```
import pymc as pm
import matplotlib.pyplot as plt
import arviz as az
import pandas as pd
from scipy import special, stats
```

from google.colab import files
uploaded = files.upload()

algae.csv(text/csv) - 17987 bytes, last modified: 10/29/2023 - 100% done
 algaeTest.csv(text/csv) - 20683 bytes, last modified: 10/29/2023 - 100% done

Saving algae.csv to algae.csv Saving algaeTest.csv to algaeTest.csv

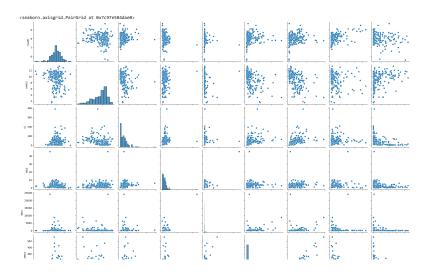
or each of these predictor variable observations, the data set also contains measurements of 7 different harmful algae, labeled a1-a7 but not named. For this work we will use the a1 measurements as the response variable. With algae.csy data and the stated goal of the EU perform the following tasks and answer the stated questions. (a) Perform exploratory data analysis (EDA) to include as minimum a review of the summary statistics for the data. Briefly explain how this EDA informs the next steps of your work.

#algae_train = pd.read_csv('algae.csv', header=0)
algae_test = pd.read_csv('algaeTest.csv', header=0)

algae_train = pd.read_csv('algae.csv')

predictors = algae_train.columns[0:11]

for p in predictors: sum_stats = (algae_train.loc[:, [p, 'al']].groupby(p).agg(["mean", "std", "count"]))





EDA Comments: Having a general idea of the trends in the data, it appears there might be an instance fo colinearity, so I should remove oPO4 when it comes to cleaning it later. Otherwise, this seems alright. And ofc I can see that certain chemeical can contribute to algae growth as

large

b) Clean the data by identifying missing values and use the following strategy to estimate missing values: 1. For continuous (real-valued) data use the mean; 2. For discrete (integer valued) use the median; and 3. For categorical or nominal data use the mode. Then scale the predictor variables (subtract the mean and divide by the standard deviation) and code the categorical variables. Explain why you are doing this. Reparameterize the response variable using a log trans form and explain why you are doing this.

```
#drop unneeded vars
#algae_train.drop(['a2','a3','a4','a5','a6','a7'], axis = 1, inplace = True)
        201
pd.isnull(algae_train).any()
 algae_train['season']
          spring
autumn
spring
autumn
          autumn
spring
autumn
winter
```

Name: season, Length: 200, dtype: object

-~ | |



sns.boxplot(data=algae_train, x="size", y="a1", hue = 'size')
plt.figure()
sns.boxplot(data=algae_train, x="speed", y="a1", hue = 'size')

print(sum stats)

```
# there are no discrete columns, and the categorical columns have no missing data
    for col in predictors[3:]:
    algae_train[col] = algae_train[col].replace(np.nan, np.mean(algae_train[col]))
algae_train[col] -= np.mean(algae_train[col])
algae_train[col] /= np.std(algae_train[col])
aspe__valn(to1) /= np.stu(algae_tvaln(to1))
season_vals = {\'unimer': 0, 'spring': 1, 'summer': 2, 'autumn': 3}
size_vals = {\'small': 0, 'medium': 1, 'large': 2}
speed_vals = {\'low': 0, 'medium': 1, 'high': 2}
algae_train['season'] = algae_train['season'].replace(season_vals)
algae_train['size'] = algae_train['size'].replace(size_vals)
algae_train['speed'] = algae_train['speed'].replace(speed_vals)
```

algae_train['a1'].loc[algae_train['a1']>0] = np.log(algae_train['a1'].loc[algae_train['a1']>0])

<ipython-input-9-ce9099891a6c>:13: SettingWithCopyWarning:
A value is trying to be set on a copy of a slice from a DataFrame

