

## Understanding of the impact of chemicals on amphibians: a meta-analytic review

Andrés Egea-Serrano<sup>1</sup>, Rick A. Relyea<sup>2</sup>, Miguel Tejedo<sup>3</sup> & Mar Torralva<sup>1</sup>

<sup>1</sup>Facultad de Biología, Departamento de Zoología y Antropología Física, Universidad de Murcia, 30100 Murcia, Spain

<sup>2</sup>Department of Biological Sciences, University of Pittsburgh, 101 Clapp Hall, Pittsburgh, Pennsylvania 15260

<sup>3</sup>Department of Evolutionary Ecology, Estación Biológica de Doñana, CSIC, Avda. Américo Vespucio s/n, 41092 Sevilla, Spain

### Keywords

Amphibians, ecotoxicology, meta-analysis, phylogenetic signal, publication bias, synergism.

### Correspondence

Andrés Egea-Serrano, Facultad de Biología, Departamento de Zoología y Antropología Física, Universidad de Murcia, 30100 Murcia, Spain. Tel: +34 868 88 4961; Fax: +34 868 88 3963; E-mail: aegea@um.es

This research was partially funded by project (CGL2004-01872/BOS and CGL2009-12767-C02-02) from Dirección General de Investigación Científica y Técnica (MICINN), conceded to Miguel Tejedo. Andrés Egea-Serrano was supported by a Caja de Ahorros del Mediterráneo predoctoral fellowship. Rick Relyea has been supported by the National Science Foundation.

Received: 22 February 2012; Accepted: 29 February 2012

**Ecology and Evolution 2012; 2(7): 1382–1397**

doi: 10.1002/ece3.249

### Abstract

Many studies have assessed the impact of different pollutants on amphibians across a variety of experimental venues (laboratory, mesocosm, and enclosure conditions). Past reviews, using vote-counting methods, have described pollution as one of the major threats faced by amphibians. However, vote-counting methods lack strong statistical power, do not permit one to determine the magnitudes of effects, and do not compare responses among predefined groups. To address these challenges, we conducted a meta-analysis of experimental studies that measured the effects of different chemical pollutants (nitrogenous and phosphorous compounds, pesticides, road deicers, heavy metals, and other wastewater contaminants) at environmentally relevant concentrations on amphibian survival, mass, time to hatching, time to metamorphosis, and frequency of abnormalities. The overall effect size of pollutant exposure was a medium decrease in amphibian survival and mass and a large increase in abnormality frequency. This translates to a 14.3% decrease in survival, a 7.5% decrease in mass, and a 535% increase in abnormality frequency across all studies. In contrast, we found no overall effect of pollutants on time to hatching and time to metamorphosis. We also found that effect sizes differed among experimental venues and among types of pollutants, but we only detected weak differences among amphibian families. These results suggest that variation in sensitivity to contaminants is generally independent of phylogeny. Some publication bias (i.e., selective reporting) was detected, but only for mass and the interaction effect size among stressors. We conclude that the overall impact of pollution on amphibians is moderately to largely negative. This implies that pollutants at environmentally relevant concentrations pose an important threat to amphibians and may play a role in their present global decline.

### Introduction

The negative impact of anthropogenic activities on biodiversity is becoming increasingly conspicuous and amphibians are currently the most globally threatened group of vertebrates (approximately 41% of all species [Hoffmann et al. 2010]). Emergent diseases, habitat destruction, introduction of exotic species, and the pollution of both terrestrial and aquatic habitats have all been described as important threatening factors (Stuart et al. 2004; Wake and Vredenburg 2008). Given these identified threats, it is critical that we determine the influence and magnitude of their effects on

amphibian populations to help develop proper management and conservation strategies.

A variety of pollutants occur in natural habitats including fertilizers, pesticides, heavy metals, and road deicers. This broad array of pollutants is increasingly introduced into the environment by direct application, runoff from crop and forest applications or mines, urban and industrial sewage, and atmospheric deposition (Vitousek et al. 1997; Linder and Grillitsch 2000; Sparling 2000; Ritter and Bergstrom 2001). In short, the presence of pollutants is widespread (Carpenter et al. 1998; Kolpin et al. 2002; Gilliom et al. 2007) and is expected to increase in the near future (Tilman

*et al.* 2001; Galloway *et al.* 2003). What we lack is an overall assessment of how different types of pollutants affect amphibians.

Documented effects of pollutants on amphibians range from lethal effects to sublethal effects including decreased growth and development and increased developmental abnormality frequency, susceptibility to diseases, and behavioral alterations (e.g., Bridges 1999; Ortiz *et al.* 2004; Relyea 2005; Griffis-Kyle 2007; Karraker *et al.* 2008; Shinn *et al.* 2008; Snodgrass *et al.* 2008; Relyea 2009). Due to the great diversity of pollutants and their modes of action, it is not surprising that they affect amphibians differentially. Additionally, several factors including pathogenic organisms and ultraviolet-B radiation are increasingly common in natural environments (Daszak *et al.* 2001; McKenzie *et al.* 2003) and these stressors can interact with chemical pollutants (e.g., Hatch and Blaustein 2000; Romansic *et al.* 2006; Macías *et al.* 2007). As a result, analyzing patterns in how these stressors interact with pollutants is of great relevance when considering the effects of pollution on amphibian populations (Hayes *et al.* 2010).

To date, a large proportion of studies examining the effects of pollutants on amphibians has been performed under laboratory conditions whereas fewer have been conducted under more natural conditions, such as outdoor mesocosms (Boone and James 2005). Although laboratory studies may use ecologically relevant concentrations, outcomes observed under such conditions may not be applicable to more natural conditions (Boone and Bridges 2003) because actual concentrations in the environment can be affected by several factors including plant uptake, denitrification, and sediment trapping (e.g., Ritter and Bergstrom 2001), or because the aforementioned interaction among stressors may mask the effect of a given chemical. As a result, it is possible that laboratory studies overestimate or underestimate the impact of chemical pollutants on amphibians (Boone and Bridges 2003; Gómez-Mestre and Tejedo 2003). This emphasizes the need to compare the effects of pollutants on amphibians across experimental venues (Skelly 2002, but see Chalcraft *et al.* 2005).

It is also reasonable to hypothesize that there may be important species- or family-level differences in sensitivity, although studies to date have been equivocal on this point (Marco *et al.* 1999; Bridges and Semlitsch 2000; Shinn *et al.* 2008; Snodgrass *et al.* 2008; Jones *et al.* 2009). What is needed is an examination of pollutant effects within a phylogenetic framework. The impact of pollutants may also vary with the developmental stage at which individuals are initially exposed (Bridges 2000; Greulich and Pflugmacher 2003; Griffis-Kyle 2005; Ortiz-Santaliestra *et al.* 2006). Collectively, it is clear that to fully understand the effects of pollutants on amphibians, we need to consider a wide variety of factors including type of pollutant, the presence of additional stressors, exper-

imental venue, phylogenetic relationships, and ontogenetic stage.

Several reviews have been published examining the effects of different pollutants on amphibians (Cowman and Mazanti 2000; Linder and Grillitsch 2000; Sparling 2000; Camargo *et al.* 2005; Relyea and Hoverman 2006; Marco and Ortiz-Santaliestra 2009; Kerby *et al.* 2010). These reviews have taken the approach of summarizing studies by the method of vote counting (counting the number of significant vs. non-significant outcomes) or by simply summarizing LC<sub>50</sub> values studies of pollutants on amphibian survival (where LC<sub>50</sub> is the lethal concentration that kills 50% of a population). In LC<sub>50</sub> reviews, the summarized studies are largely restricted to single-species lab studies (i.e., individuals extracted from their natural environment) and survival is the only response variable. Moreover, since the analysis of LC<sub>50</sub> values likely bias the actual impact of chemicals on amphibians in natural environments (LC<sub>50</sub> values often are much higher than actual concentrations in the field; e.g., Egea-Serrano *et al.* 2009a), more information on the impact of ecologically relevant concentrations on survival and sublethal endpoints is essential to determine properly the sensitivity of amphibians to chemical pollution. In vote-counting studies, there are always concerns that the conclusions obtained may not be correct (e.g., due to low sample size) and that the estimates may be highly biased since they have poor statistical power (Rosenberg *et al.* 2000). In addition, vote counting does not provide a reliable way to determine the magnitude of the effect and compare responses among predefined groups (Gurevitch *et al.* 2000).

An alternative methodology to averaging LC<sub>50</sub> studies or using vote counting is the use of meta-analytic techniques. Meta-analytic techniques incorporate the magnitude of effects and the sample size of each study to derive test statistics of overall effect sizes. Meta-analyses can also compare effect sizes among predefined groups, including phylogenetic groups, types of environmental manipulations, and experimental venues. These techniques can also be used to examine 2 × 2 factorial manipulations (Gurevitch *et al.* 2000). For example, meta-analyses have recently been used to examine the overall effect of ultraviolet-B radiation on amphibians and other aquatic organisms, as well as its interaction with other environmental factors (Bancroft *et al.* 2007, 2008).

Using meta-analytic techniques, the objectives of our study were the following: (1) to determine the overall effect of environmentally relevant concentrations of some chemical pollutants on amphibian survival, mass, developmental time, and abnormality frequency; (2) to assess the interactive effects of pollutants and other stressors on amphibians; (3) and to determine whether there are significant differences in pollution effects among predefined groups (i.e., amphibian lineages, experimental venues, developmental stages, and types of pollutants) for the abovementioned response variables.

