Residuals and Diagnostics for Ordinal Regression Models: An Introduction to the sure package

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Abstract Residual diagnostics is an important topic in the classroom, but it is less often used in practice. Part of the reason for this is that more complex models, like cumulative link models and logistic regression, do not produce standard residuals that are easily interpreted as those in ordinary linear regression. In this paper, we introduce the concept of surrogate residuals and demonstrate their use through the R package **sure**.

Introduction

Categorical outcomes are encountered frequently in practice across different fields. For example, in medical studies, the outcome of interest is often binary (e.g., presence or absence of a particular disease after applying a treatment). It is also relatively common for a categorical outcome to have a natural ordering. For instance, in an opinion poll, the response may be satisfaction, with categories low, medium, and high. In this case, the response is ordered: low < medium < high.

The *cumulative link* model is a natural choice for modelling an ordinal outcome. Consider an ordinal categorical outcome $\mathcal Y$ with ordered categories $1 < 2 < \cdots < J$. In a cumulative link model, the cumulative probabilities are linked to the linear predictor according to

$$G^{-1}\left(\Pr\left\{\mathcal{Y}\leq j\right\}\right) = \alpha_j + f\left(X, \boldsymbol{\beta}\right),\tag{1}$$

where G is a continuous cumulative distribution function, α_j are the category-specific intercepts, X is a matrix of covariates, and β is a vector of fixed regression coefficients. The intercept parameters satisfy $-\infty = \alpha_0 < \alpha_1 < \dots < \alpha_{J-1} < \alpha_J = \infty$. We should point out that some authors (and software) use the alternate formulation

$$G^{-1}\left(\Pr\left\{\mathcal{Y}\geq j\right\}\right) = \alpha_{j}^{\star} + f\left(X, \boldsymbol{\beta}^{\star}\right),\tag{2}$$

This formulation provides coefficients that are consistent with the ordinary logistic regression model. The estimated coefficients from model (2) will have the opposite sign as those in model (1).

Another way to interpret the cumulative link model is through a *latent* continuous random variable $\mathcal{Z} = -f(X, \beta) + \epsilon$, where ϵ is a continuous random variable with location parameter 0, scale parameter 1, and cumulative distribution function $G(\cdot)$. We then construct an ordered factor according to the rule

$$y = j$$
 if $\alpha_{i-1} < z \le \alpha_i$.

For $\epsilon \sim N\left(0,1\right)$, this leads to the usual probit model for ordinal responses

$$\Pr\left\{\mathcal{Y} \leq j\right\} = \Pr\left\{\mathcal{Z} \leq \alpha_j\right\} = \Pr\left\{-f\left(\mathbf{X}, \boldsymbol{\beta}\right) + \epsilon \leq \alpha_j\right\} = \Phi\left(\alpha_j + f\left(\mathbf{X}, \boldsymbol{\beta}\right)\right).$$

Common choices for the link function $G^{-1}\left(\cdot\right)$ and the implied (standard) distribution for ϵ are described in Table 1.

Link	Distribution of ϵ	G(y)	$G^{-1}\left(p\right)$
logit ¹ probit log-log	logistic standard normal Gumbel (max)	$\exp(y) / [1 + \exp(y)]$ $\Phi(y)$ $\exp[-\exp(-y)]$	$\log [p/(1-p)]$ $\Phi^{-1}(p)$ $-\log [-\log (p)]$
complimentary log-log cauchit	Gumbel (min) Cauchy	$1 - \exp\left[-\exp\left(y\right)\right]$ $\pi^{-1}\arctan\left(y\right) + 1/2$	$\log \left[-\log \left(1-p \right) \right]$ $\tan \left(\pi p - \pi/2 \right)$

Table 1: Common link functions.

There are a number of R packages that can be used to fit cumulative link models (1) and (2). The recommended package MASS (Venables and Ripley, 2002) contains the function polr (proportional odds logistic regression) which, despite the name, can be used with all of the link functions described

in Table 1. The VGAM package (Yee, 2017) has the vglm function for fitting vector generalized linear models, which includes the broad class of cumulative link models. By default, vglm uses the same parameterization as in Equation (1), but provides the option for using the parameterization seen in Equation (2) as well; this will result in the estimated coefficients having the opposite sign. Package ordinal (Christensen, 2015) has the clm function for fitting cumulative link models. The popular rms package (Harrell Jr, 2017) has two functions: 1rm for fitting logistic regression and cumulative link models using the logit link, and orm for fitting ordinal regression models. Both of these functions use the parameterization seen in Equation (2).

