
Time Series and Dynamic Systems Analyses in Drug Discovery

PS-BiOmics&Pathology-BEDA

Roche Pharma Research and Early Development, Pharmaceutical Sciences, Roche Innovation Center Basel

Tony Kam-Thong on behalf of the Bioinformatics and Exploratory Data Analysis team

Guest Lecture Mathematical and Computational Biology in Drug Discovery

University of Basel

7th of May 2021

Outline

- Time series data analysis
 - Disease progression and response to treatment
 - Mixed effects model for repeated measurements
- Dynamic systems
 - Background knowledge
 - RNA velocity estimate from Single Cell RNASeq data
 - Going beyond discrete cell types by inferring cell states and their transitions
- Disclaimer
 - Non-exhaustive list of examples
 - Many others: Stochastic processes, time to event analysis, PKPD ...

Background



Jitao David Zhang^{1,2}, Lisa Sach-Peltason¹, Christian Kramer¹, Ken Wang¹ and Martin Ebeling¹

¹Pharma Early Research and Development, Roche Innovation Center Basel, F. Hoffmann-La Roche, Grenzachstrasse 124, 4070 Basel, Switzerland

²University of Basel, Department of Mathematics and Computer Science, Spiegelgasse 1, 4051 Basel, Switzerland

- Present along the multiscale modelling path

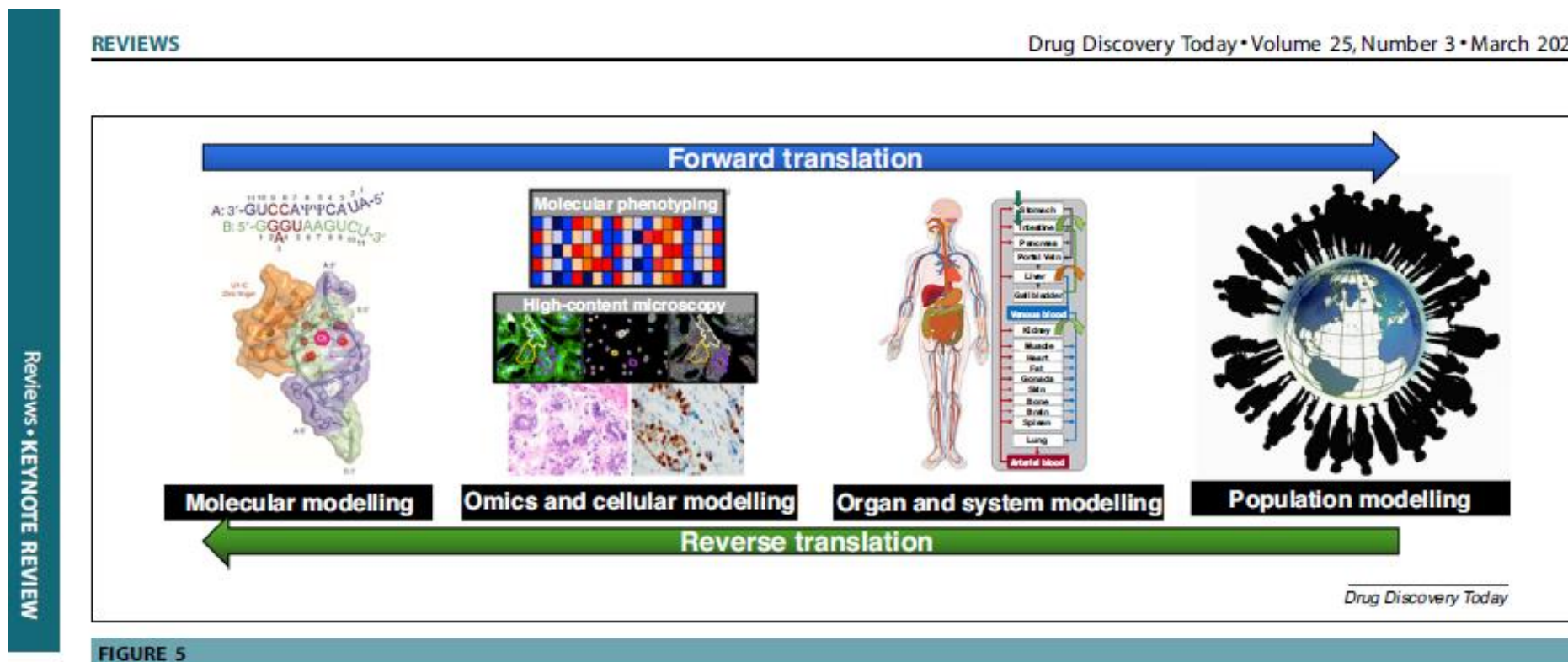
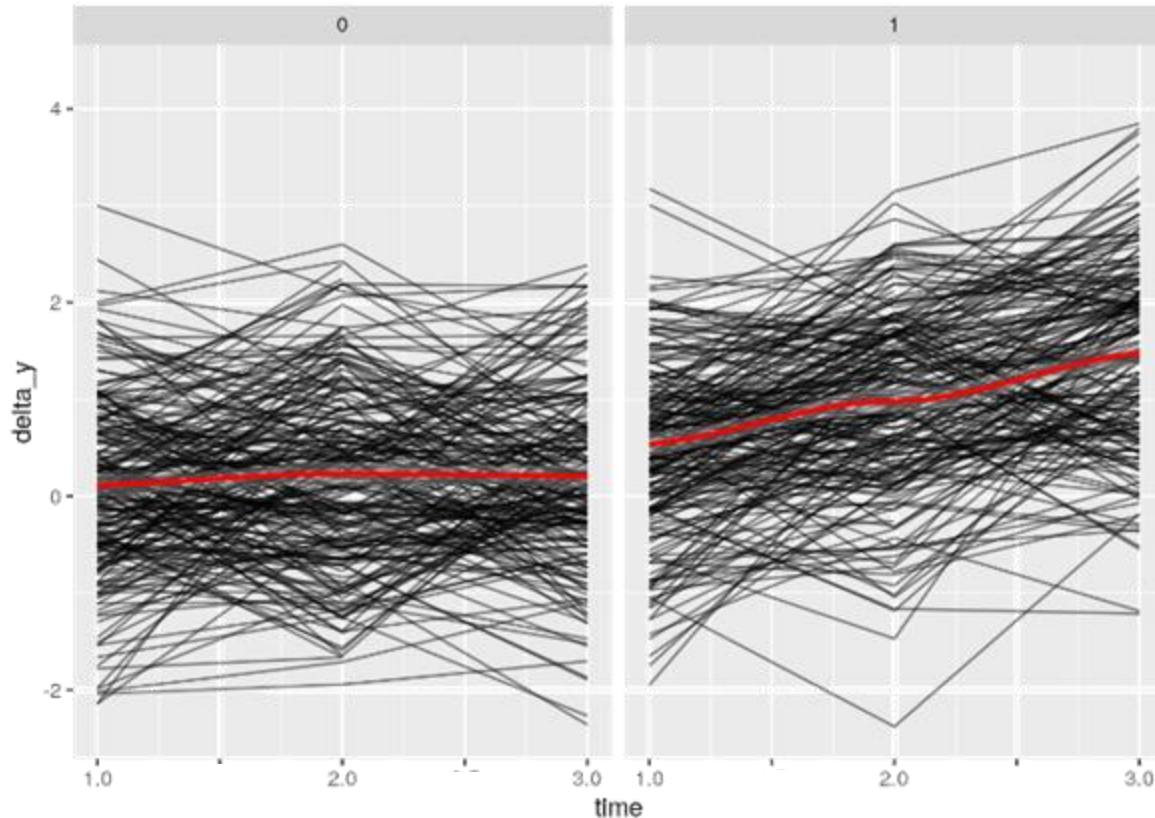


FIGURE 5

Background – Time series data analysis

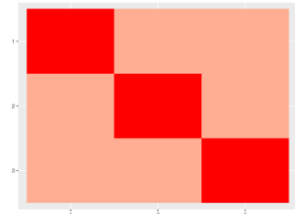
- Many of the data that we encounter include a temporal component and are inherently part of a dynamic system
 - Disease progression and drug response: Readout from patients over time → Mixed effects Model for Repeated Measurements (time is treated as ordinal categorical variable)
- Simulated change from baseline



- Examples of within-subject covariance structures (between 3 time points)

Compound Symmetry

$$\begin{bmatrix} \sigma^2 + \sigma_1^2 & \sigma_1^2 & \sigma_1^2 \\ \sigma_1^2 & \sigma^2 + \sigma_1^2 & \sigma_1^2 \\ \sigma_1^2 & \sigma_1^2 & \sigma^2 + \sigma_1^2 \end{bmatrix}$$



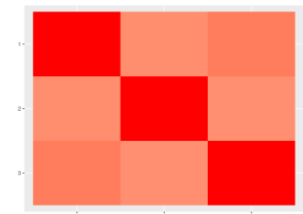
Autoregressive

$$\sigma^2 \begin{bmatrix} 1 & \rho & \rho^2 \\ \rho & 1 & \rho \\ \rho^2 & \rho & 1 \end{bmatrix}$$



Unstructured

$$\begin{bmatrix} \sigma_1^2 & \sigma_{12} & \sigma_{13} \\ \sigma_{12} & \sigma_2^2 & \sigma_{23} \\ \sigma_{13} & \sigma_{23} & \sigma_3^2 \end{bmatrix}$$



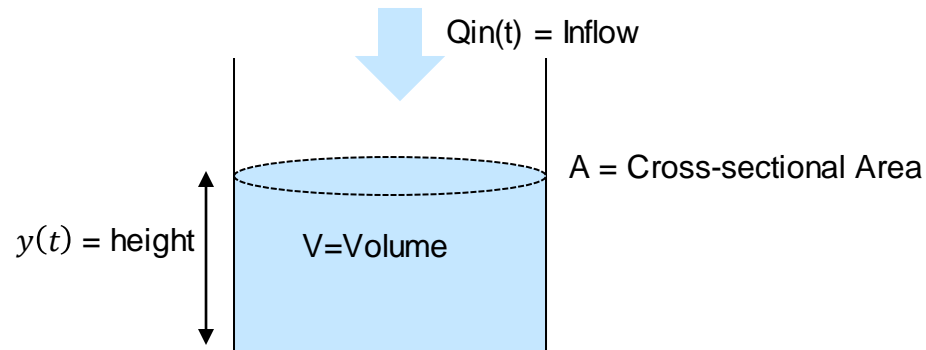
Many others ...

