

# Demo of new simple1 version

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## Overview

This demo shows:

1. basic usage,
2. variable substrates,
3. time-variable inputs,
4. speciation (proton-transfer reactions),
5. inhibition,
6. COD balance,

## Prep

```
devtools::load_all()
```

```
## i Loading ABM
```

## Function arguments

The argument list currently looks like this:

```
abm <- function(  
  days = 365,                # Number of days to run  
  delta_t = 1,               # Time step for output  
  times = NULL,              # Optional vector of times for output  
  mng_pars,  
  man_pars,  
  init_pars = list(conc_init = c(man_pars$comp_fresh, man_pars$VFA_fresh)),  
  grp_pars,  
  mic_pars,  
  sub_pars,  
  chem_pars,  
  inhib_pars = NULL,  
  mt_pars = NULL,  
  ctrl_pars = list(respir = TRUE,  
                    pH_inhib = FALSE,  
                    approx_method = 'early',  
                    par_key = '\\\\.',  
                    rates_calc = 'instant'),  
  var_pars = list(var = NULL),  
  add_pars = NULL,  
  pars = NULL,
```

```

startup = 0,                # Number of times complete simulation should be run before returning result
starting = NULL,           # Output from previous simulation to be starting condition for new one
value = 'ts',              # Type of output
warn = TRUE) {

```

The main changes in arguments are:

- new `sub_pars` for defining substrates
- new `ctrl_pars` for some “control” parameters
- new `var_pars` for *any* parameters that change over time

I have removed the default `*_pars` objects for now.

## 1. Basic behavior

The simplest usage is with constant slurry production rate and a fixed schedule. We need to set some parameters, first management parameters.

```

mng_pars = list(slurry_prod_rate = 10000,
                slurry_mass = 1000,
                storage_depth = 2,
                resid_depth = 0.1,
                area = 100,
                empty_int = 100,
                temp_C = 20,
                wash_water = 0,
                wash_int = NA,
                rest_d = 0,
                resid_enrich = 1)

```

Next substrate parameters, a new argument. This defines substrates. We could have any number with any names. Note that hydrolysis uses CTM again (like anything here, that could be changed).

```

sub_pars <- list(subs = c('VSd'),
                T_opt_hyd = c(VSd = 60),
                T_min_hyd = c(VSd = 0),
                T_max_hyd = c(VSd = 90),
                hydrol_opt = c(VSd = 0.1),
                sub_fresh = c(VSd = 50),
                sub_init = c(VSd = 50))

```

Microbial parameters are similar to other ABM versions, but inhibition is set separately now.

```

grp_pars <- list(grps = c('m0', 'm1', 'm2', 'sr1'),
                yield = c(default = 0.05, sr1 = 0.065),
                xa_fresh = c(all = 0.05),
                xa_init = c(all = 0.05),
                dd_rate = c(all = 0.02),
                ks = c(default = 1, sr1 = 0.5),
                qhat_opt = c(m0 = 1, m1 = 1, m2 = 2, sr1 = 9),
                T_opt = c(m0 = 18, m1 = 18, m2 = 28, sr1 = 44),
                T_min = c(m0 = 0, m1 = 6.41, m2 = 6.41, sr1 = 0),
                T_max = c(m0 = 25, m1 = 25, m2 = 38, sr1 = 51))

```

The `dd_rate_xa` parameter is for “death and decay”.

```

mic_pars <- list(dd_rate_xa = 0.02)

```

These last two arguments are similar to other versions. VFA is hard-wired and so has its own elements.

```
man_pars <- list(VFA_fresh = c(VFA = 2), pH = 7, dens = 1000)
```

```
chem_pars <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
                              VFA = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
                              CO2_aer = 1/0.436, CO2_sr = 1/1.2,
                              C_xa = 1/0.3753125))
```

```
out1 <- abm(365,
            mng_pars = mng_pars,
            man_pars = man_pars,
            grp_pars = grp_pars,
            mic_pars = mic_pars,
            sub_pars = sub_pars,
            chem_pars = chem_pars)
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 1.7%
```

Output is similar to other versions. (The value argument does not currently work.)

```
head(out1)
```

```
##      time      m0      m1      m2      sr1      VSd      VFA slurry_mass
## 1      0  50.0000  50.0000  50.0000  50.0000  50000.0  2000.00      1000
## 2      1  554.0098  553.8533  558.3748  544.0431  542318.4  29018.66     11000
## 3      2 1066.2767 1065.6732 1083.1940 1028.3035 1022076.9  67371.44     21000
## 4      3 1588.3161 1586.9430 1627.0197 1502.9749 1489597.5 116611.44     31000
## 5      4 2121.2034 2118.7114 2191.8594 1968.2472 1945194.0 176326.70     41000
## 6      5 2665.7726 2661.7877 2779.4361 2424.3064 2389172.2 246127.08     51000
##      CH4_emis_cum CO2_emis_cum slurry_load COD_load CH4_emis_rate temp_C pH m0_eff
## 1      0.0000      0.0000      0      0      25.52844      20 7      0
## 2     163.6272     130.5362     10000  522000     308.65512      20 7      0
## 3     628.8119     501.6449     20000 1044000     626.59326      20 7      0
## 4    1425.4337    1137.1629     30000 1566000     970.52564      20 7      0
## 5    2577.0208    2055.8603     40000 2088000    1335.99741      20 7      0
## 6    4103.8067    3273.8785     50000 2610000    1720.64113      20 7      0
##      m1_eff m2_eff sr1_eff VSd_eff VFA_eff slurry_mass_eff slurry_depth m0_conc
## 1      0      0      0      0      0      0      0.01 0.05000000
## 2      0      0      0      0      0      0      0.11 0.05036453
## 3      0      0      0      0      0      0      0.21 0.05077508
## 4      0      0      0      0      0      0      0.31 0.05123600
## 5      0      0      0      0      0      0      0.41 0.05173667
## 6      0      0      0      0      0      0      0.51 0.05227005
##      m1_conc m2_conc sr1_conc VSd_conc VFA_conc m0_eff_conc m1_eff_conc
## 1 0.05000000 0.05000000 0.05000000 50.00000 2.000000      NaN      NaN
## 2 0.05035030 0.05076135 0.04945846 49.30167 2.638060      NaN      NaN
## 3 0.05074634 0.05158067 0.04896683 48.67033 3.208164      NaN      NaN
## 4 0.05119171 0.05248451 0.04848306 48.05153 3.761660      NaN      NaN
## 5 0.05167589 0.05345999 0.04800603 47.44376 4.300651      NaN      NaN
## 6 0.05219192 0.05449875 0.04753542 46.84651 4.826021      NaN      NaN
##      m2_eff_conc sr1_eff_conc VSd_eff_conc VFA_eff_conc
## 1      NaN      NaN      NaN      NaN
## 2      NaN      NaN      NaN      NaN
## 3      NaN      NaN      NaN      NaN
```

```
## 4      NaN      NaN      NaN      NaN
## 5      NaN      NaN      NaN      NaN
## 6      NaN      NaN      NaN      NaN
```

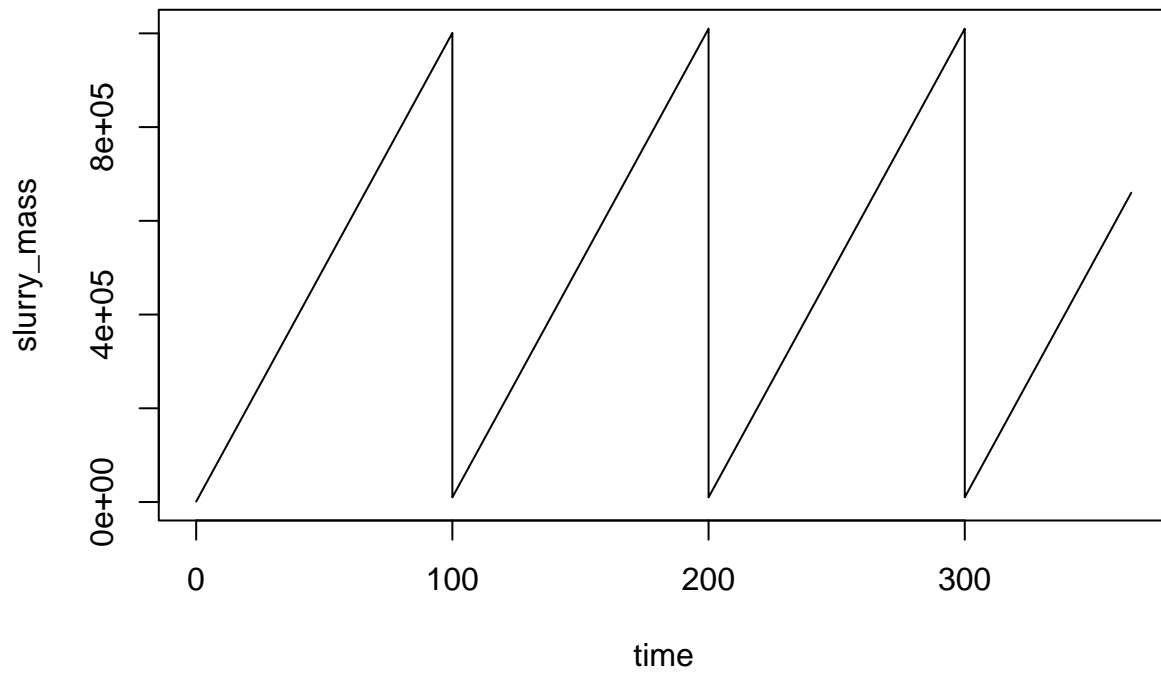
```
tail(out1)
```

```
##      time      m0      m1      m2      sr1      VSd      VFA slurry_mass
## 364  360 74909.98 72497.35 337403.6 17643.06 15468816 4796289      610000
## 365  361 77056.84 74543.26 351549.2 17788.74 15576604 4684393      620000
## 366  362 79234.75 76617.42 366160.3 17931.53 15682002 4557322      630000
## 367  363 81441.36 78717.58 381233.0 18071.50 15785084 4414981      640000
## 368  364 83673.78 80840.93 396758.5 18208.69 15885918 4257395      650000
## 369  365 85928.37 82984.01 412722.1 18343.17 15984571 4084743      660000
##      CH4_emis_cum CO2_emis_cum slurry_load  COD_load CH4_emis_rate temp_C pH
## 364      26846605      21417315      3600000 187920000      125134.1    20  7
## 365      26973866      21518840      3610000 188442000      129394.2    20  7
## 366      27105400      21623774      3620000 188964000      133674.8    20  7
## 367      27241214      21732122      3630000 189486000      137950.5    20  7
## 368      27381289      21843868      3640000 190008000      142189.3    20  7
## 369      27525568      21958969      3650000 190530000      146351.3    20  7
##      m0_eff  m1_eff  m2_eff  sr1_eff  VSd_eff  VFA_eff  slurry_mass_eff
## 364 441740.7 422210.1 2239891 63286.93 53531054 3419880      2991000
## 365 441740.7 422210.1 2239891 63286.93 53531054 3419880      2991000
## 366 441740.7 422210.1 2239891 63286.93 53531054 3419880      2991000
## 367 441740.7 422210.1 2239891 63286.93 53531054 3419880      2991000
## 368 441740.7 422210.1 2239891 63286.93 53531054 3419880      2991000
## 369 441740.7 422210.1 2239891 63286.93 53531054 3419880      2991000
##      slurry_depth  m0_conc  m1_conc  m2_conc  sr1_conc  VSd_conc  VFA_conc
## 364      6.1 0.1228032 0.1188481 0.5531206 0.02892305 25.35872 7.862769
## 365      6.2 0.1242852 0.1202311 0.5670148 0.02869152 25.12355 7.555472
## 366      6.3 0.1257694 0.1216150 0.5812069 0.02846275 24.89207 7.233845
## 367      6.4 0.1272521 0.1229962 0.5956766 0.02823671 24.66419 6.898408
## 368      6.5 0.1287289 0.1243707 0.6103976 0.02801337 24.43987 6.549838
## 369      6.6 0.1301945 0.1257334 0.6253365 0.02779268 24.21905 6.189004
##      m0_eff_conc  m1_eff_conc  m2_eff_conc  sr1_eff_conc  VSd_eff_conc  VFA_eff_conc
## 364      0.14769 0.1411602 0.7488771 0.02115912 17.89738 1.14339
## 365      0.14769 0.1411602 0.7488771 0.02115912 17.89738 1.14339
## 366      0.14769 0.1411602 0.7488771 0.02115912 17.89738 1.14339
## 367      0.14769 0.1411602 0.7488771 0.02115912 17.89738 1.14339
## 368      0.14769 0.1411602 0.7488771 0.02115912 17.89738 1.14339
## 369      0.14769 0.1411602 0.7488771 0.02115912 17.89738 1.14339
```

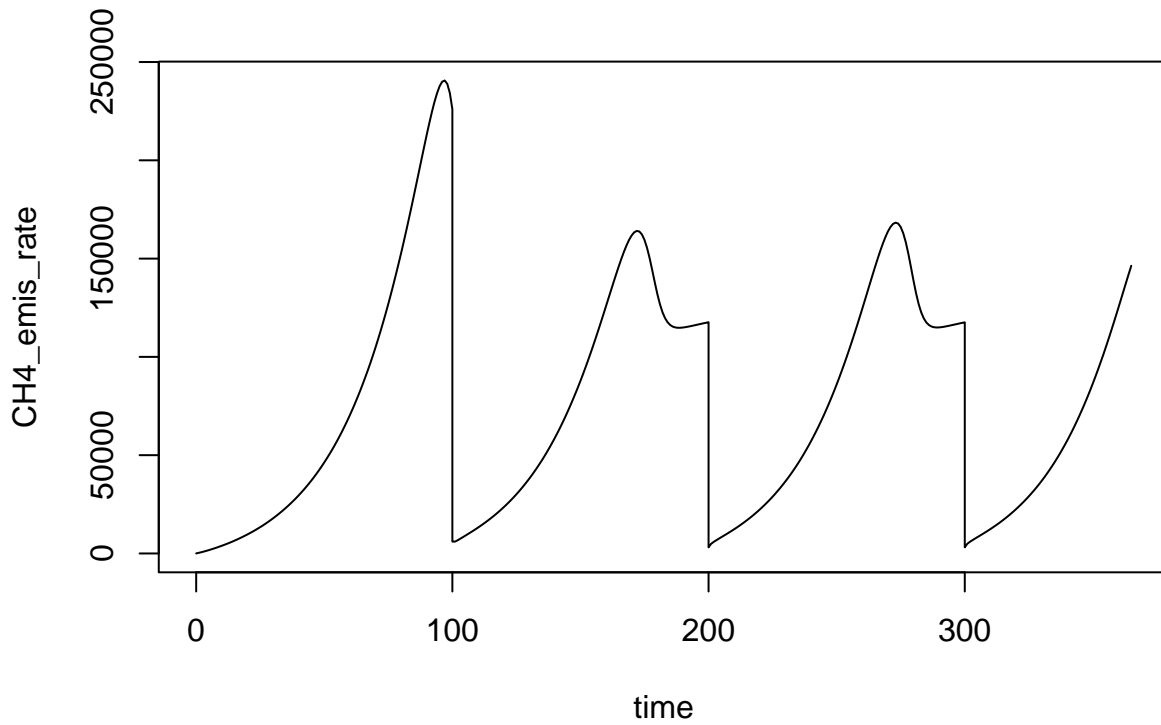
The effluent columns are cumulative. Is this what we had before? I did it for COD balance checking. We will have to discuss what is needed.

