

MIRACL WORKSHOP

MAGED GOUBRAN

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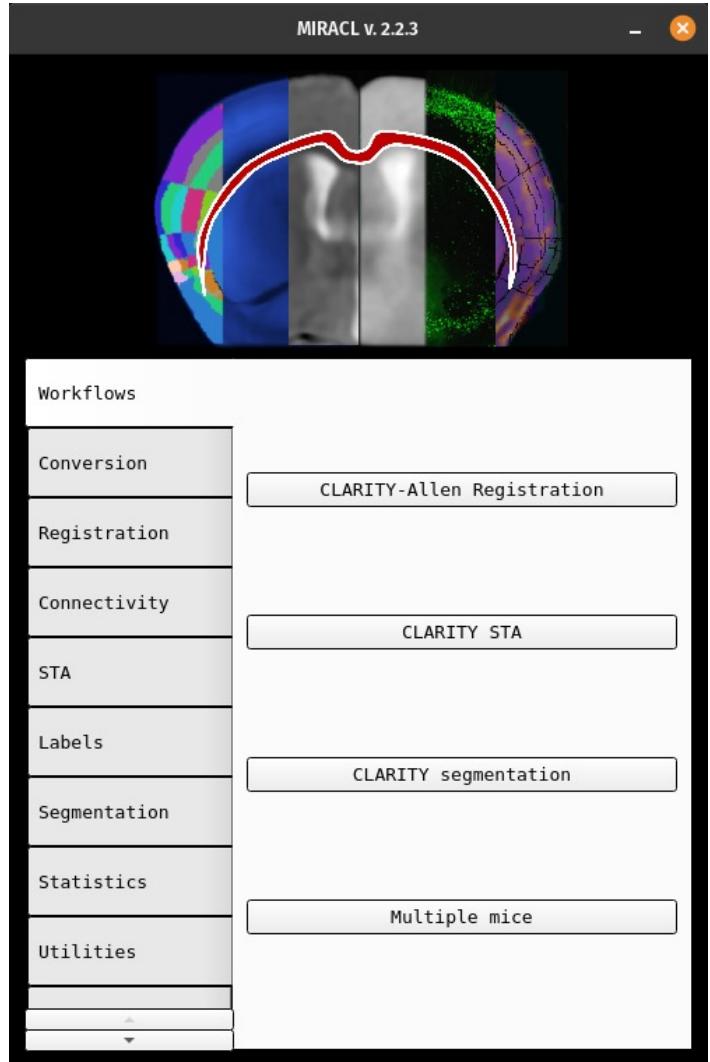


UNIVERSITY OF
TORONTO

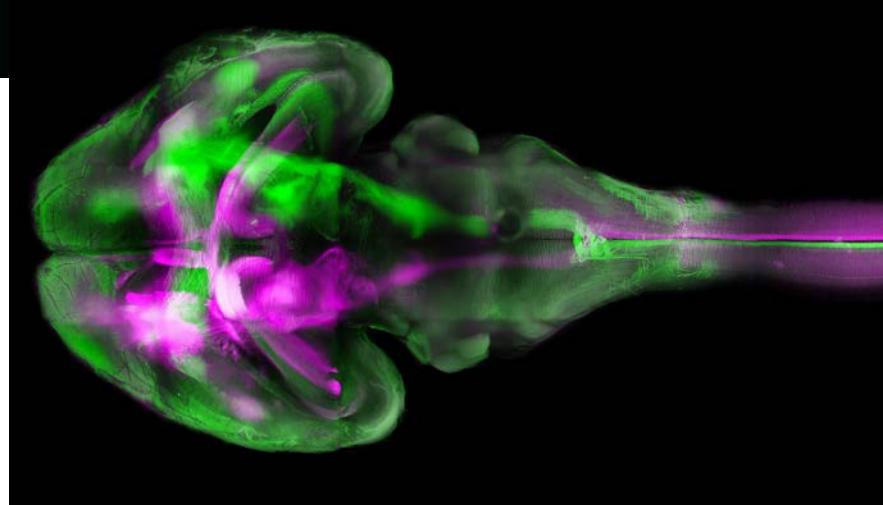
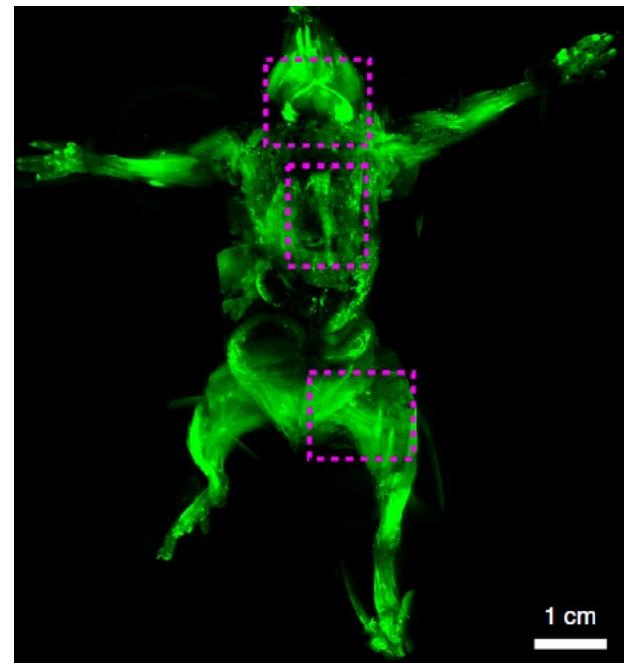
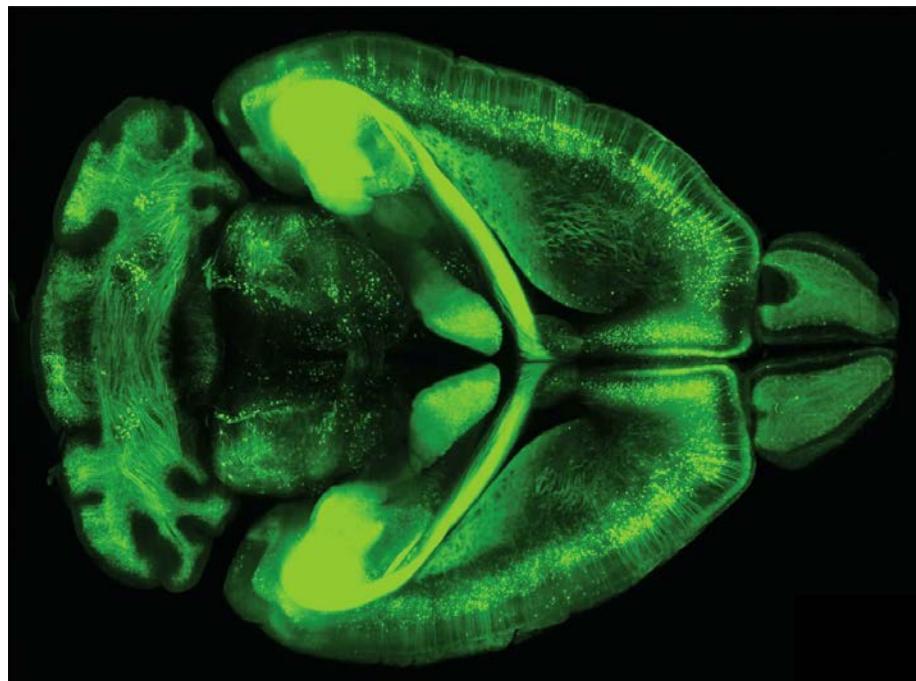


WORKSHOP OVERVIEW

- Background & motivation behind MIRACL
- Overview of different functionality
- Example work using MIRACL
- ACE: New AI model & sub-regional analyses
- MRI analyses & diffusion comparison
- Future work
- Google collab notebooks
 - Registration
 - ACE
 - Regional & sub-regional stats
 - Tract tracing



TISSUE CLEARING



Park et al. *Nature Biotechnology* 2019

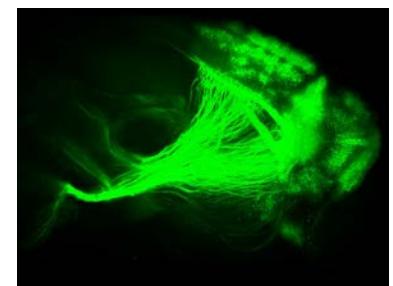
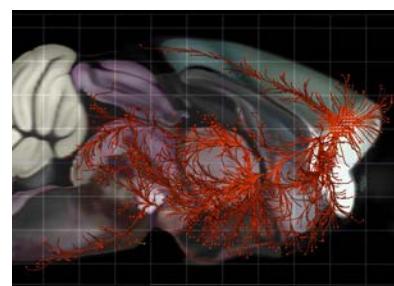
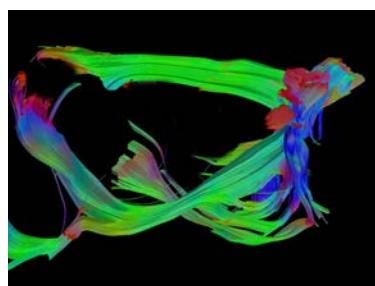
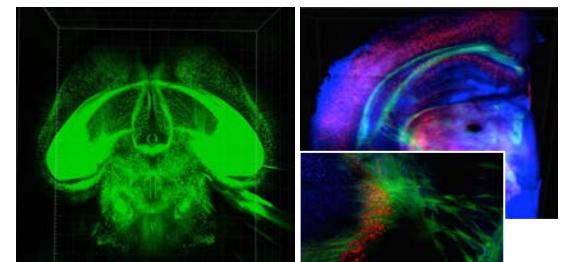
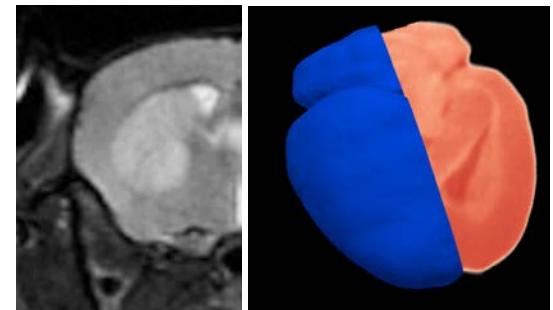
Pan et al. *Nature Methods* 2016

Chung et al. *Nature* 2015



MOTIVATION / CHALLENGES

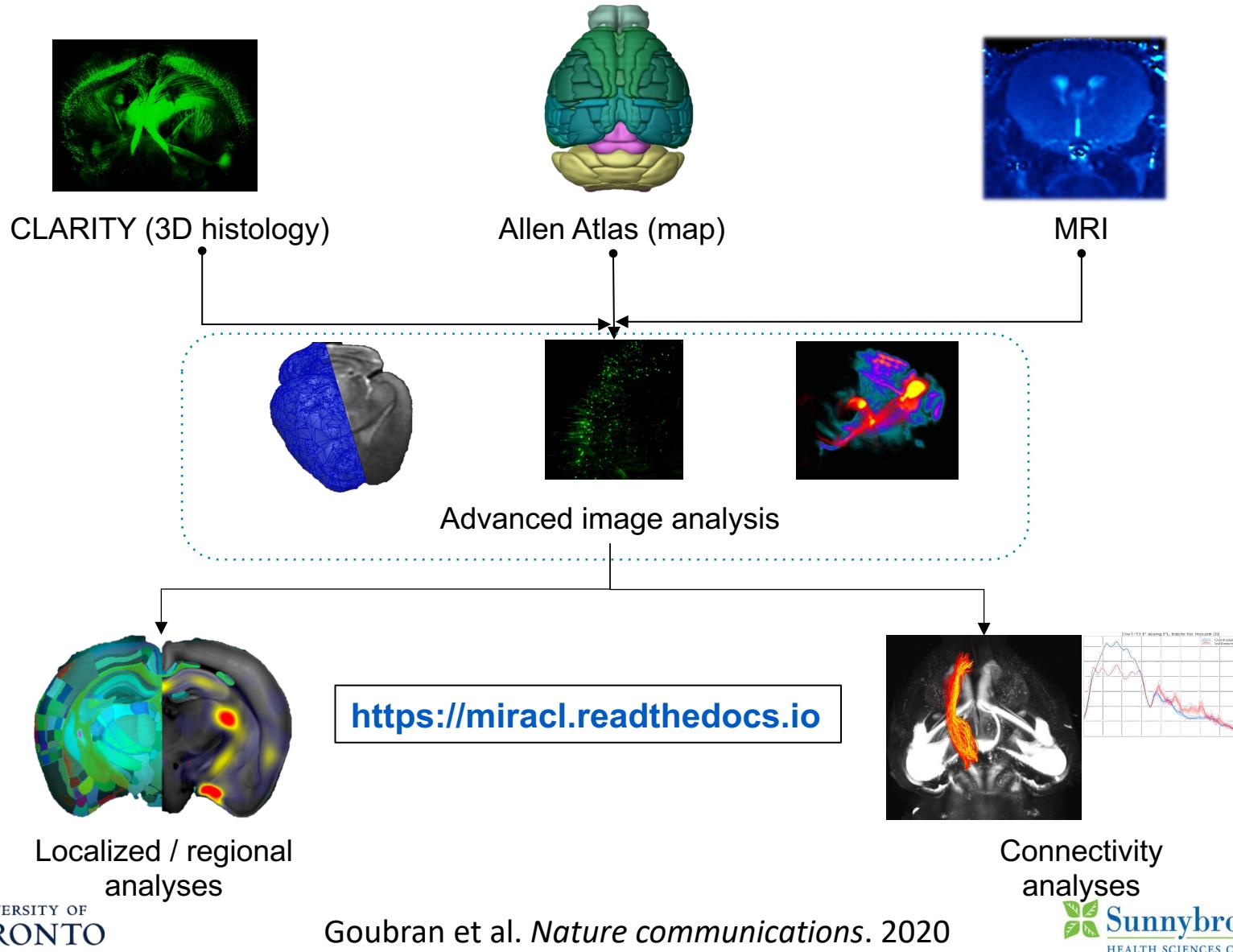
- Accurate mapping between mesoscopic & microscopic domains
- Accurate cellular & fiber quantification
- Automated network & tract-level connectivity analysis
- Automated analysis of cellular alterations across the whole-brain