Background-MMRM Model for clinical drug response

- Data can be noisy, sparse (low sampling rate) and missing
- Ignoring or misspecification of the covariance structure can lead to incorrect/inflated statistical significance of the fixed effects (time, treatment and time:treatment interaction)

Background – Dynamic Systems: Illustrative example

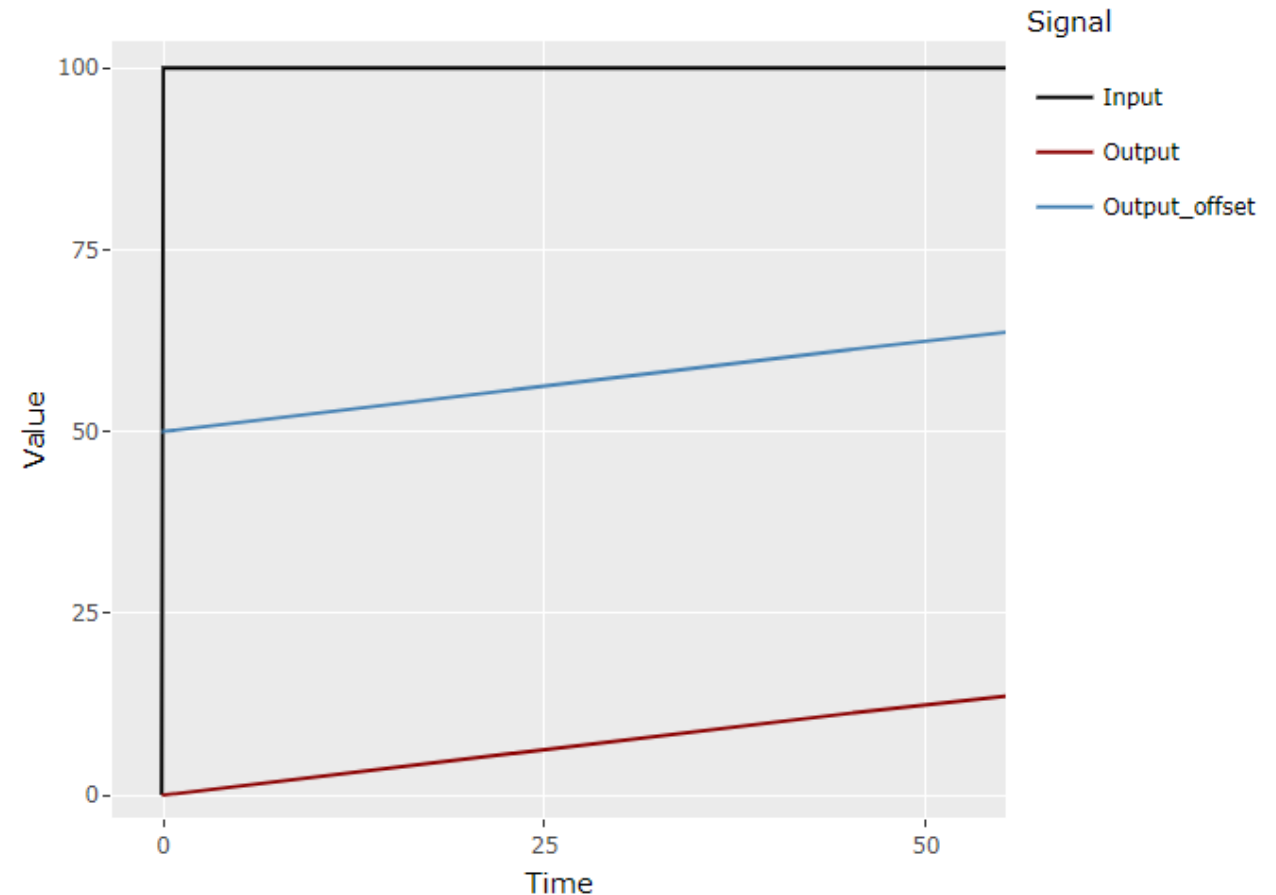
- Filling a water bucket



- Rate of change of volume = inflow rate

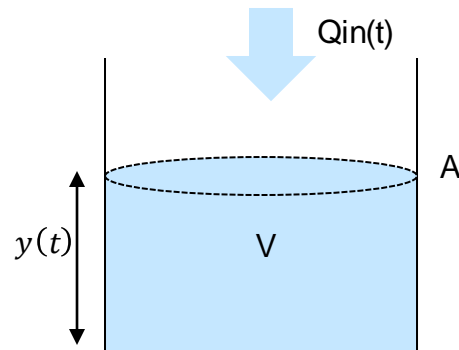
$$\frac{d(V(t))}{dt} = \frac{d(Ay(t))}{dt} = A \frac{d(y(t))}{dt} = Q_{in}(t)$$

$$y(t) = y_{t0} + \frac{G}{A} \int_0^t u(\tau) d\tau = y_{t0} + \frac{G}{A} t$$



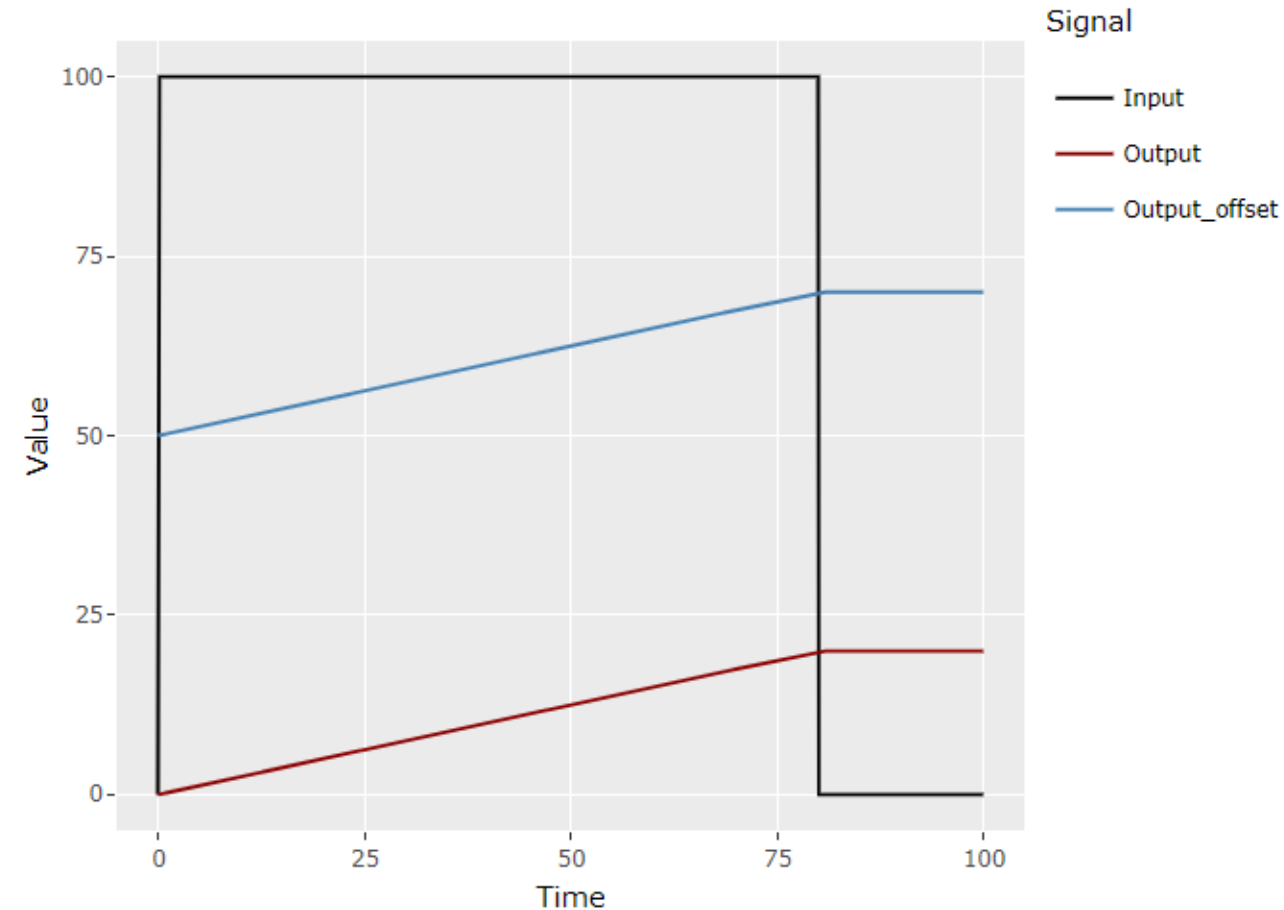
Background – Dynamic Systems

- Filling a water bucket (off switch)



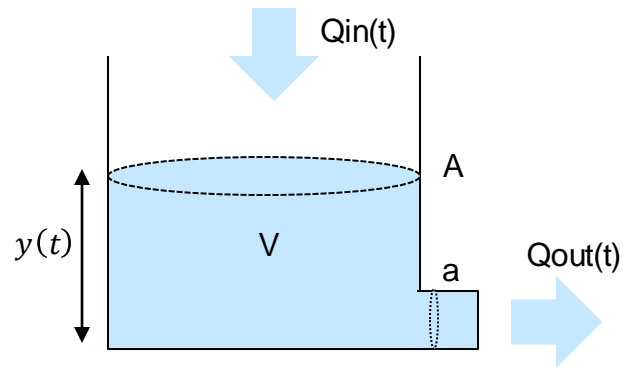
- $\frac{d(V(t))}{dt} = \frac{d(Ay(t))}{dt} = A \frac{d(y(t))}{dt} = Q_{in}(t)$
- $y(t) = y_{t0} + \frac{G}{A} \int_0^t u(\tau) - u(\tau - t_{off}) d\tau$
- $y(t) = y_{t0} + \frac{G}{A} (\int_0^{t_{off}} u(\tau) - u(\tau - t_{off}) d\tau + \int_{t_{off}}^t u(\tau) - u(\tau - t_{off}) d\tau)$
- $y(t) = \begin{cases} t < t_{off}: y_{t0} + \frac{G}{A} t \\ t \geq t_{off}: y_{t0} + \frac{G}{A} t_{off} \end{cases}$

- Constant rise (accumulation, no steady state) until tap switched off
- Not accounting for spills, overflow and evaporation!



Background – Dynamic Systems

- Leaky bucket



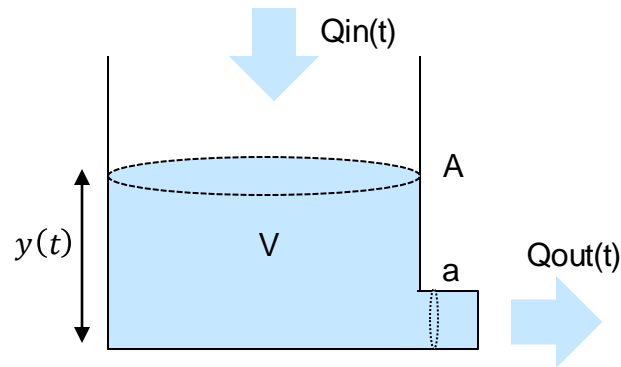
- Rate of change of volume = inflow rate – outflow rate

- $$A \frac{d(y(t))}{dt} = Q_{in} - Q_{out} = Q_{in} - f(y(t)) = Q_{in} - a\sqrt{2g} y(t)$$

