

Review

Traditional Uses, Phytochemistry and Pharmacological Activities of Annonaceae

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Abstract: In 1789, the Annonaceae family was catalogued by de Jussieu. It encompasses tropical and subtropical plants which are widespread in distribution across various continents such as Asia, South and Central America, Australia and Africa. The genus of *Annona* is one of 120 genera of the Annonaceae family and contains more than 119 species of trees and shrubs. Most species are found in tropical America, where over 105 species have been identified. Due to its edible fruits and medicinal properties, *Annona* is the most studied genus of Annonaceae family. To date, only a limited number of these species have economic value, including *A. squamosa* L. (sugar apple), *A. cherimola* Mill. (Cherimoya), *A. muricata* L. (guanabana or soursop), *A. atemoya* Mabb. (atemoya), a hybrid between *A. cherimola* and *A. squamosa*, *A. reticulata* L. (custard apple), *A. glabra* L. (pond-apple) and *A. macrophyllata* Donn. Sm. (ilama). Phytochemically, several classes of secondary metabolites, including acetogenins, essential oils, alkaloids, terpenoids and flavonoids. The pharmacological activities of *Annona* species leaves and seeds include antibacterial, anticancer, antidiabetic and anti-inflammatory properties.

Keywords: Annonaceae; *Annona*; custard apple; phytochemistry; bioactivity; ethnomedicinal pharmacological activity



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1. Introduction

In 1789, the Annonaceae family was catalogued by de Jussieu [1,2]. It encompasses tropical and subtropical plants, which are widespread in distribution across various continents such as Asia, South and Central America, Australia and Africa [3]. It is one of the largest Mangnoliidae families and the number of its genera and species is still debated [4–6]. Bailey and Popenoe believe that it has between 40 and 50 genera and from 500 to 600 species [6]; however, many studies have indicated that the Annonaceae family is comprised of more than 2400 species distributed in approximately 120 genera [4,5]. The family of Annonaceae involves trees, lianas and bushes arranged in four large subfamilies: Malmeoideae, Annonoideae, Ambavioideae and Anaxagoreoideae [7,8]. Economically, species of Annonaceae are important as a source of edible fruits, for instance, the pawpaw (*Asimina*), custard apple, sweetsop, soursop and cherimoya [1]. It has also been reported that some oils from the seeds might be used for the production of edible oils and as an ingredient in soaps, and the woods of some species have been reported for alcohol production [3]. Chemical studies of Annonaceae species have reported the isolation of a wide diversity of phytochemical components, including acetogenins, alkaloids and flavonoids from the bark, fruits, leaves, seeds and pulp of Annonaceae [9]. This review aims to provide a comprehensive summary of the botanical features, phytochemistry, pharmacological properties, and the traditional and ethnomedicinal uses of the Annonaceae family and, specifically, *Annona* species.

2. Botanical Features of Annonaceae Species

2.1. Distribution and Classification

Annonaceae has been listed as a diverse family of aromatic trees, bushes or shrubs, and climbers or lianas, which are predominantly found in the tropical and subtropical regions, with a limited number growing in temperate zones [1,10]. In tropical America, the Annonaceae species are usually shrubby and most grow in open grasslands [1]. In contrast, species that are climbers mostly grow in the tropical area of the old world [1]. In temperate zones like North America, the only genus reported is *Asimina* [1,3]. In Brazil, more than 385 species have been reported, with the majority of them reported in the Amazonian region [2]. According to the Takhtajan system of flowering plant classification, the majority of Annonaceae plants can be found in both Asia and Australasia with approximately 51 genera and more than 950 species, while 40 genera with approximately 450 species are confined to Africa and Madagascar, and about 38 genera and 740 species are native to the American continent [3]. The first classification of the Annonaceae family was described by Dunal in 1817 and was limited to only fruit morphology [11]. Subsequently, a new classification of the Annonaceae family based on flower characteristics was introduced by Diels and Alder in 1932 [11]. However, a later classification by Fries in 1959 was found to be more comprehensive and authentic, using a combination of fruit morphology and flora characteristics [11]. The Annonaceae family are characterised by the presence of a variety of primitive and archaic features, leading to them being described by Darwin as “living fossil” due to their ability to survive the mass extinction [1,11]. Under the Takhtajan system, the Annonaceae family is related to Magnoliaceae, which is one of the largest families of Magnoliales with other families such as Degeneriaceae, Canellaceae, Himantandraceae and Myristicaceae [1,11].

2.2. Diagnostic Features

From one species to another, the botanical features of Annonaceae families vary greatly based on their origin, geography, and climate. Based on morphology and habitat, the Annonaceae family is known among the homogeneous plant families [1,4]. The aromatic flowers are commonly open before other parts are entirely developed. The flowers are terminal, axillary, hermaphrodite, singular or grouped and regular [1,11]. The stamens are typically abundant, spirally arranged and hypogenous [1,11]. The leaves are characterised by having a glaucous or metallic sheen, and they are alternate, exstipulate and regular [1,11]. The fruits are typically made up of clusters of berries with an edible fleshy receptacle, particularly in the *Annona* genera and they are extensively consumed due to their high nutritional value [1,11]. Finally, the seeds are enlarged and have a copious, irregular-surfaced endosperm with a minute embryo [1,11].

2.3. Traditional Uses

Annonaceae species are famous in tropical regions and used traditionally across tropical regions due to their widespread distribution. Various parts of the species are used traditionally, including leaves, seeds, bark, fruit, stem, roots and twigs. A range of different methods for preparation is reported, such as infusions, pastes and decoctions [11]. For instance, the fresh fruit of *Annona dioica* is used for wound healing in Brazil [11]. The dried leaves of *Annona muricata* are used orally for analgesic effects in some parts of Indonesia [11]. In Burkina-Faso, the bark and roots of *Annona muricata* are used for dysentery and as an anthelmintic medicine, whereas the leaves are utilized for both fever and dysentery [12]. In the northwestern part of Brazil, both leaves and twigs of *Duguetia chrysocarpa* are ground and the extract of this mixture are utilized for treating gastrointestinal ulcers as well as a remedy for bowel disease [11]. A decoction of the stem bark of *Annickia chlorantha* is used orally as a remedy for the treatment of wounds and fever in Cameroon [13]. Further data on the traditional uses of the most widely used Annonaceae species are presented in Table 1.

Table 1. Uses of most commonly used Annonaceae family in traditional medicines.

| Annonaceae Species | Region | Local Name | Medicinal Uses | Part Used | Mode of Usage | References |
|---|------------------------------|---|--|------------------------------|---|------------|
| <i>Alphonsea javanica</i> Scheff. | Indonesia | Aku Battu | Rheumatism and edema | Leave | Ethanol extract | [14] |
| <i>Annickia chlorantha</i> (Oliv.) | Cameroon | African yellow wood (c) Moambe Jaune | Treatment of sores Antipyretic Antiemetic Stimulant Tuberculosis Treatment of jaundice Urinary tract infection | Bark | Powder Crushed bark and drink extract Decoction Decoction in baths Decoction Decoction | [12,13] |
| <i>Annickia affinis</i> (Exell) Versteegh & Sosef | Cameroon | African yellow wood | Wound healing Antiemetic Antipyretic | Stem bark | Decoction of stem bark | [13] |
| <i>Anonidium floribundum</i> Pellegr | Cameroon | Ebom, Libanga Ebom | Poison antidote Dysentery Antipyretic | Roots Root/Bark Leaves | Decoction taken orally | [12,15] |
| <i>Anonidium mannii</i> (Oliv.) | Cameroon | Ebome; Npole Wapo'o, Ebome Afan | Antipyretic | Stem bark | Decoction of stem bark | [13] |
| <i>Boutiquea platypetala</i> (Engl.) | Cameroon | Not reported | To treat fresh wounds | Leaves | Pounded fresh leaves | [12] |
| <i>Cananga odorata</i> (Lam.) Hook and Thomson | Malaysia and India | Kenanga utan, Perfume tree, Cananga oil, Ylang ylang | Rheumatism Ophthalmic inflammation and wound healing | Bark | Bark extract eye drops for inflammation and decoction are used to wash fresh wounds | [11] |
| <i>Duguetia chrysocarpa</i> Maas | Brazil | Pindaiba-da-mata | Bowl and rheumatism inflammation | Leave and twigs | Leaves and twigs extract taken to relieve inflammation | [16] |
| <i>Enicosanthellum pulchrum</i> (King) Heusden | Malaysia | Disepalum | Rheumatism fever, edema and asthma | Leave | Decoction can be used for asthma and rheumatism | [17] |
| <i>Enantia chlorantha</i> var. <i>soyauxii</i> Engler and Diels | Africa | African yellow wood | Arthritis and wound healing | Bark | Powdered bark with citrus lemon used as dressing | [11] |
| <i>Friesodielsia enghiana</i> (Diels.) Verdc | Cameroon | Lonkoso | Analgesic | Bark | Decoction of bark is taken orally | [15] |
| <i>Friesodielsia gracilipes</i> (Benth.) Steenis | Cameroon | Ntonda | Treatment of sores, skin infection, ulcers, and jaundice | Bark and wood | Decoction of bark and wood | [12] |
| <i>Fissistigma oldhamii</i> (Hemsl.) Merr | Southern China | Oldhamii | Rheumatoid arthritis | Stems and roots | Powdered of stems and roots and orally ingested | [11] |
| <i>Greenwayodendron suaveolens</i> (Engl and Diels) Verdc | Not reported | Otunga | Aphrodisiac and Vermifuge Rheumatic pains, fevers, headache, stomach-ache | Root Leaves and bark | Chew roots Pulverized leaves or bark and mixed with seeds of <i>Aframomum melegueta</i> | [15] |
| <i>Isolona hexaloba</i> (Pierre) Engl & Diels | Democratic Republic of Congo | Bodzungu | Malaria | Stem bark | Decoction of stem bark | [18] |
| <i>Monodora myristica</i> (Gaertn.) Dunal | Ivory coast | M Kpo. Abidjan district | Eye diseases and hemorrhoids, febrile pains and headache | Fruits Seed | Fruits and seeds consumed whole or ground to be used in soup and stews | [19] |
| <i>Monodora tenuifolia</i> Benth | Not reported | African nutmeg Ebom osoé Grandes feuilles | Toothache Dysentery and fevers | Root Bark and root | Clean the roots, boil and rinse the mouth Prepared as a decoction and used as an enema | [12] |
| <i>Polyalthia suaveolens</i> Engl and Diels | Cameroon | Diels; Otungui; Ntunga | Analgesic, Antiepileptic Antipyretic Treatment of jaundice | Stem bark | Decoction of stem bark | [13] |
| <i>Polyalthia longifolia</i> (Sonn.) Thwaites | India | Ashoka | Fever | Bark | Decoction of bark | [20] |
| <i>Xylopia aethiopica</i> (Dunal) A.Rich | Sudan | Ethiopia or Negro pepper | Rheumatism, colic pain, headache, and neuralgia | Fruits | Ethanol fruit extract or dried fruits are used as whole | [21] |
| <i>Xylopia aromatic</i> Lam. Mart | Columbia | Monkey pepper | Pulmonary inflammation and hemorrhoids | Roots Leaves | Insertion of root pieces into rectum and leaves burnt and smoke inhaled | [22] |

Table 1. Cont.

| Annonaceae Species | Region | Local Name | Medicinal Uses | Part Used | Mode of Usage | References |
|---|--------------------------------|------------------------------|---|-----------|-------------------------------|------------|
| <i>Xylopia parvifolia</i> Hook.f. and Thomson | East and Central Africa, India | Netawu/ Athu ketiya | Gastrointestinal ulcers Analgesic | Roots | Decoction Finely dried powder | [23] |
| <i>Xylopia staudtii</i> Engl & Diels | Not reported | Ntom, Odjobi Bush pepper (c) | Cold and headache treatment | Bark | Powder | [12] |
| <i>Monodora tenuifolia</i> Benth | Cameroon | Ebome osso | Joint and muscle pain, promotion of breast milk production and headache | Stem bark | Decoction of stem bark powder | [13] |
| <i>Uvaria acuminata</i> Oliv | Cameroon | Nosonaback | Typhoid and Yellow fever Headache and epilepsy | Stem bark | Decoction of stem bark | [13] |

3. Phytochemistry of Annonaceae Family

A wide array of chemical compounds from various parts of Annonaceae plants have been discovered, isolated and characterised. The results of both phytochemical investigations and biological studies on various plants from this family have led to the identification of a wide diversity of compounds such as annonaceous acetogenins, flavonoids, alkaloids and essential oils, as summarized in (Table 2). These phytochemical constituents have been found to exhibit a broad range of biological activities such as immunosuppressive, anti-neoplastic, cytotoxic, antimicrobial, anti-inflammatory effects (Table 3). However, it is the *Annona* genera that are the most widely used as a food source and in traditional medicines.

Table 2. Representative phytochemicals isolated from plants of Annonaceae.

| Species | Part | Compounds | Class | References |
|--|---------------------|---|-------|------------|
| <i>Anaxagoma dolichocarpa</i> Sprague and Sandwith | Fruits | <i>p</i> -Cymene Spathulenol Caryophyllene oxide Guaiene | ESO | [5] |
| <i>Anomianthus dulcis</i> (Dunal) J. Sinclair | Stem | (–)-Anolobine (–)-Anonaine | ALK | [24] |
| <i>Artabotrys pierreanus</i> Engl. & Diels | Stem bark | Cyperene Caryophyllene oxide Cyperermone Cadallene | ESO | [5] |
| <i>Artabotrys hexapetalus</i> (L.f.) Bhandari | Aerial parts | 9-Oxo-asimicinone Artapetalin-A Artapetalin-B | ACT | [25,26] |
| <i>Goniothalamus giganteus</i> Hook.f. & Thomson | Bark | Pyranicin Pyragonicin Goniotrionin | ACT | [27] |
| <i>Miliusa balansae</i> Finet & Gagnep | Leaves and branches | Ombuine Chrysosplenol Pachypodol Chrysosplenol C | FLA | [28] |

ALK (Alkaloids), ACT (Acetogenins), ESO (Essential oils) and FLA (Flavonoids).

Table 3. Pharmacological activities of some isolated compounds from Annonaceae species.

| Species | Part Used | Isolated Compounds | Pharmacological Activity | Mechanism of Action | References |
|--|--------------------------|---|-----------------------------|---|------------|
| <i>Alphonsea javanica</i> Scheff | Leaves | (+)-Altholactone (+)-Goniothalmin | Anti-inflammatory | Inhibited lipopolysaccharide (LPS) induced NO production in RAW 264.7 macrophages with IC ₅₀ = 0.8 µM. | [14] |
| <i>Artabotrys hexapetalus</i> (L.f.) Bhandari | Roots, stems, and leaves | Artabonatine B Squamolone | Anticancer | Exhibited activity against 2,2,15 and Hep G2 cell lines with IC ₅₀ 11.0 and 9.1 µg/mL. Displayed activity against Hep G2 cell lines with IC ₅₀ 2.8 µg/mL. | [29] |
| <i>Cananga odorata</i> (Lam.) Hook.f. & Thomson | Fruits | Cleistopholine | Cytotoxic | Exhibited cytotoxicity against both Hep 2,2,15 and Hep G2 cell lines with IC ₅₀ 0.54 and 0.22 µg/mL, respectively. | [30] |
| <i>Goniothalamus tamirensis</i> Pierre ex Finet & Gagnep | Stem bark | Dielsiquinone | Cytotoxic | Displayed cytotoxic activity against U251, RPMI, MCF7, HT029 and A549. with ED ₅₀ 0.37, 0.11, 0.11 1.12 and 0.11, respectively. | [31] |
| <i>Guatteria blepharophylla</i> Mart | Bark | Isocoreximine | Anti-proliferative activity | Exhibited activity against UACC-62, NCI-H460, HT-29 and MCF-7 with TGI > 764.52 µM. | [32] |
| <i>Rollinia sylvatica</i> A.St.-Hil | Leaves | Hinesol z-Caryophyllene beta-Maaliene | Anti-inflammatory | Leukocytes migration was significantly reduced at concentrations of 36.04–45.37 µg/mL. | [33] |

4. Annona Genera

The genus *Annona* is one of the 120 genera of the Annonaceae family and contains more than 119 species of trees and shrubs, most of them distributed in tropical areas of the Americas and Africa [6]. The majority of these species are found in tropical America, with more than 105 species (26 of them are endemic) and 10 species distributed in tropical Africa [10,34]. It has been reported that this genus is the second or the third largest genus in the Annonaceae family [35]. Its generic name derives from the Latin Hispaniolan Taino “annual harvest” [6,35]. Due to its edible fruits and medicinal properties, *Annona* is the most important genus of Annonaceae family [2]. Numerous *Annona* species furnish edible fruits like *Annona muricata* (“graviola”), *Annona crassiflora* (“araticum”) and *Annona squamosa* (“fruta do conde”) [2]. Most of the fruits are consumed either in fresh form or used in desserts, juices and ice cream preparations [34]. Despite *Annona* having many species, only limited species of this family are economically important such as *A. squamosa* L. (sugar apple), *A. cherimola* Mill. (Cherimoya), *A. muricata* L. (guanabana or soursop), *A. atemoya* Mabb. (atemoya), a hybrid between *A. cherimola* and *A. squamosa*, *A. reticulata* L. (custard apple), *A. glabra* L. (pond-apple) and *A. macrophyllata* Donn. Sm. (ilama) [6]. Phytochemically, several classes of secondary metabolites such as acetogenins,

essential oils, alkaloids, terpenoids and flavonoids have been described in this genus [34,36]. A variety of pharmacological activities have been reported from various parts of *Annona* species specially leaves and seeds including applications against antibacterial [37], antinociceptive [38], anticancer [39], anticonvulsant [40], antidiarrhea [41], antidiabetic [42], anti-malarial [39], anti-inflammatory [43], antioxidant [44], antileishmanial [45], antiulcer [46] and antidepressant [47].

4.1. Botanical Features

Generally, *Annona* species are small trees or shrubs with a height from 5 to 11 m depending on various factors including soil, climate, species, and crop management [2]. In relation to the botanical characteristics of *Annona* species, the majority of them are moderately erect with brown bark that is frequently furrowed (Table 4) [10]. The stems are rust-coloured (ferruginous) and covered with densely matted hairs (tomentose) when young, becoming smooth and hairless (glabrous) as they mature [6,10]. It has thin lateral roots and a taproot that is not generally pronounced [2]. With regard to the flowers, they are hermaphrodites, solitary or fascicle containing from two to four flowers. The flowers are usually fragrant, with six petals and three green sepals, in a circular arrangement of two verticils [6]. Flowering of the plant usually starts at 3 to 4 years and flower opening usually occurs by separation of the apex of external petals [6,10]. Finally, the leaves may be shiny or hairy and have an impressed vein on the upper side, and the fruits are syncarpous and comprised of seeds and many carpels [6,10].

Table 4. Botanical information of some *Annona* species.

| Species | Synonyms | Local Names | Geographic Distribution | References |
|--------------------------|--|--|--|------------|
| <i>A. cherimola</i> | <i>A. tripetala</i> Aiton <i>A. pubescens</i> Salisb | Chirimoya Chirimolia Cerimoya Cherimoyer Momona | South Africa, China Egypt Eritrea Myanmar Philippines India France Italy Mexico, Ecuador Portugal Peru | [6,48] |
| <i>A. coriacea</i> | <i>A. coriacea</i> var. amplexicaulis S.Moore, <i>A. coriacea</i> var. cuneate, <i>A. coriacea</i> var. pygmaea Warm | Marolo Araticum Marolino | Brazil (Cerrado, Caatinga) | [10,49] |
| <i>A. cornifolia</i> | <i>A. walkeri</i> S. Moore | Araticum-mirim | Brazil | [50] |
| <i>A. crassiflora</i> | <i>A. macrocarpa</i> Barb <i>A. rodriguesii</i> Barb | Araticum Pinha-docerrado Cerrado pinecone Marolo Cabeça de negro | Brazil | [51] |
| <i>A. macrophyllata</i> | <i>A. diversifolia</i> Saff | Ilama, Papauce Anona blanca | Mexico China India | [52,53] |
| <i>A. montana</i> Macfad | <i>A. Montana</i> f. marcgravii (Mart.) Porto | Mountain soursop False graviola jacá do Pará Araticum grande Shan di fan li zhi | Southern Asia, South America Amazon Rainforest and Atlantic Forest | [10] |
| <i>A. muricata</i> | <i>A. macrocarpa</i> Barb <i>A. muricata</i> Guanabarus <i>A. cearensis</i> Morales | Brazilian pawpaw Soursop, ci guo fan li zhi, Graviola Araticum grande Mullu Raama Phala, Corossol Catuche | Tropical regions of Americas Malaysia, Myanmar, Pakistan, India, Indonesia, China | [6,10] |

Table 4. Cont.

| Species | Synonyms | Local Names | Geographic Distribution | References |
|-----------------------------|---|--|---|------------|
| <i>A. reticulata</i> | <i>A. excelsa</i> Kunt <i>A. laevis</i> Kunth <i>A. longifolia</i> Moc <i>A. riparia</i> Kunth | Custard apple Bullock's heart | Indonesia, West Indies, Bangladesh, China, India | [54] |
| <i>A. sclerophylla</i> Saff | <i>Sulcata</i> Urb <i>A. spinescens</i> Mart | Not reported | Brazil | [10] |
| <i>A. senegalensis</i> | <i>A. senegalensis</i> var. <i>arenaria</i> Sillans <i>A. senegalensis</i> var. Bail. Wild <i>A. senegalensis</i> var. <i>glabrescens</i> Oliv <i>A. senegalensis</i> var. <i>cuneata</i> Oliv <i>A. arenaria</i> Thonn <i>A. chrysophylla</i> <i>A. chrysophylla</i> var. <i>porpetac</i> Bail <i>A. porpetac</i> Bail | Wild soursop Sour soup Abo Uburuochaand Gwandar | Nigeria | [6] |
| <i>A. squamosa</i> | <i>A. asiatica</i> L. <i>A. squamosa</i> f. <i>Parvifolia</i> Kuntze <i>A. cinerea</i> Dunal <i>Guanabanus squamosus</i> (L.) M.Gómez | Custard apple Sweetsop Tiep baay Amritaphala Chirimoya Fruta do conde Squamosus Gomez Guanabanus | Egypt, Sudan, Pakistan, Thailand, China, India, Costa Rica, | [6,10] |

4.1.1. *Annona Cherimola*

Annona cherimola Mill (*Cherimoya*) belongs to the genus *Annona* in the Annonaceae family in magnolias order, which means “cold seeds”, and is a small tree that produces heart-shaped and conical edible fruit [55]. It is a steep, semi-momentary and a low bunched tree that is widespread in Ecuador and Peru and distributed throughout Asia, South Europe, America and Africa [56]. In Mexican traditional medicine, this plant has been used to treat various diseases such as diabetes, cough, fever, headache, worms and inflammation either alone or in combination with other plant species [48,57–60]. Recently, various parts of *A. cherimola* have been phytochemically profiled and contain various polyphenols and alkaloids. The leaves were found to be a source of bioactive compounds with potential for use as treatments for skin and eye diseases and gastric, cardiovascular and intestinal disorders [55].

