

Class 09: Structural Bioinformatics 1

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What is the PDB anyway?

The main database of biomolecular structures is called the PDB and is available at www.rcsb.org

Lets begin by seeing what is in this database:

```
pdbstats <- read.csv("PDB.csv")
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
head(pdbstats)
```

	Molecular.Type	X.ray	EM	NMR	Multiple.methods	Neutron	Other
1	Protein (only)	152,809	9,421	12,117	191	72	32
2	Protein/Oligosaccharide	9,008	1,654	32	7	1	0
3	Protein/NA	8,061	2,944	281	6	0	0
4	Nucleic acid (only)	2,602	77	1,433	12	2	1
5	Other	163	9	31	0	0	0
6	Oligosaccharide (only)	11	0	6	1	0	4
	Total						
1		174,642					
2		10,702					
3		11,292					
4		4,127					
5		203					
6		22					

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
n.xray <- sum(as.numeric(gsub(",", "", pdbstats$X.ray)))
n.em <- sum(as.numeric(gsub(",", "", pdbstats$EM)))
sum_remove_comma <- function(x){sum(as.numeric(gsub(",", "", x)))}
n.total <- sum_remove_comma(pdbstats$Total)
```

```
p.xray <- (n.xray/n.total) * 100
p.em <- (n.em/n.total) * 100
round(p.xray, 2)
```

```
[1] 85.9
```

```
round(p.em, 2)
```

```
[1] 7.02
```

There are `r` `n.xray` protein structures (`r` `round(p.xray, 2)` and `r` `n.em` `round(p.xray, 2)`) EM structures in the current PDB database

Q2: What proportion of structures in the PDB are protein?

```
as.numeric(gsub(",", "", pdbstats$Total))/n.total
```

```
[1] 0.8689175473 0.0532469600 0.0561824587 0.0205335642 0.0010100105
[6] 0.0001094593
```

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB? 240

It is not straightforward to find all HIV-1 protease structures using plain text searching on the database

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

Hydrogen is too small to appear at this resolution

Q5: There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have

HOH 308

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend “Ball & Stick” for these side-chains). Add this figure to your Quarto document.

A picture of HIV-1 Protease form Molstar

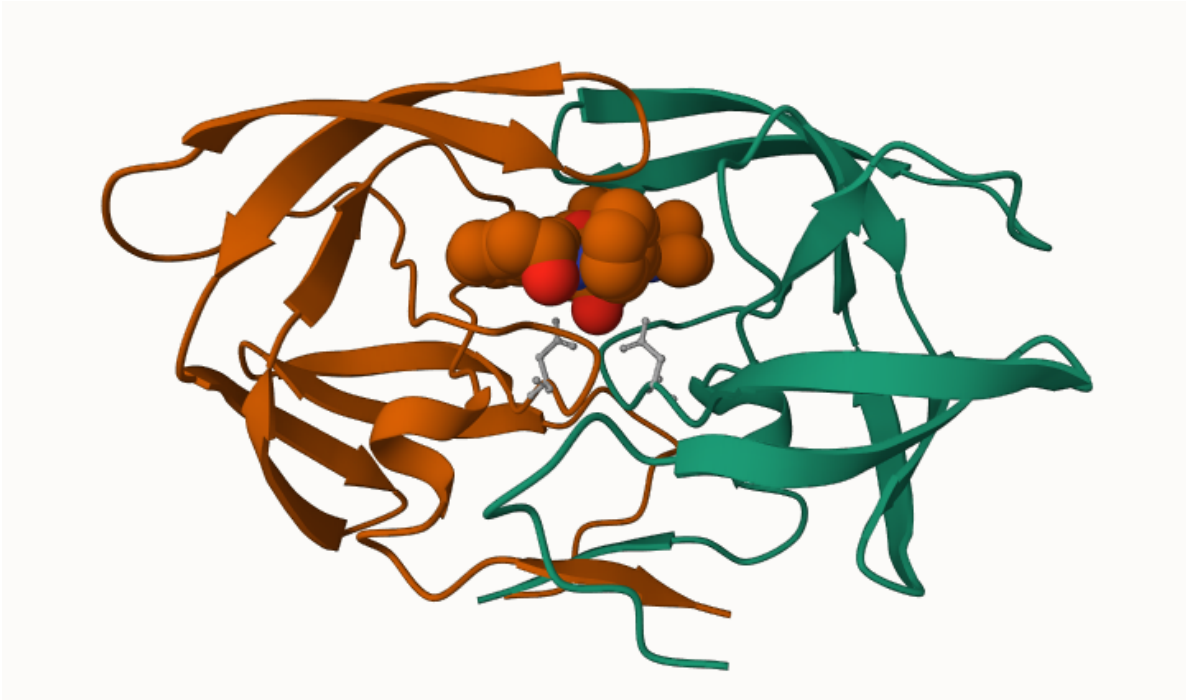


Figure 1: An image I like whilst learning how to break Molstar

Working with structure data in R

We will use the `bio3d` package for this:

