trial.



A Bayesian approach is implemented to assess the true CR rates. Specifically, the experimental monotherapies will need to demonstrate non-inferiority to the standard of care by a clinically acceptable margin. Based on previous data it was determined that the minimum CR rate with standard of care is 30%. This value was taken as the non-inferiority margin. So, if the treatments have a CR rate $\geq 30\%$ they will be considered non-inferior.

CR rates for each treatment arm will be established from the posterior probability distribution generated using a beta-binomial conjugate analysis. A minimally informative Beta(1,1) prior will be used in combination with the data observed during the trial to produce the posterior. In terms of decision making the following rule for the final analysis was specified:

$$P(CR \geq 30\%) \geq 0.8 \quad (\text{A.1})$$

That is to say that if there is a high probability (≥ 0.8) that the true CR rate is $\geq 30\%$ there will be a GO decision, which in the context of this trial means the treatment arm would be deemed non-inferior. The minimum number of

observed CRs out of the 18 patients needed to warrant a GO decision is seven. If seven responses are observed the median Bayesian estimate of the CR rate would be 40% with a 0.82 probability that the true CR rate is $\geq 30\%$.

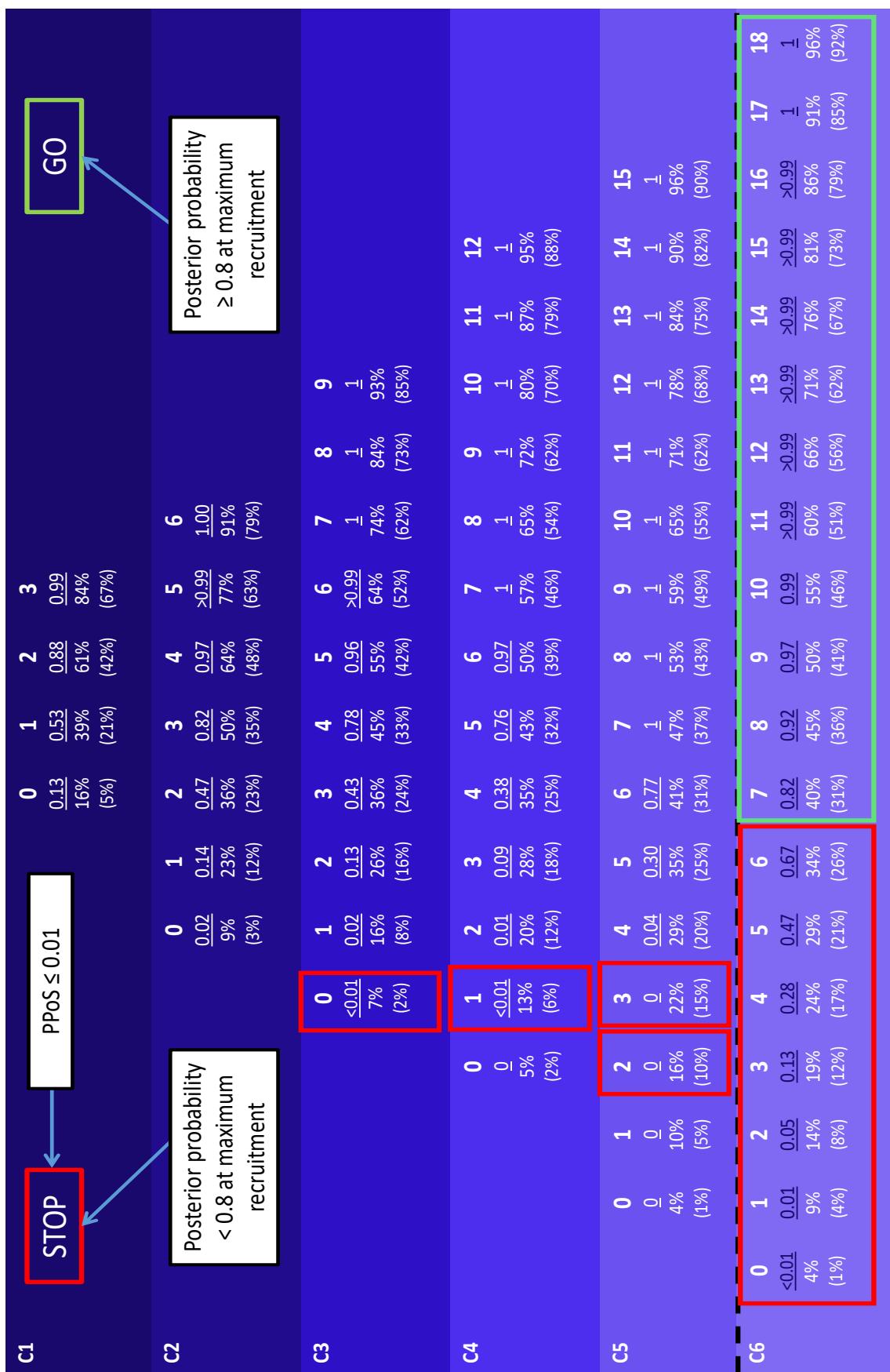
Stopping rules have also been implemented. At each interim analysis the predicted probability of success (PPoS) will be calculated. This is the probability that a GO decision would be made at the final analysis based on current data that has been accrued. Here the stopping rule is such that if the PPoS is less than 0.01 we would recommend stopping recruitment to that arm.

It is important to note that whilst the trial has two arms it is not designed with the intention of making comparisons between the two treatments. Rather the trial aims to find if there is enough evidence that one of these treatments provides a sufficient response rate to warrant a GO decision.

LK produced ETPs throughout the design of this trial. Figure A.2 shows the ETP for the design specified here. The ETPs produced here helped determine what data we wanted to present as well as how we structure the data in each of the cells. Here each cell shows the number of responses, the PPoS or the posterior probability that the response rate is $\geq 30\%$, the Bayesian estimate of CR rate and a lower one-sided 80% credible interval.

Calculations for these plots were conducted in STATA and the ETP was produced in Microsoft PowerPoint. One benefit of this approach is that the ETP can be easily customised and labelled. However, this meant that any changes in the design that affected the decision-making resulted in the ETP having to be updated manually. As a tool the ETP is effective at illustrating a final design but without the ability to easily generate them during the design of a trial they somewhat lose their purpose. This served as further motivation to produce some code or a tool that would allow for ETPs to be automatically created.

FIGURE A.2: ETP for the MonoGerm trial.



A.2 Glo-BNHL

This trial was designed by Lucinda Billingham (LB) with the assistance of Shanna Maycock (SM). During initial stages of the design Grace Holt (GH) also assisted with the implementation of ETPs. This trial is currently still in set-up so specific details of the trial may be subject to change.

The Glo-BNHL² study is a platform trial that aims to investigate the safety and effectiveness of novel treatments in children, adolescents and young adults with relapsed and/or refractory B-cell non-Hodgkin Lymphoma (r/r BNHL). This trial is an international collaboration and hopes to generate substantial evidence that could change practice in this rare cancer population.

Inclusion of novel agents into the platform will be determined by an international Trial Steering Committee (TSC). Currently the platform consists of three separate treatment arms, each focusing on a different novel agent in a distinct group of patients. Each treatment arm will be treated independently which allows for them to be analysed separately. There may also be separate eligibility criteria for each arm. Patients will be enrolled into any arm where they are eligible. The three arms that will be available at the start of the trial are as follows:

- Treatment Arm I: Bispecific Antibody (BsAb)
- Treatment Arm II: Antibody-Drug Conjugate (ADC) with standard chemotherapy
- Treatment Arm III: Chimeric Antigen Receptor (CAR) T-cells

This trial utilises an adaptive Bayesian design, which enables GO/No GO decisions specific to the distinct populations in each treatment arm. The Bayesian

²A Global Study of Novel Agents in Paediatric and Adolescent Relapsed and Refractory B-cell Non-Hodgkin Lymphoma

approach has many benefits here as it facilitates decision making with small sample sizes. Decisions will be made based on the estimate of the probability that a novel agent is clinically effective. The specific criteria will vary between each treatment arm. This design and approach also allows for continuous evaluation of each novel agent. Treatment arms may be removed if the treatment is shown to be ineffective based on the trial data. Additionally, treatment arms may also be added or amended in the future if recommended by the TSC.

Treatment arm I aims to estimate the clinical efficacy of BsAb treatment in patients with r/r BNHL in first (only one prior line of therapy) or subsequent relapse (more than one prior line of therapy). Due to this treatment arm I is split into two groups, Ia and Ib for patients with one relapse or more than one relapse respectively. These two subsequent arms will be recruited into and analysed separately. In terms of treatment these patients will receive odronextamab given as an intravenous infusion weekly for 12 weeks, then every two weeks until nine months, and every four weeks thereafter until progression or for a maximum of two years. The outcome measure for patients in treatment arms Ia and Ib is the occurrence of an objective response (OR) i.e. complete response (CR) or partial response (PR) after 12 weeks of treatment assessed by Independent Central Review. However, for interim analyses local response assessments will be used.

Treatment arm II aims to estimate the clinical efficacy of ADC treatment with modified R-ICE (rituximab, ifosfamide, carboplatin, etoposide and dexamethasone) chemotherapy in patients with r/r B-NHL in first or subsequent relapse. Patients will receive loncastuximab tesirine given as a 30 minute intravenous infusion with each cycle of modified R-ICE for a maximum of three cycles. Here the outcome measure is occurrence of CR within a maximum of three cycles of treatment.

Treatment arm III aims to estimate the efficacy of CAR T-cell therapy in r/r

B-NHL patients who have CAR T-cell product available. The specific treatments patients will receive is yet to be defined. The outcome measure is the occurrence of OR following CAR T-cell infusion.

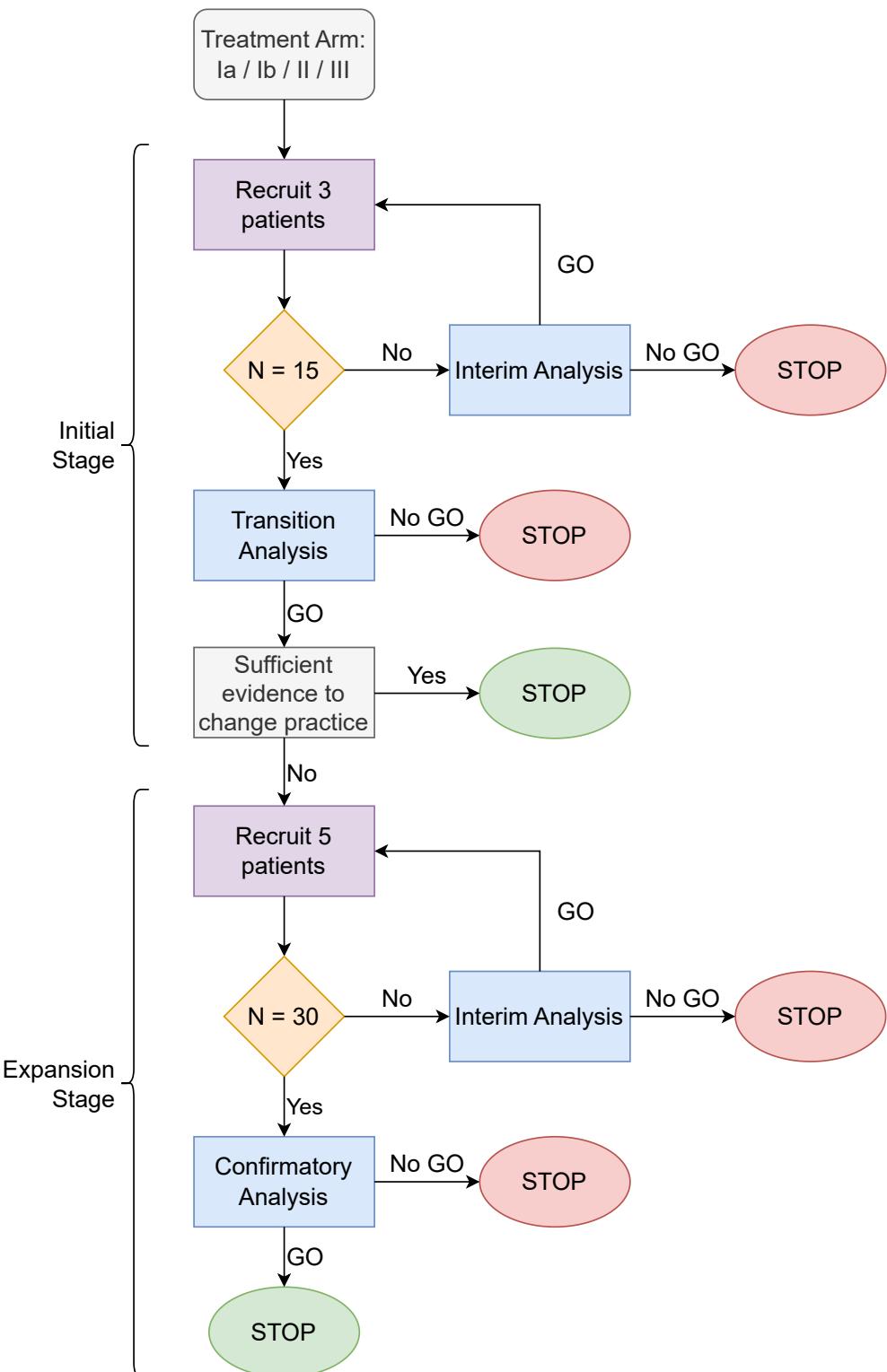
Each treatment arm and subsequent treatment arm (i.e. Ia and Ib) will aim to recruit 15 evaluable patients during the initial stage. Once this recruitment is complete a transition analysis is performed leading to three possible outcomes. If the analysis results in a No GO decision recruitment to that treatment arm will stop. If the analysis result is a GO decision there are two options either there is sufficient evidence to change practice so the trial will stop recruiting or this will trigger an expansion stage in which a further 15 patients will be recruited. Following the expansion stage a confirmatory analysis will be conducted on all 30 evaluable patients.

Interim analyses will be conducted after every three patients during the initial stage and after every five patients during the expansion stage. There will also be the option to stop recruitment to a treatment arm based on the data observed in the interim analysis. Figure A.3 shows a flowchart of the decision making process for each arm in the trial.

A beta-binomial conjugate analysis will be conducted for each treatment arm. Observed trial data will be combined with a minimally informative Beta(1,1) prior to produce a posterior probability distribution for the treatment effect θ , which represents either the OR/CR rate dependent on treatment arm. The posterior probability distribution is then used to inform decision making. GO/No Go decision criteria are specified separately for each treatment arm.

For treatment arm Ia the decision criteria at the transition analysis is $P(\theta > 40\%) \geq 0.8$. So, if the true OR rate was greater than 40% with a probability of at least 0.8 based on the data collected in the trial there would be a GO decision. This corresponds to observing at least eight responses out of 15. For the confirmatory analysis a GO decision is made if $P(\theta > 40\%) \geq 0.95$. At the

FIGURE A.3: Flowchart of the decision making process in Glo-BNHL.



final analysis we would need the probability that the true OR rate was greater than 40% to be at least 0.95. Here a GO decision would only be made if 17 out of 30 patients had an OR. Table A.1 details the criteria for each treatment arm which can be interpreted in a similar manner.

TABLE A.1: Summary of decision criteria for Glo-BNHL.

Treatment Arm	Transition Analysis		Confirmatory Analysis	
	Decision Criteria	Min No. Responses for GO	Decision Criteria	Min No. Responses for GO
Ia	$P(\theta >40\%) \geq 0.8$	8/15	$P(\theta >40\%) \geq 0.95$	17/30
Ib	$P(\theta >10\%) \geq 0.8$	3/15	$P(\theta >10\%) \geq 0.95$	6/30
II	$P(\theta >20\%) \geq 0.8$	5/15	$P(\theta >20\%) \geq 0.95$	10/30
III	$P(\theta >10\%) \geq 0.8$	3/15	$P(\theta >10\%) \geq 0.95$	6/30

For the interim analyses there are separate stopping rules. These are the same across the treatment arms but differ in the initial stage compared to the expansion stage. During the initial stage, the predicted probability of success at the transition analysis (PPoSt) is calculated and decisions are made based on the following criteria:

1. $PPoSt < 0.01$ - recommend stopping for futility
2. $0.01 \leq PPoSt < 0.05$ - consider stopping for futility
3. $0.05 \leq PPoSt < 0.15$ - consider whether sufficient benefit in continuing
4. $PPoSt \geq 0.15$ - recommend continuing

So, if the probability of success at the transition analysis is less than 1% the recommendation would be to stop recruitment to the treatment arm and if it was greater than or equal to 15% the recommendation would be to continue recruitment. If PPoSt is between 1-5% or 5-15% stopping should be also be considered for either futility or if there sufficient benefit in continuing respectively.

At the expansion stage, the predicted probability of success at the confirmatory analysis is used (PPoSc). If this is below 10% ($PPoSc < 0.1$) the recommendation would be to stop recruitment to that treatment arm due to futility.

It should be noted that these decision rules are only recommendations and the independent data monitoring committee (DMC) will make decisions based on not only primary outcomes but secondary outcomes, recruitment and safety data.

For this trial ETPs were produced separately for each treatment arm and for each stage. This resulted in eight ETPs, one for the initial stage showing the outcome of the transition analysis and one for the expansion stage showing the outcome of the confirmatory analysis for each treatment arm (Ia, Ib, II, III). ETPs were utilised throughout the design of the trial with initial versions originally created by GH using STATA and Microsoft PowerPoint. These were then further developed by SM who performed calculations in R but still utilised PowerPoint to create the ETPs. Figure A.4 and A.5 shows the ETPs for the initial and expansion stage of treatment arm Ia respectively. These ETPs created by SM helped us determine how our function and app visually presented ETPs and we utilised a similar colour scheme.

Similar to MonoGerm the process of creating these ETPs in PowerPoint can be time consuming. This is even more of an issue in the Glo-BNHL study due to the multiple treatment arms and stages. Additionally, this trial highlighted another issue when you have multiple statisticians working on a trial who use different software packages. In this case it would mean work would have to be recreated in R and STATA to conduct the calculations required for ETPs. This served as further motivation for the development of an application which requires no specific stats software knowledge. As statisticians would easily be able to recreate ETPs. SM went on to further extend the function we developed to automatically generate ETPs specific to Glo-BNHL.

FIGURE A.4: ETP for the initial stage of treatment arm Ia in Glo-BNHL.

