

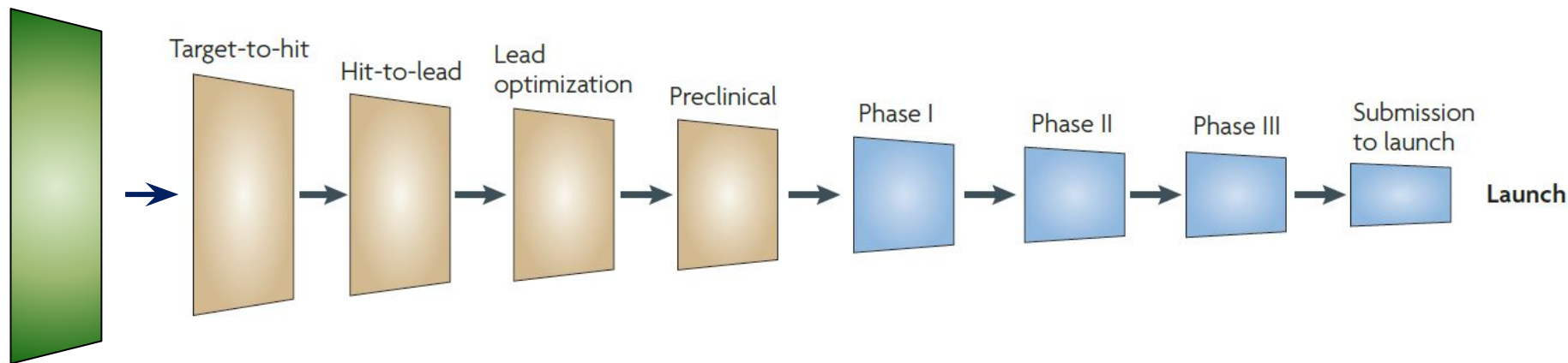
What can we do if there are no good targets

*Mathematical and Computational Biology in Drug Discovery
(MCBDD) Module II*

*Dr. Jitao David Zhang
March-April 2021*

The linear view of drug discovery builds on target-based approaches

Target identification & assessment

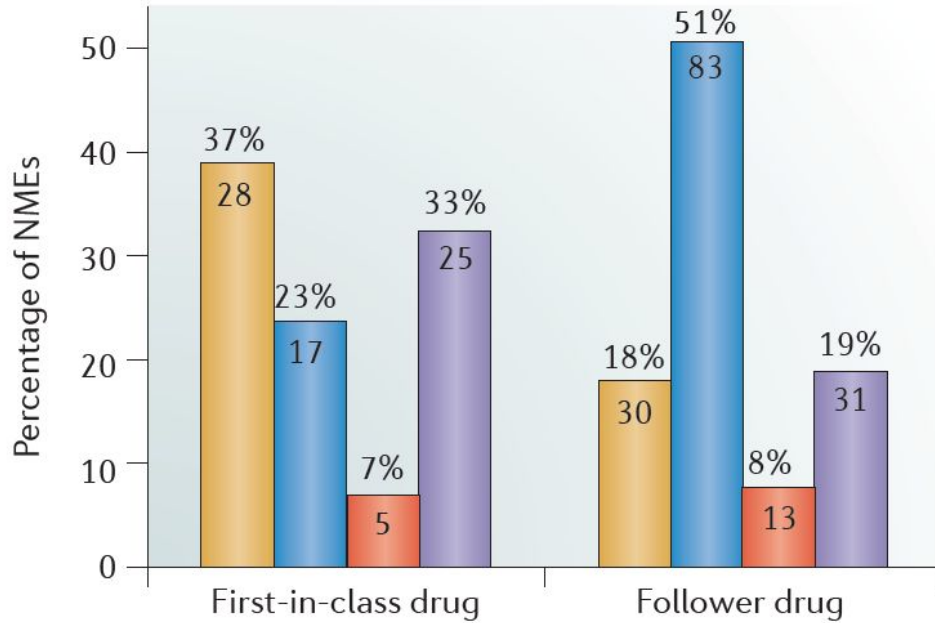
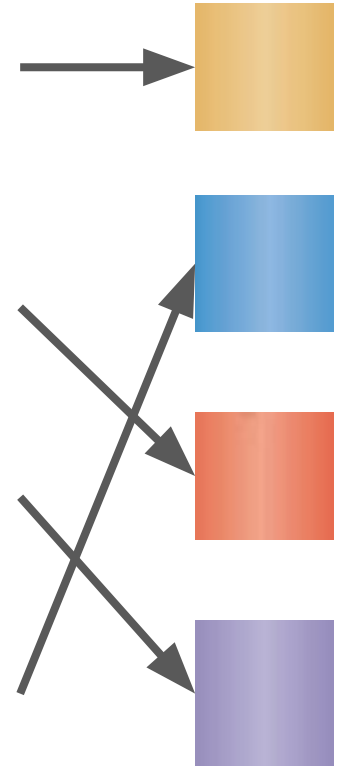


Five strategies when no good target is found

1. Phenotypic drug discovery
2. Natural products
3. Biologics
4. Interaction-based (multispecific) drug discovery
5. Drug repurposing or combination studies

Connect the lines!

- Phenotypic screening
- Modified natural products
- Biologics
- Target-based screening



Phenotypic screenings by agent and readout

Agent

High-throughput screening
libraries ($\geq 10^6$ molecules)

Genetic libraries ($\sim 10^4$)

Natural products and chemo-
genomic libraries ($\sim 10^3$)

Custom libraries ($\sim 10^0 - 10^2$)

Feasibility

Reporters

Gene
expression

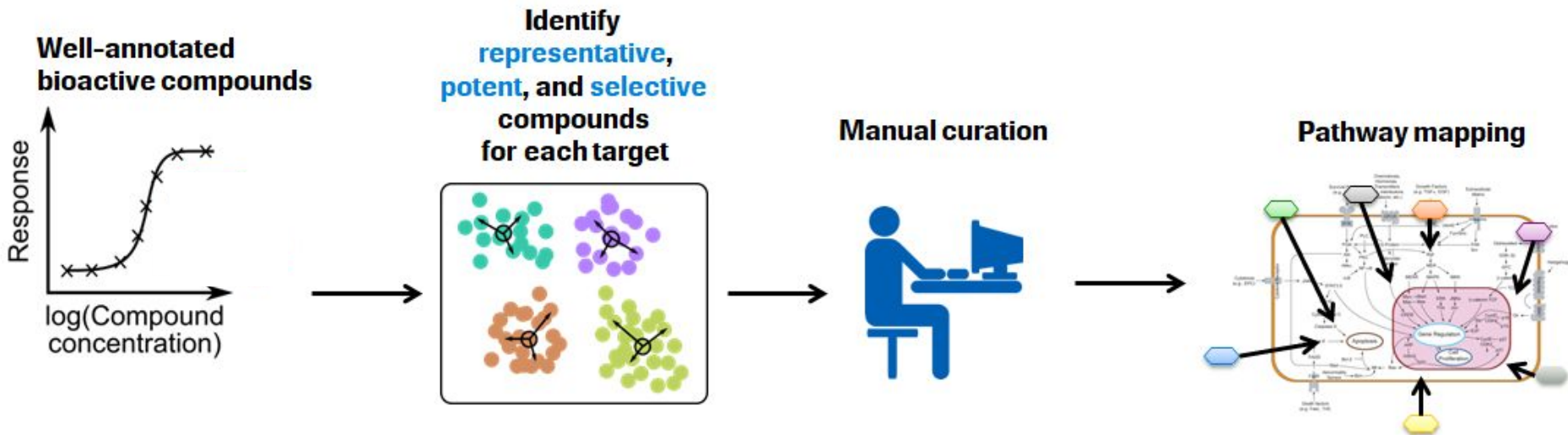
Cellular
morphology

Organ/tissue
phenotype

Organism
phenotype

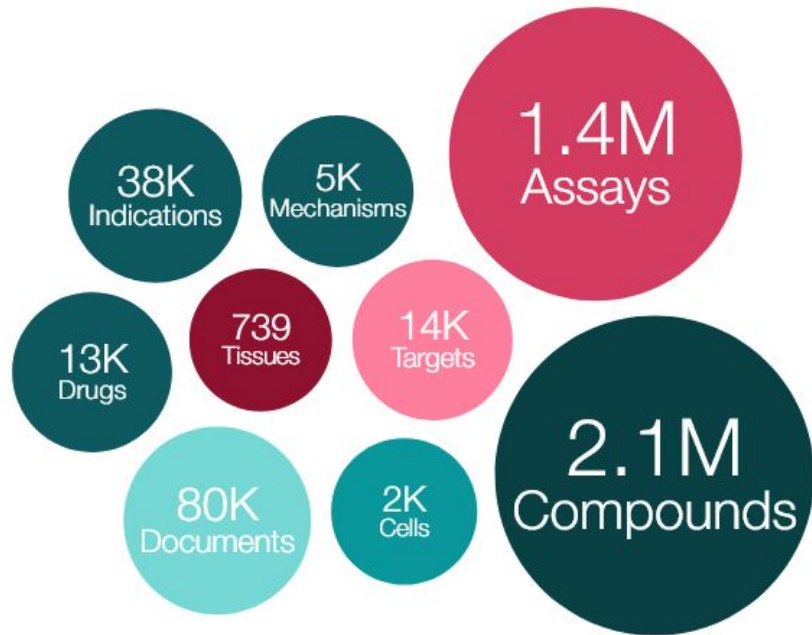
Readout

The Small-molecule Pathway Research Kit (SPARK)



The ChEMBL database

- An example of query: [aspirin](#).
- Systematic and programmatic accession via [ChEMBL API](#) ([source code](#)).
- We can use **dose-response data** to annotate the *triplets* of compound, assay activity, and targets.

