Aspects of Bayesian Networks

Presented for the Aalborg R user group

April 2021

Søren Højsgaard©

Department of Mathematical Sciences

Aalborg University, Denmark

April 8, 2021

Printed: April 8, 2021 File: bayesnet-slides.tex

Contents

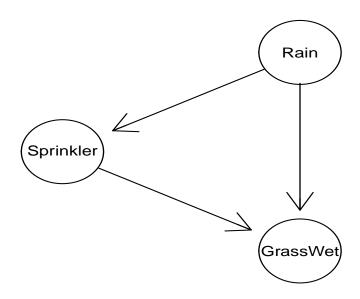
1	Example: The sprinkler network	3
	1.1 The setting	. 4
	1.2 Conditional probability tables (CPTs)	. 6
	1.3 A small digression: Operations on arrays	. 8
	1.4 Operations on arrays - overview	
	1.5 Using Bayes' formula	
	1.6 Under the hood	
	the fidea the fidea	. 10
2	Example: The chest clinic narrative	14
3	The curse of dimensionality	18
4	Message passing – excerpt from chest clinic	22
	4.1 Computing clique marginals	. 26
	4.2 Conditioning	
5	An introduction to the gRain package	39
	5.1 Specify BN from list of CPTs	. 40
	5.2 Building network	
	5.3 Querying the network	
	5.4 Setting evidence	
	5.5 Specify BN from DAG and data**	
	5.5 Specify Bit from DAG and data	. 40
6	Wrapping up	50
	5.1 Book: Graphical Models with R	. 51
	5.2 Package versions	

1 Example: The sprinkler network

1.1 The setting

Two events can cause grass to be wet: Either the sprinkler is on or it is raining. rain has a direct effect on the use of the sprinkler: when it rains, the sprinkler is usually not turned on.

What is the probability that it has been raining given that the grass is wet?



This can be modeled with a Bayesian network. The variables (R)ain, (S)prinkler, (G)rassWet have two possible values: (y)es and (n)o.

In a model building context we start in by defining conditional probabilities specifying the terms

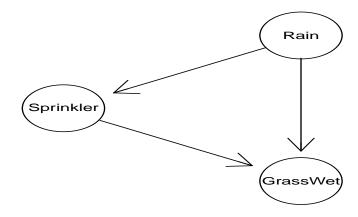
$$P(G|S,R), \quad P(S|R), \quad P(R)$$

We call these terms conditional probability tables (or CPTs). Then we construct a joint pmf by

$$P(G, S, R) \leftarrow P(G|S, R)P(S|R)P(R)$$

Notice this: Terms on the right hand side above have the form

relative to the DAG (directed acyclic graph):



1.2 Conditional probability tables (CPTs)

For compact printing of arrays define utility function

```
> flatten <- function(x){
   ftable(x, row.vars=1)
}</pre>
```

Conditional probability tables (CPTs) in R are arrays. Arrays can be created e.g. with array() or as follows:

```
> yn <- c("yes", "no")
> domain <- list(rain = yn, sprinkler = yn, wet_grass = yn)
> ## P(R)
> p.R <- tabNew("rain", levels=domain, values=c(.2, .8))
> p.R
rain
yes no
0.2 0.8
```

```
> ## P(S|R)
> p.S_R <- tabNew(c("sprinkler", "rain"), levels = domain,</pre>
               values=c(.01, .99, .4, .6))
> p.S_R %>% flatten
         rain yes no
sprinkler
           0.01 0.40
yes
             0.99 0.60
no
> ## P(G|S,R)
> p.G_SR <- tabNew(~wet_grass:sprinkler:rain, levels = domain,
                values=c(.99, .01, .8, .2, .9, .1, 0, 1))
> p.G_SR %>% flatten
         sprinkler yes no
         rain yes no yes
                                   no
wet_grass
                   0.99 0.90 0.80 0.00
yes
                   0.01 0.10 0.20 1.00
no
```

1.3 A small digression: Operations on arrays

Think of T_1 as function of variables (a,b) and T_2 as function of (b,c).

