## 6\_UMAP\_and\_Clustering\_isolated\_domain

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```
[1]: from tqdm import tqdm
import mdtraj as md
import numpy as np
import matplotlib
import matplotlib.pyplot as plt
import umap
import hdbscan
import seaborn as sns
```

## 1 UMAP and clustering of isolated C-domain

Here the analysis for the closeness centralities for the C-domain (CD) of FAT10 is shown. They were calculated in 5\_Calculating\_Closeness\_Centralities\_Isolated\_Domain.ipynb for the CD only, ignoring contacts with the rest of the protein.

#### 1.1 Loading closeness centralities

```
[2]: closeness_CD = np.load("./Closeness_Centralities_C_domain_FAT10_res_88_159.npy")
```

#### 1.2 Loading the trajectory

```
[19]: topology_file = "./start_frame_FAT10.pdb"
traj_file = "./FAT10_123_traj_center_dt100.xtc"

traj = md.load(traj_file, top = topology_file)
```

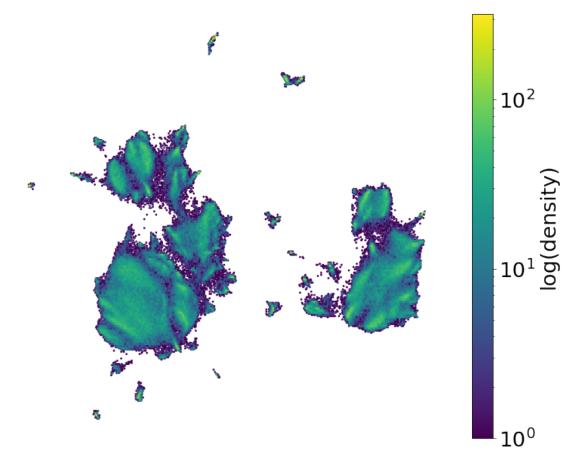
## 1.3 Calculating the UMAP embedding

To calculate the emebdding, UMAP (Uniform Manifold Approximation and Projection for Dimension Reduction) was used in its python implementation with all parameters left at default settings. Please refer to the documentation for more details.

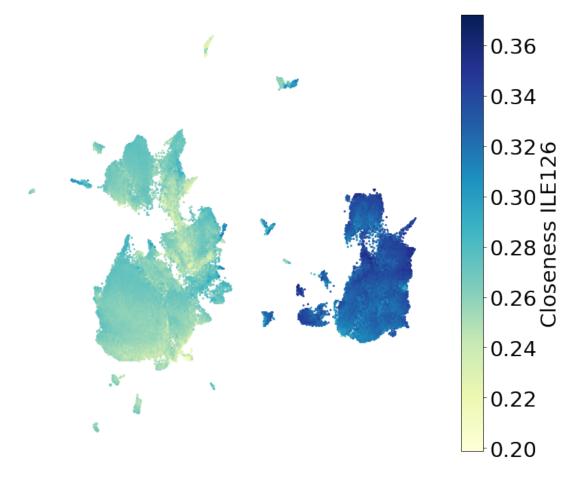
```
[3]: reducer = umap.UMAP()
embedding = reducer.fit_transform(closeness_CD/closeness_CD.max()) #__

\therefore normalizing to have max = 1 in case this was not done upon calculation.
```

## 1.4 Plotting UMAP colored by log(density)



## 1.5 Coloring UMAP by closeness centrality of Ile126



#### 1.6 Note: UMAP is non-deterministic.

This means the resulting embeddings can deviate between runs. The embeddings may appear rotated or flipped or differ slightly in other details. This can lead to differing results when using HDBSCAN, such as deviations in cluster numbering. To ensure consistency with the analyses in the paper, we provide the coordinates of the UMAP that was used in the paper and proceed in the analyses with those.

```
[13]: embedding = np.load("./UMAP_low_d_coordinates_isolated_domain.npy")
```

### 1.7 HDBSCAN Clustering

The parameters were selected to divide the UMAP into several large regions to get an overview of the most dominant differences. Based on the density, the clustering could be made in a substantially more fine-grained manner.

For details on the HDBSCAN parameters, please refer to https://hdbscan.readthedocs.io/en/latest/parameter\_selection.html.

