

Aligning reads: tools and theory



Genome

chrX: 52139280 152139290 152139300 152139310 152139320 152139330  
--->CGCCGTCCCTCAGAAATGGAAACCTCGCTTCTCTCTGCCCCACAATGCGCAAGTCAG

Sequence read

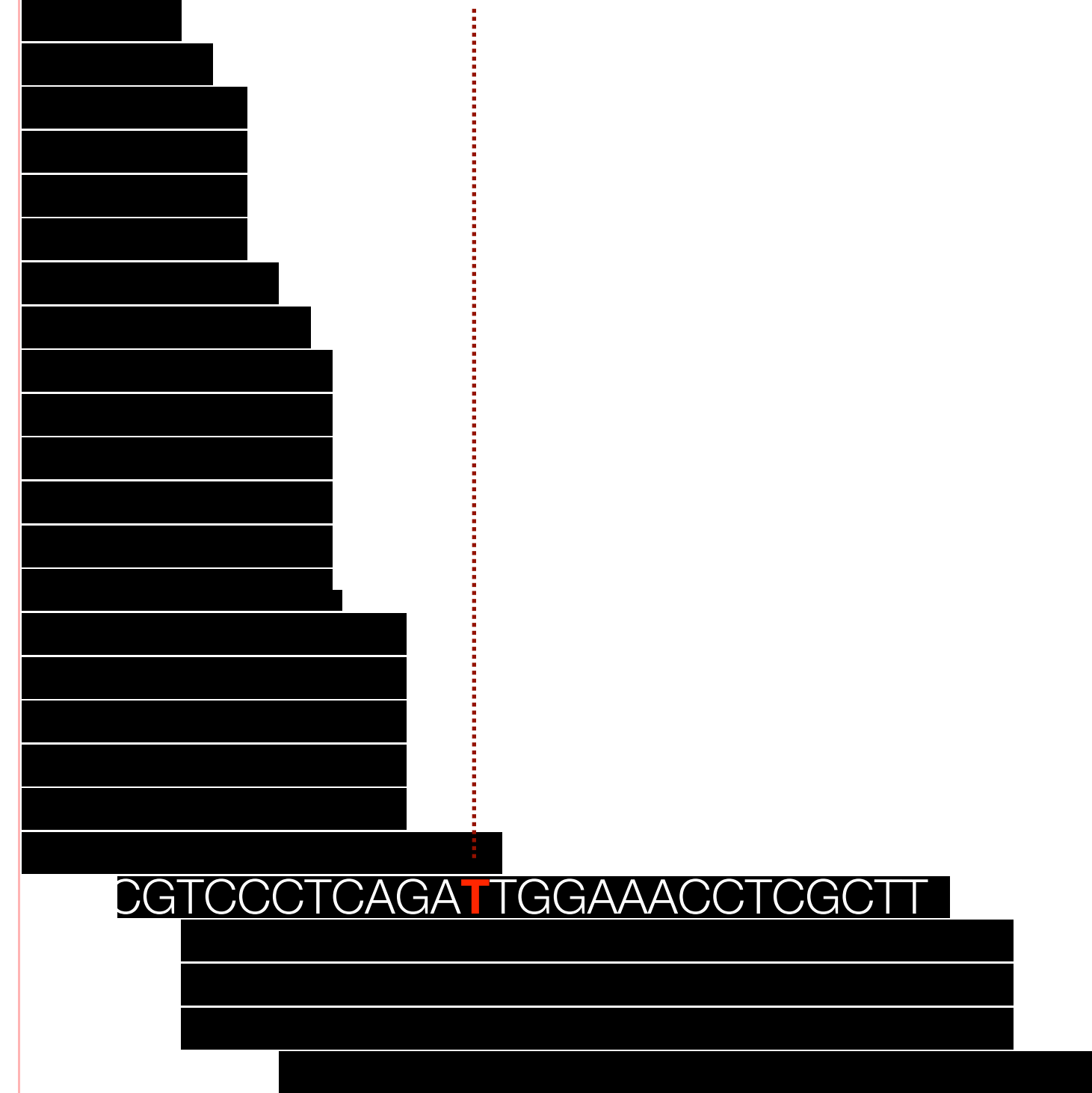
CGTCCCTCAGAAATGGAAACCTCGCTT

A simple case of string matching

Genome

chrX: 52139280 152139290 152139300 152139310 152139320 152139330  
--->CGCCGTCCCTCAGAAATGGAAACCTCGCTTCTCTCTGCCCCACAATGCGCAAGTCAG

