

# Demo of new simple1 version

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## Overview

This demo shows:

1. basic usage,
2. variable substrates,
3. time-variable inputs,
4. speciation (proton-transfer reactions),
5. inhibition,
6. volatilization, and
7. COD balance,

## Prep

```
devtools::load_all()
```

```
## i Loading ABM
```

### 1. Basic behavior

The simplest usage is with constant slurry production rate and a fixed schedule. We need to set some parameters, first management parameters.

```
mng_pars <- list(slurry_prod_rate = 10000,  
  slurry_mass = 1000,  
  storage_depth = 2,  
  resid_depth = 0.1,  
  area = 100,  
  empty_int = 100,  
  temp_C = 20,  
  wash_water = 0,  
  wash_int = NA,  
  rest_d = 0,  
  resid_enrich = 1)
```

Next substrate parameters, a new argument. This defines substrates. We could have any number with any names. Note that hydrolysis uses CTM again (like anything here, that could be changed).

```
sub_pars <- list(subs = c('VSd'),  
  T_opt_hyd = c(VSd = 60),  
  T_min_hyd = c(VSd = 0),  
  T_max_hyd = c(VSd = 90),
```

```

hydrol_opt = c(VSd = 0.1),
sub_fresh = c(VSd = 50),
sub_init = c(VSd = 50))

```

Microbial parameters are similar to other ABM versions, but inhibition is set separately now (and not shown in this simple example).

```

grp_pars <- list(grps = c('m0', 'm1', 'm2', 'sr1'),
  yield = c(default = 0.05, sr1 = 0.065),
  xa_fresh = c(all = 0.05),
  xa_init = c(all = 0.05),
  dd_rate = c(all = 0.02),
  ksv = c(default = 1, sr1 = 0.5),
  kss = c(sr1 = 0.5),
  qhat_opt = c(m0 = 1, m1 = 1, m2 = 2, sr1 = 9),
  T_opt = c(m0 = 18, m1 = 18, m2 = 28, sr1 = 44),
  T_min = c(m0 = 0, m1 = 6.41, m2 = 6.41, sr1 = 0),
  T_max = c(m0 = 25, m1 = 25, m2 = 38, sr1 = 51))

```

The `dd_rate_xa` parameter is for “death and decay”.

```
mic_pars <- list(dd_rate_xa = 0.02)
```

These last two arguments are similar to other versions. VFA is hard-wired and so has its own elements. The name should be CH3COOH.

```

man_pars <- list(VFA_fresh = c(CH3COOH = 2), pH = 7, dens = 1000)
chem_pars <- list(COD_conv = c(CH4 = 1/0.2507))

```

```
devtools::load_all()
```

```
## i Loading ABM
```

```

out1 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars)

```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
```

```
## Size-variable parameter problem: Missing element(s) in kss.
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
```

```
## pars$COD_conv, : COD balance is off by 1.7%
```

Output is similar to other versions. (The `value` argument does not currently work.)

```
head(out1)
```

```

##   time      m0      m1      m2      sr1      VSd  CH3COOH slurry_mass
## 1    0  50.0000  50.0000  50.0000  50.0000  50000.0   2000.00      1000
## 2    1  554.0098  553.8533  558.3748  544.0431  542318.4  29018.66     11000
## 3    2 1066.2767 1065.6732 1083.1940 1028.3035 1022076.9  67371.44     21000
## 4    3 1588.3161 1586.9430 1627.0197 1502.9749 1489597.5 116611.44     31000
## 5    4 2121.2034 2118.7114 2191.8594 1968.2472 1945194.0 176326.70     41000
## 6    5 2665.7726 2661.7877 2779.4361 2424.3064 2389172.2 246127.08     51000
##   CH4_emis_cum slurry_load COD_load CH4_emis_rate temp_C pH m0_eff m1_eff

```

```

## 1      0.0000      0      0      25.52844      20 7      0      0
## 2     163.6272     10000    522000     308.65512     20 7      0      0
## 3     628.8119     20000   1044000     626.59326     20 7      0      0
## 4    1425.4337     30000   1566000     970.52564     20 7      0      0
## 5    2577.0208     40000   2088000    1335.99741     20 7      0      0
## 6    4103.8067     50000   2610000    1720.64113     20 7      0      0
##   m2_eff sr1_eff VSd_eff CH3COOH_eff slurry_mass_eff slurry_depth   m0_conc
## 1      0      0      0      0      0      0.01 0.05000000
## 2      0      0      0      0      0      0.11 0.05036453
## 3      0      0      0      0      0      0.21 0.05077508
## 4      0      0      0      0      0      0.31 0.05123600
## 5      0      0      0      0      0      0.41 0.05173667
## 6      0      0      0      0      0      0.51 0.05227005
##   m1_conc   m2_conc   sr1_conc VSd_conc CH3COOH_conc m0_eff_conc
## 1 0.05000000 0.05000000 0.05000000 50.00000      2.000000      NaN
## 2 0.05035030 0.05076135 0.04945846 49.30167      2.638060      NaN
## 3 0.05074634 0.05158067 0.04896683 48.67033      3.208164      NaN
## 4 0.05119171 0.05248451 0.04848306 48.05153      3.761660      NaN
## 5 0.05167589 0.05345999 0.04800603 47.44376      4.300651      NaN
## 6 0.05219192 0.05449875 0.04753542 46.84651      4.826021      NaN
##   m1_eff_conc m2_eff_conc sr1_eff_conc VSd_eff_conc CH3COOH_eff_conc
## 1      NaN      NaN      NaN      NaN      NaN
## 2      NaN      NaN      NaN      NaN      NaN
## 3      NaN      NaN      NaN      NaN      NaN
## 4      NaN      NaN      NaN      NaN      NaN
## 5      NaN      NaN      NaN      NaN      NaN
## 6      NaN      NaN      NaN      NaN      NaN

```

```
tail(out1)
```

```

##   time      m0      m1      m2      sr1      VSd CH3COOH slurry_mass
## 364 360 74909.98 72497.35 337403.6 17643.06 15468816 4796289      610000
## 365 361 77056.84 74543.26 351549.2 17788.74 15576604 4684393      620000
## 366 362 79234.75 76617.42 366160.3 17931.53 15682002 4557322      630000
## 367 363 81441.36 78717.58 381233.0 18071.50 15785084 4414981      640000
## 368 364 83673.78 80840.93 396758.5 18208.69 15885918 4257395      650000
## 369 365 85928.37 82984.01 412722.1 18343.17 15984571 4084743      660000
##   CH4_emis_cum slurry_load COD_load CH4_emis_rate temp_C pH   m0_eff
## 364 26846605      3600000 187920000      125134.1    20 7 441740.7
## 365 26973866      3610000 188442000      129394.2    20 7 441740.7
## 366 27105400      3620000 188964000      133674.8    20 7 441740.7
## 367 27241214      3630000 189486000      137950.5    20 7 441740.7
## 368 27381289      3640000 190008000      142189.3    20 7 441740.7
## 369 27525568      3650000 190530000      146351.3    20 7 441740.7
##   m1_eff m2_eff sr1_eff VSd_eff CH3COOH_eff slurry_mass_eff slurry_depth
## 364 422210.1 2239891 63286.93 53531054      3419880      2991000      6.1
## 365 422210.1 2239891 63286.93 53531054      3419880      2991000      6.2
## 366 422210.1 2239891 63286.93 53531054      3419880      2991000      6.3
## 367 422210.1 2239891 63286.93 53531054      3419880      2991000      6.4
## 368 422210.1 2239891 63286.93 53531054      3419880      2991000      6.5
## 369 422210.1 2239891 63286.93 53531054      3419880      2991000      6.6
##   m0_conc   m1_conc   m2_conc   sr1_conc VSd_conc CH3COOH_conc m0_eff_conc
## 364 0.1228032 0.1188481 0.5531206 0.02892305 25.35872      7.862769      0.14769
## 365 0.1242852 0.1202311 0.5670148 0.02869152 25.12355      7.555472      0.14769
## 366 0.1257694 0.1216150 0.5812069 0.02846275 24.89207      7.233845      0.14769

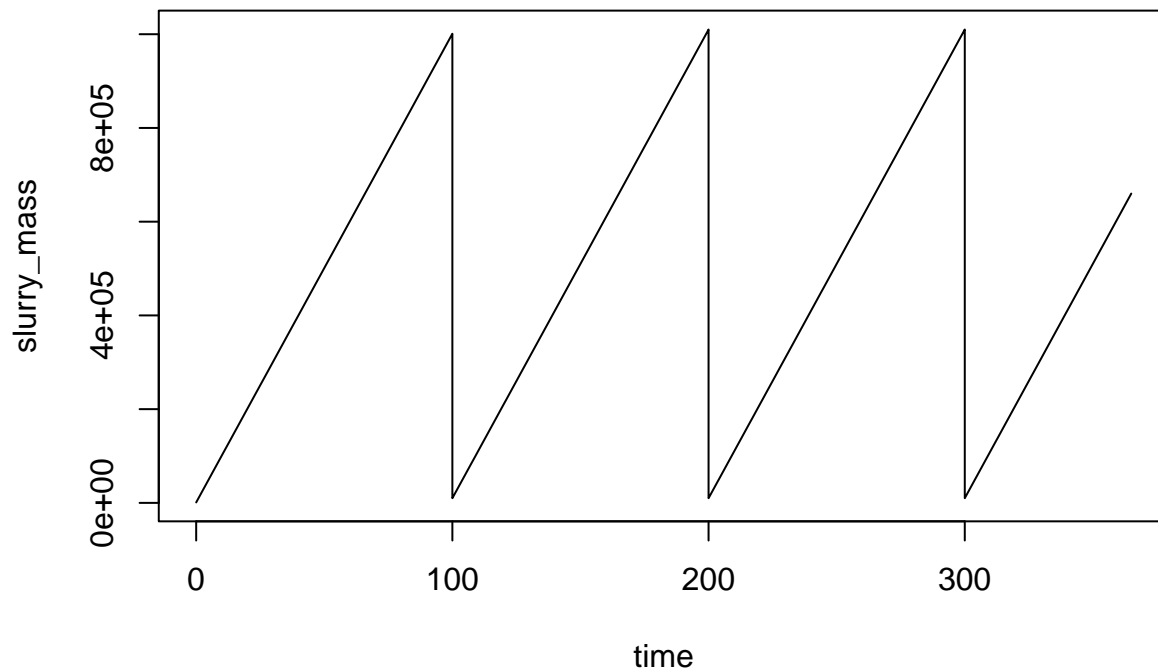
```

```
## 367 0.1272521 0.1229962 0.5956766 0.02823671 24.66419      6.898408      0.14769
## 368 0.1287289 0.1243707 0.6103976 0.02801337 24.43987      6.549838      0.14769
## 369 0.1301945 0.1257334 0.6253365 0.02779268 24.21905      6.189004      0.14769
##      m1_eff_conc m2_eff_conc sr1_eff_conc VSd_eff_conc CH3COOH_eff_conc
## 364  0.1411602   0.7488771   0.02115912    17.89738      1.14339
## 365  0.1411602   0.7488771   0.02115912    17.89738      1.14339
## 366  0.1411602   0.7488771   0.02115912    17.89738      1.14339
## 367  0.1411602   0.7488771   0.02115912    17.89738      1.14339
## 368  0.1411602   0.7488771   0.02115912    17.89738      1.14339
## 369  0.1411602   0.7488771   0.02115912    17.89738      1.14339
```

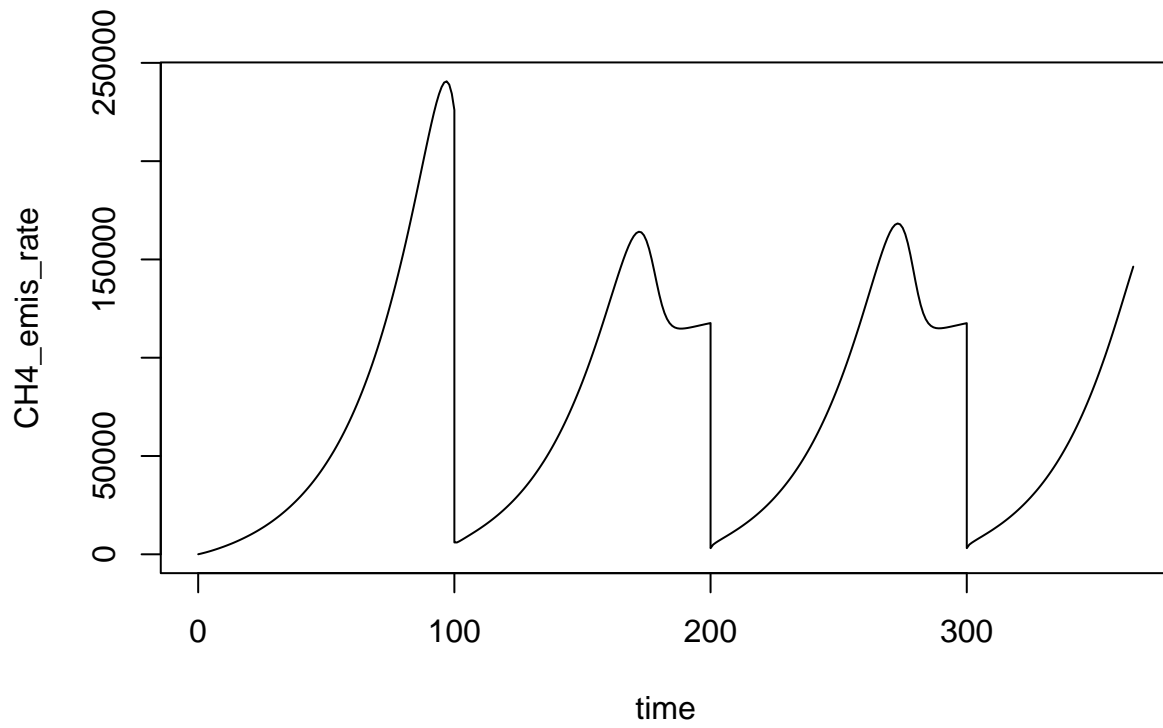
The effluent columns are cumulative. Is this what we had before? I did it for COD balance checking. We will have to discuss what is needed.

Here are some results.

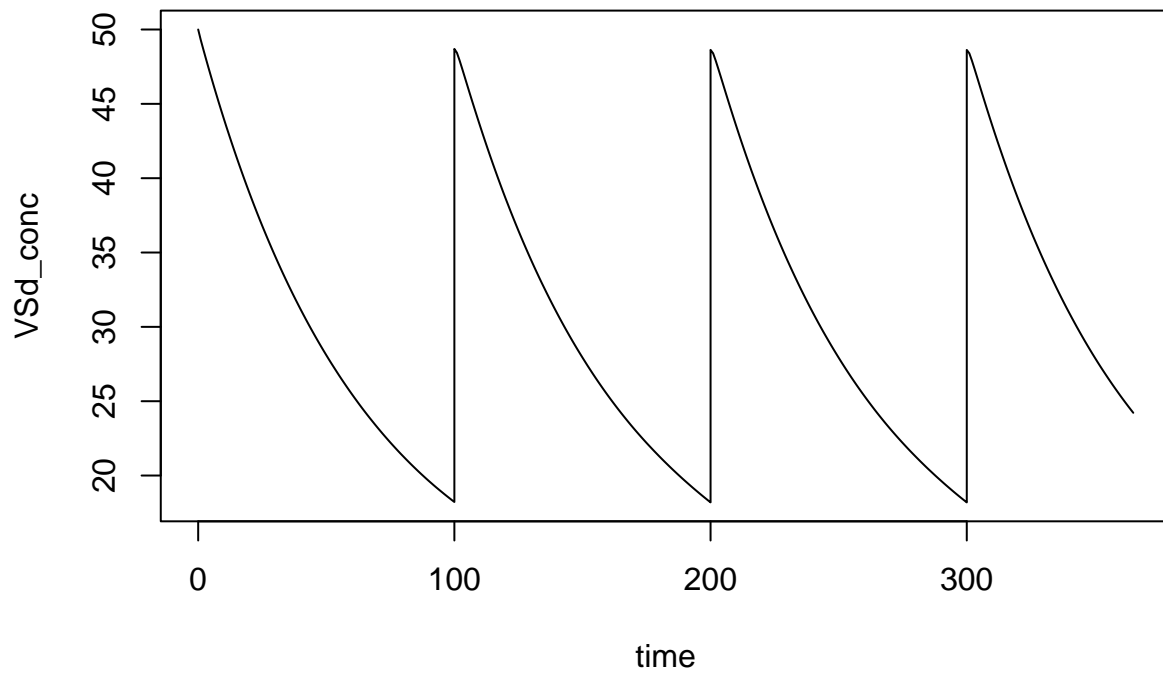
```
plot(slurry_mass ~ time, data = out1, type = 'l')
```



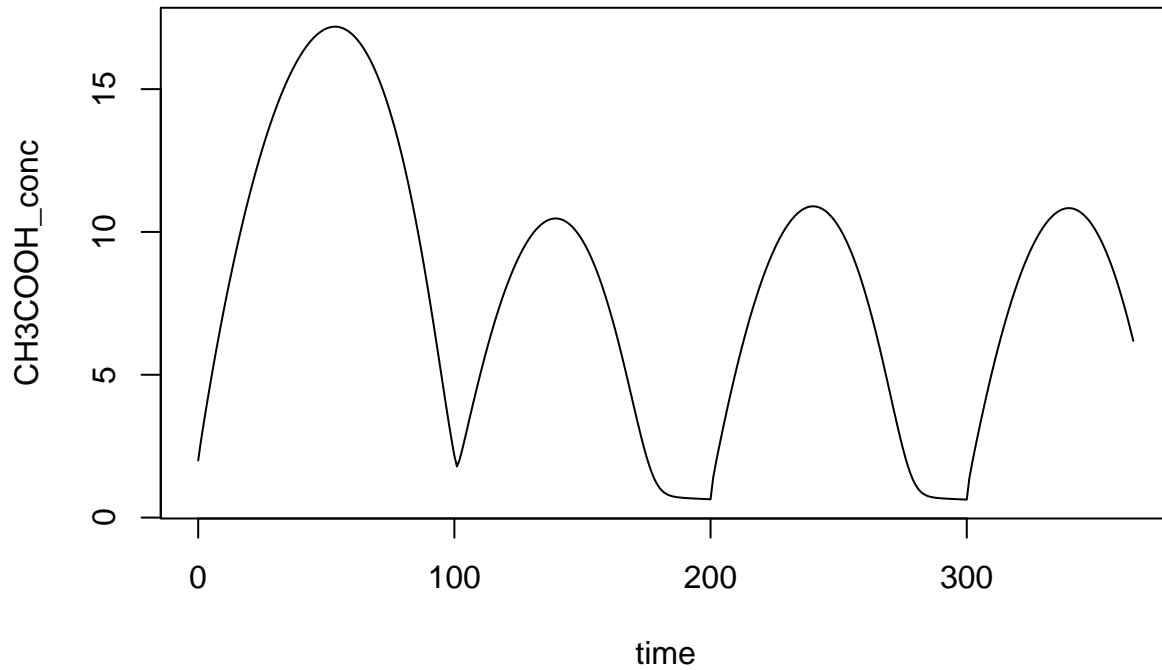
```
plot(CH4_emis_rate ~ time, data = out1, type = 'l')
```



```
plot(VSd_conc ~ time, data = out1, type = 'l')
```

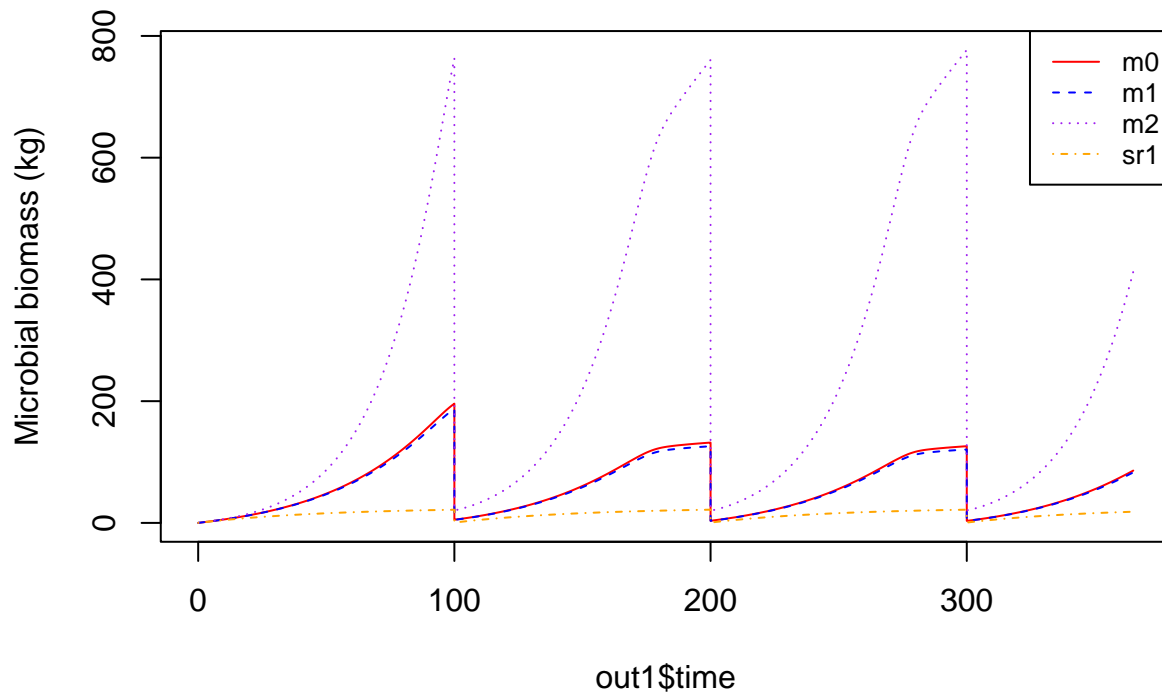


```
plot(CH3COOH_conc ~ time, data = out1, type = 'l')
```



And methanogens.

