

Demo of new simple1 version

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Overview

This demo shows:

1. basic usage,
2. variable substrates,
3. time-variable inputs,
4. speciation (proton-transfer reactions),
5. inhibition,
6. volatilization, and
7. COD balance,

Prep

```
devtools::load_all()
```

```
## i Loading ABM
```

1. Basic behavior

The simplest usage is with constant slurry production rate and a fixed schedule. We need to set some parameters, first management parameters.

```
mng_pars <- list(slurry_prod_rate = 10000,  
  slurry_mass = 1000,  
  storage_depth = 2,  
  resid_depth = 0.1,  
  area = 100,  
  empty_int = 100,  
  temp_C = 20,  
  wash_water = 0,  
  wash_int = NA,  
  rest_d = 0,  
  resid_enrich = 1)
```

Next substrate parameters, a new argument. This defines substrates. We could have any number with any names. Note that hydrolysis uses CTM again (like anything here, that could be changed).

```
sub_pars <- list(subs = c('VSd'),  
  T_opt_hyd = c(VSd = 60),  
  T_min_hyd = c(VSd = 0),  
  T_max_hyd = c(VSd = 90),
```

```

hydrol_opt = c(VSd = 0.1),
sub_fresh = c(VSd = 50),
sub_init = c(VSd = 50))

```

Microbial parameters are similar to other ABM versions, but inhibition is set separately now (and not shown in this simple example).

```

grp_pars <- list(grps = c('m0', 'm1', 'm2', 'sr1'),
  yield = c(default = 0.05, sr1 = 0.065),
  xa_fresh = c(all = 0.05),
  xa_init = c(all = 0.05),
  dd_rate = c(all = 0.02),
  ksv = c(default = 1, sr1 = 0.5),
  kss = c(sr1 = 0.5),
  qhat_opt = c(m0 = 1, m1 = 1, m2 = 2, sr1 = 9),
  T_opt = c(m0 = 18, m1 = 18, m2 = 28, sr1 = 44),
  T_min = c(m0 = 0, m1 = 6.41, m2 = 6.41, sr1 = 0),
  T_max = c(m0 = 25, m1 = 25, m2 = 38, sr1 = 51))

```

The `dd_rate_xa` parameter is for “death and decay”.

```
mic_pars <- list(dd_rate_xa = 0.02)
```

These last two arguments are similar to other versions. VFA is hard-wired and so has its own elements. The name should be CH₃COOH.

```

man_pars <- list(VFA_fresh = c(CH3COOH = 2), pH = 7, dens = 1000)
chem_pars <- list(COD_conv = c(CH4 = 1/0.2507))

```

```
devtools::load_all()
```

```
## i Loading ABM
```

```

out1 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars)

```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
```

```
## Size-variable parameter problem: Missing element(s) in kss.
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
```

```
## pars$COD_conv, : COD balance is off by 1.7%
```

Output is similar to other versions. (The `value` argument does not currently work.)

```
head(out1)
```

```

##   time      m0      m1      m2      sr1      VSd  CH3COOH slurry_mass
## 1    0  50.0000  50.0000  50.0000  50.0000  50000.0   2000.00       1000
## 2    1  554.0098  553.8533  558.3748  544.0431  542318.4  29018.66      11000
## 3    2 1066.2767 1065.6732 1083.1940 1028.3035 1022076.9  67371.44      21000
## 4    3 1588.3161 1586.9430 1627.0197 1502.9749 1489597.5 116611.44      31000
## 5    4 2121.2034 2118.7114 2191.8594 1968.2472 1945194.0 176326.70      41000
## 6    5 2665.7726 2661.7877 2779.4361 2424.3064 2389172.2 246127.08      51000
##   CH4_emis_cum slurry_load COD_load CH4_emis_rate temp_C pH m0_eff m1_eff

```

```

## 1      0.0000      0      0      25.52844      20 7      0      0
## 2     163.6272     10000    522000     308.65512     20 7      0      0
## 3     628.8119     20000   1044000     626.59326     20 7      0      0
## 4    1425.4337     30000   1566000     970.52564     20 7      0      0
## 5    2577.0208     40000   2088000    1335.99741     20 7      0      0
## 6    4103.8067     50000   2610000    1720.64113     20 7      0      0
##   m2_eff sr1_eff VSd_eff CH3COOH_eff slurry_mass_eff slurry_depth   m0_conc
## 1      0      0      0      0      0      0.01 0.05000000
## 2      0      0      0      0      0      0.11 0.05036453
## 3      0      0      0      0      0      0.21 0.05077508
## 4      0      0      0      0      0      0.31 0.05123600
## 5      0      0      0      0      0      0.41 0.05173667
## 6      0      0      0      0      0      0.51 0.05227005
##   m1_conc   m2_conc   sr1_conc VSd_conc CH3COOH_conc m0_eff_conc
## 1 0.05000000 0.05000000 0.05000000 50.00000      2.000000      NaN
## 2 0.05035030 0.05076135 0.04945846 49.30167      2.638060      NaN
## 3 0.05074634 0.05158067 0.04896683 48.67033      3.208164      NaN
## 4 0.05119171 0.05248451 0.04848306 48.05153      3.761660      NaN
## 5 0.05167589 0.05345999 0.04800603 47.44376      4.300651      NaN
## 6 0.05219192 0.05449875 0.04753542 46.84651      4.826021      NaN
##   m1_eff_conc m2_eff_conc sr1_eff_conc VSd_eff_conc CH3COOH_eff_conc
## 1      NaN      NaN      NaN      NaN      NaN
## 2      NaN      NaN      NaN      NaN      NaN
## 3      NaN      NaN      NaN      NaN      NaN
## 4      NaN      NaN      NaN      NaN      NaN
## 5      NaN      NaN      NaN      NaN      NaN
## 6      NaN      NaN      NaN      NaN      NaN

```

```
tail(out1)
```

```

##   time      m0      m1      m2      sr1      VSd CH3COOH slurry_mass
## 364 360 74909.98 72497.35 337403.6 17643.06 15468816 4796289      610000
## 365 361 77056.84 74543.26 351549.2 17788.74 15576604 4684393      620000
## 366 362 79234.75 76617.42 366160.3 17931.53 15682002 4557322      630000
## 367 363 81441.36 78717.58 381233.0 18071.50 15785084 4414981      640000
## 368 364 83673.78 80840.93 396758.5 18208.69 15885918 4257395      650000
## 369 365 85928.37 82984.01 412722.1 18343.17 15984571 4084743      660000
##   CH4_emis_cum slurry_load COD_load CH4_emis_rate temp_C pH   m0_eff
## 364 26846605      3600000 187920000      125134.1    20 7 441740.7
## 365 26973866      3610000 188442000      129394.2    20 7 441740.7
## 366 27105400      3620000 188964000      133674.8    20 7 441740.7
## 367 27241214      3630000 189486000      137950.5    20 7 441740.7
## 368 27381289      3640000 190008000      142189.3    20 7 441740.7
## 369 27525568      3650000 190530000      146351.3    20 7 441740.7
##   m1_eff m2_eff sr1_eff VSd_eff CH3COOH_eff slurry_mass_eff slurry_depth
## 364 422210.1 2239891 63286.93 53531054      3419880      2991000      6.1
## 365 422210.1 2239891 63286.93 53531054      3419880      2991000      6.2
## 366 422210.1 2239891 63286.93 53531054      3419880      2991000      6.3
## 367 422210.1 2239891 63286.93 53531054      3419880      2991000      6.4
## 368 422210.1 2239891 63286.93 53531054      3419880      2991000      6.5
## 369 422210.1 2239891 63286.93 53531054      3419880      2991000      6.6
##   m0_conc   m1_conc   m2_conc   sr1_conc VSd_conc CH3COOH_conc m0_eff_conc
## 364 0.1228032 0.1188481 0.5531206 0.02892305 25.35872      7.862769      0.14769
## 365 0.1242852 0.1202311 0.5670148 0.02869152 25.12355      7.555472      0.14769
## 366 0.1257694 0.1216150 0.5812069 0.02846275 24.89207      7.233845      0.14769

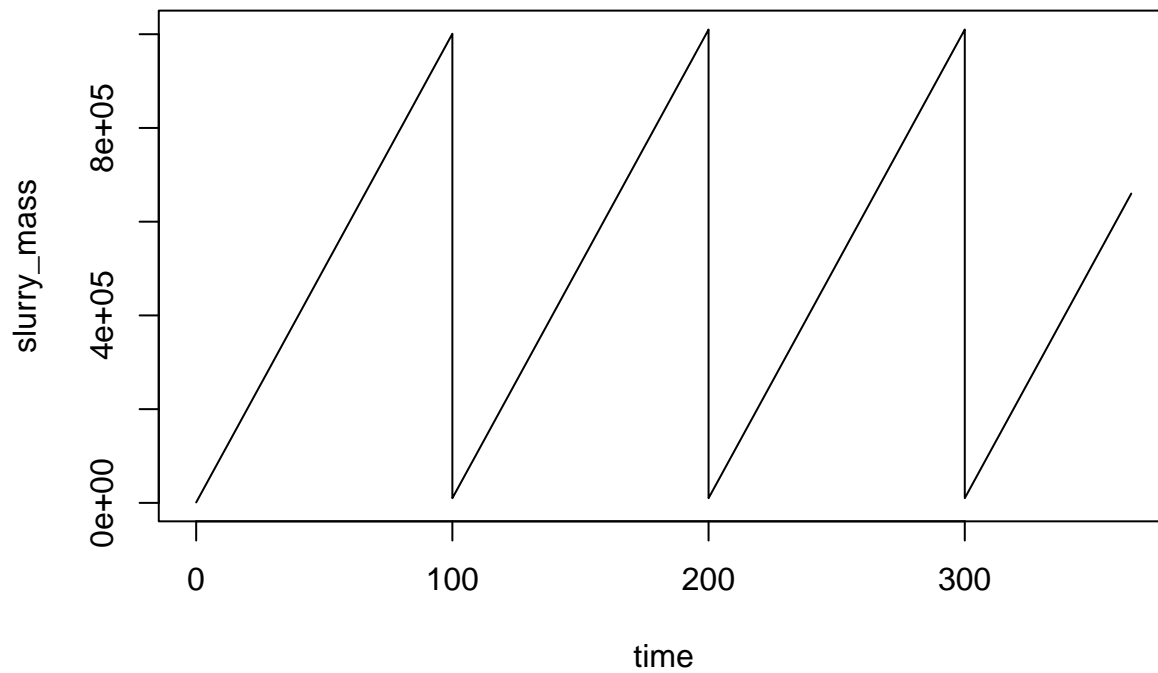
```

```
## 367 0.1272521 0.1229962 0.5956766 0.02823671 24.66419      6.898408      0.14769
## 368 0.1287289 0.1243707 0.6103976 0.02801337 24.43987      6.549838      0.14769
## 369 0.1301945 0.1257334 0.6253365 0.02779268 24.21905      6.189004      0.14769
##      m1_eff_conc m2_eff_conc sr1_eff_conc VSd_eff_conc CH3COOH_eff_conc
## 364  0.1411602   0.7488771   0.02115912   17.89738      1.14339
## 365  0.1411602   0.7488771   0.02115912   17.89738      1.14339
## 366  0.1411602   0.7488771   0.02115912   17.89738      1.14339
## 367  0.1411602   0.7488771   0.02115912   17.89738      1.14339
## 368  0.1411602   0.7488771   0.02115912   17.89738      1.14339
## 369  0.1411602   0.7488771   0.02115912   17.89738      1.14339
```

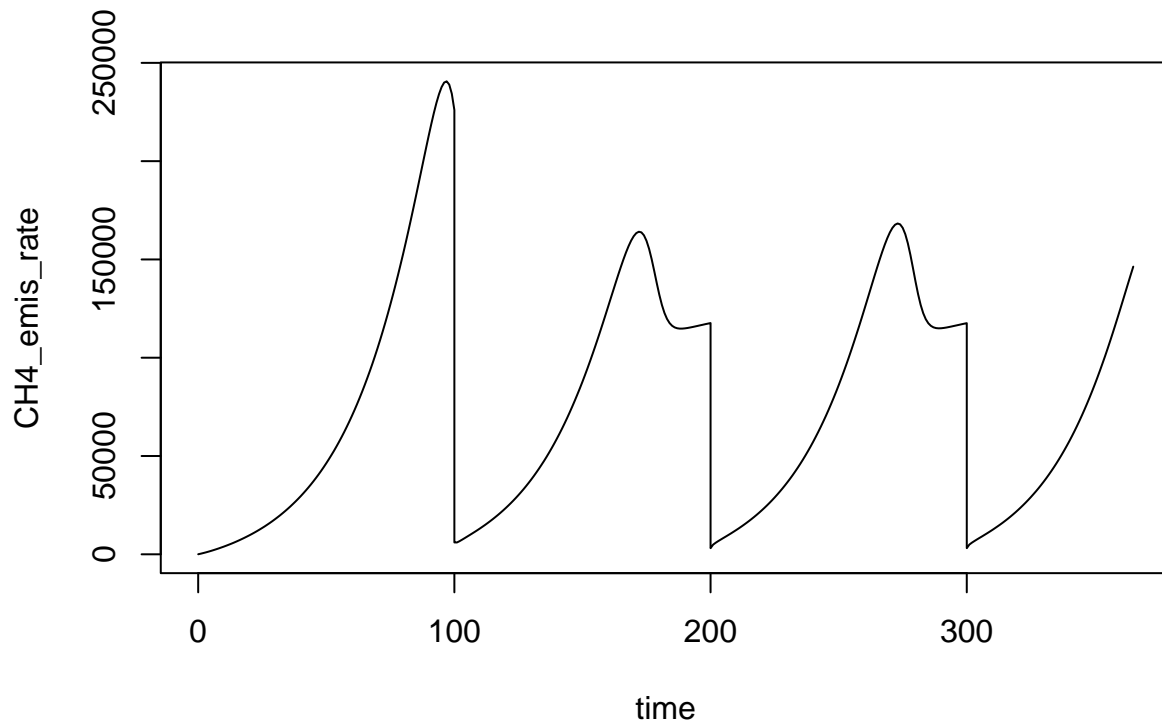
The effluent columns are cumulative. Is this what we had before? I did it for COD balance checking. We will have to discuss what is needed.

Here are some results.

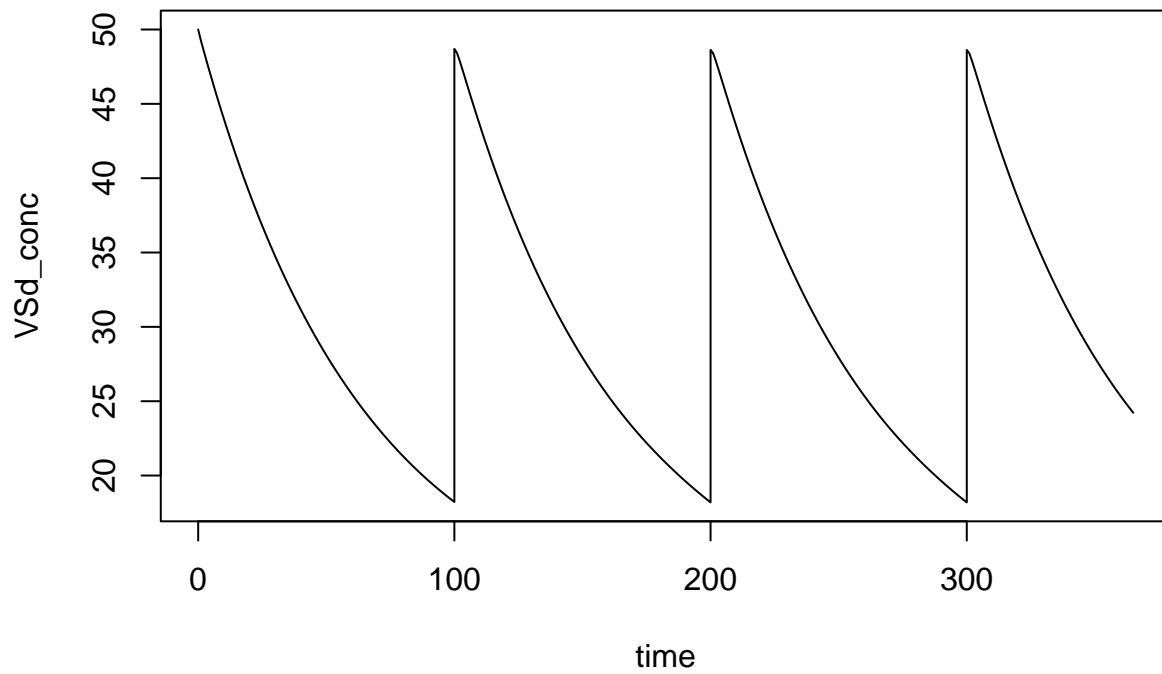
```
plot(slurry_mass ~ time, data = out1, type = 'l')
```



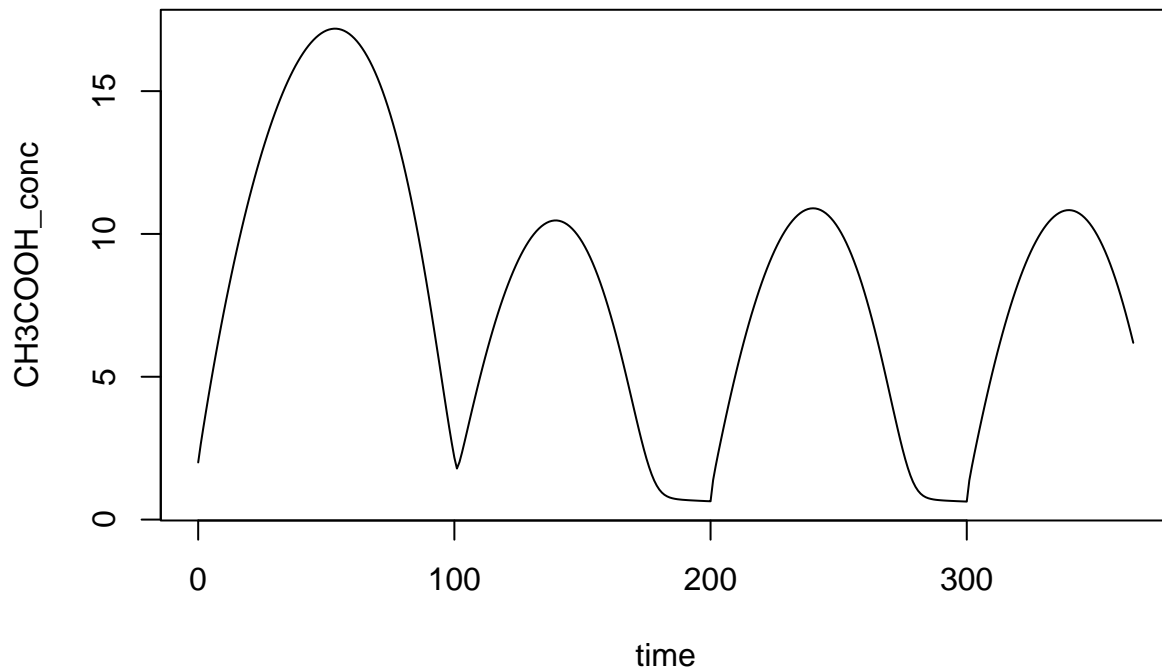
```
plot(CH4_emis_rate ~ time, data = out1, type = 'l')
```



```
plot(VSd_conc ~ time, data = out1, type = 'l')
```

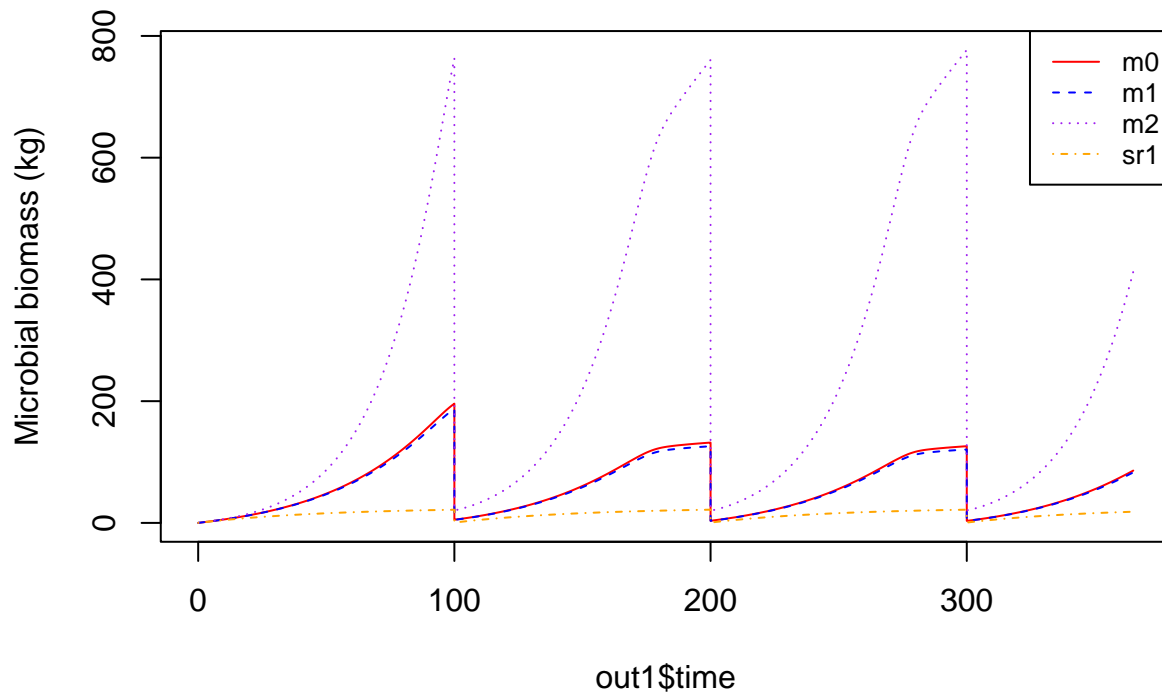


```
plot(CH3COOH_conc ~ time, data = out1, type = 'l')
```



And methanogens.

```
line_colors <- c('red', 'blue', 'purple','orange')
matplot(out1$time, out1[, nn <- c('m0','m1','m2','sr1')]/1000,
        type = 'l', lty = c(1:length(nn)), col = line_colors, ylab = 'Microbial biomass (kg)')
legend("topright", legend = nn, lty = c(1:length(nn)), col = line_colors, lwd = 1, cex = 0.8)
```



2. Substrate flexibility

Particulate substrates are defined in `sub_pars` now and there are no specific substrates hard-wired in the code. VFA is the only intermediate, and it is hard-wired. Here we will use three substrates. Parameter values have no connection to reality in this example.