Sequence reads



Difficult in practice

- Volume of data: ~3 Gbp
- ~50% of genome is **repeat regions** that cannot be covered by reads
  - Simple repeats, tandem, interspersed
  - Transposons
  - Segmental duplications where mapping is unclear
- **Gaps or unfinished regions**
  - peri-centromere, sub-telomere
  - ~5Mb unique to ethnic groups (e.g., African, Asian)

Challenges:

Human genome is large and complex<sup>4</sup>

- Short reads: 50-150 bp (versus a very long reference)
  - Non-unique alignment
  - Sensitive to sequencing errors
- Massive amount of short reads: one lane produces  $\geq 150$  million 100 nucleotide reads
- Small insert size: 200-500 bp libraries

## Challenges: short read NGS data

**Reference**    ATCTCCATAGGACTAGGAAGTAG

Substitution    ATCTCCATAG**C**ACTAGGAAGTAG

Deletion        ATCTCCATAGGAC**-**AGAAGTAG

Insertion        ATCTCCATAGGACTAGGAAGT**T**AG

3bp deletion    ATCTC**---**AGGACTAGGAAGTAG

Challenges: non-exact matching

# Local alignment vs Global alignment

- ▶ **Local alignment** matches the query with a *substring* (k-mer) of the reference
  - ▶ Tailored towards finding *regions of highly similar sequence* and aligning around those by working outwards to align the rest

## Local Alignment

```
5' ACTACTAGATTACTTACGGATCAGGTACTTTAGAGGCTTGCAACCA 3'
      |||| ||||| ||||| ||||| |||||
5' TACTCACGGATGAGGTACTTTAGAGGC 3'
```

## Global Alignment

```
5' ACTACTAGATTACTTACGGATCAGGTACTTTAGAGGCTTGCAACCA 3'
||||| ||||| ||||| ||||| |||||
5' ACTACTAGATT----ACGGATC--GTACTTTAGAGGCTAGCAACCA 3'
```

- ▶ A **global alignment** performs end-to-end alignment between the query and the reference

