# **Package**

# February 13, 2019

Type Package
Title Analyze and comapre ONT MinION and GridION X5 1D Sequencing Data
Version 1.0
Author Davide Bolognini, BS, PhD Fellow [aut, cre]
Maintainer Davide Bolognini <davidebolognini7@gmail.com></davidebolognini7@gmail.com>
<b>Depends</b> R ( $>= 3.1.3$ )
Imports parallel, ggplot2 (>= 2.2.1), seqinr, reshape2, scales, RColorBrewer, grid, gridExtra, rhdf5(>= 2.14), ShortRead (>= 1.24.0)  Encoding UTF-8  LazyData true  RoxygenNote 6.0.1
R topics documented:
FastqFilterG
NanoCompare
NanoFastqG
NanoFastqM
NanoPrepareG
NanoPrepareM
NanoStatsG
NanoStatsM
NanoTableG
NanoTableM
Index 10

2 FastqFilterG

FastqFilterG	Filter your GridION X5 .fastq files	

# **Description**

Use this function if you have .fastq and sequencing summary files and you want to filter your .fastq files in order to obtain only the high-quality ones. FastqFilterG can optionally return a .fasta file for the high-quality sequences. FastqFilterG can optionally return a total .fastq file (equivalent to a 'cat' shell command) and translate into .fasta

# Usage

```
FastqFilterG(Data, DataOut, FASTQTOT = FALSE, FASTA = FALSE, Cores = 1,
    Label)
```

### **Arguments**

Data	Path to .fastq and sequencing summary files returned from GridION X5 (can find .fastq and sequencing summary files recursively)
DataOut	Where the .fastq (and, optionally, .fasta) file will be saved
FASTQTOT	Logical. If TRUE, combine all the .fastq together and store in the DataOut folder. Default to FALSE
FASTA	Logical. If FALSE, return only .fastq file else, if TRUE, return both .fastq and .fasta files. Default to FALSE
Cores	Number of cores to use to accelerate sequencing summary files reading. Default to 1
Label	Llabel to use, together with the Flow Cell identifier, to identify the experiment

# Value

High-quality .fastq file and, optionally, high quality .fasta file, total .fastq file and total .fasta file. If one or more .fastq file is "ill-formatted", FastqFilterG stops and prints the number of the "guilty" .fastq file.

# **Examples**

```
#do not run
DataPath<-"/data/basecalled/ExperimentName/FlowCellId"
FastqFilterG(Data=DataPath, DataOut="Path/To/DataOut",FASTQTOT=FALSE,FASTA=FALSE)</pre>
```

NanoCompare 3

NanoCompare	Compare ONT experiments	

# Description

NanoCompare plots comparison statistics for MinION and GridION X5 experiments analyzed with the other functions from this package

# Usage

```
NanoCompare(DataIn, DataOut, Labels, GCC = TRUE)
```

# Arguments

DataIn Character vector containing paths to folders containing analyzed MinION and/or

GridION X5 experiments

DataOut Where to save NanoCompare results

Labels Character vector containing ordered labels used to identify experiments in DataIn

### Value

### Plots:

- Violins.pdf;
- Histograms.pdf;

# **Examples**

```
#do not run
DataIn<-c("Path/To/AnalyzedFolder1","Path/To/AnalyzedFolder2",...)
Labels<-c("Label1","Label2","Label3") #labels used
NanoCompare(DataIn=DataIn,DataOut="Path/To/DataOut",Labels=Labels,GCC=TRUE) #compare</pre>
```

NanoFastqG	Extracts .fastq informations from your GridION X5 basecalled passed
	.fast5 files

# **Description**

NanoFastqG returns a .fastq file (and, optionally, a .fasta file) for your high-quality reads

# Usage

```
NanoFastqG(DataPass, DataOut, Label, Cores = 1, FASTA = FALSE)
```

4 NanoFastqM

### **Arguments**

DataPass Path to passed .fast5 files folder

DataOut Where .fastq (and, optionally, .fasta) file will be saved

Label Label used, together with the Flow Cell identifier extracted from the inputted

data, to identify .fastq (and, optionally, .fasta) file

Cores Number of cores to be used: 1 by default

FASTA Logical. If FALSE, return only .fastq file else, if TRUE, return both .fastq and

.fasta files. Default to FALSE

### Value

.fastq file and, optionally, .fasta file for passed .fast5 files

### **Examples**

#do not run

NanoFastqG(DataPass="Path/To/DataPass", DataOut="/Path/To/DataOutExp", Cores=6, FASTA=FALSE) NanoFastqG(DataPass="Path/To/DataPass", DataOut="/Path/To/DataOutExp", Cores=6, FASTA=TRUE)