## Material and Methods

### Data collection

We used three methods to identify the studies to include in the meta-analysis. First, we searched four electronic databases (ISI Web of Science, BIOSIS Previews, ScienceDirect, Scirus) using 10 search words (fertilizers, pesticides, heavy metals, wastewater contaminants, deicers, polychlorinated biphenyls, hydrocarbons, dioxins, furans, and estrogens) for dates prior to December 2008. These search words are considered to include the major types of chemicals affecting amphibians (e.g., Sparling et al. 2000). Second, we examined the citations from the studies dealing with amphibians resulting from the abovementioned search, especially from two recent reviews (Camargo et al. 2005; Marco and Ortiz-Santaliestra 2009). From all of the studies obtained, we only included data in the meta-analysis if they met the following criteria: (1) the studies reported data on amphibian survival at the end of the experiments, time to hatching, time to metamorphosis, mass at the end of the experiments, or abnormality frequency at the end of the experiments; (2) the studies stated that concentrations used in the experiments were ecologically relevant (i.e., found in natural water bodies, regardless of whether they are unusual or typical concentrations, as reported by the authors of the original studies in their publications or in personal communications); (3) the studies clearly specified how many days the experiments lasted; (4) the studies provided means, sample sizes, and measures of variance (i.e., standard deviation, SD, or standard error, SE) for both a control group (i.e., not exposed to contaminant) and an experimental group (i.e., exposed to contaminant); (5) the studies reported the effect for a pollutant in isolation and not in combination with other factors (e.g., a pollutant combined with resource competition, predators, etc.); (6) in the case that a published study reported data for more than one species, population, pollutant, or pollutant concentration, each outcome was considered to be independent in the meta-analyses. When a given study showed the effects of both realistic and nonrealistic concentrations, data were filtered to exclude the latter ones. Therefore, all concentrations considered in the present study were ecologically relevant. Since studies dealing with polychlorinated biphenyls, hydrocarbons, dioxins, furans, and estrogens, and meeting all the abovementioned criteria were not found, the meta-analysis could not be performed on these types of chemicals.

For each study, we obtained the mean, SD, and sample size ( $n$ ) for both the control and the experimental group. When means and measures of variance were presented graphically, we digitized the graph to estimate the values (ImageProPlus version 4.5.0.29 for Windows). If SEs were reported, these values were transformed into SD according to the equation:

$SD = SE \cdot \sqrt{n}$ . For those studies that did not clearly include the required data, we attempted to contact authors to obtain the data. In addition, for each study we also compiled information regarding family, developmental stage, experimental venue, and type of pollutant.

To assess the effect of pollutants combined with other stressors (both biotic and abiotic), we conducted a factorial meta-analysis (Gurevitch et al. 2000). The factorial meta-analysis examines the magnitude of the effect of the two main factors and their interaction. Data used in the factorial meta-analysis came from publications meeting the above criteria and also showing a  $2 \times 2$  factorial structure (Gurevitch et al. 2000). The original objective was to examine the effect of the interaction between pollutants and other stressors that eventually could include a second pollutant. Consequently, we used a *first group of stressing factors* (STRESS-1) (nitrogenous compounds, pesticides, and wastewater pollutants) and a *second group of stressing factors* (STRESS-2) (competitors, pH, predators [either caged or uncaged], ultraviolet radiation, other wastewater pollutants, and mold). Other types of factors or combinations among chemicals were not included in the analysis because of the scarcity of studies meeting all the selection criteria. When for a given study both groups of stressing factors involved wastewater pollutants, each pollutant was arbitrarily assigned only to either STRESS-1 or STRESS-2. Therefore, each comparison was included only once in the calculation of effect size. Due to the scarcity of experiments addressing the impact of the interaction among pollutants and other stressors for most response variables, the factorial meta-analysis could only be performed on amphibian survival.

### Data analysis

#### Meta-analysis

For all studies reporting the impact of a control treatment (pollutant absent) and at least one polluted treatment (pollutant present) in the absence of other stressors, we used Hedge's  $d^+$  as the metric of standardized effect size. Hedge's  $d^+$  provides a measure of the overall magnitude of the treatment effect while adjusting for small sample sizes (Rosenberg et al. 2000). Since the absence of pollutant was considered as control, negative effect sizes would indicate reduced survival, mass, time to hatching, time to metamorphosis, or abnormality frequency related to pollutant exposure. To calculate Hedge's  $d^+$  for each study, mean, SD, and  $n$  for both the control and the experimental group were used after dividing mean and SD by the number of days that the corresponding experiments lasted. This approach allowed us to correct for differences among studies in the duration of exposure. Since for most studies and traits data were reported only for the end of the experiment, a linear evolution of the effect of

pollutants over time had to be assumed. This transformation was applied to all studied traits, except for time to hatching and time to metamorphosis, which explicitly consider duration of exposure. Since the results obtained after conducting the analyses described below on corrected and uncorrected Hedge's  $d^+$  were qualitatively similar, results for uncorrected data are not reported in the present study.

For each trait, data were analyzed using categorical random effects models to calculate the grand mean effect size. Additionally, any difference among predefined groups was analyzed using mixed-effects models. Such groups included: (1) amphibian family; (2) developmental stage at the time of pollutant application (embryonic, larval, or postmetamorphic individuals); (3) experimental venue (laboratory, mesocosm, or field enclosures, as well whether the animals were collected in the field [i.e., field experiments]); and (4) type of pollutant (nitrogenous compounds, phosphorous compounds, pesticides, road deicers, heavy metals, and other wastewater contaminants [i.e., perchlorate, boron, acetaminophen, caffeine, and triclosan]). When mixed-effects models were conducted, we calculated mean effect sizes and 95% confidence limits for each class. Additionally, heterogeneity statistics were calculated to quantify between-group ( $Q_B$ ) and within-group ( $Q_W$ ) variation. Effect sizes were considered significant if the 95% confidence intervals did not cross zero. The magnitude of the overall effect size is generally interpreted as "small" if  $d^+ = 0.2$ , "medium" if  $d^+ = 0.5$  and "large" if  $d^+ \geq 0.8$  (Cohen 1969). Effect sizes within analyses (e.g., nitrogenous compounds effect vs. pesticides effect) were considered different from one another if their 95% confidence intervals did not overlap. All statistical analyses were performed using MetaWin 2.1 statistical program (Rosenberg *et al.* 2000). This program eliminates groups with fewer than two valid cases from the analyses when running mixed-effects models. Therefore, mean and 95% confidence interval could not be reported for some of the categories included in the categorical random effect models performed to calculate grand mean effect sizes.

### Factorial meta-analysis

As in the previous analyses, we calculated Hedge's  $d^+$  standardized effect size and its corresponding sampling variance following Gurevitch *et al.* (2000). For each study, the following effect sizes were estimated: (1) the average effect of the exposure to STRESS-1; (2) the average effect of the exposure to STRESS-2; (3) their interaction (STRESS-1  $\times$  STRESS-2).

### Phylogenetic comparative analysis

To determine whether effects of pollutants were significantly associated with amphibian phylogeny, we conducted tests for serial independence (TFSI) on continuous characters (Abouheif 1999) for each response variable, for both simple

and factorial meta-analyses. The diagnosis is based on a measurement of the autocorrelation of a trait across phylogeny, in the form of a  $C$ -statistic, resulting from similarity between adjacent phylogenetic observations. The topology and associated numerator distribution were randomized 2000 times and the  $C$ -statistic was calculated for each randomized topology to build the null hypothesis. The observed  $C$ -statistic was compared to the randomized distribution to calculate its level of significance. Significant phylogenetic autocorrelation was defined when the observed  $C$ -statistic falls to the right of the distribution of the randomized  $C$ -statistics and  $P \leq 5\%$  (Abouheif 1999). Such a result would imply that related species show similar responses regardless of the effect of pollutants.

To conduct the phylogenetic analyses, a topology was constructed following Frost *et al.* (2006), combined with additional detail from several family phylogenetic assessments: Salamandridae (Zajc and Arntzen 2000; Weisrock *et al.* 2006; Zhang *et al.* 2008), Ambystomatidae (Shaffer *et al.* 1991; Jones *et al.* 1993), Pelobatidae and Pelodytidae (García-París *et al.* 2003), Myobatrachidae (Schäuble *et al.* 2000; Read *et al.* 2001), Hylidae (Faivovich *et al.* 2005), Bufonidae (Pauly *et al.* 2004), and Ranidae (Veith *et al.* 2003; Hillis and Wilcox 2005; Scott 2005). Branch length information was not available for our composite phylogenies, but all analyses performed can be conducted using only topology without knowledge of branch lengths. Before conducting the phylogenetic analyses, we assigned the same value to all branch lengths and a single effect size was calculated for each species. The phylogenetic comparative analyses were conducted after replacing the names of the different species at the tip of each branch in the topology constructed by the corresponding single effect sizes. Such effect sizes were the tip data (Abouheif 1999) to which TFSI were applied.

### Publication bias

To assess whether publication bias existed for the datasets used in our study (defined as the selective publication of studies reporting certain types of results over those showing other results; Begg 1994), different approaches were used. For each response variable, for both simple and factorial meta-analyses, we calculated Rosenberg's fail-safe number using a fail-safe number calculator that is applicable to random-effect models (Rosenberg 2005; <http://www.public.asu.edu/~mrosenb/software.html#failsafe>). Rosenberg's fail-safe number is the number of nonsignificant, unpublished, or missing studies that would need to be added to a meta-analysis to change the results from significant to nonsignificant (Rosenberg 2005). The results obtained are considered robust when the fail-safe number is larger than  $5n + 10$  (where  $n$  = the number of studies; Rosenberg 2005). Additionally, we used the funnel plot technique and Spearman's

rank correlation to determine the relationships between effect size and sample size for each response variable. If selective reporting is absent, plots of effect size against sample size should fit a funnel pattern and Spearman's rank correlations should not be significant. All three procedures were used to test for publication bias (Begg and Mazumdar 1994; Palmer 1999).

## Results

Among the published studies examined that met our data selection criteria, only 48 were suitable (Table S1). However, not all studies provided data for all response variables. As a result, survival and mass had more data available (survival: 35 studies, 256 point samples; mass: 19 studies, 181 point samples) than time to hatching (three studies, 23 point samples), time to metamorphosis (nine studies, 37 point samples), and abnormality frequency (four studies, 33 point samples; Table S1). For the factorial meta-analysis of survival, only 12 published studies met our data criteria and these 12 studies provided 45 datapoints (Table S1).

## Meta-analysis

### Survival

Survival was significantly heterogeneous (i.e., not all effect sizes were equal) (Table 1). Across all studies, pollutants had a medium ( $0.5 < d < 0.8$ ) negative effect on survival (Fig. 1), which translated to a 14.3% ( $\pm 1.7$  SE) decrease in survival. Ambystomatidae, Bufonidae, Pipidae, and Ranidae

families showed significantly reduced survival (Fig. 2), but we found no significant differences either among families or developmental stages. Conversely, significant differences in survival were detected for experimental venue. Although survival was reduced in laboratory and enclosure conditions, the decline in laboratory studies was of medium magnitude whereas the decline in enclosure studies was large. The type of pollutant also mattered; road deicers were more lethal than nitrogenous compounds whereas pesticides and wastewater pollutants caused an intermediate reduction in survival. Heavy metals and phosphorous compounds did not affect survival.