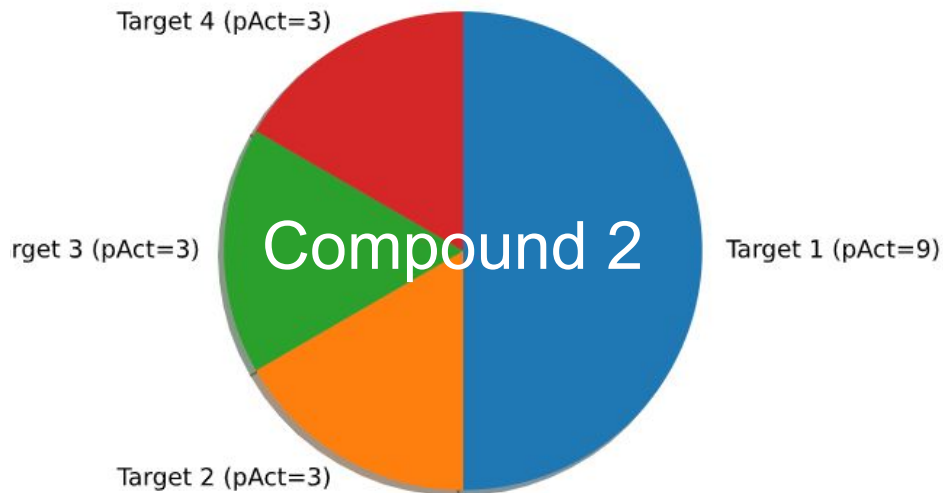
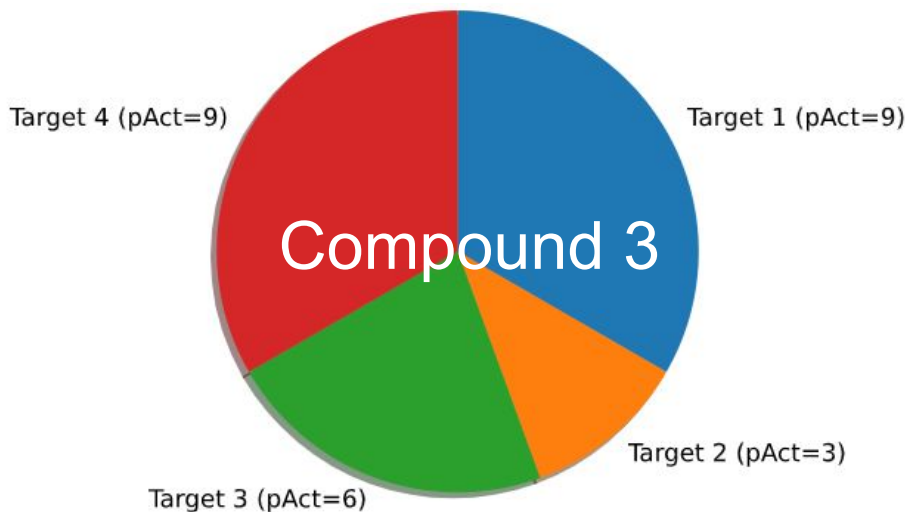
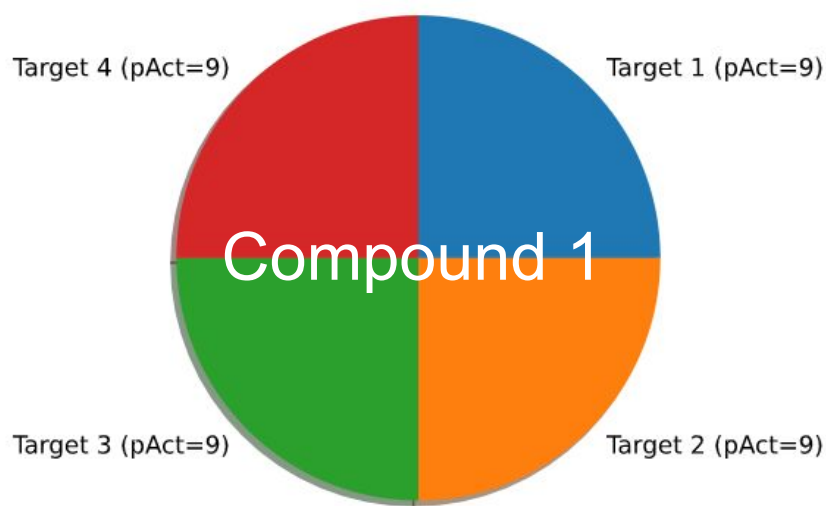


March 2021

Discussion

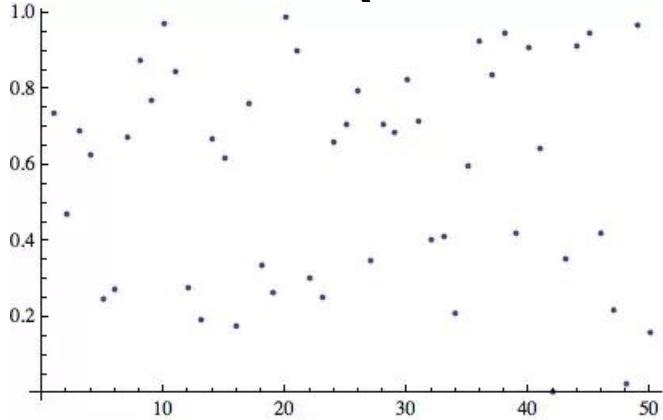
1. Why do we care selecting
representative, potent, and selective
compounds for each target?
2. How to define following terms
mathematically ...
 - a. Representativity?
 - b. Potency?
 - c. Selectivity?

A toy example about how to quantify a compound's potency and selectivity

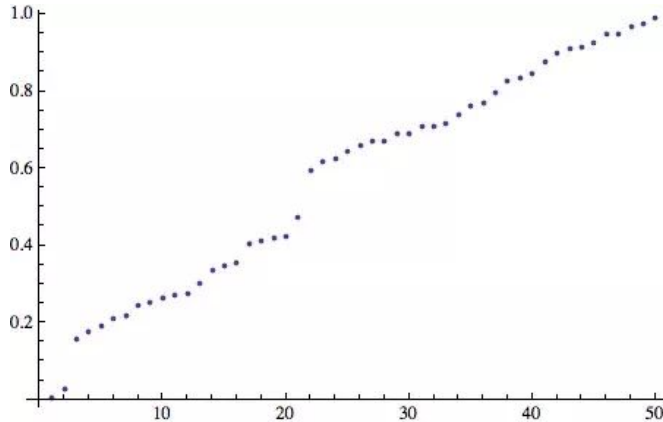


The Gini Index (a.k.a. Gini Coefficient)

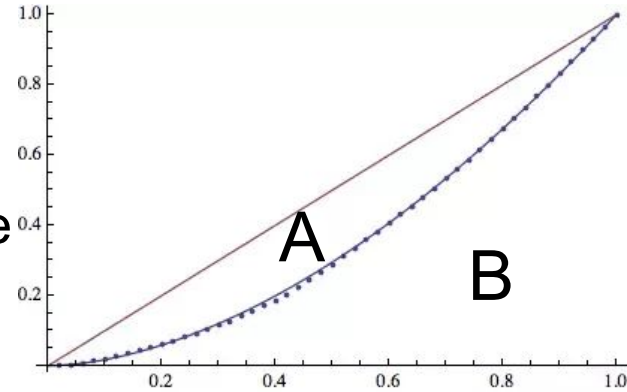
A random
vector of
50 values



Sorted
from low
to high



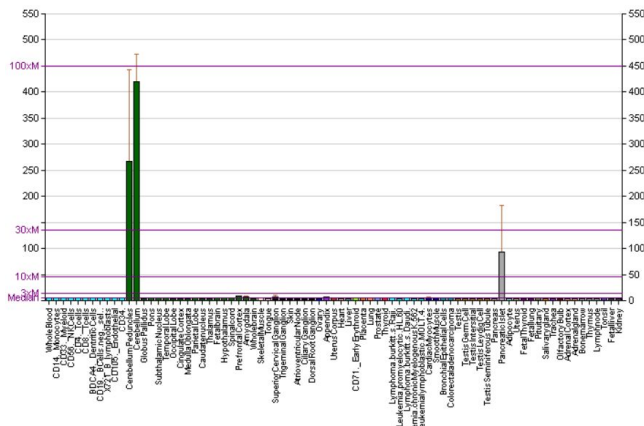
The Gini
Index is
calculated
based on the
cumulative
distribution



$$G = A / (A + B)$$

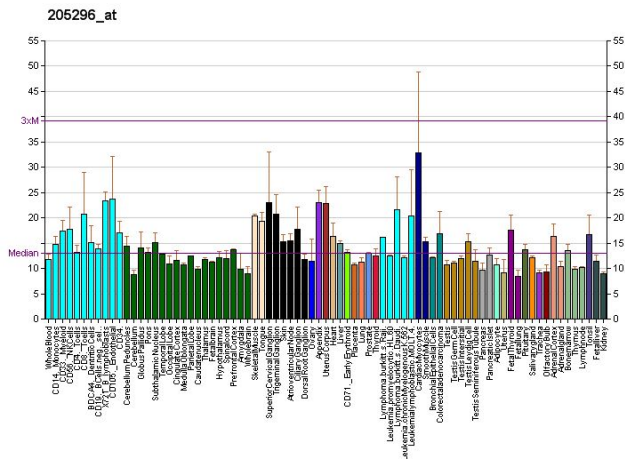
The Gini Index quantifies inequality/ selectivity