```
line_colors <- c('red', 'blue', 'purple','orange')
matplot(out1$time, out1[, nn <- c('m0','m1','m2','sr1')]/1000,
        type = 'l', lty = c(1:length(nn)), col = line_colors, ylab = 'Microbial biomass (kg)')
legend("topright", legend = nn, lty = c(1:length(nn)), col = line_colors, lwd = 1, cex = 0.8)
```



## 2. Substrate flexibility

Particulate substrates are defined in `sub_pars` now and there are no specific substrates hard-wired in the code. VFA is the only intermediate, and it is hard-wired. Here we will use three substrates. Parameter values have no connection to reality in this example.

```
sub_pars2 <- list(subs = c('cellulose', 'protein', 'lipids'),
  T_opt_hyd = c(all = 60),
  T_min_hyd = c(all = 0),
  T_max_hyd = c(all = 90),
  hydrol_opt = c(lipids = 0.1, protein = 0.01, cellulose = 0.05),
  sub_fresh = c(lipids = 3, protein = 20, cellulose = 35),
  sub_init = c(lipids = 3, protein = 20, cellulose = 35))
```

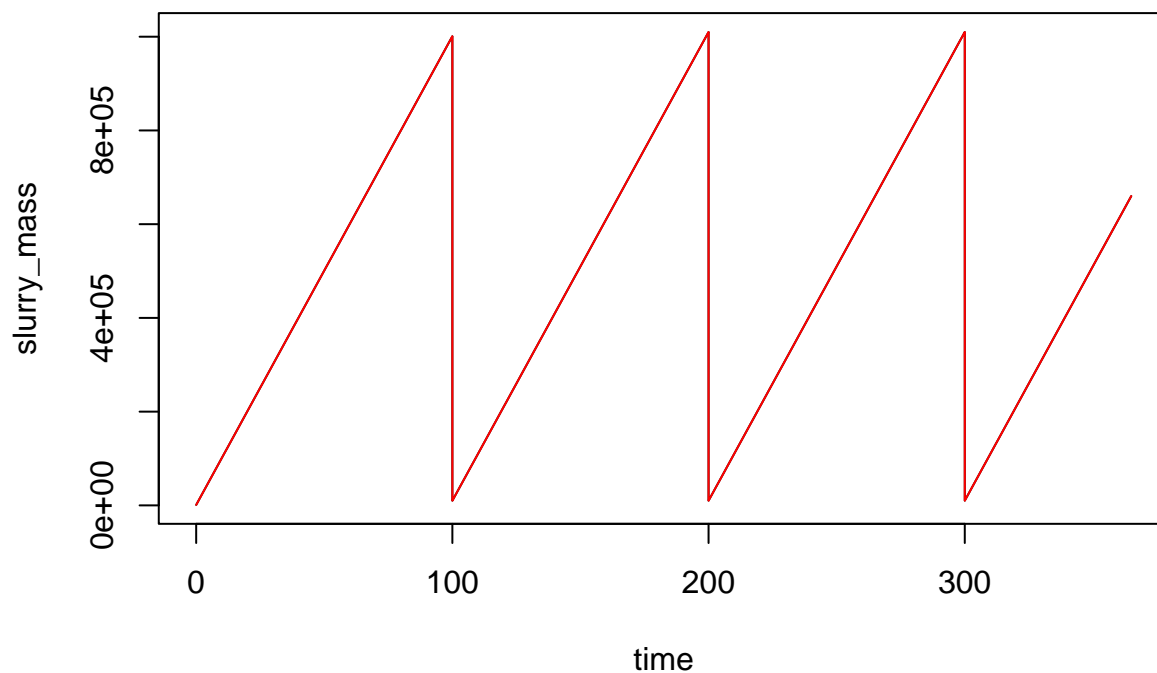
```
devtools::load_all()
```

```
## i Loading ABM
```

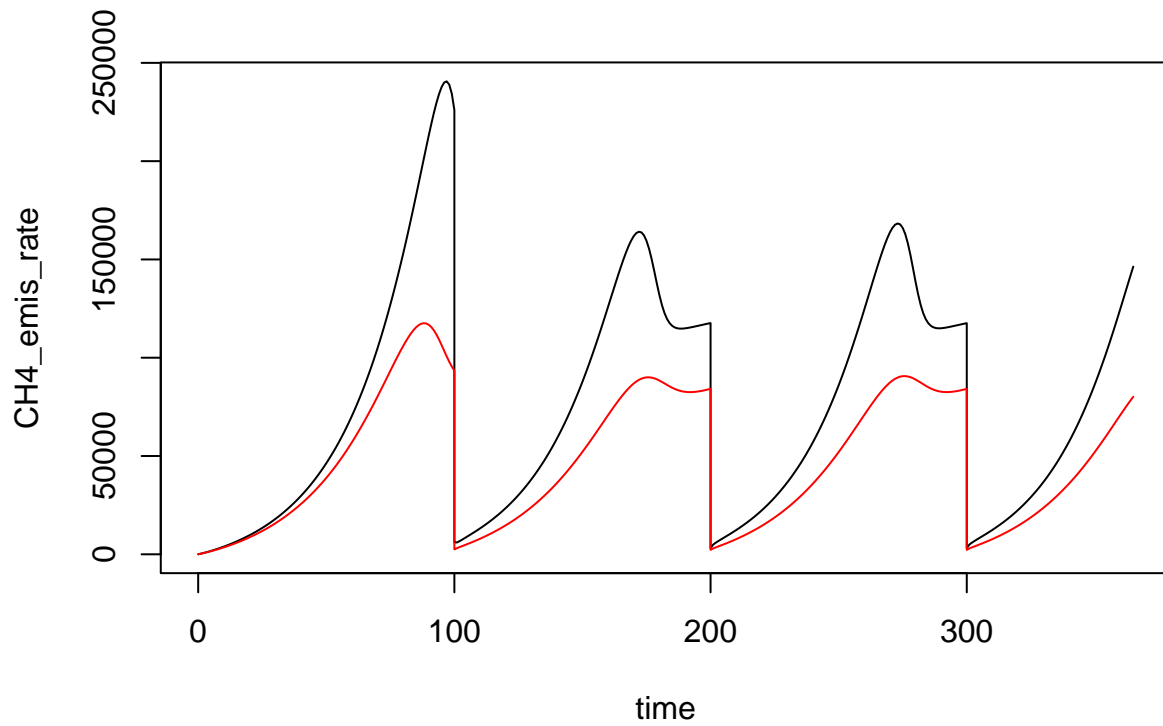
```
out2 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars2,
  chem_pars = chem_pars)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

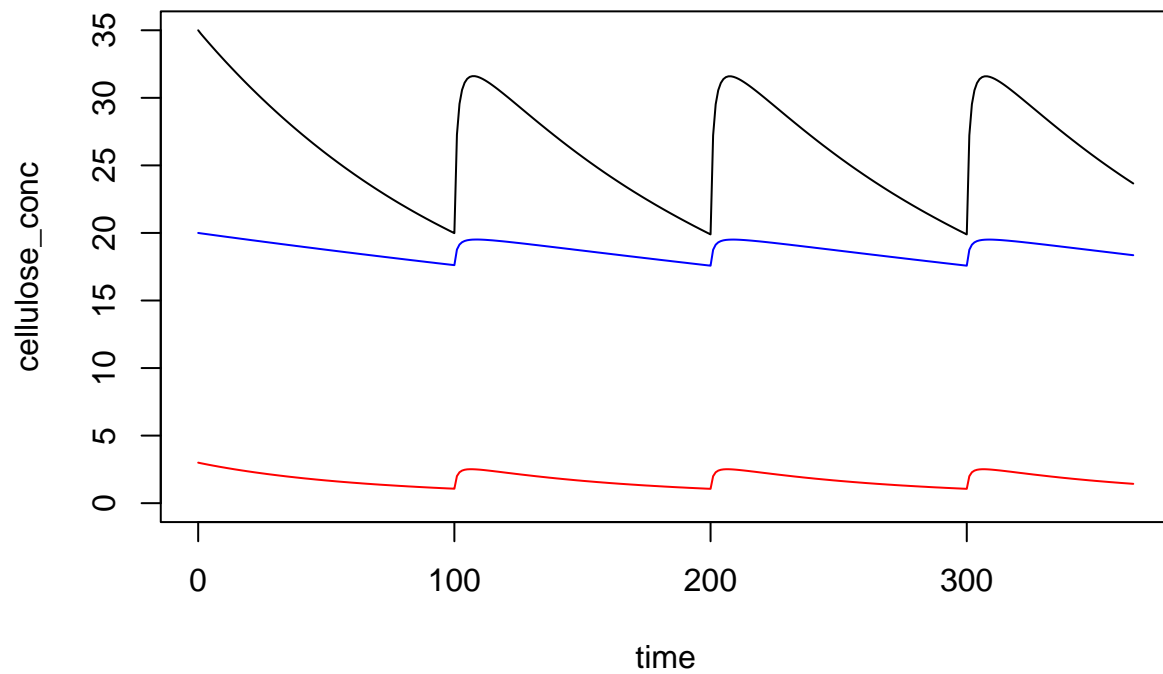
```
plot(slurry_mass ~ time, data = out2, type = 'l')
lines(slurry_mass ~ time, data = out1, col = 'red')
```



```
plot(CH4_emis_rate ~ time, data = out1, type = 'l')
lines(CH4_emis_rate ~ time, data = out2, col = 'red')
```

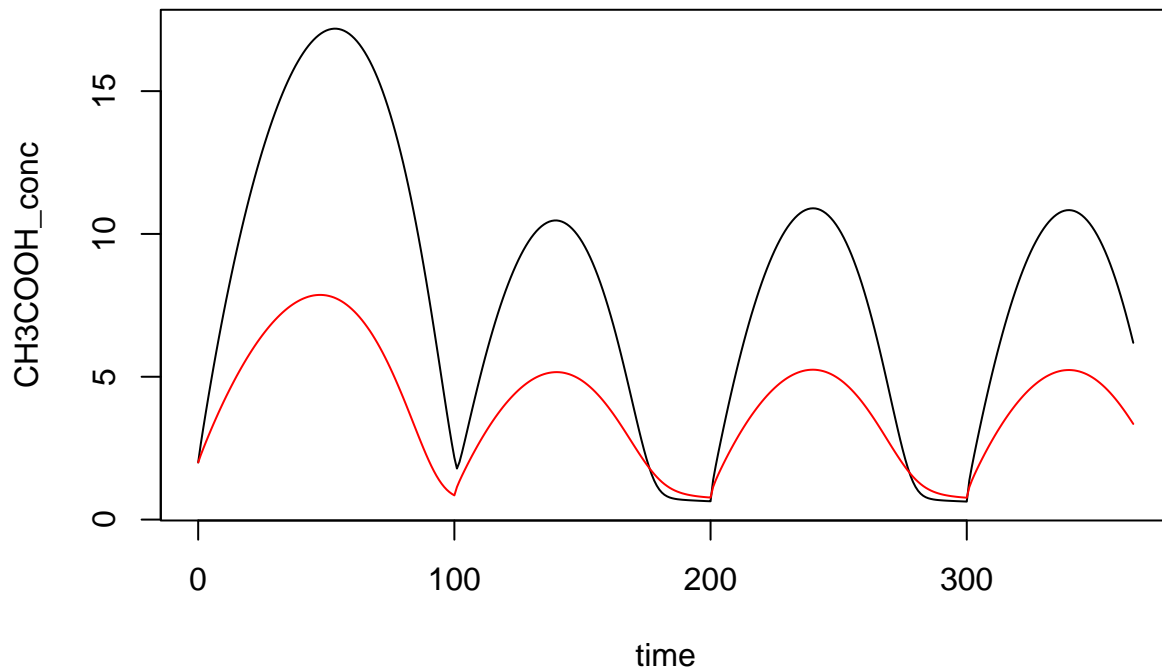


```
plot(cellulose_conc ~ time, data = out2, type = 'l', ylim = c(0, 35))
lines(lipids_conc ~ time, data = out2, type = 'l', col = 'red')
lines(protein_conc ~ time, data = out2, type = 'l', col = 'blue')
```



```
plot(CH3COOH_conc ~ time, data = out1, type = 'l')
lines(CH3COOH_conc ~ time, data = out2, col = 'red')
```



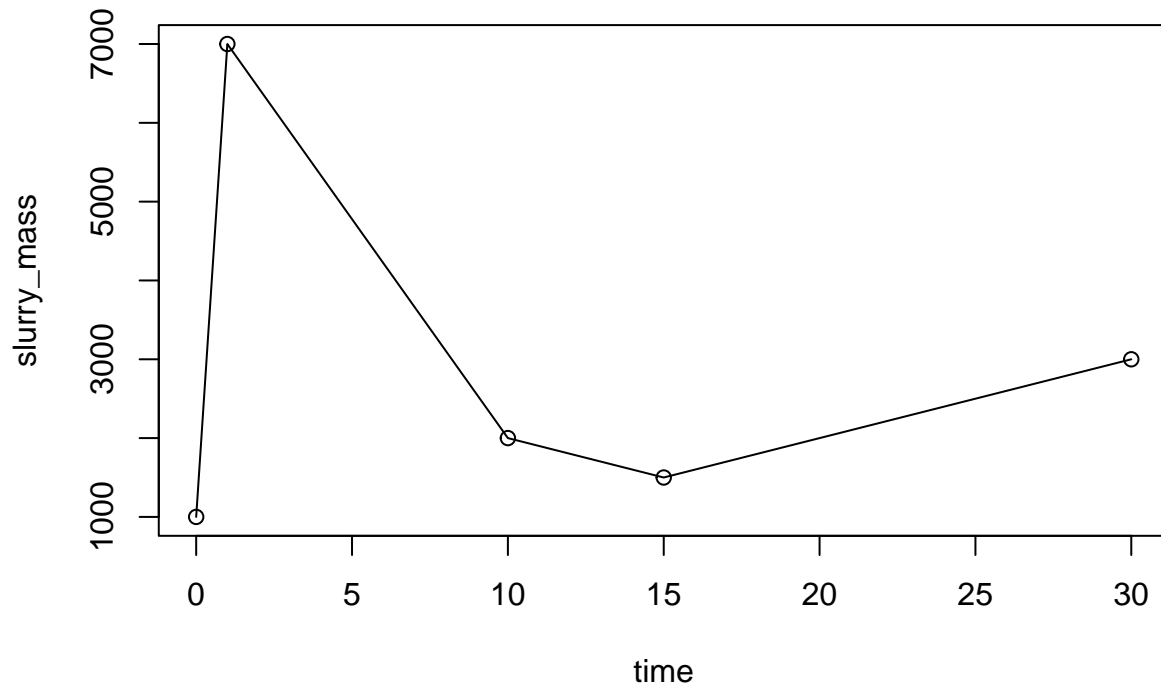


This flexibility comes from an approach similar to what we used for microbial groups.

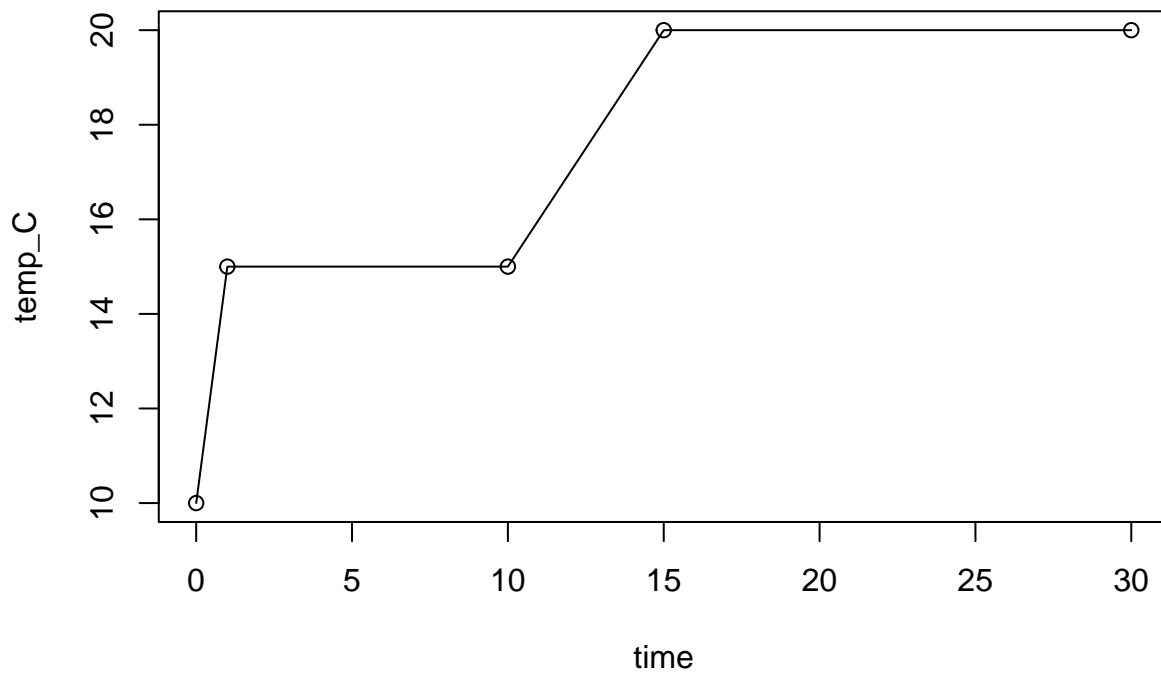
### 3. Time-variable inputs part 1

The `abm()` function can handle variability over time in any inputs now. Here slurry mass and temperature will vary.

```
var_dat <- data.frame(time = c(0, 1, 10, 15, 30),  
                      slurry_mass = c(1000, 7000, 2000, 1500, 3000),  
                      temp_C = c(10, 15, 15, 20, 20))  
  
plot(slurry_mass ~ time, data = var_dat, type = 'o')
```



```
plot(temp_C ~ time, data = var_dat, type = 'o')
```



This data frame goes in the `var_pars` argument, which must be a list, even though it might have a single element named `var`. The `var` element is the only required one. The `var` data frame must have a `slurry_mass` column if it is used—it is not possible to use an `abm_regular()`-like approach with variable temperature etc.