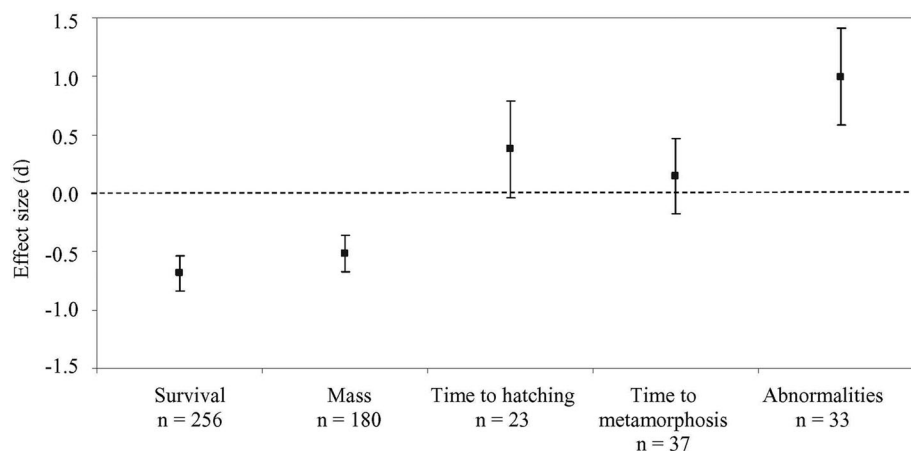
### Mass

Significant heterogeneity was detected for mass (Table 1). This response variable was significantly affected by pollutants, with an overall negative effect that was medium ( $d \sim 0.5$ ) in size (7.5% mean decrease [ $\pm 1.8$  SE] across all studies) (Fig. 1). A significant family effect was also revealed. Hylidae, Pipidae, and Ranidae exhibited significantly reduced mass, with Pipidae having a smaller effect size than the other two families. Nevertheless, there were not significant pairwise differences between families, as revealed by the overlap of the confidence intervals (Fig. 2). Significant differences were also found among developmental stage, experimental venue, and type of pollutant. Mass was lower when the exposure to pollution occurred at embryonic stages, when experiments were conducted under laboratory conditions, and when the pollutant consisted of nitrogenous compounds.

**Table 1.** Heterogeneity statistics for each model in the survival, mass, time to hatching, time to metamorphosis, and abnormality frequency analyses. NA, not applicable; df, degrees of freedom; BG, between groups (referring to the variation in effect size explained by the model,  $Q_B$ ). For clarity, the residual error heterogeneity ( $Q_W$ ) corresponding to the different statistical models is not shown. With the exception of time to hatching and time to metamorphosis (for all the models) and abnormality frequency (only for family and pollutant models), the residual error heterogeneity was significant, which implies that there is still heterogeneity among effect sizes not explained by the model (Rosenberg *et al.* 2000).

Statistical model	Survival ( $n = 270$ )			Mass ( $n = 187$ )			Time to hatching ( $n = 23$ )		
	df	Q	P	df	Q	P	df	Q	P
Full model (no structure)	<b>255</b>	<b>445.042</b>	<b>0.000001</b>	<b>179</b>	<b>215.491</b>	<b>0.032</b>	22	22.427	0.435
Family <sup>BG</sup>	7	6.222	0.514	<b>4</b>	<b>10.002</b>	<b>0.040</b>	<b>1</b>	<b>17.747</b>	<b>0.00003</b>
Developmental stage <sup>BG</sup>	1	0.812	0.368	<b>1</b>	<b>54.294</b>	<b>0.000001</b>	NA	NA	NA
Experimental venue <sup>BG</sup>	<b>2</b>	<b>27.988</b>	<b>0.000001</b>	<b>2</b>	<b>12.516</b>	<b>0.002</b>	NA	NA	NA
Pollutant <sup>BG</sup>	<b>5</b>	<b>15.078</b>	<b>0.010</b>	<b>3</b>	<b>16.843</b>	<b>0.0008</b>	1	0.423	0.515
Statistical model	Time to metamorphosis ( $n = 37$ )			Abnormality frequency ( $n = 39$ )					
	df	Q	P	df	Q	P			
Full model (no structure)	36	42.054	0.225	<b>32</b>	<b>50.754</b>	<b>0.019</b>			
Family <sup>BG</sup>	<b>3</b>	<b>9.004</b>	<b>0.029</b>	<b>6</b>	<b>15.405</b>	<b>0.017</b>			
Developmental stage <sup>BG</sup>	<b>1</b>	<b>4.430</b>	<b>0.035</b>	1	2.184	0.139			
Experimental venue <sup>BG</sup>	1	0.793	0.373	NA	NA	NA			
Pollutant <sup>BG</sup>	<b>3</b>	<b>8.989</b>	<b>0.029</b>	<b>2</b>	<b>20.928</b>	<b>0.00003</b>			





**Figure 1.** Effect size (mean and 95% confidence interval) for full models for the effect of pollutants on amphibian survival, mass, time to hatching, time to metamorphosis, and abnormality frequency. The number of point samples used to calculate each mean is shown. Means with confidence intervals that overlap the line at zero are not significantly different from zero.

### Developmental time

Although both time to hatching (2.3% mean increase [ $\pm 3.0$  SE]) and time to metamorphosis (3.5% mean increase [ $\pm 3.9$  SE]) tended to increase across all studies, the overall analysis of these response variables revealed a lack of both significant heterogeneity and effect (Table 1; Fig. 1). However, we did detect significant differences among families in their time to hatching. Only Ambystomatidae exhibited a significant delay in hatching time when exposed to pollutants (Fig. 2). No significant differences in either developmental stage were detected for the other predefined groups (Fig. 2).

### Abnormality frequency

The overall abnormality frequency was significantly heterogeneous and increased when amphibians were exposed to pollutants, which had a large effect size ( $d > 0.8$ ; Table 1; Fig. 1). This translated to a 535% mean increase ( $\pm 287.7$  SE) in abnormality frequency across all studies. Among the predefined groups, the only significant differences were among families and pollution types. Increased frequency of abnormalities was observed in the families Ambystomatidae and Ranidae, although the increase observed in these two families was not significantly different from the other families (Fig. 2). Abnormality frequency was affected most strongly by wastewater pollutants and, to a lesser extent, nitrogenous compounds. Though no significant developmental stage effect was detected, increased abnormality frequency was observed when pollutants were applied during embryonic stages.

### Factorial meta-analysis

In the factorial meta-analysis, which was only conducted on amphibian survival, we detected significant heterogeneity

for STRESS-1 and STRESS-2 effect sizes ( $P < 0.0002$  in all cases), but not for the interaction term STRESS-1  $\times$  STRESS-2 ( $Q_{44} = 45.45$ ;  $P = 0.41$ ). The average overall effect on survival of the exposure to the first group of stressors (nitrogenous compounds, pesticides, and wastewater pollutants; STRESS-1) was significantly weaker than the effect of the second group of stressors (competitors, pH, predators, ultraviolet radiation, other wastewater pollutants, and mold; STRESS-2; Fig. 3). The overall interaction effect size did not differ from zero (Fig. 3).

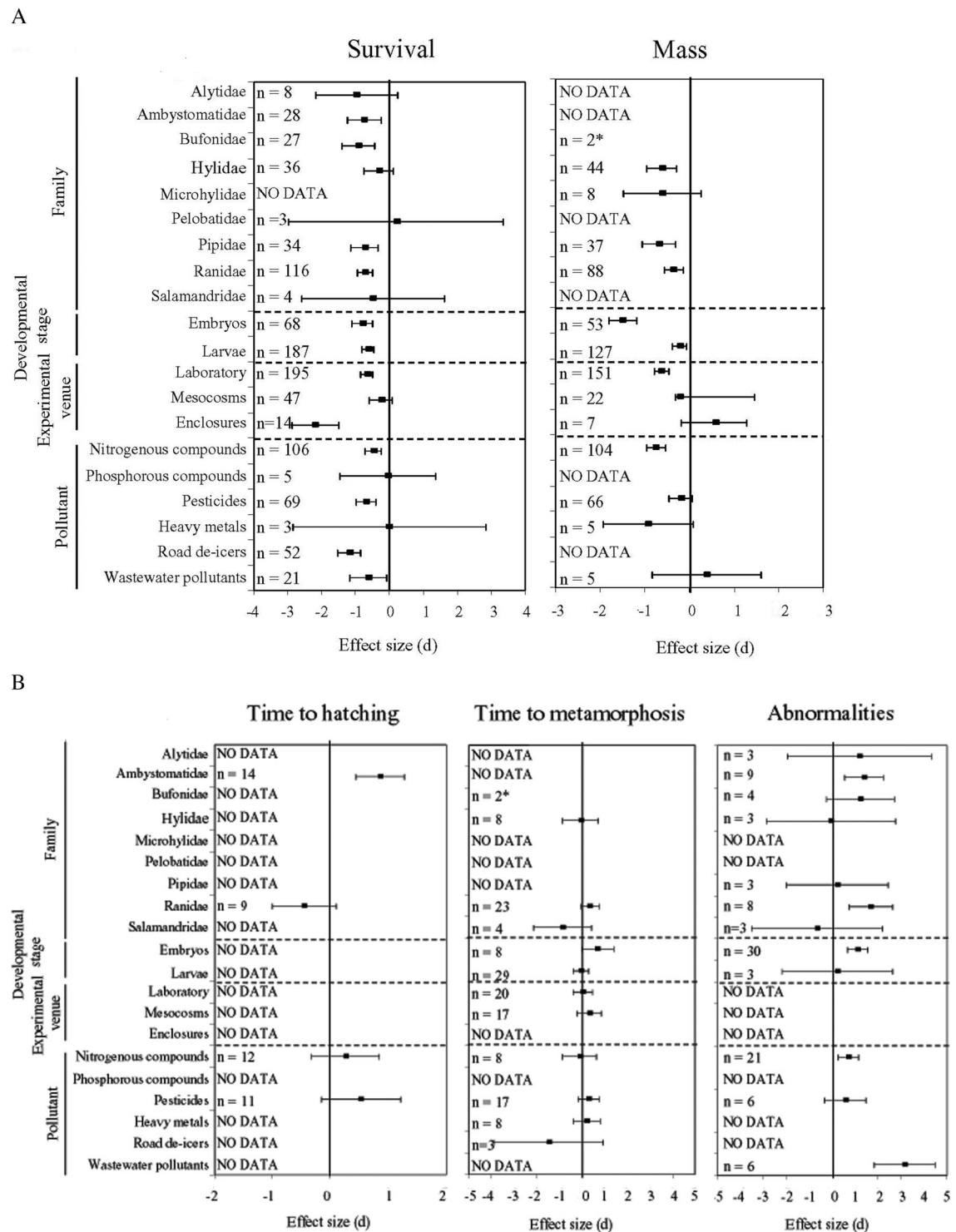
### Phylogenetic comparative analysis

The phylogenetic comparative analysis found weak evidence of phylogenetic patterns in sensitivity to pollutants. The TFSI detected significant phylogenetic autocorrelation among the tip data for the effect size in time to hatching (mean C-statistic = 0.461;  $P = 0.026$ ). However, the test did not detect phylogenetic signal for the remaining response variables.