Here are some results.

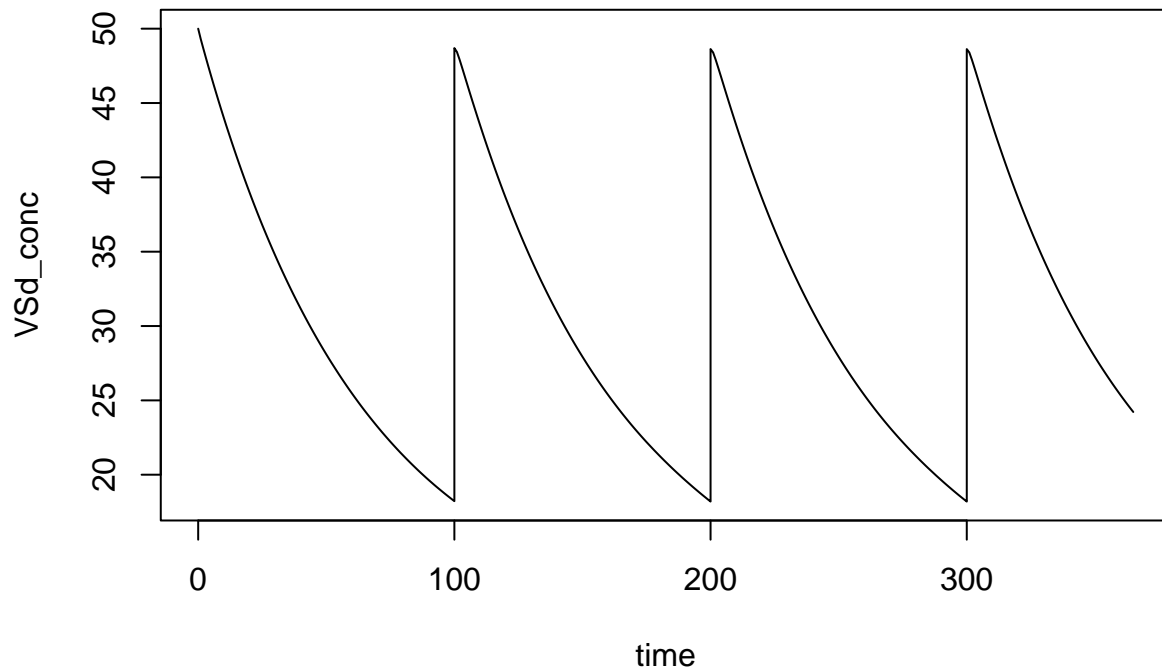
```
plot(slurry_mass ~ time, data = out1, type = 'l')
```



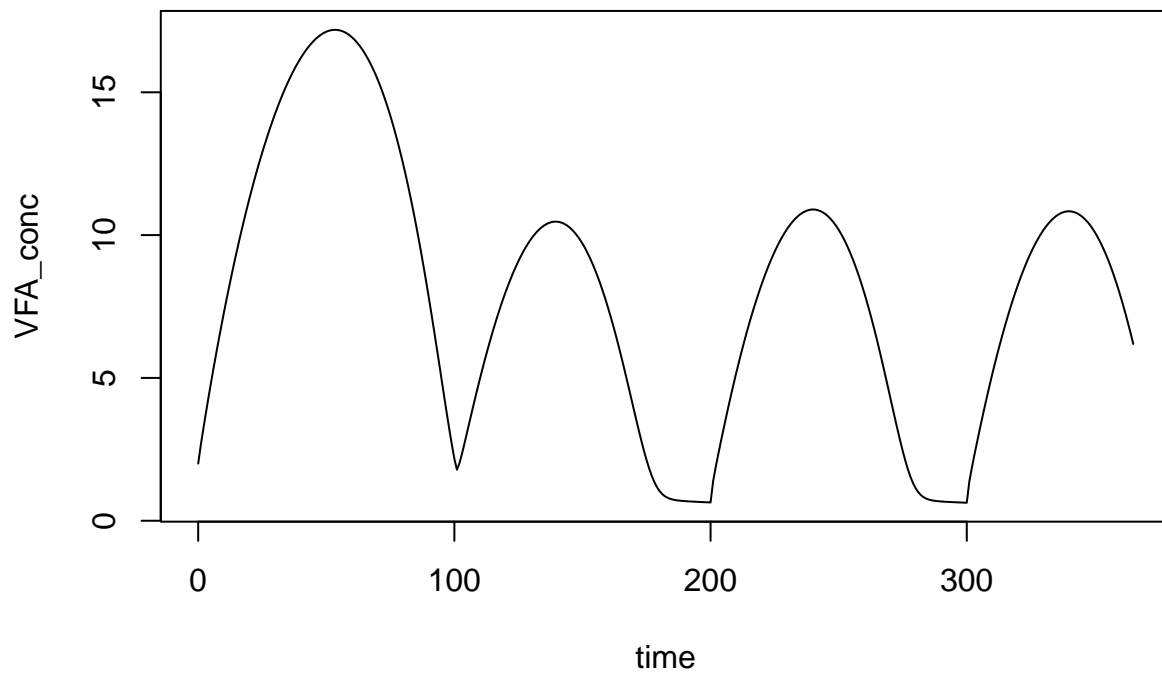
```
plot(CH4_emis_rate ~ time, data = out1, type = 'l')
```



```
plot(VSd_conc ~ time, data = out1, type = 'l')
```

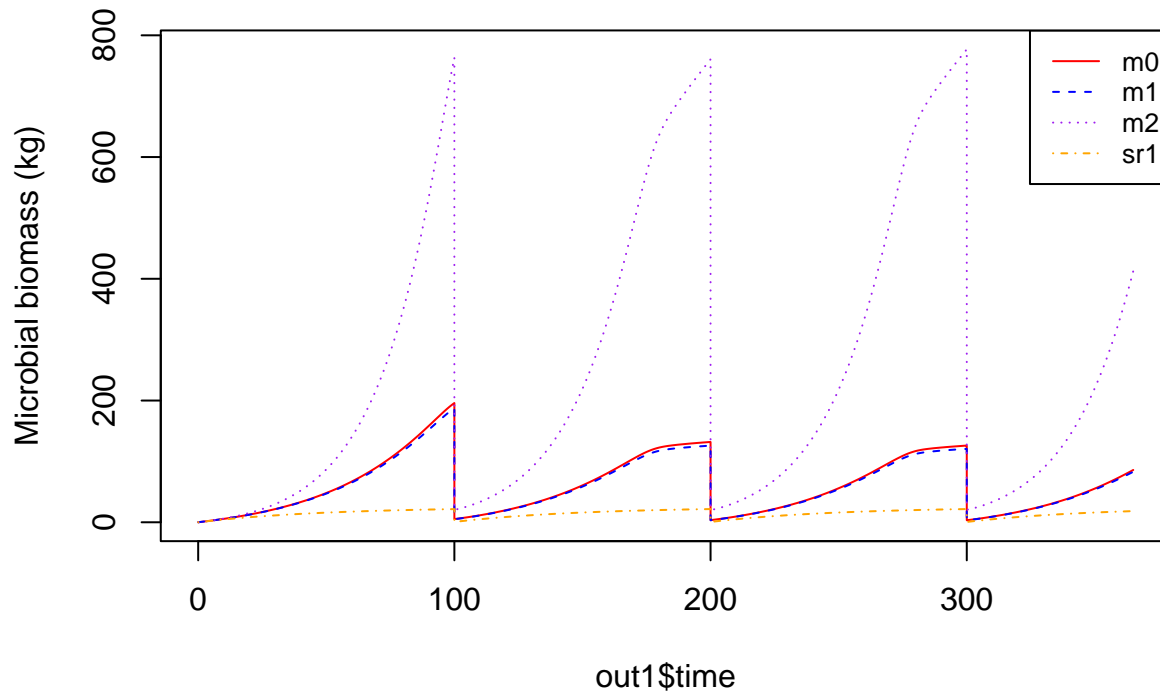


```
plot(VFA_conc ~ time, data = out1, type = 'l')
```



And methanogens.

```
line_colors <- c('red', 'blue', 'purple', 'orange')
matplot(out1$time, out1[, nn <- c('m0', 'm1', 'm2', 'sr1')]/1000,
        type = 'l', lty = c(1:length(nn)), col = line_colors, ylab = 'Microbial biomass (kg)')
legend("topright", legend = nn, lty = c(1:length(nn)), col = line_colors, lwd = 1, cex = 0.8)
```



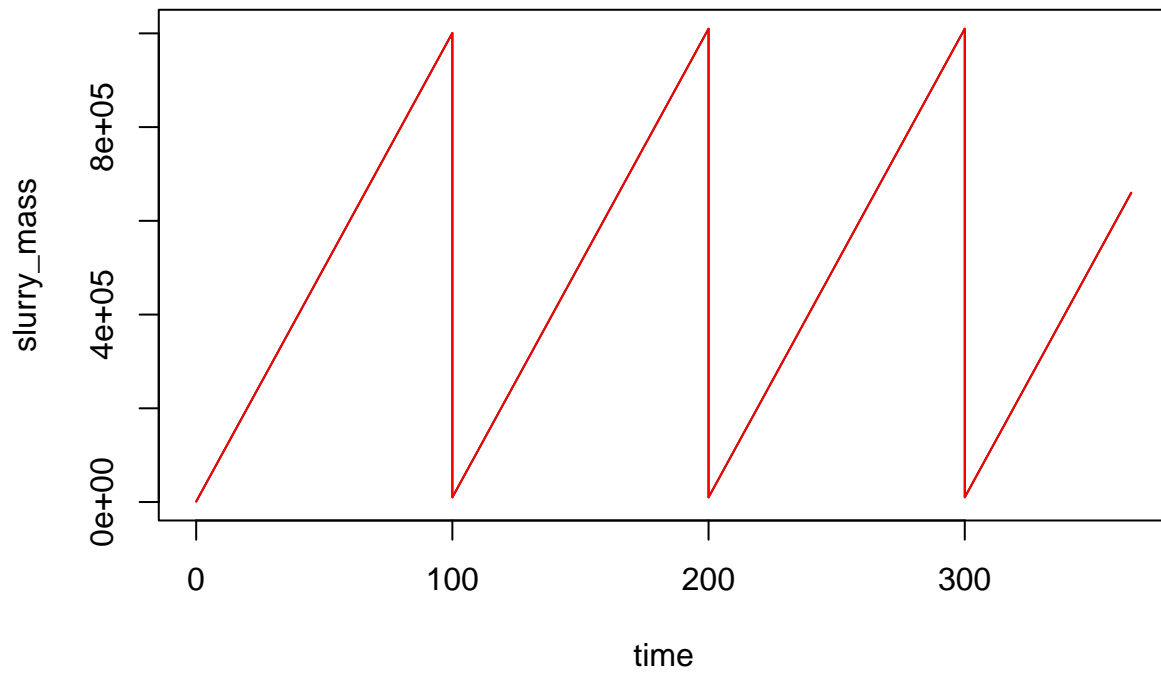
## 2. Substrate flexibility

Particulate substrates are defined in `sub_pars` now and there are no specific substrates hard-wired in the code. VFA is the only intermediate, and it is hard-wired. Here we will use three substrates. Parameter values have no connection to reality.

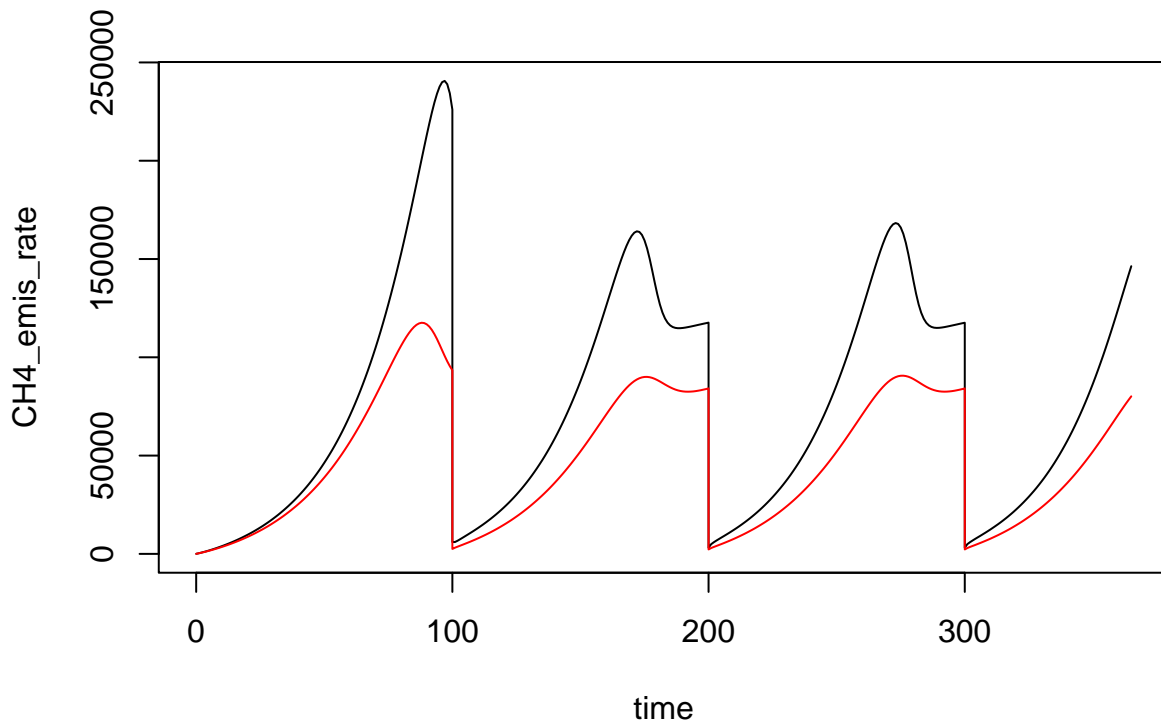
```
sub_pars2 <- list(subs = c('cellulose', 'protein', 'lipids'),
  T_opt_hyd = c(all = 60),
  T_min_hyd = c(all = 0),
  T_max_hyd = c(all = 90),
  hydrol_opt = c(lipids = 0.1, protein = 0.01, cellulose = 0.05),
  sub_fresh = c(lipids = 3, protein = 20, cellulose = 35),
  sub_init = c(lipids = 3, protein = 20, cellulose = 35))
```

```
out2 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars2,
  chem_pars = chem_pars)
```

```
plot(slurry_mass ~ time, data = out2, type = 'l')
lines(slurry_mass ~ time, data = out1, col = 'red')
```

