# Metabolomic Data Analysis with MetaboAnalyst 6.0

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July 1, 2024

#### 1 Background

MSEA or Metabolite Set Enrichment Analysis is a way to identify biologically meaningful patterns that are significantly enriched in quantitative metabolomic data. In conventional approaches, metabolites are evaluated individually for their significance under conditions of study. Those compounds that have passed certain significance level are then combined to see if any meaningful patterns can be discerned. In contrast, MSEA directly investigates if a set of functionally related metabolites without the need to preselect compounds based on some arbitrary cut-off threshold. It has the potential to identify subtle but consistent changes among a group of related compounds, which may go undetected with the conventional approaches.

Essentially, MSEA is a metabolomic version of the popular GSEA (Gene Set Enrichment Analysis) software with its own collection of metabolite set libraries as well as an implementation of user-friendly web-interfaces. GSEA is widely used in genomics data analysis and has proven to be a powerful alternative to conventional approaches. For more information, please refer to the original paper by Subramanian A, and a nice review paper by Nam D, Kim SY. <sup>1</sup>. <sup>2</sup>

#### 2 MSEA Overview

Metabolite set enrichment analysis consists of four steps - data input, data processing, data analysis, and results download. Different analysis procedures are performed based on different input types. In addition, users can also browse and search the metabolite set libraries as well as upload their self-defined metabolite sets for enrichment analysis. Users can also perform metabolite name mapping between a variety of compound names, synonyms, and major database identifiers.

## 3 Data Input

There are three enrichment analysis algorithms offered by MSEA. Accordingly, three different types of data inputs are required by these three approaches:

- A list of important compound names entered as a one column data (Over Representation Analysis (ORA));
- A single measured biofluid (urine, blood, CSF) sample- entered as tab separated two-column data with the first column for compound name, and the second for concentration values (Single Sample Profiling (SSP));

<sup>&</sup>lt;sup>1</sup>Subramanian Gene set enrichment analysis: A knowledge-based approach for interpreting genome-wide expression profiles., Proc Natl Acad Sci USA. 2005 102(43): 15545-50

<sup>&</sup>lt;sup>2</sup>Nam D, Kim SY. Gene-set approach for expression pattern analysis, Briefings in Bioinformatics. 2008 9(3): 189-197.

• A compound concentration table - entered as a comma separated (.csv) file with the each sample per row and each metabolite concentration per column. The first column is sample names and the second column for sample phenotype labels (Quantitative Enrichment Analysis (QEA))

You selected Over Representation Analysis (ORA) which requires a list of compound names as input.

#### **Data Process**

The first step is to standardize the compound labels. It is an essential step since the compound labels will be subsequently compared with compounds contained in the metabolite set library. MSEA has a built-in tool to convert between compound common names, synonyms, identifiers used in HMDB ID, PubChem, ChEBI, BiGG, METLIN, KEGG, or Reactome. Table 1 shows the conversion results. Note: 1 indicates exact match, 2 indicates approximate match, and  $\theta$  indicates no match. A text file contain the result can be found the downloaded file name map.csv