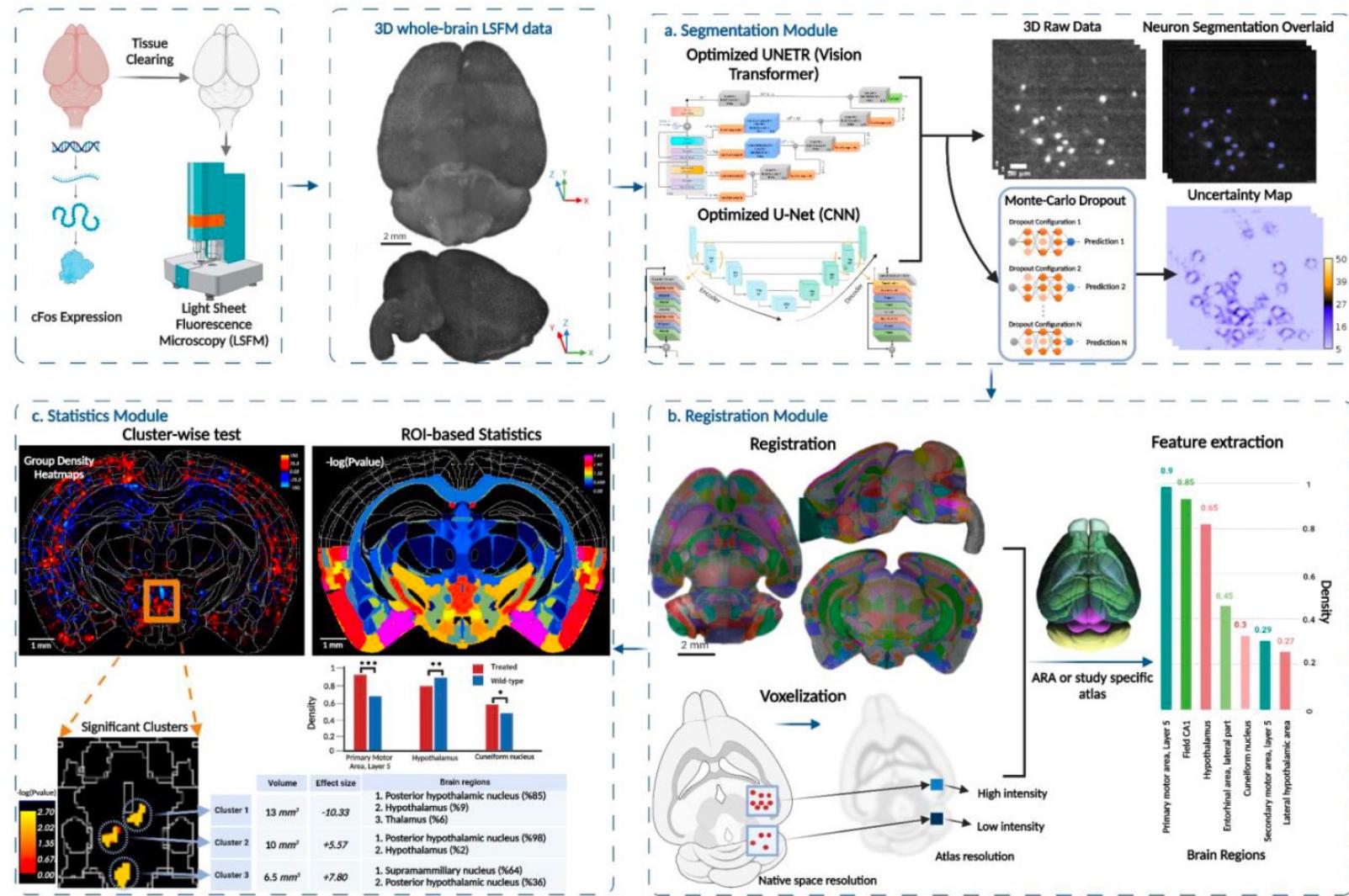
PIPELINE OVERVIEW

with Jennifer McNab, Karl Deisseroth & Michael Zeineh

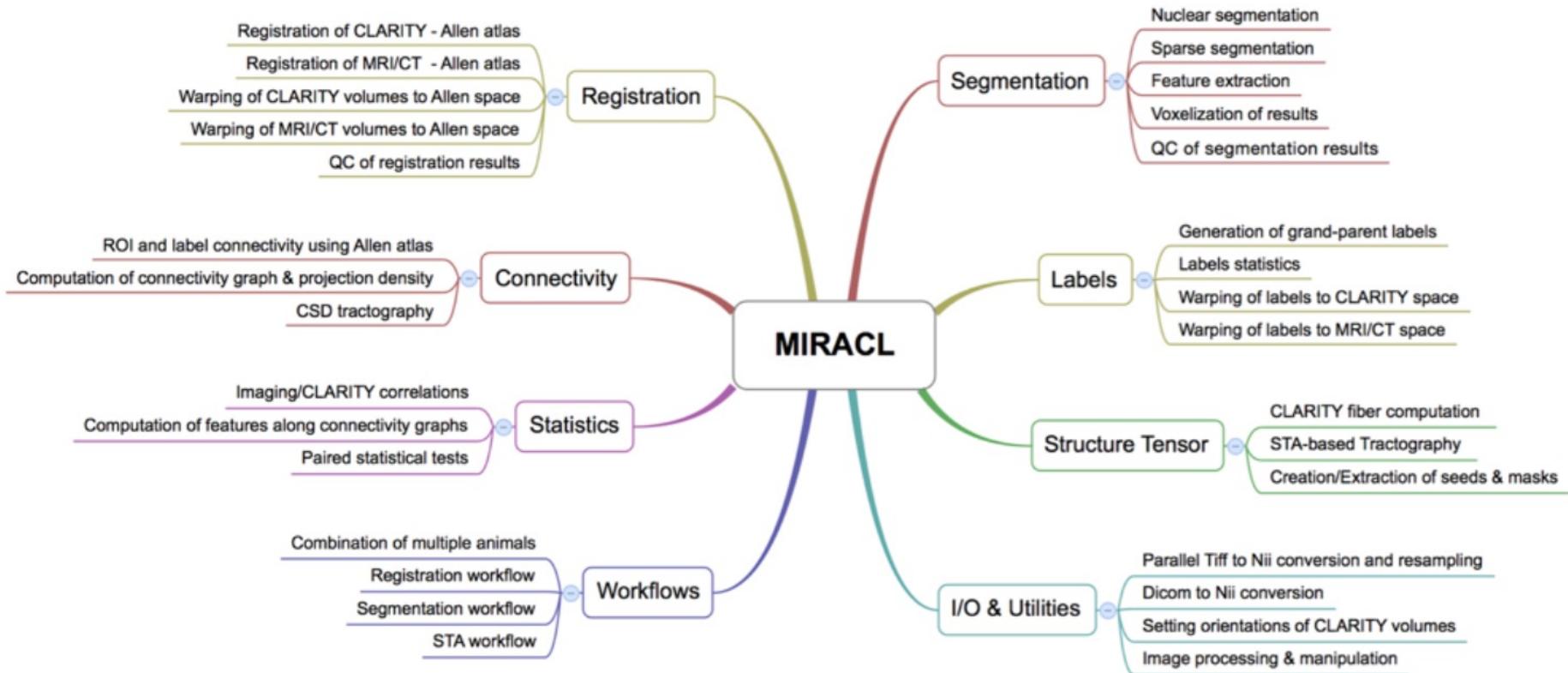
Multi-modal Image Registration And Connectivity anaLysis



ACE: AI-BASED CARTOGRAPHY OF (NEURONAL) ENSEMBLES

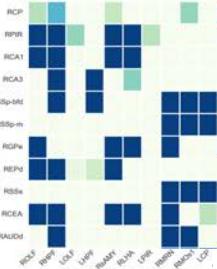


OVERVIEW OF FUNCTIONALITY

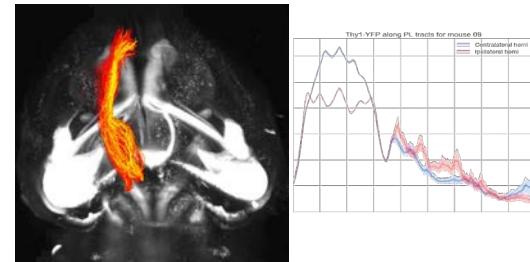


EXAMPLES OF FUNCTIONALITY / OUTPUTS

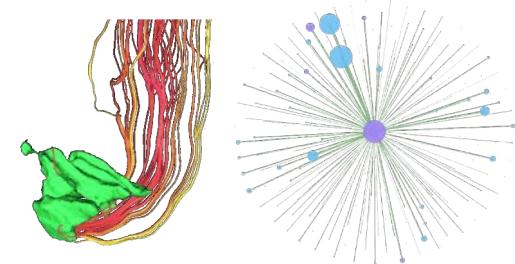
Tract & network-based analyses



Features across network

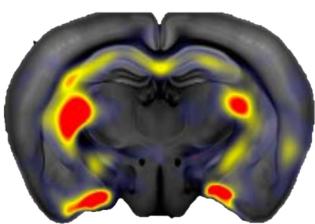


Tract-level histological analyses

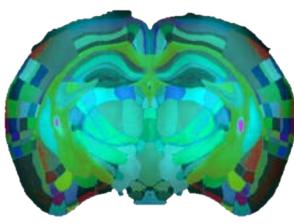


Terminals-based connectivity analyses

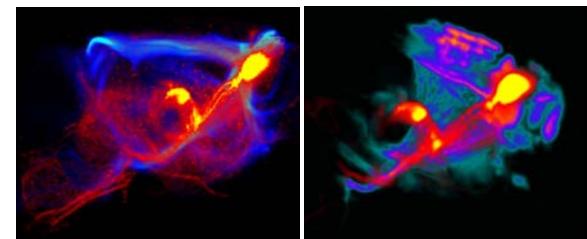
Label & voxel-based analyses



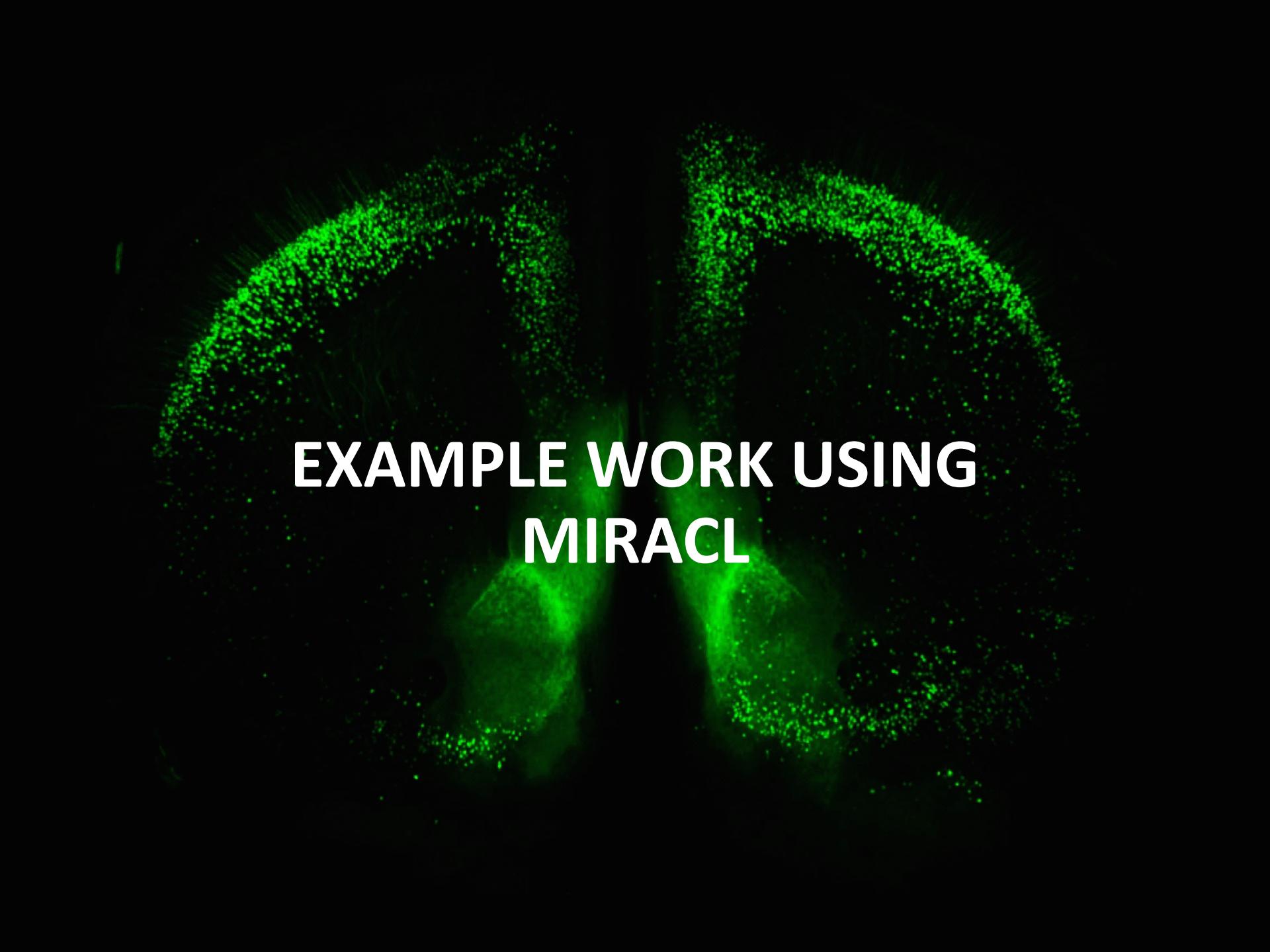
Multimodal group-level statistics



Correlations between modalities

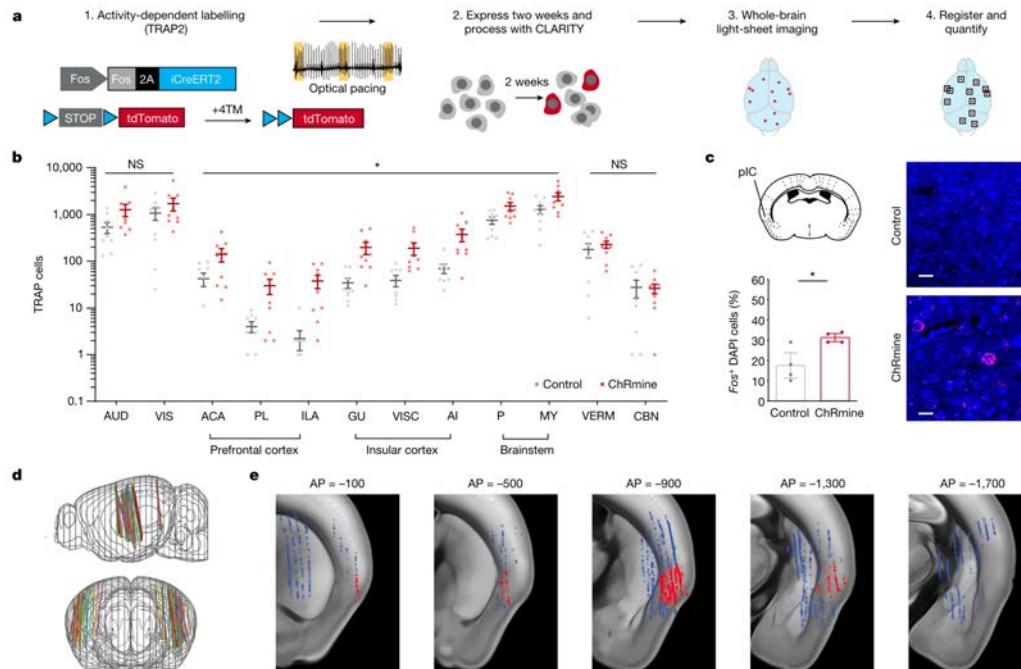
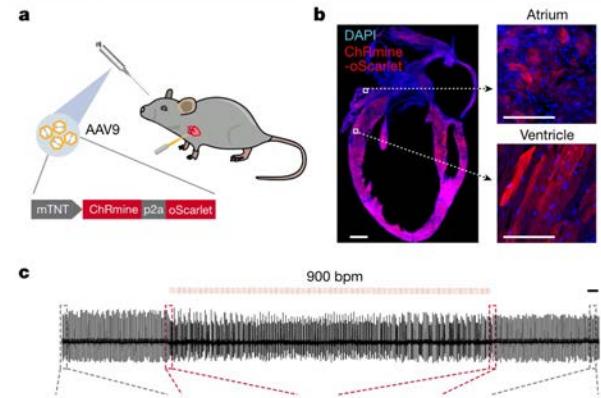
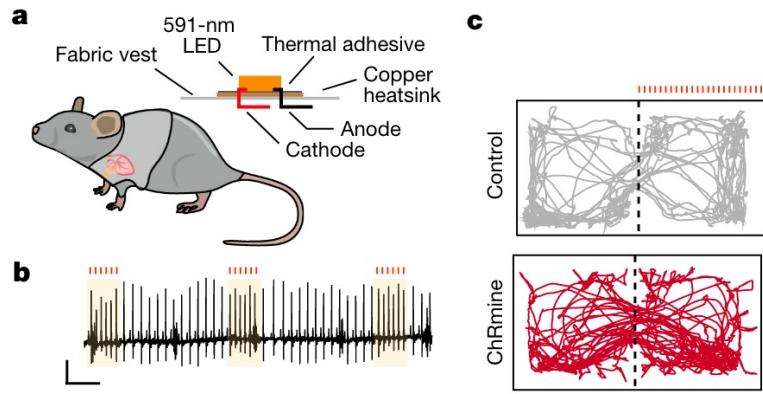