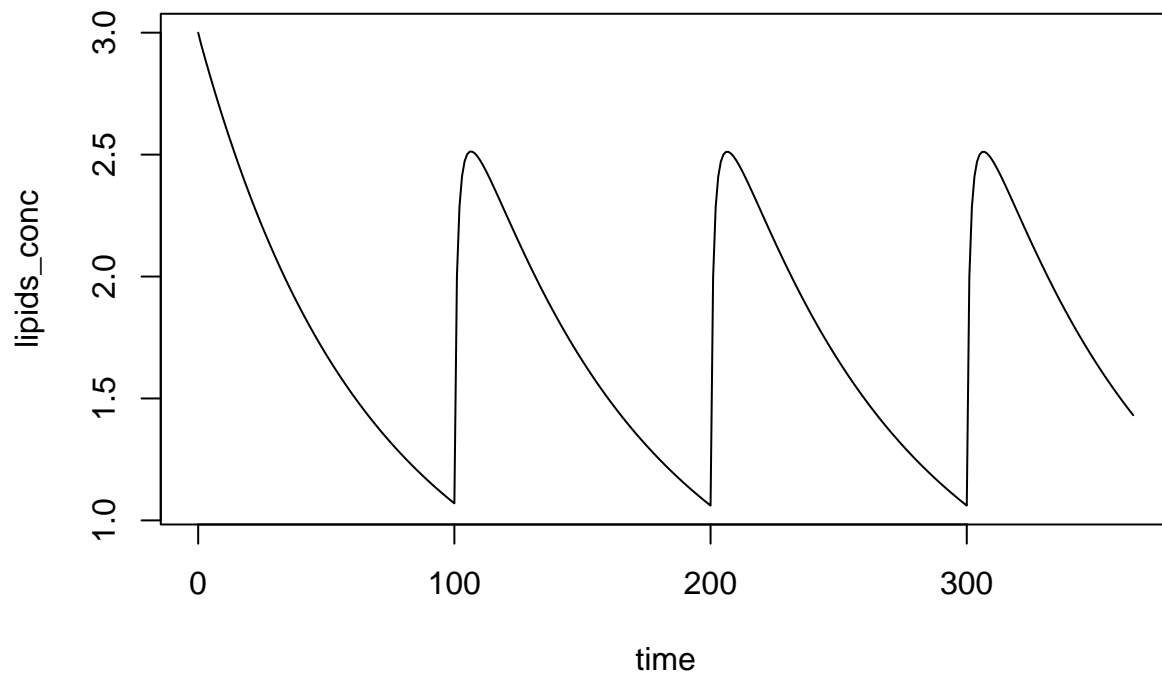


```
plot(CH4_emis_rate ~ time, data = out1, type = 'l')  
lines(CH4_emis_rate ~ time, data = out2, col = 'red')
```

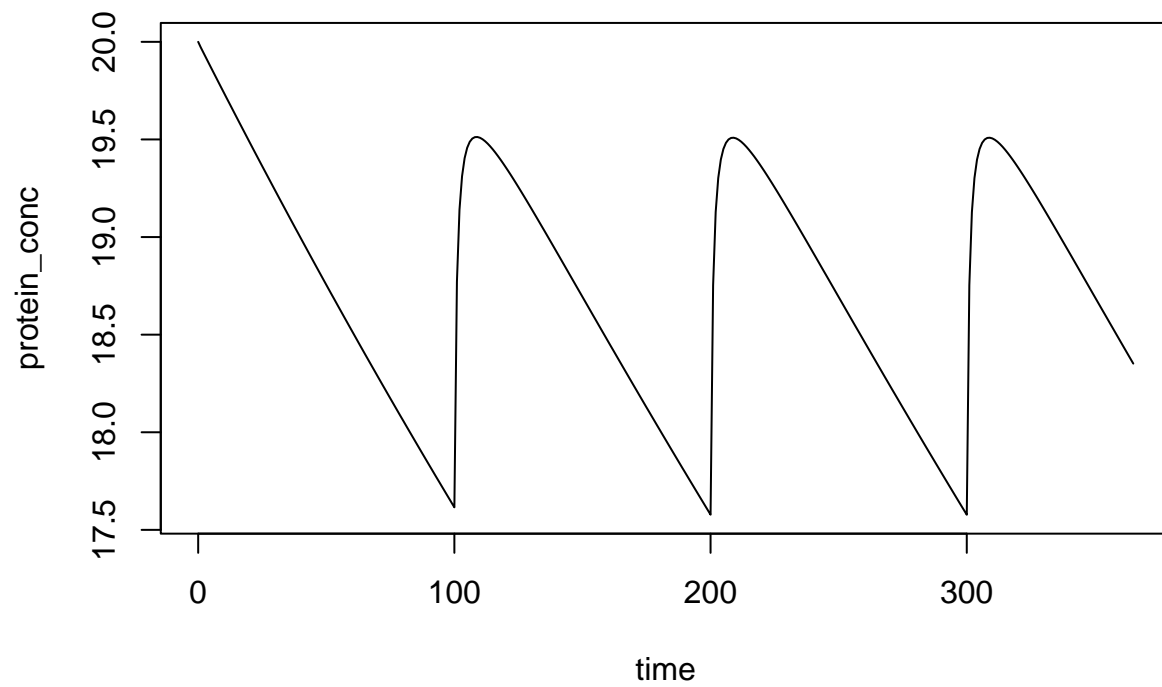


```
plot(lipids_conc ~ time, data = out2, type = 'l')
```

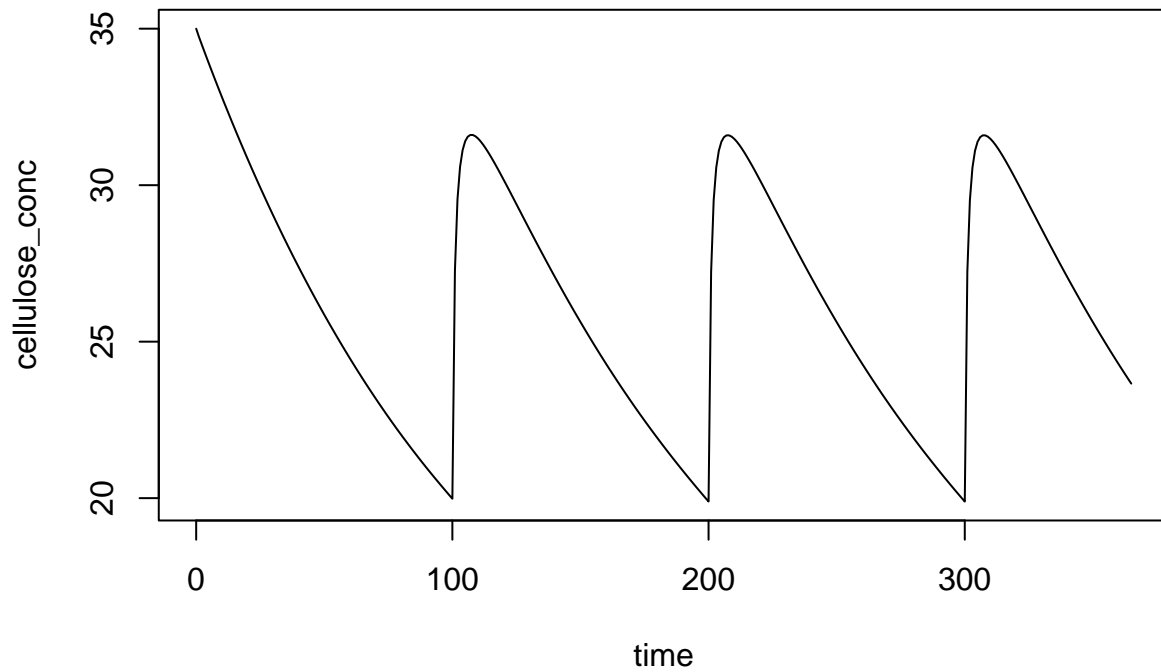




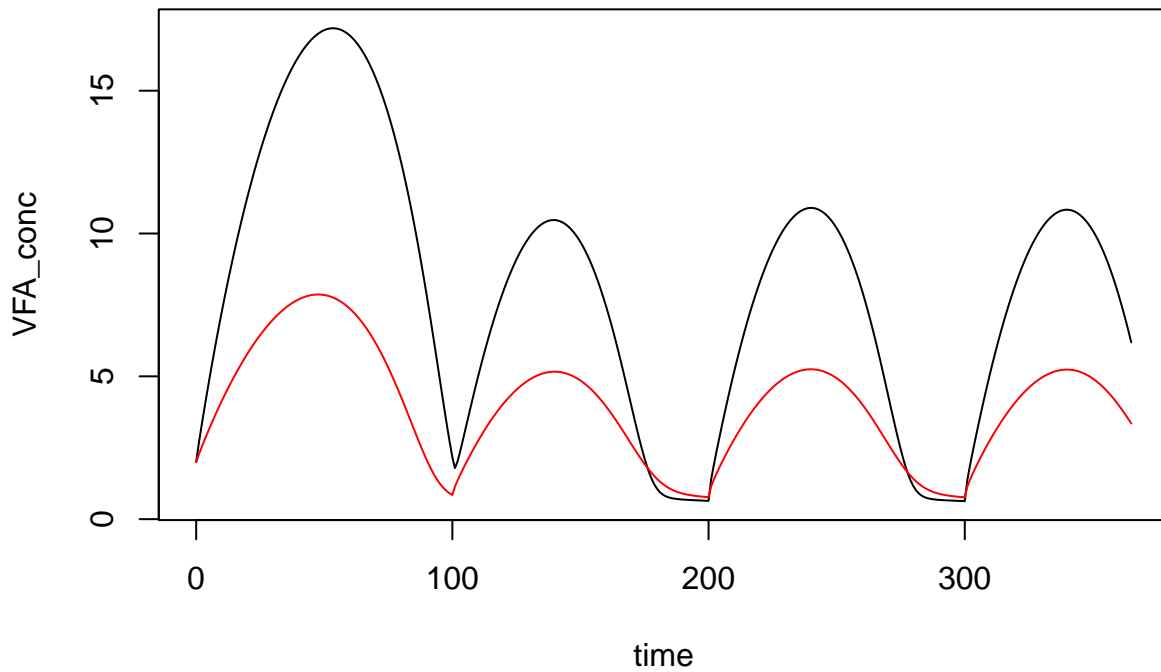
```
plot(protein_conc ~ time, data = out2, type = 'l')
```



```
plot(cellulose_conc ~ time, data = out2, type = 'l')
```



```
plot(VFA_conc ~ time, data = out1, type = 'l')
lines(VFA_conc ~ time, data = out2, col = 'red')
```



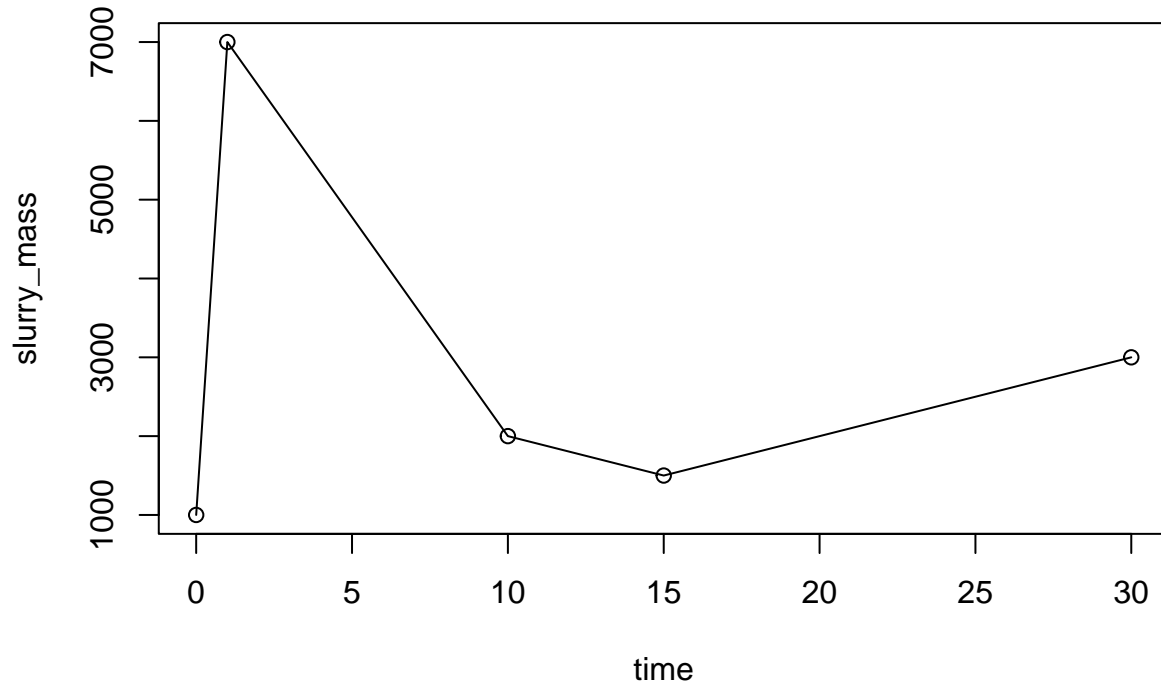
This flexibility comes from an approach similar to what we used for microbial groups.

### 3. Time-variable inputs part 1

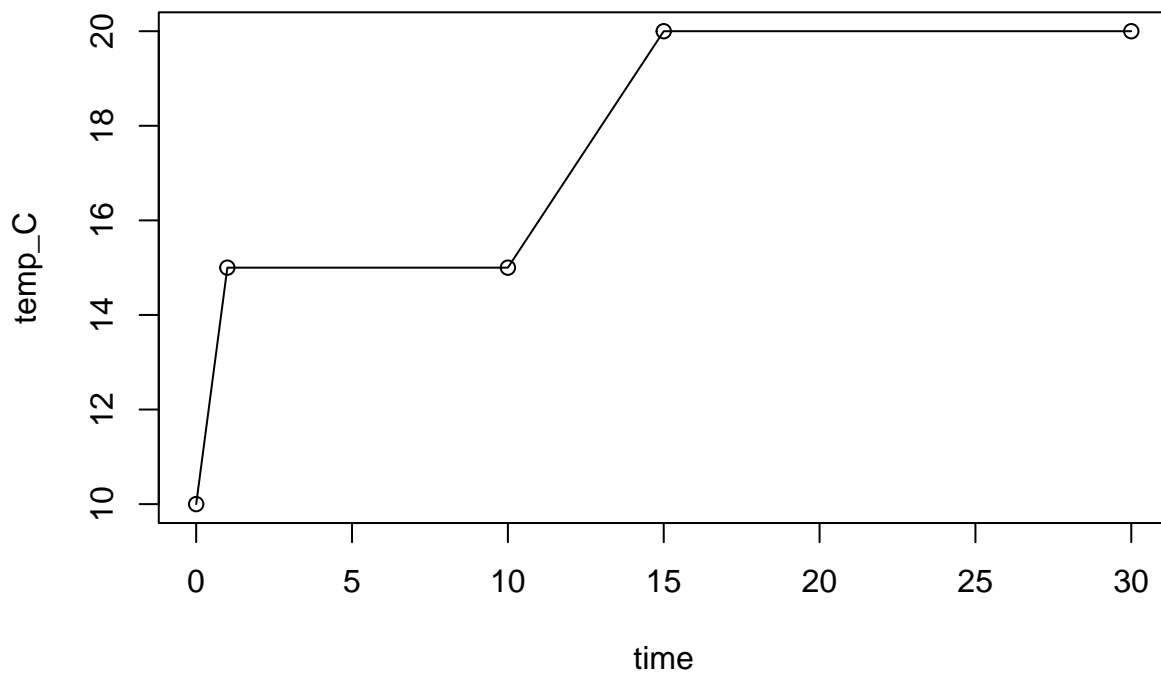
The `abm()` function can handle variability over time in any inputs now. Here slurry mass and temperature will vary.

```
var_dat <- data.frame(time = c(0, 1, 10, 15, 30),
                      slurry_mass = c(1000, 7000, 2000, 1500, 3000),
                      temp_C = c(10, 15, 15, 20, 20))

plot(slurry_mass ~ time, data = var_dat, type = 'o')
```



```
plot(temp_C ~ time, data = var_dat, type = 'o')
```