Table 1. Resi

						Table 1: Resi
	Query	Match	HMDB	PubChem	KEGG	SMILES
1	Montiporic Acid C	NA	NA	NA	NΑ	NA
2	Montiporic Acid B	NA	NA	NA	NA	NA
3	Montiporic Acid D	NA	NA	NA	NA	NA
4	Montiporic Acid A	NA	NA	NA	NA	NA
5	Cresol	p-Cresol	${ m HMDB0001858}$	2879	C01468	CC1=CC=C(O)C=C1
6	Phenethylamine	Phenylethylamine	${ m HMDB0012275}$	1001	C05332	NCCC1=CC=CC=C1
7	Uridine	Uridine	${\rm HMDB0000296}$	6029	C00299	OC[C@H]10[C@H]([C@H
8	Glucose-6-phosphate	Glucose 6-phosphate	HMDB0001401	5958	C00092	OC1O[C@H](COP(O)(O
9	Guanosine	Guanosine	HMDB0000133	6802	C00387	NC1=NC2=C(N=CN2[C
10	Indole	Indole	HMDB0000738	798	C00463	N1C=CC2=C1C=CC=C
11	Carbamoyl phosphate	Carbamoyl phosphate	HMDB0001096	278	C00169	NC(=O)OP(O)(O)=O
12	NADPH	NADPH	HMDB0000221	5884	C00005	NC(=O)C1=CN(C=CC1
13	UMP	Uridine 5'-monophosphate	${ m HMDB0000288}$	6030	C00105	O[C@H]1[C@@H](O)[C@
14	ADP	ADP	HMDB0001341	6022	C00008	NC1=NC=NC2=C1N=C
15	O-Decanoyl-L-carnitine	Decanoylcarnitine	HMDB0000651	11953821	C03299	CCCCCCCCC(=0)O[C
16	Glycyl-L-proline	Glycylproline	${ m HMDB0000721}$	3013625		NCC(=O)N1CCC[C@H]
17	Adenosine	Adenosine	HMDB0000050	60961	C00212	NC1=C2N=CN([C@@H]
18	Mannose-6-phosphate	Mannose 6-phosphate	${ m HMDB0001078}$	439198	C00275	O[C@@H]1O[C@H](COP
19	Betaine	Betaine	HMDB0000043	247	C00719	C[N+](C)(C)CC(O)=O
20	Threonine	L-Threonine	HMDB0000167	6288	C00188	C[C@@H](O)[C@H](N)C
21	Serine	Serine	HMDB0000187	5951	C00065	N[C@@H](CO)C(O)=O
$^{22}$	Lysine-Glut amine	Lysylglutamine	HMDB0028949	196305		NCCCC[C@H](N)C(=O)
23	Ribothymidine	Ribothymidine	HMDB0000884	445408	~	CC1=CN([C@@H]2O[C@
$^{24}$	NG-dimethy l-L-arginine	Asymmetric dimethylarginine	HMDB0001539	123831	C03626	$N[C@@H](CCC\setminus N=C(/N))$
$^{25}$	Tryptamine	Tryptamine	HMDB0000303	1150	C00398	NCCC1=CNC2=C1C=C
26	Glucose	D-Glucose	HMDB0000122	5793	C00031	OC[C@H]10[C@@H](O)[
27	Cellobiose	Cellobiose	HMDB0000055	10712	C00185	OC[C@H]10[C@@H](O[C
28	Arginine-Alanine	Arginy lalanine	HMDB0028702	7020333		C[C@H](NC(=O)[C@@H
29	Arginine-Glutamine	Arginy lglutamine	HMDB0028707	7019985	Coorse	N[C@@H](CCCNC(N)=1)
30	Pterin	Pterin	HMDB0000802	73000	C00715	NC1=NC2=NC=CN=C2
$\frac{31}{32}$	Adipic acid Sorbitol	Adipic acid Sorbitol	HMDB0000448	196	C06104 C00794	OC(=0)CCCCC(0)=0
			HMDB0000247	5780		OC[C@H](O)[C@@H](O)
$\frac{33}{34}$	Isoleucine NAD	Isoleucine NAD	HMDB0000172 HMDB0000902	$6306 \\ 5892$	C00407 C00003	CC[C@H](C)[C@H](N)CO NC(=O)C1=C[N+](=CC)
$\frac{54}{35}$	N N N-Trimet hyllysine	NAD NA	NA	NA	NA	NA
36	Homoarginine	NA Homo-L-arginine	HMDB0000670	9085	NA C01924	
$\frac{30}{37}$	Tyrosine	L-Tyrosine	HMDB0000158	9085 6057	C01924 C00082	N[C@@H](CCCCNC(N)=N[C@@H](CC1=CC=C(G))
38	Diethanolamine	L-Tyrosine Diethanolamine	HMDB0000138	8113	C00082	OCCNCCO
39	ATP	Adenosine triphosphate	HMDB0004437	5957	C00002	NC1=NC=NC2=C1N=C
40	CDP-Choline	Citicoline	HMDB0000338	13805	C00002	C[N+](C)(C)CCOP(O)(=
41	Sedoheptulose	Sedoheptulose	HMDB0001413	441483	C00307	OC[C@H]10[C@](O)(CO
42	Lysine	Lysine	HMDB0003219	5962	C02070	NCCCC[C@H](N)C(O) =
43	Taurine	Taurine	HMDB0000182	1123	C00047	NCCS(O)(=O)=O
44	Sucrose	Sucrose	HMDB0000251	5988	C000243	OC[C@H]10[C@@](CO)(
45	Riboflav in	Riboflavin	HMDB0000238	493570	C00055	CC1=C(C)C=C2N(C[CC])
46	Acetyllysine	N-alpha-Acetyl-L-lysine	HMDB0000244	92907	C12989	CC(=O)N[C@@H](CCCO)
47	Raffinose	Raffinose	HMDB0003213	439242	C00492	OC[C@H]10[C@@](CO)(
48	Valine	L-Valine	HMDB0000213	6287	C00432	CC(C)[C@H](N)C(O)=C
49	Phenylalanine	Phenylalanine	HMDB0000159	6140	C00133	N[C@@H](CC1=CC=CC
50	CDP-ethanolamine	CDP-ethanolamine	HMDB00001564	123727	C00570	NCCOP(O)(=0)OP(O)(
51	N-Acetyl-Glucosamine	N-Acetyl-D-Glucosamine 6-Phosphate	HMDB0001062	440996	C00317	CC(=O)N[C@H]1C(O)O
52	ADP-ribose	Adenosine diphosphate ribose	HMDB0001178	192	C00301	NC1=C2N=CN(C3OC(C
53	Glutamine	Glutamine	HMDB0001110	5961	C00064	N[C@@H](CCC(N)=O)C
54	Pyruvate	Pyruvic acid	HMDB0000243	1060	C00022	CC(=O)C(O)=O
55	FAD	FAD	HMDB0000218	643975	C00016	CC1=CC2=C(C=C1C)N
56	Trigonelline	Trigonelline	HMDB0001248	5570	C01004	C[N+]1=CC=CC(=C1)C
57	3-Phenylbutyric acid	3-Phenylbutyric acid	HMDB0000375	20724	201001	CC(CC(O)=O)C1=CC=
58	Acetyl glycine	Phenylacetylglycine	HMDB0001333	68144	C05598	OC(=O)CNC(=O)CC1=
		,, <b>-8-</b> J - <b>8-</b> J - <b></b>			233330	, 0,01.0( 0)001-

