

Chromatin ImmunoPrecipitation followed by sequencing (ChIP-seq):

Probing for epigenetic marks

NGS analysis for gene regulation and epigenomics

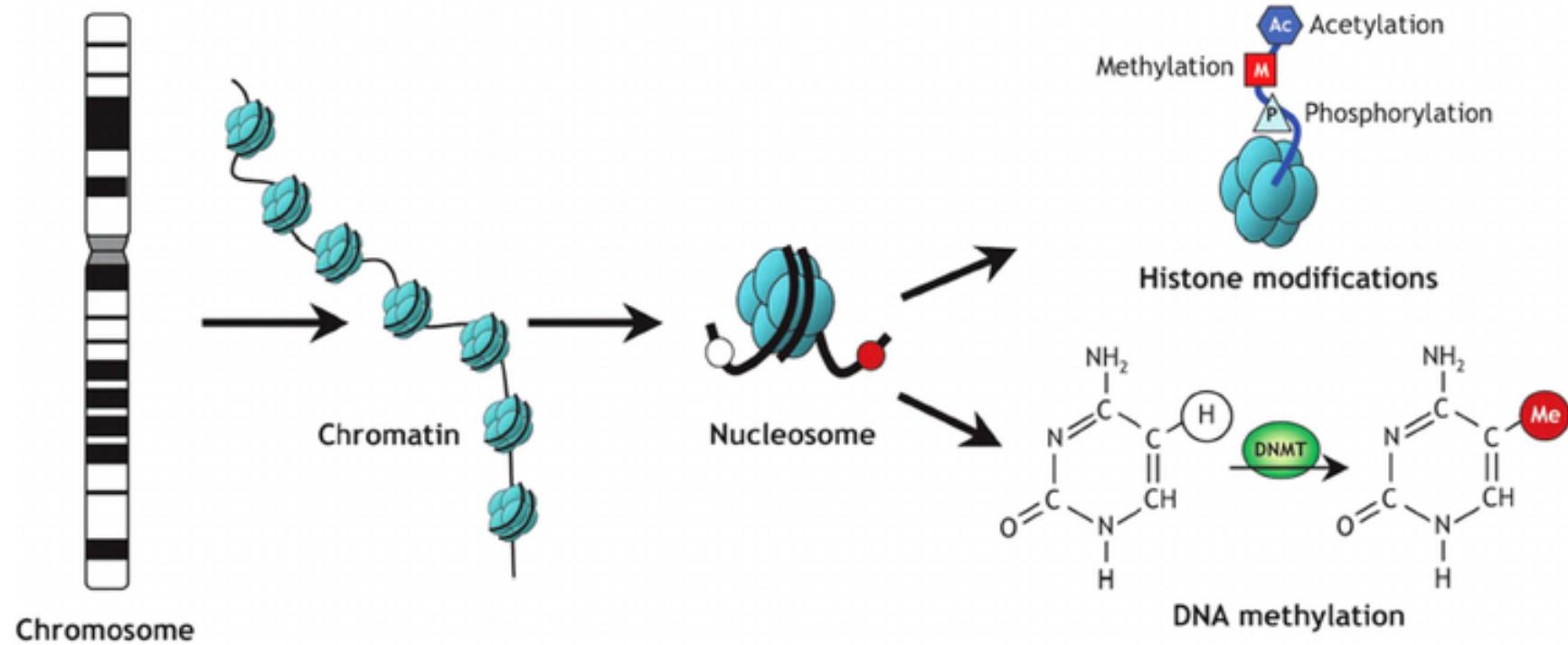
Physalia 2021

Why probing binding of proteins on DNA is important

- Many different proteins bind on/interacts with DNA: this constitutes the chromatin

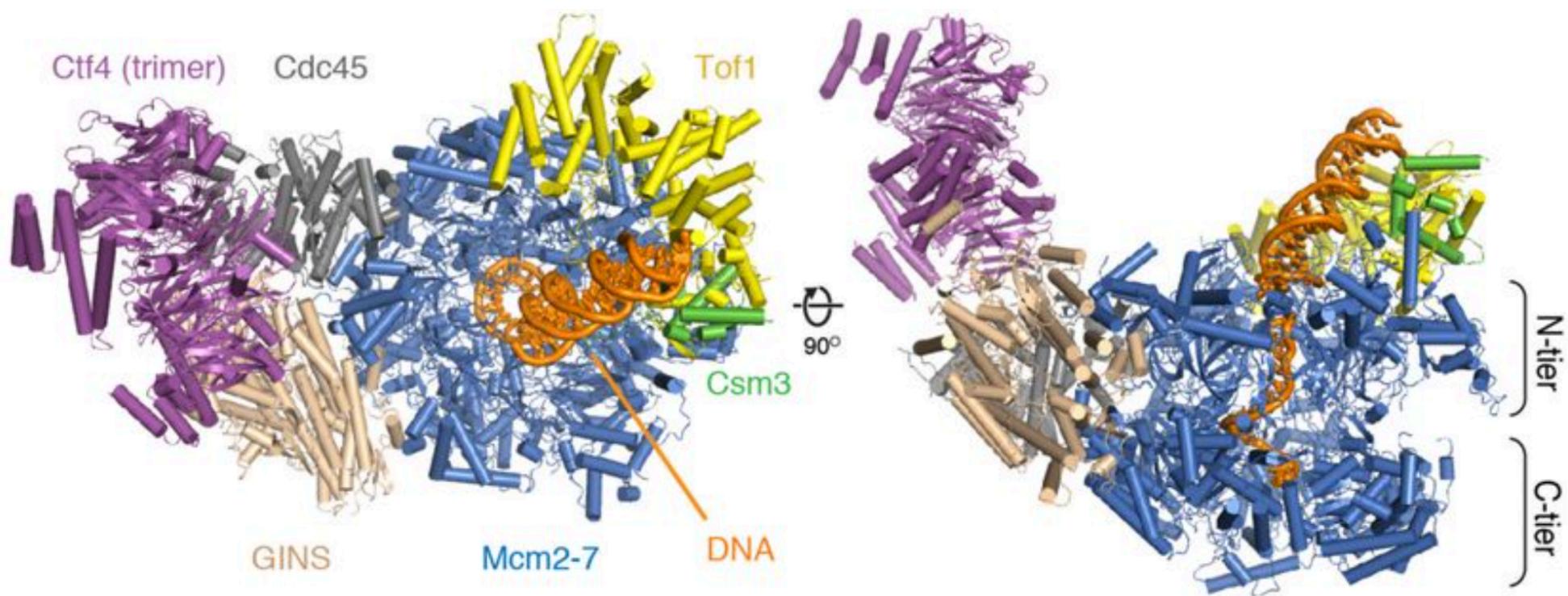
Why probing binding of proteins on DNA is important

- Basic chromatin constituents



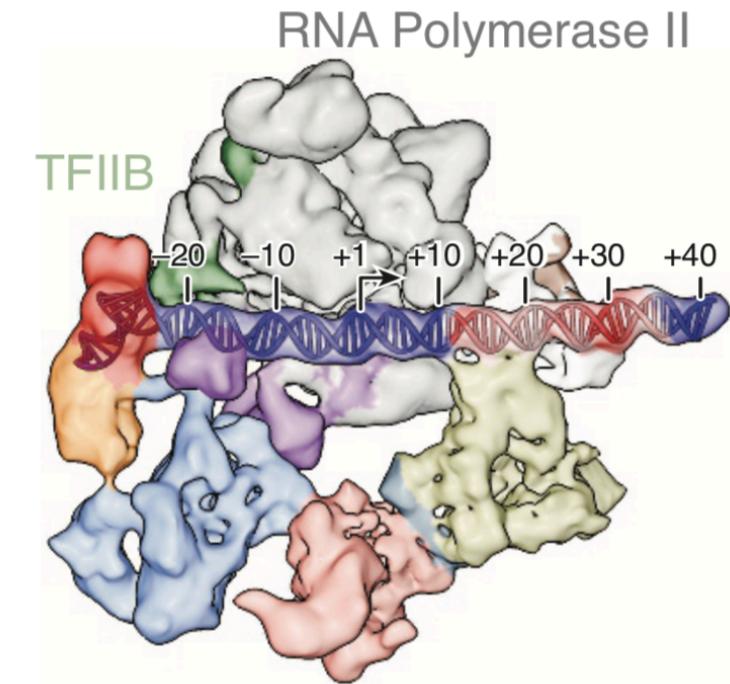
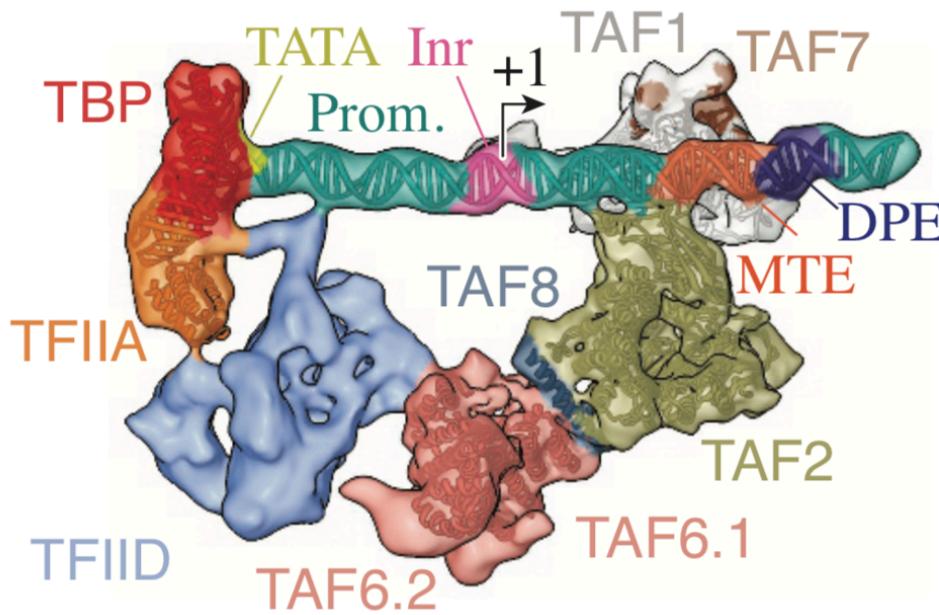
Why probing binding of proteins on DNA is important