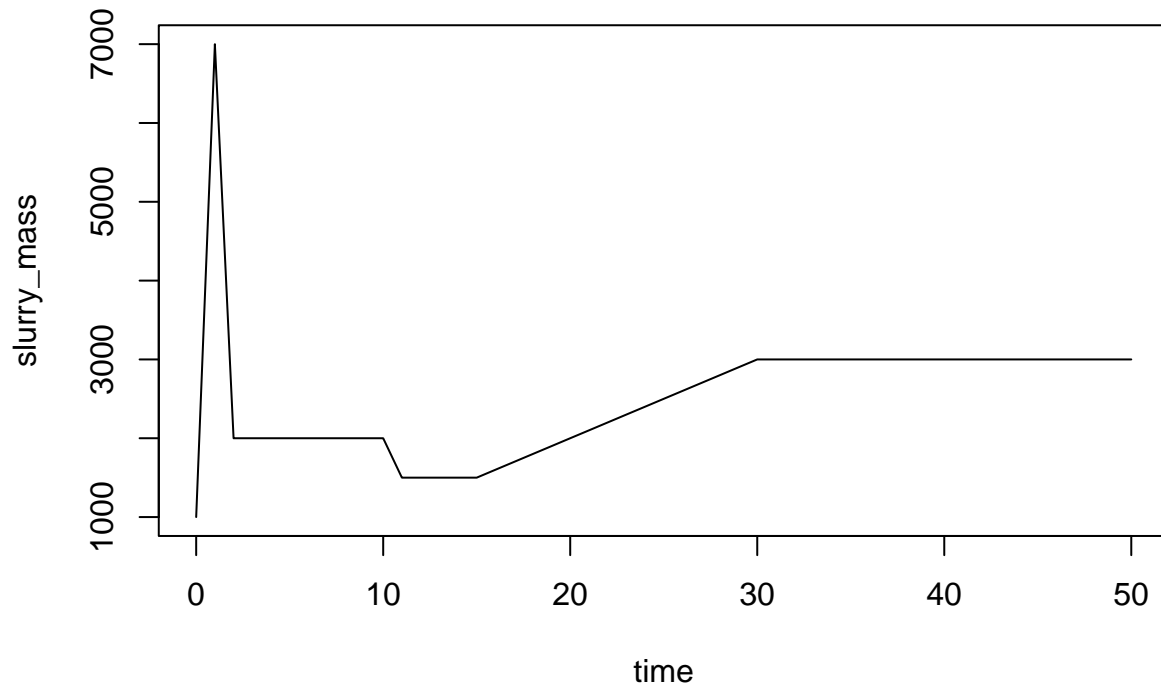


This data frame goes in the `var_pars` argument, which must be a list, even though it might have a single element named `var`. The `var` element is the only required one. The `var` data frame must have a `slurry_mass` column if it is used—it is not possible to use an `abm_regular()`-like approach with variable temperature etc.

```
var_pars <- list(var = var_dat)
```

```
out3a <- abm(50,  
  mng_pars = mng_pars,  
  man_pars = man_pars,  
  grp_pars = grp_pars,  
  mic_pars = mic_pars,  
  sub_pars = sub_pars,  
  chem_pars = chem_pars,  
  var_pars = var_pars)
```

```
plot(slurry_mass ~ time, data = out3a, type = 'l')
```



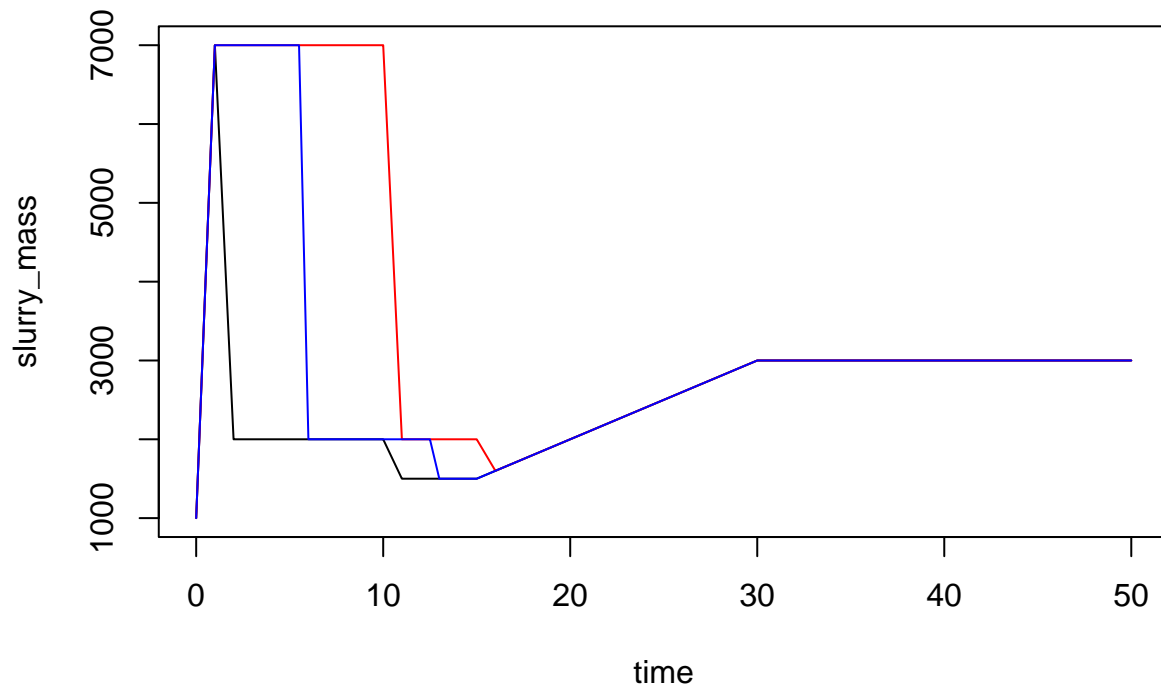
The “late” and “mid” options are still available, but now through `ctrl_pars`. Here we can change the value through `add_pars`

```
out3b <- abm(50,  
  mng_pars = mng_pars,  
  man_pars = man_pars,  
  grp_pars = grp_pars,  
  mic_pars = mic_pars,  
  sub_pars = sub_pars,  
  chem_pars = chem_pars,  
  var_pars = var_pars,  
  add_pars = list(approx_method = 'late'))
```

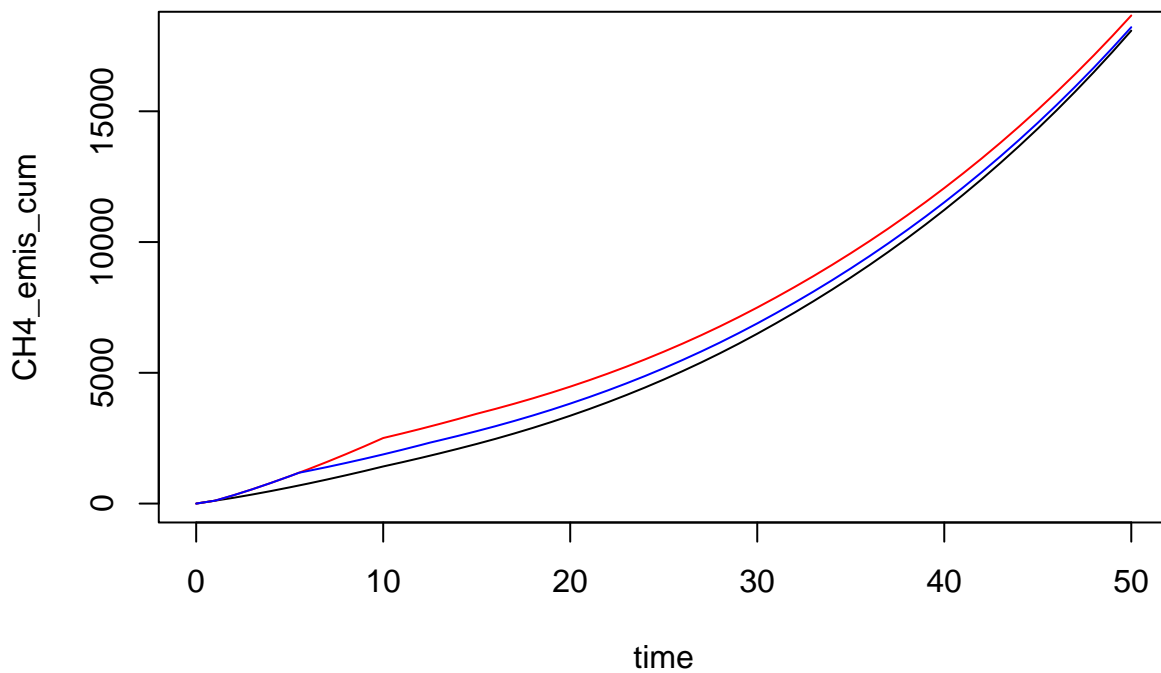
```
out3c <- abm(50,  
  mng_pars = mng_pars,  
  man_pars = man_pars,  
  grp_pars = grp_pars,  
  mic_pars = mic_pars,  
  sub_pars = sub_pars,  
  chem_pars = chem_pars,
```

```
var_pars = var_pars,  
add_pars = list(approx_method = 'mid'))
```

```
plot(slurry_mass ~ time, data = out3a, type = 'l')  
lines(slurry_mass ~ time, data = out3b, col = 'red')  
lines(slurry_mass ~ time, data = out3c, col = 'blue')
```



```
plot(CH4_emis_cum ~ time, data = out3a, type = 'l')  
lines(CH4_emis_cum ~ time, data = out3b, col = 'red')  
lines(CH4_emis_cum ~ time, data = out3c, col = 'blue')
```



## 4. Time-variable inputs part 2

Here we'll vary fresh substrate concentrations over time.

First the data frame with slurry mass.

```
var_dat <- data.frame(time = c(0, 1, 10, 15, 30, 50),
                      slurry_mass = c(1000, 7000, 2000, 5000, 3000, 10000))
```

var\_dat

```
##   time slurry_mass
## 1    0         1000
## 2    1         7000
## 3   10         2000
## 4   15         5000
## 5   30         3000
## 6   50        10000
```

Then add `sub_fresh` values. Each row needs a list containing a named vector. This is somewhat unusual data frame usage, and there is a user-friendly alternative based on additional data frames in the `var` argument (see next section). But I've kept this demo.

var\_dat

```
##   time slurry_mass
## 1    0         1000
## 2    1         7000
## 3   10         2000
## 4   15         5000
## 5   30         3000
## 6   50        10000
```

```
var_dat$sub_fresh <- rep(list(c(VSd = 50)), nrow(var_dat))
var_dat$sub_fresh[3] <- list(c(VSd = 100))
var_dat$sub_fresh[4] <- list(c(VSd = 10))
var_dat$sub_fresh[5] <- list(c(VSd = 0))
var_dat$sub_fresh[6] <- list(c(VSd = 200))
var_dat
```

```
##   time slurry_mass sub_fresh
## 1    0         1000        50
## 2    1         7000        50
## 3   10         2000       100
## 4   15         5000        10
## 5   30         3000         0
## 6   50        10000       200
```

var\_dat[1, 3]

```
## [[1]]
## VSd
## 50
```

var\_dat[5, 3]

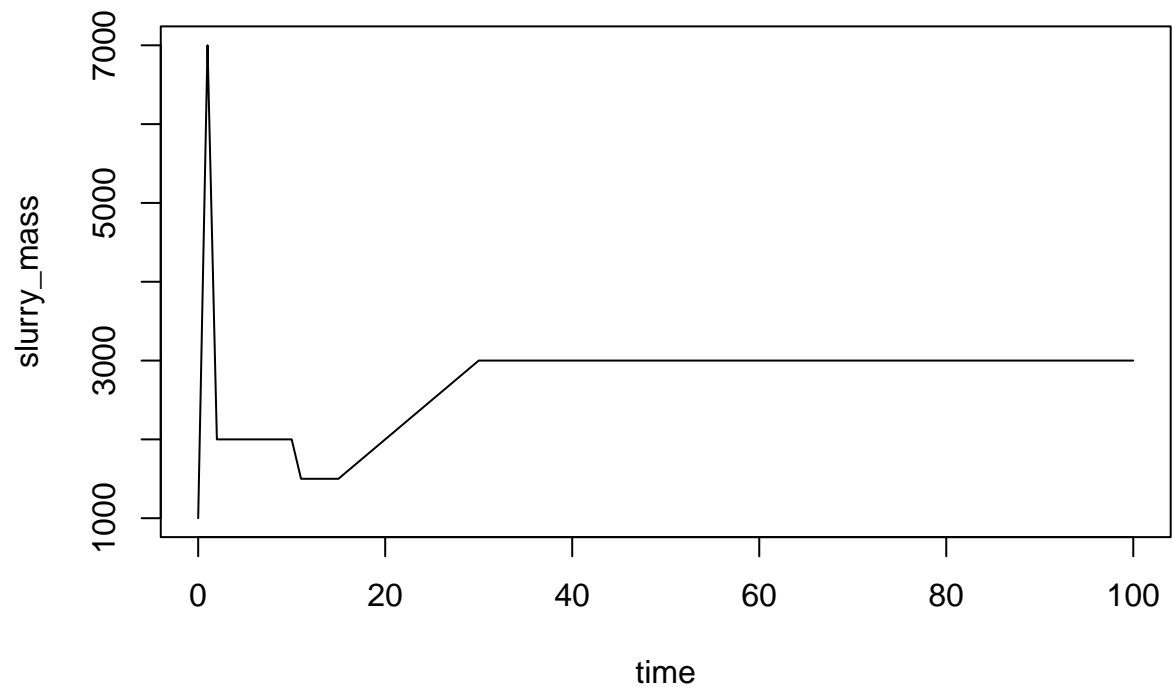
```
## [[1]]
## VSd
## 0
```

```
devtools::load_all()
```