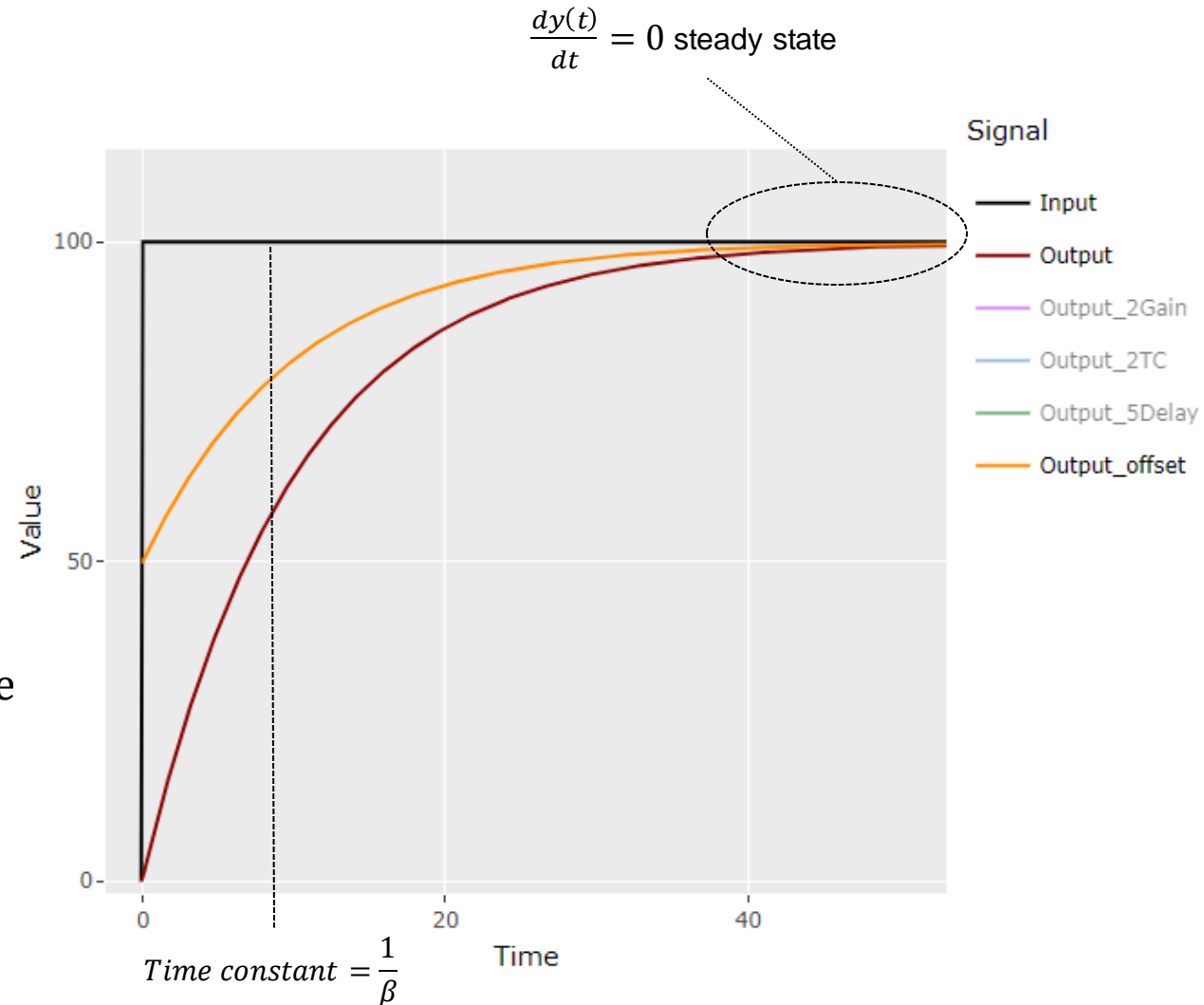
↙
Simplification for context relevance

Background – Dynamic Systems

- Leaky bucket

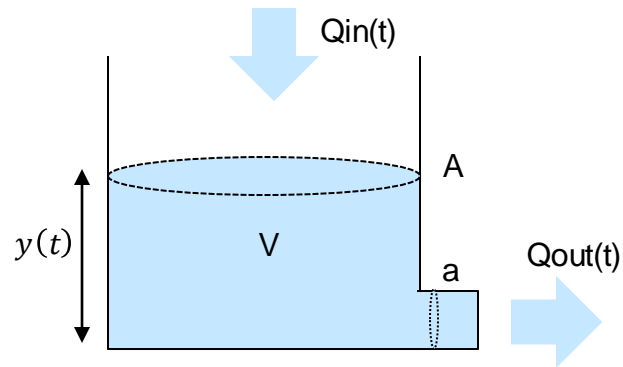


- Rate of change of volume = inflow rate – outflow rate
- $A \frac{d(y(t))}{dt} = Q_{in} - Q_{out} = Q_{in} - f(y(t)) = Q_{in} - a\sqrt{2g} y(t)$
- $\frac{d(y(t))}{dt} = \frac{G}{A} u(t) - \frac{a\sqrt{2g}}{A} y(t) = \alpha - \beta y(t)$
- $y(t) = y_0 e^{-\beta t} + \frac{\alpha}{\beta} (1 - e^{-\beta t})$

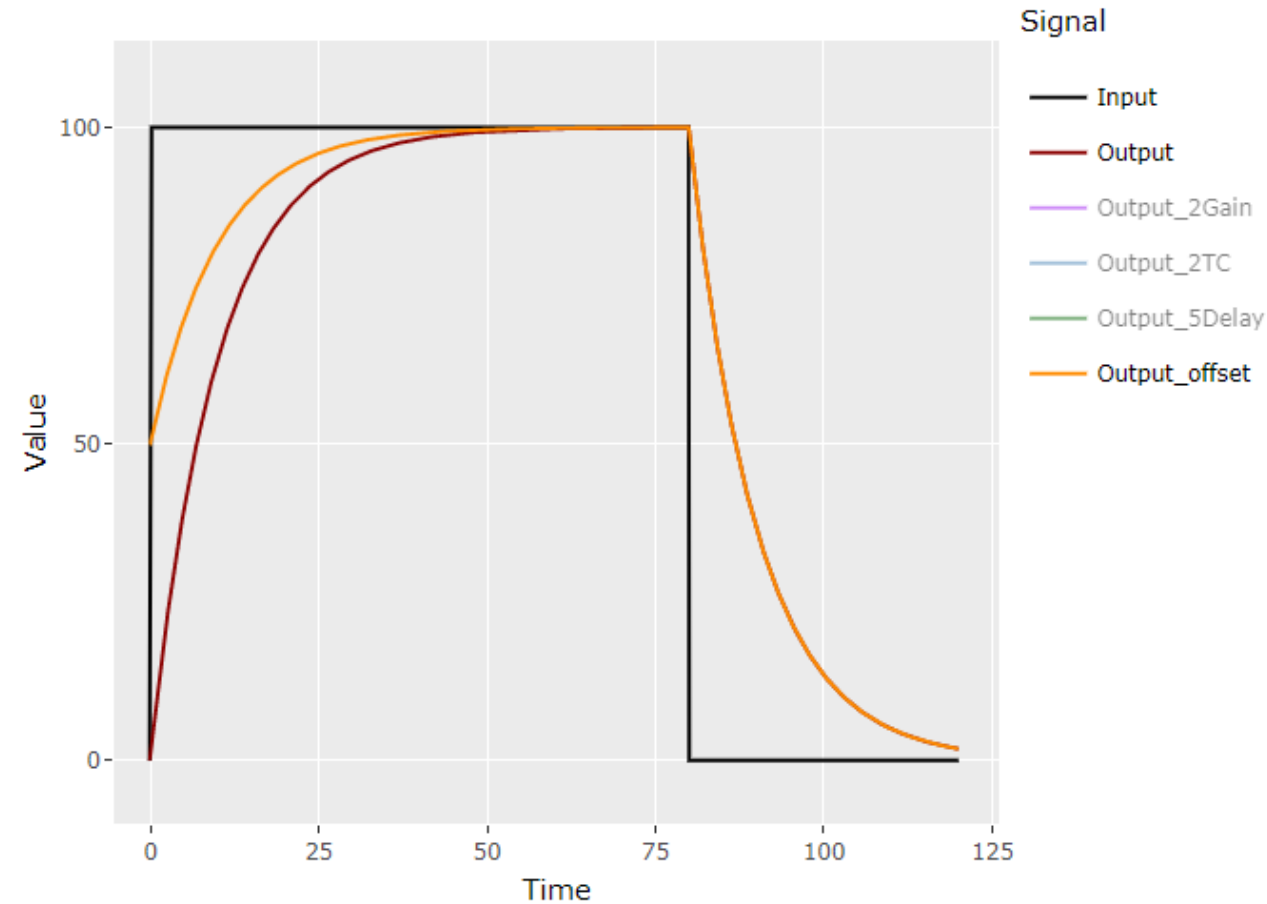


Background – Dynamic Systems

- Leaky bucket (with off switch)

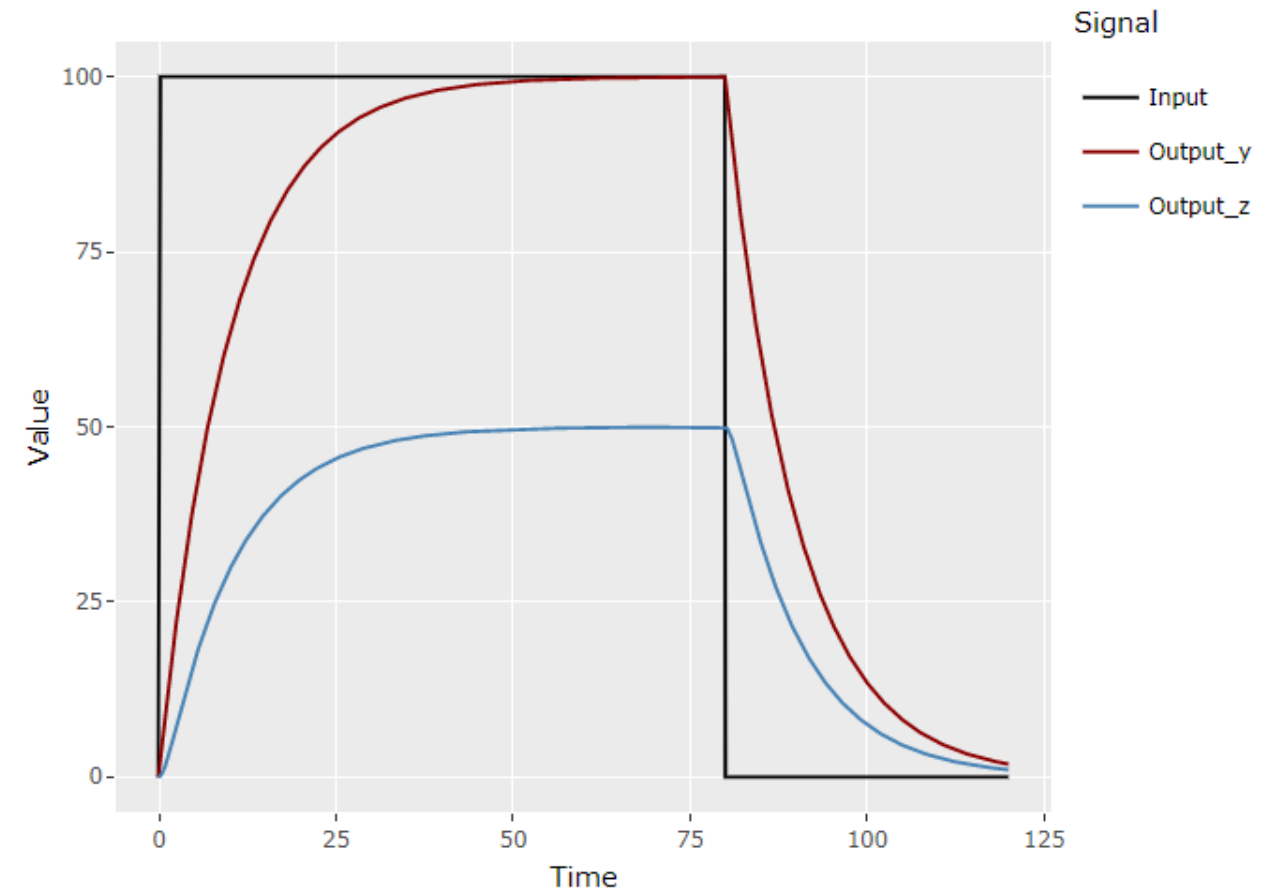
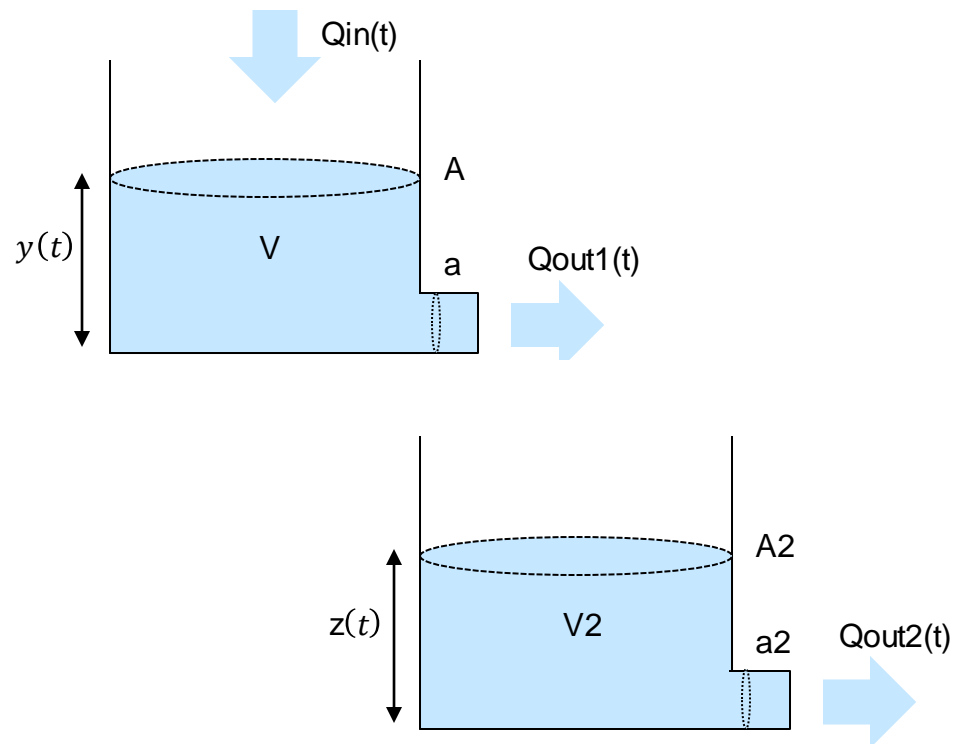