NEUROD1

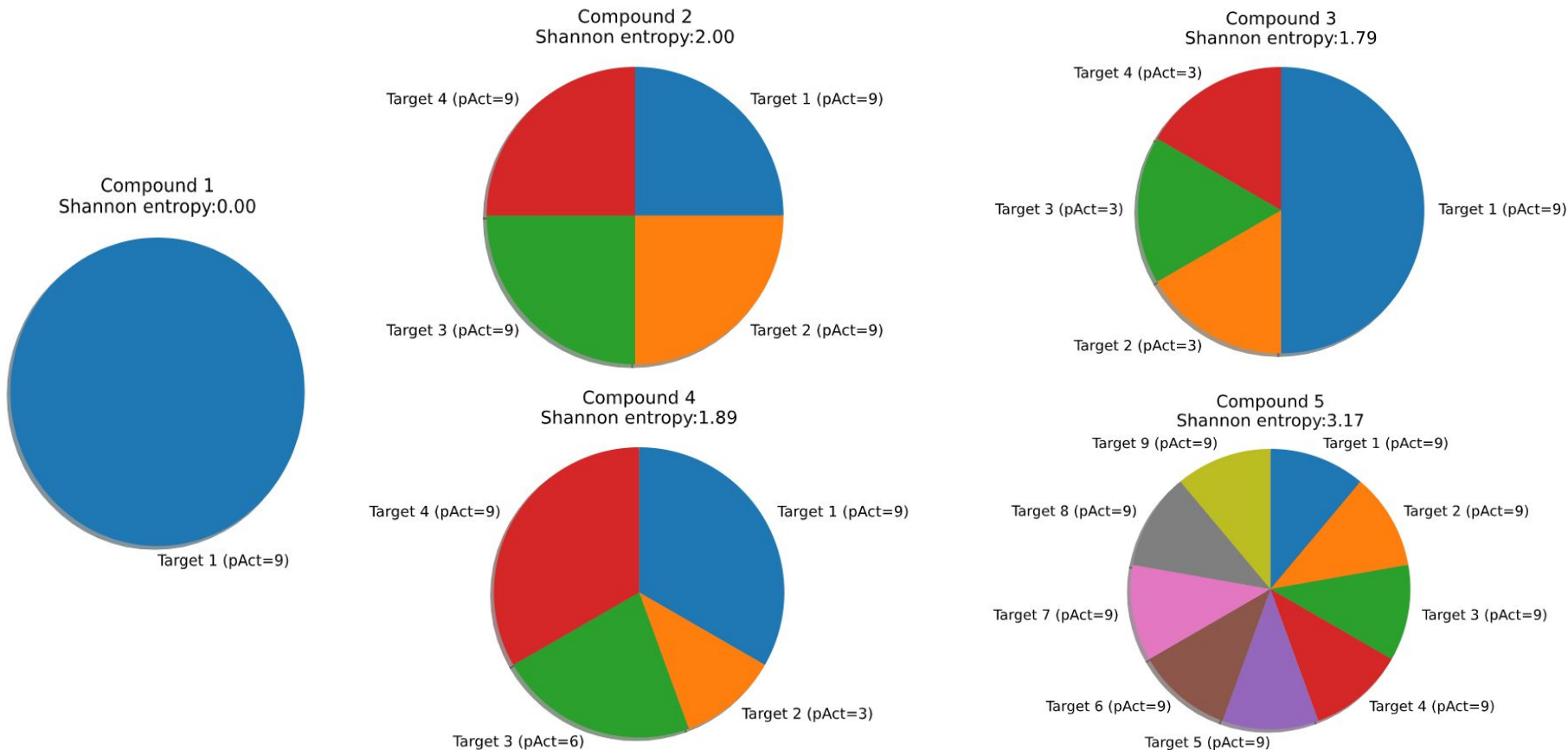


The Gini Index of expression of *NEUROD1* across tissues is near 1, whereas that of *RBL1* is near 0.

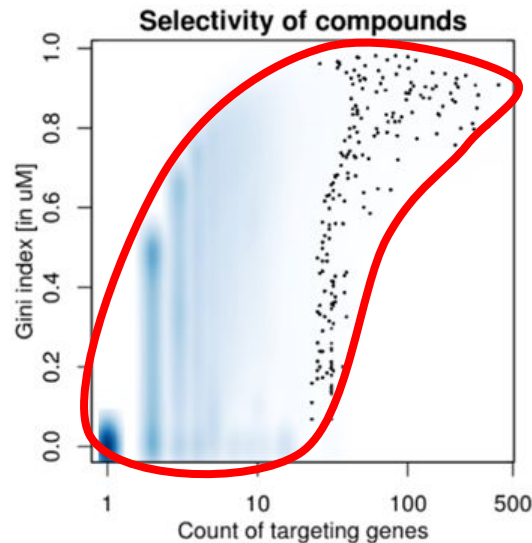
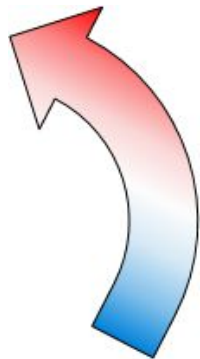
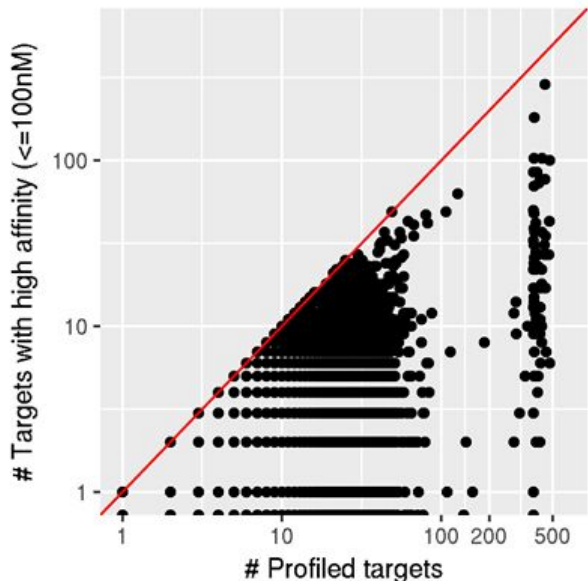
RBL1



An alternative metric: Shannon's Entropy



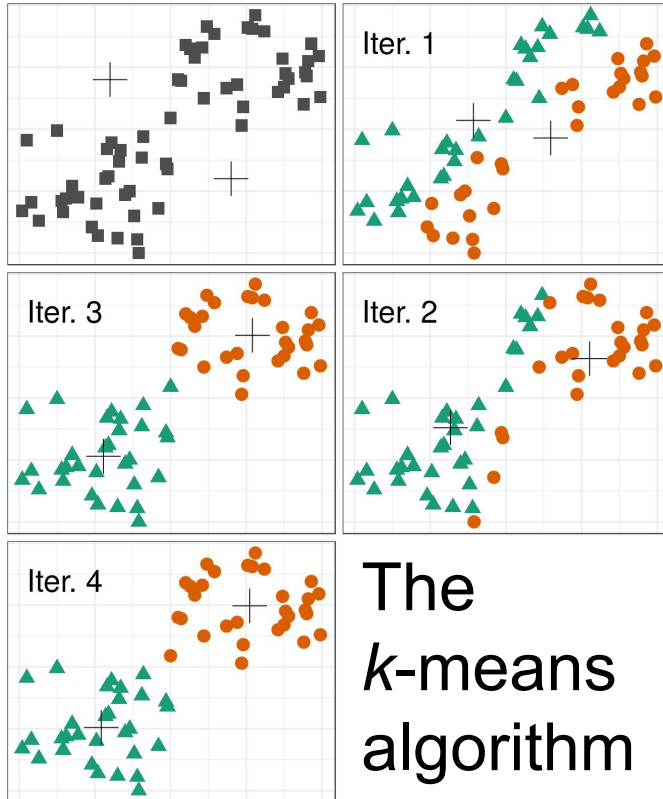
Count of targets and selectivity of ChEMBL molecules



With some exceptions, most compounds are profiled against <100 targets. We distinguish between specific and pleiotropic compounds.

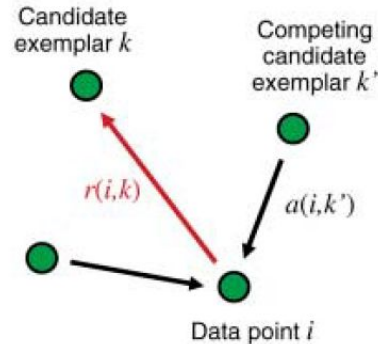
The **shark-fin shape** curve suggests that frequently profiled compounds tend to be more selective (and *vice versa*).