59	CMP	Cytidine monophosphate	${\rm HMDB0000095}$	6131	C00055	NC1=NC(=O)N(C=C1)
60	O-Phosphorylethanolamine	O-Phosphoet hanolamine	${ m HMDB0000224}$	1015	C00346	NCCOP(O)(O) = O
61	GDP-Mannose	Guanosine diphosphate mannose	HMDB0001163	18396	C00096	NC1=NC2=C(N=CN2[C
62	Malonic acid	Malonic acid	HMDB0000691	867	C00383	OC(=O)CC(O)=O
63	4-Guanidinobutanoic acid	4-Guanidinobutanoic acid	HMDB0003464	500	C01035	NC(=N)NCCCC(O)=O
64	GDP	Guanosine diphosphate	HMDB0001201	8977	C00035	NC1=NC2=C(N=CN2[C
65	Fructose	D-Fructose	${ m HMDB0000660}$	439709	C00095	OC[C@H]10[C@](O)(CO
66	Glutamate	Glutamic acid	HMDB0000148	33032	C00025	N[C@@H](CCC(O)=O)C
67	L-Octanoylcarnitine	Octanoylcarnitine	${ m HMDB0000791}$	11953814	C02838	CCCCCCC(=0)O[C@H]
68	Nicotinamide riboside	Nicotinamide riboside	${ m HMDB0000855}$	439924	C03150	NC(=O)C1=C[N+](=CC
69	1-Met hylhist idin e	1-Methylhistidine	HMDB0000001	92105	C01152	CN1C=NC(C[C@H](N)C
70	Proline	Proline	${ m HMDB0000162}$	145742	C00148	OC(=O)[C@@H]1CCCN;
71	Glutaric acid	Glutaric acid	HMDB0000661	743	C00489	OC(=O)CCCC(O)=O
72	5-Hydroxytryptophan	5-Hydroxy-L-tryptophan	${ m HMDB0000472}$	439280	C00643	N[C@@H](CC1=CNC2=0)
73	UDP-N-acetyl-glucosamine	Uridine diphosphate-N-acetylglucosamine	${ m HMDB0000290}$	445675	C00043	CC(=O)N[C@@H]1[C@@
74	UDP-D-Glucose	Uridine diphosphate glucose	${ m HMDB0000286}$	8629	C00029	OC[C@H]10[C@H](OP(
75	Pantothenate	Pantothenic acid	${ m HMDB0000210}$	6613	C00864	CC(C)(CO)[C@@H](O)C
$^{76}$	NADP	NADP	${ m HMDB0000217}$	5885	C00006	NC(=O)C1=C[N+](=CC
77	Homocitrulline	Homocitrulline	${ m HMDB0000679}$	65072	C02427	N[C@@H](CCCCNC(N)=
78	Acetyl CoA	Acetoacetyl-CoA	HMDB0001484	92153	C00332	CC(=O)CC(=O)SCCNC
79	Glutathione	Glutathione	${ m HMDB0000125}$	124886	C00051	N[C@@H](CCC(=O)N[C]
80	S-Adenosyl-homocysteine	S-Adenosylhomocysteine	${ m HMDB0000939}$	439155	C00021	N[C@@H](CCSC[C@H]1C
81	Pirbuterol	Pirbuterol	${ m HMDB0015407}$	4845	C07807	CC(C)(C)NCC(O)C1=N
82	Orotate	Orotic acid	${ m HMDB0000226}$	967	C00295	OC(=O)C1=CC(=O)NC