- $A \frac{d(y(t))}{dt} = Q_{in} - Q_{out} = Q_{in} - f(y(t)) = Q_{in} - a\sqrt{2g} y(t)$
- $\frac{d(y(t))}{dt} = \frac{G}{A} (u(t) - u(t - t_{off})) - \beta y(t)$
- $$y(t) = \begin{cases} t < t_{off} : y_0 e^{-\beta t} + \frac{\alpha}{\beta} (1 - e^{-\beta t}) \\ t \geq t_{off} : \{y_0 e^{-\beta t_{off}} + \frac{\alpha}{\beta} (1 - e^{-\beta t_{off}})\} e^{-\beta(t-t_{off})} \end{cases}$$



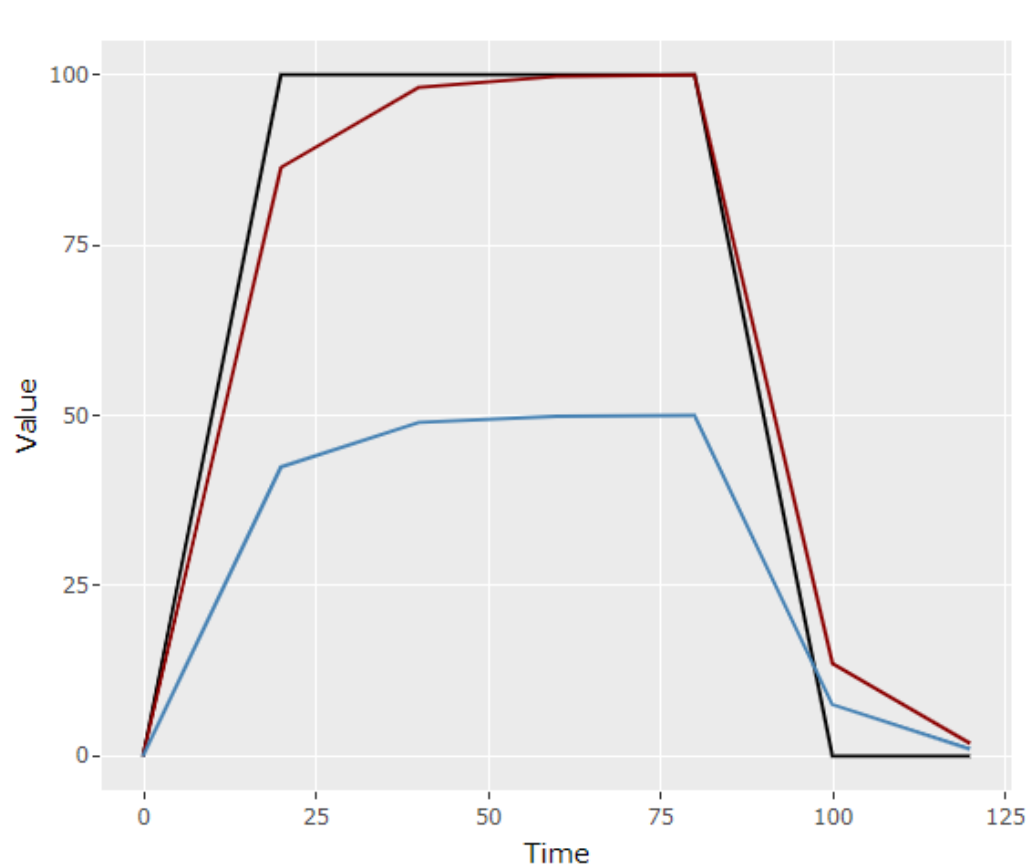
Background – Dynamic Systems

- Leaky buckets

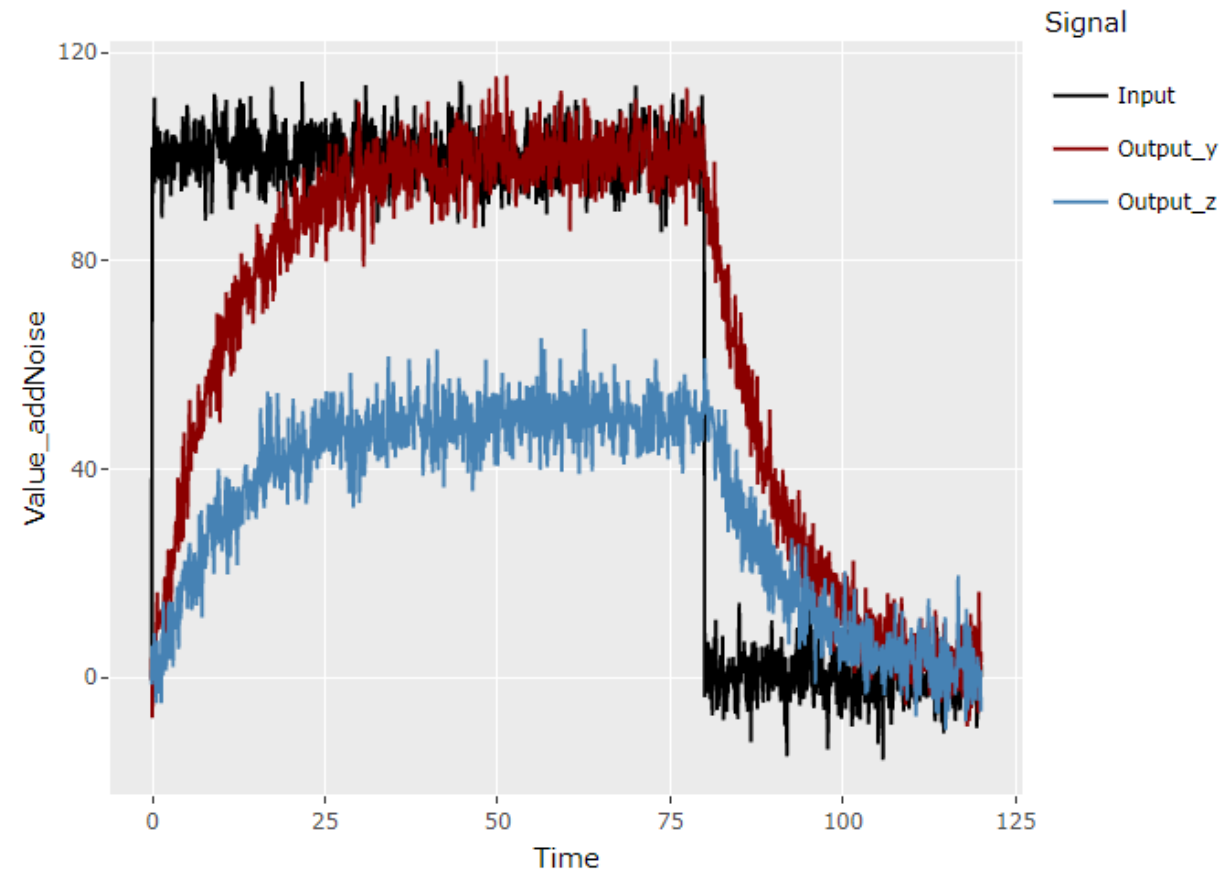


Background – Dynamic Systems - Complications

- Sparsity (Downsampling)

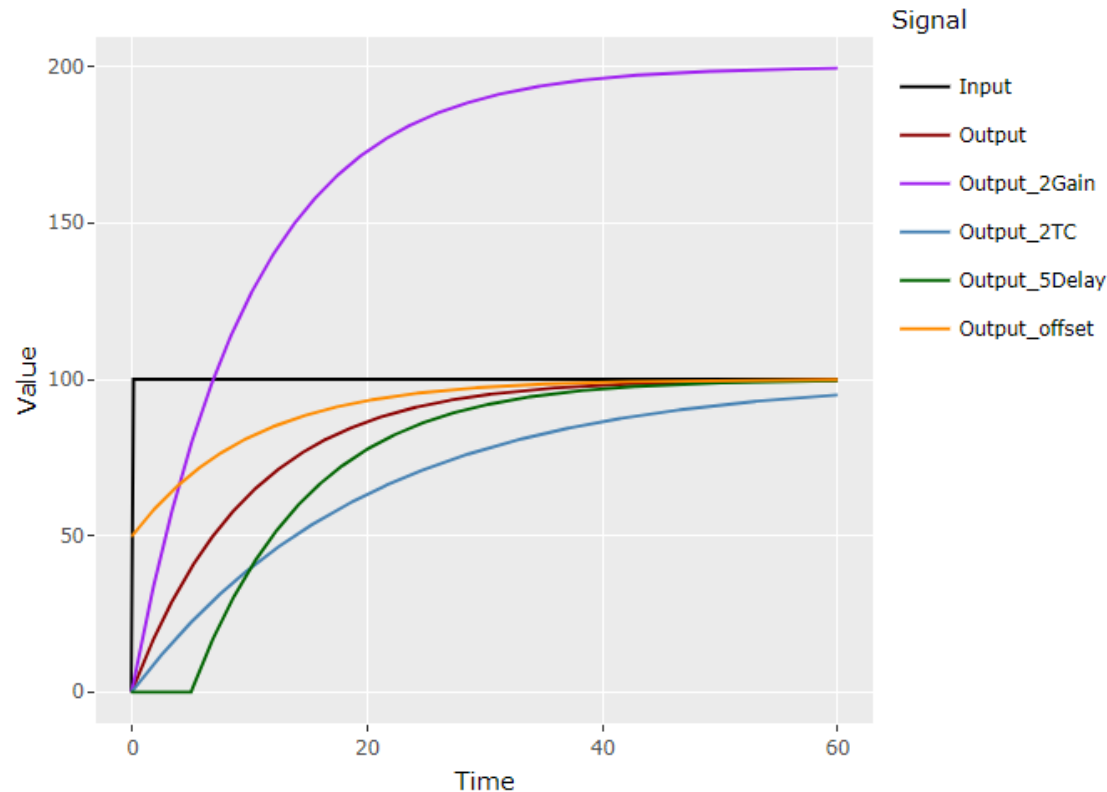


- Noisy (Additive Noise)