```
sub_pars2 <- list(subs = c('cellulose', 'protein', 'lipids'),
  T_opt_hyd = c(all = 60),
  T_min_hyd = c(all = 0),
  T_max_hyd = c(all = 90),
  hydrol_opt = c(lipids = 0.1, protein = 0.01, cellulose = 0.05),
  sub_fresh = c(lipids = 3, protein = 20, cellulose = 35),
  sub_init = c(lipids = 3, protein = 20, cellulose = 35))
```

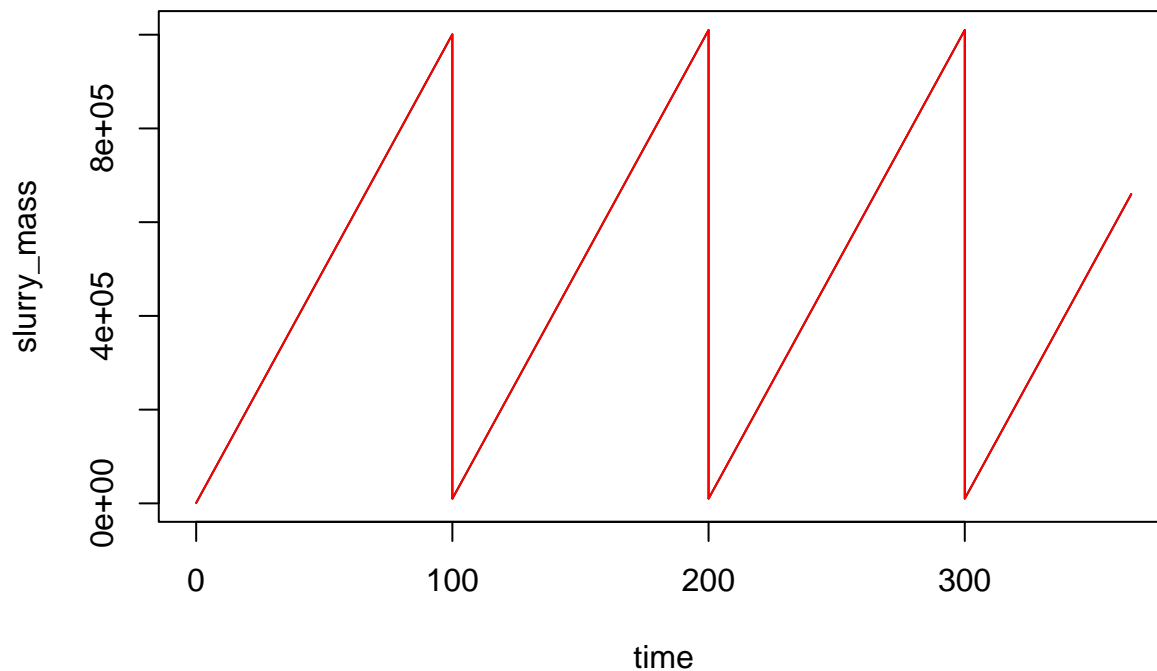
```
devtools::load_all()
```

```
## i Loading ABM
```

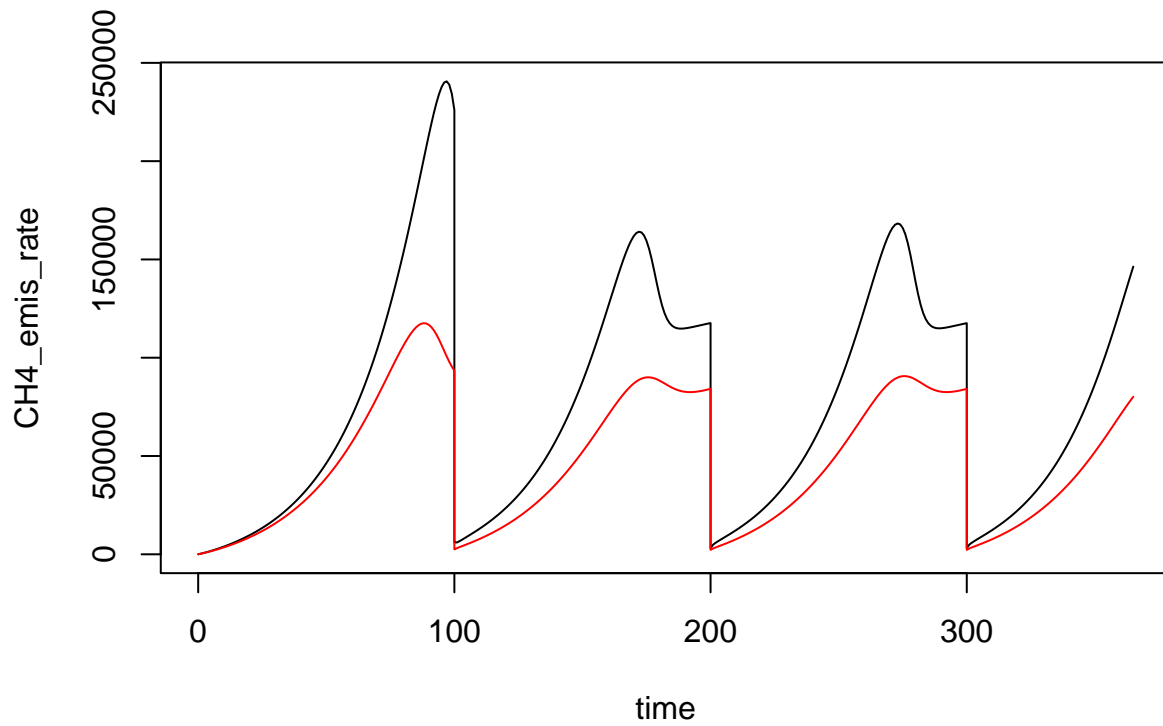
```
out2 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars2,
  chem_pars = chem_pars)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

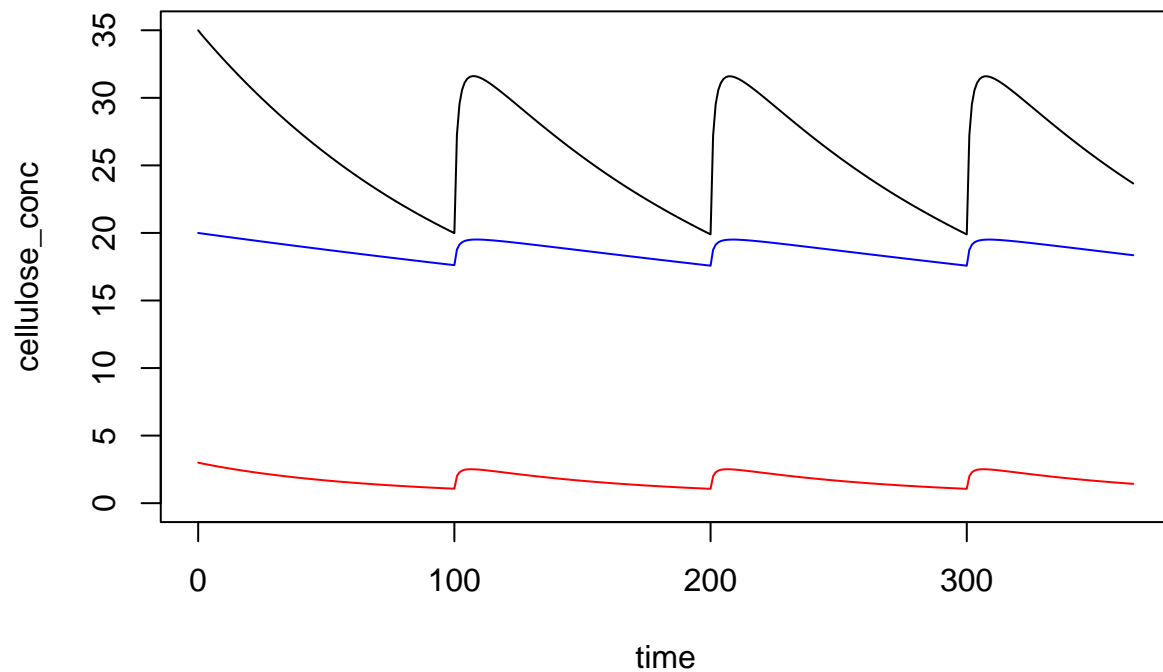
```
plot(slurry_mass ~ time, data = out2, type = 'l')
lines(slurry_mass ~ time, data = out1, col = 'red')
```



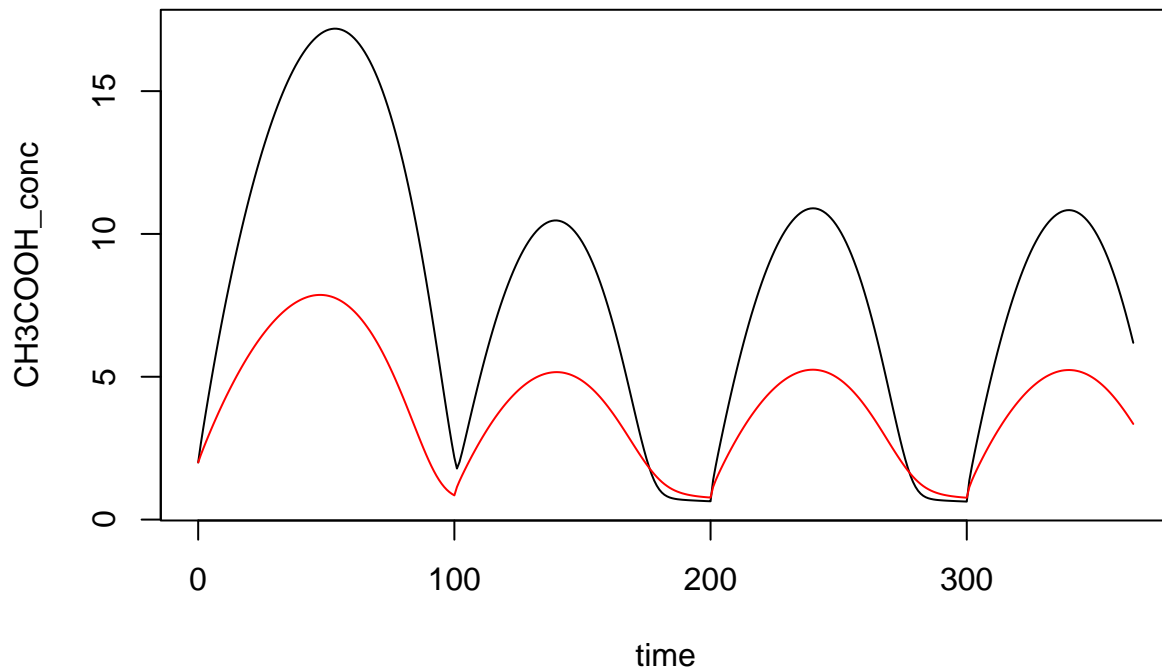
```
plot(CH4_emis_rate ~ time, data = out1, type = 'l')
lines(CH4_emis_rate ~ time, data = out2, col = 'red')
```



```
plot(cellulose_conc ~ time, data = out2, type = 'l', ylim = c(0, 35))
lines(lipids_conc ~ time, data = out2, type = 'l', col = 'red')
lines(protein_conc ~ time, data = out2, type = 'l', col = 'blue')
```



```
plot(CH3COOH_conc ~ time, data = out1, type = 'l')
lines(CH3COOH_conc ~ time, data = out2, col = 'red')
```

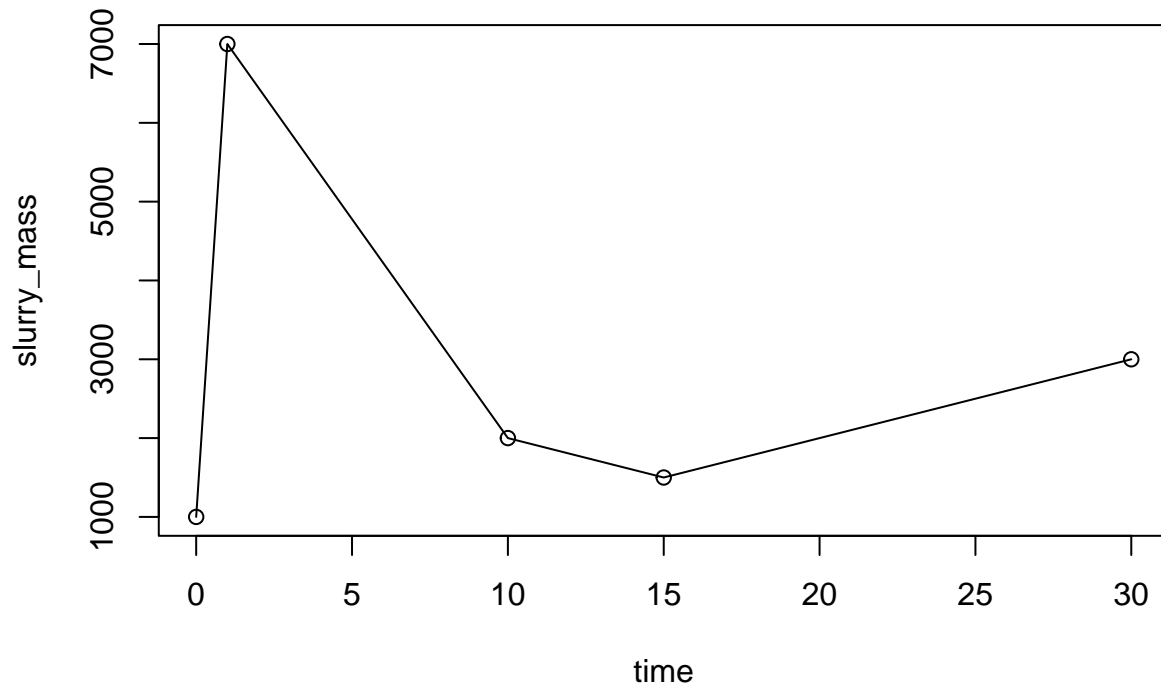



This flexibility comes from an approach similar to what we used for microbial groups.

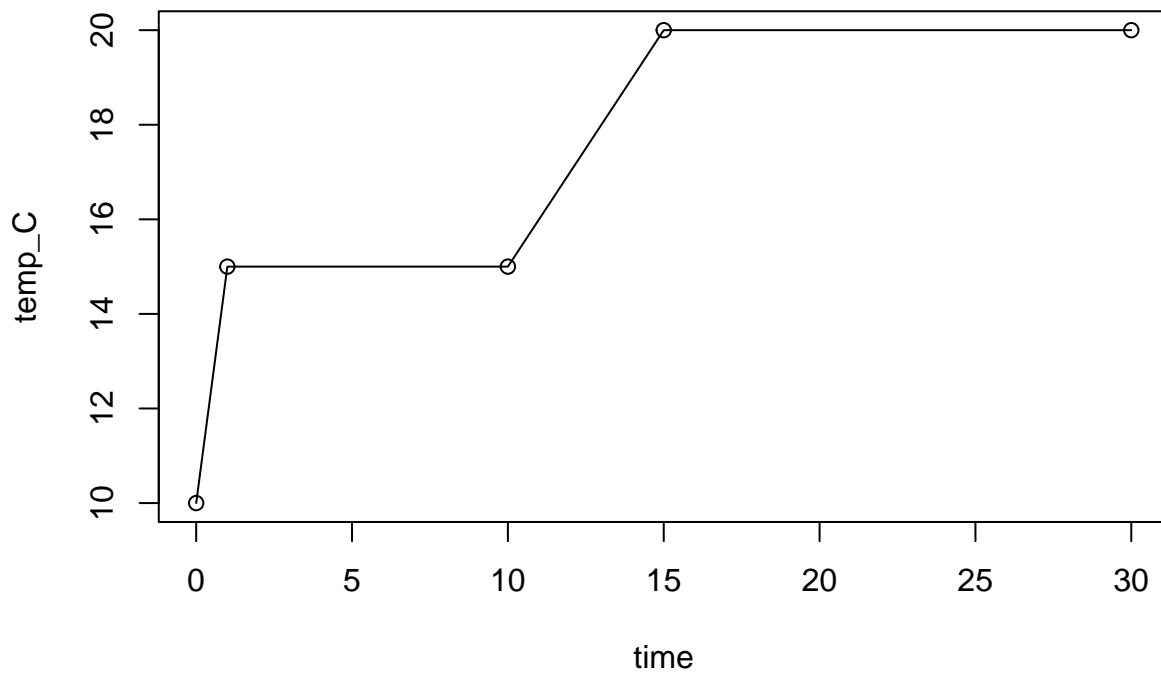
3. Time-variable inputs part 1

The `abm()` function can handle variability over time in any inputs now. Here slurry mass and temperature will vary.

```
var_dat <- data.frame(time = c(0, 1, 10, 15, 30),  
                      slurry_mass = c(1000, 7000, 2000, 1500, 3000),  
                      temp_C = c(10, 15, 15, 20, 20))  
  
plot(slurry_mass ~ time, data = var_dat, type = 'o')
```



```
plot(temp_C ~ time, data = var_dat, type = 'o')
```



This data frame goes in the `var_pars` argument, which must be a list, even though it might have a single element named `var`. The `var` element is the only required one. The `var` data frame must have a `slurry_mass` column if it is used—it is not possible to use an `abm_regular()`-like approach with variable temperature etc.

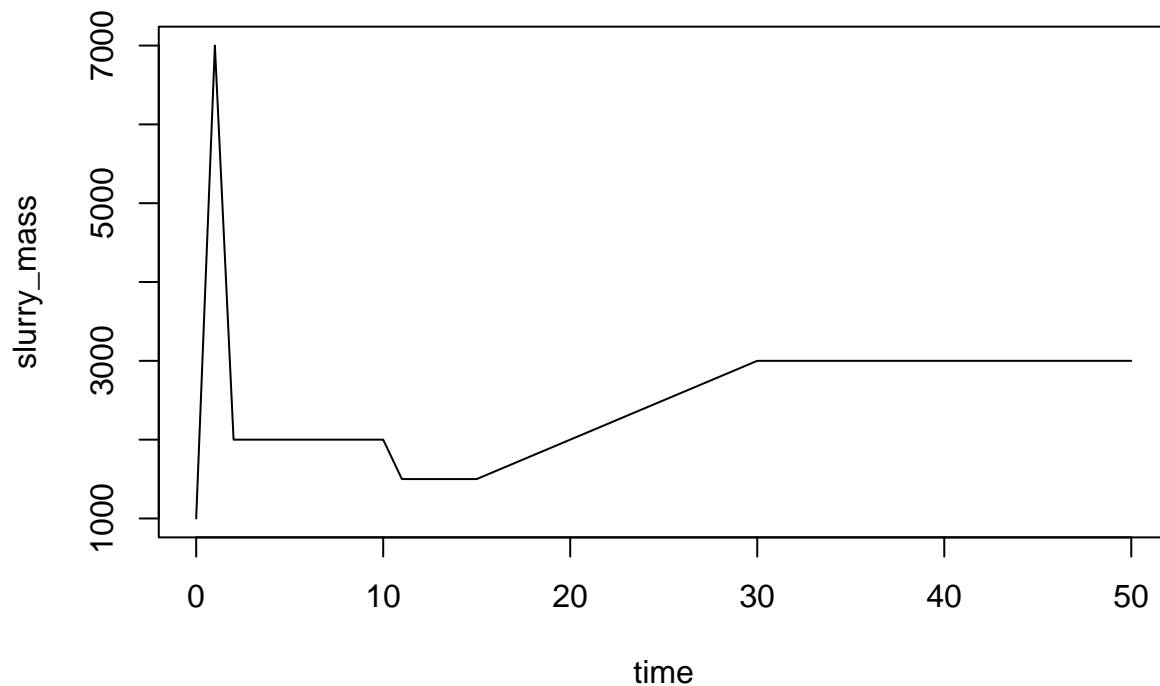
```
var_pars <- list(var = var_dat)
```

```
out3a <- abm(50,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
```

```
mic_pars = mic_pars,
sub_pars = sub_pars,
chem_pars = chem_pars,
var_pars = var_pars)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

```
plot(slurry_mass ~ time, data = out3a, type = 'l')
```



The “late” and “mid” options are still available, but now through `ctrl_pars`. Here we can change the value through `add_pars`

```
out3b <- abm(50,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars,
  var_pars = var_pars,
  add_pars = list(approx_method = 'late'))
```

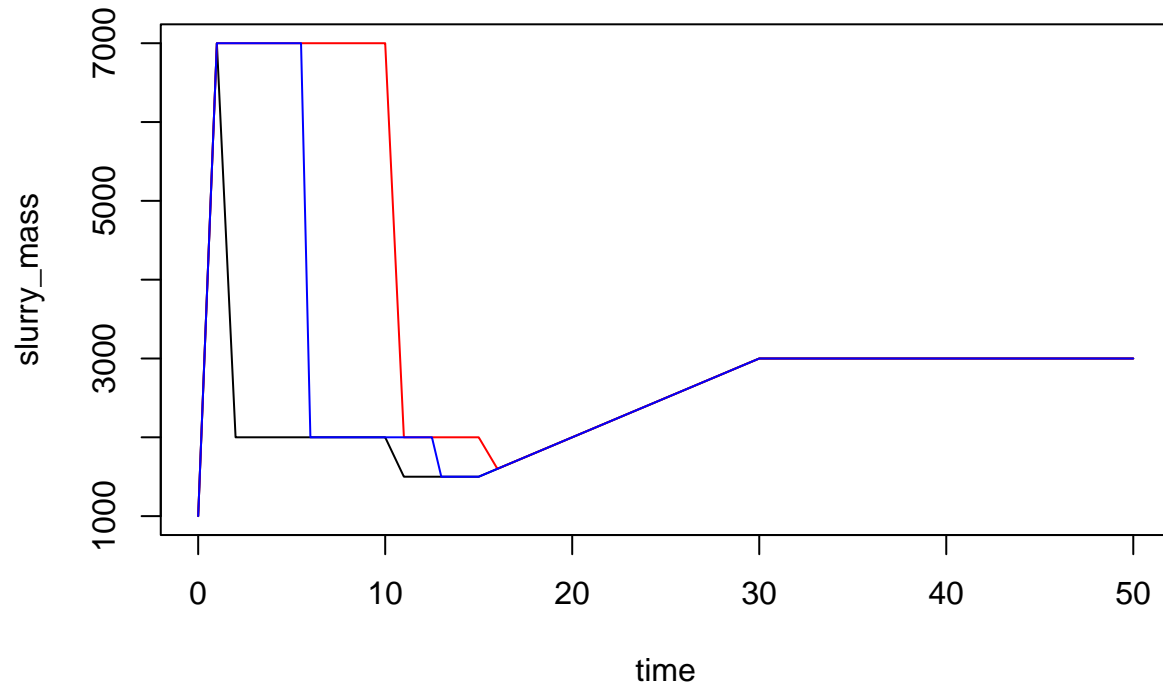
```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

```
out3c <- abm(50,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars,
  var_pars = var_pars,
```

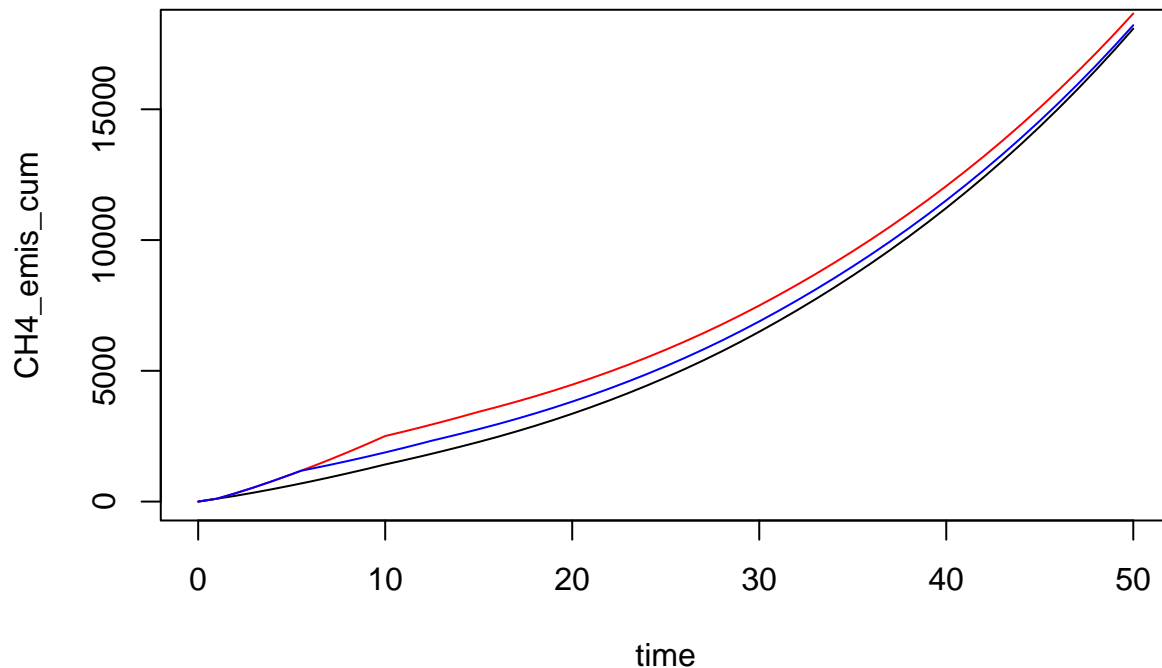
```
add_pars = list(approx_method = 'mid'))
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):  
## Size-variable parameter problem: Missing element(s) in kss.
```

```
plot(slurry_mass ~ time, data = out3a, type = 'l')  
lines(slurry_mass ~ time, data = out3b, col = 'red')  
lines(slurry_mass ~ time, data = out3c, col = 'blue')
```



```
plot(CH4_emis_cum ~ time, data = out3a, type = 'l')  
lines(CH4_emis_cum ~ time, data = out3b, col = 'red')  
lines(CH4_emis_cum ~ time, data = out3c, col = 'blue')
```



4. Time-variable inputs part 2

Here we'll vary fresh substrate concentrations over time.