For a continuous outcome \mathcal{Y} , the residual is traditionally defined as the difference between the observed and fitted values. For ordinal outcomes, the residuals are more difficult to define, and few definitions have been proposed in the literature. Liu et al. (2009) proposed using the cumulative sums of residuals derived from collapsing the ordered categories into multiple binary outcomes. Unfortunately, this method leads to multiple residuals for the ordinal outcome and therefore is difficult to interpret. Li and Shepherd (2012) showed that the sign-based statistic (SBS)

$$R_{SBS} = E\left\{sign\left(y - \mathcal{Y}\right)\right\} = Pr\left\{y > \mathcal{Y}\right\} - Pr\left\{y < \mathcal{Y}\right\},\tag{3}$$

can be used as a residual for proportional odds regression models; these are referred to later by Li and Shepherd as *probability-based residuals*, but we will follow Liu and Zhang (2017) and refer to them as SBS residuals. For an overview of the theoretical and graphical properties of the SBS residual (3), see Liu and Zhang (2017). These are available in the **PResiduals** package (Dupont et al., 2016). A limitation with the SBS residuals is that they are based on a discrete outcome and hence, are discrete themselves. This makes using them in various diagnostic plots far less useful.

Surrogate-based residuals

Liu and Zhang (2017) propose a new type of residual that is based on a continuous variable S that acts as a surrogate for the ordinal outcome \mathcal{Y} . This surrogate residual is defined as

$$R_{S} = S - E\left(S|X\right),\tag{4}$$

where S is a continuous variable based on the conditional distribution of the latent variable Z given Y. In particular, given Y = y, Liu and Zhang (2017) show that S follows a truncated distribution obtained by truncating the distribution of $Z = -f(X, \beta) + \epsilon$ using the interval (α_{y-1}, α_y) . The benefit of the surrogate-based residual (4) is that it is based on a continuous variable S, hence, S is also continuous.

Furthermore, it can be shown (Liu and Zhang, 2017) that if the hypothesized model agrees with the true model, then R_S will have the following properties:

symmetry around zero $E(R_S|X) = 0$;

homogeneity $Var(R_S|X) = c$, a constant that is independent of X;

reference distribution the empirical distribution of R_S approximates an explicit distribution that is related to the link function $G^{-1}(\cdot)$.

These properties allow for a thorough examination of the residuals to check model adequacy and misspecification of the mean structure and link function.

Jittering for general models

The latent method discussed in Section 2.2 applies to cumulative link models for ordinal outcomes. For more general models, we can define a surrogate using a technique called *jittering*. Suppose the true model for an ordinal outcome \mathcal{Y}

$$\mathcal{Y} \sim F_a(y; X, \boldsymbol{\beta}),$$
 (5)

where $F(\cdot)$ is a discrete cumulative distribution function. This model is general enough to cover the cumulative link models (1) and (2), and nearly any parametric or nonparametric model for ordinal outcomes.

Liu and Zhang (2017) suggest defining the surrogate *S* using either of the following two approaches:

- 1. jittering on the outcome scale: $S|\mathcal{Y} = y \sim \mathcal{U}[y, y + 1]$;
- 2. jittering on the probability scale: $S|\mathcal{Y} = y \sim \mathcal{U}[F_a(y-1), F_a(y)]$.

Once a surrogate is obtained, we define the surrogate residuals in the same way as Equation (4). In either case, if the hypothesized model is correct, then symmetry around zero still holds; that is $E(R_S|X) = 0$. For the latter case, if the hypothesized model is correct then $R_S|X \sim \mathcal{U}(-1/2,1/2)$. In other words, jittering on the probability scale has the additional property that the conditional distribution of R_S given X has an explicit form. This allows for a full examination of the distributional information of the residual.

Bootstrapping

Since surrogate residuals are based on sampling, additional error is introduced. One way to minimize this sampling error and help stabilize any patterns in diagnostic plots is to use the bootstrap (Efron, 1979).

The procedure for bootstrapping surrogate residuals is similar to the model-based bootstrap algorithm used in linear regression. To obtain the b-th bootstrap replicate of the residuals, Liu and Zhang (2017) suggest the following algorithm:

Step 1 Perform a standard case-wise bootstrap of the original data to obtain the bootstrap sample $\{(X_{1b}^{\star}, \mathcal{Y}_{1b}^{\star}), \dots, (X_{nk}^{\star}, \mathcal{Y}_{nk}^{\star})\}.$

Step 2 Using the procedure outlined in the previous section, obtain a sample of surrogate residuals $R_{S_{1h}}^{\star}, \dots, R_{S_{nh}}^{\star}$ using the bootstrapped data obtained in **Step 1**.