Unsupervised clustering



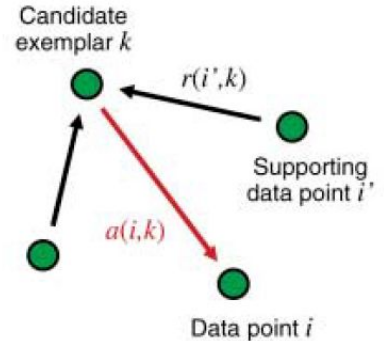
B

Sending responsibilities



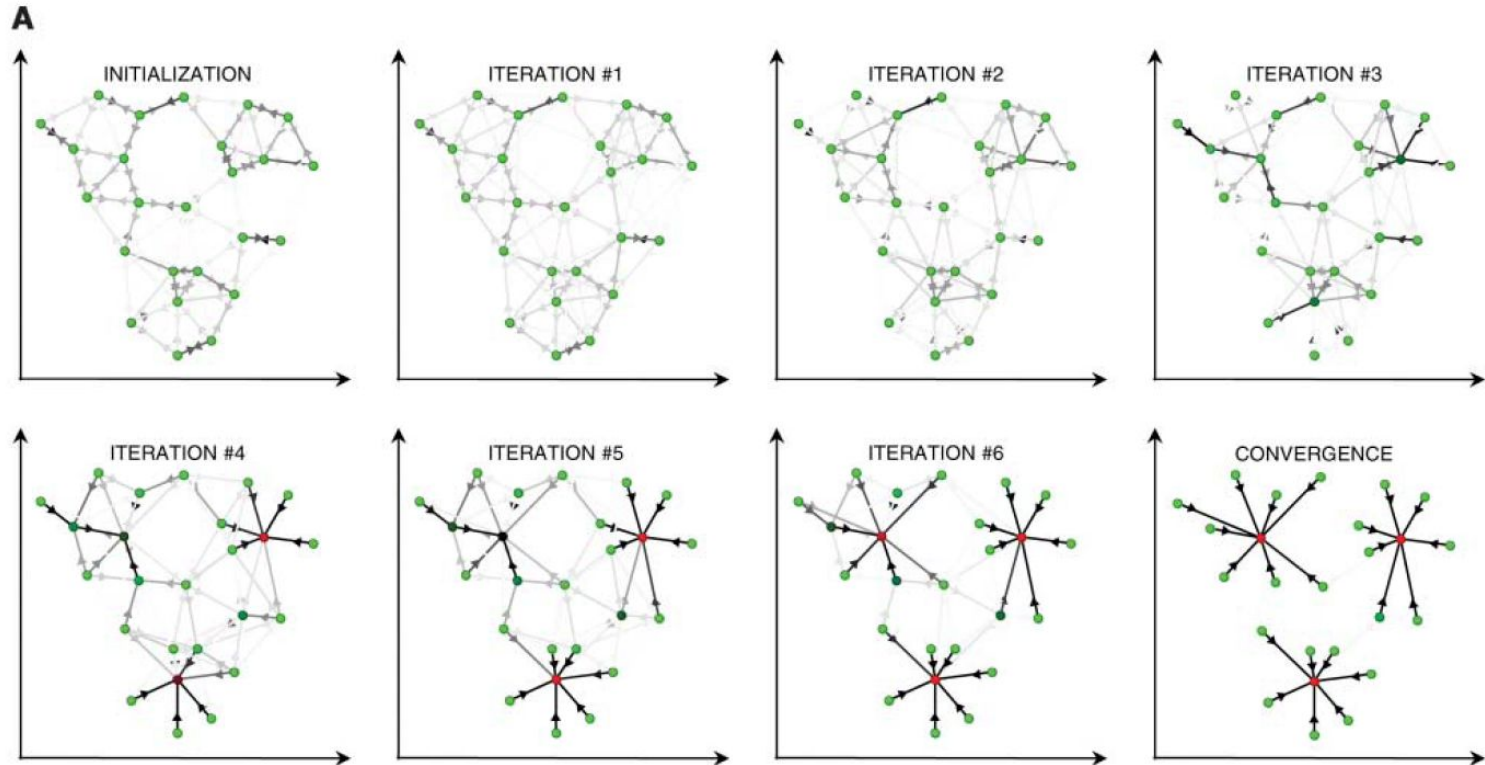
C

Sending availabilities



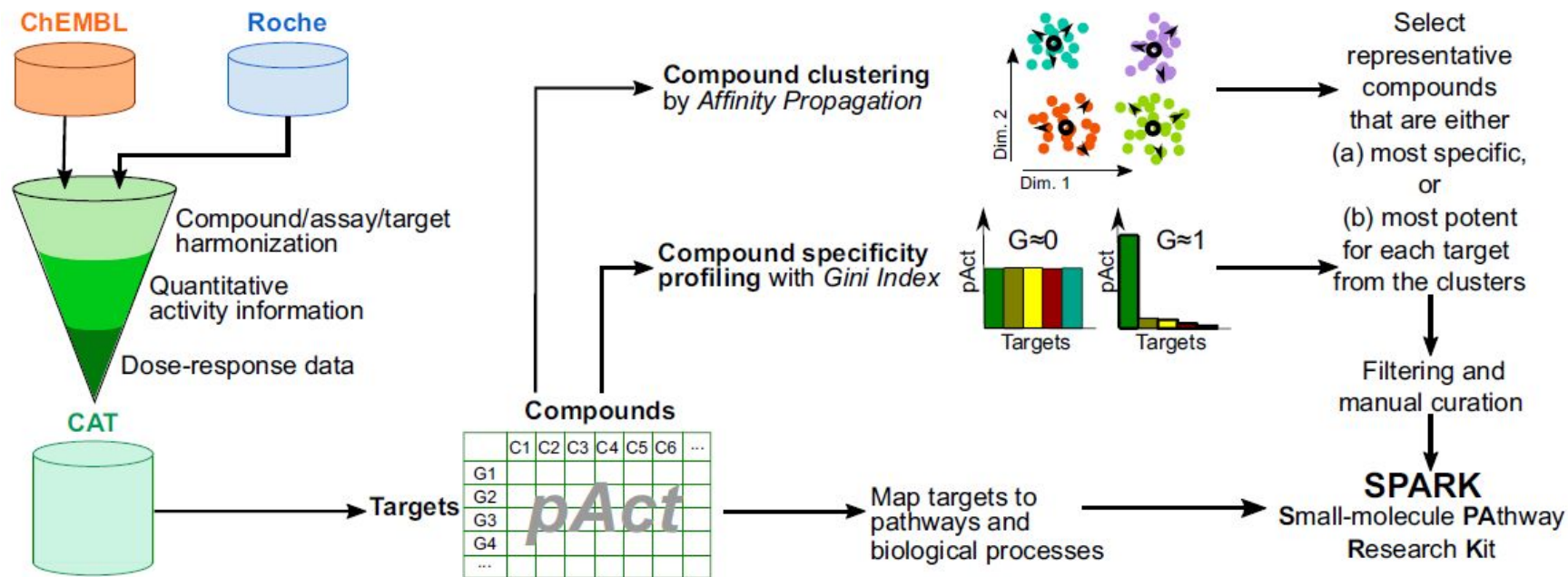
Affinity Propagation updates **responsibilities** and **availabilities** iteratively

Affinity Propagation in action



[A movie of iterations](#)

Construction of SPARK in detail



Harmonization

... of public and Roche internal data

Machine learning

... to select compounds

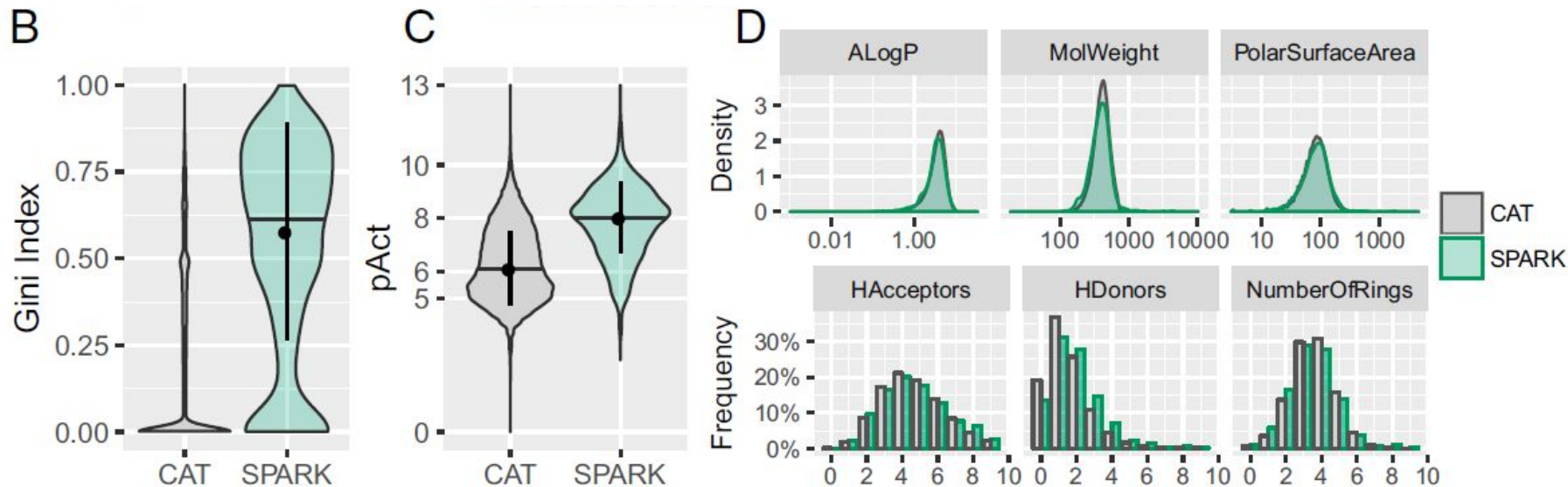
Pathways

... mapped to compounds

Curation

... to enrich quality compounds

SPARK covers the chemical space evenly with representative, potent, and specific compounds