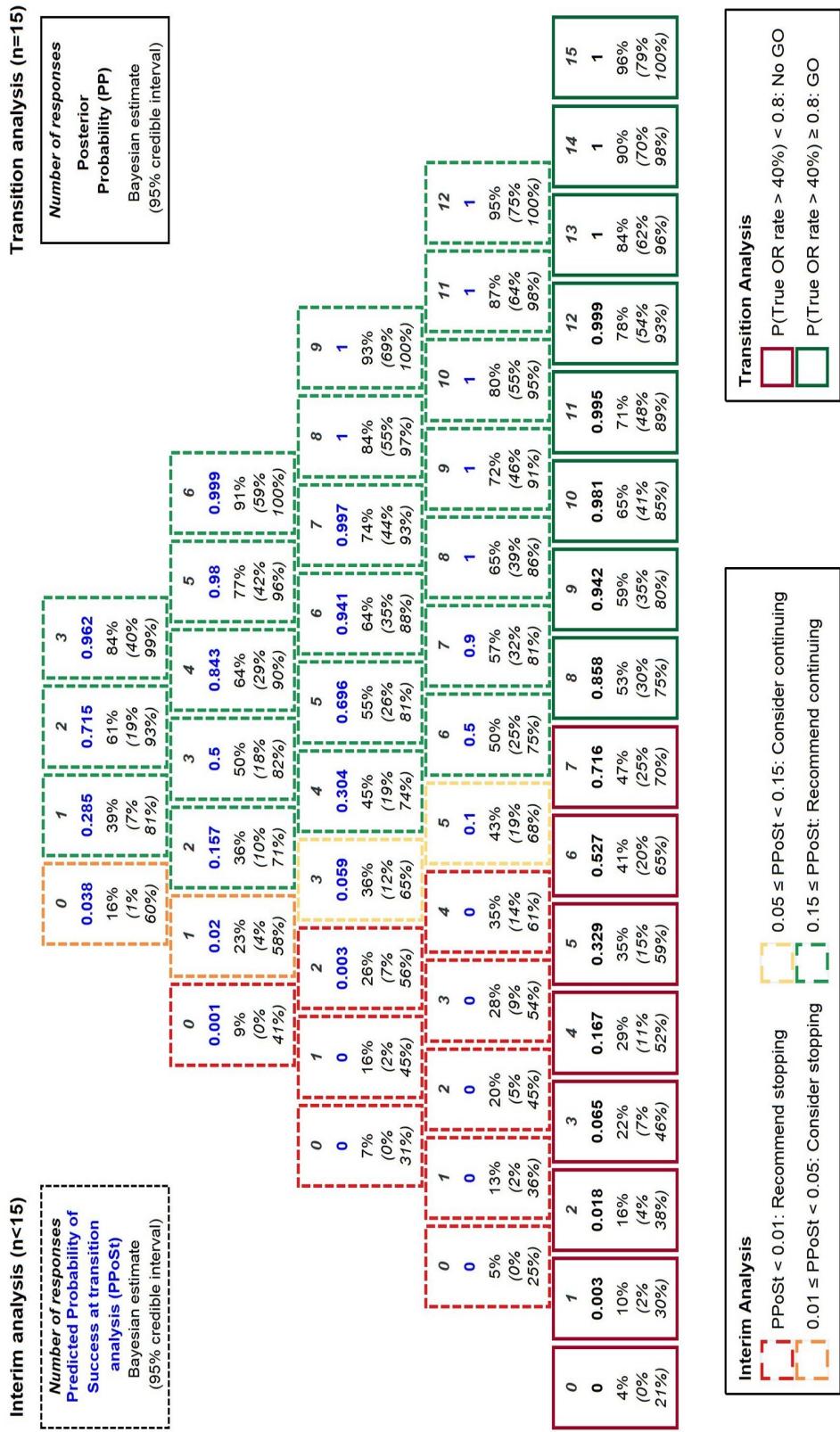
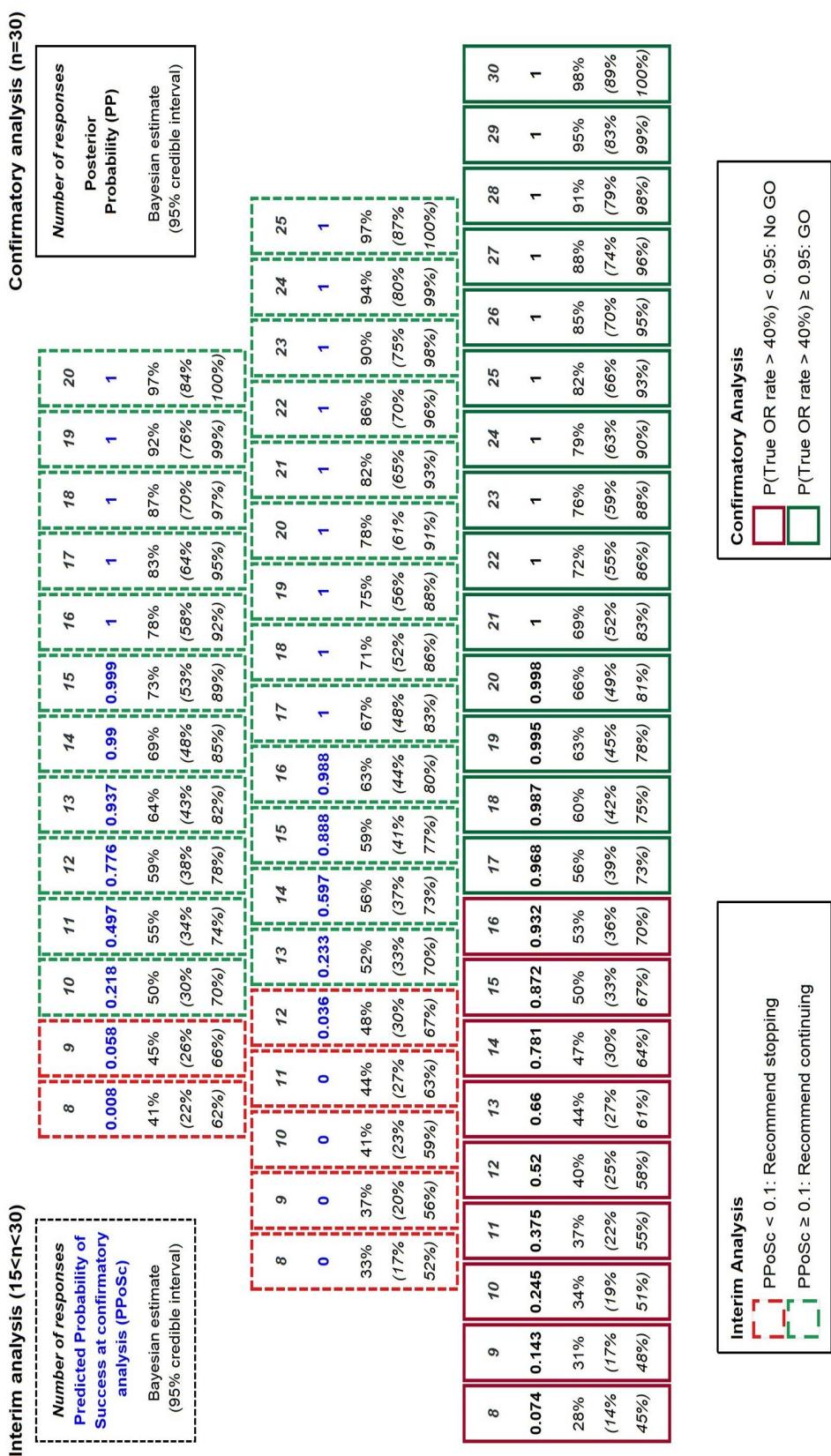


FIGURE A.5: ETP for the expansion stage of treatment arm Ia in Glo-BNHL.



A.3 DETERMINE

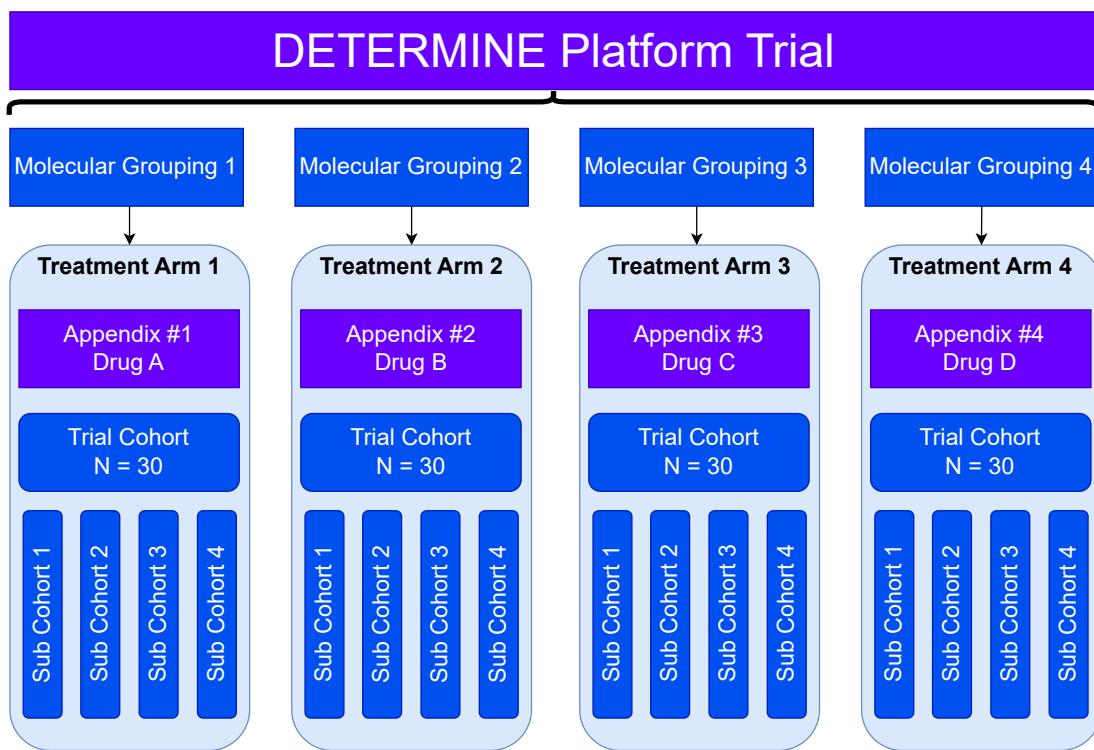
DETERMINE³ is a trial that was also designed by Lucinda Billingham (LB). It is an umbrella-basket platform trial with multiple treatment arms running in parallel. The trial aims to evaluate the efficacy of targeted therapies in rare cancers with actionable genomic alterations, including common cancers with rare actionable alterations. This trial is currently open to recruitment and may be expanded in the future as more drugs are bought onto the platform.

DETERMINE will recruit patients of all ages including, paediatric, TYA (teenage and young adult) and adults, who have rare tumours that contain an actionable genetic alteration that can be targeted therapeutically. The genetic alteration must have been identified previously from a tissue biopsy or ctDNA (circulating tumour DNA). Patients will then be stratified into molecular groups based on their tumour profile and allocated to the most suitable treatment arm by the Molecular Tumour Board (MTB). The umbrella part of the design consists of multiple non-randomised treatment arms, each evaluating a licensed targeted anti-cancer drug or drug combination in a specific molecularly-defined group of patients. Each molecularly-defined group allocated to a specific treatment arm will contain multiple baskets of different tumour types, age groups and molecular subtypes. This is visualised in Figure A.6.

A main trial cohort of 30 evaluable patients will be recruited into each treatment arm. This will include patients with different tumour types, ages and molecular subtypes. If specific subgroups within this main cohort are experiencing significant benefit from treatment sub-cohorts will be formed to investigate treatment effectiveness in these subgroups. It is possible for each treatment arm to have multiple sub-cohorts and they can recruit in parallel to

³Determining Extended Therapeutic indications for Existing drugs in Rare Molecularly-defined Indications using a National Evaluation platform trial

FIGURE A.6: Umbrella-basket platform trial design in DETERMINE.



the main cohort. Each sub-cohort will be subject to the same statistical analysis and will also aim to recruit 30 patients. Currently there are five treatment arms, details of these are provided in Table A.2.

TABLE A.2: Current treatment arms in DETERMINE

Treatment Arm	IMP(s)	Molecular Grouping	Route and Formulation
1	Alectinib	ALK gene fusion positive solid tumours	Oral capsules
2	Atezolizumab	Solid tumours with high tumour mutational burden (TMB) or microsatellite instability-high (MSI-high) or proven constitutional mismatch repair deficiency (CMMRD) disposition	Intravenous (IV) infusion
3	Entrectinib	NTRK or ROS1 gene fusion positive solid tumours	Oral capsules; Dosing depends on body surface area (BSA)
4	Trastuzumab in combination with pertuzumab	Solid tumours with HER2 amplification or mutations	IV infusion
5	Vemurafenib in combination with cobimetinib	Solid tumours with BRAF V600 mutations	Oral; 960 mg tablets

Co-primary outcomes of objective response (OR) and durable clinical benefit (DCB) will be used to assess efficacy of the treatment in each of the cohorts

in each molecular grouping. These are both classified as binary variables. OR is defined dependent on specific disease criteria. DCB is defined as the absence of disease progression for at least 24 weeks from the start of trial treatment, this will also be measured based on disease specific criteria. Patients will receive treatment based on the license schedule for the drug in their treatment arm. Patients continue treatment until either they reach progressive disease (PD), unacceptable adverse events or withdraw from the trial.

All outcomes in each treatment arm, cohort and sub cohort will be analysed using a Beta-Binomial conjugate analysis. Posterior probability distributions for OR and DCB will be generated using minimally informative priors, Beta(0.1,0.9) and data that is collected on the trial. However decision making will be done using the BOP-2 design [112], [113]. This design makes GO or No GO decisions by assessing posterior probabilities for a set of outcomes, which are optimised to either maximise power or minimise the number of patients. The design also allows for the control of type I and type II error rates. In DETERMINE, for each co-primary outcome, a rate of 10% or less would represent a treatment effect that is not clinically relevant and a rate of 30% would represent a clinically meaningful treatment effect. These can be thought of as a null and alternative hypothesis. The use of this design was made possible due to the development of web applications by the authors at the MD Anderson Cancer Centre [103].

Interim analyses will be conducted throughout the trial, however formal decision making will first be conducted after 10 evaluable patients have been recruited and then every 5 patients from that time-point in each cohort/sub-cohort. If probability that both the true OR and DCB rates are lower than the critical threshold of 10% the trial would recommend stopping. The design is optimised to minimise the type II error i.e. to minimise the probability of not rejecting the null hypothesis when treatment is effective. This is done whilst

controlling the type I error rate at 0.1. Under the BOP-2 design this provides the following stopping boundaries of $0/10$, $\leq 1/15$, $\leq 2/20$, $\leq 3/25$ at each of the planned analyses at 10, 15, 20 and 25 patients. For the final analysis at 30 patients there would be a GO decision if $\geq 6/30$ patients had either a OR or DCB.

The ETPs we have shown so far all have been based on a Beta-Binomial conjugate analysis with decisions made using PPoS and the posterior distribution. However, they can also be utilised here. LB created the ETP shown in Figure A.7. Here we can see each cell still pertains to a specific number of responses out of a certain number of patients. Bayesian estimates, credible intervals and posterior probabilities of the treatment effect being greater than our null and alternative hypothesis. The cells have then been colour coded based on the decision criteria elicited from the BOP-2 design. This trial also shows that ETPs are a flexible tool that can also be applied to trials not explicitly using a Beta-Binomial conjugate analysis for decision making.

FIGURE A.7: ETP for the DETERMINE trial.

Each cell contains: number of DCB/OR, Bayesian estimate of DCB/OR rate, 95% CrI, PP>10%, PP>30%												
n=5	0	1	2	3	4	5	6	7	8	9	10	11
	0	15	33	52	70	89	56	65	74	83	92	101
	0.16	1.55	6.73	16.86	30.95	50.99	>0.99	>0.99	>0.99	>0.99	>0.99	>0.99
	0.050	0.641	0.933	0.993								
	0.006	0.198	0.57	0.858	0.975	0.998						
n=10	0	1	2	3	4	5	6	7	8	9	10	11
	0	8	17	27	36	46	56	65	74	83	92	101
	0.9	0.32	3.46	7.57	13.66	19.75	27.82	36.88	>0.99	>0.99	>0.99	>0.99
	0.021	0.393	0.765	0.940	0.989	>0.99	>0.99					
	0.000	0.035	0.168	0.410	0.674	0.864	0.959	0.991				
n=15	0	1	2	3	4	5	6	7	8	9	10	11
	5	12	18	25	31	38	44	51	57	63	69	75
	0.23	2.33	5.41	8.49	12.56	17.62	22.68	27.74	33.79	>0.99	>0.99	>0.99
	0.238	0.582	0.835	0.951	0.989	0.998	>0.99	>0.99	>0.99			
	0.006	0.041	0.140	0.317	0.538	0.739	0.880	0.955	0.987			
n=20	0	1	2	3	4	5	6	7	8	9	10	11
	9	14	19	23	28	33	38	43	48	53	58	63
	1.26	3.32	6.38	9.44	12.50	16.55	20.60	23.64	28.69	32.73	>0.99	>0.99
	0.422	0.701	0.880	0.962	0.990	0.998	>0.99	>0.99	>0.99	>0.99		
	0.009	0.040	0.117	0.254	0.436	0.626	0.786	0.895	0.956	0.985		
n=25	0	1	2	3	4	5	6	7	8	9	10	11
	11	15	19	23	27	31	35	39	43	46	50	54
	3.27	5.32	7.37	10.41	12.46	15.50	18.54	21.58	25.62	28.65	32.69	>0.99
	0.563	0.782	0.911	0.970	0.992	0.998	>0.99	>0.99	>0.99	>0.99	>0.99	
	0.010	0.037	0.099	0.206	0.357	0.529	0.692	0.822	0.909	0.959	0.984	
n=30	0	1	2	3	4	5	6	7	8	9	10	11
	12	16	19	22	26	29	32	35	39	42	45	49
	4.27	6.31	8.35	10.39	13.43	15.46	18.50	20.53	23.56	26.60	29.63	32.66
	0.668	0.838	0.934	0.977	0.993	0.998	>0.99	>0.99	>0.99	>0.99	>0.99	>0.99
	0.011	0.033	0.083	0.170	0.295	0.447	0.604	0.743	0.850	0.921	0.963	0.985

Appendix B

Code

This section contains code which was used as part of this thesis. Code has been separated by chapter and labelled as appropriate. All code and documentation relating to this thesis can be found online at the following link - <https://github.com/A-mit-Patel/PhD>

B.1 TITE-PO-CRM in ADePT-DDR chapter code

The following code is the function that is used to conduct a dose recommendation for TITE-POCRM in ADePT-DDR.

```
## Background ----

# This function is to facilitate the use of the TITE-PO
- CRM methodology for
# dose-finding trials. This is an altered version of
the pocrm.imp function
# from the pocrm package.

## Parameters ----

# prior.s -      A matrix of values corresponding to the
possible orderings
```

```

#           of the toxicity probabilities from the
# skeleton

# prior.o -      A vector of prior probabilities for the
# ordering. (Must sum up to
#           1 and be the same length as the number
# of possible orders)

# target -       The target DLT rate

# dlt -          Vector of patient outcomes indicating
# if patient experienced a
#           DLT (1 indicates DLT, 0 indicates no
# DLT)

# dose -         A vector of dose-levels assigned to
patients (Length of dose
#           must be the same as the length of tox)

# weight -       A vector of weights for each patient.
# Weights must take values
#           between 0 and 1 (Length of weigh must
be the same as length of
#           tox and dose)

# stop.low -     Recommend stopping the trial if
toxicity is too high at lowest
#           dose (TRUE/FALSE default is FALSE)

# stop.target -  DLT rate required to trigger stopping
rule. Must be specified if
#           stop.low == TRUE (Must be higher than
target)