First the data frame with slurry mass.

```
var_dat <- data.frame(time = c(0, 1, 10, 15, 30, 50),
                      slurry_mass = c(1000, 7000, 2000, 5000, 3000, 10000))
var_dat
```

```
##   time slurry_mass
## 1    0         1000
## 2    1         7000
## 3   10         2000
## 4   15         5000
## 5   30         3000
## 6   50        10000
```

Then add `sub_fresh` values. Each row needs a list containing a named vector. This is somewhat unusual data frame usage, and there is a user-friendly alternative based on additional data frames in the `var` argument (see next section). But I've kept this demo.

```
var_dat
##   time slurry_mass
## 1    0         1000
## 2    1         7000
## 3   10         2000
## 4   15         5000
## 5   30         3000
## 6   50        10000

var_dat$sub_fresh <- rep(list(c(VSd = 50)), nrow(var_dat))
var_dat$sub_fresh[3] <- list(c(VSd = 100))
```

```
var_dat$sub_fresh[4] <- list(c(VSd = 10))
var_dat$sub_fresh[5] <- list(c(VSd = 0))
var_dat$sub_fresh[6] <- list(c(VSd = 200))
var_dat
```

```
##   time slurry_mass sub_fresh
## 1    0         1000         50
## 2    1         7000         50
## 3   10         2000        100
## 4   15         5000         10
## 5   30         3000          0
## 6   50        10000        200
```

```
var_dat[1, 3]
```

```
## [[1]]
## VSd
## 50
```

```
var_dat[5, 3]
```

```
## [[1]]
## VSd
## 0
```

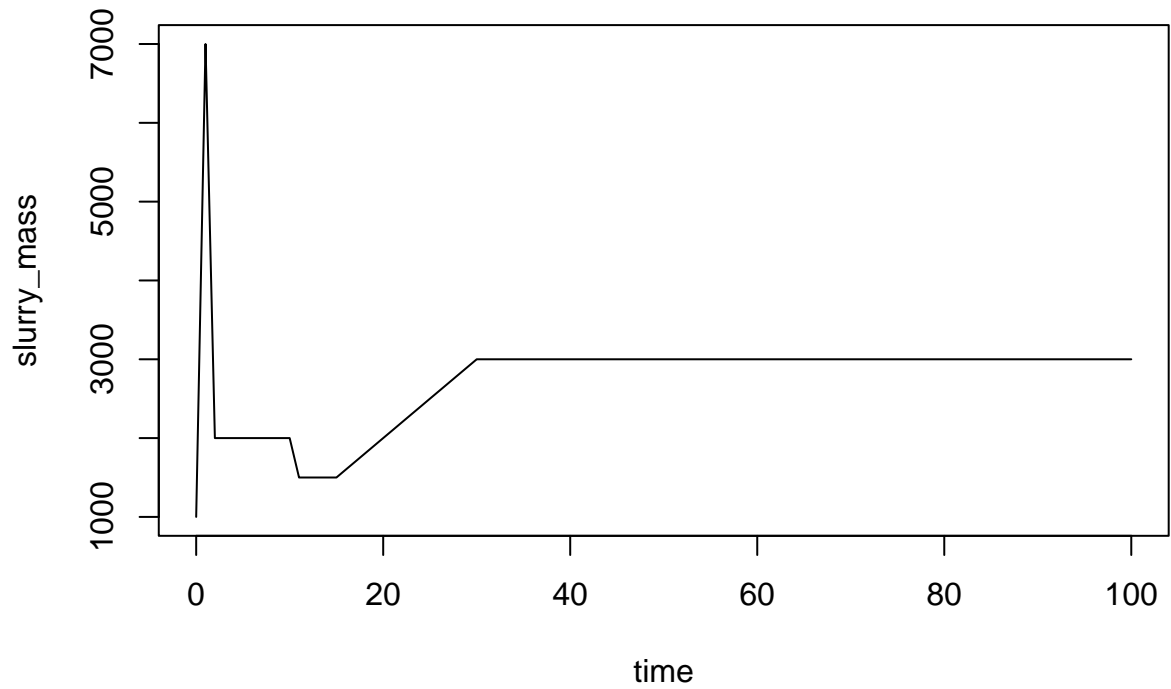
```
devtools::load_all()
```

```
## i Loading ABM
```

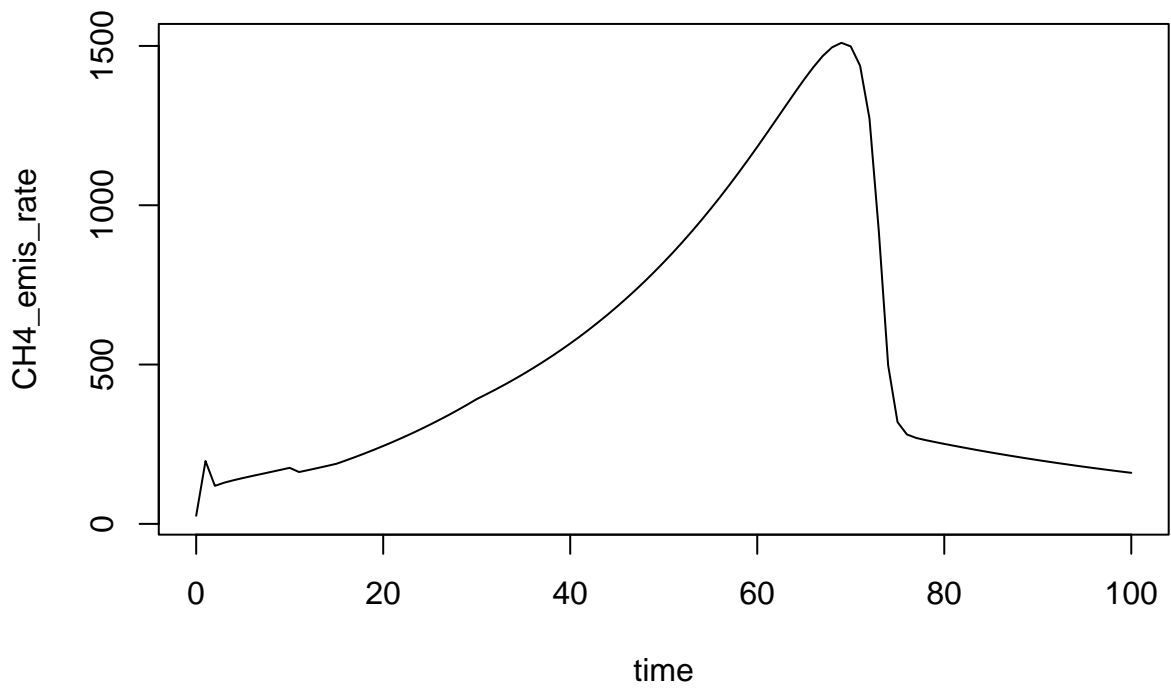
```
out4a <- abm(100,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars,
  var_pars = var_pars)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

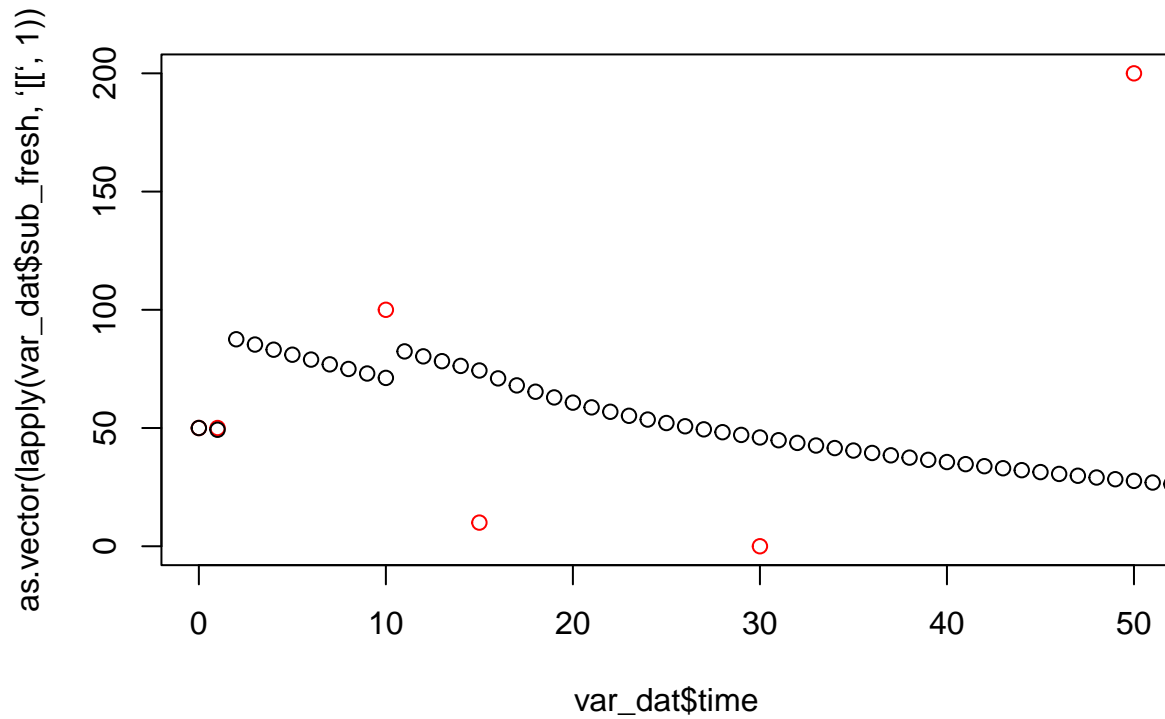
```
plot(slurry_mass ~ time, data = out4a, type = 'l')
```



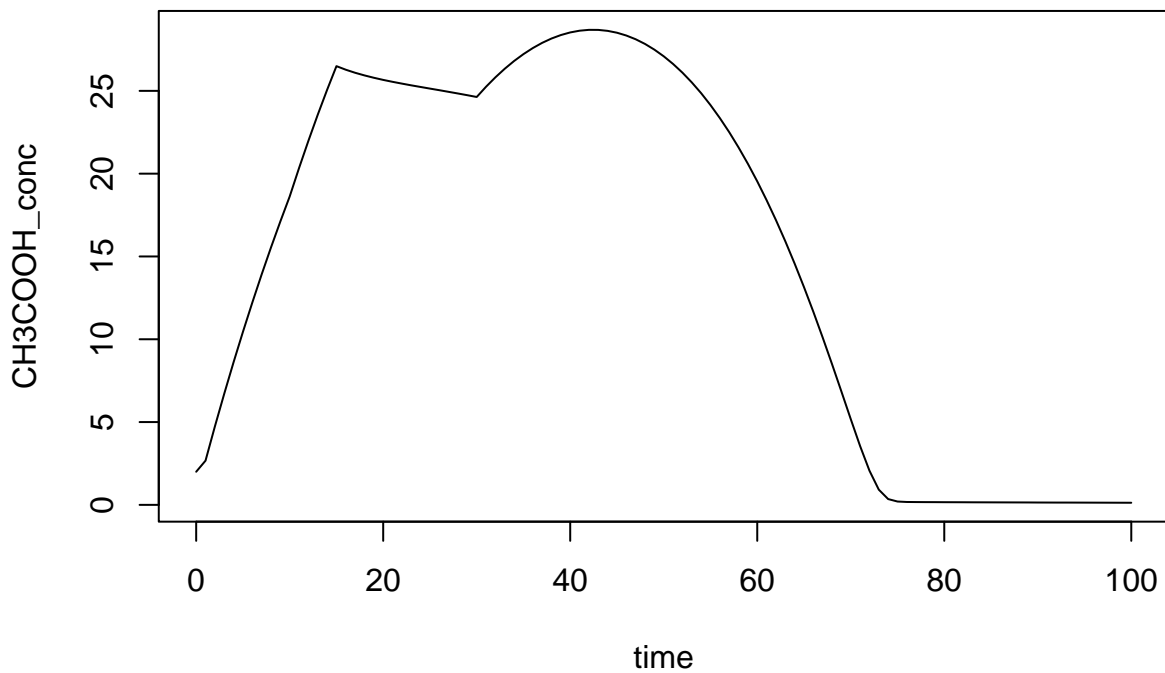
```
plot(CH4_emis_rate ~ time, data = out4a, type = 'l')
```



```
plot(var_dat$time, as.vector(lapply(var_dat$sub_fresh, `[`, 1)), type = 'p', col = 'red')
lines(VSd_conc ~ time, data = out4a, type = 'p')
```



```
plot(CH3COOH_conc ~ time, data = out4a, type = 'l')
```



Let's vary some microbial parameters as well. And pH.

```
var_dat <- data.frame(time = c(0, 1, 10, 15, 30, 50),
                      slurry_mass = c(1000, 7000, 2000, 5000, 3000, 10000),
                      pH = 7 - 0:5/10)
var_dat
```

```
##   time slurry_mass  pH
## 1    0         1000 7.0
```



```
## 2    1      7000 6.9
## 3   10      2000 6.8
## 4   15      5000 6.7
## 5   30      3000 6.6
## 6   50     10000 6.5
```

VSd.

```
var_dat$sub_fresh <- rep(list(c(VSd = 50)), nrow(var_dat))
var_dat$sub_fresh[3] <- list(c(VSd = 100))
```

Some microbial parameters for a shift in temperature optima, “adaptation” for example.

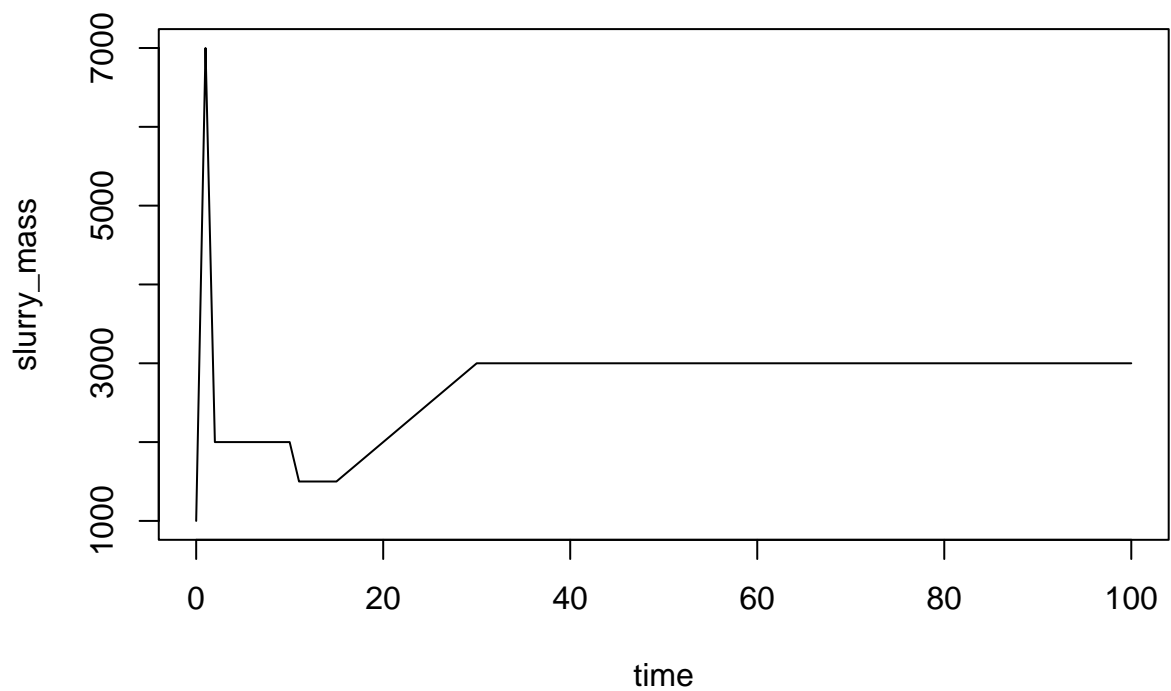
```
for (i in 1:nrow(var_dat)) {
  var_dat$T_opt[i] <- list(grp_pars$T_opt + 2 * i)
}
var_dat
```

```
##    time slurry_mass  pH sub_fresh      T_opt
## 1     0       1000 7.0      50 20, 20, 30, 46
## 2     1       7000 6.9      50 22, 22, 32, 48
## 3    10       2000 6.8     100 24, 24, 34, 50
## 4    15       5000 6.7      50 26, 26, 36, 52
## 5    30       3000 6.6      50 28, 28, 38, 54
## 6    50      10000 6.5      50 30, 30, 40, 56
```

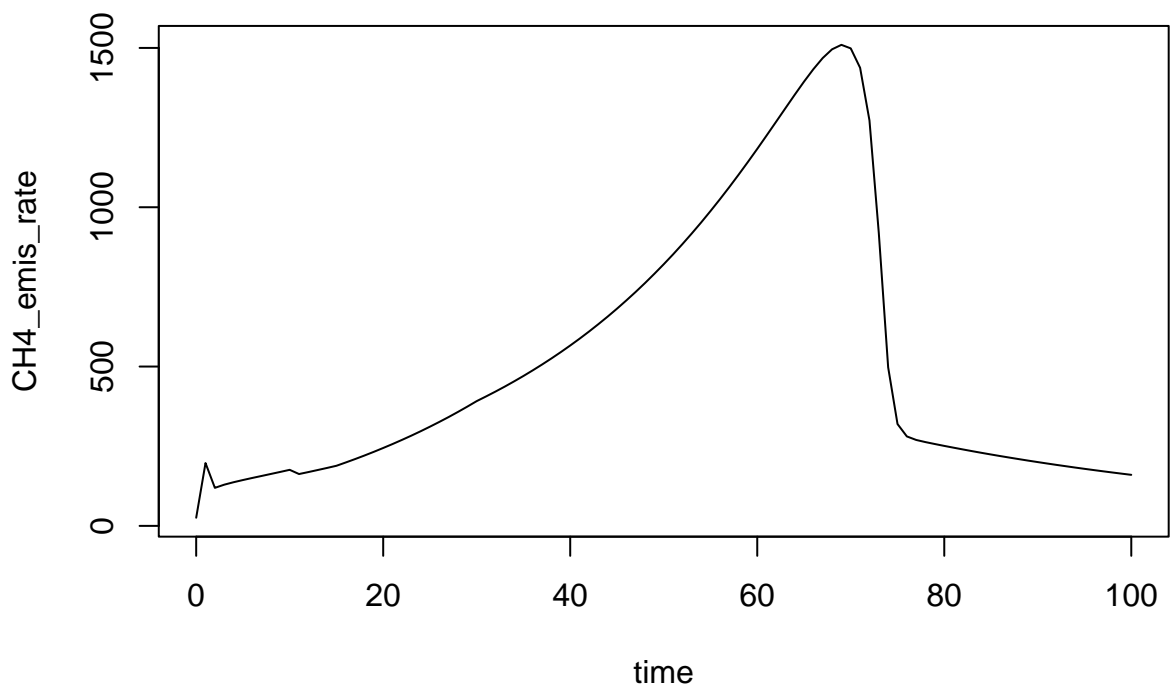
```
out4b <- abm(100,
  mng_pars = mng_pars,
  man_pars = man_pars,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars,
  var_pars = var_pars)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

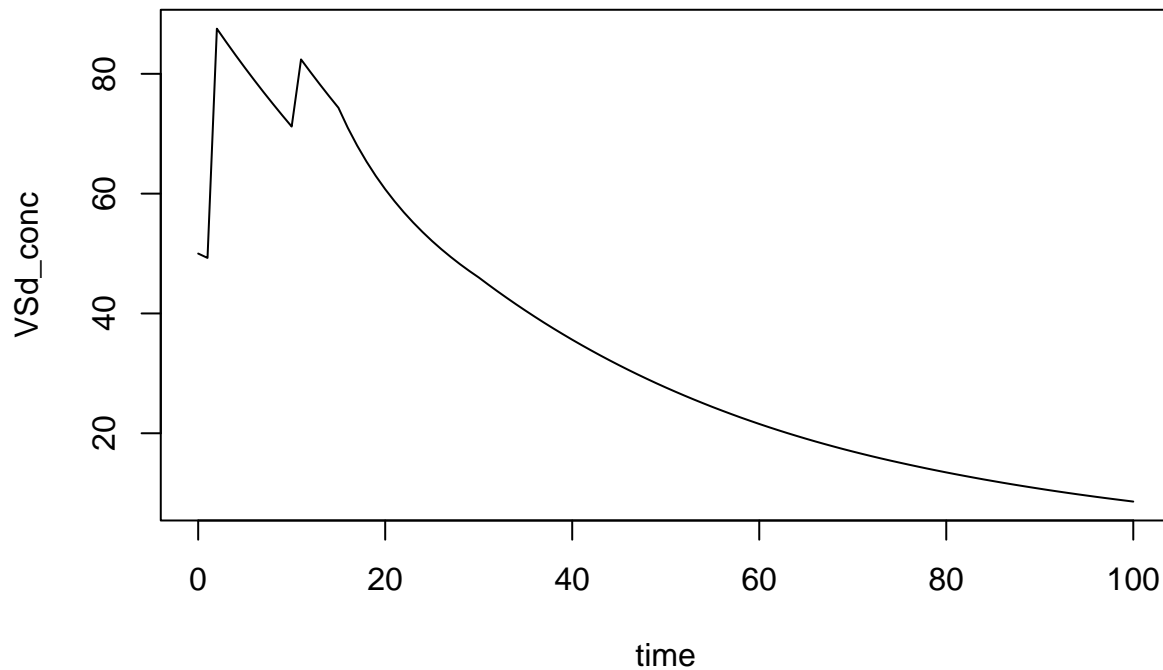
```
plot(slurry_mass ~ time, data = out4b, type = 'l')
```



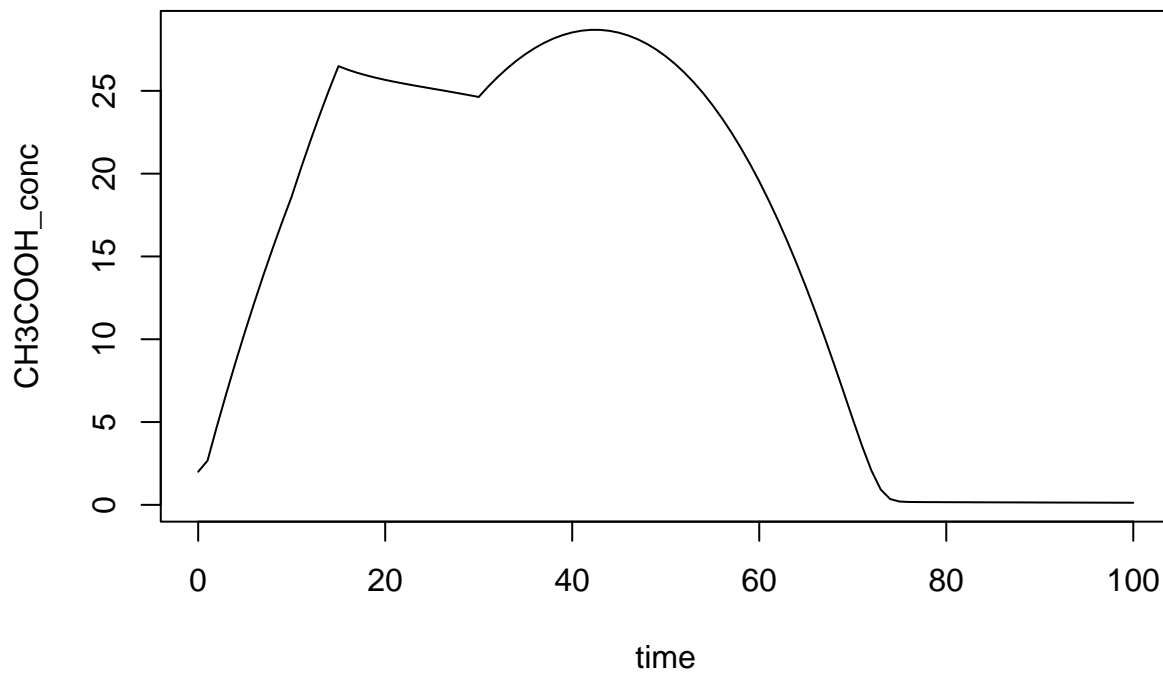
```
plot(CH4_emis_rate ~ time, data = out4b, type = 'l')
```



```
plot(VSd_conc ~ time, data = out4b, type = 'l')
```



```
plot(CH3COOH_conc ~ time, data = out4b, type = 'l')
```



5. Time-variable inputs part 3

The list-in-data frame approach is clunky. Here is an alternative.

```
var_dat <- data.frame(time = c(0, 1, 10, 15, 30, 50),
                      slurry_mass = c(1000, 7000, 2000, 5000, 3000, 10000))
var_dat
```

```
##   time slurry_mass
```

```
## 1    0      1000
## 2    1      7000
## 3   10      2000
## 4   15      5000
## 5   30      3000
## 6   50     10000
```

Make a separate data frame for each other argument (any name, but note column names!).

```
sub_fresh_dat = data.frame(time = c(0, 1, 10, 15, 30, 50),
                           VSd = c(50, 100, 0, 0, 200, 50))
```

```
T_opt_dat = data.frame(time = c(0, 1, 10, 15, 30, 50),
                        m1 = 20 + 0:5 * 2,
                        m2 = 20 + 0:5 * 2,
                        m3 = 30 + 0:5 * 2,
                        sr1 = 46 + 0:5 * 2)
```

and combine them in a list, using the parameter element names for element names (e.g., `sub_fresh` is the name of an element in `sub_pars`).

```
var_pars <- list(var = var_dat, sub_fresh = sub_fresh_dat, T_opt = T_opt_dat)
```