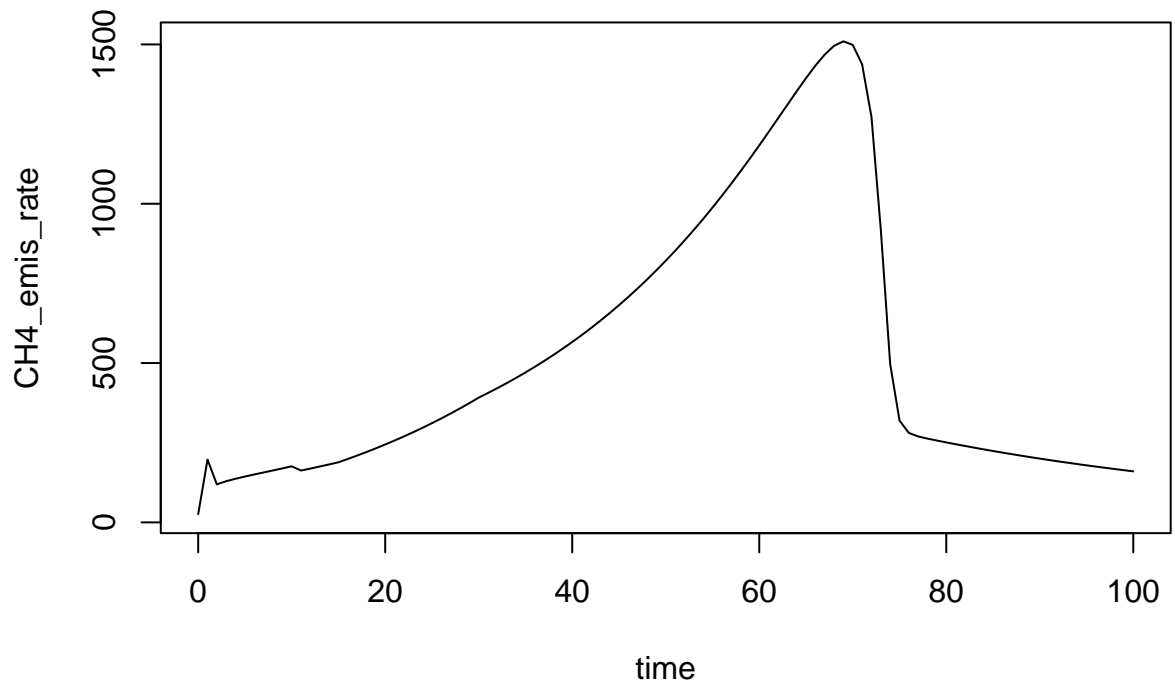
```
## i Loading ABM
```

```
out4a <- abm(100,  
  mng_pars = mng_pars,  
  man_pars = man_pars,  
  grp_pars = grp_pars,  
  mic_pars = mic_pars,  
  sub_pars = sub_pars,  
  chem_pars = chem_pars,  
  var_pars = var_pars)
```

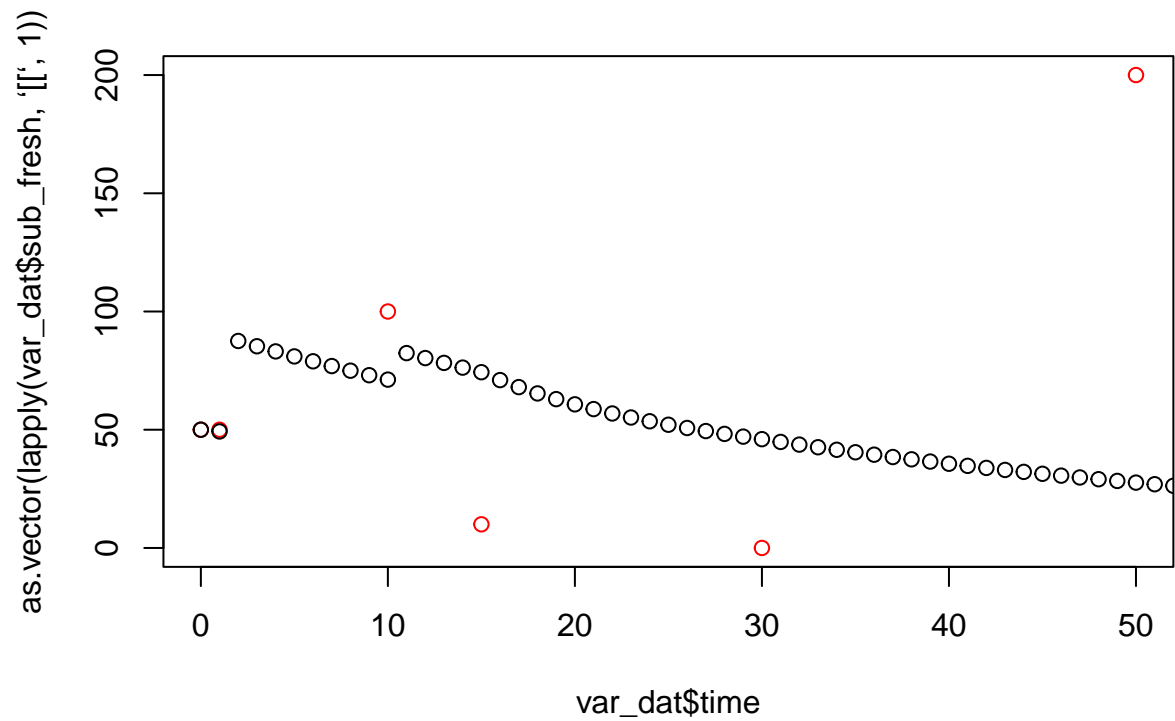
```
plot(slurry_mass ~ time, data = out4a, type = 'l')
```



```
plot(CH4_emis_rate ~ time, data = out4a, type = 'l')
```

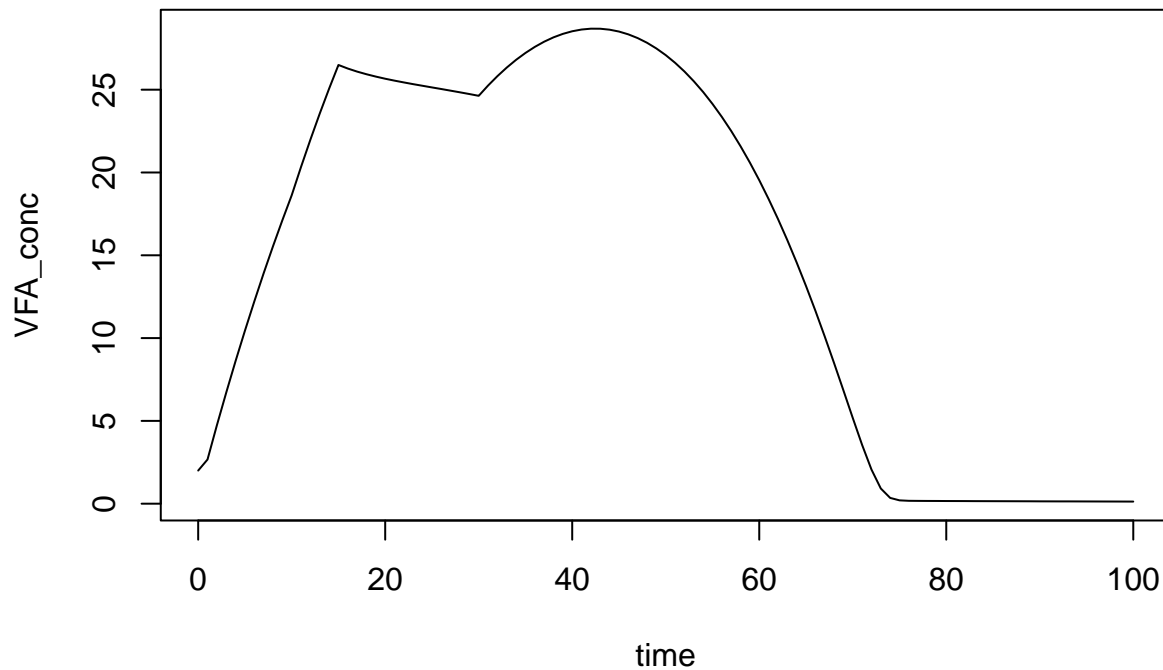


```
plot(var_dat$time, as.vector(lapply(var_dat$sub_fresh, `[`, 1)), type = 'p', col = 'red')
lines(VSd_conc ~ time, data = out4a, type = 'p')
```



```
plot(VFA_conc ~ time, data = out4a, type = 'l')
```





Let's vary some microbial parameters as well. And pH.

```
var_dat <- data.frame(time = c(0, 1, 10, 15, 30, 50),
                      slurry_mass = c(1000, 7000, 2000, 5000, 3000, 10000),
                      pH = 7 - 0:5/10)
var_dat
```

```
##   time slurry_mass  pH
## 1    0         1000 7.0
## 2    1         7000 6.9
## 3   10         2000 6.8
## 4   15         5000 6.7
## 5   30         3000 6.6
## 6   50        10000 6.5
```

VSd.

```
var_dat$sub_fresh <- rep(list(c(VSd = 50)), nrow(var_dat))
var_dat$sub_fresh[3] <- list(c(VSd = 100))
```

Some microbial parameters for a shift in temperature optima, "adaptation" for example.

```
for (i in 1:nrow(var_dat)) {
  var_dat$T_opt[i] <- list(grp_pars$T_opt + 2 * i)
}
var_dat
```

```
##   time slurry_mass  pH sub_fresh      T_opt
## 1    0         1000 7.0         50 20, 20, 30, 46
## 2    1         7000 6.9         50 22, 22, 32, 48
## 3   10         2000 6.8        100 24, 24, 34, 50
## 4   15         5000 6.7         50 26, 26, 36, 52
## 5   30         3000 6.6         50 28, 28, 38, 54
## 6   50        10000 6.5         50 30, 30, 40, 56
```

```

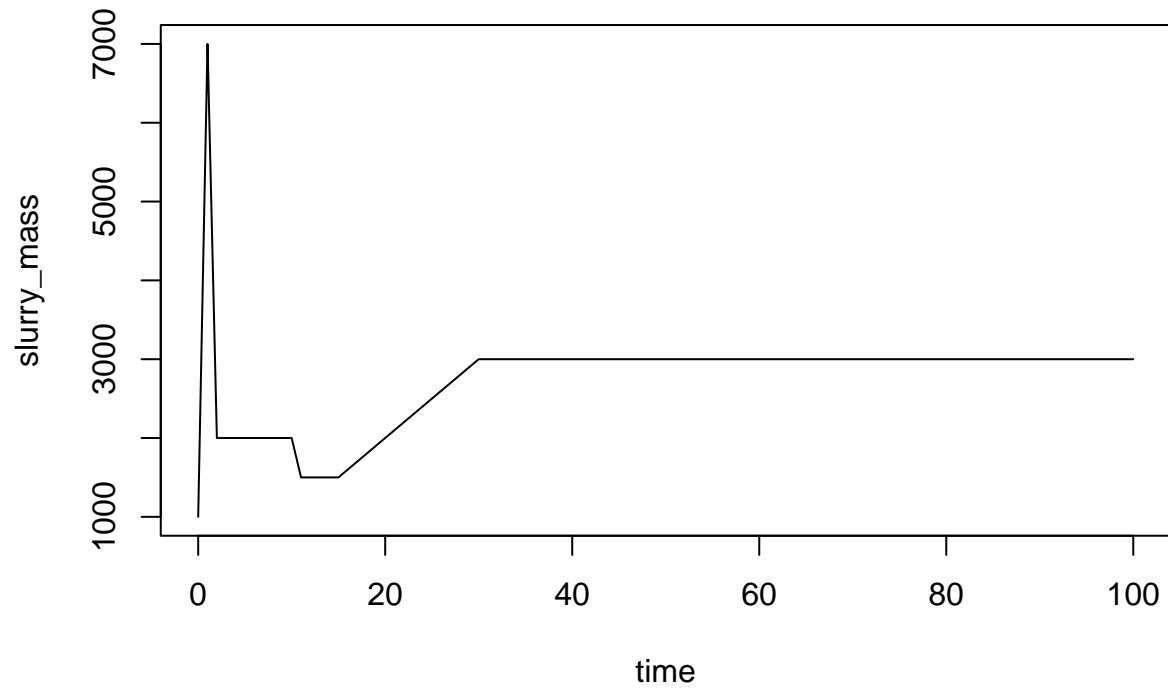
out4b <- abm(100,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars,
  var_pars = var_pars)

```

```

plot(slurry_mass ~ time, data = out4b, type = 'l')

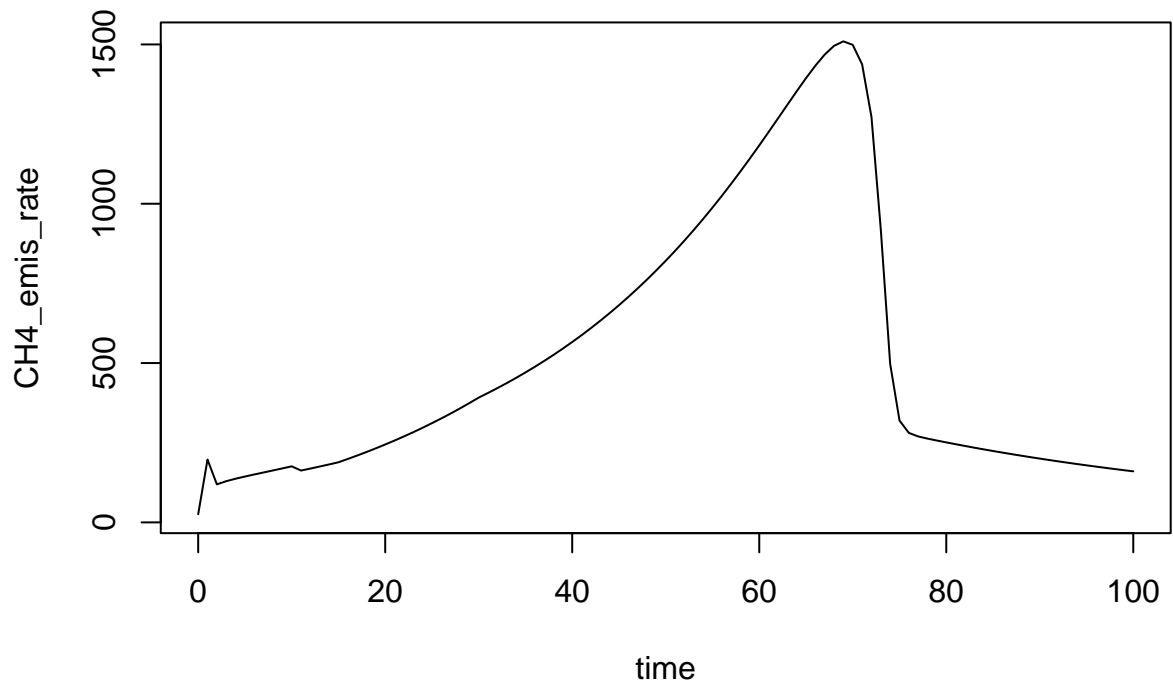
```