Connectivity maps comparison across scales



EXAMPLE WORK USING MIRACL

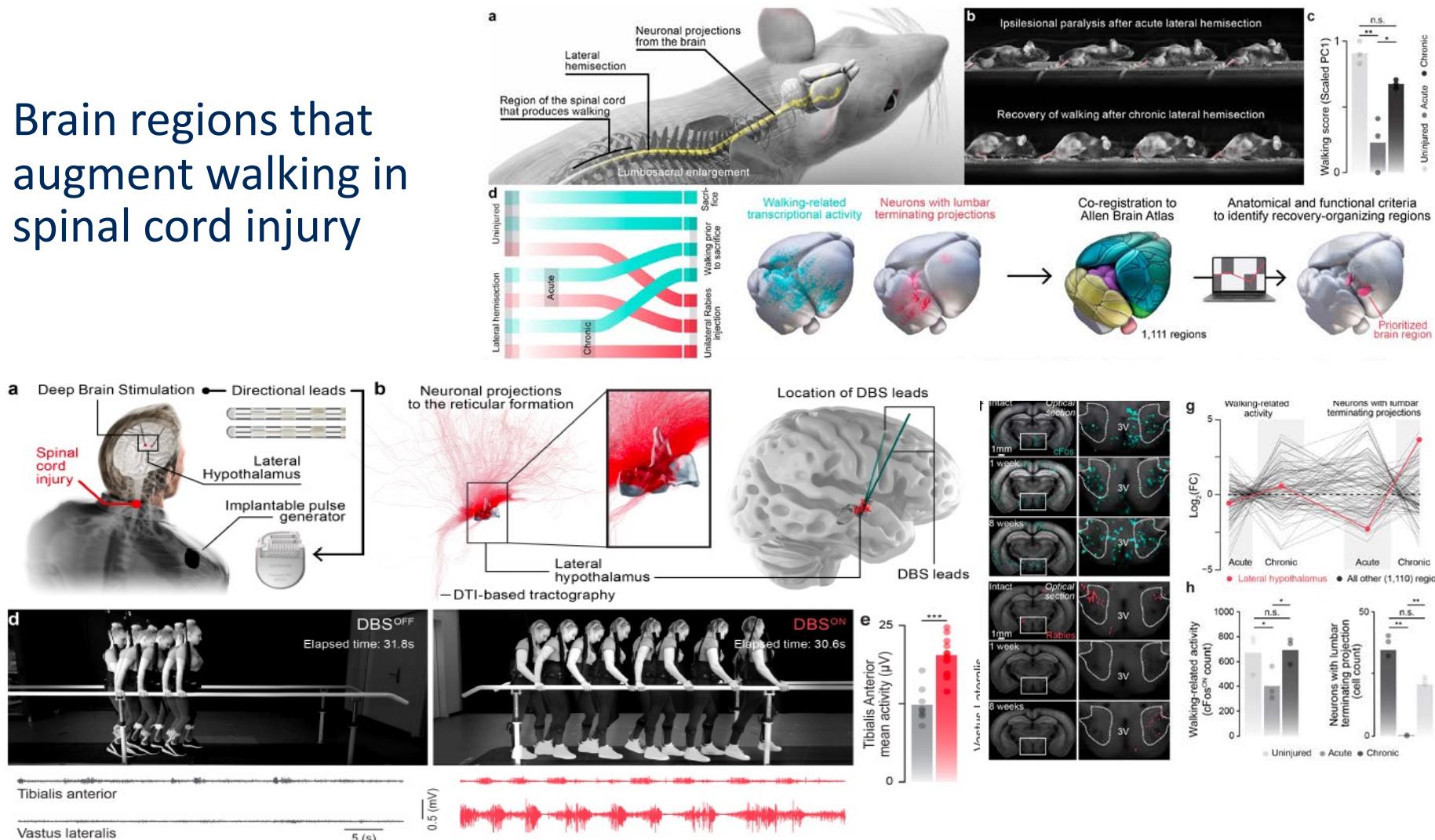
BRAIN REGIONS ORCHESTRATING ANXIETY



Hsueh et al. *Nature* 2023

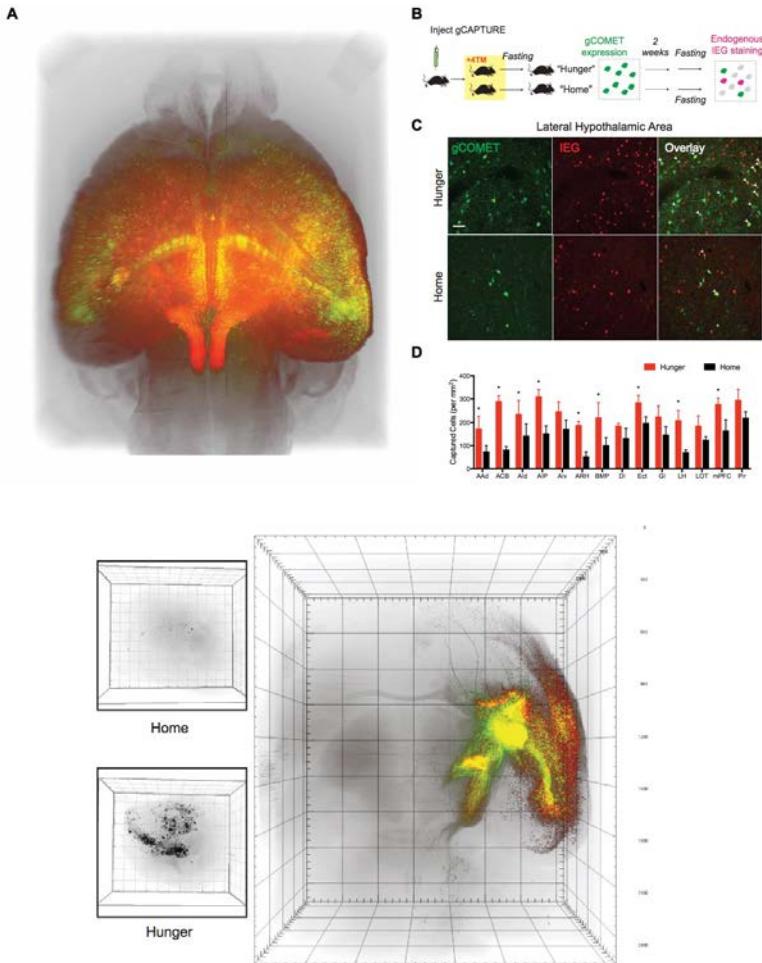
RECOVERING WALKING AFTER SPINAL CORD INJURY

Brain regions that augment walking in spinal cord injury

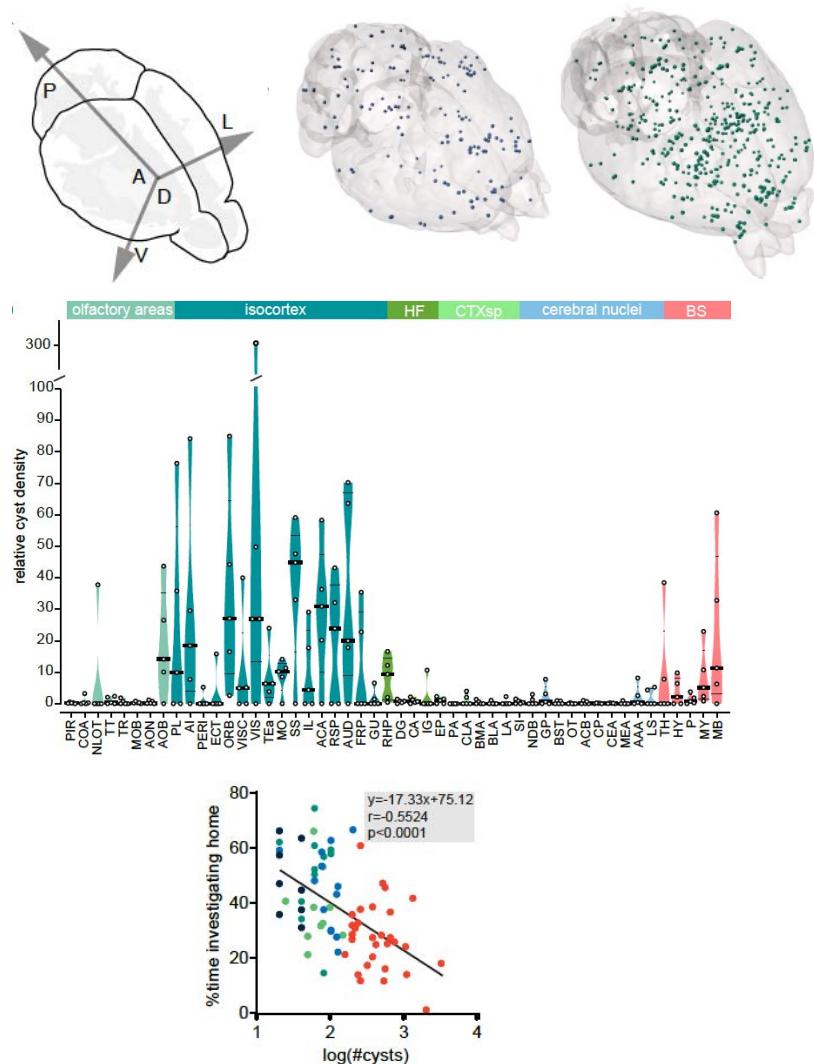


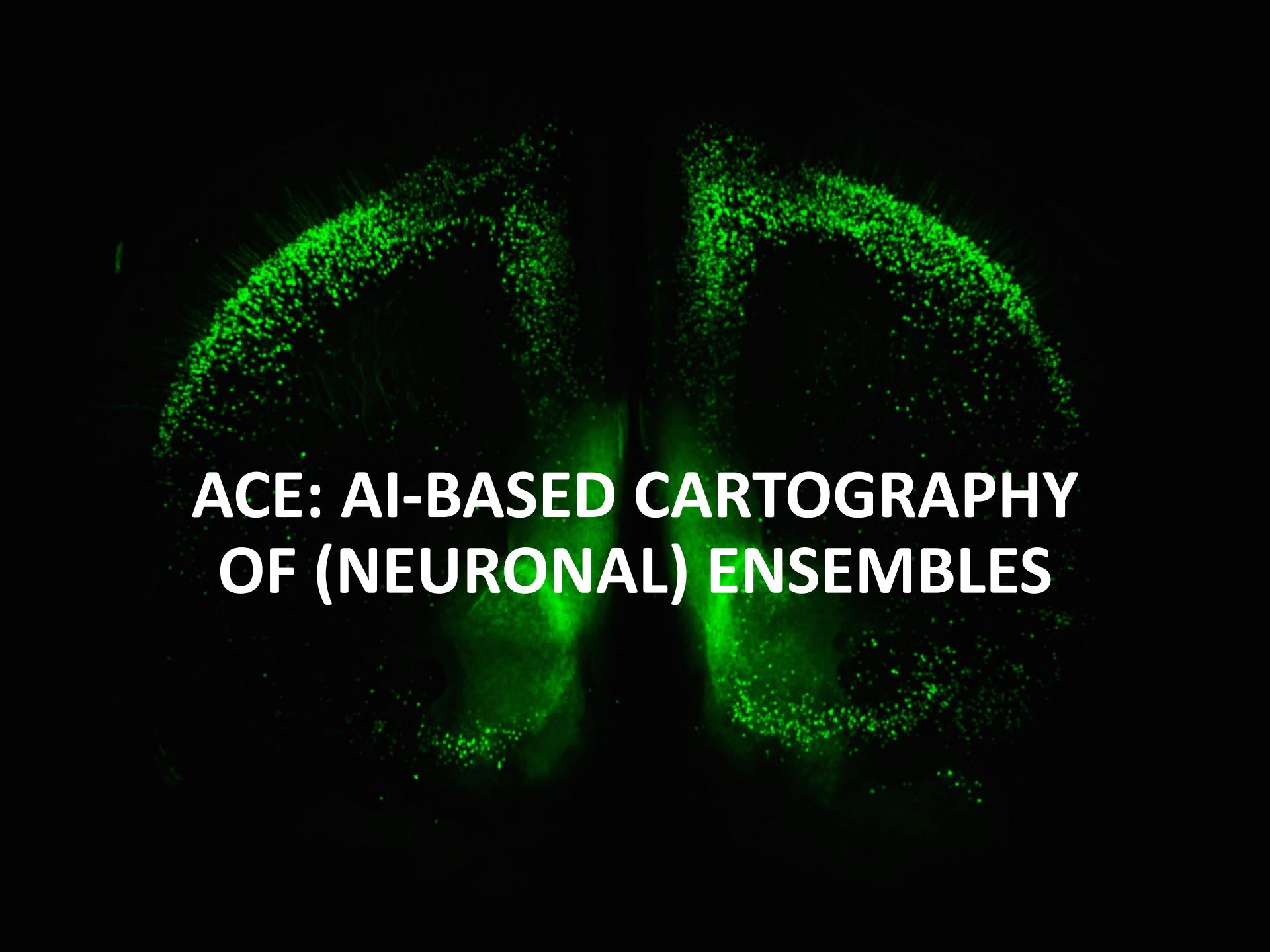
OTHER EXAMPLE STUDIES

Projection mapping / Tractography



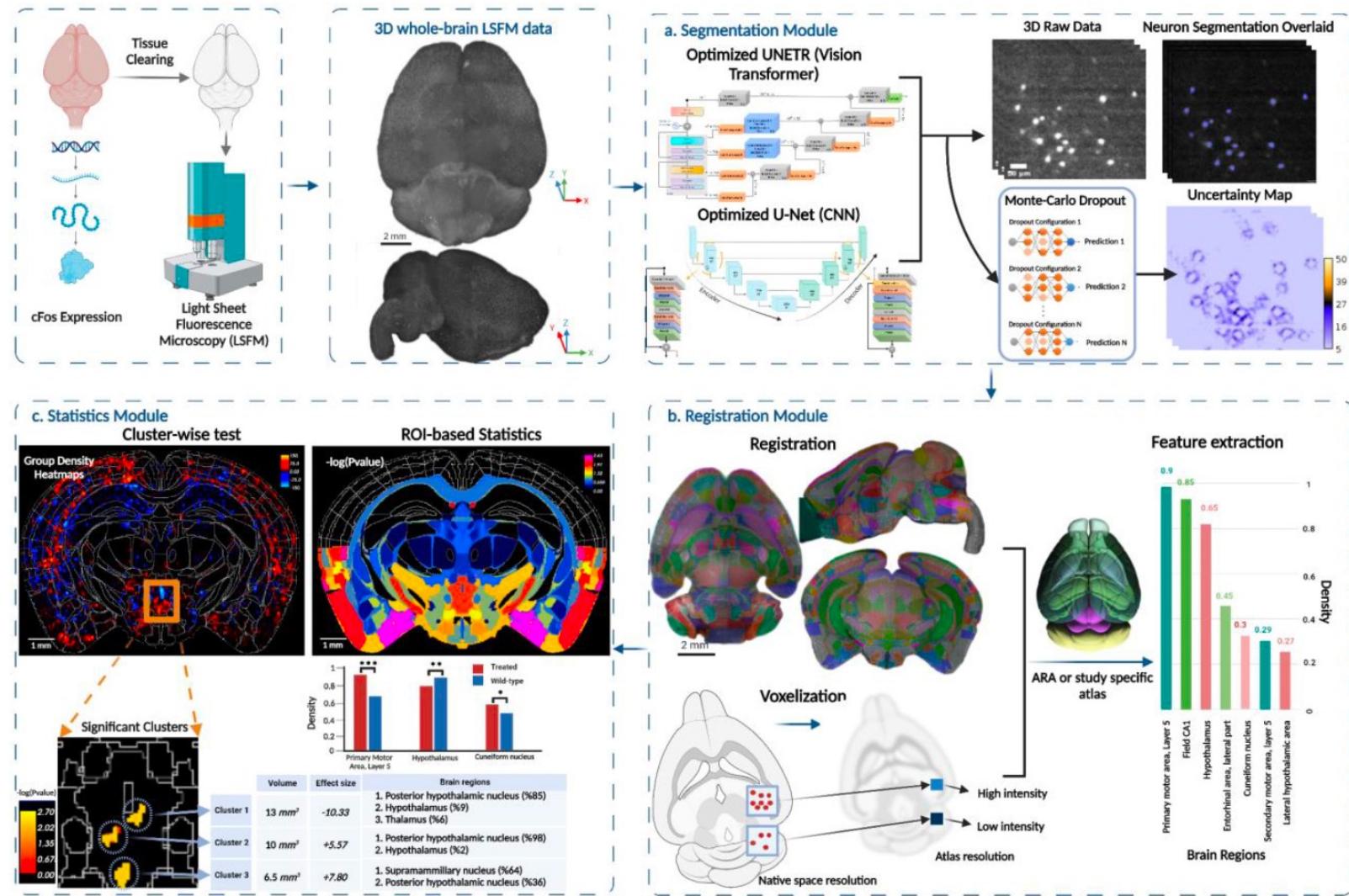
Regional mapping / Cartography





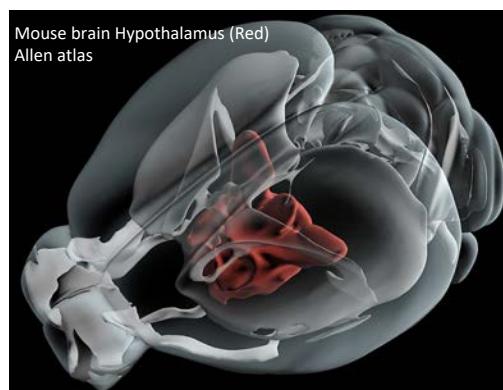
ACE: AI-BASED CARTOGRAPHY OF (NEURONAL) ENSEMBLES