- Replication machinery



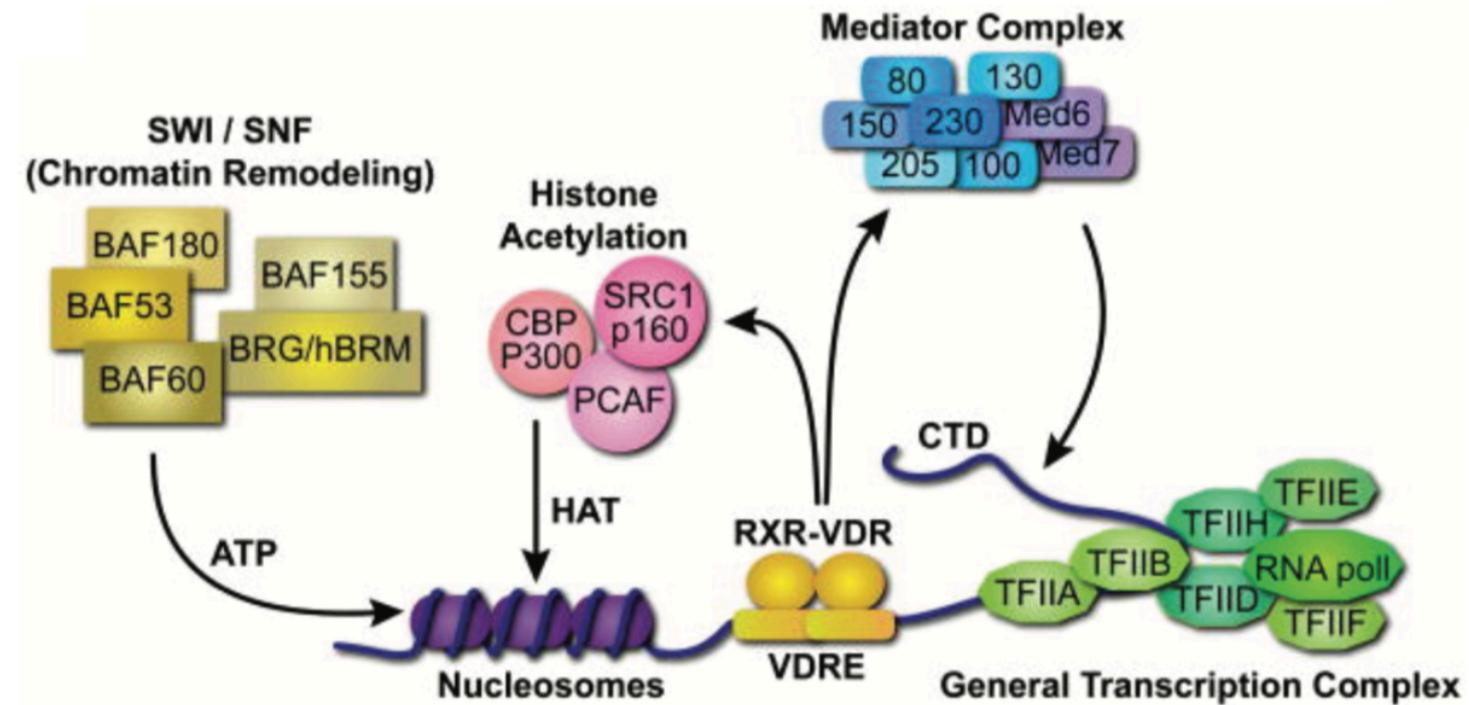
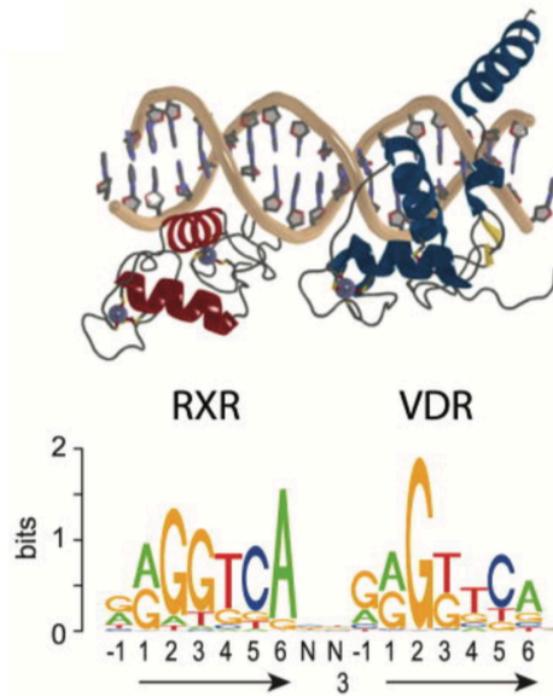
Why probing binding of proteins on DNA is important

- Transcription machinery



Why probing binding of proteins on DNA is important

- Regulatory proteins: transcription factors & co



Chromatin immunoprecipitation: an old tool rejuvenated by high-throughput sequencing

- Chromatin IP is not a new approach. It has been around for the past three decades

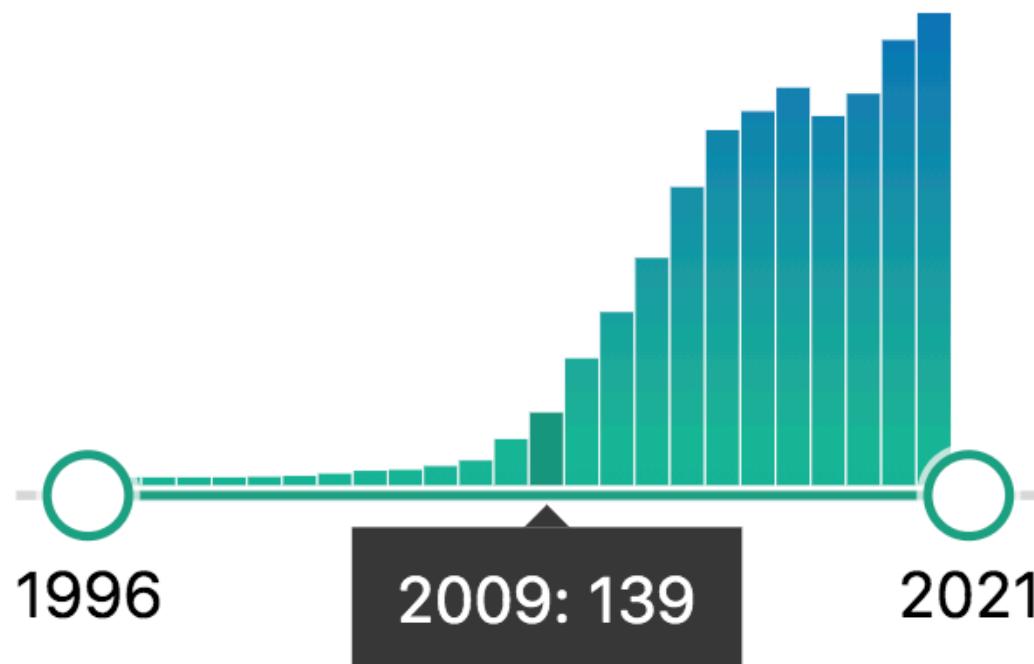


Summary

We have used formaldehyde-mediated proteinDNA crosslinking within intact cells to examine the *in vivo* chromatin structure of the *D. melanogaster* heat shock protein 70 (*hsp70*) genes. In agreement with previous *in vitro* studies, we find that the heat shockmediated transcriptional induction of the *hsp70* genes perturbs their chromatin structure, resulting in fewer proteinDNA contacts crosslinkable *in vivo* by formaldehyde. However, contrary to earlier *in vitro* evidence that histones may be absent from actively transcribed genes, we show directly, by immunoprecipitation of *in vivo*crosslinked chromatin fragments, that at least histone H4 remains bound to *hsp70* DNA *in vivo*, irrespective of its rate of transcription. The formaldehyde-based *in vivo* mapping techniques described in this work are generally applicable, and can be used both to probe proteinDNA interactions within specific genes and to determine the genomic location of specific chromosomal proteins.

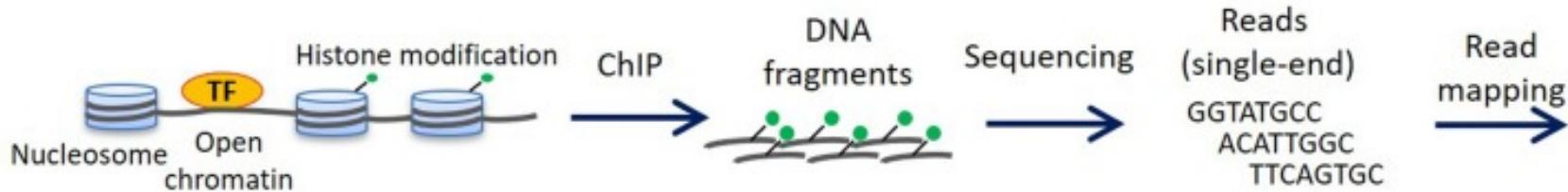
Chromatin immunoprecipitation: an old tool rejuvenated by high-throughput sequencing

