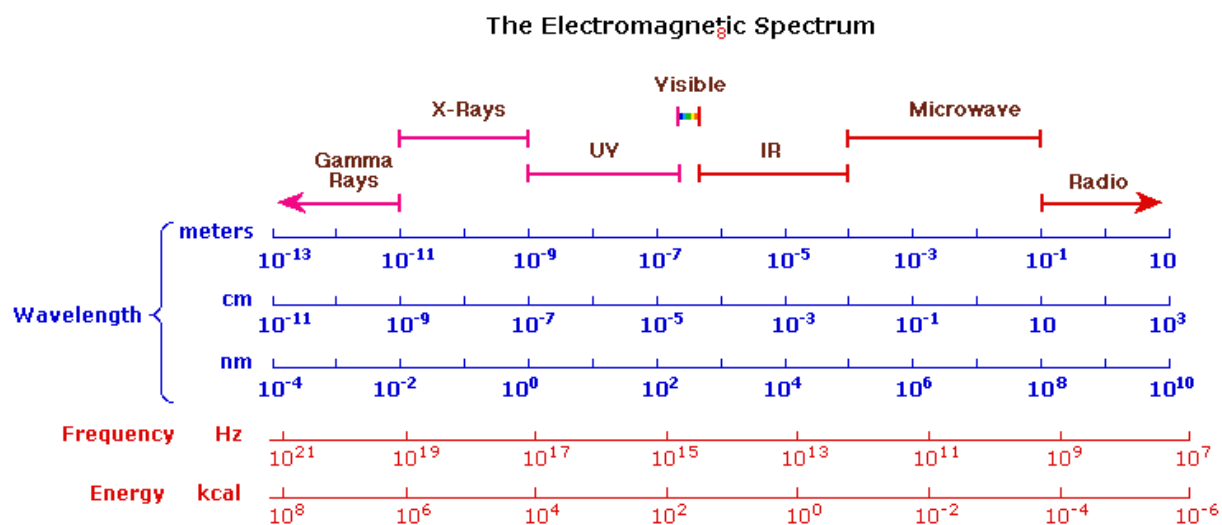


1. UV- Visible spectroscopy

The Electromagnetic Spectrum

The visible spectrum constitutes a small part of the total radiation spectrum. Most of the radiation that surrounds us cannot be seen, but can be detected by dedicated sensing instruments. This electromagnetic spectrum ranges from very short wavelengths (including gamma and x-rays) to very long wavelengths (including microwaves and broadcast radio waves). The following chart displays many of the important regions of this spectrum and demonstrates the inverse relationship between wavelength and frequency (shown in the top equation below the chart).



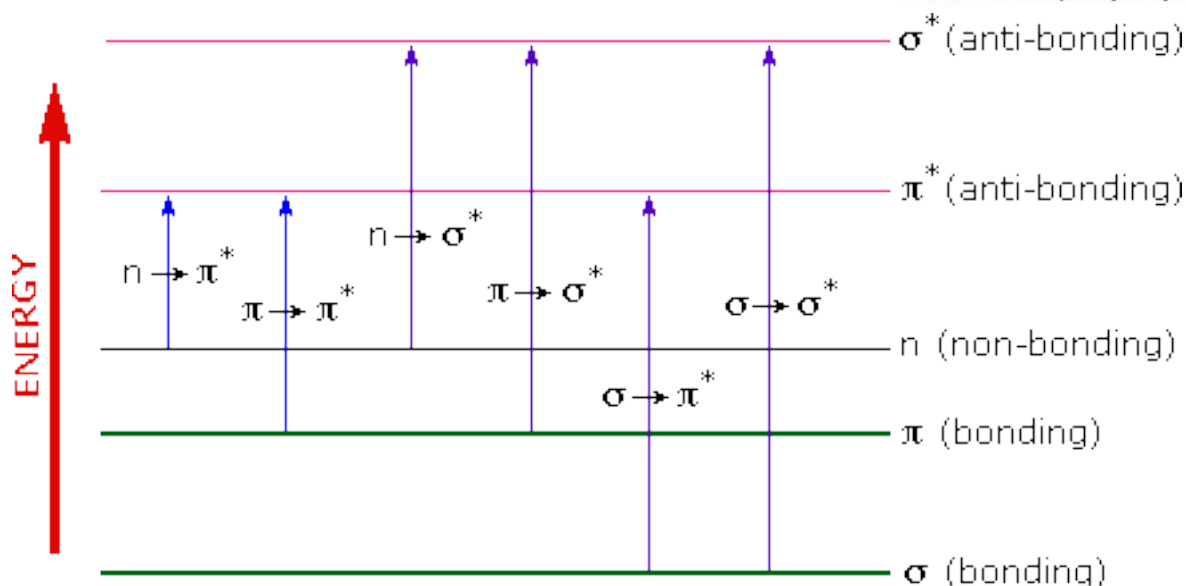
The energy associated with a given segment of the spectrum is proportional to its frequency. The equation below describes this relationship, which provides the energy carried by a photon of a given wavelength of radiation.

$\nu = c/\lambda$ ν =frequency, λ =wavelength, c =velocity of light ($c=3 \cdot 10^{10}$ cm/sec)
 $\Delta E = h\nu$ E =energy, ν =frequency, h =Planck's constant ($h=6.6 \cdot 10^{-27}$ erg sec)

Electronic Transition

The visible region of the spectrum comprises photon energies of 36 to 72 kcal / mole, and the near ultraviolet region, out to 200 nm, extends this energy range to 143 kcal / mole. Ultraviolet radiation having wavelengths less than 200 nm is difficult to handle, and is seldom used as a routine tool for structural analysis.

The energies noted above are sufficient to promote or excite a molecular electron to a higher energy orbital. Consequently, absorption spectroscopy carried out in this region is sometimes called "electronic spectroscopy".



A diagram showing the various kinds of electronic excitation that may occur in organic molecules is shown above. Of the six transitions outlined, only the two lowest energy ones (leftmost, colored blue) are achieved by the energies available in the 200 to 800 nm spectrum. As a rule, energetically favored electron promotion will be from the highest occupied molecular orbital (HOMO) to the lowest unoccupied molecular orbital (LUMO), and the resulting species is called an excited state.

When sample molecules are exposed to light having an energy that matches a possible electronic transition within the molecule, some of the light energy will be absorbed as the electron is promoted to a higher energy orbital. An optical spectrometer records the wavelengths at which absorption occurs, together with the degree of absorption at each wavelength. The resulting spectrum is presented as a graph of absorbance (A) versus wavelength, as in the isoprene spectrum shown below. Since isoprene is colorless, it does not absorb in the visible part of the spectrum and this region is not displayed on the graph. Absorbance usually ranges from 0 (no absorption) to 2 (99% absorption), and is precisely defined in context with spectrometer operation.

When a beam of light passes through an absorbing medium for example a solution, a part of the light is absorbed and the rest is transmitted. The amount of light absorbed depends on the concentration of the solution and the length traversed by the light through the solution. The quantitative relation between the amount of light absorbed, concentration and the length of the absorbing medium is governed by the laws- the Lambert's Law, the Beer's Law.

Lambert's Law

The law states that “Equal fractions of the incident light are absorbed by successive layers of equal thickness of the absorbing medium”.

If a monochromatic light passes through a transparent medium the rate of decrease in intensity with the thickness of medium is proportional to intensity of the incident light i.e.

$$-dI/I \propto dl$$

$$dI/I = -k_1 dl \text{-----(i)}$$

$$-\int_{I_0}^{I_t} \frac{dI}{I} = K \int_0^l C dl$$

Where I is intensity of the incident light of wavelength λ , l is the thickness of the medium and k_1 is proportionality factor. The negative sign indicates that due to absorption, the intensity of light decreases as it passes through the absorbing medium.

If I_0 is the initial intensity of incident light on the absorbing medium of length l and I_t is the intensity of transmitted beam then the integration of eqn (i) becomes

$$\ln(I_t / I_0) = -k_1 \cdot l$$

$$\ln(I_0 / I_t) = k_1 \cdot l$$

$$\log_{10}(I_0 / I_t) = \frac{k_1 \times l}{2.303}$$

Beer's Law

The law states that “Equal fractions of the incident light are absorbed by successive layers having equal concentration of the absorbing medium”.

The intensity of a beam of monochromatic light decreases exponentially as the concentration of the absorbing substance increases arithmetically. i.e.

$$-dI/I \propto dc$$

$$dI/I = -k_2 dc \text{-----(iii)}$$

$$-\int_{I_0}^{I_t} \frac{dI}{I} = K \int_0^c C dc$$

If I_0 is the initial intensity of incident light on the absorbing medium of concentration c and I_t is the intensity of transmitted beam then the integration of eqn (iii) becomes

$$\ln(I_t / I_0) = -k_2 \cdot c$$

$$\ln(I_0 / I_t) = k_2 \cdot c$$

$$\log_{10}(I_0 / I_t) = \frac{k_2 \times c}{2.303} \text{-----(iv)}$$

Beer- Lambert's law:

A combination of Lambert's Law and Beer's Law results in Beer- Lambert's Law it states that "Equal fractions of the incident light are absorbed by successive layers of equal thickness and equal concentration of the absorbing medium"

Combining (ii) and (iv),

$$\log_{10}(I_0/I_t) = \frac{k}{2.303} \times c \times l$$

$$\log_{10}(I_0/I_t) = \epsilon c l$$

$\log(I_0/I_t)$ is called as absorbance, if c is expressed in mol dm^{-3} and l in cm

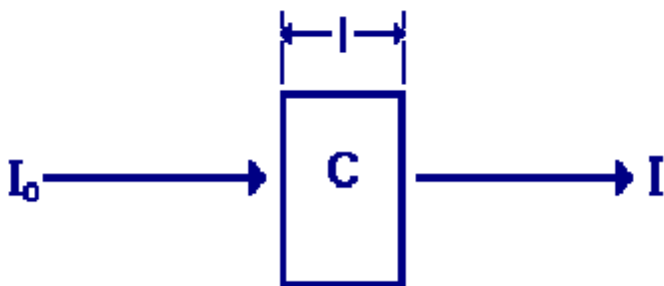
then ϵ is called as molar absorptivity or absorption coefficient.