4.1.2. *Annona Squamosa*

Annon squamosa L., commonly known as custard apple, is a tropical, endemic species of the West Indies, Ecuador, Peru, Brazil, South and Central America, Mexico, Bahamas, Bermuda, and Egypt [61]. This plant is extensively cultivated in various states of India, including Maharashtra, Gujarat, Madhya Pradesh, Chhattisgarh, Assam, Uttar Pradesh, Bihar, Rajasthan, Andhra Pradesh, and Tamil Nadu. The total area of cultivation has been reported by the Indian Council of Agricultural Research (ICAR) as 40,000 ha [62]. Its tree grows as a small sapling from 3 m to 8 m, with large branches having brownish or light brownish bark and it has thin leaves and is known for its edible fruit [61]. In the Aligarh district village in Uttar Pradesh, *A. squamosa* is well-known for its antidiabetic properties [63]. Its seeds, bark and leaves possess various pharmacological properties, mainly anti-tumour properties [64].

4.1.3. *Annona Muricata*

Annona muricata is a commonly known as soursop and graviola and is native to Central and South America. It is a small tree 5–10 m tall and 15–83 cm in diameter; it has low branches and edible fruit that are used commercially for the production of candy, juice and

sherbets [65]. Traditionally, the aerial parts of this plant have been used to treat various diseases like diabetes and malaria and nowadays, it is widely used by people diagnosed with cancer [66]. Moreover, this species possesses several pharmacological properties, including vasodilator, cardio-depressive, antispasmodic, antimutagen, anticonvulsant, antiviral, antidiabetic and antihypertensive effects [67]. Both the leaves and seeds of *A. muricata* have been evaluated for their constituents resulting in the identification and isolation of more than 50 mono-THF acetogenins, alkaloids, terpenoids, saponins, flavonoids, coumarins, cardiac glycosides, phenols, tannins and anthraquinones [67].

4.1.4. *Annona Reticulata*

Annona reticulata Linn is a traditionally important plant utilized in traditional medicines [68]. It is indigenous to the West Indies and widely distributed in tropical and subtropical regions of the world [54]. It is a small tree with a height between 6 and 7.5 m and contains numerous lateral branches [54]. It has a cylindrical stem that contains lenticels and very short coffee-colored hairs [54]. The leaves of *A. reticulata* are lanceolate, membranous, oblong, and rounded or curate at the base. Fruits are edible, rough, somewhat heart-shaped and yellow in color that shifts to yellowish-red on ripening, and the seed is smooth and blackish in color [69]. Traditionally, *A. reticulata* has been utilized for the treatment of epilepsy, dysentery, cardiac problem, constipation, haemorrhage, bacterial infection, parasite and worm infestations, fever, ulcers and as an insecticide [68,69]. Its leaves are used for helminthiasis treatment while bark is a powerful astringent and used as a tonic [68,69].

4.1.5. *Annona Coriacea*

Annona coriacea Mart. is a species belonging to the *Annona* genera, commonly known as “marolo”, “araticum” and “araticum-liso” [70]. This plant is distributed across Paraguay and Brazil, with little available information about its ethnomedicinal uses [71]. It is a small tree (3–6 m) and its edible fruit consist of an ovoid-obtuse syncarp and weighing up to 1.5 kg [72]. The leaves are glabrous on the ventral surface, obovate, and the base is frequently cordate and margin undulate [73]. The flowers are terminal, thick, solitary and having fleshy petals with colors shifting between orange and pink [73]. The leaves are traditionally used as carminatives, anthelmintics, antirheumatics and in the treatment of stomatitis, headaches, abscesses, neuralgia, rheumatism, ulcers and dermatitis [74,75]. Both seeds and fruits are toxic when crushed and exhibited effects against ectoparasites like lice [75].

4.1.6. *Annona Senegalensis*

Annona senegalensis is a small tree 2–6 m tall that is commonly known as wild custard apple and wild soursop [76]. This plant is native to tropical east and northeast, west and west-central, and southern Africa and islands in the western Indian Ocean [76]. Its leaves are simple, alternate, oblong, green to bluish-green, ovate or elliptic, and mainly lack hairs on the upper surface and brownish hairs on the lower surface [77]. This plant has been used in traditional medicine as a pain reliever, antioxidant, antidiarrheal, antitrypanosomal, antimalarial, anti-inflammatory, antimicrobial, antiparasitic, anticonvulsant and as an anti-snake venom [78]. It has been reported that the leaves of *A. senegalensis* are used for the treatment of tuberculosis, yellow fever and smallpox, whereas stem bark is reported for the treatment of injury from venomous animals [79]. The root was also reported for treating erectile dysfunction, tuberculosis, gastritis, reproductive deficiency and in the management of malaria and diabetes [80].

4.1.7. *Annona Vepretorum* Mart

Annona vepretorum is commonly recognized as ‘bruteira’, is a small tree of 2.5–10 m high native to the Brazilian biome Caatinga [81]. The fruits of *A. vepretorum* can be consumed either raw or as juice for nutritional purposes [82]. Traditionally, a decoction of the leaves have been used to bathe in for the treatment of allergies, yeast, skin diseases

and microbial infections, whereas the root is traditionally used to treat snake and bee bites, inflammatory conditions and heart pain [81].

4.1.8. *Annona Salzmannii*

Annona salzmannii is a tree of 6–20 m high that known as “araticum-da-mata” and “araticum”apé” [83]. It is commonly cultivated in Brazil especially in the States of Bahia, Pernambuco, and Paraíba [83]. Its root, seeds and leaves are used in folk medicine for treating several illnesses like ulcers, dysentery and inflammatory conditions [83]. The leaves and bark of *A. salzmannii* are utilized for the treatment of tumors, diabetes and inflammatory conditions [84].

4.1.9. *Annona Crassiflor*

Annona crassiflor is known as araticum of cerrado or cerradão [85]. It is a small tree that bears a typical fruit known as araticum of cerrado or cerradão [86]. The fruits are highly consumed “in natura” by native people and can be used to make juice, jelly and ice-cream [85,86]. In folk medicine, the seeds are used to treat scalp infections, and infusions of the leaves and seeds are utilized for their antidiarrheal and antitumor properties [85]. For more details about the botanical characteristics and traditional uses of *Annona* species, see Tables 4 and 5.

Table 5. Traditional uses of common *Annona* species.

| Species | Region | Local Name | Medicinal Uses | Part Used | Mode of Usage | References |
|-----------------------------------|--|---|---|---|---|------------|
| <i>A. ambotay</i> Aubl | French Guiana | Not reported | Treating fever | Leaves and bark | Leaves and bark crushed and rubbed on body | [18] |
| <i>A. cherimola</i> Mill. | Tropical America Asia Gabon Cultivated in Spain and Australia | Cherimola Cherimoya Chirimoya Custard apple Mao ye fan li zhi | Abortion Anti-anxiety Cough Diarrhea Hypercholesterolemia Infections Painful inflammations Parasitic Sedative | Aerial parts Fruit Leaf Root Seed Stem | Not reported | [87,88] |
| <i>A. coriacea</i> Mart. | Brazilian (Cerrado, Caatinga) | Araticum Marolino Marolo | Anthelmintic Chronic diarrhea Inflammation Leishmaniasis Malaria Rheuma | Leaves Root Seeds | Not reported | [89] |
| <i>A. cornifolia</i> St-Hil | Bolivian and Brazilian savannah | Not reported | Antiulcerative (green fruit) | Seeds | Not reported | [90] |
| <i>A. crassiflora</i> Mart. | Brazil (Cerrado) | Araticum of the Cerrado Araticum-mirim Marolo Panã | Analgesic Antimicrobial Antirheumatic Carminative Digestive Rheumatism, Anti-inflammatory Wound healing | Leaves Root bark Root wood Seeds Fruit | Not reported | [43,91,92] |
| <i>A. cuneata</i> (Oliv.) R.E. Fr | Congo | Not reported | Asthenia Female sterility Hernia Parasitic infections Venereal diseases | Root bark Stem bark | Not reported | [93] |
| <i>A. dioica</i> A. St.-Hil. | Brazil (Cerrado, Pantanal) | Ceraticum and araticum | Diarrhea Rheumatism | Fruits Leaves | Dried leave paste and fresh fruit decoction | [11,33] |

Table 5. Cont.

| Species | Region | Local Name | Medicinal Uses | Part Used | Mode of Usage | References |
|--------------------------------------|--|---|--|--|---|------------|
| <i>A. diversifolia</i> Saff | Tropical forest of Central America China | Ilama Papausa White anona, Yi ye fan li zhi | Arthritic pain Anti-spasmodic | Leaves Seeds | Not reported | [40,53] |
| <i>A. foetida</i> Mart | Brazil | Araticum-da-caatinga | Malaria | Bark and leaves | Decoction of bark and leaf | [94] |
| <i>A. glabra</i> L | Caribbean | Mamain | Fever | Leaves | Not reported | [95] |
| <i>A. glauca</i> Schumacher & Thonn | West Tropical Africa (Senegal, Ghana, Suriname) | Dangan Mampihege, Mandé sunsun Tangasu | Arachnoides Blennorrhoea Diuretic Fish-poisons Insecticides | Roots Seeds | Not reported | [96] |
| <i>A. haematantha</i> Miq | French Guiana South American tropical rainforest | Not reported | Fever | Leaves Bark Roots | Leaves and bark crushed rubbed on body | [18,97] |
| <i>A. montana</i> Macfad. | South America Southern Asia The Amazon Brazil (Mata Atlântica, Pantanal) | Mountain soursop Shan Di fan li zhi false Graviola Araticum grande Jacá do Pará | Against snake bite Against obesity | Leaf Pulp juice Seed Stem Twig | Not reported | [98,99] |
| <i>A. muricata</i> Linn | Brazil | Araticum Condessa Graviola | Anthelmintic Analgesic, neuralgia, rheumatism, arthritis pain | Fruit, juice, and crushed seeds Fruit and leaves | Juice of fruit Water extraction of the leaf | [66] |
| <i>A. pickelii</i> (Diels) H. Rainer | Mexico Caribbean Central America Venezuela Colombia BelizeCentral America South America Southern Asia Africa Madagascar | Sincollo Soncoyo Bullock's-heart Custard apple Anona blanca Anona Niiu xin fan li zhi | Contraceptive Blood dysentery Cold Stomachache Fainting spinal disorders Fever Hysteria Influenza Mental depression Skin diseases Unhealthy ulcers Wounds | Leaf Leaf Root Stem Seed Aerial parts Bark Fruit Leaf Root Seed Stem bark | Not reported | [100,101] |
| <i>A. reticulata</i> Linn | West Indies | Ramphal | Bronchitis Asthma Bowel inflammation | Fruit Seeds Leaves | Decoction of fruit Oral ingestion of powdered seeds Oral ingestion of the leaf powder | [102] |
| <i>A. senegalensis</i> Persoon | Nigeria | Ukopko (Idoma) | Anti-inflammatory Analgesic Anthelmintic Cancer Diarrhea Epilepsy Infectious diseases Inflammations Sleeping sickness Snakebite Cardiovascular diseases Diabetes Febrile seizures Gout Mental disorders Painful | Leaves Seed Stem Bark Root bark | Roots and bark are ground together and their decoction is used | [103] |
| <i>A. squamosa</i> Linn | Cameroon | Sugar apple (English); Kedahan (Yambetta) | Vomiting, abscesses, muscle aches, fever, and skin disease | Leaves | Decoction of leaves | [13] |
| <i>A. vepretorum</i> Mart | Brazil | Araticum Bruteira | Analgesic and anti-inflammatory | Leaves | Methanolic leaf extract | [104] |

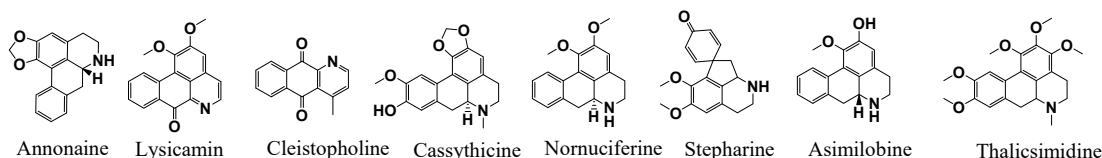
4.2. Traditional and Ethnomedicinal Uses of Annona Genus

Traditionally, the *Annona* species have been used widely. For instance, antidiarrheal effects have been reported for *A. reticulata*, *A. muricata* and *A. salzmannii*, whereas *A. cherimola*, *A. squamosa* and *A. reticulata* have been reported for their antiparasitic effects (Table 5) [10]. Moreover, both *A. vepretorum* and *A. salzmannii* have been also reported for anti-inflammatory effects [10]. *A. purpurea* and *A. reticulata* have been used to treat fever, while anticancer effects have been reported for *A. senegalensis* and *A. muricata* [105,106]. Furthermore, *A. foetida*, *A. muricata* and *A. glabra* have been traditionally used to treat rheumatism [107], while *A. reticulata*, *A. salzmannii*, *A. foetida* and *A. squamosa* have been described for treating ulcers [4]. In Indonesia, the fruit juice of *A. muricata* has used as a diuretic and to treat liver ailments and leprosy [108], whereas leaves was used to treat spasms, boils and as an aphrodisiac [36]. The leaves of *A. diversifolia* have been used as anti-inflammatory, anticonvulsant and analgesic agents [52]. Ethnobotanically, despite reports of the toxicity of *A. muricata* seeds, the powder of toasted seeds has been reported to be used as an emetic and cathartic in the traditional Mexican pharmacopeia [36]. To South-east Asian people, the immature fruit of *A. reticulata* was used to treat both dysentery and diarrhea, and a decoction of roots was used to cure toothache and as an antipyretic [109]. Additionally, a decoction of leaves has been used internally against worms and topically to treat abscesses and boils [109]. Finally, the leaves of *A. squamosa* have been used as tonic and cold remedy in tropical America and systemically to cure dysentery in India [108].

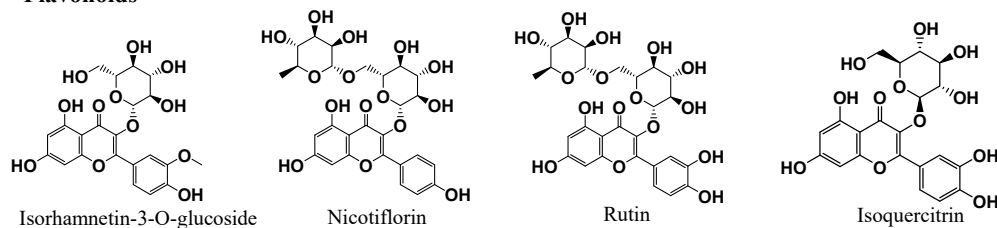
5. Phytochemistry of Annona Species

A wide range of secondary metabolites, including acetogenins, flavonoids, alkaloids and essential oils (Figure 1), from nearly every part of *Annona* plants, have been discovered, isolated and characterised (Table 6). The plants of the *Annona* genera are also found to be rich in minerals and vitamins, for instance, calcium, potassium, magnesium, sodium, copper, zinc, selenium, phosphorus, iron, vitamin C, pantothenic acid B₅, thiamine and riboflavin [6].

Alkaloids



Flavonoids



Terpenoids

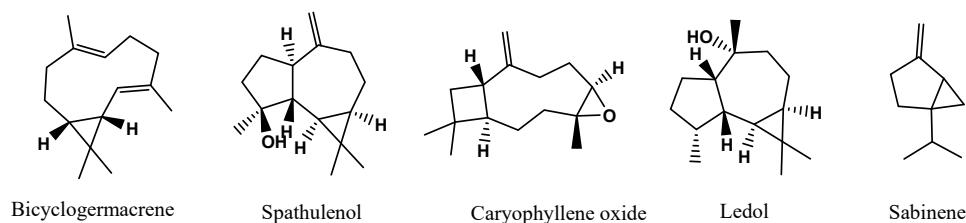
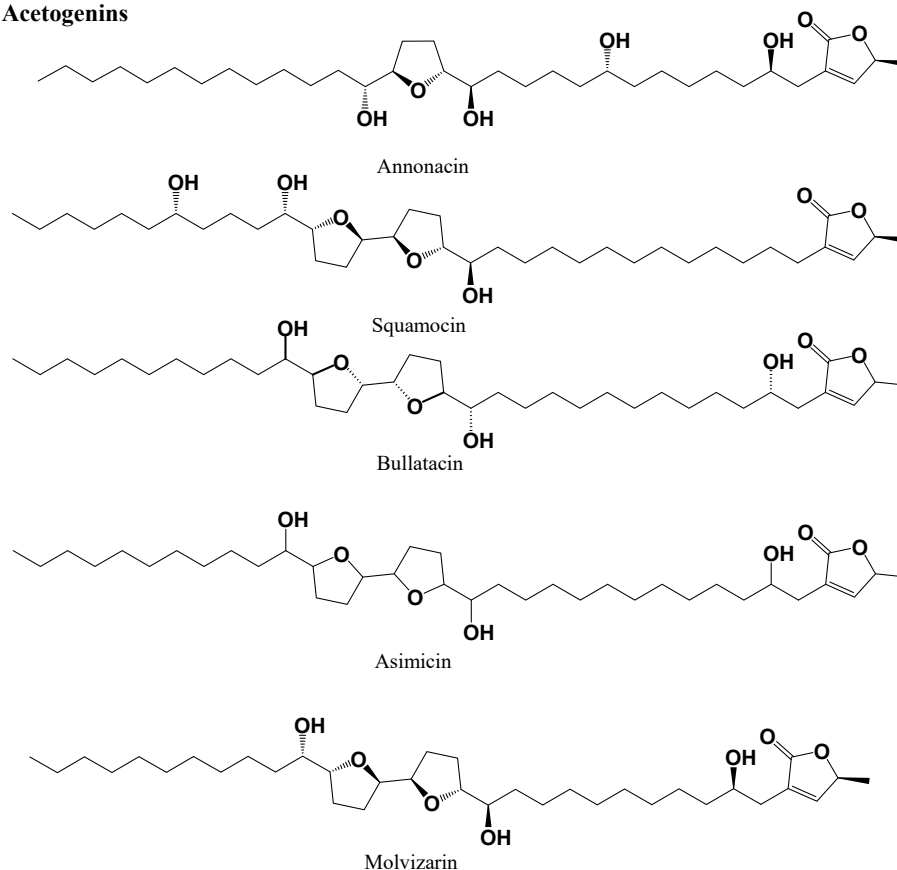


Figure 1. Cont.