The product $T = T_1T_2$ is a function of (a, b, c) defined as

$$T(a,b,c) \leftarrow T_1(a,b)T_2(b,c)$$

1.4 Operations on arrays - overview

```
> ## Multiplication
> T1 %a*% T2
> tabMult(T1, T2)
> ## Division
> T1 %a/% T2
> tabDiv(T1, T2)
> ## Division; 0/0 = 0
> T1 %a/0% T2
> tabDiv0(T1, T2)
> ## Addition
> T1 %a+% T2
> tabAdd(T1, T2)
> ## Subtraction
> T1 %a-% T2
> tabSubt(T1, T2)
> ## Equality
> T1 %a==% T2
> tabEqual(T1, T2)
> ## Marginalization
> T1 %a_% set
> tabMarg(T1, set)
```

```
Joint pmf:
> ## P(G,S,R)
> p.GSR <- p.G_SR %a*% p.S_R %a*% p.R
> p.GSR %>% flatten
          wet_grass
                         yes
                                          no
          rain
                         yes
                                  no
                                         yes
                                                   no
sprinkler
                    0.00198 0.28800 0.00002 0.03200
yes
                     0.15840 0.00000 0.03960 0.48000
no
> sum(p.GSR) # check
[1] 1
```

1.5 Using Bayes' formula

Question: What is the probability that it is raining given that the grass is wet?

Answer: Use Bayes formula:

$$P(R|G = y) = \frac{P(R, G = y)}{P(G = y)}$$

$$= \frac{\sum_{S=y,n} P(R, S, G = y)}{\sum_{R=y,n;S=y,n} P(R, S, G = y)}$$

This question - and others - can be answered with tabDist:
> tabDist(p.GSR, marg="rain", cond=list(wet_grass="yes"))
rain
 yes no
0.36 0.64

1.6 Under the hood

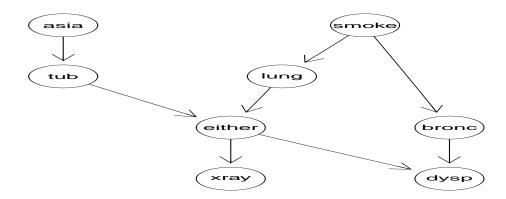
```
In detail we have the following computations:
> ## 1) Marginalize: P(S, G, R) -> P(R, G) -> P(G)
> p.RG <- p.GSR %amarg% ~rain + wet_grass; p.RG</pre>
     wet_grass
rain yes no
 yes 0.16 0.04
 no 0.29 0.51
> p.G <- p.RG %amarg% ~wet_grass; p.G</pre>
wet_grass
yes no
0.45 0.55
> ## 2) Condition \rightarrow P(R|G)
> p.R_G <- p.RG %a/% p.G; p.R_G
     wet_grass
rain yes no
 yes 0.36 0.072
 no 0.64 0.928
> ## 3) Pick the slice -> P(R|G=yes)
> p.R_G %aslice% list(wet_grass="yes")
yes
     no
0.36 0.64
```

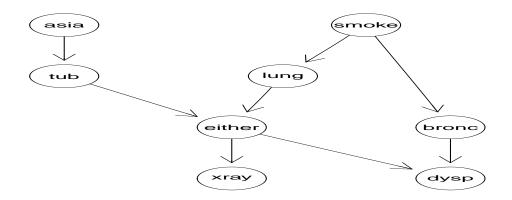
2 Example: The chest clinic narrative

Lauritzen and Spiegehalter (1988) present the following narrative:

- Shortness—of—breath (*dyspnoea*) may be due to *tuberculosis*, *lung* cancer or *bronchitis*, or none of them, or more than one of them.
- A recent visit to Asia increases the chances of tuberculosis, while smoking is known to be a risk factor for both lung cancer and bronchitis.
- The results of a single chest X–ray do not discriminate between lung cancer and bronchitis, as neither does the presence or absence of dyspnoea.

The narrative can be pictured as a DAG (Directed Acyclic Graph)





With an informal notation, a joint distribution for all variables

$$V = \{Asia, Tub, Smoke, Lung, Either, Bronc, Xray, Dysp\}$$

$$\equiv \{a, t, s, l, e, b, x, d\}$$

can be obtained as

$$p(V) = \prod_{v} p(v|pa(v))$$

which here boils down to

$$p(V) = p(a)p(t|a)p(s)p(t|s)p(b|s)p(e|t, l)p(d|e, b)p(x|e).$$

All variables are binary with levels "yes", "no".