Hence,

$$A = \epsilon c l = \log(I_0/I_t) = \log(1/T) = -\log(T)$$

where T is transmittance.

Conditions: The law is only true for monochromatic light, which is light of a single wavelength or narrow band of wavelengths, and provided that the physical or chemical state of the substance does not change with concentration.

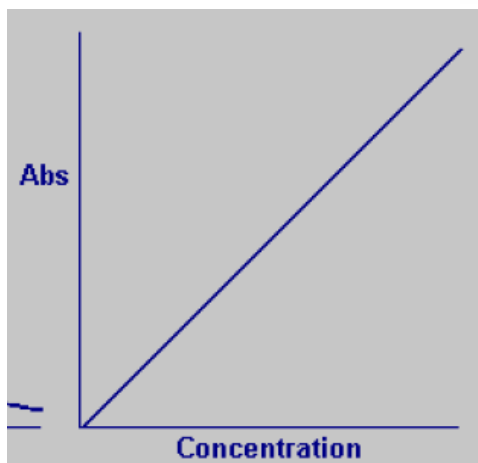


I_0 is the intensity of the incident radiation and I is the intensity of the transmitted radiation. The ratio I/I_0 is called transmittance. This is sometimes expressed as a percentage and referred to as %transmittance.

Mathematically, absorbance is related to percentage transmittance T by the expression:

$$A = \log_{10}(I_0/I) = \log_{10}(100/T) = \epsilon \times c \times L$$

Where, L is the length of the radiation path through the sample, c is the concentration of absorbing molecules in that path, and ϵ is the molar extinction coefficient - a constant dependent only on the nature of the molecule and the wavelength of the radiation. If concentration is expressed in mol/dm^3 and L is expressed in dm then unit for ϵ is dm^2/mol .



Absorbance vs concentration

Limitations of the Beer-Lambert law

1. The linear relationship between absorbance and concentration of solution is not observed at concentration above 10^{-2} M hence concentrated solution do not obey Beer-Lambert's Law
2. The law does not obeyed if the absorbing species reacts with the solvent, dissociates or associates in solution. The molecules of the absorbing species should remain as simple molecules and should not undergo any change in molecular condition.
3. The light incident on the absorbing medium should be monochromatic otherwise minor deviations from Beer-Lambert's Law are observed. Hence monochromators have to be used to produce monochromatic beams.
4. The molar absorptivity depends on the refractive index of the absorbing medium. The refractive index changes with the concentration of the absorbing medium. At high concentration these changes are considerable but at concentrations below 10^{-2} M these changes can be neglected.
5. Temperature fluctuations and entry of stray light into the absorbing system also lead to deviations from Beer-Lambert's law.
6. The Fluorescence or phosphorescence can cause a positive deviation in % T and negative deviation for A in the system.
7. Non-monochromatic radiation, deviations can be minimized by using a relatively flat part of the absorption spectrum such as the maximum of an absorption band.

Ultraviolet-visible spectrophotometer

The instrument used in ultraviolet-visible spectroscopy is called a UV/Vis spectrophotometer. It measures the intensity of light passing through a sample (I), and compares it to the intensity of light before it passes through the sample (I_0). The ratio I/I_0 is called the transmittance, and is usually expressed as a percentage (%T). The absorbance, A, is based on the transmittance:

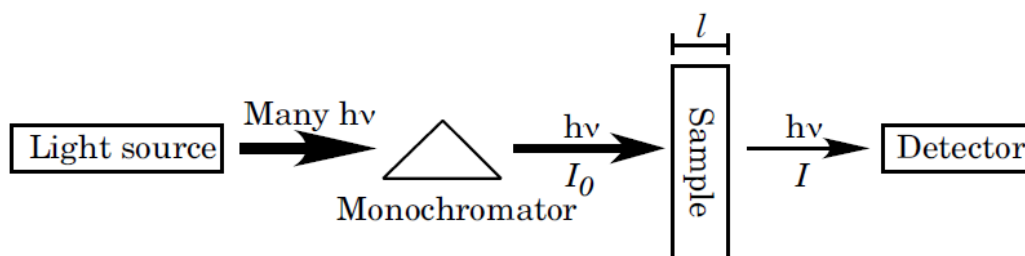
$$A = -\log(\%T/100\%)$$

The UV- visible spectrophotometer can also be configured to measure reflectance. In this case, the spectrophotometer measures the intensity of light reflected from a sample (I), and compares it to the intensity of light reflected from a reference material (I_0) (such as a white tile). The ratio I/I_0 is called the reflectance, and is usually expressed as a percentage (%R).

The basic parts of a spectrophotometer are a light source, a holder for the sample, a diffraction grating in a monochromator or a prism to separate the different wavelengths of light, and a detector.

Single beam spectrophotometer

A single beam spectrophotometer is comprised of a light source, a monochromator, a sample holder, and a detector. An ideal instrument has a light source that emits with equal intensity at all wavelengths, a monochromator that is equally efficient in splitting light into narrow groups of wavelength for all wavelengths, and a detector that is sensitive and responds equally to all wavelengths.



Light sources

Because no single light source with the appropriate characteristics exists, most spectrophotometers use two lamps, with one for the ultraviolet region and one for the visible region. The visible lamp is usually a tungsten lamp (300- 2500 nm), while the ultraviolet lamp is a deuterium lamp (190- 400 nm). An alternative, relatively rarely used in spectrophotometers, although commonly used in other types of spectroscopic instruments, is a xenon arc lamp (160- 2,000 nm).

Monochromators

Although prisms can be used as monochromators, most instruments use diffraction gratings. Light shining on the closely spaced grooves of a diffraction grating at an angle is separated into different wavelengths in a consistent manner, assuming that the grooves are consistently produced.

Detector

The detector is typically a photomultiplier tube, a photodiode, a photodiode array or a charge-coupled device (CCD). Single photodiode detectors and photomultiplier tubes are used with scanning monochromators, which filter the light so that only light of a single wavelength reaches the detector at one time.

The most commonly used detector is a photomultiplier tube (PMT). An incoming photon hits a thin metal film inside a vacuum tube. The metal film is maintained at a large negative potential, and emits electrons. These collide with a series of dynodes maintained at progressively lower potentials; each dynode emits several electrons in response to each incoming electron, resulting in a large amplification of the signal. An alternative type of detector uses photodiodes. Photodiodes are inexpensive but not very sensitive. Their low cost has allowed arrays of photodiodes to be set up to allow simultaneous detection of many wavelengths. A charge coupled device (CCD) is a sensitive array detector. CCDs store charges released in response to photon impacts.

Cuvettes (Sample Holder)

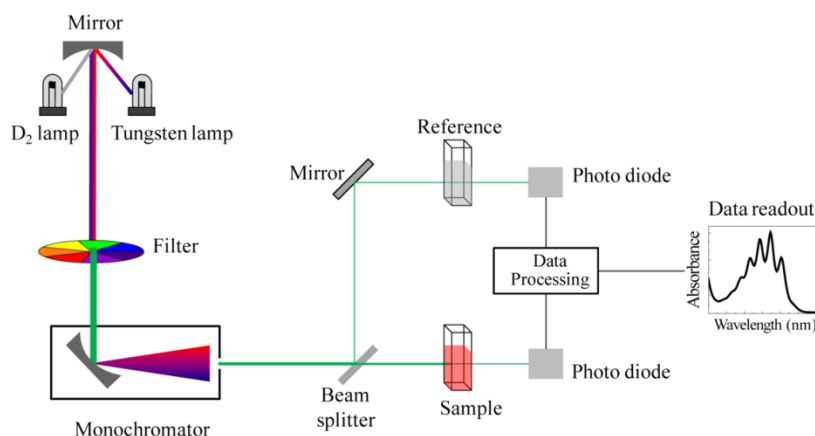
Most samples studied using visible and ultraviolet spectroscopy is liquid. The sample must therefore be placed in a transparent container to allow measurement. These containers are called cuvettes. Cuvettes are generally made from transparent plastic, glass, or quartz. Different cuvettes have different optical properties.

Working: The sample holder is filled with the solvent (blank). The absorbance value of the solvent is adjusted to zero for a particular wavelength, obtained by the rotation of the monochromator. The sample is then taken in the sample holder and its absorbance value is determined for the same wavelength. The procedure is repeated for different wavelengths obtained by the rotation of the monochromator. The absorbance values can be read either on a dial or a digital display. The λ_{max} values for the sample can be thus found.

Double Beam Spectrophotometer

A beam of visible light from a incandescent tungsten lamp passes through a colour filter which selects a narrow band of wavelengths. A mirror splits this narrow band into two beams - one passing through the sample and the other passing through the solvent (blank) hence the name Double beam Spectrophotometer.

The sample absorbs a part of beam whereas the solvent transmits it completely. The two beams then fall on the respective photocells, where photoelectrons are emitted and recorded.



Working: The solvent is first taken in both cuvettes, and zero level adjusted. The sample is then placed in the sample cuvette. It absorbs a part of the light and the transmitted beam emerging from it falls on the photodiode. This beam has less intensity than when only solvent was present in the sample cuvette. Hence there will be proportionate decrease in the electric current produced in photodiode and recorded.

Advantages of Double beam spectrophotometer over single beam spectrophotometer:

1. Changes in the intensity of incident light due to voltage fluctuations in the power supply are compensated by splitting the incident beam into identical beams by the use of a mirror and two balanced photodiodes. Any error due to solvent or impurity is balanced out as identical beams pass through the blank and sample as the absorbance of blank is initially adjusted to zero.
2. The readings are not affected by changes in sensitivity of photodiodes as the zero method is used.
3. The scale of instrument is linear with the concentration of the sample solution.