Acetogenins

Figure 1. Structure of selected compounds identified in *Annona* species.Table 6. Compounds isolated from plants of *Annona* genus.

| Species | Part | Isolated Compounds | References |
|--------------------------------|-------|---|------------|
| <i>A. amazonica</i> R.E. Fries | Stems | Cassythicine Liriodenine (ALK) | [110] |
| <i>A. cherimola</i> | Root | Corytenchine, Isocoreximine (ALK) | [111–116] |
| | Fruit | α -Pinene, α -Thujene, Terpinen-4-ol, Germacrene D (ESO) | |
| | Seed | 2,4- <i>cis</i> -Annocherinones, Annocherin, 2,4- <i>trans</i> -Isoannonacins, Annocherimolin, Annomolin, Annomocherin, Annomontacin, Annonacin, Asimicin, Tucumanin, 2,4- <i>trans</i> -Annocherinones, 2,4- <i>cis</i> -Isoannonacins, <i>cis</i> -Annonacin, Annogalene, Annosenegalin, Annomolon A, Annomolon B, Cherimolacyclopeptide C (ACT) | |
| | Stem | Annocherine A and B, Artabonatin B, Romucosine H, Cherianoine (ALK) | |
| <i>A. coriacea</i> Mart. | Bulb | Crolechinic acid, Crolechinic acid (methyl ester), Annonene, Annonalide (ESO) | [117–123] |
| | Seed | Gigantecin, Coriapentocin A and B, Bullacin (ACT) | |
| | Leaf | Quercetin-3-O- β -(6''-O- β -glucosyl)-glucoside, Quercetin-3-O- β -(6''-O- α -rhamnosyl)-galactoside, Trigonelline, Rutin, Hyperin, Hyperin, Isorhamnetin-3-O- β -glucoside, Isorhamnetin-3-O- β -galactoside, Isoquercitrin, Isoquercitrin, Nicotiflorin, Biorobin, Keioside, Cacticin, Isorhamnetin-3-O- β -glucoside, Narcissin, Rutin (FLA) | |
| | Root | Coriacin, Coriadienin, Coriaheptocin A and B, Coriacyclodienin, Coriacycloenin, 4-Deoxycoriacin, Annoheptocin A and B (ACT) | |
| <i>A. crassiflora</i> | Leaf | Kaempferol-3-O- β -diglucoside, Kaempferol-3-O- β -glucoside, Quercetin-3-O- β -D-galactopyranoside, Epicatechin, Quercetin-3-O- β -L-arabinopyranoside (FLA) | [124] |

Table 6. Cont.

| Species | Part | Isolated Compounds | References |
|---|---------------------|--|------------|
| <i>A. foetida</i> | Bark | Annomontine, <i>N</i> -Hydroxyannomontine, Liriodenine, <i>O</i> -methylmoschatoline (ALK) | [125–127] |
| | Leaf | (<i>E</i>)-caryophyllene, Bicyclogermacrene, α -Copaene (ESO) | |
| | Branch | Atherospermidine (ALK) | |
| <i>A. glabra</i> | Fruit | 16 α -17-Dihydroxy-ent-kauran-19-oic acid, 16 α -Hydro-ent-kauran-17-oic acid, 16 β -Hydroxy-17-acetoxy-ent-kauran-19-oic acid, 16 α -Hydro-19-al-ent-kauran-17-oic acid, 16 β -Hydro-ent-kauran-17-oic acid, 19-nor-ent-kauran-4 α -ol-17-oic acid, Annoglabasin A and B, ent-Kaur-15-ene-17,19-diol, ent-Kaur-16-en-19-ol, ent-Kaur-16-en-19-oic acid, Methyl-16 α -hydro-19-al-ent-kauran-17-oate (ALK) | [128–132] |
| | Fruit & stem | Annoglabasin A, B, C, D, E and F, (–)-Anonaine, (–)-Asimilobine, (–)-Kikemanine, (–)-Nornuciferine, (+)-Stepharine, Blumenol A, Liriodenine, <i>N</i> - <i>p</i> -Coumaroyltyramine, (–)- <i>N</i> -Formylanonaine, (+)-Nordomesticine, Annobrine, Dehydrocorydalmine, Lysicamine, <i>N</i> -trans-Feruloyltyramine (ALK), 6- <i>O</i> -Palmitoyl- β -sitosteryl-D-glucoside, β -Sitosteryl-glucoside, Stigmasteryl-D-glucoside, β -Sitosterol, Stigmasterol (STE) | |
| | Seed | Isodesacetyluricin (ACT) | |
| | Leaf | Bullatanocin, Glabracins A and B, Javoricin, Glacins A and B (ACT), 3- <i>O</i> - α -L-Arabinopyranoside, 3- <i>O</i> - β -D-Glucopyranoside (GLU) (–)-Actinodaphnine, (–)-Asimilobine, (–)-Anolobine, (–)- <i>N</i> -Methylactinodaphnine, (–)-Roemeroline, (+)-Boldine, (+)-Norisodomesticine, (+)-Stepharine, Liriodenine, (–)-Pallidine, (+)-1 <i>S</i> ,2 <i>S</i> -Reticuline <i>N</i> -oxide, (+)-Magnoflorine, (+)-Reticuline (ALK) Quercetin, Quercetin–3- <i>O</i> - β -D-galactopyranoside (FLA) | |
| <i>Annona leptopetala</i> (R.E.Fr.) H. Rainer | Leaves and branches | Laurotetanine, Nornuciferine, Corypalmine, Norannuradhapurine Anonaine (ALK) | [133] |
| <i>A. montana</i> | Leaf | Annotatine, Annotetine, Liriodenine, Argentinine (ALK), β -Sitosterol- β -D-glucoside, β -Sitosterol (STE), Montanacin-K, L, C, D, B and E, Annonacin-10-one, Annonacin-A, <i>cis</i> -Annonacin-10-one, Annonacin, <i>cis</i> -Annonacin (ACT) | [134–137] |
| | Seeds | Montalicens G, Montalicens H Monlicins A & B, Murisolin, 4-Deoxyannomontacin, Muricatacin (ACT) | |
| | Stem | <i>N</i> -trans-Feruloyltyramine, <i>N</i> - <i>p</i> -Coumaroyltyramine, <i>N</i> -trans-Caffeoyltyramine (PHE) | |
| <i>A. muricata</i> | Seed | 2,4- <i>cis</i> -Gigantetrocinone, 2,4- <i>trans</i> -Isoaiinonacin, 2,4- <i>trans</i> -Gigantetrocinone, 2,4- <i>trans</i> -Isoannnonacin-10-one, Gigantetrocin-A, Muricatenol, Annomontacin, Gigantetronenin, Annonacin A, Annoreticum-9-one, <i>cis</i> -Annomontacin, Murisolin, Muricin H, Xylomaticin, Muricin I, <i>cis</i> -Annonacin, <i>cis</i> -Goniothalamycin, <i>cis</i> -Annonacin-10-one, Arianacin, Javoricin, Donhexocin, Murihexol, Cohibins C, Cohibins D, Gigantetrocin B, Longifolicin, Muricin A, B, C, D, E, F and G, Annomuricin B and C (ACT) | [138–146] |
| | Stem bark | Muricatin A, B and C (ACT) | |
| | Fruit | Epomuricenins-A and B, Epomuricins-A and B, Epomusenins-A and B, Muricin J, K and L (ACT) Asimilobine, Nornuciferine, Annonaine (ALK) | |
| | Fruit & Root | Sabadelin (ACT) | |
| | Leaf | Annonacin, Annomuricin C, Muricatocin C, (2,4- <i>cis</i>)-10 <i>R</i> -annonacin-A-one, (2,4- <i>trans</i>)-10 <i>R</i> -annonacin-A-one, Annohexocin, Annomutacin, Annopentocins A, B and C, Annomuricine, Muricapentocin, Annomuricins A and B, <i>cis</i> -annomuricin-D-ones, trans-annomuricin-D-ones, Muricatocins A and B, Murihexocin A and B, Muricoreacin, Murihexocin C (ACT) (<i>R</i>)-4- <i>O</i> -methylcoclaurine, (<i>R</i>)- <i>O</i> , <i>O</i> -dimethylcoclaurine, (<i>R</i>)-Anonaine, Annonamine, (<i>S</i>)-Norcorydine, Anonaine, Isolaureline, Xylopinine (ALK) Catechine, Epicatechine, Gallic acid, Chlorogenic acid, Kaempferol, Kaempferol-3- <i>O</i> -rutinoside, Quercetin-3- <i>O</i> -rutinoside, Quercetin-3- <i>O</i> -glucoside, Quercetin-3- <i>O</i> -neohispidoside, Quercetin-3- <i>O</i> -robinoside, Annoionols A and B, Annoionoside (FLA) | |
| | Leaf & seed | Annonacin, Annocatacin A and B, Annonacinone, Annocatalin, <i>cis</i> -Corossolone, Goniothalamycin, Isoannnonacin, Corossolone (ACT) | |
| | Pericarp | Annonacin, Annonacin A, Annomuricin A (ACT) | |
| <i>A. muricata</i> | Root | Annonacin, Muridienins-1, 2,3 and 4, Chatenaytrienins-1, 2 and 3, Muricadienin, Montecristin, <i>cis</i> -Panatellin, <i>cis</i> -Reticulatacin-10-one, <i>cis</i> -Uvariamicin IV, Coronin, <i>cis</i> -reticulatacin, <i>cis</i> -Solamin, Cohibins A and B (ACT) | |

Table 6. Cont.

| Species | Part | Isolated Compounds | References |
|-----------------------------|--------------|---|--------------|
| <i>A. purpurea</i> | Leaf | Lirinidine, 7-Formyl-dehydrothalicimidine, 7-Hydroxy-dehydrothalicimidine, N-Methylasimilobine, N-Methylaurotetanine, Thalicsimidine, Norpurpureine (ALK) | [105,147] |
| | Root | Annomontine (ALK) | |
| <i>A. reticulata</i> | Leaf | Dopamine, Salsolinol, Spathenelol, Muurolene, Coclaurine, Copaene, Eudesmol (ESO), Squamone, Solamin, Rolliniastatin 2, Annorecticin-9-one, Annomonicin, Annonaretin A (ACT) | [54,148–152] |
| | Stem bark | Dopamine, Salsolinol (ESO), Reticullacinone, Rolliniastatin-2, (ACT) | |
| | Root | Liriodenine, Norushinsunine, Neoannonin, Reticuline (ALK) Spathenelol, Copaene, Eudesmol, Muurolene (ESO) | |
| | Bark | Reticulatacin, Liriodenine, Copaene, Coclaurine (ALK) Patchoulane (ESO) Molvizarin, Bullatacin (ACT) | |
| | Seed | Squamocin, <i>cis</i> -/ <i>trans</i> -isomurisolenin, Bullatacin, <i>cis</i> -/ <i>trans</i> -Bullatacinone, Annorecticin, Annorecticin-9-one, Solamin, Annomonicin, Isoannonaretin, Rolliniastatin-1, 2 Squamone, Annonaretin, 2, 4- <i>cis</i> -Isoannonaretin, Solamin, Murisolin, Reticullacinone, Annomonicin, Sitosterol, Annorecticin (ACT), Myrcene, Limonene, Germacrene D (ESO) | |
| | Fruit | Terpinen-4-ol, Germacrene D, Limonene, Pinene, Myrcene (ESO) | |
| | Root bark | Anonaine, Michelalbaine, Reticuline, Oxoushinsunine (ALK) | |
| | Leaf | (–)-Roemerine, α -Humulene, γ -Cadinene, Germacrene D, β -Caryophyllene (ESO) | |
| <i>A. senegalensis</i> | Aerial parts | (–)-Anonaine, (–)-Asimilobine, (+)-Nornantenine (ALK) (+)-Catechin (FLA) | [153,154] |
| | Seed | Annogalene, Annosenegalin (ACT) | |
| <i>Annona sericea</i> Dunal | Leaf | Nornantenine, Nornuciferine, Isoboldine, Lysicamine, Hydroxynornuciferine (ALK) | [155] |
| <i>A. squamosa</i> | Leaf | (–) Anonaine, O-Methylarmepavine, β -Caryophyllene, β -Cedrene, (E)-Caryophyllene, Germacrene D, Bicyclogermacrene, Quercetin-3-O-glucoside (ESO) | [156,157] |
| | Bark | 2,4- <i>cis</i> -Mosinone A, 2,4- <i>trans</i> -Mosinone A, Annorecticin-9-one, Mosin B and C, Bullatacin, Bullatacinone, Squamone (ACT) | |
| | Pulp fruit | α -Pinene, Limonene, Sabinene (ESO) | |
| | Stem | 11 <i>ent</i> -Kauranes, 10-nor- <i>ent</i> -Kaurane-4 α ,16 β ,17-triol, 16 α ,17-Dihydroxy- <i>ent</i> -kauran-19-oic acid, 16 β ,17-Dihydroxy- <i>ent</i> -kauran-19-al, 16 β -Hydro- <i>ent</i> -kauran-17,19-dioic acid, 17-Hydroxy-16 β - <i>ent</i> -kauran-19-oic acid, <i>ent</i> -Kaur-16-en-19-oic acid, 16 α ,17-Dihydroxy- <i>ent</i> -kauran-19-al, 16 α -Hydro-19-al- <i>ent</i> -kauran-17-oic acid, 16 β ,17-Dihydroxy- <i>ent</i> -kauran-19-oic acid, 16 β -Hydroxy-17-acetoxy- <i>ent</i> -kauran-19-oic acid, 4 α -Hydroxy-19-nor- <i>ent</i> -kauran-17-oic acid (ALK) | |
| | Seed | Neoannonin-B, Annosquamins A, B and C, Annosquacin-I, Annosquamin A, B and C, Annosquatin A and B, Annotemoyin-1 and 2, Cherimolin-1 and 2, Diepomuricanin A and B, Dieporeticenin, Dieposabadelin, Squadiolin A, B and C, D, E, F, G, H, I, J, K, L, M and N, Squamostanin A, B, C, D, E and F, Cyclosquamosin A, B, C, D, E, F, G, H and I, Squamin A and B (ACT) | |
| <i>A. vepretorum</i> | Leaf | Spathulenol, Bicyclogermacrene, α -Phellandrene (ESO) | [116,158] |

ALK (Alkaloids), ACT (Acetogenins), ESO (Essential oils). STE (Sterols), FLA (Flavonoids), PHE (Phenolics).

6. Pharmacological Properties of *Annona* Species

The species of the *Annona* genera have been reported to elicit a diversity of biological activities such as antitumor, anti-inflammatory, antioxidant, antinociceptive, antiprotazoal, antipyretic, antiulcer, antihyperglycemic, anthelmintic, antileishmanial, antimalarial, antidiarrheal, antifungal and antimicrobial promoted by whole extracts, fractions, or pure compounds (Table 7).

Table 7. Pharmacological activities of *Annona* species.

| Species | Biogeographical Distribution | Used Part | Traditional Use | Pharmacological Activities | Extract/Compound Evaluated | References |
|------------------------|--|---|---|---|---|------------|
| <i>A. ambotay</i> | South American tropical rainforest | Trunkwood | Antipyretic | Antimicrobial | Alkaloids | [159] |
| <i>A. bullata</i> | Endemic of Cuba | Bark | Not reported | Antitumoral | 32-Hydroxybullatacinone | [160] |
| <i>A. cherimola</i> | Tropical America, Asia, Spain, Australia, Gabon | Aerial parts Fruit Leaf Root Seed Stem | Abortion Anti-anxiety Cough Diarrhea Hypercholesterolemia Infections Painful inflammations Parasitic Sedative | Antidepressant Antifungal Antiprotozoal Antitumoral Antihypercholesterolemic Antitumor Antiviral Insecticidal Vasodilator | Acetogenins: Molvizarin, Squamocin, Cherimolin-1, Motrilin, Aherradurin, Tucumanin, Annomocherin, Annonacin, Annonomontacin, Alkaloids: Roemerine, Anonaine, Dehydroroemerine | [37,161] |
| <i>A. coriacea</i> | Brazilia (Cerrado and Caatinga) | Leaf Root Seed | Leishmaniasis Malaria Rheuma Anthelmintic Chronic diarrhea Inflammation | Antifungal Anti-inflammatory Antitumoral Insecticidal Leishmanicidal Trypanocidal | Acetogenins: Annoheptocins A-B, coriacin, 4-Deoxycoriacin, Coriaheptocins A-B, Coriadienin, Gigantecin | [10] |
| <i>A. muricata</i> | America, Asia, Africa | Bark Leaf Fruit Root Root bark Seed Stem bark | Anthelmintic Antiscorbutic Asthma Cancer Cough Cystitis Diabetes Diuretic | Anti-arthritis Antidepressant, Antidiabetic Anti-inflammatory Antimicrobial Antimalarial Antiviral Hepatoprotective | Acetogenins: Solamin, Muricin H, Muricin I, cis-Annonacin, Muricins A-G, Muricoreacin, Muricapentocin, Gigantetrocin A, Annonopentocins A-C | [162,163] |
| <i>A. salzmannii</i> | Brazil | Bark Leaf | Not reported | Antioxidant Antimicrobial | Alkaloids: Reticuline, Anonaine, Laurelliptine, Isoboldine | [101] |
| <i>A. senegalensis</i> | Madagascar, Comoros, Cape Verde, Tropical Africa | Leaves Seed Stem Bark Root bark | Anti-inflammatory and Analgesic Anthelmintic Cancer Diarrhea Epilepsy Infectious diseases Inflammations Sleeping sickness Snakebite | Anticonvulsant Antidiabetic Antidiarrheal anthelmintic Anti-inflammatory Antimalarial Antimicrobial Antioxidant Insecticidal Hepatoprotective Antitumoral | Aqueous extract. Ethanolic extract Terpenoids, coumarins, flavonoids, tannins, alkaloids, quinones. Methanolic extract containing Annonosnegalin, Annonalene. | [10,164] |
| <i>A. squamosa</i> | Tropical America, Asia Australia | Seed Stem Bark Root bark | Analgesic Anthelmintic Antirheumatic Cancer Digestive Headache Anti-inflammatory Antimicrobial Carminative | Antibacterial Antidiabetic Antilipidemic Antioxidant Antimalarial Antigenotoxicity Antileishmanial | Acetogenins: Squadiolins A and B, Squafosacin B, Bullatacinona, Squamona, Tetrahydrosquamone Monoterpenes: Limonene, β -Cubebene, β -Caryophyllene, Spathulenol, Caryophyllene oxide | [10] |

6.1. Antibacterial Activity

The antibacterial activities of *Annona* species have been reported in many studies, for example, both methanolic and ethanolic leaf extracts of *A. muricata* exhibited antimicrobial activity against *Staphylococcus aureus*, and this activity was attributed due to the presence of flavonoids, alkaloids and steroids in the extract [165,166]. In contrast, an aqueous extract of the peel of *A. muricata* did not show any activity [165,166]. The root of *A. reticulata* has also been investigated for its antibacterial activity against Gram-positive *Staphylococcus aureus*, *Bacillus cereus* and *Bacillus subtilis*, and Gram-negative *Pseudomonas aeruginosa*, *Escherichia coli* and *Salmonella typhi* [54]. The root extract was found to possess pronounced activity against *Bacillus cereus* as well as notable inhibition against all tested strains [54]. Moreover, the leaves of *A. cherimola* have been reported for antibacterial activity against *Staphylococcus aureus* and *Bacillus subtilis* with growth inhibition zone diameters of 11 mm and 14 mm, respectively [37]. The aqueous and methanolic seed extracts of *A. squamosa* have reported activity against *Staphylococcus aureus* with Minimum Inhibitory Concentrations (MIC) of 50 mg/mL and Minimum Bactericidal Concentrations (MBC) of 100 mg/mL [167]. The activity of the isolated compounds from *Annona* species has been reported in various studies; for instance, the fatty alcohol 11-hydroxy-16-hentriacontanone

isolated from leaves of *A. squamosa* has a reported activity against Gram-positive and Gram-negative bacterial strains, with MIC values of 25–50 µg/mL [168]. Additionally, the alkaloids liriodenine, annonaine, asimilobine, reticuline and cleistopholine isolated from *A. salzmannii* demonstrated activity against a range of Gram-positive bacteria, including *Kocuria rhizophila*, *Staphylococcus aureus*, *Staphylococcus epidermidis* and *Enterococcus faecalis* with MIC values from 25 to 500 µg/mL [36]. Notably, annonaine and asimilobine had activity equal to or better than the control chloramphenicol (MIC 50 µg/mL) against many of the species tested [36].

6.2. Anticancer and Antiproliferative Activity

Various studies have reported the anticancer activity of either crude extracts or isolated compounds from *Annona* species. For example, the leaves extract of both *A. squamosa* and *A. reticulata* exhibited potent antiproliferative effects against two human T-lymphotropic virus type = 1 infected cell lines (MT-1 and MT-2) with EC₅₀ values from 0.1 to 1 µg/mL [169]. In in vitro studies, the ethanolic extract of *A. muricata* leaves was reported for its cytotoxicity against promonocytic leukemic cells (U-937) with an LC₅₀ = 7.8 µg/mL [170]. Isocoreximine isolated from *A. cherimola* demonstrated cytotoxicity against multiple cancer cell lines. At a concentration of 50 µg/mL, isocoreximine inhibited cell viability of the breast cancer cell line (MCF-7) by 85.76%, human colorectal carcinoma cell line (HCT-15) by 63.05%, human prostate tumor cell line (PC-3) by 78.71%, human astrocytoma cell line (U-251) by 65.23% and human leukemia cell line (K-562) by 94.15% [112].

The antiproliferative activity of methanolic extracts from the leaves and seeds of *A. coriacea* was tested in vitro against a range of human tumor cell lines; including melanoma (UACC-62), non-small cell lung cancer cells (NCI-H460), colon cancer cell line (HT29), breast cancer (MCF-7) and leukemia (K-562). The seed extracts displayed potent antitumor activity with GI₅₀ values between 0.02 and 3.83 µg/mL, and the leaf extracts exhibited anticancer activity at concentrations ranging from 0.02 to 0.08 µg/mL [35,171]. The cytotoxicity of annonacin, found in many *Annona* species, has been reported against various cell lines derived from cervical cancer (HeLa and HeLa S3) with IC₅₀ 0.219 and 0.426 µg/mL, and ovarian cancer (PA-1 and SKOV3) with IC₅₀s of 0.452 and 0.411 µg/mL [172]. The cytotoxicity of annonacin has also been demonstrated against bladder cancer (T24), breast cancer (MCH7) and skin cancer (BCC-1) with IC₅₀s 0.324, 0.433 and 0.427 µg/mL, respectively [172]. The cytotoxicity of five other acetogenins (squamocin M, annofolin, isolongimicin B, glaucanisin, and annotacin) isolated from *A. cornifolia* against human breast cancer (MCF-7) was reported, with IC₅₀s of approximately 0.3 µM [173]. In an in vitro study, the annoaceous acetogenins laherradurin and cherimolin-2 isolated from *A. diversifolia* were shown to have ED₅₀s of 0.015 and 0.05 µg/mL, respectively, against the cervical cancer cell line (HeLa) [174].