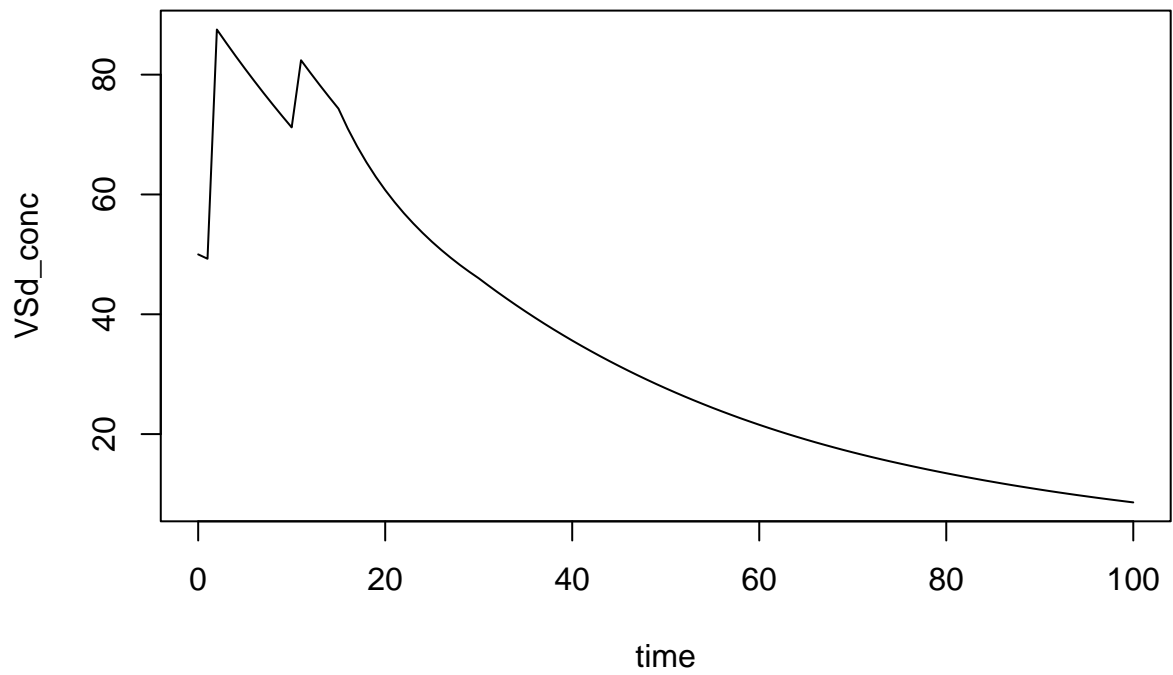
```

plot(CH4_emis_rate ~ time, data = out4b, type = 'l')

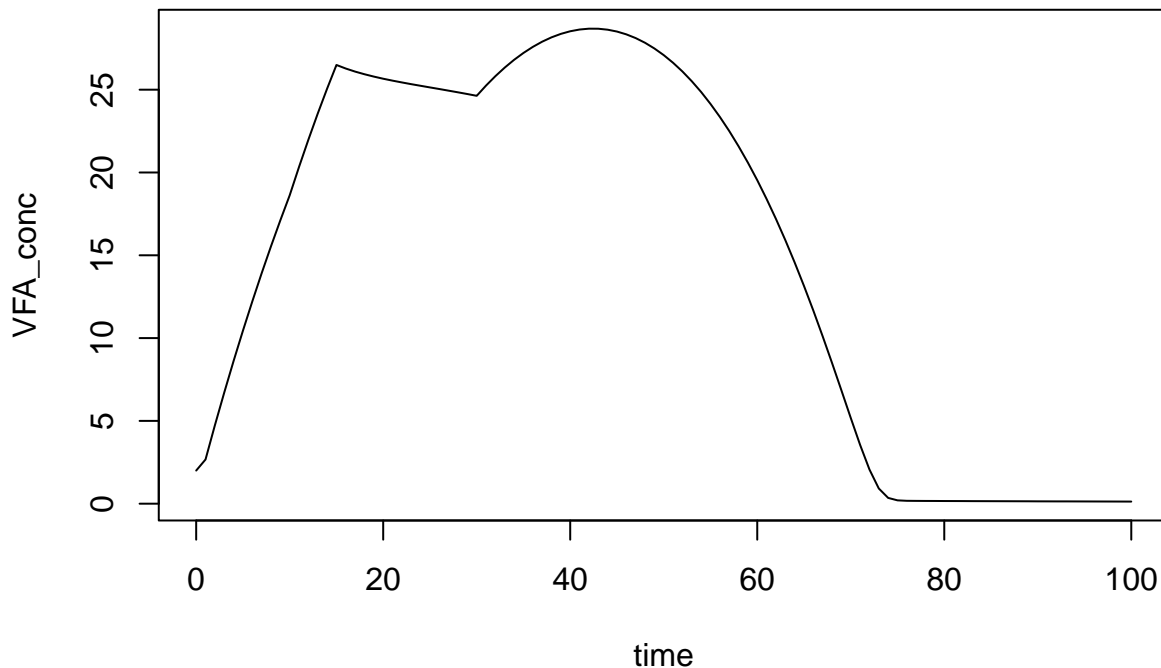
```



```
plot(VSd_conc ~ time, data = out4b, type = 'l')
```



```
plot(VFA_conc ~ time, data = out4b, type = 'l')
```



## 5. Time-variable inputs part 3

The list-in-data frame approach is clunky. Here is an alternative.

```
var_dat <- data.frame(time = c(0, 1, 10, 15, 30, 50),
                      slurry_mass = c(1000, 7000, 2000, 5000, 3000, 10000))
var_dat
```

```
##   time slurry_mass
## 1    0         1000
## 2    1         7000
## 3   10         2000
## 4   15         5000
## 5   30         3000
## 6   50        10000
```

Make a separate data frame for each other argument (any name, but note column names!).

```
sub_fresh_dat = data.frame(time = c(0, 1, 10, 15, 30, 50),
                            VSd = c(50, 100, 0, 0, 200, 50))
```

```
T_opt_dat = data.frame(time = c(0, 1, 10, 15, 30, 50),
                        m1 = 20 + 0:5 * 2,
                        m2 = 20 + 0:5 * 2,
                        m3 = 30 + 0:5 * 2,
                        sr1 = 46 + 0:5 * 2)
```

and combine them in a list, using the parameter element names for element names (e.g., `sub_fresh` is the name of an element in `sub_pars`).

```
var_pars <- list(var = var_dat, sub_fresh = sub_fresh_dat, T_opt = T_opt_dat)
```

```
devtools::load_all()
```

```
## i Loading ABM
```

```

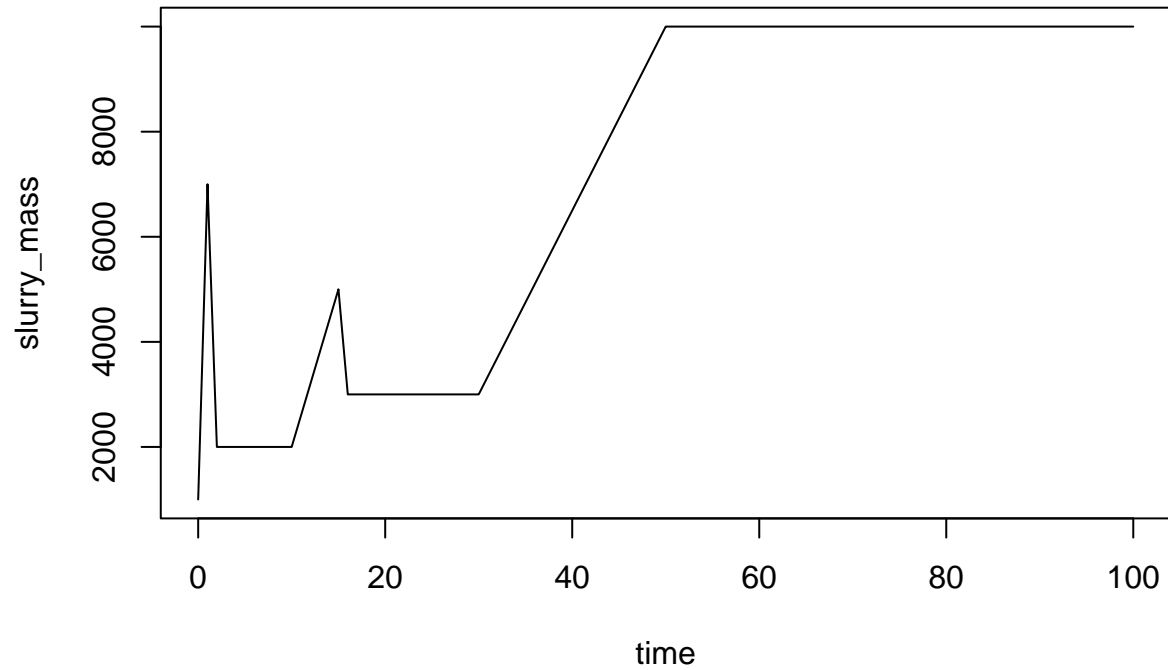
out5 <- abm(100,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars,
  var_pars = var_pars)

```

```

plot(slurry_mass ~ time, data = out5, type = 'l')

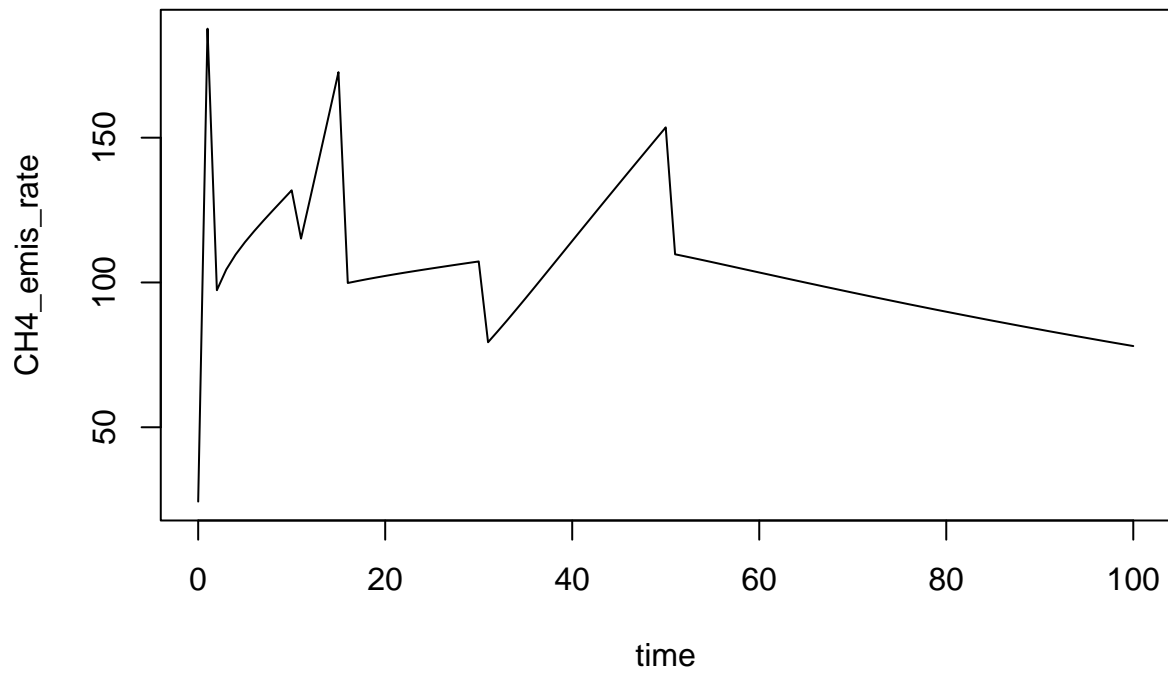
```



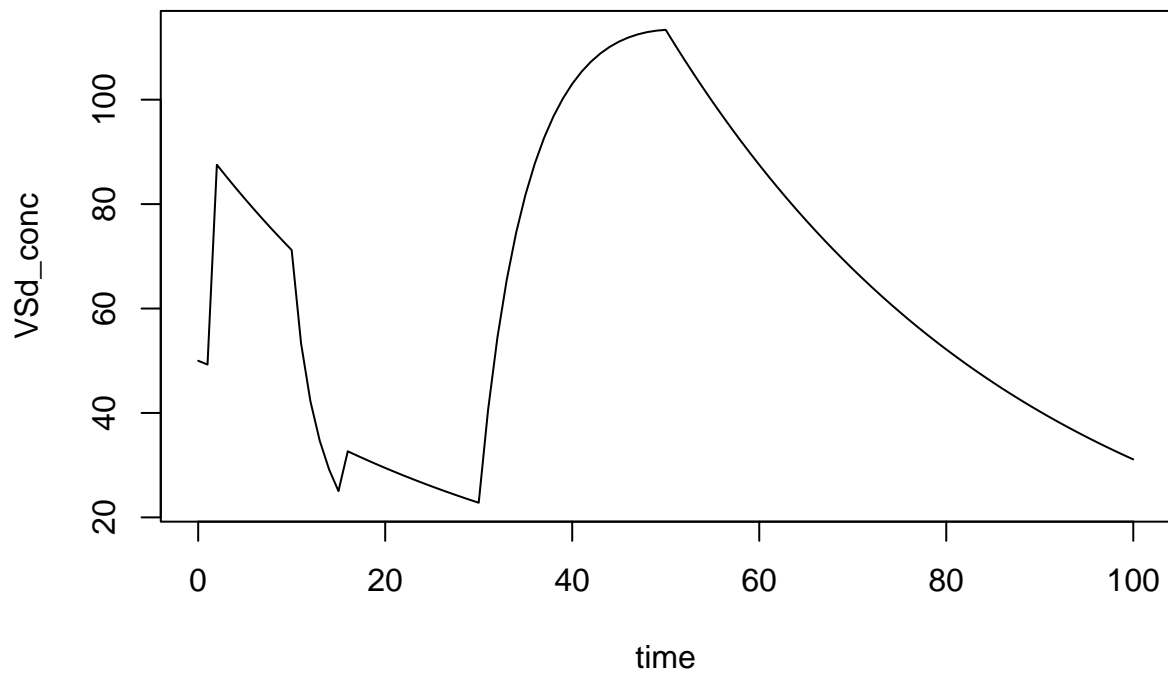
```

plot(CH4_emis_rate ~ time, data = out5, type = 'l')

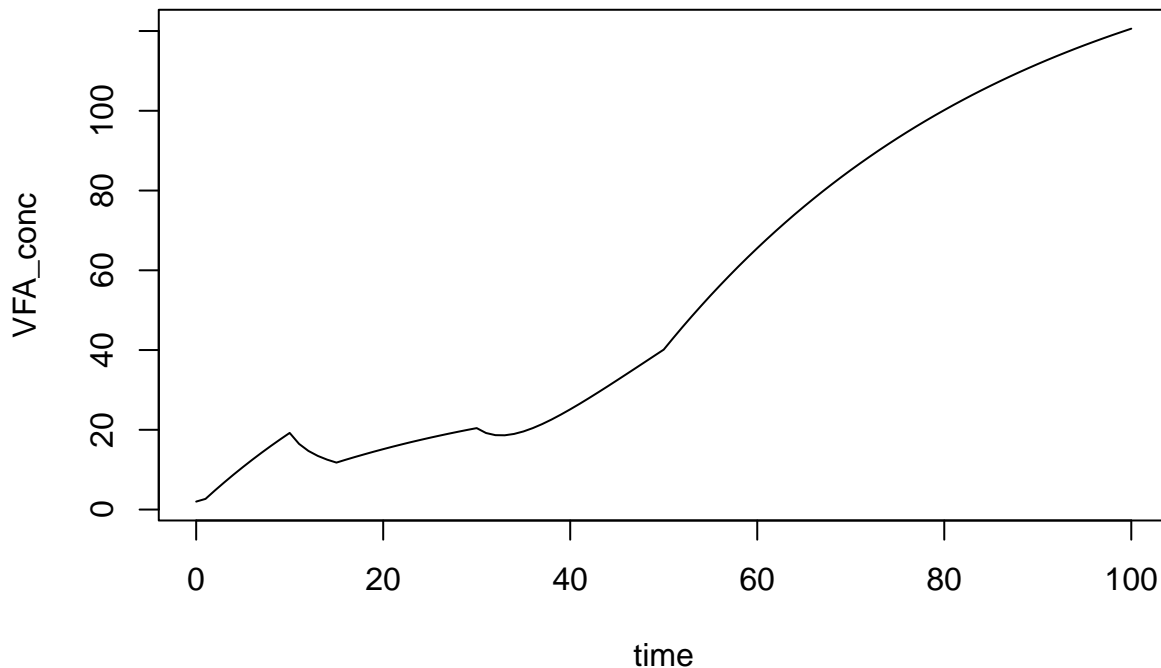
```



```
plot(VSd_conc ~ time, data = out5, type = 'l')
```



```
plot(VFA_conc ~ time, data = out5, type = 'l')
```



