Day 8: Beta Diversity (Guerrero Negro)

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All of the code in this page is meant to be run in R unless otherwise specified.

Install biom package and vegan package if not installed.

```
install.packages(c('biom','vegan'),repo='http://cran.wustl.edu')
```

Load biom package, load data

```
library('biom')
library('vegan')

# load biom file
otus.biom <- read_biom('otu_table_json.biom')

# Extract data matrix (OTU counts) from biom table
otus <- as.matrix(biom_data(otus.biom))

# transpose so that rows are samples and columns are OTUs
otus <- t(otus)

# load mapping file
map <- read.table('map.txt', sep='\t', comment='', head=T, row.names=1)</pre>
```

It is extremely important to ensure that your OTU table and metadata table sample IDs are lined up correctly.

```
# see rownames of map and otus
rownames(map)
    [1] "GNO1P.484257"
                         "GN01P.o.484256" "GN02P.o.484250" "GN02P.484248"
   [5] "GNO3P.484253"
##
                         "GNO3P.o.484249" "GNO4P.484258"
                                                            "GNO4P.o.484251"
  [9] "GN05P.o.484260" "GN05P.484261"
                                           "GN06P.o.484262" "GN06P.484247"
## [13] "GN07P.o.484246" "GN07P.484259"
                                           "GN08P.484265"
                                                            "GNO8P.o.484263"
## [17] "GN09P.484254"
                         "GN09P.o.484264" "GN10P.o.484252" "GN10P.484255"
rownames (otus)
   [1] "GNO1P.484257"
##
                         "GN07P.o.484246" "GN01P.o.484256" "GN06P.484247"
  [5] "GN05P.484261"
                         "GN08P.484265"
                                           "GN05P.o.484260" "GN06P.o.484262"
## [9] "GN08P.o.484263" "GN07P.484259"
                                           "GN04P.484258"
                                                             "GN04P.o.484251"
## [13] "GNO9P.484254"
                                                            "GNO3P.484253"
                         "GN09P.o.484264" "GN02P.484248"
## [17] "GNO2P.o.484250" "GNO3P.o.484249"
# find the overlap
common.ids <- intersect(rownames(map), rownames(otus))</pre>
# get just the overlapping samples
otus <- otus[common.ids,]</pre>
map <- map[common.ids,]</pre>
```

See dimensions of OTU table

```
dim(otus)
```

```
## [1] 18 1750
```

See dimensions of mapping file

```
dim(map)
```

```
## [1] 18 60
```

Get five different distances metrics

```
# get Euclidean distance
d.euc <- dist(otus)

# get Bray-Curtis distances (default for Vegan)
d.bray <- vegdist(otus)

# get Chi-square distances using vegan command
# we will extract chi-square distances from correspondence analysis
my.ca <- cca(otus)
d.chisq <- as.matrix(dist(my.ca$CA$u[,1:2]))

# load unweighted and weighted unifrac
d.uuf <- read.table('beta/unweighted_unifrac_otu_table.txt', sep='\t',head=T,row=1)
d.wuf <- read.table('beta/weighted_unifrac_otu_table.txt', sep='\t',head=T,row=1)

# ensure that these last two matrices have the same samples in the
# same order as the metadata table
d.uuf <- d.uuf[common.ids, common.ids]
d.wuf <- d.wuf[common.ids, common.ids]</pre>
```

Now run principal coordinates embedding on the distance metrics

```
# Run PCoA (not PCA)
pc.euc <- cmdscale(d.euc, k=2)

# Bray-Curtis principal coords
pc.bray <- cmdscale(d.bray,k=2)

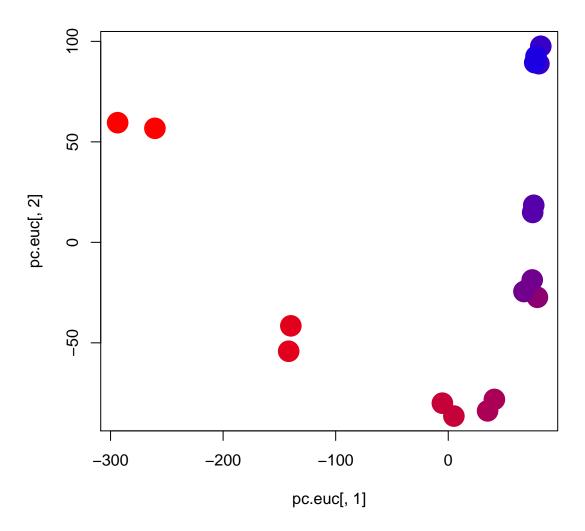
# get first two dimensions of chi-square coordinates:
pc.chisq <- my.ca$CA$u[,1:2]

# get first two dimensions of unifrac distances:
pc.uuf <- cmdscale(d.uuf, k=2)
pc.wuf <- cmdscale(d.wuf, k=2)</pre>
```