```
library(bio3d)
```

Read a PDB file from the online database

```
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

pdb

```
Call: read.pdb(file = "1hsg")
```

```
Total Models#: 1
```

```
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
```

```
Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
```

```
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 172 (residues: 128)
```

```
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
```

```
Protein sequence:
```

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

```
head(pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40
	segid	elesy	charge										
1	<NA>	N	<NA>										
2	<NA>	C	<NA>										
3	<NA>	C	<NA>										
4	<NA>	O	<NA>										
5	<NA>	C	<NA>										
6	<NA>	C	<NA>										

What is the first residue 3 letter code and 1 letter code

```
pdb$atom$resid[1]
```

```
[1] "PRO"
```

```
aa321(pdb$atom$resid[1])
```

```
[1] "P"
```

Q7: How many amino acid residues are there in this pdb object?

198

Q8: Name one of the two non-protein residues?

HOH

Q9: How many protein chains are in this structure?

2

Predicting functional motions of a single structure

Lets read a new PDB structure of Adenylate Kinase and perform normal mode analysis

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file

PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

Call: read.pdb(file = "6s36")

Total Models#: 1

Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

```
Protein Atoms#: 1654 (residues/Calpha atoms#: 214)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)
Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]
```

Protein sequence:

```
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLRAAVKSGSELGKQAKDIMDAGKLV
DELVIALVKERIAQEDCRNGFLLDGFPRTPQADAMKEAGINVDYVLEFDVPDELIVDKI
VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

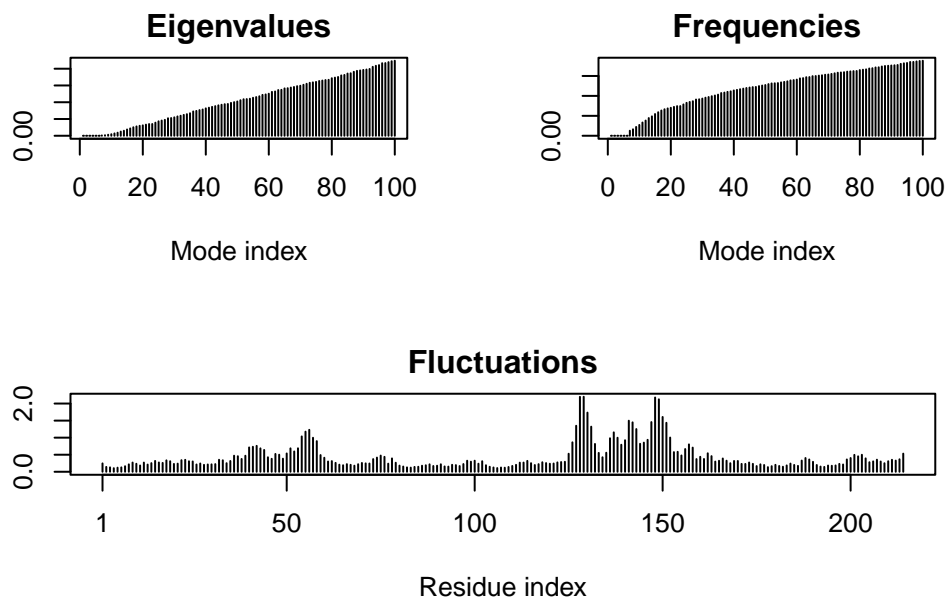
```
+ attr: atom, xyz, seqres, helix, sheet,
      calpha, remark, call
```