The second step is to check concentration values. For SSP analysis, the concentration must be measured in umol for blood and CSF samples. The urinary concentrations must be first converted to  $umol/mmol\_creatinine$  in order to compare with reported concentrations in literature. No missing or negative values are allowed in SSP analysis. The concentration data for QEA analysis is more flexible. Users can upload either the original concentration data or normalized data. Missing or negative values are allowed (coded as NA) for QEA.

## 5 Selection of Metabolite Set Library

Before proceeding to enrichment analysis, a metabolite set library has to be chosen. There are seven built-in libraries offered by MSEA:

- Metabolic pathway associated metabolite sets (currently contains 99 entries);
- Disease associated metabolite sets (reported in blood) (currently contains 344 entries);
- Disease associated metabolite sets (reported in urine) (currently contains 384 entries)
- Disease associated metabolite sets (reported in CSF) (currently contains 166 entries)
- Metabolite sets associated with SNPs (currently contains 4598 entries)
- Predicted metabolite sets based on computational enzyme knockout model (currently contains 912 entries)
- Metabolite sets based on locations (currently contains 73 entries)
- Drug pathway associated metabolite sets (currently contains 461 entries)

In addition, MSEA also allows user-defined metabolite sets to be uploaded to perform enrichment analysis on arbitrary groups of compounds which researchers want to test. The metabolite set library is simply a two-column comma separated text file with the first column for metabolite set names and the second column for its compound names (**must use HMDB compound name**) separated by "; ". Please note, the built-in libraries are mainly from human studies. The functional grouping of metabolites may not be valid. Therefore, for data from subjects other than human being, users are suggested to upload their self-defined metabolite set libraries for enrichment analysis.

## 6 Enrichment Analysis

Over Representation Analysis (ORA) is performed when a list of compound names is provided. The list of compound list can be obtained through conventional feature selection methods, or from a clustering algorithm, or from the compounds with abnormal concentrations detected in SSP, to investigate if some biologically meaningful patterns can be identified.

ORA was implemented using the *hypergeometric test* to evaluate whether a particular metabolite set is represented more than expected by chance within the given compound list. One-tailed p values are provided after adjusting for multiple testing. **Figure 2** below summarizes the result.