Plot Euclidean distances with gradient colors

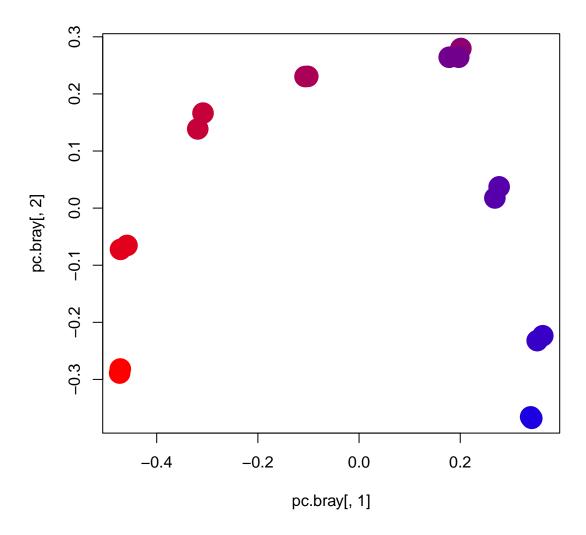
```
# makes a gradient from red to blue
my.colors <- colorRampPalette(c('red','blue'))(10)

# plot Euclidean PCoA coords using color gradient
# based on layer (1...10)
layer <- map[,'LAYER']
plot(pc.euc[,1], pc.euc[,2], col=my.colors[layer], cex=3, pch=16)</pre>
```



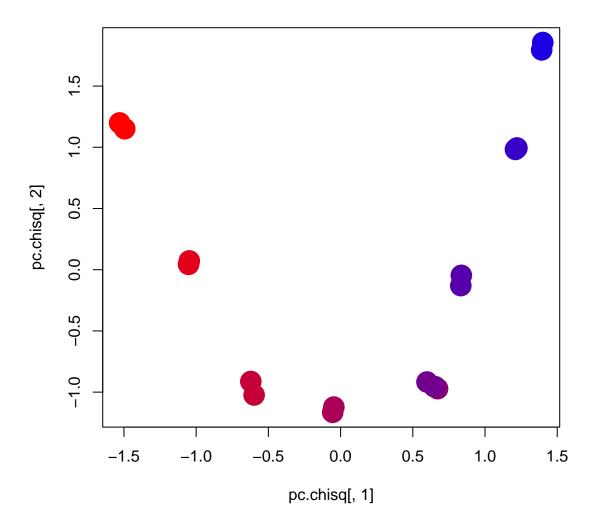
Plot Bray-Curtis distances with gradient colors

```
# Plot Bray-Curtis PCoA
plot(pc.bray[,1], pc.bray[,2], col=my.colors[layer], cex=3, pch=16)
```



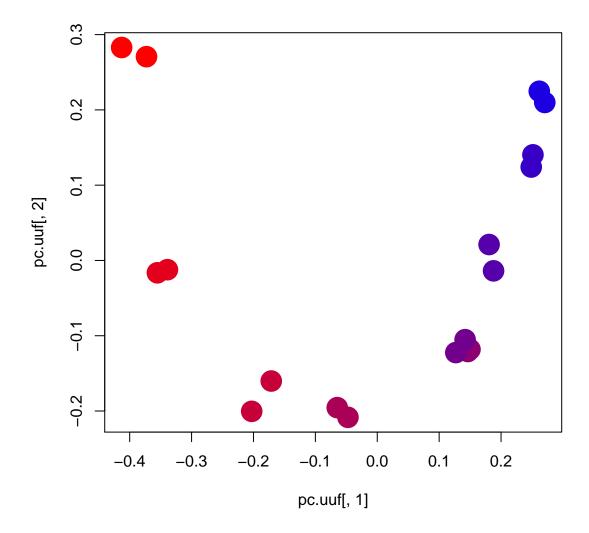
Plot Chi-square distances with gradient colors

```
# Plot Chi-square PCoA
plot(pc.chisq[,1], pc.chisq[,2], col=my.colors[layer], cex=3, pch=16)
```