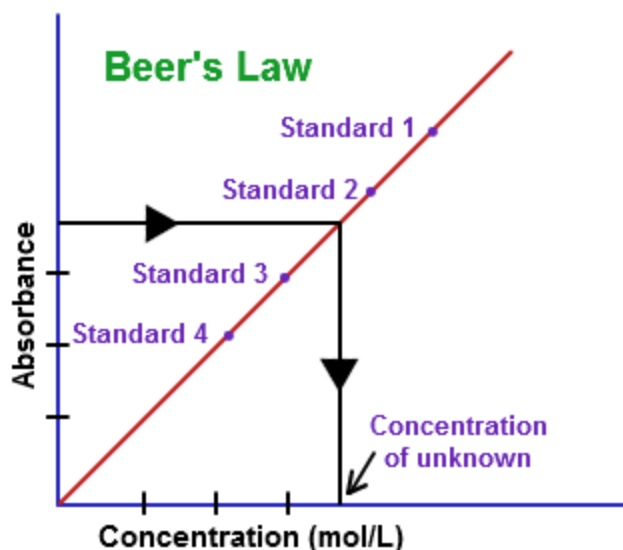
Applications:

- 1) **Determination of λ_{max} and identifying the functional groups:** It is possible to identify a particular group in a molecule by determining its λ_{max} value. In spectrophotometers, the beams of characteristic wavelength are produced, these are absorbed by sample, which are indicative of functional groups as different functional groups in the molecule absorb their own λ_{max} values.

Function group	Example	λ_{max} in nm	Solvent
- COOH	Acetic acid	208	Ethyl alcohol
- COCl	Acetyl chloride	220	Hexane
- CONH ₂	Acetamide	178	Hexane
- NO ₂	Nitromethane	201	Methyl alcohol

- 2) **Quantitative Analysis (Calibration curve method):** UV-Vis spectroscopy is routinely used in analytical chemistry for the quantitative determination of different analytes, such as transition metal ions, highly conjugated organic compounds, and biological macromolecules. Spectroscopic analysis is commonly carried out in solutions but solids and gases may also be studied.

In this, a series of solutions of different concentrations of the compound is prepared. Usually the concentrations vary from 10^{-3} M to 10^{-2} M. The absorbance is then determined for these standard solutions at the wavelength which absorbed maximally by the coloured compound (λ_{\max}). A graph of absorbance versus the concentration is then plotted.



From the calibration curve the concentration of the unknown solution can be found out as shown.

- 3) **Photometric titrations:** Spectrophotometric measurements can be employed to advantage in locating the equivalence point of a titration, provided the analyte, reagent or titration product absorbs radiation. Alternatively absorbing indicator can provide the absorbance change necessary for the location of equivalence point.
- 4) A UV-Vis spectrophotometer may be used as a detector for HPLC. The presence of an analyte gives a response assumed to be proportional to the concentration. For accurate results, the instrument's response to the analyte in the unknown should be compared with the response to a standard; this is very similar to the use of calibration curves. The response (e.g., peak height) for a particular concentration is known as the response factor.
- 5) UV-Vis spectroscopy is also used in the semiconductor industry to measure the thickness and optical properties of thin films on a wafer. UV-Vis spectrometers are used to measure the reflectance of light, and can be analyzed via the Forouhi-Bloomer dispersion equations to determine the Index of Refraction (n) and the Extinction Coefficient (k) of a given film across the measured spectral range.

2 pH Metry

Definition of pH

pH is an abbreviation of “pondus hydrogenii” and was proposed by the Danish scientist S. P. L. Sørensen in 1909 in order to express the very small concentrations of hydrogen ions.

In 1909, pH was defined as the negative base 10 logarithm of the hydrogen ion concentration. However, as most chemical and biological reactions are governed by the hydrogen ion activity, the definition was quickly changed. As a matter of fact, the first potentiometric methods used actually resulted in measurements of ion activity.

The definition based on hydrogen ion activity is the definition we use today:

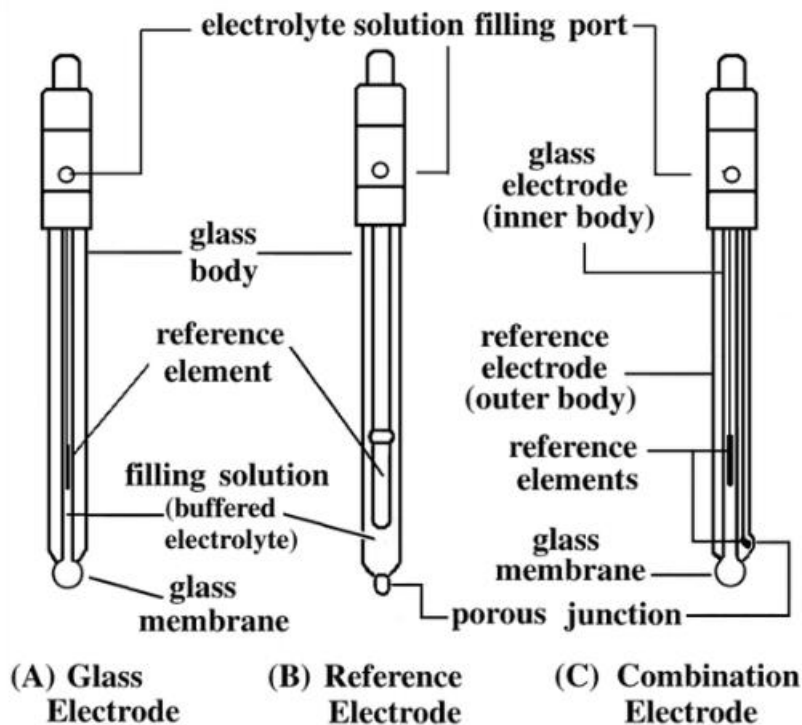
$$\text{pH} = -\log_{10} a_{\text{H}^+}$$

pH Theory:

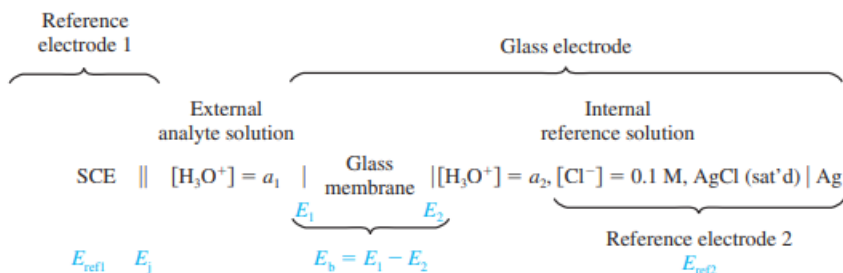
pH is measured using a setup with two electrodes: the indicator electrode and the reference electrode. These two electrodes are often combined into one - a combined electrode. When the two electrodes are immersed in a solution, a small galvanic cell is established. The potential developed is dependent on both electrodes. Ideal measuring conditions exist when only the potential of the indicator electrode changes in response to varying pH while the potential of the reference electrode remains constant.

Glass Electrode:

Construction: It consists of a glass bulb membrane, which gives it its name and an electrically insulating tubular body, which separates an internal solution and a silver/silver chloride electrode from the studied solution. The Ag/AgCl electrode is connected to a lead cable terminated with some connector that can hook up to a special voltmeter, the pH meter.



It is represented as



As shown in Figure, the boundary potential consists of two potentials, E_1 and E_2 , each of which is associated with one of the two glass surfaces. The boundary potential is simply the difference between these potentials:

$$E_b = E_1 - E_2$$

If the two membrane surfaces have the same number of negatively charged sites (as they normally do) from which H^+ can dissociate,

we find that

$$E_b = E_1 - E_2$$

$$= 0.0592 \log a_1 / a_2$$

Thus, the boundary potential E_b depends only on the hydrogen ion activities of the solutions on either side of the membrane. For a glass pH electrode, the hydrogen ion activity of the internal solution a_2 is held constant

The Asymmetry Potential (E_{asy}): When identical solutions are placed on the two sides of a glass membrane, the boundary potential should in principle be zero (see Figure 23-6b). In fact, however, we frequently encounter a small asymmetry potential that changes gradually with time. The sources of the asymmetry potential

The Glass Electrode Potential: As noted earlier, the potential of a glass indicator electrode E_{ind} has three components: (1) the boundary potential, (2) the potential of the internal Ag/AgCl reference electrode E_{ref} , and (3) the small asymmetry potential E_{asy} .

$$E_{ind} = E_b + E_{ref2} + E_{asy}$$

$$E_{ind} = L' + 0.0592 \log a_1 + E_{ref2} + E_{asy}$$

or

$$E_{ind} = L + 0.0592 \log a_1 = L - 0.0592 \text{ pH}$$

where L is a combination of the three constant terms. That is,

$$L = L' + E_{ref2} + E_{asy}$$

Working: The pH meter measures the potential difference and its changes across the glass membrane. The potential difference must be obtained between two points; one is the electrode contacting the internal solution. A second point is obtained by connecting to a reference electrode, immersed in the studied solution.

Reference Electrode:

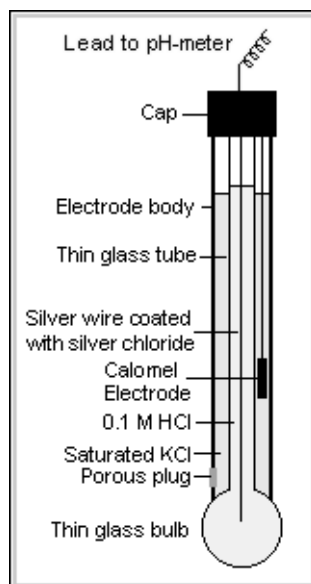
The reference electrode is a silver wire coated with silver chloride in contact with a defined electrolyte solution, see above figure. In many reference electrodes a gel is used instead of a liquid as the internal filling. These gels also contain KCl to maintain the

reference potential and add sufficient conductivity. As described in 2.2 the reference potential is constant as long as the internal electrolyte is constant. The reference potential also varies slightly with temperature.

The tube containing all the reference elements and solutions/gel is in contact with the sample to measure through a junction (diaphragm). It is essential to maintain a free flow of ions through the junction. Otherwise the reference electrode will not respond properly to pH changes in the sample.