In clinical studies, the anticancer activity of *A. muricata* has been reported in a small number of studies. A patient diagnosed with breast cancer has maintained stable disease activity with no reported side effects after using an aqueous extract of *A. muricata* leaves for more than five years [175]. Another patient with metastatic ovarian cancer experienced disease stability after starting to take a complementary medication containing *A. muricata* as a tablet [176]. Finally, the effect of *A. muricata* leaves extract revealed higher cytotoxicity in the supplemented group with colorectal cancer compared with the placebo group in a randomized controlled trial [177].

6.3. Antidiabetic and Antilipidemic Activity

Multiple studies have investigated the antidiabetic activity of various extracts from *Annona* plants such as *A. cherimola*, *A. squamosa*, *A. muricata* and *A. reticulata*. The ethanolic leaf extract of *A. cherimola* (300 mg/kg) was administered to alloxan-induced type 2 diabetic rats, and four hours later, blood glucose level had decreased from 331.5 mg/dL to 149.2 mg/dL [58]. The young leaves of *A. squamosa*, often in combination with black pepper (*Piper nigrum*), have been used in northern Indian traditional medicine as an anti-diabetic, and are

still in use today. Administration of aqueous *A. squamosa* leaf extract to streptozotocin-nicotinamide type-2 diabetic rats resulted in decreased blood glucose and increased levels of serum insulin [178]. Another traditional Indian medicine used as an antidiabetic and anti-lipidemic is a polyherbal formulation of *A. squamosa* fruits and *Nigella sativa* seeds. The polyherbal formulation administered over a one-month period, dose-dependently decreased blood glucose and increased insulin in streptozotocin-induced diabetic rats, with a dose of 200 mg/kg showing similar to the effects of a dose of 250 mg/kg tolbutamide [179]. A single dose of 100 or 200 mg/kg of aqueous leaf extract of *A. muricata* did not inhibit blood glucose levels in normal rats; however, the same doses administered to the diabetic rats effectively lowered blood glucose levels by 31.77% and 45.77%, respectively [180]. Finally, a dose of 100 mg/kg of both methanolic extract and the residual fractions of *A. reticulata* leaves decreased blood glucose levels from 432.33 to 371.67 mg/dL and 417.83 to 402.50 mg/dL, respectively, in streptozotocin-induced diabetic rats [181]. *A. cherimola* leaf extract was also found to decrease HbA1c by 7% and lead to a significant decrease in urine glucose over a 28-day subchronic study in streptozotocin-induced diabetic mice [182]. These studies support the traditional use of *Annona* species as antidiabetics, suggesting that further identification of the active constituent(s) with antidiabetic properties and clinical studies of a longer duration are warranted.

Limited studies have also investigated the antilipidemic activity of some *Annona* species. The polyherbal formulation of *A. squamosa* fruits and *Nigella sativa* seeds (200 mg/kg) administered to streptozotocin-induced diabetic rats for one month also resulted in significant inhibition of the formation of both lipid peroxide and tissue lipids [179]. Administration of an extract of *A. muricata* extracts resulted in reductions in the serum total cholesterol, low-density lipoprotein cholesterol and triglycerides in diabetic rats [183]. The tea infusion of leaves from *A. cherimola* (1.5 g) also elicited a reduction in the total cholesterol, triglycerides, and low-density lipoprotein by 15.4, 21.9 and 63.2%, respectively, in streptozotocin-induced diabetic rats [182].

6.4. Anti-Inflammatory Activity

The anti-inflammatory effects of *Annona* plants have been reported in many studies; for instance, after one day of orally administered doses of 200 and 400 mg/kg of *A. squamosa* root extract in an acute carrageenan-induced rat paw edema model, significant inhibition was produced with 24% and 47% inhibition respectively compared to diclofenac sodium inhibiting inflammation by 72% [184]. In an in vitro study, the chloroform extract of *A. muricata* leaves significantly inhibited activity of phospholipase A₂ [185]. With doses of 0.2–0.6 mg/mL, the enzyme activity was inhibited by 23.91%–43.48% [185]. In the same study, the chloroform extract of *A. muricata* leaves at 0.5 and 1.0 mg/mL also inhibited prostaglandin synthase activity by 87.46% and 82.92%, respectively, compared to the positive control indomethacin at 1 mg/mL which reduced enzyme activity by 87.46% [185].

Extracts of *A. senegalensis* roots were assessed for anti-inflammatory activity through in vitro inhibition of protein denaturation, hyaluronidase and xanthine oxidase. The ethyl acetate fraction was found to have the greatest activity inhibiting protein denaturation (70.6%), hyaluronidase (72.2%) and xanthine oxidase (78.7%) at a concentration of 100 µg/mL [186]. The ethanolic extract of *A. muricata* leaves exhibited anti-inflammatory effects in the carrageenan-induced rat paw acute edema model. Paw edema was reduced after orally administered doses of 200 and 400 mg/kg with (23.16% and 29.33%) and (29.50% and 37.33%), respectively, after 60 and 90 min of treatment [187]. Additionally, *A. muricata* fruit has been shown to exert an anti-inflammatory effect in a xylene-induced ear edema test [188]. Additional information regards pharmacological activities of *Annona* species, see (Table 7).

The lyophilized fruit extract at 50 mg/kg and 100 mg/kg inhibited the xylene-induced ear edema by 82.35% and 76.47%, respectively, compared to prednisolone, which reduced ear edema by 47.06% [188]. At intraperitoneal doses of 25, 50 and 100 mg/kg, the ethanolic extract of *A. vepretorum* leaves, inhibited carrageenan-induced leukocyte migration to the

peritoneal cavity by 62%, 76% and 98%, compared to dexamethasone (2 mg/kg, i.p.). which reduced leukocyte migration by 89% [104]. The flavonoids, quercetin and kaempferol isolated from leaves of *A. dioica* exhibited potent dose- and time-dependent anti-inflammatory activity in a carrageenan-induced paw oedema model with IC₅₀s of 8.53 and 10.57 µg/mL, respectively. The crude methanolic extract of *A. dioica* also reduced myeloperoxidase activity 6 h after the induction of paw oedema with a maximal inhibition of 51% at a dose of 300 mg/kg. [189]. Finally, hinesol, z-caryophyllene and beta-maaliene isolated from leaves of *A. sylvatica* also inhibited leukocyte migration at concentrations from 36.04 to 45.37 µg/mL in both carrageenan- and complete Freund's adjuvant-induced mouse paw edema [33].

6.5. Antioxidant Activity

An extract of *A. coriacea* seeds was investigated for its antioxidant activity using free radical 2,2-diphenyl-1-picrylhydrazil (DPPH) and bleaching of β-carotene, and a moderate antioxidant effect was reported of 31.53% in the DPPH test and 51.59% for the β-carotene bleaching test [190]. The pulp of *A. coriacea* fruit displayed a weaker antioxidant activity compared to the seeds, with 13.49% for the DPPH test and 32.32% for the β-carotene assay [190]. Additionally, various parts of *A. muricata*, including bark, leaves and stem, exhibited antioxidant activity using DPPH assay and the EC₅₀ value was recorded as 90 mg/g for bark, 290 mg/g for leaves, and 116 mg/g for the stem, compared to ascorbic acid with 157.5 mg/g [44]. Finally, an ethanolic extract of *A. squamosa* leaves was also reported as having antioxidant activity when evaluating DPPH, nitric oxide and superoxide radical assays. The activity was reported as 75.12%, 34.69% and 10.29%, respectively, at a concentration of 100 µg/mL [191].

6.6. Antileishmanial Activity

Many extracts and pure compounds from *Annona* plants have been tested against *Leishmania*, such as methanolic seed and leaf extracts of *A. squamosa*, for activity against *L. amazonensis*, with resulting showing IC₅₀s of 46.54 µg/mL and 28.32 µg/mL, respectively [192]. Alkaloids and acetogenins isolated from both leaves and seeds of *A. squamosa* were also reported for their activity against promastigote forms of *L. chagasi*, with the EC₅₀ value reported as 23.3 µg/mL for alkaloids and from 25.9–37.6 µg/mL for acetogenins [192]. Alkaloids like liriodenin isolated from the leaves of *A. mucosa* exhibited antileishmanial activity against promastigote forms of *L. braziliensis*, *L. guyanensis* and *L. amazonensis* with IC₅₀s of 55.92 µg/mL, 0.84 µg/mL 1.43 µg/mL respectively [193]. Finally, O-methylarmepavine isolated from leaves *A. squamosa* displayed antileishmanial activity against both promastigote and amastigote forms of *L. chagasi* with EC₅₀s of 23.3 µg/mL and 25.3 µg/mL, respectively [45].

6.7. Antiviral Activity

The antiviral activity of various *Annona* species was reported in several studies using either whole extract or pure compounds. For instance, 16β,17-dihydroxy-ent-kauran-19-oic acid was isolated from the fruits of *A. squamosa* and showed significant activity against human immunodeficiency virus (HIV) replication using H9 lymphocyte cell assay with EC₅₀ value of 0.8 µg/mL [194]. The ethanolic extract of *A. squamosa* seeds at 0.15 µg/mL, 0.25 µg/mL and 0.35 µg/mL also exhibited dose-dependent antiviral activity against the Avian influenza virus with the percentage of antiviral activity at 33.33%, 43.06% and 59.72%, respectively [195]. The leaves of *A. squamosa* extract were also tested against dengue virus type-2 (DENV-2) in Vero cells using Viral ToxGlo™ assay. At a concentration of 6.25 µg/mL, DENV-2 replication was reduced with IC₅₀ 73.78 µg/mL in Vero cells [196]. The methanolic extracts from the peels of *A. squamosa* and *A. reticulata* demonstrated antiviral activity against human immunodeficiency virus 1 (HIV-1) using a non-radioactive immune/colorimetric assay. Both *A. squamosa* and *A. reticulata* revealed high antiviral activity by inhibition of HIV-1 reverse transcriptase with values of 96.45% and 78.63% [197].

Moreover, *A. cherimola* was also evaluated for its antiviral activity against herpes simplex virus type 2 (HSV-2) using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) assay. The leaf extract inhibited HSV-2 replication and showed antiherpetic activity with a therapeutic index 8.40 [198]. Finally, the ethanolic stem extraction of *A. muricata* demonstrated antiviral activity against herpes simplex virus type 1 (HSV-1) with a minimum inhibitory concentration (MIC) of 1 mg/mL [199].

7. Pharmacological Activity of Isolated Compounds from *Annona* Species and their Mechanism of Action

The in vitro and in vivo biological activity of compounds that have been isolated from various parts of *Annona* plants will be discussed. Squamins C–F were isolated from the seeds of *A. globifera* and tested in vitro against trophozoites of *Acanthamoeba* spp. strains such as *A. castellanii* Neff, *A. polyphaga*, *A. griffin* and *A. quina* (Table 8) [200]. All tested compounds exhibited antiamoeboid activity against the strains by inducing programmed cell death [200]. The same compounds were also tested for their cytotoxicity against murine macrophage cell line J774A.1 (ATCCTIB-67) and showed no cytotoxicity effect with CC₅₀ values greater than 200 μ M [200].

Table 8. Antiamoebic activity of squamins C–F versus *Acanthamoeba* spp. Strains.

| Compounds | <i>A. castellanii</i> Neff IC ₅₀ (μ M) | <i>A. polyphaga</i> IC ₅₀ (μ M) | <i>A. griffin</i> IC ₅₀ (μ M) | <i>A. quina</i> IC ₅₀ (μ M) |
|-----------|--|---|---|---|
| Squamin C | 20.77 \pm 3.48 | 71.78 \pm 0.41 | 38.81 \pm 7.34 | 24.28 \pm 0.64 |
| Squamin D | 18.38 \pm 1.14 | 71.57 \pm 0.14 | 39.53 \pm 5.90 | 26.52 \pm 0.87 |
| Squamin E | 21.00 \pm 0.86 | 62.19 \pm 15.52 | 44.75 \pm 2.06 | 25.82 \pm 0.99 |
| Squamin F | 18.02 \pm 3.28 | 64.08 \pm 12.42 | 50.49 \pm 6.92 | 30.32 \pm 0.27 |

Rollinacin and rolliniastatin-1 isolated from the seed of *A. mucosa* were also reported for their larvicidal effect against *Aedes aegypti* and *Aedes albopictus* larvae [201]. Rolliniastatin-1 exhibited the best larvicidal effect against both *Aedes aegypti* and *Aedes albopictus* with LC₅₀s of 0.43 and 0.20 μ g/mL^{−1}, respectively. Rollinacin displayed similar activity against *Aedes aegypti* and *Aedes albopictus* with LC₅₀s of 0.78 μ g/mL and 1.128 μ g/mL, respectively [201]. However, the larvicidal mechanism action of these compounds was not reported. Annonacin isolated from the seed of *A. muricata* was evaluated for its larvicidal activity on *Aedes aegypti* and *Aedes albopictus* larvae [202]. The greater larvicidal activity was reported against *Aedes aegypti* with a LC₅₀ of 2.65 μ g/mL compared to *Aedes albopictus* with LC₅₀ of 8.34 μ g/mL. The mechanism of action was reported as being inhibition of their metabolic enzymes, particularly proteases and amylases that are important for the development of *Aedes* spp. larvae [202]. Twelve acetogenins isolated from the seed of *A. cornifolia* were tested for their antioxidant activity against DPPH [90]. These acetogenins were identified as 9-hydroxyfolianin, 4-desoxylongimicin, squamocin M, squamocin L, folianin A, folianin B, annofolin, isolongimicina B, bullatacin, asimicin, cornifolin and anotacin, and showed a strong DPPH radical scavenging with IC₅₀s ranging from 0.99 \pm 0.18 to 1.95 \pm 0.34 μ g/mL compared to ascorbic acid with an IC₅₀ 1.62 \pm 0.35 μ g/mL [90]. It has been suggested that the antiradical activity of acetogenins may be related to the α,β -unsaturated lactone ring moiety, which is also present in ascorbic acid [90]. Furthermore, the antioxidant activity of pure compounds from the bark of *A. salzmanni* after isolation of five alkaloids identified as liriodenine, anonaine, asimilobine, reticuline and cleistopholine [203]. The antioxidant activity was assessed through the Oxygen Radical Absorbance Capacity (ORAC) assay and asimilobine was found to be the most active alkaloid with ORAC value of 2.09 [203]. The rest of the compounds exhibited antioxidant activity with ORAC values ranging from 0.25 to 0.85 [203]. These same compounds were also examined for their antimicrobial activity against *Kocuria rhizophila*, *Staphylococcus aureus*, *Staphylococcus epidermidis* and *Enterococcus faecalis* with MIC values from 25 to 100 μ g/mL [203]. In an in vitro study, the antimicrobial activity of isolated alkaloids from aerial parts of *A. senegalensis* was assessed in a microdilution assay [154].

These alkaloids were identified as anonaine and asimilobine, and demonstrated against *Streptococcus mutans* with MIC values of 0.12 and 0.25 mg/mL, respectively [154]. For trypanocidal activity, three alkaloids liriodenine, annomontine, and O-methylmoschatoline were isolated from the branch of *A. foetida* and tested against both epimastigote and trypomastigote forms of *Trypanosoma cruzi* [101]. A potent trypanocidal effect was demonstrated against epimastigote forms with IC₅₀s ranging from 92.0 ± 18.4 to 198.0 ± 4.2 µg/mL, and from 3.8 ± 1.8 to 4.2 ± 1.9 µg/mL for trypomastigote forms [101]. Additionally, N-hydroxyannomontine isolated from the bark of *A. foetida*, demonstrated antileishmanial activity versus *Leishmania. braziliensis* and *L. guyanensis* with IC₅₀ values of 252.7 ± 2.2 and 437.5 ± 2.5 µM, respectively [204].

Many compounds isolated from various *Annona* species have demonstrated cytotoxicity against different cancer cell lines (Table 9). Three alkaloids were isolated from leaves of *A. crassiflora* identified as crassiflorine, xylopine and stephalagine and tested for their activity against colon carcinoma cells (HCT-116) using MTT assay [204]. The cytotoxicity activity for the tested compounds was reported with IC₅₀ values of 143.4 µM, 30.2 µM and 48.5 µM, respectively [204]. Muricin J–K isolated from the fruit of *A. muricata* exhibited anticancer activity against human prostate cancer cell lines (PC-3) through inhibition of the mitochondrial complex I in an in vitro study [205]. The anticancer activity was also reported for the alkaloid coclaurin isolated from aerial parts of *A. squamosa*. Cytotoxicity studies against human breast cancer cells (MCF-7), human colon cancer cells (HCT116) and human liver cancer cells (HEPG-2) reported IC₅₀ values of 15.345 µg/mL, 8.233 µg/mL and 1.674 µg/mL, respectively [206]. Bullatacin isolated from *A. cherimola* demonstrated inhibition of tumor growth at a dose of 15 µg/kg in mice bearing HepS and S180 xenografts and tumor growth was reduced by 63.4% and 65.8%, respectively [207]. In the same study, annonacin administrated orally (10 mg/kg) to hybrid mice (BDF-1) models significantly reduced lung cancer by 57.9% [207]. However, the mechanism of action of these acetogenins was not described in this study. Finally, stephalagin, an alkaloid isolated from the peel of *A. crassiflora* fruit, was reported as pancreatic lipase inhibitor with an IC₅₀ of 8.35 µg/mL^{−1} in vitro study [208].

Table 9. Anticancer activity of isolated compounds from *Annona* species and their mode of action.

| <i>Annona</i> Species | Plant Part | Isolated Components | Cell Line or Animal Model | Mechanism of Action | References |
|-----------------------|---------------|---|--|--|------------|
| <i>A. cherimola</i> | Seeds | Annomolin and Annocherimolin | Prostate tumor cell line (PC-3), breast (MCF-7) and colon (HT-29) cancer cell lines | Exhibited potent cytotoxicity | [209] |
| | Leaves | Asimilobine | Acute myeloid leukemia cell line | Upregulation of Bax, downregulation of Bcl2, and cleavage of PARP | [210] |
| <i>A. crassiflora</i> | Crude extract | Catechin | Cervical cancer cell | Apoptosis via intrinsic pathway | [211] |
| <i>A. glabra</i> | Fruits | Annoglabasin H | Lung adenocarcinoma cell line (LU-1), human breast carcinoma (MCF-7), human melanoma (SK-Mel2) | Exhibited significant cytotoxic activity | [212] |
| | | Annoglabayin | Human liver cancer cell line (Hep G2) | Apoptosis via mitochondrial pathway | [132] |
| | Leaves | Cunabic acid and ent-kauran-19-al-17-oic acid | Liver cancer (HLC) cell line SMMC-7721 | Apoptosis via down-regulation of BCL-2 gene and upregulation of bax gene | [213] |
| | | Asinicin | Human monocytic leukemia cells (CRL-12253) | Mitochondria mediated anticancer and antiproliferative effects | [214] |
| | | Annoglacin A and B | Human breast carcinoma (MCF-7) and Pancreatic carcinoma (PACA-2) cell lines | Suppressed proliferation | [215] |
| | | Icariside D2 | Human leukemia cell line (HL-60) | Induced apoptosis and decreased phosphorylation of AKT in cells | [216] |

Table 9. Cont.

| <i>Annona</i> Species | Plant Part | Isolated Components | Cell Line or Animal Model | Mechanism of Action | References |
|-------------------------------|------------|--|--|--|------------|
| <i>A. muricata</i> | Leaves | Annomuricin | Breast cancer cell | Suppressed breast cancer proliferation and induced apoptosis | [217] |
| | | Muricoreacin, Murihexocin | Colon cancer cell (HT-29, HCT-116) | Up-regulation of Bax, downregulation of Bcl-2 proteins and activated initiator and executioner caspases | [218] |
| | | Annomuricine, Muricapentocin | Pancreatic carcinoma (PACA-2) and colon adenocarcinoma (HT-29) cell | Exhibited repressive effect | [219] |
| | | Muricatocins A and B | Lung tumor cell line (A-549) | Enhanced cytotoxic activity | [146] |
| | Fruits | Muricin M and Muricin N | Prostate cancer (PC-3) cells | Exhibited cytotoxicity | [220] |
| <i>A. purpurea</i> | Roots | Annopurpuricins A–D | HeLa and HepG2 cells | Mitochondrial membrane depolarization and apoptosis | [221] |
| <i>A. reticulata</i> | Fruits | Catechin | Breast cancer cell line (MCF-7) | Inhibition via apoptosis | [222] |
| | Seeds | Annonacin | T24 bladder cancer cells | Bax expression was induced, caspase-3 activity enhanced and caused apoptosis | [172] |
| | | Bullatacin | Leukemia cell line (K562) and breast cancer cell line (MCF-7) | Cell death via apoptosis | [223] |
| | Leaves | Annomonicin | Colon cancer (HCT15), human lung cancer (Hop65) and human hepatoma (HEPG2) cell lines | Exhibited cytotoxic effect | [224] |
| | | Rolliniastatin | Breast cancer cell (T-47D) | Caspases dependent apoptosis | [225] |
| <i>A. senegalensis</i> | Leaves | (–) Roemerine | Breast cancer MDA-MB-231 cells | Exhibited dose-dependent cytotoxicity via targeting the ribosomal protein eL42 and arresting the crosslinking reaction with tRNAox | [226] |
| | Bark | Kaurenoic acid | Pancreatic tumor (PANC-1) cell lines and Henrietta Lacks' cervical cancer cell line (HeLa) | Exhibited significant cytotoxic activity | [227] |
| | Stem | Ent-kaurenoids | Breast cancer (MCF-7) cells, prostate cancer (PC-3) cells | Exhibited significant cytotoxic activity | [228] |
| <i>A. squamosa</i> | Leaves | Annorecticuin | Breast cancer cell (MCF-7) | Induced Apoptosis | [229] |
| | Seeds | Dieporeticenin B, Squamocin, Annosquatin III | Nasopharyngeal cancer (KB) cells, breast cancer (MCF-7) cells | Exerted inhibitory activity | [230] |
| | | Asimilobine | Human colon cancer cell (WiDr) | Increased expression of caspase-3 | [231] |
| | | Annosquatin A, B | Human leukemia cell line (K-562), human colon carcinoma (COLO-205) | Reduced intracellular glutathione levels and regulation of Bcl-2 and PS externalization | [232] |
| | | Annosquacins A–D, Annosquatin A, B | Human breast cancer cell line (MCF-7), human lung adenocarcinoma cell line (A-549) | Exhibited cytotoxic activity | [233] |
| | | (–)-Anonaine | H22 solid tumor cell | Inhibition of IL-6/Jak/Stat3 pathway | [234] |
| | Bark | Coclaurine | DMBA painted hamsters | Enhanced lipid peroxidation | [235] |
| | Fruits | (–)-Ent-kaur-16-en-19 oic acid, 16 α ,17 dihydroxy-ent-kauran-19-oic acid | Dalton's lymphoma cells, HeLa cells | Exhibited cytotoxic activity | [236] |
| <i>A. sylvatica</i> A.St.-Hil | Leaves | Quercetin Kaempferol | Anti-inflammatory | Leukocytes migration was significantly reduced at IC ₅₀ 8.53 and 10.57 $\mu\text{g/mL}$, respectively. | [189] |
| <i>A. vepretorum</i> Mart. | Leaves | Bicyclogermacrene | Antimicrobial | Against <i>Candida tropicalis</i> with a MIC value of 100 $\mu\text{g} \cdot \text{mL}^{-1}$. | [237] |

8. Toxicity and Interactions

Generally, the safety of natural medicines can be assessed according to their effects and drug-drug interactions. An epidemiological study has reported that consumption of fruits of Annonaceae led to the prevalence of atypical parkinsonism in Guadeloupe due to the presence of acetogenins in the plant fruits [238]. According to Champy et al. 2005, the amount of annonacin per a single fruit is approximately 15 mg, and an adult who consumes a daily intake of one fruit for one year is equivalent to the amount of annonacin injected into rats, which induced the brain lesions [239]. It has been suggested that the toxicity might be related to the capacity of the tetrahydrofuran ring to chelate calcium ions [35]. Moreover, the fruit of *A. squamosa* has been analysed for its quantity of squamocin using HPLC-MS and reported 13.5–36.4 mg of squamocin for each fruit, and that long-term consumption of *A. squamosa* fruit may be a risk factor in the development of neurodegenerative disorders [240]. Additionally, the use of a dietary supplement sold in the USA containing an extract of *A. muricata* has been found to exhibit neurotoxic effects in human neuron cultures [241].