- It gained a lot of traction when high-throughput sequencing emerged

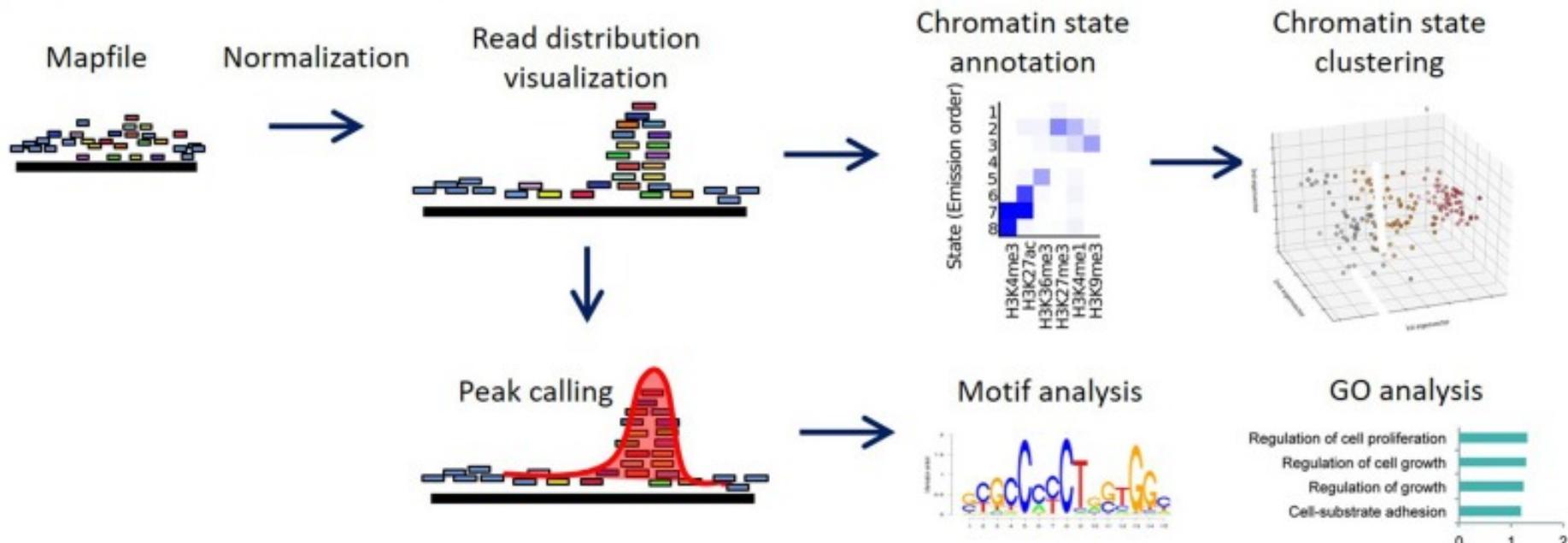


Classical workflow

(A) Sample preparation and sequencing

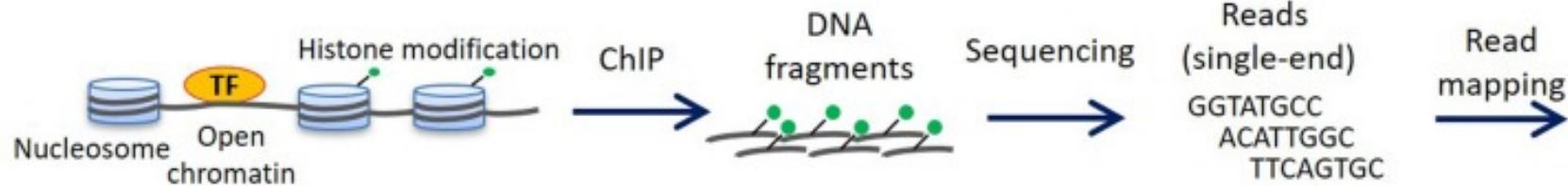


(B) Computational analysis

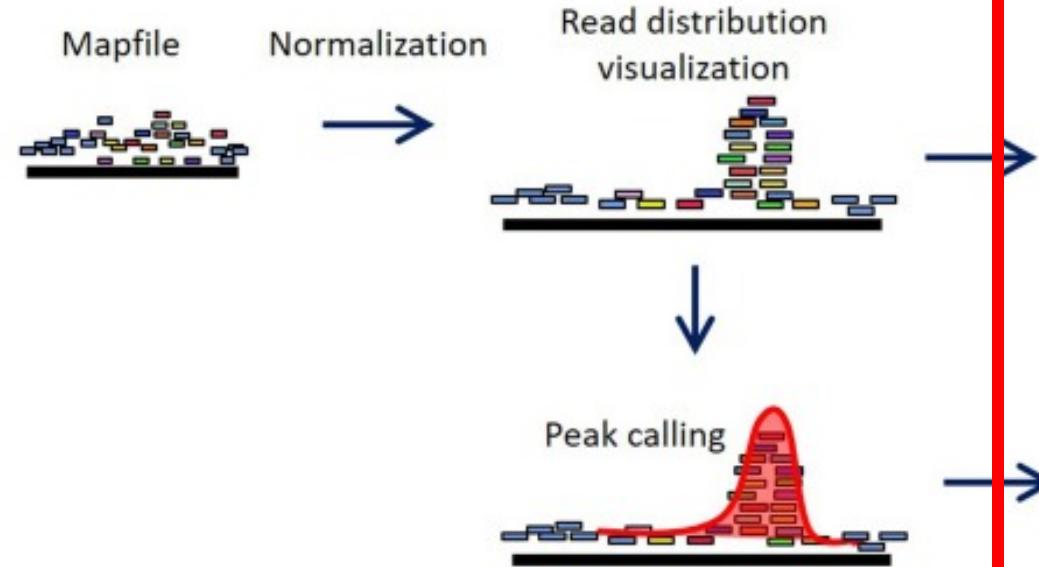


Classical workflow

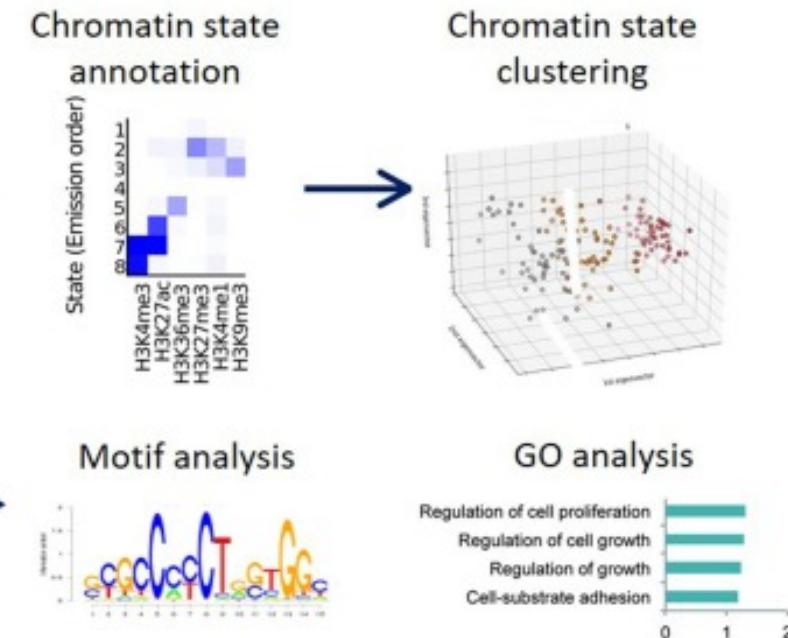
(A) Sample preparation and sequencing



(B) Computational analysis

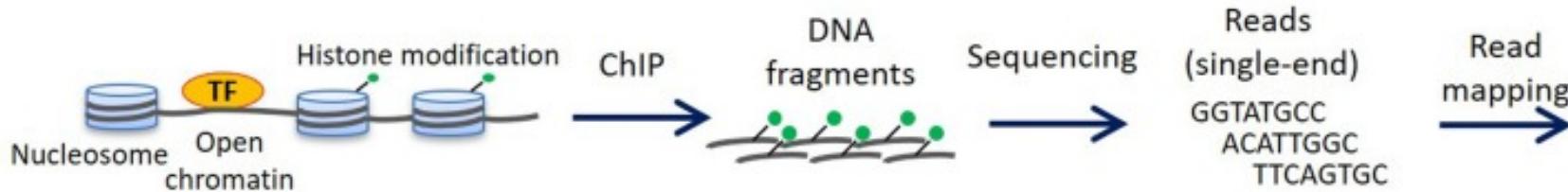


Ex 03-1

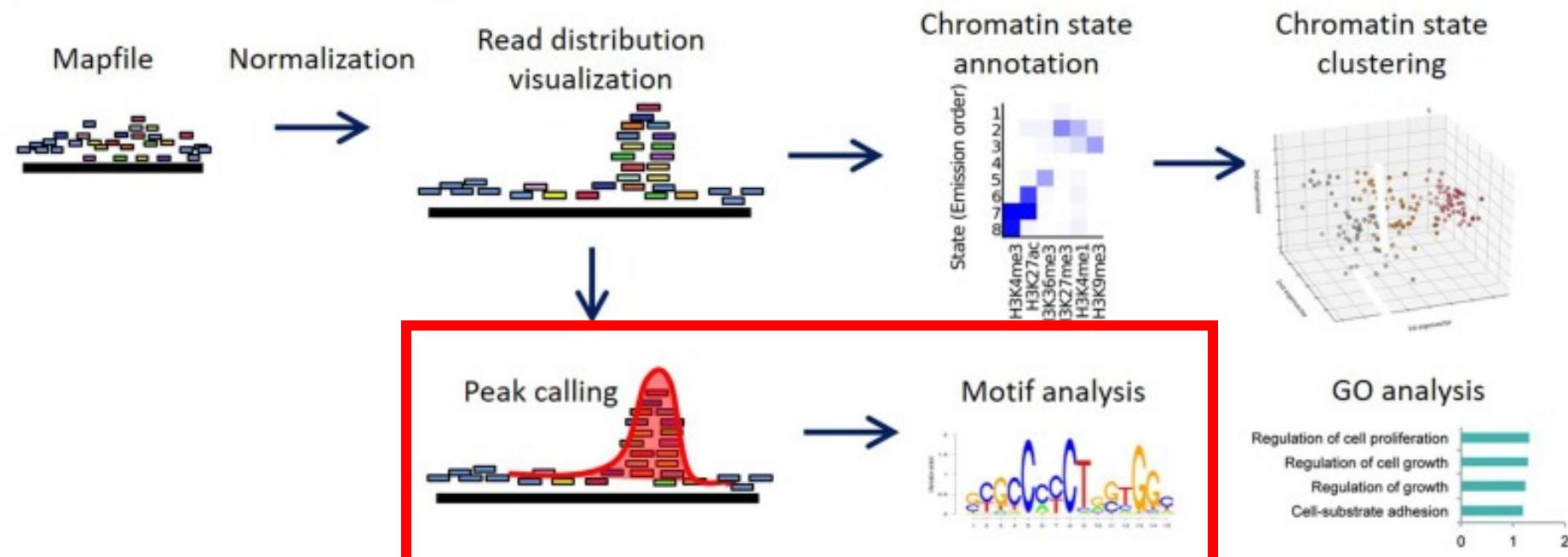


Classical workflow

(A) Sample preparation and sequencing

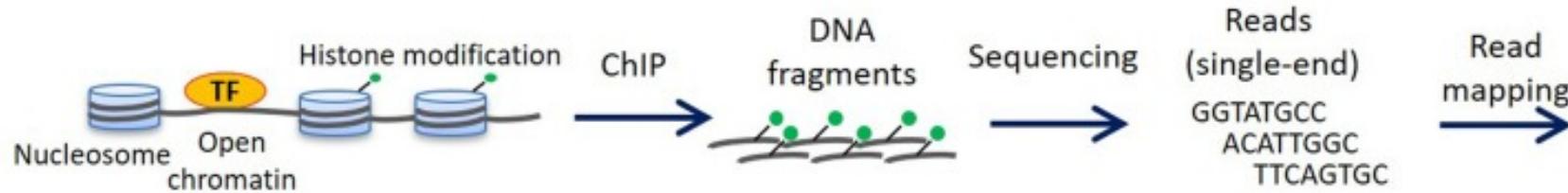


(B) Computational analysis

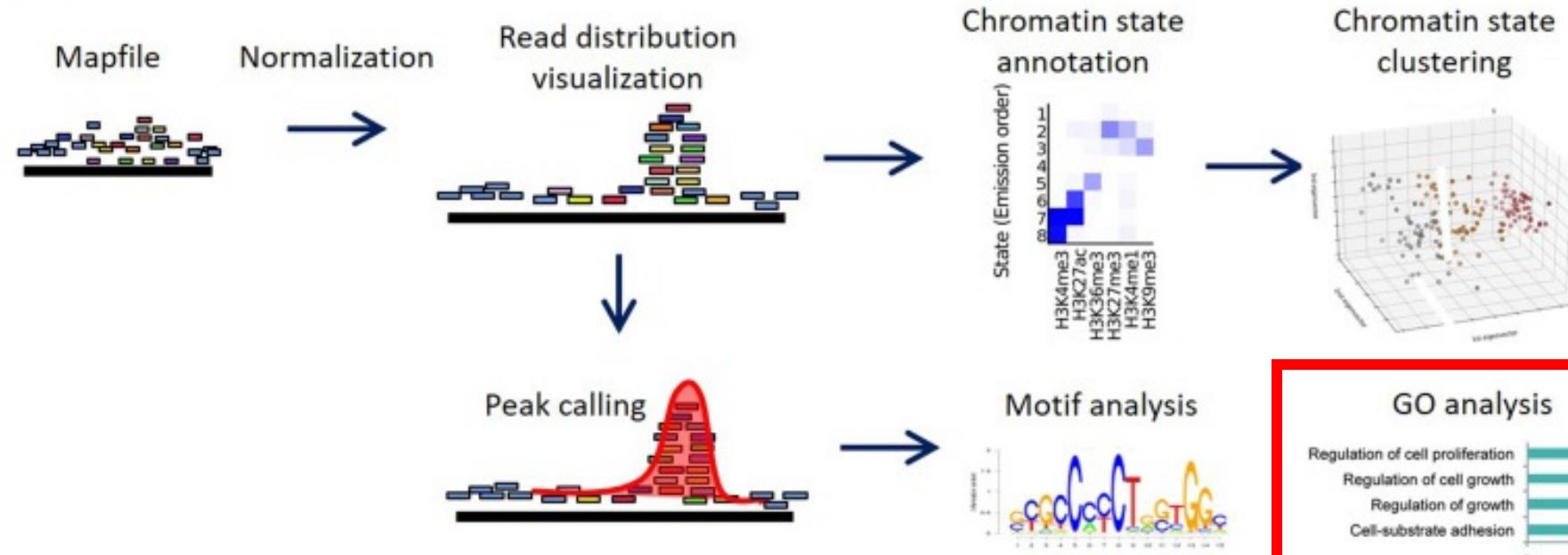


Classical workflow

(A) Sample preparation and sequencing



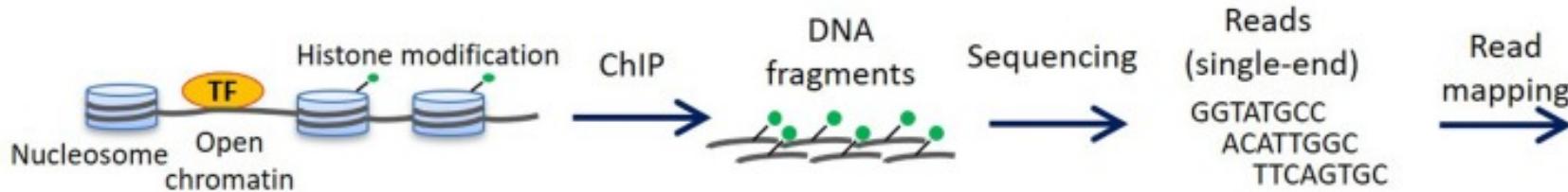
(B) Computational analysis



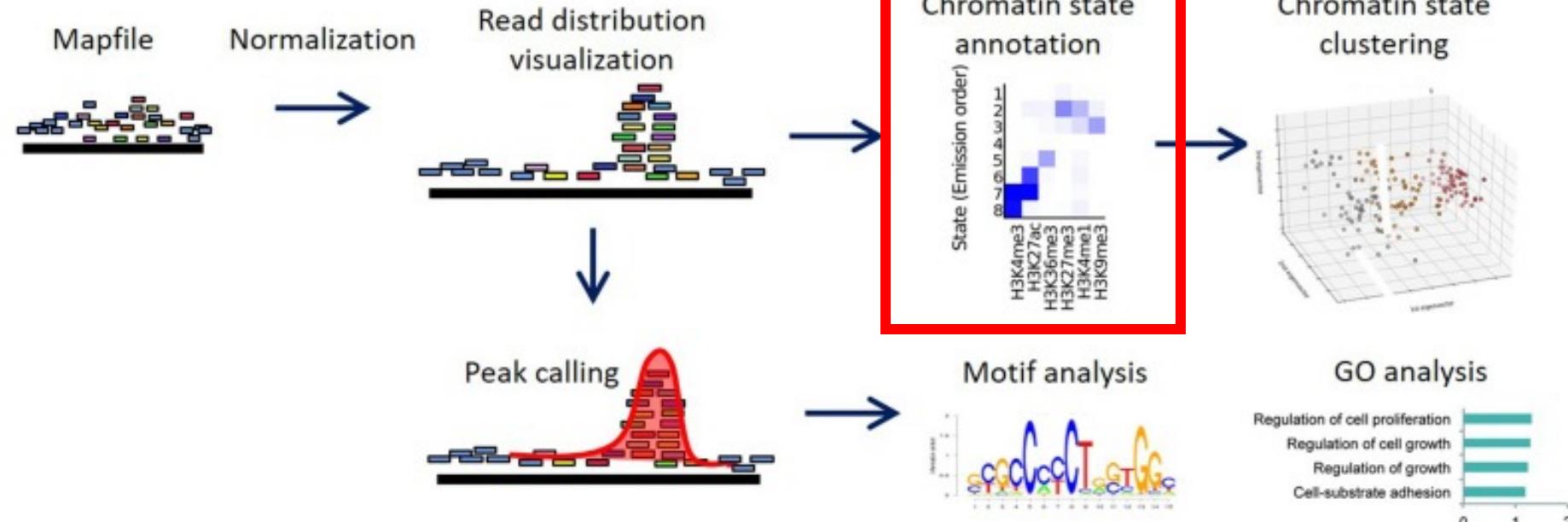