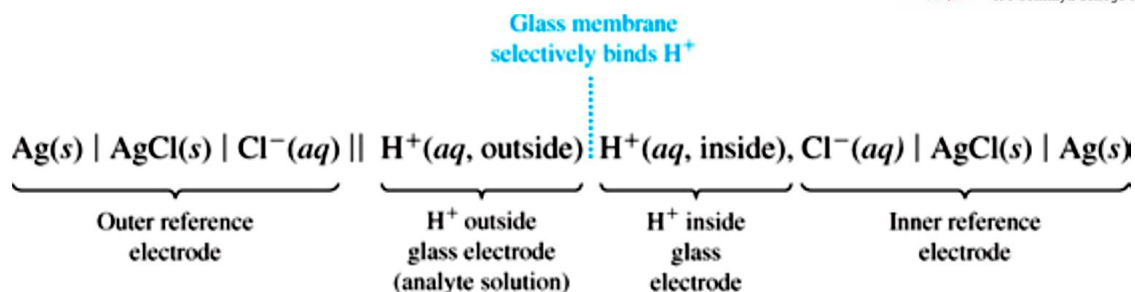
$$E_{ref} = E_0 - 0.59 \times \log [Cl^-]$$

Combination Glass Electrode:



Construction: The fig shows the internal components of the pH electrode. The heart of the electrode is a thin bulb of pH sensitive glass, which is blown out of the end of a length of glass tubing. The pH sensitive glass (glass membrane) is sealed to the electrode and contains a solution of potassium chloride at pH 7. A pH measurement system consists of a pH probe, reference probe, temperature sensor, pH meter and the sample to be measured. In Combined glass electrode the three probes are combined in one electrode. When the pH probe is in contact with a solution a potential forms between the pH probe and the reference probe. The meter measures the potential and converts it, using the calibration curve parameters, into a pH value.

A typical combination glass electrode is represented as



Working: The outside surface of the glass membrane is in contact with sample being measured and the inside surface contacts the filling solution. A complex mechanism at each glass liquid interface defines the potential of the pH glass electrode, while the inner pH glass/ filling solution potential is constant, the outside potential varies based on the H^+ ions concentration in sample. This equilibrium depends also on temperature.

$$E_{ind} = L' + 0.0592 \log a_1 + E_{ref2} + E_{asy}$$

or

$$E_{ind} = L + 0.0592 \log a_1 = L - 0.0592 \text{pH}$$

To Measure pH of solution i.e. Measurement of pH

In order to measure the pH of a sample first the standardization of pH meter is required to be done before analyzing the sample.

a) Standardization of pH meter:

A two point standardization method is used to standardize the pH meter, it involves immersing the pH assembly i.e. glass electrode into a standard reference pH buffer (pH=4.0) and recording the reading if the meter reading is more or less than the expected value (4.0) then it is adjusted to pH 4.0 using a screw knob.

Standardization at only one pH value does not assure the validity of reading at other pH values considerably. Hence a second standard reference buffer pH = 9.2 is used. The pH meter reading is recorded using this second buffer solution and the reading is adjusted to pH 9.2 using a screw knob.

During both the steps, the glass electrode is rinsed with distilled water. Immerse the glass electrode previously in water for several hours. Start the measurement more than 5 minutes after switching on. Rinse well the detecting unit with water, and blot the water gently with a piece of filter paper very fine.

b) To measure pH of solution:

Wash well the detecting unit with water, and blot the water gently with a piece of filter paper. Place glass electrode in solution you wish to measure pH. Be sure that it is stirring slowly during measure and pH adjustment and take readings.

c) Precautions:

When analysis is complete, put pH meter in stand-by mode. Remove electrode from solution and rinse thoroughly with water. Blot dry and put back in yellow pH storage buffer. Place parafilm over the hole and around the bottle to minimize evaporation.

3. Conductometry

Conductometry is a general method, where two electrode systems are used for simultaneous measurement of all electroactive compounds in a solution. It is a measurement of electrolytic conductivity to monitor a progress of chemical reaction.

Conductometric Titration:

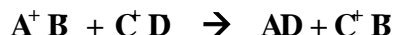
Conductance is a property of solutions of electrolytes. It is the measure of the number of ions present in the solution, as well as the current carrying capacity of the ions. When the solution contains one single electrolyte, the measured conductance of the solution can be related to concentration of the electrolyte.

Conductometric titration is a type of titration in which the electrolytic conductivity of the reaction mixture is continuously monitored as one reactant is added. The equivalence point is the point at which the conductivity undergoes a sudden change. Marked increase or decrease in conductance is associated with the changing concentrations of the two most highly conducting ions, viz. the hydrogen and hydroxyl ions. The method can be used for titrating coloured solutions or homogeneous suspension e.g.: wood pulp suspension, which cannot be used with normal indicators.

Principle:

When solution of one electrolyte is added to another electrolyte without appreciable volume change, the conductance of the solution will alter, if an ionic reaction occurs. If no ionic reaction takes place then the conductance of the solution will simply increase. On the other hand, if an ionic reaction occurs, the ion added may replace another ion and hence bring about change in the conductance.

Let A^+B^- be the ions of titrand and C^+D^- be ions of the titrant, the ionic reaction in the titration is combination of A^+ and D^- , AD formed may be insoluble or weakly ionized.



Thus as the titration proceeds, A^+ are replaced by C^+ . The conductance of the solution increases or decreases depending on whether conductance of C^+ is greater than or less than that of A^+ . After equivalence point the ionic reaction does not occur and hence, the conductance of the solution will raise due to the excess addition of titrant C^+D^- .

The principle of conductometric titration is changes in the conductance of the solution due to difference in the ionic conductance or due to production of more number of ions in the solution. Both factors permit, location of the equivalence point by conductance measurements.

Procedure:

A definite volume of the solution to be estimated is pipetted out in a beaker. A dip type conductivity cell is placed in a beaker. Addition of distilled water may be necessary if the cell does not dip completely in the solution. The cell is connected to a conductometer and the conductance of the solution is measured. The titrant is filled in the burette. The titrant is added in

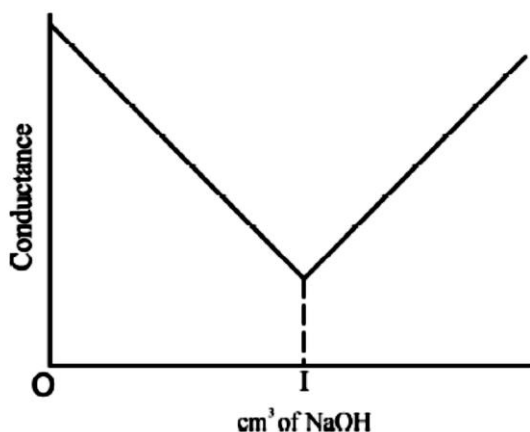
the small portions, generally 0.5 mL at a time. The solution is stirred after each addition. The solution is allowed to stand for a minute or two after stirring before conductance is measured. Addition of titrant is continued till about seven to eight readings beyond the equivalence point are obtained. The plot of conductance against volume of the titrant added is used to locate the equivalence point.

Titration Curves:

Some Typical Conductometric Titration Curves are:

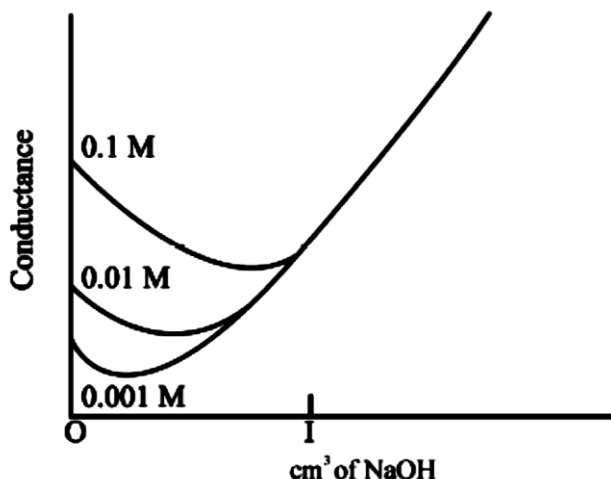
1) Strong Acid with a Strong Base, [HCl Vs NaOH]

Before NaOH is added, the conductance is high due to the presence of highly mobile hydrogen ions. When the base is added, the conductance falls due to the replacement of hydrogen ions by the added cations as H^+ ions react with OH^- ions to form undissociated water. This decrease in the conductance continues till the equivalence point. At the equivalence point, the solution contains only NaCl. After the equivalence point, the conductance increases due to the large conductivity of OH^- ions.



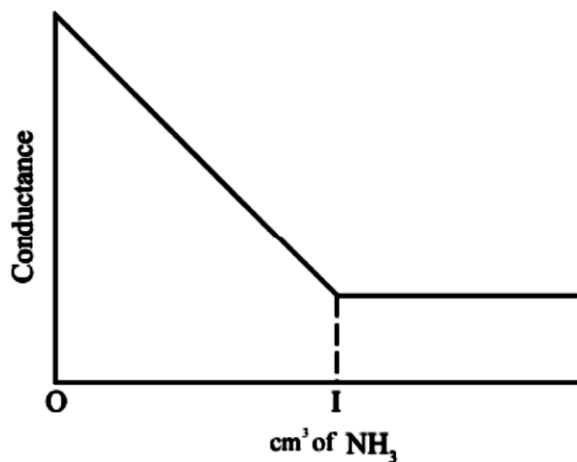
Conductometric titration of a strong acid (HCl) vs. a strong base (NaOH)

- 2) **Weak Acid with a Strong Base, [CH₃COOH Vs NaOH]** Initially the conductance is low due to the feeble ionization of acetic acid. On the addition of base, there is a decrease in conductance not only due to the replacement of H^+ by Na^+ but also suppresses the dissociation of acetic acid due to the common ion acetate. But very soon, the conductance increases on adding NaOH as NaOH neutralizes the undissociated CH₃COOH to CH₃COONa which is the strong electrolyte. This increase in conductance continues to rise up to the equivalence point. The graph near the equivalence point is curved due to the hydrolysis of salt CH₃COONa. Beyond the equivalence point, conductance increases more rapidly with the addition of NaOH due to the highly conducting OH^- ions.