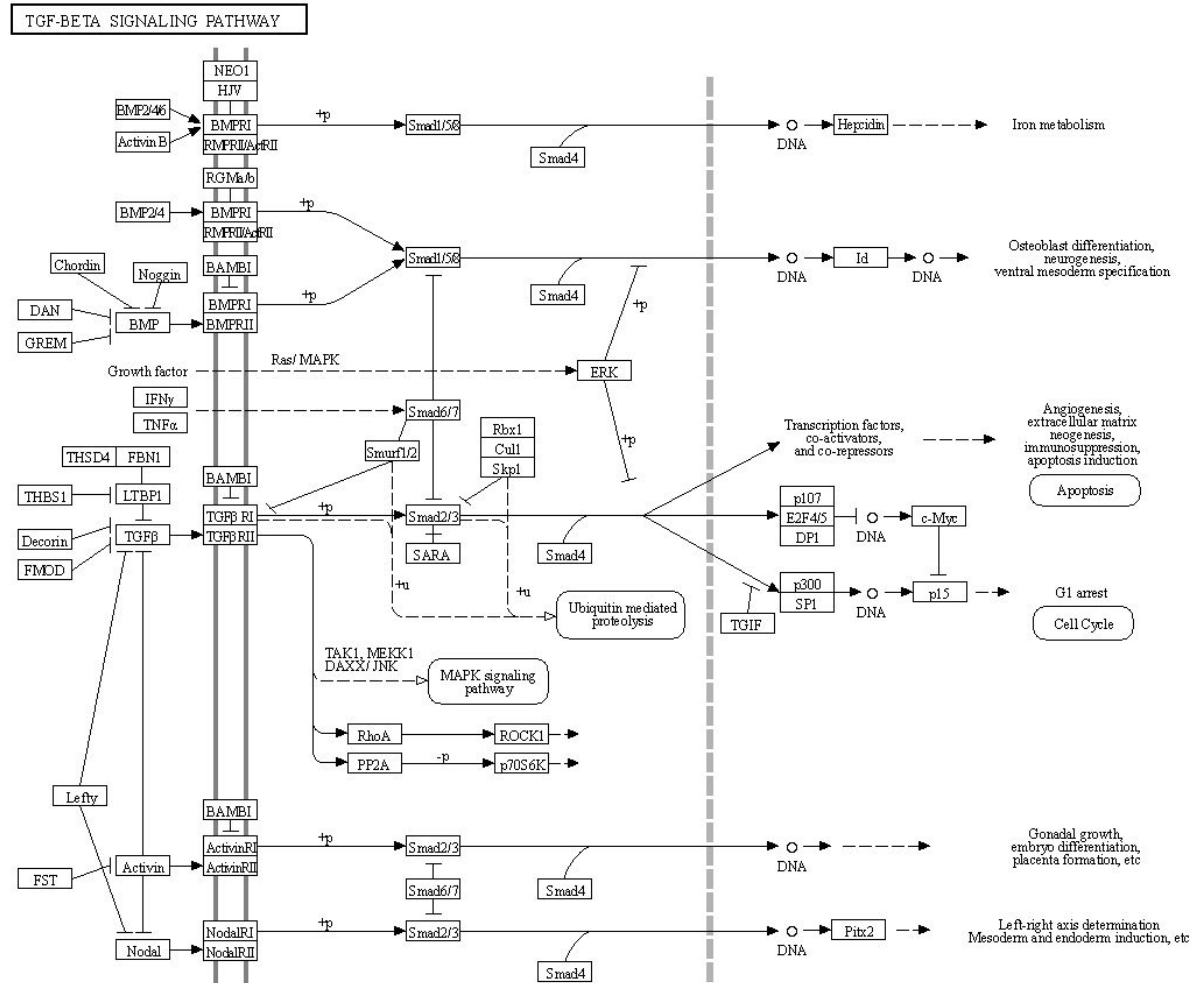


Roudnicky *et al.*, PNAS, 2020,
<https://www.pnas.org/content/early/2020/08/04/1911532117>

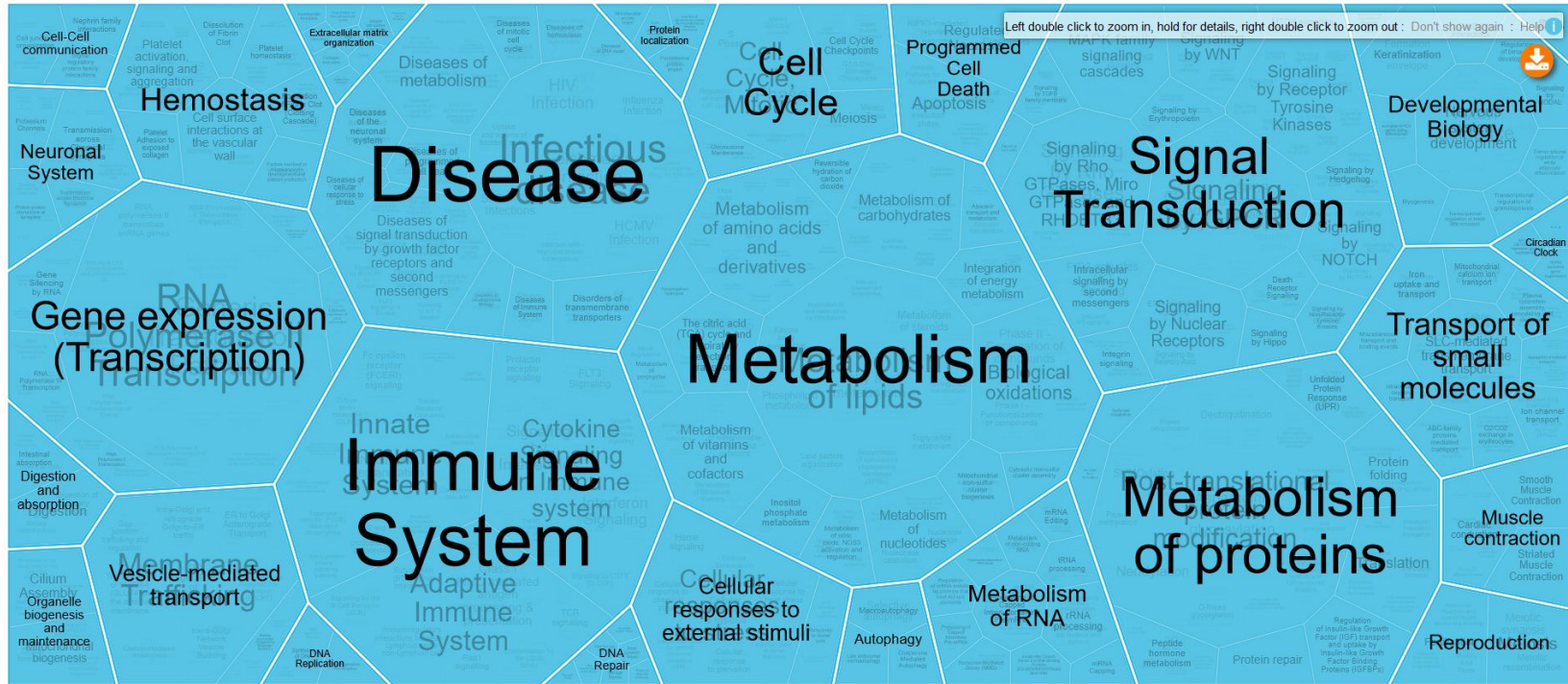
Mapping genes to biological pathways

Option 1: [KEGG pathways](#), with the example of [TGF- \$\beta\$ signaling pathway](#).

[A RESTful API](#) is available for academic use, with clients in Python and R.



Overview of pathways captured by Reactome



The Voronoi (Reacfoam) view of all pathways in Reactome

Mapping genes to biological processes

- Gene Ontology
- UniProtKB keywords
- Example:
TGFR2_HUMAN
(TGF-beta receptor type
-2, P37173)

Keywords

Molecular function	Kinase, Receptor, Serine/threonine-protein kinase, Transferase
Biological process	Apoptosis, Differentiation, Growth regulation
Ligand	ATP-binding, Magnesium, Manganese, Metal-binding, Nucleotide-binding