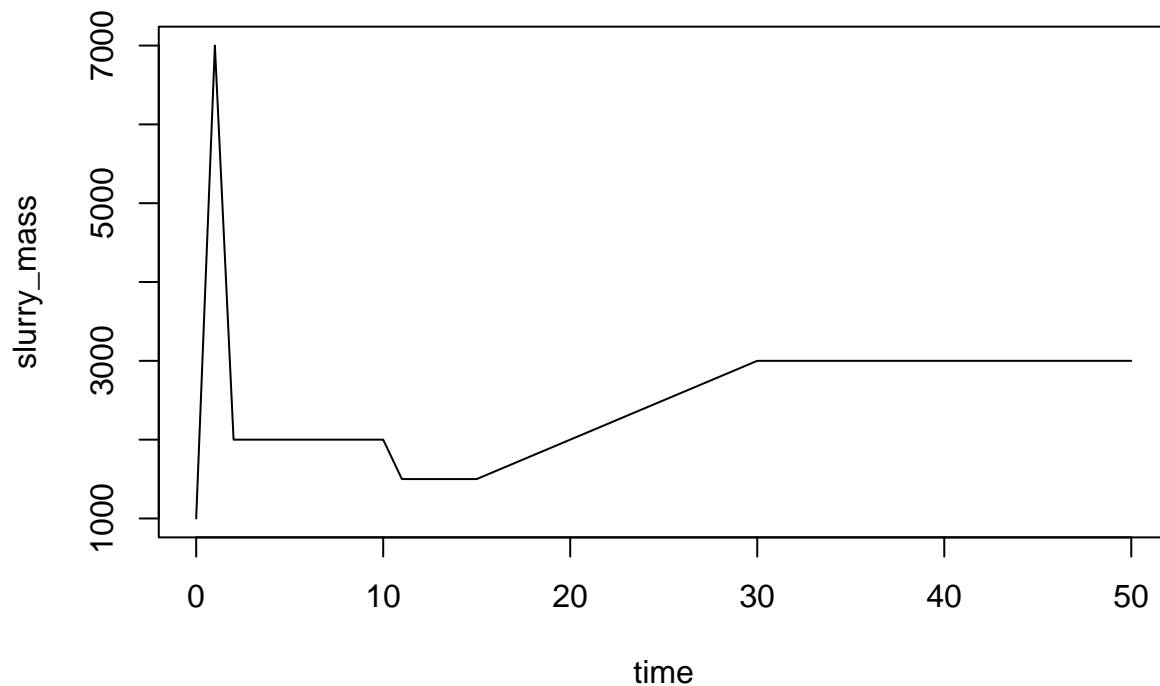
```
var_pars <- list(var = var_dat)
```

```
out3a <- abm(50,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
```

```
mic_pars = mic_pars,
sub_pars = sub_pars,
chem_pars = chem_pars,
var_pars = var_pars)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

```
plot(slurry_mass ~ time, data = out3a, type = 'l')
```



The “late” and “mid” options are still available, but now through `ctrl_pars`. Here we can change the value through `add_pars`

```
out3b <- abm(50,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars,
  var_pars = var_pars,
  add_pars = list(approx_method = 'late'))
```

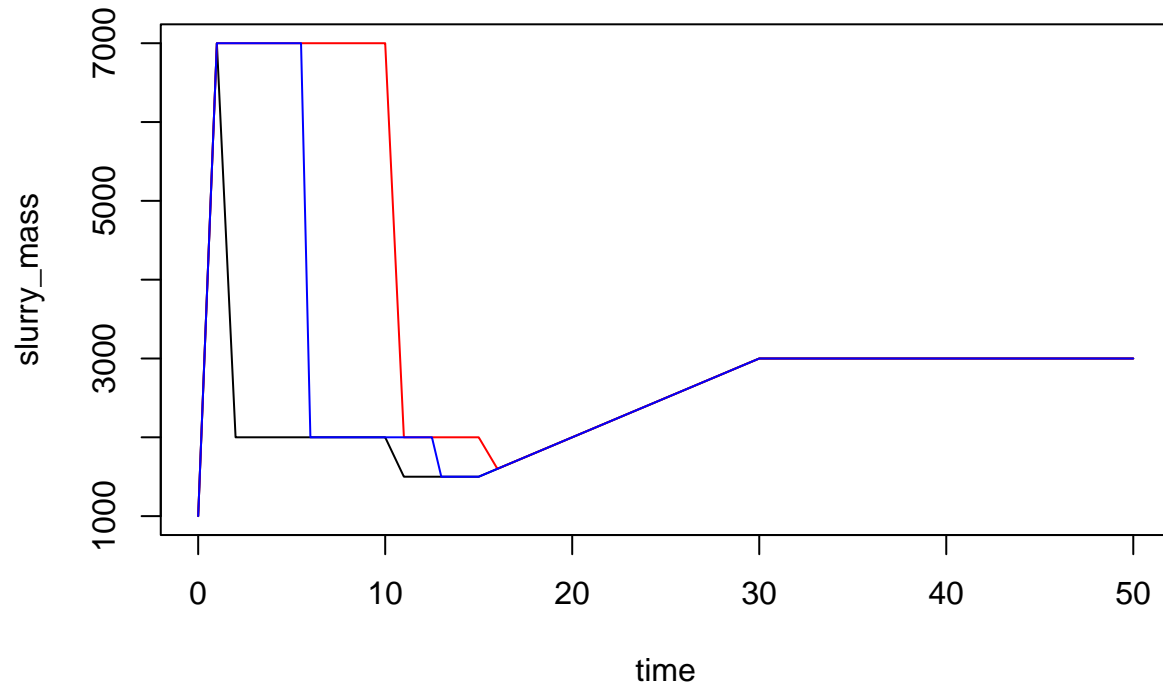
```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

```
out3c <- abm(50,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars,
  var_pars = var_pars,
```

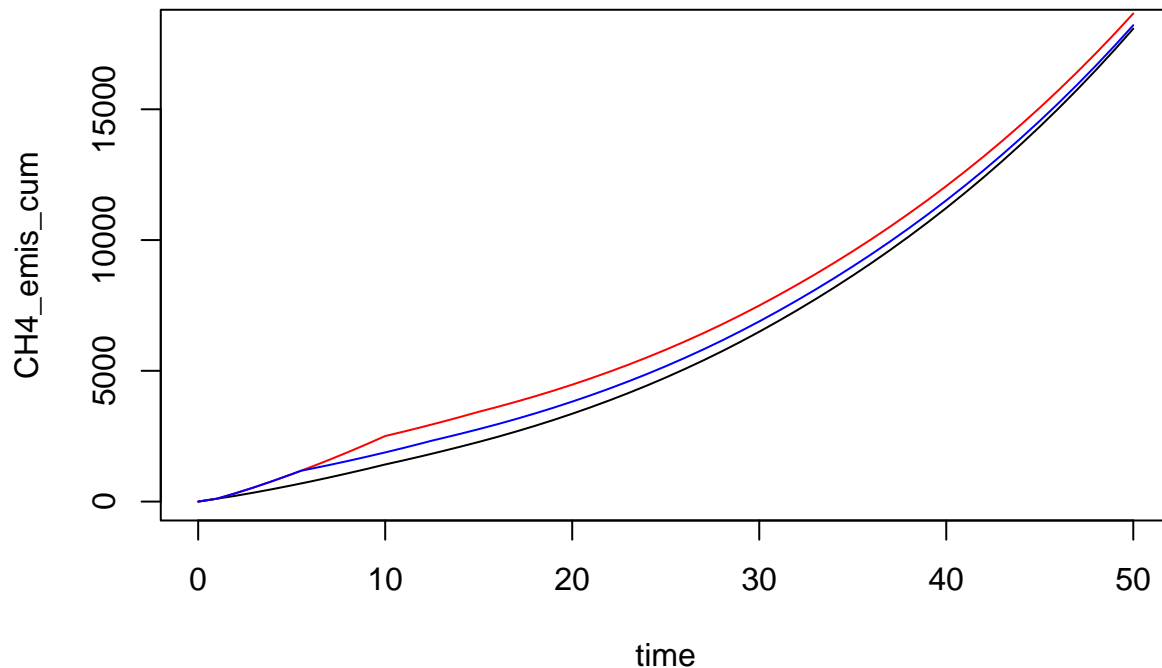
```
add_pars = list(approx_method = 'mid'))
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):  
## Size-variable parameter problem: Missing element(s) in kss.
```

```
plot(slurry_mass ~ time, data = out3a, type = 'l')  
lines(slurry_mass ~ time, data = out3b, col = 'red')  
lines(slurry_mass ~ time, data = out3c, col = 'blue')
```



```
plot(CH4_emis_cum ~ time, data = out3a, type = 'l')  
lines(CH4_emis_cum ~ time, data = out3b, col = 'red')  
lines(CH4_emis_cum ~ time, data = out3c, col = 'blue')
```



## 4. Time-variable inputs part 2

Here we'll vary fresh substrate concentrations over time.

First the data frame with slurry mass.

```
var_dat <- data.frame(time = c(0, 1, 10, 15, 30, 50),
                      slurry_mass = c(1000, 7000, 2000, 5000, 3000, 10000))
var_dat
```

```
##   time slurry_mass
## 1    0         1000
## 2    1         7000
## 3   10         2000
## 4   15         5000
## 5   30         3000
## 6   50        10000
```

Then add `sub_fresh` values. Each row needs a list containing a named vector. This is somewhat unusual data frame usage, and there is a user-friendly alternative based on additional data frames in the `var` argument (see next section). But I've kept this demo.

```
var_dat
##   time slurry_mass
## 1    0         1000
## 2    1         7000
## 3   10         2000
## 4   15         5000
## 5   30         3000
## 6   50        10000

var_dat$sub_fresh <- rep(list(c(VSd = 50)), nrow(var_dat))
var_dat$sub_fresh[3] <- list(c(VSd = 100))
```

```

var_dat$sub_fresh[4] <- list(c(VSd = 10))
var_dat$sub_fresh[5] <- list(c(VSd = 0))
var_dat$sub_fresh[6] <- list(c(VSd = 200))
var_dat

```

```

##   time slurry_mass sub_fresh
## 1    0         1000         50
## 2    1         7000         50
## 3   10         2000        100
## 4   15         5000         10
## 5   30         3000          0
## 6   50        10000        200

```

```
var_dat[1, 3]
```

```

## [[1]]
## VSd
## 50

```

```
var_dat[5, 3]
```

```

## [[1]]
## VSd
## 0

```

```
devtools::load_all()
```

```
## i Loading ABM
```

```

out4a <- abm(100,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars,
  var_pars = var_pars)

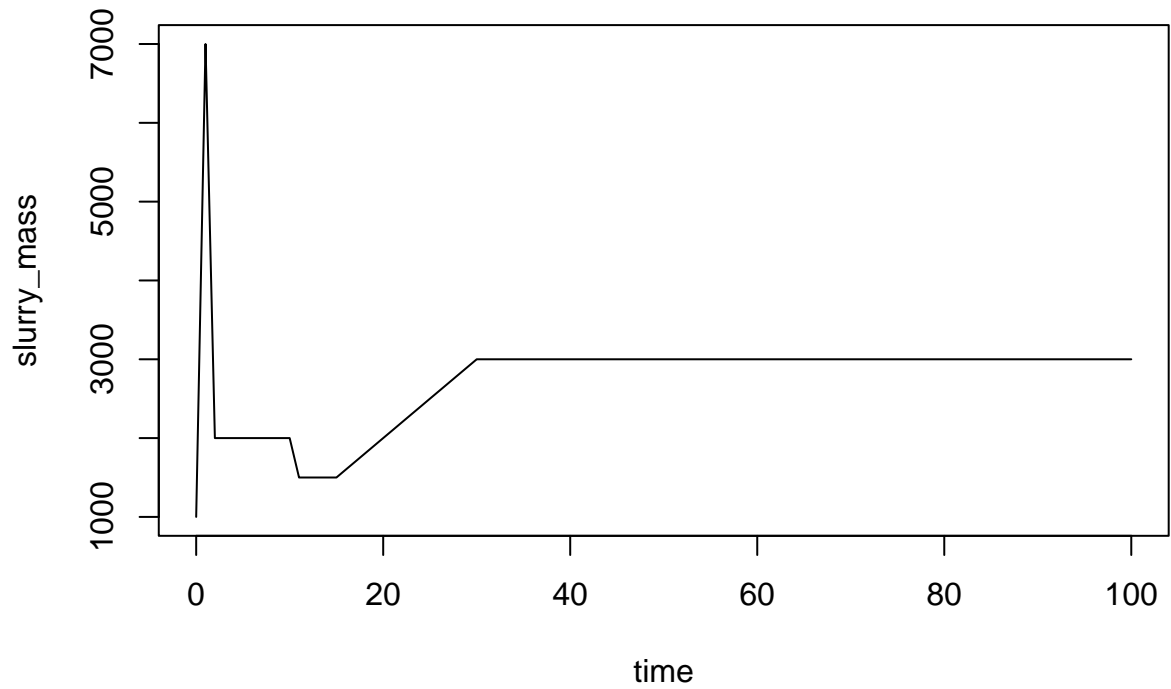
```

```

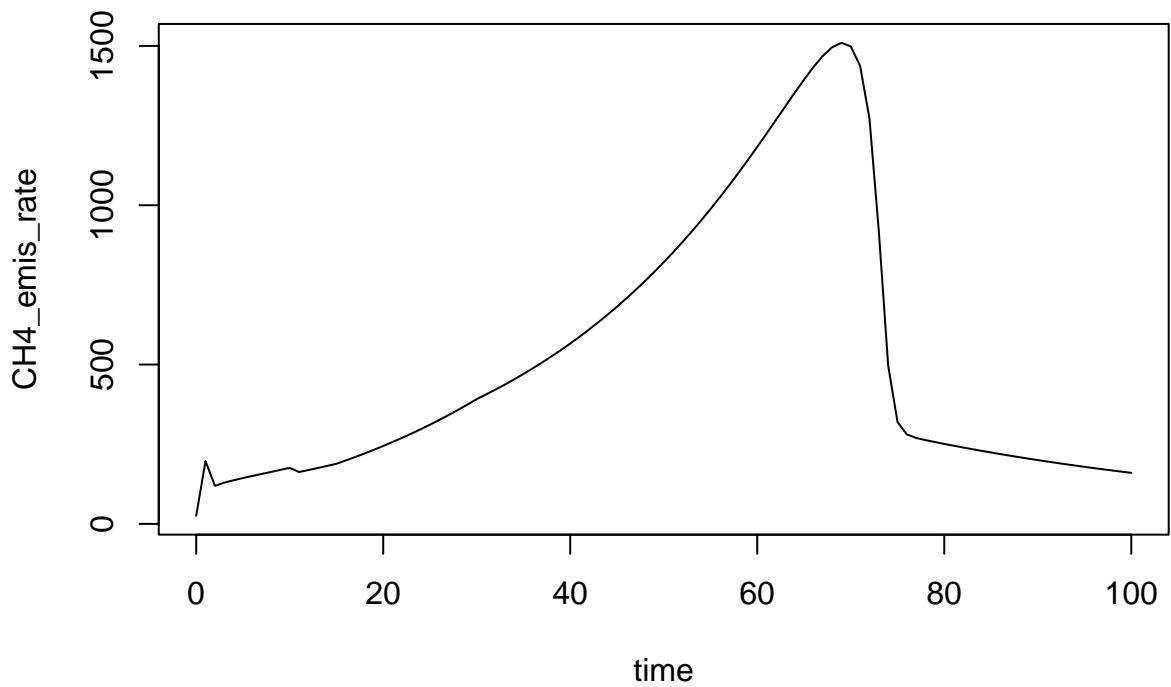
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.

```

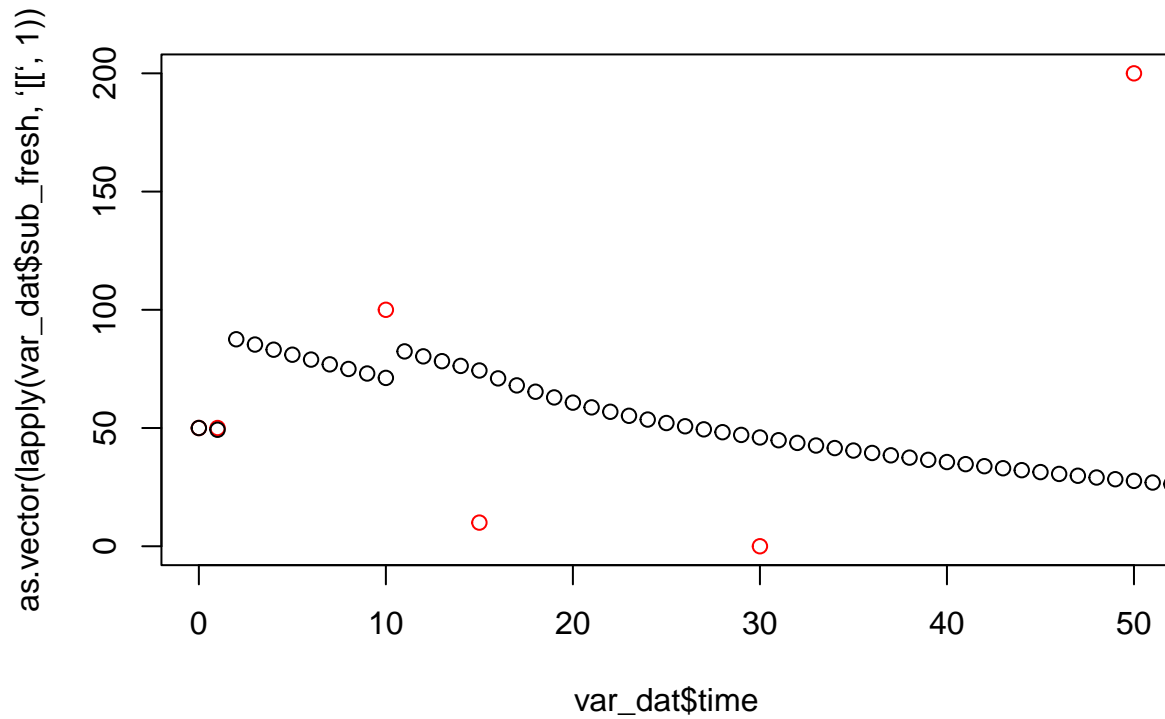
```
plot(slurry_mass ~ time, data = out4a, type = 'l')
```



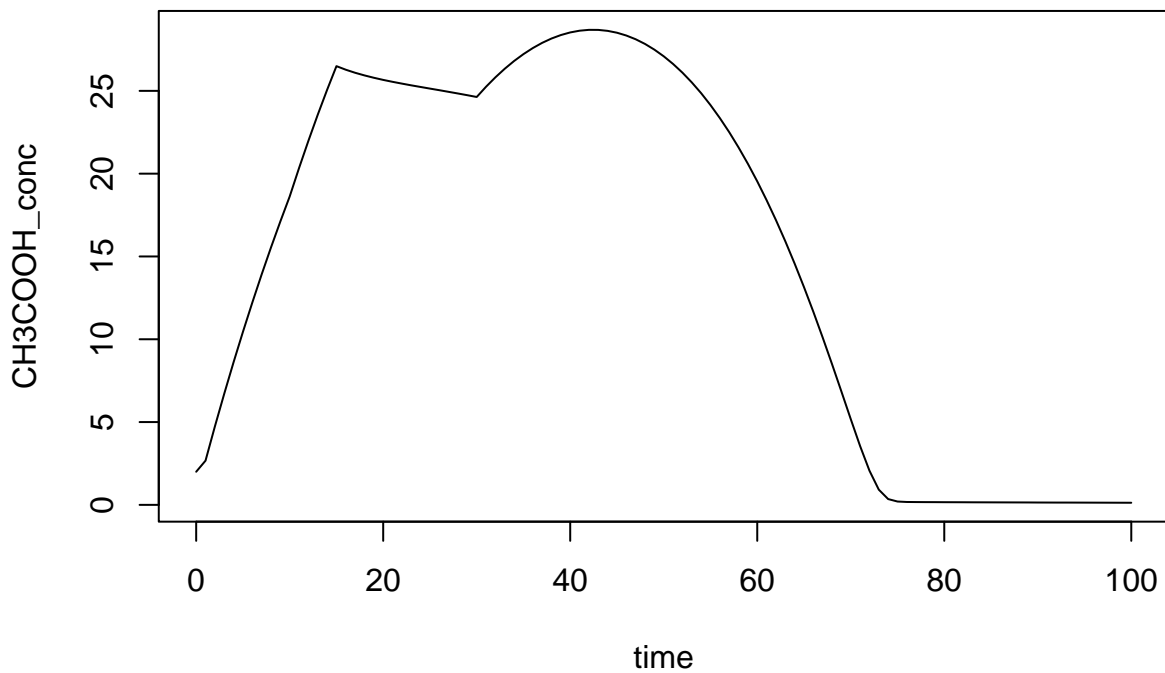
```
plot(CH4_emis_rate ~ time, data = out4a, type = 'l')
```



```
plot(var_dat$time, as.vector(lapply(var_dat$sub_fresh, `[`, 1)), type = 'p', col = 'red')
lines(VSd_conc ~ time, data = out4a, type = 'p')
```



```
plot(CH3COOH_conc ~ time, data = out4a, type = 'l')
```



Let's vary some microbial parameters as well. And pH.

```
var_dat <- data.frame(time = c(0, 1, 10, 15, 30, 50),
                      slurry_mass = c(1000, 7000, 2000, 5000, 3000, 10000),
                      pH = 7 - 0:5/10)
var_dat
```

```
##   time slurry_mass  pH
## 1    0         1000 7.0
```



```
## 2    1      7000 6.9
## 3   10      2000 6.8
## 4   15      5000 6.7
## 5   30      3000 6.6
## 6   50     10000 6.5
```

VSd.

```
var_dat$sub_fresh <- rep(list(c(VSd = 50)), nrow(var_dat))
var_dat$sub_fresh[3] <- list(c(VSd = 100))
```

Some microbial parameters for a shift in temperature optima, “adaptation” for example.

