# EnzyChrom<sup>TM</sup> NADP<sup>+</sup>/NADPH Assay Kit (ECNP-100)

Ultrasensitive Colorimetric Determination of NADP<sup>+</sup>/NADPH at 565 nm

#### **DESCRIPTION**

Pyridine nucleotides play an important role in metabolism and, thus, there is continual interest in monitoring their concentration levels. Quantitative determination of NADP+/NADPH has applications in research pertaining to energy transformation and redox state of cells or tissue.

Simple, direct and automation-ready procedures for measuring NADP+/NADPH concentration are very desirable. BioAssay Systems' EnzyChrom™ NADP+/NADPH assay kit is based on a glucose dehydrogenase cycling reaction, in which the formed NADPH reduces a formazan (MTT) reagent. The intensity of the reduced product color, measured at 565 nm, is proportionate to the NADP+/NADPH concentration in the sample. This assay is highly specific for NADP\*/NADPH and is not interfered by NAD\*/NADH. Our assay is a convenient method to measure NADP, NADPH and their ratio.

#### **APPLICATIONS**

Direct Assays: NADP+/NADPH concentrations and ratios in cell or tissue extracts.

#### **KEY FEATURES**

Sensitive and accurate. Detection limit 0.1 µM, linearity up to 10 µM NADP+/NADPH in 96-well plate assay.

Convenient. The procedure involves adding a single working reagent, and reading the optical density at time zero and 30 min at room temperature. No 37°C heater is required.

High-throughput. Can be readily automated as a high-throughput 96well plate assay for thousands of samples per day.

### KIT CONTENTS (100 tests in 96-well plates)

Assay Buffer: 10 mL Glucose (1 M): 1.5 mL MTT Solution: 1.5 mL Enzyme Mix: 120 μL NADP Standard: 0.5 mL 1 mM

NAD(P)/NAD(P)H Extraction Buffers: each 12 mL

Storage conditions. The kit is shipped on ice. Store all reagents at -20°C. Shelf life: 6 months after receipt.

**Precautions**: reagents are for research use only. Normal precautions for laboratory reagents should be exercised while using the reagents. Please refer to Material Safety Data Sheet for detailed information.

# **PROCEDURES**

- 1. Sample Preparation. For tissues weigh ~20 mg tissue for each sample, wash with cold PBS. For cell samples, wash cells with cold PBS and pellet ~10<sup>5</sup> cells for each sample. Homogenize samples (either tissue or cells) in a 1.5 mL eppindorf tube with either 100  $\mu\text{L}$ NADP extraction buffer for NADP determination or 100 µL NADPH extraction buffer for NADPH determination. Heat extracts at 60°C for 5 min and then add 20  $\mu L$  Assay Buffer and 100  $\mu L$  of the opposite extraction buffer to neutralize the extracts. Briefly vortex and spin the samples down at 14,000 rpm for 5 min. Use supernatant for NADP/NADPH assays. Determination of both NADP and NADPH concentrations requires extractions from two
- 2. Calibration Curve. Prepare 500 µL 10 µM NADP Premix by mixing 5 μL 1 mM Standard and 495 μL distilled water.

No	Premix + H <sub>2</sub> O	Vol (μL)	[NADP] (µM)
1	100μL + 0μL	100	10
2	80μL + 20μL	100	8
3	60μL + 40μL	100	6
4	40μL + 60μL	100	4
5	30μL + 70μL	100	3
6	20μL + 80μL	100	2
7	10μL + 90μL	100	1
8	0μL + 100μL	100	0

Dilute standard as shown in the Table. Transfer 40  $\mu L$  standards into wells of a clear bottom 96-well plate.

Samples: add 40 µL sample per well in separate wells.

- 3. Reagent Preparation. For best results allow Enzyme to come to RT (15-30 min) before preparing the Working Reagent. For each well of reaction, prepare Working Reagent by mixing 60 µL Assay Buffer, 1  $\mu$ L Enzyme Mix, 10  $\mu$ L Glucose and 14  $\mu$ L MTT. Fresh reconstitution is recommended.
- 4. Reaction. Add 80 μL Working Reagent per well quickly. Tap plate to mix briefly and thoroughly.
- 5. Read optical density (OD<sub>0</sub>) for time "zero" at 565 nm (520-600nm) and OD30 after a 30-min incubation at room temperature.
- 6. Calculation. Subtract ODo from OD30 for the standard and sample wells. Use the ΔOD values to determine sample NADP/NADPH concentration from the standard curve.

$$[NADP(H)] = \frac{\Delta OD_{SAMPLE} - \Delta OD_{BLANK}}{Slope (\mu M^{-1})} \times n \quad (\mu M)$$

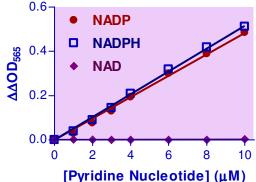
Note: If the sample  $\triangle OD$  values are higher than the  $\triangle OD$  value for the  $10 \mu M$  standard, dilute sample in distilled water and repeat this assay. Multiply the results by the dilution factor.

# MATERIALS REQUIRED, BUT NOT PROVIDED

Pipetting (multi-channel) devices. Clear-bottom 96-well plates (e.g. Corning Costar) and plate reader.

# **GENERAL CONSIDERATIONS**

- 1. At these concentrations, the standard curves for NADP and NADPH are identical. Since NADPH in solution is unstable, we provide only NADP as the standard.
- 2. This assay is based on an enzyme-catalyzed kinetic reaction. Addition of Working Reagent should be quick and mixing should be brief but thorough. Use of multi-channel pipettor is recommended.
- 3. The following substances interfere and should be avoided in sample preparation. EDTA (>0.5 mM), ascorbic acid, SDS (>0.2%), sodium azide, NP-40 (>1%) and Tween-20 (>1%).



Standard Curve in 96-well plate assay

# **PUBLICATIONS**

- 1. Kwak, S., et al. (2020). Redirection of the glycolytic flux enhances isoprenoid production in Saccharomyces cerevisiae. Biotechnology Journal 15(2): e1900173.
- 2. Zhang, Y., et al. (2021). Hepatic stellate cells specific liposomes with the Toll-like receptor 4 shRNA attenuates liver fibrosis. Journal of Cellular and Molecular Medicine 25(2): 1299-1313.
- 3. Hu, Li, et al (2019). Melatonin decreases M1 polarization via attenuating mitochondrial oxidative damage depending on UCP2 pathway in prorenin-treated microglia. PloS one 14.2: e0212138.