```

```
# stop.conf - Confidence level to trigger stopping
# rule. Must be specified if
# stop.low == TRUE

## Packages ---
# These packages aren't require to run this function
# but contain helpful
# functions which will facilitate the use of this
# function. getprior in the
# dfcrm pacakge to obtain a skeleton and getwm from
# pocrm to obtain the matrix
# of skeleton values (prior.s)

library(dfcrm)
library(pocrm)

tite.pocrm <- function(prior.s,
                       prior.o,
                       target,
                       dlt,
                       dose,
                       weight,
                       stop.low = FALSE,
                       stop.target = NA,
                       stop.conf = NA)

{
  # Empty vector to store estimates
```

```
pred <- rep(0, nrow(prior.s))

# Empty vector to store likelihood
lik <- rep(0, nrow(prior.s))

# n is the number of patients
n <- length(dlt)

# Loop through each order
for (k in 1:nrow(prior.s)) {

  # Apply skeleton values to patients doses
  beta <- prior.s[k,][dose]

  # Likelihood calculation
  ll <- function(a) {

    la <- 0

    # Loop through each patient
    for (i in 1:n) {

      la <- la + dlt[i] * a * log(weight[i]*beta[i])
      +
      (1-dlt[i]) * log((1 - weight[i])*beta[i]^a)
    }
    la
  }

  # Estimates for each order
}
```

```
pred[k] <- optimize(f = ll, interval = c(0, 500),
maximum = T)$maximum

# Likelihood for each order

lik[k] <- ll(pred[k])

}

# Posterior probabilities of each order

pord <- (exp(lik) * prior.o)/sum(exp(lik) * prior.o)

# Select order with greatest posterior

ord <- which.is.max(round(pord,7))

# Estimate of the model parameter from the selected
order

bhat <- pred[ord]

# Updated estimates of the toxicity probabilities

rpred <- prior.s[ord, ]^bhat

# Select dose level closest to the target

next.lev <- which.is.max(-(abs(rpred - target)))

# Obtain the current dose level

current.lev <- tail(dose, n = 1)

# Calculation of the inverse variance

beta <- prior.s[ord,][dose]

inverse.var <- 0

for (i in 1:n) {
```

```

inverse.var <- inverse.var + ((1-dlt[i])*(weight[i]
*beta[i]^bhat)*(log((weight[i]*beta[i])))^2)/
(1-((weight[i]*beta[i]^bhat)))^2

}

inverse.var <- ifelse(inverse.var == 0, 0, 1/inverse.
var)

# If stopping rule is true check toxicity at lowest
dose

if (stop.low == TRUE){

# Sample to check if lowest dose too toxic
samp <- rnorm(n = 100000, mean = bhat, sd = sqrt(
inverse.var))

tox.low <- prior.s[ord, 1]^samp
prob.tox.low <- mean(tox.low > stop.target)

if(prob.tox.low > stop.conf){
next.lev <- "stop"
}

}

# 95% Confidence Intervals
crit <- qnorm(0.975)

```

```

ptox.L <- prior.s[ord,]^bhat + crit*sqrt(inverse.var
))

ptox.U <- prior.s[ord,]^bhat - crit*sqrt(inverse.var
))

# Output
out <- list(ord.prob = round(pord,3),
            order.est = ord,
            b.est = round(bhat, 3),
            ptox.est = rpred,
            dose.rec = next.lev,
            ptox.U = ptox.U,
            ptox.L = ptox.L,
            post.var = 1/inverse.var,
            se = sqrt((inverse.var^2)/n))

}


```

The code below is for various functions that were used to run the simulations in Chapter 2.

```

# Simulation function for PO-TITE-CRMs
titepocrm_sim <- function (r, alpha, prior.o, x0, stop,
                           n, theta, nsim,
                           tox.range, cohort, obswin,
                           minfu, win2, recrate,
                           mtd.lim, tox.lim, tox.cert)
{
  start_time <- Sys.time()

```

```

sim <- sim1 <- apred <- lik <- pord <- ord <- ahat <-
rpred <- next.lev <- n1 <- N <- NULL

d <- ncol(alpha)
s <- nrow(alpha)

if (nsim > 1) {
  lpocrm <- function(r, alpha, prior.o, x0, stop, n,
  theta) {
    if (is.vector(alpha))
      alpha = t(as.matrix(alpha))
    nord.tox = nrow(alpha)
    mprior.tox = prior.o
    # Function for calculating the likelihood
    bcrml <- function(a, p1, y, w) {
      lik = 0
      for (j in 1:length(y)) {
        lik = lik + y[j] * a * log(w[j]*p1[j]) +
          (1 - y[j]) * log((1 - w[j]*p1[j]^a))
      }
      return(lik)
    }
    # specifies the number of doses
    ncomb = ncol(alpha)
    # Initialise empty vectors for use later on
    # y (number of dlt's per dose), npts(number of
    # patients per dose)
    y = npts = ptox.hat = numeric(ncomb)
  }
}

```

```

# The dose level to be selected (add 1 to
# indicate position to stop)
comb.select = numeric(ncomb)

# Starting dose taken from first element of the
# dose escalation scheme
comb.curr = x0[1]

stoprule = 0
stop.count <- 0

# Specifies the dose escalation scheme
stage1 <- c(x0, rep(ncol(alpha), n - length(x0)))

# Counter for the number of cohorts
cohort.count <- 1

# Empty vectors to store dose received and if dlt
# was observed
duration <- dose <- dlt <- dlt_time <- fu <- tox
<- weights <- vector()

# Time to recruit new patients
rectime <- obswin/recrate

while(length(tox) < n){

  # Takes draws from a binomial distribution to
  # determine if patients in the cohort had a
  # toxic outcome
  cohort_tox <- stats::rbinom(n = cohort, size =
    1, prob = r[comb.curr])

  # Dummy variable to store DLT which will be
  # updated once the time for DLT has passed
}

```

```

cohort_dlt <- rep(0, cohort)

# Patients in the same cohort receive the same
# dose

cohort_comb <- rep(comb.curr, cohort)

# Follow up time based on the time taken to
# recruit a patient

# First patient assumed to be recruited
# instantly

# Add the min fu time to indicate when analysis
# is done

cohort_fu <- (rectime*(cohort-1)) - (rectime *
0:(cohort - 1)) + minfu

# Time of DLT is recorded and set to be at any
# time during the obswin

cohort_dlt_time <- vector()

for (i in 1:cohort) {

  if(cohort_tox[i] == 0){

    cohort_dlt_time[i] <- NA

  }

  else {

    cohort_dlt_time[i] <- runif(1,0, obswin)

  }

}

# Stores the time DLT occurs for new cohort

dlt_time <- c(dlt_time, cohort_dlt_time)

# Stores new cohorts toxicity

tox <- c(tox, cohort_tox)

```

```

# Stores dose level for each new cohort
dose <- c(dose, cohort_comb)

# Store dummy dlt for new cohort
dlt <- c(dlt, cohort_dlt)

# Add on follow up time and recruitment time
# for previous cohorts
fu <- fu + (rectime*(cohort-1)) + minfu
#duration <- duration + (cohort * rectime) +
minfu

# store new cohorts follow up time
fu <- c(fu, cohort_fu)
#duration <- c(duration, cohort_fu)

# Loop through all the patients who will have a
DLT
for(i in 1:length(fu)){
  if(tox[i] == 1){
    # If the follow up time is greater than the
    # time the dlt was determined to occur
    if(fu[i] >= dlt_time[i]){
      # Change the follow up time to the obswin
      # to indicate the dlt has now happened
      fu[i] <- obswin
      # As the dlt has happened it should now
      # be stored for calculation in the
      # likelihood
      dlt[i] <- 1
    }
  }
}

```

```

        }

    }

fu <- pmin(fu, obswin)

# Calculated weights where the min fu accounts
# for 60% and is 80% by window 2
for (i in 1:length(fu)) {
  weights[i] <- 0.6 +
  0.2*(min(win2 - minfu, fu[i]-minfu)/(win2 -
  minfu))+ 0.2*(max(fu[i] - win2, 0)/(
  obswin-win2))
}

# These values are used to calculate patient
# allocation and %DLT
y[comb.curr] = y[comb.curr] + sum(cohort_tox)
npts[comb.curr] = npts[comb.curr] + cohort

if (sum(dlt) == length(dlt)) {
  comb.curr <- ifelse(comb.curr == 1, comb.curr
  ,
  comb.curr - 1)
}

else if (sum(dlt) == 0) {
  comb.curr <- ifelse(comb.curr == ncomb, comb.
curr,
  stage1[cohort.count + 1])
}

```

```
    }

    else {

        break

    }

# Stopping rule ends stage 1 when the same dose
is recommended to a sixth cohort. As this
follows the escalation scheme only stops
when the max dose is prescribed to sixth
cohort.

if(sum(dose == comb.curr) == mtd.lim ){

    break

}

cohort.count <- cohort.count +1

}

while (length(dlt) <= n) {

    if (sum(dlt) == 0 ) {

        stop = 0

        break

    }

    else {

        like.tox = est.tox = rep(0, nord.tox)
```

```

for (k in 1:nord.tox) {

  beta <- alpha[k,][dose[1:length(dose)]]

  est.tox[k] <- optimize(f = bcrm1, interval
= c(0,500),
                        p1 = beta, y = dlt,
                        w = weights,
                        maximum = T)$maximum

  like.tox[k] <- optimize(f = bcrm1, interval
= c(0,500),
                        p1 = beta, y = dlt,
                        w = weights,
                        maximum = T)$

  objective

}

postprob.tox = (exp(like.tox) * mprior.tox)/
sum(exp(like.tox) * mprior.tox)

if (nord.tox > 1) {
  mtox.sel = which.is.max(round(postprob.tox
,8))
}

else {
  mtox.sel = 1
}

```

```
ptox.hat = alpha[mtox.sel, ]^est.tox[mtox.sel
]

loss = abs(ptox.hat - theta)

comb.curr = which.is.max(round(-loss,8))

# obtain the last dose level from the data
last.lev <- tail(dose, n = 1)

if (comb.curr %in% dose | comb.curr < last.
lev){

  # If the next recommended lev has already
  # been tested or is lower than
  # the current dose level than recommend
  # next level

  comb.curr <- comb.curr
}

else {

  comb.curr <- max(dose) + 1
}

if (npts[comb.curr] == stop) {

  stoprule <- 0
  break
}

# Calculate asymptotic variance
beta <- alpha[mtox.sel,][dose]
inverse.var <- 0

for (i in 1:length(dlt)) {
```

```
inverse.var <- inverse.var + ((1-dlt[i])*(
  weights[i]*beta[i]^est.tox[mtox.sel])*(

  log((weights[i]*beta[i])))^2)/(1-((

  weights[i]*beta[i]^est.tox[mtox.sel])))

^2

}

post.alpha.mean <- est.tox[mtox.sel]

post.alpha.var <- ifelse(inverse.var == 0, 0
  , 1/inverse.var)

post.alpha.samp <- rnorm(n = 100000, mean =
  post.alpha.mean,
  sd = sqrt(post.alpha
  .var))

post.prob.tox.samp <- alpha[mtox.sel,1]^post.
  alpha.samp

prob.too.toxic <- mean(post.prob.tox.samp >
  tox.lim)

if(is.na(prob.too.toxic)) {

  prob.too.toxic <- 1

}
```

```
# stopping rule for when lowest dose is too
# toxic AND lowest dose has been tested ptox
# .hat[1] > 0.5 & 1 %in% dose
#
# stopping rule if dose is being recommended
# for a xth time
if (length(dlt) == n | (prob.too.toxic > tox.
cert) & 1 %in% dose | sum(dose == comb.
curr) == mtd.lim) {
stoprule = 0

# once all patients recruited calculate
# likelihoods at max weight
like.tox = est.tox = rep(0, nord.tox)
for (k in 1:nord.tox) {

#set all the weights at 1
weights <- rep(1, length(tox))
#ensures that all those predicted to have
#toxic event is included in dlt data
#as we are assuming full follow up once
#the last patients are recruited
dlt <- tox
beta <- alpha[k,][dose[1:length(dose)]]
est.tox[k] <- optimize(f = bcrm1,
interval = c(0,500),
```

```

      p1 = beta, y = tox
      , w = weights,
maximum = T)$

maximum

like.tox[k] <- optimize(f = bcrm1,
interval = c(0,500),
p1 = beta, y =
tox, w =
weights,
maximum = T)$

objective

}

postprob.tox = (exp(like.tox) * mprior.tox)
/sum(exp(like.tox) * mprior.tox)

if (nord.tox > 1) {
mtox.sel = which.is.max(round(postprob.
tox,8))
}
else {
mtox.sel = 1
}
ptox.hat = alpha[mtox.sel, ]^est.tox[mtox.
sel]
loss = abs(ptox.hat - theta)

```

```
comb.curr = which.is.max(round(-loss,8))

# obtain the last dose level from the data
last.lev <- tail(dose, n = 1)

if (comb.curr %in% dose | comb.curr < last.lev){

  # If the next recommended level has
  # already been tested or is lower than
  # the current dose level than recommend
  # next level

  comb.curr <- comb.curr

}

else {

  comb.curr <- max(dose) + 1

}

# Calculate asymptotic variance
beta <- alpha[mtox.sel,][dose]
inverse.var <- 0

for (i in 1:length(tox)) {

  inverse.var <- inverse.var + ((1-tox[i]
    ])*(weights[i]*beta[i]^est.tox[mtox.
    sel])* (log((weights[i]*beta[i])))^2
  / (1-((weights[i]*beta[i]^est.tox[
    mtox.sel])))^2
}
```

```
}

post.alpha.mean <- est.tox[mtox.sel]
post.alpha.var <- ifelse(inverse.var ==
  0, 0, 1/inverse.var)
post.alpha.samp <- rnorm(n = 100000, mean
  = post.alpha.mean,
  sd = sqrt(post.
alpha.var))

post.prob.tox.samp <- alpha[mtox.sel,1]^
  post.alpha.samp
prob.too.toxic <- mean(post.prob.tox.samp
  > tox.lim)

if(is.na(prob.too.toxic)) {
  prob.too.toxic <- 1
}

break

} else {
```

```
# Takes draws from a binomial distribution
# to determine if patients in the cohort
# had a toxic outcome
cohort_tox <- stats::rbinom(n = cohort,
                             size = 1, prob = r[comb.curr])
cohort_dlt <- rep(0, cohort)
# Patients in the same cohort receive the
# same dose
cohort_comb <- rep(comb.curr, cohort)
# Follow up time based on the time taken to
# recruit a patient
# First patient assumed to be recruited
# instantly
# Add the min fu time to indicate when
# analysis is done
cohort_fu <- (rectime*(cohort-1))-(rectime
               * 0:(cohort - 1))+minfu
# Time of DLT is recorded and set between
# the time the patient has been followed
# up and the remaining observation window
cohort_dlt_time <- vector()
for (i in 1:cohort) {
  if(cohort_tox[i] == 0){
    cohort_dlt_time[i] <- NA
  }
  else {
```

```

cohort_dlt_time[i] <- runif(1,0, obswin
)
}

# Stores the time DLT occurs for new cohort
dlt_time <- c(dlt_time, cohort_dlt_time)

# Stores new cohorts toxicity
tox <- c(tox, cohort_tox)

# Stores dose level for each new cohort
dose <- c(dose, cohort_comb)

# Store dummy dlt for new cohort
dlt <- c(dlt, cohort_dlt)

# Add on follow up time and recruitment
# time for previous cohorts
fu <- fu + (rectime*(cohort-1)) + minfu
#duration <- duration + (cohort * rectime)
+ minfu

# store new cohorts follow up time
fu <- c(fu, cohort_fu)
#duration <- c(duration, cohort_fu)

for(i in 1:length(fu)){
  if(tox[i] == 1){
    if(fu[i] >= dlt_time[i]){
      fu[i] <- obswin
      dlt[i] <- 1
    }
  }
}

```

```

}

fu <- pmin(fu, obswin)

# Calculated weights where the min fu
accounts for 60% and 80% by win2

for (i in 1:length(fu)) {

  weights[i] <- 0.6 +
  0.2*(min(win2 - minfu, fu[i]-minfu)/(

    win2 - minfu))+ 0.2*(max(fu[i]- win2
    , 0)/(obswin-win2))

}

y[comb.curr] = y[comb.curr] + sum(cohort_
tox)

npts[comb.curr] = npts[comb.curr] + cohort
}

}

if (stoprule == 0) {

  if (!exists("prob.too.toxic")){prob.too.toxic
  <- 0}

  if((prob.too.toxic > tox.cert)& 1 %in% dose){

    stop.count <- 1

  }

  else {
}
}

```

```

    comb.select[comb.curr] = comb.select[comb.

        curr] + 1

    }

}

# Calculate the duration of the trial

num_cohorts <- length(tox) / cohort

duration <- cohort_fu[1] + (rectime*(cohort-1) +
    minfu) * (num_cohorts-1) + (obswin - minfu)

return(list(MTD.selection = comb.select, tox.data
    = y,
    patient.allocation = npts, duration =
        duration,
    stop = stop.count))

}

lpocrm.sim <- function(nsim) {

ncomb = length(r)

y <- npts <- matrix(nrow = nsim, ncol = ncomb)

comb.select <- matrix(nrow = nsim, ncol = ncomb )

duration <- stop.count <- trialsize <- rep(0, nsim)

```

```
nstop = 0
for (i in 1:nsim) {
  #print(i)
  result <- lpocrm(r, alpha, prior.o, x0, stop, n,
                    theta)
  comb.select[i, ] = result$MTD.selection
  y[i, ] = result$tox.data
  npts[i, ] = result$patient.allocation
  trialsize[i] = sum(result$patient.allocation)
  duration[i] = result$duration # convert duration
  to months
  stop.count[i] = result$stop
}
return(list(true.prob = r,
           time = Sys.time() - start_time,
           MTD.selection = round(colMeans(comb.
                                         select),2),
           patient.allocation = 100* round(
             colMeans(npts)/mean(trialsize),2),
           percent.DLT = sum(colMeans(y))/mean(
             trialsize),
           months = 12*mean(duration)/365,
           stop = mean(stop.count),
           max.n.count = length(trialsize[
             trialsize == max(trialsize)]),
           summary.tralsize = summary(trialsize),
           mean.n = mean(trialsize),
```

```

mean.n.perdose = colMeans(npts),
acceptable = sum(colMeans(comb.select)[
  which(round(abs(r - theta), 2) <=
tox.range)]))

}

if (nsim == 1) {
  twostgcrm(r, x0, stop, n, theta)
}
else {
  lpocrm.sim(nsim)
}
}

# Simulation function for PO-CRMs
pocrm_sim <- function (r, alpha, prior.o, x0, stop, n,
theta, nsim,
tox.range, cohort,
mtd.lim, tox.lim, tox.cert)
{
  start_time <- Sys.time()
  sim <- sim1 <- apred <- lik <- pord <- ord <- ahat <-
  rpred <- next.lev <- n1 <- N <- NULL
  d <- ncol(alpha)
  s <- nrow(alpha)
  if (nsim > 1) {
    mean.n.perdose = colMeans(npts),
    acceptable = sum(colMeans(comb.select)[
      which(round(abs(r - theta), 2) <=
      tox.range)]))
  }
}
```

```
lpocrm <- function(r, alpha, prior.o, x0, stop, n,
theta) {
  if (is.vector(alpha))
    alpha = t(as.matrix(alpha))
  nrow.tox = nrow(alpha)
  mprior.tox = prior.o

  bcrml <- function(a, p1, y, n) {
    lik = 0
    for (j in 1:length(p1)) {
      lik = lik + y[j] * a * log(p1[j]) + (n[j] -
y[j])
      *
      log
      ((1 -
p1 [
j]^
a))
    }
    return(lik)
  }

# specifies the number of doses
ncomb = ncol(alpha)
# Initialise empty vectors for use later on
```

```

# y (number of dlt's per dose), npts(number of
# patients per dose)
y = npts = ptox.hat = numeric(ncomb)

# The dose level to be selected (add 1 to
# indicate position to stop)
comb.select = numeric(ncomb)

# Starting dose taken from first element of the
# dose escalation scheme
comb.curr = x0[1]

stoprule = 0

# Specifies the dose escalation scheme
stage1 <- c(x0, rep(ncol(alpha), n - length(x0)))

# Counter for the number of cohorts
cohort.count <- 1

# Empty vectors to store dose received and if dlt
# was observed
dose <- dlt <- dlt_time <- fu <- tox <- weights
<- vector()

# Time to recruit new patients
#rectime <- obswin/recrate

while(length(dlt) < n){

  # Takes draws from a binomial distribution to
  # determine if patients in the cohort had a
  # toxic outcome
  cohort_tox <- stats::rbinom(n = cohort, size =
  1, prob = r[comb.curr])
}

