Statistical Methods for High Dimensional Biology STAT/BIOF/GSAT 540

Lecture 9 – Linear Models Part III

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FEB 01 2016

outline

Review of previous lecture

 Linear regression III – general formulation and quantitative covariates

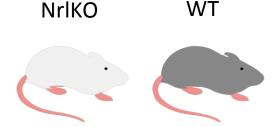
Analyzing patterns across all genes

Developing mouse retina – time course for the experiment

So sample collections:

4 developmental stages

2 genotypes: wild-type , Nrl KO



Experimental design

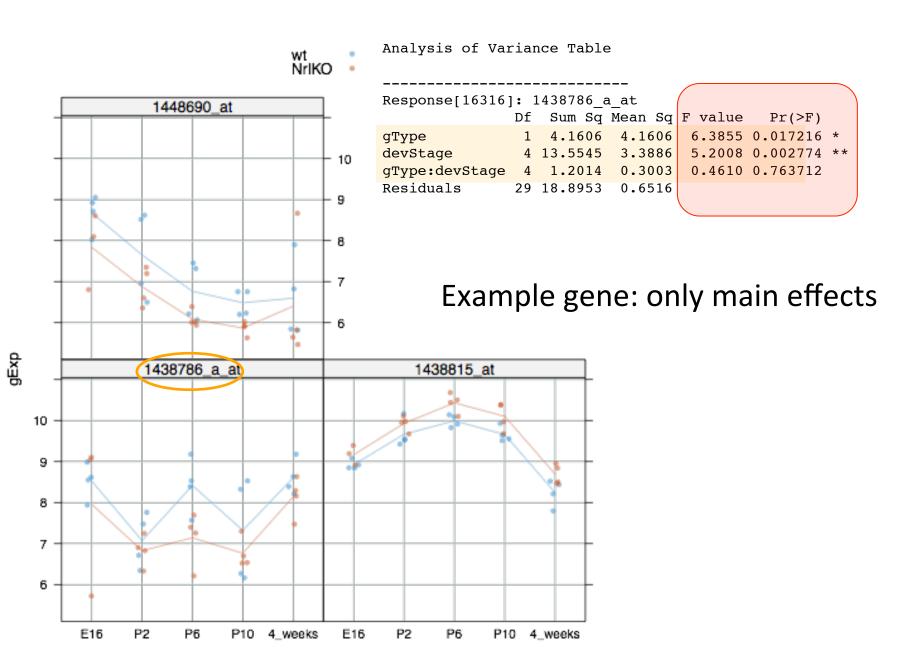
devStage	wt	NrlKO
E16	4	3
P2	4	4
P6	4	4
P10	4	4
4 _weeks	4	4

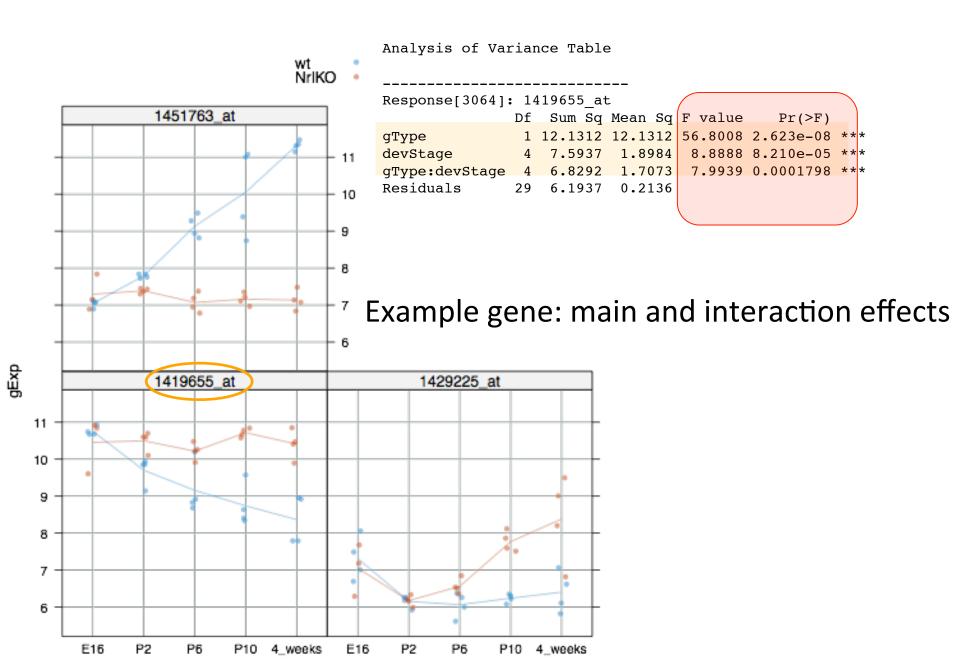
- 1) What are the genes whose expression levels differ between the two genotypes?
- 2) What are the genes whose expression levels differ across developmental stages?
- 3) What are the genes whose expression *trajectories* across development differs between the two genotypes?

Two-way ANOVA

Genotype effect Interaction effect Gene i Genotype j Dev Stage k
$$y_{ijk} = \theta + \tau_j + \beta_k + (\tau\beta)_{jk} + \varepsilon_{ijk}$$
 Reference (i.e., intercept) "devStage" effect

devStage gType	E16	P2	P6	PIO	4_weeks
wt	heta	$oldsymbol{eta}_{P2}$	$oldsymbol{eta_{P6}}$	$oldsymbol{eta}_{P10}$	$oldsymbol{eta}_{4_weeks}$
NrlKO	τ _{NrlKO}	(τ β) _{NrlKO,P2}	$(auoldsymbol{eta})_{NrlKO,P6}$	(τ β) NrlKO,P10	(τ β) NrlKO,4_weeks





F tests in regression

small model is nested within big -- it's a special case where some parameters are equal to zero

model	example	# params = DF	RSS
small	lm(y ~ gType + devStage)	$p_{small} = 6$	RSS _{small}
big	Im(y ~ gType * devStage)	Pbig = I0	RSS _{big}

$$y_{ijk} = \theta + \tau_j + \beta_k + (\tau \beta)_{jk} + \varepsilon_{ijk} \text{ "big"}$$

$$y_{ijk} = \theta + \tau_j + \beta_k + (\tau \beta)_{jk} + \varepsilon_{ijk} \text{ "small"}$$

F tests in regression

small model is nested within big -- it's a special case where some parameters are equal to zero

model	example	# params = DF	RSS
small	lm(y ~ gType + devStage)	p _{small} = 6	RSS _{small}
big	Im(y ~ gType * devStage)	P _{big} = I 0	RSS _{big}

$$y_{ijk} = \theta + \tau_j + \beta_k + (\tau \beta)_{jk} + \varepsilon_{ijk} \text{ "big"}$$

$$y_{ijk} = \theta + \tau_j + \beta_k + (\tau \beta)_{jk} + \varepsilon_{ijk} \text{ "small"}$$

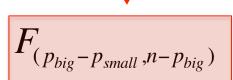
Plugging the observed F into the null distribution for F with given DOF, gives us the tail probability under the null

by definition:

$$p_{small} < p_{big}$$

 $RSS_{small} \ge RSS_{big}$

$$F = \frac{\left(\frac{RSS_{small} - RSS_{big}}{p_{big} - p_{small}}\right)}{\frac{RSS_{big}}{n - p_{big}}} \sim_{H}$$



Let's talk about linear regression as a very general framework, how do we get the parameters?

$Y = X\alpha + \varepsilon$

$$\begin{bmatrix} y_1 \\ y_2 \\ \vdots \\ y_n \end{bmatrix} = \begin{bmatrix} 1 & x_1 \\ 1 & x_2 \\ \vdots & \vdots \\ 1 & x_n \end{bmatrix} + \begin{bmatrix} \varepsilon_1 \\ \varepsilon_2 \\ \vdots \\ \varepsilon_n \end{bmatrix} = \begin{bmatrix} \alpha_0 \cdot 1 + \alpha_1 \cdot x_1 \\ \alpha_0 \cdot 1 + \alpha_1 \cdot x_2 \\ \vdots \\ \alpha_0 \cdot 1 + \alpha_1 \cdot x_n \end{bmatrix} + \begin{bmatrix} \varepsilon_1 \\ \varepsilon_2 \\ \vdots \\ \varepsilon_n \end{bmatrix} = \begin{bmatrix} \alpha_0 + \alpha_1 x_1 + \varepsilon_1 \\ \alpha_0 + \alpha_1 x_2 + \varepsilon_2 \\ \vdots \\ \alpha_0 + \alpha_1 x_1 + \varepsilon_1 \end{bmatrix}$$

$$y_i = \alpha_0 + \alpha_1 x_i + \varepsilon_i$$

Here we are just fitting a line but using matrix notation to handle all n observations at once, more elegantly.