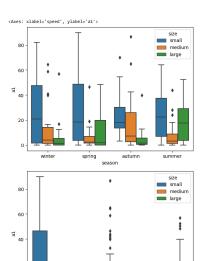
See the caveats in the documentation: https://pandas.pydata.org/pandas-docs/stable/user_guide/inde algae_train['a1'].loc[algae_train['a1']>0] = np.log(algae_train['a1'].loc[algae_train['a1']>0]) indexing.html#returning-a-view-versus-a-copy

pd.isnull(algae_train).any()

```
| Season | False | Fal
                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                   False
```

//1.59998 U.UUU NAN 1 std count 0.200 0.300 0.400 0.500 0.600 66.000000 NaN 47.600000 24.688999 31.800000 NaN 42.716667 24.978904 41.633333 12.287121 88.255 0.000000 92.667 7.200000 93.683 12.300000 98.817 1.200000 110.456 0.000000 [131 rows x 3 columns]

EGA: checking pairwise comparisions between each variable of interest
sns.pairplot(algae_train,vars= ['mxPH' ,'mm02' ,'Cl' ,'N03' ,'NH4' ,'oP04' ,'P04' ,'Chla', 'a1'])



False False

algae_train

	season	size	speed	mxPH	mn02	C1	NO3	NH4	oP04	P04	Chla	a1	a2	a3	a4	a5	a6	a7
0	0	0	1	-0.019710	0.287463	3.770155e-01	0.788574	0.039380	0.347256	0.250877	1.817928e+00	0.000000	0.0	0.0	0.0	34.2	8.3	0.0
1	1	0	1	0.568222	-0.470990	3.100197e-01	-0.532115	-0.067408	3.926568	3.287449	-6.393585e- 01	0.336472	7.6	4.8	1.9	6.7	0.0	2.1
2	3	0	1	0.148270	0.961644	-7.943460e- 02	0.546315	-0.079388	0.575746	0.384111	8.218554e-02	1.193922	53.6	1.9	0.0	0.0	0.0	9.7
3	1	0	1	0.097876	-1.819352	7.408577e-01	-0.261574	-0.206962	-0.137187	0.006389	-6.343127e- 01	1.131402	41.0	18.9	0.0	1.4	0.0	1.4
4	3	0	1	0.081078	-0.049627	2.573017e-01	1.903289	-0.137386	-0.169912	-0.314805	-1.751483e- 01	2.219203	2.9	7.5	0.0	7.5	4.1	1.0
195	3	2	1	0.652213	-0.302445	-5.768510e- 01	0.146906	-0.214371	-0.220735	-0.454077	-5.869834e- 01	2.541602	21.7	5.6	0.0	1.0	0.0	0.0
196	1	2	1	0.484232	0.624554	-6.439566e- 01	-0.021982	-0.193022	-0.422968	-0.484118	-4.694676e- 01	2.890372	7.0	1.7	0.0	4.8	10.3	1.0
197	3	2	1	0.316251	-0.892353	2.116468e+00	-0.081213	-0.226508	0.052431	0.018262	8.964706e-01	0.000000	15.9	2.4	1.0	0.0	0.0	0.0
198	0	2	1	-0.019710	-0.639535	1.560767e-16	0.000000	0.000000	0.000000	0.000000	-8.963074e- 17	0.000000	12.5	3.7	1.0	0.0	0.0	4.9
199	2	2	1	0.820193	-1.018762	8.614062e-01	-0.128704	-0.243472	-0.106032	0.020582	2.184211e-01	0.875469	10.5	9.0	7.8	0.0	0.0	5.8
200 rd	ows × 18 c	olumn	3															

So the variables are scaled by the SD and mean to standardize the data. Effectively, doing so transforms all values into z-scores and makes it easier to identify outliers as well as each data point's position relative to the mean.

Additinally, one hot encoding enables the transformation of categorical variables into discrete variables and makes it possible to incorporate them into the pymc model in the next step

Log transformations, as used on the response variable here, help to unskew the data and make its distribution more symmetrical, enabling stronger analysis.