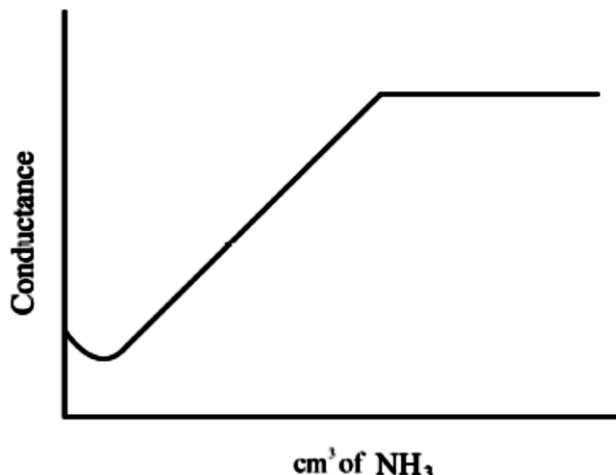
Conductometric titration of a weak acid (acetic acid) vs. a strong base (NaOH)

- 3) **Strong Acid with a Weak Base, e.g. sulphuric acid with dilute ammonia:** Initially the conductance is high and then it decreases due to the replacement of H^+ . But after the endpoint has been reached the graph becomes almost horizontal, since the excess aqueous ammonia is not appreciably ionised in the presence of ammonium sulphate.



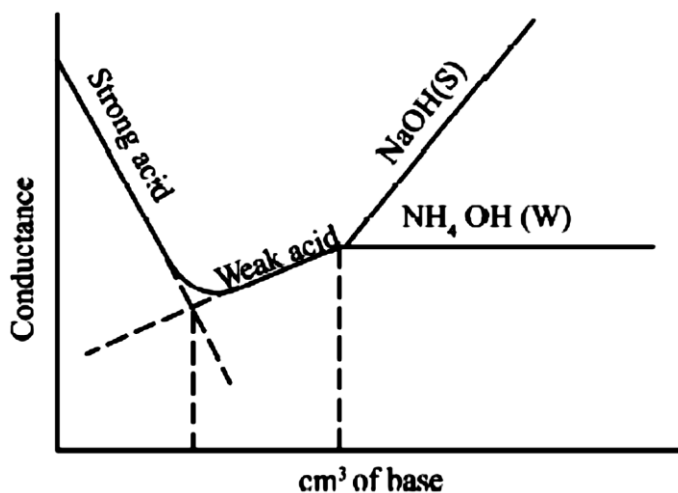
Conductometric titration of a strong acid (H_2SO_4) vs. a weak base (NH_4OH)

- 4) **Weak Acid with a Weak Base:** The nature of curve before the equivalence point is similar to the curve obtained by titrating weak acid against strong base. After the equivalence point, conductance virtually remains same as the weak base which is being added is feebly ionized and, therefore, is not much conducting.



Conductometric titration of a weak acid (acetic acid) vs. a weak base (NH₄OH)

- 5) **Mixture of a Strong Acid and a Weak Acid vs. a Strong Base or a Weak Base:** In this curve there are two break points. The first break point corresponds to the neutralization of strong acid. When the strong acid has been completely neutralized only then the weak acid starts neutralizing. The second break point corresponds to the neutralization of weak acid and after that the conductance increases due to the excess of OH⁻ ions in case of a strong base as the titrant. However, when the titrant is a weak base, it remains almost constant after the end point similar to Fig



Conductometric titration of a mixture of a strong acid (HCl) and a weak acid (CH₃COOH) vs. a strong base (NaOH) or a weak base (NH₄OH)

Advantages of Conductometric titrations:

Conductometric titration are found to possess several advantages over normal titrimetry namely,

1. Coloured solutions can be titrated
2. The method works equally well for dilute solutions also, as it is based on the changes in the conductance, rather than the absolute value of the conductance.
3. In conductometric titrations, it is not necessary to make observations around the equivalence point, with small increments of the titrant added. The observations, far away from the equivalence point, on either side are of importance.
4. An extremely weak acid or a weak base can be titrated conductometrically, which may not be possible in normal titrimetry.
5. A mixture of weak and strong acids, can also be titrated with relative ease. Thus, making the simultaneous determination possible.

Limitations:

1. In dilute solutions, obtuse curves are obtained. With obtuse curves it is difficult to locate the equivalence point accurately.
2. The overall accuracy of the conductometric titrations is limited as the technique does not permit addition of small increments of the titrant.

4 IR spectroscopy

In contrast to ultraviolet spectroscopy IR spectrum provides a rich array of absorption bands which can provide accurate structural information about a molecule. It provides the methods for studying materials in all three physical states i.e. solid, liquid and gas. Analytically useful IR spectrum covers the following range of electromagnetic spectrum

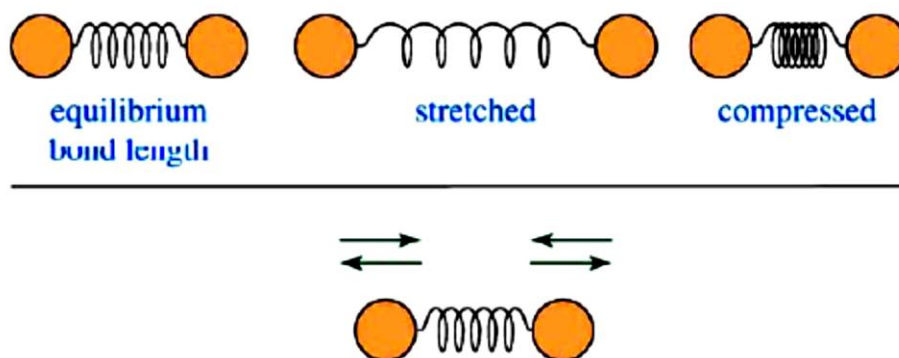
Near IR 15000 cm^{-1} to 3000 cm^{-1}

Mid IR 4000 cm^{-1} to 400 cm^{-1}

Far IR 200 cm^{-1} to 10 cm^{-1}

Most used 4000 cm^{-1} to 670 cm^{-1}

Infrared radiation is largely thermal energy. It induces stronger molecular vibrations in covalent bonds, which can be viewed as springs holding together two masses, or atoms. Specific bonds respond to (absorb) specific frequencies.

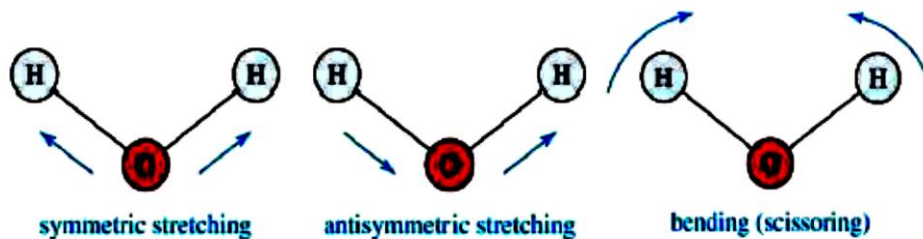


VI BRATI ONAL MODES:

The information contained in IR spectrum originates from molecular vibrations. These are either fundamental vibrational modes that are associated with the vibrations of specific functional group, or molecule, vibrational overtones or summational modes of fundamental vibrations.

A molecule resembles a system of balls of varying masses corresponding to atoms of a molecule and spring of varying lengths corresponding to various chemical bonds. There are two fundamental vibrational modes.

1. **Stretching** in which the distance between the two atoms increases or decreases but the atoms remain in the same bond axis.
 2. **Bending** in which the position of the atom changes relative to the bond axis.
- Covalent bonds can vibrate in several modes, including stretching, rocking, and scissoring



The various stretching and bending vibrations occur at certain frequencies. When an IR radiation of same frequency is incident on the molecule, the energy is absorbed and the amplitude of that vibration increases correspondingly. When the molecule returns to ground state the absorbed energy is released as heat.

A nonlinear molecule containing n atoms has $3n-6$ possible vibrational modes through which IR radiation may be absorbed. For example, methane has 9 and benzene has 30 possible fundamental absorption bands respectively. In order that a particular vibration results in an absorption band, the vibration must cause a change in the dipole moment of the molecule.

Which substances give a signal in IR spectrum?

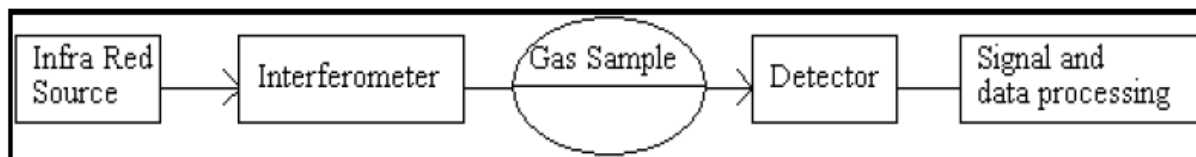
Ans: The molecules that contain polar bonds i.e. molecules composed of atoms of different elements, organic compounds and inorganic compounds (H_2O , CO_2 , NO_2 , HCl , salts...) can give a signal in IR spectrum. Whereas pure chemical elements in molecular or crystal state e.g. Ar , O_2 , O_3 , N_2 , Cl_2 , S_8 , silicon, graphite, Diamond etc cannot give a signal in IR spectrum.

Basic Principle:

When a sample is placed in a beam of infrared radiation, the sample will absorb radiation at frequencies corresponding to molecular vibrational frequencies, but will transmit all other frequencies. The frequencies of radiation absorbed are measured by an infrared spectrometer, and the resulting plot of absorbed energy vs. frequency is called the infrared spectrum of the material. Identification of a substance is possible because different materials have different vibrations and yield different infrared spectra. Furthermore, from the frequencies of the absorption it is possible to determine whether various chemical groups are present or absent in a chemical structure.

Instrumentation of FTIR:

The basic components of an FTIR are shown schematically in fig



1. **The Source:-** Infrared energy is emitted from a glowing black body source. This beam passes through an aperture which controls the amount of energy presented to the sample (and, ultimately, to the detector).

2. **The Interferometer:-** The beam enters the interferometer where the “spectral encoding” takes place. The resulting interferogram signal then exits the interferometer.

3. **The Sample:-** The gaseous sample can be directly analysed. Liquid can also be used directly but in diluted form in NaCl plates. Solid compound can be mixed with KBr and formed a pellet and used.