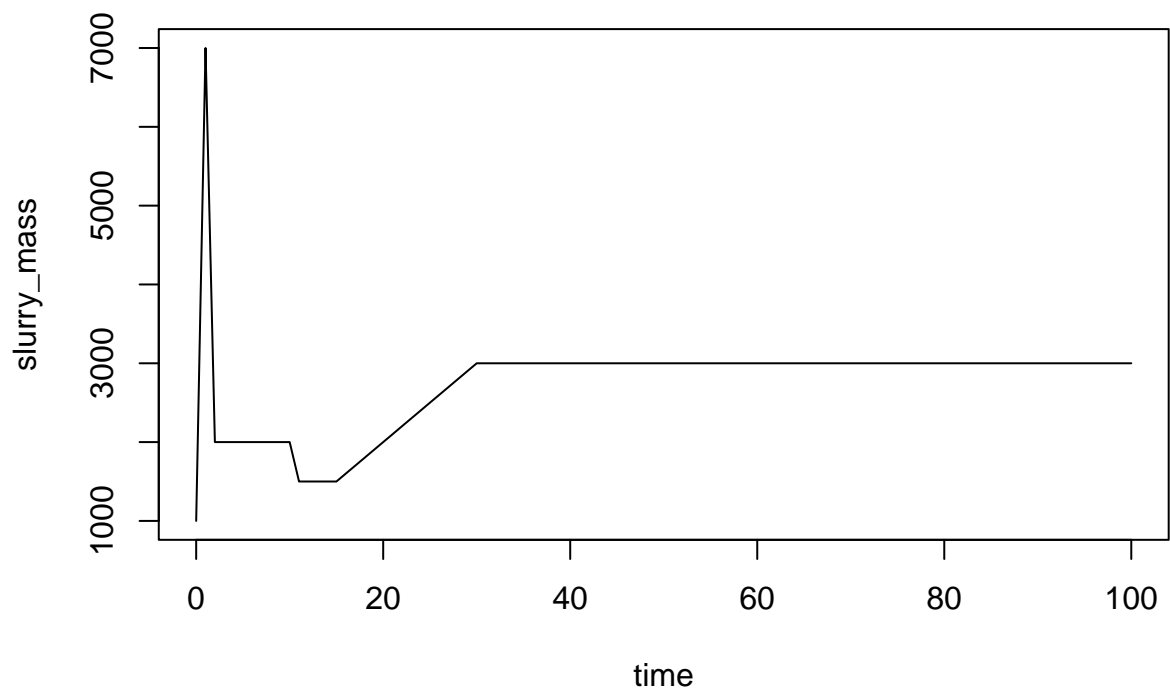
```
for (i in 1:nrow(var_dat)) {
  var_dat$T_opt[i] <- list(grp_pars$T_opt + 2 * i)
}
var_dat
```

```
##    time slurry_mass  pH sub_fresh      T_opt
## 1     0      1000 7.0      50 20, 20, 30, 46
## 2     1      7000 6.9      50 22, 22, 32, 48
## 3    10      2000 6.8     100 24, 24, 34, 50
## 4    15      5000 6.7      50 26, 26, 36, 52
## 5    30      3000 6.6      50 28, 28, 38, 54
## 6    50     10000 6.5      50 30, 30, 40, 56
```

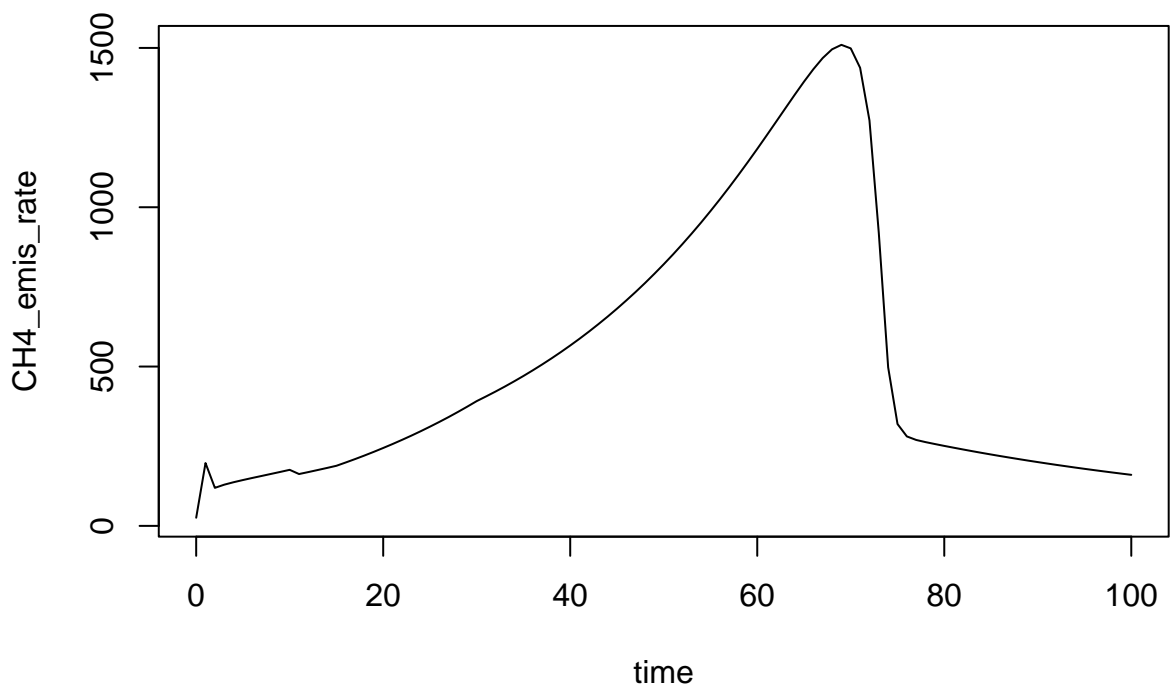
```
out4b <- abm(100,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars,
  var_pars = var_pars)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

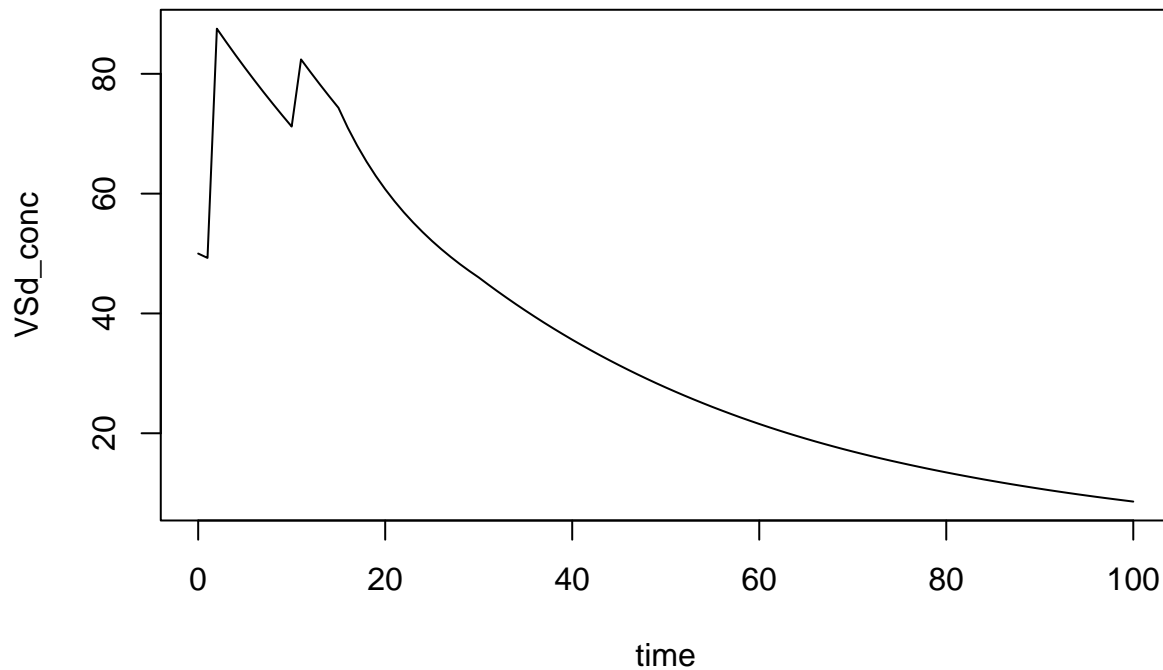
```
plot(slurry_mass ~ time, data = out4b, type = 'l')
```



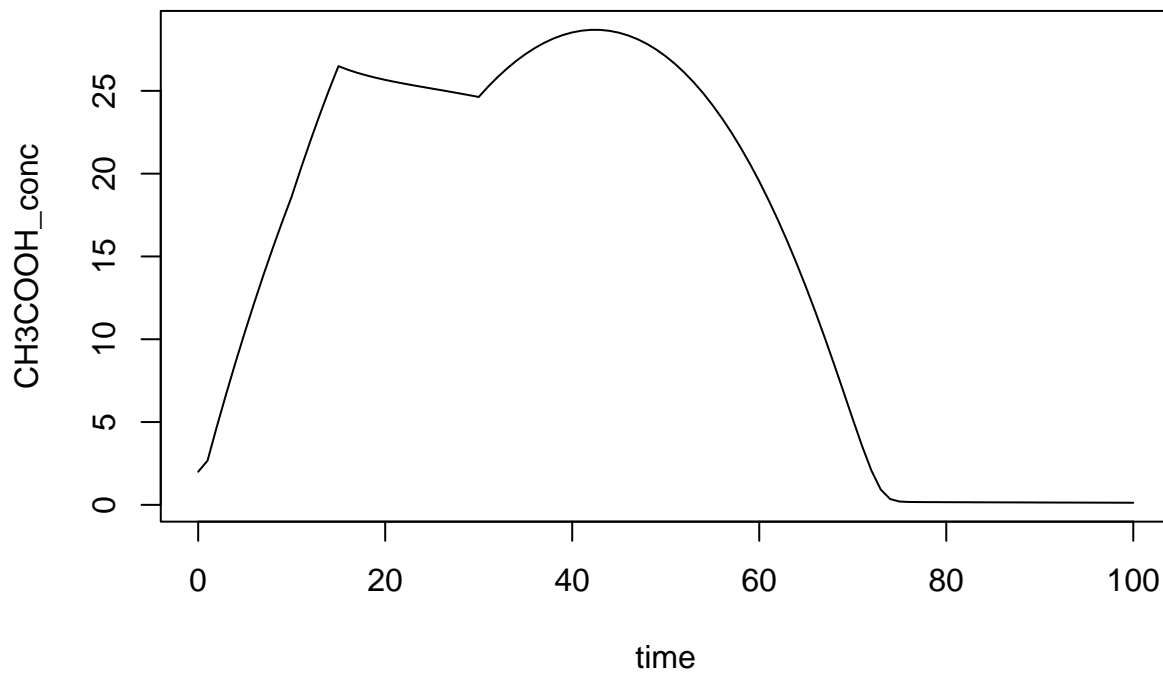
```
plot(CH4_emis_rate ~ time, data = out4b, type = 'l')
```



```
plot(VSd_conc ~ time, data = out4b, type = 'l')
```



```
plot(CH3COOH_conc ~ time, data = out4b, type = 'l')
```



## 5. Time-variable inputs part 3

The list-in-data frame approach is clunky. Here is an alternative.

```
var_dat <- data.frame(time = c(0, 1, 10, 15, 30, 50),
                      slurry_mass = c(1000, 7000, 2000, 5000, 3000, 10000))
var_dat
```

```
##   time slurry_mass
```

```
## 1    0      1000
## 2    1      7000
## 3   10      2000
## 4   15      5000
## 5   30      3000
## 6   50     10000
```

Make a separate data frame for each other argument (any name, but note column names!).

```
sub_fresh_dat = data.frame(time = c(0, 1, 10, 15, 30, 50),
                           VSd = c(50, 100, 0, 0, 200, 50))
```

```
T_opt_dat = data.frame(time = c(0, 1, 10, 15, 30, 50),
                       m1 = 20 + 0:5 * 2,
                       m2 = 20 + 0:5 * 2,
                       m3 = 30 + 0:5 * 2,
                       sr1 = 46 + 0:5 * 2)
```

and combine them in a list, using the parameter element names for element names (e.g., `sub_fresh` is the name of an element in `sub_pars`).

```
var_pars <- list(var = var_dat, sub_fresh = sub_fresh_dat, T_opt = T_opt_dat)
```

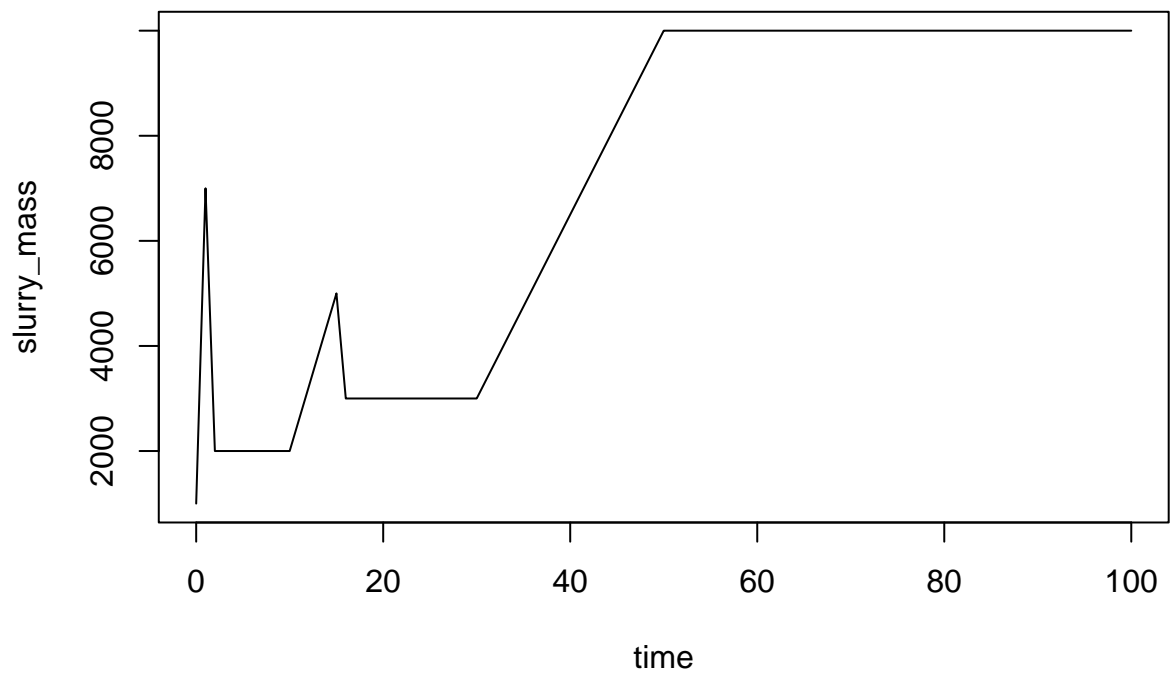
```
devtools::load_all()
```

```
## i Loading ABM
```

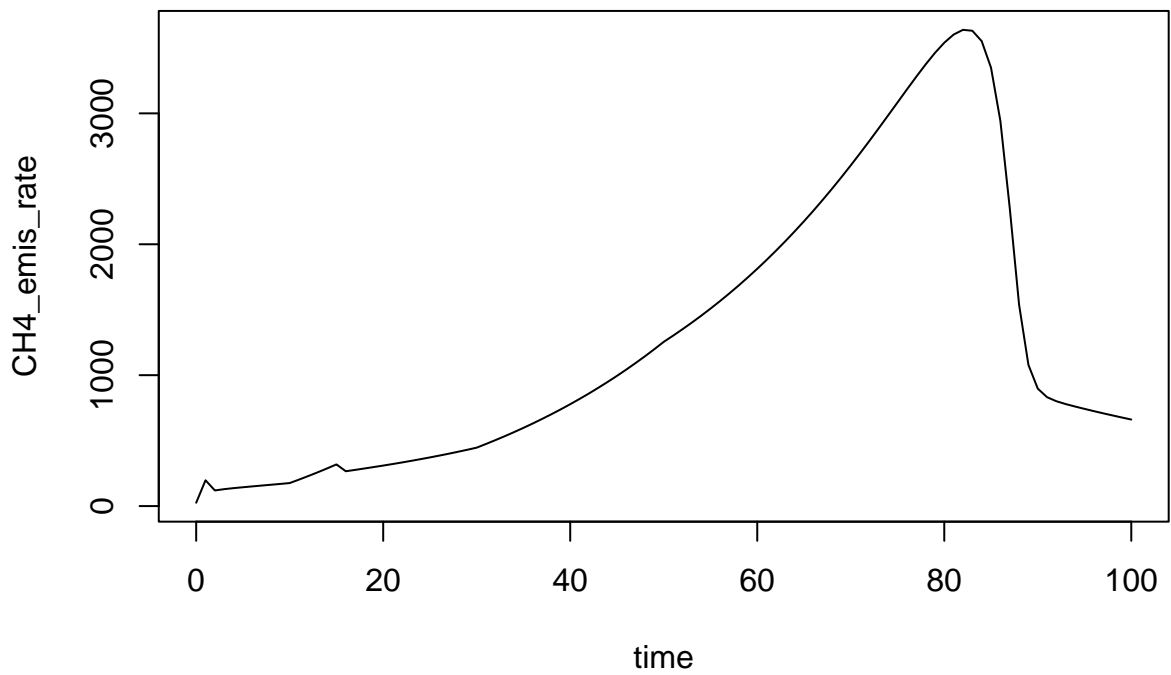
```
out5 <- abm(100,
            mng_pars = mng_pars,
            man_pars = man_pars,
            grp_pars = grp_pars,
            mic_pars = mic_pars,
            sub_pars = sub_pars,
            chem_pars = chem_pars,
            var_pars = var_pars)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

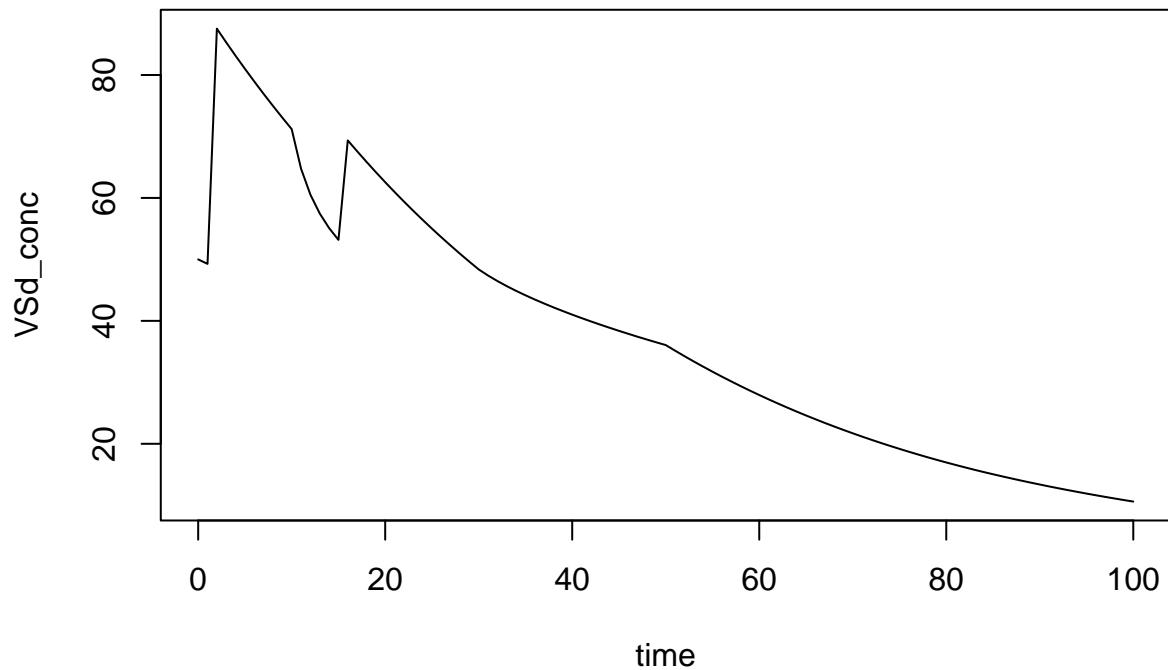
```
plot(slurry_mass ~ time, data = out5, type = 'l')
```



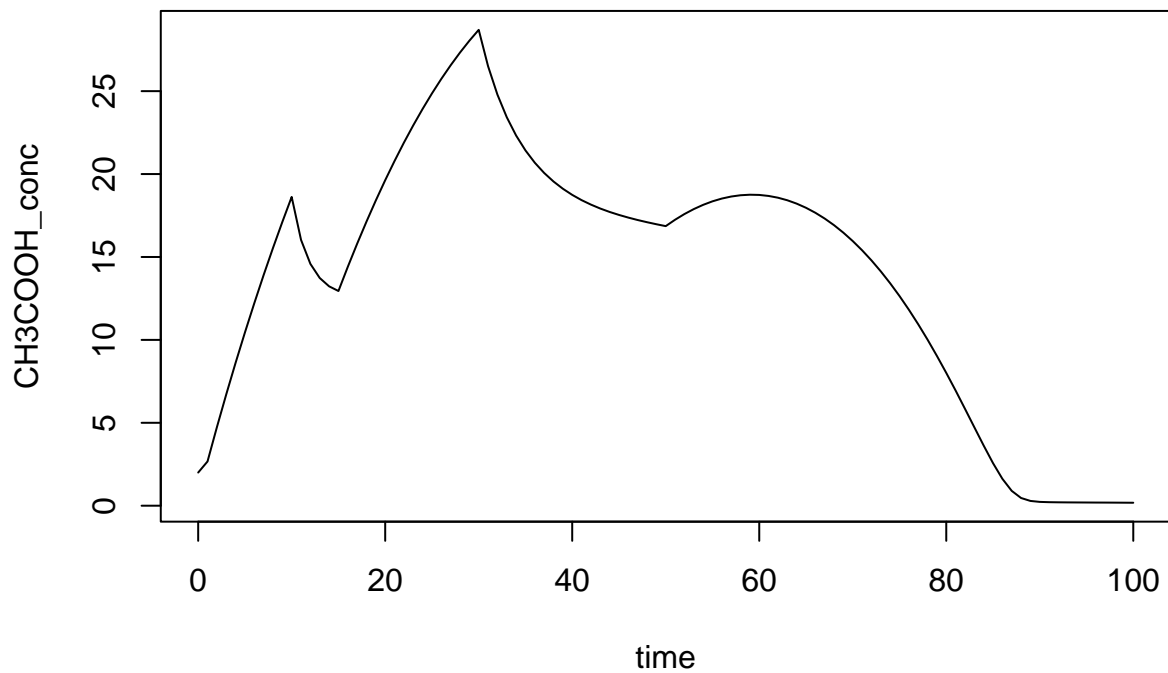
```
plot(CH4_emis_rate ~ time, data = out5, type = 'l')
```



```
plot(VSd_conc ~ time, data = out5, type = 'l')
```



```
plot(CH3COOH_conc ~ time, data = out5, type = 'l')
```



