

QuantiChrom™ Formaldehyde Assay Kit (DFOR-100) Quantitative Fluorimetric Determination of Formaldehyde

DESCRIPTION

FORMALDEHYDE (methanal) is the simplest aldehyde. It is widely employed in industry for wide range of applications. Formaldehyde is also used as a disinfectant and is a commonly utilized tissue fixative and embalming agent. Formaldehyde is naturally present in all tissues and body fluids. Recently it has been shown that some cancer types exhibit elevated formaldehyde levels. Increased formaldehyde concentration in urine has been associated with prostate and bladder cancer. Thus, measuring formaldehyde in urine can be a very useful tool when studying cancer.

BioAssay Systems' newly designed Formaldehyde Assay Kit provides a convenient fluorimetric means to measure formaldehyde in biological samples. In the assay, formaldehyde is derivatized with acetoacetanilide in the presence of ammonia. The resulting fluorescent product is then quantified fluorimetrically ($\lambda_{exc/em} = 370/470\text{nm}$). The assay is simple, sensitive, stable and high-throughput adaptable. The assay can detect as low as 1.5 μM formaldehyde in biological samples.

KEY FEATURES

Safe. Non-radioactive assay.

Sensitive and accurate. As low as 1.5 μM (45 ppb) formaldehyde can be quantified.

Homogeneous and convenient. "Mix-incubate-measure" type assay. No wash and reagent transfer steps are involved.

Robust and amenable to HTS: Can be readily automated on HTS liquid handling systems for processing thousands of samples per day.

APPLICATIONS

Formaldehyde determination in urine and other biological samples.

KIT CONTENTS

Reagent A: 5 mL **Reagent B:** 3 mL **Standard:** 100 μL
10% TCA: 5 mL **Neutralizer:** 2 x 1.5 mL

Storage conditions: The kit is shipped at room temperature. Store all reagents at 4°C. Shelf life of 18 months after receipt.

Precautions: reagents are for research use only. Normal precautions for laboratory reagents should be exercised while using the reagents. Please refer to Material Safety Data Sheet for detailed information.

ASSAY PROCEDURE

Use black flat-bottom plates. Prior to assay, bring all reagents to room temperature.

- Standards. Mix 5 μL the provided 10 mM Formaldehyde with 495 μL dH₂O to make a 100 μM Premix. Dilute standard as follows.

No	Premix + H ₂ O	Formaldehyde (μM)
1	100 μL + 0 μL	100
2	60 μL + 40 μL	60
3	30 μL + 70 μL	30
4	0 μL + 100 μL	0

Transfer 50 μL standards into separate wells of the plate.

- Sample Preparation. Urine samples should be diluted 2-5 fold with dH₂O. If urine samples contain visible particulates, then the samples should be cleared by either filtration or centrifugation (14000 rpm, 5 min).

Samples high in protein (cell lysate, serum, etc.) need to be deproteinized and neutralized prior to assaying. To deproteinize, add 50 μL 10% TCA per 100 μL sample. Vortex and centrifuge for 5 min at 14000 rpm. Transfer 100 μL of clear supernatant to a clean tube and neutralize with 25 μL Neutralizer. *Note: Measured ΔRFU 's for deproteinized samples need to be multiplied by 1.875 to compensate for the resulting dilution of the sample.*

Samples not measured the same day should be stored frozen, preferably at -80°C.

- Add 50 μL of each prepared sample to two separate wells of the plate (one well will be used as a Sample Blank).
- Prepare Working Reagent for each standard and sample well by mixing 33 μL Reagent A and 22 μL Reagent B. For the Sample Blanks, make the following Working Reagent: 33 μL Reagent A + 22 μL dH₂O. Add 50 μL of the appropriate Working Reagent to each well. Tap plate to mix. Incubate at room temperature for 30 min protected from light.
- Read fluorescence intensity at $\lambda_{exc} = 370\text{ nm}$ and $\lambda_{em} = 470\text{ nm}$.

CALCULATION

Plot the RFU measured at 30 min for each standard against the standard concentrations. Determine the slope using linear regression fitting. The Formaldehyde concentration of a Sample is calculated as

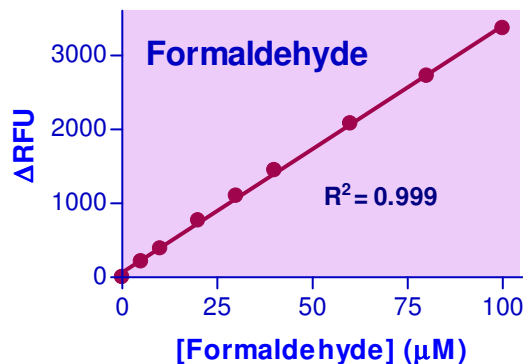
$$[\text{Formaldehyde}] = \frac{\text{RFU}_{\text{SAMPLE}} - \text{RFU}_{\text{BLANK}} - \text{RFU}_{\text{WATER}}}{\text{Slope}} \times n \text{ (}\mu\text{M)}$$

where $\text{RFU}_{\text{SAMPLE}}$, $\text{RFU}_{\text{BLANK}}$ and $\text{RFU}_{\text{WATER}}$ are the measured fluorescence values of the sample, sample blank and water (std #4) respectively. **Slope** is the slope of the standard curve in μM^{-1} and **n** is the dilution factor ($n = 1.875$ for deproteinized samples). Note: if the Sample Formaldehyde concentration is higher than the 100 μM prior to applying the dilution factor, dilute sample in water and repeat the assay. Multiply result by the dilution factor.

Conversion factor: 1 μM formaldehyde is equivalent to 30 ppb.

MATERIALS REQUIRED, BUT NOT PROVIDED

Pipetting devices, centrifuge tubes, black flat bottom 96-well plates (e.g. Corning Costar) and plate reader.



Formaldehyde Standard Curve in Water

PUBLICATIONS

- Ai, L., et al. (2019). Endogenous formaldehyde is a memory-related molecule in mice and humans. *Communications Biology*, 2(1), 446.
- Zhai, R et al (2016). Evidence for Conversion of Methanol to Formaldehyde in Nonhuman Primate Brain. *Analytical Cell Pathology (Amsterdam)*: 4598454.
- Yang, M et al (2014). Alzheimer's disease and methanol toxicity (part 2): lessons from four rhesus macaques (*Macaca mulatta*) chronically fed methanol. *J Alzheimers Dis.* 41(4): 1131-1147.