```

```
# Dummy variable to store DLT which will be
# updated once the time for DLT has
# passed
cohort_dlt <- cohort_tox

# Patients in the same cohort receive the same
# dose
cohort_comb <- rep(comb.curr, cohort)

# Stores new cohorts toxicity
tox <- c(tox, cohort_tox)

# Stores dose level for each new cohort
dose <- c(dose, cohort_comb)

# Store dummy dlt for new cohort
dlt <- c(dlt, cohort_dlt)

y[comb.curr] = y[comb.curr] + sum(cohort_tox)
npts[comb.curr] = npts[comb.curr] + cohort

if (sum(dlt) == length(dlt)) {
  comb.curr <- ifelse(comb.curr == 1, comb.curr
  ,
  comb.curr - 1)
}

else if (sum(dlt) == 0) {
  comb.curr <- ifelse(comb.curr == ncomb, comb.
curr,
```

```
stage1[cohort.count + 1])  
}  
  
else {  
  break  
}  
  
# Stopping rule ends stage 1 when the same dose  
is recommended to a sixth cohort. As this  
follows the escalation scheme only stops  
when the max dose is prescribed to sixth  
cohort.  
  
if(sum(dose == comb.curr) == mtd.lim ){  
  break  
}  
  
cohort.count <- cohort.count +1  
  
}  
  
  
  
  
  
  
while (length(dlt) <= n) {  
  if (sum(dlt) == 0 ) {  
    stop = 0  
    break  
  }  
  else {
```

```

like.tox = est.tox = rep(0, nord.tox)

for (k in 1:nord.tox) {

  est.tox[k] <- optimize(f = bcrm1, interval
    = c(0,100),
    p1 = alpha[k, ], y =
    y, n = npts,
    maximum = T)$maximum

  like.tox[k] <- optimize(f = bcrm1, interval
    = c(0,100),
    p1 = alpha[k, ], y
    = y, n = npts,
    maximum = T)$

  objective
}

postprob.tox = (exp(like.tox) * mprior.tox)/
sum(exp(like.tox) * mprior.tox)

if (nord.tox > 1) {
  mtox.sel = which.is.max(round(postprob.tox
    ,8))
}

else {
  mtox.sel = 1
}

```

```

ptox.hat = alpha[mtox.sel, ]^est.tox[mtox.sel
]

loss = abs(ptox.hat - theta)

comb.curr = which.is.max(round(-loss,8))

# obtain the last dose level from the data

last.lev <- tail(dose, n = 1)

if (comb.curr %in% dose | comb.curr < last.

lev){

# If the next recommended lev has already
been tested or is lower than
# the current dose level than recommend
next level

comb.curr <- comb.curr

}

else {

  comb.curr <- max(dose) + 1

}

if (npts[comb.curr] == stop) {

  stoprule <- 0

  break

}

# Calculate asymptotic variance

beta <- alpha[mtox.sel,][dose]

inverse.var <- 0

for (i in 1:length(dlt)) {

```

```

    inverse.var <- inverse.var + ((1-dlt[i])*(

        beta[i]^est.tox[mtox.sel])*(log((beta[i

]))^2)/(1-((beta[i]^est.tox[mtox.sel]))

)^2

}

post.alpha.mean <- est.tox[mtox.sel]
post.alpha.var <- 1/inverse.var
post.alpha.samp <- rnorm(n = 100000, mean =
post.alpha.mean,
sd = sqrt(post.alpha
.var))

post.prob.tox.samp <- alpha[mtox.sel,1]^post.

alpha.samp
prob.too.toxic <- mean(post.prob.tox.samp >
tox.lim)

if(is.na(prob.too.toxic)) {

prob.too.toxic <- 1
}

# stopping rule for when lowest dose is too
# toxic AND lowest dose has been tested ptox
# .hat[1] > 0.5 & 1 %in% dose
#
# stopping rule if dose is being recommended
# for a xth time

```

```
if (length(dlt) == n |(prob.too.toxic > tox.
cert) & 1 %in% dose | sum(dose == comb.
curr) == mtd.lim) {

stoprule = 0

break
}

else {

# Takes draws from a binomial distribution
# to determine if patients in the cohort
# had a toxic outcome

cohort_tox <- stats::rbinom(n = cohort,
size = 1, prob = r[comb.curr])

cohort_dlt <- rep(0, cohort)

# Patients in the same cohort receive the
# same dose

cohort_comb <- rep(comb.curr, cohort)

# Stores new cohorts toxicity
tox <- c(tox, cohort_tox)

# Stores dose level for each new cohort
dose <- c(dose, cohort_comb)

# Store dummy dlt for new cohort
dlt <- c(dlt, cohort_dlt)
```

```

y[comb.curr] = y[comb.curr] + sum(cohort_
tox)

npts[comb.curr] = npts[comb.curr] + cohort
}

}

}

if (stoprule == 0) {
  if (!exists("prob.too.toxic")){prob.too.toxic
    <- 0}
  if((prob.too.toxic > tox.cert) & 1 %in% dose){
    #comb.select[ncomb+1] <- 1
  }
  else {
    comb.select[comb.curr] = comb.select[comb.
      curr] + 1
  }
}

stop.count <- 0
if((prob.too.toxic > tox.cert) & 1 %in% dose){
  stop.count <- 1
}

duration <- 474 + 504*((length(tox)-1)/3)
return(list(MTD.selection = comb.select, tox.data
= y,

```

```
    patient.allocation = npts, stop =
      stop.count,
      duration = duration))

}

lpocrm.sim <- function(nsim) {
  ncomb = length(r)

  y <- npts <- matrix(nrow = nsim, ncol = ncomb)
  comb.select <- matrix(nrow = nsim, ncol = ncomb)

  duration <- stop.count <- trialsize <- rep(0, nsim)
  nstop = 0

  for (i in 1:nsim) {
    print(i)

    result <- lpocrm(r, alpha, prior.o, x0, stop, n,
                      theta)

    comb.select[i, ] = result$MTD.selection
    y[i, ] = result$tox.data
    npts[i, ] = result$patient.allocation
    trialsize[i] = sum(result$patient.allocation)
    stop.count[i] = result$stop
    duration[i] = result$duration
  }

  return(list(true.prob = r,
```

```
        time = Sys.time() - start_time,
        MTD.selection = round(colMeans(comb.
            select), 2),
        patient.allocation = 100 * round(
            colMeans(npts)/mean(trialsizes), 2),
        percent.DLT = sum(colMeans(y))/mean(
            trialsizes),
        stop = mean(stop.count),
        months = 12*mean(duration)/365,
        mean.n = mean(trialsizes),
        mean.n.perdose = colMeans(npts),
        acceptable = sum(colMeans(comb.select)[
            which(round(abs(r - theta), 2) <=
            tox.range)])))
    }
    if (nsim == 1) {
        twostgcrm(r, x0, stop, n, theta)
    }
    else {
        lpocrm.sim(nsim)
    }
}

# Simulation function for two-stage TITE-CRM
applied_titecrmmts_sim_v2 <- function (true_tox, prior,
    target, max_sample_size,
```

```
    num_sims , win2 ,
    cohort = 1 ,
    obswin , minfu ,
    recrate , initdes ,
    n.mtd ,
    dose_func =
    applied_
    titecrm , ...)

{

iterations <- list()
start_time <- Sys.time()
for (s in 1:num_sims) {

  stop <- FALSE
  stop_reason <- NULL
  rectime <- obswin/recrate

  level <- initdes[1]
  stage1 <- c(initdes , rep(length(prior) , max_sample_size -
  length(initdes)))
  duration <- dose <- dlt <- dlt_time <- fu <- tox <-
  weights <- vector()

  # Counter for the number of cohorts
  cohort.count <- 1

  # specifies the number of doses
  ncomb <- length(true_tox)
  while (length(tox) < max_sample_size) {
```

```
# Takes draws from a binomial distribution to
# determine if patients in the cohort had a
# toxic outcome
cohort_tox <- stats::rbinom(n = cohort, size = 1,
                             prob = true_tox[level])
# Dummy variable to store DLT which will be
# updated once the time for DLT has
# passed
cohort_dlt <- rep(0, cohort)
# Patients in the same cohort receive the same
# dose
cohort_level <- rep(level, cohort)
# Follow up time based on the time taken to
# recruit a patient
# First patient assumed to be recruited instantly
# Add the min fu time to indicate when analysis
# is done
cohort_fu <- (rectime*(cohort-1)) - (rectime *
                                         0:(cohort - 1)) + minfu
# Time of DLT is recorded and set to be at any
# time during the obswin
cohort_dlt_time <- vector()
for (i in 1:cohort) {
  if(cohort_tox[i] == 0){
    cohort_dlt_time[i] <- NA
  }
  else {
```

```

        cohort_dlt_time[i] <- runif(1,0, obswin)
    }
}

# Stores the time DLT occurs for new cohort
dlt_time <- c(dlt_time, cohort_dlt_time)

# Stores new cohorts toxicity
tox <- c(tox, cohort_tox)

# Stores dose level for each new cohort
dose <- c(dose, cohort_level)

# Store dummy dlt for new cohort
dlt <- c(dlt, cohort_dlt)

# Add on follow up time and recruitment time for
# previous cohorts
fu <- fu + (rectime*(cohort-1)) + minfu

#duration <- duration + (cohort * rectime) +
minfu

# store new cohorts follow up time
fu <- c(fu, cohort_fu)

#duration <- c(duration, cohort_fu)

# Loop through all the patients who will have a
# DLT

for(k in 1:length(fu)){
  if(tox[k] == 1){
    # If the follow up time is greater than the
    # time the dlt was determined to occur
    if(fu[k] >= dlt_time[k]){

```

```
# Change the follow up time to the obswin
# to indicate the dlt has now happened
fu[k] <- obswin
# As the dlt has happened it should now be
# stored for calculation in the likelihood
dlt[k] <- 1
}

}
}

fu <- pmin(fu, obswin)
# Calculated weights where the min fu accounts
# for 60% and is 80% bywindow 2
for (a in 1:length(fu)) {
  weights[a] <- 0.6 +
  0.2*(min(win2 - minfu, fu[a]-minfu)/(win2 -
  minfu))+
  0.2*(max(fu[a] - win2, 0)/(obswin-win2))
}

if (sum(dlt) == length(dlt)) {
  level <- ifelse(level == 1, level,
  level - 1)
}
else if (sum(dlt) == 0) {
  level <- ifelse(level == ncomb, level,
```

```
stage1[cohort.count + 1])  
}  
  
else {  
    break  
}  
  
# Stopping rule ends stage 1 when the same dose  
is recommended to a sixth cohort. As this  
follows the escalation scheme only stops when  
the max dose is prescribed to sixth cohort.  
  
if(sum(dose == level) == n.mtd ){  
    break  
}  
  
cohort.count <- cohort.count +1  
}  
  
  
while ( !stop & length(tox) <= max_sample_size) {  
    if (length(dlt) == max_sample_size & sum(dlt) ==  
        0 ) {  
        stop = TRUE  
        break  
    }  
    else{  
        x <- dose_func(prior = prior, target = target,  
                        weights = weights ,
```

```
          tox = dlt, level = dose,
          followup = fu, obswin =
          obswin,
          ...)

fdose <- level <- x$mtd

stop <- ifelse(is.null(x$stop), FALSE, x$stop)
stop_reason <- x$stop_reason

if(length(dlt) == max_sample_size | stop ==
TRUE ){
  x <- dose_func(prior = prior, target = target
  , tox = tox,
    level = dose, followup = rep(
      obswin, length(tox)),
      obswin = obswin, weights = rep
      (1, length(tox)),
      ...)

  fdose <- x$mtd

  stop <- ifelse(is.null(x$stop), FALSE, x$stop
  )

  stop_reason <- x$stop_reason
  break
}

else{
```

```
cohort_tox = stats::rbinom(n = cohort, size =
  1,
  prob = true_tox[
    level])

cohort_dlt <- rep(0, cohort) ##
cohort_level = rep(level, cohort)
cohort_fu = (rectime * (cohort - 1)) - (
  rectime *
  c(
    (0:(cohort -
      1) +
      minfu

cohort_dlt_time <- c()
for (j in 1:cohort) {
  if(cohort_tox[j] == 0){
    cohort_dlt_time[j] <- NA
  }
}
```

```

    else {
      cohort_dlt_time[j] <- runif(1,0, obswin)
    }
  }

# cohort_fu[cohort_tox == 1] <- obswin

# Stores the time DLT occurs for new cohort
dlt_time <- c(dlt_time, cohort_dlt_time) ##

# Store dummy dlt for new cohort
dlt <- c(dlt, cohort_dlt) ##

tox <- c(tox, cohort_tox)

dose <- c(dose, cohort_level)

fu <- fu + (rectime*(cohort-1)) + minfu

fu <- c(fu, cohort_fu)

#duration <- duration + (cohort * rectime) +
minfu

for (l in 1:length(tox)) {
  if(tox[l] == 1 & fu[l] >= dlt_time[l]){
    fu[l] <- obswin
    dlt[l] <- 1
  }
}

fu <- pmin(fu, obswin)

for (b in 1:length(fu)) {
  weights[b] <- 0.6 +

```

```
    0.2*(min(win2 - minfu, fu[b]-minfu)/(win2
        - minfu))+
    0.2*(max(fu[b]- win2, 0)/(obswin-win2))
}

cohort.count <- cohort.count+1

}

}

print(s)

num_cohorts <- length(tox) / cohort
duration <- cohort_fu[1] + (rectime*(cohort-1) +
    minfu) * (cohort.count-1) +(obswin - minfu)
iterations[[s]] <- list(tox = tox, level = dose,
    mtd = fdose,
    stop = stop, stop_reason =
    stop_reason,
    duration = duration,
    cohorts = cohort.count)
}

dose_selections = sapply(iterations, function(x) x$  
mtd)
doses_given = unlist(sapply(iterations, function(x) x  
$level))
```

```

duration = sapply(iterations, function(x) x$duration)

summary = list(true_tox = true_tox, prior = prior,
target = target,
max_sample_size = max_sample_size,
initdes = initdes,
months = 12*mean(duration)/365,
time = Sys.time() - start_time,
num_sims = num_sims, cohort = cohort,
prob_stop = table(substr(unlist(sapply
(iterations, function(x)
x$stop_reason)), 1, 15))/num_sims,
mtd = sapply(1:length(prior),
function(d)
sum(dose_selections == d, na.rm =
TRUE)/num_sims),
doses_given = sapply(1:length(prior),
function(d)
sum(doses_given == d, na.rm = TRUE)/
num_sims), prob_dose_given =
sapply(1:length(prior),

```



```
return(list(summary = summary, iterations =
iterations))
}

# Simulations for no cohorts and removal of minimum
# followup

# Not used

titepocrm_sim_nomin <- function (r, alpha, prior.o, x0,
stop, n, theta, nsim,
tox.range, cohort, obswin,
minfu, win2, recrate,
mtd.lim, tox.lim, tox.cert)
{

sim <- sim1 <- apred <- lik <- pord <- ord <- ahat <-
rpred <- next.lev <- n1 <- N <- NULL
d <- ncol(alpha)
s <- nrow(alpha)
if (nsim > 1) {
lpocrm <- function(r, alpha, prior.o, x0, stop, n,
theta) {
if (is.vector(alpha))
alpha = t(as.matrix(alpha))
nord.tox = nrow(alpha)
mprior.tox = prior.o
# Function for calculating the likelihood
}
```

```

bcrml <- function(a, p1, y, w) {
  lik = 0
  for (j in 1:length(y)) {
    lik = lik + y[j] * a * log(w[j]*p1[j]) +
      (1 - y[j]) * log((1 - w[j]*p1[j]^a))
  }
  return(lik)
}

# specifies the number of doses
ncomb = ncol(alpha)

# Initialise empty vectors for use later on
# y (number of dlt's per dose), npts(number of
# patients per dose)
y = npts = ptox.hat = numeric(ncomb)

# The dose level to be selected (add 1 to
# indicate position to stop)
comb.select = numeric(ncomb)

# Starting dose taken from first element of the
# dose escalation scheme
comb.curr = x0[1]

stoprule = 0
stop.count <- 0

# Specifies the dose escalation scheme
stage1 <- c(x0, rep(ncol(alpha), n - length(x0)))

# Counter for the number of cohorts
cohort.count <- 1

```

```
# Empty vectors to store dose received and if dlt
# was observed

duration <- dose <- dlt <- dlt_time <- fu <- tox
<- weights <- vector()

# Time to recruit new patients

rectime <- obswin/recrate

while(length(tox) < n){

  # Takes draws from a binomial distribution to
  # determine if patients in the cohort had a
  # toxic outcome

  cohort_tox <- stats::rbinom(n = cohort, size =
    1, prob = r[comb.curr])

  # Dummy variable to store DLT which will be
  # updated once the time for DLT has passed

  cohort_dlt <- rep(0, cohort)

  # Patients in the same cohort receive the same
  # dose

  cohort_comb <- rep(comb.curr, cohort)

  # Follow up time based on the time taken to
  # recruit a patient

  # First patient assumed to be recruited
  # instantly

  # Add the min fu time to indicate when analysis
  # is done

  cohort_fu <- (rectime*(cohort-1)) - (rectime *
    0:(cohort - 1)) + rectime}
```

```

# Time of DLT is recorded and set to be at any
# time during the obswin
cohort_dlt_time <- vector()
for (i in 1:cohort) {
  if(cohort_tox[i] == 0){
    cohort_dlt_time[i] <- NA
  }
  else {
    cohort_dlt_time[i] <- runif(1,0, obswin)
  }
}

# Stores the time DLT occurs for new cohort
dlt_time <- c(dlt_time, cohort_dlt_time)

# Stores new cohorts toxicity
tox <- c(tox, cohort_tox)

# Stores dose level for each new cohort
dose <- c(dose, cohort_comb)

# Store dummy dlt for new cohort
dlt <- c(dlt, cohort_dlt)

# Add on follow up time and recruitment time
# for previous cohorts
fu <- fu + (cohort * rectime)

#duration <- duration + (cohort * rectime) +
#minfu

# store new cohorts follow up time
fu <- c(fu, cohort_fu)

#duration <- c(duration, cohort_fu)

```

```

# Loop through all the patients who will have a
# DLT

for(i in 1:length(fu)){
  if(tox[i] == 1){

    # If the follow up time is greater than the
    # time the dlt was determined to occur
    if(fu[i] >= dlt_time[i]){

      # Change the follow up time to the obswin
      # to indicate the dlt has now happened
      fu[i] <- obswin

      # As the dlt has happened it should now
      # be stored for calculation in the
      # likelihood
      dlt[i] <- 1
    }
  }
}

fu <- pmin(fu, obswin)

# Calculated weights where the min fu accounts
# for 60% and is 80% by window 2
for (i in 1:length(fu)) {
  weights[i] <- 0.6*(min(fu[i], minfu)/minfu) +
  0.2*max((min(win2 - minfu, fu[i]-minfu)/(win2 - minfu)), 0) +
  0.2*max((max(fu[i] - win2, 0)/(obswin - win2)),
  , 0)
}