Normal mode analysis (NMA) is a structural bioinformatics method to predict protein flexibility and potential functional motions (a.k.a. conformational changes).

```
m <- nma(adk)
```

```
Building Hessian...      Done in 0.04 seconds.
Diagonalizing Hessian... Done in 0.35 seconds.
```

```
plot(m)
```



To view a “movie” of these predicted motions we can generate a molecular “trajectory” with the `mktrj()` function.

```
mktrj(m, file="adk_m7.pdb")
```

Comparative structure analysis of ADK

Q10. Which of the packages above is found only on BioConductor and not CRAN?

msa > Q11. Which of the above packages is not found on BioConductor or CRAN?:

Q12. True or False? Functions from the `devtools` package can be used to install packages from GitHub and BitBucket?

```
library(bio3d)
aa <- get.seq("lake_A")
```

Warning in `get.seq("lake_A")`: Removing existing file: `seqs.fasta`

Fetching... Please wait. Done.

aa

```
      1      .      .      .      .      .      .      60
pdb|1AKE|A  MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMLRAAVKSGSELGKQAKDIMDAGKLV
      1      .      .      .      .      .      .      60

      61      .      .      .      .      .      .      120
pdb|1AKE|A  DELVIALVKERIAQEDCRNGFLLDGFPRTPQADAMKEAGINVDYVLEFDVPDELIVDRI
      61      .      .      .      .      .      .      120

     121      .      .      .      .      .      .      180
pdb|1AKE|A  VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG
     121      .      .      .      .      .      .      180

     181      .      .      .      214
pdb|1AKE|A  YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
     181      .      .      .      214
```

Call:

```
read.fasta(file = outfile)
```

Class:

```
fasta
```

Alignment dimensions:

```
1 sequence rows; 214 position columns (214 non-gap, 0 gap)
```

```
+ attr: id, ali, call
```

Q13. How many amino acids are in this sequence, i.e. how long is this sequence?

214

```
# Blast or hmmer search
#b <- blast.pdb(aa)
```

I could save and load my blast results next time so I don't need to run the search every time.

```
#saveRDS(b, file="blast_results.RDS")
```

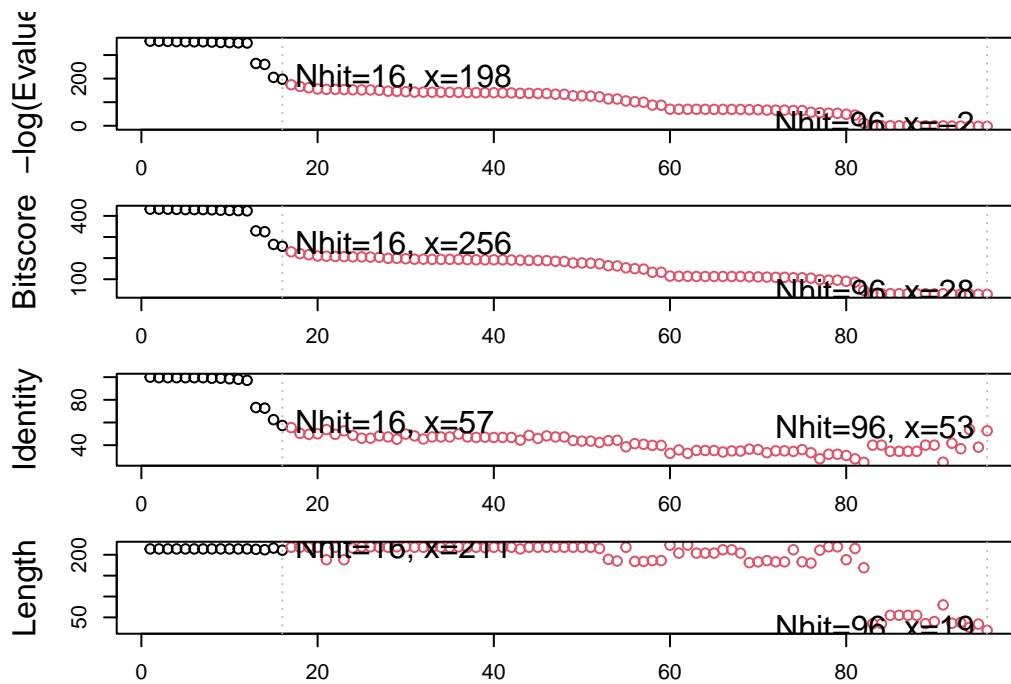
```
b <- readRDS("blast_results.RDS")
```



```
# Plot a summary of search results
hits <- plot(b)
```

```
* Possible cutoff values: 197 -3
    Yielding Nhits: 16 96

* Chosen cutoff value of: 197
    Yielding Nhits: 16
```



```
# List out some 'top hits'
head(hits$ pdb.id)
```

```
[1] "1AKE_A" "4X8M_A" "6S36_A" "6RZE_A" "4X8H_A" "3HPR_A"
```

```
# Download related PDB files
files <- get.pdb(hits$ pdb.id, path="pds", split=TRUE, gzip=TRUE)
```

```
Warning in get.pdb(hits$ pdb.id, path = "pds", split = TRUE, gzip = TRUE):
pds/1AKE.pdb exists. Skipping download
```