#### **Enrichment Overview (top 25)**

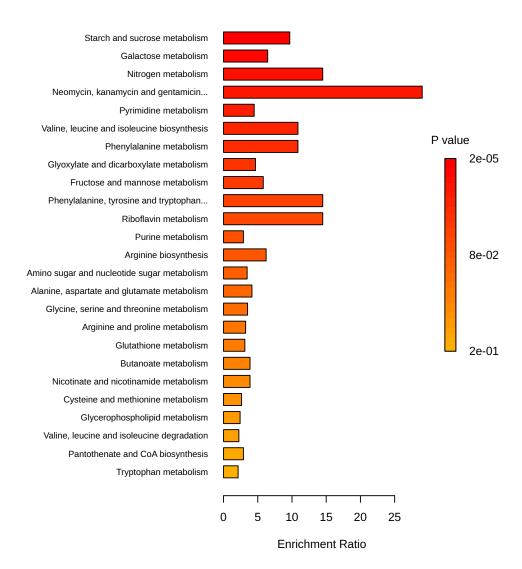


Figure 1: Summary Plot for Over Representation Analysis (ORA)

Table 2: Result from Over Representation Analysis

		-		-		
	total	expected	hits	Raw p	Holm p	FDR
Starch and sucrose metabolism	18	0.62	6	1.69E-05	1.36E-03	1.36E-03
Galactose metabolism	27	0.93	6	2.13E-04	1.68E-02	8.50E-03
Nitrogen metabolism	6	0.21	3	7.18E-04	5.60E-02	1.91E-02
Neomycin, kanamycin and gentamicin	2	0.07	2	1.16E-03	8.97E-02	2.19E-02
biosynthesis						
Pyrimidine metabolism	39	1.34	6	1.70E-03	1.29E-01	2.19E-02
Valine, leucine and isoleucine biosynthe-	8	0.28	3	1.91E-03	1.43E-01	2.19E-02
sis						
Phenylalanine metabolism	8	0.28	3	1.91E-03	1.43E-01	2.19E-02
Glyoxylate and dicarboxylate	31	1.07	5	3.44E-03	$2.51\mathrm{E}\text{-}01$	3.44E-02
metabolism						
Fructose and mannose metabolism	20	0.69	4	4.04E-03	2.91E-01	3.59E-02
Phenylalanine, tyrosine and tryptophan	4	0.14	2	6.68E-03	4.74E-01	4.86E-02
biosynthesis						
Riboflavin metabolism	4	0.14	2	6.68E-03	4.74E-01	4.86E-02
Purine metabolism	70	2.41	7	8.59E-03	5.93E-01	5.73E-02
Arginine biosynthesis	14	0.48	3	1.07E-02	7.30E-01	6.60E-02
Amino sugar and nucleotide sugar	42	1.45	5	1.29E-02	8.65E-01	7.38E-02
metabolism						
Alanine, aspartate and glutamate	28	0.96	4	1.39E-02	9.18E-01	7.41E-02
metabolism						
Glycine, serine and threonine metabolism	33	1.14	4	2.45E-02	1.00E + 00	1.22E-01
Arginine and proline metabolism	36	1.24	4	3.26E-02	1.00E + 00	1.54E-01
Glutathione metabolism	28	0.96	3	6.89E-02	1.00E+00	3.06E-01
Butanoate metabolism	15	0.52	2	9.18E-02	1.00E + 00	3.67E-01
Nicotinate and nicotinamide metabolism	15	0.52	2	9.18E-02	1.00E + 00	3.67E-01
Cysteine and methionine metabolism	33	1.14	3	1.02E-01	1.00E + 00	3.88E-01
Glycerophospholipid metabolism	36	1.24	3	1.24E-01	1.00E + 00	4.51E-01
Valine, leucine and isoleucine degrada-	39	1.34	3	1.48E-01	1.00E + 00	4.97 E-01
tion						
Pantothenate and CoA biosynthesis	20	0.69	2	1.49E-01	1.00E+00	4.97E-01
Tryptophan metabolism	41	1.41	3	1.65E-01	1.00E + 00	5.27E-01
Pyruvate metabolism	23	0.79	2	1.86E-01	1.00E+00	5.73E-01
Taurine and hypotaurine metabolism	8	0.28	1	2.45E-01	1.00E+00	7.26E-01
Ascorbate and aldarate metabolism	9	0.31	1	2.71E-01	1.00E + 00	7.63E-01
Lysine degradation	30	1.03	2	2.77E-01	1.00E+00	7.63E-01
Biotin metabolism	10	0.34	1	2.96E-01	1.00E+00	7.81E-01
Sphingolipid metabolism	32	1.10	2	3.03E-01	1.00E+00	7.81E-01