Big pay-offs ensue

$Y = X\alpha + \varepsilon$

Estimation of the parameter α

Two viewpoints:

- maximum likelihood estimation, assuming ε_i are iid $N(0, \sigma^2)$
- "ordinary least squares" (OLS), i.e. minimizing the sum of the squared residuals

both lead to the same estimator of α :

$$\hat{\alpha} = (X^T X)^{-1} X^T y = \min^{-1} \sum_{i=1}^{n} (y_i - x_i \alpha)^2$$

Linear regression: maximum likelihood estimation

$$Y = X\alpha + \varepsilon$$
 $\varepsilon_i \sim N(0, \sigma^2)$

$$P(Y \mid \theta) = N(Y \mid X\alpha, \sigma^2) = \frac{1}{2\sqrt{\pi}\sigma} e^{\left(-\frac{(X\alpha - Y)^T (X\alpha - Y)}{2\sigma}\right)}$$

Need to maximize the log of the likelihood function to solve for the parameter(s)

Linear regression: least squares formulation

Minimize the squared error in prediction:

$$Y = X\alpha + \varepsilon$$

minimize
$$(Y - X\alpha)^T (Y - X\alpha)$$

Greatest Hits of Regression Results (normal iid errors)

$$Y = X\alpha + \varepsilon$$
 regression model

$$\hat{\alpha} = (X^T X)^{-1} X^T Y$$
 the MLE and OLS estimator of α

$$\hat{Y} = X\hat{\alpha}$$
 the fitted or predicted values

$$\hat{Y} = X(X^T X)^{-1} X^T Y = HY$$
 where $H = X(X^T X)^{-1} X^T$ is called the "hat matrix"

$$\hat{\varepsilon} = Y - \hat{Y} = Y - X\hat{\alpha}$$
 the residuals (note NOT the same as the errors ε)

$$\hat{\sigma}^2 = \frac{1}{n-p} \hat{\varepsilon}^T \hat{\varepsilon}$$
 the estimated error variance (p is the dimension of \alpha)

$$\hat{V}(\hat{\alpha}) = \hat{\sigma}^2 (X^T X)^{-1}$$
 the estimated covariance matrix of $\hat{\alpha}$ estimated standard errors for the estimated regression coefficients — $\hat{se}(\hat{\alpha}_j)$ — are obtained by taking the square root of the diagonal elements of $\hat{V}(\hat{\alpha})$

Testing the significance of the parameter estimates in linear regression

$$Y = X\alpha + \varepsilon$$

How test $H_0: \alpha_i = 0$?

With a t-statistic. Under H_0 , we have (at least approximately) that:

$$\frac{\hat{\alpha}_{j}}{\widehat{se}(\hat{\alpha}_{j})} \sim t_{n-p}$$

so a p-value is obtained by computing a tail probability for the observed value of $\hat{\alpha}_j$ from a t_{n-p} distribution.

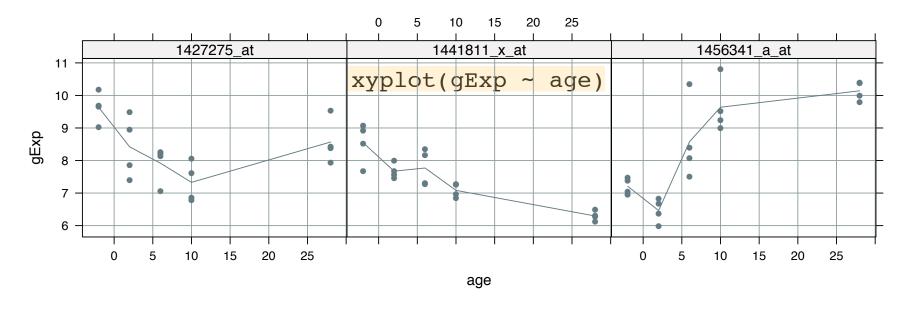
 $\hat{\sigma}^2 = \frac{1}{n-p} \hat{\varepsilon}^T \hat{\varepsilon} \text{ the estimated error variance } (p \text{ is the dimension of } \alpha)$ $\hat{V}(\hat{\alpha}) = \hat{\sigma}^2 (X^T X)^{-1} \text{ the estimated covariance matrix of } \hat{\alpha}$ estimated standard errors for the estimated regression coefficients -- $\hat{se}(\hat{\alpha}_j)$ -- are obtained by taking the square root of the diagonal elements of $\hat{V}(\hat{\alpha})$

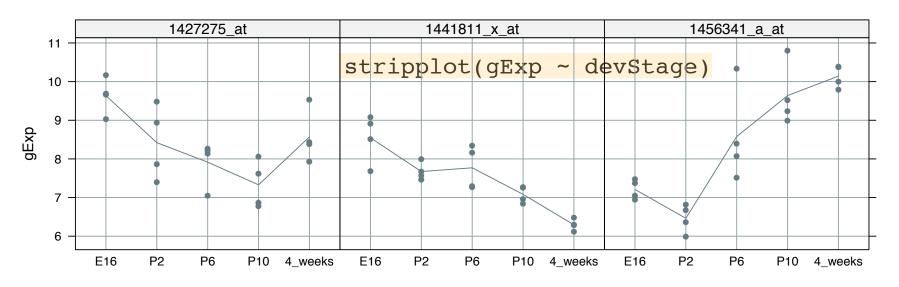
Investigating a *quantitative* covariate in our example dataset

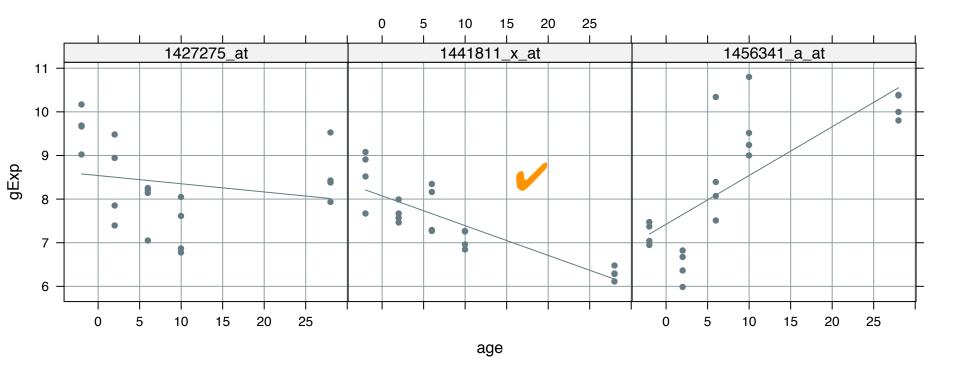
```
> ## recode() is from add-on package 'car'
> prDes$age <-
     recode(prDes$devStage,
            "'E16'=-2; 'P2'=2; 'P6'=6; 'P10'=10; '4 weeks'=28",
            as.factor.result = FALSE)
> peek(prDes)
         sample devStage gType age
Sample 22
             22
                    E16
                           wt -2
            16 E16 NrlKO -2
Sample 16
Sample 5
         5 P2 NrlKO
Sample 31
             31 P6
                           wt
         15
36 4 w
Sample 15
                    P10 NrlKO 10
Sample 36
             36 4 weeks
                           wt 28
Sample 2
         2 4 weeks NrlKO 28
> str(prDes)
'data.frame': 39 obs. of 4 variables:
$ sample : num 20 21 22 23 16 17 6 24 25 26 ...
 $ devStage: Factor w/ 5 levels "E16", "P2", "P6", ...: 1 1 1 1 1 1 1 2 2 2 ...
          : Factor w/ 2 levels "wt", "NrlKO": 1 1 1 1 2 2 2 1 1 1 ...
          : num -2 -2 -2 -2 -2 -2 2 2 2 ...
 $ age
```

meet our new quantitative covariate or predictor ... age, which is a new version of the factor devStage

for starters, let's just work with wild type data for 3 example probesets







linear looks reasonable for I, but not the other two

Remember: $Y=f(x)=a_0+bx$

- The nature of the regression function $f(x; \alpha)$ is one of the defining characteristics of a regression model
 - **-** f linear in α ⇒ linear model
 - **-** f not linear in α ⇒ nonlinear model