This procedure is repeated a total of B times. For residual vs. covariate (i.e., R vs. X) plots and residual vs. fitted value (i.e., R vs. $f(X, \beta)$) plots, we simply scatter all $B \times n$ residuals on the same plot. For large data sets, we find it useful to also lower the opacity of the data points to help alleviate any issues with over plotting. This approach is valid since the bootstrap samples are drawn independently from each other. For Q-Q plots, on the other hand, Liu and Zhang (2017) suggest using the median of the B bootstrap distributions which is the implementation used in the **sure** package.

Surrogate-based residuals in R

The sure package (Greenwell et al., 2017) supports a variety of R packages for fitting cumulative link and other types of models. The supported packages and their corresponding functions are described in Table 2.

Package	Function(s)	Model	Parameterization
stats	glm	binary regression	NA
MASS	polr	cumulative link	$Pr\left\{ \mathcal{Y}\leq j\right\}$
rms	1rm	cumulative link	$Pr\left\{ \mathcal{Y} \geq j \right\}$
	1rm	logistic regression	NA
	orm	cumulative link	$Pr\left\{ \mathcal{Y}\geq j\right\}$
ordinal	clm	cumulative link	$Pr\left\{\mathcal{Y}\leq j\right\}$
VGAM	vglm	cumulative link	$Pr\left\{\mathcal{Y} \leq j\right\}^2$

Table 2: Modelling packages supported by **sure** and the corresponding parameterization they use for fitting cumulative link models.

The **sure** package currently only exports four functions:

- resids—for constructing surrogate residuals;
- surrogate—for generating the surrogate response values used in the residuals;
- autoplot—produce various diagnostic plots using ggplot2 graphics (Wickham, 2009);
- gof—simulate p-values from a goodness-of-fit test.

In addition, the package also includes three simulated data sets: df1, df2, and df3. These data sets are used to demonstrate various uses of the surrogate residual approach throughout this paper.

Detecting a misspecified mean structure

For illustration, the data frame df1 contains n = 2000 observations from the following cumulative link model:

$$Pr\{\mathcal{Y} \le j\} = \Phi\left(\alpha_j + \beta_1 X + \beta_2 X^2\right), \quad j = 1, 2, 3, 4,$$
 (6)

where $\alpha_1 = -16$, $\alpha_2 = -12$, $\alpha_3 = -8$, $\beta_1 = 8$, $\beta_2 = -1$, and $X \sim \mathcal{U}(1,7)$. These simulated data are available in the df1 data frame from the **sure** package and are loaded automatically with the package; see ?df1 for details. Below, we fit a (correctly specified) probit model using the polr function from the **MASS** package.

```
# Load required package(s)
library(MASS)

# Fit a cumulative link model with probit link
fit.polr <- polr(y ~ x + I(x ^ 2), data = df1, method = "probit")</pre>
```

The code chunk below obtains the SBS residuals (3) from the previously fitted probit model fit.polr using the **PResiduals** package and constructs a couple of diagnostic plots. The results are displayed in Figure 1.

```
# Load required package(s)
library(PResiduals)
# Obtain the SBS/probability-scale residuals
pres <- presid(fit.polr)</pre>
# Residual vs. covariate plot
p1 \leftarrow ggplot(data.frame(x = df1$x, y = pres), aes(x, y)) +
 geom_point(alpha = 0.5) +
 geom_smooth(color = "red", se = FALSE) +
 ylab("Probability-scale residual")
# Q-Q plot of the residuals
p2 <- ggplot(data.frame(y = pres), aes(sample = y)) +</pre>
 stat_qq(distribution = qunif, dparams = list(min = -1, max = 1), alpha = 0.5) +
 xlab("Sample quantile") +
 ylab("Theoretical quantile")
# Figure 1
grid.arrange(p1, p2, ncol = 2)
```

(**Note:** the reference distribution for the SBS residual is the $\mathcal{U}(-1,1)$ distribution.) As can be seen in Figure 1, the SBS residuals, which are inherently discrete, often display unusual patterns in diagnostic plots, making them less useful as a diagnostic tool. There is a pattern for each of the J=4 classes!