The building blocks $p(z_v|z_{pa(v)})$, for example

are represented as multidimensional arrays (here a $2 \times 2 \times 2$ array).

In real world applications, such arrays can become very large and will often contain many zeros. 3 The curse of dimensionality

In principle (and in practice in this small toy example) we can find e.g. $p(b|a^+,d^+)$ by brute force calculations.

Recall: We have a collection of conditional probability tables (CPTs) of the form p(v|pa(v)):

$$\{p(a), p(t|a), p(s), p(l|s), p(b|s), p(e|t, l), p(d|e, b), p(x|e)\}$$

Brute force computations:

1) Form the joint distribution p(V) by multiplying the CPTs

$$p(V) \leftarrow p(a)p(t|a)p(s)p(t|s)p(b|s)p(e|t,l)p(d|e,b)p(x|e).$$

This gives p(V) represented by a table with giving a table with $2^8=256$ entries.

2) Find the marginal distribution p(a,b,d) by marginalizing p(V)=p(a,t,s,k,e,b,x,d)

$$p(a, b, d) = \sum_{t, s, k, e, b, x} p(t, s, k, e, b, x, d)$$

This is table with $2^3 = 8$ entries.

3) Lastly notice that $p(b|a^+, d^+) \propto p(a^+, b, d^+)$.

Hence from p(a,b,d) we must extract those entries consistent with $a=a^+$ and $d=d^+$ and normalize the result.

Alternatively (and easier): Set all entries not consistent with $a=a^+$ and $d=d^+$ in p(a,b,d) equal to zero.

In chest clinic example the joint state space is $2^8 = 256$.

With 80 variables each with 10 levels, the joint state space is $10^{80} \approx$ the number of atoms in the universe!

Still, **gRain** has been successfully used in a genetics network with 80.000 nodes... How can this happen?

The trick is to NOT to calculate the joint distribution

$$p(V) = p(a)p(t|a)p(s)p(t|s)p(b|s)p(e|t,t)p(d|e,b)p(x|e).$$

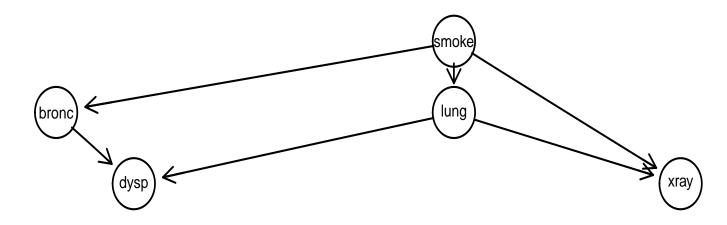
explicitly because that leads to working with high dimensional tables.

Instead we do local computations on on low dimensional tables and "send messages" between them.

The challenge is to organize these local computations. gRain does that for us.

Message passing – excerpt from chest clinic

Consider a small excerpt from the chest clinic example:



```
> yn <- c("yes","no")
> s <- tabNew(~smoke, values=c(5,5),
        levels=yn, normalize="first")
> b.s <- tabNew(~bronc | smoke, values=c(6,4,3,7),
        levels=yn, normalize="first")
> l.s <- tabNew(~lung | smoke, values=c(1,9,1,99),
        levels=yn, normalize="first")
> x.sl <- tabNew(~xray | smoke:lung, values=c(1,0,1,0,.1,.9,0,1),
        levels=yn, normalize="first")</pre>
```

> d.bl <- tabNew(~dysp | bronc:lung, values=c(9,1,7,3,8,2,1,9),
 levels=yn, normalize="first")</pre>

The joint pmf is the product of the cpt's

> p.joint <- s %a*% b.s %a*% l.s %a*% x.sl %a*% d.bl

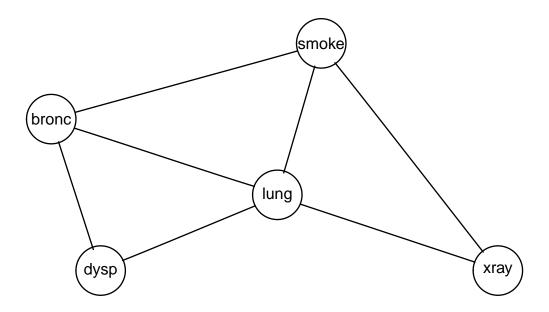
but we DO NOT want to compute the joint pmf in real-world applications. It is prohibitive in terms of storage and computing time.