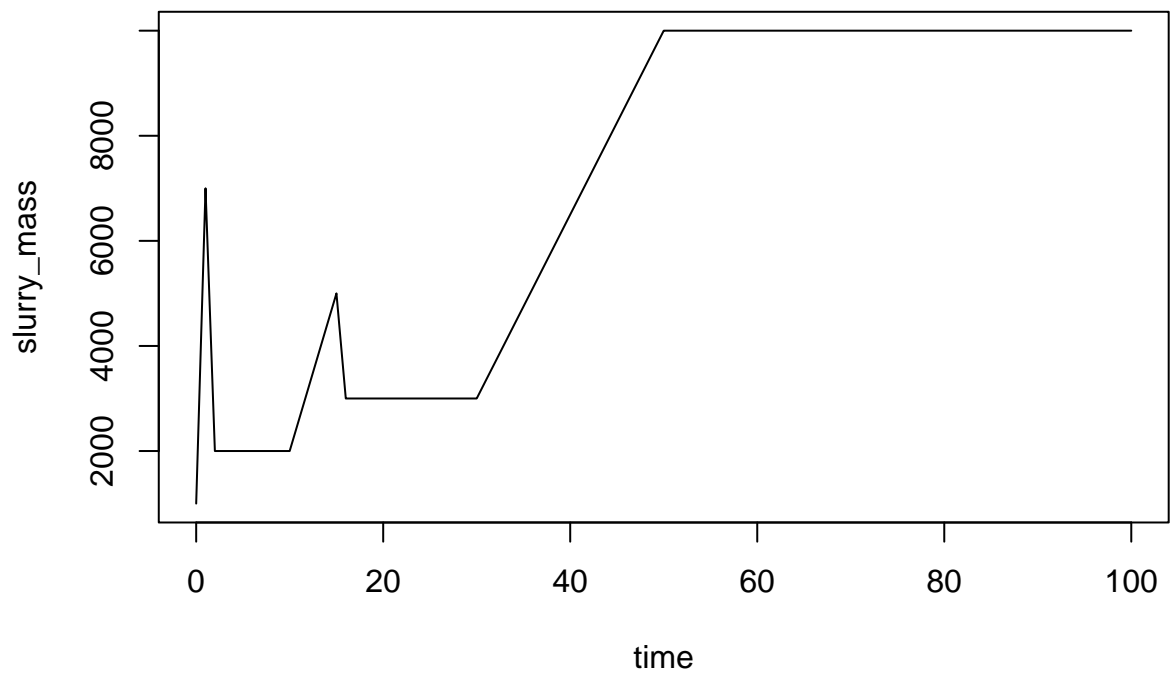
```
devtools::load_all()
```

```
## i Loading ABM
```

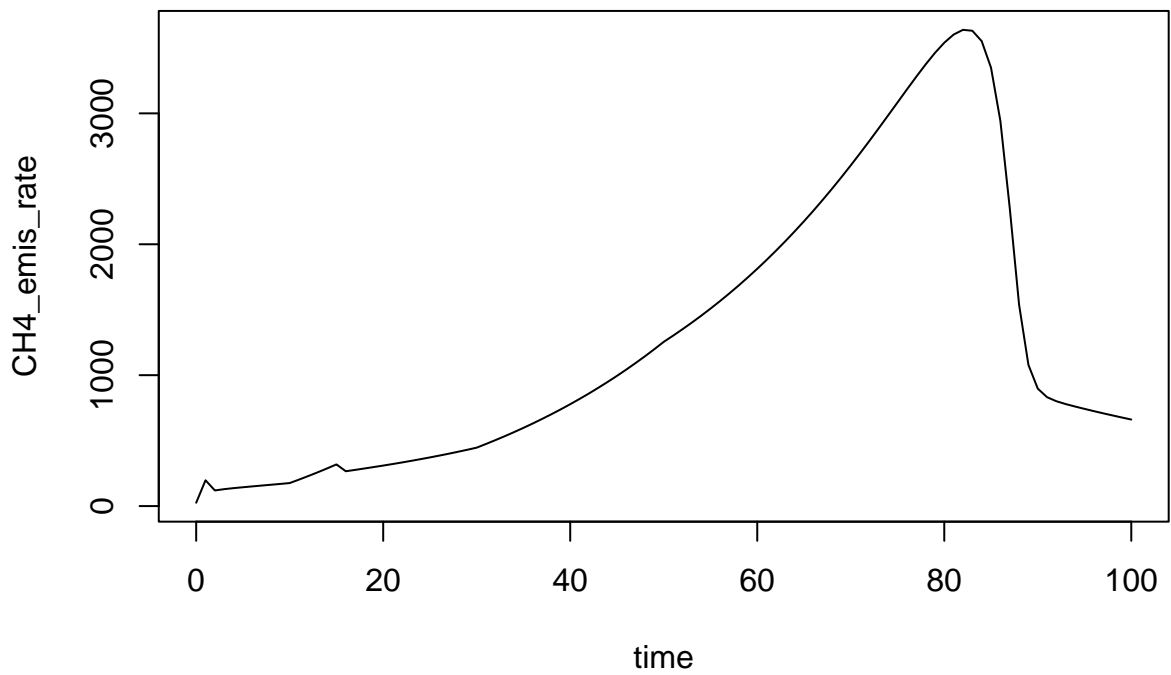
```
out5 <- abm(100,
            mng_pars = mng_pars,
            man_pars = man_pars,
            grp_pars = grp_pars,
            mic_pars = mic_pars,
            sub_pars = sub_pars,
            chem_pars = chem_pars,
            var_pars = var_pars)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

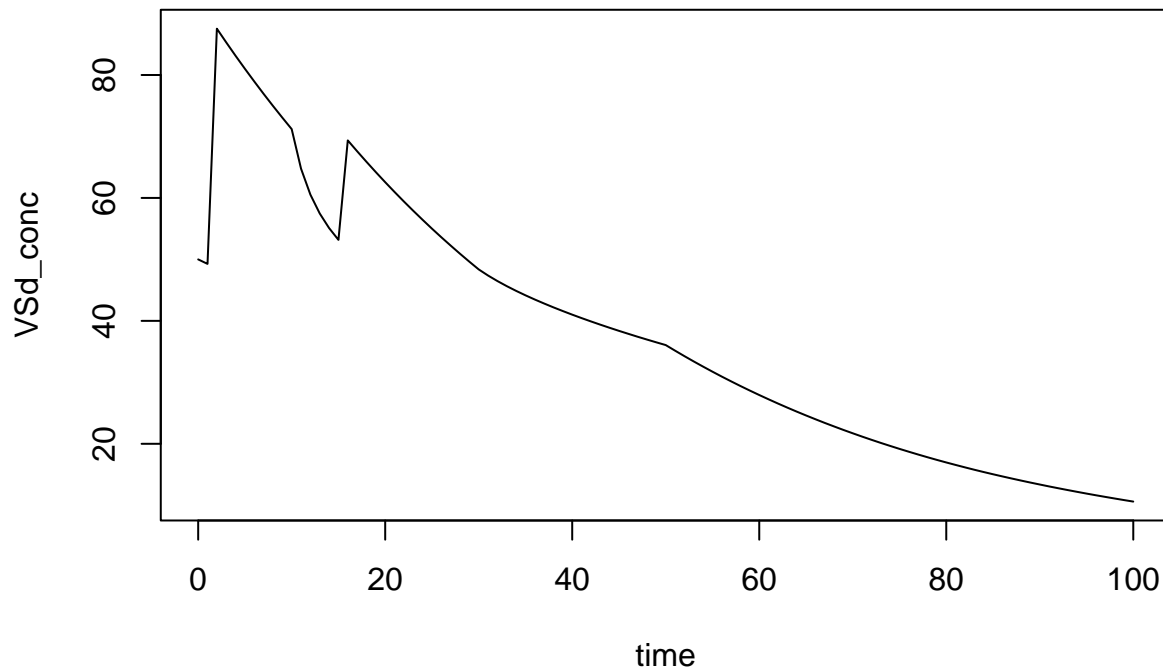
```
plot(slurry_mass ~ time, data = out5, type = 'l')
```



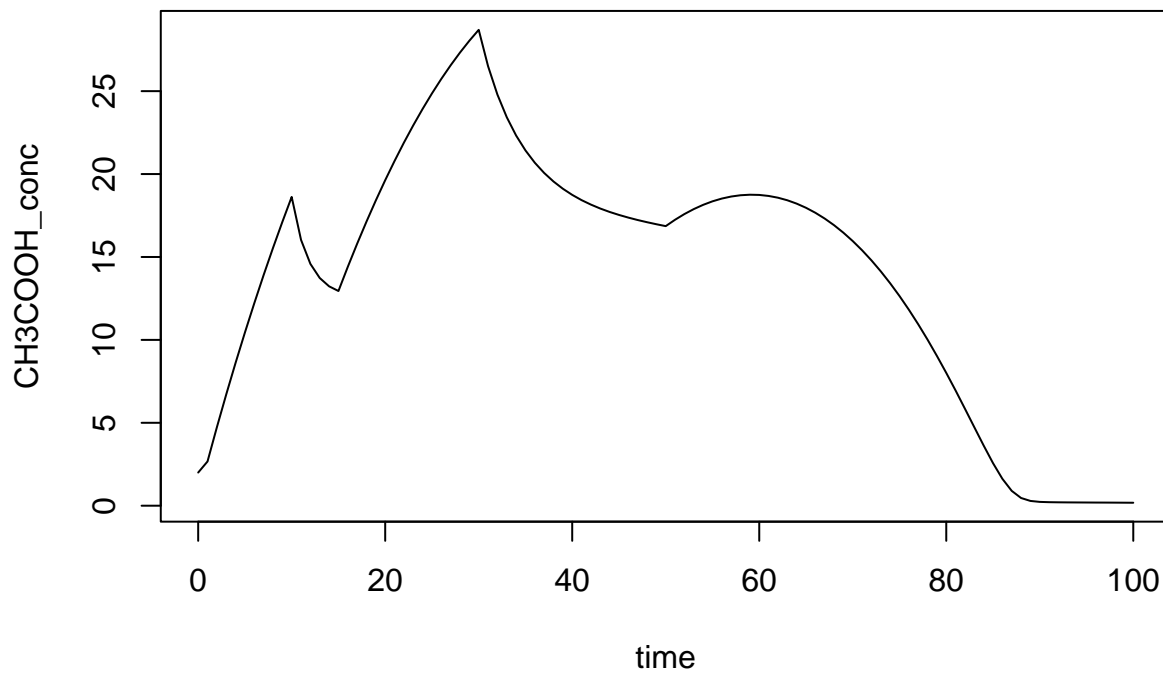
```
plot(CH4_emis_rate ~ time, data = out5, type = 'l')
```



```
plot(VSd_conc ~ time, data = out5, type = 'l')
```



```
plot(CH3COOH_conc ~ time, data = out5, type = 'l')
```



6. Flexible solutes and sulfate reduction

Any conservative solute can be added in `man_pars`, using any names. I am moving toward using a “master species” approach, so it makes sense to use the chemical formula of the primary species, with `p` or `m` for a charge symbol. The `comp` part of the name below is for “component”.

```
man_pars6 <- list(comps = c('H2S', 'SO4m2', 'NH4p'),
  comp_fresh = c(H2S = 0.01, SO4m2 = 0.2, NH4p = 2.5),
  VFA_fresh = c(CH3COOH = 2),
```

```
pH = 7, dens = 1000)
```

Note that CH3COOH is still special—it has a fixed name in the code and is not conservative.

```
devtools::load_all()
```

```
## i Loading ABM
```

```
out6a <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars6,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 2%
```

```
tail(out6a)
```

```
##      time      m0      m1      m2      sr1      VSd      H2S      S04m2      NH4p
## 364 360 74751.48 72348.85 334784.1 22188.55 15469950 115596.8 12503.17 1525000
## 365 361 76892.18 74389.05 348806.4 22374.50 15577739 117589.8 12610.22 1550000
## 366 362 79063.75 76457.33 363289.4 22556.57 15683138 119579.7 12720.29 1575000
## 367 363 81263.87 78551.47 378229.0 22734.81 15786218 121566.3 12833.71 1600000
## 368 364 83489.65 80668.67 393616.7 22909.25 15887047 123549.1 12950.88 1625000
## 369 365 85737.49 82805.50 409438.1 23079.95 15985694 125527.7 13072.29 1650000
##      CH3COOH slurry_mass CH4_emis_cum slurry_load COD_load CH4_emis_rate temp_C
## 364 4760816      610000      26704791      3600000 187920000      124223.5      20
## 365 4650827      620000      26831123      3610000 188442000      128446.1      20
## 366 4525825      630000      26961690      3620000 188964000      132688.7      20
## 367 4385715      640000      27096499      3630000 189486000      136926.3      20
## 368 4230525      650000      27235530      3640000 190008000      141127.4      20
## 369 4060430      660000      27378729      3650000 190530000      145252.5      20
##      pH      m0_eff      m1_eff      m2_eff      sr1_eff      VSd_eff      H2S_eff      S04m2_eff      NH4p_eff
## 364 7 440845.9 421385.9 2227298 78898.1 53534259 561218.1 66891.92 7477500
## 365 7 440845.9 421385.9 2227298 78898.1 53534259 561218.1 66891.92 7477500
## 366 7 440845.9 421385.9 2227298 78898.1 53534259 561218.1 66891.92 7477500
## 367 7 440845.9 421385.9 2227298 78898.1 53534259 561218.1 66891.92 7477500
## 368 7 440845.9 421385.9 2227298 78898.1 53534259 561218.1 66891.92 7477500
## 369 7 440845.9 421385.9 2227298 78898.1 53534259 561218.1 66891.92 7477500
##      CH3COOH_eff slurry_mass_eff slurry_depth      m0_conc      m1_conc      m2_conc
## 364 3372964      2991000      6.1 0.1225434 0.1186047 0.5488264
## 365 3372964      2991000      6.2 0.1240196 0.1199823 0.5625909
## 366 3372964      2991000      6.3 0.1254980 0.1213608 0.5766498
## 367 3372964      2991000      6.4 0.1269748 0.1227367 0.5909829
## 368 3372964      2991000      6.5 0.1284456 0.1241056 0.6055642
## 369 3372964      2991000      6.6 0.1299053 0.1254629 0.6203607
##      sr1_conc VSd_conc      H2S_conc      S04m2_conc      NH4p_conc      CH3COOH_conc      m0_eff_conc
## 364 0.03637467 25.36057 0.1895030 0.02049700      2.5      7.804616 0.1473908
## 365 0.03608791 25.12539 0.1896609 0.02033907      2.5      7.501334 0.1473908
## 366 0.03580408 24.89387 0.1898091 0.02019094      2.5      7.183849 0.1473908
```

```
## 367 0.03552313 24.66596 0.1899473 0.02005267      2.5      6.852680 0.1473908
## 368 0.03524500 24.44161 0.1900756 0.01992443      2.5      6.508500 0.1473908
## 369 0.03496962 24.22075 0.1901935 0.01980650      2.5      6.152166 0.1473908
##      m1_eff_conc m2_eff_conc sr1_eff_conc VSd_eff_conc H2S_eff_conc
## 364 0.1408846 0.7446668 0.0263785 17.89845 0.1876356
## 365 0.1408846 0.7446668 0.0263785 17.89845 0.1876356
## 366 0.1408846 0.7446668 0.0263785 17.89845 0.1876356
## 367 0.1408846 0.7446668 0.0263785 17.89845 0.1876356
## 368 0.1408846 0.7446668 0.0263785 17.89845 0.1876356
## 369 0.1408846 0.7446668 0.0263785 17.89845 0.1876356
##      S04m2_eff_conc NH4p_eff_conc CH3COOH_eff_conc
## 364 0.0223644      2.5      1.127704
## 365 0.0223644      2.5      1.127704
## 366 0.0223644      2.5      1.127704
## 367 0.0223644      2.5      1.127704
## 368 0.0223644      2.5      1.127704
## 369 0.0223644      2.5      1.127704
```

```
head(out6a)
```

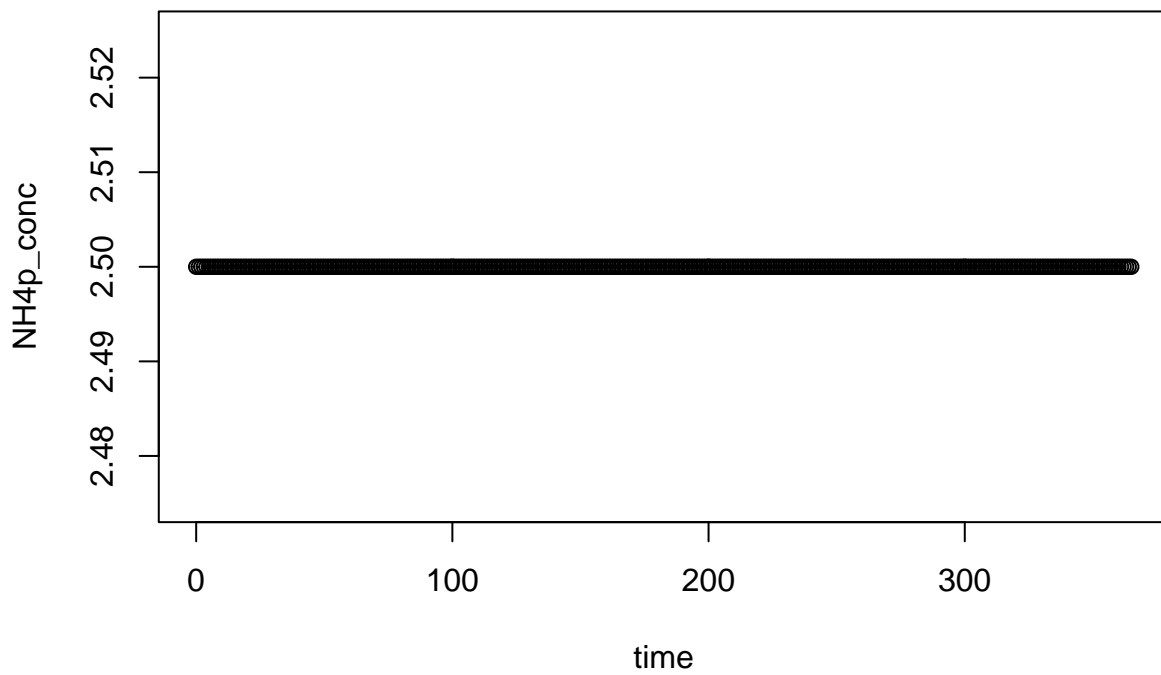
```
##      time      m0      m1      m2      sr1      VSd      H2S      S04m2
## 1      0 50.0000 50.0000 50.0000 50.0000 50000.0 10.0000 200.000
## 2      1 553.9977 553.8414 558.3574 555.0147 542318.5 268.9637 2041.036
## 3      2 1066.2080 1065.6056 1083.0938 1067.9792 1022077.4 788.7361 3621.264
## 4      3 1588.1329 1586.7628 1626.7495 1587.7198 1489599.2 1554.5640 4955.436
## 5      4 2120.8456 2118.3596 2191.3257 2112.6512 1945197.9 2545.3729 6064.627
## 6      5 2665.1817 2661.2072 2778.5446 2641.0154 2389179.5 3737.1169 6972.883
##      NH4p      CH3COOH slurry_mass CH4_emis_cum slurry_load COD_load CH4_emis_rate
## 1 2500 2000.00      1000      0.0000      0      0      25.52844
## 2 27500 28849.48      11000      163.4287      10000 522000 308.15048
## 3 52500 66757.26      21000      627.6723      20000 1044000 625.17882
## 4 77500 115293.28      31000      1422.3638      30000 1566000 968.06890
## 5 102500 174068.44      41000      2570.9586      40000 2088000 1332.46915
## 6 127500 242718.37      51000      4093.6840      50000 2610000 1716.05164
##      temp_C pH m0_eff m1_eff m2_eff sr1_eff VSd_eff H2S_eff S04m2_eff NH4p_eff
## 1      20 7      0      0      0      0      0      0      0      0
## 2      20 7      0      0      0      0      0      0      0      0
## 3      20 7      0      0      0      0      0      0      0      0
## 4      20 7      0      0      0      0      0      0      0      0
## 5      20 7      0      0      0      0      0      0      0      0
## 6      20 7      0      0      0      0      0      0      0      0
##      CH3COOH_eff slurry_mass_eff slurry_depth      m0_conc      m1_conc      m2_conc
## 1      0      0      0.01 0.05000000 0.05000000 0.05000000
## 2      0      0      0.11 0.05036343 0.05034921 0.05075976
## 3      0      0      0.21 0.05077181 0.05074312 0.05157589
## 4      0      0      0.31 0.05123009 0.05118590 0.05247579
## 5      0      0      0.41 0.05172794 0.05166731 0.05344697
## 6      0      0      0.51 0.05225847 0.05218053 0.05448127
##      sr1_conc VSd_conc      H2S_conc S04m2_conc NH4p_conc CH3COOH_conc m0_eff_conc
## 1 0.05000000 50.00000 0.01000000 0.2000000      2.5      2.000000      NaN
## 2 0.05045588 49.30168 0.02445125 0.1855488      2.5      2.622680      NaN
## 3 0.05085615 48.67035 0.03755886 0.1724411      2.5      3.178917      NaN
## 4 0.05121677 48.05159 0.05014723 0.1598528      2.5      3.719138      NaN
## 5 0.05152808 47.44385 0.06208226 0.1479177      2.5      4.245572      NaN
## 6 0.05178462 46.84666 0.07327680 0.1367232      2.5      4.759184      NaN
```


##	m1_eff_conc	m2_eff_conc	sr1_eff_conc	VSd_eff_conc	H2S_eff_conc	S04m2_eff_conc
## 1	NaN	NaN	NaN	NaN	NaN	NaN
## 2	NaN	NaN	NaN	NaN	NaN	NaN
## 3	NaN	NaN	NaN	NaN	NaN	NaN
## 4	NaN	NaN	NaN	NaN	NaN	NaN
## 5	NaN	NaN	NaN	NaN	NaN	NaN
## 6	NaN	NaN	NaN	NaN	NaN	NaN

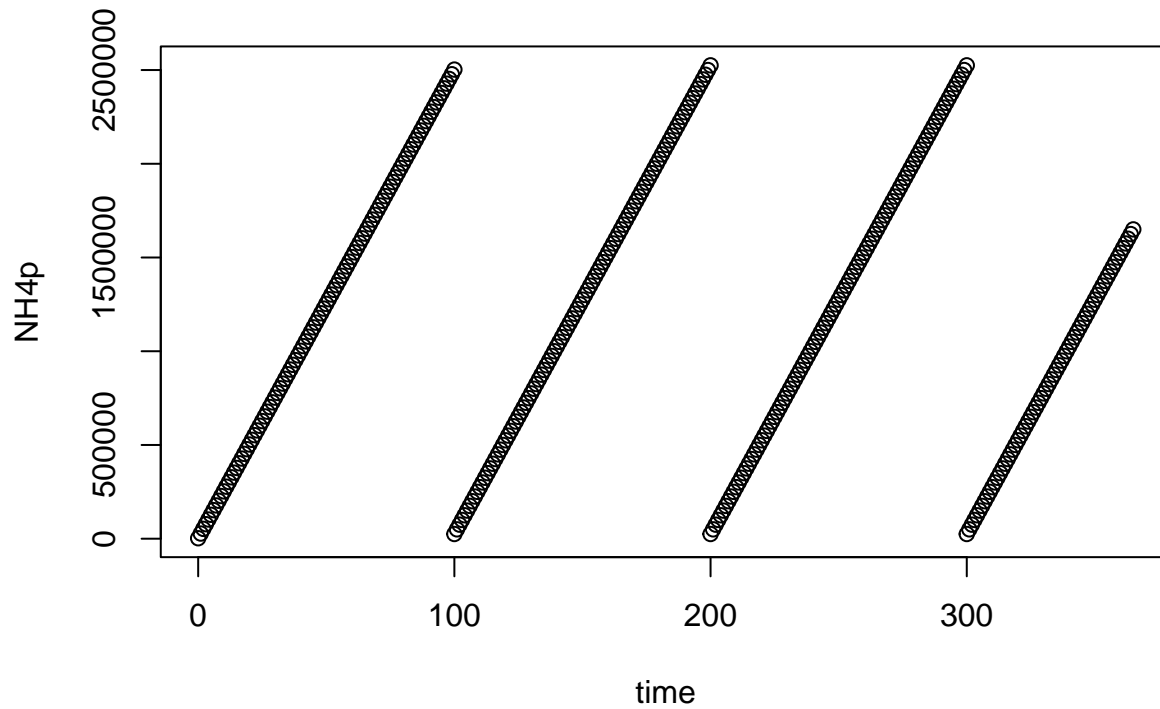
##	NH4p_eff_conc	CH3COOH_eff_conc
## 1	NaN	NaN
## 2	NaN	NaN
## 3	NaN	NaN
## 4	NaN	NaN
## 5	NaN	NaN
## 6	NaN	NaN

Conservative components are boring in output (without inhibition or volatilization).

