

# Advanced brain imaging methods: Practical 2

Remi Gau

24<sup>th</sup> February 2015

## 1 Running a GLM with SPM

### 1.1 Fixed effect analysis: First level with an block design

First follow the instruction in SPM\_GLM\_Block.pdf (on canvas) extracted from the SPM manual on how to model the GLM of a block design experiment, how to estimate the model and view the results in SPM. For the data to input either use the one you have pre-processed in the practical 1 or unzip the file in BlockDesignData.zip (on canvas). I have created the render file mentionned at the end of this pdf, so you will not need to have to do it.

Once you have viewed the results of the 2 t-contrasts (listening>rest and rest>listening), create a new F-contrast by just specifying '1' as input in the contrast manager. Compare the results of that F-contrast to the the 2 t-contrasts you have previously created: how do they differ in terms of patterns and intensity (i.e p values).

If you want to know in which structure a specific peak or cluster is located you can use the `spm_atlas` function with the X/Y/Z MNI coordinates of a voxel of interest as follows:

```
spm_atlas('query','neuromorphometrics',[X Y Z]')
```

You can also use the anatomy toolbox (downloadable from the SPM website) that will give you more information about the probability of an activation to be in a given brain structure.

### 1.2 Fixed effect analysis: First level with an event-related design

Now we are going to model an event-related experiment. The instructions are in the file SPM\_GLM\_Event.pdf (on canvas). This is a repetition priming experiment performed using event-related fMRI. Briefly, this is a 2 by 2 factorial study with factors "fame" and "repetition" where famous and non-famous faces were presented twice against a checkerboard baseline. The subject was asked to make fame judgements by making key presses. There are thus four event-types of interest; first and second presentations of famous and non-famous faces, which we denote N1, N2, F1 and F2.

For the data to input unzip the file in EventDesignData.zip (on canvas), though I would recommend you to do the pre-processing of those data yourself at least once: event-related designs necessitate a pre-processing extra-step (i.e slice timing) that we have not seen in the last practical.

### 1.3 Random effect analysis: second level

To finish this part on how to use SPM to model GLM and estimate them, we are going to perform a second level analysis (at the groupe level). The instructions are in the file SPM.RFX.pdf (on canvas).

## 2 GLM: 2 samples t-test

### 2.1 Having a look at the data

Unzip the file DataPractical2.zip. The file 'T-Test.mat' contains simulated data from 2 different groups A and B presented with two different condition from an experiment. You can load the content of this file into matlab by double clicking on it or by typing:

```
load T-Test.mat
```

You should now see two new variables,  $Y_A$  and  $Y_B$ , in the matlab workspace. You can plot the distribution of the group A by typing `hist(Y_A,x)` where x is the number of bin you want your distribution to have.

### 2.2 Design Matrix

If you want to perform a two-sample t-test on this data using a GLM, what should the design matrix ( $X$ ) be? Create this design matrix in matlab. For this you can use the functions:

- `ones(a,b)` which will create a matrix with a lines and b columns containing only ones,
- `zeros(a,b)` which will do the same but with zeros,
- `repmat(x,a,b)` which will repeat the matrix x, a times vertically and b times horizontally,
- you can concatenate vertically 2 vectors A and B into a new vector C like this `C=[A;B]` ,
- you can concatenate horizontally 2 vectors A and B into a new vector C like this `C=[A,B]` ,

You can display your design matrix using the `imagesc` function. If you want to have the design matrix in white and black type `colormap('Gray')` before displaying it. You can add a color bar to your figure with `colorbar` .

## 2.3 Estimating Beta

The ordinary least squares method gives the following unbiased estimator of  $\beta$ :

$$\hat{\beta} = (X^T * X)^{-1} * X^T * Y$$

where  $X$  is the design matrix and  $Y$  the concatenated data.

What is the matlab code equivalent to this equation that will compute  $\beta$ ? Remember that in matlab:

- to get the transpose  $X^T$  of a matrix  $X$ , you can simply type `X'`,
- to get the inverse  $X^{-1}$  of a matrix  $X$ , you can simply type `inv(X)`.

## 2.4 Residual and statistics

You can now compute  $\hat{Y}$  the predicted value for each subject with the following equation:

$$\hat{Y} = X\beta$$

The residual for each subject (i.e the errors of your model) can then be computed with this equation:

$$\varepsilon = Y - \hat{Y}$$

You can have a look at the errors to make sure they are normally distributed using the `hist` command or, even better if you have acces to matlab statistical toolbox, the `normplot` command.

Using the function `std`, compute the  $\sigma_\varepsilon$ , the standard deviation of the errors, and then you should be able to find the t-value with the following equation:

$$t = \frac{c^T \beta}{\sqrt{\sigma_\varepsilon^2 c^T \text{inv}(X' * X) c}}$$

where  $c$  is a vertical vector that defines the contrast you want to test.

To get the square root of a number  $a$ , you can just do `sqrt(a)` and to get the square of  $a$ , typing `a^2` will do the job.