The beam enters the sample compartment where it is transmitted through or reflected off of the surface of the sample, depending on the type of analysis being accomplished. This is where specific frequencies of energy, which are uniquely characteristic of the sample, are absorbed.

4. **The Detector:-** The beam finally passes to the detector for final measurement. The detectors used are specially designed to measure the special interferogram signal.

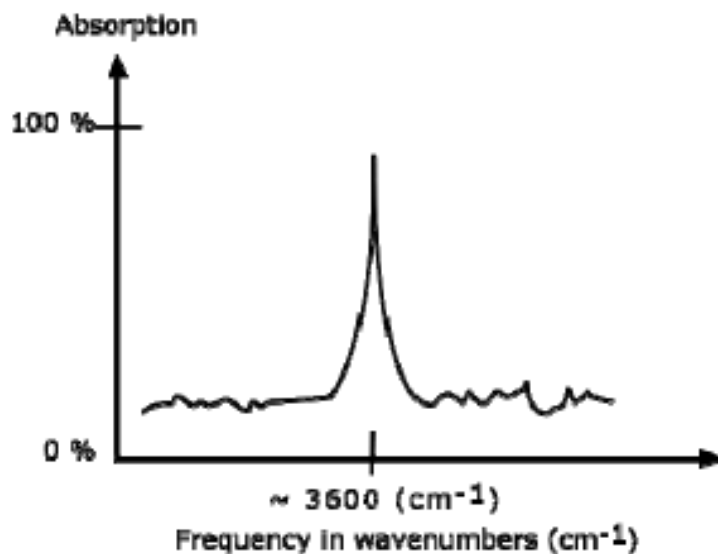
5. **The Computer:-** The measured signal is digitized and sent to the computer where the Fourier transformation takes place. The final infrared spectrum is then presented to the user for interpretation and any further manipulation.

Working:

The infrared source emits a broad band of different wavelength of infrared radiation. The IR source used is a SiC ceramic at a temperature of 1550 K. The IR radiation goes through an interferometer that modulates the infrared radiation. The interferometer performs an optical inverse Fourier transform on entering IR radiation. The modulated IR beam passes through the gas sample where it is absorbed to various extents at different wavelengths by the various molecules present. Finally, the intensity of the IR beam is detected by a detector, which is a liquid nitrogen cooled MCT (Mercury–Cadmium–Telluride) detector. The detected signal is digitised and Fourier transformed by the computer to get the IR spectrum of the sample gas.

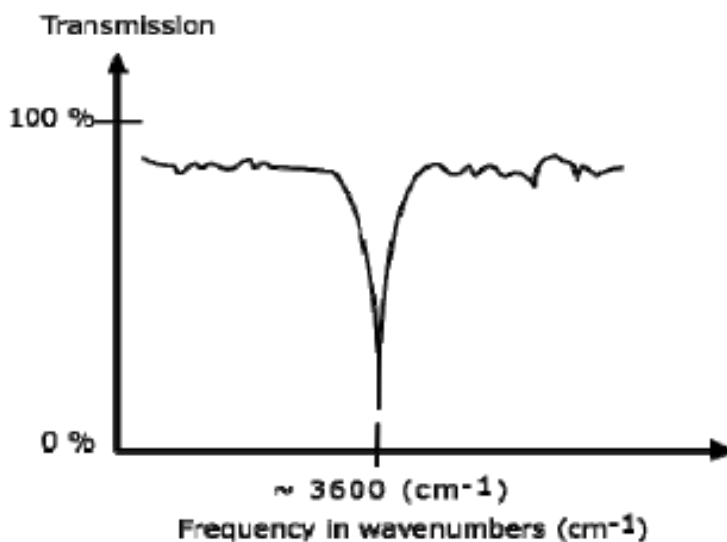
AN IR SPECTRUM IN ABSORPTION MODE:

The IR spectrum is basically a plot of transmitted (or absorbed) frequencies vs intensity of the transmission (or absorption). Frequencies appear in the x-axis in units of inverse centimeters (wavenumbers), and intensities are plotted on the y-axis in percentage units.



The graph above shows a spectrum in **absorption** mode.

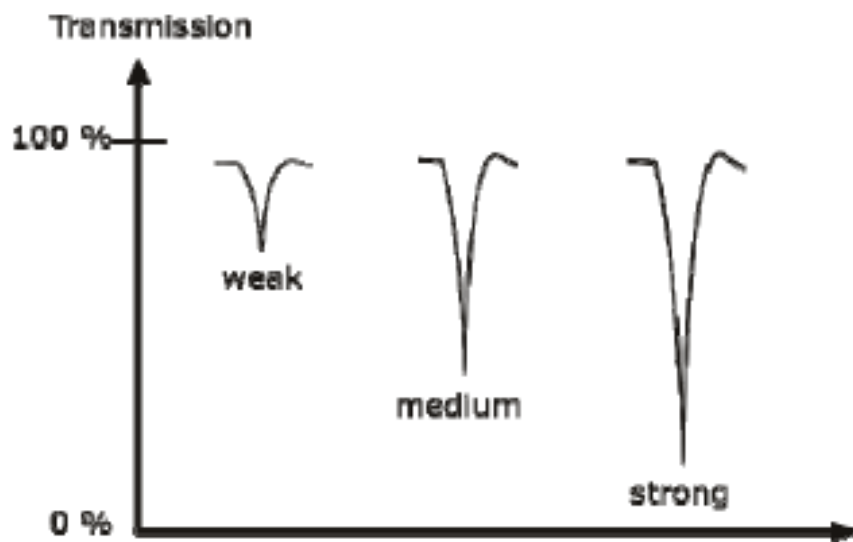
AN IR SPECTRUM IN TRANSMISSION MODE:



The graph above shows a spectrum in transmission mode. This is the most commonly used representation and the one found in most chemistry and spectroscopy books.

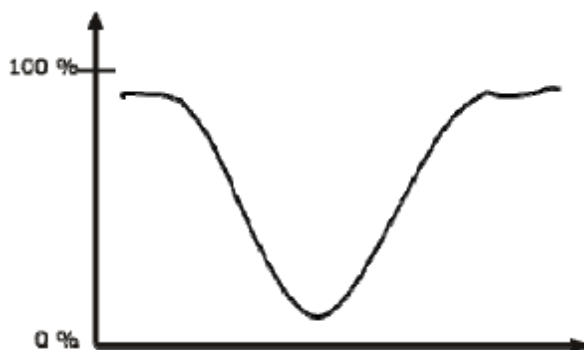
CLASSIFICATION OF IR BANDS

IR bands can be classified as strong (s), medium (m), or weak (w), depending on their relative intensities in the infrared spectrum. A strong band covers most of the y-axis. A medium band falls to about half of the y-axis, and a weak band falls to about one third or less of the y-axis.



Infrared band shapes come in various forms. Two of the most common are narrow and broad. Narrow bands are thin and pointed, like a dagger. Broad bands are wide and smoother.

A typical example of a broad band is that displayed by O-H bonds, such as those found in alcohols and carboxylic acids, as shown below



INFORMATION OBTAINED FROM IR SPECTRA

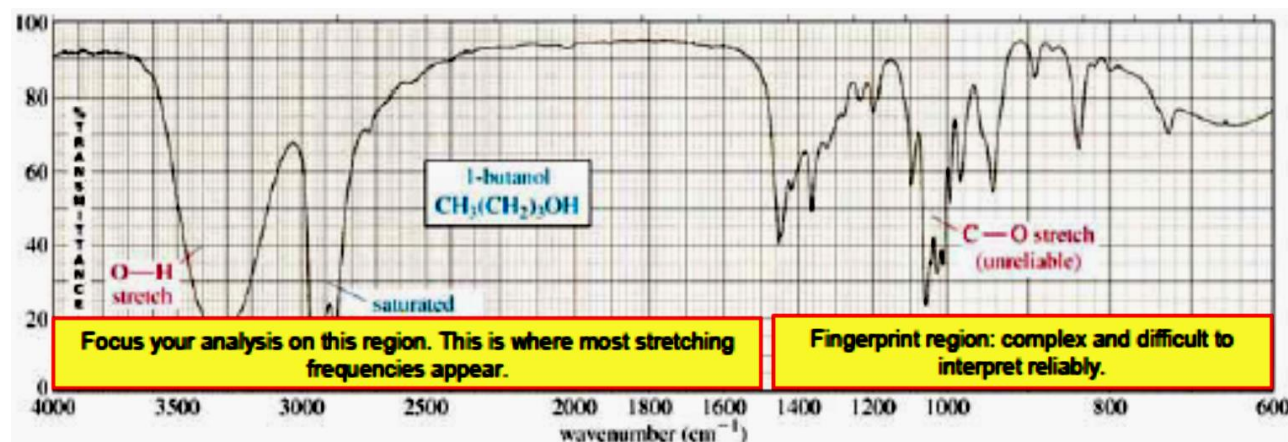
- IR is most useful in providing information about the presence or absence of specific functional groups.
- IR can provide a molecular fingerprint that can be used when comparing samples. If two pure samples display the same IR spectrum it can be argued that they are the same compound.
- IR does not provide detailed information or proof of molecular formula or structure. It provides information on molecular fragments, specifically functional groups.
- Therefore it is very limited in scope, and must be used in conjunction with other techniques to provide a more complete picture of the molecular structure.

THE FINGERPRINT REGION

Although the entire IR spectrum can be used as a fingerprint for the purposes of comparing molecules, the 600 - 1400 cm^{-1} range is called the fingerprint region.

This is normally a complex area showing many bands, frequently overlapping each other.

It is much more difficult to pick out individual bonds in this region than it is in the "cleaner" region at higher wavenumbers. The importance of the fingerprint region is that each different compound produces a different pattern of troughs in this part of the spectrum.