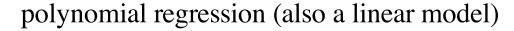
nonlinear parametric regression

$$Y = \frac{1}{1 + e^{(\phi - x)/\xi}} + \varepsilon$$

simple linear regression (a linear model)

$$Y = \alpha_0 + \alpha_1 x + \varepsilon$$

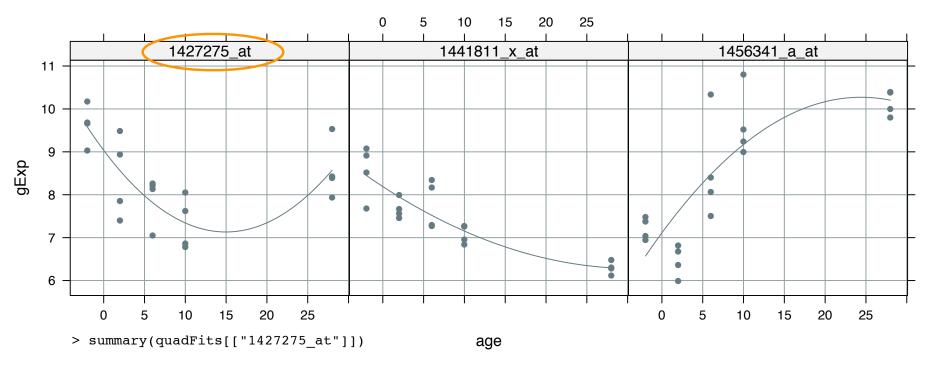
What we just did.



What we're $Y = \alpha_0 + \alpha_1 x + \alpha_2 x^2 + \varepsilon$ about to do.

fairly good fit for 2 of 3 now! 1427275_at 1456341 a at 1441811_x_at 11 -gExp age 1427275_at 1441811_x_at 1456341_a_at 11 -gExp

age



Call:

 $lm(formula = gExp \sim age + I(age^2), data = z)$

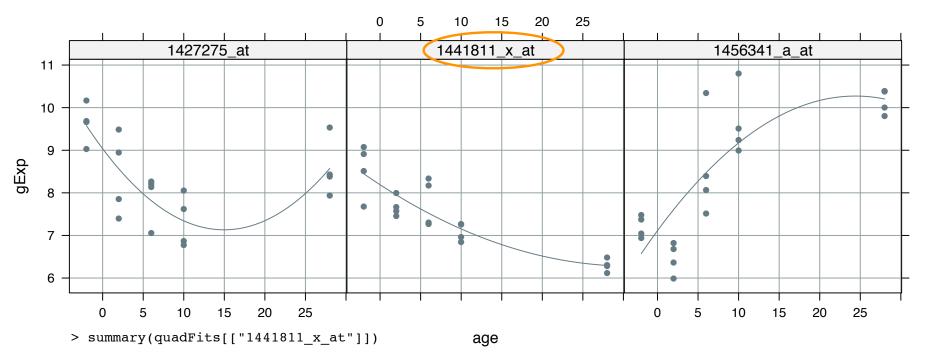
Residuals:

Min 1Q Median 3Q Max -1.16275 -0.55506 0.09503 0.40804 0.95803

Coefficients:

Estimate Std. Error t value Pr(>|t|)
(Intercept) 9.036401 0.212313 42.562 < 2e-16 ***
age -0.254305 0.048125 -5.284 6.07e-05 ***
I(age^2) 0.008490 0.001661 5.110 8.71e-05 ***
--Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.6444 on 17 degrees of freedom Multiple R-squared: 0.6218, Adjusted R-squared: 0.5773 F-statistic: 13.98 on 2 and 17 DF, p-value: 0.0002572



Call:

 $lm(formula = gExp \sim age + I(age^2), data = z)$

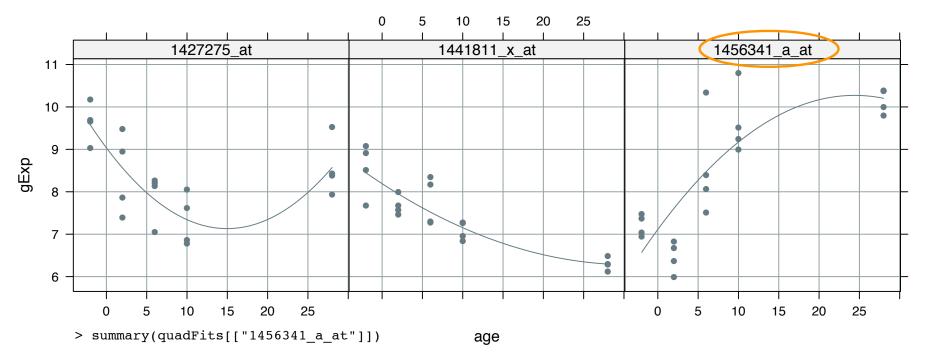
Residuals:

Min 1Q Median 3Q Max -0.76946 -0.25477 -0.00589 0.13662 0.82202

Coefficients:

Estimate Std. Error t value Pr(>|t|)
(Intercept) 8.190766 0.140969 58.103 < 2e-16 ***
age -0.123836 0.031953 -3.876 0.00121 **
I(age^2) 0.002006 0.001103 1.819 0.08660 .
--Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.4279 on 17 degrees of freedom Multiple R-squared: 0.774, Adjusted R-squared: 0.7475 F-statistic: 29.12 on 2 and 17 DF, p-value: 3.23e-06



Call:

 $lm(formula = gExp \sim age + I(age^2), data = z)$

Residuals:

Min 1Q Median 3Q Max -1.6211 -0.5010 -0.0050 0.3955 1.8651

Coefficients:

Estimate Std. Error t value Pr(>|t|)
(Intercept) 7.112481 0.310922 22.875 3.3e-14 ***
age 0.258892 0.070477 3.673 0.00188 **
I(age^2) -0.005303 0.002433 -2.180 0.04363 *
--Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.9437 on 17 degrees of freedom Multiple R-squared: 0.6737, Adjusted R-squared: 0.6353 F-statistic: 17.55 on 2 and 17 DF, p-value: 7.337e-05

F tests in regression

Remember this?

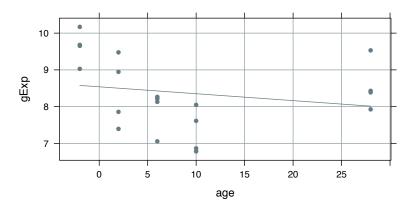
small model is nested within big, e.g., it's a special case where some parameters are equal to zero

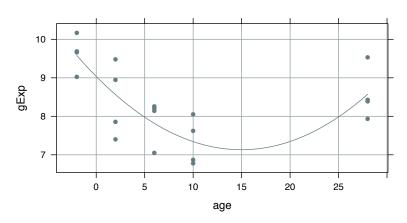
model	example	# params = DF	RSS
small	lm(y ~ gType + devStage)	p _{small} = 6	RSS _{small}
big	Im(y ~ gType * devStage)	Pbig = I0	RSS _{big}

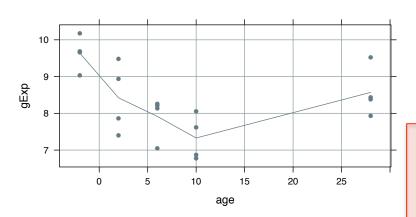
$$\begin{aligned} y_{ijk} &= \theta + \tau_j + \beta_k + (\tau \beta)_{jk} + \varepsilon_{ijk} \text{ "big"} \\ y_{ijk} &= \theta + \tau_j + \beta_k + (\tau \beta)_{jk} + \varepsilon_{ijk} \text{ "small"} \end{aligned}$$

 $p_{small} < p_{big}$ $RSS_{small} \ge RSS_{big}$

$$F = \frac{\begin{pmatrix} RSS_{small} - RSS_{big} \\ \hline p_{big} - p_{small} \end{pmatrix}}{\frac{RSS_{big}}{n - p_{big}}} \sim_{H_0} F_{(p_{big} - p_{small}, n - p_{big})}$$