## 6. Flexible solutes

Any conservative solute can be added in `man_pars`, using any names. I am moving toward using a “master species” approach, so it makes sense to use the chemical formula of the primary species, with `p` or `m` for a charge symbol. The `comp` part of the name below is for “component”.

```
man_pars6 <- list(comps = c('H2S', 'SO4m2', 'NH4p'),
                  comp_fresh = c(H2S = 0.01, SO4m2 = 0.2, NH4p = 2.5),
                  VFA_fresh = c(VFA = 2),
                  pH = 7, dens = 1000)
```

Note that VFA is still special—it has a fixed name in the code and is not conservative.

```
devtools::load_all()
```

```
## i Loading ABM
```

```
out6a <- abm(365,
             mng_pars = mng_pars,
             man_pars = man_pars6,
             grp_pars = grp_pars,
             mic_pars = mic_pars,
             sub_pars = sub_pars,
             chem_pars = chem_pars)
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 1.7%
```

```
tail(out6a)
```

```
##      time      m0      m1      m2      sr1      VSd H2S SO4m2  NH4p
## 364  360 74909.98 72497.35 337403.6 17643.06 15468816 6100 122000 1525000
## 365  361 77056.84 74543.26 351549.2 17788.74 15576604 6200 124000 1550000
## 366  362 79234.75 76617.42 366160.3 17931.53 15682002 6300 126000 1575000
## 367  363 81441.36 78717.58 381233.0 18071.50 15785084 6400 128000 1600000
```

```

## 368 364 83673.78 80840.93 396758.5 18208.69 15885918 6500 130000 1625000
## 369 365 85928.37 82984.01 412722.1 18343.17 15984571 6600 132000 1650000
##      VFA slurry_mass CH4_emis_cum CO2_emis_cum slurry_load COD_load
## 364 4796289      610000      26846605      21417315      3600000 187920000
## 365 4684393      620000      26973866      21518840      3610000 188442000
## 366 4557322      630000      27105400      21623774      3620000 188964000
## 367 4414981      640000      27241214      21732122      3630000 189486000
## 368 4257395      650000      27381289      21843868      3640000 190008000
## 369 4084743      660000      27525568      21958969      3650000 190530000
##      CH4_emis_rate temp_C pH      m0_eff      m1_eff      m2_eff      sr1_eff      VSd_eff      H2S_eff
## 364      125134.1      20      7 441740.7 422210.1 2239891 63286.93 53531054 29910
## 365      129394.2      20      7 441740.7 422210.1 2239891 63286.93 53531054 29910
## 366      133674.8      20      7 441740.7 422210.1 2239891 63286.93 53531054 29910
## 367      137950.5      20      7 441740.7 422210.1 2239891 63286.93 53531054 29910
## 368      142189.3      20      7 441740.7 422210.1 2239891 63286.93 53531054 29910
## 369      146351.3      20      7 441740.7 422210.1 2239891 63286.93 53531054 29910
##      S04m2_eff NH4p_eff VFA_eff slurry_mass_eff slurry_depth      m0_conc      m1_conc
## 364      598200 7477500 3419880      2991000      6.1 0.1228032 0.1188481
## 365      598200 7477500 3419880      2991000      6.2 0.1242852 0.1202311
## 366      598200 7477500 3419880      2991000      6.3 0.1257694 0.1216150
## 367      598200 7477500 3419880      2991000      6.4 0.1272521 0.1229962
## 368      598200 7477500 3419880      2991000      6.5 0.1287289 0.1243707
## 369      598200 7477500 3419880      2991000      6.6 0.1301945 0.1257334
##      m2_conc      sr1_conc VSd_conc H2S_conc S04m2_conc NH4p_conc VFA_conc
## 364 0.5531206 0.02892305 25.35872      0.01      0.2      2.5 7.862769
## 365 0.5670148 0.02869152 25.12355      0.01      0.2      2.5 7.555472
## 366 0.5812069 0.02846275 24.89207      0.01      0.2      2.5 7.233845
## 367 0.5956766 0.02823671 24.66419      0.01      0.2      2.5 6.898408
## 368 0.6103976 0.02801337 24.43987      0.01      0.2      2.5 6.549838
## 369 0.6253365 0.02779268 24.21905      0.01      0.2      2.5 6.189004
##      m0_eff_conc m1_eff_conc m2_eff_conc sr1_eff_conc VSd_eff_conc H2S_eff_conc
## 364      0.14769      0.1411602      0.7488771      0.02115912      17.89738      0.01
## 365      0.14769      0.1411602      0.7488771      0.02115912      17.89738      0.01
## 366      0.14769      0.1411602      0.7488771      0.02115912      17.89738      0.01
## 367      0.14769      0.1411602      0.7488771      0.02115912      17.89738      0.01
## 368      0.14769      0.1411602      0.7488771      0.02115912      17.89738      0.01
## 369      0.14769      0.1411602      0.7488771      0.02115912      17.89738      0.01
##      S04m2_eff_conc NH4p_eff_conc VFA_eff_conc
## 364      0.2      2.5      1.14339
## 365      0.2      2.5      1.14339
## 366      0.2      2.5      1.14339
## 367      0.2      2.5      1.14339
## 368      0.2      2.5      1.14339
## 369      0.2      2.5      1.14339

```

```
head(out6a)
```

```

##      time      m0      m1      m2      sr1      VSd H2S S04m2      NH4p
## 1      0 50.0000 50.0000 50.0000 50.0000 50000.0 10 200 2500
## 2      1 554.0098 553.8533 558.3748 544.0431 542318.4 110 2200 27500
## 3      2 1066.2767 1065.6732 1083.1940 1028.3035 1022076.9 210 4200 52500
## 4      3 1588.3161 1586.9430 1627.0197 1502.9749 1489597.5 310 6200 77500
## 5      4 2121.2034 2118.7114 2191.8594 1968.2472 1945194.0 410 8200 102500
## 6      5 2665.7726 2661.7877 2779.4361 2424.3064 2389172.2 510 10200 127500
##      VFA slurry_mass CH4_emis_cum CO2_emis_cum slurry_load COD_load

```



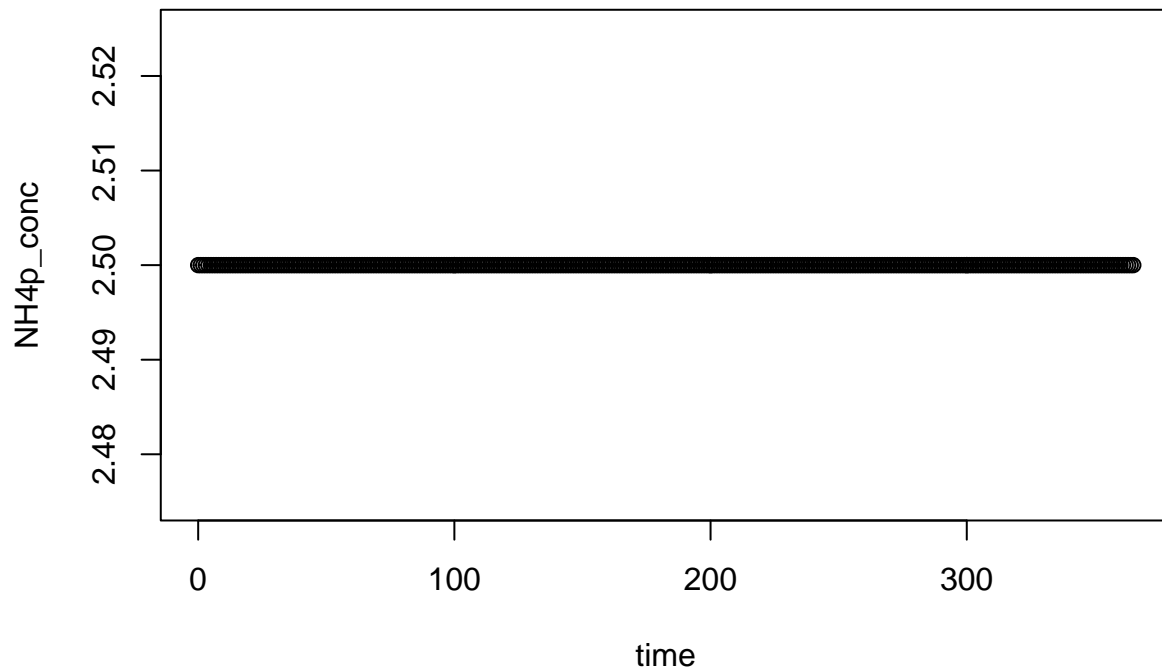
```

## 1 2000.00 1000 0.0000 0.0000 0 0
## 2 29018.66 11000 163.6272 130.5362 10000 522000
## 3 67371.44 21000 628.8119 501.6449 20000 1044000
## 4 116611.44 31000 1425.4337 1137.1629 30000 1566000
## 5 176326.70 41000 2577.0208 2055.8603 40000 2088000
## 6 246127.08 51000 4103.8067 3273.8785 50000 2610000
## CH4_emis_rate temp_C pH m0_eff m1_eff m2_eff sr1_eff VSd_eff H2S_eff
## 1 25.52844 20 7 0 0 0 0 0 0
## 2 308.65512 20 7 0 0 0 0 0 0
## 3 626.59326 20 7 0 0 0 0 0 0
## 4 970.52564 20 7 0 0 0 0 0 0
## 5 1335.99741 20 7 0 0 0 0 0 0
## 6 1720.64113 20 7 0 0 0 0 0 0
## S04m2_eff NH4p_eff VFA_eff slurry_mass_eff slurry_depth m0_conc m1_conc
## 1 0 0 0 0 0.01 0.05000000 0.05000000
## 2 0 0 0 0 0.11 0.05036453 0.05035030
## 3 0 0 0 0 0.21 0.05077508 0.05074634
## 4 0 0 0 0 0.31 0.05123600 0.05119171
## 5 0 0 0 0 0.41 0.05173667 0.05167589
## 6 0 0 0 0 0.51 0.05227005 0.05219192
## m2_conc sr1_conc VSd_conc H2S_conc S04m2_conc NH4p_conc VFA_conc
## 1 0.05000000 0.05000000 50.00000 0.01 0.2 2.5 2.000000
## 2 0.05076135 0.04945846 49.30167 0.01 0.2 2.5 2.638060
## 3 0.05158067 0.04896683 48.67033 0.01 0.2 2.5 3.208164
## 4 0.05248451 0.04848306 48.05153 0.01 0.2 2.5 3.761660
## 5 0.05345999 0.04800603 47.44376 0.01 0.2 2.5 4.300651
## 6 0.05449875 0.04753542 46.84651 0.01 0.2 2.5 4.826021
## m0_eff_conc m1_eff_conc m2_eff_conc sr1_eff_conc VSd_eff_conc H2S_eff_conc
## 1 NaN NaN NaN NaN NaN NaN NaN
## 2 NaN NaN NaN NaN NaN NaN NaN
## 3 NaN NaN NaN NaN NaN NaN NaN
## 4 NaN NaN NaN NaN NaN NaN NaN
## 5 NaN NaN NaN NaN NaN NaN NaN
## 6 NaN NaN NaN NaN NaN NaN NaN
## S04m2_eff_conc NH4p_eff_conc VFA_eff_conc
## 1 NaN NaN NaN
## 2 NaN NaN NaN
## 3 NaN NaN NaN
## 4 NaN NaN NaN
## 5 NaN NaN NaN
## 6 NaN NaN NaN

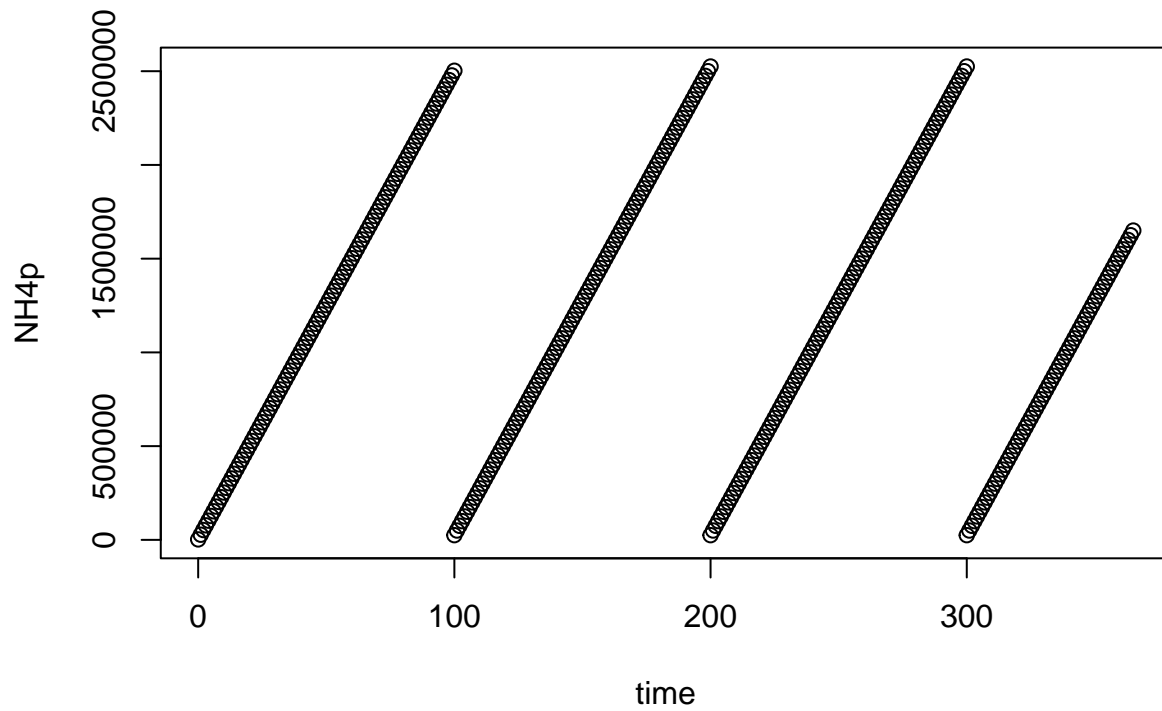
```

Conservative components are boring in output (without inhibition or volatilization).

```
plot(NH4p_conc ~ time, data = out6a)
```



```
plot(NH4p ~ time, data = out6a)
```



We can see dilution effects at least if some washing water is added.

```
mng_pars6 = list(slurry_prod_rate = 10000,
                  slurry_mass = 1000,
                  storage_depth = 2,
                  resid_depth = 0.1,
                  area = 100,
                  empty_int = 100,
                  temp_C = 20,
```