FUNCTIONAL GROUPS AND IR TABLES:

Characteristic IR Absorption Frequencies of Organic Functional Groups

Functional Group	Type of Vibration	Characteristic Absorptions (cm^{-1})	Intensity
Alcohol			
O-H	(stretch, H bonded)	3200-3600	strong, broad
O-H	(stretch, free)	3500-3700	strong, sharp
C-O	(stretch)	1050-1150	strong
Alkane			
C-H	stretch	2850-3000	strong
-C-H	bending	1350-1480	variable
Alkene			
=C-H	stretch	3010-3100	medium
=C-H	bending	675-1000	strong
C=C	stretch	1620-1680	variable
Alkyl Halide			

C-F	stretch	1000-1400	strong
C-Cl	stretch	600-800	strong
C-Br	stretch	500-600	strong
C-I	stretch	500	strong
Alkyne			
C-H	stretch	3300	strong sharp
$\text{—C}\equiv\text{C—}$	stretch	2100-2260	variable, not present in symmetrical alkynes
Amine			
N-H	stretch	3300-3500	medium (primary amines have two bands; secondary have one band, often very weak)
C-N	stretch	1080-1360	medium weak
N-H	bending	1600	medium
Aromatic			
C-H	stretch	3000-3100	medium
C=C	stretch	1400-1600	medium weak, multiple bands
Analysis of C-H out-of-plane bending can often distinguish substitution patterns			
Carbonyl		Detailed Information on Carbonyl IR	
C=O	stretch	1670-1820	strong
(conjugation moves absorptions to lower wave numbers)			
Ether			
C-O	stretch	1000-1300 (1070-1150)	strong
Nitrile			
CN	stretch	2210-2260	medium
Nitro			
N-O	stretch	1515-1560 & 1345-1385	strong two bands

IR Absorption Frequencies of Functional Groups Containing a Carbonyl (C=O)

Functional	Type	of Characteristic Absorptions	Intensity
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Group	Vibration	(cm ⁻¹)	
Carbonyl			
C=O	stretch	1670- 1820	strong
(conjugation moves absorptions to lower wave numbers)			
Acid			
C=O	stretch	1700- 1725	strong
O- H	stretch	2500- 3300	strong very broad
C- O	stretch	1210- 1320	strong
Aldehyde			
C=O	stretch	1740- 1720	strong
=C- H	stretch	2820- 2850 & 2720- 2750	medium two peaks
Amine			
C=O	stretch	1640- 1690	strong
N- H	stretch	3100- 3500	unsubstituted have two bands
N- H	bending	1550- 1640	
Anhydride			
C=O	stretch	1800- 1830 & 1740- 1775	two bands
Ester			
C=O	stretch	1735- 1750	strong
C- O	stretch	1000- 1300	two bands or more
Ketone			
Acyclic	stretch	1705- 1725	strong
Cyclic	Stretch	3- membered - 1850	strong
		4- membered - 1780	
		5- membered - 1745	
		6- membered - 1715	
		7- membered - 1705	
α, β unsaturated	Stretch	1665- 1685	strong
aryl ketone	Stretch	1680- 1700	strong

How to analyse the IR spectrum?

When analysing the IR spectrum of an unknown molecule, first efforts on determining the presence or absence of a few major functional groups. The $\text{C}=\text{O}$, $\text{O}-\text{H}$, $\text{N}-\text{H}$, $\text{C}-\text{O}$, $\text{C}=\text{C}$, $-\text{C}\equiv\text{C}-$, $-\text{CN}$ and NO_2 peaks. These peaks are most conspicuous and give immediate structural information if they are present. Do not try to make a detailed analysis of the $\text{C}-\text{H}$ absorption near 3000cm^{-1} , almost all the compounds have these absorptions.

Follow the steps

1. **Is carbonyl group is present?** The $-\text{C}=\text{O}$ group give rise to a strong absorption in the region $1820-1660\text{cm}^{-1}$. The peak is often strongest in the spectrum and of medium width.
2. **If $\text{C}=\text{O}$ is present**, then check the following types
- 3.

Acids	Is $\text{O}-\text{H}$ also present? Broad absorption near $3400-2400\text{cm}^{-1}$ usually overlaps with $\text{C}-\text{H}$
Amides	Is $\text{N}-\text{H}$ also present? Medium absorption near 3400cm^{-1} , sometimes double peaks with same size.
Esters	Is $\text{C}-\text{O}$ present? Strong intensity absorption near $1300-1000\text{cm}^{-1}$
Anhydrides	Two $\text{C}=\text{O}$ absorption near 1810 and 1760cm^{-1}
Aldehydes	Is aldehyde $\text{C}-\text{H}$ present? Two weak absorption near 2850 and 2750cm^{-1}
Ketones	The preceding five choices have been eliminated

4. **If $\text{C}=\text{O}$ absent**, then check the following options

Alcohols, Phenols	Check for $\text{O}-\text{H}$ broad absorption near $3400-3300\text{cm}^{-1}$, confirm this by finding $\text{C}-\text{O}$ near $1300-1000\text{cm}^{-1}$
Amines	Check for $\text{N}-\text{H}$ Medium absorptions near 3400cm^{-1}
Ethers	Check for $\text{C}-\text{O}$ near $1300-1000\text{cm}^{-1}$ and absence of $\text{O}-\text{H}$ near 3400cm^{-1}

5. **Double bonds and/or aromatic rings**

1. $\text{C}=\text{C}$ is weak absorption near 1650cm^{-1}
2. Medium strong absorption in the region $1600-1450\text{cm}^{-1}$, these often imply on aromatic ring
3. Confirm the double bond or aromatic ring by consulting the $\text{C}-\text{H}$ region, aromatic and vinyl $\text{C}-\text{H}$ occurs to left of 3000cm^{-1} , aliphatic $\text{C}-\text{H}$ occurs to right of this value.

4. **Triple bonds**

1) $\text{C}\equiv\text{N}$ is medium sharp absorption near 2250cm^{-1}
2) $\text{C}\equiv\text{C}$ is weak, sharp absorption near 2150cm^{-1}
3) Check also for acetylenic $\text{C}-\text{H}$ near 3300cm^{-1}

5. Nitro groups

Two strong absorption at $1600\text{--}1530\text{ cm}^{-1}$ and $1390\text{--}1300\text{ cm}^{-1}$

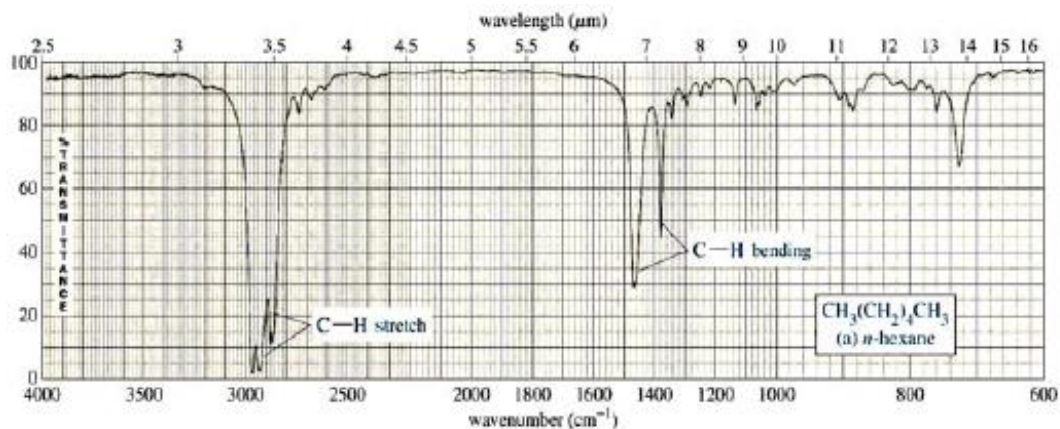
6. Hydrocarbons

None of the preceding found

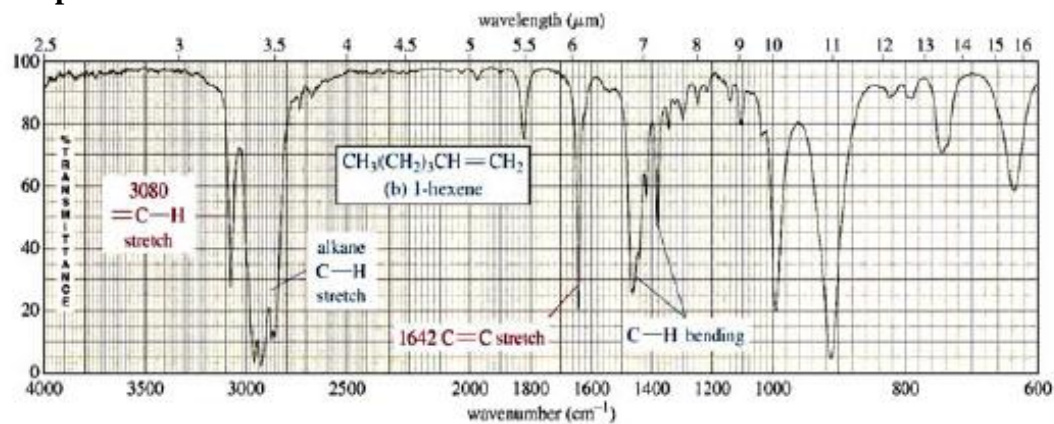
Major absorptions are in C-H region near 3000 cm^{-1}

Very simple structure the only another absorption appear near 1460 and 1375 cm^{-1}