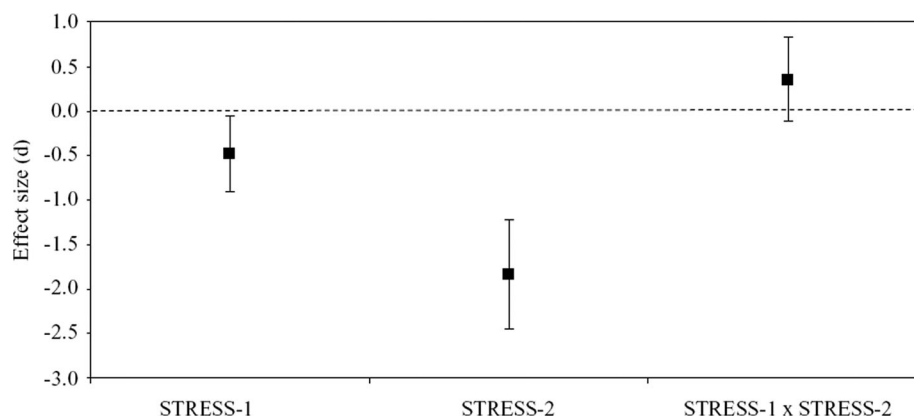
### Publication bias

The weighted Rosenberg's fail-safe number was large for survival (2304.9) and mass (992.0), whereas it was smaller for the rest of variables ( $< 57.5$  in all cases). For the factorial meta-analysis of survival, Rosenberg's fail-safe number was low for all the effect sizes analyzed ( $< 35.8$  in all cases).

We also correlated effect size and sample size using Spearman's rank correlation analyses to formally test for publication bias. We found no significant correlation for time to metamorphosis ( $r_s = 0.013$ ;  $P = 0.939$ ), time to hatching ( $r_s = -0.087$ ;  $P = 0.693$ ), abnormality frequency ( $r_s = 0.245$ ;  $P = 0.169$ ), or survival ( $r_s = 0.076$ ;  $P = 0.229$ ). For the factorial meta-analysis, no significant correlation was found for



**Figure 2.** Effect (mean and 95% confidence interval) of pollutants on survival, mass, time to hatching, time to metamorphosis, and abnormality frequency for the categories considered for the a priori defined groups. The number of point samples used to calculate each mean is shown for each analysis. The expression “no data” denotes both those classes for which actually no data were available and those for which fewer than two valid studies (see Material and Methods for more detail). Effect sizes were considered significant if 95% confidence intervals did not overlap with zero. Effect sizes within analyses were considered different from one another if their 95% confidence intervals did not overlap. Notice the different scales for each variable shown. \*For clarity, statistics corresponding to categories showing small sampling size ( $n \leq 2$ ) are not shown in the graphic.



**Figure 3.** Effect size (mean and 95% confidence interval) for full models for the parameters calculated for the factorial meta-analysis of the effect of the interaction between pollutants and additional stressors on amphibian survival ( $n = 45$ ). Effect sizes were considered significant if 95% confidence intervals did not overlap with zero. STRESS-1: average overall effect sizes of exposure to STRESS-1; STRESS-2: average overall effect sizes of exposure to STRESS-2; STRESS-1  $\times$  STRESS-2: average interaction effect size between the exposure to STRESS-1 and to STRESS-2.

the overall effect size of exposure to STRESS-1 ( $r_s = 0.168$ ;  $P = 0.270$ ). The only significant correlations were for mass ( $r_s = 0.373$ ;  $P = 0.0001$ ) in the meta-analysis and for survival under the exposure to STRESS-2 (STRESS-2;  $r_s = -0.447$ ;  $P = 0.002$ ) and for the interaction effect size between the exposure to STRESS-1 and to STRESS-2 ( $r_s = 0.276$ ;  $P = 0.039$ ) in the subsequent factorial meta-analysis. Collectively, these results suggest that there was not publication bias for most of the response variables, although it cannot be ruled out in three of the eight cases tested.

## Discussion

The meta-analysis revealed that the exposure of amphibians to chemical pollutants as a group at environmentally relevant concentrations causes medium effect sizes on reduced survival and mass and a large effect on increased frequency of abnormalities. This overall analysis supports the conclusions derived from vote-counting reviews on the general effect of pollutants on amphibians (Cowman and Mazanti 2000; Linder and Grillitsch 2000; Sparling 2000; Camargo et al. 2005; Relyea and Hoverman 2006; Marco and Ortiz-Santaliestra 2009). Although changes caused by pollutants on survival and mass are lower than 15%, any reduction in these traits, or the presence of deformities, could reduce juvenile recruitment affecting amphibian population sizes (Hayes et al. 2010), which agrees with the hypothesis that pollution is one of the major threats faced by amphibians (e.g., Beebee and Griffiths 2005; Hayes et al. 2010). However, our meta-analytic approach highlighted the fact that the impact of pollution was primarily on survival, mass, and abnormality frequency and not on amphibian developmental time (i.e., time to hatching and to metamorphosis). Although some evidences of publication bias were detected for some of the

variables analyzed, the effect of such a bias is possibly small. Therefore, the results obtained appear to be a reliable review of the published evidences on the impact of pollution on amphibians analyzed.

Although it can be difficult to know when negative effects on individuals may also affect population dynamics (e.g., Schmidt 2004), understanding how individuals are affected by such single and multiple stressors is essential because individual traits such as larval mortality may have deleterious effects at the population level (Gamradt and Kats 1996; Sih et al. 2004; Vredenburg 2004; Hayes et al. 2010). The overall effects of pollutants detected in the meta-analysis certainly could lead to population declines both directly by reducing individual survival and indirectly by decreasing mass or by increasing the frequency of abnormalities. We discuss these issues in more detail below.

## Survival

Embryonic and larval amphibian survival was affected by pollutants, although significant differences among experimental venues and types of pollutants were observed. The negative overall effect of pollutants on survival is due to physiological alterations, such as increased methemoglobin concentrations, modification of enzyme activities, and even DNA damage (e.g., Huey and Beiting 1980a; Ralph and Petras 1997; Widder and Bidwell 2006).

Road deicers had a significantly larger effect relative to nitrogenous compounds. This fact may be due to the high electrical conductivity of water considered in those studies examining the effect of road deicers (e.g., Karraker 2007; Karraker et al. 2008). Although such conductivity levels were found in natural ponds, it is likely that, overall, they were much higher than in the case of pollution by nitrogenous



compounds. Consequently, osmotic stress was more deadly than the effect of these latter pollutants. The lack of effect of phosphorous compounds on amphibians agrees with previous findings (Smith 2007, but see Hamer *et al.* 2004). However, the lack of effect of heavy metal exposure contradicts the conclusions of past studies (Linder and Grillitsch 2000), although this may be due to the small sample size and low statistical power incurring in type II errors. Moreover, the reported study revealed that the lack of effect of heavy metals on survival may be mediated by the elevated mortality in control treatment (Chen *et al.* 2006), also increasing type II errors. Further research is needed to verify the effects of heavy metals on amphibian survival.

The exposure to pollutants in field enclosure experiments exhibited higher mortality than in laboratory or mesocosms experiments. Since high salinity, acidity, or ultraviolet-B radiation, and even additional pollutants become additional stressors present in natural ponds and streams, these factors may be the cause of the higher mortality under enclosure conditions (e.g., Hatch and Blaustein 2000; Boone *et al.* 2005; Macías *et al.* 2007; Egea-Serrano *et al.* 2009b; Egea-Serrano 2010; Ortiz-Santaliestra *et al.* 2010).

Evidence of ontogenic variation in vulnerability to pollutants has been previously reported for individual pollutants (e.g., Bridges 2000; Bridges and Semlitsch 2000; Greulich and Pflugmacher 2003; Ortiz-Santaliestra *et al.* 2006). Differences in sensitivity to pollutants between embryos and larvae have often been attributed to the gelatinous egg matrix in which embryos live and the complete tissue and organ differentiation of larvae. The jelly coat may protect embryos from some chemicals (Berrill *et al.* 1998; Pauli *et al.* 1999), but it can react with other chemicals and become more toxic (Räsänen *et al.* 2003a; Marquis *et al.* 2006a). Incomplete development of the nervous system may protect embryos from those chemicals affecting nervous system (Ortiz-Santaliestra *et al.* 2006). Additionally, nitrate in the gut may be transformed into nitrosamines, which are carcinogenic (Committee on Nitrate Accumulation 1972), and symbiotic gut bacteria are involved in digestion (affecting food ingestion by larvae) and in the transformation of nitrate into nitrite (compound increasing methemoglobin concentration) (Huey and Beiting 1980b; Hecnar 1995). Thus, incomplete gut differentiation can protect embryos from some chemicals. On the other hand, later larval stages may be more tolerant to chemicals than embryos because of increased detoxifying ability (Bucciarelli *et al.* 1999) and because of their internal gills (Duellman and Trueb 1994) and increased skin thickness, which protect them against osmoregulatory alterations (McDiarmid and Altig 1999). All these aspects did not make us to expect significant differences among developmental stages. Thus, while individual studies have found differences in tolerance among developmental stages, our analysis across all studies indicates there is not a general pattern of differential sen-

sitivity to pollutants between embryos and larvae. Some of the original studies included in the analysis lasted only until embryos hatched. Consequently, carry-over effects were not taken into account, which may underestimate the observed effect when the exposure to pollutants started at embryonic stages. Therefore, future research identifying the effect of embryonic exposure on tadpole performance is relevant to establish the actual role of pollutants on amphibian population decline.