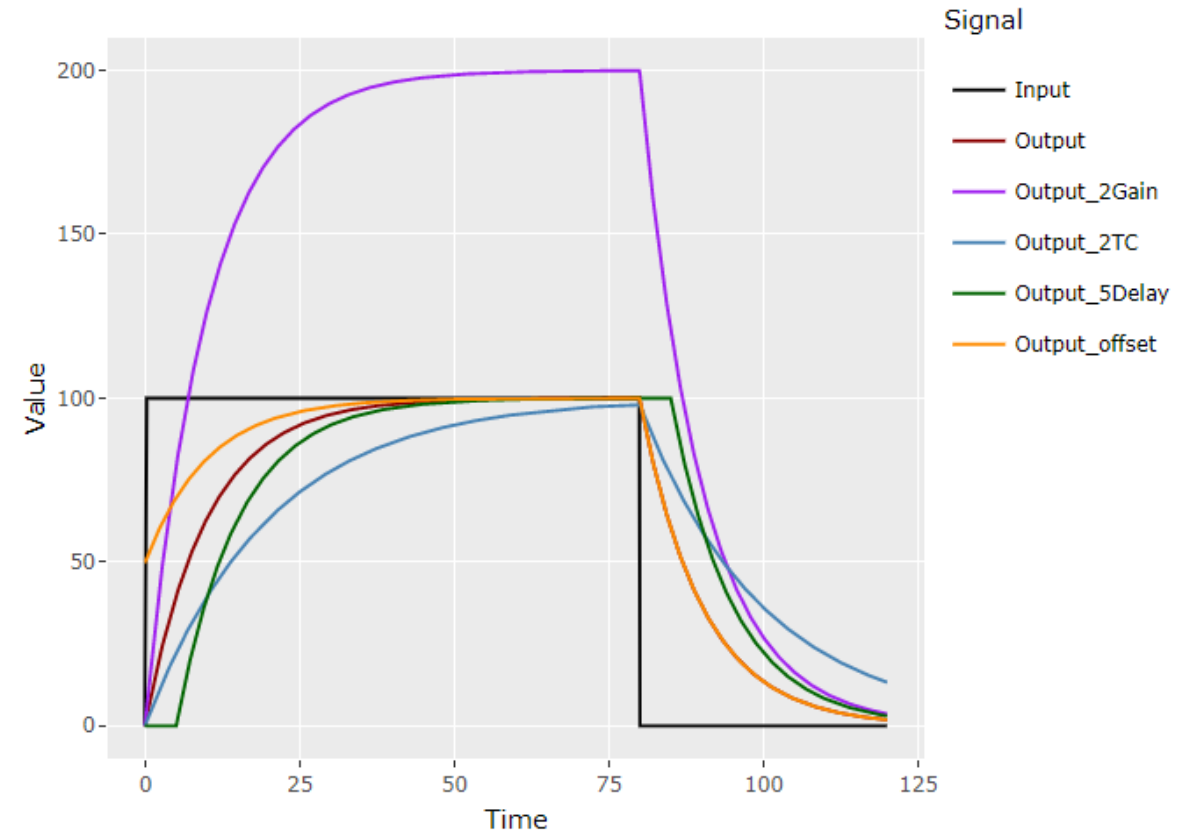


Background – Dynamic Systems: Exploring different Parameters

- Step function responses



- Square function responses

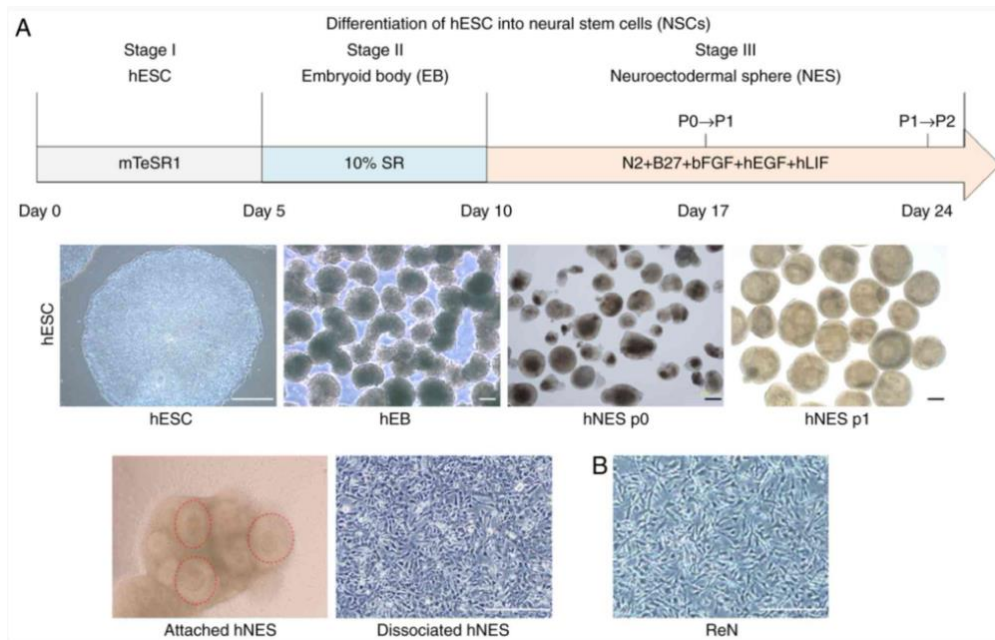


Full Report:

https://github.com/tkamth/Timeresponse/blob/main/UniBasel_DynamicSystem.html

RNA Velocity

- scRNASeq is just a time snapshot but it can capture cells in different states (e.g. proliferation, differentiation)
- Can we infer “cell states” based on transcriptomic profiles?
 - Similar to morphological differences observed from a single histology slide?

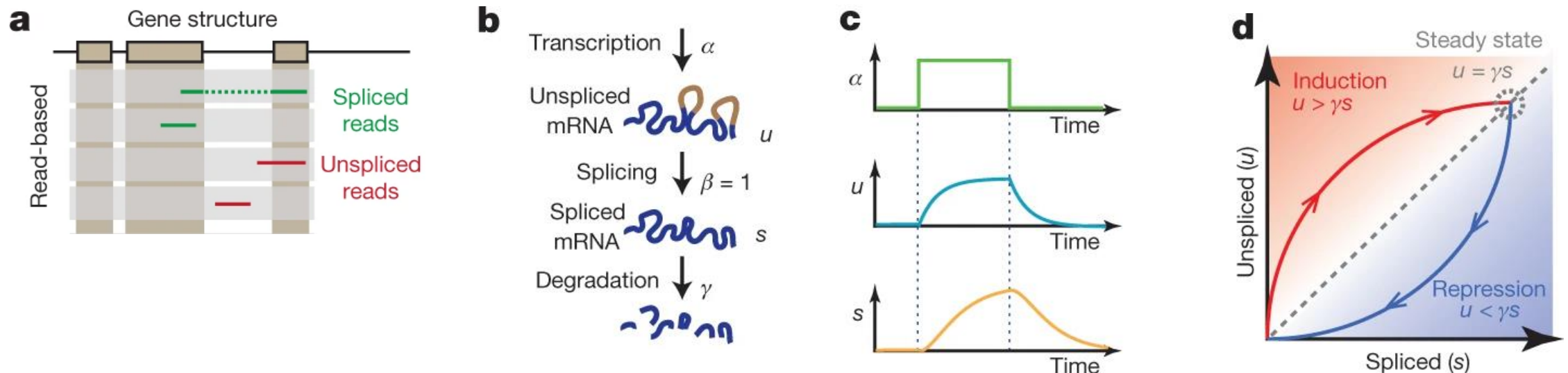


RNA velocity of single cells

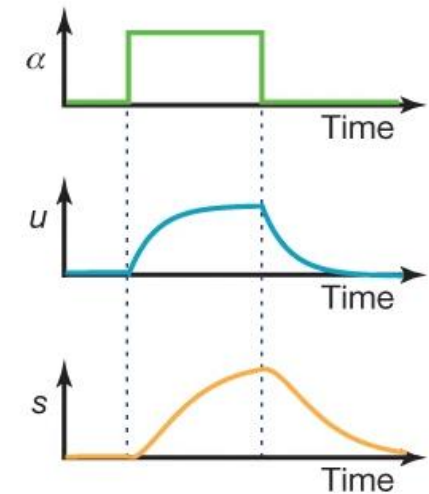
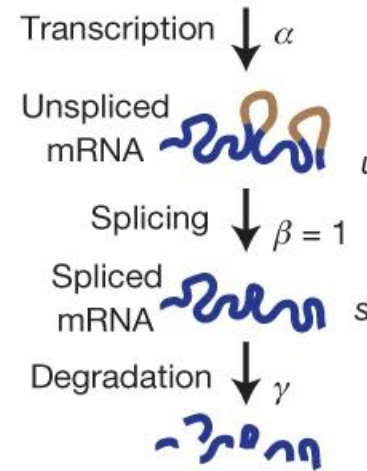
Gioele La Manno^{1,2}, Ruslan Soldatov³, Amit Zeisel^{1,2}, Emelie Braun^{1,2}, Hannah Hochgerner^{1,2}, Viktor Petukhov^{3,4}, Katja Lidschreiber⁵, Maria E. Kastriti⁶, Peter Lönnerberg^{1,2}, Alessandro Furlan¹, Jean Fan³, Lars E. Borm^{1,2}, Zehua Liu³, David van Bruggen¹, Jimin Guo³, Xiaoling He⁷, Roger Barker⁷, Erik Sundström⁸, Gonçalo Castelo-Branco¹, Patrick Cramer^{5,9}, Igor Adameyko⁶, Sten Linnarsson^{1,2*} & Peter V. Kharchenko^{3,10*}

La Manno et al. Nature 2018

- Velocyto
 - Analysis of expression dynamics in scRNASeq data.
 - Enables estimations of RNA velocities of single cells by distinguishing spliced and unspliced mRNAs
 - Spliced: polyA selection ignores noncoding RNA
 - Unspliced: artifact due to internal priming of intronic sequences



RNA Velocity



- Model transcription as a square wave (On/Off)
- Transcriptional induction for a gene \rightarrow increase of newly transcribed precursor unspliced mRNAs (until steady state)
- Repression/absence of transcription for a gene \rightarrow decrease of unspliced mRNAs

RNA Velocity

- System of 1st order Ordinary Differential Equations per gene

$$\frac{du}{dt} = \alpha(t) - \beta(t) u(t)$$

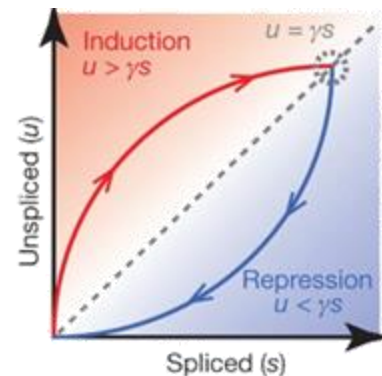
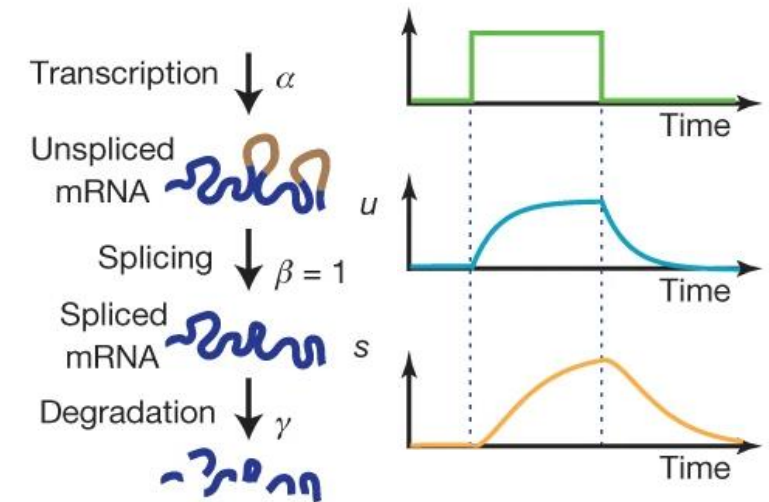
$$\frac{ds}{dt} = \beta(t) u(t) - \gamma(t) s(t)$$

- Assumptions
 - time invariant rates
 - common splicing rate across genes (normalizing/measuring rates by units of splicing rate by setting beta=1)

$$\frac{du}{dt} = \alpha - u(t)$$

$$\frac{ds}{dt} = u(t) - \gamma s(t)$$

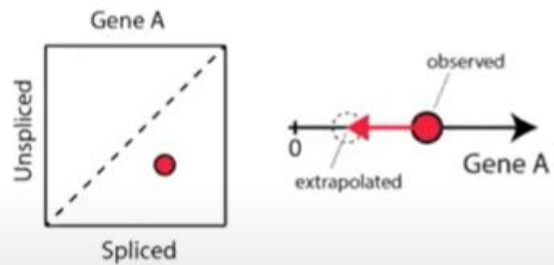
- Assuming data has cells in steady-state $ds/dt = 0$, $\longrightarrow \gamma = \frac{u}{s}$
- Velocity $v = u - \gamma s$ (vertical distance of the observed u from steady state slope)