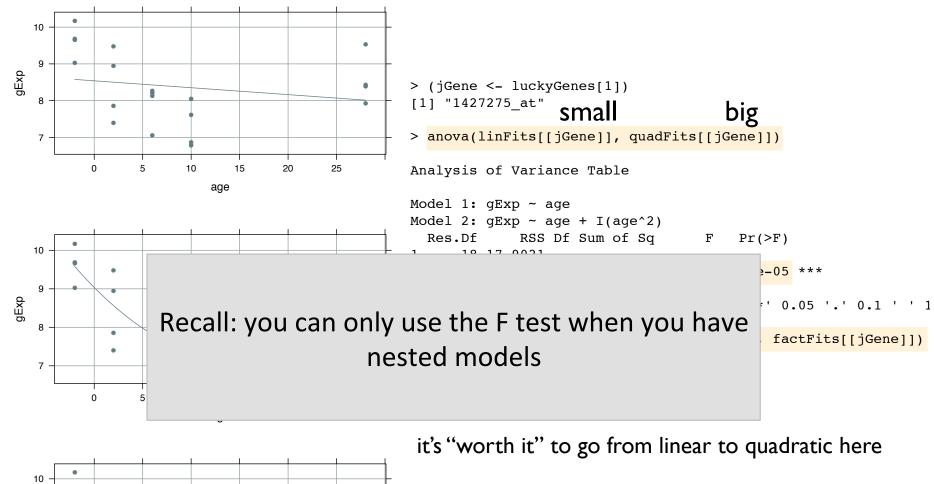


```
> (jGene <- luckyGenes[1])</pre>
[1] "1427275 at"
                                      big
> anova(linFits[[jGene]], quadFits[[jGene]])
Analysis of Variance Table
Model 1: gExp ~ age
Model 2: gExp ~ age + I(age^2)
             RSS Df Sum of Sq
                                        Pr(>F)
      18 17.9021
                       10.843 26.113 8.71e-05 ***
         7.0591 1
                0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Signif. codes:
> AIC(linFits[[jGene]], quadFits[[jGene]], factFits[[jGene]])
                  df
                           AIC
linFits[[jGene]]
                   3 60.54129
quadFits[[jGene]]
                   4 43.92930
factFits[[jGene]]
                   6 47.54810
```

it's "worth it" to go from linear to quadratic here

but hard to justify going from quadratic to oneway ANOVA

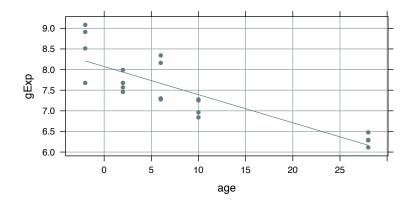
possible links to read more about using AIC to compare non-nested models: stackexchange and Wikipedia

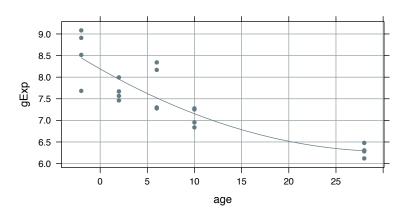


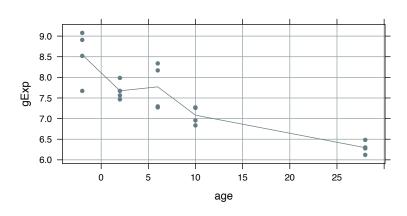
10 9 8 7 0 5 10 15 20 25 age

but hard to justify going from quadratic to oneway ANOVA

possible links to read more about using AIC to compare non-nested models: stackexchange and Wikipedia







```
> (jGene <- luckyGenes[3])</pre>
[1] "1441811 x at"
small big
anova(linFits[[jGene]], quadFits[[jGene]])
Analysis of Variance Table
Model 1: gExp ~ age
Model 2: gExp ~ age + I(age^2)
            RSS Df Sum of Sq
                                   F Pr(>F)
      18 3.7176
      17 3.1120 1
                      0.60559 3.3081 0.0866 .
Signif. codes:
                0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
> AIC(linFits[[jGene]], quadFits[[jGene]], factFits[[jGene]])
                           AIC
linFits[[jGene]]
                    3 29.10466
                                meh
                   4 27.54851
quadFits[[jGene]]
factFits[[jGene]]
                    6 27.12587
```

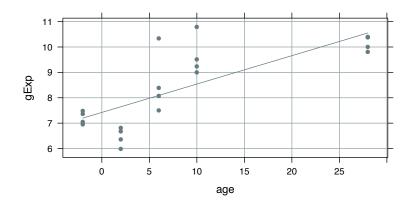
not clear it's "worth it" to go from linear to quadratic here

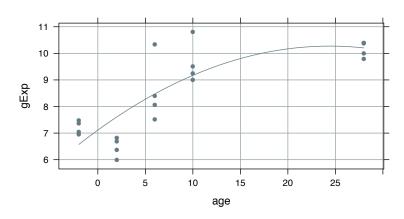
even less payoff to go from quadratic to one-way ANOVA

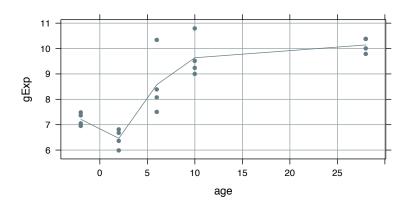
Occam's Razor and the KISS principle → stick w/ simple linear model

Occam's razer

- Principle of parsimony: states that between competing hypotheses, the one with the fewest assumptions.
 - Roughly speaking: do not make things more complicated than needs to be







```
> (jGene <- luckyGenes[2])</pre>
[1] "1456341 a at"
small
> anova(linFits[[jGene]], quadFits[[jGene]])
Analysis of Variance Table
Model 1: gExp ~ age
Model 2: gExp ~ age + I(age^2)
            RSS Df Sum of Sq
                                   F Pr(>F)
      18 19.370
                       4.2308 4.7509 0.04363 *
      17 15.139 1
Signif. codes:
                0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
> AIC(linFits[[jGene]], quadFits[[jGene]], factFits[[jGene]])
                   df
                           AIC
linFits[[jGene]]
                    3 62.11743
quadFits[[jGene]]
                   4 59.18864
factFits[[jGene]]
                    6 48.70210
```

it's probably "worth it" to go from linear to quadratic here (?)

going from quadratic to one-way ANOVA seems justified

Break to talk about projects

Project teams

FISCH

Studying metabolites in hyperphenylalaninemia patients using Mass Spec

GLLAD

Looking at RNA microarray gene expression data in HIV/HCV co-infected population, HCV monoinfected, a healthy comparison group to explain differences in clinical outcome in co-infected and mono-infected patients

GutCHECK

Studying the correlation of microbial composition and metabolic pathways in type 2 diabetes (T2D) or inflammatory bowel disease (IBD) patients of different ethnicity and geographical origin.

IUGRoup

Comparing DNA methylation patterns in intrauterine growth restriction (IUGR) and in healthy placenta

Project teams

JaWSPR

Looking at expression signature in mice with different alcohol tolerance

META-VEG

Analyzing grape berries and their microflora by looking at RNAseq dataset of grapes throughout development

Leptin

Investigating the mechanism of leptin therapy in type 1 diabetes

PHASAX

Characterizing the effect of the immune microenvironment on tumour evolution in high-grade serous ovarian cance

Treed

Looking at DNA methylation in male and female mice

Next steps

- Start thinking about your proposal (outline; specifics; task assignment)
- You'll get assigned a TA and an instructor
- Consult with your assigned TA/instructor when you have a good sketch of the outline/aims/tasks etc

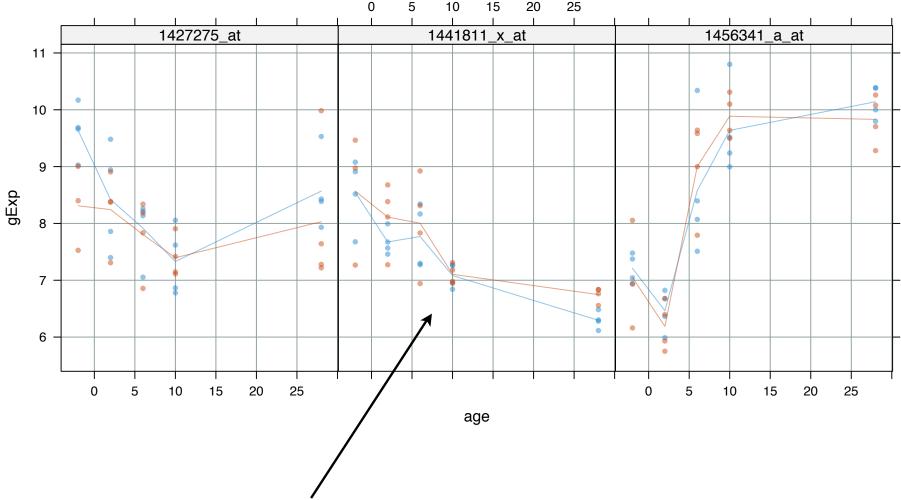
Back to lecture

increase the complexity ...