The general lack of variation in survival effects among amphibian families and weak phylogenetic signal disagrees with individual studies that have suggested the existence of a phylogenetic signal in the impact of particular pesticide or nitrogenous compound (e.g., Marco *et al.* 1999; Bridges and Semlitsch 2000; Shinn *et al.* 2008; Snodgrass *et al.* 2008; Jones *et al.* 2009; Relyea and Jones 2009). Thus, while some chemicals can have species-specific effects, when we combined all studies, we found no general phylogenetic pattern of sensitivity. However, the fact that some families (Alytidae, Pelobatidae, and Pipidae) were represented by a single species may contribute to explain the general lack of a pattern observed. Moreover, population-level variation in chemical tolerance can occur (e.g., Shinn *et al.* 2008; Egea-Serrano *et al.* 2009b) and high levels of such a variation may be obscuring species-level patterns. Indeed, this is an issue in any phylogenetic analysis and researchers commonly must work under the assumption that population-level variation is lower than species-level variation. To assess the magnitude of this potential problem, future research should examine both species- and population-level tolerance within a phylogenetic framework.

## Mass

Overall, pollutants significantly reduced amphibian mass, although this effect depended on developmental stage, experimental venue, and pollutant type. No significant differences among amphibian families were detected, which would support the lack of phylogenetic autocorrelation detected for this trait. As mentioned earlier, this nonsignificant effect could be influenced by that for families Microhylidae and Pipidae, all data came from a single species and/or interpopulation variation in tolerance, a factor that should be analyzed in future research.

The overall negative effect of pollutants on mass can be a combination of direct and indirect effects. Direct effects include reduced foraging efficiency or increased physiological stress due to detoxification pathways (Wright and Wright 1996; Egea-Serrano *et al.* 2009b). Indirect effects occur when pollutants may impact either positively or negatively mass by affecting algal growth in more realistic experimental venues (Boone *et al.* 2007; Relyea 2009; Egea-Serrano 2010).

Pollutant effect on mass was significantly more negative when the exposure began in embryonic stages than when it began in larval stages. Such effects may be mediated by simply longer exposure to pollutants when experiments began at earlier stages (mean  $\pm$  1 SE; embryos:  $751 \pm 107$  h,  $n = 76$ ; larvae:  $446 \pm 34$  h,  $n = 286$ ;  $F_{1,360} = 12.514$ ;  $P = 0.0001$ ) or may be due to a higher sensitivity of embryos to pollution or some lag effect that reduce their subsequent growth rates (Herkovits and Fernández 1978; McDiarmid and Altig 1999; Marquis *et al.* 2006a). Declines in mass, especially when amphibians are exposed as embryos, may subsequently result in additional detrimental effects by making individuals more vulnerable to gape-size predators for longer periods of time (Semlitsch and Gibbons 1988), reducing competitive abilities, increasing larval development duration (Snodgrass *et al.* 2004), or affecting future survival and fitness (Berven and Gill 1983; Smith 1987; Semlitsch *et al.* 1988; Reques and Tejedo 1997; Altwegg and Reyer 2003).

The smaller decline in mass observed in mesocosm and enclosure studies compared to lab studies may be attributed to the indirect effect of pollutants. Under the more realistic conditions of mesocosms and enclosures, these chemicals can affect algal growth (Boone *et al.* 2007; Relyea 2009; Egea-Serrano 2010) in ways that can positively and negatively affect amphibian growth. In addition, there is typically a reduction in larval crowding conditions in mesocosms and enclosures compared to laboratory studies, which should reduce competitive stress, and allow individuals to get more energy to invest in detoxification pathways. This reduction in density in mesocosms and enclosures may counterbalance negative effects of pollutants on mass, and weaken their impact, an outcome already suggested for other stressors such as predation risk (Peacor and Werner 2000).

A second explanation for the observed differences among experimental venues is that water is frequently changed and pollutants are reapplied in lab experiments. In contrast, mesocosm or enclosure experiments typically use a single application of pollutants. Therefore, pollutants in mesocosm and enclosure experiments have a greater opportunity to break down over time. As a result, amphibians in these venues should experience lower average exposures, resulting in less-severe effects on mass.

Across all studies, nitrogenous compounds generally reduced amphibian mass more than pesticides. This overall effect contradicts a number of individual studies reporting that ecologically relevant concentrations of some pesticides and nitrogenous compounds caused similar effects on mass (e.g., Boone *et al.* 2005; Boone and Bridges-Britton 2006). Nitrogenous compounds may increase water salinity (e.g., Egea-Serrano 2010) and lead to oxygen depletion because of eutrophication processes (see review Camargo and Alonso 2006). The interaction between the nitrogen toxicity and these additional stressing effects may exacerbate the effects of

nitrogen toxicity isolated and reduce larval mass synergistically (e.g., Ortiz-Santaliestra 2008), which would explain the results obtained.

## Developmental time

Overall, pollutants did not affect time to hatching or time to metamorphosis, although their effects varied among families. Although some individual studies have detected significant effect of pollutants on time to hatching (e.g., Ingermann *et al.* 1997; Räsänen *et al.* 2003a, b; Rohr *et al.* 2003, 2004; Griffis-Kyle 2007), many studies have reported no impact (Berrill *et al.* 1994, 1998; Berrill and Bertram 1997; Pauli *et al.* 1999; Greulich and Pflugmacher 2003; Griffis-Kyle 2007). The frequent lack of pollutant effect on time to hatching may be due to the protective role of the embryonic jelly coat (Räsänen *et al.* 2003c; Marquis *et al.* 2006b; Edginton *et al.* 2007) or incomplete embryonic organ development (Hecnar 1995). However, the hypothesized protective mechanisms were not effective for other response variables, including survival, suggesting that pathways involved in embryonic development do not affect other traits.

Significant heterogeneity among ambystomatids and ranids was observed in relation to time to hatching, which is supported by the significant autocorrelation detected for this variable. Ranids were unaffected by pollutants whereas ambystomatids delayed their embryonic development, likely due to the impact of chemicals on the rate of cellular division. The lack of effect of pollutants on time to hatching detected for ranids is consistent with the results obtained for time to metamorphosis for this family, the only one for which data for both variables were available. Although for some species larval period may be affected by pollutants (e.g., Hayes *et al.* 2006), the overall lack of impact of these compounds on time to metamorphosis suggests a general lack of escape strategy from polluted aquatic environments.

## Abnormality frequency

Exposure to pollutants increased the overall abnormality frequency and there were differences among pollutant types. The exposure to wastewater and nitrogenous pollutants significantly increased the incidence of abnormality frequency, likely due to the alteration of those enzymes involved in development or to DNA damage (Dunson and Connell 1982; Ralph and Petras 1997). In the particular case of the wastewater boron cation, it has been argued that cations may more severely affect the hatching enzyme responsible for enlarging the perivitelline membrane surrounding the embryo than anions (e.g., nitrate) (Dunson and Connell 1982; Laposata and Dunson 1998), thereby increasing the incidence of malformations. Physical abnormalities are correlated with reduced speed and anomalous movements (Laposata and Dunson 1998) that may increase mortality by both predation (e.g.,

Watkins 1996; Dayton and Fitzgerald 2001) and increased metabolic costs (Rowe *et al.* 2002). Therefore, the effect of pollutants on abnormality frequency may severely affect individual fitness.

### The effect of multiple stressors on survival: do pollutants act synergistically?

The combination of chemicals such as fertilizers or pesticides with other factors (e.g., predator cues, ultraviolet-B radiation, or even other chemicals) has been reported to have synergistic effects on amphibian survival, growth, and development (e.g., Hatch and Blaustein 2000; Relyea and Mills 2001; Boone *et al.* 2005; Hayes *et al.* 2006; Macías *et al.* 2007; Egea-Serrano *et al.* 2009b). However, other studies have not found synergistic effects (Boone *et al.* 2005; Boone and Bridges-Britton 2006). Our factorial analysis across all studies found no overall synergistic effect of pollutants when combined with other stressors. This is important since it implies that while individual chemicals or stressors can interact, there is not an overall pattern that additional stressors consistently increase the lethality of pollutants.

### Conclusions and future research directions

The results of our meta-analysis found that pollution is a major threat to amphibians by having large effects on abnormality frequency and medium effects on survival and mass. Such effects may explain the link between pollutants and population declines described for several areas around the world (Davidson *et al.* 2001; Sparling *et al.* 2001; Fellers *et al.* 2004; Hamer *et al.* 2004). In contrast, developmental time was not significantly affected by pollutants and the addition of other stressors did not generally exacerbate the effects of the chemicals. To our knowledge, this study is the first quantitative summary analysis to examine the impact of pollutants on amphibians at ecologically relevant concentrations. However, we have to be cautious when interpreting the results of a meta-analysis as evidence for the overall impact of pollution on amphibians in the wild. We stress that more research is needed to achieve more robust conclusions on the traits studied, especially, for development traits and also for factorial designs exhibiting low Rosenberg's fail-safe number. In particular, because pollutants in the environment are expected to be experienced in the presence of other stressors, the study of interactions among factors is of great relevance to fully understand the role of pollutants in amphibian declines (Hayes *et al.* 2006).

The response variables selected for our meta-analysis are related to amphibian fitness. Nevertheless, they are just a sample of the possible effects that pollutants may have on amphibians. For instance, activity level, habitat use, courtship, and swimming performance are all affected by pollution (e.g., Bridges 1997, 1999; Marco and Blaustein 1999; Shinn *et al.*

2008; Ortiz-Santaliestra *et al.* 2009). Therefore, since the ability of individuals to escape from predators (or other stressors such as ultraviolet-B radiation), to feed or even to reproduce, may be affected, it is likely that the impact of pollutants is even higher than that reported in our study and, consequently, such traits should be considered in future meta-analyses.

A final remark is that the response variables were all recorded at the individual level. Nevertheless, they are the expression of the alterations of the physiology of the individuals affected (e.g., Huey and Beiting 1980a; Widder and Bidwell 2006). Moreover, different species may be differently susceptible to pollution and other stressors, and the interaction among all these factors may lead to impoverished communities dominated by tolerant species (Christensen *et al.* 2006). All of these considerations underscore the relevance of running meta-analyses at both physiological and community scales to get a broad and comprehensive view of the causes and consequences of chemical pollution on amphibians.