```
plot(NH4p_conc ~ time, data = out6a)
```

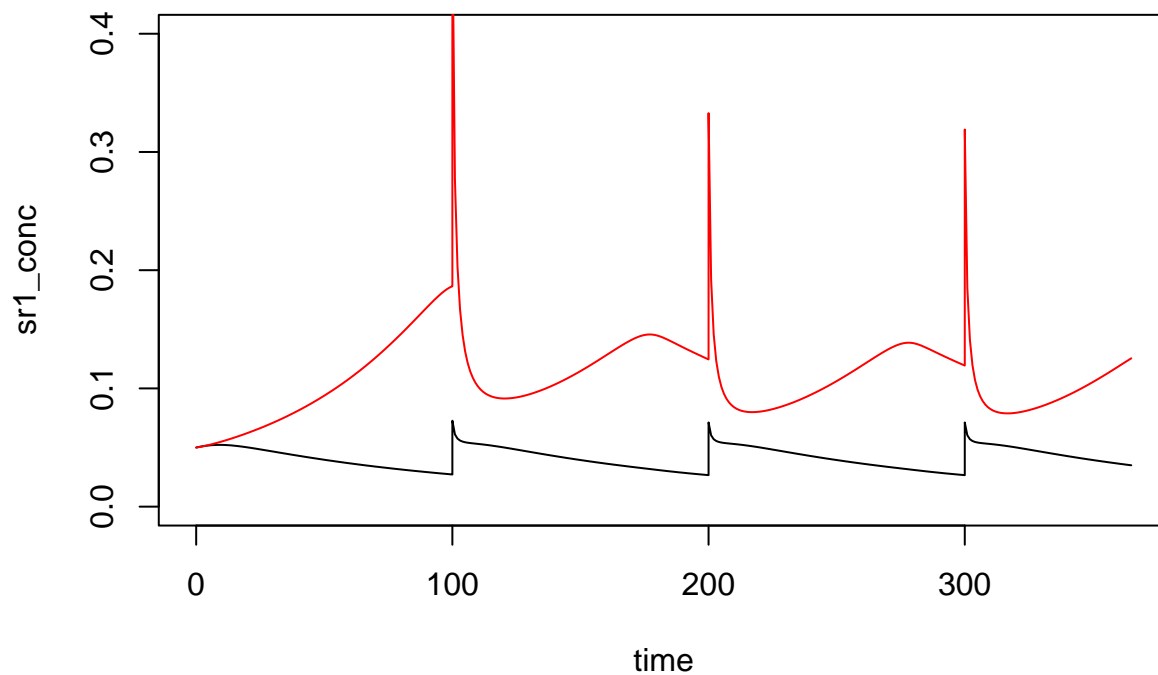


```
plot(NH4p ~ time, data = out6a)
```



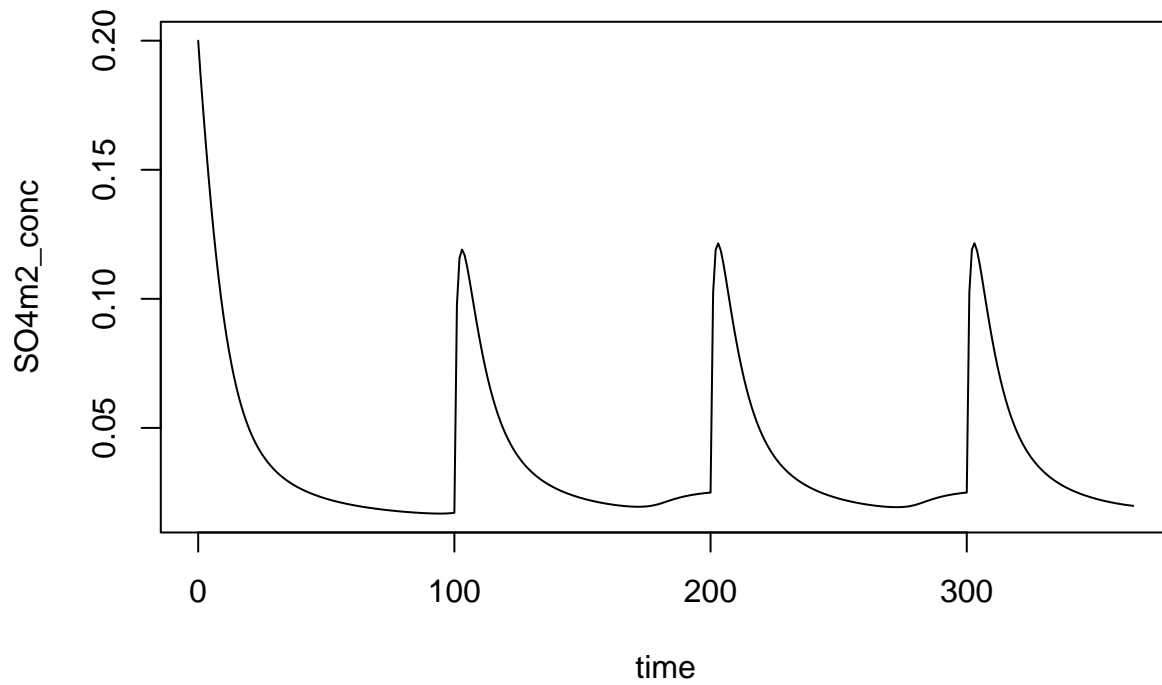
But now with sulfate and sulfide, sulfate reducers can grow.

```
plot(sr1_conc ~ time, data = out6a, type = 'l', ylim = c(0, 0.4))
lines(m1_conc ~ time, data = out6a, col = 'red')
```

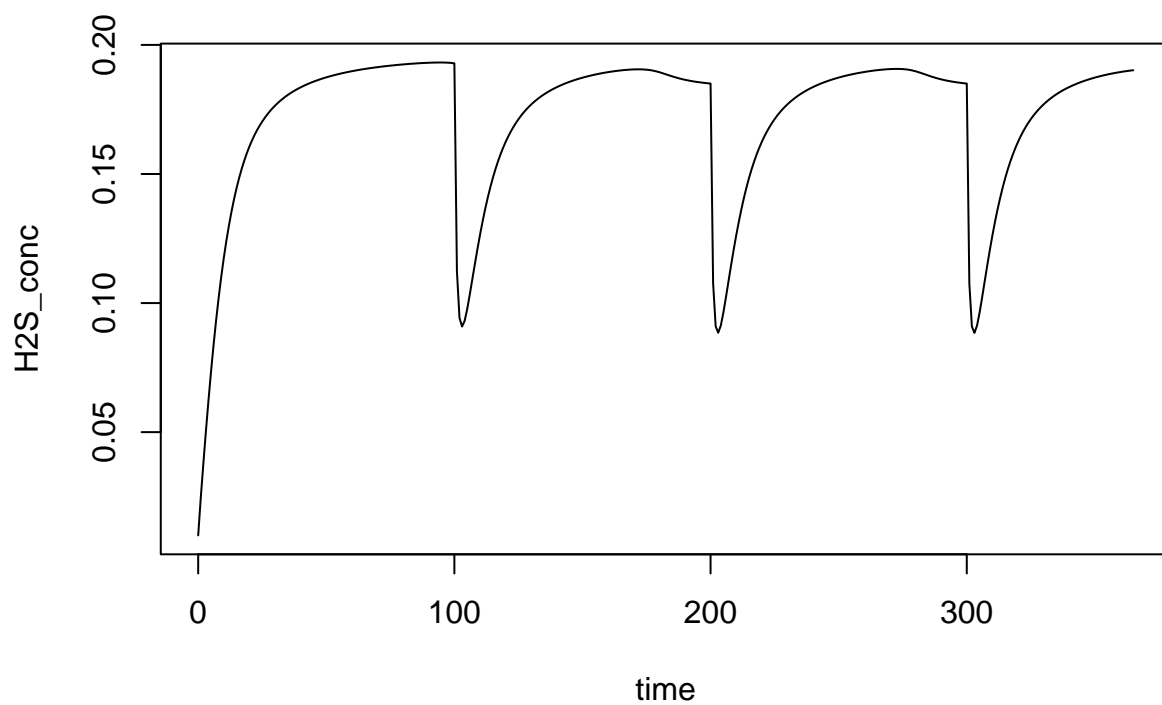


And if H₂S is a component, it will be produced.

```
plot(S04m2_conc ~ time, data = out6a, type = 'l')
```



```
plot(H2S_conc ~ time, data = out6a, type = 'l')
```



We can see dilution effects at if some washing water is added.

```
mng_pars6 = list(slurry_prod_rate = 10000,
                  slurry_mass = 1000,
                  storage_depth = 2,
                  resid_depth = 0.1,
                  area = 100,
                  empty_int = 100,
                  temp_C = 20,
```

```

        wash_water = 100000,
        wash_int = 100,
        rest_d = 0,
        resid_enrich = 1)

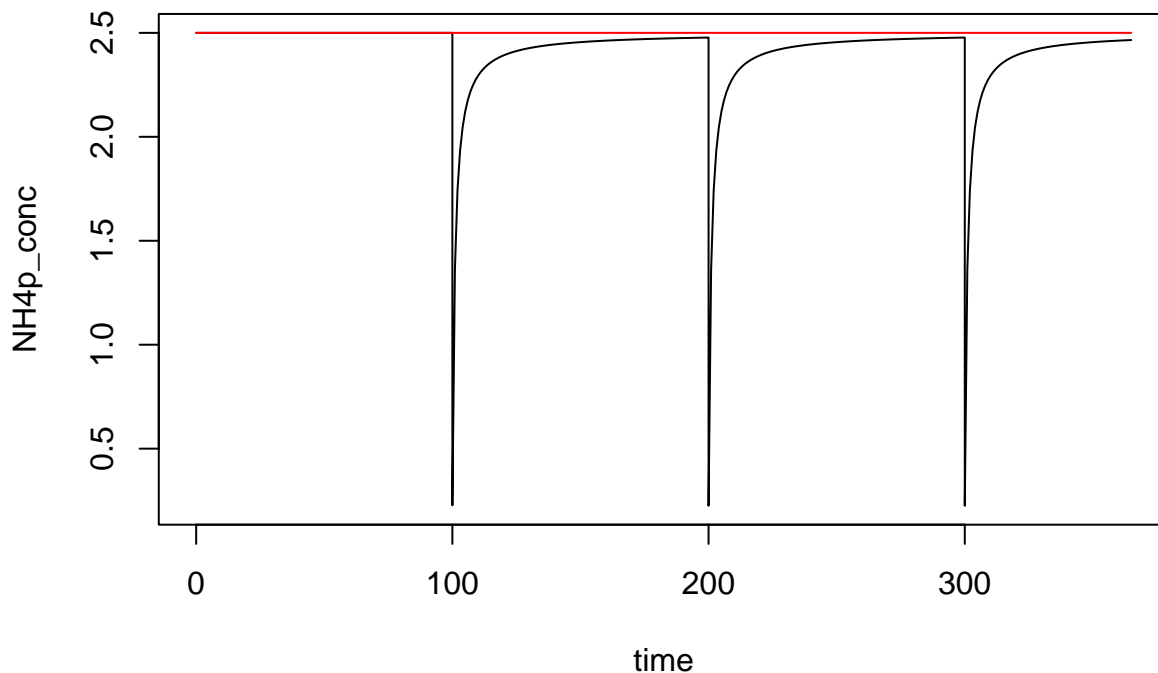
out6b <- abm(365,
  mng_pars = mng_pars6,
  man_pars = man_pars6,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars)

## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.

## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 32%

plot(NH4p_conc ~ time, data = out6b, type = 'l')
lines(NH4p_conc ~ time, data = out6a, col = 'red')

```



To include sulfate reduction, there must be an `sr` microbial group and `S04m2` must be a chemical component. Omit either to exclude sulfate reduction.

```

man_pars6c <- list(comps = c('H2S', 'NH4p'),
  comp_fresh = c(H2S = 0.01, NH4p = 2.5),
  VFA_fresh = c(CH3COOH = 2),
  pH = 7, dens = 1000)

grp_pars6c <- list(grps = c('m0', 'm1', 'm2'),
  yield = c(default = 0.05),
  xa_fresh = c(all = 0.05),
  xa_init = c(all = 0.05),
  dd_rate = c(all = 0.02),

```

```

ksv = c(default = 1),
qhat_opt = c(m0 = 1, m1 = 1, m2 = 2),
T_opt = c(m0 = 18, m1 = 18, m2 = 28),
T_min = c(m0 = 0, m1 = 6.41, m2 = 6.41),
T_max = c(m0 = 25, m1 = 25, m2 = 38))

```

```
devtools::load_all()
```

```
## i Loading ABM
```

```

out6c <- abm(365,
  mng_pars = mng_pars6,
  man_pars = man_pars6c,
  grp_pars = grp_pars6c,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars)

```

```

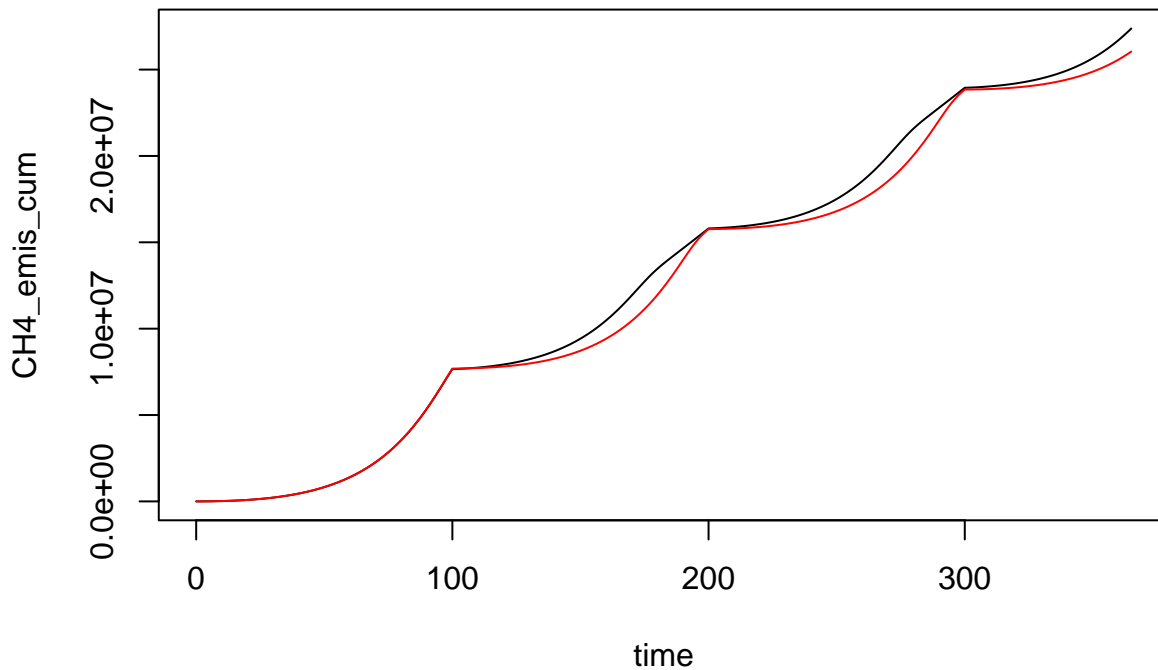
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 32%

```

```

plot(CH4_emis_cum ~ time, data = out6a, type = 'l')
lines(CH4_emis_cum ~ time, data = out6c, col = 'red')

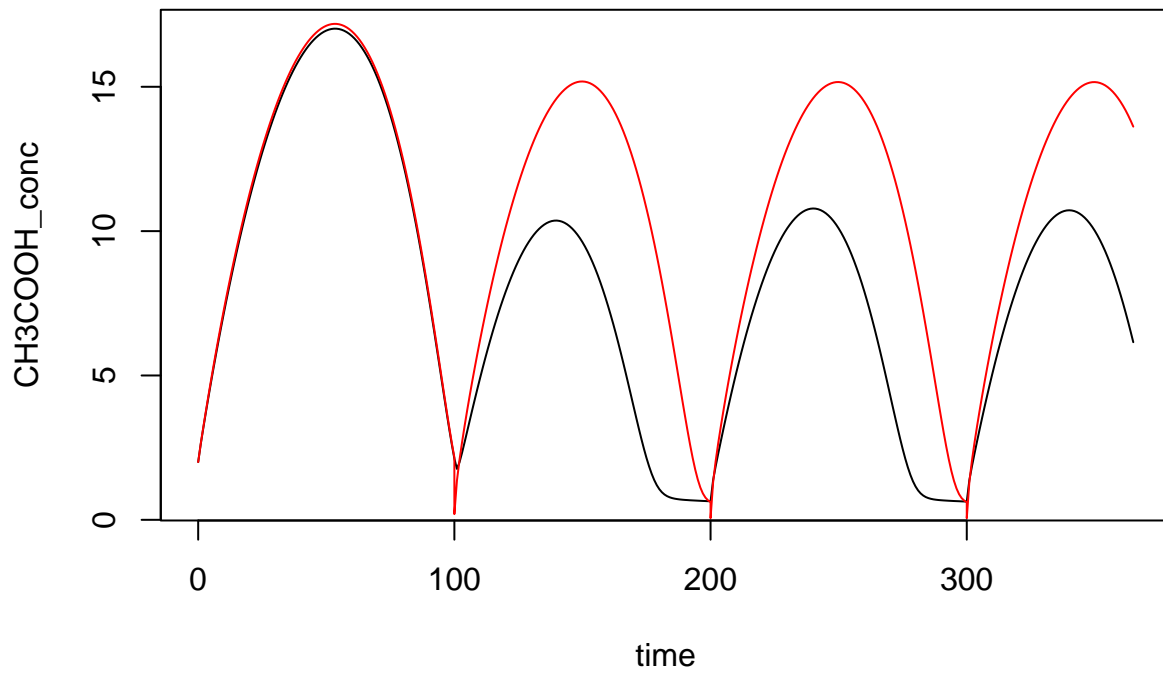
```



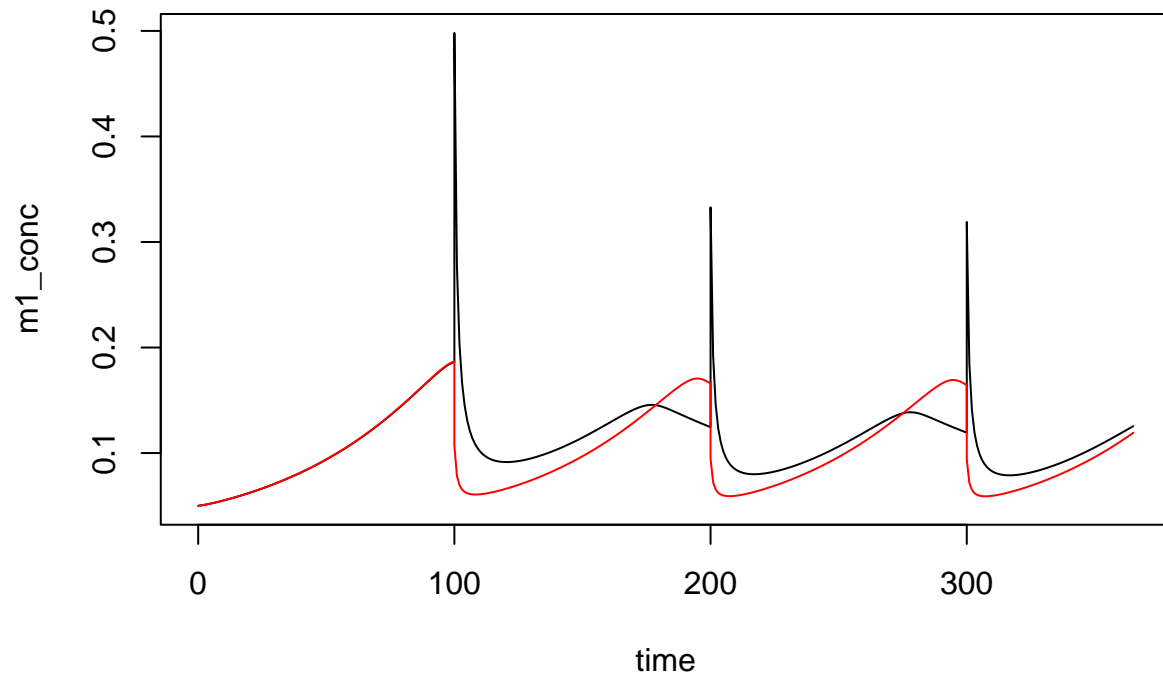
```

plot(CH3COOH_conc ~ time, data = out6a, type = 'l')
lines(CH3COOH_conc ~ time, data = out6c, col = 'red')

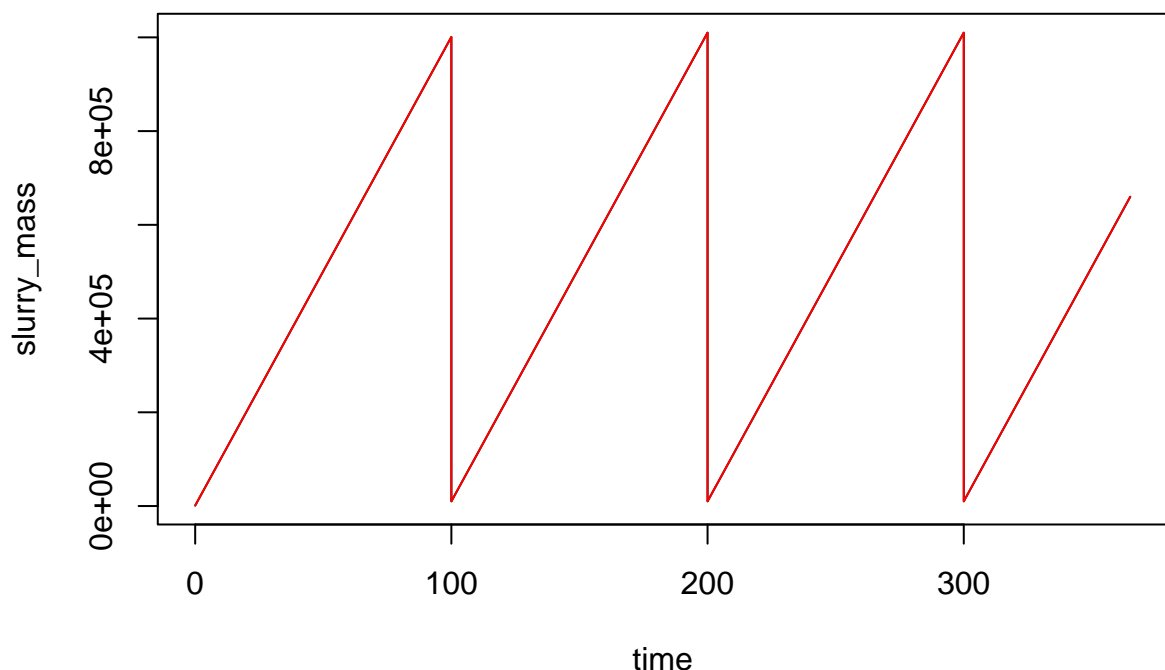
```



```
plot(m1_conc ~ time, data = out6a, type = 'l')
lines(m1_conc ~ time, data = out6c, col = 'red')
```



```
plot(slurry_mass ~ time, data = out6a, type = 'l')
lines(slurry_mass ~ time, data = out6c, col = 'red')
```



7. Speciation

Acid-base reactions are needed for inhibition and for CO₂ emission. They can be added for any component. The `chem_pars` argument can accept temperature-dependent log *ka* expressions. Use `temp_K` for absolute temperature in those expressions.

```
man_pars7 <- list(comps = c('H2S', 'NH4p'),
  comp_fresh = c(H2S = 0.01, NH4p = 2.5),
  VFA_fresh = c(CH3COOH = 2),
  pH = 7, dens = 1000)

chem_pars7 <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
  VFA = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
  CO2_aer = 1/0.436, CO2_sr = 1/1.2,
  C_xa = 1/0.3753125),
  specs = c('NH3', 'HSm', 'CH3COOm'),
  mspec = c(NH3 = 'NH4p', HSm = 'H2S', CH3COOm = 'CH3COOH'),
  lka = c(NH3 = '- 0.09046 - 2729.31/temp_K',
    HSm = '- 3448.7/temp_K + 47.479 - 7.5227 * log(temp_K)',
    CH3COOm = '- 4.8288 + 21.42/temp_K')
)
```

Here `comps` are the chemical “components”, or “master species”, as described a bit above. All are automatically included as chemical species in `packPars()`. In the `chem_pars` argument, `specs` are the other (non-master) species that the master species are in equilibrium with. And `mspec` are the associated master species. So the NH₃ species comes from NH₄⁺. When there is speciation, the master species are always taken as the protonated ones. So the species in `specs` always have one less proton than their associated master species. Only 2 species are supported for any component (master species and one more).