The interactions of *Annona* species with other drugs have been reported in other studies; for instance, administration of capsules of *A. muricata* leaves in combination with glibenclamide resulted in improved glycaemic control compared to patients who received only glibenclamide [242]. An additional study reported that a combination of aqueous custard apple leaf extract and glipizide enabled a decrease in the dose of glipizide by up to half in rats with type-2 diabetes and reduced the risk of requiring insulin therapy [243]. These outcomes suggest the potential use of certain *Annona* species in conjunction with antidiabetic medications to maximize the efficacy of a lower therapeutic dose.

9. Conclusions

This review provides a comprehensive summary of the botanical features, ethnomedical uses, pharmacology and phytochemistry of the main species of Annonaceae family and, in particular, the *Annona* genera used in traditional medicine practices. Of the many members of the Annonaceae family, the *Annona* species is heavily used in traditional medicines across the world. Among the 30 reviewed *Annona* species, six species *A. squamosa*, *A. muricata*, *A. cherimola*, *A. senegalensis*, *A. reticulata* and *A. coriacea* are the most widely studied for their pharmacological activities and phytochemical profiles of their bark, leaves, fruits and seeds. Various pharmacological properties have been reported, including antidiabetic, hepatoprotective, anti-inflammatory, antiprotazoal, antitumor, antioxidant, antimicrobial and anticonvulsant activity.

With regard to the phytochemistry of *Annona* species, the main classes of constituents identified to date are acetogenins, alkaloids, phenols and essential oils. The alkaloids are mainly present in the leaves, whereas acetogenins are present in the seeds and found in smaller quantities in the pulp and leaves of *Annona* species. The chemical profiles of the acetogenins present in different species have been extensively studied and their anticancer activity investigated, with low concentrations exhibiting chemotoxicity against several cancer cell lines. These preclinical results, along with the reported case studies, suggest that further clinical studies evaluating the role of acetogenins in the treatment of various types of cancers are warranted. Importantly, formulations, including the parts of the *Annona* species used, agricultural practices and the extraction methods vary considerably, leading to likely variations in the phytochemical and pharmacological profiles. In this respect, further characterization of standardized formulations of *Annona* species is required to predict likely clinical effects. Additional interesting results on the antidiabetic effects of fruits from *Annona* species also warrant further investigation as nutraceuticals to assist in the therapy of diabetes.

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References

- Lúcio, A.S.S.C.; da Silva Almeida, J.R.G.; Da-Cunha, E.V.L.; Tavares, J.F.; Barbosa Filho, J.M. Alkaloids of the Annonaceae: Occurrence and a compilation of their biological activities. *Alkaloids: Chem. Biol.* **2015**, *74*, 233–409.
- Rabelo, S.V.; Quintans, J.d.S.S.; Costa, E.V.; da Silva Almeida, J.R.G.; Júnior, L.J.Q. *Annona Species (Annonaceae) Oils*; Academic Press: Cambridge, MA, USA, 2015; pp. 221–229.
- Leboeuf, M.; Cavé, A.; Bhaumik, P.; Mukherjee, B.; Mukherjee, R. The phytochemistry of the Annonaceae. *Phytochemistry* **1980**, *21*, 2783–2813. [[CrossRef](#)]
- Biba, V.; Amily, A.; Sangeetha, S.; Remani, P. Anticancer, antioxidant and antimicrobial activity of Annonaceae family. *World J. Pharm. Pharm. Sci.* **2014**, *3*, 1595–1604.
- Fournier, G.; Leboeuf, M.; Cavé, A. Annonaceae essential oils: A review. *J. Essent. Oil Res.* **1999**, *11*, 131–142. [[CrossRef](#)]
- Pinto, A.D.Q.; Cordeiro, M.C.R.; De Andrade, S.; Ferreira, F.R.; Filgueiras, H.d.C.; Alves, R.; Kinpara, D.I. *Annona Species*; University of Southampton, International Centre for Underutilised Crops: Southampton, UK, 2005.
- Van Heusden, E. Flowers of annonaceae. *Blumea Suppl.* **1992**, *7*, 1–218.
- Couvreur, T.L.; Maas, P.J.; Meinke, S.; Johnson, D.M.; Kessler, P.J. Keys to the genera of Annonaceae. *Bot. J. Linn. Soc.* **2012**, *169*, 74–83. [[CrossRef](#)]
- Kessler, P. Annonaceae. In *Flowering Plants: Dicotyledons*; Springer: Berlin/Heidelberg, Germany, 1993; pp. 93–129.
- Quílez, A.; Fernández-Arche, M.; García-Giménez, M.; De la Puerta, R. Potential therapeutic applications of the genus *Annona*: Local and traditional uses and pharmacology. *J. Ethnopharmacol.* **2018**, *225*, 244–270. [[CrossRef](#)]
- Attiq, A.; Jalil, J.; Husain, K. Annonaceae: Breaking the wall of inflammation. *Front. Pharmacol.* **2017**, *8*, 752. [[CrossRef](#)]
- Bele, M.Y.; Focho, D.A.; Egbe, E.A.; Chuyong, B.G. Ethnobotanical survey of the uses Annonaceae around mount Cameroon. *Afr. J. Plant Sci.* **2011**, *5*, 237–247.
- Tsabang, N.; Fokou, P.V.T.; Tchokouaha, L.R.Y.; Noguem, B.; Bakarnga-Via, I.; Nguépi, M.S.D.; Nkongmeneck, B.A.; Boyom, F.F. Ethnopharmacological survey of Annonaceae medicinal plants used to treat malaria in four areas of Cameroon. *J. Ethnopharmacol.* **2012**, *139*, 171–180. [[CrossRef](#)]
- Johnson, T.A.; Sohn, J.; Ward, A.E.; Cohen, T.L.; Lorig-Roach, N.D.; Chen, H.; Pilli, R.A.; Widjaja, E.A.; Hanafi, M.; Kardono, L.B. (+)-Altholactone exhibits broad spectrum immune modulating activity by inhibiting the activation of pro-inflammatory cytokines in RAW 264.7 cell lines. *Bioorg. Med. Chem.* **2013**, *21*, 4358–4364. [[CrossRef](#)] [[PubMed](#)]
- Focho, D.; Egbe, E.; Chuyong, G.; Fongod, A.; Fonge, B.; Ndam, W.; Youssoufa, B. An ethnobotanical investigation of the annonaceae on Mount Cameroon. *J. Med. Plant Res.* **2010**, *4*, 2148–2158.
- Ayu, C.A.; Weta, I.W.; Aman, I.G.M. Moringa (*Moringa oleifera*) leaves extract gel improved wound healing by increasing fibroblasts, neovascularization and in male Wistar rats. *IJAAM* **2020**, *4*, 28–32.
- Nordin, N.; Salama, S.M.; Golbabapour, S.; Hajrezaie, M.; Hassandarvish, P.; Kamalidehghan, B.; Majid, N.A.; Hashim, N.M.; Omar, H.; Fadaienasab, M. Anti-ulcerogenic effect of methanolic extracts from *Enicosanthellum pulchrum* (King) Heusden against ethanol-induced acute gastric lesion in animal models. *PLoS ONE* **2014**, *9*, e111925.
- Grenand, F.; Grenand, P. La Côte d’Amapá: De la bouche de l’Amazone à la baie d’Oyapock, à travers la tradition orale palikur. *Bol. Do Mus. Para. Emilio Goeldi* **1987**, 1–77.
- Moukette, B.M.; Pieme, C.A.; Njimou, J.R.; Biapa, C.P.N.; Marco, B.; Ngogang, J.Y. In vitro antioxidant properties, free radicals scavenging activities of extracts and polyphenol composition of a non-timber forest product used as spice: *Monodora myristica*. *Biol. Res.* **2015**, *48*, 15. [[CrossRef](#)]
- Krishnamurthi, A. *The Wealth of India: Raw Materials*; CSIR: Delhi, India, 1969; Volume VIII. Ph-Re.
- Woode, E.; Ameyaw, E.O.; Boakye-Gyasi, E.; Abotsi, W.K. Analgesic effects of an ethanol extract of the fruits of *Xylopia aethiopica* (Dunal) A. Rich (Annonaceae) and the major constituent, xylopic acid in murine models. *J. Pharm. Bioallied Sci.* **2012**, *4*, 291–301. [[CrossRef](#)]
- Woguem, V.; Fogang, H.P.; Maggi, F.; Tapondjou, L.A.; Womeni, H.M.; Quassinti, L.; Bramucci, M.; Vitali, L.A.; Petrelli, D.; Lupidi, G. Volatile oil from striped African pepper (*Xylopia parviflora*, Annonaceae) possesses notable chemopreventive, anti-inflammatory and antimicrobial potential. *Food Chem.* **2014**, *149*, 183–189. [[CrossRef](#)]
- Nishiyama, Y.; Moriyasu, M.; Ichimaru, M.; Iwasa, K.; Kato, A.; Mathenge, S.G.; Mutiso, P.B.C.; Juma, F.D. Secondary and tertiary isoquinoline alkaloids from *Xylopia parviflora*. *Phytochemistry* **2006**, *67*, 2671–2675. [[CrossRef](#)]

24. Pornputtapitak, W. Chemical Constituents of the Branches of *Anomianthus Dulcis* and the Branches of *Dalbergia Cochinchinensis* Pierre. Ph.D. Thesis, Silpakorn University, Nakorn Pathom, Thailand, 2008.
25. Hopp, D.C.; Alali, F.Q.; Gu, Z.-m.; McLaughlin, J.L. Three new bioactive bis-adjacent THF-ring acetogenins from the bark of *Annona squamosa*. *Bioorg. Med. Chem.* **1998**, *6*, 569–575. [\[CrossRef\]](#)
26. Wong, H.-F.; Brown, G.D. β -Methoxy- γ -methylene- α , β -unsaturated- γ -butyrolactones from *Artabotrys hexapetalus*. *Phytochemistry* **2002**, *59*, 99–104. [\[CrossRef\]](#)
27. Alali, F.Q.; Rogers, L.; Zhang, Y.; McLaughlin, J.L. Unusual bioactive annonaceous acetogenins from *Goniothalamus giganteus*. *Tetrahedron* **1998**, *54*, 5833–5844. [\[CrossRef\]](#)
28. Huong, D.; Luong, D.; Thao, T.; Sung, T. A new flavone and cytotoxic activity of flavonoid constituents isolated from *Miliusa balansae* (Annonaceae). *Pharmazie* **2005**, *60*, 627–629. [\[CrossRef\]](#) [\[PubMed\]](#)
29. Hsieh, T.-J.; Chang, F.-R.; Chia, Y.-C.; Chen, C.-Y.; Lin, H.-C.; Chiu, H.-F.; Wu, Y.-C. The alkaloids of *Artabotrys uncinatus*. *J. Nat. Prod.* **2001**, *64*, 1157–1161. [\[CrossRef\]](#)
30. Hsieh, T.-J.; Chang, F.-R.; Chia, Y.-C.; Chen, C.-Y.; Chiu, H.-F.; Wu, Y.-C. Cytotoxic constituents of the fruits of *Cananga odorata*. *J. Nat. Prod.* **2001**, *64*, 616–619. [\[CrossRef\]](#)
31. Soonthornchareonnon, N.; Suwanborirux, K.; Bavovada, R.; Patarapanich, C.; Cassady, J.M. New cytotoxic 1-azaanthraquinones and 3-aminonaphthoquinone from the stem bark of *Goniothalamus marcanii*. *J. Nat. Prod.* **1999**, *62*, 1390–1394. [\[CrossRef\]](#)
32. Costa, E.V.; Marques, F.d.A.; Pinheiro, M.L.B.; Braga, R.M.; Delarmelina, C.; Duarte, M.C.T.; Ruiz, A.L.T.; Carvalho, J.E.d.; Maia, B.H. Chemical constituents isolated from the bark of *Guatteria blepharophylla* (Annonaceae) and their antiproliferative and antimicrobial activities. *J. Braz. Chem. Soc.* **2011**, *22*, 1111–1117. [\[CrossRef\]](#)
33. Formagio, A.S.; Vieira, M.d.C.; Dos Santos, L.A.; Cardoso, C.A.; Foglio, M.A.; de Carvalho, J.E.; Andrade-Silva, M.; Kassuya, C.A. Composition and evaluation of the anti-inflammatory and anticancer activities of the essential oil from *Annona sylvatica* A. St.-Hil. *J. Med. Food* **2013**, *16*, 20–25. [\[CrossRef\]](#)
34. Oliveira, G.N.D.S.A.; Dutra, L.M.; Paz, W.H.P.; da Silva, F.M.A.; Costa, E.V.; da Silva Almeida, J.R.G. Chemical constituents from the leaves and branches of *Annona coriacea* Mart. (Annonaceae). *Biochem. Syst. Ecol.* **2021**, *97*, 104297.
35. Tundis, R.; Xiao, J.; Loizzo, M.R. *Annona* species (Annonaceae): A rich source of potential antitumor agents? *Ann. N. Y. Acad. Sci.* **2017**, *1398*, 30–36. [\[CrossRef\]](#)
36. Nugraha, A.S.; Damayanti, Y.D.; Wangchuk, P.; Keller, P.A. Anti-infective and anti-cancer properties of the *Annona* species: Their ethnomedicinal uses, alkaloid diversity, and pharmacological activities. *Molecules* **2019**, *24*, 4419. [\[CrossRef\]](#) [\[PubMed\]](#)
37. Takahashi, J.A.; Pereira, C.R.; Pimenta, L.P.; Boaventura, M.A.D.; Silva, L.G.E. Antibacterial activity of eight Brazilian Annonaceae plants. *Nat. Prod. Res.* **2006**, *20*, 21–26. [\[CrossRef\]](#) [\[PubMed\]](#)
38. Carballo, A.I.; Martínez, A.L.; González-Trujano, M.E.; Pellicer, F.; Ventura-Martínez, R.; Díaz-Reval, M.I.; López-Muñoz, F. Antinociceptive activity of *Annona diversifolia* Saff. leaf extracts and palmitone as a bioactive compound. *Pharmacol. Biochem. Behav.* **2010**, *95*, 6–12. [\[CrossRef\]](#) [\[PubMed\]](#)
39. Ajaiyeoba, E.; Falade, M.; Ogbale, O.; Okpako, L.; Akinboye, D. In vivo antimalarial and cytotoxic properties of *Annona senegalensis* extract. *J. Tradit. Complement. Med.* **2006**, *3*, 137–141. [\[CrossRef\]](#)
40. González-Trujano, M.E.; Tapia, E.; López-Meraz, L.; Navarrete, A.; Reyes-Ramírez, A.; Martínez, A. Anticonvulsant effect of *Annona diversifolia* Saff. and palmitone on penicillin-induced convulsive activity. A behavioral and EEG study in rats. *Epilepsia* **2006**, *47*, 1810–1817. [\[CrossRef\]](#)
41. Afroz, N.; Hoq, M.A.; Jahan, S.; Islam, M.M.; Ahmed, F.; Shahid-Ud-Daula, A.; Hasanuzzaman, M. Methanol soluble fraction of fruits of *Annona muricata* possesses significant antidiarrheal activities. *Heliyon* **2020**, *6*, e03112. [\[CrossRef\]](#)
42. Panda, S.; Kar, A. Antidiabetic and antioxidative effects of *Annona squamosa* leaves are possibly mediated through quercetin-3-O-glucoside. *Biofactors* **2007**, *31*, 201–210. [\[CrossRef\]](#)
43. Rocha, R.S.; Kassuya, C.A.L.; Formagio, A.S.N.; Mauro, M.d.O.; Andrade-Silva, M.; Monreal, A.C.D.; Cunha-Laura, A.L.; Vieira, M.d.C.; Oliveira, R.J. Analysis of the anti-inflammatory and chemopreventive potential and description of the antimutagenic mode of action of the *Annona crassiflora* methanolic extract. *Pharm. Biol.* **2016**, *54*, 35–47. [\[CrossRef\]](#)
44. Essama, S.R.; Nyegue, M.; Foe, C.N.; Silihe, K.K.; Tamo, S.B.; Etoa, F. Antibacterial and antioxidant activities of hydro-ethanol extracts of barks, leaves and stems of *Annona muricata*. *Am. J. Pharmacol. Sci.* **2015**, *3*, 126–131.
45. Brígido, H.P.C.; Correa-Barbosa, J.; da Silva-Silva, J.V.; Costa, E.V.S.; Percário, S.; Dolabela, M.F. Antileishmanial activity of *Annona* species (Annonaceae). *SN Appl. Sci.* **2020**, *2*, 1524. [\[CrossRef\]](#)
46. Castillo-Juárez, I.; González, V.; Jaime-Aguilar, H.; Martínez, G.; Linares, E.; Bye, R.; Romero, I. Anti-helicobacter pylori activity of plants used in Mexican traditional medicine for gastrointestinal disorders. *J. Ethnopharmacol.* **2009**, *122*, 402–405. [\[CrossRef\]](#) [\[PubMed\]](#)
47. Martínez-Vázquez, M.; Estrada-Reyes, R.; Escalona, A.A.; Velázquez, I.L.; Martínez-Mota, L.; Moreno, J.; Heinze, G. Antidepressant-like effects of an alkaloid extract of the aerial parts of *Annona cherimolia* in mice. *J. Ethnopharmacol.* **2012**, *139*, 164–170. [\[CrossRef\]](#) [\[PubMed\]](#)
48. Andrade-Cetto, A.; Heinrich, M. Mexican plants with hypoglycaemic effect used in the treatment of diabetes. *J. Ethnopharmacol.* **2005**, *99*, 325–348. [\[CrossRef\]](#) [\[PubMed\]](#)
49. de Lima, M.R.F.; de Souza Luna, J.; dos Santos, A.F.; de Andrade, M.C.C.; Sant’Ana, A.E.G.; Genet, J.-P.; Marquez, B.; Neuville, L.; Moreau, N. Anti-bacterial activity of some Brazilian medicinal plants. *J. Ethnopharmacol.* **2006**, *105*, 137–147. [\[CrossRef\]](#) [\[PubMed\]](#)