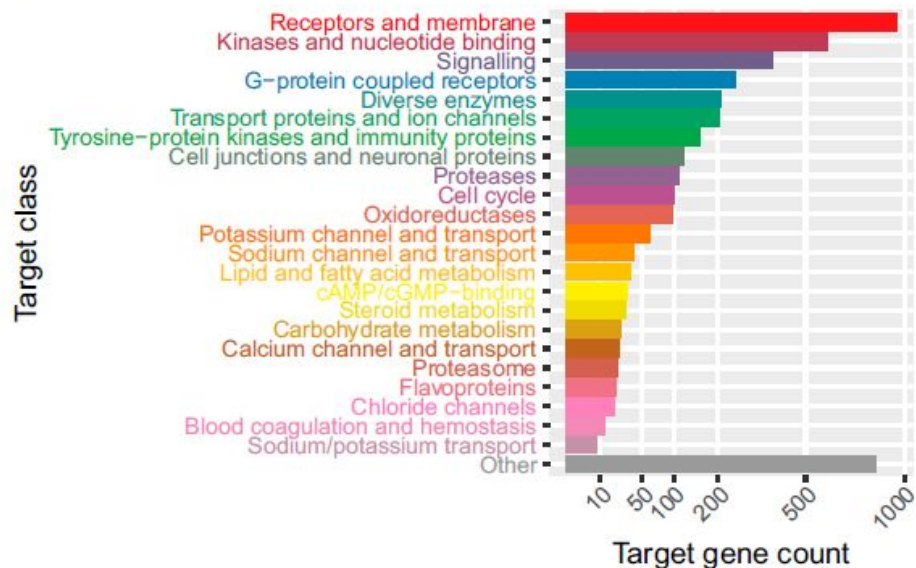


GO - Biological process

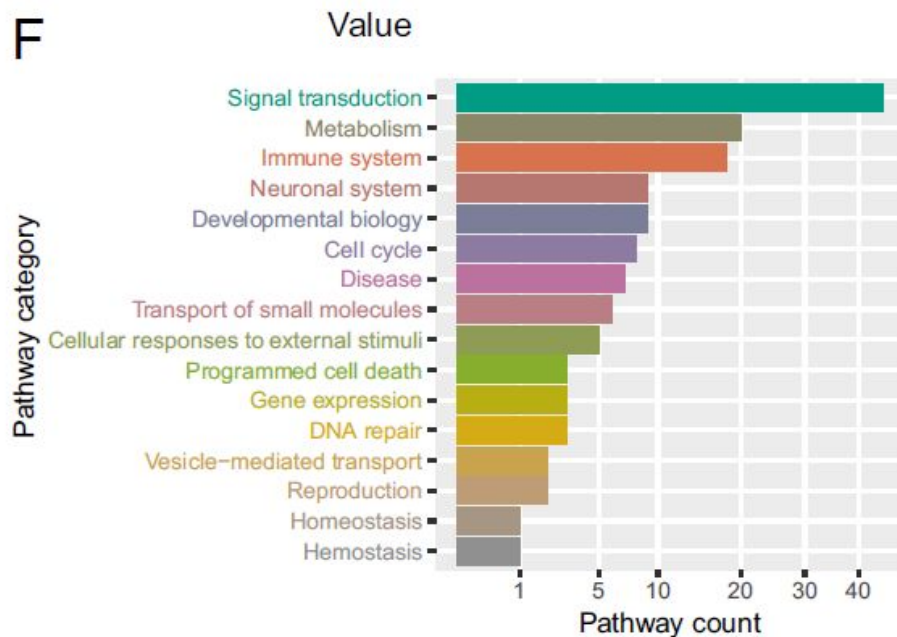
- activation of protein kinase activity Source: BHF-UCL
- aging Source: Ensembl
- animal organ regeneration Source: Ensembl
- apoptotic process Source: UniProtKB
- atrioventricular valve morphogenesis Source: BHF-UCL
- blood vessel development Source: BHF-UCL
- brain development Source: BHF-UCL

SPARK covers the target space evenly with representative, potent, and specific compounds

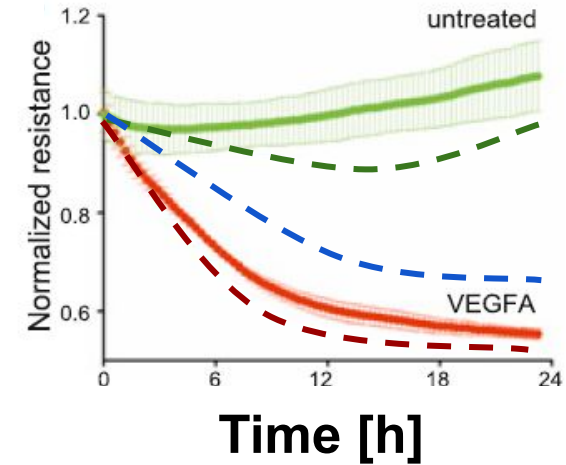
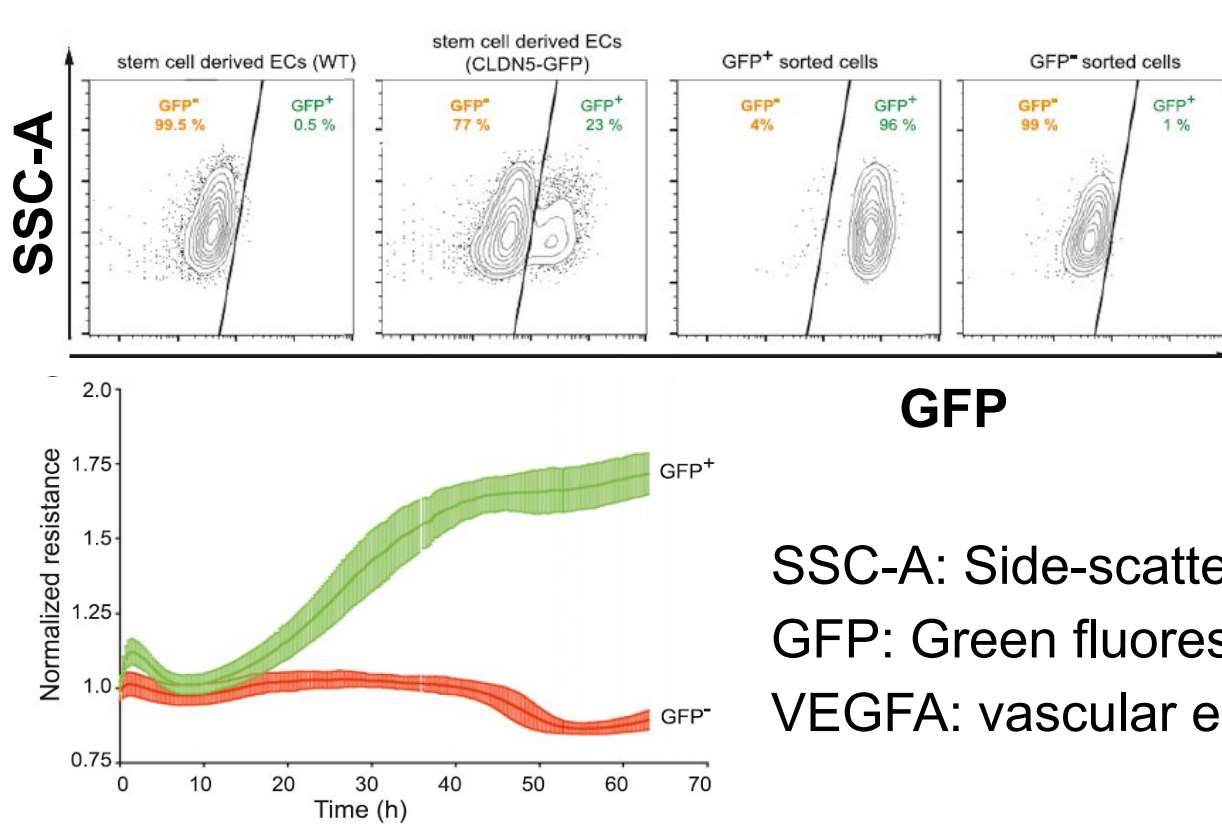
E



F

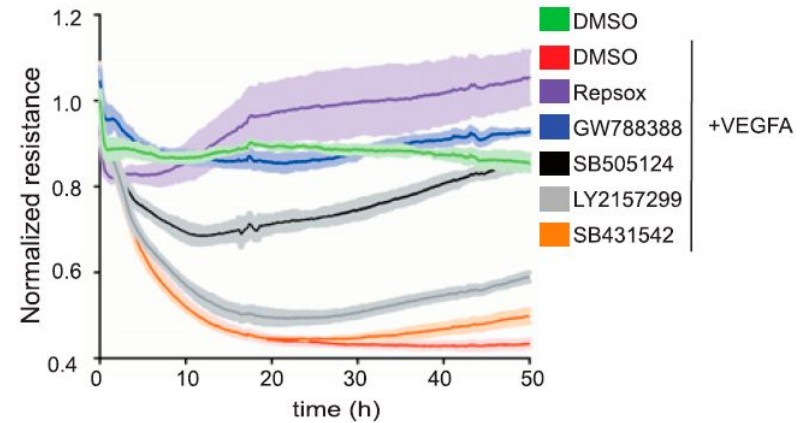
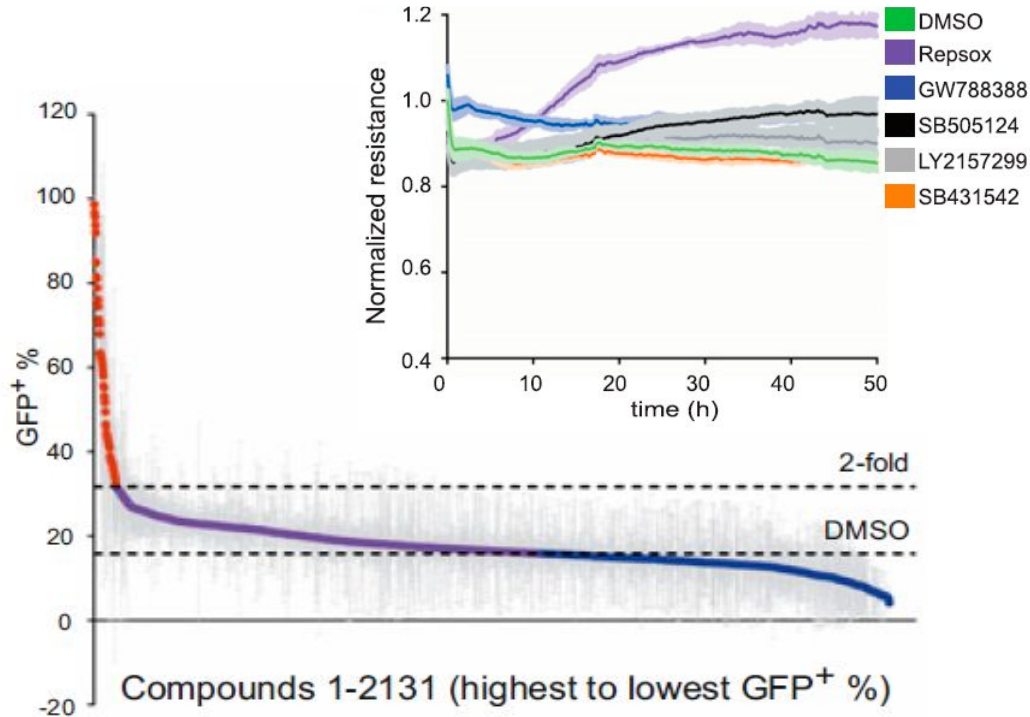