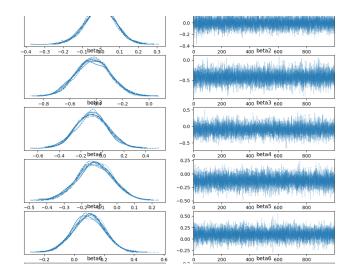
(c) Build a robust Bayesian regression model to predict the response and explain your choices for the priors

graphics_path = '/Users/addyg/Desktop/UVA_Courses/DS6040 Bayes/'

```
with pm.Model() as robust:
beta0 = pm.Normal('beta0', mu=0, sigma=10)
   beta1 = pm.Normal('beta1', mu=0, sigma=10)
    beta2 = pm.Normal('beta2', mu=0, sigma=10)
beta3 = pm.Normal('beta3', mu=0, sigma=10)
   beta4 = pm.Normal('beta4', mu=0, sigma=10)
   beta5 = pm.Normal('beta5', mu=0, sigma=10)
beta6 = pm.Normal('beta6', mu=0, sigma=10)
    beta7 = pm.Normal('beta7', mu=0, sigma=10)
    beta8 = pm.Normal('beta8', mu=0, sigma=10)
beta9 = pm.Normal('beta9', mu=0, sigma=10)
   beta10 = pm.Normal('beta10', mu=0, sigma=10)
    heta11 = pm.Normal('beta11', mu=0, sigma=10'
```

σ = pm.Uniform("σ", 10**-5, 10)

ph = pm.MutableData("ph", algae_train.iloc[:,3])
mn = pm.MutableData("mn", algae_train.iloc[:,4])
cl = pm.MutableData("cl", algae_train.iloc[:,5])



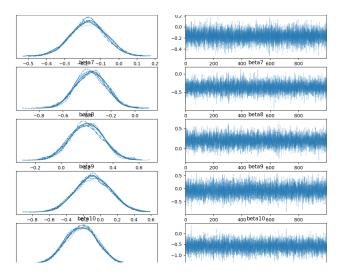


trace_robust.extend(pm.sample_posterior_predictive(trace_robust))

<Axes: title={'center': 'a1 / a1'}>

fig, ax = plt.subplots() az.plot_pc(trace_robust, color = 'b', observed_rug=True, ax=ax)
ax.axvline(algae_train['ai'].mean(), ls="--", color="r", label="Observed mean") ax.legend(fontsize=10);

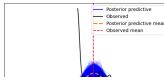
no3 = pm.MutableData("no3", algae_train.iloc[:,6])
nh4 = pm.MutableData("nh4", algae_train.iloc[:,7]) opo4 = pm.MutableData("opo4", algae train.iloc[:.8]) po4 = pm.MutableData("po4", algae_train.iloc[:,9]) chla = pm.MutableData("chla", algae_train.iloc[:,9]) u = pm.Deterministic("u", beta0 + beta1*algae train.iloc(:, 0] + beta2*algae train.iloc(:, 1] + beta3*algae train.iloc(:, 2] + beta4*pl + beta5*mn + beta6*cl + beta7*no3 + beta8*nh4 + beta9*opo4 + beta10*po4 + beta11*chla) alg = pm.StudentT('a1', mu=\mu, sigma=\sigma\), nu=\mu,observed=algae_train['a1']) trace = pm.sampla(cores = 8, return_inferencedata = False, random_seed=1234)
trace_robust = pm.to_inference_data(trace=trace, log_likelihood=True)
robust_pp = pm.sample_posterior_predictive(trace_robust, extend_inferencedata=True, random seed=1234) y_pred_test = robust_pp['posterior_predictive']['al']
az.plot_trace(trace_robust, var_names=['beta0", "beta1", "beta2", "beta3", "beta4", "beta5", "beta6", "beta6", "beta7", "beta8", "beta9", "beta10" "beta11", "o", "v"])



100 00% [8000/8000 00:02<00:00]