```

```

}

# These values are used to calculate patient
# allocation and %DLT

y[comb.curr] = y[comb.curr] + sum(cohort_tox)
npts[comb.curr] = npts[comb.curr] + cohort

if (sum(dlt) == length(dlt)) {
  comb.curr <- ifelse(comb.curr == 1, comb.curr
    ,
    comb.curr - 1)
}

else if (sum(dlt) == 0) {
  comb.curr <- ifelse(comb.curr == ncomb, comb.
    curr,
    stage1[cohort.count + 1])
}

else {
  break
}
}

# Stopping rule ends stage 1 when the same dose
# is recommended to a sixth cohort. As this
# follows the escalation scheme only stops
# when the max dose is prescribed to sixth
# cohort.

if(sum(dose == comb.curr) == mtd.lim ){

```

```
break
}

cohort.count <- cohort.count +1

}

while (length(dlt) <= n) {
  if (sum(dlt) == 0 ) {
    stop = 0
    break
  }
  else {
    like.tox = est.tox = rep(0, nord.tox)
    for (k in 1:nord.tox) {
      beta <- alpha[k,][dose[1:length(dose)]]
      est.tox[k] <- optimize(f = bcrm1, interval
        = c(0,500),
        p1 = beta, y = dlt,
        w = weights,
        maximum = T)$maximum
      like.tox[k] <- optimize(f = bcrm1, interval
        = c(0,500),
        p1 = beta, y = dlt,
        w = weights,
```

```

maximum = T)$
objective
}

postprob.tox = (exp(like.tox) * mprior.tox)/
sum(exp(like.tox) * mprior.tox)

if (nord.tox > 1) {
  mtox.sel = which.is.max(round(postprob.tox
,8))
}
else {
  mtox.sel = 1
}
ptox.hat = alpha[mtox.sel, ]^est.tox[mtox.sel
]
loss = abs(ptox.hat - theta)
comb.curr = which.is.max(round(-loss,8))
# obtain the last dose level from the data
last.lev <- tail(dose, n = 1)
if (comb.curr %in% dose | comb.curr < last.
lev){
# If the next recommended lev has already
been tested or is lower than
# the current dose level than recommend
next level
}
```

```
    comb.curr <- comb.curr
  }
else {
  comb.curr <- max(dose) + 1
}

if (npts[comb.curr] == stop) {
  stoprule <- 0
  break
}
# Calculate asymptotic variance
beta <- alpha[mtox.sel,][dose]
inverse.var <- 0

for (i in 1:length(dlt)) {
  inverse.var <- inverse.var + ((1-dlt[i])*(weights[i]*beta[i]^est.tox[mtox.sel])*(log((weights[i]*beta[i])))^2)/(1-((weights[i]*beta[i]^est.tox[mtox.sel])))^2
}

post.alpha.mean <- est.tox[mtox.sel]
post.alpha.var <- ifelse(inverse.var == 0, 0
, 1/inverse.var)
post.alpha.samp <- rnorm(n = 100000, mean =
post.alpha.mean,
```

```

sd = sqrt(post.alpha
           .var))

post.prob.tox.samp <- alpha[mtox.sel,1]^post.
alpha.samp

prob.too.toxic <- mean(post.prob.tox.samp >
tox.lim)

if(is.na(prob.too.toxic)) {
  prob.too.toxic <- 1
}

# stopping rule for when lowest dose is too
# toxic AND lowest dose has been tested ptox
# .hat[1] > 0.5 & 1 %in% dose

# stopping rule if dose is being recommended
# for a xth time

if (length(dlt) == n | (prob.too.toxic > tox.
cert) & 1 %in% dose | sum(dose == comb.
curr) == mtd.lim) {

stoprule = 0

# once all patients recruited calculate
# likelihoods at max weight

```

```
like.tox = est.tox = rep(0, nord.tox)

for (k in 1:nord.tox) {

  #set all the weights at 1
  weights <- rep(1, length(tox))
  #ensures that all those predicted to have
  #toxic event is included in dlt data
  # as we are assuming full follow up once
  #the last patients are recruited

  dlt <- tox

  beta <- alpha[k,][dose[1:length(dose)]]

  est.tox[k] <- optimize(f = bcrm1,
    interval = c(0,500),
    p1 = beta, y = tox
    , w = weights,
    maximum = T)$
    maximum

  like.tox[k] <- optimize(f = bcrm1,
    interval = c(0,500),
    p1 = beta, y =
    tox, w =
    weights,
    maximum = T)$
    objective

}
```

```
postprob.tox = (exp(like.tox) * mprior.tox)
               /sum(exp(like.tox) * mprior.tox)

if (nord.tox > 1) {
  mtox.sel = which.is.max(round(postprob.
    tox,8))
}

else {
  mtox.sel = 1
}

ptox.hat = alpha[mtox.sel, ]^est.tox[mtox.
  sel]

loss = abs(ptox.hat - theta)

comb.curr = which.is.max(round(-loss,8))

# obtain the last dose level from the data
last.lev <- tail(dose, n = 1)

if (comb.curr %in% dose | comb.curr < last.
  lev){
  # If the next recommended level has
  # already been tested or is lower than
  # the current dose level than recommend
  # next level
  comb.curr <- comb.curr
}

else {
```

```
    comb.curr <- max(dose) + 1
}

# Calculate asymptotic variance
beta <- alpha[mtox.sel,][dose]
inverse.var <- 0

for (i in 1:length(tox)) {
  inverse.var <- inverse.var + ((1-tox[i])*
    (weights[i]*beta[i]^est.tox[mtox.sel])
    *(log((weights[i]*beta[i])))^2)/(1-(
    weights[i]*beta[i]^est.tox[mtox.sel]))
    )^2
}

post.alpha.mean <- est.tox[mtox.sel]
post.alpha.var <- ifelse(inverse.var == 0,
  0 , 1/inverse.var)
post.alpha.samp <- rnorm(n = 100000, mean =
  post.alpha.mean ,
  sd = sqrt(post.
  alpha.var))

post.prob.tox.samp <- alpha[mtox.sel,1]^
  post.alpha.samp
prob.too.toxic <- mean(post.prob.tox.samp >
  tox.lim)
```

```

if(is.na(prob.too.toxic)) {
  prob.too.toxic <- 1
}

break
}

else {
  # Takes draws from a binomial distribution
  # to determine if patients in the cohort
  # had a toxic outcome
  cohort_tox <- stats::rbinom(n = cohort,
    size = 1, prob = r[comb.curr])
  cohort_dlt <- rep(0, cohort)
  # Patients in the same cohort receive the
  # same dose
  cohort_comb <- rep(comb.curr, cohort)
  # Follow up time based on the time taken to
  # recruit a patient
  # First patient assumed to be recruited
  # instantly
  # Add the min fu time to indicate when
  # analysis is done
}

```

```
cohort_fu <- (rectime*(cohort-1))-(rectime
  * 0:(cohort - 1)) +rectime
# Time of DLT is recorded and set between
# the time the patient has been followed
# up and the remaining observation window
cohort_dlt_time <- vector()
for (i in 1:cohort) {
  if(cohort_tox[i] == 0){
    cohort_dlt_time[i] <- NA
  }
  else {
    cohort_dlt_time[i] <- runif(1,0, obswin
      )
  }
}
# Stores the time DLT occurs for new cohort
dlt_time <- c(dlt_time, cohort_dlt_time)
# Stores new cohorts toxicity
tox <- c(tox, cohort_tox)
# Stores dose level for each new cohort
dose <- c(dose, cohort_comb)
# Store dummy dlt for new cohort
dlt <- c(dlt, cohort_dlt)
# Add on follow up time and recruitment
# time for previous cohorts
fu <- fu + (cohort * rectime)
```

```

#duration <- duration + (cohort * rectime)
+ minfu

# store new cohorts follow up time
fu <- c(fu, cohort_fu)

#duration <- c(duration, cohort_fu)

for(i in 1:length(fu)){
  if(tox[i] == 1){
    if(fu[i] >= dlt_time[i]){
      fu[i] <- obswin
      dlt[i] <- 1
    }
  }
}

fu <- pmin(fu, obswin)

# Calculated weights where the min fu
accounts for 60% and 80% by win2
for (i in 1:length(fu)) {
  weights[i] <- 0.6*(min(fu[i], minfu)/minfu
  ) +
  0.2*max((min(win2 - minfu, fu[i]-minfu)
  /(win2 - minfu)), 0) +
  0.2*max((max(fu[i]- win2, 0)/(obswin -
  win2)), 0)
}

```

```

y[comb.curr] = y[comb.curr] + sum(cohort_
tox)

npts[comb.curr] = npts[comb.curr] + cohort
}

}

if (stoprule == 0) {
  if (!exists("prob.too.toxic")){prob.too.toxic
    <- 0}

  if((prob.too.toxic > tox.cert)& 1 %in% dose){
    stop.count <- 1
  }

  else {
    comb.select[comb.curr] = comb.select[comb.
      curr] + 1
  }
}

# Calculate the duration of the trial

num_cohorts <- length(tox) / cohort
duration <- cohort_fu[1] + (rectime*(cohort)) * (
  num_cohorts -1) + (obswin)
return(list(MTD.selection = comb.select, tox.data
= y,

```

```

    patient.allocation = npts, duration =
        duration,
        stop = stop.count))
}

lpocrm.sim <- function(nsim) {
  ncomb = length(r)
  y <- npts <- matrix(nrow = nsim, ncol = ncomb)
  comb.select <- matrix(nrow = nsim, ncol = ncomb )
  duration <- stop.count <- trialsize <- rep(0, nsim)
  nstop = 0
  for (i in 1:nsim) {
    #print(i)
    result <- lpocrm(r, alpha, prior.o, x0, stop, n,
                      theta)
    comb.select[i, ] = result$MTD.selection
    y[i, ] = result$tox.data
    npts[i, ] = result$patient.allocation
    trialsize[i] = sum(result$patient.allocation)
    duration[i] = result$duration # convert duration
    to months
    stop.count[i] = result$stop
  }
  return(list(true.prob = r,
              MTD.selection = round(colMeans(comb.
                select),2),

```

```

    patient.allocation = 100 * round(
        colMeans(npts)/mean(trialsizes),2),
    percent.DLT = sum(colMeans(y))/mean(
        trialsizes),
    months = 12*mean(duration)/365,
    stop = mean(stop.count),
    mean.n = mean(trialsizes),
    mean.n.perdose = colMeans(npts),
    acceptable = sum(colMeans(comb.select)[
        which(round(abs(r - theta), 2) <=
        tox.range)])))
}

if (nsim == 1) {
    twostgcrm(r, x0, stop, n, theta)
}
else {
    lpocrm.sim(nsim)
}
}

```

B.2 Extension to Phase I/II design chapter code

The code below is for various functions that were used to run the simulations in Chapter 3.

```

####install required R packages

library(binom)

library(nnet)

```

```
library(dfcrm)
# pwr calcs
library(pwr)
library(matrixStats)

# Written by KB to split out method calculation and
# simulation

post.tox <- function(a, p, y, n) {
  s2=1.34
  lik=1
  for(j in 1:length(p)){
    pj=p[j]**exp(a)
    lik=lik*pj^y[j]*(1-pj)^(n[j]-y[j]);
  }
  return(lik*exp(-0.5*a*a/s2));
}

# the posterior mean of ptox
posttoxf <- function(a, p, y, n, j) {
  p[j]^(exp(a))*post.tox(a, p, y, n);
}

post.eff <- function(b, q, z, n) {
  s2=1.34
  lik=1
  for(j in 1:length(q)){
    qj=q[j]**exp(b)
```

```

lik=lik*qj^z[j]*(1-qj)^(n[j]-z[j]);
}

return(lik*exp(-0.5*b*b/s2));
}

# the posterior mean of peff
postefff <- function(b, q, z, n, j) {
  q[j]^(exp(b))*post.eff(b, q, z, n);
}

wages.tait = function(y, z, n, p.skel, q.skel, n.ar,
  comb.curr = NULL,
  mprior.tox = NULL, mprior.eff =
  NULL,
  safety.confidence = 0.95,
  futility.confidence = 0.95,
  check.tox.at.dose.level,
  lowest.is.placebo = FALSE,
  placebo.rand.prob = NULL) {

# y, number of toxicity events at each dose
# z, number of efficacy events at each dose
# n, number of patients allocated to each dose
# p.skel, toxicity skeleton(s) as vector or matrix
# q.skel, efficacy skeleton(s) as vector or matrix
# n.ar, the number of patients to use in the adaptive
randomisation stage
}

```

```
# comb.curr, deprecated, ignored
# mprior.tox, prior for each toxicity ordering
# mprior.eff, prior for each efficacy ordering
# safety.confidence is the % CI to construct when
#       checking whether lowest dose
#           is too toxic
# futility.confidence is the % CI to construct when
#       checking whether current
#           dose is too toxic
# check.tox.at.dose.level is the index of the dose on
#       which design
#       should invoke the logic to detect excess toxicity
#           . In the majority of
#       cases this will be dose-level 1. If you have a
#           trivial lowest dose ("d=0")
#       so that toxicity is nearly impossible, you will
#           want to check for excess
#       tox at dose 2 instead (lest you short-circuit the
#           early stopping criteria)
#       This is KB's embellishment.

# lowest.is.placebo is an indicator to identify if
#       the lowest dose is set to
#           be a control/placebo dose
# placebo.rand.prob is the probability of allocating
#       to the placebo/control
#       dose level
```

```
comb.curr = NULL

if(is.vector(p.skel)) p.skel = t(as.matrix(p.skel));
if(is.vector(q.skel)) q.skel = t(as.matrix(q.skel));

ncomb = ncol(p.skel);      # number of doses or
                           combinations

ptox.hat = numeric(ncomb); # estimate of toxicity
                           prob

peff.hat = numeric(ncomb); # estimate of efficacy
                           prob

nord.tox = nrow(p.skel);

if(is.null(mprior.tox))
  mprior.tox = rep(1 / nord.tox, nord.tox)

nord.eff = nrow(q.skel);
if(is.null(mprior.eff))
  mprior.eff = rep(1 / nord.eff, nord.eff)

marginal.tox = rep(0, nord.tox);
for(k in 1:nord.tox) {
  marginal.tox[k] = integrate(post.tox, lower=-Inf,
                               upper=Inf, p=p.skel[,],
                               y=y, n=n)$value;
}
postprob.tox = (marginal.tox * mprior.tox) / sum(
  marginal.tox * mprior.tox);
```

```
marginal.eff = rep(0, nord.eff);
for(k in 1:nord.eff) {
  marginal.eff[k] = integrate(post.eff, lower=-Inf,
    upper=Inf, q=q.skel[k,],
    z=z, n=n)$value;
}
postprob.eff = (marginal.eff * mprior.eff) / sum(
  marginal.eff * mprior.eff);

# toxicity model selection, identify the model with
# the highest posterior prob
if(nord.tox > 1) {
  mtox.sel = which.is.max(postprob.tox);
}
else {
  mtox.sel = 1;
}

# efficacy model selection, identify the model with
# the highest posterior prob
if(nord.eff > 1){
  meff.sel = which.is.max(postprob.eff);
}
else {
  meff.sel = 1;
}
```

```
# calculate posterior mean of toxicity probability at
# each combo
for(j in 1:ncomb){
  ptox.hat[j] = integrate(posttoxf, lower=-Inf, upper
=Inf, p.skel[mtox.sel,],
                         y, n, j)$value / marginal.
                         tox[mtox.sel];
}

# calculate posterior mean of efficacy probability at
# each combo
for(j in 1:ncomb){
  peff.hat[j] = integrate(postefff, lower=-Inf, upper
=Inf, q.skel[meff.sel,],
                         z, n, j)$value / marginal.
                         eff[meff.sel];
}

# determine set of safe doses based on toxicity
aset = which(ptox.hat <= tul)
if(length(aset)==0) {
  aset = which.min(ptox.hat)
}

peff.hat.aset = rep(0, ncomb)
peff.hat.aset[aset] = peff.hat[aset]
```

```

# determine randomisation probabilities based on
# efficacy

ar.prob = peff.hat.aset / sum(peff.hat.aset)

if(lower.is.placebo == TRUE){
  ar.prob = c(placebo.rand.prob,
             ar.prob[-1]*(1-placebo.rand.prob)/sum(
               ar.prob[-1]))
}

if(length(aset) == 1) {
  # The best and recommended dose can only be this
  comb.best <- aset
  comb.curr <- aset
}
else {
  # The best dose is always that in admissible set
  # with maximal Pr(Eff)
  comb.best <- which.max(peff.hat.aset)
  # In AR stage, sample current dose from admissible
  # doses with weighted prob;
  # After AR stage, current dose is best dose
  ifelse(sum(n) < n.ar,
        comb.curr <- sample(1:ncomb, 1, prob=ar.prob
                           ),
        comb.curr <- comb.best)
}

```

```
}

max.dose.given <- max(seq(1, ncomb)[n > 0])

if(comb.curr > max.dose.given + 1) {

  # Recommending dose above maximum given, so prevent
  # skipping

  comb.curr <- max.dose.given + 1

  skipping_prevented = TRUE

} else {

  skipping_prevented = FALSE

}

#####
# Stopping rules

stop = 0

tox.dl = check.tox.at.dose.level

safety = binom.confint(y[tox.dl], n[tox.dl], conf.

  level = safety.confidence,
  methods="exact")$lower

if(safety > tul){

  stop = 1

}

if(sum(n) > n.ar){

  # We are not in the AR phase, so stopping for
  # futility is used.

  futility = binom.confint(z[comb.curr], n[comb.curr
  ] ,
```

```

conf.level = futility.

confidence,

methods = "exact")$upper

if(n[comb.curr] > 0 & futility < ell){

  # Recommended dose has been given; upper efficacy
  # estimate is below limit

  # stop = 2 # KB - surely an error? The stop
  # value is added below to a previous value
  # so it should only be 0 or 1? Maybe the logic NW
  # wanted was ">0 => stop".

  stop = 1

}

}

else {

  futility = NULL

}

return(list(
  # marginal.tox = marginal.tox,
  ToxBayesFactor = postprob.tox,
  ProbTox = ptox.hat,
  ToxSkeleton = mtox.sel,
  # marginal.eff = marginal.eff,
  EffBayesFactor = postprob.eff,
  EffSkeleton = meff.sel,
  ProbEff = peff.hat,

```

```

AdaptiveRandProb = ar.prob,
RecommendedDose = comb.curr,
OptimalDose = comb.best,
Dose1ToxLowerBound = safety,
LowestActiveDoseToxLowerBound = safety,
RecommendedDoseEffUpperBound = futility,
AdmissibleSet = aset,
Stop = stop,
SkippingPrevented = skipping_prevented,
Note = "The parameter Dose1ToxLowerBound is a
misnomer.

      Use LowestActiveDoseToxLowerBound instead."
))

}

```

```

####Load the function 'bpocrm'
wt.sim.one <- function(p0, q0, p.skel, q.skel, tul, ell
, cohortsize, ncohort,
start.comb, n.ar, mprior.tox =
NULL, mprior.eff = NULL,
safety.confidence = 0.95,
futility.confidence = 0.95,
check.tox.at.dose.level,
lowest.is.placebo = FALSE,
placebo.rand.prob = NULL) {