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4X8M.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6S36.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6RZE.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4X8H.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3HPR.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1E4V.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/5EJE.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/1E4Y.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3X2S.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6HAP.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/6HAM.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4K46.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4NP6.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/3GMT.pdb exists. Skipping download

Warning in get.pdb(hits\$pdb.id, path = "pdbs", split = TRUE, gzip = TRUE):
pdbs/4PZL.pdb exists. Skipping download

	0%
====	6%
=====	12%
=====	19%
=====	25%
=====	31%
=====	38%
=====	44%
=====	50%
=====	56%
=====	62%
=====	69%
=====	75%
=====	81%
=====	88%
=====	94%
=====	100%

Lets align the PDBs

```
pdbbs <- pdbaln(files, fit = TRUE, exefile="msa")
```

Reading PDB files:

```
pdbbs/split_chain/1AKE_A.pdb
pdbbs/split_chain/4X8M_A.pdb
pdbbs/split_chain/6S36_A.pdb
pdbbs/split_chain/6RZE_A.pdb
pdbbs/split_chain/4X8H_A.pdb
pdbbs/split_chain/3HPR_A.pdb
pdbbs/split_chain/1E4V_A.pdb
pdbbs/split_chain/5EJE_A.pdb
pdbbs/split_chain/1E4Y_A.pdb
pdbbs/split_chain/3X2S_A.pdb
pdbbs/split_chain/6HAP_A.pdb
pdbbs/split_chain/6HAM_A.pdb
pdbbs/split_chain/4K46_A.pdb
pdbbs/split_chain/4NP6_A.pdb
pdbbs/split_chain/3GMT_A.pdb
pdbbs/split_chain/4PZL_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
..   PDB has ALT records, taking A only, rm.alt=TRUE
.    PDB has ALT records, taking A only, rm.alt=TRUE
..   PDB has ALT records, taking A only, rm.alt=TRUE
..   PDB has ALT records, taking A only, rm.alt=TRUE
....  PDB has ALT records, taking A only, rm.alt=TRUE
.     PDB has ALT records, taking A only, rm.alt=TRUE
....
```

Extracting sequences

```
pdb/seq: 1   name: pdbbs/split_chain/1AKE_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 2   name: pdbbs/split_chain/4X8M_A.pdb
pdb/seq: 3   name: pdbbs/split_chain/6S36_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 4   name: pdbbs/split_chain/6RZE_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 5   name: pdbbs/split_chain/4X8H_A.pdb
pdb/seq: 6   name: pdbbs/split_chain/3HPR_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
```

```

pdb/seq: 7   name: pdbc/split_chain/1E4V_A.pdb
pdb/seq: 8   name: pdbc/split_chain/5EJE_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 9   name: pdbc/split_chain/1E4Y_A.pdb
pdb/seq: 10  name: pdbc/split_chain/3X2S_A.pdb
pdb/seq: 11  name: pdbc/split_chain/6HAP_A.pdb
pdb/seq: 12  name: pdbc/split_chain/6HAM_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 13  name: pdbc/split_chain/4K46_A.pdb
      PDB has ALT records, taking A only, rm.alt=TRUE
pdb/seq: 14  name: pdbc/split_chain/4NP6_A.pdb
pdb/seq: 15  name: pdbc/split_chain/3GMT_A.pdb
pdb/seq: 16  name: pdbc/split_chain/4PZL_A.pdb