**Reference**    ATCTCCATAGGACTAGGAAGTAG

Substitution    ATCTCCATAG**C**ACTAGGAAGTAG

Deletion        ATCTCCATAGGAC**-**AGGAAGTAG

Insertion        ATCTCCATAGGACTAGGAAGT**T**AG

3bp deletion    ATCTC**---**AGGACTAGGAAGTAG

General concepts: edit distance

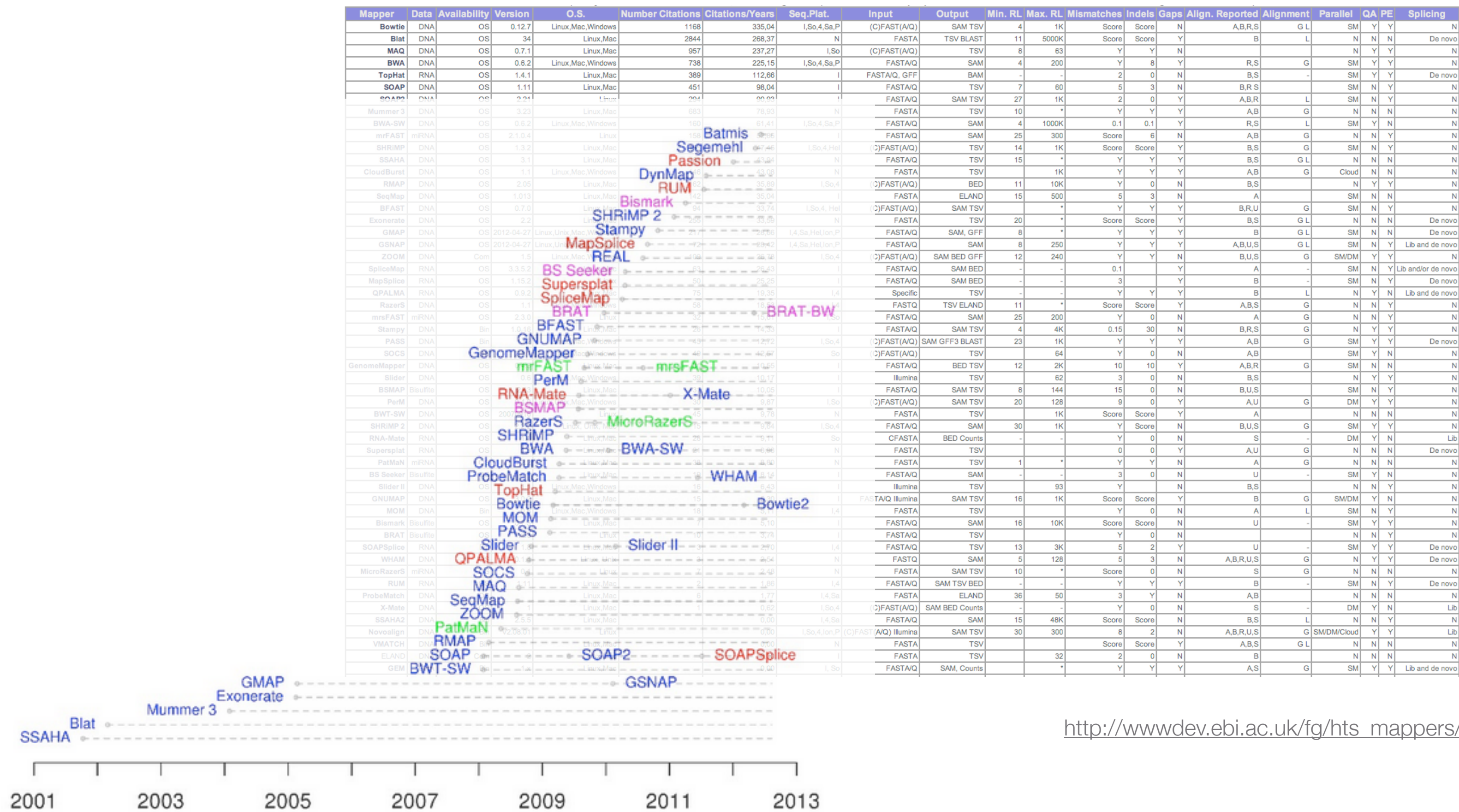


Reference CGTCCCTCAGATTGGAA—CCTCGCTT

Read TCCCTCAGAATGGAAACCTCGCT

Edit distance =3

General concepts: edit distance



[http://wwwdev.ebi.ac.uk/fg/hts\\_mappers/](http://wwwdev.ebi.ac.uk/fg/hts_mappers/)

# Short-read aligners: choices



# Building an index

- ▶ Having an index of the reference genome provides an efficient way to search
- ▶ Once index is built, it can be queried any number of times
- ▶ Indexes are genome and tool-specific
- ▶ Different types of indices (i.e hash-tables, suffix arrays, Burrows-Wheeler Transform)



- ▶ Bowtie2: indexes with an FM Index to keep low memory footprint. Supports gapped, local and paired-end alignment
- ▶ BWA: indexes with the Burrows-Wheeler Transform (BWT). Has three algorithms for varying read lengths.
- ▶ SOAP2: uses a 2-way BWT for indexing. Fast and accurate alignment of Illumina sequencing reads. Not open source.
- ▶ MAQ: first aligns reads to reference sequences and then calls the consensus. Designed for Illumina reads.

Commonly used aligners for ChIP-seq



# Alignment considerations for ChIP-seq

- ▶ Percentage of uniquely mapped reads
  - ▶ 70% or higher is good. 50% or lower is concerning.
  - ▶ Percentages vary between organisms
- ▶ Duplicates
  - ▶ Usually due to over-amplification or short read length
- ▶ Multi-mappers
  - ▶ Reduce by setting allowable mismatches according to sequencing platform
  - ▶ For situations when protein binds repetitive DNA, use paired-end sequencing

**Keep only uniquely mapped reads for peak calling!**

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