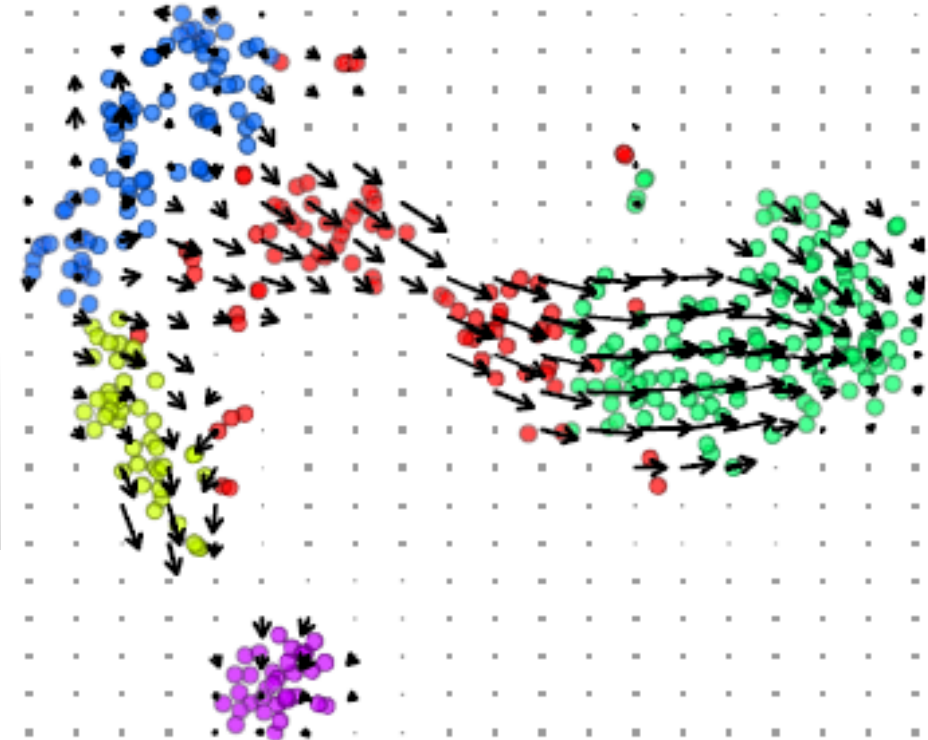
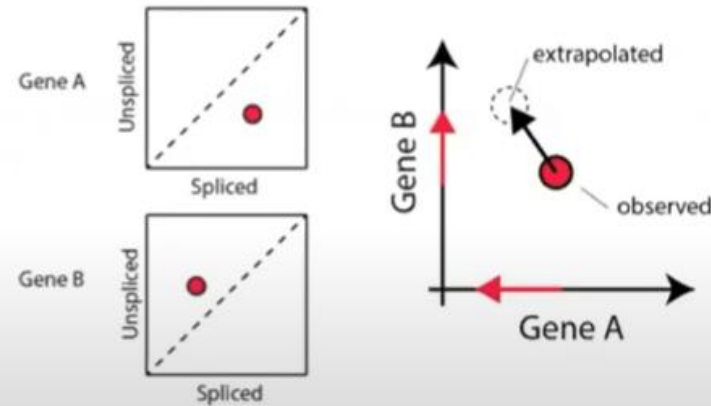
RNA Velocity

- Velocity (magnitude + direction) from single to multiple genes

Single gene

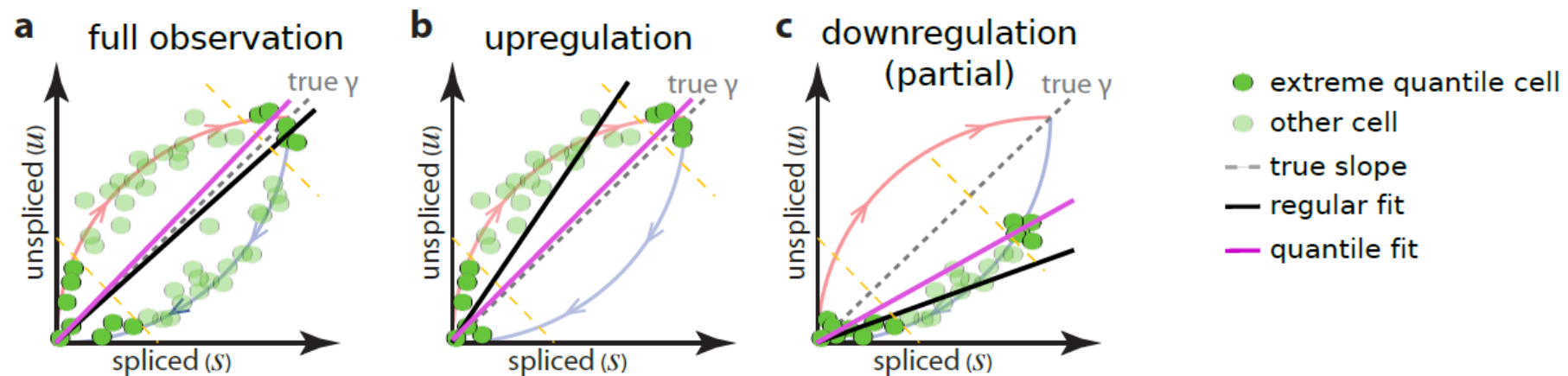


Many genes



RNA Velocity

Achieve more accurate estimation of gene-specific steady-state coefficient γ \rightarrow regression based on the cells found in the extreme quantiles of expression



Extension

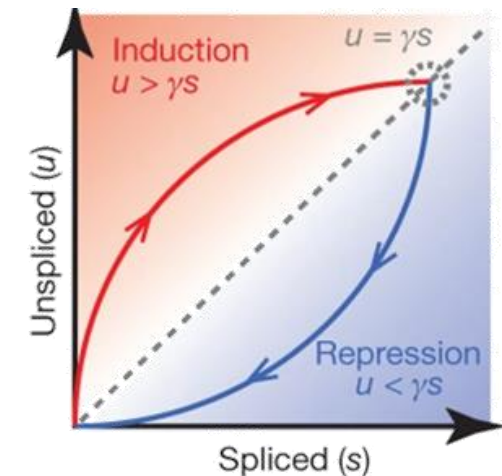
Generalizing RNA velocity to transient cell states through dynamical modeling

Volker Bergen^{1,2}, Marius Lange^{1,2}, Stefan Peidli², F. Alexander Wolf¹ and Fabian J. Theis^{1,2}

Bergen et al. Nature Biotech. 2020

scVelo: Generalizing RNA velocity to transient cell states through dynamical modeling

- Extension to estimate velocity without assuming either presence of steady states or common splicing rate across genes
- Maintains the weaker assumptions of constant gene-specific splicing
- Same transcriptional states are modeled to account for all possible configurations of gene activity
 - 2 steady states
 - 2 dynamic transient states (induction and repression)



Extension

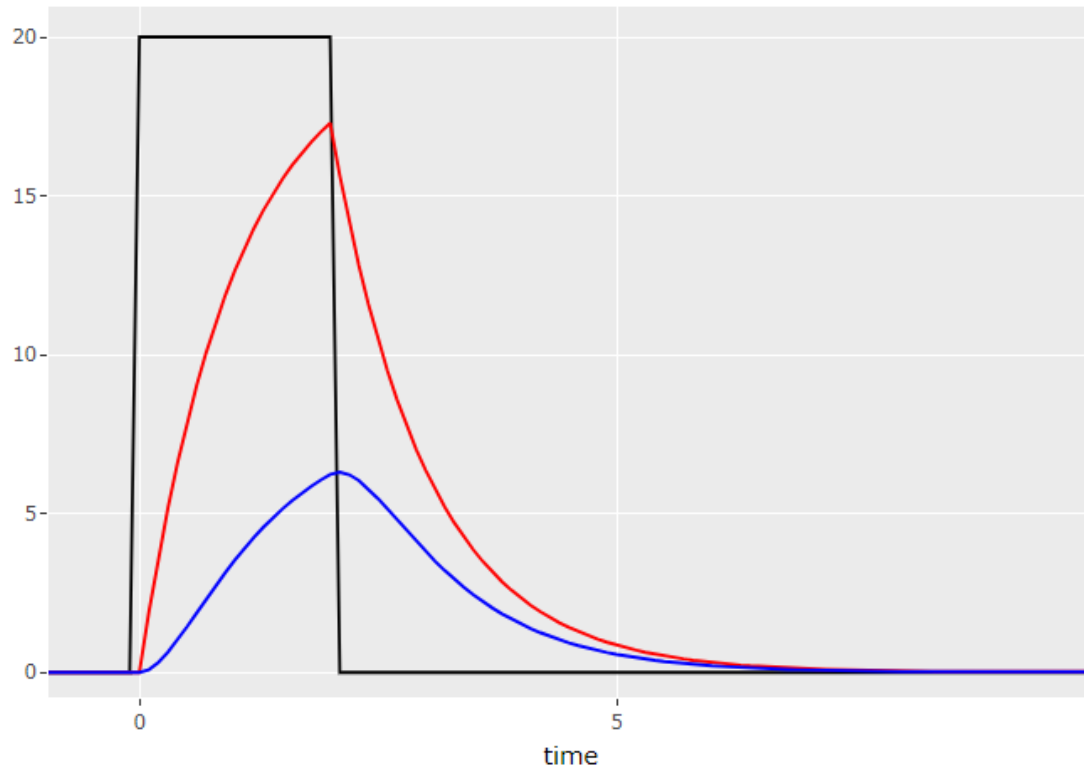
- Dynamic model
 - Define distinct states (induction and regression → either transcription is on or off)
 - Integrating the differential equations and setting $\tau = t - t_0^{(k)}$ (where k is the state when transcription is either on or off)

$$u(t) = u_0 e^{-\beta\tau} + \frac{\alpha^{(k)}}{\beta} (1 - e^{-\beta\tau}),$$

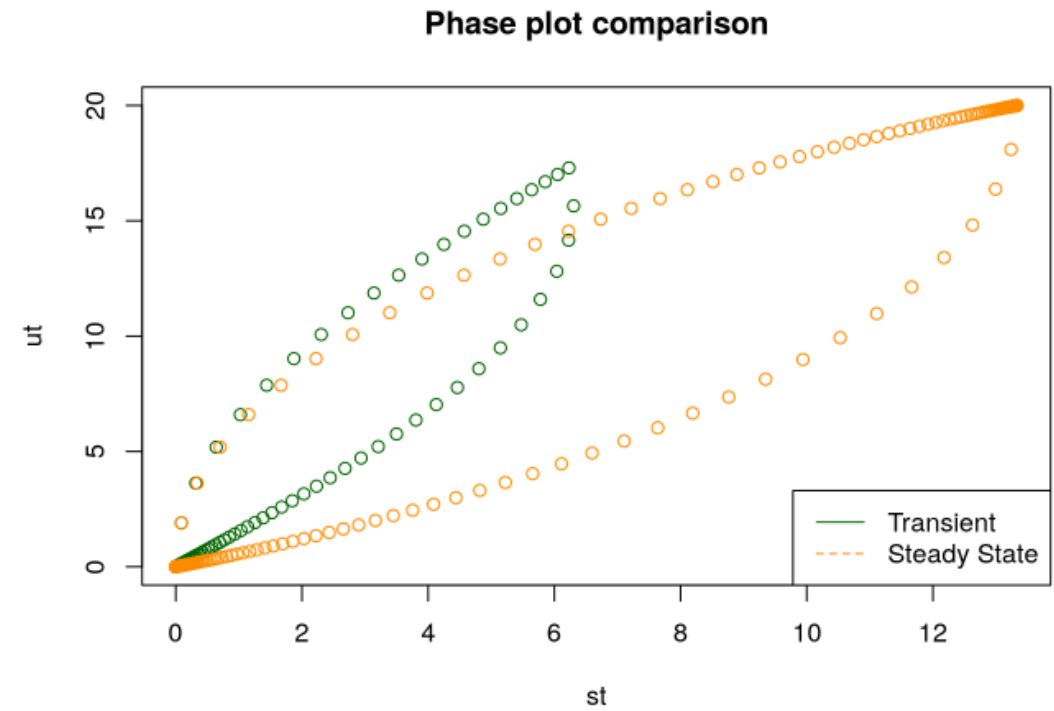
$$s(t) = s_0 e^{-\gamma\tau} + \frac{\alpha^{(k)}}{\gamma} (1 - e^{-\gamma\tau}) + \frac{\alpha^{(k)} - \beta u_0}{\gamma - \beta} (e^{-\gamma\tau} - e^{-\beta\tau})$$