Ex 05-1

Classical workflow

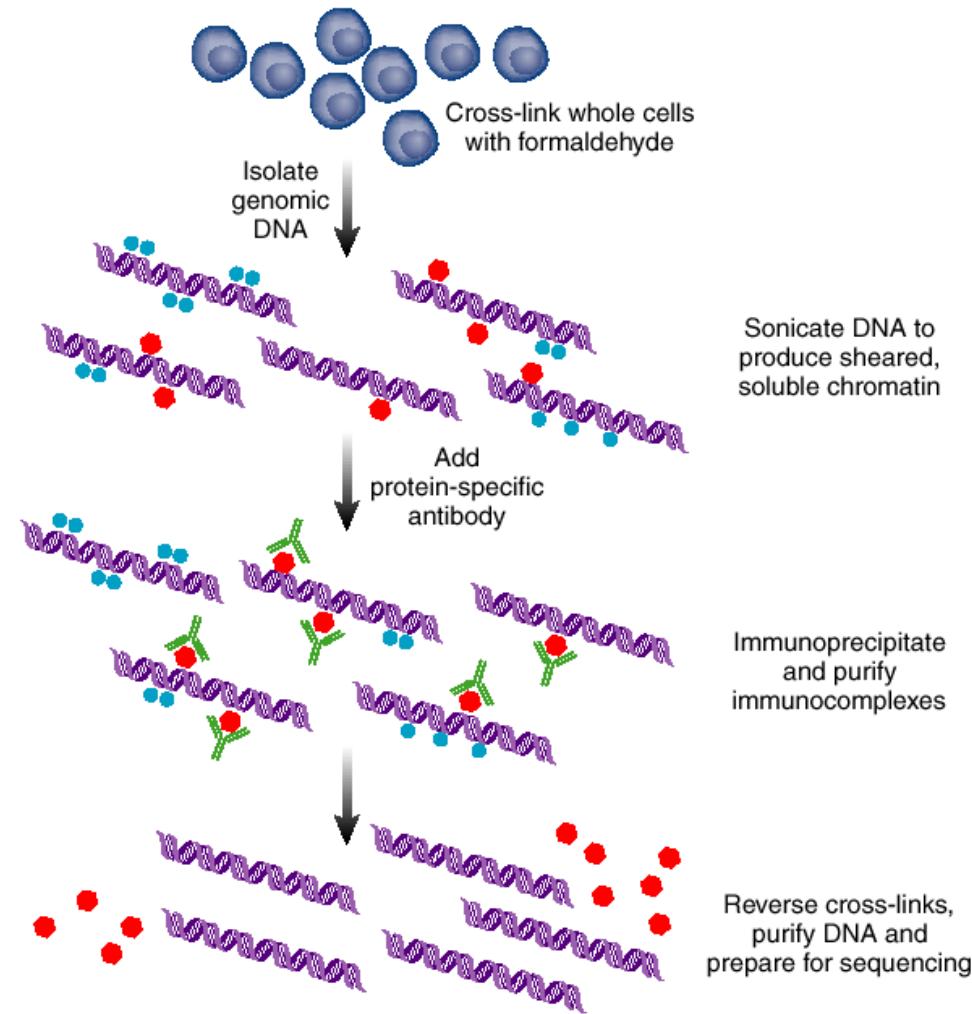
(A) Sample preparation and sequencing



(B) Computational analysis



Classical workflow: IP step



Current ChIP-seq approaches

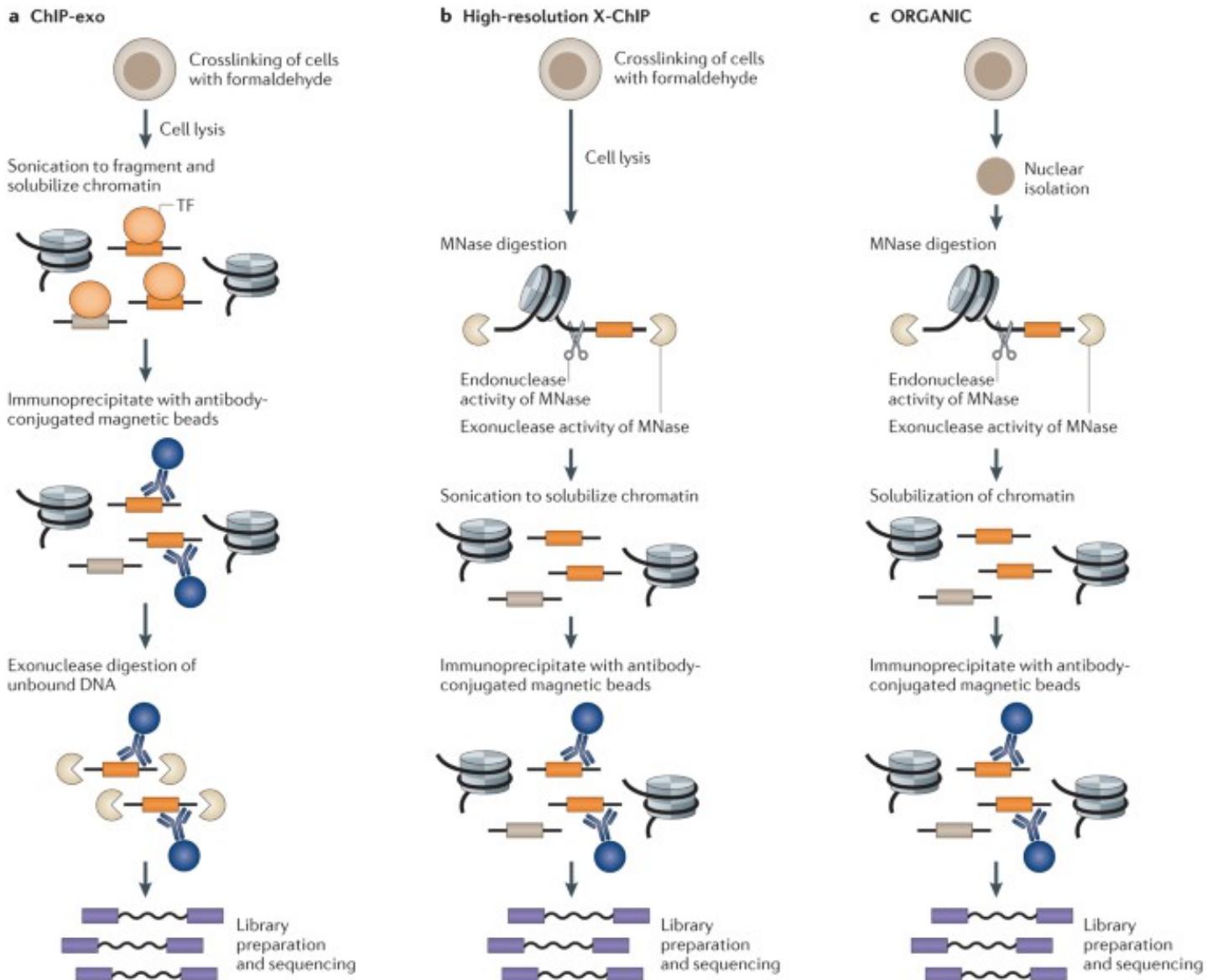
- ChIP-seq originally referred to a precise protocol, involving formaldehyde crosslinking, mechanical fragmentation and antibody-based chromatin pull-down.
- It became more of a “generic” term to describe any chromatin immunoprecipitation experimental approach backed by high-throughput sequencing.

Current ChIP-seq approaches

- ChIP-seq
- Low input ChIP-seq
- Native ChIP-seq
- Indirect ChIP-seq with DamID
- ChIP-seq with chemical-based fragmentation
- Cut&Run, Cut&Tag
- Single-cell ChIP-seq
- ...

Current ChIP-seq approaches

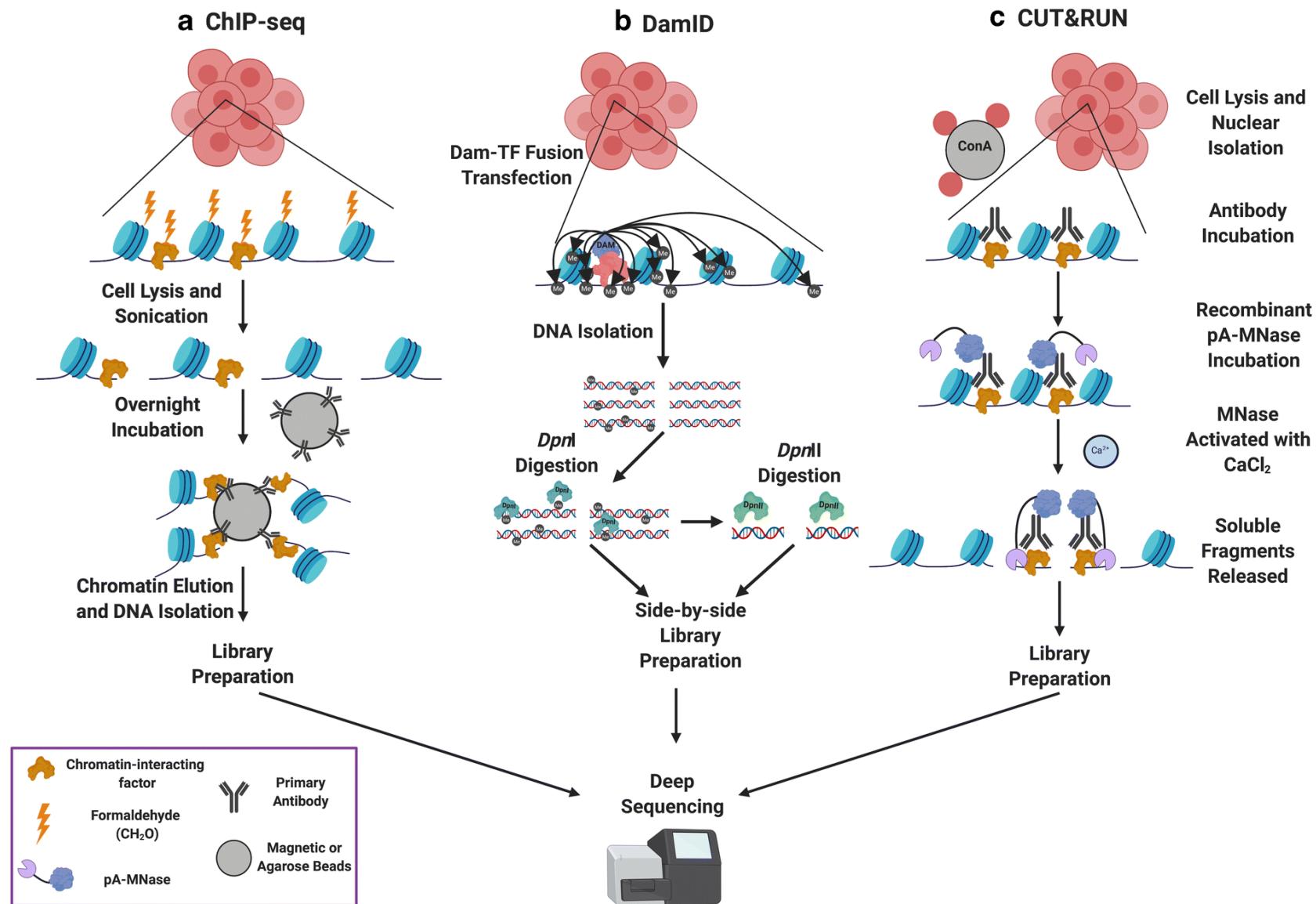