## 6. Flexible solutes

Any conservative solute can be added in `man_pars`, using any names. I am moving toward using a “master species” approach, so it makes sense to use the chemical formula of the primary species, with `p` or `m` for a charge symbol. The `comp` part of the name below is for “component”.

```
man_pars6 <- list(comps = c('H2S', 'SO4m2', 'NH4p'),
  comp_fresh = c(H2S = 0.01, SO4m2 = 0.2, NH4p = 2.5),
  VFA_fresh = c(CH3COOH = 2),
```

```
pH = 7, dens = 1000)
```

Note that CH3COOH is still special—it has a fixed name in the code and is not conservative.

```
devtools::load_all()
```

```
## i Loading ABM
```

```
out6a <- abm(365,  
  mng_pars = mng_pars,  
  man_pars = man_pars6,  
  grp_pars = grp_pars,  
  mic_pars = mic_pars,  
  sub_pars = sub_pars,  
  chem_pars = chem_pars)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):  
## Size-variable parameter problem: Missing element(s) in kss.
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =  
## pars$COD_conv, : COD balance is off by 1.4%
```

```
tail(out6a)
```

```
##      time      m0      m1      m2      sr1      VSd H2S SO4m2 NH4p  
## 364 360 74916.39 72503.00 337818.8 82704.67 15488476 6100 122000 1525000  
## 365 361 77063.33 74548.96 351980.7 85137.23 15597076 6200 124000 1550000  
## 366 362 79241.30 76623.17 366608.7 87616.74 15703312 6300 126000 1575000  
## 367 363 81447.98 78723.37 381698.7 90142.39 15807257 6400 128000 1600000  
## 368 364 83680.45 80846.76 397241.9 92713.00 15908980 6500 130000 1625000  
## 369 365 85935.10 82989.88 413223.8 95326.92 16008549 6600 132000 1650000  
##      CH3COOH slurry_mass CH4_emis_cum slurry_load COD_load CH4_emis_rate temp_C  
## 364 4795391      610000      26879564      3600000 187920000      125252.0      20  
## 365 4683511      620000      27006946      3610000 188442000      129516.0      20  
## 366 4556462      630000      27138603      3620000 188964000      133800.5      20  
## 367 4414148      640000      27274545      3630000 189486000      138080.1      20  
## 368 4256596      650000      27414751      3640000 190008000      142322.7      20  
## 369 4083986      660000      27559165      3650000 190530000      146488.5      20  
##      pH m0_eff m1_eff m2_eff sr1_eff VSd_eff H2S_eff SO4m2_eff NH4p_eff  
## 364 7 442067.8 422515.1 2243571 529520.2 53725160 29910 598200 7477500  
## 365 7 442067.8 422515.1 2243571 529520.2 53725160 29910 598200 7477500  
## 366 7 442067.8 422515.1 2243571 529520.2 53725160 29910 598200 7477500  
## 367 7 442067.8 422515.1 2243571 529520.2 53725160 29910 598200 7477500  
## 368 7 442067.8 422515.1 2243571 529520.2 53725160 29910 598200 7477500  
## 369 7 442067.8 422515.1 2243571 529520.2 53725160 29910 598200 7477500  
##      CH3COOH_eff slurry_mass_eff slurry_depth m0_conc m1_conc m2_conc  
## 364 3444790      2991000      6.1 0.1228138 0.1188574 0.5538013  
## 365 3444790      2991000      6.2 0.1242957 0.1202403 0.5677108  
## 366 3444790      2991000      6.3 0.1257798 0.1216241 0.5819186  
## 367 3444790      2991000      6.4 0.1272625 0.1230053 0.5964042  
## 368 3444790      2991000      6.5 0.1287392 0.1243796 0.6111414  
## 369 3444790      2991000      6.6 0.1302047 0.1257422 0.6260966  
##      sr1_conc VSd_conc H2S_conc SO4m2_conc NH4p_conc CH3COOH_conc m0_eff_conc  
## 364 0.1355814 25.39094      0.01      0.2      2.5      7.861296      0.1477993  
## 365 0.1373181 25.15657      0.01      0.2      2.5      7.554051      0.1477993  
## 366 0.1390742 24.92589      0.01      0.2      2.5      7.232480      0.1477993
```

```
## 367 0.1408475 24.69884 0.01 0.2 2.5 6.897107 0.1477993
## 368 0.1426354 24.47535 0.01 0.2 2.5 6.548610 0.1477993
## 369 0.1444347 24.25538 0.01 0.2 2.5 6.187857 0.1477993
## m1_eff_conc m2_eff_conc sr1_eff_conc VSd_eff_conc H2S_eff_conc
## 364 0.1412622 0.7501073 0.1770378 17.96227 0.01
## 365 0.1412622 0.7501073 0.1770378 17.96227 0.01
## 366 0.1412622 0.7501073 0.1770378 17.96227 0.01
## 367 0.1412622 0.7501073 0.1770378 17.96227 0.01
## 368 0.1412622 0.7501073 0.1770378 17.96227 0.01
## 369 0.1412622 0.7501073 0.1770378 17.96227 0.01
## S04m2_eff_conc NH4p_eff_conc CH3COOH_eff_conc
## 364 0.2 2.5 1.151719
## 365 0.2 2.5 1.151719
## 366 0.2 2.5 1.151719
## 367 0.2 2.5 1.151719
## 368 0.2 2.5 1.151719
## 369 0.2 2.5 1.151719
```

```
head(out6a)
```

```
## time m0 m1 m2 sr1 VSd H2S S04m2 NH4p
## 1 0 50.0000 50.0000 50.0000 50.000 50000.0 10 200 2500
## 2 1 554.0098 553.8532 558.3748 555.424 542318.5 110 2200 27500
## 3 2 1066.2767 1065.6731 1083.1939 1071.000 1022077.5 210 4200 52500
## 4 3 1588.3159 1586.9428 1627.0195 1597.789 1489599.4 310 6200 77500
## 5 4 2121.2032 2118.7111 2191.8591 2136.557 1945198.4 410 8200 102500
## 6 5 2665.7723 2661.7874 2779.4357 2687.920 2389180.8 510 10200 127500
## CH3COOH slurry_mass CH4_emis_cum slurry_load COD_load CH4_emis_rate temp_C
## 1 2000.00 1000 0.0000 0 0 25.52844 20
## 2 29018.42 11000 163.6268 10000 522000 308.65442 20
## 3 67370.97 21000 628.8106 20000 1044000 626.59218 20
## 4 116610.76 31000 1425.4312 30000 1566000 970.52434 20
## 5 176325.85 41000 2577.0170 40000 2088000 1335.99603 20
## 6 246126.14 51000 4103.8015 50000 2610000 1720.63977 20
## pH m0_eff m1_eff m2_eff sr1_eff VSd_eff H2S_eff S04m2_eff NH4p_eff
## 1 7 0 0 0 0 0 0 0 0
## 2 7 0 0 0 0 0 0 0 0
## 3 7 0 0 0 0 0 0 0 0
## 4 7 0 0 0 0 0 0 0 0
## 5 7 0 0 0 0 0 0 0 0
## 6 7 0 0 0 0 0 0 0 0
## CH3COOH_eff slurry_mass_eff slurry_depth m0_conc m1_conc m2_conc
## 1 0 0 0.01 0.05000000 0.05000000 0.05000000
## 2 0 0 0.11 0.05036453 0.05035029 0.05076134
## 3 0 0 0.21 0.05077508 0.05074634 0.05158066
## 4 0 0 0.31 0.05123600 0.05119170 0.05248450
## 5 0 0 0.41 0.05173666 0.05167588 0.05345998
## 6 0 0 0.51 0.05227004 0.05219191 0.05449874
## sr1_conc VSd_conc H2S_conc S04m2_conc NH4p_conc CH3COOH_conc m0_eff_conc
## 1 0.05000000 50.00000 0.01 0.2 2.5 2.000000 NaN
## 2 0.05049309 49.30168 0.01 0.2 2.5 2.638039 NaN
## 3 0.05099999 48.67036 0.01 0.2 2.5 3.208141 NaN
## 4 0.05154158 48.05159 0.01 0.2 2.5 3.761638 NaN
## 5 0.05211115 47.44386 0.01 0.2 2.5 4.300630 NaN
## 6 0.05270431 46.84668 0.01 0.2 2.5 4.826003 NaN
```



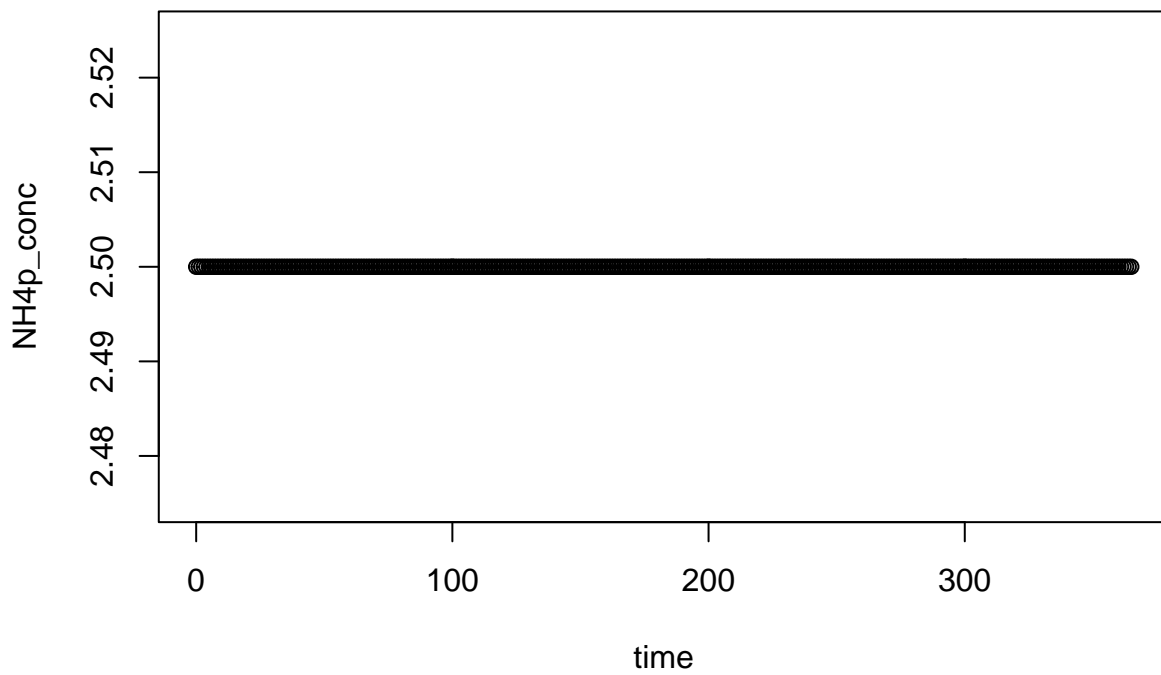
##	m1_eff_conc	m2_eff_conc	sr1_eff_conc	VSd_eff_conc	H2S_eff_conc	S04m2_eff_conc
## 1	NaN	NaN	NaN	NaN	NaN	NaN
## 2	NaN	NaN	NaN	NaN	NaN	NaN
## 3	NaN	NaN	NaN	NaN	NaN	NaN
## 4	NaN	NaN	NaN	NaN	NaN	NaN
## 5	NaN	NaN	NaN	NaN	NaN	NaN
## 6	NaN	NaN	NaN	NaN	NaN	NaN

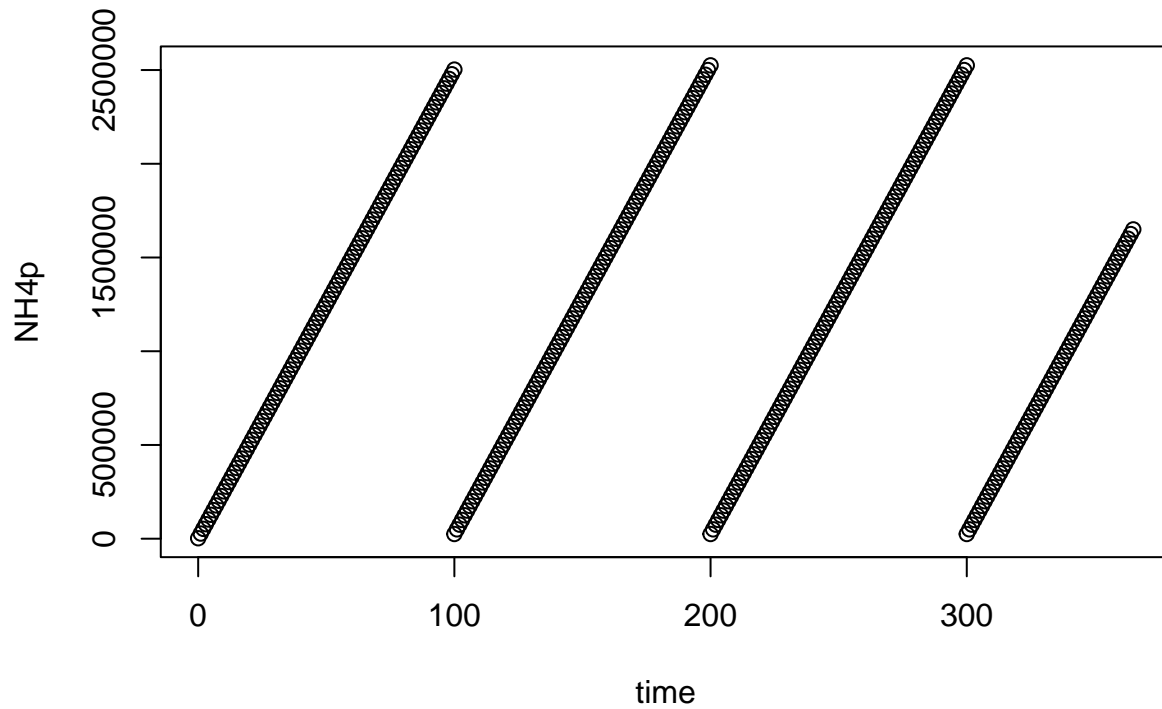
##	NH4p_eff_conc	CH3COOH_eff_conc
## 1	NaN	NaN
## 2	NaN	NaN
## 3	NaN	NaN
## 4	NaN	NaN
## 5	NaN	NaN
## 6	NaN	NaN

Conservative components are boring in output (without inhibition or volatilization).

```
plot(NH4p_conc ~ time, data = out6a)
```

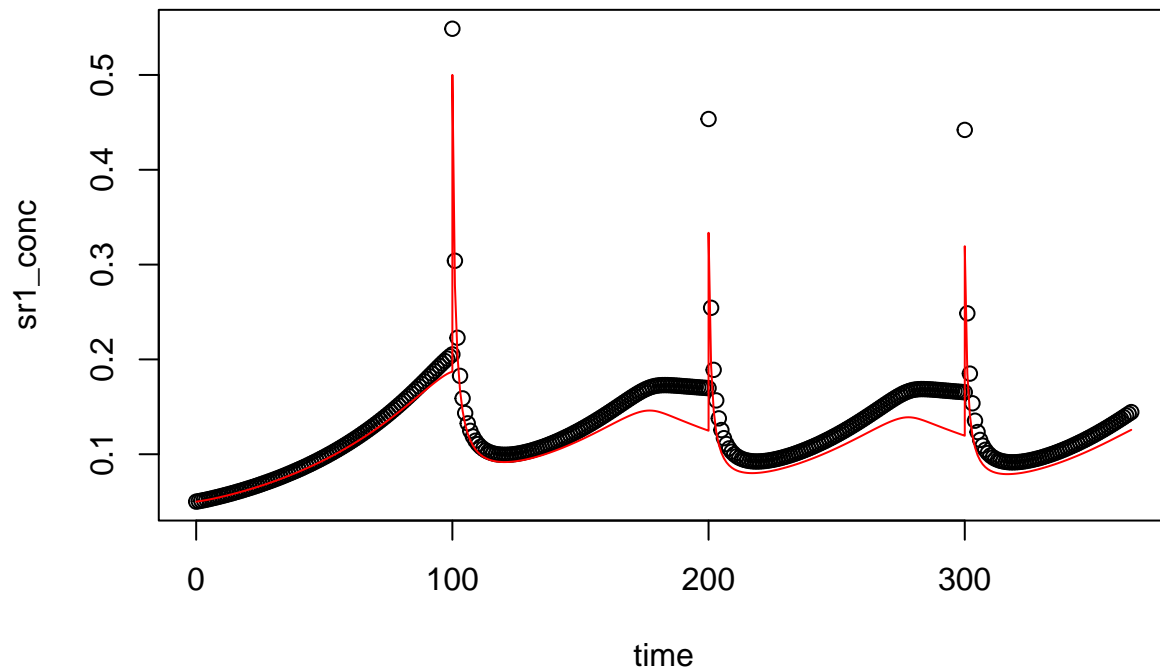


```
plot(NH4p ~ time, data = out6a)
```



But now with sulfate and sulfide, sulfate reducers can grow.

```
plot(sr1_conc ~ time, data = out6a)
lines(m1_conc ~ time, data = out6a, col = 'red')
```



We can see dilution effects at least if some washing water is added.

```
mng_pars6 = list(slurry_prod_rate = 10000,
                  slurry_mass = 1000,
                  storage_depth = 2,
                  resid_depth = 0.1,
```

```

        area = 100,
        empty_int = 100,
        temp_C = 20,
        wash_water = 100000,
        wash_int = 100,
        rest_d = 0,
        resid_enrich = 1)

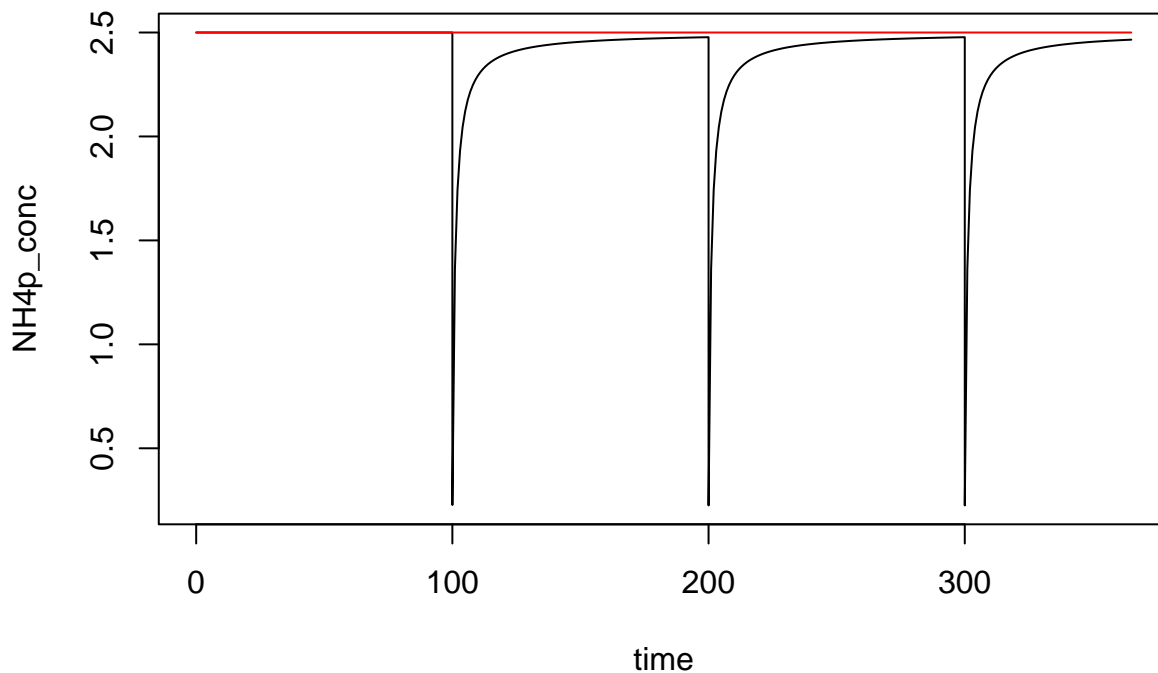
out6b <- abm(365,
  mng_pars = mng_pars6,
  man_pars = man_pars6,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars)

## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.

## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 32%

plot(NH4p_conc ~ time, data = out6b, type = 'l')
lines(NH4p_conc ~ time, data = out6a, col = 'red')