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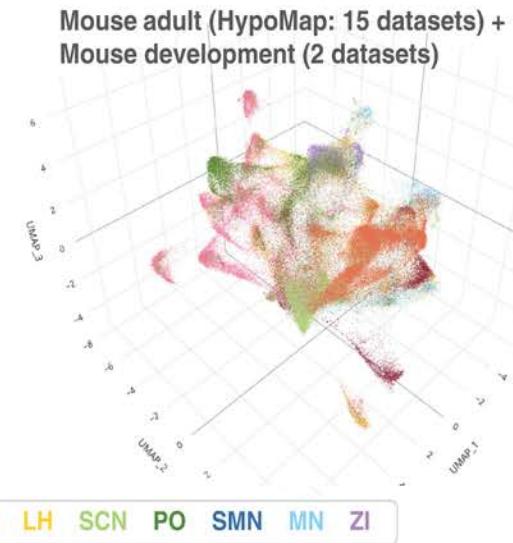
MOTIVATION BEHIND ACE

- Methodological gaps:
 - Current computational pipelines are focused on *regional* analysis, yet
 - Within-region activity is quite heterogeneous (*cf.* results of single cell genomics and transcriptomics analyses)



TM ARC PVH VMH DMH LH SCN PO SMN MN ZI

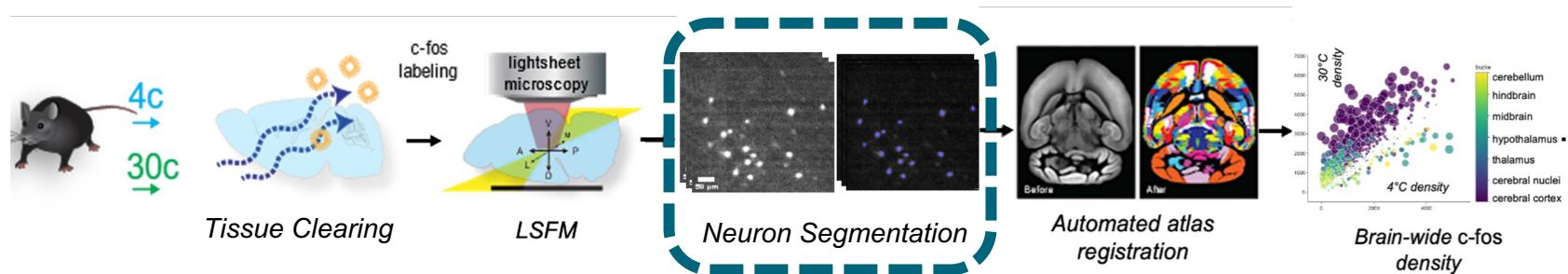
different hypothalamic neuronal subtypes,
identified by single cell transcriptomics¹



- How do we map local/sub-regional/laminar-specific activity across the brain?

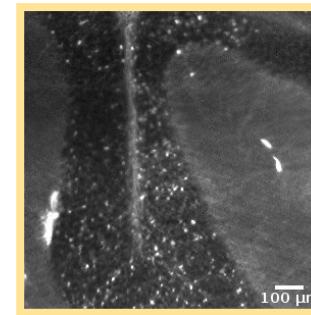
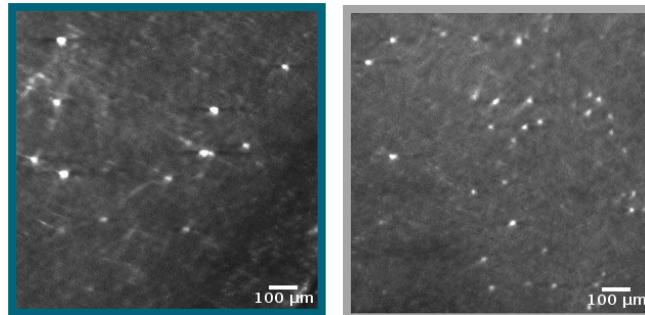
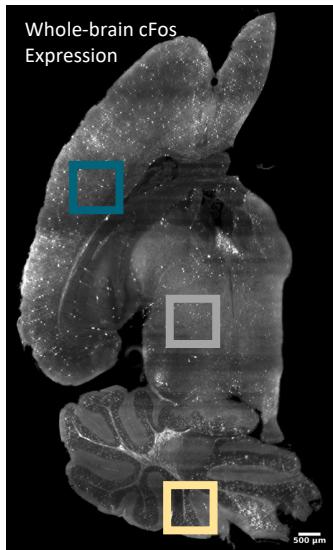
¹ Herb, et al. *Science Advances*, 2023

CHALLENGES OF WHOLE-BRAIN MAPPING OF NEURONAL ACTIVITY



➤ Current segmentation tools:

- 2D-based techniques^{1, 2}
- A small area of the brain³
- Conventional processing methods⁴ with limited generalizability

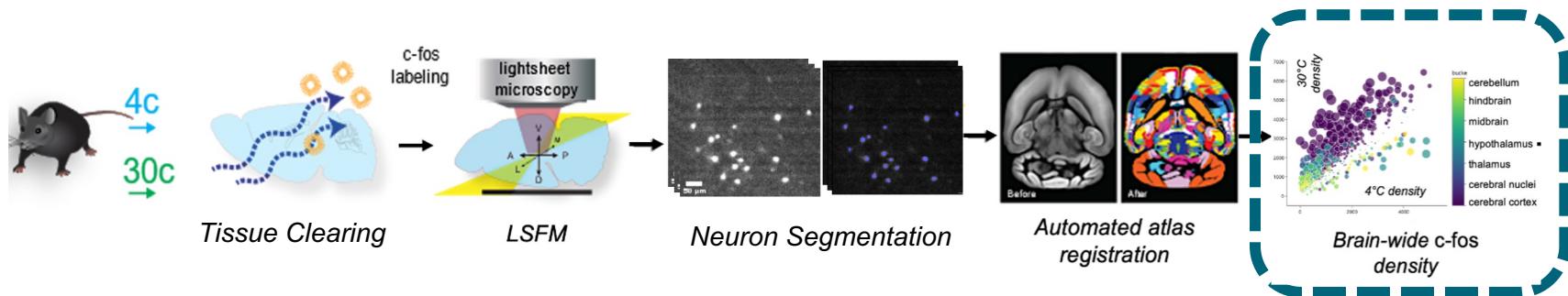


¹Stringer, et al *Nat. Methods*, 2021 ³Herbel, et al. *Nat. Methods*, 2018

²Schmidt et al. *MICCAI*, 2018

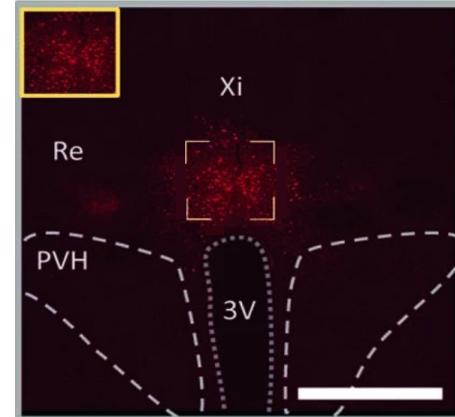
⁴Berg, et al. *Nat. Methods*, 2019

CHALLENGES OF WHOLE-BRAIN MAPPING OF NEURONAL ACTIVITY



- Current pipelines (ClearMap¹ and MIRACL²) rely on pre-defined atlas regions:
 - Requires a priori knowledge or data-specific expertise to choose ROIs
 - Provides no insight into subregional changes

- In prior work³, using manual identification of ROIs
 - Xiphoid: a key area for mediating food-seeking behaviors in response to cold stress
 - Xiphoid does not have pre-defined boundaries in the Allen atlas³



¹ Renier, et al. *Cell*, 216

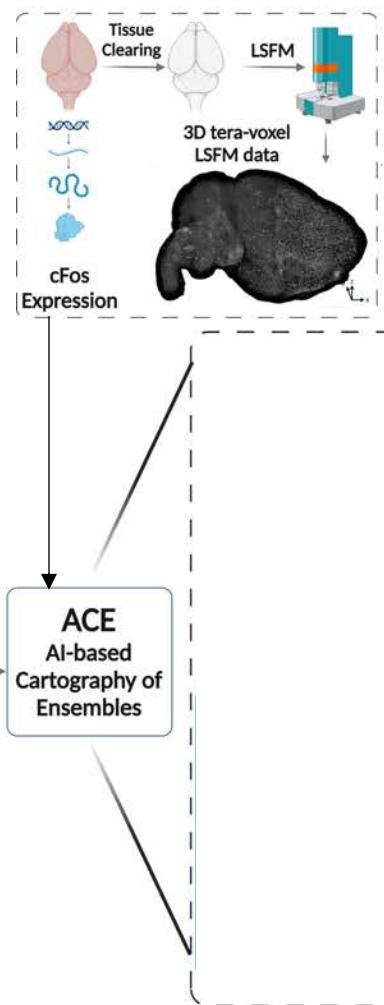
² Goubran et al. *Nat Commun*, 2019

³ Lal et al. *Nat*, 2023

Aim

A deep learning pipeline for **3D** mapping of **local** neuronal activity in
tera-voxel light sheet microscopy data

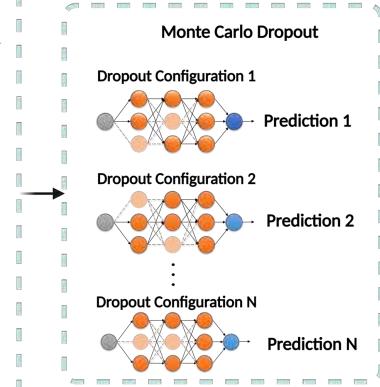
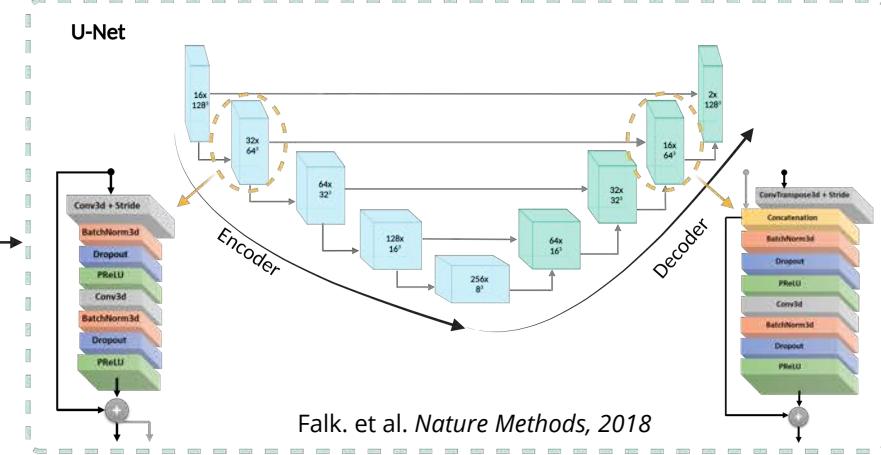
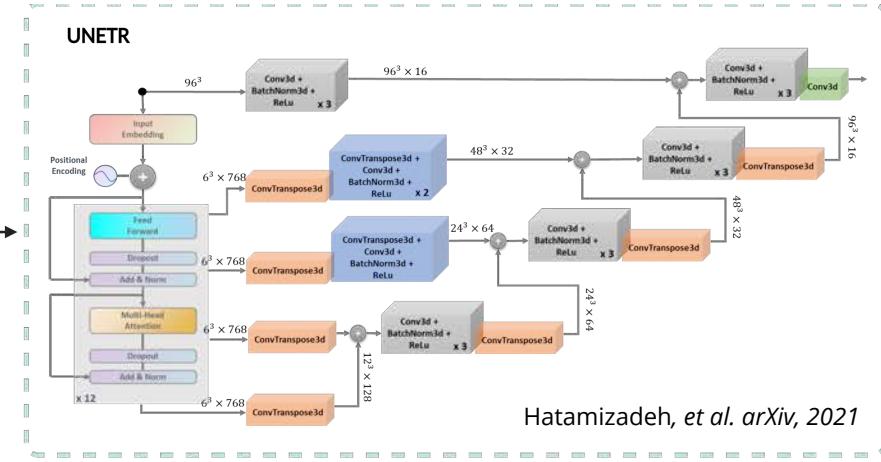
AI-BASED CARTOGRAPHY OF ENSEMBLES (ACE)



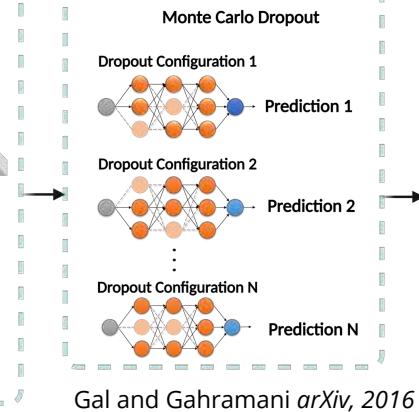
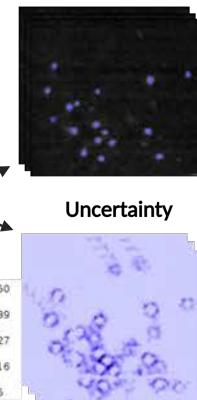
ACE'S SEGMENTATION CORE



3D Image Patch



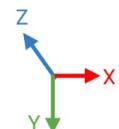
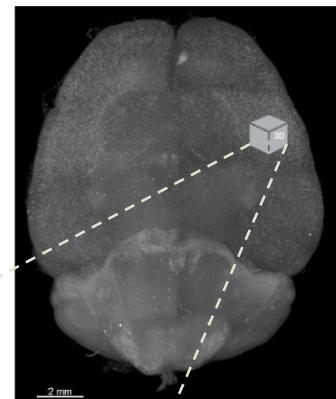
Segmentation



MODEL TRAINING & DATASETS

- **LSFM data acquired from 18 TG mice (TRAP2-Ai9 mice: tamoxifen dependent expression of cFos¹⁾**

Whole brain cFos expression

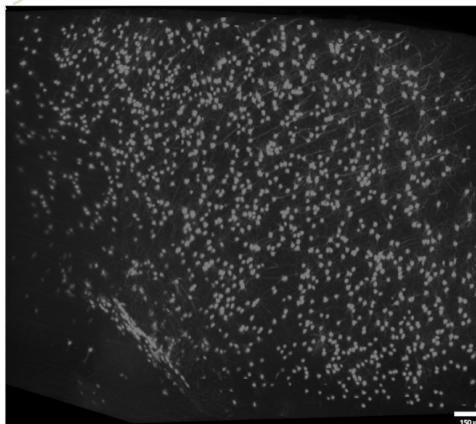


- **Image specifications:**

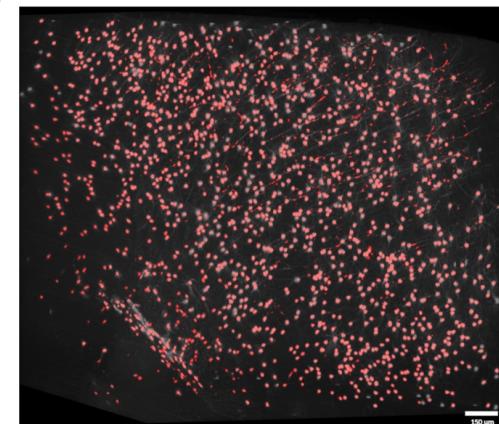
- Matrix size: ~ 2800x5500x2100 Voxels
- File Size: ~ 1 TB per subject
- Resolution: 3.5x3.5x4 µm
- 10/3/5 subjects to train/validate/test
- each patch: 96^3 voxels ~ $0.35 \times 0.35 \times 0.35$ mm³
- **N=30,400 patches** > 5x existing models