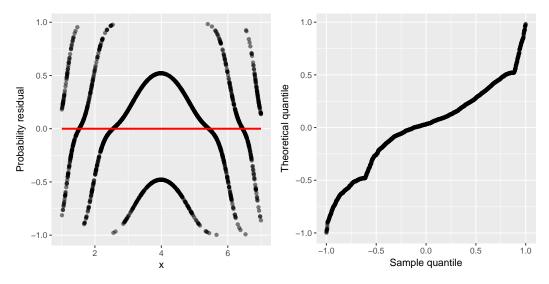


Figure 1: SBS residual plots for the (correctly specified) probit model fit to the df1 data set. *Left*: Residual vs. covariate plot. *Right*: Q-Q plot of the residuals. Nonparametric smooths are indicated by red curves.

Similarly, we can use the resids function in package **sure** to obtain the surrogate-based residuals discussed in Section 2.2. This is illustrated in the following code chunk. the results are displayed in Figure 2.

```
# Load required package(s)
library(ggplot2)
library(sure)

# Obtain surrogate-based residuals
set.seed(101) # for reproducibility
sres <- resids(fit.polr)

# Residual vs. covariate plot
p1 <- autoplot(sres, what = "covariate", x = df1$x, xlab = "x")

# Q-Q plot of the residuals
p2 <- autoplot(sres, what = "qq", distribution = pnorm)

# Figure ?
grid.arrange(p1, p2, ncol = 2)</pre>
```

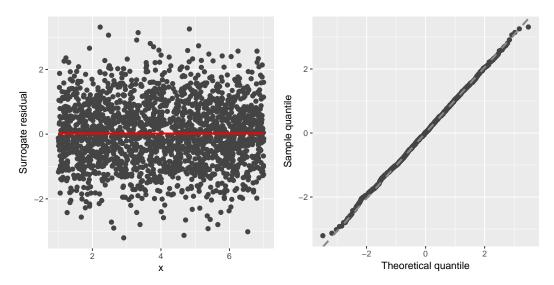


Figure 2: Surrogate-based residual plots for the (correctly specified) probit model fit to the df1 data set. *Left*: Residual vs. covariate plot. *Right*: Q-Q plot of the residuals. Nonparametric smooths are indicated by red curves.

We also wrote autoplot methods for various classes of models listed in Table 2, so you can just give autoplot the fitted model directly. The benefit of this approach is that the fitted values and reference distirbution (used in Q-Q plots) are automatically extracted. For example, to reproduce the Q-Q plot in Figure 2, we could have just used

```
set.seed(101) # for reproducibility
autoplot(fit.polr, what = "qq") # same as top right of Figure 1
```

Suppose that we did not include the quadratic term in our fitted model. We could expect a residual-vs-*x* plot to clearly indicate that such a (correct) quadratic term is missing. Below we update the previously fitted model by removing the quadratic term, then update the residual-vs-covariate plots (code not shown). The updated residual plots are displayed in Figure 3.

```
fit.polr <- update(fit.polr, y \sim x) # remove quadratic term
```

The SBS residuals gives some indication of a misspecified mean structure, but this only becomes more clear with increasing *J*, and the plot is still discrete. This is overcome by the surrogate residuals which produces a residual plot not unlike those seen in ordinary linear regression models.

Detecting heteroscedasticty

One issue that often raises concerns in statistical inference is that of heteroscedasticity; that is, when the error term has non constant variance. Heteroscedasticity can bias the statistical inference and lead to improper standard errors, confidence intervals, and *p*-values. Therefore, it is imperative to identify heteroscedacticity whenever present and take appropriate action (e.g., transformations, etc.).

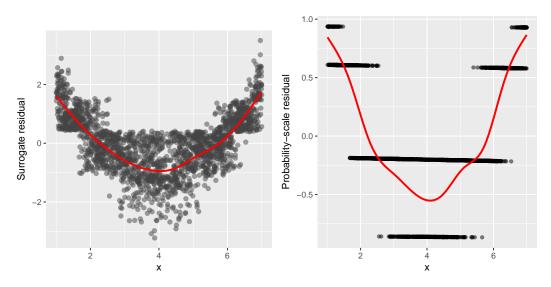


Figure 3: Residual-vs-covariate plots for a probit model with a misspecified mean structure fit to the simulated data from model (6). *Left*: Surrogate residuals. *Right*: SBS residuals. Nonparametric smooths are indicated by red curves.

In ordinary linear regression, this topic has been covered extensively. For categorical models, on the other hand, not much has been proposed in the literature.

As discussed in Section 2.2, one of the properties of the surrogate-based residual R_S is that, if the model is specified correctly, then $Var(R_S|X) = c$, where c is a constant.