I quantitative covariate: age

AND I categorical covariate:

genotype = wt vs. Nrl knockout

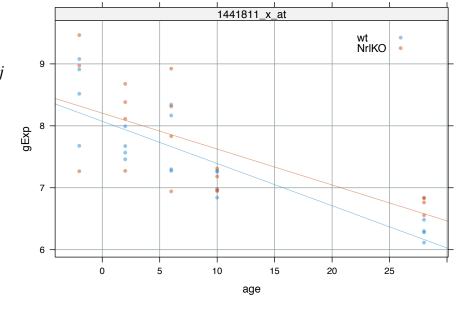


let's focus on this one for a model with just intercept and slope, possibly different for wt and NrIKO

$$y_{ij} = \alpha_{0,wt} + \tau_{0,j} + (\alpha_{1,wt} + \tau_{1,j})age_i + \varepsilon_{ij}$$
where $j \in \{wt, NrlKO\}$

$$i = 1, 2, ..., n_j$$

$$\tau_{0,wt} = \tau_{1,wt} \equiv 0$$



Call:

lm(formula = gExp ~ gType * age, data = jDat)

Residuals:

Min 1Q Median 3Q Max -1.05383 -0.41194 -0.02491 0.31295 1.14417

Coefficients:

Estimate Std. Error t value Pr(>|t|)
(Intercept) 8.07337 0.16552 48.776 < 2e-16 ***

gTypeNrlKO 0.13148 0.24070 0.546 0.588
age -0.06818 0.01215 -5.612 2.51e-06 ***
gTypeNrlKO:age 0.01019 0.01744 0.584 0.563

The intercept for the knockouts is:

$$\alpha_{0,wt}$$
 + $au_{0,\Delta Nrl}$

and the slope for knockouts is:

$$\alpha_{1,wt} + \tau_{1,\Delta Nrl}$$

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.5651 on 35 degrees of freedom Multiple R-squared: 0.607, Adjusted R-squared: 0.5733 F-statistic: 18.02 on 3 and 35 DF, p-value: 3.047e-07

as always, different parametrizations are possible!

$$y_{ij} = \alpha_{0,j} + \alpha_{1,j} age_i + \varepsilon_{ij}$$
where $j \in \{wt, NrlKO\}$

$$i = 1, 2, ..., n_j$$

Call:

lm(formula = gExp ~ gType/age - 1, data = jDat)

Residuals:

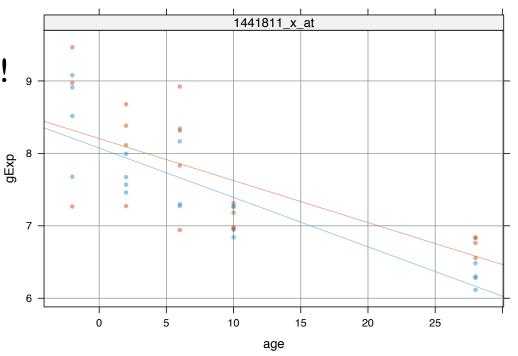
Min 1Q Median 3Q Max -1.05383 -0.41194 -0.02491 0.31295 1.14417

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)	
gTypewt	8.07337	0.16552	48.776	< 2e-16	***
gTypeNrlKO	8.20485	0.17476	46.949	< 2e-16	***
gTypewt:age	-0.06818	0.01215	-5.612	2.51e-06	***
gTypeNrlKO:age	-0.05799	0.01251	-4.636	4.80e-05	***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.5651 on 35 degrees of freedom Multiple R-squared: 0.9951, Adjusted R-squared: 0.9945 F-statistic: 1761 on 4 and 35 DF, p-value: < 2.2e-16



(intercept, slope) for wild type:

$$(\alpha_{0,wt},\alpha_{1,wt})$$

(intercept, slope) for the knockouts:

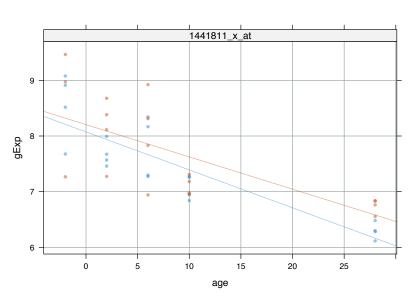
$$(\alpha_{0,\Delta Nrl}, \alpha_{1,\Delta Nrl})$$

as always, you can switch between parametrizations via multiplication by an appropriate contrast matrix!

```
y_{ij} = \alpha_{0,wt} + \tau_{0,i} + (\alpha_{1,wt} + \tau_{1,i})age_i + \varepsilon_{ij}
 where j \in \{wt, NrlKO\}
 i = 1, 2, ..., n_i
 \tau_{0,wt} = \tau_{1.wt} \equiv 0
  (contMat \leftarrow rbind(c(1, 0, 0, 0),
                      c(1, 1, 0, 0),
                       c(0, 0, 1, 0),
                       c(0, 0, 1, 1))
      [,1] [,2] [,3] [,4]
[1,]
[2,]
[3,]
[4,]
> cbind(coefDefault = coef(jFit),
         coefAlt = coef(jFitAlt),
         matrixResult = as.vector(contMat %*% coef(jFit)))
                                   coefAlt matrixResult
                 coefDefault
(Intercept)
                  8.07337352 8.07337352
                                               8.07337352
qTypeNrlKO
                 0.13147574 8.20484926
                                             8.20484926
                 -0.06817881 <mark>-0.06817881</mark>
                                              -0.06817881
age
qTypeNrlKO:age 0.01018928 -0.05798953
                                             -0.05798953
```

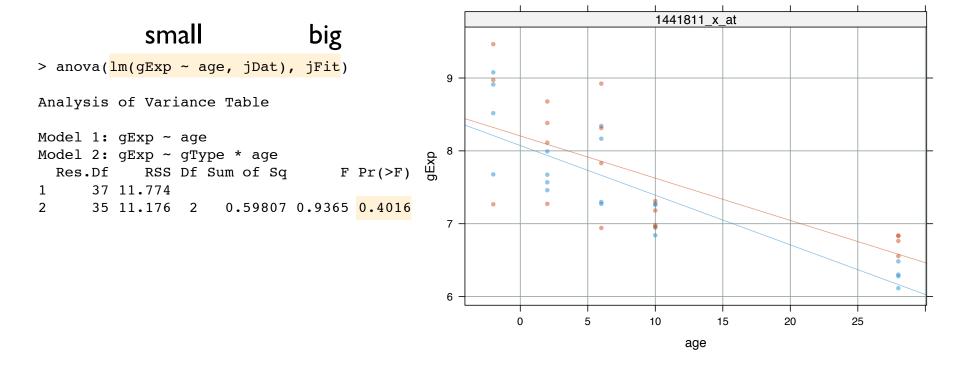
$$y_{ij} = \alpha_{0,j} + \alpha_{1,j} age_i + \varepsilon_{ij}$$
where $j \in \{wt, NrlKO\}$