- **Semi-automated ground truth generation of cFos positive neurons (silver standard):**

- 3D Watershed Algorithm² - precision
- Ilastik³ - sensitivity
- Shape filter⁴ – large object removal



3D image patch **1.8×1.8×2 mm**



Ground truth (red)

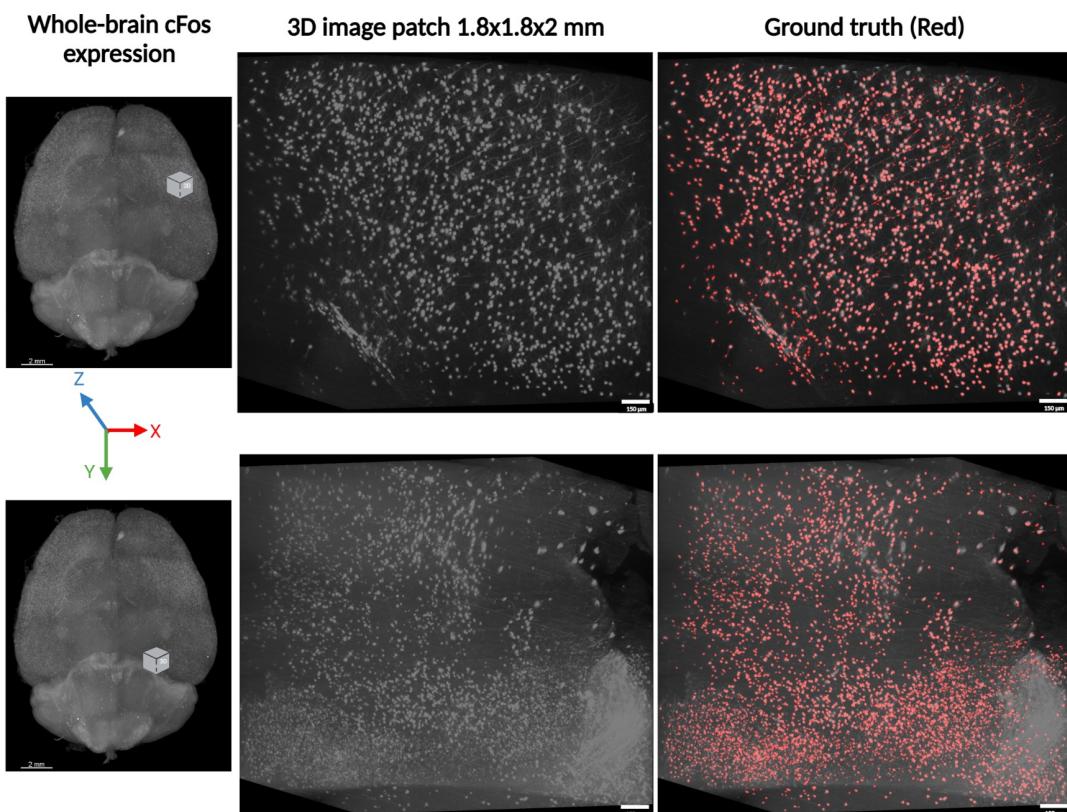
¹ DeNardo et al. *Nat Neuroscience*, 2019

³ Berg, et al. *Nat. Methods*, 2019

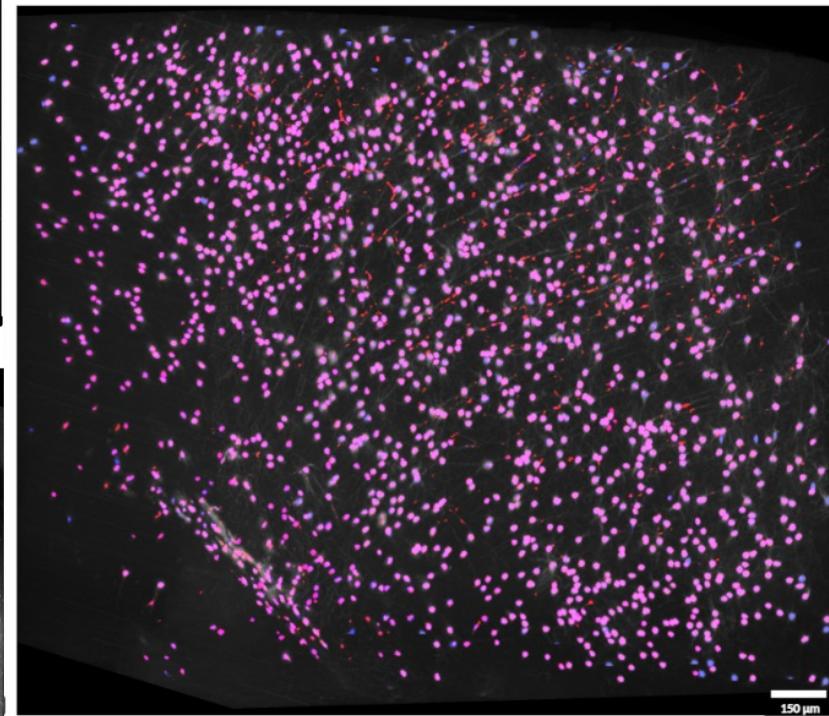
² Goubran et al. *Nat Commun*, 2019

⁴ Schindelin, et al. *Nat. Methods*, 2012

SEGMENTATION RESULTS



Overlay (Purple) / Dice = 0.78



COMPARISON TO STATE-OF-THE-ART METHODS

$$DSC(GT, P) = 2 \times \frac{|GT \cap P|}{|GT| + |P|}$$

$$HD(X, Y) = \max\{\sup_{x \in X} \inf_{y \in Y} d(x, y), \sup_{y \in Y} \inf_{x \in X} d(y, x)\}$$

$$Precision = \frac{TP}{TP+FP}$$

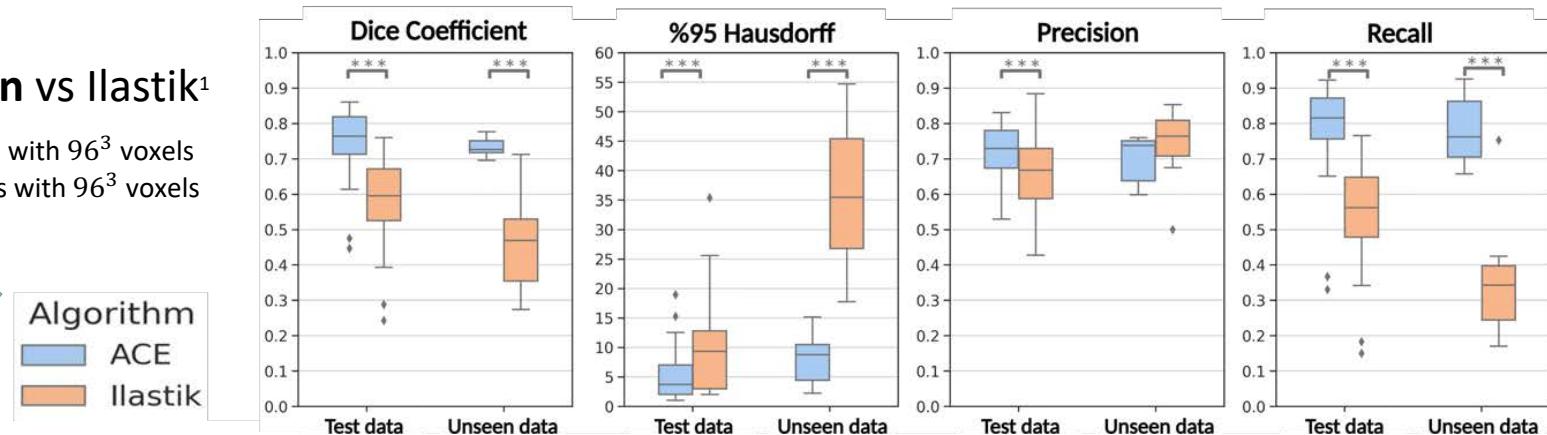
$$Recall = \frac{TP}{TP+FN}$$

$$F1\text{-score} = \frac{2 \times Precision \times Recall}{Precision + Recall}$$

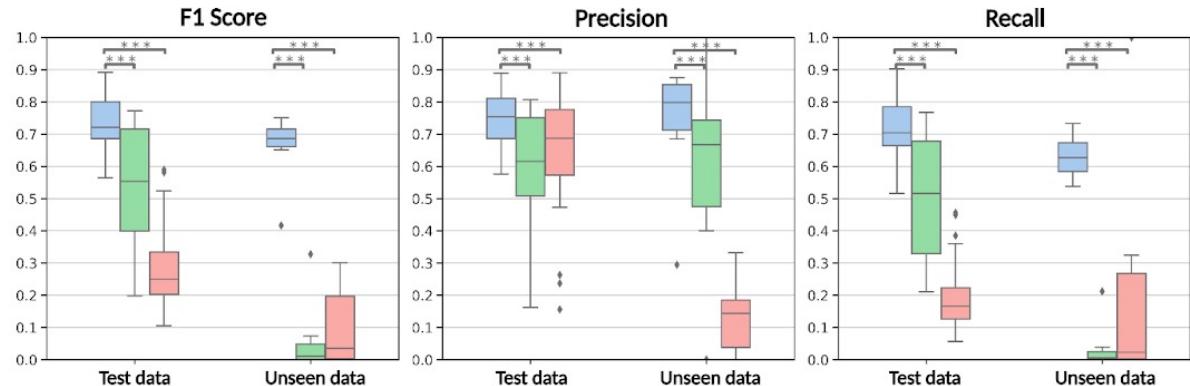
ACE Segmentation vs Illestik¹

N_{test} : 12,160 unique patches with 96^3 voxels

N_{unseen} : 1820 unique patches with 96^3 voxels

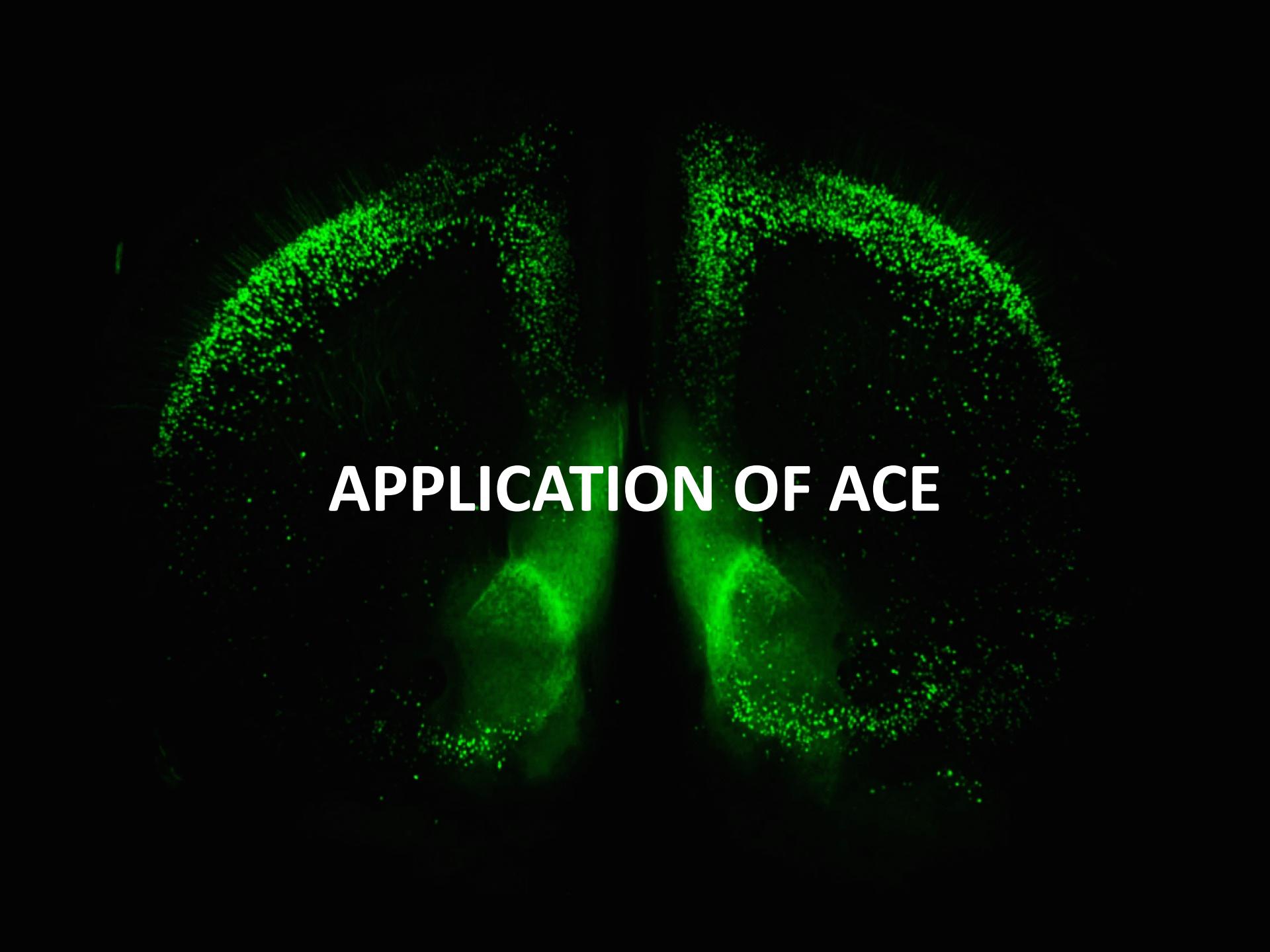


ACE Detection mode vs Cellfinder²



¹ Berg et al. Nat Method, 2019

² Tyson et al. PLOS COM. BIO., 2019



APPLICATION OF ACE

MAPPING OF NEURONAL ENSEMBLES UNDERLYING COLD-INDUCED FOOD SEEKING

- Aim: map neuronal activations underlying food-seeking behavior in response to cold stress¹
- Wild-type C57BL/6 mice exposed to either 4°C (cold) or 30°C (thermoneutral) for 6 hrs
- *c-Fos* staining
- N = 4/group
- Image size: ~ 8000x6000x3000 voxels
- Image resolution: 1.75x1.75x4 μm

A. Overview of experiments and analysis

Acclimation at 4°C vs 30°C



¹ Lal, et al. *Nature*, 2023

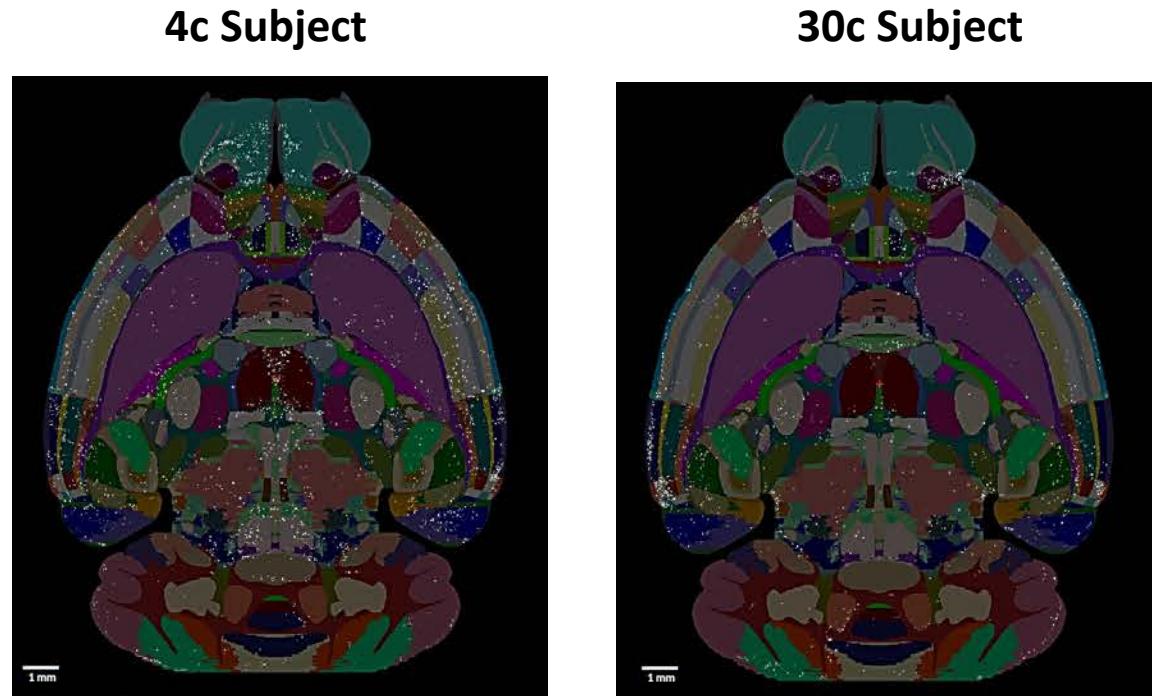
MAPPING OF NEURONAL ENSEMBLES UNDERLYING COLD-INDUCED FOOD SEEKING

- segment:
 - ACE UNET/UNETR ensemble of ensembles

- register:
 - Interface to MIRACL¹

- voxelize & warp to the Allen² atlas (10 um)