Screening with stem-cell-derived endothelial cells with a reporter added by genome editing



SSC-A: Side-scatter area of flow cytometry;
GFP: Green fluorescent protein;
VEGFA: vascular endothelial growth factor A

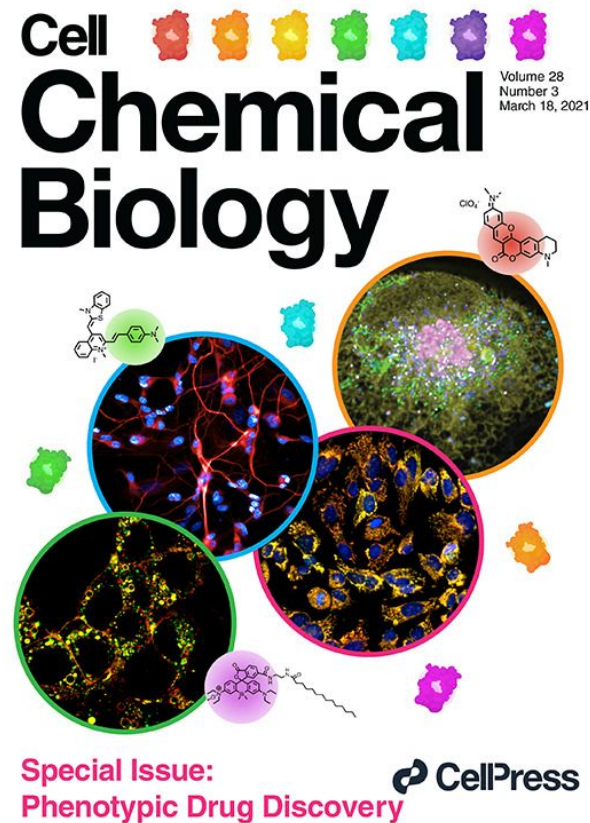
Compounds targeting the TGF- β pathway such as RepSox modulates endothelial cells



Further *in vitro* and *in vivo* experiments establish RepSox as a tool compound modulating retinopathy.

Conclusions about chemogenomic library

- Phenotypic drug discovery can lead to first-in-class drugs with novel mechanisms;
- Unsupervised machine learning and data modelling contribute to build chemogenomic libraries;
- We can link drug candidates via targets to biological pathways and processes.



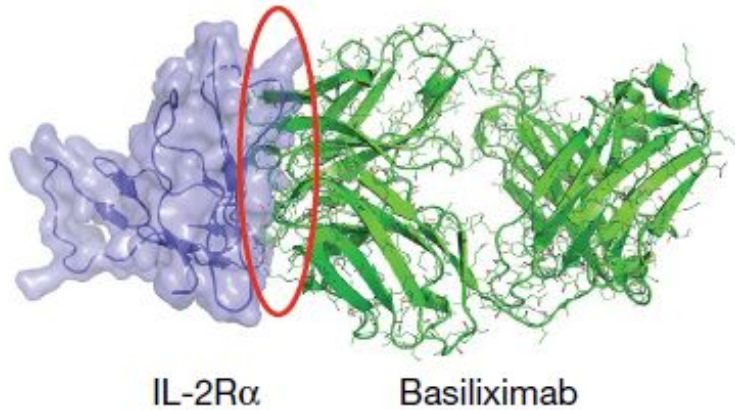
Offline activities of Module II

Please use your favourite programming language (shell scripts, python, R, for instance) and APIs (Application Programming Interfaces) of databases to perform following operations. Submit your code.

1. Retrieve all approved drugs from the ChEMBL database, sort them by approval year and name ([a Python example is here](#); documentations of the ChEMBL API can be found [here](#));
2. For each approved drug **since 2011** that you identified in step (1), retrieve a list of UniProt accession numbers, namely protein targets associated with the drug;
3. For each protein with a UniProt accession number that you identified in step (2), retrieve UniProt keywords associated with it. [You can use the UniProt API, documented here](#). [Python](#) and [R](#) clients are also available.

Multispecific Drug Use or Target Interactions

- b** Conventional drug:
- Forms 1 drug–target interface
 - Can act throughout body
 - Only works if its binding to target alters function of target



- c** Obligate multispecific drug:
- Forms 2 or more drug–target interfaces
- ↓

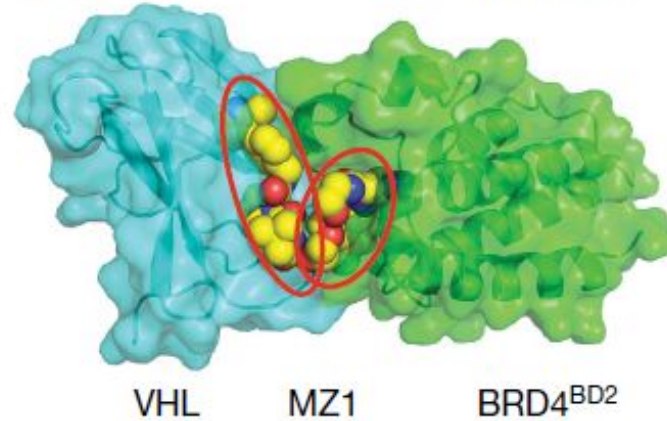
Class 1 ‘tetherbodies’

 - Enrich drug at relevant site of action

↓

Class 2 ‘matchmakers’

 - Link drug to a biological effector



References

1. Swinney, David C., and Jason Anthony. 2011. "How Were New Medicines Discovered?" *Nature Reviews Drug Discovery* 10 (7): 507–19. <https://doi.org/10.1038/nrd3480>.
2. Marx, Uwe, Tommy B. Andersson, Anthony Bahinski, Mario Beilmann, Sonja Beken, Flemming R. Cassee, Murat Cirit, et al. 2016. "Biology-Inspired Microphysiological System Approaches to Solve the Prediction Dilemma of Substance Testing." *ALTEX - Alternatives to Animal Experimentation* 33 (3): 272–321. <https://doi.org/10.14573/altex.1603161>.
3. Dickey, Seth W., Gordon Y. C. Cheung, and Michael Otto. 2017. "Different Drugs for Bad Bugs: Antivirulence Strategies in the Age of Antibiotic Resistance." *Nature Reviews Drug Discovery* 16 (7): 457–71. <https://doi.org/10.1038/nrd.2017.23>.
4. Lewis, Kim. 2013. "Platforms for Antibiotic Discovery." *Nature Reviews Drug Discovery* 12 (5): 371–87. <https://doi.org/10.1038/nrd3975>.
5. Warner, Katherine Deigan, Christine E. Hajdin, and Kevin M. Weeks. 2018. "Principles for Targeting RNA with Drug-like Small Molecules." *Nature Reviews Drug Discovery* 17 (8): 547–58. <https://doi.org/10.1038/nrd.2018.93>.
6. Berg, Ellen L. 2021. "The Future of Phenotypic Drug Discovery." *Cell Chemical Biology* 28 (3): 424–30. <https://doi.org/10.1016/j.chembiol.2021.01.010>.
7. Chin, Marcus Y., Jether Amos Espinosa, Grace Pohan, Sarine Markossian, and Michelle R. Arkin. 2021. "Reimagining Dots and Dashes: Visualizing Structure and Function of Organelles for High-Content Imaging Analysis." *Cell Chemical Biology* 28 (3): 320–37. <https://doi.org/10.1016/j.chembiol.2021.01.016>.
8. Conway, Louis P., Weichao Li, and Christopher G. Parker. 2021. "Chemoproteomic-Enabled Phenotypic Screening." *Cell Chemical Biology* 28 (3): 371–93. <https://doi.org/10.1016/j.chembiol.2021.01.012>.
9. Dahlin, Jayme L., Douglas S. Auld, Ina Rothenaigner, Steve Haney, Jonathan Z. Sexton, J. Willem M. Nissink, Jarrod Walsh, et al. 2021. "Nuisance Compounds in Cellular Assays." *Cell Chemical Biology* 28 (3): 356–70. <https://doi.org/10.1016/j.chembiol.2021.01.021>.
10. Ha, Jaeyoung, Hankum Park, Jongmin Park, and Seung Bum Park. 2021. "Recent Advances in Identifying Protein Targets in Drug Discovery." *Cell Chemical Biology* 28 (3): 394–423. <https://doi.org/10.1016/j.chembiol.2020.12.001>.