- Direct approaches



Zentner & Henikoff,
Nat. Rev. Genet.
2014

Current ChIP-seq approaches

- Indirect approaches



A plethora of commercial kits to help with ChIP-seq

- Kits help to reduce expenses when trying out a new method.
- They also help to “gently” dive in the methodology of the experimental steps.
- They (usually) provide a streamlined, easy version of an originally complex approach.
- Sometimes include downstream analysis workflows

A plethora of commercial kits to help with ChIP-seq

- Diagenode
- ActiveMotif
- Abcam
- Illumina
- ThermoFisher
- Qiagen
- TakaraBio
- Novogene
- Epigentek

A plethora of commercial kits to help with ChIP-seq

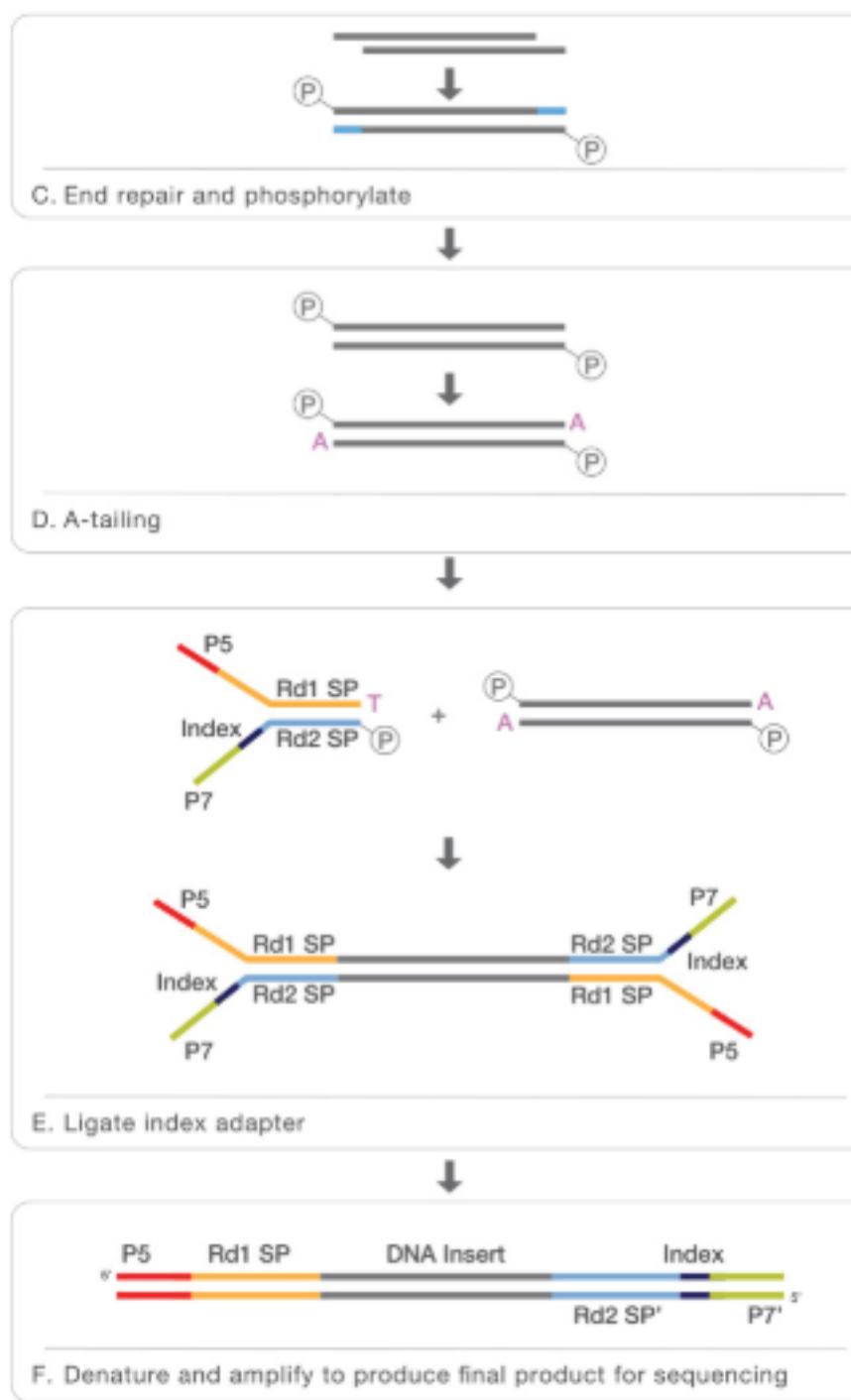
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The required input, reagents, yield and/or quality will vary depending on the kit.