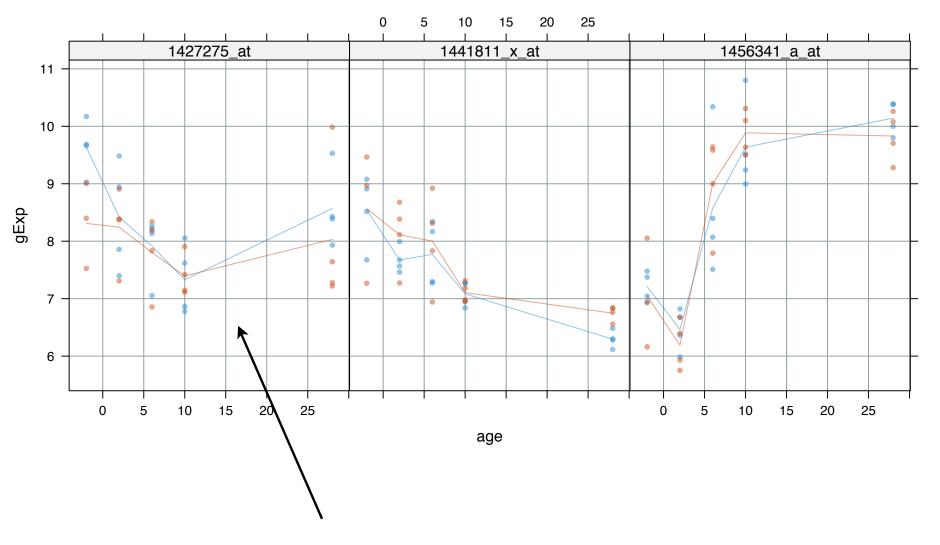
$$i = 1, 2, ..., n_j$$



as always, you can assess the relevance of several terms at once -- such as everything involving genotype -- with an F test

it's not clear that genotype affects the intercept or the slope





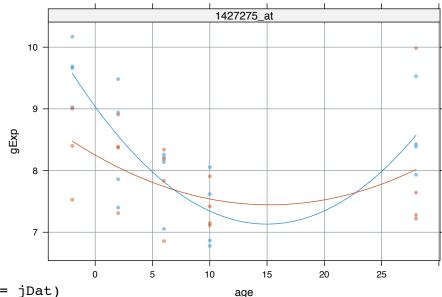
let's focus here for a model including a quadratic age term

$$y_{ij} = \alpha_{0,wt} + \tau_{0,j} + (\alpha_{1,wt} + \tau_{1,j})age_i + (\alpha_{2,wt} + \tau_{2,j})age_i^2 + \varepsilon_{ij}$$

where $j \in \{wt, NrlKO\}$

$$i = 1, 2, ..., n_j$$

$$\tau_{0,wt} = \tau_{1,wt} = \tau_{2,wt} \equiv 0$$



> summary(jFit)

Call:

Residuals:

Min 1Q Median 3Q Max -1.16275 -0.55816 0.08203 0.42020 1.96803

Coefficients:

Estimate Std. Error t value Pr(>|t|) 0.234853 38.477 < 2e-16 *** (Intercept) 9.036401 0.350249 - 2.241qTypeNrlKO -0.784969 0.0319 * 0.053234 -4.777 3.55e-05 *** age -0.254305 $I(age^2)$ 0.008490 0.001838 4.620 5.63e-05 *** 0.078232 1.894 qTypeNrlKO:age 0.0670 . 0.148195 0.002673 - 1.8710.0702 . gTypeNrlKO:I(age^2) -0.005001

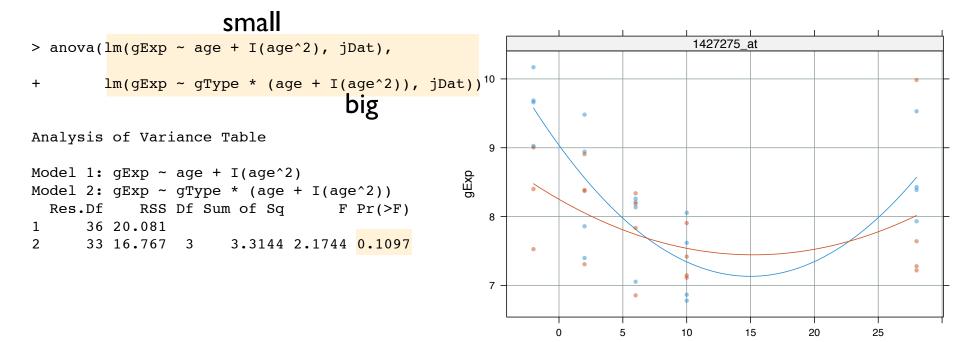
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 0.7128 on 33 degrees of freedom Multiple R-squared: 0.4755, Adjusted R-squared: 0.3961 F-statistic: 5.984 on 5 and 33 DF, p-value: 0.0004804

as always, you can assess the relevance of several terms at once -- such as everything involving genotype -- with an F test

borderline evidence that genotype affects something about the parabola (location or shape)

age



linear model framework is extremely general!

one extreme (simple): two-sample common variance t-test

another extreme (flexible): a polynomial, potentially different for each level of some factor

dichotomous variable? OK! categorical variable? OK! quantitative variable? OK! various combinations of the above? OK!

don't be afraid to build models with more than I covariate don't be intimidated by all the "contrast" talk

$lm(yMat \sim x)$

$$Y = X\alpha + \varepsilon$$

$$\begin{bmatrix} y_{11} & y_{1G} \\ y_{21} & y_{2G} \\ \vdots & \cdots & y_{nG} \end{bmatrix} = X \begin{bmatrix} \alpha_1 & \alpha_1 & \alpha_G \\ \alpha_1 & \cdots & \alpha_G \end{bmatrix} + \begin{bmatrix} \varepsilon_{11} & \varepsilon_{1G} \\ \varepsilon_{21} & \varepsilon_{2G} \\ \vdots & \cdots & \varepsilon_{nG} \end{bmatrix}$$

built-in function lm() can do "multivariate regression" = many dependent vars ("responses") aka "multivariate multiple regression"

From lm() documentation:

If response is a matrix a linear model is fitted separately by least-squares to each column of the matrix.

lm returns an object of class "lm" or for multiple responses of class c("mlm", "lm").

Industrial scale model fitting is good because things like this are not recomputed 30K times unnecessarily*

 $Y = X\alpha + \varepsilon$ regression model

 $\hat{\alpha} = (X^T X)^{-1} X^T Y$ the MLE and OLS estimator of α unnecessarily*

 $\hat{\sigma}^2 = \frac{1}{n-p} \hat{\varepsilon}^T \hat{\varepsilon}$ the estimated error variance

 $\hat{V}(\hat{\alpha}) = \hat{\sigma}^2 (X^T X)^{-1}$ the estimated covariance matrix of $\hat{\alpha}$

How test $H_0: \alpha_i = 0$?

With a t-statistic. Under H_0 , we have (at least approximately) that:

$$\frac{\hat{\alpha}_{j}}{\widehat{se}(\hat{\alpha}_{i})} \sim t_{n-p}$$

so a p-value is obtained by computing a tail probability for the observed value of $\hat{\alpha}_i$ from a t_{n-p} distribution.

* under the hood, lm() is doing something more clever and numerically stable than this

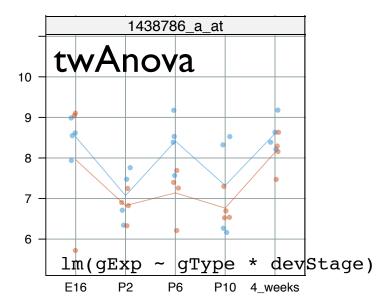
I have fit all the models we've considered to all ~30K probesets.

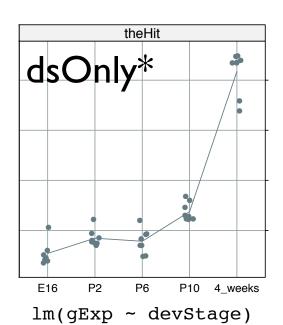
Let's examine some of the results en masse.