4.1 Computing clique marginals

Instead we do certain local computations - outlined in the following.

First: moralize dag; then triangulate:

- > g1 <- dg1 %>% moralize %>% triangulate
- > plot(g1)



Next, g1 is the basis of our computations. There are three cliques (maximal complete subsets in g1):

```
> getCliques(g1)  %>% str
List of 3
$ : chr [1:3] "lung" "bronc" "smoke"
$ : chr [1:3] "lung" "bronc" "dysp"
$ : chr [1:3] "lung" "xray" "smoke"
```

Form clique potentials by grouping CPTs (here we do carry out the multiplications)

```
> q1.bsl <- s %a*% b.s
> q2.dbl <- d.bl
> q3.xsl <- l.s %a*% x.sl</pre>
```

Now the clique potentials are the basis of our computations and we can forget about the CPTs (and the DAG).

Joint state-space of pmf is $2^5=32$. The clique potentials are smaller $3\times 2^3=24$. In real-world cases the difference is much larger.

```
> q1.bsl %>% flatten
    smoke yes no
bronc
yes 0.30 0.15
no 0.20 0.35
> q2.dbl %>% flatten
   bronc yes no
    lung yes no yes no
dysp
yes 0.9 0.8 0.7 0.1
no 0.1 0.2 0.3 0.9
> q3.xsl %>% flatten
    smoke yes
            no
    lung yes no yes no
xray
      0.10 0.09 0.01 0.00
yes
        0.00 0.81 0.00 0.99
no
```

The trick in message passing: Manipulate the qs such that they end up containing corresponding distributions.

```
> # From q3 to q1:
> # Marginalize "onto" (s,1):
> # Divide q3
> # Multiply q1 - does not change domain of q1
> q3.sl <- q3.xsl %amarg% ~smoke + lung</pre>
> q3.xs1 <- q3.xs1 %a/0% q3.s1
> q1.bsl <- q1.bsl %a*% q3.sl</pre>
> q1.bsl %>% flatten
      smoke yes
                             no
      lung yes no yes
                                    no
bronc
yes 0.0300 0.2700 0.0015 0.1485
           0.0200 0.1800 0.0035 0.3465
no
```

```
> # From q1 to q2:
> # Work on q1: marginalize "onto" (b, 1)
> # Divide q1
> # Multiply q2 - does not change domain of q2
> q1.bl <- q1.bsl %amarg% ~bronc + lung</pre>
> q1.bsl <- q1.bsl %a/0% q1.bl</pre>
> q2.dbl <- q2.dbl %a*% q1.bl</pre>
> q2.dbl %>% flatten
    bronc yes
                            no
    lung yes no yes no
dysp
yes 0.0284 0.3348 0.0165 0.0527
no 0.0032 0.0837 0.0071 0.4738
> q2.dbl %>% sum # check!
[1] 1
```

no 0.0015 0.1485 0.0035 0.3465

> q1.bsl %>% sum

[1] 1

Empirical proof about clique marginals: In this toy example we can compute the joint pmf:

```
> p.joint <- s %a*% b.s %a*% l.s %a*% x.sl %a*% d.bl
```

Next marginalize and compare with clique potentials:

```
> p1.bsl <- p.joint %amarg% ~bronc + smoke + lung
> p2.dbl <- p.joint %amarg% ~dysp + bronc + lung
> p3.xsl <- p.joint %amarg% ~xray + smoke + lung</pre>
```

Are ps and qs identical?

```
> p1.bsl %a==% q1.bsl
[1] TRUE
> p2.dbl %a==% q2.dbl
[1] TRUE
> p3.xsl %a==% q3.xsl
[1] TRUE
```

4.2 Conditioning

Conditioning is trivial:

Suppose xray='yes'.