```

```
# p0, true toxicity probabilities
# q0, true efficacy probabilities
# p.skel, toxicity skeleton(s)
# q.skel, efficacy skeleton(s)
# tul, toxicity upper limit
# ell, efficacy lower limit
# cohortsize, obvious
# ncohort, number of cohorts
# start.comb, starting dose-level
# n.ar, number of patients in adaptive randomisation
# stage
# mprior.tox, Prior weights to toxicity models,
# uniform if omitted
# mprior.eff, Prior weights to efficacy models,
# uniform if omitted
# safety.confidence is the % CI to construct when
# checking whether lowest dose
# is too toxic
# futility.confidence is the % CI to construct when
# checking whether current
# dose is too toxic
# check.tox.at.dose.level is the index of the dose on
# which design
# should invoke the logic to detect excess toxicity
# . In the majority of
```

```
# cases this will be dose-level 1. If you have a
# trivial lowest dose ("d=0")
#
# so that toxicity is nearly impossible, you will
# want to check for excess
#
# tox at dose 2 instead (lest you short-circuit the
# early stopping criteria)
#
# This is KB's embellishment.

# if a single ordering is inputed as a vector,
# convert it to a matrix
if(is.vector(p.skel)) p.skel = t(as.matrix(p.skel));

# if a single ordering is inputed as a vector,
# convert it to a matrix
if(is.vector(q.skel)) q.skel = t(as.matrix(q.skel));

## run a trial
ncomb = ncol(p.skel);    # number of combos
y = rep(0, ncomb);      # number of toxicity/responses at
# each dose level
z = rep(0, ncomb);      # number of efficacy at each
# dose level
n = rep(0, ncomb);      # number of treated patients at
# each dose level
comb.curr = start.comb; # current dose level
ptox.hat = numeric(ncomb); # estimate of toxicity
prob
```

```

peff.hat = numeric(ncomb); # estimate of efficacy

prob

comb.select = rep(0, ncomb); # a vector of indicators

for dose selection

stop = 0; # indicate if trial stops early

tox.skel = rep(0, nrow(p.skel))

eff.skel = rep(0, nrow(q.skel))

for(i in 1:ncohort)

{

  # generate data for a new cohort of patients

  y[comb.curr] = y[comb.curr] + rbinom(1, cohortsize,
    p0[comb.curr]);

  z[comb.curr] = z[comb.curr] + rbinom(1, cohortsize,
    q0[comb.curr]);

  n[comb.curr] = n[comb.curr] + cohortsize;

  # TODO (... something - but what?!)

  cohort.update = wages.tait(y, z, n, p.skel, q.skel,
    n.ar,

                                comb.curr = comb.curr,
                                mprior.tox = mprior.tox,
                                mprior.eff = mprior.

                                eff,
                                safety.confidence =
                                safety.confidence,

```

```
futility.confidence =
    futility.confidence,
    check.tox.at.dose.level
    = check.tox.at.dose.
        level,
lowest.is.placebo =
    lowest.is.placebo,
placebo.rand.prob =
    placebo.rand.prob
)

# cat('ToxSkel', cohort.update$ToxSkeleton, '\n')
# cat('EffSkel', cohort.update$EffSkeleton, '\n')

stop = cohort.update$Stop
comb.curr = cohort.update$RecommendedDose
tox.skel[cohort.update$ToxSkeleton] = tox.skel[
    cohort.update$ToxSkeleton] +1
eff.skel[cohort.update$EffSkeleton] = eff.skel[
    cohort.update$EffSkeleton] +1

if(stop > 0)
    break
}

if(stop == 0){
    comb.select[comb.curr] = comb.select[comb.curr] +
        1;
```

```

}

# cat('FinalToxSkel', cohort.update$ToxSkeleton, '\n
')

# cat('FinalEffSkel', cohort.update$EffSkeleton, '\n
')

return(list(comb.select = comb.select, tox.data = y,
           eff.data = z,
           pt.allocation = n, stop = stop,
           ToxSkel = tox.skel, FinalToxSkel = cohort
             .update$ToxSkeleton,
           EffSkel = eff.skel, FinalEffSkel = cohort
             .update$EffSkeleton))

}

# Nolan originally called this bpocrm
bpocrm = wt.sim.one

## ## Load the function 'bpocrm.sim'
wt.sim <- function(p0, q0, p.skel, q.skel, tul, ell,
                    cohortsize, ncohort,
                    start.comb, n.ar, ntrial,
                    mprior.tox = NULL, mprior.eff = NULL
                    ,
                    safety.confidence = 0.95, futility.
                    confidence = 0.95,

```

```
check.tox.at.dose.level,
verbose = TRUE, really.verbose =
FALSE,
full_output = TRUE,
lowest.is.placebo = FALSE, placebo.
rand.prob = NULL,
effect.sizes = NULL) {

# p0, true toxicity probabilities
# q0, true efficacy probabilities
# p.skel, toxicity skeleton(s)
# q.skel, efficacy skeleton(s)
# tul, toxicity upper limit
# ell, efficacy lower limit
# cohortsize, obvious
# ncohort, number of cohorts
# start.comb, starting dose-level
# nar, number of patients in adaptive randomisation
stage
# ntrial, number of trials to simulate
# mprior.tox, Prior weights to toxicity models,
uniform if omitted
# mprior.eff, Prior weights to efficacy models,
uniform if omitted
# safety.confidence is the % CI to construct when
checking whether lowest dose
# is too toxic
```

```

# futility.confidence is the % CI to construct when
# checking whether current
# dose is too toxic
# check.tox.at.dose.level is the index of the dose on
# which design
# should invoke the logic to detect excess toxicity
# . In the majority of
# cases this will be dose-level 1. If you have a
# trivial lowest dose ("d=0")
# so that toxicity is nearly impossible, you will
# want to check for excess
# tox at dose 2 instead (lest you short-circuit the
# early stopping criteria)
# This is KB's embellishment.

```

```
StartTime = Sys.time()
```

```

# if a single ordering is inputed as a vector,
# convert it to a matrix
if(is.vector(p.skel)) p.skel = t(as.matrix(p.skel));
# if a single ordering is inputed as a vector,
# convert it to a matrix
if(is.vector(q.skel)) q.skel = t(as.matrix(q.skel));

ncomb = length(p0)
comb.select <- y <- z <- n <- matrix(nrow = ntrial, ncol
= ncomb)
```

```
tox.skel <- matrix(nrow = ntrial, ncol = nrow(p.skel))
)
final.tox.skel <- rep(0, nrow(p.skel))
print(final.tox.skel)

eff.skel <- matrix(nrow = ntrial, ncol = nrow(q.skel))
)
final.eff.skel <- rep(0, nrow(q.skel))

nstop = 0

h <- effect.sizes

npower <- matrix(nrow = ntrial, ncol = length(h))

for(i in 1:ntrial){

  if(verbose & i %% floor(ntrial / 10) == 0) print(i)

  result <- bpocrm(p0, q0, p.skel, q.skel, tul, ell,
  cohortsize, ncohort,
  start.comb, n.ar, mprior.tox =
  mprior.tox,
  mprior.eff = mprior.eff,
  safety.confidence = safety.
  confidence,
  futility.confidence = futility.
  confidence,
  check.tox.at.dose.level = check.
  tox.at.dose.level,
  lowest.is.placebo = lowest.is.
  placebo,
```

```

placebo.rand.prob = placebo.rand.

prob)

if(result$stop == 0) {

  comb.select[i,] = result$comb.select

} else {

  comb.select[i,] = 0

}

y[i, ] <- result$tox.data

z[i, ] <- result$eff.data

n[i, ] <- result$pt.allocation

tox.skel[i, ] <- result$ToxSkel

final.tox.skel[result$FinalToxSkel] <- final.tox.

skel[result$FinalToxSkel]+1

eff.skel[i, ] <- result$EffSkel

final.eff.skel[result$FinalEffSkel] <- final.eff.

skel[result$FinalEffSkel]+1

nstop <- nstop + result$stop

}

if(result$stop == 0 & result$comb.select[1] != 1){

  if(result$pt.allocation[1] > 0 &

  result$pt.allocation[result$comb.select == 1]

  >0 ){

    for (j in 1:length(h)) {

```

```
    npower[i,j] <- pwr.2p2n.test(h = h[j], n1 =
      result$pt.allocation[1],
      n2 = result$pt.
      allocation[
      result$comb.
      select == 1],
      sig.level = 0.1)
      $power
    }
  }

}

if(really.verbose) {
  cat("True_tox_probability:", round(p0,3), sep="\\u00u
  ", "\\n");
  cat("True_eff_probability:", round(q0,3), sep="\\u00u
  ", "\\n");
  cat("selection_percentage:", formatC(colMeans(comb
  .select)*100, digits=1,
  format="f"),
  sep="\\u00u
  ",
  , "\\n");
}
```

```
cat("number of toxicities: ", formatC(colMeans(y  
) , digits=1, format="f"),  
sep=" ", "\n");  
  
cat("number of responses: ", formatC(colMeans(z)  
, digits=1, format="f"),  
sep=" ", "\n");  
  
cat("number of patients treated: ", formatC(  
colMeans(n), digits=1,  
format  
=  
"  
f  
"  
)  
,  
sep  
=  
"  
 "  
 "  
 "  
 "  
"  
,  
"\n");
```

```
cat("percentage of stop:", nstop/ntrial*100, "\\n");
}

# Return object
l <- list(
  TrueTox = p0,
  TrueEff = q0,
  ToxSkel = p.skel,
  EffSkel = q.skel,
  TUL = tul,
  ELL = ell,
  NumSims = ntrial,
  NumPatients = cohortsize * ncohort,
  CohortSize = cohortsize,
  NumCohorts = ncohort,
  SizeARPhase = n.ar,
  SafetyConfidence = safety.confidence,
  FutilityConfidence = futility.confidence,
  TreatedAtDose = round(colMeans(n), 1),
  ToxAtDose = round(colMeans(y), 1),
  EffAtDose = round(colMeans(z), 1),
  ProbSelect = round(colMeans(comb.select), 4),
  NStop = nstop,
  ProbStop = nstop / ntrial,
  EffectSizes = h,
  PowerMean = round(colMeans(npower, na.rm = T), 4),
```

```

PowerSD = round(colsds(npower, na.rm = T), 4),
NPower = sum(!is.na(npower[,1])),
CohortToxSkel = tox.skel,
ProbFinalToxSkel = final.tox.skel / ntrial,
CohortEffSkel = eff.skel,
ProbFinalEffSkel = final.eff.skel / ntrial,
SimulationTime = Sys.time() - StartTime
)

# Full output includes patients
if(full_output) {
  l$FullTreatedAtDose = n
  l$FullToxAtDose = y
  l$FullEffAtDose = z
  l$FullRecommendation = comb.select
  l$FullPower = npower
  # l$FullStopTrial = stop_trial
}

return(l)
}

# Nolan originally called this bpocrm.sim
bpocrm.sim = wt.sim

```

B.3 TITE-DTP chapter code

The code below was used to generate the DTPs used as examples in Chapter 4.

```
library(dfcrm)
library(escalation)
library(dplyr)
library(kableExtra)
library(dtprm)
library(tidyr)
library(diagram)
library(DiagrammeRsvg)
library(rsvg)

# Set-up
skeleton <- getprior(target = 0.25, nu = 4, nlevel = 5,
halfwidth = 0.05)
target <- 0.25

model <- get_dfcrm(skeleton = skeleton, target = target
)

paths <- model %>%
get_dose_paths(cohort_sizes = c(3,3,3), next_dose = 2)

#DTPs
spread_paths(as_tibble(paths)) %>%
select(o0 = 'outcomes0', d0 = 'next_dose0',
o1 = 'outcomes1', d1 = 'next_dose1',
o2 = 'outcomes2', d2 = 'next_dose2',
o3 = 'outcomes3', d3 = 'next_dose3') %>%
```

```

print(n=100)

# Node plot
graph_paths(paths, RColorBrewer_palette = 'Set1') %>%
  export_svg() %>%
  chartToRaw %>%
  rsvg_pdf('C:/Users/Amit/Documents/GitHub/PhD/Thesis/
  Figures/TITE-DTP-InitialExampleDTPNode.pdf')

# Code for the LaTeX table
spread_paths(as_tibble(paths)) %>%
  select(o0 = 'outcomes0', d0 = 'next_dose0',
         o1 = 'outcomes1', d1 = 'next_dose1',
         o2 = 'outcomes2', d2 = 'next_dose2',
         o3 = 'outcomes3', d3 = 'next_dose3') %>%
  print(n=100) %>%
  data.frame() %>%
  mutate(o0 = 1:64) %>%
  kable('latex', booktabs = T, linesep = "", align = "c",
        '',
        col.names = c('Pathway', 'Dose', 'Outcomes', '',
                     'Dose', 'Outcomes', 'Dose',
                     'Outcomes', 'Dose'),
        caption = '\\label{tab_tite-dtp:
  InitialDTPExample}Initial DTP for the first
  three cohorts of our example CRM.')
) %>%

```

```
kable_styling(latex_options = c("striped", "HOLD_"
  position", "scale_down"),
  position = "center", font_size = 8) %>%
add_header_above(c(, 'Cohort\u20d71' = 2, 'Cohort\u20d72' =
  2, 'Cohort\u20d73' = 2,
  'Cohort\u20d74' = 1)) %>%
cat()

# Check using dtpcrm
calculate_dtps(2, cohort_sizes = c(3, 3, 3), dose_func
= applied_crm,
prior = skeleton, target =
target, no_skip_esc = FALSE)

# matches results from escalation

# DTP flow diagram

calculate_dtps(2, cohort_sizes = c(3, 3, 3), dose_func
= applied_crm,
prior = skeleton, target = target, no_
skip_esc = FALSE) %>%
dtpflow()

# #######
```

```

# DTPs with stopping rules

model1 <- get_dfcrm(skeleton = skeleton, target =
target) %>%
  dont_skip_doses(when_escalating = TRUE) %>%
  stop_when_too_toxic(dose = 1, tox_threshold = 0.35,
confidence = 0.9)

paths <- model1 %>%
  get_dose_paths(cohort_sizes = c(3,3,3), next_dose = 2)

# DTPs
spread_paths(as_tibble(paths)) %>%
  select(o0 = 'outcomes0', d0 = 'next_dose0',
         o1 = 'outcomes1', d1 = 'next_dose1',
         o2 = 'outcomes2', d2 = 'next_dose2',
         o3 = 'outcomes3', d3 = 'next_dose3') %>%
  print(n=100)

# Node plot
graph_paths(paths, RColorBrewer_palette = 'Set1') %>%
  export_svg() %>%
  chartToRaw %>%
  rsvg_pdf('C:/Users/Amit/Documents/GitHub/PhD/Thesis/
Figures/TITE-DTP-UpdatedExampleDTPNode.pdf')

# Code for the LaTeX table

```

```

spread_paths(as_tibble(paths)) %>%
  select(o0 = 'outcomes0', d0 = 'next_dose0',
         o1 = 'outcomes1', d1 = 'next_dose1',
         o2 = 'outcomes2', d2 = 'next_dose2',
         o3 = 'outcomes3', d3 = 'next_dose3') %>%
  print(n=100) %>%
  data.frame() %>%
  mutate(o0 = 1:55) %>%
  replace_na(list(d2 = 'STOP', d3 = 'STOP')) %>%
  kable('latex', booktabs = T, linesep = "", align = "c",
        '',
        col.names = c('Pathway', 'Dose', 'Outcomes', 'Dose',
                     'Outcomes', 'Dose',
                     'Outcomes', 'Dose),
        caption = '\\label{tab_tite-dtp:\\
          UpdatedDTPExample}Updated DTPs for the first\\
          three cohorts of our example CRM with\\
          additional rules.')

) %>%
kable_styling(latex_options = c("striped", "HOLD_\\
position", "scale_down"),
              position = "center", font_size = 4) %>%
add_header_above(c('' , 'Cohort\\_1' = 2, 'Cohort\\_2' =
2, 'Cohort\\_3' = 2,
'Cohort\\_4' = 1)) %>%
cat()

```

```
# Check using dtpcrm

stop_func <- function(x) {
  x = stop_for_excess_toxicity_empiric(x, tox_lim =
  0.35, prob_cert = 0.90,
  dose = 1, nsamps
  = 10000)
}

calculate_dtps(2, cohort_sizes = c(3, 3, 3), dose_func
= applied_crm,
prior = skeleton, target = target, stop_
func = stop_func,
global_coherent_esc = FALSE)

# matches results from escalation

# DTP flow diagram

calculate_dtps(2, cohort_sizes = c(3, 3, 3), dose_func
= applied_crm,
prior = skeleton, target = target, stop_
func = stop_func,
global_coherent_esc = FALSE) %>%
dtpflow()

prior <- c(0.05, 0.10, 0.20, 0.35, 0.50, 0.70)
target <- 0.2
```

```

level <- c(2, 2, 2, 3, 3, 3, 4, 4)
y <- c(0, 0, 0, 0, 0, 0, 0, 1)
foo <- crm(skeleton, target, y, level)
summary(foo)

x0 <- c(2, 2, 2, 3, 3, 3, 4, 4, rep(5, 24)) # initial design
foo <- cohere(skeleton, target, x0)
foo$message

#
#####
# DTPs with stopping rules for next 3 cohorts after 2
NNN 3NNTNNT

model2 <- get_dfcrm(skeleton = skeleton, target =
target) %>%
  dont_skip_doses(whenEscalating = TRUE) %>%
  stopWhenTooToxic(dose = 1, toxThreshold = 0.35,
confidence = 0.9)

paths <- model2 %>%
  getDosePaths(cohortSizes = c(3, 3, 3),
previousOutcomes = '2NNN\u22253NNTNNT')

# DTPs
spreadPaths(as_tibble(paths)) %>%

```

```

select(o0 = 'outcomes0', d0 = 'next_dose0',
       o1 = 'outcomes1', d1 = 'next_dose1',
       o2 = 'outcomes2', d2 = 'next_dose2',
       o3 = 'outcomes3', d3 = 'next_dose3') %>%
print(n=100)

# Node plot
graph_paths(paths, RColorBrewer_palette = 'Set1') %>%
export_svg() %>%
chartToRaw %>%
rsvg_pdf('C:/Users/Amit/Documents/GitHub/PhD/Thesis/
Figures/TITE-DTP-UsingDuringTrialDTPNode.pdf')

# Code for the LaTeX table
spread_paths(as_tibble(paths)) %>%
select(o0 = 'outcomes0', d0 = 'next_dose0',
       o1 = 'outcomes1', d1 = 'next_dose1',
       o2 = 'outcomes2', d2 = 'next_dose2',
       o3 = 'outcomes3', d3 = 'next_dose3') %>%
print(n=100) %>%
data.frame() %>%
mutate(o0 = 1:64) %>%
#replace_na(list(d2 = 'STOP', d3 = 'STOP')) %>%
kable('latex', booktabs = T, linesep = "", align = "c",
      ,
      col.names = c('Pathway', 'Dose', 'Outcomes', ,
Dose', 'Outcomes', 'Dose', ,