D-Amino acid metabolism	15	0.52	1	4.10E-01	1.00E+00	$1.00 \mathrm{E} \! + \! 00$
Tyrosine metabolism	42	1.45	2	4.29E-01	1.00E+00	$1.00 \mathrm{E} \! + \! 00$
Histidine metabolism	16	0.55	1	4.31E-01	1.00E+00	$1.00 \mathrm{E} \! + \! 00$
Ubiquinone and other terpenoid-quinone	18	0.62	1	4.70E-01	1.00E+00	$1.00 \mathrm{E} \! + \! 00$
biosynthesis						
Terpenoid backbone biosynthesis	18	0.62	1	4.70E-01	1.00E+00	$1.00 \mathrm{E} \! + \! 00$
Pentose and glucuronate interconversions	19	0.65	1	4.88E-01	1.00E+00	1.00E + 00
Citrate cycle (TCA cycle)	20	0.69	1	5.06E-01	1.00E + 00	$1.00 \mathrm{E} \! + \! 00$
Pentose phosphate pathway	23	0.79	1	5.56E-01	1.00E+00	1.00E + 00
Glycolysis / Gluconeogenesis	26	0.90	1	6.01E-01	1.00E + 00	$1.00\mathrm{E}\!+\!00$
Lipoic acid metabolism	28	0.96	1	6.28E-01	1.00E+00	1.00E + 00
Inositol phosphate metabolism	30	1.03	1	6.54E-01	1.00E+00	$1.00 \mathrm{E} \! + \! 00$
Porphyrin metabolism	31	1.07	1	6.66E-01	1.00E+00	$1.00 \mathrm{E} \! + \! 00$
Fatty acid degradation	39	1.34	1	7.49E-01	1.00E+00	$1.00 \mathrm{E} \! + \! 00$
N-Glycan biosynthesis	40	1.38	1	7.58E-01	1.00E+00	$1.00 \mathrm{E} \! + \! 00$
Primary bile acid biosynthesis	46	1.58	1	8.05E-01	1.00E+00	$1.00 \mathrm{E} \! + \! 00$
Fatty acid biosynthesis	47	1.62	1	8.12E-01	1.00E+00	1.00E + 00
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### 7 Appendix: R Command History

```
[1] "mSet<-InitDataObjects(\"conc\", \"msetora\", FALSE)"
 [2] "cmpd.vec<-c(\"Montiporic Acid C\",\"Montiporic Acid B\",\"Montiporic Acid D\",\"Montiporic Acid D\",\"
 [3] "mSet<-Setup.MapData(mSet, cmpd.vec);"
 [4] "mSet<-CrossReferencing(mSet, \"name\");"
 [5] "mSet<-CreateMappingResultTable(mSet)"
 [6] "mSet<-PerformDetailMatch(mSet, \"Cresol\");"</pre>
 [7] "mSet<-GetCandidateList(mSet);"
 [8] "mSet<-SetCandidate(mSet, \"Cresol\", \"p-Cresol\");"
 [9] "mSet<-PerformDetailMatch(mSet, \"O-Decanoyl-L-carnitine\");"
[10] "mSet<-GetCandidateList(mSet);"
[11] "mSet<-SetCandidate(mSet, \"0-Decanoyl-L-carnitine\", \"Decanoylcarnitine\");"
[12] "mSet<-PerformDetailMatch(mSet, \"Lysine-Glutamine\");"
[13] "mSet<-GetCandidateList(mSet);"
[14] "mSet<-SetCandidate(mSet, \"Lysine-Glutamine\", \"Lysylglutamine\");"
[15] "mSet<-PerformDetailMatch(mSet, \"NG-dimethyl-L-arginine\");"
[16] "mSet<-GetCandidateList(mSet);"</pre>
[17] "mSet<-SetCandidate(mSet, \"NG-dimethyl-L-arginine\", \"Asymmetric dimethylarginine\");"