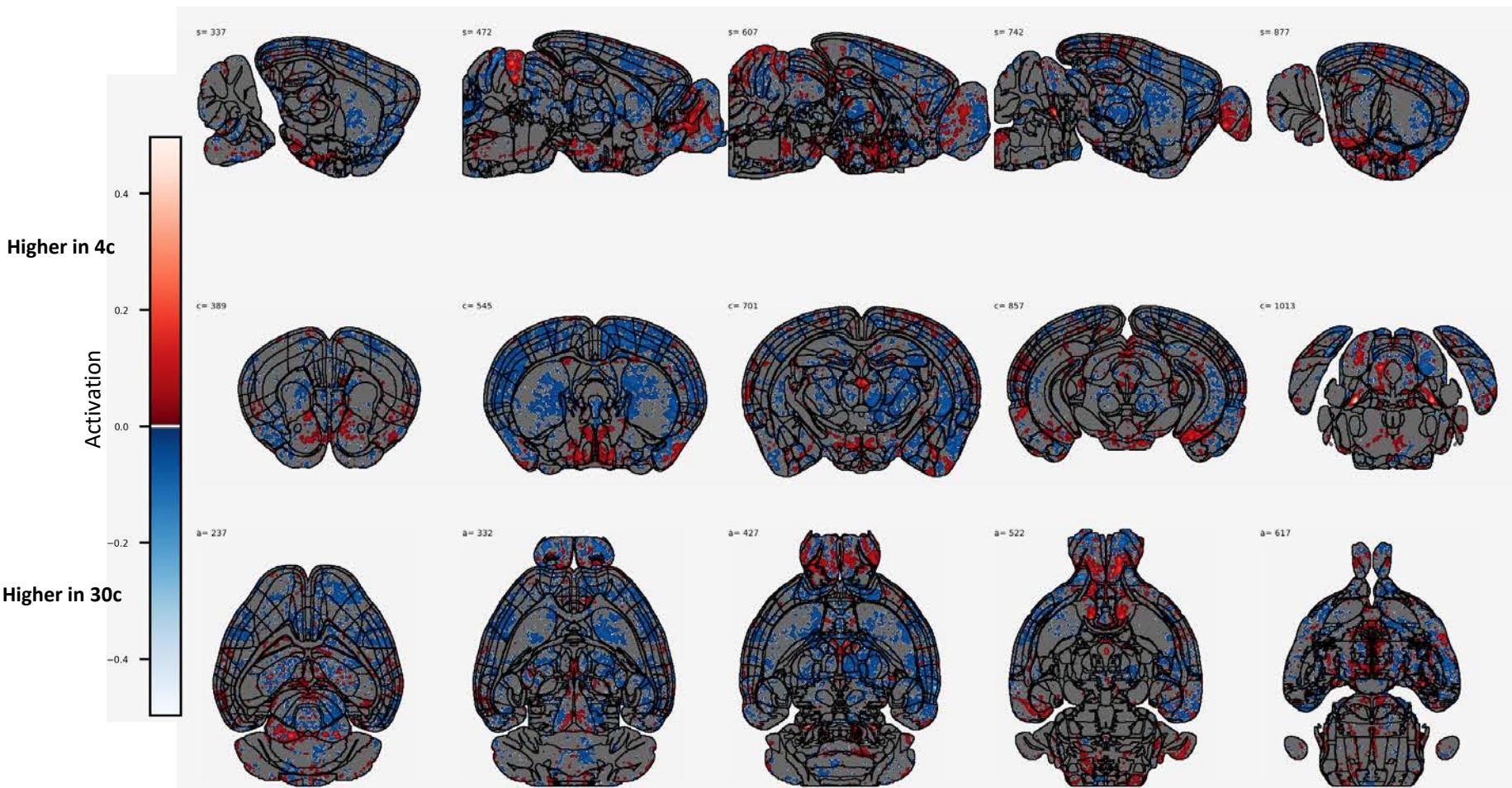
- Generate group density heat maps



¹Goubran *et al.* *Nat Commun*, 2019

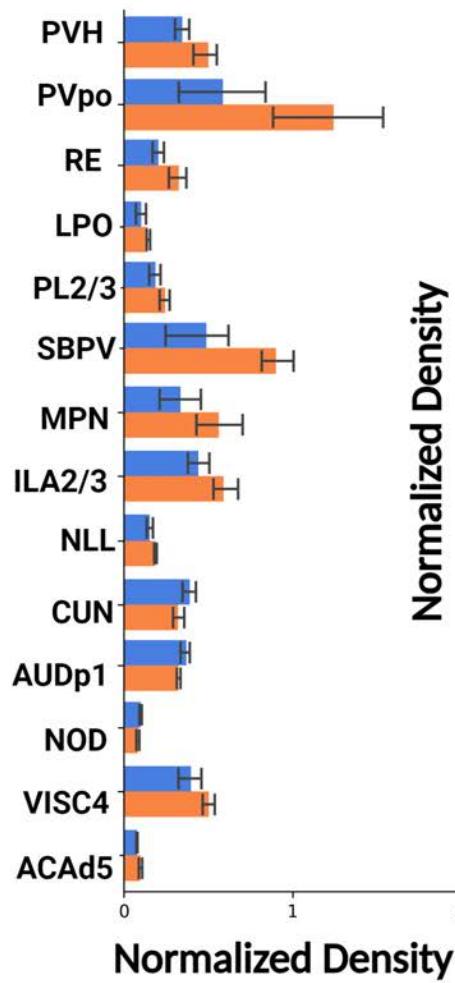
²Wang, *et al.* *Cell*, 2020

GROUP NEURONAL (C-FOS) DENSITY HEATMAPS

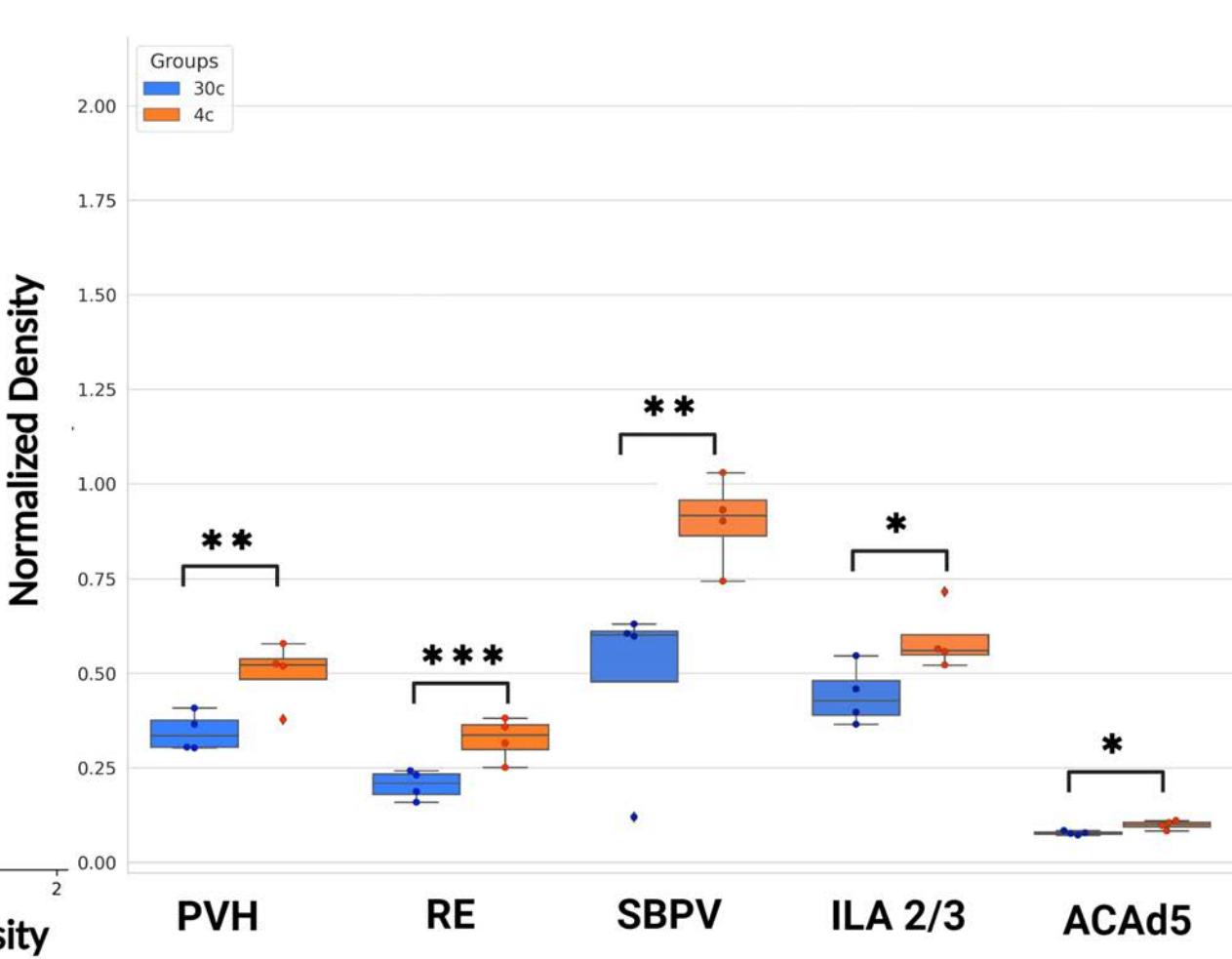


ROI-BASED STATISTICAL ANALYSIS (USING ARA LABELS)

Trending regions

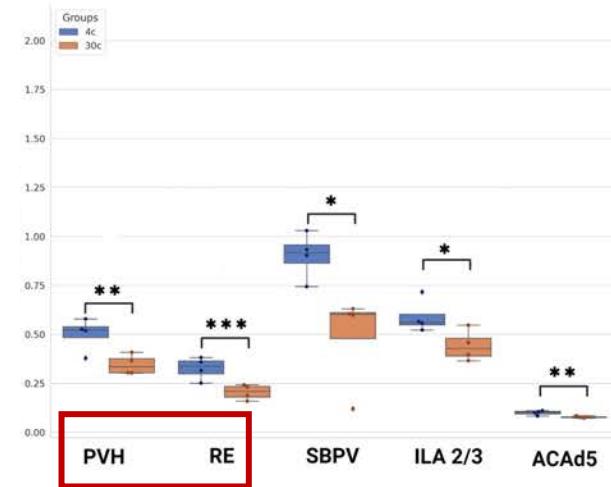


Significant regions



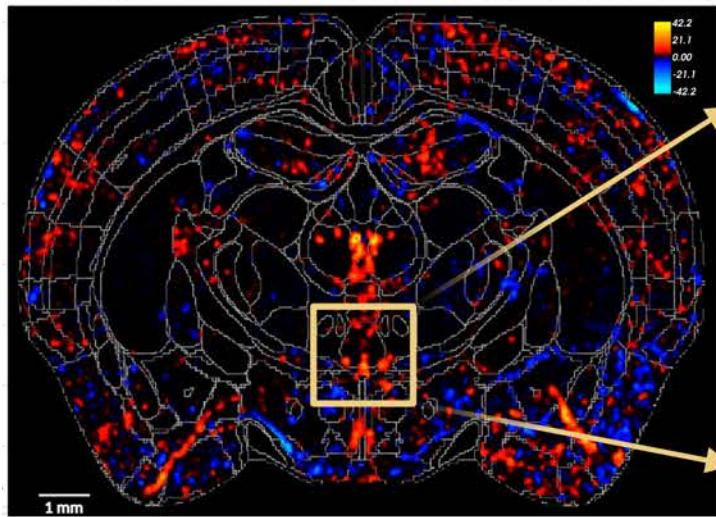
CLUSTER-WISE ANALYSIS DETECTS LOCAL NEURONAL ENSEMBLES UNDERLYING FOOD-SEEKING BEHAVIOR DURING COLD STRESS

- map *localized* differences (w excellent sensitivity) in c-Fos in an atlas-agnostic manner by a cluster-wise threshold-free cluster enhancement permutation test using a group-wise ANOVA
- detect a cluster in Xiphoid - a small nucleus in midline thalamus:
 - no pre-defined boundary in Allen Atlas
 - proximal to **paraventricular hypothal nuc (PVH)** & **nuc of reunions (RE)**
 - selectively activated by prolonged cold exposure¹