References (continued)



11. Hsu, Ku-Lung. 2021. "Shining a Light on Phenotypic Drug Discovery." *Cell Chemical Biology* 28 (2): 115–17. <https://doi.org/10.1016/j.chembiol.2021.01.020>.
12. Hughes, Rebecca E., Richard J. R. Elliott, John C. Dawson, and Neil O. Carragher. 2021. "High-Content Phenotypic and Pathway Profiling to Advance Drug Discovery in Diseases of Unmet Need." *Cell Chemical Biology* 28 (3): 338–55. <https://doi.org/10.1016/j.chembiol.2021.02.015>.
13. Vandana, J. Jeya, Lauretta A. Lacko, and Shuibing Chen. 2021. "Phenotypic Technologies in Stem Cell Biology." *Cell Chemical Biology* 28 (3): 257–70. <https://doi.org/10.1016/j.chembiol.2021.02.001>.
14. Ziegler, Slava, Sonja Sievers, and Herbert Waldmann. 2021. "Morphological Profiling of Small Molecules." *Cell Chemical Biology* 28 (3): 300–319. <https://doi.org/10.1016/j.chembiol.2021.02.012>.
15. Moffat, John G., Fabien Vincent, Jonathan A. Lee, Jörg Eder, and Marco Prunotto. 2017. "Opportunities and Challenges in Phenotypic Drug Discovery: An Industry Perspective." *Nature Reviews Drug Discovery* 16 (8): 531–43. <https://doi.org/10.1038/nrd.2017.111>.
16. Drawnel, Faye Marie, Jitao David Zhang, Erich Küng, Natsuyo Aoyama, Fethallah Benmansour, Andrea Araujo Del Rosario, Sannah Jensen Zoffmann, et al. 2017. "Molecular Phenotyping Combines Molecular Information, Biological Relevance, and Patient Data to Improve Productivity of Early Drug Discovery." *Cell Chemical Biology* 18 (24(5)): 624–34. <https://doi.org/10.1016/j.chembiol.2017.03.016>.
17. Roudnicky, Filip, Jitao David Zhang, Bo Kyoung Kim, Nikhil J. Pandya, Yanjun Lan, Lisa Sach-Peltason, Heloise Ragelle, et al. 2020. "Inducers of the Endothelial Cell Barrier Identified through Chemogenomic Screening in Genome-Edited HPSC-Endothelial Cells." *Proceedings of the National Academy of Sciences*, August. <https://doi.org/10.1073/pnas.1911532117>.
18. Zoffmann, Sannah, Maarten Vercruysse, Fethallah Benmansour, Andreas Maunz, Luise Wolf, Rita Blum Marti, Tobias Heckel, et al. 2019. "Machine Learning-Powered Antibiotics Phenotypic Drug Discovery." *Scientific Reports* 9 (1): 1–14. <https://doi.org/10.1038/s41598-019-39387-9>.
19. Nichols, Robert J., Saunak Sen, Yoe Jin Choo, Pedro Beltrao, Matylda Zietek, Rachna Chaba, Sueyoung Lee, et al. 2011. "Phenotypic Landscape of a Bacterial Cell." *Cell* 144 (1): 143–56. <https://doi.org/10.1016/j.cell.2010.11.052>.
20. Smith, Kevin, Filippo Piccinini, Tamas Balassa, Krisztian Koos, Tivadar Danko, Hossein Azizpour, and Peter Horvath. 2018. "Phenotypic Image Analysis Software Tools for Exploring and Understanding Big Image Data from Cell-Based Assays." *Cell Systems* 6 (6): 636–53. <https://doi.org/10.1016/j.cels.2018.06.001>.

References (continued)

21. Scheeder, Christian, Florian Heigwer, and Michael Boutros. 2018. "Machine Learning and Image-Based Profiling in Drug Discovery." *Current Opinion in Systems Biology, Pharmacology and drug discovery*, 10 (August): 43–52. <https://doi.org/10.1016/j.coisb.2018.05.004>.
22. Schirle, Markus, and Jeremy L. Jenkins. 2016. "Identifying Compound Efficacy Targets in Phenotypic Drug Discovery." *Drug Discovery Today* 21 (1): 82–89. <https://doi.org/10.1016/j.drudis.2015.08.001>.
23. Wilkinson, Isabel V. L., Georg C. Terstappen, and Angela J. Russell. 2020. "Combining Experimental Strategies for Successful Target Deconvolution." *Drug Discovery Today* 25 (11): 1998–2005. <https://doi.org/10.1016/j.drudis.2020.09.016>.
24. Comess, Kenneth M., Shaun M. McLoughlin, Jon A. Oyer, Paul L. Richardson, Henning Stöckmann, Anil Vasudevan, and Scott E. Warder. 2018. "Emerging Approaches for the Identification of Protein Targets of Small Molecules - A Practitioners' Perspective." *Journal of Medicinal Chemistry* 61 (19): 8504–35. <https://doi.org/10.1021/acs.jmedchem.7b01921>.
25. Sidders, Ben, Anna Karlsson, Linda Kitching, Rubben Torella, Paul Karila, and Anne Phelan. 2018. "Network-Based Drug Discovery: Coupling Network Pharmacology with Phenotypic Screening for Neuronal Excitability." *Journal of Molecular Biology, Theory and Application of Network Biology Toward Precision Medicine*, 430 (18, Part A): 3005–15. <https://doi.org/10.1016/j.jmb.2018.07.016>.
26. Aulner, Nathalie, Anne Danckaert, JongEun Ihm, David Shum, and Spencer L. Shorte. 2019. "Next-Generation Phenotypic Screening in Early Drug Discovery for Infectious Diseases." *Trends in Parasitology* 35 (7): 559–70. <https://doi.org/10.1016/j.pt.2019.05.004>.
27. Boess, Franziska, Barbara Lenz, Juergen Funk, Urs Niederhauser, Simon Bassett, Jitao David Zhang, Thomas Singer, and Adrian B. Roth. 2017. "Use of Early Phenotypic in Vivo Markers to Assess Human Relevance of an Unusual Rodent Non-Genotoxic Carcinogen in Vitro." *Toxicology* 379 (March): 48–61. <https://doi.org/10.1016/j.tox.2017.01.018>.
28. Feng, Yan, Timothy J. Mitchison, Andreas Bender, Daniel W. Young, and John A. Tallarico. 2009. "Multi-Parameter Phenotypic Profiling: Using Cellular Effects to Characterize Small-Molecule Compounds." *Nature Reviews Drug Discovery* 8 (7): 567–78. <https://doi.org/10.1038/nrd2876>.
29. Jones, Lyn H., and Mark E. Bunnage. 2017. "Applications of Chemogenomic Library Screening in Drug Discovery." *Nature Reviews Drug Discovery* 16 (January): 285–296. <https://doi.org/10.1038/nrd.2016.244>.
30. Kulesa, Anthony, Jared Kehe, Juan E. Hurtado, Prianca Tawde, and Paul C. Blainey. 2018. "Combinatorial Drug Discovery in Nanoliter Droplets." *Proceedings of the National Academy of Sciences* 115 (26): 6685–90. <https://doi.org/10.1073/pnas.1802233115>.

References (continued)

31. Vlachogiannis, Georgios, Somaieh Hedayat, Alexandra Vatsiou, Yann Jamin, Javier Fernández-Mateos, Khurum Khan, Andrea Lampis, et al. 2018. "Patient-Derived Organoids Model Treatment Response of Metastatic Gastrointestinal Cancers." *Science* 359 (6378): 920–26. <https://doi.org/10.1126/science.aao2774>.
32. Wawer, Mathias J., Kejie Li, Sigrun M. Gustafsdottir, Vebjorn Ljosa, Nicole E. Bodycombe, Melissa A. Marton, Katherine L. Sokolnicki, et al. 2014. "Toward Performance-Diverse Small-Molecule Libraries for Cell-Based Phenotypic Screening Using Multiplexed High-Dimensional Profiling." *Proceedings of the National Academy of Sciences* 111 (30): 10911–16. <https://doi.org/10.1073/pnas.1410933111>.
33. Zhang, Jitao David, Erich Küng, Franziska Boess, Ulrich Certa, and Martin Ebeling. 2015. "Pathway Reporter Genes Define Molecular Phenotypes of Human Cells." *BMC Genomics* 16: 342. <https://doi.org/10.1186/s12864-015-1532-2>.
34. Ziegler, Slava, Verena Pries, Christian Hedberg, and Herbert Waldmann. 2013. "Target Identification for Small Bioactive Molecules: Finding the Needle in the Haystack." *Angewandte Chemie International Edition* 52 (10): 2744–92. <https://doi.org/10.1002/anie.201208749>.
35. Antolin, Albert A., Joseph E. Tym, Angeliki Komianou, Ian Collins, Paul Workman, and Bissan Al-Lazikani. 200017. "Objective, Quantitative, Data-Driven Assessment of Chemical Probes." *Cell Chemical Biology* 0 (0). <https://doi.org/10.1016/j.chembiol.2017.11.4>.
36. Fellmann, Christof, Benjamin G. Gowen, Pei-Chun Lin, Jennifer A. Doudna, and Jacob E. Corn. 2017. "Cornerstones of CRISPR–Cas in Drug Discovery and Therapy." *Nature Reviews Drug Discovery* 16 (2): 89–100. <https://doi.org/10.1038/nrd.2016.238>.
37. Hart, Charles P. 2005. "Finding the Target after Screening the Phenotype." *Drug Discovery Today* 10 (7): 513–19. [https://doi.org/10.1016/S1359-6446\(05\)03415-X](https://doi.org/10.1016/S1359-6446(05)03415-X).
38. Stanton, Benjamin Z., Emma J. Chory, and Gerald R. Crabtree. 2018. "Chemically Induced Proximity in Biology and Medicine." *Science* 359 (6380): [eaa05902](https://doi.org/10.1126/science.aao5902). <https://doi.org/10.1126/science.aao5902>.
39. Deshaies, Raymond J. 2020. "Multispecific Drugs Herald a New Era of Biopharmaceutical Innovation." *Nature* 580 (7803): 329–38. <https://doi.org/10.1038/s41586-020-2168-1>.
40. Tom Gregory, <https://www.quora.com/Is-Gini-coefficient-outdated>

References (continued)

41. [EBI online tutorial of Reactome](#), DOI: 10.6019/TOL.Reactome_exbp-t.2017.00001.1
42. Frey, Brendan J., and Delbert Dueck. 2007. "Clustering by Passing Messages Between Data Points." *Science* 315 (5814): 972–76.
<https://doi.org/10.1126/science.1136800>.
43. [Webinar introduction to apcluster](#)
44. [Affinity Propagation algorithm visualized](#)