- 1. Multiply entries corresponding to xray='no' in the qs by 0.
- 2. Repeat all steps above. Afterwards clique potentials will contain conditionl distributions.

```
> q1.bsl <- s %a*% b.s
> q2.dbl <- d.bl
> q3.xsl <- l.s %a*% x.sl</pre>
```

Now repeat all steps above:

```
> q3.sl <- q3.xsl %amarg% ~smoke + lung</pre>
> q3.xs1 <- q3.xs1 %a/0% q3.s1
> q1.bsl <- q1.bsl %a*% q3.sl</pre>
> q1.bsl %>% flatten
     smoke yes
                             no
     lung yes no yes
                                   no
bronc
yes 0.0300 0.0270 0.0015 0.0000
          0.0200 0.0180 0.0035 0.0000
no
> q1.bl <- q1.bsl %amarg% ~bronc + lung</pre>
> q1.bsl <- q1.bsl %a/0% q1.bl</pre>
> q2.dbl <- q2.dbl %a*% q1.bl</pre>
> q2.dbl %>% flatten
    bronc yes
                            no
    lung yes no yes no
dysp
yes 0.0284 0.0216 0.0165 0.0018
no 0.0032 0.0054 0.0071 0.0162
> q2.db1 <- q2.db1 / sum(q2.db1)
> q2.dbl %>% sum # check!
[1] 1
> q2.bl <- q2.dbl %amarg% ~bronc + lung</pre>
```

```
> q1.bsl <- q1.bsl %a*% q2.bl</pre>
> q1.bsl %>% flatten
     bronc yes
                no
     lung yes no yes no
smoke
yes 0.300 0.270 0.200 0.180
no 0.015 0.000 0.035 0.000
> q1.bsl %>% sum
\lceil 1 \rceil 1
> q1.sl <- q1.bsl %amarg% ~smoke + lung</pre>
> q3.xsl <- q3.xsl %a*% q1.sl</pre>
> q3.xsl %>% flatten
    smoke yes no
    lung yes no yes no
xray
yes 0.50 0.45 0.05 0.00
no 0.00 0.00 0.00 0.00
> q3.xsl %>% sum
[1] 1
```

Empirical proof

```
> p.cond <- tabDist(p.joint, cond=list(xray="yes"))</pre>
> p.cond %>% flatten
      dysp yes
                                             no
      bronc yes
                              no
                                            yes
                                                           no
      lung yes
                       no
                             yes
                                     no
                                            yes
                                                    no
                                                          yes
                                                                  no
smoke
            0.2700 0.2160 0.1400 0.0180 0.0300 0.0540 0.0600 0.1620
yes
            0.0135 0.0000 0.0245 0.0000 0.0015 0.0000 0.0105 0.0000
no
Next marginalize and compare with clique potentials:
> p1.bsl <- p.cond %amarg% ~bronc + smoke + lung
> p2.dbl <- p.cond %amarg% ~dysp + bronc + lung
> p3.xsl <- p.cond %amarg% ~ smoke + lung</pre>
```

Are ps and qs identical?

```
> p1.bsl %a==% q1.bsl
[1] TRUE
> p2.dbl %a==% q2.dbl
[1] TRUE
> p3.xsl %a==% (q3.xsl %amarg% ~smoke + lung)
[1] TRUE
```

5 An introduction to the **gRain** package

5.1 Specify BN from list of CPTs

Specify chest clinic network. Can be done in many ways; one is from a list of CPTs:

```
> cpt.list <- compileCPT(list(a, t.a, s, l.s, b.s, e.lt, x.e, d.be))
> cpt.list
cpt_spec with probabilities:
P( asia )
P( tub | asia )
P( smoke )
P( lung | smoke )
P( bronc | smoke )
P( either | lung tub )
P( xray | either )
P( dysp | bronc either )
```

```
> cpt.list$asia
asia
yes no
0.01 0.99
> cpt.list$tub
    asia
tub yes no
 yes 0.05 0.01
 no 0.95 0.99
> ftable(cpt.list$either, row.vars=1) # Notice: logical variable
      lung yes
                   no
      tub yes no yes no
either
yes
no
```

5.2 Building network

```
> # Create network from CPT list:
> bn <- grain(cpt.list)
> # Compile network (details follow)
> bn <- compile(bn)
> bn
Independence network: Compiled: TRUE Propagated: FALSE
   Nodes: chr [1:8] "asia" "tub" "smoke" "lung" "bronc" "either" "xray"
```