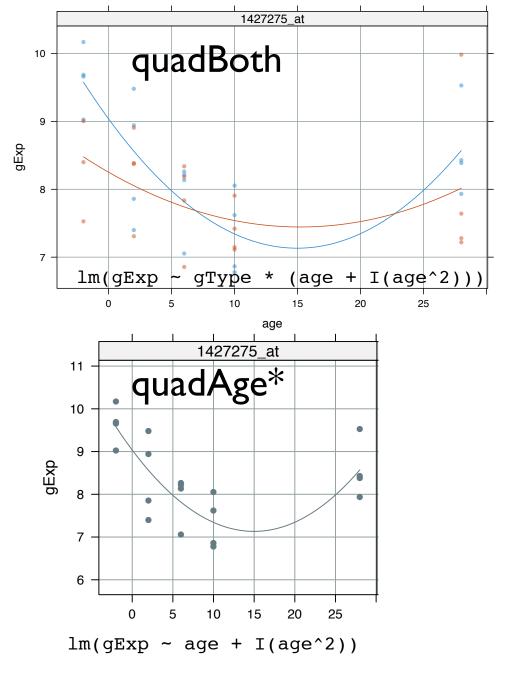
Let this drive home the point that ...

- background variability
- intercepts
- Nrl knockout effects
- devStage effects
- age effects, both linear and quadratic
- •and interactions of all the above

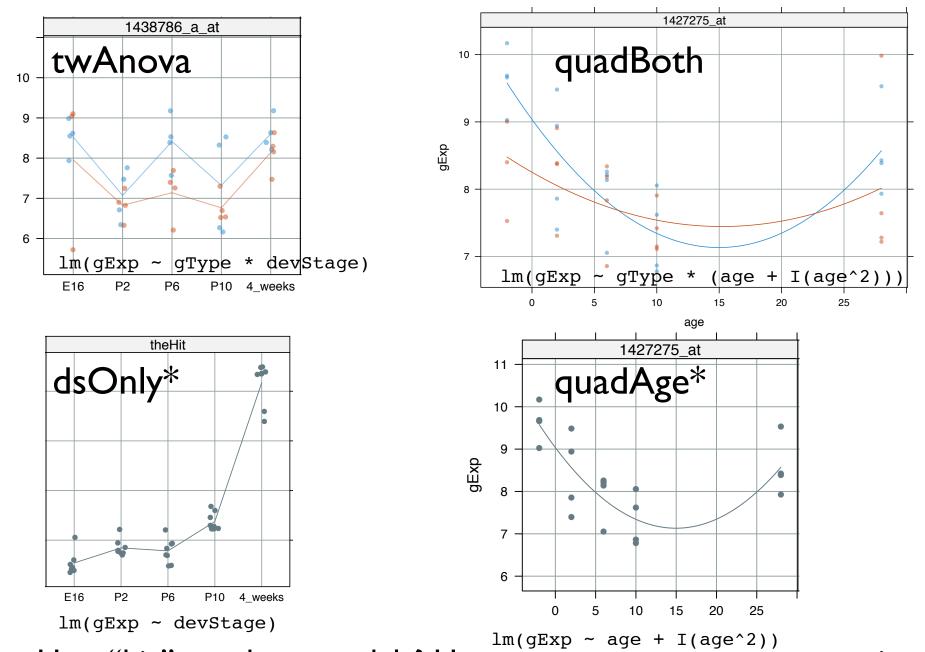
differ for each gene.



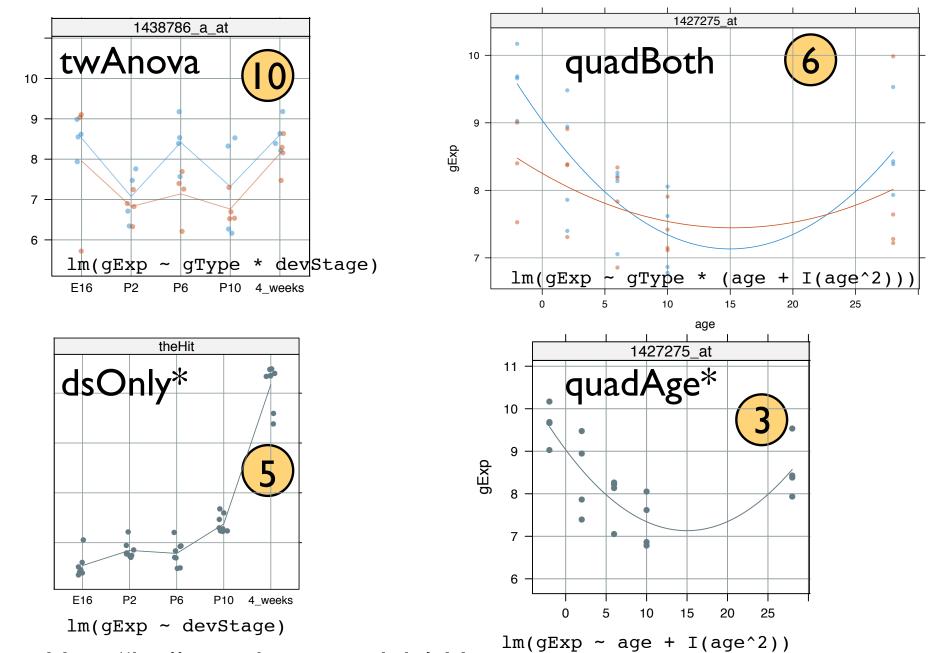




^{*} Figures slightly misleading. Model is fit to all the data, wild type and Nrl knockout, but gType is not used as a covariate.



How "big" are these models? How many parameters are we using to specify the mean structure?

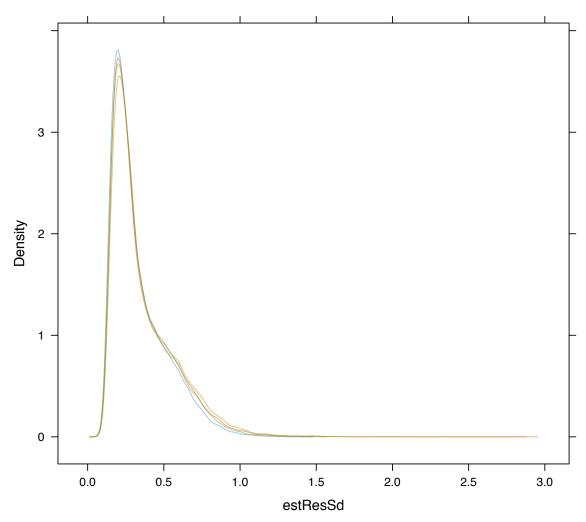


How "big" are these models? How many parameters are we using to specify the mean structure?

$$y_i = f(x_i; \alpha) + \varepsilon_i, \text{var}(\varepsilon) = \sigma^2$$

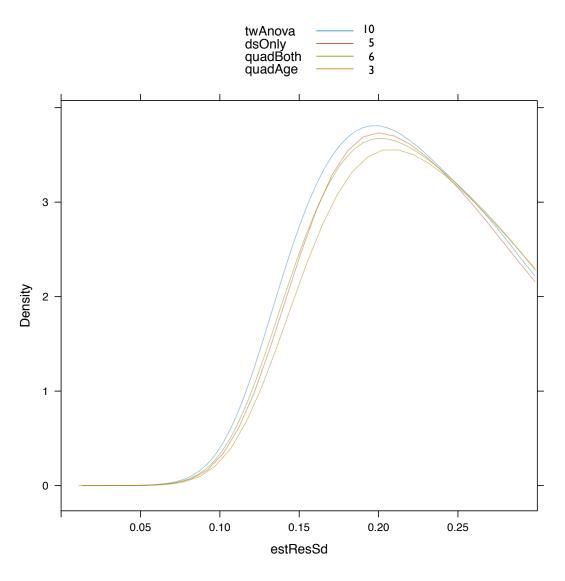
Let's look at estimates of the error standard deviation.