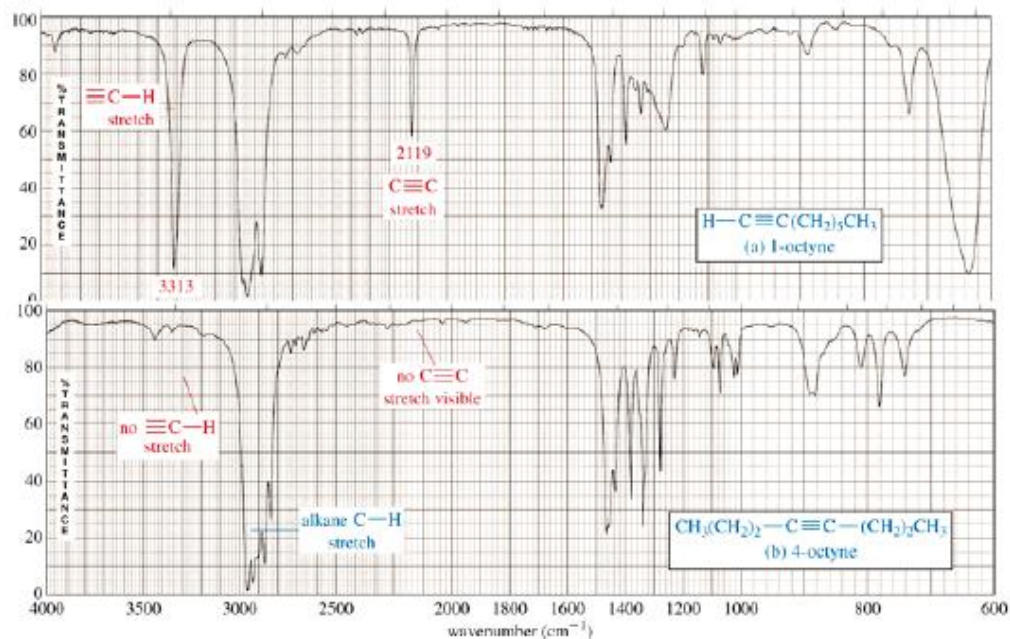
IR spectrum of n-Hexane



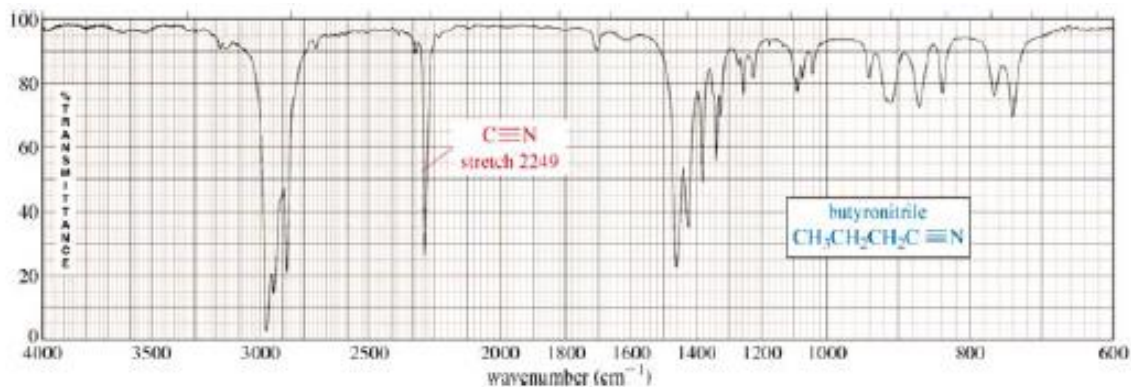
IR spectrum of 1-Hexene



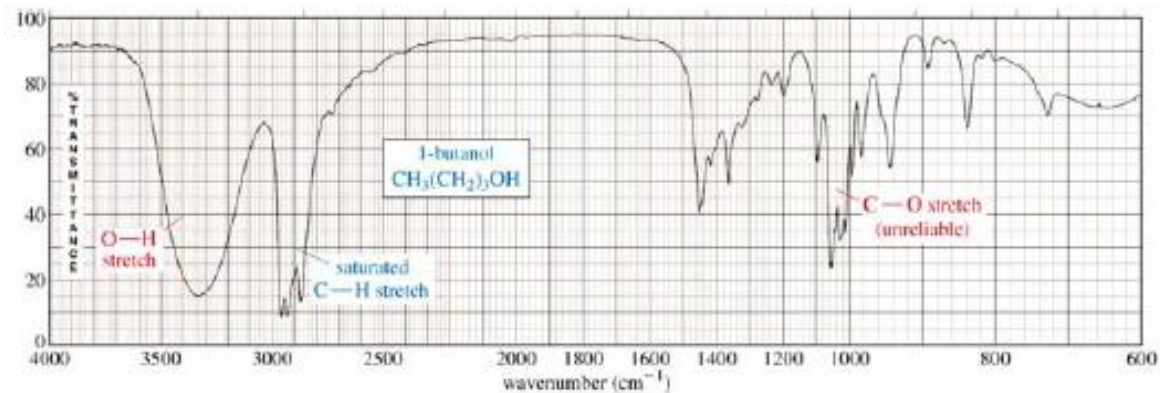
IR spectrum of 1-Octyne and 4-Octyne



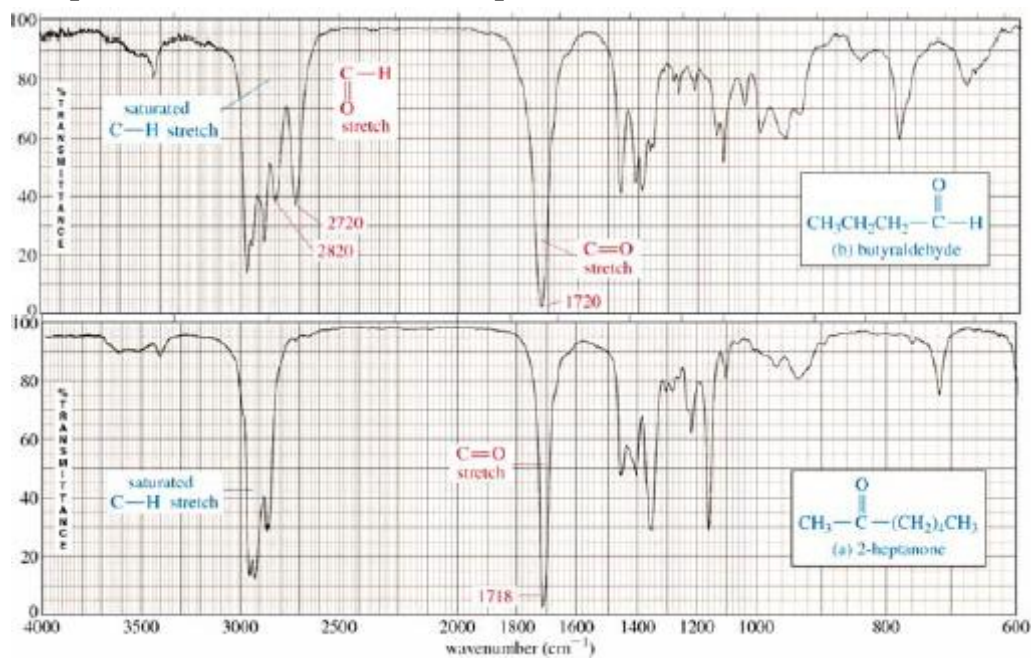
IR spectrum of Butyronitrile



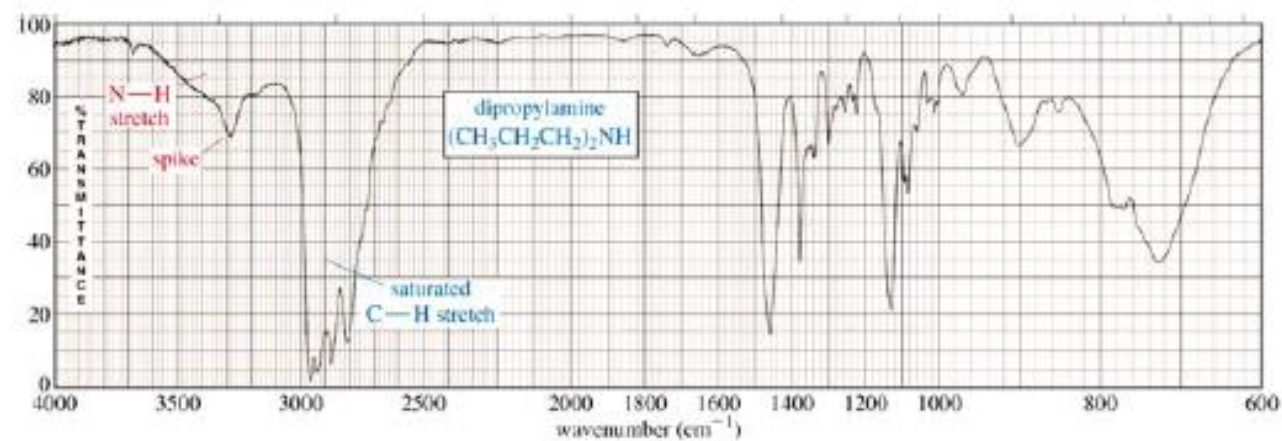
IR spectrum of 1-Butanol



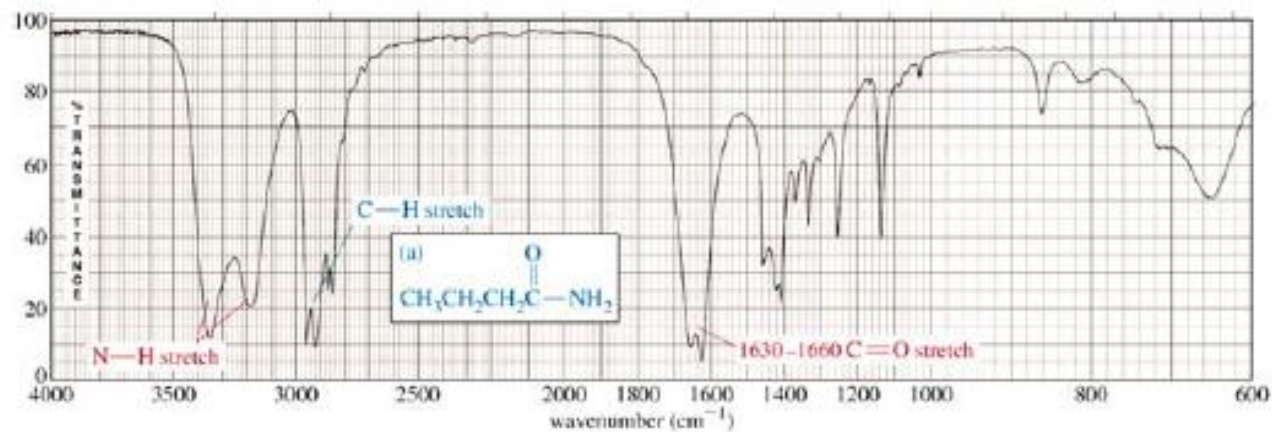
IR spectrum of 1- Butanal and 2- Hept anone



IR spectrum of n- Hexanoic acid



IR spectrum of 1- Butanamide



IR spectrum of 3-Bromoaniline