5.3 Querying the network

```
> # Query network to find marginal probabilities of diseases
> disease <- c("tub", "lung", "bronc")</pre>
> bn %>% qgrain(nodes=disease)
$tub
tub
yes no
0.01 0.99
$lung
lung
 yes no
0.055 0.945
$bronc
bronc
yes no
0.45 0.55
```

5.4 Setting evidence

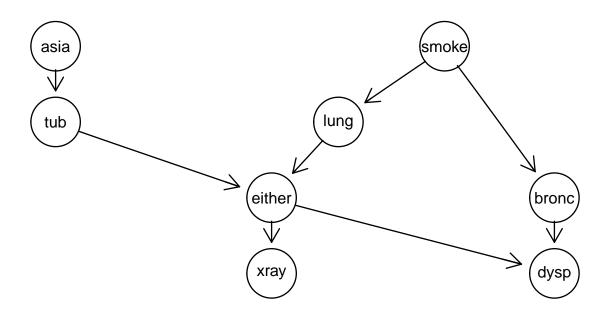
```
> # Set evidence and query network again
> bn.ev <- bn %>% setEvidence(evi=list(asia="yes", dysp="yes"))
> bn.ev %>% qgrain(nodes=disease)
$tub
tub
  yes
      no
0.088 0.912
$lung
lung
yes no
0.1 0.9
$bronc
bronc
yes no
0.81 0.19
```

A little shortcut: Most uses of **gRain** involves 1) setting evidence into a network and 2) querying nodes. This can be done in one step:

```
> qgrain(bn,
    evidence=list(asia="yes", dysp="yes"),
    nodes=disease)
$tub
tub
 yes
      no
0.088 0.912
$lung
lung
yes no
0.1 0.9
$bronc
bronc
yes
       no
0.81 0.19
```

5.5 Specify BN from DAG and data**

If the structure of the DAG is known and we have data, we can just do:

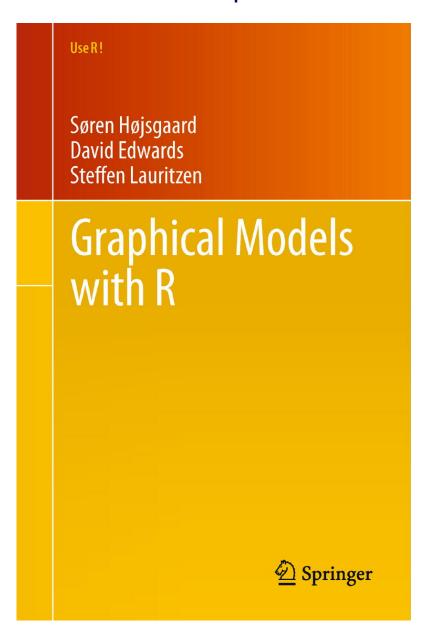


```
> data(chestSim1000, package="gRbase")
> head(chestSim1000)
 asia tub smoke lung bronc either xray dysp
   no
       no
            no
                 no
                     yes
                            no
                                 no
                                    yes
   no
      no
          yes no
                   yes
                                 no yes
                            no
3
                   no
                            no no
   no
      no
          yes no
                                   no
4
                            no no no
   no
      no
         no no no
5
  no no
         yes no yes
                            no no yes
   no
       no
           yes yes yes yes
                                    yes
> bn2 <- grain(dg, data=chestSim1000, smooth=.1)</pre>
> bn2
Independence network: Compiled: TRUE Propagated: FALSE
 Nodes: chr [1:8] "asia" "tub" "smoke" "lung" "bronc" "either" "xray"
```

The CPTs are estimated as the relative frequencies.

6 Wrapping up

6.1 Book: Graphical Models with R



6.2 Package versions

We shall in this tutorial use the R-packages gRbase, gRain and gRim.

Go to http://people.math.aau.dk/~sorenh/software/gR for installation instructions.

The tutorial is based on these versions of the packages (which are available on github):

```
> packageVersion("gRbase")
[1] '1.8.6.9001'
> packageVersion("gRain")
[1] '1.3.6'
> packageVersion("gRim")
[1] '0.2.5'
```