50. Pio-Correa, M. *Dicionário de Plantas úteis do Brasil e das Plantas Exóticas Cultivadas*; Embrapa-CPAC: Brasília, Brazil, 1974.
51. Luzia, D.M.; Jorge, N. Bioactive substance contents and antioxidant capacity of the lipid fraction of *Annona crassiflora* Mart. seeds. *Ind. Crops Prod.* **2013**, *42*, 231–235. [\[CrossRef\]](#)
52. González-Trujano, M.; Navarrete, A.; Reyes, B.; Hong, E. Some pharmacological effects of the ethanol extract of leaves of *Annona diversifolia* on the central nervous system in mice. *Phytother. Res.* **1998**, *12*, 600–602. [\[CrossRef\]](#)
53. González-Trujano, M.E.; Navarrete, A.; Reyes, B.; Cedillo-Portugal, E.; Hong, E. Anticonvulsant properties and bio-guided isolation of palmitone from leaves of *Annona diversifolia*. *Planta Med.* **2001**, *67*, 136–141. [\[CrossRef\]](#)
54. Jamkhande, P.G.; Wattamwar, A.S. *Annona reticulata* Linn. (Bullock's heart): Plant profile, phytochemistry and pharmacological properties. *J. Tradit. Complement. Med.* **2015**, *5*, 144–152. [\[CrossRef\]](#)
55. Mannino, G.; Gentile, C.; Porcu, A.; Agliassa, C.; Caradonna, F.; Berte, C.M. Chemical profile and biological activity of cherimoya (*Annona cherimola* Mill.) and atemoya (*Annona atemoya*) Leaves. *Molecules* **2020**, *25*, 2612. [\[CrossRef\]](#)
56. Rao, V.; Dasaradhan, P.; Krishnaiah, K. Antifertility effect of some indigenous plants. *Indian J. Med. Res.* **1979**, *70*, 517–520.
57. Calzada, F.; Correa-Basurto, J.; Barbosa, E.; Mendez-Luna, D.; Yépez-Mulia, L. Antiprotozoal constituents from *Annona cherimola* miller, a plant used in Mexican traditional medicine for the treatment of diarrhea and dysentery. *Pharmacogn. Mag.* **2017**, *13*, 148. [\[PubMed\]](#)
58. Calzada, F.; Solares-Pascasio, J.I.; Ordoñez-Razo, R.; Velazquez, C.; Barbosa, E.; García-Hernández, N.; Mendez-Luna, D.; Correa-Basurto, J. Antihyperglycemic activity of the leaves from *Annona cherimola* miller and rutin on alloxan-induced diabetic rats. *Pharmacogn. Res.* **2017**, *9*, 1. [\[CrossRef\]](#) [\[PubMed\]](#)
59. Amoo, I.A.; Emenike, A.E.; Akpambang, V.O.E. Compositional evaluation of *Annona cherimoya* (custard apple) fruit. *Trends Appl. Sci. Res.* **2008**, *2*, 216–220.
60. Vasarri, M.; Barletta, E.; Vinci, S.; Ramazzotti, M.; Francesconi, A.; Manetti, F.; Degl'Innocenti, D. *Annona cherimola* Miller fruit as a promising candidate against diabetic complications: An in vitro study and preliminary clinical results. *Foods* **2020**, *9*, 1350. [\[CrossRef\]](#)
61. Kalidindi, N.; Thimmaiah, N.V.; Jagadeesh, N.V.; Nandee, R.; Swetha, S.; Kalidindi, B. Antifungal and antioxidant activities of organic and aqueous extracts of *Annona squamosa* Linn. leaves. *J. Food Drug Anal.* **2015**, *23*, 795–802. [\[CrossRef\]](#)
62. Sundaramahalingam, M.; Karthikumar, S.; Kumar, R.S.; Samuel, K.J.; Shajahan, S.; Sivasubramanian, V.; Sivashanmugam, P.; Varalakshmi, P.; Syed, A.; Marraiki, N. An intensified approach for transesterification of biodiesel from *Annona squamosa* seed oil using ultrasound-assisted homogeneous catalysis reaction and its process optimization. *Fuel* **2021**, *291*, 120195. [\[CrossRef\]](#)
63. Atique, A.; Iqbal, M.; Ghouse, A. Use of *Annona squamosa* and *Piper nigrum* against diabetes. *Fitoterapia* **1985**, *56*, 190–192.
64. Ahmed, R.H.A.; Mariod, A.A. *Annona squamosa*: Phytochemical Constituents, Bioactive Compounds, Traditional and Medicinal Uses. In *Wild Fruits: Composition, Nutritional Value and Products*; Springer: Berlin/Heidelberg, Germany, 2019; pp. 143–155.
65. Benavides González, A. Caracterización numérica de germoplasma de guanábana (*Annona muricata* L.) muestreado in situ en el Pacífico y Norte de Nicaragua. *Rev. La Calera* **2011**, *4*, 29–35.
66. Badrie, N.; Schauss, A.G. Soursop (*Annona muricata* L.): Composition, Nutritional Value, Medicinal Uses, and Toxicology. In *Bioactive Foods in Promoting Health*; Elsevier: Amsterdam, The Netherlands, 2010; pp. 621–643.
67. Adjanohoun, J.; Aboubakar, N.; Dramane, K.; Ebot, M.; Ekpere, J.; Enow-Orock, E.; Focho, D.; Gbile, Z.; Kamanyi, A.; Kamsu-Kom, J. *Traditional Medicine and Pharmacopoeia: Contribution to Ethnobotanical and Floristic Studies in Cameroon*; Organization of African Unity Scientific, Technical and Research Commission; Technical Research Commission Centre National de Production de Manuels Scolaires: Porto-Novo, Benin, 1996; p. 133.
68. Zaman, K. Pharmacognostical and Phytochemical Studies on the Leaf and Stem Bark of *Annona reticulata* Linn. *J. Pharmacogn. Phytochem.* **2013**, *1*, 1–7.
69. Nirmal, S.; Gaikwad, S.; Dhasade, V.; Dhikale, R.; Kotkar, P.; Dighe, S. Anthelmintic activity of *Annona reticulata* leaves. *Res. J. Pharm. Biol. Chem.* **2010**, *1*, 115–118.
70. Camilo, C.J.; Caldas, R.P.; Damasceno, S.S.; Lopes, C.M.; Almeida, S.C.; da Costa, J.G.; Rodrigues, F.F. Chemical research and evaluation of the antioxidant activity of the flowers of *Annona coriacea* Mart. (Annonaceae). *Curr. Bioact. Compd.* **2017**, *13*, 66–70. [\[CrossRef\]](#)
71. Leite, D.O.; de FA Nonato, C.; Camilo, C.J.; de Carvalho, N.K.; da Nobrega, M.G.; Pereira, R.C.; da Costa, J.G. *Annona* genus: Traditional uses, phytochemistry and biological activities. *Curr. Pharm. Des.* **2020**, *26*, 4056–4091. [\[CrossRef\]](#) [\[PubMed\]](#)
72. Paulino-Neto, H.F. Polinização e biologia reprodutiva de Araticum-liso (*Annona coriacea* Mart.: Annonaceae) em uma área de cerrado paulista: Implicações para fruticultura. *Rev. Bras. Frutic* **2014**, *36*, 132–140. [\[CrossRef\]](#)
73. Maas, P.J.; de Kamer, H.M.-v.; Junikka, L.; de Mello-Silva, R.; Rainer, H. Annonaceae from central-eastern Brazil. *Rodriguésia* **2001**, *52*, 65–98. [\[CrossRef\]](#)
74. Monteiro, Á.B.; de Souza Rodrigues, C.K.; do Nascimento, E.P.; dos Santos Sales, V.; de Araújo Delmondes, G.; da Costa, M.H.N.; de Oliveira, V.A.P.; de Moraes, L.P.; Boligon, A.A.; Barbosa, R. Anxiolytic and antidepressant-like effects of *Annona coriacea* (Mart.) and caffeic acid in mice. *Food Chem. Toxicol.* **2020**, *136*, 111049. [\[CrossRef\]](#)
75. Souza, R.K.D.; da Silva, M.A.P.; de Menezes, I.R.A.; Ribeiro, D.A.; Bezerra, L.R.; de Almeida Souza, M.M. Ethnopharmacology of medicinal plants of carrasco, northeastern Brazil. *J. Ethnopharmacol.* **2014**, *157*, 99–104. [\[CrossRef\]](#)
76. Yisa, J.; Egila, J.; Darlinton, A. Chemical composition of *Annona senegalensis* from Nupe land, Nigeria. *Afr. J. Biotechnol.* **2010**, *9*, 4106–4109.

77. Orwa, C.; Mutua, A.; Kindt, R.; Jamnadass, R.; Simons, A. *Agroforestry Database: A Tree Reference and Selection Guide. Version 4*; World Agroforestry Centre: Quénia, Nairobi, 2009.
78. Mustapha, A.A. *Annona senegalensis* Persoon: A multipurpose shrub, its phytotherapeutic, phytopharmacological and phytomedicinal uses. *Int. J. Sci. Technol.* **2013**, *2*, 862–865.
79. Dambatta, S.H.; Aliyu, B. A survey of major ethno medicinal plants of Kano north, Nigeria, their knowledge and uses by traditional healers. *Bayero J. Pure Appl. Sci* **2011**, *4*, 28–34. [\[CrossRef\]](#)
80. Igoli, J.; Ogaji, O.; Tor-Ayiin, T.; Igoli, N. Traditional medicine practice amongst the Iggede people of Nigeria. Part II. *Afr. J. Tradit. Complement. Altern. Med.* **2005**, *2*, 134–152. [\[CrossRef\]](#)
81. Dutra, L.M.; Bomfim, L.M.; Rocha, S.L.; Nepel, A.; Soares, M.B.; Barison, A.; Costa, E.V.; Bezerra, D.P. ent-Kaurane diterpenes from the stem bark of *Annona vepretorum* (Annonaceae) and cytotoxic evaluation. *Bioorg. Med. Chem. Lett.* **2014**, *24*, 3315–3320. [\[CrossRef\]](#) [\[PubMed\]](#)
82. Souza, V.C.; Lorenzi, H. *Botânica Sistemática: Guia Ilustrado Para Identificação das Famílias de Angiospermas da Flora Brasileira, Baseado em APG II*; Nova Odessa Instituto Plantarum: Nova Odessa, Brazil, 2005; p. 640.
83. Pontes, A.F.; Barbosa, M.R.d.V.; Maas, P.J. Flora Paraibana: Annonaceae Juss. *Acta Bot. Brasilica* **2004**, *18*, 281–293. [\[CrossRef\]](#)
84. Corrêa, M.P. Dicionário das plantas úteis do Brasil e das exóticas cultivadas. In *Dicionário das Plantas Úteis do Brasil e Das Exóticas Cultivadas*; Smithsonian Libraries and Archives: Washington, DC, USA, 1984; p. 687.
85. Almeida, S.d.; Proença, C.E.; Sano, S.M.; Ribeiro, J.F. Cerrado: Espécies Vegetais Úteis. *Planaltina Embrapa-CPAC* **1998**, 464.
86. Lorenzi, H. *Arvores Brasileiras manual de Identificação e Cultivo de Plantas Arbóreas do Brasil*; Instituto Plantarum de Estudos da Flora: Nova Odessa, Brazil, 1998.
87. Vanhove, W. *Descriptors for Cherimoya (Annona Cherimola Mill.)*; Bioversity International: Rome, Italy, 2008.
88. Scheldeman, X. Distribution and Potential of Cherimoya (*Annona cherimola* Mill.) and Highland Papayas (*Vasconcellea* spp.) in Ecuador. Ph.D. Thesis, Ghent University, Ghent, Belgium, 2002.
89. Silva, M.V.; Costa, T.R.; Costa, M.R.; Ferreira, E.C.; Fernandes, O.F.; Santos, S.C.; Lião, L.M.; Ferri, P.H.; Paula, J.R.; Ferreira, H.D. Growth inhibition effect of Brazilian cerrado plant extracts on *Candida* species. *Pharm. Biol.* **2001**, *39*, 138–141. [\[CrossRef\]](#)
90. Lima, L.A.S.; Pimenta, L.P.; Boaventura, M.A.D. Acetogenins from *Annona cornifolia* and their antioxidant capacity. *Food Chem.* **2010**, *122*, 1129–1138. [\[CrossRef\]](#)
91. Vilar, J.B.; Ferreira, F.L.; Ferri, P.H.; Guillo, L.A.; Chen Chen, L. Assessment of the mutagenic, antimutagenic and cytotoxic activities of ethanolic extract of araticum (*Annona crassiflora* Mart. 1841) by micronucleus test in mice. *Braz. J. Biol.* **2008**, *68*, 141–147. [\[CrossRef\]](#) [\[PubMed\]](#)
92. Rodrigues, A.M.D.S.; De Paula, J.E.; Dégallier, N.; Molez, J.-F.; Espindola, L.S. Larvicidal activity of some Cerrado plant extracts against *Aedes aegypti*. *J. Am. Mosq. Control Assoc.* **2006**, *22*, 314–317. [\[CrossRef\]](#)
93. Khallouki, F.; Haubner, R.; Ulrich, C.M.; Owen, R.W. Ethnobotanical survey, chemical composition, and antioxidant capacity of methanolic extract of the root bark of *Annona cuneata* Oliv. *J. Med. Food* **2011**, *14*, 1397–1402. [\[CrossRef\]](#)
94. King, S.R. Medicinal plants of Brazil. *Phytomedicine* **2001**, *8*, 495.
95. Padmaja, V.; Thankamany, V.; Hara, N.; Fujimoto, Y.; Hisham, A. Biological activities of *Annona glabra*. *J. Ethnopharmacol.* **1995**, *48*, 21–24. [\[CrossRef\]](#)
96. Waechter, A.I.; Yaluff, G.; Inchausti, A.; de Arias, A.R.; Hocquemiller, R.; Cavé, A.; Fournet, A. Leishmanicidal and trypanocidal activities of acetogenins isolated from *Annona glauca*. *Phytother. Res.* **1998**, *12*, 541–544. [\[CrossRef\]](#)
97. Waechter, A.I.; Ferreira, M.E.; Fournet, A.; de Arias, A.R.; Nakayama, H.; Torres, S.; Hocquemiller, R.; Cavé, A. Experimental treatment of cutaneous leishmaniasis with argentiactone isolated from *Annona haematantha*. *Planta Med.* **1997**, *63*, 433–435. [\[CrossRef\]](#) [\[PubMed\]](#)
98. Chuang, P.-H.; Hsieh, P.-W.; Yang, Y.-L.; Hua, K.-F.; Chang, F.-R.; Shiea, J.; Wu, S.-H.; Wu, Y.-C. Cyclopeptides with anti-inflammatory activity from seeds of *Annona montana*. *J. Nat. Prod.* **2008**, *71*, 1365–1370. [\[CrossRef\]](#) [\[PubMed\]](#)
99. Barbalho, S.M.; Soares de Souza, M.d.S.; Bueno, P.C.d.S.; Guiguer, E.L.; Farinazzi-Machado, F.M.V.; Araújo, A.C.; Meneguim, C.O.; Pascoal Silveira, E.; de Souza Oliveira, N.; da Silva, B.C. *Annona montana* fruit and leaves improve the glycemic and lipid profiles of Wistar rats. *J. Med. Food* **2012**, *15*, 917–922. [\[CrossRef\]](#)
100. Costa, E.V.; Dutra, L.M.; Salvador, M.J.; Ribeiro, L.H.G.; Gadelha, F.R.; de Carvalho, J.E. Chemical composition of the essential oils of *Annona pickelii* and *Annona salzmanii* (Annonaceae), and their antitumour and trypanocidal activities. *Nat. Prod. Res.* **2013**, *27*, 997–1001. [\[CrossRef\]](#)
101. Costa, E.V.; Dutra, L.M.; de Jesus, H.C.R.; de Lima Nogueira, P.C.; de Souza Moraes, V.R.; Salvador, M.J.; de Holanda Cavalcanti, S.C.; dos Santos, R.L.C.; do Nascimento Prata, A.P. Chemical composition and antioxidant, antimicrobial, and larvicidal activities of the essential oils of *Annona salzmanii* and *A. pickelii* (Annonaceae). *Nat. Prod. Commun.* **2011**, *6*, 907–912. [\[CrossRef\]](#)
102. Auddy, B.; Ferreira, M.; Blasina, F.; Lafon, L.; Arredondo, F.; Dajas, F.; Tripathi, P.; Seal, T.; Mukherjee, B. Screening of antioxidant activity of three Indian medicinal plants, traditionally used for the management of neurodegenerative diseases. *J. Ethnopharmacol.* **2003**, *84*, 131–138. [\[CrossRef\]](#)
103. Ofukwu, R.; Ayoola, A.; Akwuobu, C. Medicinal plants used in the treatment of tuberculosis in humans and animals by Idom tribe of North Central Nigeria. *Niger. Vet. J.* **2008**, *29*, 25–30. [\[CrossRef\]](#)

104. Silva, J.C.; Araújo, C.d.S.; de Lima-Saraiva, S.R.G.; de Oliveira-Junior, R.G.; Diniz, T.C.; Wanderley, C.W.d.S.; Pallheta-Júnior, R.C.; Mendes, R.L.; Guimarães, A.G.; Quintans-Júnior, L.J. Antinociceptive and anti-inflammatory activities of the ethanolic extract of *Annona vepretorum* Mart.(Annonaceae) in rodents. *BMC Complement. Med. Ther.* **2015**, *15*, 197. [CrossRef]
105. Chang, F.-R.; Wei, J.-L.; Teng, C.-m.; Wu, Y.-C. Two new 7-dehydroaporphine alkaloids and antiplatelet action aporphines from the leaves of *Annona purpurea*. *Phytochemistry* **1998**, *49*, 2015–2018. [CrossRef]
106. Pathak, K.; Zaman, K. An overview on medicinally important plant—*Annona reticulata* Linn. *Int. J. Pharm. Pharm. Res.* **2013**, *5*, 299–301.
107. Siebra, C.A.; Nardin, J.M.; Florão, A.; Rocha, F.H.; Bastos, D.Z.; Oliveira, B.H.; Weffort-Santos, A.M. Potencial antiinflamatório de *Annona glabra*, Annonaceae. *Rev. Bras. Farmacogn.* **2009**, *19*, 82–88. [CrossRef]
108. Morton, J.F.; Dowling, C.F. *Fruits of Warm Climates*; JF Morton: Miami, FL, USA, 1987.
109. Jansen, P.; Lemmens, R.; Oyen, L. *Plant Resources of South-East Asia: Basic List of Species Commodity Grouping. Final Version*; Pudoc: Wageningen, The Netherlands, 1991.
110. Pinheiro, M.L.B.; Xavier, C.M.; de Souza, A.D.; Rabelo, D.d.M.; Batista, C.L.; Batista, R.L.; Costa, E.V.; Campos, F.R.; Barison, A.; Valdez, R.H. Acanthoic acid and other constituents from the stem of *Annona amazonica* (Annonaceae). *J. Braz. Chem. Soc.* **2009**, *20*, 1095–1102. [CrossRef]
111. Pino, J.A. Volatile components of Cuban *Annona* fruits. *J. Essent. Oil Res.* **2000**, *12*, 613–616. [CrossRef]
112. Martínez-Vázquez, M.; Diana, G.; Estrada-Reyes, R.; González-Lugo, N.M.; Apan, T.R.; Heinze, G. Bio-guided isolation of the cytotoxic corytenchne and isocoreximine from roots of *Annona cherimolia*. *Fitoterapia* **2005**, *76*, 733–736. [CrossRef] [PubMed]
113. Woo, M.-H.; Kim, D.-H.; Fotopoulos, S.S.; McLaughlin, J.L. Annocherin and (2, 4)-cis- and trans-annocherones, monotetrahydrofuran annonaceous acetogenins with a C-7 carbonyl group from *Annona cherimolia* seeds. *J. Nat. Prod.* **1999**, *62*, 1250–1255. [CrossRef] [PubMed]
114. Chen, C.-Y.; Chang, F.-R.; Chiu, H.-F.; Wu, M.-J.; Wu, Y.-C. Aromin-A, an Annonaceous acetogenin from *Annona cherimolia*. *Phytochemistry* **1999**, *51*, 429–433. [CrossRef]
115. Chen, C.-Y.; Chang, F.-R.; Pan, W.-B.; Wu, Y.-C. Four alkaloids from *Annona cherimolia*. *Phytochemistry* **2001**, *56*, 753–757. [CrossRef]
116. Sahpaz, S.; Carmen, M.; Hocquemiller, R.; Zafra-Polo, M.C.; Cortes, D. Annosenegalin and annogalene: Two cytotoxic monotetrahydrofuran acetogenins from *Annona senegalensis* and *Annona cherimolia*. *Phytochemistry* **1996**, *42*, 103–107. [CrossRef]
117. Meneses da Silva, E.; Roblot, F.; Laprévote, O.; Varenne, P.; Cavé, A. Coriacin and 4-deoxycoriacin, two new mono-THF acetogenins from the roots of *Annona coriacea*. *Nat. Prod. Lett.* **1995**, *7*, 235–242. [CrossRef]
118. Meneses da Silva, E.L.; Roblot, F.; Mahuteau, J.; Cavé, A. Coriadienin, the first annonaceous acetogenin with two double bonds isolated from *Annona coriacea*. *J. Nat. Prod.* **1996**, *59*, 528–530. [CrossRef] [PubMed]
119. Da Silva, E.L.M.; Roblot, F.; Laprévote, O.; Sérani, L.; Cavé, A. Coriaheptocins A and B, the first heptahydroxylated acetogenins, isolated from the roots of *Annona coriacea*. *J. Nat. Prod.* **1997**, *60*, 162–167. [CrossRef]
120. Cavé, A.; Figadère, B.; Laurens, A.; Cortes, D. Acetogenins from annonaceae. *Fortschr. Chem. Org. Naturst.* **1997**, 81–288.
121. Ferrari, M.; Pelizzoni, F.; Ferrari, G. New diterpenoids with clerodane skeleton. *Phytochemistry* **1971**, *10*, 3267–3269. [CrossRef]
122. Mussini, P.; Orsini, F.; Pelizzoni, F.; Ferrari, G. Constituents of *Annona coriacea*. The structure of a new diterpenoid. *J. Chem. Soc. Perkin Trans. I* **1973**, 2551–2557. [CrossRef]
123. Novaes, P.; Ferreira, M.J.P.; dos Santos, D.Y.A.C. Flavonols from *Annona coriacea* Mart.(Annonaceae). *Biochem. Syst. Ecol.* **2018**, *78*, 77–80. [CrossRef]
124. Lage, G.A.; Medeiros, F.d.S.; Furtado, W.d.L.; Takahashi, J.A.; Filho, J.D.d.S.; Pimenta, L.P.S. The first report on flavonoid isolation from *Annona crassiflora* Mart. *Nat. Prod. Res.* **2014**, *28*, 808–811. [CrossRef]
125. Costa, E.V.; Pinheiro, M.L.B.; Souza, A.D.L.d.; Barison, A.; Campos, F.R.; Valdez, R.H.; Ueda-Nakamura, T.; Nakamura, C.V. Trypanocidal activity of oxoaporphine and pyrimidine- β -carboline alkaloids from the branches of *Annona foetida* Mart. (Annonaceae). *Molecules* **2011**, *16*, 9714–9720. [CrossRef]
126. Costa, E.V.; Pinheiro, M.L.B.; Silva, J.R.d.A.; Maia, B.H.L.d.N.S.; Duarte, M.C.T.; Amaral, A.C.F.; Machado, G.M.d.C.; Leon, L.L. Antimicrobial and antileishmanial activity of essential oil from the leaves of *Annona foetida* (Annonaceae). *Química Nova* **2009**, *32*, 78–81. [CrossRef]
127. Costa, E.V.; Pinheiro, M.L.B.; Xavier, C.M.; Silva, J.R.; Amaral, A.C.F.; Souza, A.D.; Barison, A.; Campos, F.R.; Ferreira, A.G.; Machado, G.M. A pyrimidine- β -carboline and other alkaloids from *Annona foetida* with antileishmanial activity. *J. Nat. Prod.* **2006**, *69*, 292–294. [CrossRef]
128. Chang, F.R.; Chen, C.Y.; Hsieh, T.J.; Cho, C.P.; Wu, Y.C. Chemical constituents from *Annona glabra* III. *J. Chin. Chem. Soc.* **2000**, *47*, 913–920. [CrossRef]
129. Chang, F.-R.; Yang, P.-Y.; Lin, J.-Y.; Lee, K.-H.; Wu, Y.-C. Bioactive kaurane diterpenoids from *Annona glabra*. *J. Nat. Prod.* **1998**, *61*, 437–439. [CrossRef] [PubMed]
130. Lee, S.-S.; Wu, D.-Y.; Tsai, S.-F.; Chen, C.-K. Anti-acetylcholinesterase alkaloids from *Annona glabra* leaf. *Nat. Prod. Commun.* **2015**, *10*, 891–893. [CrossRef] [PubMed]
131. Wu, T.-Y.; Yang, I.-H.; Tsai, Y.-T.; Wang, J.-Y.; Shiurba, R.; Hsieh, T.-J.; Chang, F.-R.; Chang, W.-C. Isodesacetylvaricin, an annonaceous acetogenin, specifically inhibits gene expression of cyclooxygenase-2. *J. Nat. Prod.* **2012**, *75*, 572–576. [CrossRef] [PubMed]