NanoFastqM Extracts .fastq informations from your MinION passed .fast5 files

# Description

NanoFastqM returns a .fastq file (and, optionally, a .fasta file) for your high-quality reads

### Usage

NanoFastqM(DataPass, DataOut, Label, Cores = 1, FASTA = FALSE)

# **Arguments**

DataPass Path to passed .fast5 files folder

DataOut Where .fastq (and, optionally, .fasta) file will be saved Label Label used to identify .fastq (and, optionally, .fasta) file

Cores Number of cores to be used: 1 by default

FASTA Logical. If FALSE, return only .fastq file else, if TRUE, return both .fastq and

.fasta files. Default to FALSE

### Value

.fastq file and, optionally, .fasta file for passed .fast5 files

### **Examples**

```
#do not run
```

NanoFastqM(DataPass="Path/To/DataPass", DataOut="/Path/To/DataOutExp", Cores=6, FASTA=FALSE) NanoFastqM(DataPass="Path/To/DataPass", DataOut="/Path/To/DataOutExp", Cores=6, FASTA=TRUE) NanoPrepareG 5

NanoPrepareG Prepares GridION X5 data for your analyses with NanoR	_
--	---

### **Description**

NanoPrepareG generates an object of class list that contains informations required by other functions from NanoR when analyzing GridION X5 data.

### Usage

```
NanoPrepareG(BasecalledFast5 = FALSE, Data, DataFail = NA, DataSkip = NA,
Cores = 1, Label)
```

### **Arguments**

	_		_				_
Basecal		led	H	а	51	t	5

Logical. TRUE if dealing with basecalled .fast5 files. Defaulto to FALSE

Path to GridION X5 folder containing fastq and sequencing summary files (if

BasecalledFast5 = FALSE) or to basecalled .fast5 files (if BasecalledFast5 =

TRUE)

DataFail Path to failed .fast5 files folder
DataSkip Path to skipped .fast5 files folder

Cores Number of cores to be used to accelerate sequencing summary files reading

(useful only if BasecalledFast5 = FALSE)

Label Label used, together with the Flow Cell identifier extracted from the inputted

data, to identify your experiment: do not use underscore characters ("\_").

# Details

NanoPrepareg can find desired inputs recursively. DataSkip and DataFail can be omitted.

## Value

Object of class list containing informations required by NanoTableG and NanoStatsG functions.

### **Examples**

```
#do not run
#when working with sequencing summary files and .fastq files
Data<-"/data/basecalled/ExperimentName/FlowCellId"
NanoGList<-NanoPrepareG(BasecalledFast5=FALSE, Data=Data, Label="Ex", Cores=3)
#when working with basecalled .fast5 files
Pass<-"Path/to/workspace/pass"
Fail<-"Path/to/workspace/fail"
NanoGList<-NanoPrepareG(BasecalledFast5=TRUE, Data=Pass, DataFail=Fail, Label="Ex")</pre>
```

NanoPrepareM

NanoPrepareM	Prepares MinION data for your analyses with NanoR
·	

# **Description**

NanoPrepareM generates an object of class list that contains informations required by other functions from NanoR when analyzing MinION data

# Usage

```
NanoPrepareM(DataPass, DataFail = NA, DataSkip = NA, Label)
```

# Arguments

DataPass	Path to MinION passed .fast5 files folder
DataFail	Path to MinION failes .fast5 files folder
DataSkip	Path to MinION skipped .fast5 files folder
Label	Label to identify your MinION experiment

# **Details**

NanoPreareM can find .fast5 files recursivel. DataFail and DataSkip can be omitted (MinKNOW generates passed, failes and skipped .fast5 files folders but failed and skipped .fast5 files are taken into account only for calculating their number and percentage)

# Value

Object of class list

# **Examples**

```
#do not run
PathPass<-"/Path/To/PassFast5"
Lab<-"Exp"
NanoMList<-NanoPrepareM(DataPass=PathPass, Label=Lab)</pre>
```

NanoStatsG 7

NanoStatsG	Plots statistics for your GridION X5 .fast5 files

# Description

NanoStatsG plots statistics for passed .fast5 files and returns 4 tables used by NanoCompare

### Usage

NanoStatsG(NanoPrepareGList, NanoGTable, DataOut)

### **Arguments**

NanoPrepareGList

Object of class list returned by NanoPrepareG

NanoGTable Table returned by NanoTableG

DataOut Where NanoStatsG results will be saved. Use the same directory specified for

NanoTableG function and be sure that it doesn't already contain NanoStatsM

(or NanoStatsG) results

### Value

### Plots:

- Cumulative\_Reads\_&\_Cumulative\_Basepairs.pdf;
- Reads\_Basepairs\_Length\_Quality.pdf;
- Length\_versus\_Quality.pdf;
- Pass\_Fail\_Skip\_and\_GC\_Content.pdf or Pass\_Fail\_Skip\_NO\_GC\_Content.pdf;
- Channels\_Activity.pdf or Channels\_and\_Muxes\_Activity.pdf. Not-working channels and muxes are grey-colored.