Once someone has experience in (1) molecular biology practice and (2) making high-throughput sequencing libraries, it becomes more straightforward / cost-effective to perform ChIP-seq using in-house protocols.

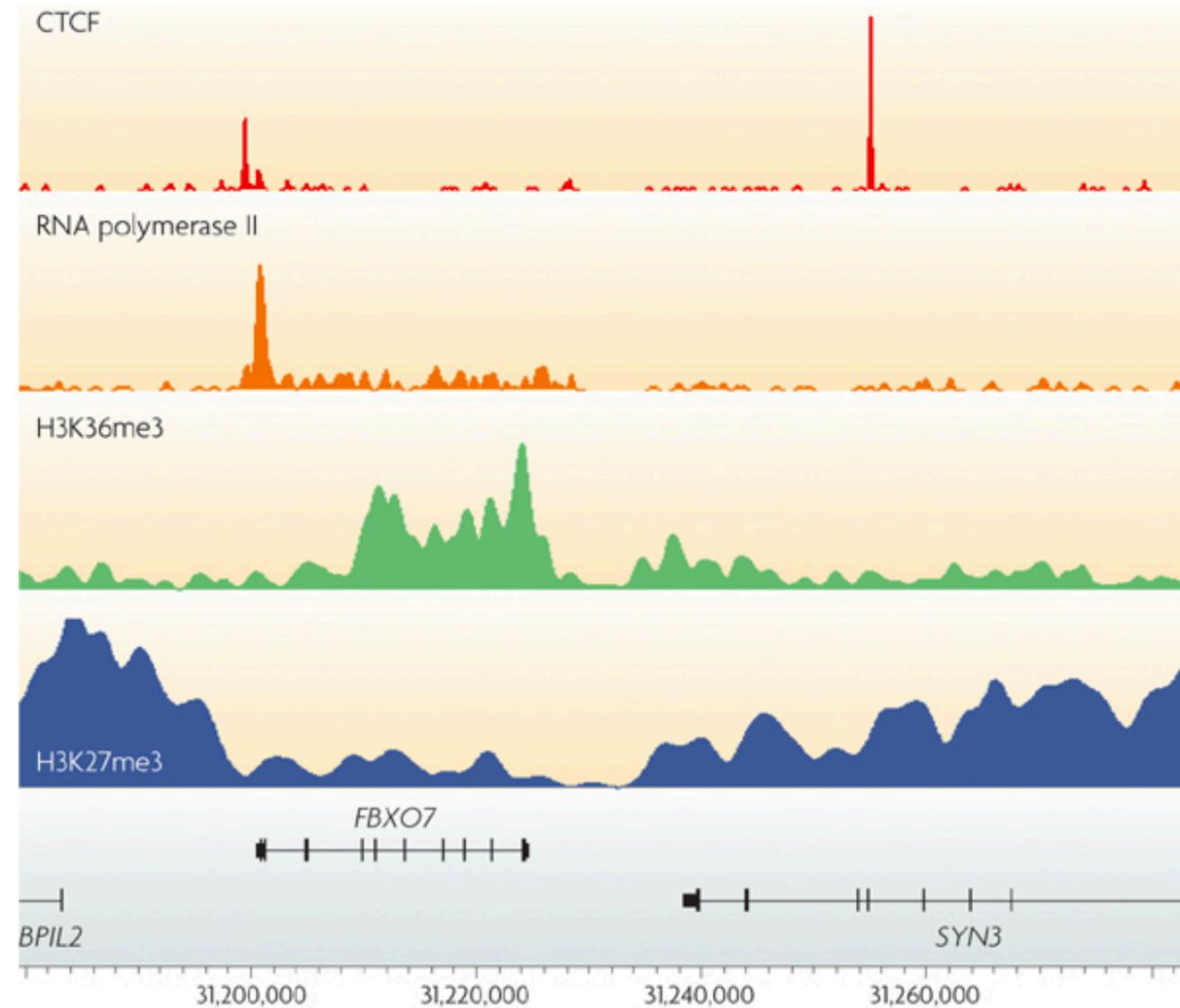
ChIP-seq outputs

- **[Biological material]**
High-throughput sequencing libraries



ChIP-seq outputs

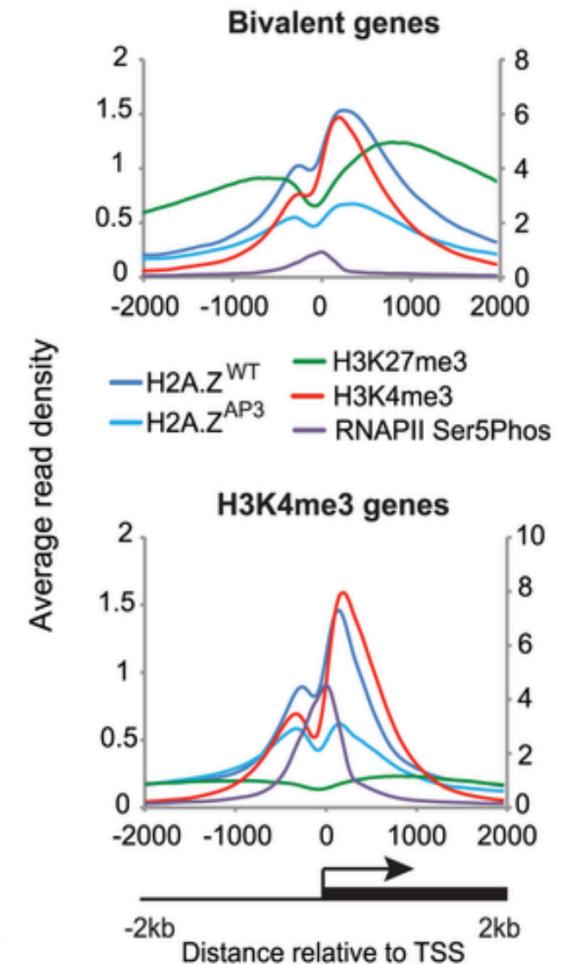
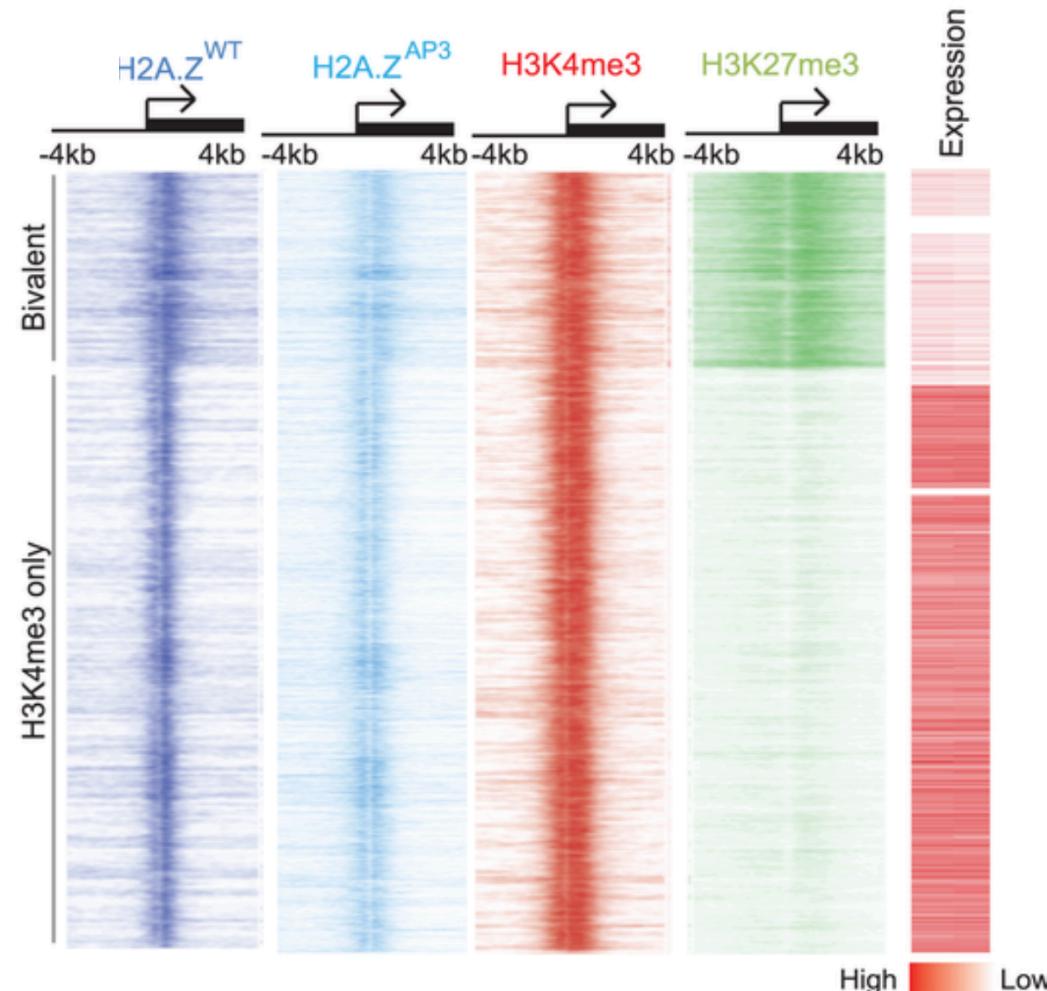
- Genome-wide coverage tracks
 - Can be directly viewed in a genome browser



Park et al.,
Nat. Rev. Genet.
2009

ChIP-seq outputs

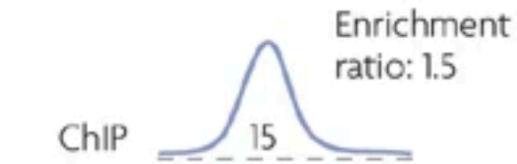
- Genome-wide coverage tracks
 - Can also be aligned at genomic features of interest (e.g. TSSs)