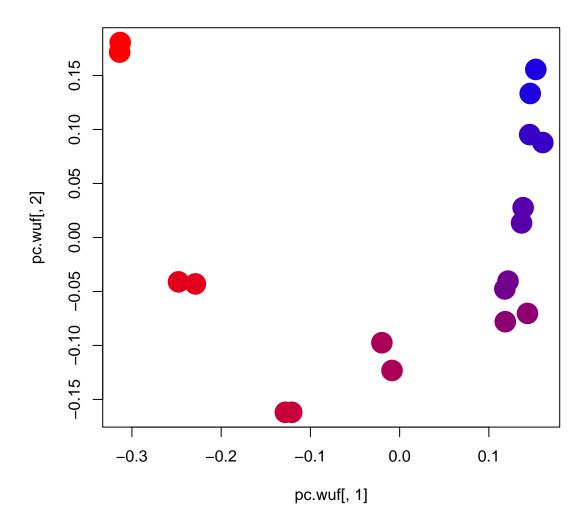
Plot unweighted UniFrac distances with gradient colors

```
plot(pc.uuf[,1], pc.uuf[,2], col=my.colors[layer], cex=3, pch=16)
```



Plot weighted UniFrac distances with gradient colors

```
plot(pc.wuf[,1], pc.wuf[,2], col=my.colors[layer], cex=3, pch=16)
```

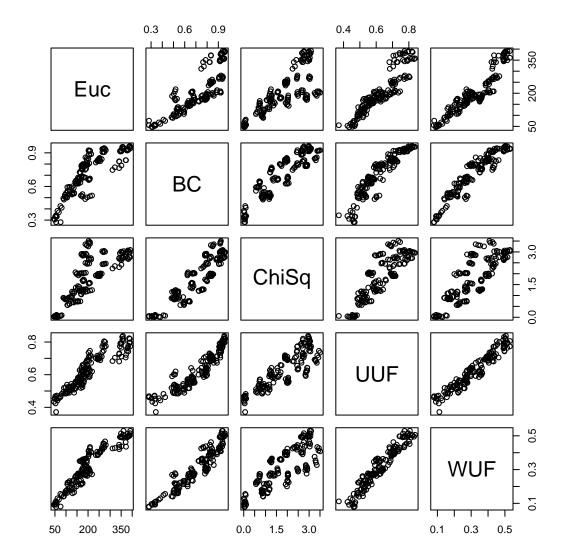


Note: to make a PDF:

```
pdf("chisq.pdf",width=5,height=5)
plot(pc.chisq[,1], pc.chisq[,2], col=my.colors[map[,'LAYER']], cex=3, pch=16)
dev.off()
```

Let's plot pairwise comparisons of the different distance metrics

```
d.vector.matrix <- cbind(as.numeric(d.euc), as.numeric(d.bray), as.numeric(as.dist(d.chisq)), as.numeri
colnames(d.vector.matrix) <- c('Euc', 'BC', 'ChiSq', 'UUF', 'WUF')
pairs(d.vector.matrix)</pre>
```



And plot the pairwise pearson correlations

cor(d.vector.matrix)

```
## Euc 1.000000 0.8324774 0.7664577 0.9159583 0.9344522 ## BC 0.8324774 1.000000 0.8976043 0.9298203 0.9454640 ## ChiSq 0.7664577 0.9159583 0.8192654 ## UUF 0.9159583 0.9298203 0.8535358 1.000000 0.9631557 ## WUF 0.9344522 0.9454640 0.8192654 0.9631557 1.0000000
```