/usr/local/lib/python3.18/dist-packages/arviz/pitos/ppcipt.py:24: FutureWarning: color has been deprecated in favor of colors warnings.warm("color has been deprecated in favor of colors", FutureWarning)
//usr/local/lib/python3.18/dist-packages/Python0.core/events.py:85: UserWarning: Creating legend with loc="best" can be slow with large amounts of data. func(*args, **kwargs) (usr/local/lib/python3.10/dist-packages/IPython/core/pylabtools.py:151: UserWarning: Creating legend with loc="best" can be slow with large amounts of data.

fig.canvas.print figure(bytes io, **kw) Posterior predictive - Observed -- Posterior predictive mean --- Observed mean



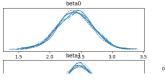
In the above plots, it's apparent that the Bayesian P-values that this robust model created distributions that run clearly to expected distribution of Bayes p-values. The posterior predictive plot demonstrates that the sampling worked well, since the observed and predicted values are similar.

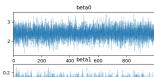
(e) Plot the posterior distributions for the parameters and discuss your results. Plot the response variable vs. one of the predictors using counterfactuals and discuss this plot.

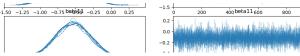
az.plot posterior(trace robust)

100.00% [16000/16000 02:33<00:00 Sampling 8 chains, 0 divergences]

```
[<Axes: title={'center': 'beta1'}>
 <Axes: title={'center': 'beta8'}>1.
 dtype=object)
        beta0
                            beta0
```







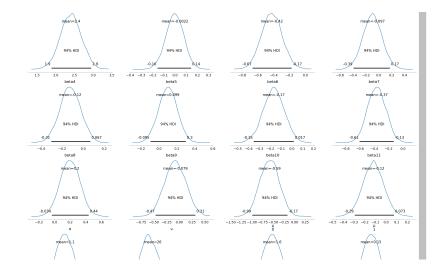
For this model, I chose to use normal priors for the beta distributions because the data has already been scaled in part b and a normal distribution is a reasonable expectation.

10000

(d) Evaluate the sampling and choice of priors using Bayesian p-value and posterior predictive plots. Briefly explain your results.

```
1.0 -Press and analysis the propriet of the second statement to the contract of the contract o
az.plot_bpv(trace_robust, figsize = (10,8))
```

```
/usr/local/lib/pvthon3.10/dist-packages/arviz/plots/plot utils.pv:271: UserWarning: rcParams['plot.max subplots'] (40) is smaller than the number of vari
   <Axes: title={'center': 'beta5'}>,
<Axes: title={'center': 'beta6'}>,
<Axes: title={'center': 'beta7'}>],
                                                      cAses: title=('conter': 'beta7')],
[cAses: title=('conter': 'beta7')],
[cAses: title=('conter': 'beta19'),
cAses: title=('conter': 'beta19'),
[cAses: title=('conter': 'beta19')],
[cAses: title=('conter': 'beta19')],
[cAses: title=('conter': 'bun01'),
[cAses: title=('conter': 'yun01'),
[case
                                                      caxes: title={ center : 'µ\n1'}>],
caxes: title={ center : 'µ\n1'}>],
caxes: title={ center : 'µ\n2'}>,
caxes: title={ center : 'µ\n2'}>,
caxes: title={ center : 'µ\n4'}>,
                                                                   <Axes: title={'center': '\u00e4\n7'}>,
<Axes: title={'center': '\u00e4\n8'}>,
                                                               <Axes: title={'center': 'u\n9'}>1.
                                                   <Axes: title={'center': '\u00e4\n15'}>,
<Axes: title={'center': '\u00e4\n16'}>,
                                                      caxes: title={ center ': "µn17'}>],
caxes: title={ center ': "µn17'}>],
caxes: title={ center ': "µn18'}>,
caxes: title={ center ': "µn18'}>,
caxes: title={ center ': "µn28'}>,
caxes: title={ center ': "µn28'}>,
caxes: title={ center ': "µn22'}>,
[caxes: title={ center ': "µn22'}>,
                                                                   <Axes: title={'center': 'µ\n23'}>,
                                                                   <Axes: title={'center': 'µ\n24'}>,
<Axes: title={'center': 'µ\n25'}>]], dtype=object)
```



```
trace_robust,
var_names=["µ"],
    return_inferencedata=True,
predictions=True.
    extend_inferencedata=True
    random_seed=1234,
_, ax = plt.subplots(figsize=(12, 6))
  ounterfactual = algae_pp_pred.predictions_constant_data['no3']
model_preds = algae_pp_pred.predictions
  counterfactual
  *az.hdi(model_preds)["µ"].transpose("hdi", ...)
ax.plot(
  counterfactual
  model preds["u"].mean(("chain", "draw")).
  color="C1"
  alpha=0.8,
  label="Expected prob.".
 ax.set_xlabel("no3 Z Score")
ax.set vlabel("Algae Z Score")
ax.set title("Counterfactual Predictions")
ax.legend(fontsize=10, frameon=True, framealpha=0.5);
```

```
132 rows * 18 columns

algae_test["msPH'] = np.square(algae_test["msPH'])

with np.Model() as robust_test:

beta0 = pm.Normal("beta0", mu=0, sigma=10)

beta1 = np.Normal("beta1", mu=0, sigma=10)

beta3 = np.Normal("beta3", mu=0, sigma=10)

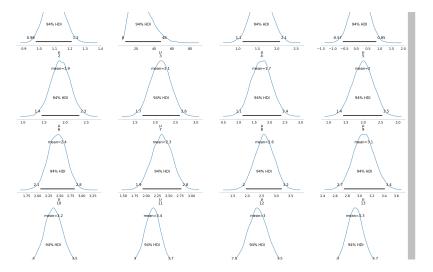
beta4 = pm.Normal("beta4", mu=0, sigma=10)

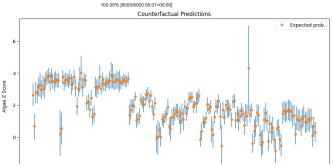
beta5 = np.Normal("beta5", mu=0, sigma=10)

beta6 = pm.Normal("beta6", mu=0, sigma=10)

beta6 = pm.Normal("beta6", 0, 10)

so = pm.Normal("beta6", 0, 10)
```