132. Chen, C.-H.; Hsieh, T.-J.; Liu, T.-Z.; Chern, C.-L.; Hsieh, P.-Y.; Chen, C.-Y. Annoglabayin, a novel dimeric kaurane diterpenoid, and apoptosis in Hep G2 Cells of Annomontacin from the Fruits of *Annona glabra*. *J. Nat. Prod.* **2004**, *67*, 1942–1946. [\[CrossRef\]](#)
133. de Castro Rodrigues, C.M.S.; Dutra, L.M.; Barison, A.; Costa, E.V.; da Silva Almeida, J.R.G. Isoquinoline alkaloids from the leaves of *Annona leptopetala* (Annonaceae). *Biochem. Syst. Ecol.* **2016**, *69*, 222–225. [\[CrossRef\]](#)
134. Liaw, C.-C.; Chang, F.-R.; Chen, S.-L.; Wu, C.-C.; Lee, K.-H.; Wu, Y.-C. Novel cytotoxic monotetrahydrofuranic Annonaceous acetogenins from *Annona montana*. *Bioorg. Med. Chem.* **2005**, *13*, 4767–4776. [\[CrossRef\]](#)
135. Wu, Y.-C.; Chang, G.-Y.; Chang-Yih, D.; Shang-Kwei, W. Cytotoxic alkaloids of *Annona montana*. *Phytochemistry* **1993**, *33*, 497–500. [\[CrossRef\]](#)
136. Wu, Y.-C.; Chang, G.-Y.; Ko, F.-N.; Teng, C.-M. Bioactive constituents from the stems of *Annona montana*. *Planta Med.* **1995**, *61*, 146–149. [\[CrossRef\]](#)
137. Hidalgo, J.R.; Gilabert, M.; Cabedo, N.; Cortes, D.; Neske, A. Montanacin-L and montanacin-K two previously non-described acetogenins from *Annona montana* twigs and leaves. *Phytochem. Lett.* **2020**, *38*, 78–83. [\[CrossRef\]](#)
138. Melot, A.; Fall, D.; Gleye, C.; Champy, P. Apolar Annonaceous acetogenins from the fruit pulp of *Annona muricata*. *Molecules* **2009**, *14*, 4387–4395. [\[CrossRef\]](#) [\[PubMed\]](#)
139. Sun, S.; Liu, J.; Kadouh, H.; Sun, X.; Zhou, K. Three new anti-proliferative annonaceous acetogenins with mono-tetrahydrofuran ring from graviola fruit (*Annona muricata*). *Bioorg. Med. Chem. Lett.* **2014**, *24*, 2773–2776. [\[CrossRef\]](#) [\[PubMed\]](#)
140. Hasrat, J.; De Bruyne, T.; De Backer, J.-P.; Vauquelin, G.; Vlietinck, A. Isoquinoline derivatives isolated from the fruit of *Annona muricata* as 5-HT_{1A} receptor agonists in rats: Unexploited antidepressive (lead) products. *J. Pharm. Pharmacol.* **1997**, *49*, 1145–1149. [\[CrossRef\]](#) [\[PubMed\]](#)
141. Wu, F.-E.; Zhao, G.-X.; Zeng, L.; Zhang, Y.; Schwedler, J.T.; McLaughlin, J.L.; Sastrodihardjo, S. Additional bioactive acetogenins, annomutacin and (2, 4-trans and cis)-10R-annonacin-A-ones, from the leaves of *Annona muricata*. *J. Nat. Prod.* **1995**, *58*, 1430–1437. [\[CrossRef\]](#) [\[PubMed\]](#)
142. Zeng, L.; Wu, F.-E.; Gu, Z.-m.; McLaughlin, J.L. Murihexocins A and B, two novel mono-THF acetogenins with six hydroxyls, from *Annona muricata* (Annonaceae). *Tetrahedron Lett.* **1995**, *36*, 5291–5294. [\[CrossRef\]](#)
143. Zeng, L.; Wu, F.-E.; McLaughlin, J.L. Annohexocin, a novel mono-THF acetogenin with six hydroxyls, from *Annona muricata* (Annonaceae). *Bioorg. Med. Chem. Lett.* **1995**, *5*, 1865–1868. [\[CrossRef\]](#)
144. Zeng, L.; Wu, F.-E.; Oberlies, N.H.; McLaughlin, J.L.; Sastrodihardjo, S. Five new monotetrahydrofuran ring acetogenins from the leaves of *Annona muricata*. *J. Nat. Prod.* **1996**, *59*, 1035–1042. [\[CrossRef\]](#)
145. Nawwar, M.; Ayoub, N.; Hussein, S.; Hashim, A.; El-Sharawy, R.; Wende, K.; Harms, M.; Lindequist, U. Flavonol triglycoside and investigation of the antioxidant and cell stimulating activities of *Annona muricata* Linn. *Arch. Pharm. Res.* **2012**, *35*, 761–767. [\[CrossRef\]](#)
146. Wu, F.-E.; Zeng, L.; Gu, Z.-M.; Zhao, G.-X.; Zhang, Y.; Schwedler, J.T.; McLaughlin, J.L.; Sastrodihardjo, S. Muricatocins A and B, two new bioactive monotetrahydrofuran Annonaceous acetogenins from the leaves of *Annona muricata*. *J. Nat. Prod.* **1995**, *58*, 902–908. [\[CrossRef\]](#)
147. del Carmen Rejón-Orantes, J.; González-Esquinca, A.R.; de la Mora, M.P.; Roldan, G.R.; Cortes, D. Annomontine, an alkaloid isolated from *Annona purpurea*, has anxiolytic-like effects in the elevated plus-maze. *Planta Med.* **2011**, *77*, 322–327. [\[CrossRef\]](#)
148. Bhalke, R.D.; Chavan, M.J. Analgesic and CNS depressant activities of extracts of *Annona reticulata* Linn. bark. *Phytopharm.* **2011**, *1*, 160–165.
149. Islam, M.R.; Rahman, S.M.; Ahmed, M.; Das, P.R.; Tabibul, M.; Islam, M.H.K.; Ahmad, I.; Rahmatullah, M. Antinociceptive activity studies with methanol extract of *Annona reticulata* L. (Annonaceae) and *Carissa carandas* L. (Apocynaceae) leaves in Swiss albino mice. *ANAS* **2012**, *6*, 1313–1318.
150. Thang, T.D.; Kuo, P.-C.; Huang, G.-J.; Hung, N.H.; Huang, B.-S.; Yang, M.-L.; Luong, N.X.; Wu, T.-S. Chemical constituents from the leaves of *Annona reticulata* and their inhibitory effects on NO production. *Molecules* **2013**, *18*, 4477–4486. [\[CrossRef\]](#) [\[PubMed\]](#)
151. Chavan, M.J.; Kolhe, D.R.; Wakte, P.S.; Shinde, D.B. Analgesic and antiinflammatory activity of kaur-16-en-19-oic acid from *Annona reticulata* L. Bark. *Phytother. Res.* **2012**, *26*, 273–276. [\[CrossRef\]](#)
152. Chavan, M.J.; Wakte, P.S.; Shinde, D.B. Analgesic and anti-inflammatory activities of the sesquiterpene fraction from *Annona reticulata* L. bark. *J. Nat. Prod.* **2012**, *26*, 1515–1518. [\[CrossRef\]](#)
153. Ameen, O.; Usman, L.; Oganija, F.; Hamid, A.; Muhammed, N.; Zubair, M.; Adebayo, S. Chemical composition of leaf essential oil of *Annona senegalensis* Pers. (Annonaceae) growing in North Central Nigeria. *Int. J. Biol. Chem. Sci.* **2011**, *5*, 375–379. [\[CrossRef\]](#)
154. Lall, N.; Kishore, N.; Bodiba, D.; More, G.; Tshikalange, E.; Kikuchi, H.; Oshima, Y. Alkaloids from aerial parts of *Annona senegalensis* against *Streptococcus mutans*. *Nat. Prod. Res.* **2017**, *31*, 1944–1947. [\[CrossRef\]](#)
155. Campos, F.R.; Batista, R.L.; Batista, C.L.; Costa, E.V.; Barison, A.; dos Santos, A.G.; Pinheiro, M.L.B. Isoquinoline alkaloids from leaves of *Annona sericea* (Annonaceae). *Biochem. Syst. Ecol.* **2008**, *36*, 804–806. [\[CrossRef\]](#)
156. Li, X.-H.; Hui, Y.-H.; Rupprecht, J.; Liu, Y.-M.; Wood, K.; Smith, D.; Chang, C.-J.; McLaughlin, J. Bullatacin, bullatacinone, and squamone, a new bioactive acetogenin, from the bark of *Annona squamosa*. *J. Nat. Prod.* **1990**, *53*, 81–86. [\[CrossRef\]](#)
157. Fujimoto, Y.; Eguchi, T.; Kakinuma, K.; Ikekawa, N.; Sahai, M.; Gupta, Y.K. Squamocin, a new cytotoxic bis-tetrahydrofuran containing acetogenin from *Annona squamosa*. *Chem. Pharm. Bull.* **1988**, *36*, 4802–4806. [\[CrossRef\]](#)

158. Meira, C.S.; Guimarães, E.T.; Macedo, T.S.; da Silva, T.B.; Menezes, L.R.; Costa, E.V.; Soares, M.B. Chemical composition of essential oils from *Annona vepretorum* Mart. and *Annona squamosa* L. (Annonaceae) leaves and their antimalarial and trypanocidal activities. *J. Essent. Oil Res.* **2015**, *27*, 160–168. [\[CrossRef\]](#)
159. De Oliveira, A.B.; De Oliveira, G.G.; Carazza, F.; Maia, J.G.S. Geovanine, a new azaanthracene alkaloid from *Annona ambotay* aubl. *Phytochemistry* **1987**, *26*, 2650–2651. [\[CrossRef\]](#)
160. Gu, Z.M.; Fang, X.P.; Hui, Y.H.; McLaughlin, J.L. 10-, 12-, and 29-Hydroxybullatacinones: New cytotoxic annonaceous acetogenins from *Annona bullata* rich (annonaceae). *Nat. Toxins* **1994**, *2*, 49–55. [\[CrossRef\]](#) [\[PubMed\]](#)
161. Garcia, V.N.; Gonzalez, A.; Fuentes, M.; Aviles, M.; Rios, M.; Zepeda, G.; Rojas, M. Antifungal activities of nine traditional Mexican medicinal plants. *J. Ethnopharmacol.* **2003**, *87*, 85–88. [\[CrossRef\]](#)
162. Chan, P.; Ah, R.; Mh, K. Anti-arthritis activities of *Annona muricata* L. leaves extract on complete Freund's adjuvant (CFA)-induced arthritis in rats. *Planta Med.* **2010**, *76*, 166. [\[CrossRef\]](#)
163. Adeyemi, D.O.; Komolafe, O.A.; Adewole, O.S.; Obuotor, E.M.; Adenowo, T.K. Anti hyperglycemic activities of *Annona muricata* (Linn). *Afr. J. Tradit. Complem* **2009**, *6*, 62.
164. Okoye, T.C.; Akah, P.A.; Omeje, E.O.; Okoye, F.B.; Nworu, C.S. Anticonvulsant effect of kaurenoic acid isolated from the root bark of *Annona senegalensis*. *Pharmacol. Biochem. Behav.* **2013**, *109*, 38–43. [\[CrossRef\]](#)
165. Viera, G.H.F.; Mourão, J.A.; Ângelo, Â.M.; Costa, R.A.; Vieira, R.H.S.d.F. Antibacterial effect (in vitro) of *Moringa oleifera* and *Annona muricata* against Gram positive and Gram negative bacteria. *Rev. Inst. Med. Trop* **2010**, *52*, 129–132. [\[CrossRef\]](#)
166. Bento, E.B.; Matias, E.F.; Brito Jr, F.E.; Oliveira, D.R.; Coutinho, H.D.; Costa, J.G.; Kerntopf, M.R.; Menezes, I.R. Association between food and drugs: Antimicrobial and synergistic activity of *Annona muricata* L. *Int. J. Food Prop.* **2013**, *16*, 738–744. [\[CrossRef\]](#)
167. Nasser, M.; El-Mestrah, M.; As-sadi, F.; Cheaito, L.; Hijazi, A.; Chokr, A.; Hassan, R. Antibacterial, antioxidant and antiproliferative activities of the hydroalcoholic extract of the Lebanese *Annona squamosa* L. seeds. *Int. Res. J. Pharm* **2017**, *8*, 1–7.
168. Shanker, K.S.; Kanjilal, S.; Rao, B.; Kishore, K.H.; Misra, S.; Prasad, R. Isolation and antimicrobial evaluation of isomeric hydroxy ketones in leaf cuticular waxes of *Annona squamosa*. *Phytochem. Anal.* **2007**, *18*, 7–12. [\[CrossRef\]](#) [\[PubMed\]](#)
169. Nakano, D.; Ishitsuka, K.; Kamikawa, M.; Matsuda, M.; Tsuchihashi, R.; Okawa, M.; Okabe, H.; Tamura, K.; Kinjo, J. Screening of promising chemotherapeutic candidates from plants against human adult T-cell leukemia/lymphoma (III). *J. Nat. Med.* **2013**, *67*, 894–903. [\[CrossRef\]](#) [\[PubMed\]](#)
170. Osorio, E.; Arango, G.J.; Jiménez, N.; Alzate, F.; Ruiz, G.; Gutiérrez, D.; Paco, M.A.; Giménez, A.; Robledo, S. Antiprotozoal and cytotoxic activities in vitro of Colombian Annonaceae. *J. Ethnopharmacol.* **2007**, *111*, 630–635. [\[CrossRef\]](#) [\[PubMed\]](#)
171. Formagio, A.; Vieira, M.; Volobuff, C.; Silva, M.; Matos, A.; Cardoso, C.; Foglio, M.; Carvalho, J. In vitro biological screening of the anticholinesterase and antiproliferative activities of medicinal plants belonging to Annonaceae. *Braz. J. Med. Biol.* **2015**, *48*, 308–315. [\[CrossRef\]](#)
172. Yuan, S.-S.F.; Chang, H.-L.; Chen, H.-W.; Yeh, Y.-T.; Kao, Y.-H.; Lin, K.-H.; Wu, Y.-C.; Su, J.-H. Annonacin, a mono-tetrahydrofuran acetogenin, arrests cancer cells at the G1 phase and causes cytotoxicity in a Bax-and caspase-3-related pathway. *Life Sci.* **2003**, *72*, 2853–2861. [\[CrossRef\]](#)
173. Lima, L.A.; Alves, T.; Zani, C.L.; Sales, P.A.; Romanha, A.J.; Johann, S.; Cisalpino, P.S.; Pimenta, L.P.; Boaventura, M.A.D. In vitro cytotoxic, antifungal, trypanocidal and leishmanicidal activities of acetogenins isolated from *Annona cornifolia* A. St.-Hil. (Annonaceae). *An. Acad. Bras. Cienc.* **2014**, *86*, 829–839. [\[CrossRef\]](#)
174. Schlie-Guzmán, M.A.; García-Carrancá, A.; González-Esquinca, A.R. In vitro and in vivo antiproliferative activity of laherradurin and cherimolin-2 of *Annona diversifolia* saff. *Phytother. Res.* **2009**, *23*, 1128–1133. [\[CrossRef\]](#)
175. Hansra, D.M.; Silva, O.; Mehta, A.; Ahn, E. Patient with metastatic breast cancer achieves stable disease for 5 years on graviola and xeloda after progressing on multiple lines of therapy. *Adv. Breast Cancer Res.* **2014**, *3*, 84–87. [\[CrossRef\]](#)
176. Beale, P.; Syed, S.; Pham, T.T.D. Graviola in metastatic ovarian cancer: A case report of sustained disease stability. *Clin. Oncol. Res.* **2019**, *2*, 1–3. [\[CrossRef\]](#)
177. Indrawati, L.; Ascobat, P.; Bela, B.; Abdullah, M.; Surono, I.S. The effect of an 'Annona muricata' leaf extract on nutritional status and cytotoxicity in colorectal cancer: A randomized controlled trial. *Asia Pac. J. Clin. Nutr.* **2017**, *26*, 606–612.
178. Shirwaikar, A.; Rajendran, K.; Kumar, C.D.; Bodla, R. Antidiabetic activity of aqueous leaf extract of *Annona squamosa* in streptozotocin–nicotinamide type 2 diabetic rats. *J. Ethnopharmacol.* **2004**, *91*, 171–175. [\[CrossRef\]](#) [\[PubMed\]](#)
179. Singh, S.; Manvi, F.; Nanjwade, B.; Nema, R.K. Antihyperlipidemic screening of polyherbal formulation of *Annona squamosa* and *Nigella sativa*. *Inter. J. Toxicol. Pharmacol. Res.* **2010**, *2*, 1–5.
180. Florence, N.T.; Benoit, M.Z.; Jonas, K.; Alexandra, T.; Désiré, D.D.P.; Pierre, K.; Théophile, D. Antidiabetic and antioxidant effects of *Annona muricata* (Annonaceae), aqueous extract on streptozotocin-induced diabetic rats. *J. Ethnopharmacol.* **2014**, *151*, 784–790. [\[CrossRef\]](#) [\[PubMed\]](#)
181. Rout, S.P.; Kar, D.M.; Mohapatra, S.B.; Swain, S.P. Anti-hyperglycemic effect *Annona reticulata* L. leaves on experimental diabetic rat model. *Asian J. Pharm. Clin. Res.* **2013**, *6*, 56–60.
182. Martínez-Solís, J.; Calzada, F.; Barbosa, E.; Valdés, M. Antihyperglycemic and antilipidemic properties of a tea infusion of the leaves from *Annona cherimola* miller on streptozocin-induced type 2 diabetic mice. *Molecules* **2021**, *26*, 2408. [\[CrossRef\]](#)
183. Adeyemi, D.; Komolafe, O.; Adewole, S.; Obuotor, E. Anti hyperlipidemic activities of *Annona muricata* (Linn). *Internet J. Alternative Med.* **2009**, *7*, 1–9.