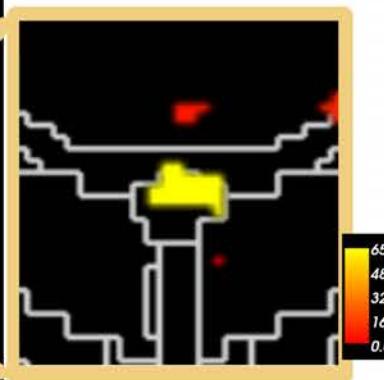


¹ Lal, et al. *Nature*, 2023

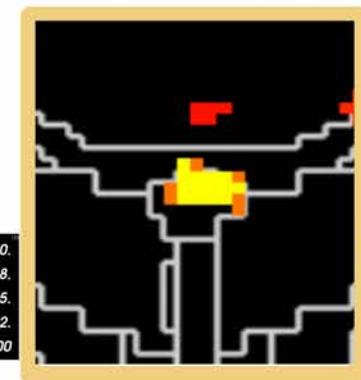
Group difference density heatmap



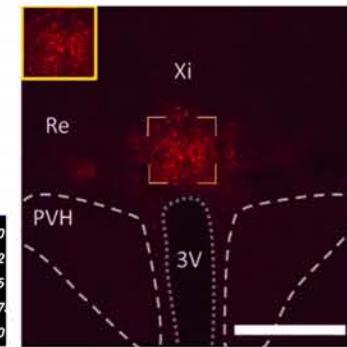
f statistics



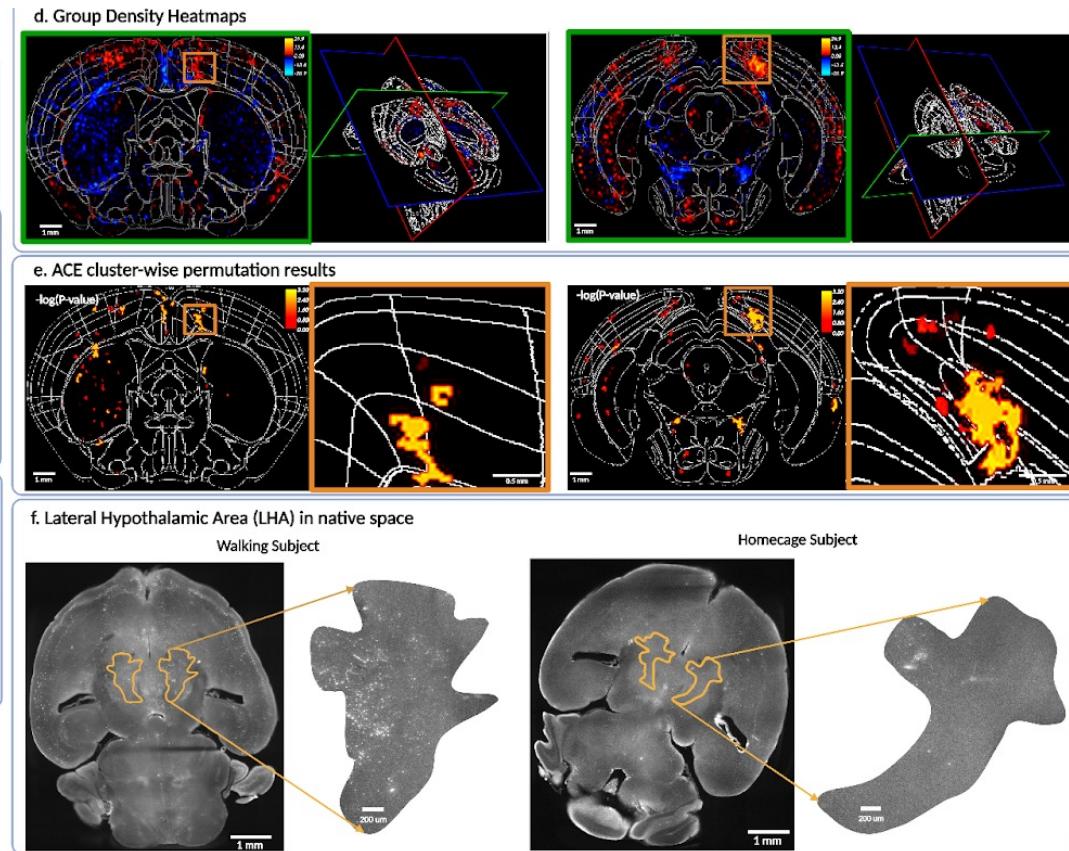
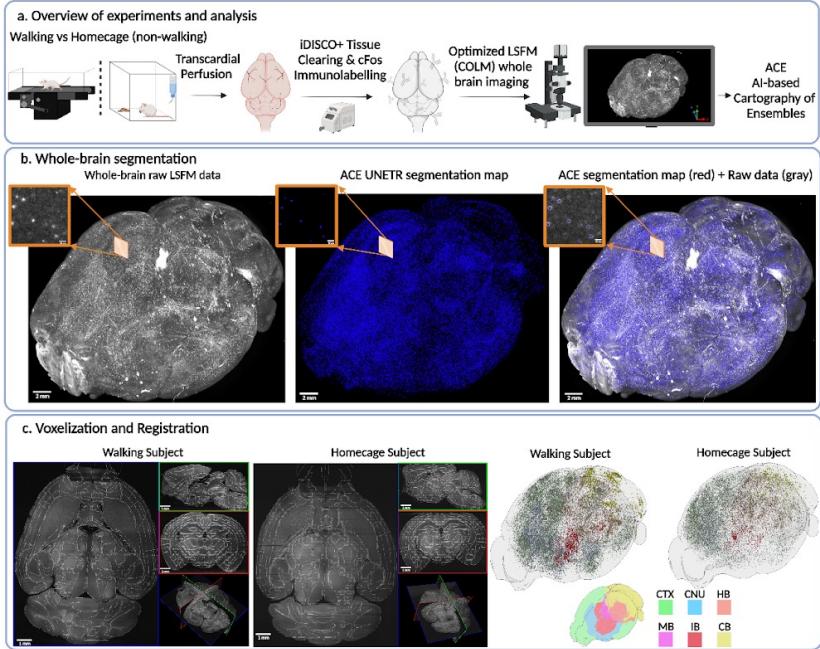
-log(P-value)

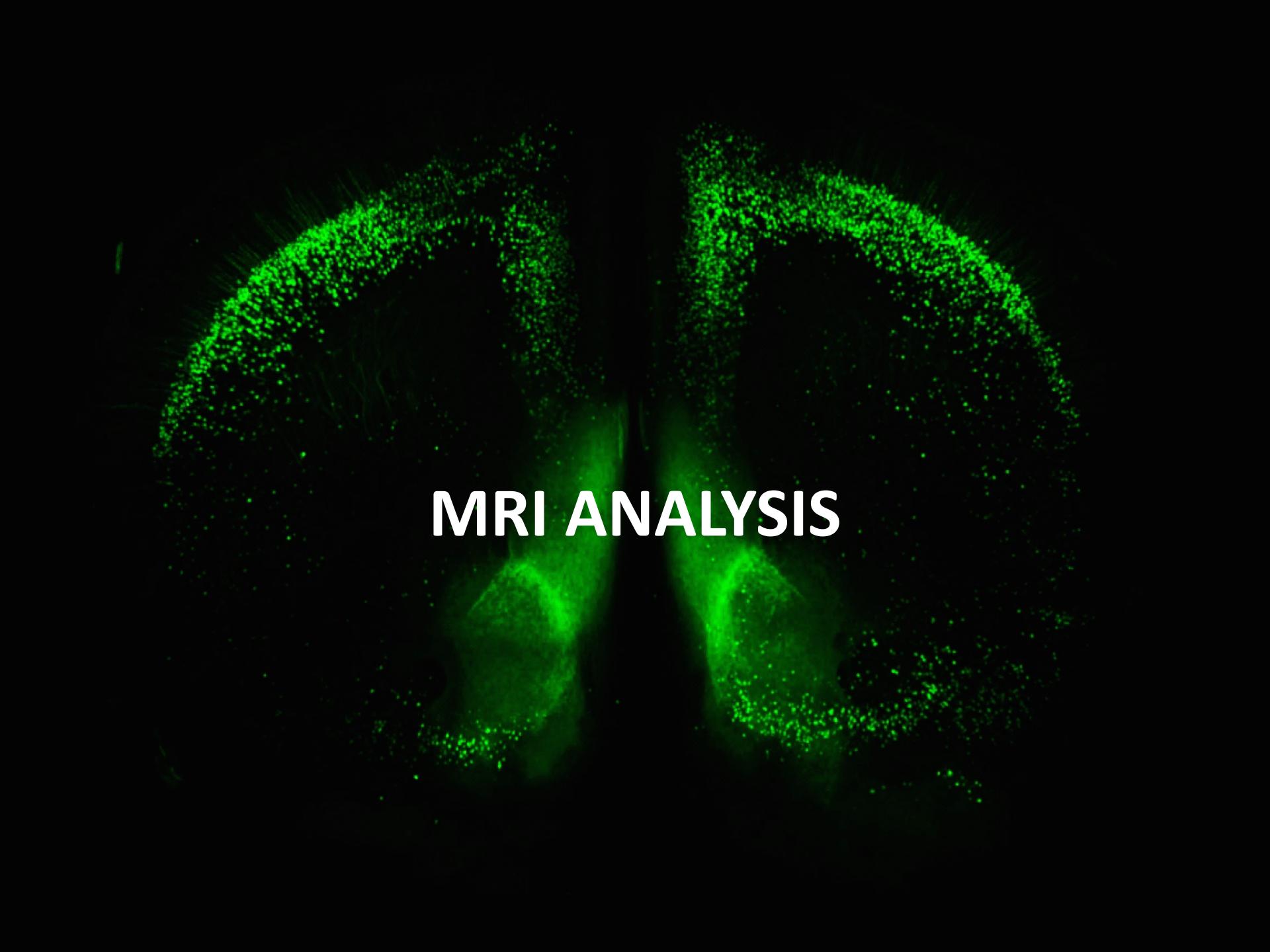


Lal et al. *Nat.* 2023



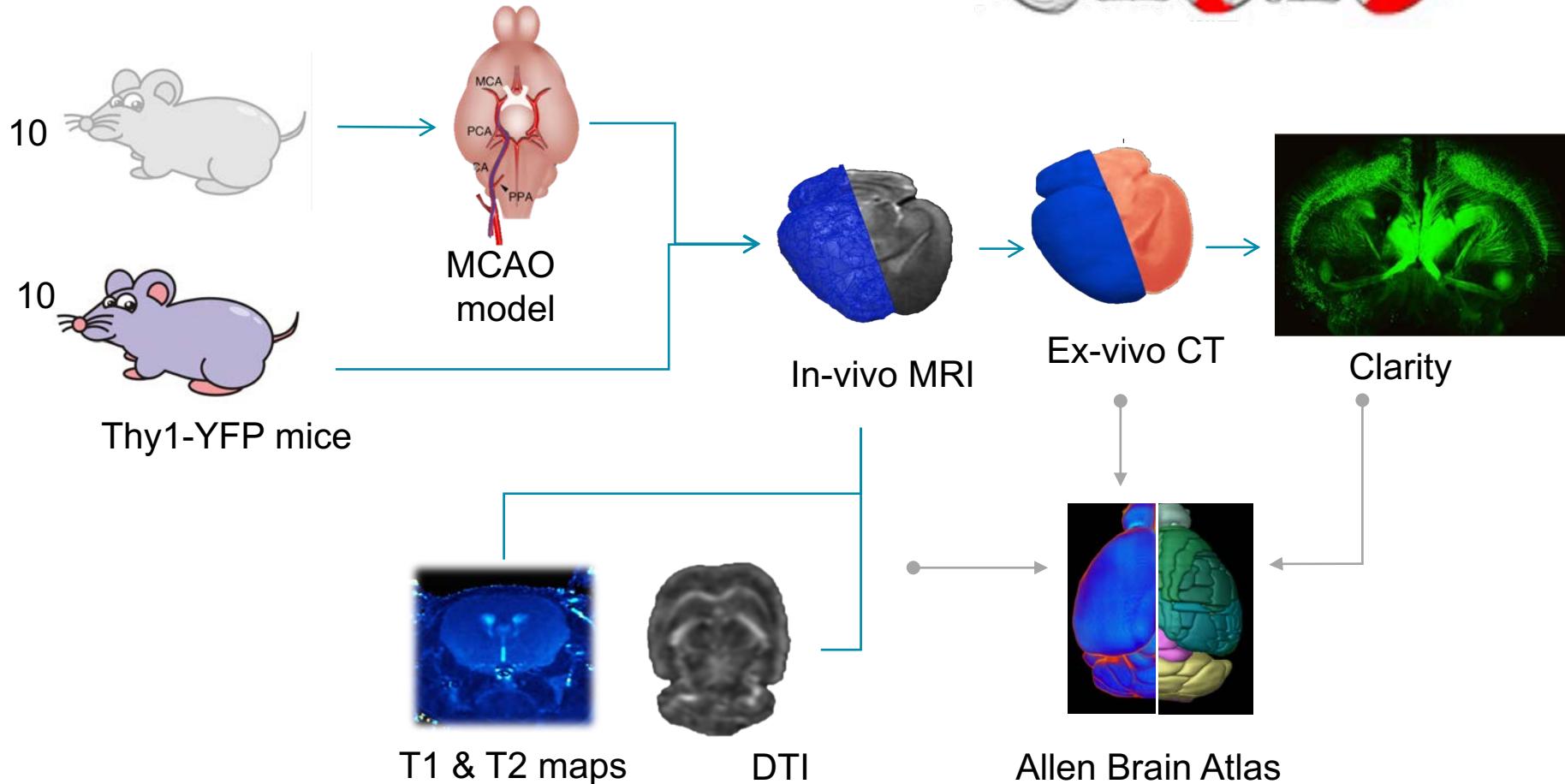
APPLICATION OF ACE: WALKING VS HOMECAGE



A grayscale MRI scan of a human brain, viewed from a slightly elevated angle. The image shows internal structures like the cerebral cortex, white matter tracts, and ventricles. The background is dark, making the bright tissue structures stand out.

MRI ANALYSIS

STROKE MODEL & STUDY DESIGN

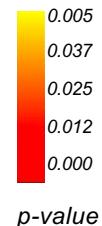
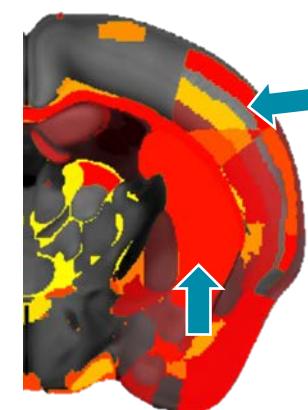
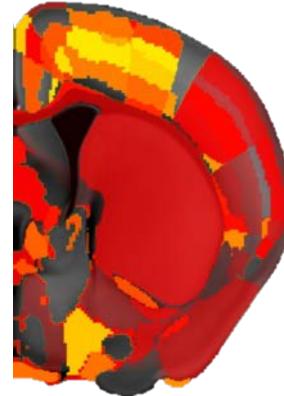
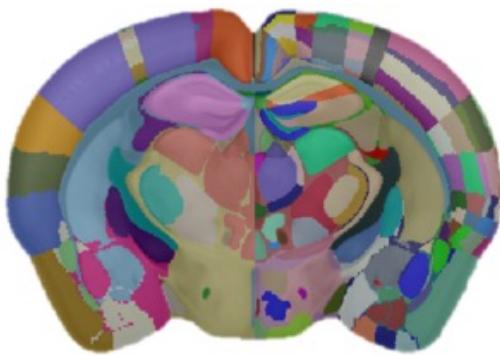


IPSILESIONAL VS. CONTRALESIONAL STATISTICS

0.005

F

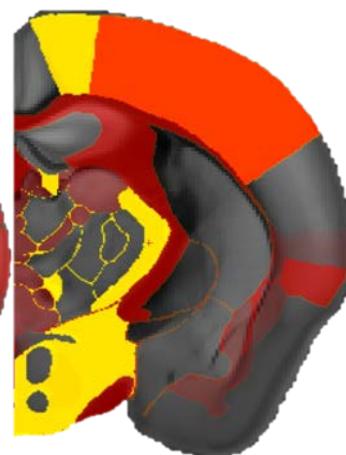
Label-wise paired t-test (ipsi vs. contra)