For this example, we generated n = 2000 observations from the following ordered probit model:

$$Pr\left\{ \mathcal{Y} \leq j \right\} = \Phi\left\{ \left(\alpha_j + \beta X \right) / \sigma_X \right\}, \quad j = 1, 2, 3, 4, 5,$$

where $\alpha_1 = -36$, $\alpha_2 = -6$, $\alpha_3 = 34$, $\alpha_4 = 64$, $\beta = -4$, $X \sim \mathcal{U}(2,7)$, and $\sigma_X = X^2$. Notice how the variability is an increasing function of X. These data are available in the df2 data frame that is automatically loaded with the **sure** package; see ?df2 for details.

The following block of code uses the orm function from the popular **rms** package to fit a probit model to the simulated data. **Note** that we had to set x = TRUE in the call to orm in order to use the presid function later.

```
# Load required package(s)
library(rms)

# Fit a cumulative link model with probit link
fit.orm <- orm(y ~ x, data = df2, family = "probit", x = TRUE)</pre>
```

If heteroscedasticity is present, we would expect this to show up in various diagnostic plots, such as a residual vs. covariate plot in this case. Below we obtain the SBS and surrogate residuals as before and plot them against *X*. The results are displayed in Figure 4.

```
pres <- presid(fit.orm) # SBS residuals
set.seed(102) # for reproducibility
sres <- resids(fit.orm) # surrogate residuals

# Residual vs. covariate plots
p1 <- autoplot(sres, what = "covariate", x = df2$x, xlab = "x")
p2 <- ggplot(data.frame(x = df2$x, y = presid(fit.orm)), aes(x, y)) +
geom_point(size = 2, alpha = 0.25) +
geom_smooth(col = "red", se = FALSE) +
ylab("Probability scale residual")

# Figure ?
grid.arrange(p1, p2, ncol = 2)</pre>
```

In Figure 4, it is clear from the plot of the surrogate residuals (left side of Figure 4) that the variance increases with *X*, a clear sign of heteroscedasticity. As a matter fact, the plot suggests that the true

link function has a varying scale parameter, $\sigma = \sigma(X)$. The plot of the SBS residuals (right side of Figure 4), on the other hand, gives no indication of an issue with nonconstant variance.

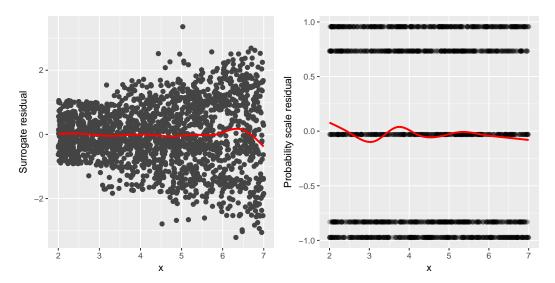


Figure 4: Residual vs. covariate plots for the simulated data. *Left*: Surrogate residuals. *Right*: SBS residuals.

Detecting a misspecified link function

For this example, we simulated n = 2000 observations from the following model

$$Pr(\mathcal{Y} \le j) = \Phi(\alpha_j + \beta_1 X + \beta_2 X^2), \quad j = 1, 2, 3, 4$$

where $G(\cdot)$ is the cumulative distribution function for the gumbel distribution. The data are available in the data frame df3 available with the package; see ?df3 for details.

below we fit a model with various link functions. For this model, however, the correct link function to use is the log-log link. From these models, we construct Q-Q plots of the residuals using R=100 bootstrap replicates. The results are displayed in Figure 5.

```
# Fit models with various link functions to the simulated data fit.probit <- polr(y \sim x + I(x ^{\circ} 2), data = df3, method = "probit") fit.logistic <- polr(y \sim x + I(x ^{\circ} 2), data = df3, method = "logistic") fit.loglog <- polr(y \sim x + I(x ^{\circ} 2), data = df3, method = "loglog") # correct link fit.cloglog <- polr(y \sim x + I(x ^{\circ} 2), data = df3, method = "cloglog") # Construct Q-Q plots of the surrogate residuals for each model p1 <- autoplot(fit.probit, nsim = 100, what = "qq") p2 <- autoplot(fit.logistic, nsim = 100, what = "qq") p3 <- autoplot(fit.loglog, nsim = 100, what = "qq") p4 <- autoplot(fit.cloglog, nsim = 100, what = "qq") # Figure ? grid.arrange(p1, p2, p3, p4, ncol = 2) # bottom left plot is correct model
```

From the Q-Q plots in Figure 5, it is clear the the model with the log-log link (which corresponds to gumbel errors in the latent variable formulation) is the most appropriate, while the other plots indicate deviations from the hypothesized model.