### **Examples**

```
#do not run
#knows how to deal with different inputs type autonomously
NanoStatsG(NanoPrepareGList=NanoGList, NanoGTable=NanoGTable, DataOut="/Path/To/DataOutEx")
```

NanoStatsM Plots statistics for your MinION .fast5 files

# **Description**

NanoStatsM plots statistics for passed .fast5 files and returns 4 tables used by NanoCompare

### Usage

NanoStatsM(NanoPrepareMList, NanoMTable, DataOut)

8 NanoTableG

### **Arguments**

NanoPrepareMList

Object of class list returned by NanoPrepareM

NanoMTable Table returned by NanoTableM

DataOut Where NanoStatsM results will be saved. Use the same directory specified for

NanoTableM function and be sure that it doesn't already contain NanoStatsM

(or NanoStatsG) results

### Value

#### Plots:

- Cumulative\_Reads\_&\_Cumulative\_Basepairs.pdf;

- Reads\_Basepairs\_Length\_Quality.pdf;

Length\_versus\_Quality.pdf;

- Pass\_Fail\_Skip\_and\_GC\_Content.pdf or Pass\_Fail\_Skip\_NO\_GC\_Content.pdf;

- Channels and Muxes Activity.pdf. Inactive channels and muxes are grey-colored

### **Examples**

#do not run

NanoStatsM(NanoPrepareMList=NanoMList, NanoMTable=NanoMTable, DataOut="/Path/To/DataOutExp")

NanoTableG

Generates an information table for your GridION X5 .fast5 files

# **Description**

NanoTableG generates a table that contains useful informations for each read identified by NanoPrepareG function. When analyzing GridION X5 basecalled .fast5 files, metadata extraction can be accelerated using multiple cores. Sometimes .fastq files returned by GridION X5 can be ill-formatted: in this case, NanoTableG will stop and you can run this function again after set GCC to FALSE. This problem can be avoided if basecalled .fast5 files are used.

## Usage

NanoTableG(NanoPrepareGList, DataOut, Cores = 1, GCC = TRUE)

### **Arguments**

NanoPrepareGList

Object of class list returned by NanoPrepareG function

DataOut Where the table will be saved. Do not use a directory that already contains a

NanoTableM (or NanoTableG) result

Cores Number of cores to be used: 1 by default. Does not affect time when dealing

with .fastq and sequencing summary files.

GCC Logical. If TRUE, NanoTableM computes GC content for each read. Default to

**TRUE** 

NanoTableM 9

### Value

Table with 7 columns

### **Examples**

```
#do not run
DataOut <- "/Path/To/DataOutEx"
#when working with sequencing summary files and .fastq files
NanoGTable<-NanoTableG(NanoPrepareGList=NanoGList, DataOut=DataOut) #set GCC to "FALSE" on error
#when working with basecalled .fast5 files
NanoGTable<-NanoTableG(NanoPrepareGList=NanoGList, DataOut=DataOut, GCC=TRUE, Cores=6)</pre>
```

NanoTableM

Generates a metadata table for your MinION .fast5 files

## Description

NanoTableM generates a table that contains useful informations for each read of the "pass" .fast5 files folder given to NanoPrepareM function. As NanoTableM can take some time (it depends on the number of reads it has to deal with), this function can be accelerated using multiple cores

# Usage

```
NanoTableM(NanoPrepareMList, DataOut, Cores = 1, GCC = TRUE)
```

# **Arguments**

NanoPrepareMList

Object of class list returned by NanoPrepareM

DataOut Where the table will be saved. Do not use a directory that already contains a

NanoTableM (or NanoTableG) result

Cores Number of cores to be used: 1 by default

GCC Logical. If TRUE, NanoTableM computes GC content for each read. Default to

**TRUE** 

### Value

Table with 7 columns

# **Examples**

#do not run

NanoMTable < -NanoTable M(NanoPrepareMList=NanoMList, DataOut="/Path/To/DataOutExp", Cores=6, GCC=TRUE)

# **Index**

FastqFilterG, 2
NanoCompare, 3
NanoFastqG, 3
NanoFastqM, 4
NanoPrepareG, 5
NanoPrepareM, 6
NanoStatsG, 7
NanoStatsM, 7
NanoTableG, 8
NanoTableM, 9