```



## 7. Speciation

Acid-base reactions are needed for inhibition and for CO<sub>2</sub> emission. They can be added for any component. The `chem_pars` argument can accept temperature-dependent log *ka* expressions. Use `temp_K` for absolute temperature in those expressions.

```

man_pars7 <- list(comps = c('H2S', 'NH4p'),
  comp_fresh = c(H2S = 0.01, NH4p = 2.5),

```

```

VFA_fresh = c(CH3COOH = 2),
pH = 7, dens = 1000)

chem_pars7 <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
VFA = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
CO2_aer = 1/0.436, CO2_sr = 1/1.2,
C_xa = 1/0.3753125),
specs = c('NH3', 'HSm', 'CH3COOm'),
mspec = c(NH3 = 'NH4p', HSm = 'H2S', CH3COOm = 'CH3COOH'),
lka = c(NH3 = '- 0.09046 - 2729.31/temp_K',
HSm = '- 3448.7/temp_K + 47.479 - 7.5227 * log(temp_K)',
CH3COOm = '- 4.8288 + 21.42/temp_K')
)

```

Here **comps** are the chemical “components”, or “master species”, as described a bit above. All are automatically included as chemical species in **packPars()**. In the **chem\_pars** argument, **specs** are the other (non-master) species that the master species are in equilibrium with. And **mspec** are the associated master species. So the NH3 species comes from NH4+. When there is speciation, the master species are always taken as the protonated ones. So the species in **specs** always have one less proton than their associated master species. Only 2 species are supported for any component (master species and one more).

```
devtools::load_all()
```

```
## i Loading ABM
```

```

out7 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars6,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars7)

```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
```

```
## Size-variable parameter problem: Missing element(s) in kss.
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
```

```
## pars$COD_conv, : COD balance is off by 1.4%
```

```
head(out7)
```

```

##   time      m0      m1      m2      sr1      VSd H2S SO4m2  NH4p
## 1    0  50.0000  50.0000  50.0000  50.000  50000.0  10   200   2500
## 2    1  554.0098  553.8532  558.3748  555.424  542318.5 110  2200  27500
## 3    2 1066.2767 1065.6731 1083.1939 1071.000 1022077.5 210  4200  52500
## 4    3 1588.3159 1586.9428 1627.0195 1597.789 1489599.4 310  6200  77500
## 5    4 2121.2032 2118.7111 2191.8591 2136.557 1945198.4 410  8200 102500
## 6    5 2665.7723 2661.7874 2779.4357 2687.920 2389180.8 510 10200 127500
##   CH3COOH slurry_mass CH4_emis_cum slurry_load COD_load CH4_emis_rate temp_C
## 1   2000.00         1000         0.0000         0         0         25.52844     20
## 2  29018.42        11000        163.6268       10000     522000        308.65442     20
## 3  67370.97        21000        628.8106       20000    1044000        626.59218     20
## 4 116610.76        31000       1425.4312       30000    1566000        970.52434     20
## 5 176325.85        41000       2577.0170       40000    2088000       1335.99603     20
## 6 246126.14        51000       4103.8015       50000    2610000       1720.63977     20
##   pH m0_eff m1_eff m2_eff sr1_eff VSd_eff H2S_eff SO4m2_eff NH4p_eff
## 1    7     0     0     0     0     0     0     0     0

```

```

## 2 7 0 0 0 0 0 0 0 0
## 3 7 0 0 0 0 0 0 0 0
## 4 7 0 0 0 0 0 0 0 0
## 5 7 0 0 0 0 0 0 0 0
## 6 7 0 0 0 0 0 0 0 0
## CH3COOH_eff slurry_mass_eff slurry_depth m0_conc m1_conc m2_conc
## 1 0 0 0.01 0.05000000 0.05000000 0.05000000
## 2 0 0 0.11 0.05036453 0.05035029 0.05076134
## 3 0 0 0.21 0.05077508 0.05074634 0.05158066
## 4 0 0 0.31 0.05123600 0.05119170 0.05248450
## 5 0 0 0.41 0.05173666 0.05167588 0.05345998
## 6 0 0 0.51 0.05227004 0.05219191 0.05449874
## sr1_conc VSd_conc H2S_conc SO4m2_conc NH4p_conc CH3COOH_conc m0_eff_conc
## 1 0.05000000 50.00000 0.01 0.2 2.5 2.000000 NaN
## 2 0.05049309 49.30168 0.01 0.2 2.5 2.638039 NaN
## 3 0.05099999 48.67036 0.01 0.2 2.5 3.208141 NaN
## 4 0.05154158 48.05159 0.01 0.2 2.5 3.761638 NaN
## 5 0.05211115 47.44386 0.01 0.2 2.5 4.300630 NaN
## 6 0.05270431 46.84668 0.01 0.2 2.5 4.826003 NaN
## m1_eff_conc m2_eff_conc sr1_eff_conc VSd_eff_conc H2S_eff_conc SO4m2_eff_conc
## 1 NaN NaN NaN NaN NaN NaN
## 2 NaN NaN NaN NaN NaN NaN
## 3 NaN NaN NaN NaN NaN NaN
## 4 NaN NaN NaN NaN NaN NaN
## 5 NaN NaN NaN NaN NaN NaN
## 6 NaN NaN NaN NaN NaN NaN
## NH4p_eff_conc CH3COOH_eff_conc
## 1 NaN NaN
## 2 NaN NaN
## 3 NaN NaN
## 4 NaN NaN
## 5 NaN NaN
## 6 NaN NaN

```

But chemical species don't matter unless they are used in inhibition or emission.

## 8. Inhibition

Any chemical species can inhibit any microbial group. Inhibition parameters (currently initial and complete concentrations, with linear response, why not?) are entered in a matrix.

```

ilwr <- matrix(
  c(5, 0.2, 10, 0.5,
    5, 0.2, 10, 0.5,
    5, 0.2, 10, 0.5,
    5, 0.2, 10, 0.5),
  nrow = 4,
  byrow = TRUE,
  dimnames = list(
    c('m0', 'm1', 'm2', 'sr1'),
    c('NH4p', 'NH3', 'CH3COOm', 'CH3COOH')
  )
)

```

```

iupr <- matrix(
  c(9, 0.9, 30, 1,
    9, 0.9, 30, 1,
    9, 0.9, 30, 1,
    9, 0.9, 30, 1),
  nrow = 4,
  byrow = TRUE,
  dimnames = list(
    c('m0', 'm1', 'm2', 'sr1'),
    c('NH4p', 'NH3', 'CH3COOm', 'CH3COOH')
  )
)

inhib_pars <- list(
  ilwr = ilwr,
  iupr = iupr
)

inhib_pars

## $ilwr
##      NH4p NH3 CH3COOm CH3COOH
## m0      5 0.2      10      0.5
## m1      5 0.2      10      0.5
## m2      5 0.2      10      0.5
## sr1     5 0.2      10      0.5
##
## $iupr
##      NH4p NH3 CH3COOm CH3COOH
## m0      9 0.9      30      1
## m1      9 0.9      30      1
## m2      9 0.9      30      1
## sr1     9 0.9      30      1

man_pars8 <- list(comps = c('H2S', 'NH4p'),
  comp_fresh = c(H2S = 0.01,
    NH4p = 2.5),
  VFA_fresh = c(CH3COOH = 2),
  pH = 7, dens = 1000)

chem_pars8 <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
  CH3COOH = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
  CO2_aer = 1/0.436, CO2_sr = 1/1.2,
  C_xa = 1/0.3753125),
  specs = c('NH3', 'HSm', 'CH3COOm'),
  mspec = c(NH3 = 'NH4p', HSm = 'H2S', CH3COOm = 'CH3COOH'),
  lka = c(NH3 = '- 0.09046 - 2729.31/temp_K',
    HSm = '- 3448.7/temp_K + 47.479 - 7.5227* log(temp_K)',
    CH3COOm = '-4.8288 + 21.42/temp_K')
)

out8 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars7,

```

```

    grp_pars = grp_pars,
    mic_pars = mic_pars,
    sub_pars = sub_pars,
    chem_pars = chem_pars7,
    inhib_pars = inhib_pars
)

```

```

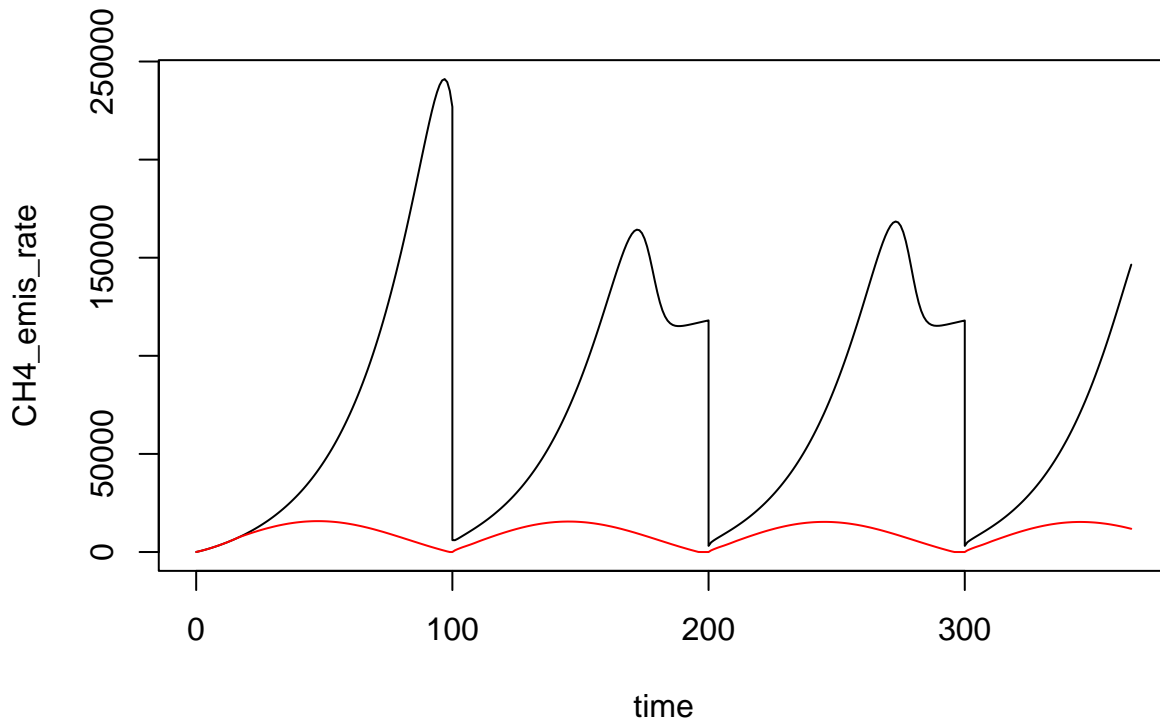
## Warning in expandParams(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.

```

```

plot(CH4_emis_rate ~ time, data = out7, type = 'l')
lines(CH4_emis_rate ~ time, data = out8, col = 'red')

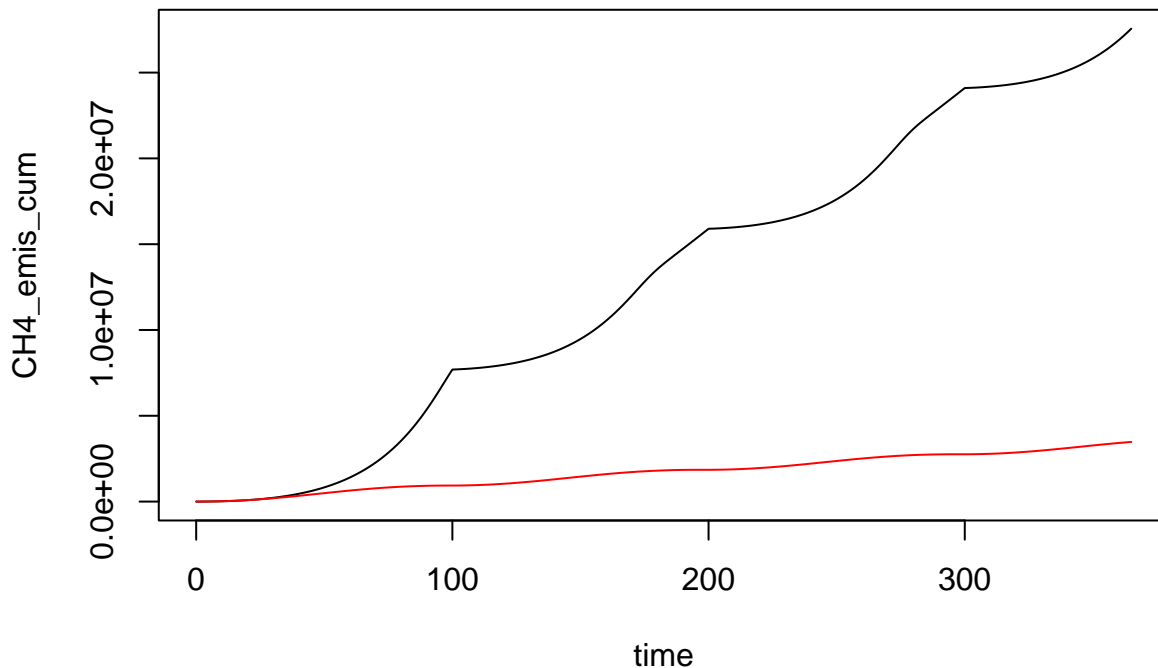
```



```

plot(CH4_emis_cum ~ time, data = out7, type = 'l')
lines(CH4_emis_cum ~ time, data = out8, col = 'red')

```



## 9. Volatilization

Any chemical species can volatilize.

```
man_pars9 <- list(comps = c('H2S', 'NH4p'),
  comp_fresh = c(H2S = 0.01, NH4p = 2.5),
  VFA_fresh = c(CH3COOH = 2),
  pH = 7, dens = 1000)
```

We need to set mass transfer coefficient values (m/d) for any species that volatilizes. These are overall (possibly two-film) values for in liquid-phase units.

```
chem_pars9 <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
  VFA = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
  CO2_aer = 1/0.436, CO2_sr = 1/1.2,
  C_xa = 1/0.3753125),
  specs = c('NH3', 'HSm', 'CH3COOm'),
  mspec = c(NH3 = 'NH4p', HSm = 'H2S', CH3COOm = 'CH3COOH'),
  lka = c(NH3 = '- 0.09046 - 2729.31/temp_K',
    HSm = '- 3448.7/temp_K + 47.479 - 7.5227 * log(temp_K)',
    CH3COOm = '-4.8288 + 21.42/temp_K'),
  kl = c(NH3 = 0.01, H2S = 0.01, CH3COOH = 0.01) * 86400)
```

```
devtools::load_all()
```

```
## i Loading ABM
```

```
out9a <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars9,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars9,
```



```

    inhib_pars = inhib_pars
)

## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.

## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 2%

```

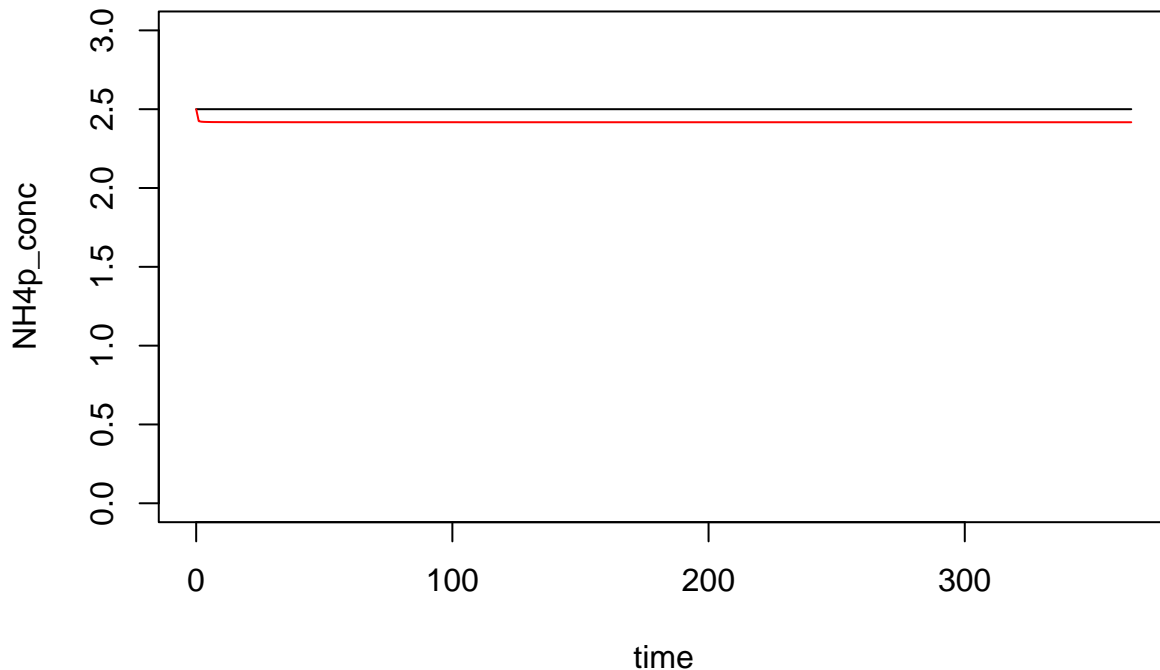
The state variable vector and output data frame automatically expand for the new values.