184. Hemlatha, K.; Satyanarayana, D. Anti-inflammatory activity of *Annona squamosa* Linn. *Biomed. Pharmacol. J.* **2015**, *2*, 17–20.
185. Oyekachukwu, A.; Elijah, J.; Eshu, O.; Nwodo, O. Anti-inflammatory effects of the chloroform extract of *Annona muricata* leaves on phospholipase A2 and prostaglandin synthase activities. *Transl. Biomed.* **2017**, *8*, 137.
186. Konaté, K.; Sanou, A.; Aworet-Samseny, R.R.; Benkhalti, F.; Sytar, O.; Brestic, M.; Souza, A.; Dicko, M.H. Safety profile, in vitro anti-inflammatory activity, and in vivo antiulcerogenic potential of root barks from *Annona senegalensis* Pers. (Annonaceae). *Evid. Based Complement. Altern. Med.* **2021**, *2021*, 4441375. [\[CrossRef\]](#)
187. De Sousa, O.V.; Vieira, G.D.-V.; De Pinho, J.d.J.R.; Yamamoto, C.H.; Alves, M.S. Antinociceptive and anti-inflammatory activities of the ethanol extract of *Annona muricata* L. leaves in animal models. *Int. J. Mol. Sci.* **2010**, *11*, 2067–2078. [\[CrossRef\]](#)
188. Ishola, I.O.; Awodele, O.; Olusayero, A.M.; Ochieng, C.O. Mechanisms of analgesic and anti-inflammatory properties of *Annona muricata* Linn.(Annonaceae) fruit extract in rodents. *J. Med. Food* **2014**, *17*, 1375–1382. [\[CrossRef\]](#) [\[PubMed\]](#)
189. Formagio, A.S.; Kassuya, C.A.; Neto, F.F.; Volobuff, C.R.; Iriguchi, E.K.; Vieira, M.d.C.; Foglio, M.A. The flavonoid content and antiproliferative, hypoglycaemic, anti-inflammatory and free radical scavenging activities of *Annona dioica* St. Hill. *BMC Complement. Altern. Med.* **2013**, *13*, 14. [\[CrossRef\]](#) [\[PubMed\]](#)
190. Benites, R.; Formagio, A.; Argandoña, E.; Volobuff, C.; Trevizan, L.; Vieira, M.; Silva, M. Contents of constituents and antioxidant activity of seed and pulp extracts of *Annona coriacea* and *Annona sylvatica*. *Braz. J. Biol.* **2015**, *75*, 685–691. [\[CrossRef\]](#) [\[PubMed\]](#)
191. Baskar, R.; Rajeswari, V.; Kumar, T.S. In vitro antioxidant studies in leaves of *Annona* species. *Indian J. Exp. Biol.* **2007**, *45*, 480–485. [\[PubMed\]](#)
192. Vila-Nova, N.S.; Morais, S.M.d.; Falcão, M.J.C.; Machado, L.K.A.; Beviláqua, C.M.L.; Costa, I.R.S.; Brasil, N.V.G.P.d.S.; Andrade Júnior, H.F.d. Leishmanicidal activity and cytotoxicity of compounds from two Annonacea species cultivated in Northeastern Brazil. *Rev. Soc. Bras. Med. Trop* **2011**, *44*, 567–571. [\[CrossRef\]](#) [\[PubMed\]](#)
193. De Lima, J.P.; Pinheiro, M.L.; Santos, A.M.G.; Pereira, J.L.d.S.; Santos, D.M.; Barison, A.; Silva-Jardim, I.; Costa, E.V. In vitro antileishmanial and cytotoxic activities of *Annona mucosa* (Annonaceae). *Rev. Virtual. Quim.* **2012**, *4*, 692–702.
194. Wu, Y.-C.; Hung, Y.-C.; Chang, F.-R.; Cosentino, M.; Wang, H.-K.; Lee, K.-H. Identification of ent-16 β , 17-dihydroxykauran-19-oic acid as an anti-HIV principle and isolation of the new diterpenoids annosquamosins A and B from *Annona squamosa*. *J. Nat. Prod.* **1996**, *59*, 635–637. [\[CrossRef\]](#)
195. Yunita, M.N.; Raharjo, A.P.; Wibawati, P.A.; Agustono, B. Antiviral activity of ethanolic extract of srikaya seeds (*Annona squamosa* L.) against avian influenza virus. *Indian Vet. J.* **2019**, *96*, 26–29.
196. Ansori, A.N.M.; Fadholly, A.; Proboningrat, A.; Antonius, Y.; Hayaza, S.; Susilo, R.J.K.; Inayatillah, B.; Sibero, M.T.; Naw, S.W.; Posa, G.A.V. Novel antiviral investigation of *Annona squamosa* Leaf extract against the Dengue Virus Type-2: In vitro study. *Pharmacogn. J.* **2021**, *13*, 456–462. [\[CrossRef\]](#)
197. Chel-Guerrero, L.D.; Gómez-Cansino, R.; Gúzman-Gutierrez, S.L.; Campos-Lara, M.G.; Saury-Duch, E.; Díaz De León Sánchez, F.; Reyes-Chilpa, R.; Mendoza-Espinoza, J.A. In vitro antiviral activity and phytochemical screen in the extracts of peels from four species of tropical fruits collected in Merida Yucatan, Mexico. *Phyton* **2018**, *87*, 68–71.
198. Betancur-Galvis, L.; Saez, J.; Granados, H.; Salazar, A.; Ossa, J. Antitumor and antiviral activity of Colombian medicinal plant extracts. *Mem. Inst. Oswaldo Cruz* **1999**, *94*, 531–535. [\[CrossRef\]](#) [\[PubMed\]](#)
199. Padma, P.; Pramod, N.; Thyagarajan, S.; Khosa, R. Effect of the extract of *Annona muricata* and *Petunia nyctaginiflora* on Herpes simplex virus. *J. Ethnopharmacol.* **1998**, *61*, 81–83. [\[CrossRef\]](#)
200. Rodríguez-Expósito, R.L.; Sosa-Rueda, J.; Reyes-Battle, M.; Sifaoui, I.; Cen-Pacheco, F.; Daranas, A.H.; Díaz-Marrero, A.R.; Piñero, J.E.; Fernández, J.J.; Lorenzo-Morales, J. Antiamoeboid activity of squamins C–F, cyclooctapeptides from *Annona globiflora*. *Int. J. Parasitol. Drugs Drug Resist.* **2021**, *17*, 67–79. [\[CrossRef\]](#) [\[PubMed\]](#)
201. Rodrigues, A.M.; da Silva, A.A.; de Freitas, J.C.C.; Martins, V.E.P.; Ferreira, M.A.P.; Ferreira, A.C.S.; Cabeça, C.L.S.; Rodrigues, A.L.M.; Alves, D.R.; de Morais, S.M. Larvicidal activity of *Annona mucosa* Jacq. extract and main constituents rolliniastatin 1 and rollinacin against *Aedes aegypti* and *Aedes albopictus*. *Ind. Crops Prod.* **2021**, *169*, 113678. [\[CrossRef\]](#)
202. Rodrigues, A.M.; Silva, A.A.S.; Pinto, C.C.C.; Santos, D.L.d.; Freitas, J.C.C.d.; Martins, V.E.P.; Morais, S.M.d. Larvicidal and enzymatic inhibition effects of *Annona muricata* seed extract and main constituent annonacin against *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae). *Pharmaceuticals* **2019**, *12*, 112. [\[CrossRef\]](#)
203. Costa, E.V.; da Cruz, P.E.O.; de Lourenço, C.C.; de Souza Moraes, V.R.; de Lima Nogueira, P.C.; Salvador, M.J. Antioxidant and antimicrobial activities of aporphinoids and other alkaloids from the bark of *Annona salzmannii* A. DC.(Annonaceae). *Nat. Prod. Res.* **2013**, *27*, 1002–1006. [\[CrossRef\]](#)
204. Pena-Hidalgo, M.; Furtado, L.C.; Costa-Lotufo, L.V.; Ferreira, M.J.; Santos, D.Y. Alkaloids from the Leaves of *Annona crassiflora* and their cytotoxic activity. *Rev. Bras. Farmacogn* **2021**, *31*, 244–248. [\[CrossRef\]](#)
205. Daddiouaissa, D.; Amid, A. Anticancer activity of acetogenins from *Annona muricata* fruit. *Int. Med. J. Malays.* **2018**, *17*, 1–6. [\[CrossRef\]](#)
206. Al-Ghazzawi, A.M. Anti-cancer activity of new benzyl isoquinoline alkaloid from Saudi plant *Annona squamosa*. *BMC. Chemistry.* **2019**, *13*, 13. [\[CrossRef\]](#)
207. Durán, A.G.; Gutiérrez, M.T.; Mejías, F.J.; Molinillo, J.M.; Macías, F.A. An Overview of the Chemical Characteristics, Bioactivity and Achievements Regarding the Therapeutic Usage of Acetogenins from *Annona cherimola* Mill. *Molecules* **2021**, *26*, 2926. [\[CrossRef\]](#)

208. Pereira, M.N.; Justino, A.B.; Martins, M.M.; Peixoto, L.G.; Vilela, D.D.; Santos, P.S.; Teixeira, T.L.; da Silva, C.V.; Goulart, L.R.; Pivatto, M. Stephalagine, an alkaloid with pancreatic lipase inhibitory activity isolated from the fruit peel of *Annona crassiflora* Mart. *Ind. Crops Prod.* **2017**, *97*, 324–329. [\[CrossRef\]](#)
209. Kim, D.H.; Ma, E.S.; Suk, K.D.; Son, J.K.; Lee, J.S.; Woo, M.H. Annomolin and Annocherimolin, new cytotoxic annonaceous acetogenins from *Annona cherimolia* Seeds. *J. Nat. Prod.* **2001**, *64*, 502–506. [\[CrossRef\]](#) [\[PubMed\]](#)
210. Ammourey, C.; Younes, M.; El Khoury, M.; Hodroj, M.H.; Haykal, T.; Nasr, P.; Sily, M.; Taleb, R.I.; Sarkis, R.; Khalife, R.; et al. The pro-apoptotic effect of a terpene-rich *Annona cherimola* leaf extract on leukemic cell lines. *BMC Complement Altern. Med.* **2019**, *19*, 365. [\[CrossRef\]](#)
211. Silva, V.A.O.; Alves, A.L.V.; Rosa, M.N.; Silva, L.R.V.; Melendez, M.E.; Cury, F.P.; Gomes, I.N.F.; Tansini, A.; Longato, G.B.; Martinho, O.; et al. Hexane partition from *Annona crassiflora* Mart. promotes cytotoxicity and apoptosis on human cervical cancer cell lines. *Invest. New Drugs* **2019**, *37*, 602–615. [\[CrossRef\]](#) [\[PubMed\]](#)
212. Hien, N.; Hang, D.; Ha, T.; Nhiem, N.; Hien, T.; Thu, V.; Van Minh, C.; Kiem, P. Ent-kaurane diterpenes from *Annona glabra* and their cytotoxic activities. *Nat. Prod. Commun.* **2014**, *9*, 1681–1682.
213. Zhang, Y.H.; Peng, H.Y.; Xia, G.H.; Wang, M.Y.; Han, Y. Anticancer effect of two diterpenoid compounds isolated from *Annona glabra* Linn. *Acta Pharmacol. Sin.* **2004**, *25*, 937–942.
214. Liu, Y.; Liu, D.; Wan, W.; Zhang, H. In vitro mitochondria-mediated anticancer and antiproliferative effects of *Annona glabra* leaf extract against human leukemia cells. *J. Photochem. Photobiol. B: Biol.* **2018**, *189*, 29–35. [\[CrossRef\]](#)
215. Liu, X.-X.; Alali, F.Q.; Pilarinou, E.; McLaughlin, J.L. Two bioactive mono-tetrahydrofuran acetogenins, annoglacins A and B, from *Annona glabra*. *Phytochemistry* **1999**, *50*, 815–821. [\[CrossRef\]](#)
216. Hien, N.T.T.; Nhiem, N.X.; Yen, D.T.H.; Hang, D.T.T.; Tai, B.H.; Quang, T.H.; Tuan Anh, H.L.; Kiem, P.V.; Minh, C.V.; Kim, E.-J.; et al. Chemical constituents of the *Annona glabra* fruit and their cytotoxic activity. *Pharm. Biol.* **2015**, *53*, 1602–1607. [\[CrossRef\]](#)
217. Syed Najmuddin, S.U.F.; Romli, M.F.; Hamid, M.; Alitheen, N.B.; Nik Abd Rahman, N.M.A. Anti-cancer effect of *Annona muricata* Linn Leaves Crude Extract (AMCE) on breast cancer cell line. *BMC Complement Altern. Med.* **2016**, *16*, 311. [\[CrossRef\]](#)
218. Zorofchian Moghadamtousi, S.; Karimian, H.; Rouhollahi, E.; Paydar, M.; Fadaeinasab, M.; Abdul Kadir, H. *Annona muricata* leaves induce G1 cell cycle arrest and apoptosis through mitochondria-mediated pathway in human HCT-116 and HT-29 colon cancer cells. *J. Ethnopharmacol.* **2014**, *156*, 277–289. [\[CrossRef\]](#) [\[PubMed\]](#)
219. Kim, G.S.; Zeng, L.; Alali, F.; Rogers, L.L.; Wu, F.-E.; McLaughlin, J.L.; Sastrodihardjo, S. Two new mono-tetrahydrofuran ring acetogenins, annomuricin E and muricapentocin, from the leaves of *Annona muricata*. *J. Nat. Prod.* **1998**, *61*, 432–436. [\[CrossRef\]](#) [\[PubMed\]](#)
220. Sun, S.; Liu, J.; Zhou, N.; Zhu, W.; Dou, Q.P.; Zhou, K. Isolation of three new annonaceous acetogenins from Graviola fruit (*Annona muricata*) and their anti-proliferation on human prostate cancer cell PC-3. *Bioorg. Med. Chem. Lett.* **2016**, *26*, 4382–4385. [\[CrossRef\]](#) [\[PubMed\]](#)
221. Hernández-Fuentes, G.A.; García-Argáez, A.N.; Peraza Campos, A.L.; Delgado-Enciso, I.; Muñiz-Valencia, R.; Martínez-Martínez, F.J.; Toninello, A.; Gómez-Sandoval, Z.; Mojica-Sánchez, J.P.; Dalla Via, L. Cytotoxic Acetogenins from the Roots of *Annona purpurea*. *Int. J. Mol. Sci.* **2019**, *20*, 1870. [\[CrossRef\]](#) [\[PubMed\]](#)
222. Rayar, A.; Manivannan, R. Antioxidant and anticancer activities of (+)-catechin isolated from *Annona reticulata* Linn. *Int. J. Recent Sci. Res.* **2016**, *7*, 9451–9456.
223. Ravimanickam, T.N.Y. Yaminipriya D 3 and Yogananth N 4, Evaluation of antitumor properties of *Annona reticulata* (L.) seed on cancer cell lines. *J. Pharm. Res.* **2018**, *7*, 132–135.
224. Gingine, A.P.; Sv, G.; Jamkh, P.G. In vitro evaluation of Anticancer Activity of Methnaolic Extract of *Annona reticulata* Linn. (Ramphal) Leaves on Different Human Cancer Cell Lines. *J. Anal. Pharmaceut. Res.* **2016**, *3*, 87.
225. Roham, P.H.; Kharat, K.R.; Mungde, P.; Jadhav, M.A.; Makhija, S.J. Induction of mitochondria mediated apoptosis in human breast cancer cells (T-47D) by *Annona reticulata* L. leaves methanolic extracts. *Nutr. Cancer* **2016**, *68*, 305–311. [\[CrossRef\]](#)
226. Agbo'Saga, F.; Ahissou, H.; Baouz, S. Bioguided search for anticancer natural products in the plant *Annona senegalensis*: A preliminary efficacy assessment. *MOJ Biorg Org Chem* **2019**, *3*, 65–70.
227. Fatope, M.O.; Audu, O.T.; Takeda, Y.; Zeng, L.; Shi, G.; Shimada, H.; McLaughlin, J.L. Bioactive ent-Kaurene Diterpenoids from *Annona senegalensis*. *J. Nat. Prod.* **1996**, *59*, 301–303. [\[CrossRef\]](#)
228. Okoye, T.C.; Akah, P.A.; Nworu, C.S.; Ezike, A.C. Kaurenoic acid isolated from the root bark of *Annona senegalensis* induces cytotoxic and antiproliferative effects against PANC-1 and HeLa cells. *European J. Med. Plants* **2014**, 579–589. [\[CrossRef\]](#)
229. Vivek, R.; Thangam, R.; Muthuchelian, K.; Gunasekaran, P.; Kaveri, K.; Kannan, S. Green biosynthesis of silver nanoparticles from *Annona squamosa* leaf extract and its in vitro cytotoxic effect on MCF-7 cells. *Process Biochemistry* **2012**, *47*, 2405–2410. [\[CrossRef\]](#)
230. Ma, C.; Wang, Q.; Shi, Y.; Li, Y.; Wang, X.; Li, X.; Chen, Y.; Chen, J. Three new antitumor annonaceous acetogenins from the seeds of *Annona squamosa*. *Nat. Prod. Res.* **2017**, *31*, 2085–2090. [\[CrossRef\]](#) [\[PubMed\]](#)
231. Fadholly, A.; Proboningrat, A.; Iskandar, R.P.D.; Rantam, F.A.; Sudjarwo, S.A. In vitro anticancer activity *Annona squamosa* extract nanoparticle on WiDr cells. *J. Adv. Pharm. Technol. Res.* **2019**, *10*, 149–154. [\[PubMed\]](#)
232. Pardhasaradhi, B.V.V.; Reddy, M.; Ali, A.M.; Kumari, A.L.; Khar, A. Differential cytotoxic effects of *Annona squamosa* seed extracts on human tumour cell lines: Role of reactive oxygen species and glutathione. *J. Biosci.* **2005**, *30*, 237–244. [\[CrossRef\]](#)
233. Chen, Y.; Chen, J.-w.; Wang, Y.; Xu, S.-s.; Li, X. Six cytotoxic annonaceous acetogenins from *Annona squamosa* seeds. *Food Chem.* **2012**, *135*, 960–966. [\[CrossRef\]](#)

234. Chen, Y.; Chen, Y.; Shi, Y.; Ma, C.; Wang, X.; Li, Y.; Miao, Y.; Chen, J.; Li, X. Antitumor activity of *Annona squamosa* seed oil. *J. Ethnopharmacol.* **2016**, *193*, 362–367. [[CrossRef](#)]
235. Suresh, K.; Manoharan, S.; Panjamurthy, K.; Kavitha, K. Chemopreventive and antilipidperoxidative efficacy of *Annona squamosa* bark extracts in experimental oral carcinogenesis. *Pak. J. Biol. Sci.* **2006**, *9*, 2600–2605. [[CrossRef](#)]
236. Joy, B.; Remani, P. Antitumor constituents from *Annona squamosa* fruit pericarp. *Med. Chem. Res.* **2008**, *17*, 345–355. [[CrossRef](#)]
237. Costa, E.V.; Dutra, L.M.; Nogueira, P.C.d.L.; Moraes, V.R.d.S.; Salvador, M.J.; Ribeiro, L.H.G.; Gadelha, F.R. Essential oil from the leaves of *Annona vepretorum*: Chemical composition and bioactivity. *Nat. Prod. Commun.* **2012**, *7*, 265–266. [[CrossRef](#)]
238. Caparros-Lefebvre, D.; Steele, J.; Kotake, Y.; Ohta, S. Geographic isolates of atypical Parkinsonism and tauopathy in the tropics: Possible synergy of neurotoxins. *J. Mov. Disord.* **2006**, *21*, 1769–1771. [[CrossRef](#)] [[PubMed](#)]
239. Champy, P.; Melot, A.; Guérineau Eng, V.; Gleye, C.; Fall, D.; Höglinger, G.U.; Ruberg, M.; Lannuzel, A.; Laprévote, O.; Laurens, A. Quantification of acetogenins in *Annona muricata* linked to atypical parkinsonism in Guadeloupe. *Mov. Disord.* **2005**, *20*, 1629–1633. [[CrossRef](#)] [[PubMed](#)]
240. Bonneau, N.; Baloul, L.; ba Ndob, I.B.; Senejoux, F.; Champy, P. The fruit of *Annona squamosa* L. as a source of environmental neurotoxins: From quantification of squamocin to annotation of Annonaceous acetogenins by LC–MS/MS analysis. *Food Chem.* **2017**, *226*, 32–40. [[CrossRef](#)] [[PubMed](#)]
241. Höllerhage, M.; Rösler, T.W.; Berjas, M.; Luo, R.; Tran, K.; Richards, K.M.; Sabaa-Srur, A.U.; Maia, J.G.S.; Moraes, M.R.d.; Godoy, H.T. Neurotoxicity of dietary supplements from Annonaceae species. *Int. J. Toxicol.* **2015**, *34*, 543–550. [[CrossRef](#)] [[PubMed](#)]
242. Arroyo, J.; Martínez, J.; Ronceros, G.; Palomino, R.; Villarreal, A.; Bonilla, P.; Palomino, C.; Quino, M. Efecto hipoglicemiante coadyuvante del extracto etanólico de hojas de *Annona muricata* L. (guanábana), en pacientes con diabetes tipo 2 bajo tratamiento de glibenclamida. *An. De La Fac. De Med.* **2009**, *70*, 163–167. [[CrossRef](#)]
243. Kaur, R.; Afzal, M.; Kazmi, I.; Ahamd, I.; Ahmed, Z.; Ali, B.; Ahmad, S.; Anwar, F. Polypharmacy (herbal and synthetic drug combination): A novel approach in the treatment of type-2 diabetes and its complications in rats. *J. Nat. Med.* **2013**, *67*, 662–671. [[CrossRef](#)]