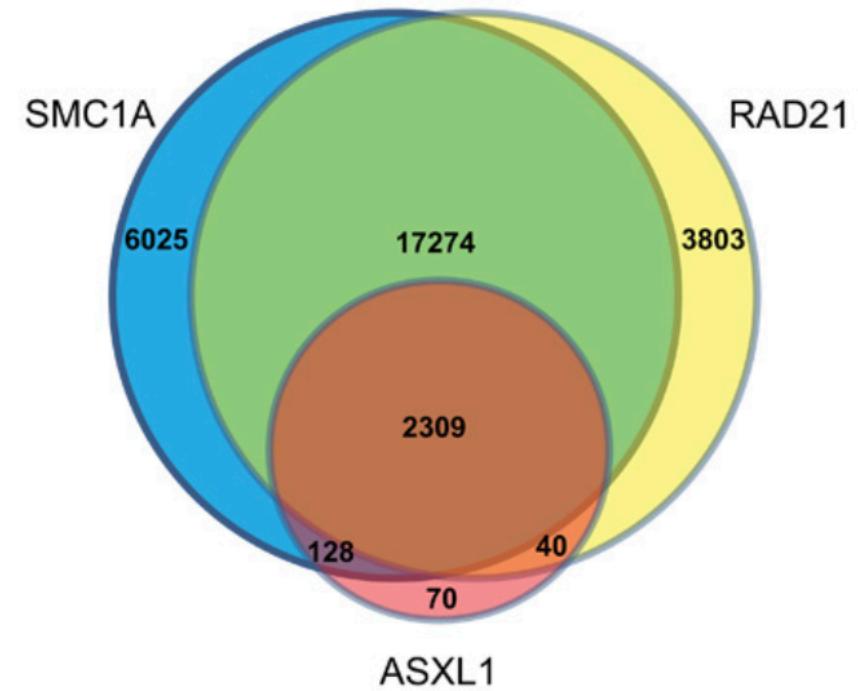
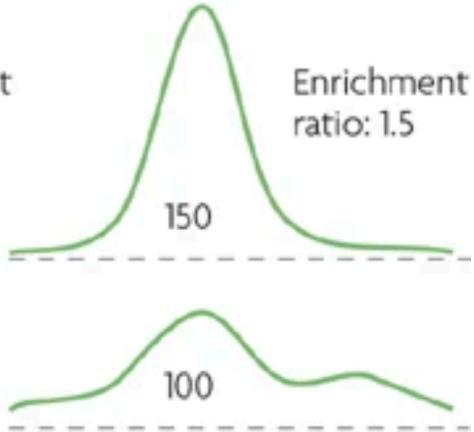
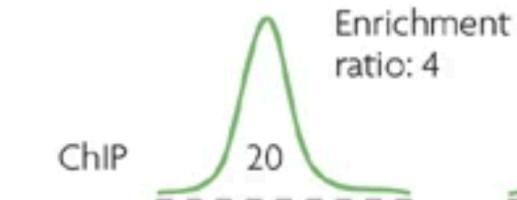
ChIP-seq outputs

- Peak sets

Ba Not statistically significant



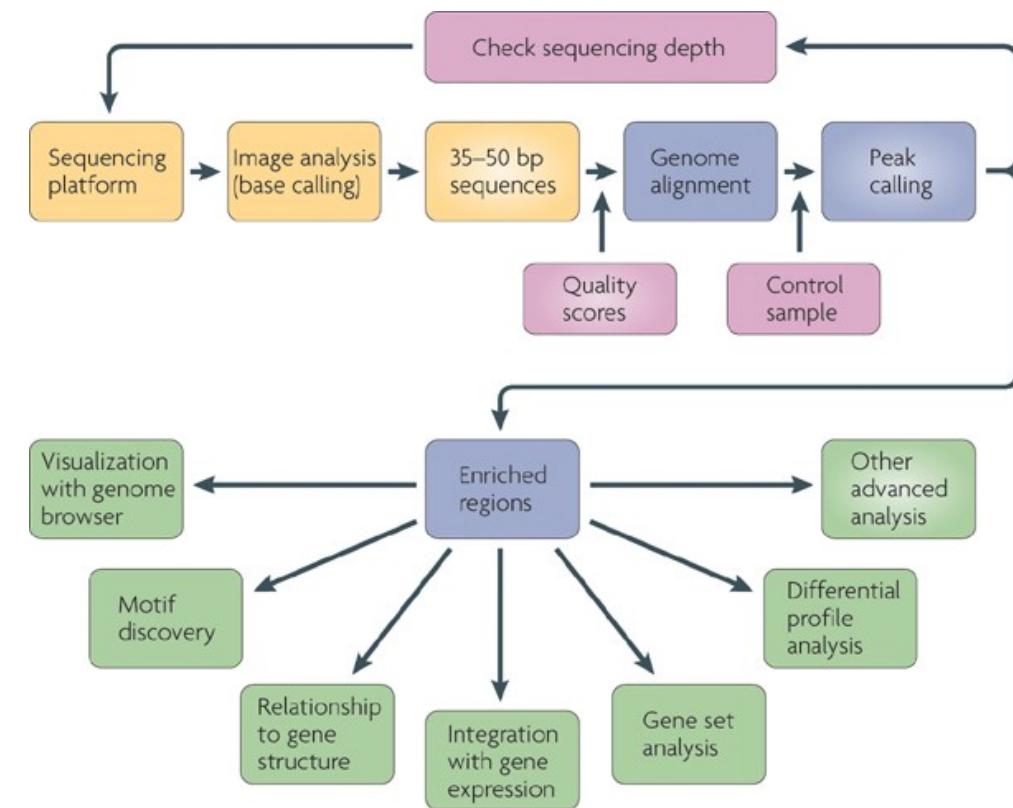
Bb Statistically significant



ChIP-seq outputs

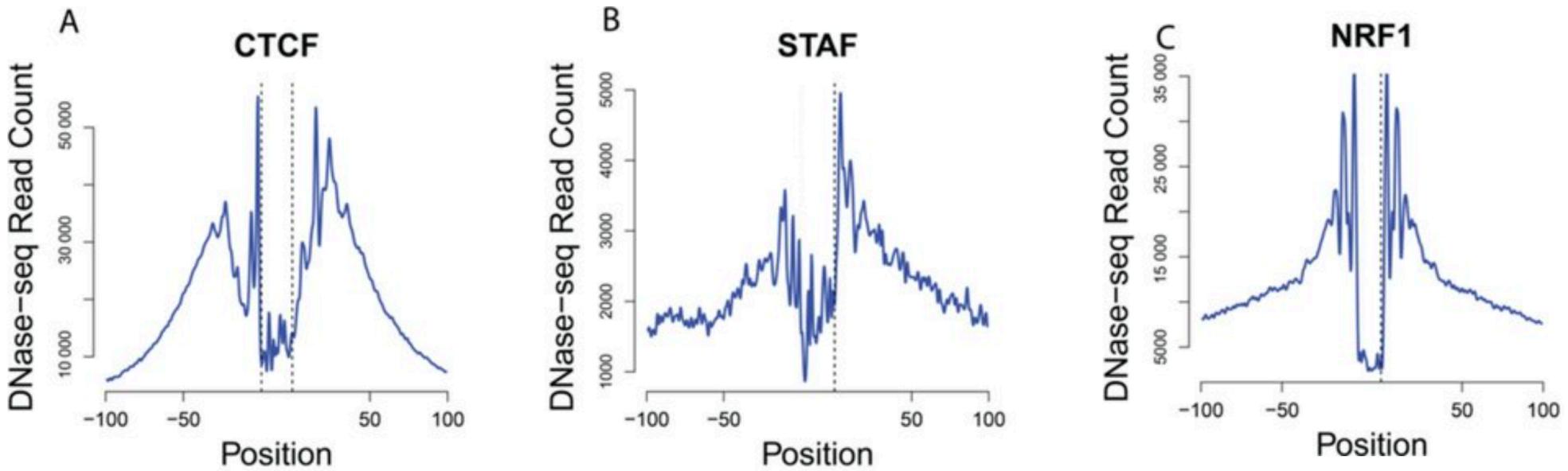
- And more from downstream analyses, according to the biological question:

- DNA binding motifs
- Differential binding of a factor
- Cooperation between factors
- Biological functions in which a factor is involved
- Temporal dynamics of factor binding
- Gene regulatory networks
- ...



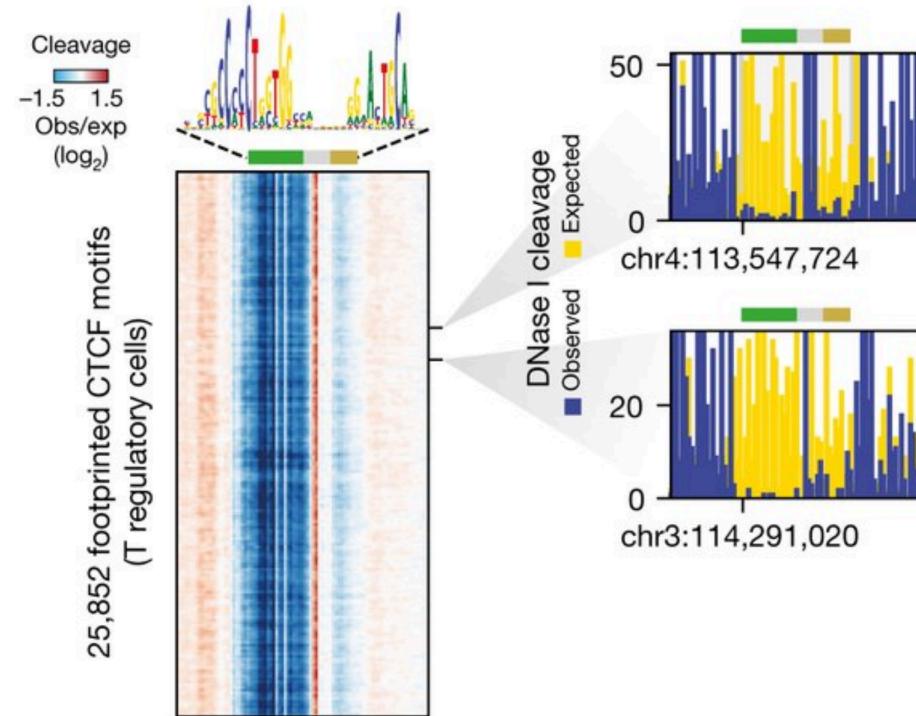
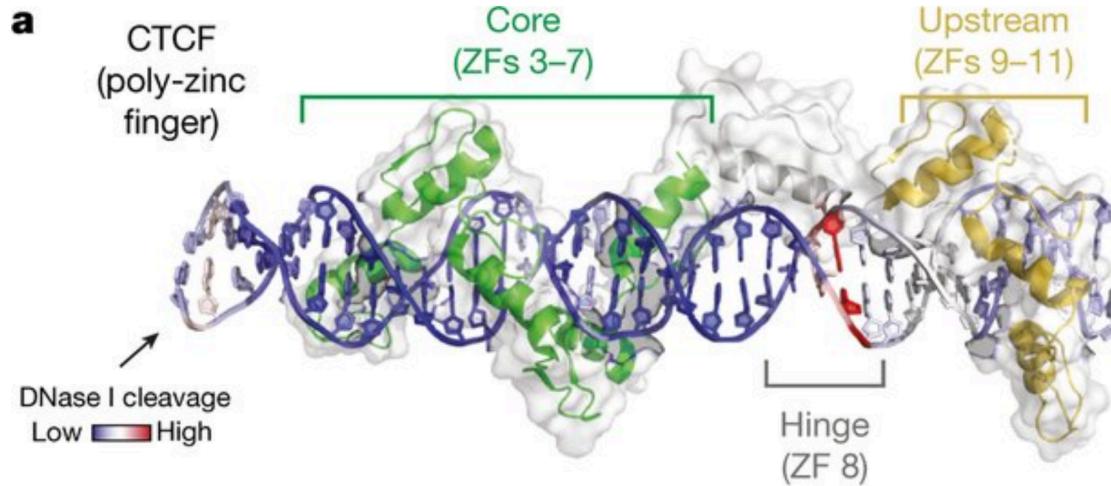
Alternative to chromatin immunoprecipitation approaches