- Two distinct set of steady states $(u_{\infty}^{(k)}, s_{\infty}^{(k)}) = \left(\frac{\alpha^{(k)}}{\beta}, \frac{\alpha^{(k)}}{\gamma}\right)$

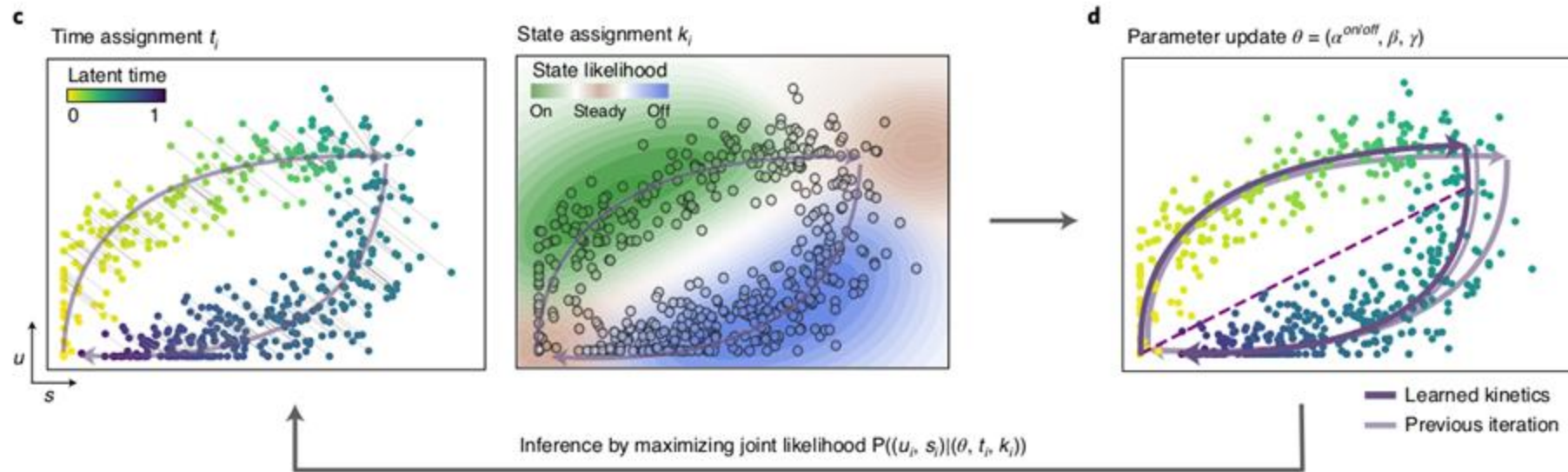
Steady state not reached



Phase plot



Extension-EM



Challenge: both time and rate parameters are unknown

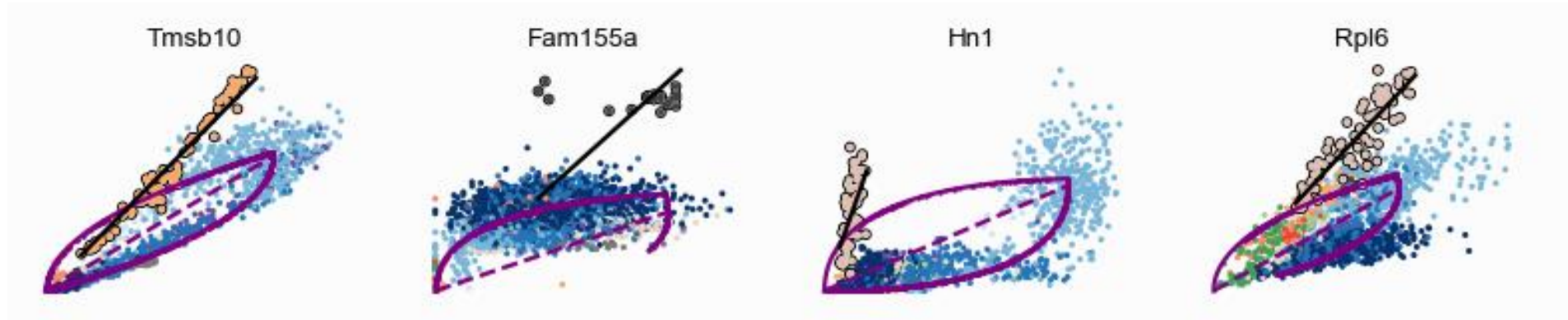
- E step: Assign a latent time to the observed value by minimizing the distance to the phase trajectory model
- M step: reaction rate parameters updated (updated to maximize the log-likelihood using downhill simplex method Nelder–Mead) → new trajectory phase

RNA Velocity

- Differential Kinetic Test
 - Distinct cell clusters may exhibit different kinetics regimes
 - Likelihood ratio (asymptotic chi-squared distribution) can be tested for significance.
 - Ratio of a single-kinetic and two-kinetic model (i.e.: supporting multiple clusters),

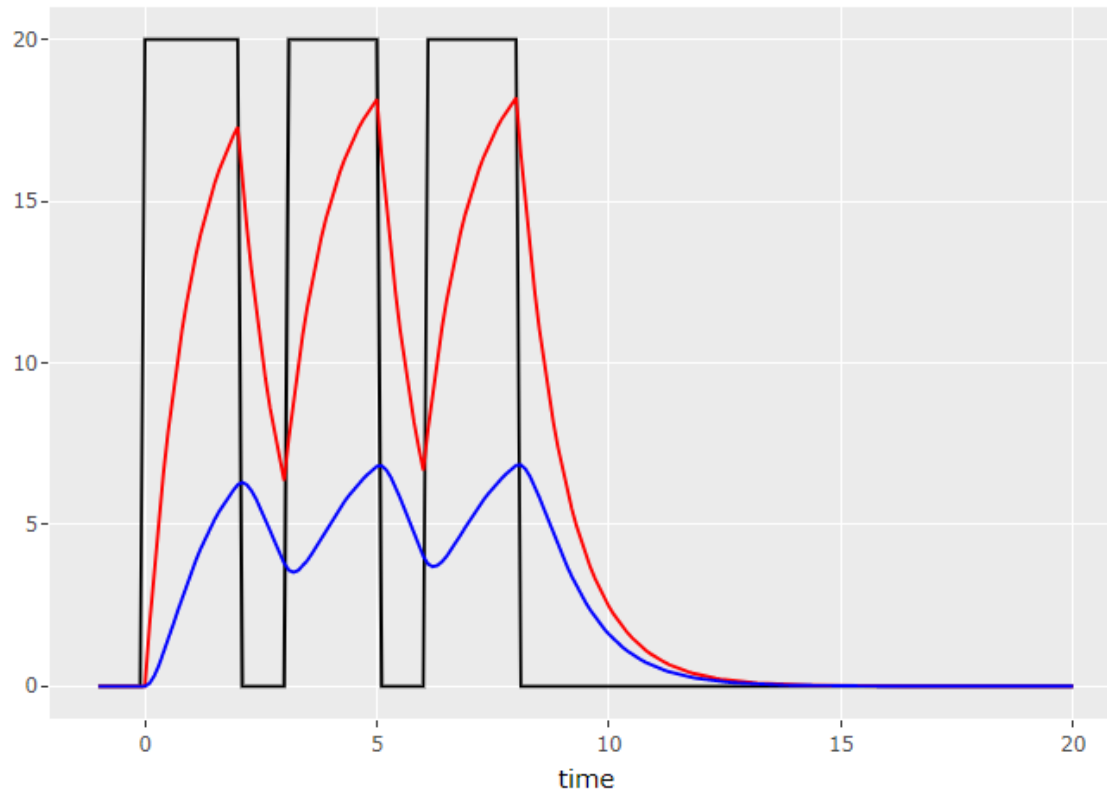
$$LR = -2 \ln \frac{\sup_{\theta} \mathcal{L}(\theta)}{\sup_{\theta, \theta'} \mathcal{L}(\theta, \theta')}$$

- Limitation: For computational reason → by default an orthogonal regression is used instead of a full phase trajectory to test whether a cluster is well explained by the overall kinetic or not

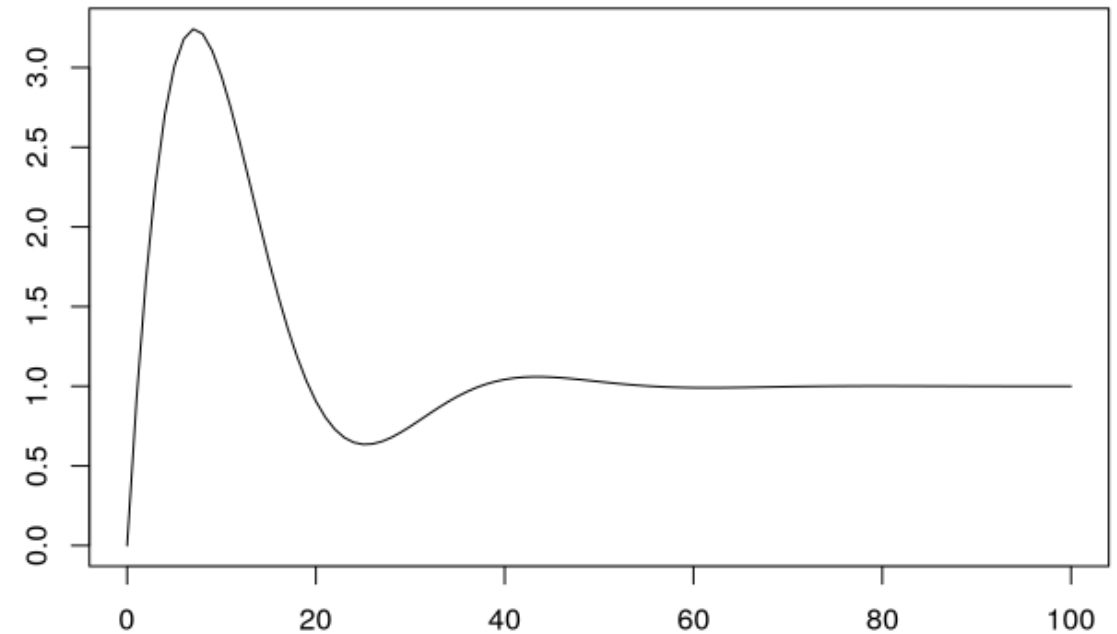


Further Extensions – Complicated inputs and models

Toggleing or more frequent switches of transcription input signal



Second order dynamic systems



Theory Summary-RNA Velocity

- Steady state / deterministic model (velocityto)
 - Approximated with a linear regression on the presumed steady states in the lower and upper quantiles.
 - 2 fundamental assumptions:
 - steady-state mRNA levels is captured in the data
 - a common splicing rate across genes
- Extension to a full Dynamic model (using a likelihood-based framework) (scvelo)
 - Solved by likelihood-based expectation-maximization framework

scVelo-Tutorial (Endocrine pancreas data)

- scRNA sequencing of pancreatic cells during embryonic development in mouse
- Elucidate how endocrine progenitors segregate into different endocrine subtypes during development
 - Ideal scenario with developmental data and multiple lineages
- Typically between 10%-25% of reads are unspliced molecules with intronic sequences