```

```

        'Outcomes', 'Dose')),

caption = '\\label{tab_tite-dtp:

UsingDuringTrialDTPs4-7}DTPs for three additional cohorts after observing outcomes for the first three cohorts.'

) %>%
kable_styling(latex_options = c("striped", "HOLD_"
position", "scale_down"),
position = "center", font_size = 4) %>%
add_header_above(c('' , 'Cohort 4' = 2, 'Cohort 5' =
2, 'Cohort 6' = 2,
'Cohort 7' = 1)) %>%
cat()

# non uniform cohorts

paths <- model2 %>%
get_dose_paths(cohort_sizes = c(2,1,2),
previous_outcomes = '2NNN|3NNTNNT')

graph_paths(paths, RColorBrewer_palette = 'Set1') %>%
export_svg() %>%
charToRaw %>%
rsvg_pdf('C:/Users/Amit/Documents/GitHub/PhD/Thesis/
Figures/TITE-DTP-NonUniformDTPNode.pdf')

# Code for the LaTeX table

spread_paths(as_tibble(paths)) %>%

```

```

select(o0 = 'outcomes0', d0 = 'next_dose0',
       o1 = 'outcomes1', d1 = 'next_dose1',
       o2 = 'outcomes2', d2 = 'next_dose2',
       o3 = 'outcomes3', d3 = 'next_dose3') %>%
print(n=100) %>%
data.frame() %>%
mutate(o0 = 1:36) %>%
#replace_na(list(d2 = 'STOP', d3 = 'STOP')) %>%
kable('latex', booktabs = T, linesep = "", align = "c",
      ,
      col.names = c('Pathway', 'Dose', 'Outcomes', ,
                    'Dose', 'Outcomes', 'Dose',
                    'Outcomes', 'Dose'),
caption = '\\label{tab_tite-dtp:NonUniformDTPs4
-7}DTPs for three additional cohorts with
varying cohort sizes after observing
outcomes for the first three cohorts.'
) %>%
kable_styling(latex_options = c("striped", "HOLD_"
position", "scale_down"),
position = "center", font_size = 4) %>%
add_header_above(c(' ', 'Cohort_4' = 2, 'Cohort_5' =
2, 'Cohort_6' = 2,
'Cohort_7' = 1)) %>%
cat()

```

The code below was used to generate the TITE-DTPs in Chapter 4.

```
library(dfcrm)
```

```
library(dplyr)
library(tidyr)
library(ggplot2)
library(kableExtra)
library(gtools)
library(rgl)
library(magick)
library(ggrepel)
library(ggpubr)

skeleton <- getprior(target = 0.25, nu = 4, nlevel = 5,
                      halfwidth = 0.05)
target <- 0.25
obswin <- 35
#
#####
# Cohorts of 1
# T
level <- 2
tox <- 1
followup <- 35
mod <- titecrm(prior = skeleton, target = target, tox =
                 tox, level = level,
                 obswin = obswin, scheme = 'linear',
                 followup = followup)
mod$mtd
```

```
# N

level <- 2
tox <- 0
followup <- 1:obswin
results <- data.frame('Observed' = followup, 'Rec' =
rep(0,obswin))
for (i in 1:obswin) {
  followup <- i
  mod <- titecrm(prior = skeleton, target = target, tox
= tox, level = level,
  obswin = obswin, scheme = 'linear',
  followup = followup)
  results$Rec[i] <- mod$mtd
}

DTP_c1 <- results %>%
  mutate( Dose = 2,
  Pathway = Observed + 1,
  Outcome = paste0('N', '(', Observed, ')'),
  Outcome = if_else(Outcome == 'N(35)', 'N',
  Outcome)) %>%
  select(Pathway, Dose, Outcome, Rec) %>%
  add_row(Pathway = 1, Dose = 2, Outcome = 'T', Rec =
1) %>%
  arrange(Pathway)
```

```

DTP_c1 %>%
  kable('latex', booktabs = T, linesep = "", align = "c",
        ,
        col.names = c('Pathway', 'Dose', 'Outcomes',
                     , 'Dose'),
        caption = '\\label{tab_tite-dtp:TITEDTP_c1
}TITE-DTP for a cohort of 1.') %>%
  kable_styling(latex_options = c("striped", "HOLD_",
                                  position", "scale_down"),
                position = "center", font_size = 5) %>%
  add_header_above(c('' , 'Cohort_1' = 2, 'Cohort_2' =
1)) %>%
  cat()
#
#####
# Cohorts of 2
## TT
level <- c(2,2)
tox <- c(1,1)
followup <- c(35,35)
mod <- titecrm(prior = skeleton, target = target, tox =
tox, level = level,
               obswin = obswin, scheme = 'linear',
               followup = followup)
mod$mtd

```

```

## NT

level <- c(2,2)

tox <- c(1,0)

followupcombo <- cbind(rep(obswin, obswin), 1:obswin)

results <- data.frame('Observed' = 1:obswin, 'Rec' =
rep(0,obswin))

for (i in 1:obswin) {

  followup <- followupcombo[i,]

  mod <- titecrm(prior = skeleton, target = target, tox
  = tox, level = level,
  obswin = obswin, scheme = 'linear',
  followup = followup)

  results$Rec[i] <- mod$mtd

}

DTP_c2NT <- results %>%
  mutate( Dose = 2,
  Pathway = Observed,
  Outcome = paste0('N', '(', Observed, ')', 'T')
  ,
  Outcome = if_else(Outcome == 'N(35)T', 'NT',
  Outcome)) %>%
  select(Pathway, Dose, Outcome, Rec) %>%
  arrange(Pathway)

DTP_c2NT %>%

```

```

kable('latex', booktabs = T, linesep = "", align = "c
",
      col.names = c('Pathway', 'Dose', 'Outcomes', ,
                    Dose'),
      caption = '\\label{tab_tite-dtp:TITEDTP_c2NT}
TITE-DTP for a cohort of 2 for scenario NT.'
) %>%
kable_styling(latex_options = c("striped", "HOLD_"
position", "scale_down"),
              position = "center", font_size = 5) %>%
add_header_above(c('', 'Cohort_1' = 2, 'Cohort_2' =
1)) %>%
cat()

## NN

level <- c(2,2)
tox <- c(0,0)

combos <- combinations(n = 35, r = 2, repeats.allowed =
T, v = 1:35)
pos <- cbind(rep(0,nrow(combos)))
results <- pos[, rep(1, each=length(tox)+1)]
for (i in 1:nrow(combos)) {
  followup <- as.numeric(combos[i,])
  weights <- followup / obswin
  mod <- titecrm(prior = skeleton, target = target, tox
= tox, level = level,

```

```
        obswin = obswin, weights = weights)

for (j in 1:ncol(results)) {
  results[,j][i] <- followup[j]
  results[,ncol(results)][i] <- mod$mtd
}

results <- data.frame(results)
colnames(results) <- c('Patient1', 'Patient2', 'Rec')

results %>% mutate(TotalFollow = Patient1+Patient2) %>%
  group_by(Rec) %>%
  summarise(n = n(), min = min(TotalFollow), max = max(
    TotalFollow))

results %>% mutate(TotalFollow = Patient1+Patient2) %>%
  filter(TotalFollow == 20 | TotalFollow == 21) %>%
  select(Patient1, Patient2, TotalFollow, Rec) %>%
  arrange(desc(TotalFollow)) %>%
  kable('latex', booktabs = T, linesep = "", align = "c",
        ,
        col.names = c('Patient_1', 'Patient_2', 'Combined_Follow-up', 'Dose_Recommendation'),
```

```
caption = '\\label{tab_tite-dtp:TITEDTP_'
c2NNprob}Different dose recommendations with
overlapping combined follow-up times.')
%>%
kable_styling(latex_options = c("striped", "HOLD_"
position),
position = "center", font_size = 11)
%>%
add_header_above(c('Follow-up' = 2, ' ' = 2)) %>%
cat()

# calculate exact number of pathways for each specific
outcome

results <- results %>%
mutate(TF = Patient1 + Patient2)

results %>%
filter(TF <= 19) %>%
nrow()

results %>%
filter(TF >= 22) %>%
nrow()

results %>%
filter(TF == 20 & Patient2 <= 17) %>%
```

```
nrow()

results %>%
  filter(TF == 20 & Patient2 >= 18) %>%
  nrow()

results %>%
  filter(TF == 21 & Patient2 <= 14) %>%
  nrow()

results %>%
  filter(TF == 21 & Patient2 >= 15) %>%
  nrow()

# To produce a plot need to run on all possible
# combinations not just unique

level <- c(2,2)
tox <- c(0,0)
combos <- 1:obswin
combos <- expand.grid(combos, combos)
pos <- cbind(rep(0,nrow(combos)))
results <- pos[, rep(1, each=length(tox)+1)]
for (i in 1:nrow(combos)) {
  followup <- as.numeric(combos[i,])
  weights <- followup / obswin
```

```
mod <- titecrm(prior = skeleton, target = target, tox
= tox, level = level,
              obswin = obswin, weights = weights)

for (j in 1:ncol(results)) {
  results[,j][i] <- followup[j]
  results[,ncol(results)][i] <- mod$mtd
}

results <- data.frame(results)
colnames(results) <- c('Patient1', 'Patient2', 'Rec')

results %>%
  ggplot(aes(x = Patient1, y = Patient2, fill = as.
factor(Rec))) +
  geom_tile() +
  scale_fill_brewer(palette = 'Paired') +
  theme_bw() +
  geom_abline(intercept = 19, slope = -1, col = 'red',
,
            linetype = 'dashed') +
  geom_abline(intercept = 22, slope = -1, col = 'red',
,
            linetype = 'dashed') +
  theme(panel.border = element_blank(), panel.grid.
major = element_blank(),
```

```

    panel.grid.minor = element_blank(),
    axis.line = element_line(colour = "black")) +
  labs(fill = 'Recommended Dose', x = 'Patient 1',
       follow-up(Days),
       y = 'Patient 2 follow-up(Days') +
  scale_x_continuous(breaks = seq(0, 60, by = 05),
                     expand = c(0, 0)) +
  scale_y_continuous(breaks = seq(0, 60, by = 05),
                     expand = c(0, 0))

# #####
# cohorts of 3

# TTT
level <- c(2,2,2)
tox <- c(1,1,1)
followup <- c(35,35,35)
mod <- titecrm(prior = skeleton, target = target, tox =
                tox, level = level,
                obswin = obswin, scheme = 'linear',
                followup = followup)
mod$mtd

# NTT
level <- c(2,2,2)
tox <- c(1,1,0)

```

```

followupcombo <- cbind(rep(obswin, obswin) ,rep(obswin,
obswin), 1:obswin)

results <- data.frame('Observed' = 1:obswin, 'Rec' =
rep(0, obswin))

for (i in 1:obswin) {

  followup <- followupcombo[i,]

  mod <- titecrm(prior = skeleton, target = target, tox
= tox, level = level,
obswin = obswin, scheme = 'linear',
followup = followup)

  results$Rec[i] <- mod$mtd
}

DTP_c3NTT <- results %>%
  mutate( Dose = 2,
Pathway = Observed,
Outcome = paste0('N', '(', Observed, ')', 'TT',
),
Outcome = if_else(Outcome == 'N(35)TT', 'NTT',
, Outcome)) %>%
  select(Pathway, Dose, Outcome, Rec) %>%
  arrange(Pathway)

DTP_c3NTT %>%
  kable('latex', booktabs = T, linesep = "", align = "c
",

```

```

col.names = c('Pathway', 'Dose', 'Outcomes', ,
             Dose'),
caption = '\\label{tab_tite-dtp:TITEDTP_c3NTT}
TITE-DTP for a cohort of three for scenario
NTT.') %>%
kable_styling(latex_options = c("striped", "HOLD_",
position", "scale_down"),
position = "center", font_size = 5) %>%
add_header_above(c('', 'Cohort_1' = 2, 'Cohort_2' =
1)) %>%
cat()

## NNT
level <- c(2,2,2)
tox <- c(1,0,0)
 combos <- combinations(n = 35, r = 2, repeats.allowed =
T, v = 1:35)
 combos <- cbind(35, combos)
 pos <- cbind(rep(0,nrow(combos)))
 results <- pos[, rep(1, each=length(tox)+1)]
for (i in 1:nrow(combos)) {
  followup <- as.numeric(combos[i,])
  weights <- followup / obswin
  mod <- titecrm(prior = skeleton, target = target, tox =
tox, level = level,
                 obswin = obswin, weights = weights)
}

```

```
for (j in 1:ncol(results)) {  
  results[,j][i] <- followup[j]  
  results[,ncol(results)][i] <- mod$mtd  
}  
  
}  
  
results <- data.frame(results)  
colnames(results) <- c('Patient1', 'Patient2', '  
Patient3', 'Rec')  
  
results %>% mutate(TotalFollow = Patient2+Patient3) %>%  
group_by(Rec) %>%  
summarise(n = n(), min = min(TotalFollow), max = max(  
TotalFollow))  
  
results %>% mutate(TotalFollow = Patient2+Patient3) %>%  
View()  
  
# NNN  
  
level <- c(2,2,2)  
tox <- c(0,0,0)  
combos <- combinations(n = 35, r = 3, repeats.allowed =  
T, v = 1:35)  
  
pos <- cbind(rep(0,nrow(combos)))
```

```
results <- pos[, rep(1, each=length(tox)+1)]  
for (i in 1:nrow(combos)) {  
  if(i == 1){  
    start.time <- Sys.time()  
    print(start.time)  
  }  
  if(i == nrow(combos)){  
    end.time <- Sys.time()  
    print(end.time)  
  }  
  followup <- as.numeric(combos[i,])  
  weights <- followup / obswin  
  mod <- titecrm(prior = skeleton, target = target, tox  
  = tox, level = level,  
  obswin = obswin, weights = weights)  
  
  for (j in 1:ncol(results)) {  
    results[,j][i] <- followup[j]  
    results[,ncol(results)][i] <- mod$mtd  
  }  
  
}  
  
results <- data.frame(results)  
colnames(results) <- c('Patient1', 'Patient2', 'Patient3  
, 'Rec')
```

```
results %>% group_by(Rec) %>%
  summarise(n=n())

results %>% mutate(TotalFollow = Patient1+Patient2+
  Patient3) %>%
  group_by(Rec) %>%
  summarise(n = n(), min = min(TotalFollow), max = max(
    TotalFollow))

results %>%
  mutate(TotalFollow = Patient1+Patient2+Patient3) %>%
  filter(TotalFollow %in% c(24,25,26)) %>%
  group_by(Rec) %>%
  summarise(n = n())
# filter(Patient3 == 15)

# Need to run all combinations for the plot
level <- c(2,2,2)
tox <- c(0,0,0)
 combos <- 1:obswin
 combos <- expand.grid(combos, combos, combos)
 pos <- cbind(rep(0,nrow(combos)))

results <- pos[, rep(1, each=length(tox)+1)]
for (i in 1:nrow(combos)) {
  if(i == 1){
    start.time <- Sys.time()
```

```
print(start.time)

}

if(i == nrow(combos)){
  end.time <- Sys.time()
  print(end.time)
}

followup <- as.numeric(combos[i,])
weights <- followup / obswin
mod <- titecrm(prior = skeleton, target = target, tox
= tox, level = level,
  obswin = obswin, weights = weights)

for (j in 1:ncol(results)) {
  results[,j][i] <- followup[j]
  results[,ncol(results)][i] <- mod$mtd
}

}

results <- data.frame(results)
colnames(results) <- c('Patient1', 'Patient2', 'Patient3
', 'Rec')
plot3d(x = results$Patient1, y = results$Patient2, z =
results$Patient3,
  col = as.factor(results$Rec), xlab = 'Patient_1_
follow-up',
  ylab = 'Patient_2_follow-up', zlab = 'Patient_3_
follow-up')
```

```
rgl.postscript("c3NNN", "pdf")
outputFile = sub(".gif$","",outputFile)
movie3d(spin3d(axis=c(0,0,1), rpm=5), duration=12, dir=
getwd(), movie="c3NNN")
#
#####
# Investigating combinations
# With a combined follow-up of 20 2, 18 leads to 5 but
# 3, 17 leads to 4
# With a combined follow-up of 21 6, 15 leads to 5 but
# 7, 14 leads to 4
level <- c(2,2)
tox <- c(0,0)
followup <- c(2, 18)
weights <- followup / obswin
mod1 <- titecrm(prior = skeleton, target = target, tox
= tox, level = level,
obswin = obswin, weights = weights)

followup <- c(3, 17)
weights <- followup / obswin
mod2 <- titecrm(prior = skeleton, target = target, tox
= tox, level = level,
obswin = obswin, weights = weights)
```

```
# Re run analysis but record estimates and variance

level <- c(2,2)

tox <- c(0,0)

combos <- combinations(n = 35, r = 2, repeats.allowed =
T, v = 1:35)

pos <- cbind(rep(0,nrow(combos)))

results <- pos[, rep(1, each=length(tox)+3)]

for (i in 1:nrow(combos)) {

  followup <- as.numeric(combos[i,])

  weights <- followup / obswin

  mod <- titecrm(prior = skeleton, target = target, tox
= tox, level = level,
                 obswin = obswin, weights = weights)

  for (j in 1:ncol(results)) {

    results[,j][i] <- followup[j]

    results[,3][i] <- mod$mtd

    results[,4][i] <- mod$estimate

    results[,5][i] <- mod$post.var

  }

}

# Rename data frame

results <- data.frame(results)
```

```

colnames(results) <- c('Patient1', 'Patient2', 'Rec', ,
  Est', 'Var')

# Check matches previous results
results %>% mutate(TotalFollow = Patient1+Patient2) %>%
  group_by(Rec) %>%
  summarise(n = n(), min = min(TotalFollow), max = max(
    TotalFollow))

# Create table with beta estimate and variance included
results %>% mutate(TotalFollow = Patient1+Patient2,
  Est = Est %>% round(4),
  Var = Var %>% round(4)) %>%
  filter(TotalFollow == 20 | TotalFollow == 21) %>%
  select(Patient1, Patient2, TotalFollow, Rec, Est, Var
  ) %>%
  arrange(desc(TotalFollow)) %>%
  kable('latex', booktabs = T, linesep = "", align = "c",
  ",
  col.names = c('Patient_1', 'Patient_2', ,
  Combined', 'Dose_Recommendation',
  'Beta', 'Variance'),
  caption = '\\label{tab_tite-dtp:TITEDTP_'
  c2NNprob}Different_dose_recommendations_with
  _overlapping_combined_follow-up_times.') %>%
  kable_styling(latex_options = c("striped", "HOLD_"
  position)),

```

```

      position = "center", font_size = 11)

%>%
add_header_above(c('Follow-up' = 3, '□' = 1, 'Posterior □ Estimates' = 2)) %>%
cat()

# Overall plot
results %>% mutate(TotalFollow = Patient1+Patient2) %>%
ggplot(aes(x = TotalFollow, y = Est, col = factor(Rec))) +
geom_point() +
geom_hline(yintercept = 0.149, col = "red", linetype =
= "longdash") +
labs(x = "Combined □ Total □ Follow-up", y = expression(
Posterior ~ Estimate ~ beta),
col = "Recommended □ Dose") +
scale_color_manual(values = c("#994d1a", "#1a6699")) +
scale_x_continuous(breaks = seq(0, 70, 5)) +
theme_bw() +
theme(legend.position = "bottom")

# Plot for 20 & 21 days
results %>% mutate(TotalFollow = Patient1+Patient2) %>%
filter(TotalFollow == 20 | TotalFollow == 21) %>%
ggplot(aes(x = factor(TotalFollow), y = Est, col =
factor(Rec))) +
geom_point() +