```
devtools::load_all()
```

```
## i Loading ABM
```

```
out7 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars6,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars7)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.

## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 2%
```

```
head(out7)
```

```
##      time      m0      m1      m2      sr1      VSd      H2S      S04m2
## 1      0  50.0000  50.0000  50.0000  50.0000  50000.0  10.0000  200.000
## 2      1  553.9977  553.8414  558.3574  555.0147  542318.5  268.9637  2041.036
## 3      2 1066.2080 1065.6056 1083.0938 1067.9792 1022077.4  788.7361  3621.264
## 4      3 1588.1329 1586.7628 1626.7495 1587.7198 1489599.2 1554.5640 4955.436
## 5      4 2120.8456 2118.3596 2191.3257 2112.6512 1945197.9 2545.3729 6064.627
## 6      5 2665.1817 2661.2072 2778.5446 2641.0154 2389179.5 3737.1169 6972.883
##      NH4p      CH3COOH slurry_mass CH4_emis_cum slurry_load COD_load CH4_emis_rate
## 1    2500    2000.00      1000      0.0000      0      0      25.52844
## 2   27500   28849.48     11000     163.4287     10000   522000     308.15048
## 3   52500   66757.26     21000     627.6723     20000  1044000     625.17882
## 4   77500  115293.28     31000    1422.3638     30000  1566000     968.06890
## 5  102500  174068.44     41000    2570.9586     40000  2088000    1332.46915
## 6  127500  242718.37     51000    4093.6840     50000  2610000    1716.05164
##      temp_C pH m0_eff m1_eff m2_eff sr1_eff VSd_eff H2S_eff S04m2_eff NH4p_eff
## 1      20  7      0      0      0      0      0      0      0      0
## 2      20  7      0      0      0      0      0      0      0      0
## 3      20  7      0      0      0      0      0      0      0      0
## 4      20  7      0      0      0      0      0      0      0      0
## 5      20  7      0      0      0      0      0      0      0      0
## 6      20  7      0      0      0      0      0      0      0      0
##      CH3COOH_eff slurry_mass_eff slurry_depth      m0_conc      m1_conc      m2_conc
## 1          0          0          0.01 0.05000000 0.05000000 0.05000000
## 2          0          0          0.11 0.05036343 0.05034921 0.05075976
## 3          0          0          0.21 0.05077181 0.05074312 0.05157589
## 4          0          0          0.31 0.05123009 0.05118590 0.05247579
## 5          0          0          0.41 0.05172794 0.05166731 0.05344697
## 6          0          0          0.51 0.05225847 0.05218053 0.05448127
##      sr1_conc VSd_conc      H2S_conc S04m2_conc NH4p_conc CH3COOH_conc m0_eff_conc
## 1 0.05000000 50.00000 0.01000000 0.2000000      2.5      2.000000      NaN
## 2 0.05045588 49.30168 0.02445125 0.1855488      2.5      2.622680      NaN
## 3 0.05085615 48.67035 0.03755886 0.1724411      2.5      3.178917      NaN
## 4 0.05121677 48.05159 0.05014723 0.1598528      2.5      3.719138      NaN
## 5 0.05152808 47.44385 0.06208226 0.1479177      2.5      4.245572      NaN
## 6 0.05178462 46.84666 0.07327680 0.1367232      2.5      4.759184      NaN
##      m1_eff_conc m2_eff_conc sr1_eff_conc VSd_eff_conc H2S_eff_conc S04m2_eff_conc
## 1      NaN      NaN      NaN      NaN      NaN      NaN
## 2      NaN      NaN      NaN      NaN      NaN      NaN
## 3      NaN      NaN      NaN      NaN      NaN      NaN
```



```
## 4      NaN      NaN      NaN      NaN      NaN      NaN
## 5      NaN      NaN      NaN      NaN      NaN      NaN
## 6      NaN      NaN      NaN      NaN      NaN      NaN
##  NH4p_eff_conc CH3COOH_eff_conc
## 1      NaN      NaN
## 2      NaN      NaN
## 3      NaN      NaN
## 4      NaN      NaN
## 5      NaN      NaN
## 6      NaN      NaN
```

But chemical species don't matter unless they are used in inhibition or emission.

8. Inhibition

Any chemical species can inhibit any microbial group. Inhibition parameters (currently initial and complete concentrations, with linear response, why not?) are entered in a matrix.

```
ilwr <- matrix(
  c(5, 0.2, 10, 0.5,
    5, 0.2, 10, 0.5,
    5, 0.2, 10, 0.5,
    5, 0.2, 10, 0.5),
  nrow = 4,
  byrow = TRUE,
  dimnames = list(
    c('m0', 'm1', 'm2', 'sr1'),
    c('NH4p', 'NH3', 'CH3COOm', 'CH3COOH')
  )
)

iupr <- matrix(
  c(9, 0.9, 30, 1,
    9, 0.9, 30, 1,
    9, 0.9, 30, 1,
    9, 0.9, 30, 1),
  nrow = 4,
  byrow = TRUE,
  dimnames = list(
    c('m0', 'm1', 'm2', 'sr1'),
    c('NH4p', 'NH3', 'CH3COOm', 'CH3COOH')
  )
)

inhib_pars <- list(
  ilwr = ilwr,
  iupr = iupr
)

inhib_pars

## $ilwr
##      NH4p NH3 CH3COOm CH3COOH
## m0      5 0.2      10      0.5
## m1      5 0.2      10      0.5
```

```

## m2      5 0.2      10      0.5
## sr1     5 0.2      10      0.5
##
## $iupr
##      NH4p NH3 CH3COOm CH3COOH
## m0      9 0.9      30      1
## m1      9 0.9      30      1
## m2      9 0.9      30      1
## sr1     9 0.9      30      1

man_pars8 <- list(comps = c('H2S', 'NH4p'),
                  comp_fresh = c(H2S = 0.01,
                                NH4p = 2.5),
                  VFA_fresh = c(CH3COOH = 2),
                  pH = 7, dens = 1000)

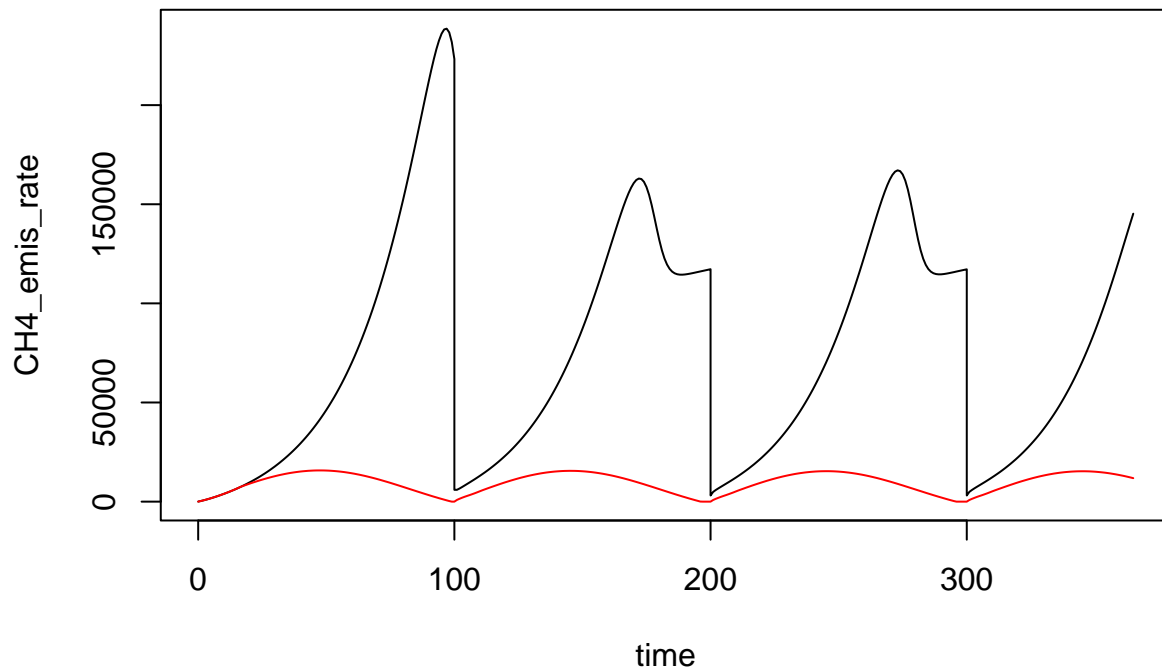
chem_pars8 <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
                                CH3COOH = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
                                CO2_aer = 1/0.436, CO2_sr = 1/1.2,
                                C_xa = 1/0.3753125),
                  specs = c('NH3', 'HSm', 'CH3COOm'),
                  mspec = c(NH3 = 'NH4p', HSm = 'H2S', CH3COOm = 'CH3COOH'),
                  lka = c(NH3 = '- 0.09046 - 2729.31/temp_K',
                          HSm = '- 3448.7/temp_K + 47.479 - 7.5227* log(temp_K)',
                          CH3COOm = '-4.8288 + 21.42/temp_K')
)

out8 <- abm(365,
            mng_pars = mng_pars,
            man_pars = man_pars7,
            grp_pars = grp_pars,
            mic_pars = mic_pars,
            sub_pars = sub_pars,
            chem_pars = chem_pars7,
            inhib_pars = inhib_pars
)

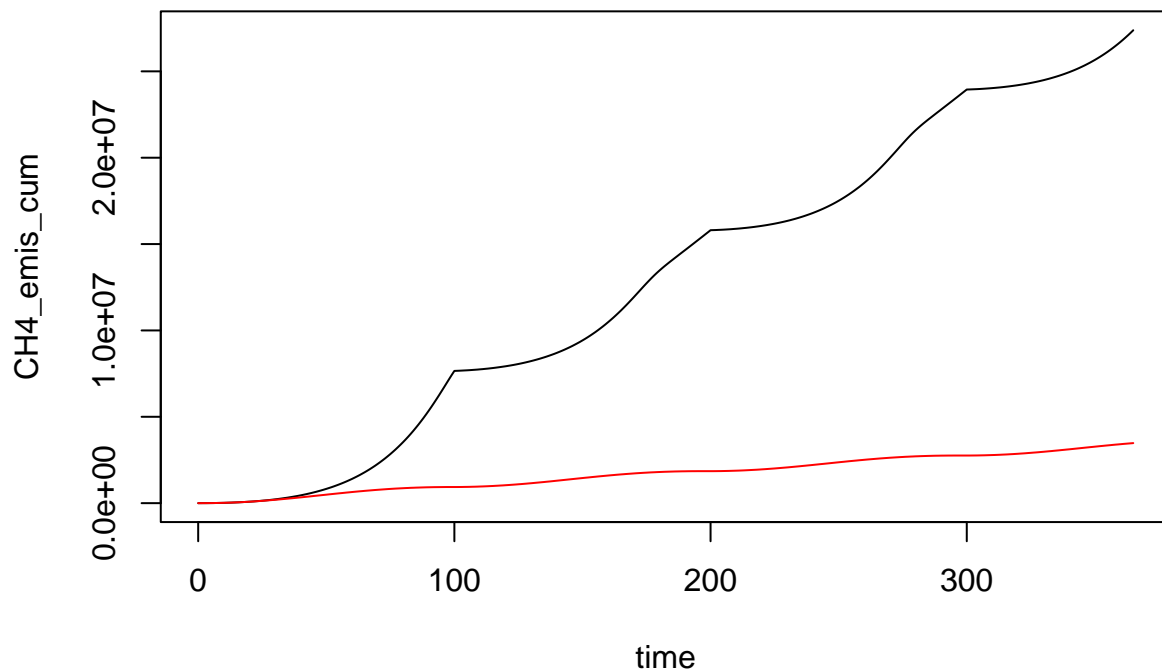
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.

plot(CH4_emis_rate ~ time, data = out7, type = 'l')
lines(CH4_emis_rate ~ time, data = out8, col = 'red')

```



```
plot(CH4_emis_cum ~ time, data = out7, type = 'l')
lines(CH4_emis_cum ~ time, data = out8, col = 'red')
```



9. Volatilization

Any chemical species can volatilize.

```
man_pars9 <- list(comps = c('H2S', 'NH4p'),
  comp_fresh = c(H2S = 0.01, NH4p = 2.5),
  VFA_fresh = c(CH3COOH = 2),
  pH = 7, dens = 1000)
```

We need to set mass transfer coefficient values (m/d) for any species that volatilizes. These are overall (possibly two-film) values for in liquid-phase units.

```
chem_pars9 <- list(COD_conv = c(CH4 = 1/0.2507),
  specs = c('NH3', 'HSm', 'CH3COOm'),
  mspec = c(NH3 = 'NH4p', HSm = 'H2S', CH3COOm = 'CH3COOH'),
  lka = c(NH3 = '- 0.09046 - 2729.31/temp_K',
    HSm = '- 3448.7/temp_K + 47.479 - 7.5227 * log(temp_K)',
    CH3COOm = '-4.8288 + 21.42/temp_K'),
  kl = c(NH3 = 0.01, H2S = 0.01, CH3COOH = 0.01) * 86400)
```

```
devtools::load_all()
```

```
## i Loading ABM
```

```
out9a <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars9,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars9,
  inhib_pars = inhib_pars
)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
```

```
## Size-variable parameter problem: Missing element(s) in kss.
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
```

```
## pars$COD_conv, : COD balance is off by 2%
```

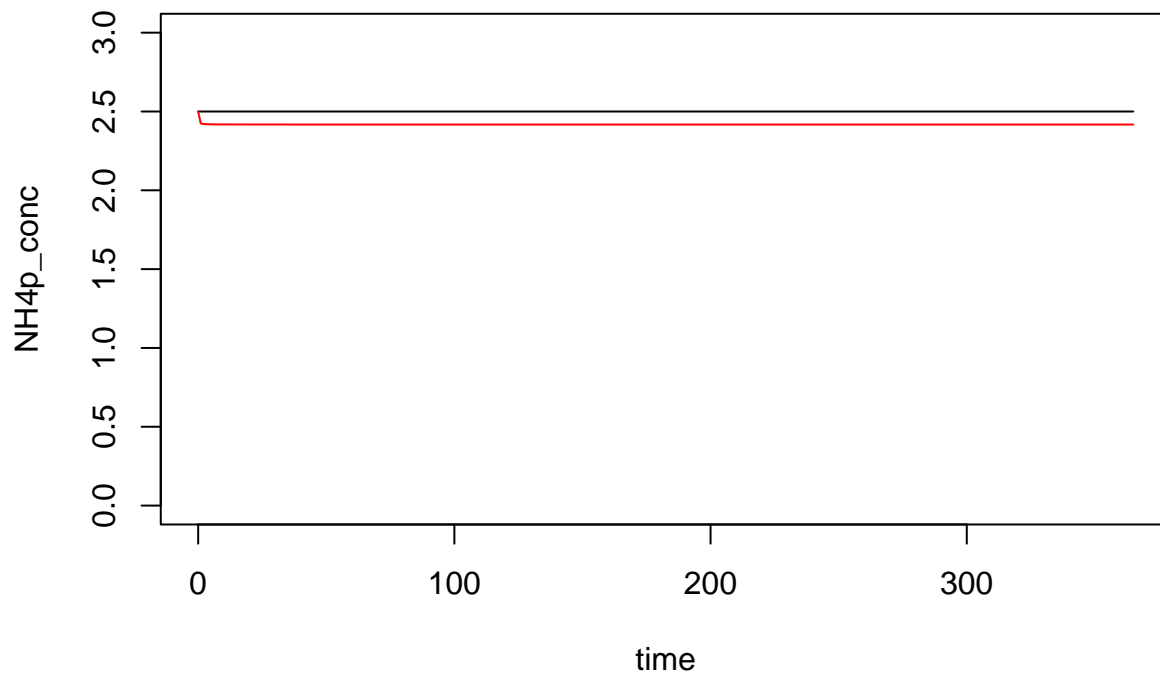
The state variable vector and output data frame automatically expand for the new values.

```
head(out9a[, grep('_emis', names(out9a))])
```

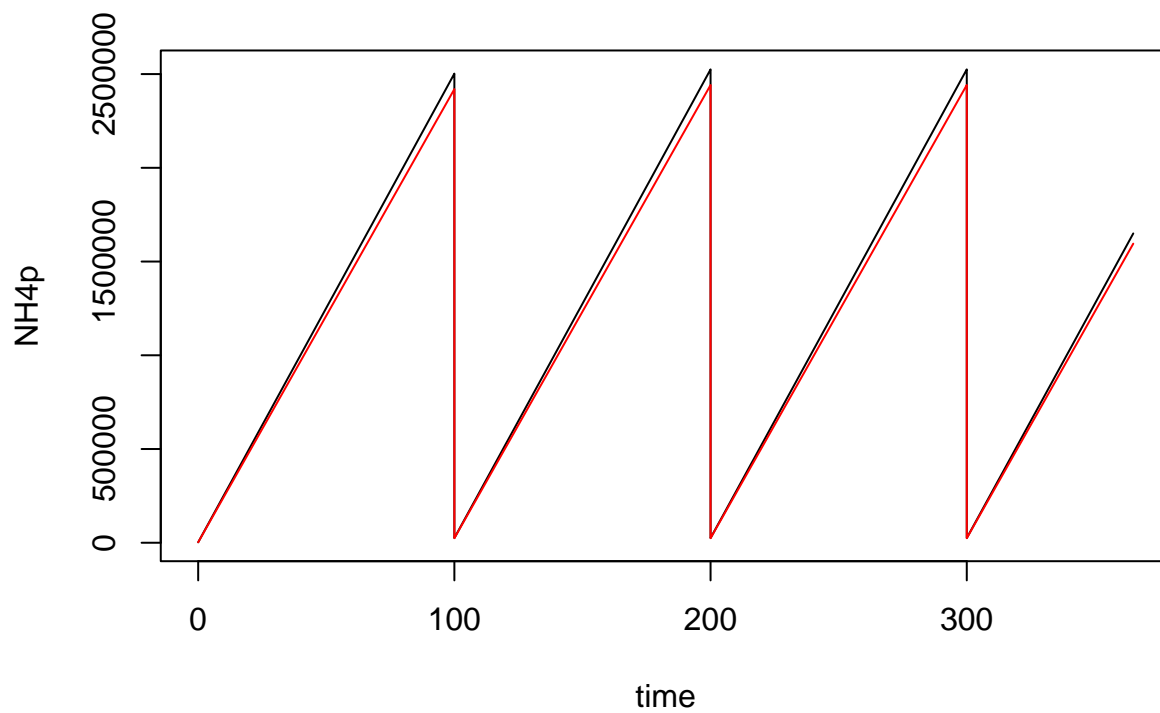
```
##   CH4_emis_cum NH3_emis_cum H2S_emis_cum CH3COOH_emis_cum CH4_emis_rate
## 1      0.0000      0.000      0.0000      0.000      25.52844
## 2     161.8334     833.273     89.6908     1106.072     305.28189
## 3     622.3551    1661.702    171.2283     2484.484     620.73997
## 4    1412.0784    2489.451    252.7655     4130.203     962.63333
## 5    2554.8558    3316.914    334.3028     6036.464    1326.29785
## 6    4071.0922    4144.219    415.8401     8196.713    1709.25431
```

```
plot(NH4p_conc ~ time, data = out8, type = 'l', ylim = c(0, 3))
```

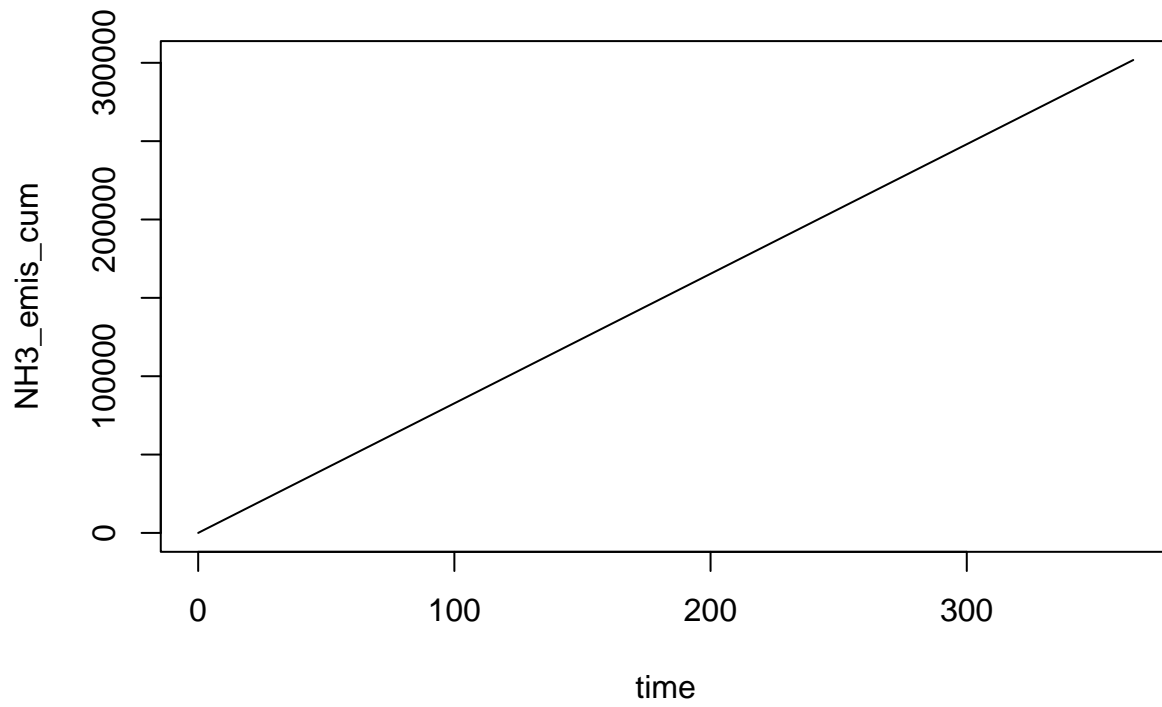
```
lines(NH4p_conc ~ time, data = out9a, col = 'red')
```



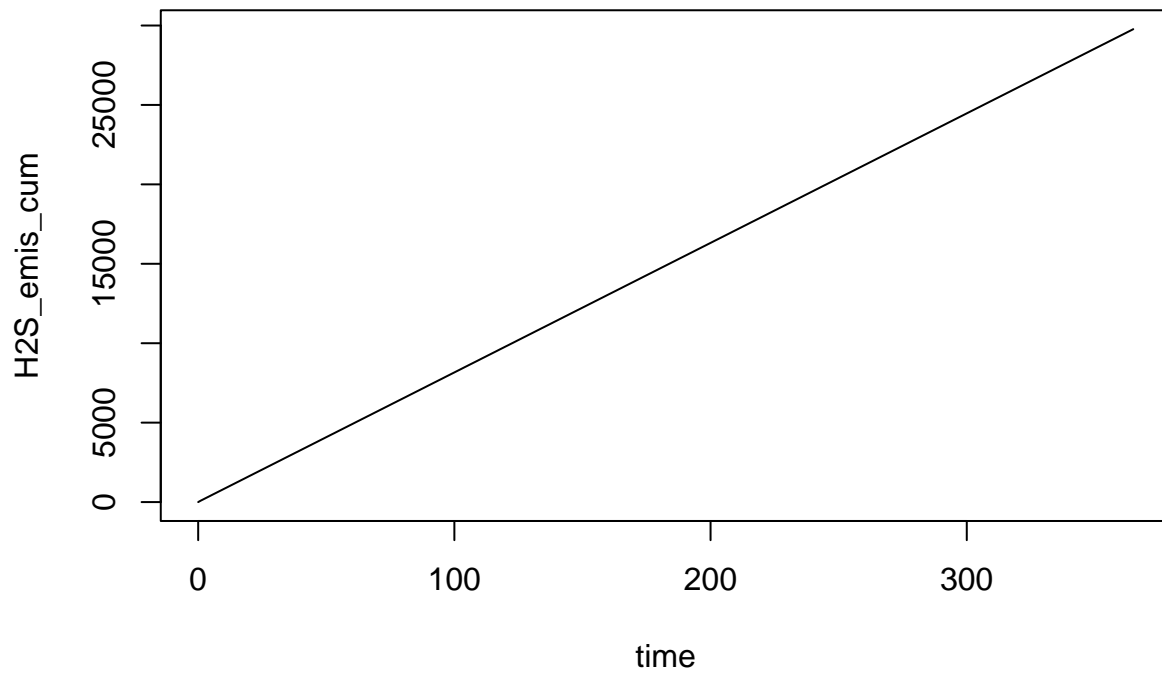
```
plot(NH4p ~ time, data = out8, type = 'l')  
lines(NH4p ~ time, data = out9a, col = 'red')
```



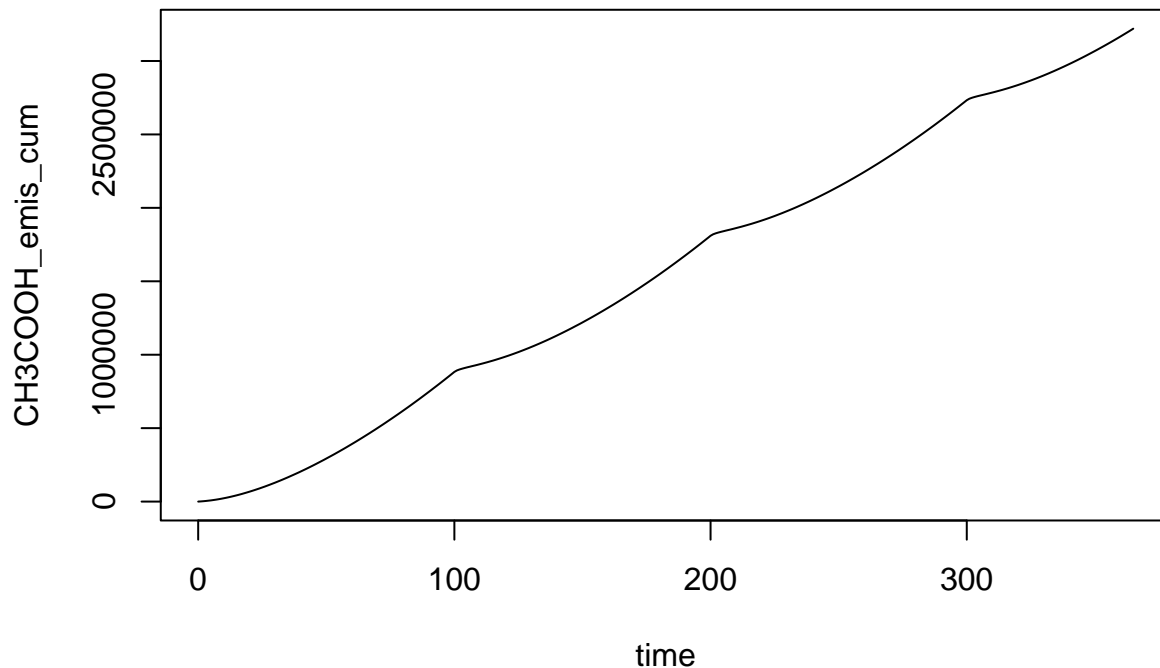
```
plot(NH3_emis_cum ~ time, data = out9a, type = 'l')
```



```
plot(H2S_emis_cum ~ time, data = out9a, type = 'l')
```



```
plot(CH3COOH_emis_cum ~ time, data = out9a, type = 'l')
```



Let's use a fixed slurry mass to exaggerate emission.