```

pdbc

```

[Truncated_Name:1] 1AKE_A.pdb      1          .          .          .          40
-----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:2] 4X8M_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:3] 6S36_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:4] 6RZE_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:5] 4X8H_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:6] 3HPR_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:7] 1E4V_A.pdb      -----MRIILLGAPVAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:8] 5EJE_A.pdb      -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:9] 1E4Y_A.pdb      -----MRIILLGALVAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:10] 3X2S_A.pdb     -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:11] 6HAP_A.pdb     -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:12] 6HAM_A.pdb     -----MRIILLGAPGAGKGTQAQFIMEKYGIPQIS
[Truncated_Name:13] 4K46_A.pdb     -----MRIILLGAPGAGKGTQAQFIMAKFGIPQIS
[Truncated_Name:14] 4NP6_A.pdb     -----NAMRIILLGAPGAGKGTQAQFIMEKFGIPQIS
[Truncated_Name:15] 3GMT_A.pdb     -----MRLILLGAPGAGKGTQANFIKEKFGIPQIS
[Truncated_Name:16] 4PZL_A.pdb     TENLYFQSNAMRIILLGAPGAGKGTQAKIIEQKYNIAHIS
                                **~*****  *****  *  *~ *  **
1          .          .          .          40

41          .          .          .          80
TGDMLRAAVKSGSELGKQAKDIMDAGKLVTDLVIALVKE
[Truncated_Name:1] 1AKE_A.pdb      TGDMLRAAVKSGSELGKQAKDIMDAGKLVTDLVIALVKE
[Truncated_Name:2] 4X8M_A.pdb      TGDMLRAAVKSGSELGKQAKDIMDAGKLVTDLVIALVKE
[Truncated_Name:3] 6S36_A.pdb      TGDMLRAAVKSGSELGKQAKDIMDAGKLVTDLVIALVKE
[Truncated_Name:4] 6RZE_A.pdb      TGDMLRAAVKSGSELGKQAKDIMDAGKLVTDLVIALVKE

```

[Truncated_Name:5] 4X8H_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVKE
[Truncated_Name:6] 3HPR_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVKE
[Truncated_Name:7] 1E4V_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVKE
[Truncated_Name:8] 5EJE_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDACKLVTDDELVIALVKE
[Truncated_Name:9] 1E4Y_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVKE
[Truncated_Name:10] 3X2S_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDCGKLVTDDELVIALVKE
[Truncated_Name:11] 6HAP_A.pdb	TGDMRLRAAVKSGSELGKQAKDIMDAGKLVTDDELVIALVRE
[Truncated_Name:12] 6HAM_A.pdb	TGDMRLRAAIKSGSELGKQAKDIMDAGKLVTDDEIIIALVKE
[Truncated_Name:13] 4K46_A.pdb	TGDMRLRAAIKAGTELGKQAKSVIDAGQLVSDDIILGLVKE
[Truncated_Name:14] 4NP6_A.pdb	TGDMRLRAAIKAGTELGKQAKAVIDAGQLVSDDIILGLIKE
[Truncated_Name:15] 3GMT_A.pdb	TGDMRLRAAVKAGTPLGVEAKTYMDEGKLVPSDLIIIGLVKE
[Truncated_Name:16] 4PZL_A.pdb	TGDMIRETIKSGSALGQELKKVLDAGELVSDEFIIKIVKD
	****~* ~* *~ ** * ~* ** * ^^ ~^^~
	41 . . . 80
	81 . . . 120
[Truncated_Name:1] 1AKE_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:2] 4X8M_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:3] 6S36_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:4] 6RZE_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:5] 4X8H_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:6] 3HPR_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:7] 1E4V_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:8] 5EJE_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:9] 1E4Y_A.pdb	RIAQEDCRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:10] 3X2S_A.pdb	RIAQEDSRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:11] 6HAP_A.pdb	RICQEDSRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:12] 6HAM_A.pdb	RICQEDSRNGFLLDGFPR TIPQADAMKEAGINVDYVLEFD
[Truncated_Name:13] 4K46_A.pdb	RIAQDDCAKGFLDGFPR TIPQADGLKEVGVVVDYVIEFD
[Truncated_Name:14] 4NP6_A.pdb	RIAQADCEKGFLLDGFPR TIPQADGLKEMGINVDYVIEFD
[Truncated_Name:15] 3GMT_A.pdb	RLKEADCANGYLFDFPR TIPQADAMKEAGVAIDYVLEID
[Truncated_Name:16] 4PZL_A.pdb	RISKNCNNGFLLDGVPR TIPQAQELDKLGVNIDYIVEVD
	*~ * *~* ** ***** ** ^ *~ ~**~* *
	81 . . . 120
	121 . . . 160
[Truncated_Name:1] 1AKE_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTG
[Truncated_Name:2] 4X8M_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTG
[Truncated_Name:3] 6S36_A.pdb	VPDELIVDKIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTG
[Truncated_Name:4] 6RZE_A.pdb	VPDELIVDAIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTG
[Truncated_Name:5] 4X8H_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTG
[Truncated_Name:6] 3HPR_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHV KFNPPKVEGKDDGTG
[Truncated_Name:7] 1E4V_A.pdb	VPDELIVDRIVGRRVHAPSGRVYHV KFNPPKVEGKDDVTG


```
Call:
  pdbaln(files = files, fit = TRUE, exefile = "msa")

Class:
  pdba, fasta

Alignment dimensions:
  16 sequence rows; 227 position columns (204 non-gap, 23 gap)

+ attr: xyz, resno, b, chain, id, ali, resid, sse, call
```

```
# Vector containing PDB codes for figure axis
ids <- basename.pdb(pdb$ids)

# Draw schematic alignment
#plot(pdb, labels=ids)
```

```
anno <- pdb.annotate(ids)
unique(anno$source)
```