- DNase footprinting methods



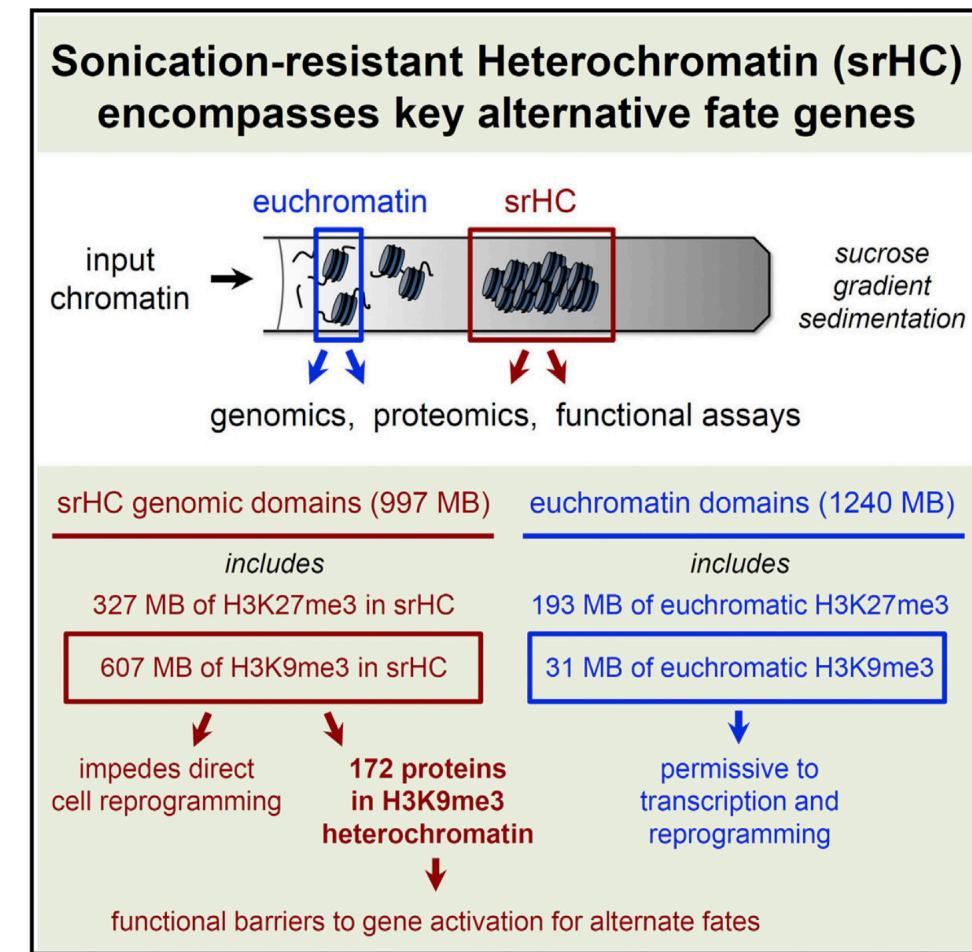
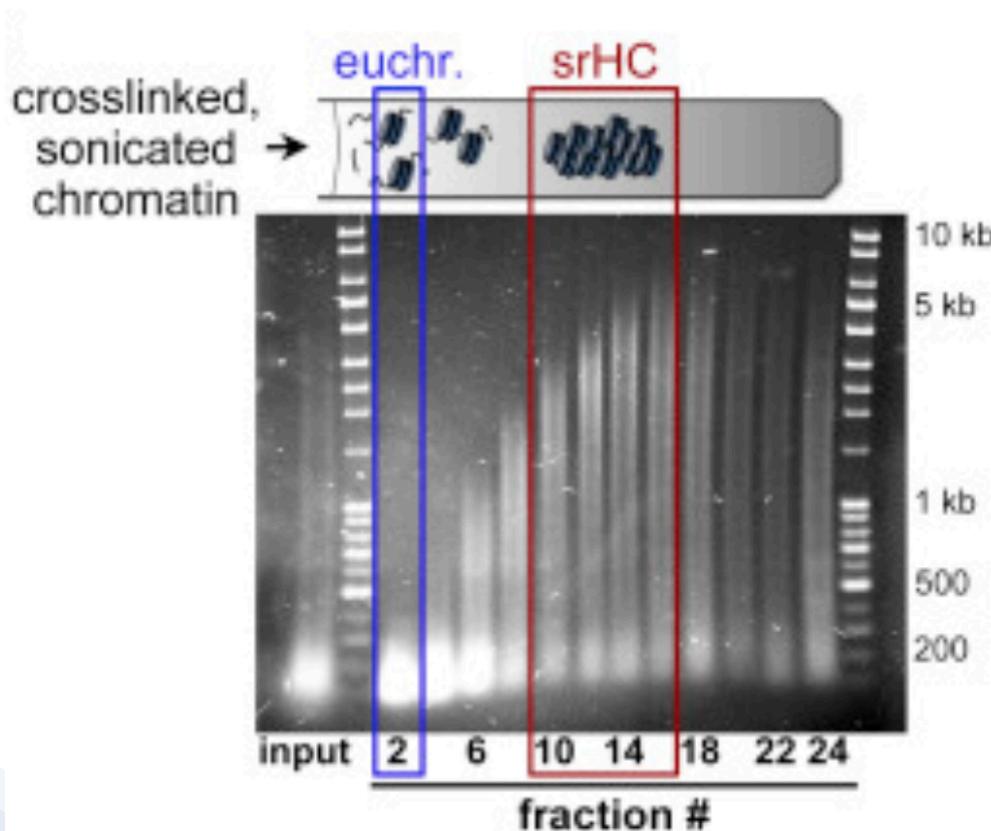
Alternative to chromatin immunoprecipitation approaches

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Alternative to chromatin immunoprecipitation approaches

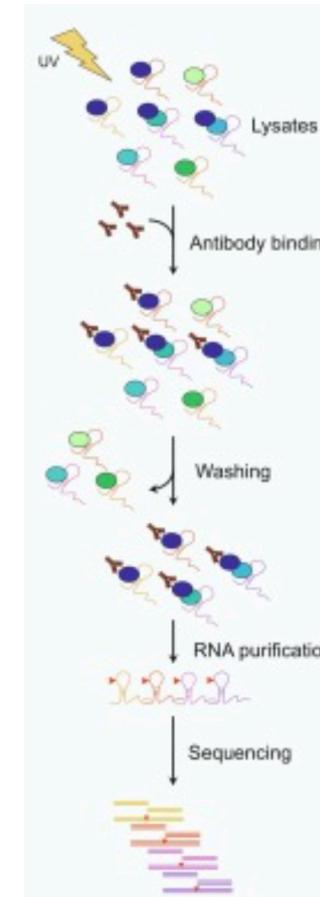
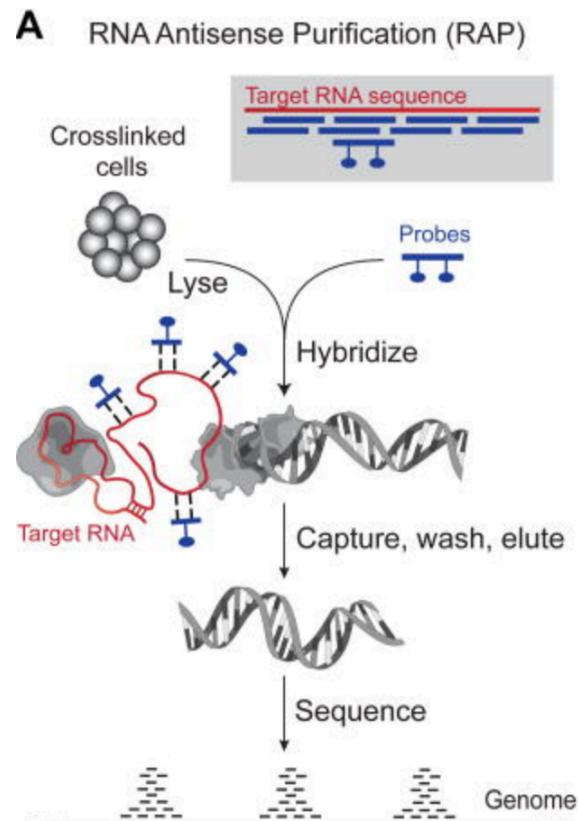
- Chromatin sedimentation methods



Going further: probing RNA-DNA or protein-RNA interactions

- Methods also based on crosslinking, this time between RNA and DNA or proteins and RNA

Engreitz et al.,
Science 2013



Wang & Zie,
Translational Epigenetics
2020

Some resources

- Github:
 - <https://github.com/danielecook/Awesome-Bioinformatics#chip-seq>
 - <https://github.com/crazyhottommy/ChIP-seq-analysis>
- Henikoff's lab methods website:
 - 20+ methods developed by the lab to profile chromatin
 - <https://research.fredhutch.org/henikoff/en/methods.html>
- Peter Park's review in Nature Review Genetics (2009)