Though, this is not ideal, a general negative trend can be observed between the no3 Z-score and the algae z-score based on the counterfactual plot.

(f) The data set, algaetest.csv, contains a test set. Build a second model that may contain fewer predictors but should contain at least one nonlinear element. Evaluate both models using mean square error for the test set.

100.00% [16000/16000 01:14<00:00 Sampling 8 chains, 0 divergences] 100.00% [8000/8000 00:02<00:00]

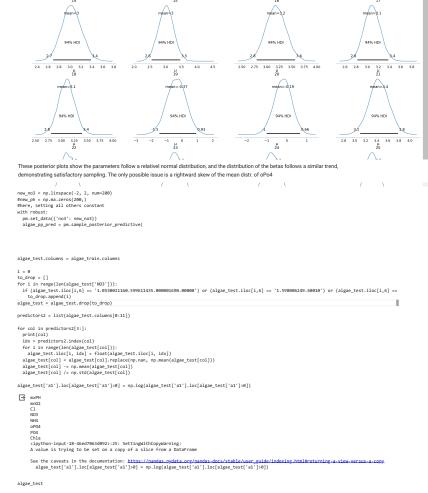
```
#Evaluating MSE
from sklearn.metrics import mean_squared_error
pred = y_pred_test.mean(dim =['chain','draw'])
mse = mean_squared_error(pred, algae_train['a1'])
mse

1.265569866571974

pred_test = test_y_pred.mean(dim =['chain','draw'])
mse_test = mean_squared_error(pred_test, algae_test['a1'])
mse_test = mean_squared_error(pred_test, algae_test['a1'])
mse_test

1.2711914667782464
```

(g) Summarize your results and give your answer to the EU's goal.



Bsed on everything that I've seen in these models, their p-values, predictive performance MSE, it seems that that these would do a decent job in predicting the likelyhood of algae growth, given each of the predictor variables and outcomes. In order to better deal with these algae blooms, the linear model aposers to do a better iob, and I would recommend the EU use such a model when motioring water content.