### Acknowledgments

We are grateful to G. Smith, M. E. Ortiz-Santaliestra, and A.-L. Mandrillon for kindly replying to our queries on their published studies. We also thank A. Richter-Boix for his comments on tests for serial independence. This research was partially funded by project (CGL2004-01872/BOS and CGL2009-12767-C02-02) from Dirección General de Investigación Científica y Técnica (MICINN), conceded to M. T. A. E. S. was supported by a Caja de Ahorros del Mediterráneo predoctoral fellowship. R. R. has been supported by the National Science Foundation.

### References

- Abouheif, E. 1999. A method to test the assumption of phylogenetic independence in comparative data. *Evol. Ecol. Res.* 1:895–909.
- Altwegg, R., and H. U. Reyer. 2003. Patterns of natural selection on size at metamorphosis in water frogs. *Evolution* 57:872–882.
- Bancroft, B. A., N. J. Baker, and A. R. Blaustein. 2007. Effects of UV-B radiation on marine and freshwater organisms: a synthesis through meta-analysis. *Ecol. Lett.* 10:332–345.
- Bancroft, B. A., N. J. Baker, and A. R. Blaustein. 2008. A meta-analysis of the effects of ultraviolet B radiation and its synergistic interactions with pH, contaminants, and disease on amphibian survival. *Conserv. Biol.* 22:987–996.
- Beebee, T. J. C., and R. A. Griffiths. 2005. The amphibian decline crisis: a watershed for conservation biology? *Biol. Conserv.* 125:271–285.
- Begg, C. B., and M. Mazumdar. 1994. Operating characteristics of a rank correlation test for publication bias. *Biometrics* 50:1088–1101.
- Begg, C. B. 1994. Publication bias. Pp 399–409 *in* H. Cooper and V. Hedges, eds. *The handbook of research synthesis*. Russell Sage Foundation, New York.

- Berrill, M., and S. Bertram. 1997. Effects of pesticides on amphibian embryos and larvae. *Herpetol. Conserv.* 1:233–245.
- Berrill, M., D. Coulson, L. Mc Gillivray, and B. Pauli. 1998. Toxicity of endosulfan to aquatic stages of anuran amphibians. *Environ. Toxicol. Chem.* 17:1738–1744.
- Berrill, M., S. Bertram, L. Mc Gillivray, M. Kolohon, and B. Pauli. 1994. Effects of low concentrations of forest use pesticides on frog embryos and tadpoles. *Environ. Toxicol. Chem.* 13:657–664.
- Berven, K. E., and D. E. Gill. 1983. Interpreting geographic variation in life-history traits. *Am. Zool.* 23:85–93.
- Boone, M. D., and C. Bridges-Britton. 2006. Examining multiple sublethal contaminants on the gray treefrog (*Hyla versicolor*): effects of an insecticide, herbicide and fertilizer. *Environ. Toxicol. Chem.* 25:3261–3265.
- Boone, M. D., and C. M. Bridges. 2003. Effects of pesticides on amphibian populations. Pp 152–167 in R. D. Semlitsch, ed. *Amphibian conservation*. Smithsonian, WA.
- Boone, M. D., and D. James. 2005. Aquatic and terrestrial mesocosms in amphibian ecotoxicology. *Appl. Herpetol.* 2:231–257.
- Boone, M. D., C. M. Bridges, J. F. Fairchild, and E. E. Little. 2005. Multiple sublethal chemicals negatively affect tadpoles of the green frog, *Rana clamitans*. *Environ. Toxicol. Chem.* 24:1267–1272.
- Boone, M. D., R. D. Semlitsch, E. E. Little, and M. C. Doyle. 2007. Multiple stressors in amphibian communities: effects of chemical contamination, bullfrogs, and fish. *Ecol. Appl.* 17:291–301.
- Bridges, C. M., and R. D. Semlitsch. 2000. Variation in pesticide tolerance of tadpoles among and within species of Ranidae and patterns of amphibian decline. *Conserv. Biol.* 14:1490–1499.
- Bridges, C. M. 1997. Tadpole swimming performance and activity affected by acute exposure to sublethal levels of carbaryl. *Environ. Toxicol. Chem.* 19:1935–1939.
- Bridges, C. M. 1999. Effects of a pesticide on tadpole activity and predator avoidance behavior. *J. Herpetol.* 33:303–306.
- Bridges, C. M. 2000. Long-term effects of pesticide exposure at various life stages of the Southern leopard frog (*Rana sphenocephala*). *Arch. Environ. Contam. Toxicol.* 39:91–96.
- Bucciarelli, T., P. Sacchetta, A. Pennelli, L. Cornelio, R. Romagnoli, S. Merino, R. Petruzelli, and C. Di Ilio. 1999. Characterization of toad glutathione transferase. *Biochim. Biophys. Acta* 1431:189–198.
- Camargo, J. A., A. Alonso, and A. Salamanca. 2005. Nitrate toxicity to aquatic animals: a review with new data for freshwater invertebrates. *Chemosphere* 58:1255–1267.
- Camargo, J. A., and A. Alonso. 2006. Ecological and toxicological effects of inorganic nitrogen pollution in aquatic ecosystems: a global assessment. *Environ. Int.* 32:831–849.
- Carpenter, S., N. F. Caraco, D. L. Correll, R. W. Howarth, A. N. Sharples, and V. H. Smith. 1998. Nonpoint pollution of surface waters with phosphorous and nitrogen. *Ecol. Appl.* 8:559–568.
- Chalcraft, D. R., C. A. Binckley, and W. J. Resetarits Jr. 2005. Experimental venue and estimation of interaction strength: comment. *Ecology* 86:1061–1067.
- Chen, T. H., J. A. Gross, and W. H. Karasov. 2006. Sublethal effects of lead on Northern leopard frog (*Rana pipiens*) tadpoles. *Environ. Toxicol. Chem.* 25:1383–1389.
- Christensen, M. R., M. D. Graham, R. D. Vinebrooke, D. L. Findlay, M. J. Paterson, and M. A. Turner. 2006. Multiple anthropogenic stressors cause ecological surprises in boreal lakes. *Glob. Change Biol.* 12:1–7.
- Cohen, J. 1969. *Statistical power analysis for the behavioral sciences*. Academic Press, New York.
- Committee on Nitrate Accumulation. 1972. *Accumulation of Nitrate*. National Academy of Sciences, Washington, DC.
- Cowman, D. F., and L. E. Mazanti. 2000. Ecotoxicology of “new generation” pesticides to amphibians. Pp 233–268 in D. W. Sparling, G. Linder, and C. A. Bishop, eds. *Ecotoxicology of amphibians and reptiles*. Society of Environmental Toxicology and Chemistry (SETAC), Pensacola, FL.
- Daszak, P., A. A. Cunningham, and A. D. Hyatt. 2001. Anthropogenic environmental change and the emergence of infectious diseases in wildlife. *Acta Tropica* 78:103–116.
- Davidson, C., H. B. Shaffer, and M. R. Jennings. 2001. Declines of the California red-legged frog: climate, UV-B, habitat and pesticides hypotheses. *Ecol. Appl.* 11:464–479.
- Dayton, G. H., and L. A. Fitzgerald. 2001. Competition, predation, and the distributions of four desert anurans. *Oecologia* 129:430–435.
- Duellman, W., and L. Trueb. 1994. *Biology of amphibians*. Johns Hopkins Univ. Press, Baltimore, MD.
- Dunson, W. A., and J. Connell. 1982. Specific inhibition of hatching in amphibians exposed to low pH. *J. Herpetol.* 16:314–316.
- Edgington, A. N., C. Rouleau, G. R. Stephenson, and H. J. Boermans. 2007. 2,4-D butoxyethyl ester kinetics in embryos of *Xenopus laevis*: the role of the embryonic jelly coat in reducing chemical absorption. *Arch. Environ. Contam. Toxicol.* 52:113–120.
- Egea-Serrano, A., M. Tejedo, and M. Torralva. 2009a. Estimating mean lethal concentrations of three nitrogenous compounds for larvae of the Iberian waterfrog, *Pelophylax perezi* (Seoane, 1885). *Revista Española de Herpetología* 23:107–116.
- Egea-Serrano, A., M. Tejedo, and M. Torralva. 2009b. Populational divergence in the impact of three nitrogenous compounds and their combination on larvae of the frog *Pelophylax perezi* (Seoane, 1885). *Chemosphere* 76:869–877.
- Egea-Serrano, A. 2010. Aspectos relevantes en la conservación de anfibios en la Región de Murcia: efectos de la contaminación por fertilizantes sobre *Pelophylax perezi* (Seoane, 1885) [Relevant aspects in amphibian conservation in the province of Murcia: effect of fertilizer pollution on *Pelophylax perezi* (Seoane, 1885)]. Ph.D. thesis, Universidad de Murcia, Murcia, 366 pp.