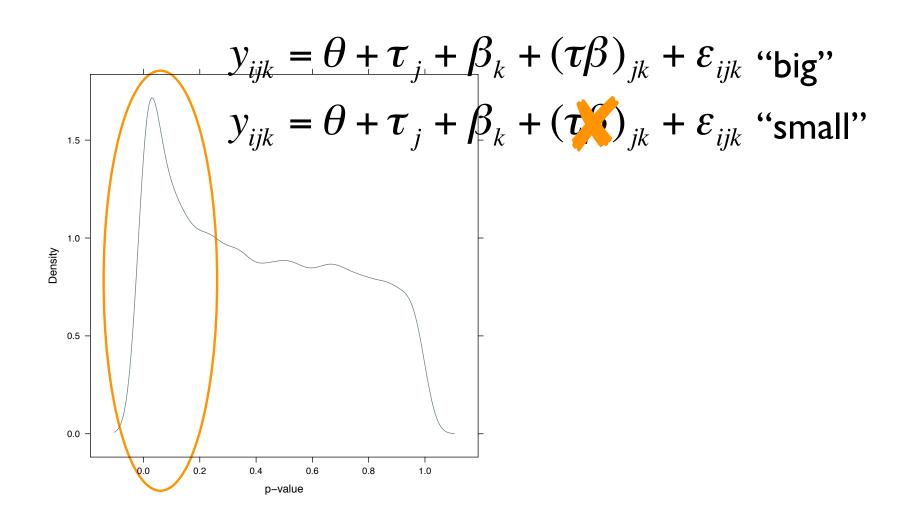
$$y_i = f(x_i; \alpha) + \varepsilon_i, \text{var}(\varepsilon) = \sigma^2$$

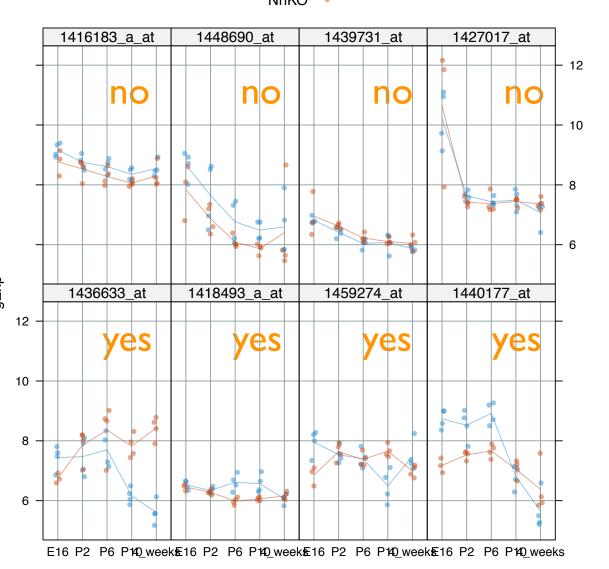
Let's look at estimates of the error standard deviation.

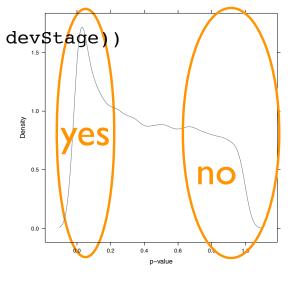


In the two-way ANOVA model, is there evidence for gType * devStage interaction? YES.

this code is fictional but conveys the point
anova(lm(gExp ~ gType * devStage), lm(gExp ~ gType + devStage))
inspecting the p-values from these F tests



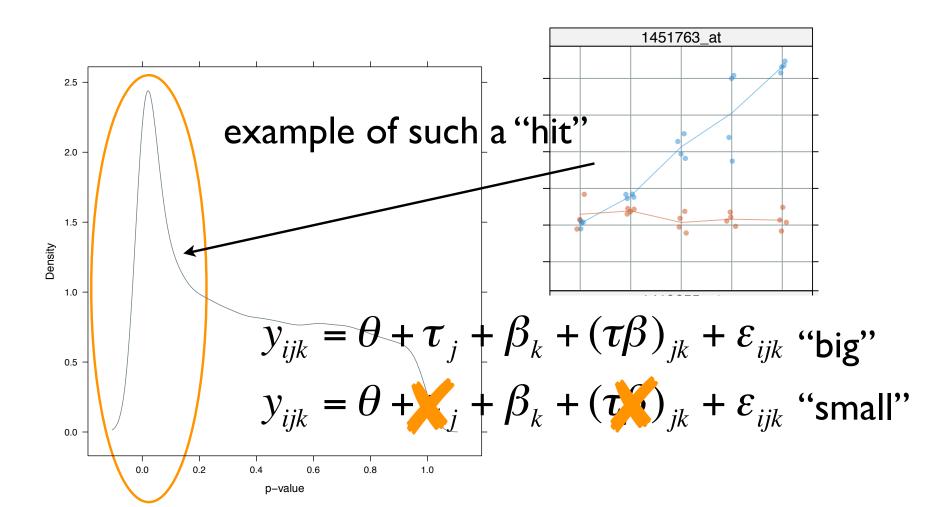




interaction?

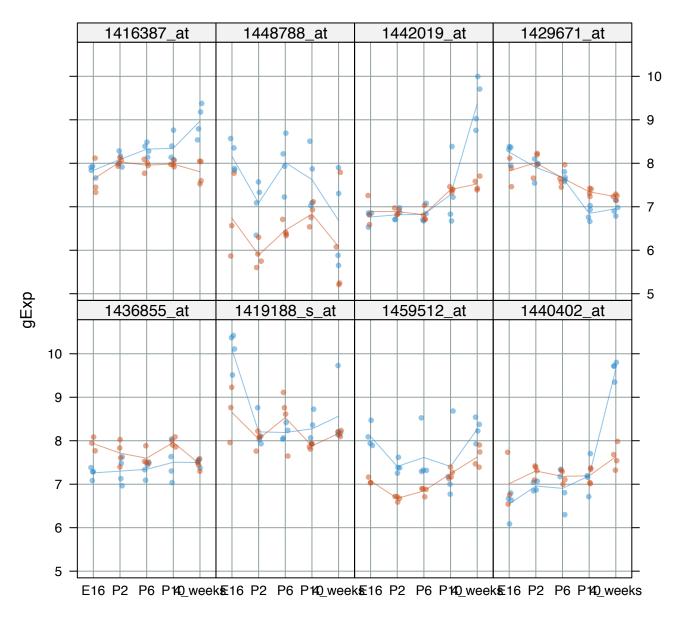
In the two-way ANOVA model, is there evidence that genotype matters? YES.

```
## this code is fictional but conveys the point
anova(lm(gExp ~ gType * devStage), lm(gExp ~ devStage))
## inspecting the p-values from these F tests
```

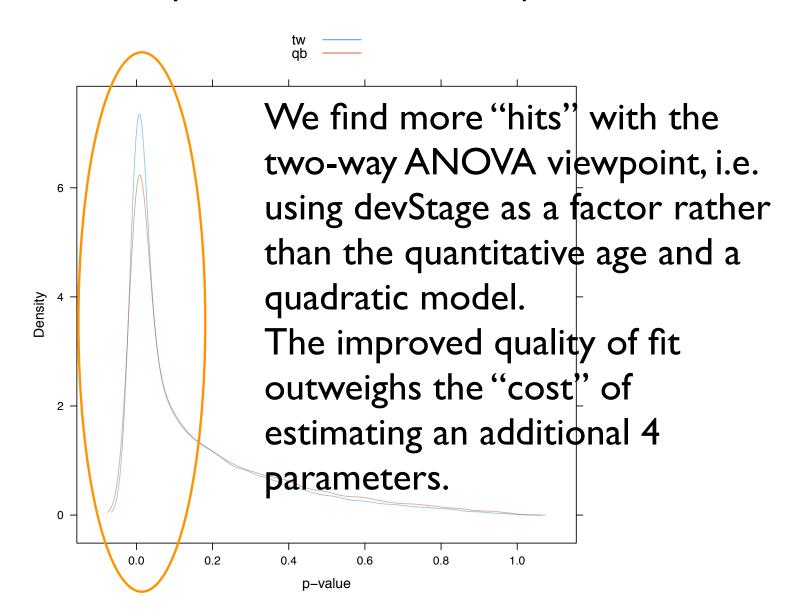


more "gType" hits within the ANOVA models

wt • NrIKO •



Looking at evidence of any differential expression at all (overall F test) in the two-way ANOVA model vs. the quadratic.



where to next? ... Wednesday

in many studies, the # replicates is small relative to # params being estimated

can lead to crazy small estimates of error variance which leads to crazy large test statistics which leads to crazy small p-values which leads to "hits" where the observed phenomenon is rather subtle

which leads to people saying the platform and/or analysis method and/or analyst is bad

moderating the variance estimates can be very helpful --> limma!

where to next? ... following Wednesday

multiple testing, large scale inference

analysis of high-throughput data results in thousands of "genewise" hypothesis tests

often, "gene-wise" analysis is relatively simple

BUT a recurring and thorny issue is how to handle thousands of p-values, each for a separate hypothesis test

how to guard against crazy # false positives?

which error rate is more relevant ... rate at which null genes are 'discovered' or rate at which 'discoveries' are null?