Checking the proportionality assumption

An important feature of the cumulative link model (1) is the proportional odds assumption, which assumes that mean structure, $f(X, \beta)$, remains the same for each of the J categories; for the logit case (see row one of Table 1), this is also referred to as the proportional odds assumption. Harrell (2001, pp. 334–335) suggests computing each observation's contribution to the first derivative of the log likelihood function with respect to β , average them within each of the J categories, and examine any trends in the

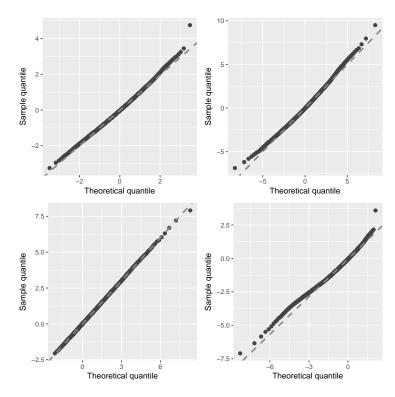


Figure 5: Q-Q plots of the residuals for various cumulative link models fit to simulated data with gumbel errors. *Top left*: A model with probit link. *Top right*: A model with logit link. *Bottom left*: A model with log-log link (i.e., the correct model). *Top left*: A model with complimentary log-log link.

residual plots, but these plots can be difficult to interpret. Fortunately, it is relatively straight forward to use the simulated surrogate response values *S* to check the proportionality assumption.

To illustrate, we generated 2000 observations from each of the following two probit models

$$Pr\left(\mathcal{Y} \leq j\right) = \Phi\left(\alpha_{j} + \beta_{1}X\right), \quad j = 1, 2, 3, \quad \text{and} \quad Pr\left(\mathcal{Y} \leq j\right) = \Phi\left(\alpha_{j} + \beta_{2}X\right), \quad j = 4, 5,$$

where $\alpha_1 = -1.5$, $\alpha_2 = 0$, $\alpha_3 = 1$, $\alpha_4 = 3$, $\beta_1 = 1$, and $\beta_2 = 1.5$. The data are available in the data frame df4 included with the package; see ?df4 for details.

Checking the proportionality assumption here amounts to checking whether or not $\beta_1 - \beta_2 = 0$. As outlined in Liu and Zhang (2017), we can generate surrogates $S_1 \sim \mathcal{N}\left(-\beta_1 X, 1\right)$ and $S_s \sim \mathcal{N}\left(-\beta_2 X, 1\right)$, both conditional on X. We then define the difference $D = S_2 - S_1$ which, conditional on X, follows a $\mathcal{N}\left((\beta_1 - \beta_2) X, 1\right)$ distribution. If $\beta_1 - \beta_2 = 0$, then D should be independent of X. This can be easily checked by plotting D against X. Below, we use the surrogate function to generate the surrogate response values directly (as opposed to the residuals) and generate the D vs. X plot shown in Figure 6.

From Figure 6, it is clear that $\beta_1 - \beta_2 \neq 0$; hence, the proportionality assumption does not hold.

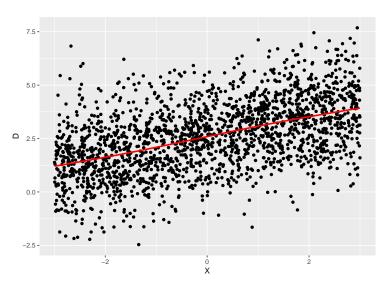


Figure 6: Scatterplot of $D = S_1 - S_2$ vs. X. A nonparametric smooth is represented by the red curve.

Summary

In this paper, we have introduced the **sure** package which implements the surrogate approach to residuals for ordinal regression models described in Liu and Zhang (2017). Using simulated data sets, we have illustrated the various properties of these residuals and how they can be used to check various assumptions in the ordinal regression model (e.g., heteroscedasticity, misspecified link functions, etc.) using a continuous residuals which produces diagnostic plots not unlike those seen in ordinary linear regression. This offers a new way of performing typical diagnostic checks for ordinal regression models that are easily interpreted by the analyst. Furthermore, this new approach to constructing residuals for ordinal regression models is still under active development and new functionality will be added to **sure** accordingly in the future.

Acknowledgments

TBD.

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