- Faivovich, J., C. F. B. Haddad, P. C. A. Garcia, D. R. Frost, J. A. Campbell, and W. C. Wheeler. 2005. Systematic review of the frog family Hylidae, with special reference to Hylinae: phylogenetic analysis and taxonomic revision. *Bull. Am. Mus. Nat. Hist.* 294:1–240.
- Fellers, G. M., L. L. McConnell, D. Pratt, and S. Datta. 2004. Pesticides in mountain yellow-legged frogs (*Rana muscosa*) from the Sierra Nevada mountains of California, USA. *Environ. Toxicol. Chem.* 23:2170–2177.
- Frost, D. R., T. Grant, J. Faivovich, R. H. Bain, A. Haas, C. F. D. Haddad, R. O. De Sá, A. Channing, M. Wilkinson, S. C. Donnellan, *et al.* 2006. The amphibian tree of life. *Bull. Am. Mus. Nat. Hist.* 297:1–371.
- Galloway, J. N., J. D. Aber, J. W. Erisman, S. P. Seitzinger, R. W. Howarth, E. B. Cowling, and B. J. Cosby. 2003. The nitrogen cascade. *Bioscience* 53:341–356.
- Gamradt, S. C., and L. B. Kats. 1996. Effects of introduced crayfish and mosquitofish on Californian newts. *Conserv. Biol.* 10:1155–1162.
- García-París, M., D. R. Buchholz, and G. Parra-Olea. 2003. Phylogenetic relationships of Pelobatoidea re-examined using mtDNA. *Mol. Phylogenet. Evol.* 28:12–23.
- Gilliom, R. J., J. E. Barbash, C. G. Crawford, P. A. Hamilton, J. D. Martin, N. Nakagaki, L. H. Nowell, J. C. Scott, P. E. Stackelberg, G. P. Thelin, *et al.* 2007. Pesticides in the nation's streams and ground water, 1992–2001. U.S. Geological Survey Circular 1291. Revised February 15, 2007.
- Greulich, K., and S. Pflugmacher. 2003. Differences in susceptibility of various life stages of amphibians to pesticide exposure. *Aquat. Toxicol.* 65:329–336.
- Griffis-Kyle, K. L. 2005. Ontogenic delays in effects of nitrite exposure on tiger salamanders (*Ambystoma tigrinum tigrinum*) and wood frogs (*Rana sylvatica*). *Environ. Toxicol. Chem.* 24:1523–1527.
- Griffis-Kyle, K. L. 2007. Sublethal effects of nitrite on eastern tiger salamander (*Ambystoma tigrinum tigrinum*) and wood frog (*Rana sylvatica*) embryos and larvae: implications for field populations. *Aquat. Toxicol.* 41:119–127.
- Gurevitch, J., J. A. Morrison, and L. V. Hedges. 2000. The interaction between competition and predation: a meta-analysis of field experiments. *Am. Nat.* 155:435–453.
- Gómez-Mestre, I., and M. Tejedo. 2003. Local adaptation of an anuran amphibian to osmotically stressful environments. *Evolution* 57:1889–1899.
- Hamer, A. J., J. A. Makings, S. J. Lane, and M. J. Mahony. 2004. Amphibian decline and fertilizers used on agricultural land in south-eastern Australia. *Agric. Ecosyst. Environ.* 102:299–305.
- Hatch, A. C., and A. R. Blaustein. 2000. Combined effects of UV-B, nitrate and low pH reduce the survival and activity level of larval Cascades frogs (*Rana cascadae*). *Arch. Environ. Contam. Toxicol.* 39:494–499.
- Hayes, T. B., P. Case, S. Chui, D. Chung, C. Haeffele, K. Haston, M. Lee, V. P. Mai, Y. Marjua, and M. Tsui. 2006. Pesticide mixtures, endocrine disruption, and amphibian declines: are we underestimating the impact? *Environ. Health Perspect.* 114:40–50.
- Hayes, T. B., P. Falso, S. Gallipeau, and M. Stice. 2010. The cause of global amphibian declines: a developmental endocrinologist's perspective. *J. Exp. Biol.* 213:921–933.
- Hecnar, S. J. 1995. Acute and chronic toxicity of ammonium nitrate fertilizer to amphibians from southern Ontario. *Environ. Toxicol. Chem.* 14:2131–2137.
- Herkovits, J., and A. Fernández. 1978. Tolerancia a toxas durante el desarrollo embrionario. *Medicina (Buenos Aires)* 39:400–408.
- Hillis, D. M., and T. P. Wilcox. 2005. Phylogeny of the New World true frogs (*Rana*). *Mol. Phylogenet. Evol.* 34:299–314.
- Hoffmann, M., C. Hilton-Taylor, A. Angulo, M. Böhm, T. M. Brooks, S. H. M. Butchart, K. E. Carpenter, J. Chanson, B. Collen, N. A. Cox, *et al.* 2010. The impact of conservation on the status of the world's vertebrates. *Science* 330:1503–1509.
- Huey, D. W., and T. L. Beiting. 1980a. Hematological responses of larval *Rana catesbeiana* to sublethal nitrite exposures. *Bull. Environ. Contam. Toxicol.* 25:574–577.
- Huey, D. W., and T. L. Beiting. 1980b. Toxicity of nitrite to larvae of the salamander *Ambystoma texanum*. *Bull. Environ. Contam. Toxicol.* 25:909–912.
- Ingermann, R. L., D. C. Bencic, and V. P. Eroschenko. 1997. Methoxychlor alters hatching and larval startle response in the salamander *Ambystoma macrodactylum*. *Bull. Environ. Contam. Toxicol.* 59:815–821.
- Jones, D. K., J. I. Hammond, and R. A. Relyea. 2009. Very highly toxic effects of endosulfan across nine species of tadpoles: lag effects and family-level sensitivity. *Environ. Toxicol. Chem.* 28:1939–1945.
- Jones, T. R., A. G. Kluge, and A. J. Wolf. 1993. When theories and methodologies clash: a phylogenetic reanalysis of the North American ambystomatid salamanders (Caudata: Ambystomatidae). *Syst. Biol.* 42:92–102.
- Karraker, N. E., J. P. Gibbs, and J. R. Vonesh. 2008. Impacts of road deicing salt on the demography of vernal pool-breeding amphibians. *Ecol. Appl.* 18:724–734.
- Karraker, N. E. 2007. Are embryonic and larval green frogs (*Rana clamitans*) insensitive to road deicing salt? *Herpetol. Conserv. Biol.* 2:35–41.
- Kerby, J. L., K. L. Richards-Hrdlicka, A. Storfer, and D. K. Skelly. 2010. An examination of amphibian sensitivity to environmental contaminants: are amphibians poor canaries? *Ecol. Lett.* 13:60–67.
- Kolpin, D. W., E. T. Furlong, M. T. Meyer, E. M. Thurman, S. D. Zaugg, L. B. Barber, and H. T. Buxton. 2002. Pharmaceuticals, hormones, and other organic wastewater contaminants in US streams, 1999–2000: a national reconnaissance. *Environ. Sci. Technol.* 36:1202–1211.
- Laposata, M. M., and W. A. Dunson. 1998. Effects of boron and nitrate on hatching success of amphibian eggs. *Arch. Environ. Contam. Toxicol.* 35:615–619.



- Linder, G., and B. Grillitsch. 2000. Ecotoxicology of metals. Pp 325–459 in D. W. Sparling, G. Linder, and C. A. Bishop, eds. Ecotoxicology of amphibians and reptiles. Society of Environmental Toxicology and Chemistry (SETAC), Pensacola, FL.
- Macías, G., A. Marco, and A. R. Blaustein. 2007. Combined exposure to UV-B radiation and nitrite negatively affects survival of amphibian early stages. *Sci. Total Environ.* 385:55–65.
- Marco, A., and A. R. Blaustein. 1999. The effects of nitrite on behavior and metamorphosis in cascades frogs (*Rana cascadae*). *Environ. Toxicol. Chem.* 18:946–949.
- Marco, A., and M. E. Ortiz-Santaliestra. 2009. Impact of reactive nitrogen on amphibians. Chapter 7 in H. Heatwole, ed. Vol. 8. Amphibian biology, Conservation of amphibians.
- Marco, A., C. Quilchano, and A. R. Blaustein. 1999. Sensitivity to nitrate and nitrite in pond breeding amphibians from the Pacific Northwest, USA. *Environ. Toxicol. Chem.* 18:2836–2839.
- Marquis, O., A. Millery, S. Guittonneau, and C. Miaud. 2006b. Toxicity of PAHs and jelly protection of eggs in the common frog *Rana temporaria*. *Amphib. Reptil.* 27:472–475.
- Marquis, O., A. Millery, S. Guittonneau, and S. Miaud. 2006a. Solvent toxicity to amphibian embryos and larvae. *Chemosphere* 63:889–892.
- McDiarmid, R. W., and R. Altig, eds. 1999. Tadpoles. The biology of anuran larvae. Univ. of Chicago Press, Chicago, IL.
- McKenzie, R. L., L. O. Björn, A. Bais, and M. Ilyasd. 2003. Change in biologically active ultraviolet radiation reaching the Earth's surface. *Photochem. Photobiol. Sci.* 2:5–15.
- Ortiz, M. E., A. Marco, M. Saiz, and M. Lizana. 2004. Impact of ammonium nitrate on growth and survival of six European amphibians. *Arch. Environ. Contam. Toxicol.* 47:234–239.
- Ortiz-Santaliestra, M. E., A. Marco, M. J. Fernández, and M. Lizana. 2006. Influence of developmental stage on sensitivity to ammonium nitrate of aquatic stages of amphibians. *Environ. Toxicol. Chem.* 25:105–111.
- Ortiz-Santaliestra, M. E., A. Marco, M. J. Fernández-Benítez, and M. Lizana. 2009. Alteration of courtship behavior because of water acidification and minor effect of ammonium nitrate in the Iberian newt (*Lissotriton boscai*). *Environ. Toxicol. Chem.* 28:1500–1505.
- Ortiz-Santaliestra, M. E., M. J. Fernández-Benítez, M. Lizana, and A. Marco. 2010. Adaptation to osmotic stress provides protection against ammonium nitrate in *Pelophylax perezi* embryos. *Environ. Pollut.* 158:934–940.
- Ortiz-Santaliestra, M. E. 2008. Efectos de la contaminación por nitrógeno sobre la reproducción y el desarrollo de anfibios. Unpublished Ph.D. thesis, University of Salamanca, Salamanca, 339 pp.
- Palmer, A. R. 1999. Detecting publication bias in meta-analyses: a case study of fluctuating asymmetry and sexual selection. *Am. Nat.* 154:220–233.
- Pauli, B. D., D. R. Coulson, and M. Berrill. 1999. Sensitivity of amphibian embryos and tadpoles to Mimic<sup>®</sup> 240 LV insecticide following single or double exposures. *Environ. Toxicol. Chem.* 18:2538–2544.
- Pauly, G. B., D. M. Hillis, and D. C. Cannatella. 2004. The history of a nearctic colonization: molecular phylogenetics and biogeography of the nearctic toads (*Bufo*). *Evolution* 58:2517–2535.
- Peacor, S. D., and E. E. Werner. 2000. Predator effects on an assemblage of consumers through induced changes in consumer foraging behavior. *Ecology* 81:1998–2010.
- Ralph, S., and M. Petras. 1997. Genotoxicity monitoring of small bodies of water using two species of tadpoles and the alkaline single cell gel (comet) assay. *Environ. Mol. Mutagen.* 29:418–430.
- Read, K., J. S. Keogh, I. A. W. Scott, J. D. Roberts, and P. Doughty. 2001. Molecular phylogeny of the Australian frog genera *Crinia*, *Geocrinia*, and allied taxa (Anura: Myobatrachidae). *Mol. Phylogenet. Evol.* 21:294–308.
- Relyea, R., and J. Hoverman. 2006. Assessing the ecology in ecotoxicology: a review and synthesis in freshwater systems. *Ecol. Lett.* 9:1157–1171.
- Relyea, R. A., and D. K. Jones. 2009. The toxicity of roundup original max<sup>®</sup> to 13 species of larval amphibians. *Environ. Toxicol. Chem.* 28:2004–2008.
- Relyea, R. A., and N. Mills. 2001. Predator-induced stress makes the pesticide carbaryl more deadly to grey treefrog tadpoles (*Hyla versicolor*). *Proc. Natl. Acad. Sci.* 98:2491–2496.
- Relyea, R. A. 2005. The lethal impacts of Roundup and predatory stress on six species of North American tadpoles. *Arch. Environ. Contam. Toxicol.* 48:351–357.
- Relyea, R. A. 2009. A cocktail of contaminants: how mixtures of pesticides at low concentrations affect aquatic communities. *Oecologia* 159:363–376.
- Reques, R., and M. Tejedo. 1997. Reaction norms for metamorphic traits in natterjack toads to larval density and pond duration. *J. Evol. Biol.* 10:829–851.
- Ritter, W. F., and L. Bergstrom. 2001. Nitrogen and water quality. Pp 59–90 in W. F. Ritter and A. Shirmohammadi, eds. Agricultural nonpoint source pollution: watershed management and hydrology. Lewis Publishers, Boca Raton.
- Rohr, J. R., A. A. Elskus, B. S. Shepherd, P. H. Crowley, T. M. McCarthy, J. H. Niedzwiecki, T. Sager, A. Sih, and B. D. Palmer. 2003. Lethal and sublethal effects of atrazine, carbaryl, endosulfan, and octylphenol on the streamside salamander (*Ambystoma barbouri*). *Environ. Toxicol. Chem.* 22:2385–2392.
- Rohr, J. R., A. A. Elskus, B. S. Shepherd, P. H. Crowley, T. M. McCarthy, J. H. Niedzwiecki, T. Sager, A. Sih, B. D. Palmer. 2004. Multiple stressors and salamanders: effects of an herbicide, food limitation, and hydroperiod. *Ecol. Appl.* 14:1028–1040.
- Romansic, J. M., K. A. Diez, E. M. Higashi, and A. R. Blaustein. 2006. Effects of nitrate and the pathogenic water mold *Saprolegnia* on survival of amphibian larvae. *Dis. Aquat. Organ.* 68:235–243.