'Grand-parent' ARA labels used for correlation analysis

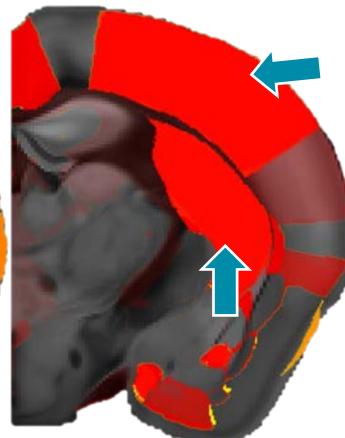
EYFP group statistics

0.005
0.037
0.025
0.012
0.000
p-value



0.005
0.037
0.025
0.012
0.000
p-value

Propidium Iodide

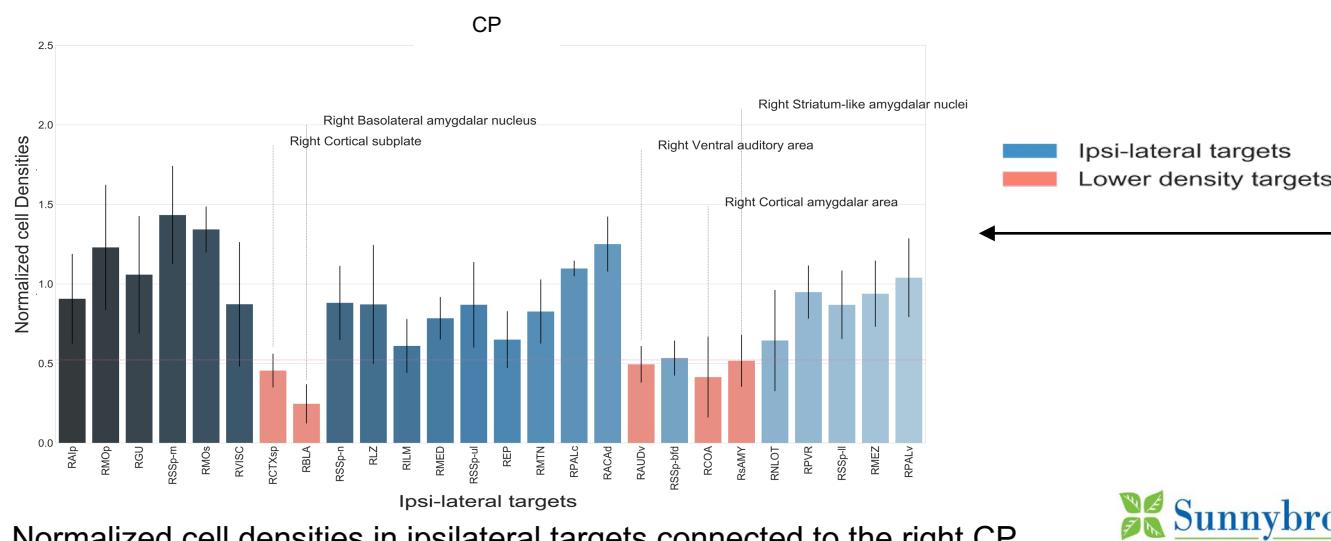
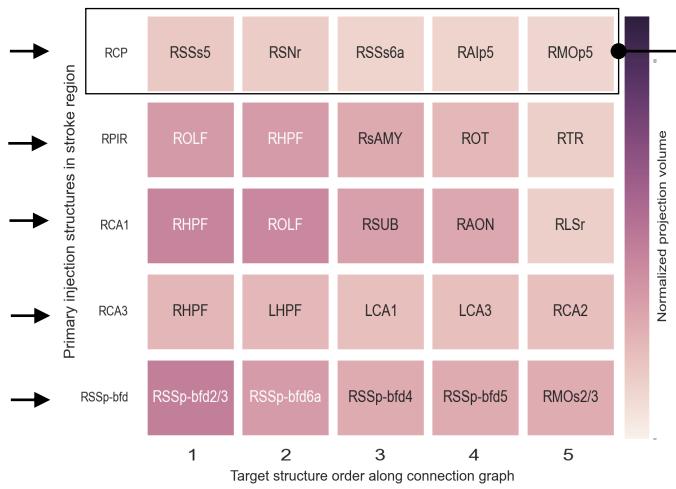
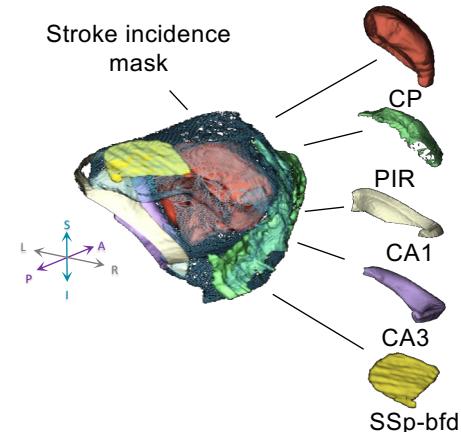
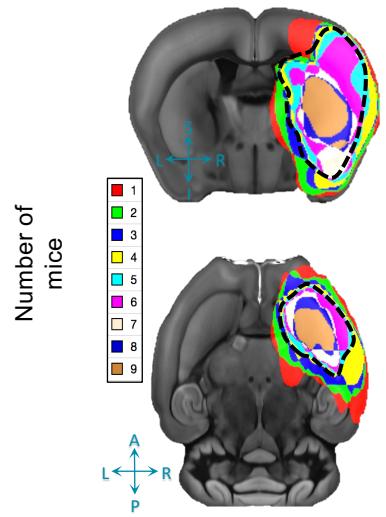


0.005
0.037
0.025
0.012
0.000
p-value

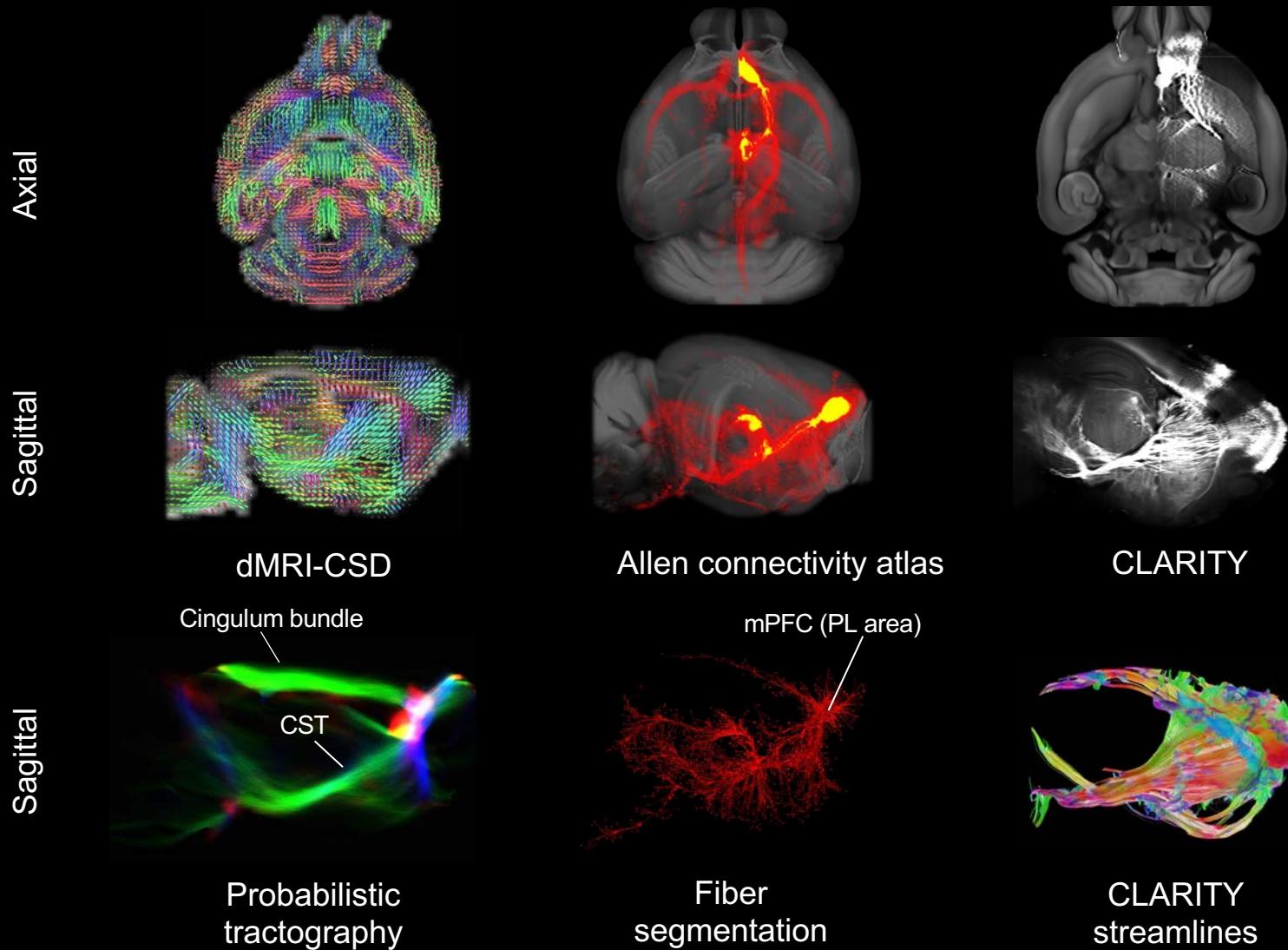
Quantitative T2

Mean Diffusivity

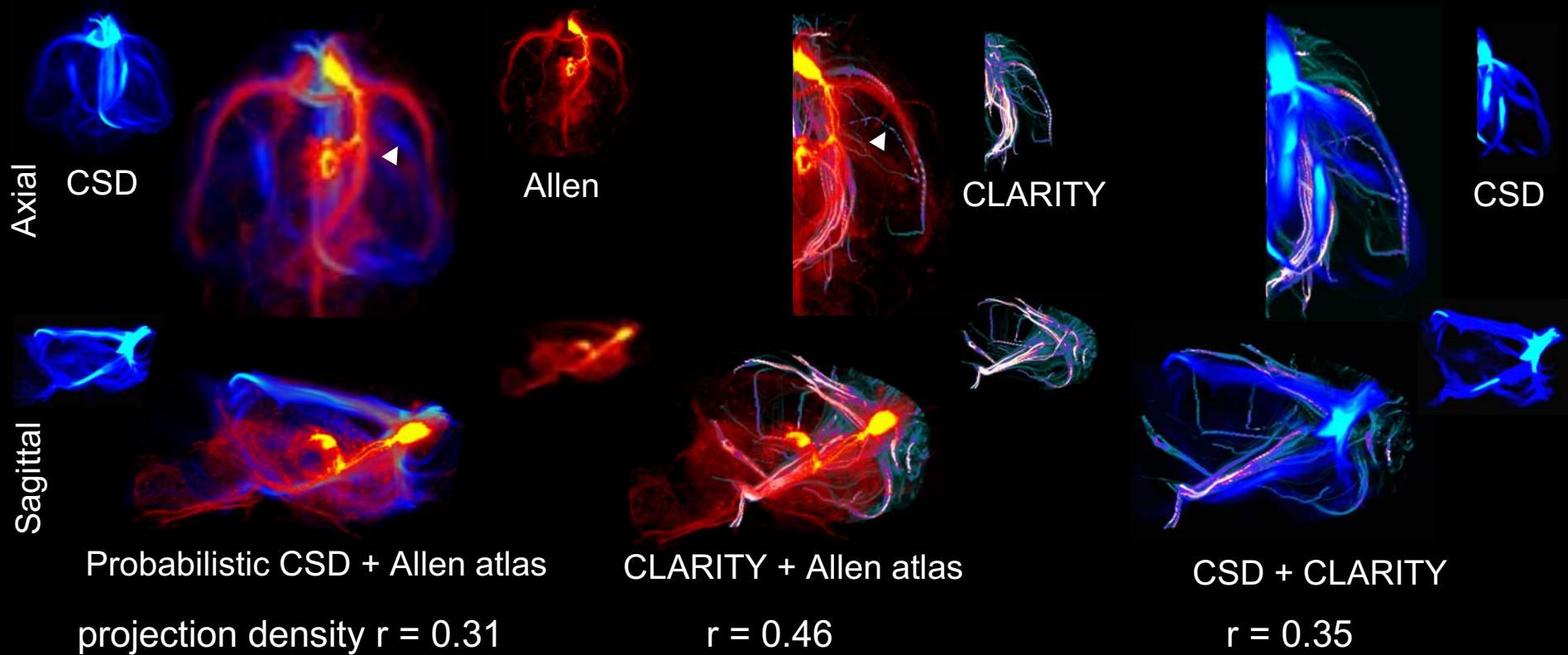
CELL DENSITY IN CONNECTED REGIONS TO THE STROKE



DMRI VS CLARITY VS ALLEN ATLAS



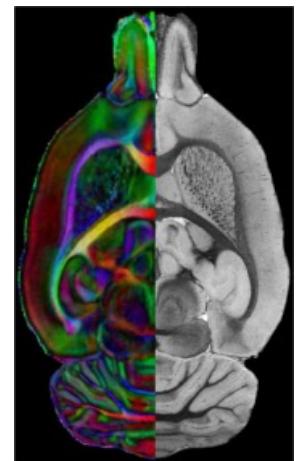
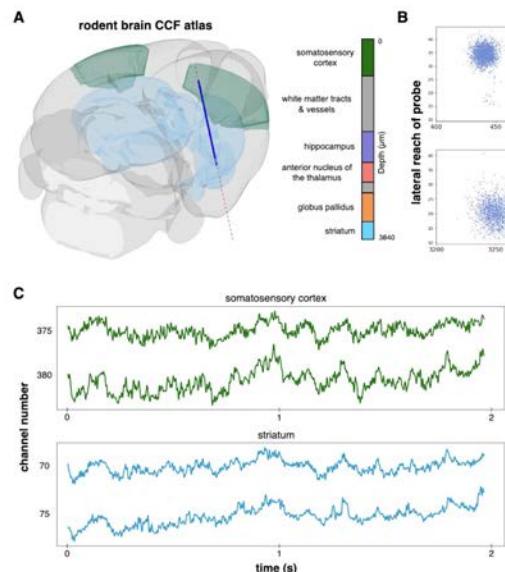
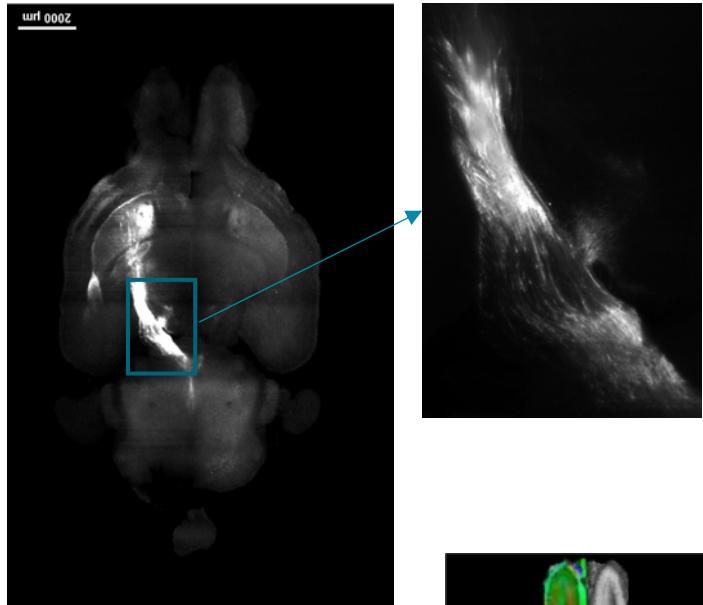
DMRI VS CLARITY VS ALLEN ATLAS



FUTURE WORK

- AI models for quantification of single-axon projections
- AI-assisted registration
- More atlases
- Integration of fMRI & Neuropixels (high-density Ephys)

Virus tracing in the rat



WEBSITE

- <https://miracl.readthedocs.io/>

- Installation

- Docker
- Singularity
- Sherlock

- Tutorials

- Troubleshooting

- Example data

Installing and running MIRACL

We provide instructions on how to install and run MIRACL using either of the following methods:

Important

Docker is our recommended method for running MIRACL on local machines and servers. We recommend Singularity to run MIRACL in a cluster environment (e.g. Compute Canada).

Attention

Support for installing MIRACL locally (i.e. on your host system directly without using Docker or Singularity) will be phased out in future versions of the software.

Docker Singularity Local Windows

We provide a build script to automatically create a Docker image for you that can be run using Docker Compose. This method does not require a manual installation of MIRACL and works on Linux, macOS and Windows (using WSL 2).

Tip

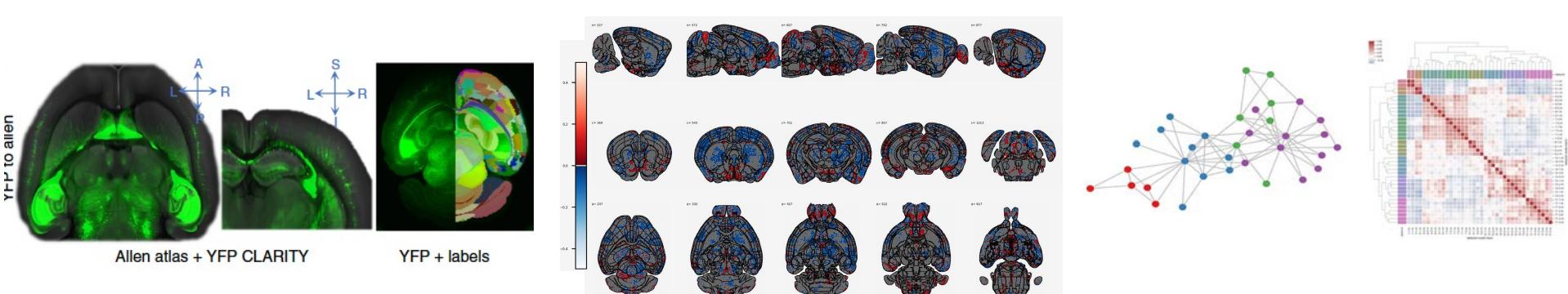
This is our recommended method for running MIRACL on local machines and servers

Docker is well suited if you want to run MIRACL on a local machine or local server. If you need to run MIRACL on a cluster, see our instructions for installing Singularity. If you don't have Docker installed on your computer, do that first. Make sure your installation includes Docker Compose as it is required to run the build script we provide. Note that Docker Compose is included as part of the Docker Desktop installation by default.

Getting started

SUMMARY

1. Registration of cleared and imaging data to atlases
2. 3D Segmentation and feature extraction of cleared data
3. Tract-specific or network-level connectivity analysis
4. Statistical analysis of cleared and imaging data
5. Comparison of dMRI/tractography, virus tracing, & connectivity atlases
6. Atlas generation and Label manipulation



THANK YOU

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