```
mng_pars9b = list(slurry_prod_rate = 0,
                  slurry_mass = 1E6,
                  storage_depth = 2,
                  resid_depth = 0.1,
                  area = 100,
                  empty_int = 100,
                  temp_C = 20,
                  wash_water = 0,
                  wash_int = NA,
                  rest_d = 0,
                  resid_enrich = 1)

chem_pars9b <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
                                VFA = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
                                CO2_aer = 1/0.436, CO2_sr = 1/1.2,
                                C_xa = 1/0.3753125),
                  specs = c('NH3', 'HSm', 'CH3COOm'),
                  mspec = c(NH3 = 'NH4p', HSm = 'H2S', CH3COOm = 'CH3COOH'),
                  lka = c(NH3 = '- 0.09046 - 2729.31/temp_K',
                          HSm = '- 3448.7/temp_K + 47.479 - 7.5227 * log(temp_K)',
                          CH3COOm = '-4.8288 + 21.42/temp_K'),
                  kl = c(NH3 = 0.01, H2S = 0.01) * 86400)

devtools::load_all()

## i Loading ABM

out9b <- abm(365,
             mng_pars = mng_pars9b,
             man_pars = man_pars9,
             grp_pars = grp_pars,
             mic_pars = mic_pars,
```

```

    sub_pars = sub_pars,
    chem_pars = chem_pars9b,
    inhib_pars = inhib_pars
)

```

```

## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.

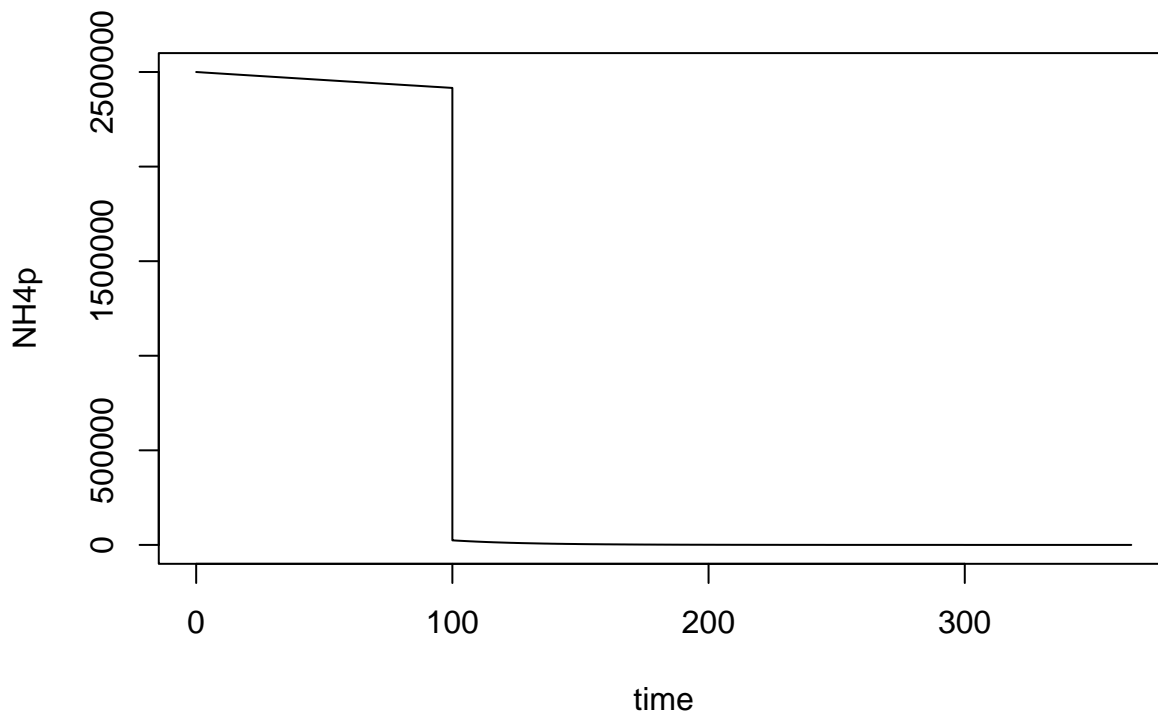
## Warning in emptyStore(y, resid_mass = pars$resid_mass, resid_enrich =
## pars$resid_enrich): Emptying skipped.
## Warning in emptyStore(y, resid_mass = pars$resid_mass, resid_enrich =
## pars$resid_enrich): Emptying skipped.

```

```

plot(NH4p ~ time, data = out9b, type = 'l')

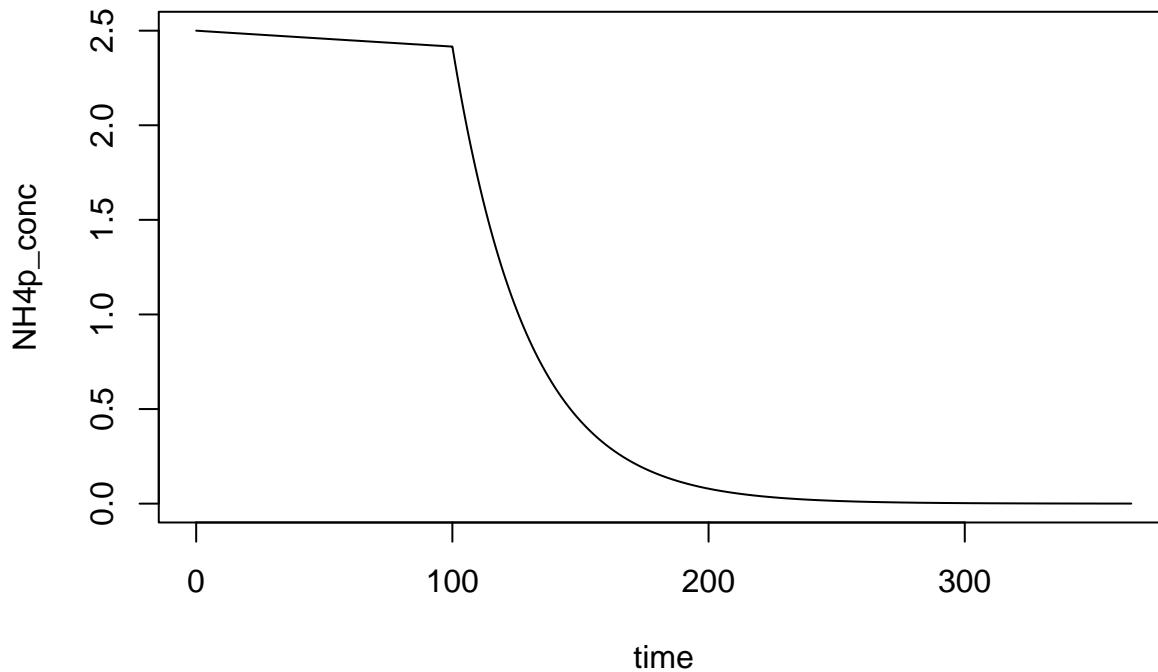
```



```

plot(NH4p_conc ~ time, data = out9b, type = 'l')

```

9. COD balance

There is now a `checkCOD()` function that runs on `abm()` results before returning them. For now the tolerance is fixed at 1%. Some of the examples above do not meet that criterion for some reason. At least one shows a real problem that needs to be identified. For the emission example above, the problem is that VFA is emitted but that loss is not included in the balance check (this might have been fixed). I need to decide about how to pass that COD information. We can make it worse by pretending the the charged form can volatilize (VFA changed to `CH3COOm` below).

```
chem_pars10 <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
                                VFA = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
                                CO2_aer = 1/0.436, CO2_sr = 1/1.2,
                                C_xa = 1/0.3753125),
  specs = c('NH3', 'HSm', 'CH3COOm'),
  mspec = c(NH3 = 'NH4p', HSm = 'H2S', CH3COOm = 'CH3COOH'),
  lka = c(NH3 = '- 0.09046 - 2729.31/temp_K',
          HSm = '- 3448.7/temp_K + 47.479 - 7.5227 * log(temp_K)',
          CH3COOm = '-4.8288 + 21.42/temp_K'),
  kl = c(NH3 = 0.01, H2S = 0.01, CH3COOm = 0.01) * 86400)
```

```
out10 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars9,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars,
  chem_pars = chem_pars10,
  inhib_pars = inhib_pars
)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 36%
```

10. Stoichiometry and nitrogen mineralization

Now substrates can produce any amount of arbitrary components (defined in `man_pars`, possibly volatilized, possibly involved in speciation in inhibition) through hydrolysis and fermentation to VFA.

```
man_pars10 <- list(comps = c('H2S', 'SO4m2', 'NH4p'),
  comp_fresh = c(H2S = 0.01, SO4m2 = 0.2, NH4p = 2.5),
  VFA_fresh = c(CH3COOH = 2),
  pH = 7, dens = 1000)
```

(Hmm, should `comps` be moved to `chem_pars`?)

Here we'll have 4 substrates. But substrates need not actually produce VFA anymore.

```
sub_pars10 <- list(subs = c('cellulose', 'protein', 'lipids', 'urea'),
  T_opt_hyd = c(all = 60),
  T_min_hyd = c(all = 0),
  T_max_hyd = c(all = 90),
  hydrol_opt = c(lipids = 0.1, protein = 0.01, cellulose = 0.05, urea = 1),
  sub_fresh = c(lipids = 3, protein = 20, cellulose = 35, urea = 10),
  sub_init = c(lipids = 3, protein = 20, cellulose = 35, urea = 10))
```

Production of any component is set in the `stoich` element of the `chem_pars` argument.

```
smat <- matrix(c(0, 0.2, 0, 0.2,
  0, 0.01, 0, 0,
  1, 1, 1, 0),
  nrow = 3,
  byrow = TRUE,
  dimnames = list(
    c('NH4p', 'H2S', 'CH3COOH'),
    c('cellulose', 'protein', 'lipids', 'urea')))
```

smat

```
##      cellulose protein lipids urea
## NH4p         0    0.20      0  0.2
## H2S          0    0.01      0  0.0
## CH3COOH      1    1.00      1  0.0
```

Substrate and other component quantities are

1. COD mass, or if COD = 0,
2. N mass, or if N = 0,
3. C mass, or if C = 0,
4. S mass, or if S = 0,
5. total mass

So the `CH3COOH` row should only have 1 or 0.

The `stoich` matrix can be calculated from substrate chemical formulas—see next example.

```
chem_pars10 <- list(COD_conv = c(CH4 = 1/0.2507, xa = 1/0.7069561,
  VFA = 1/0.9383125, S = 1/0.5015, VS = 1/0.69,
  CO2_aer = 1/0.436, CO2_sr = 1/1.2,
  C_xa = 1/0.3753125),
```

```

specs = c('NH3', 'HSm', 'CH3COOm'),
mspec = c(NH3 = 'NH4p', HSm = 'H2S', CH3COOm = 'CH3COOH'),
stoich = smat)

```

```
devtools::load_all()
```

```
## i Loading ABM
```

```

out10 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars10,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars10,
  chem_pars = chem_pars10)

```

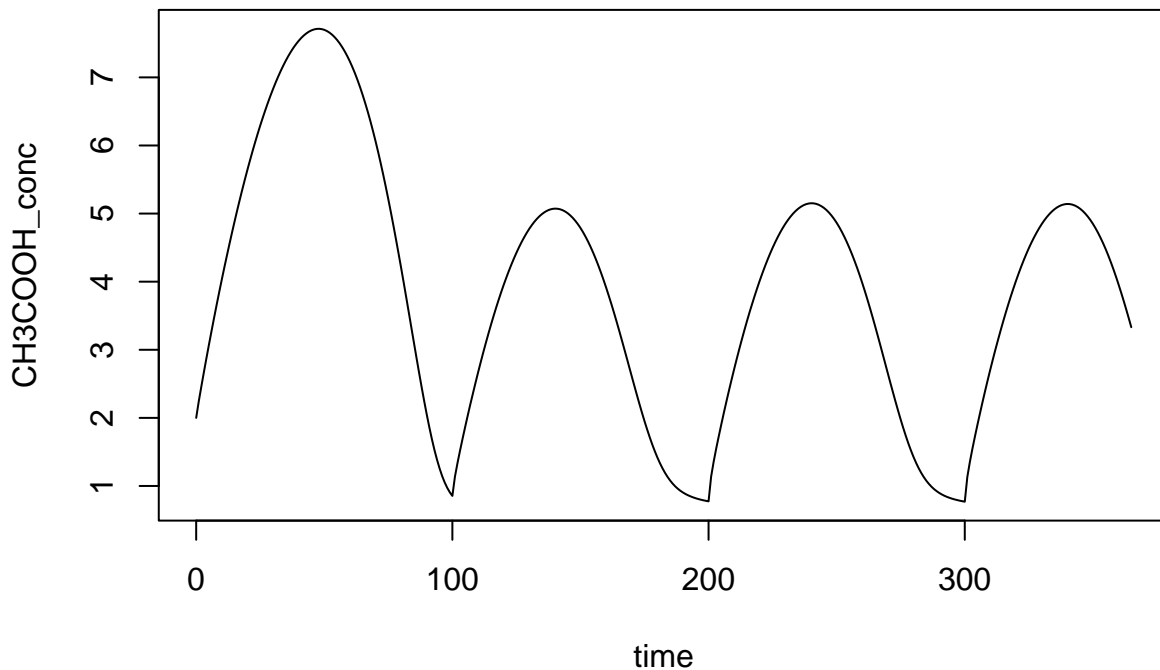
```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
```

```
## Size-variable parameter problem: Missing element(s) in kss.
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
```

```
## pars$COD_conv, : COD balance is off by 1.2%
```

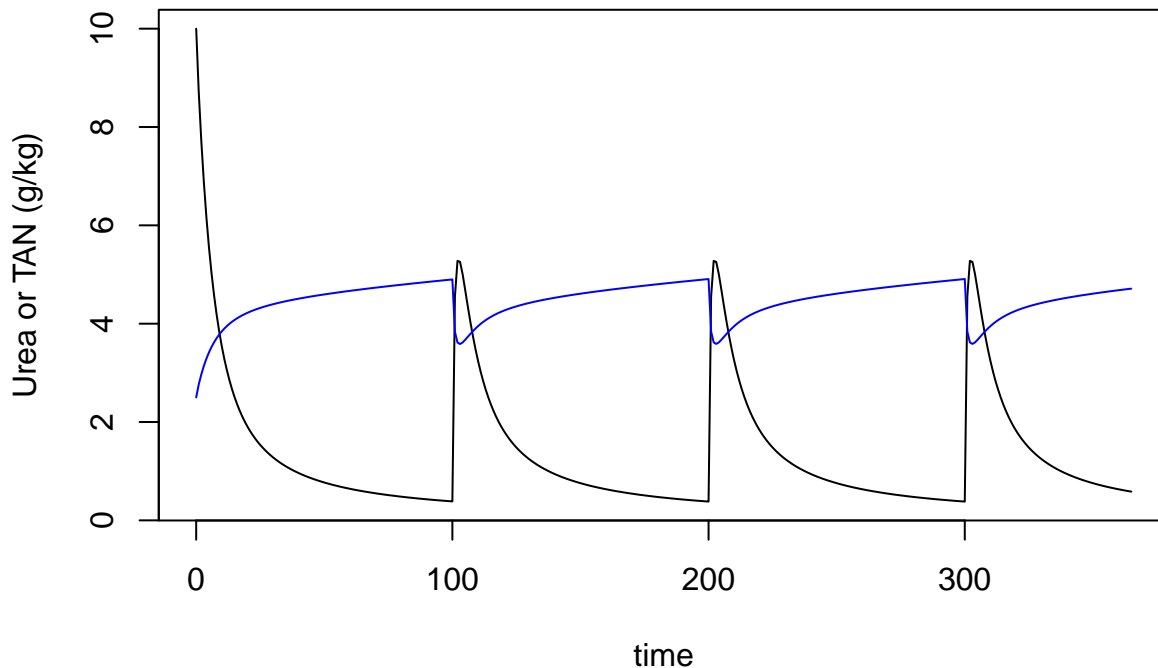
```
plot(CH3COOH_conc ~ time, data = out10, type = 'l')
```



```

plot(urea_conc ~ time, data = out10, type = 'l', ylab = 'Urea or TAN (g/kg)')
lines(NH4p_conc ~ time, data = out10, col = 'blue')

```



It might make more sense to have `stoich` calculated internally. For this, substrate formulas must be provided.

```
sub_pars10b <- list(subs = c('cellulose', 'protein', 'lipids', 'urea'),
  forms = c(cellulose = 'C6H10O5', protein = 'C4 H6.1 O1.2 N',
    lipids = 'C57 H104 O6', urea = 'CO(NH2)2'),
  T_opt_hyd = c(all = 60),
  T_min_hyd = c(all = 0),
  T_max_hyd = c(all = 90),
  hydrol_opt = c(lipids = 0.1, protein = 0.01, cellulose = 0.05, urea = 1),
  sub_fresh = c(lipids = 3, protein = 20, cellulose = 35, urea = 10),
  sub_init = c(lipids = 3, protein = 20, cellulose = 35, urea = 10))
```

Internally, the `getStoich()` function is used, which in turn calls `predFerm()`. Among these substrates, cellulose produces no CO₂ from fermentation, protein and lipids *consume* CO₂ (they are highly reduced), and urea is a special case with no COD. So it produces no VFAs.

```
predFerm(sub_pars10b$forms[1])
```

```
##      H2O CH3COOH
##      -1      3
```

```
predFerm(sub_pars10b$forms[2])
```

```
##      H2O      CO2 CH3COOH      NH3
## -2.6250 -0.1750  2.0875  1.0000
```

```
predFerm(sub_pars10b$forms[3])
```

```
##      H2O      CO2 CH3COOH
##      -28      -23      40
```

```
predFerm(sub_pars10b$forms[4])
```

```
##      CO2      NH3      H.      H2O CH3COOH      H2
##       1       2       0      -1       0       0
```

Internally, the stoichiometric coefficients are changed to the same units given above (COD, N, C, S, or total

mass). Now we must have CO2 as a component, because it will be produced.

```
man_pars10b <- list(comps = c('H2S', 'SO4m2', 'NH4p', 'CO2'),
  comp_fresh = c(H2S = 0.01, SO4m2 = 0.2, NH4p = 2.5, CO2 = 1),
  VFA_fresh = c(CH3COOH = 2),
  pH = 7, dens = 1000)
```

We don't actually need the species included above. But because of a bit of a programming quirk, we need to provide the master species for NH3.