- Rosenberg, M. S., D. C. Adams, and J. Gurevitch. 2000. Meta Win: Statistical software for meta-analysis, version 2.1. Sinauer Associates, Inc., Sunderland, MA.
- Rosenberg, M. S. 2005. The file-drawer problem revisited: a general weighted method for calculating fail-safe numbers in meta-analysis. *Evolution* 59:464–468.
- Rowe, C. L., W. A. Hopkins, and J. D. Congdon. 2002. Ecotoxicological implications of aquatic disposal of coal combustion residues in the United States: a review. *Environ. Monit. Assess.* 80:207–276.
- Räsänen, K., A. Laurila, and J. Merila. 2003a. Geographic variation in acid stress tolerance of the moor frog, *Rana arvalis*. II. Adaptive maternal effects. *Evolution* 57:363–371.
- Räsänen, K., A. Laurila, and J. Merila. 2003b. Geographic variation in acid stress tolerance of the moor frog, *Rana arvalis*. II. Local adaptation. *Evolution* 57:352–362.
- Räsänen, K., M. Pahkala, A. Laurila, and J. Merila. 2003c. Does jelly envelope protect the common frog *Rana temporaria* embryos from UV-B radiation? *Herpetologica* 59:293–300.
- Schmidt, B. R. 2004. Pesticides, mortality and population growth rate. *Trends Ecol. Evol.* 19:459–460.
- Schäuble, C. S., C. Moritz, and R. W. Slade. 2000. A molecular phylogeny for the frog genus *Limnodynastes* (Anura: Myobatrachidae). *Mol. Phylogenet. Evol.* 16:379–391.
- Scott, E. 2005. A phylogeny of ranid frogs (Anura: Ranoidea: Ranidae), based on a simultaneous analysis of morphological and molecular data. *Cladistics* 21:507–574.
- Semlitsch, R. D., and J. W. Gibbons. 1988. Fish predation in size-structured populations of treefrog tadpoles. *Oecologia* 75:321–326.
- Semlitsch, R. D., D. E. Scott, and J. H. K. Pechmann. 1988. Time and size at metamorphosis related to adult fitness in *Ambystoma tadpoideum*. *Ecology* 69:184–192.
- Shaffer, H. B., J. M. Clark, and F. Kraus. 1991. When molecules and morphology clash: a phylogenetic analysis of the North American ambystomatid salamanders (Caudata: Ambystomatidae). *Syst. Zool.* 40:284–303.
- Shinn, C., A. Marco, and L. Serrano. 2008. Inter- and intra-specific variation on sensitivity of larval amphibians to nitrite. *Chemosphere* 71:507–514.
- Sih, A., J. Kerby, A. Bell, and R. Relyea. 2004. Response to Schmidt. Pesticides, mortality and population growth rate. *Trends Ecol. Evol.* 19:460–461.
- Skelly, D. K. 2002. Experimental venue and estimation of interaction strength. *Ecology* 83:2097–2101.
- Smith, D. C. 1987. Adult recruitment in chorus frogs: effects of size and date at metamorphosis. *Ecology* 71:2313–2322.
- Smith, G. R. 2007. Lack of effect of nitrate, nitrite, and phosphate on wood frog (*Rana sylvatica*) tadpoles. *Appl. Herpetol.* 4:287–291.
- Snodgrass, J. W., R. E. Casey, D. Joseph, and J. A. Simon. 2008. Microcosm investigations of stormwater pond sediment toxicity to embryonic and larval amphibians: variation in sensitivity among species. *Environ. Pollut.* 154:291–297.
- Snodgrass, J. W., W. A. Hopkins, J. Broughton, D. Gwinn, J. A. Baionno, and J. Burger. 2004. Species-specific responses of developing anurans to coal combustion wastes. *Aquat. Toxicol.* 66:171–182.
- Sparling, D. W., G. Linder, and C. A. Bishop. 2000. Ecotoxicology of amphibians and reptiles. Society of Environmental Toxicology and Chemistry (SETAC), Pensacola, FL.
- Sparling, D. W., G. M. Fellers, and L. L. Mc Connell. 2001. Pesticides and amphibian population declines in California, USA. *Environ. Toxicol. Chem.* 20:1591–1595.
- Sparling, D. W. 2000. Ecotoxicology of organic contaminants to amphibians. Pp 461–494 in D. W. Sparling, G. Linder, and C. A. Bishop, eds. Ecotoxicology of amphibians and reptiles. Society of Environmental Toxicology and Chemistry (SETAC), Pensacola, FL.
- Stuart, S. N., J. S. Chanson, N. A. Cox, B. E. Young, A. S. L. Rodríguez, D. L. Fischman, and R. M. Waller. 2004. Status and trends of amphibians declines and extinctions worldwide. *Science* 306:1783–1786.
- Tilman, D., J. Fargione, B. Wolff, C. D'Antonio, A. Dobson, R. Howarth, D. Schindler, W. H. Schlesinger, D. Simberloff, D. Swackhamer. 2001. Forecasting agriculturally driven global environmental change. *Science* 292:281–284.
- Veith, M., J. Kosuch, and M. Vences. 2003. Climatic oscillations triggered post-Messinian speciation of Western Palearctic brown frogs (Amphibia, Ranidae). *Mol. Phylogenet. Evol.* 26:310–327.
- Vitousek, P. M., J. Aber, R. W. Howarth, G. E. Likens, P. A. Matson, D. W. Schindler, W. H. Schlesinger, G. D. Tilman. 1997. Human alteration of the global nitrogen cycle: causes and consequences. *Ecol. Appl.* 7:737–750.
- Vredenburg, V. T. 2004. Reversing introduced species effects: experimental removal of introduced fish leads to rapid recovery of declining frog. *Proc. Natl. Acad. Sci. USA* 101:7646–7650.
- Wake, D. B., and V. T. Vredenburg. 2008. Are we in the midst of the sixth mass extinction? A view from the world of amphibians. *Proc. Natl. Acad. Sci. USA* 105:11466–11473.
- Watkins, T. B. 1996. Predator-mediated selection on burst swimming performance in tadpoles of the Pacific tree frog, *Pseudacris regilla*. *Physiol. Zool.* 69:154–167.
- Weisrock, D. W., T. J. Papenfuss, J. R. Macey, S. N. Litvinchuk, R. Polymeni, I. H. Ugurtas, E. Zhao, H. Howkar, and A. Larson. 2006. A molecular assessment of phylogenetic relationships and lineage accumulation rates within the family Salamandridae (Amphibia, Caudata). *Mol. Phylogenet. Evol.* 41:368–383.
- Widder, P. D., and J. R. Bidwell. 2006. Cholinesterase activity and behavior in chlorpyrifos-exposed *Rana sphenoccephala* tadpoles. *Environ. Toxicol. Chem.* 25:2446–2454.

- Wright, P. M., and P. A. Wright. 1996. Nitrogen metabolism and excretion in bullfrog (*Rana catesbeiana*) tadpoles and adults exposed to elevated environmental ammonia levels. *Physiol. Zool.* 69:1057–1078.
- Zajc, I., and J. W. Arntzen. 2000. Evolutionary relationships among European newts (genus *Triturus*) as inferred from two mtDNA fragments. *Eur. J. Physiol.* 439(Suppl.):R21–R22.
- Zhang, P., T. J. Papenfuss, M. H. Wake, L. Qu, and D. B. Wake. 2008. Phylogeny and biogeography of the family Salamandridae (Amphibia: Caudata) inferred from complete mitochondrial genomes. *Mol. Phylogenet. Evol.* 49:586–597.

## Supporting Information

Additional Supporting Information may be found online on Wiley Online Library.

**Table S1.** Summary information for each point sample included in the meta-analyses performed for each response variable.

Please note: Wiley-Blackwell is not responsible for the content or functionality of any supporting materials supplied by the authors. Any queries (other than missing material) should be directed to the corresponding author for the article.