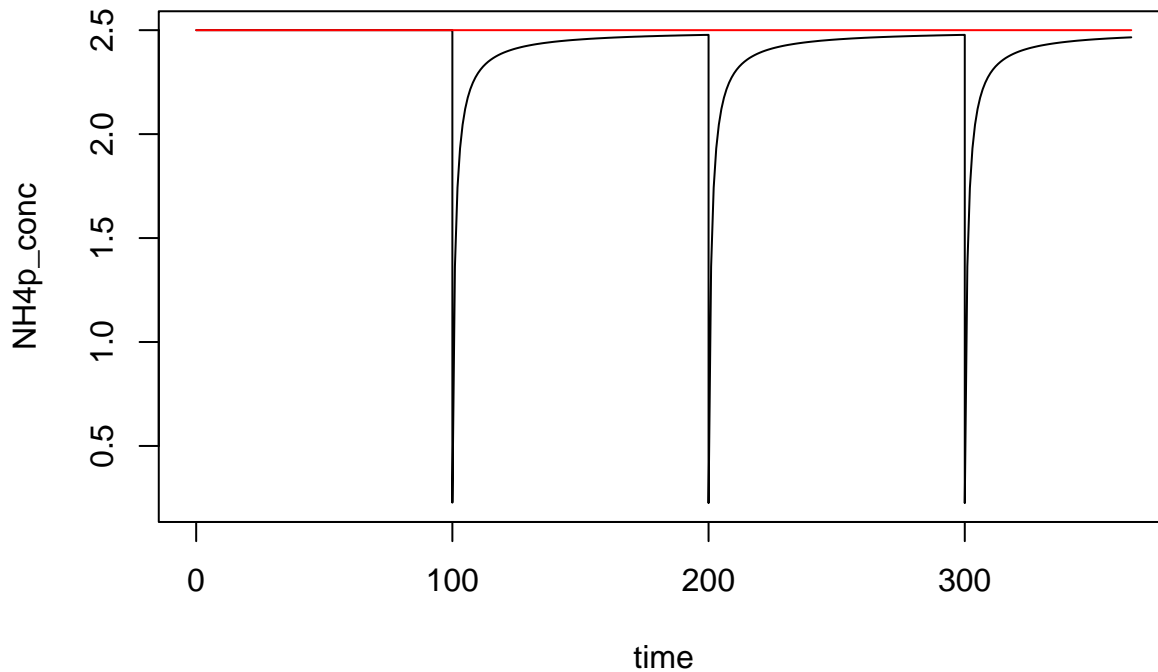
```

        wash_water = 100000,
        wash_int = 100,
        rest_d = 0,
        resid_enrich = 1)

out6b <- abm(365,
  mng_pars = mng_pars6,
  man_pars = man_pars6,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars)

## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 32%
plot(NH4p_conc ~ time, data = out6b, type = 'l')
lines(NH4p_conc ~ time, data = out6a, col = 'red')

```



## 7. Speciation

Acid-base reactions are needed for inhibition. They can be added for any component. The `chem_pars` argument can accept temperature-dependent log  $K_a$  expressions.

```

man_pars7 <- list(comps = c('H2S', 'NH4p'),
  comp_fresh = c(H2S = 0.01, NH4p = 2.5),
  VFA_fresh = c(VFA = 2),
  pH = 7, dens = 1000)

chem_pars7 <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
  VFA = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
  CO2_aer = 1/0.436, CO2_sr = 1/1.2,
  C_xa = 1/0.3753125),

```

```

specs = c('NH3', 'HSm', 'VFAm'),
mspec = c(NH3 = 'NH4p', HSm = 'H2S', VFAm = 'VFA'),
lka = c(NH3 = '- 0.09046 - 2729.31/temp_K',
        HSm = '- 3448.7/temp_K + 47.479 - 7.5227 * log(temp_K)',
        VFAm = '-4.8288 + 21.42/temp_K')
)

```

```

out7 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars6,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars7)

```

```

## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 1.7%

```

```
head(out7)
```

```

##      time      m0      m1      m2      sr1      VSd H2S S04m2  NH4p
## 1      0  50.0000  50.0000  50.0000  50.0000  50000.0  10   200   2500
## 2      1  554.0098  553.8533  558.3748  544.0431  542318.4 110  2200  27500
## 3      2 1066.2767 1065.6732 1083.1940 1028.3035 1022076.9 210  4200  52500
## 4      3 1588.3161 1586.9430 1627.0197 1502.9749 1489597.5 310  6200  77500
## 5      4 2121.2034 2118.7114 2191.8594 1968.2472 1945194.0 410  8200 102500
## 6      5 2665.7726 2661.7877 2779.4361 2424.3064 2389172.2 510 10200 127500
##      VFA slurry_mass CH4_emis_cum CO2_emis_cum slurry_load COD_load
## 1    2000.00      1000      0.0000      0.0000      0      0
## 2   29018.66     11000     163.6272     130.5362     10000  522000
## 3   67371.44     21000     628.8119     501.6449     20000 1044000
## 4 116611.44     31000    1425.4337    1137.1629     30000 1566000
## 5 176326.70     41000    2577.0208    2055.8603     40000 2088000
## 6 246127.08     51000    4103.8067    3273.8785     50000 2610000
##      CH4_emis_rate temp_C pH m0_eff m1_eff m2_eff sr1_eff VSd_eff H2S_eff
## 1      25.52844      20  7      0      0      0      0      0      0
## 2      308.65512      20  7      0      0      0      0      0      0
## 3      626.59326      20  7      0      0      0      0      0      0
## 4      970.52564      20  7      0      0      0      0      0      0
## 5     1335.99741      20  7      0      0      0      0      0      0
## 6     1720.64113      20  7      0      0      0      0      0      0
##      S04m2_eff NH4p_eff VFA_eff slurry_mass_eff slurry_depth  m0_conc  m1_conc
## 1      0      0      0      0      0.01 0.05000000 0.05000000
## 2      0      0      0      0      0.11 0.05036453 0.05035030
## 3      0      0      0      0      0.21 0.05077508 0.05074634
## 4      0      0      0      0      0.31 0.05123600 0.05119171
## 5      0      0      0      0      0.41 0.05173667 0.05167589
## 6      0      0      0      0      0.51 0.05227005 0.05219192
##      m2_conc  sr1_conc VSd_conc H2S_conc S04m2_conc NH4p_conc VFA_conc
## 1 0.05000000 0.05000000 50.00000      0.01      0.2      2.5 2.000000
## 2 0.05076135 0.04945846 49.30167      0.01      0.2      2.5 2.638060
## 3 0.05158067 0.04896683 48.67033      0.01      0.2      2.5 3.208164
## 4 0.05248451 0.04848306 48.05153      0.01      0.2      2.5 3.761660
## 5 0.05345999 0.04800603 47.44376      0.01      0.2      2.5 4.300651
## 6 0.05449875 0.04753542 46.84651      0.01      0.2      2.5 4.826021

```

```
##      m0_eff_conc m1_eff_conc m2_eff_conc sr1_eff_conc VSd_eff_conc H2S_eff_conc
## 1          NaN          NaN          NaN          NaN          NaN          NaN
## 2          NaN          NaN          NaN          NaN          NaN          NaN
## 3          NaN          NaN          NaN          NaN          NaN          NaN
## 4          NaN          NaN          NaN          NaN          NaN          NaN
## 5          NaN          NaN          NaN          NaN          NaN          NaN
## 6          NaN          NaN          NaN          NaN          NaN          NaN
##      S04m2_eff_conc NH4p_eff_conc VFA_eff_conc
## 1          NaN          NaN          NaN
## 2          NaN          NaN          NaN
## 3          NaN          NaN          NaN
## 4          NaN          NaN          NaN
## 5          NaN          NaN          NaN
## 6          NaN          NaN          NaN
```

But chemical species are actually ignored unless they are used in inhibition.

## 8. Inhibition

Any chemical species can inhibit any microbial group. Inhibition parameters (currently initial and complete concentrations, with linear response, why not?) are entered in a matrix.

```
ilwr <- matrix(
  c(5, 0.2, 10, 0.5,
    5, 0.2, 10, 0.5,
    5, 0.2, 10, 0.5,
    5, 0.2, 10, 0.5),
  nrow = 4,
  byrow = TRUE,
  dimnames = list(
    c('m0', 'm1', 'm2', 'sr1'),
    c('NH4p', 'NH3', 'VFAm', 'VFA')
  )
)

iupr <- matrix(
  c(9, 0.9, 30, 1,
    9, 0.9, 30, 1,
    9, 0.9, 30, 1,
    9, 0.9, 30, 1),
  nrow = 4,
  byrow = TRUE,
  dimnames = list(
    c('m0', 'm1', 'm2', 'sr1'),
    c('NH4p', 'NH3', 'VFAm', 'VFA')
  )
)

inhib_pars <- list(
  ilwr = ilwr,
  iupr = iupr
)

inhib_pars
```

```

## $ilwr
##      NH4p NH3 VFAm VFA
## m0      5 0.2  10 0.5
## m1      5 0.2  10 0.5
## m2      5 0.2  10 0.5
## sr1     5 0.2  10 0.5
##
## $iupr
##      NH4p NH3 VFAm VFA
## m0      9 0.9  30  1
## m1      9 0.9  30  1
## m2      9 0.9  30  1
## sr1     9 0.9  30  1

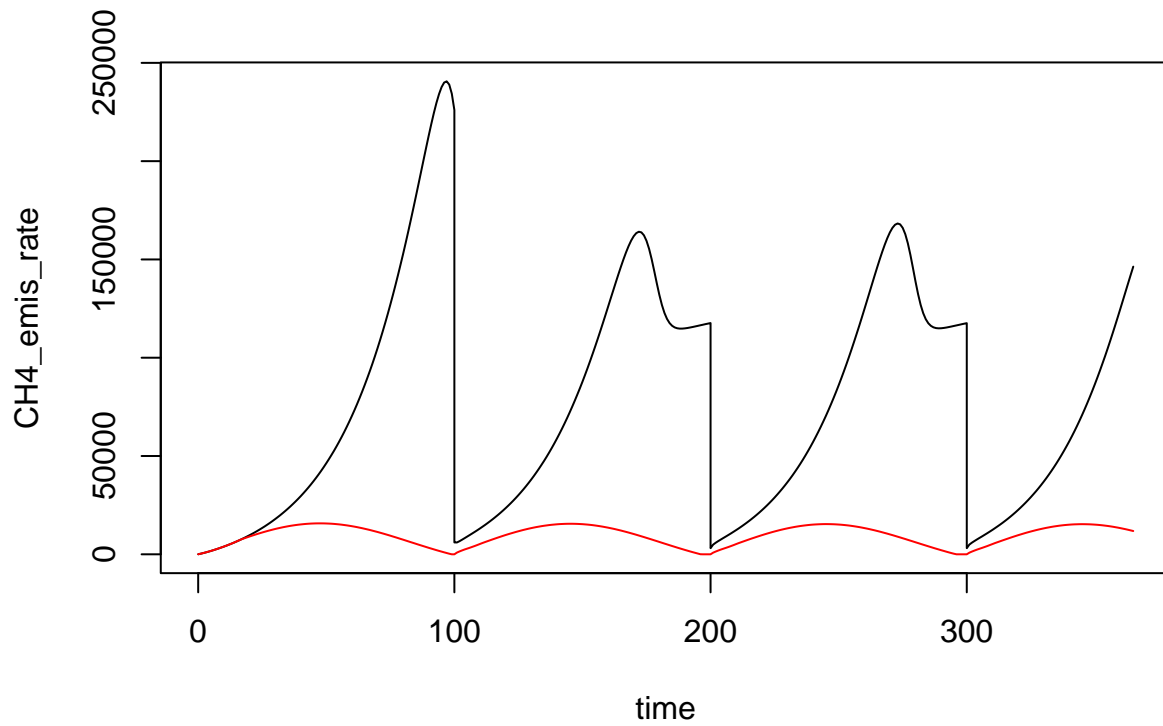
man_pars8 <- list(comps = c('H2S', 'NH4p'),
                  comp_fresh = c(H2S = 0.01,
                                NH4p = 2.5),
                  VFA_fresh = c(VFA = 2),
                  pH = 7, dens = 1000)

chem_pars8 <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
                                VFA = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
                                CO2_aer = 1/0.436, CO2_sr = 1/1.2,
                                C_xa = 1/0.3753125),
                  specs = c('NH3', 'HSm', 'VFAm'),
                  mspec = c(NH3 = 'NH4p', HSm = 'H2S', VFAm = 'VFA'),
                  lka = c(NH3 = '- 0.09046 - 2729.31/temp_K',
                          HSm = '- 3448.7/temp_K + 47.479 - 7.5227* log(temp_K)',
                          VFAm = '-4.8288 + 21.42/temp_K')
)

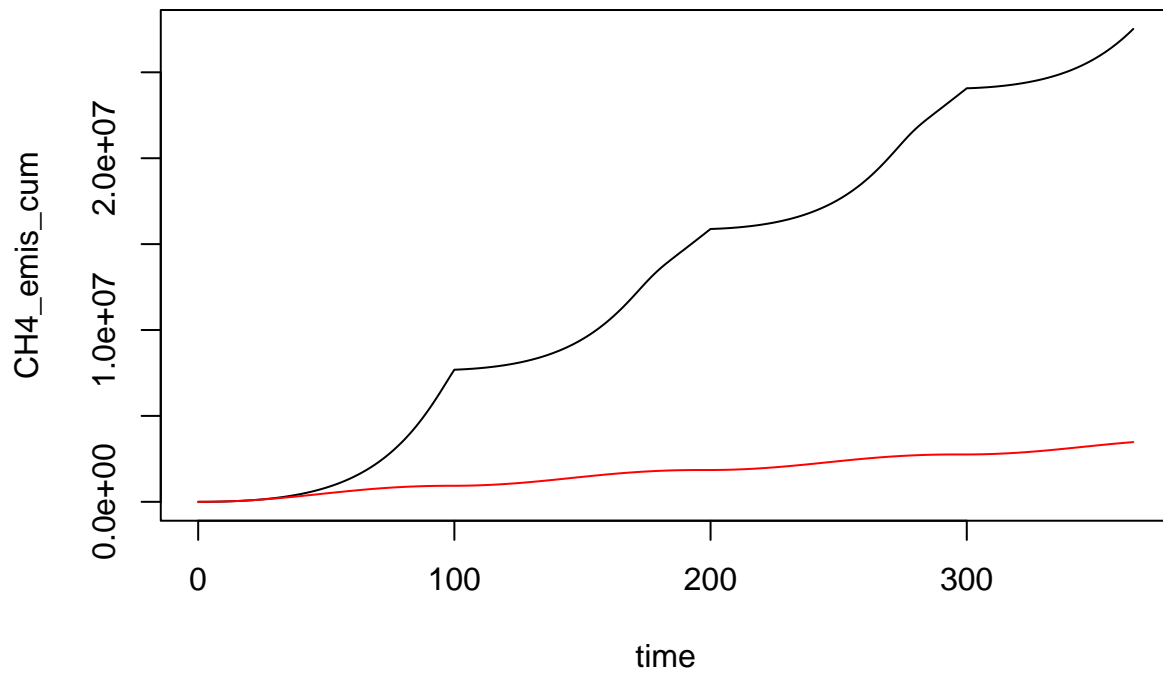
out8 <- abm(365,
            mng_pars = mng_pars,
            man_pars = man_pars7,
            grp_pars = grp_pars,
            mic_pars = mic_pars,
            sub_pars = sub_pars,
            chem_pars = chem_pars7,
            inhib_pars = inhib_pars
)

plot(CH4_emis_rate ~ time, data = out7, type = 'l')
lines(CH4_emis_rate ~ time, data = out8, col = 'red')

```



```
plot(CH4_emis_cum ~ time, data = out7, type = 'l')
lines(CH4_emis_cum ~ time, data = out8, col = 'red')
```



## 9. COD balance

There is now a `checkCOD()` function that runs on `abm()` results before returning them. For now the tolerance is fixed at 1%. Some of the examples above do not meet that criterion for some reason. At least one shows a real problem that needs to be identified.