[18] "mSet<-PerformDetailMatch(mSet, \"Arginine-Alanine\");"
[19] "mSet<-GetCandidateList(mSet);"</pre>
[20] "mSet<-SetCandidate(mSet, \"Arginine-Alanine\", \"Arginylalanine\");"
[21] "mSet<-PerformDetailMatch(mSet, \"Arginine-Glutamine\");"
[22] "mSet<-GetCandidateList(mSet);"</pre>
[23] "mSet<-SetCandidate(mSet, \"Arginine-Glutamine\", \"Arginylglutamine\");"
[24] "mSet<-PerformDetailMatch(mSet, \"N N N-Trimethyllysine\");"
[25] "mSet<-GetCandidateList(mSet);"</pre>
[26] "mSet<-PerformDetailMatch(mSet, \"N-Acetyl-Glucosamine\");"
[27] "mSet<-GetCandidateList(mSet);"</pre>
[28] "mSet<-SetCandidate(mSet, \"N-Acetyl-Glucosamine\", \"N-Acetyl-D-Glucosamine 6-Phosphate\");"
[29] "mSet<-PerformDetailMatch(mSet, \"Acetyl glycine\");"
[30] "mSet<-GetCandidateList(mSet);"
[31] "mSet<-SetCandidate(mSet, \"Acetyl glycine\", \"Phenylacetylglycine\");"
[32] "mSet<-PerformDetailMatch(mSet, \"Acetyl CoA\");"
[33] "mSet<-GetCandidateList(mSet);"
[34] "mSet<-SetCandidate(mSet, \"Acetyl CoA\", \"Acetoacetyl-CoA\");"
[35] "mSet<-SetMetabolomeFilter(mSet, F);"</pre>
[36] "mSet<-SetCurrentMsetLib(mSet, \"kegg_pathway\", 2);"
[37] "mSet<-CalculateHyperScore(mSet)"
[38] "mSet<-PlotORA(mSet, \"ora_0_\", \"net\", \"png\", 72, width=NA)"
[39] "mSet<-PlotEnrichDotPlot(mSet, \"ora\", \"ora_dot_0_\", \"png\", 72, width=NA)"
[40] "mSet<-CalculateHyperScore(mSet)"
[41] "mSet<-PlotORA(mSet, \"ora_1_\", \"net\", \"png\", 72, width=NA)"
[42] "mSet<-PlotEnrichDotPlot(mSet, \"ora\", \"ora_dot_1_\", \"png\", 72, width=NA)"
[43] "mSet<-CalculateHyperScore(mSet)"
[44] "mSet<-Plot0RA(mSet, \"ora_2\", \"net\", \"png\", 72, width=NA)"
[45] "mSet<-PlotEnrichDotPlot(mSet, \"ora\", \"ora_dot_2_\", \"png\", 72, width=NA)"
[46] "mSet<-CalculateHyperScore(mSet)"
[47] "mSet<-PlotORA(mSet, \"ora_3_\", \"net\", \"png\", 72, width=NA)"
[48] "mSet<-PlotEnrichDotPlot(mSet, \"ora\", \"ora_dot_3_\", \"png\", 72, width=NA)"
[49] "mSet<-CalculateHyperScore(mSet)"
[50] "mSet<-PlotORA(mSet, \"ora_4_\", \"net\", \"png\", 72, width=NA)"
[51] "mSet<-PlotEnrichDotPlot(mSet, \"ora\", \"ora_dot_4_\", \"png\", 72, width=NA)"
[52] "mSet<-SaveTransformedData(mSet)"
[53] "mSet<-PreparePDFReport(mSet, \"guest18173587466078868361\")\n"
```

The report was generated on Mon Jul 1  $15:57:49\ 2024$  with R version  $4.3.2\ (2023-10-31)$ , OS system: Linux, version: -Ubuntu SMP Tue Mar 5  $20:16:58\ UTC\ 2024$ .