```
[14]: clusterer = hdbscan.HDBSCAN(min_cluster_size=1500, min_samples=1500)
    clusterer.fit(embedding)
    cluster_ids = clusterer.labels_
```

#### 1.7.1 Custom color map

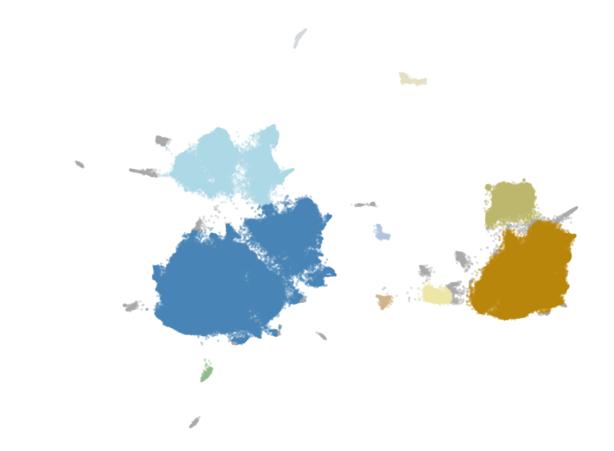
```
[15]: color_list = [matplotlib.colors.hex2color(hexcolor) for hexcolor in [
          "#A9A9A9",
                          # Neutral Grey (entry 1)
          "#E6E1C5",
                          # Light Grey-Yellow (muted darkgoldenrod)
          "#D3DADD",
                          # Light Grey-Blue (muted steelblue)
          "#BOC4DE",
                          # Light Steelblue
                          # Tan (muted brown-gold)
          "#D2B48C",
          "#BDB76B",
                          # Dark Khaki (muted gold-brown)
                          # Dark seagreen
          "#8FBC8F",
                          # Steelblue (entry 8)
          "#4682B4",
          "#ADD8E6",
                          # Light Blue
                          # Pale Goldenrod
          "#EEE8AA",
          "#B8860B",
                          # Darkgoldenrod (entry 11)
      ]]
      color_map = matplotlib.colors.LinearSegmentedColormap.

→from_list("custom_colormap", color_list)
```

## 1.8 Coloring the UMAP based on cluster IDs

gray indicates cluster $_{id} = -1$ , i.e. HDBSCAN classifies these points as noise.

```
[16]: %matplotlib inline
fig, ax = plt.subplots(figsize = (10,8),tight_layout = True)
```



1.9 Defining the residues which belong to the C-domain for writing out exemplar structures

```
[21]: CD_start_res = 87 # Starting residue of the C-Domain (0-Indexed. This is → residue LEU88 in the structure)

CD_end_res = 158 # End residue of the C-Domain (0-Indexed. This is ALA159 in → the structure).
```

# 1.10 Plotting and writing out cluster exemplar structures based on HDBSCAN clusterer.exemplars\_

This serves the purpose of obtaining several structures that are characteristic for a cluster. The cluster exemplars are colored in red in the plot. It is evident that the exemplars are not sampled from just a single location in the cluster, but several different regions, which HDBSCAN considers characteristic for the cluster. More details on writing out exemplars are explained in 4\_Clustering\_EncoderMap\_full\_length\_FAT10.ipynb.

```
[23]: n_representatives = 10
     %matplotlib inline
     plt.rcParams.update({'font.size': 25})
     fig, ax = plt.subplots(figsize = (10,8),tight_layout = True)
     %matplotlib inline
     fig, ax = plt.subplots(figsize = (10,8),tight_layout = True)
     ax.scatter(embedding[:, 0], embedding[:, 1],
               s=10,
               marker="o",
               linewidths=0,
               alpha = 0.4,
               c = cluster_ids,
               cmap = color map)
     ax.set_axis_off()
     for cluster in tqdm(np.unique(cluster_ids)):
         if (cluster > -1):
            cluster_exemplars_xy = clusterer.exemplars_[cluster]
             if (len(cluster_exemplars_xy) > n_representatives):
                reduction_factor = int(np.ceil(len(cluster_exemplars_xy)/
      →n_representatives))
                cluster_exemplars_xy = cluster_exemplars_xy[::reduction_factor]
             cluster_exemplars_id = [np.where((embedding ==_
      →cluster_exemplars_xy[point]).all(axis = 1))[0][0] for point in_
      →range(len(cluster_exemplars_xy))]
             ax.scatter(embedding[cluster_exemplars_id, 0],__
      →embedding[cluster_exemplars_id, 1],
               s = 22,
               c = "red")
             cluster_structures = traj[cluster_exemplars_id].
      atom_indices_
      cluster structures.save pdb("./Cluster Exemplars UMAP isolated domain/
      →UMAP_CD_Cluster_Exemplars_Cluster{}.pdb".format(cluster))
```