16


```
head(anno)
```

	structureId	chainId	macromoleculeType	chainLength	experimentalTechnique
1AKE_A	1AKE	A	Protein	214	X-ray
4X8M_A	4X8M	A	Protein	214	X-ray
6S36_A	6S36	A	Protein	214	X-ray
6RZE_A	6RZE	A	Protein	214	X-ray
4X8H_A	4X8H	A	Protein	214	X-ray
3HPR_A	3HPR	A	Protein	214	X-ray
	resolution	scopDomain	pfam	ligandId	
1AKE_A	2.00	Adenylate kinase	Adenylate kinase (ADK)	AP5	
4X8M_A	2.60	<NA>	Adenylate kinase (ADK)	<NA>	
6S36_A	1.60	<NA>	Adenylate kinase (ADK)	CL (3),NA,MG (2)	
6RZE_A	1.69	<NA>	Adenylate kinase (ADK)	NA (3),CL (2)	
4X8H_A	2.50	<NA>	Adenylate kinase (ADK)	<NA>	
3HPR_A	2.00	<NA>	Adenylate kinase (ADK)	AP5	
	ligandName	source			
1AKE_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE	Escherichia coli			
4X8M_A	<NA>	Escherichia coli			
6S36_A	CHLORIDE ION (3),SODIUM ION,MAGNESIUM ION (2)	Escherichia coli			
6RZE_A	SODIUM ION (3),CHLORIDE ION (2)	Escherichia coli			
4X8H_A	<NA>	Escherichia coli			
3HPR_A	BIS(ADENOSINE)-5'-PENTAPHOSPHATE	Escherichia coli K-12			

1AKE_A STRUCTURE OF THE COMPLEX BETWEEN ADENYLATE KINASE FROM ESCHERICHIA COLI AND THE INHIBITORS
 4X8M_A
 6S36_A
 6RZE_A
 4X8H_A
 3HPR_A