Once you have the t-value the following code should give you the p-value associated to it:

```
% degrees of freedom: depends only on
% the rank of the design matrix (X)
df      = rank(X)-1;

% degrees of freedom of the error ;
% Y is a vector containing all the data
% 'length' tells us how many values Y contains
dferror = length(Y) - df - 1;

% the p value is found via the cummulative distribution
% function (cdf) of a student distribution
% requires access to matlab statistical toolbox
p = tcdf(-t, dferror)
```

### 3 Modelling the hemodynamic response (HRF)

SPM uses the function `spm_hrf(dt)` to create an HRF, with  $dt$  being the temporal resolution. Uses this function to create an HRF with a 0.01 second temporal resolution.

The HRF is usually modelled by a weighted sum of gamma function. A gamma function can be described by the following equation:

$$h(t) = \frac{t^{n-1}}{\lambda^n (n-1)!} e^{-t/\lambda}$$

A gamma function is given by the following code:

```
g = ((t).^(n-1)) / (lambda^n*factorial(n-1)) .* exp(-(t)/lambda)
```

First, create a 'time' vector,  $t$  containing all the values from 0 to 32 seconds in steps of 0.01 second (e.g the code `0:1:10` would give you all the values from 0 to 10 in steps of 1). Then create a first gamma function  $h_1$  with  $n = 4$  and  $\lambda = 1$ , a second one  $h_2$  with  $n = 8$  and  $\lambda = 1$  and a third one  $h_3$  with  $n = 16$  and  $\lambda = 1$ . Finally get the HRF by making the weighted sums of those functions as follow:

$$HRF = c_1 h_1 + c_2 h_2 + c_3 h_3;$$

Try different values of  $c_1$ ,  $c_2$  and  $c_3$  and plot the results to try to find some of the values that SPM uses to create a typical HRF (hint: start with  $c_1 = 1$  and a negative  $c_2$ ).

### 4 More on the HRF: basis set

Most of the time you want to model the HRF but also its temporal and dispersion derivatives. The 3 of them together can then be used as an informed basis function set to model the BOLD signal. The following code will produce one of those basis set:

```
% Temporal resolution in seconds of the
% informed basis set you are going to create
xBF.dt = .1;
xBF.name = 'hrf (with time and dispersion derivatives)';

% Length of the HRF in seconds
xBF.length = 33;
xBF.order = 1;

% Creates the informed basis set
xBF = spm_get_bf(xBF);
```

The structure `xBF` contains the shape of the different basis function under the field `bf`. For example `xBF.bf(:,1)` contains the shape of the canonical HRF and `xBF.bf(:,2)` the temporal derivative. Using the `plot` function plot the 3 different functions.

Then you can try different weighted sum or difference of those functions to see how you can model different BOLD responses with this basis set. For example:

```
2*xBF.bf(:,1)-0.5*xBF.bf(:,2)
```

## 5 Convolving with the HRF

Create a basis function set with a temporal resolution of 1 second.

Then, using the functions `ones`, `zeros` and `repmat`, create a 'time' vector made of 8 repetitions of 16 zeros followed by one 1. This is the onset vector: a value of 1 means that at this instant in time a stimulus was presented. You can view this vector using the function `plot` or the function `stem`.

You can then convolve this vector with the HRF and its temporal derivative using the `conv` function. For example.

```
ConvolvedRegressor = conv(HRF, OnsetVector);
```

Plot the results of both convolution.

## 6 Doing a GLM on real fMRI data

Load the file `TimeCourseA1.mat` which contains the time course of the BOLD signal acquired from the left and right primary auditory cortices during an experiment where subjects were passively listening to words (These are actually the time courses you could get at the end of the practical 1). The TR for this experiment was 7 seconds. 84 acquisitions were made in blocks of 6. Successive blocks alternated between rest and auditory stimulation, starting with rest. Auditory stimulation was bi-syllabic words presented binaurally at a rate of 60 per minute.

Do a GLM analysis for each of those time courses:

It is always good to have a look at the raw data.

The following code will give you the onset vector for this experiment:

```
Onsets = 6:12:84;
OnsetVector = zeros(length(TimeCourseLeft),1);
for i=1:length(Onsets)
    OnsetVector([Onsets(i):Onsets(i)+5])=1;
end
```

Convolve this onset vector with the canonical HRF from the basis set (use a temporal resolution equal to the TR).

Create a design matrix with this convolved regressor. Should you add a column to that matrix to model a constant?

Just like the T-test at the beginning of this practical, compute the  $\beta$ , the predicted values and the residuals for this model.

Plot the predicted and the raw data on the same figure.

Plot the normplot of the residuals.

You can also check if there is any autocorrelation in the residual with the following code:

```
[AutoCorrFunc, lags] = xcorr(Residuals, 'coeff');  
plot(lags, AutoCorrFunc)  
ylabel('Autocorrelation')
```

Given those different graphs, can you think of simple ways to improve your model?