```
chem_pars10b <- list(COD_conv = c(CH4 = 1/0.2507),
  specs = c('NH3'),
  mspec = c(NH3 = 'NH4p'),
  stoich = 'calc')
```

```
devtools::load_all()
```

```
## i Loading ABM
```

```
out10b <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars10b,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars10b,
  chem_pars = chem_pars10b)
```

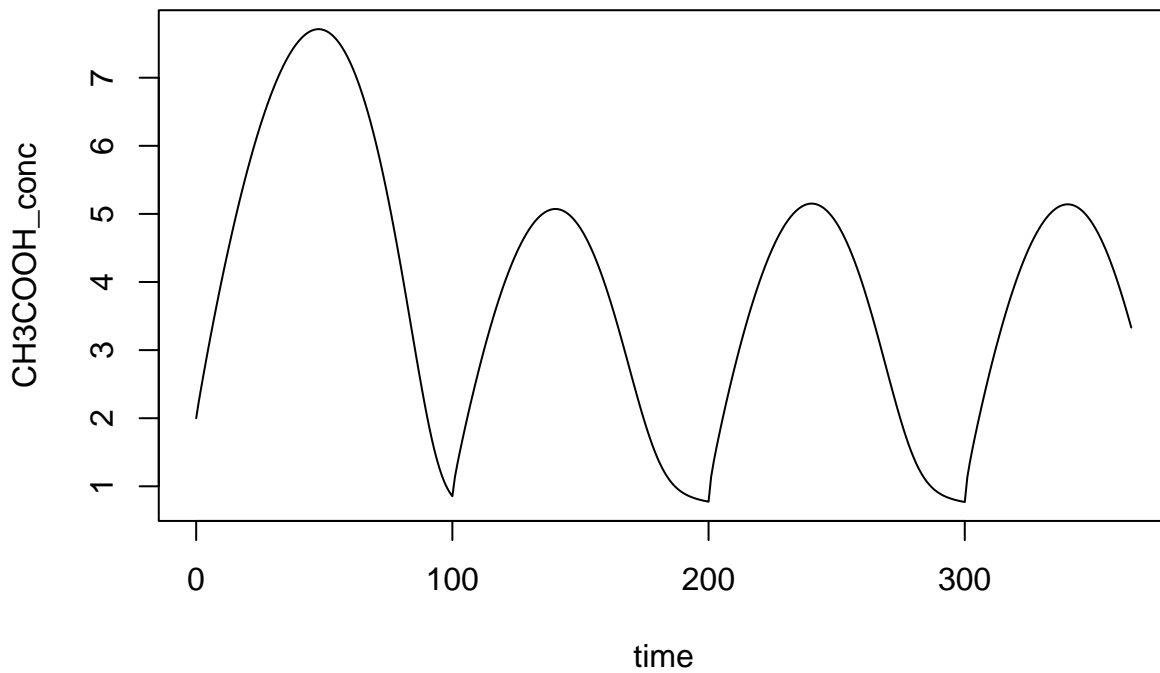
```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
```

```
## Size-variable parameter problem: Missing element(s) in kss.
```

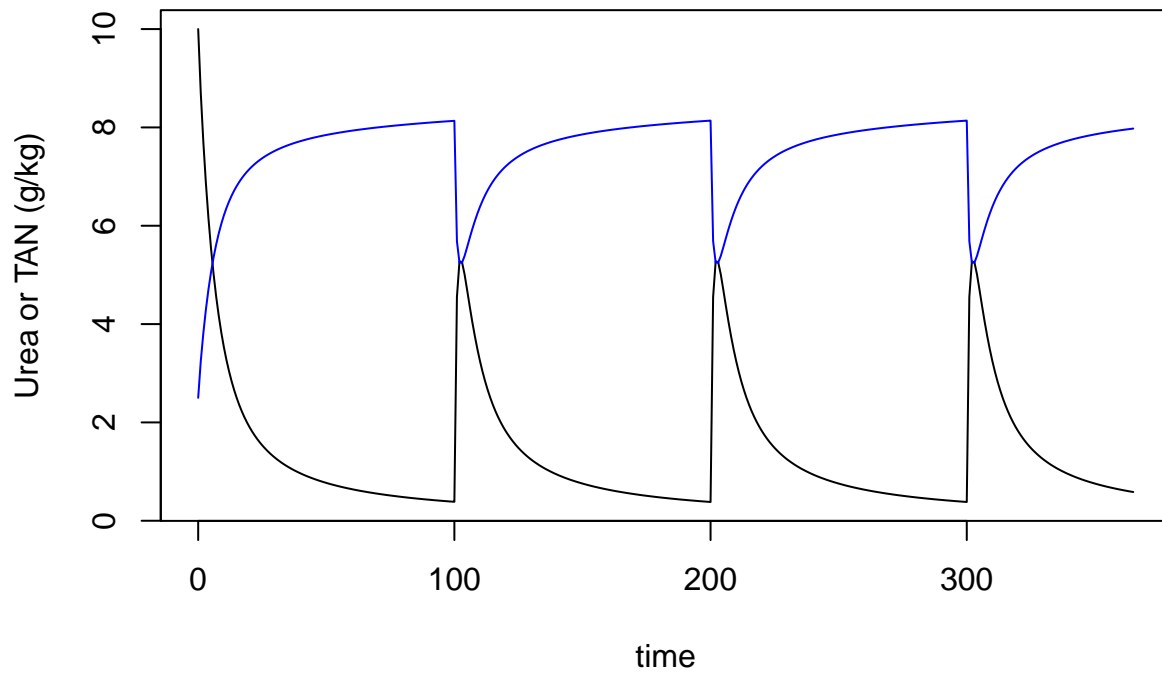
```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
```

```
## pars$COD_conv, : COD balance is off by 1.2%
```

```
plot(CH3COOH_conc ~ time, data = out10b, type = 'l')
```

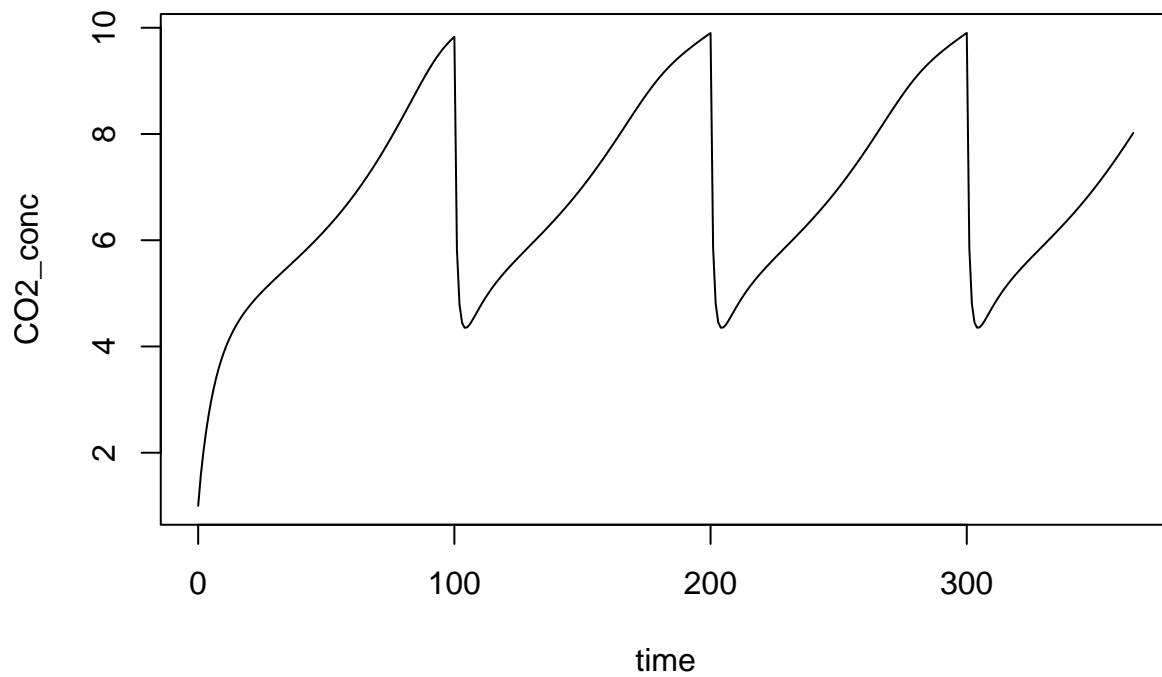


```
plot(urea_conc ~ time, data = out10b, type = 'l', ylab = 'Urea or TAN (g/kg)')
lines(NH4p_conc ~ time, data = out10b, col = 'blue')
```



Now we also have dissolved CO₂ (really TIC) concentration. Only there is no emission, so it does not mean much.

```
plot(CO2_conc ~ time, data = out10b, type = 'l')
```



11. CO2 emission

CO2 emission can be included through the volatilization route. It can be produced from both fermentation and methanogenesis. Speciation of dissolved CO2 (really H2CO3*) and HCO3- needs to be included (although a reduced mass transfer coefficient could achieve the same effect). So far, CO2 emission behavior is troublesome—it is difficult to produce plausible results.

```
sub_pars11 <- list(subs = c('VSd'),
  forms = c(VSd = 'C23H37O14N (CO2)1.5'),
  T_opt_hyd = c(all = 60),
  T_min_hyd = c(all = 0),
  T_max_hyd = c(all = 90),
  hydrol_opt = c(all = 0.1),
  sub_fresh = c(VSd = 20),
  sub_init = c(VSd = 20))
```

```
man_pars11 <- list(comps = c('CO2', 'NH4p'),
  comp_fresh = c(CO2 = 1, NH4p = 2.5),
  VFA_fresh = c(CH3COOH = 2),
  pH = 7, dens = 1000)
```

Here we will define speciation only for TIC. There is no way to include CO3-2 as a third species.

```
chem_pars11 <- list(COD_conv = c(CH4 = 1/0.2507),
  specs = c('HCO3m'),
  mspec = c(HCO3m = 'CO2', NH3 = 'NH4p'),
  lka = c(HCO3m = '-2.778 + -353.5305 -0.06092*temp_K + 21834.37/temp_K + 126.8339*1',
  stoich = 'calc',
  kl = c(CO2 = 0.01) * 86400)
```

```
devtools::load_all()
```

```
## i Loading ABM
```

```
out11 <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars11,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars11,
  chem_pars = chem_pars11)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
```

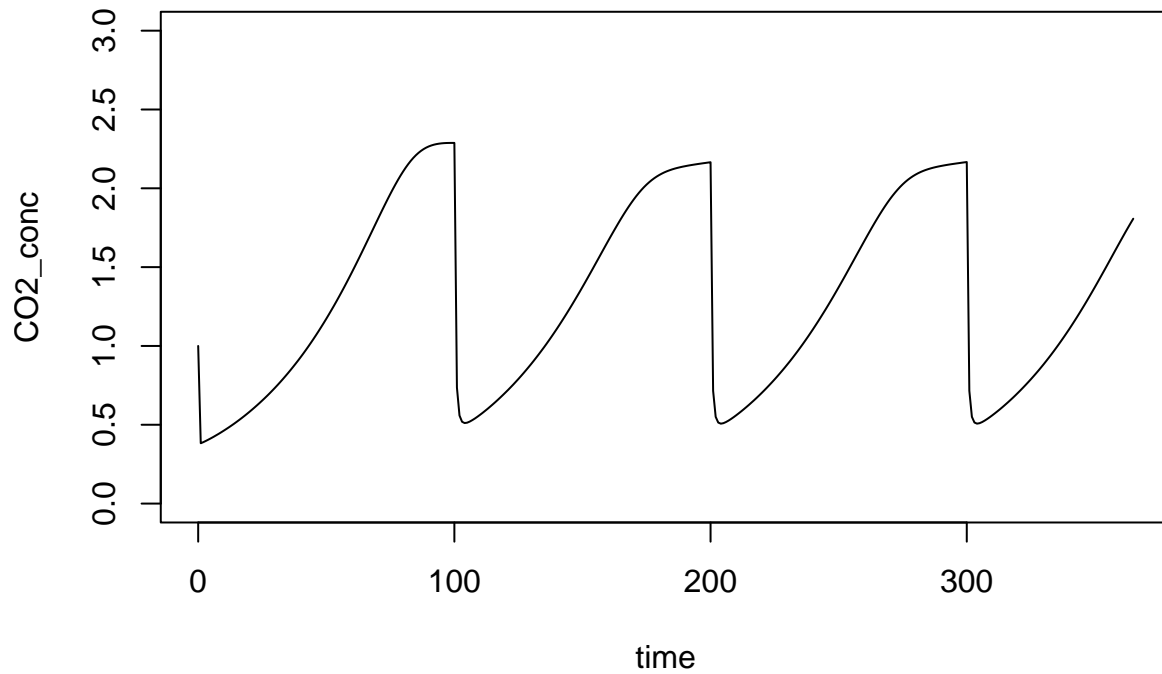
```
## Size-variable parameter problem: Missing element(s) in kss.
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
```

```
## pars$COD_conv, : COD balance is off by 1.8%
```

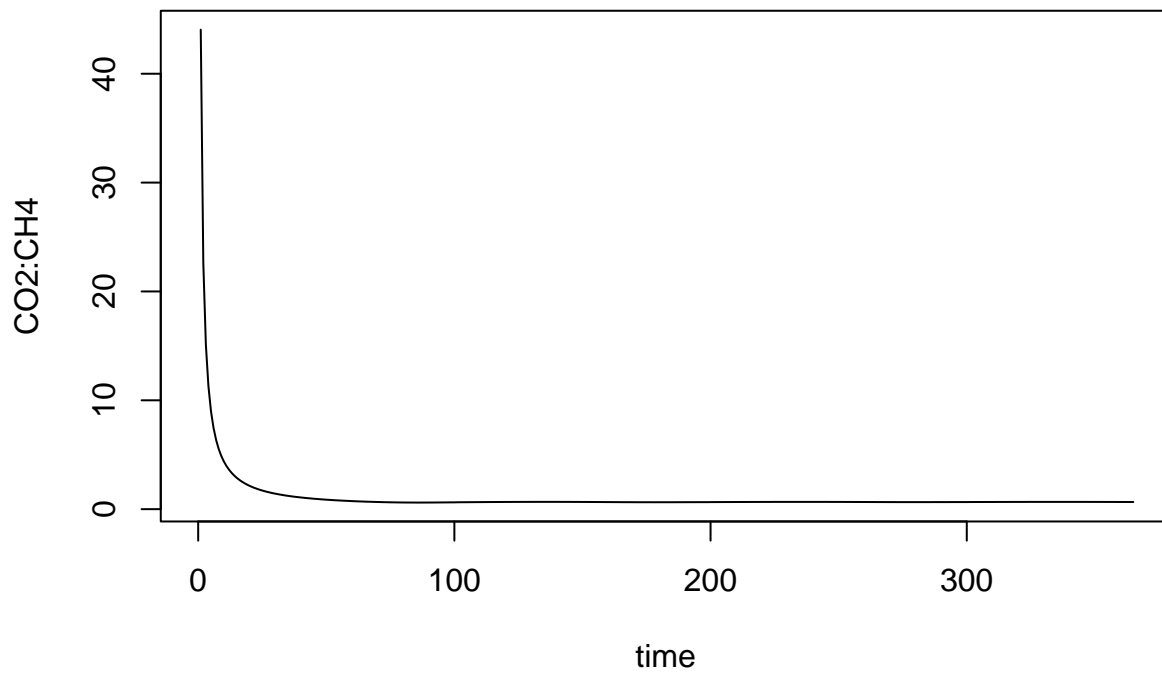
TIC in slurry shows a lot of change over time, but can be kept away from 0 by adjusting kl. The low slurry level leads to a high relative loss.

```
plot(CO2_conc ~ time, data = out11, type = 'l', ylim = c(0, 3))
```

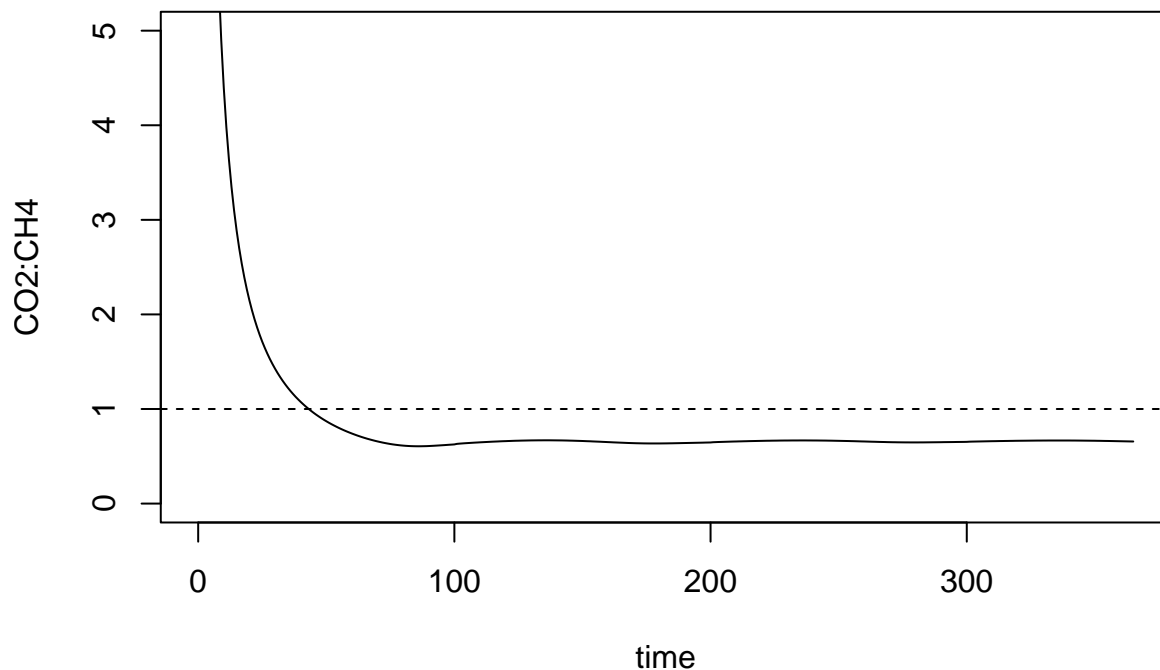


But now we can predict a CO2:CH4 ratio. This is a cumulative value.

```
plot(CO2_emis_cum / CH4_emis_cum ~ time, data = out11, type = 'l', ylab = 'CO2:CH4')
```



```
plot(CO2_emis_cum / CH4_emis_cum ~ time, data = out11, type = 'l', ylab = 'CO2:CH4', ylim = c(0, 5))
abline(h = 1, lty = 2)
```

Let's try some different substrates.

12. Respiration

To include respiration just add a `k1` value for `O2`.

```
chem_pars12a <- list(COD_conv = c(CH4 = 1/0.2507))
```

```
chem_pars12b <- list(COD_conv = c(CH4 = 1/0.2507),
                    k1 = c(O2 = 0.1) * 86400)
```

```
devtools::load_all()
```

```
## i Loading ABM
```

```
out12a <- abm(365,
             mng_pars = mng_pars,
             man_pars = man_pars,
             grp_pars = grp_pars,
             mic_pars = mic_pars,
             sub_pars = sub_pars,
             chem_pars = chem_pars12a,
             )
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 1.7%
```

```
devtools::load_all()
```

```
## i Loading ABM
```

```
out12b <- abm(365,
             mng_pars = mng_pars,
```

```

    man_pars = man_pars,
    grp_pars = grp_pars,
    mic_pars = mic_pars,
    sub_pars = sub_pars,
    chem_pars = chem_pars12b,
)

```

```

## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.

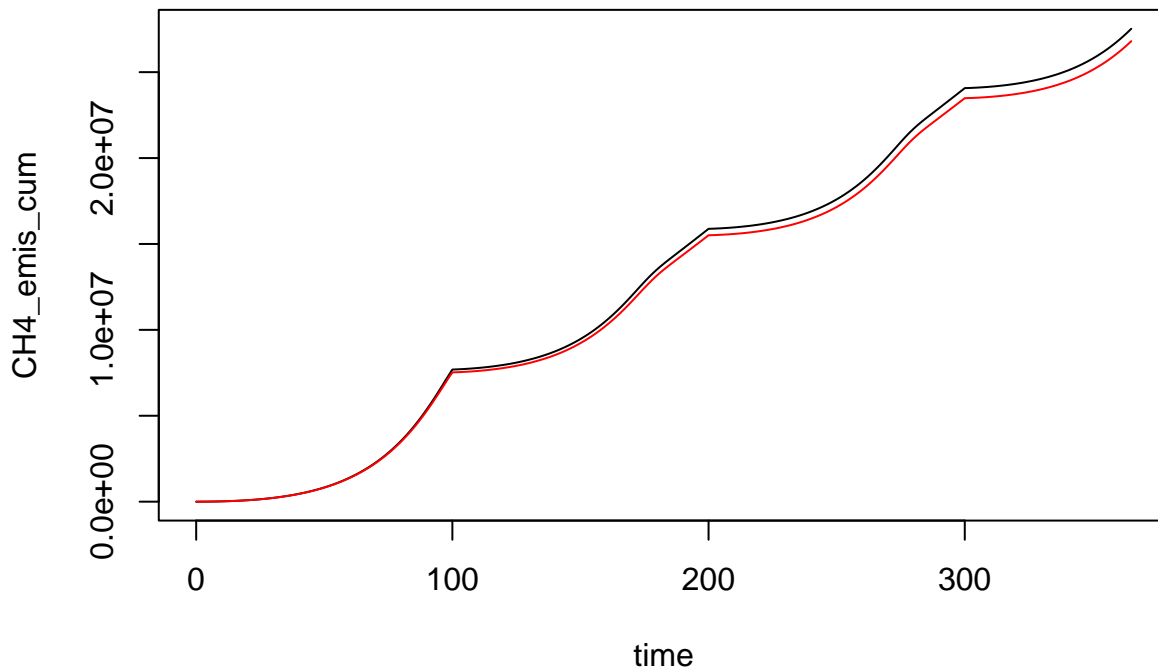
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 3.3%

```

```

plot(CH4_emis_cum ~ time, data = out12a, type = 'l')
lines(CH4_emis_cum ~ time, data = out12b, col = 'red')

```



To track CO₂ emission, we need all the inputs used in example 11.

```

man_pars12 <- list(comps = c('CO2', 'NH4p'),
  comp_fresh = c(CO2 = 1, NH4p = 3),
  VFA_fresh = c(CH3COOH = 2),
  pH = 7, dens = 1000)

```

```

chem_pars12c <- list(COD_conv = c(CH4 = 1/0.2507),
  specs = c('HCO3m'),
  mspec = c(HCO3m = 'CO2', NH3 = 'NH4p'),
  lka = c(HCO3m = '-2.778 + -353.5305 -0.06092*temp_K + 21834.37/temp_K + 126.8339*1000/temp_K'),
  stoich = 'calc',
  kl = c(CO2 = 0.01) * 86400)

```

```

chem_pars12d <- list(COD_conv = c(CH4 = 1/0.2507),
  specs = c('HCO3m'),
  mspec = c(HCO3m = 'CO2', NH3 = 'NH4p'),
  lka = c(HCO3m = '-2.778 + -353.5305 -0.06092*temp_K + 21834.37/temp_K + 126.8339*1000/temp_K'),
  stoich = 'calc',
  )

```

```
kl = c(O2 = 0.1, CO2 = 0.01) * 86400)
```

(Hmm, shouldn't stoich be part of sub_pars?)

We also need stoichiometry for CO2, so we need a substrate chemical formula.

```
sub_pars12 <- list(subs = c('VSd'),
  forms = c(VSd = 'C23H37O14N (CO2)1.5'),
  T_opt_hyd = c(all = 60),
  T_min_hyd = c(all = 0),
  T_max_hyd = c(all = 90),
  hydrol_opt = c(all = 0.1),
  sub_fresh = c(VSd = 20),
  sub_init = c(VSd = 20))
```

```
devtools::load_all()
```

```
## i Loading ABM
```

```
out12c <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars12,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars12,
  chem_pars = chem_pars12c,
)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 1.8%
```

```
devtools::load_all()
```

```
## i Loading ABM
```

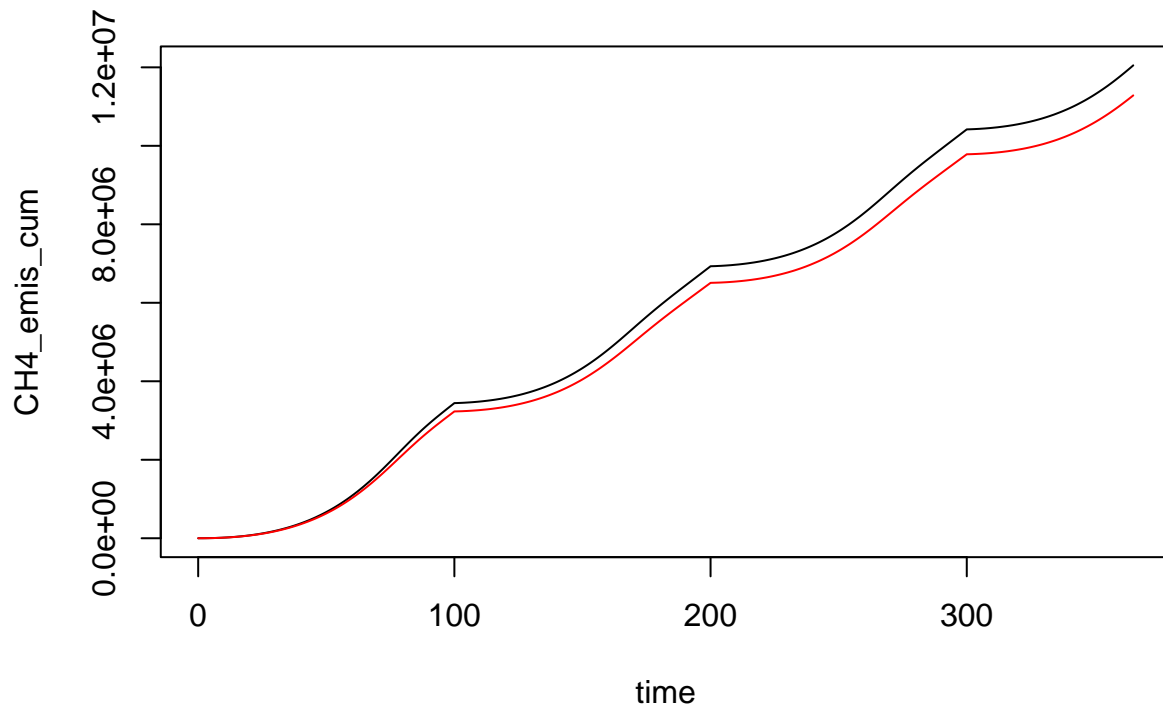
```
out12d <- abm(365,
  mng_pars = mng_pars,
  man_pars = man_pars12,
  grp_pars = grp_pars,
  mic_pars = mic_pars,
  sub_pars = sub_pars12,
  chem_pars = chem_pars12d,
)
```

```
## Warning in expandPars(pars = pars, elnms = pars$grps, parnms = grp_par_nms):
## Size-variable parameter problem: Missing element(s) in kss.
```

```
## Warning in checkCOD(dat = dat, grps = pars$grps, subs = pars$subs, COD_conv =
## pars$COD_conv, : COD balance is off by 5.6%
```

See how COD balance is worse?

```
plot(CH4_emis_cum ~ time, data = out12c, type = 'l')
lines(CH4_emis_cum ~ time, data = out12d, col = 'red')
```



```
plot(CO2_emis_cum ~ time, data = out12c, type = 'l')  
lines(CO2_emis_cum ~ time, data = out12d, col = 'red')
```