```
head(out9a[, grep('_emis', names(out9a))])
```

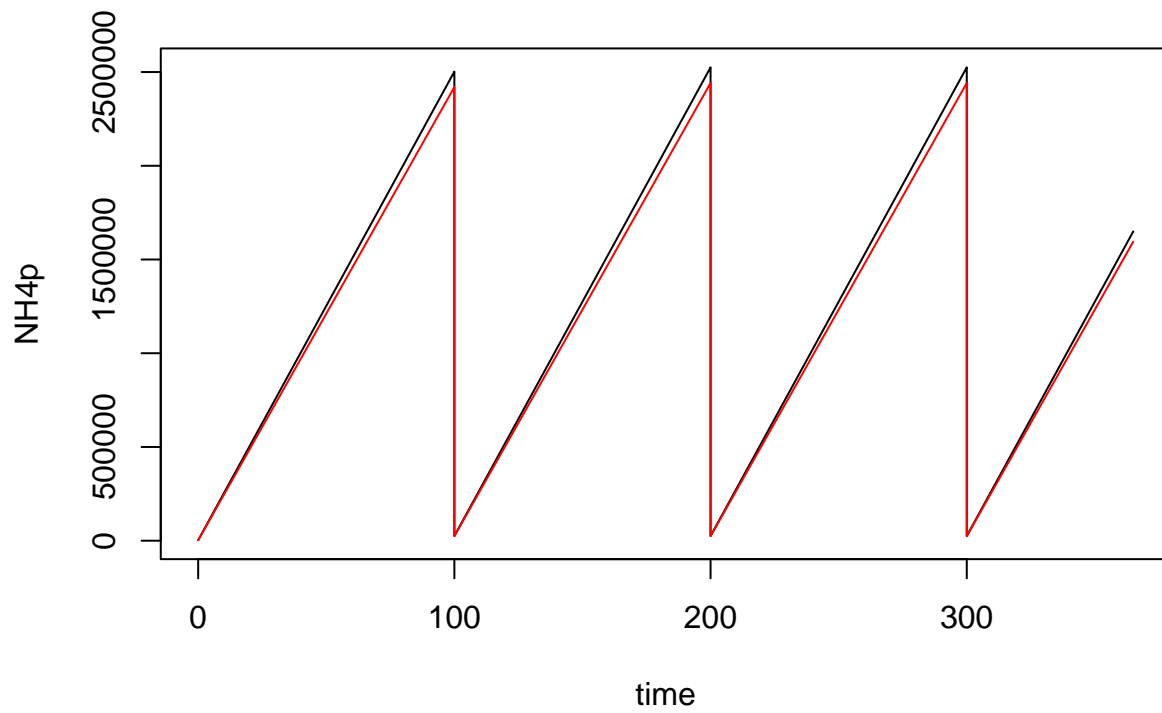
```
##   CH4_emis_cum NH3_emis_cum H2S_emis_cum CH3COOH_emis_cum CH4_emis_rate
## 1      0.0000      0.000      0.0000      0.000      25.52844
## 2     161.8334     833.273     89.6908     1106.072     305.28189
## 3     622.3551    1661.702    171.2283     2484.484     620.73997
## 4    1412.0784    2489.451    252.7655     4130.203     962.63333
## 5    2554.8558    3316.914    334.3028     6036.464    1326.29785
## 6    4071.0922    4144.219    415.8401     8196.713    1709.25431

```

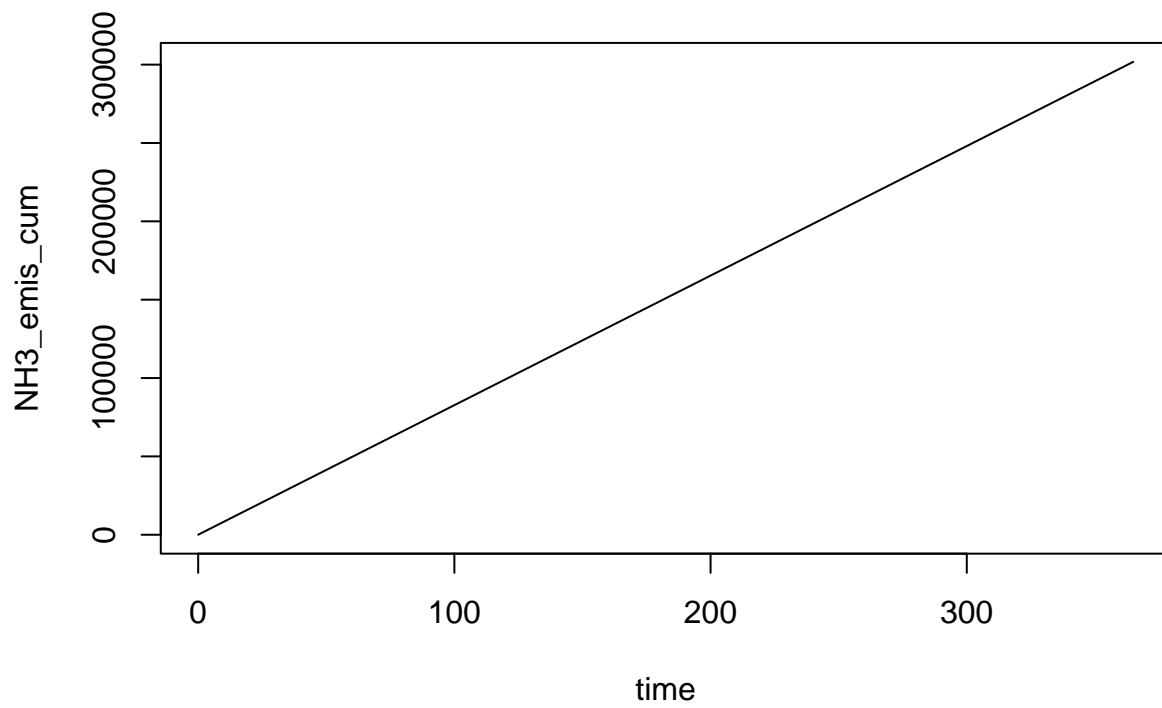
```
plot(NH4p_conc ~ time, data = out8, type = 'l', ylim = c(0, 3))
lines(NH4p_conc ~ time, data = out9a, col = 'red')
```



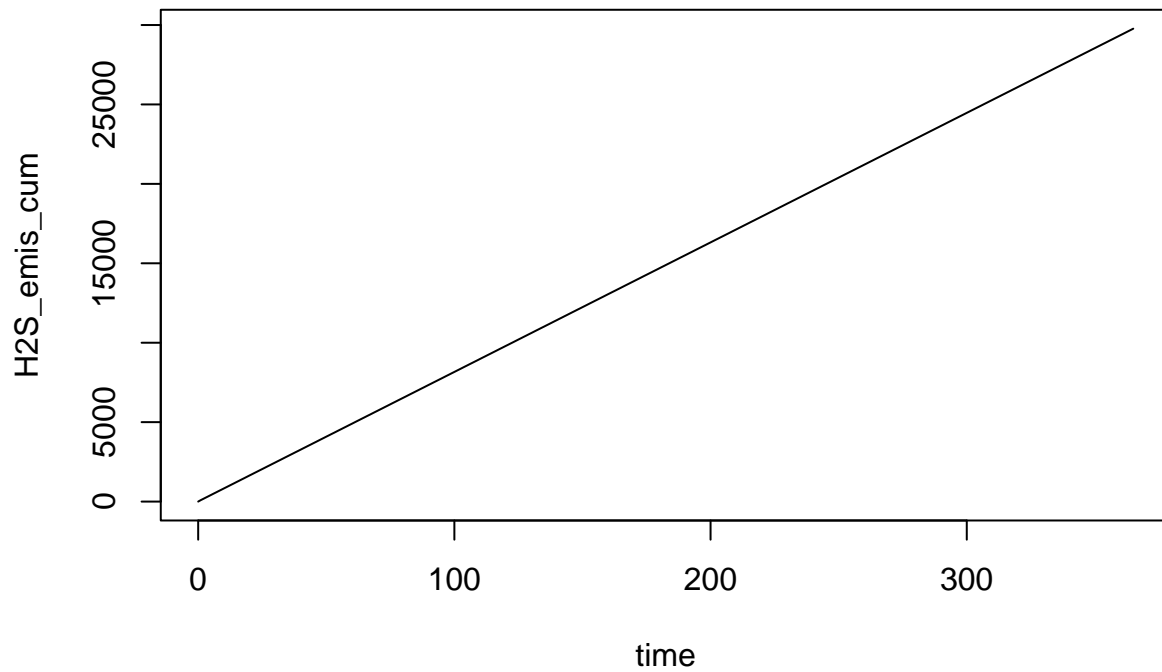
```
plot(NH4p ~ time, data = out8, type = 'l')
lines(NH4p ~ time, data = out9a, col = 'red')
```



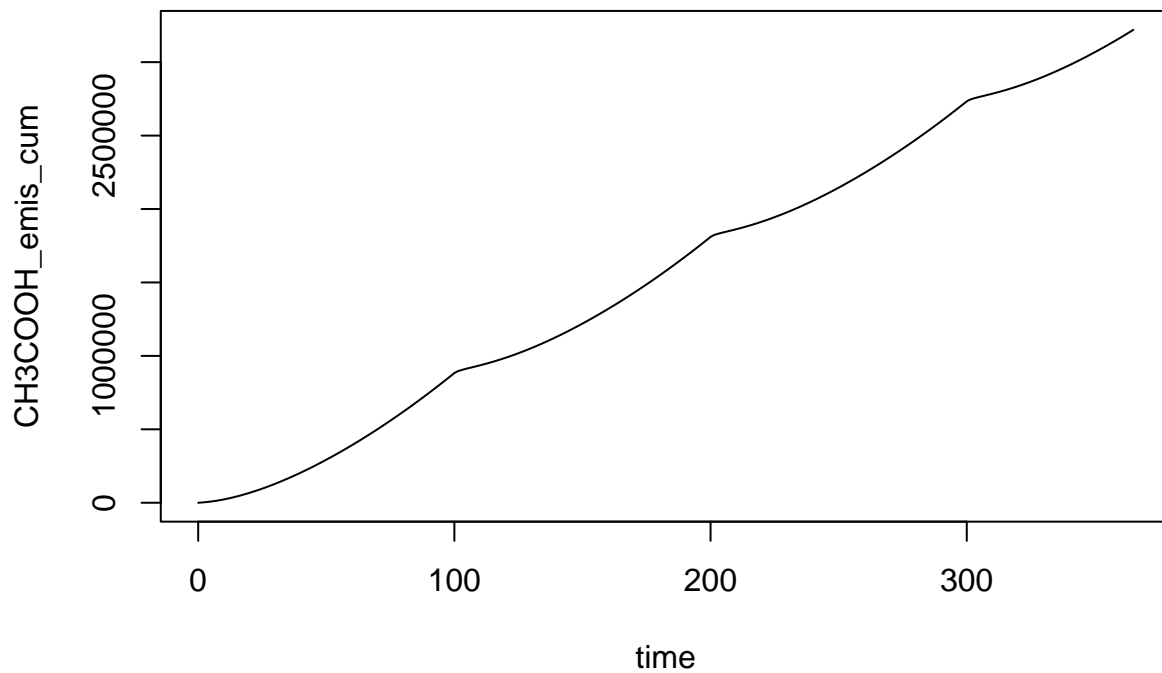
```
plot(NH3_emis_cum ~ time, data = out9a, type = 'l')
```



```
plot(H2S_emis_cum ~ time, data = out9a, type = 'l')
```



```
plot(CH3COOH_emis_cum ~ time, data = out9a, type = 'l')
```



Let's use a fixed slurry mass to exaggerate emission.

```
mng_pars9b = list(slurry_prod_rate = 0,
                  slurry_mass = 1E6,
                  storage_depth = 2,
                  resid_depth = 0.1,
                  area = 100,
                  empty_int = 100,
                  temp_C = 20,
                  wash_water = 0,
```

```

wash_int = NA,
rest_d = 0,
resid_enrich = 1)

chem_pars9b <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
                                VFA = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
                                CO2_aer = 1/0.436, CO2_sr = 1/1.2,
                                C_xa = 1/0.3753125),
  specs = c('NH3', 'HSm', 'CH3COOm'),
  mspec = c(NH3 = 'NH4p', HSm = 'H2S', CH3COOm = 'CH3COOH'),
  lka = c(NH3 = '- 0.09046 - 2729.31/temp_K',
          HSm = '- 3448.7/temp_K + 47.479 - 7.5227 * log(temp_K)',
          CH3COOm = '-4.8288 + 21.42/temp_K'),
  kl = c(NH3 = 0.01, H2S = 0.01) * 86400)

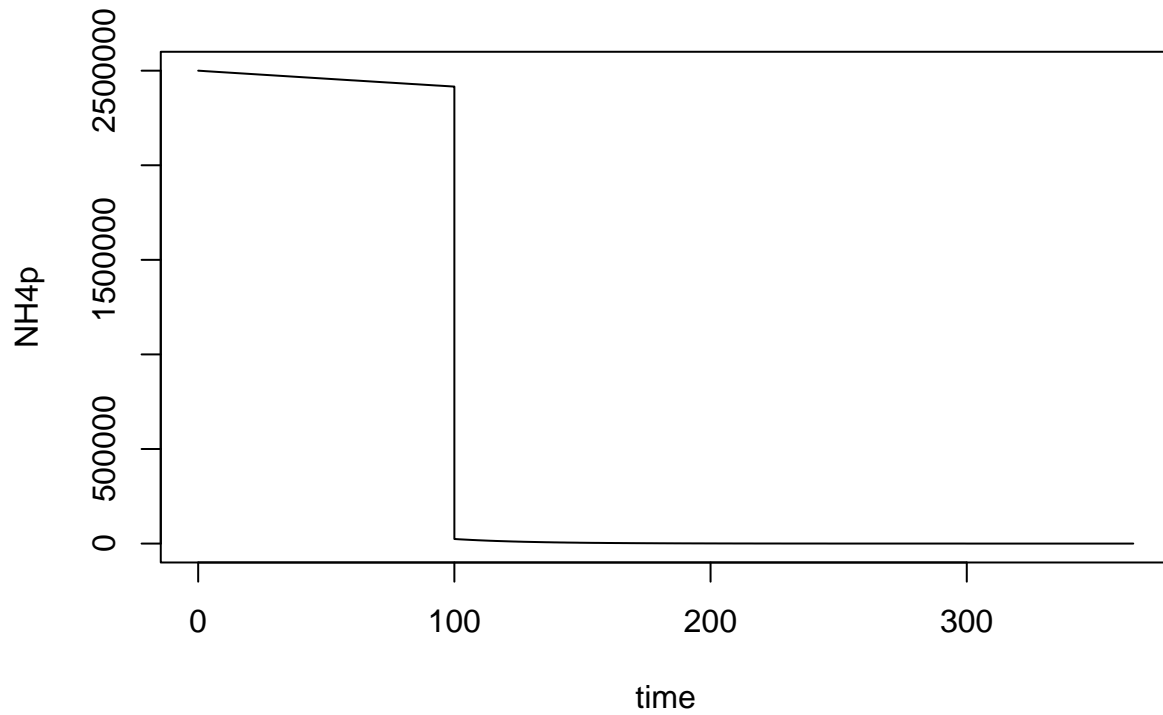
devtools::load_all()

## i Loading ABM
out9b <- abm(365,
  mng_pars = mng_pars9b,
  man_pars = man_pars9,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars9b,
  inhib_pars = inhib_pars
)

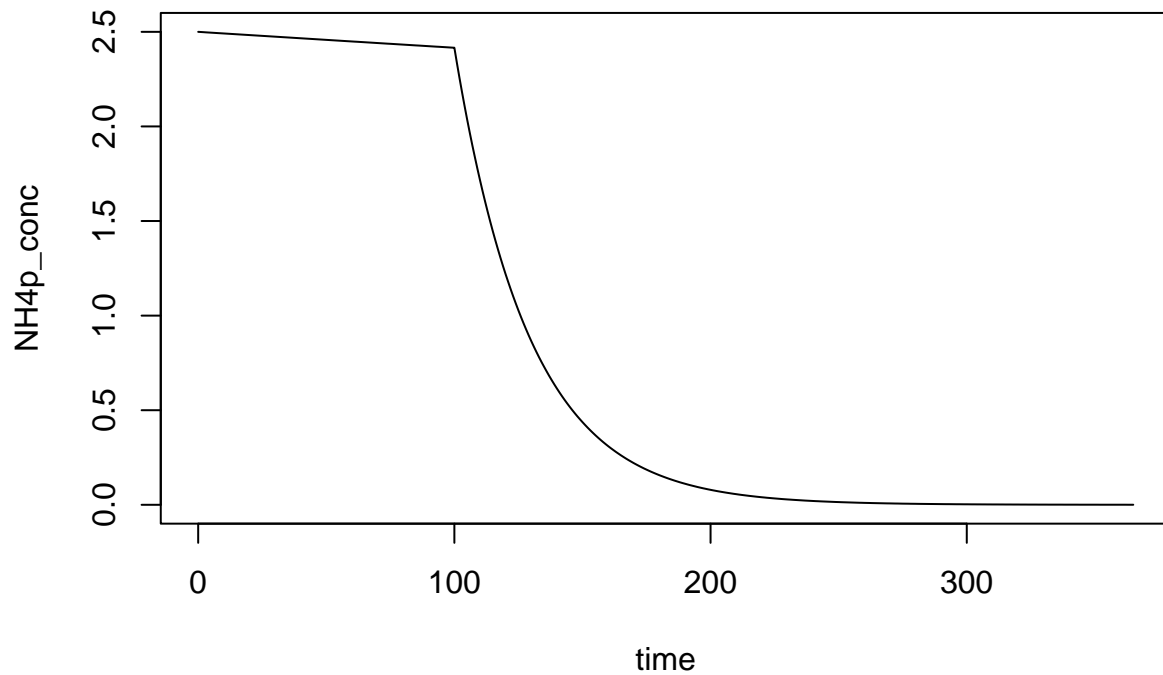
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.

## Warning in emptyStore(y, resid_mass = pars$resid_mass, resid_enrich =
## pars$resid_enrich): Emptying skipped.
## Warning in emptyStore(y, resid_mass = pars$resid_mass, resid_enrich =
## pars$resid_enrich): Emptying skipped.
plot(NH4p ~ time, data = out9b, type = 'l')

```



```
plot(NH4p_conc ~ time, data = out9b, type = 'l')
```



## 9. COD balance

There is now a `checkCOD()` function that runs on `abm()` results before returning them. For now the tolerance is fixed at 1%. Some of the examples above do not meet that criterion for some reason. At least one shows a real problem that needs to be identified. For the emission example above, the problem is that VFA is emitted but that loss is not included in the balance check (this might have been fixed). I need to decide about how to pass that COD information. We can make it worse by pretending the the charged form can volatilize (VFA

changed to CH3COOm below).

```
chem_pars10 <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
                                VFA = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
                                CO2_aer = 1/0.436, CO2_sr = 1/1.2,
                                C_xa = 1/0.3753125),
  specs = c('NH3', 'HSm', 'CH3COOm'),
  mspec = c(NH3 = 'NH4p', HSm = 'H2S', CH3COOm = 'CH3COOH'),
  lka = c(NH3 = '- 0.09046 - 2729.31/temp_K',
          HSm = '- 3448.7/temp_K + 47.479 - 7.5227 * log(temp_K)',
          CH3COOm = '-4.8288 + 21.42/temp_K'),
  kl = c(NH3 = 0.01, H2S = 0.01, CH3COOm = 0.01) * 86400)

out10 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars9,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars10,
  inhib_pars = inhib_pars
)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.

## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 36%
```

## 10. Stoichiometry and nitrogen mineralization

Now substrates can produce any amount of arbitrary components (defined in `man_pars`, possibly volatilized, possibly involved in speciation in inhibition) through hydrolysis and fermentation to VFA.

```
man_pars10 <- list(comps = c('H2S', 'S04m2', 'NH4p'),
  comp_fresh = c(H2S = 0.01, S04m2 = 0.2, NH4p = 2.5),
  VFA_fresh = c(CH3COOH = 2),
  pH = 7, dens = 1000)
```

(Hmm, should `comps` be moved to `chem_pars`?)

Here we'll have 4 substrates. But substrates need not actually produce VFA anymore.

```
sub_pars10 <- list(subs = c('cellulose', 'protein', 'lipids', 'urea'),
  T_opt_hyd = c(all = 60),
  T_min_hyd = c(all = 0),
  T_max_hyd = c(all = 90),
  hydrol_opt = c(lipids = 0.1, protein = 0.01, cellulose = 0.05, urea = 1),
  sub_fresh = c(lipids = 3, protein = 20, cellulose = 35, urea = 10),
  sub_init = c(lipids = 3, protein = 20, cellulose = 35, urea = 10))
```

Production of any component is set in the `stoich` element of the `chem_pars` argument.

```
smat <- matrix(c(0, 0.2, 0, 0.2,
                 0, 0.01, 0, 0,
                 1, 1, 1, 0),
  nrow = 3,
```

```

byrow = TRUE,
dimnames = list(
  c('NH4p', 'H2S', 'CH3COOH'),
  c('cellulose', 'protein', 'lipids', 'urea'))))

```

smat

```

##           cellulose protein lipids urea
## NH4p           0     0.20      0  0.2
## H2S             0     0.01      0  0.0
## CH3COOH         1     1.00      1  0.0

```

Substrate and other component quantities are

1. COD mass, or if COD = 0,
2. N mass, or if N = 0,
3. C mass, or if C = 0,
4. S mass, or if S = 0,
5. total mass

So the CH3COOH row should only have 1 or 0.

The stoich matrix can be calculated from substrate chemical formulas—see next example.