```

```
geom_hline(yintercept = 0.149, col = "red", linetype
= "longdash") +
geom_text_repel(mapping = aes(label = paste0("(",
Patient1, ", ", Patient2, ")"))),
show.legend = FALSE) +
labs(x = "Combined\u222aTotal\u222aFollow-up", y = expression(
Posterior ~ Estimate ~ beta),
col = "Recommended\u222aDose") +
scale_color_manual(values = c("#994d1a", "#1a6699")) +
theme_bw() +
theme(legend.position = "bottom")

skeleton
beta <- seq(0, 1, 0.0001)
df <- data.frame(beta, rec = rep(NA, times = length(
beta)))
for (i in 1:length(beta)) {
  ptox <- skeleton^exp(beta[i])
  if (all(ptox <= target)) {
    df$rec[i] <- length(skeleton)
  }
  else if (all(ptox >= target)) {
    df$rec[i] <- 1
  }
  else {
    df$rec[i] <- order(abs(ptox - target))[1]
```

```
}

}

df %>% group_by(rec) %>%
  summarise(min = min(beta) %>% round(4),
            max = max(beta) %>% round(4))

# 0.1492 -> 4, 0.1493 ->5

#####
# For 3 patients
#####

level <- c(2,2,2)
tox <- c(0,0,0)
combos <- combinations(n = 35, r = 3, repeats.allowed =
T, v = 1:35)
pos <- cbind(rep(0,nrow(combos)))
results <- pos[, rep(1, each=length(tox)+3)]

for (i in 1:nrow(combos)) {
  if(i == 1){
    start.time <- Sys.time()
    print(start.time)
  }
  if(i == nrow(combos)){
    end.time <- Sys.time()
    print(end.time)
  }
}
```

```
}

followup <- as.numeric(combos[i,])

weights <- followup / obswin

mod <- titecrm(prior = skeleton, target = target, tox
= tox, level = level,
               obswin = obswin, weights = weights)

for (j in 1:ncol(results)) {

  results[,j][i] <- followup[j]

  results[,4][i] <- mod$mtd

  results[,5][i] <- mod$estimate

  results[,6][i] <- mod$post.var

}

}

results <- data.frame(results)

colnames(results) <- c('Patient1', 'Patient2', 'Patient3',
  'Rec', 'Est', 'Var')

results %>% group_by(Rec) %>%
  summarise(n=n())

results %>% mutate(TotalFollow = Patient1+Patient2+
  Patient3) %>%
  group_by(Rec) %>%
```

```
summarise(n = n(), min = min(TotalFollow), max = max(
  TotalFollow))

results %>%
  mutate(TotalFollow = Patient1+Patient2+Patient3) %>%
  filter(TotalFollow %in% c(20,21)) %>%
  group_by(Rec) %>%
  summarise(n = n())

# Create table with beta estimate and variance included
# for 9 pathways
results %>% mutate(TotalFollow = Patient1+Patient2+
  Patient3,
  Est = Est %>% round(4),
  Var = Var %>% round(4)) %>%
filter(TotalFollow %in% c(20,21) & Rec == 5) %>%
select(Patient1, Patient2, Patient3, TotalFollow, Rec
  , Est, Var) %>%
arrange(desc(TotalFollow)) %>%
kable('latex', booktabs = T, linesep = "", align = "c",
  ",
```

```

col.names = c('Patient_1', 'Patient_2', ,
            Patient_3', 'Combined', 'Dose_Recommendation
            ',
            'Beta', 'Variance') ,
caption = '\\label{tab_tite-dtp:TITEDTP_
c3NNNprob}Follow-up_combinations_totalling_
20_or_21_days_leading_to_dose-level_5.') %>%
kable_styling(latex_options = c("striped", "HOLD_-
position", "scale_down"),
              position = "center", font_size = 11)
%>%
add_header_above(c('Follow-up' = 4, ' ' = 1, ' '
Posterior_Estimates' = 2)) %>%
cat()

# Overall plot
results %>% mutate(TotalFollow = Patient1+Patient2+
Patient3) %>%
ggplot(aes(x = TotalFollow, y = Est, col = factor(Rec
))) +
geom_point(size = 0.5) +
geom_hline(yintercept = 0.1492, col = "red", linetype
= "longdash") +
labs(x = "Combined_Total_Follow-up", y = expression(
Posterior ~ Estimate ~ beta),
col = "Recommended_Dose") +
scale_color_manual(values = c("#994d1a", "#1a6699"))+

```

```

scale_x_continuous(breaks = seq(0, 105, 5))+
theme_bw()+
theme(legend.position = "bottom")

# Plot for 20 & 21 days
results %>% mutate(TotalFollow = Patient1+Patient2+
Patient3) %>%
filter(TotalFollow == 20 | TotalFollow == 21) %>%
ggplot(aes(x = factor(TotalFollow), y = Est, col =
factor(Rec))) +
geom_point() +
geom_hline(yintercept = 0.1492, col = "red", linetype
= "longdash") +
geom_text_repel(mapping = aes(label = paste0("(",
Patient1, ",",
Patient2
, ",",
Patient3
, ")"))
show.legend = FALSE, max.overlaps =
Inf)+

labs(x = "Combined Total Follow-up", y = expression(
Posterior ~ Estimate ~ beta),
col = "Recommended Dose")+
scale_color_manual(values = c("#994d1a", "#1a6699"))+

```

```
theme_bw() +  
  theme(legend.position = "bottom")  
  
# Calculate DTP  
results <- results %>%  
  mutate(TF = Patient1 + Patient2+ Patient3)  
  
results %>%  
  filter(TF <= 19) %>%  
  nrow()  
  
results %>%  
  filter(TF >= 22) %>%  
  nrow()  
  
results %>%  
  filter(TF == 20 & Patient3 <= 17)  
  
results %>%  
  filter(TF == 20 & Patient3 >= 18)  
  
results %>%  
  filter(TF == 21 & ((Patient3 <= 15 & Patient2 <=3) |  
    Patient3 <= 14))  
  
results %>%
```

```
filter(TF == 21 & ((Patient3 >= 15 & Patient2 >=4) |  
Patient3 >= 16))
```

B.4 ETP chapter code

The code below was used to run the simulations shown in Chapter 5.

```
# Operating Characteristics - Beta-Binomial Posterior  
-----  
  
# prob_true = underlying true probability of response  
# npat = number of patients  
# prior_par = vector of parameters for beta(a,b) prior  
# distribution  
# crit_rate = required repsonse rate to be greater than  
# e.g. 0.1  
# accept_prob = probability required to be greater than  
# the required response rate with e.g. 0.8  
# nsims = number of simulations  
  
  
  
  
  
PPoS <- function(n2, apost, bpost, obj.resp.rate,  
accept.prob){  
# If remaining number of patients is 0 don't need to  
# calculate PPoS  
if(n2 == 0){  
PPoS = NA
```

```

}

else{

Pr2 <- sapply(0:n2,
              function(i) (gamma(apost+bpost)*gamma
              (n2+1)*gamma(i+apost)*
              gamma(n2-i+bpost)) /
              (gamma(apost)*gamma(bpost)*gamma(i
              +1)*gamma(n2-i+1)*
              gamma(n2+apost+bpost)) )



B <- sapply(0:n2, function(i) 1-pbeta(obj.resp.rate
, apost+i, bpost+n2-i))

temp <- cbind(Pr2, B) %>%
  data.frame() %>%
  mutate(G0 = if_else(B > accept.prob, 1, 0),
         PPP = Pr2*G0)

PPOS = sum(temp$PPP)

}

return(PPOS)
}

oc_betabinom <- function(prob_true, npat, prior_par = c
(0.5, 0.5), crit_rate, accept_prob = 0.8, nsims, nint
) {

prob <- c(rep(0, nsims))

```

```

ppc <- npat/(nint+1)

cohorts <- npat/ppc

a0 <- prior_par[1]

b0 <- prior_par[2]

for(i in 1:nsims) {

  data <- replicate(npat, rbinom(1,1,prob_true))

  post_prob <- pbeta(crit_rate, shape1 = (prior_par
  [1] + sum(data)),
  shape2 = (prior_par[2] + npat -
  sum(data)),
  lower.tail = FALSE)

  ifelse(post_prob >= accept_prob, prob[i] <- 1, prob
  [i] <- 0)

  for(j in 1:nint) {

    n <- ppc*j

    r <- data[1:n]

    ppos <- PPOS(n2 = npat-n, apost = a0 + sum(r),
    bpost = b0 + n - sum(r),
    obj.resp.rate = crit_rate,
    accept.prob = accept_prob)

    if(ppos < 0.05){

      prob[i] <- 0
    }
  }
}

```

```
        break
    }
}

Acceptance_Rate <- (sum(prob) / (nsims)) * 100

return(Acceptance_Rate)
}
```

The function below was created to produce ETPs.

```
## Required packages: ----
library(ggplot2)
library(tidyverse)

## Parameters: -----
# cohort.size -           number of patients in each cohort
# (cohort size)
# cohorts -               number of cohorts
# obj.resp.rate -         target objective response rate
# (0 - 1)
# accept.prob -           acceptably probability level to
# make a go/no go decision
# #                         at final analysis (0 - 1)
# ppos.accept.prob -      acceptably probability level for
# PPos go/no go decision at
# #                         interim (0 - 1)
```

```

# prior -           vector of parameters for beta(a, b)
#           prior distribution,
#           default is a beta(1, 1) which is
#           entered as c(1, 1)

# cred.int.side -   specify either a "two.sided" or "
#           one.sided " credible

#           interval

# cred.int.level -  specify credible interval level e.
#           g. 95% (0.95)

# align -           specify the alignment of the plot
#           options are either "left"
#           or "centre",

# size -            changes size of the text on the
#           plot default is 5

# legend.off -      turn off the legend on the plot
#           default is on == FALSE

# output.data -     function will return key data
#           along with the plot, default

#           is off == FALSE

# verbose.data -    returns data with additional
#           columns with x, y coordinates for
#           plotting, default is off == FALSE

# Function: ----

etp <- function(cohort.size,
                 cohorts,
                 obj.resp.rate,

```

```
accept.prob ,
ppos.accept.prob ,
prior = c(1,1),
cred.int.side = c("two.sided", "one.
sided"),
cred.int.level = 0.95,
align = c("centre", "left"),
size = 5,
legend.off = FALSE,
output.data = FALSE,
verbose.data = FALSE) {

# Calculate total sample size
N <- cohort.size*cohorts

# Extract prior parameters
a0 <- prior[1]
b0 <- prior[2]

# Function to calculate PPoS
PPoS <- function(n2, apost, bpost, obj.resp.rate,
accept.prob){

# If remaining number of patients is 0 don't need
to calculate PPoS

if(n2 == 0){

PPoS = NA

}

else{
```

```

Pr2 <- sapply(0:n2,
              function(i) (gamma(a$post+b$post)*
                           gamma(n2+1)*gamma(i+a$post)*
                           gamma(n2-i+b$post)) /
              (gamma(a$post)*gamma(b$post)*gamma(
                           i+1)*gamma(n2-i+1)*
                           gamma(n2+a$post+b$post)) )

B <- sapply(0:n2, function(i) 1-pbeta(obj.resp.
                                         rate, a$post+i, b$post+n2-i))

temp <- cbind(Pr2, B) %>%
  data.frame() %>%
  mutate(G0 = if_else(B > accept.prob, 1, 0),
        PPP = Pr2*G0)
PPOS = sum(temp$PPP)

}

return(PPOS)
}

# Generate data frame with all the data required for
# the etp plot

df <- data.frame(
  # produces a vector of responses for each
  # cumulative cohort
  r = rep(0:(cohort.size*cohorts), rev(c(rep(1:
                                             cohorts, each = cohort.size),
                                             
```

```
cohorts))))  
%>%  
group_by(r) %>%  
mutate(  
  # produces y co-ords for plot  
  y = cohorts$row_number(),  
  # generic weighting for size of each rectangle on  
  # the plot  
  w = rep(1, length(r)),  
  # Establish the specific cohort each row belongs  
  # to  
  Cohort = 1+y,  
  # Different fonts for the plot  
  bolditalic = rep('bold.italic', length(r)),  
  bold = rep('bold', length(r)),  
  italic = rep('italic', length(r)),  
  # For each cohort n1 = how many patients with data  
  # , n2 = how many left to recruit  
  n1 = Cohort * cohort.size,  
  n2 = N - n1,  
  # At interim parameters for probability  
  # calculations  
  apost = a0 + r,  
  bpost = b0 + n1 - r,  
  # Labels to identify what probability needs  
  # calculating and analysis stage
```

```

Prob_label = if_else(Cohort == cohorts, 'PP', ,
                     PPOS') ,
Analysis = if_else(Cohort == cohorts, 'Final', ,
                     Interim') ,
# Calculates estimates
Estimate = signif(qbeta(p = 0.5, shape1 = apost,
                         shape2 = bpost)*100, 2) ,
# Calculate two-sided credible interval
LB = signif(qbeta(p = 0 + (1-cred.int.level)/2,
                     shape1 = apost,
                     shape2 = bpost)*100, 2) ,
UB = signif(qbeta(p = 1 - (1-cred.int.level)/2,
                     shape1 = apost,
                     shape2 = bpost)*100, 2) ,
# Calculate one-sided credible interval
OSB = signif(qbeta(p = 1-cred.int.level, shape1 =
                     apost, shape2 = bpost)*100,
                     2)
)%>%
# Ungroup and do probability calcs for each row
ungroup() %>%
rowwise() %>%
mutate(
# Calculate PPoS
Prob = PPOS(n2 = n2, apost = apost, bpost = bpost
, obj.resp.rate,
accept.prob),

```

```

# If NA means that its the final cohort so need
# posterior prob

Prob = if_else(is.na(Prob),
               round(pbeta(q = obj.resp.rate,
                           shape1 = apost,
                           shape2 = bpost, lower.
                           tail = F), 7),
               Prob),

Prob = round(Prob, 3),
# Determine what the decision outcome would be

Decision = case_when(Cohort == cohorts & Prob >=
accept.prob ~ 'GO',
                      Cohort == cohorts & Prob <
accept.prob ~ 'No_GO',
                      Cohort != cohorts & Prob >=
ppos.accept.prob ~ 'GO',
                      Cohort != cohorts & Prob <
ppos.accept.prob ~ 'No_GO
'),
DecRules = factor(paste(Decision, Analysis, sep =
""),
levels = c("GO_Final", "No_GO_
Final", "GO_Interim",
"No_GO_Interim"))
)

# Produce x coords based on alignment

```

```
if (align == "centre") {  
  df <- df %>%  
    mutate(x = ifelse(y < cohorts, r+(cohorts-y)*(  
      cohort.size/2), r),  
          xmin = x - w/2,  
          xmax = x + w/2)  
  
}  
  
if (align == "left"){  
  df <- df %>%  
    mutate(x = r + 0.5,  
          xmin = r + w - 1,  
          xmax = r + w)  
}  
  
# Code to generate plot  
plot <-  
  ggplot(df, aes(xmin = xmin, xmax = xmax, ymin = y,  
                 ymax = y + 1)) +  
  # plot rectangles for each response in each cohort  
  geom_rect(aes( colour = DecRules , lty = DecRules),  
            fill= "white",  
            size = 1) +  
  # reverse the y axis  
  scale_y_reverse() +  
  scale_color_manual("Decision $\sqcup$ Rules",
```

```

values = c("dark□green", "red",
          "dark□green", "red"),
labels = c(bquote(P(theta >= .(
obj.resp.rate*100) ~ '%') >=
.(accept.
prob) :
Go),
bquote(P(theta >= .(
obj.resp.rate*100)
~ '%') <
.(accept.
prob) ~ ,
:No□Go'),
bquote(PPoS >= .(ppos.
accept.prob) ~ ':',
Go),
bquote(PPoS < .(ppos.
accept.prob) ~ ':',
No□Go)),
drop = FALSE)+

scale_linetype_manual("Decision□Rules",
values = c(1,1,2,2),
labels = c(bquote(P(theta >=
.(obj.resp.rate*100) ~ '%'
) >=

```

```

.(accept.
prob)
:Go),
bquote(P(theta >=
.(obj.resp.rate
*100) ~ '%') <
.(accept.
prob)
~ ':No
□Go'),

bquote(PPoS >= .(
ppos.accept.
prob) ~ ':Go'),

bquote(PPoS < .(
ppos.accept.
prob) ~ ':No□Go
')),

drop = FALSE)+

theme(axis.line=element_blank(),axis.text.x=element
_blank(),
axis.text.y=element_blank(),axis.ticks=
element_blank(),
axis.title.x=element_blank(),
axis.title.y=element_blank(),
panel.background=element_blank(),
panel.border=element_blank(),

```

```
    panel.grid.major=element_blank(),
    panel.grid.minor=element_blank(),
    plot.background=element_blank(),
    legend.spacing = unit(0.00, 'cm'))}

if(cred.int.side == "one.sided"){
  plot <- plot +
    geom_text(aes(label=r, x = x , y = y+0.2 ,
      fontface = bolditalic),
      size = size, colour = '#313639')+

  geom_text(data = df %>% filter(Prob_label == 'PPOS') ,
    aes(label=Prob , x = x , y = y +0.4 ,
      fontface = bold),
      colour = 'blue' , size = size)+

  geom_text(data = df %>% filter(Prob_label == 'PP') ,
    aes(label=Prob , x = x , y = y +0.4 ,
      fontface = bold),
      colour = 'black' , size = size)+

  geom_text(aes(label=paste0(Estimate , '%')) , x =
    x , y = y +0.6) ,
    size = size) +
```

```
geom_text(aes(label = paste0('(', OSB, '%', ')'))  
, x = x, y = y + 0.8 ,  
fontface = italic), size = size)  
  
data <- df %>%  
mutate(CrI = paste('(', OSB, '%', ')', sep = ''))  
}  
  
}  
  
if(cred.int.side == "two.sided") {  
plot <- plot +  
geom_text(aes(label=r, x = x , y = y+0.1 ,  
fontface = bolditalic) ,  
size = size, colour = '#313639') +  
  
geom_text(data = df %>% filter(Prob_label == '  
PPOS') ,  
aes(label=Prob , x = x , y = y +0.3 ,  
fontface = bold) ,  
colour = 'blue' , size = size) +  
  
geom_text(data = df %>% filter(Prob_label == 'PP'  
) ,  
aes(label=Prob , x = x , y = y +0.3 ,  
fontface = bold) ,  
colour = 'black' , size = size) +
```

```
geom_text(aes(label=paste0(Estimate, '%')), x = x,
          y = y + 0.5,
          size = size) +
  geom_text(aes(label = paste0('(', LB, '%'), ', UB, %', ')',
                y = y + 0.7,
                fontface = italic), size = size) +
  geom_text(aes(label = paste0(UB, '%', ')')), x = x,
            y = y + 0.9,
            fontface = italic), size = size)

data <- df %>%
  mutate(CrI = paste('(', LB, '%', ', UB, %', ')',
                    ' ', sep = ''))

}

if (isTRUE(legend.off)) {
  plot <- plot +
    theme(legend.position="none")
}

if(isTRUE(verbose.data)){
  data <- data %>%
    select(Cohort, n = n1, r, Analysis, Prob,
           Estimate, CrI, Decision,
           x, xmin, xmax, y, apost, bpost, LB, UB,
           OSB) %>%
    arrange(Cohort, r)
```

```
}

else{

  data <- data %>%
    select(Cohort, n = n1, r, Analysis, Prob,
           Estimate, CrI, Decision) %>%
    arrange(Cohort, r)

}

if(isTRUE(output.data)){
  return(list(ETP = plot, Data = data))
}

else{
  plot
}

}
```

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