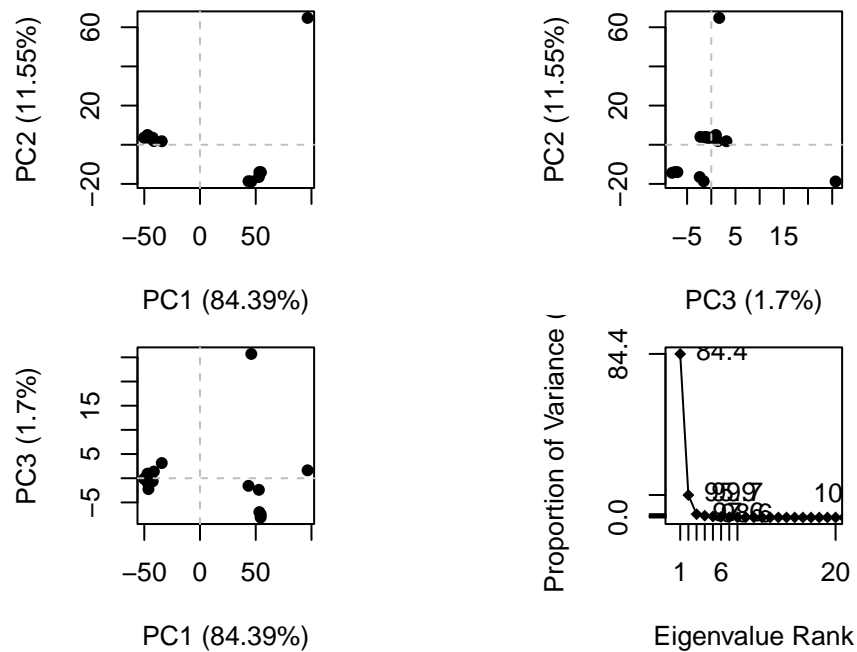
		citation	rObserved	rFree
1AKE_A		Muller, C.W., et al. J Mol Biol (1992)	0.1960	NA
4X8M_A		Kovermann, M., et al. Nat Commun (2015)	0.2491	0.3089
6S36_A		Rogne, P., et al. Biochemistry (2019)	0.1632	0.2356
6RZE_A		Rogne, P., et al. Biochemistry (2019)	0.1865	0.2350
4X8H_A		Kovermann, M., et al. Nat Commun (2015)	0.1961	0.2895
3HPR_A	Schrank, T.P., et al. Proc Natl Acad Sci U S A (2009)		0.2100	0.2432
	rWork	spaceGroup		
1AKE_A	0.1960	P 21 2 21		
4X8M_A	0.2463	C 1 2 1		
6S36_A	0.1594	C 1 2 1		
6RZE_A	0.1819	C 1 2 1		

```
4X8H_A 0.1914    C 1 2 1
3HPR_A 0.2062    P 21 21 2
```

##Principal Component Analysis

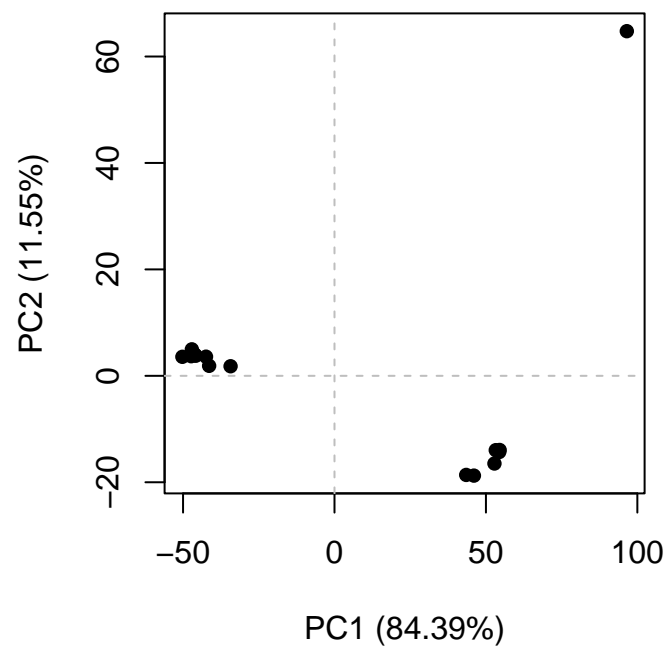
Time for PCA. We will use not the `prcomp()` function from base r but the `pca()` function from the `bio3d` package as this one is designed to work nicely with biomolecular data.

```
pc.xray <- pca(pdbbs)
plot(pc.xray)
```