```

chem_pars10 <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
                                VFA = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
                                CO2_aer = 1/0.436, CO2_sr = 1/1.2,
                                C_xa = 1/0.3753125),
  specs = c('NH3', 'HSm', 'CH3COOm'),
  mspec = c(NH3 = 'NH4p', HSm = 'H2S', CH3COOm = 'CH3COOH'),
  stoich = smat)

```

```
devtools::load_all()
```

```
## i Loading ABM
```

```

out10 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars10,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars10,
  chem_pars = chem_pars10)

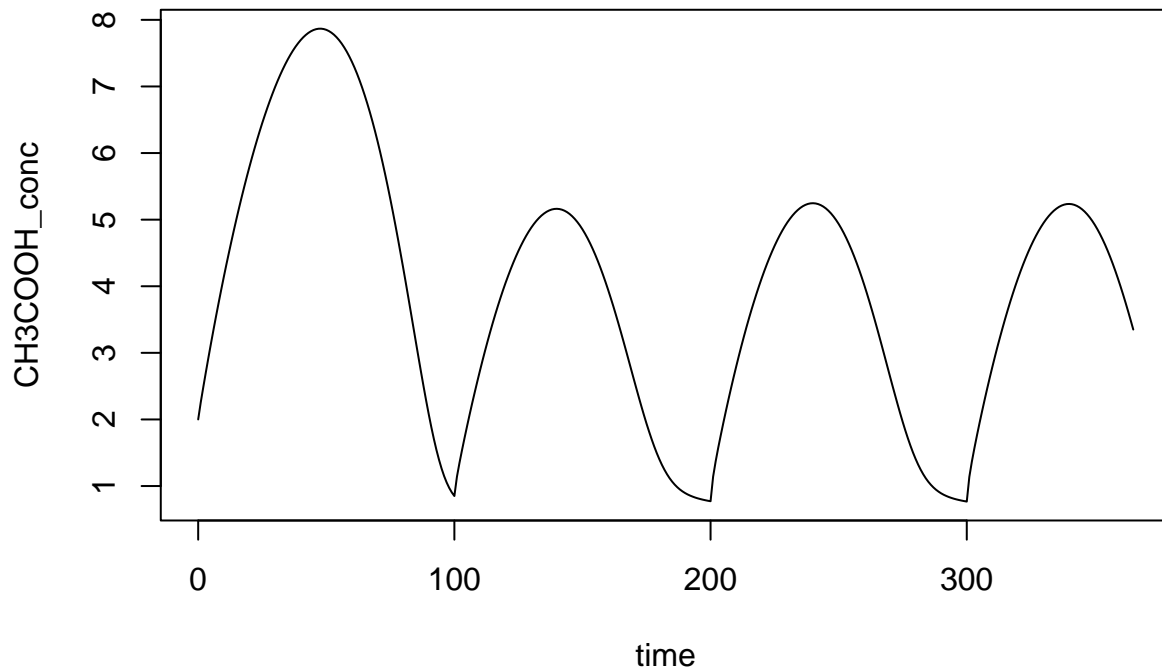
```

```

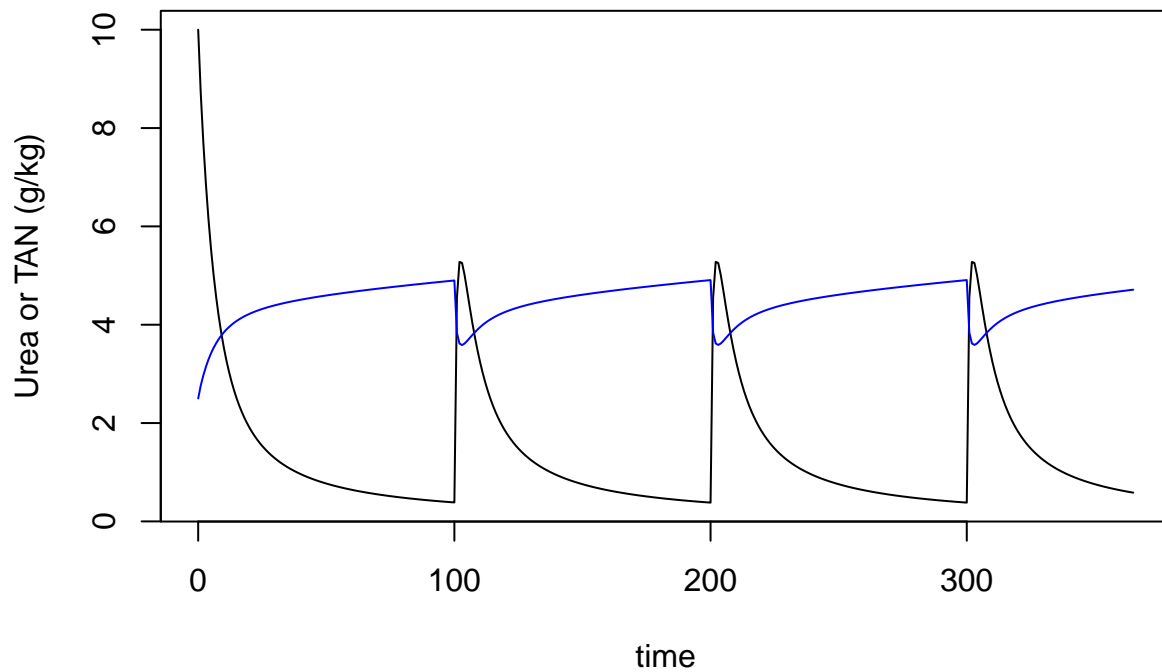
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.

```

```
plot(CH3COOH_conc ~ time, data = out10, type = 'l')
```



```
plot(urea_conc ~ time, data = out10, type = 'l', ylab = 'Urea or TAN (g/kg)')
lines(NH4p_conc ~ time, data = out10, col = 'blue')
```



It might make more sense to have `stoich` calculated internally. For this, substrate formulas must be provided.

```
sub_pars10b <- list(subs = c('cellulose', 'protein', 'lipids', 'urea'),
  forms = c(cellulose = 'C6H10O5', protein = 'C4 H6.1 O1.2 N',
    lipids = 'C57 H104 O6', urea = 'CO(NH2)2'),
  T_opt_hyd = c(all = 60),
  T_min_hyd = c(all = 0),
  T_max_hyd = c(all = 90),
  hydrol_opt = c(lipids = 0.1, protein = 0.01, cellulose = 0.05, urea = 1),
```



```
sub_fresh = c(lipids = 3, protein = 20, cellulose = 35, urea = 10),
sub_init = c(lipids = 3, protein = 20, cellulose = 35, urea = 10))
```

Internally, the `getStoich()` function is used, which in turn calls `predFerm()`. Among these substrates, cellulose produces no CO<sub>2</sub> from fermentation, protein and lipids *consume* CO<sub>2</sub> (they are highly reduced), and urea is a special case with no COD. So it produces no VFAs.

```
predFerm(sub_pars10b$forms[1])
```

```
##      H2O CH3COOH
##      -1      3
```

```
predFerm(sub_pars10b$forms[2])
```

```
##      H2O      CO2 CH3COOH      NH3
## -2.6250 -0.1750  2.0875  1.0000
```

```
predFerm(sub_pars10b$forms[3])
```

```
##      H2O      CO2 CH3COOH
##      -28      -23      40
```

```
predFerm(sub_pars10b$forms[4])
```

```
##      CO2      NH3      H.      H2O CH3COOH      H2
##       1       2       0      -1       0       0
```

Internally, the stoichiometric coefficients are changed to the same units given above (COD, N, C, S, or total mass). Now we must have CO<sub>2</sub> as a component, because it will be produced.

```
man_pars10b <- list(comps = c('H2S', 'SO4m2', 'NH4p', 'CO2'),
                    comp_fresh = c(H2S = 0.01, SO4m2 = 0.2, NH4p = 2.5, CO2 = 1),
                    VFA_fresh = c(CH3COOH = 2),
                    pH = 7, dens = 1000)
```

We don't actually need the species included above. But because of a bit of a programming quirk, we need to provide the master species for NH<sub>3</sub>.

```
chem_pars10b <- list(COD_conv = c(CH4 = 1/0.2507),
                    specs = c('NH3'),
                    mspec = c(NH3 = 'NH4p'),
                    stoich = 'calc')
```

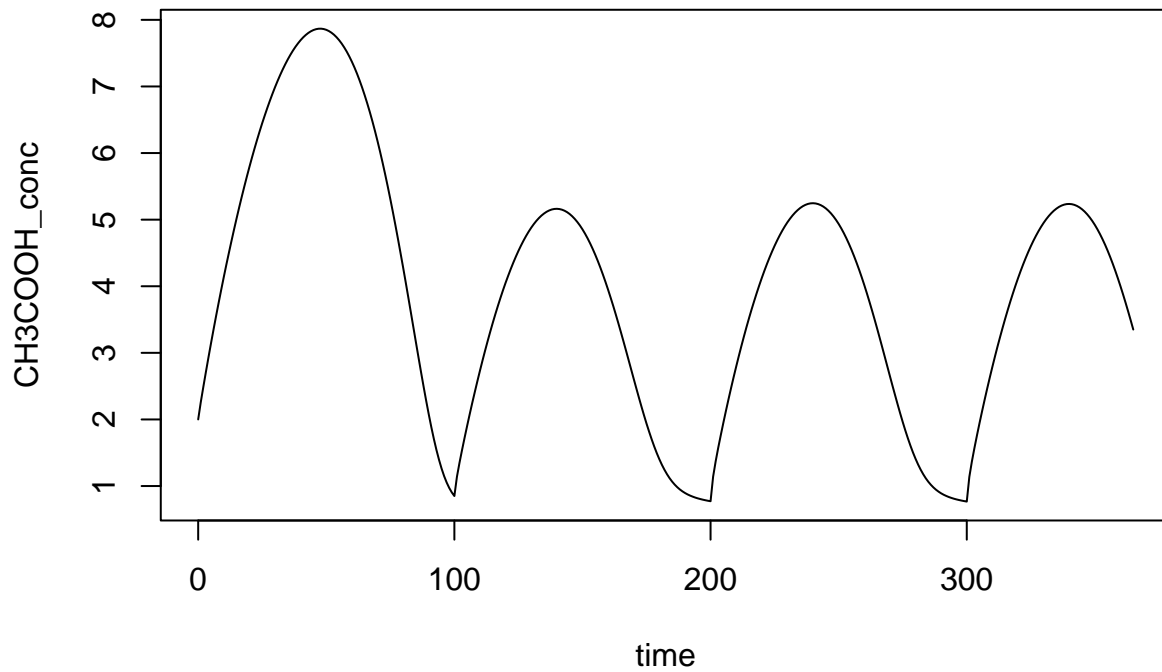
```
devtools::load_all()
```

```
## i Loading ABM
```

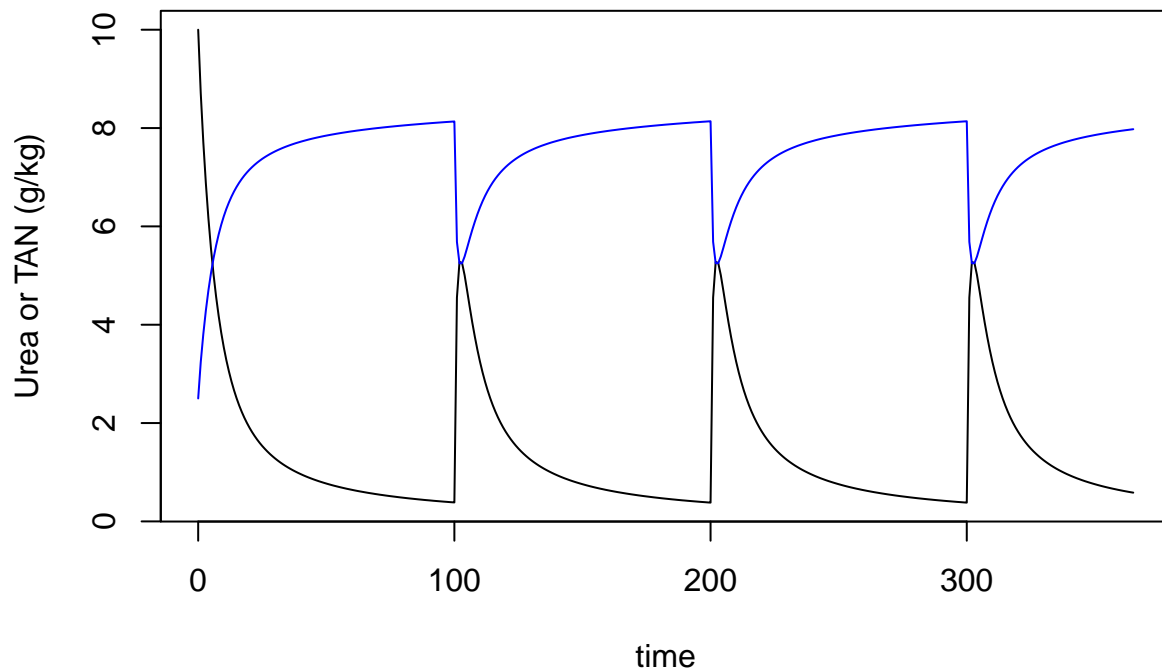
```
out10b <- abm(365,
              mng_pars = mng_pars,
              man_pars = man_pars10b,
              grp_pars = grp_pars,
              mic_pars = mic_pars,
              sub_pars = sub_pars10b,
              chem_pars = chem_pars10b)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

```
plot(CH3COOH_conc ~ time, data = out10b, type = 'l')
```

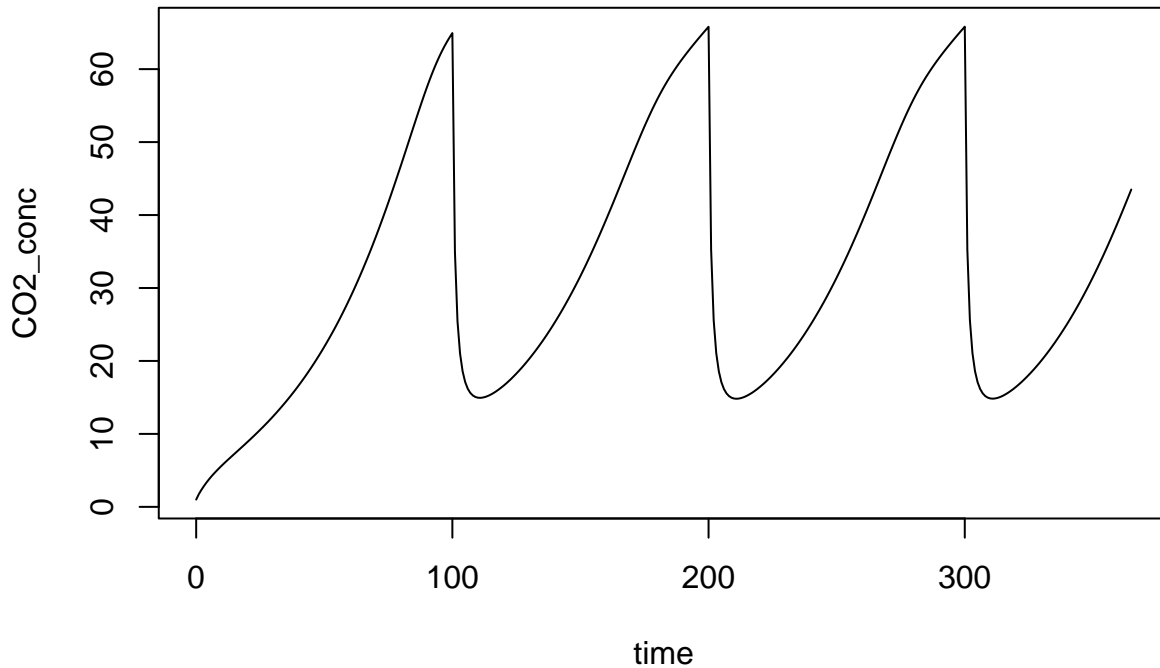


```
plot(urea_conc ~ time, data = out10b, type = 'l', ylab = 'Urea or TAN (g/kg)')
lines(NH4p_conc ~ time, data = out10b, col = 'blue')
```



Now we also have dissolved  $\text{CO}_2$  (really TIC) concentration. Only there is no emission, so it does not mean much.

```
plot(CO2_conc ~ time, data = out10b, type = 'l')
```



## 11. CO2 emission

CO2 emission can be included through the volatilization route. It can be produced from both fermentation and methanogenesis. Speciation of dissolved CO2 (really H<sub>2</sub>CO<sub>3</sub><sup>\*</sup>) and HCO<sub>3</sub><sup>-</sup> needs to be included (although a reduced mass transfer coefficient could achieve the same effect). So far, CO2 emission behavior is troublesome—it is difficult to produce plausible results.

```
sub_pars11 <- list(subs = c('VSd'),
  forms = c(VSd = 'C23H37O14N (CO2)1.5'),
  T_opt_hyd = c(all = 60),
  T_min_hyd = c(all = 0),
  T_max_hyd = c(all = 90),
  hydrol_opt = c(all = 0.1),
  sub_fresh = c(VSd = 20),
  sub_init = c(VSd = 20))
```

```
man_pars11 <- list(comps = c('CO2', 'NH4p'),
  comp_fresh = c(CO2 = 1, NH4p = 2.5),
  VFA_fresh = c(CH3COOH = 2),
  pH = 7, dens = 1000)
```

Here we will define speciation only for TIC. There is no way to include CO<sub>3</sub><sup>2-</sup> as a third species.

```
chem_pars11 <- list(COD_conv = c(CH4 = 1/0.2507),
  specs = c('HCO3m'),
  mspec = c(HCO3m = 'CO2', NH3 = 'NH4p'),
  lka = c(HCO3m = '-2.778 + -353.5305 -0.06092*temp_K + 21834.37/temp_K + 126.8339*1000/temp_K'),
  stoich = 'calc',
  kl = c(CO2 = 0.1) * 86400)
```

```
devtools::load_all()
```

```
## i Loading ABM
```

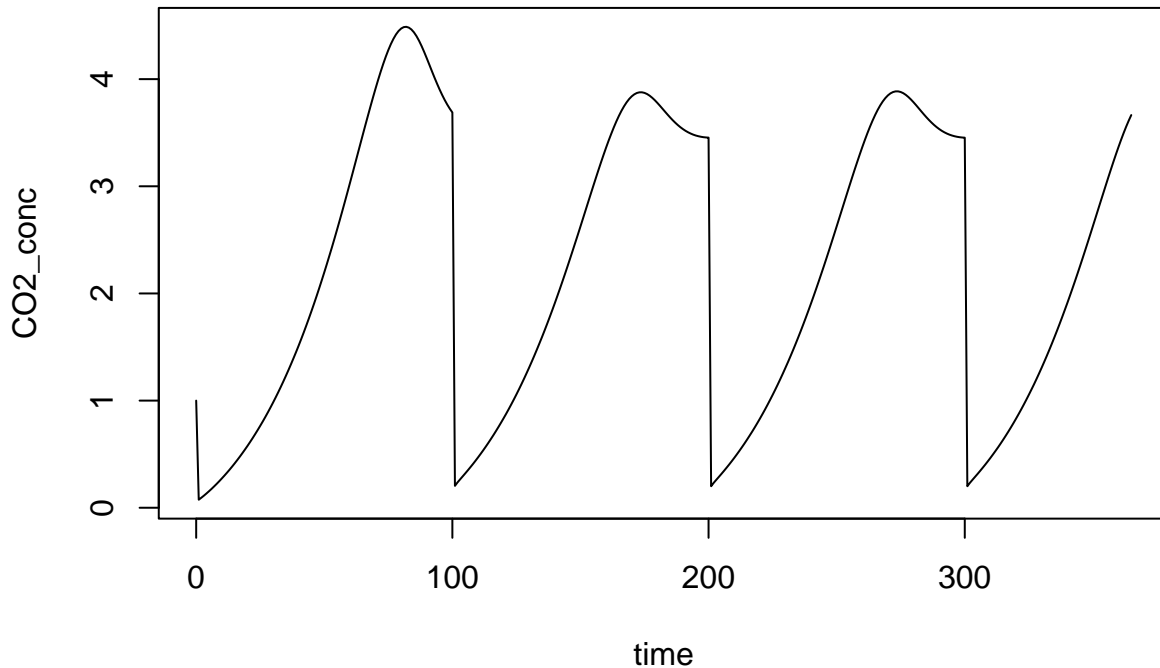
```
out11 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars11,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars11,
  chem_pars = chem_pars11)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.

## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 1.8%
```

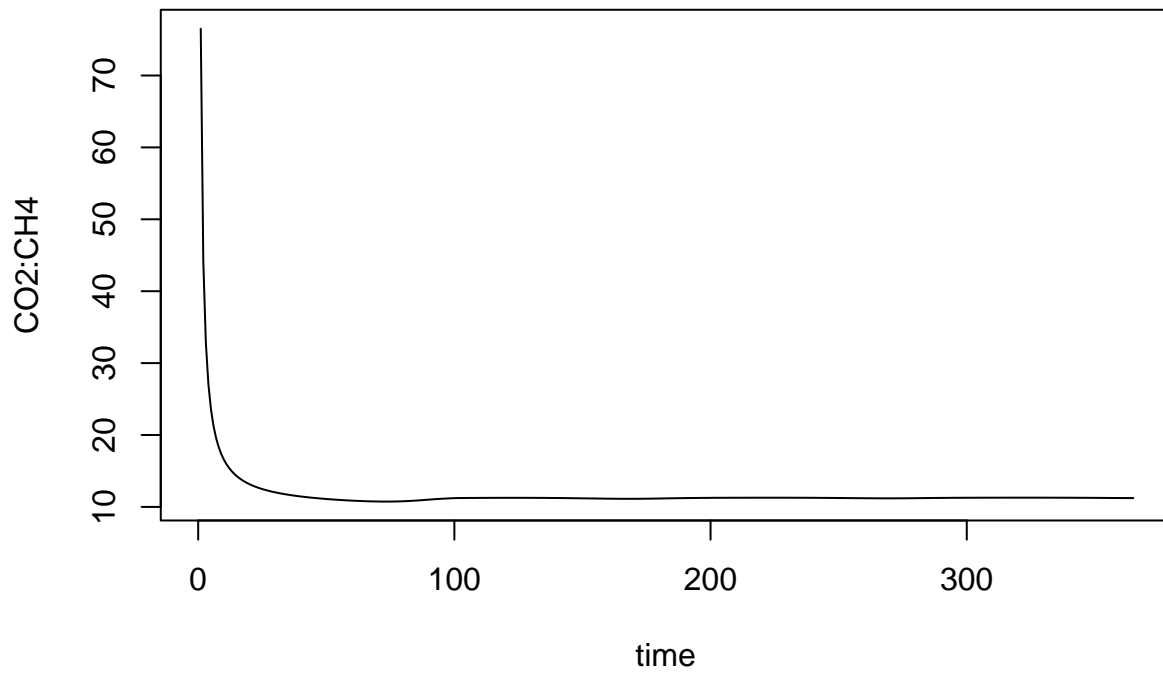
TIC in slurry shows a lot of change over time, implausible, really. Emission reduces the concentration to nearly zero when the level is low.

```
plot(CO2_conc ~ time, data = out11, type = 'l')
```



But now we can predict a CO2:CH4 ratio. This is a cumulative value.

```
plot(CO2_emis_cum / CH4_emis_cum ~ time, data = out11, type = 'l', ylab = 'CO2:CH4')
```



```
plot(CO2_emis_cum / CH4_emis_cum ~ time, data = out11, type = 'l', ylab = 'CO2:CH4', ylim = c(0, 20))
```