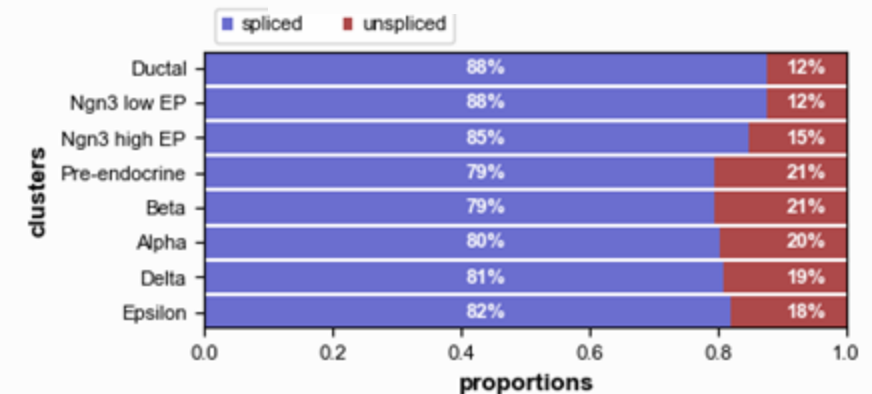
© 2019. Published by The Company of Biologists Ltd | Development (2019) 146, dev173849. doi:10.1242/dev.173849



RESEARCH ARTICLE

Comprehensive single cell mRNA profiling reveals a detailed roadmap for pancreatic endocrinogenesis

Aimée Bastidas-Ponce^{1,2,3,4,*}, Sophie Tritschler^{1,5,6,*}, Leander Dony^{5,6,7}, Katharina Scheibner^{1,2,3,4}, Marta Tarquis-Medina^{1,2,3,4}, Ciro Salinno^{1,2,3,4}, Silvia Schirge^{1,2,3}, Ingo Bartscher^{1,2,3}, Anika Böttcher^{1,2,3}, Fabian J. Theis^{5,8,†}, Heiko Lickert^{1,2,3,4,†} and Mostafa Bakhti^{1,2,3,†}

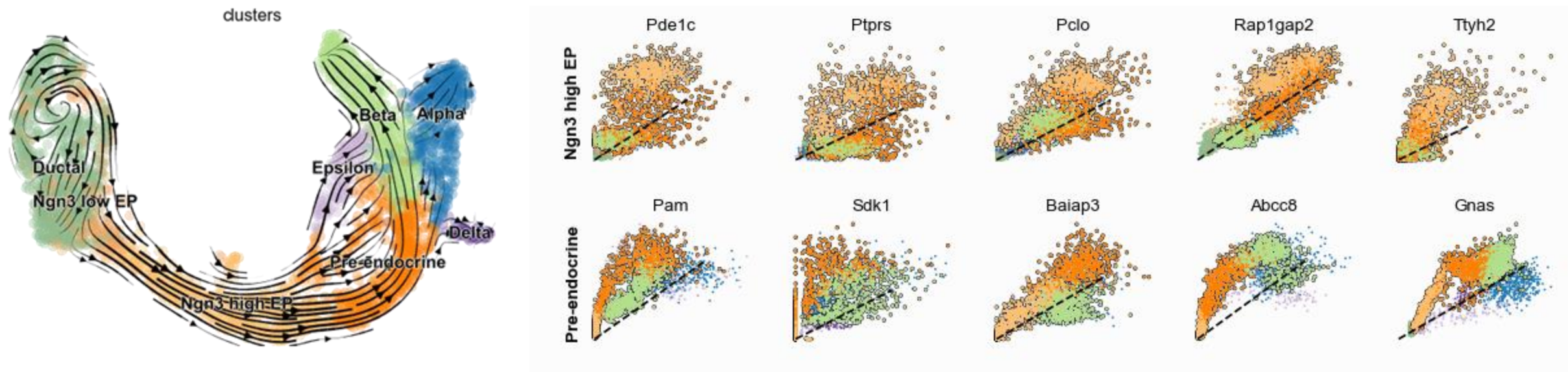


doi: 10.1242/dev.173849

<https://scvelo.readthedocs.io/>

scVelo-Tutorial (Endocrine pancreas data)

- Cluster specific differential velocity expression in two transient clusters
 - Neurogenin 3 high endocrine progenitor cells (yellow)
 - Pre-endocrine (orange)



Can we target these driver genes to alter the terminal cell states associated with healthy controls?

CellRank

New Results

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CellRank for directed single-cell fate mapping

Marius Lange, Volker Bergen, Michal Klein, Manu Setty, Bernhard Reuter, Mostafa Bakhti, Heiko Lickert, Meshal Ansari, Janine Schniering, Herbert B. Schiller, Dana Pe'er, Fabian J. Theis

doi: <https://doi.org/10.1101/2020.10.19.345983>

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Posted November 20, 2020.

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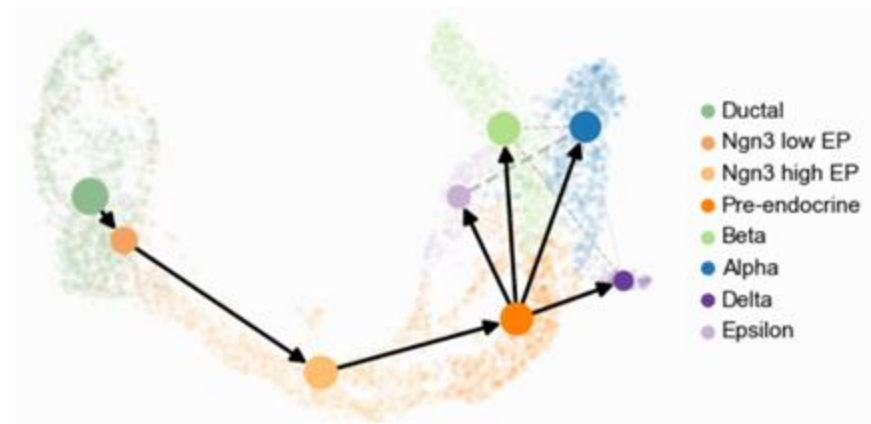
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Identifying initial and terminal states

- How likely each cell is to develop from each initial state(s) or towards each terminal state(s)
- Challenge: Interpretation of which genes are supporting the direction of the flow for that cell
 - biased of the umap projection to genes that are more highly abundant

Objective: Construct probabilistic fate maps

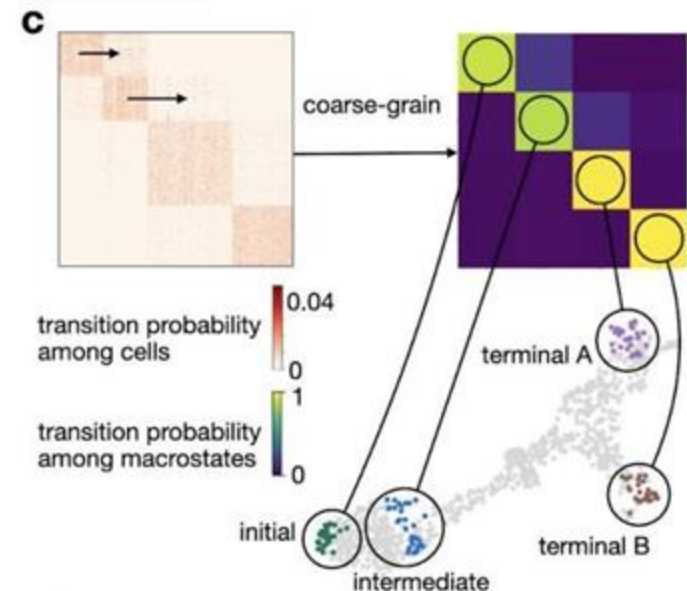
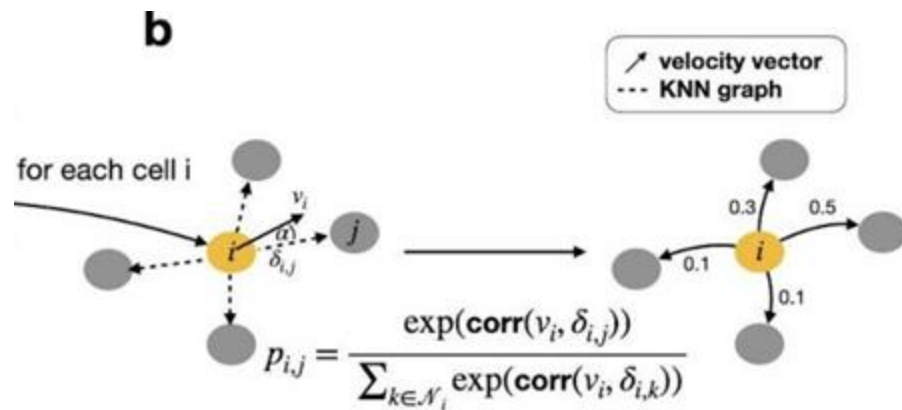


<https://doi.org/10.1101/2020.10.19.345983>

CellRank

Markov chain transition matrix

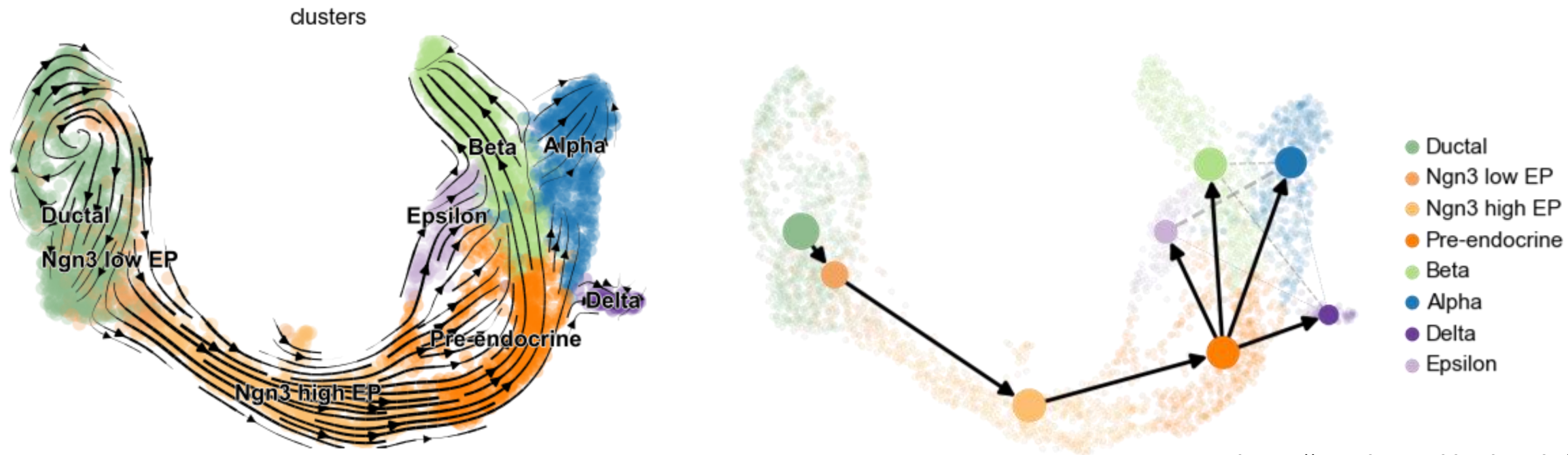
- Use each velocity vector to find likely cell transitions that are accordance with that direction
- Transition probabilities are computed using dot product projection between the potential cell-to-cell transitions and the velocity vector and stored in a matrix denoted as velocity graph.



scVelo-Tutorial (Endocrine pancreas data)

Developmental processes

- Delineates cycling population of ductal cells and endocrine progenitor cells
- Illuminates endocrine cell differentiation and lineage commitment



Conclusions

Key Messages

- Time series data are fun, powerful and omnipresent
- Data can be sparse and noisy → careful modeling has be factored in
- RNASeq velocity & trajectory analyses provide novel insight into genes important for lineage transitions (*continued effort and assessment by many scRNASeq squad colleagues*)
- Many challenges to model the data correctly while reflecting the true biological mechanism that is often oversimplified
- Novel algorithms and solvers are needed to “integrate” multiscale modelling (computational time)
- Words of wisdom:
 - When you decide to embark on a project → go full speed and deep until you reach “steady state” before the system or/and input signal switch