We can now focus in on PC1 vs PC2

```
plot(pc.xray, 1:2)
```

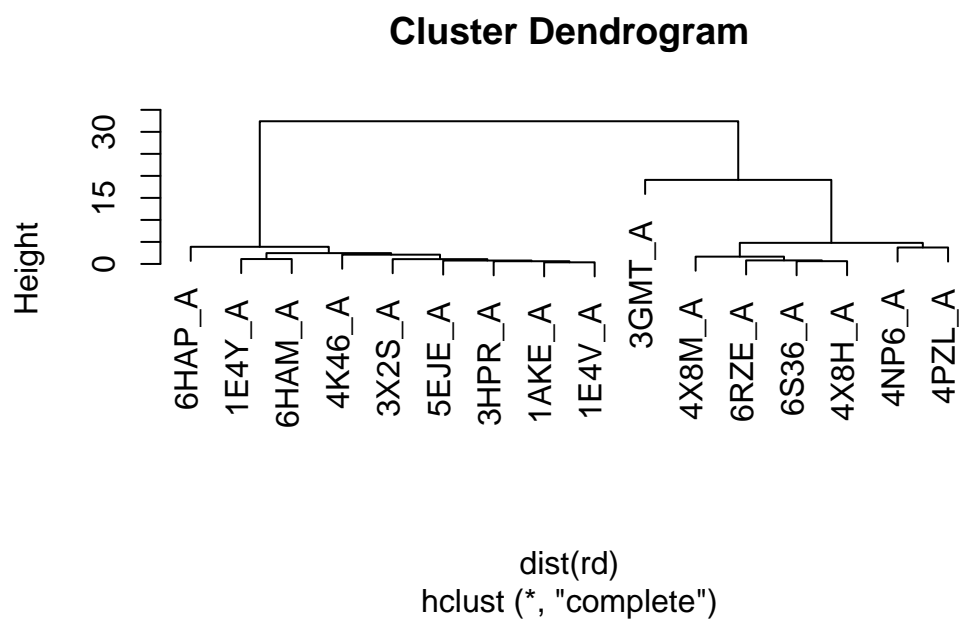


Lets cluster our structures

```
# Calculate RMSD  
rd <- rmsd(pdb)
```

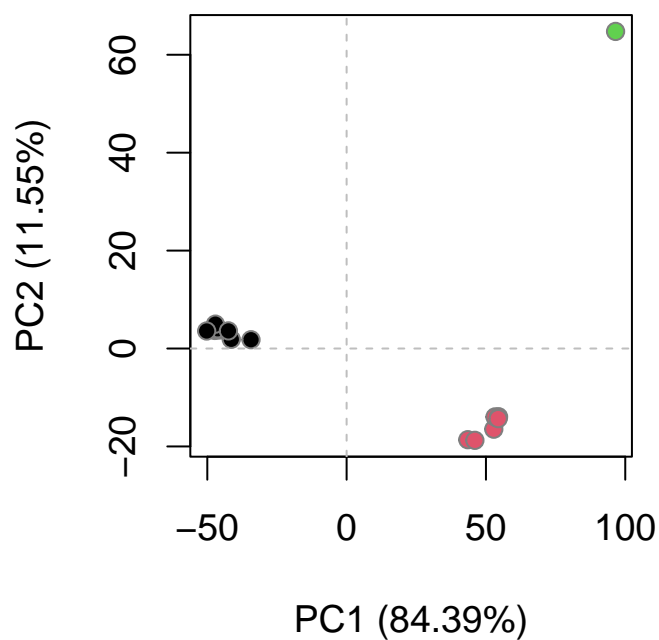
Warning in rmsd(pdb): No indices provided, using the 204 non NA positions

```
# Structure-based clustering  
hc.rd <- hclust(dist(rd))  
grps.rd <- cutree(hc.rd, k=3)  
plot(hc.rd)
```



And now my PC plot colored by clustering group

```
plot(pc.xray, 1:2, col="grey50", bg=grps.rd, pch=21, cex=1)
```



To visualize the major structural variations in the ensemble the function `mktrj()` can be used to generate a trajectory PDB file by interpolating along a give PC (eigenvector):

```
# Visualize first principal component
mktrj(pc.xray, pc=1, file="pc_1.pdb")
```

We can now open this trajectory file in Molstar to view a movie of the major differences (i.e. displacements of atoms) in the structure set as we move along PC1.