

Bioinformatic approaches to regulatory genomics and epigenomics


376-1347-00L - 2022

Pierre-Luc Germain

Plan for today

- About your lecturer
 - About your fellow students
 - What's epigenetics/epigenomics?
 - Structure of the course
 - Expectations and evaluations
-
- Introduction to the practical tools for the course:
 - R notebooks, bioconductor, git & github

About your lecturer

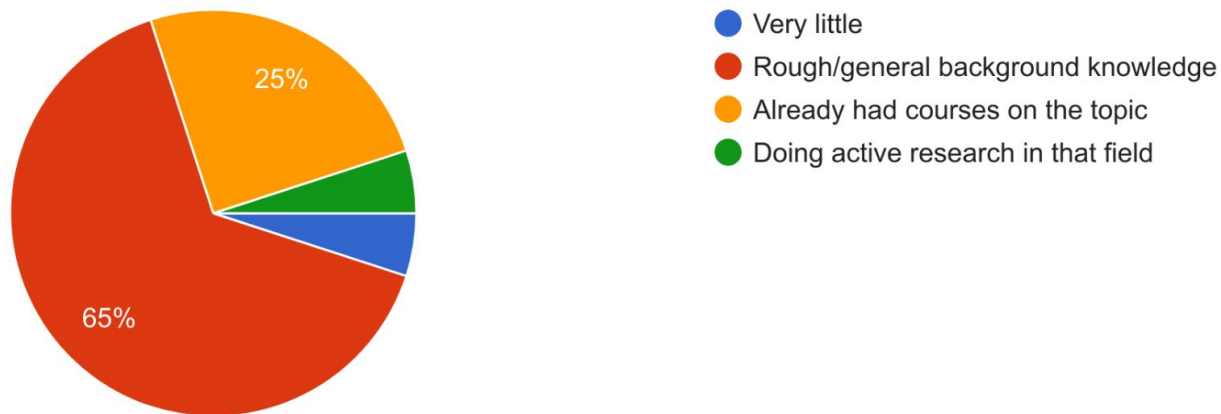
- Oberassistent & Junior PI at the D-HEST Institute for Neurosciences (ETH) & Lab of Statistical Bioinformatics (UZH)
- From Quebec, Canada 
- Background in the humanities (history and philosophy of the life sciences):
 - biological functions & evolutionary explanations, animal experimentation, modeling & extrapolation, molecular oncology, etc.
- Biological research:
 - gene expression regulation in stem cells, germline and the brain; brain and cellular responses to stress; genetics of neurodevelopmental disorders
- Bioinformatic research:
 - methods for bulk and single-cell transcriptomics and epigenomics; miRNA target prediction; TF binding & activity inference, reconstruction of transcriptional networks

About your fellow students

Chiefly master students, a few doctoral students and others

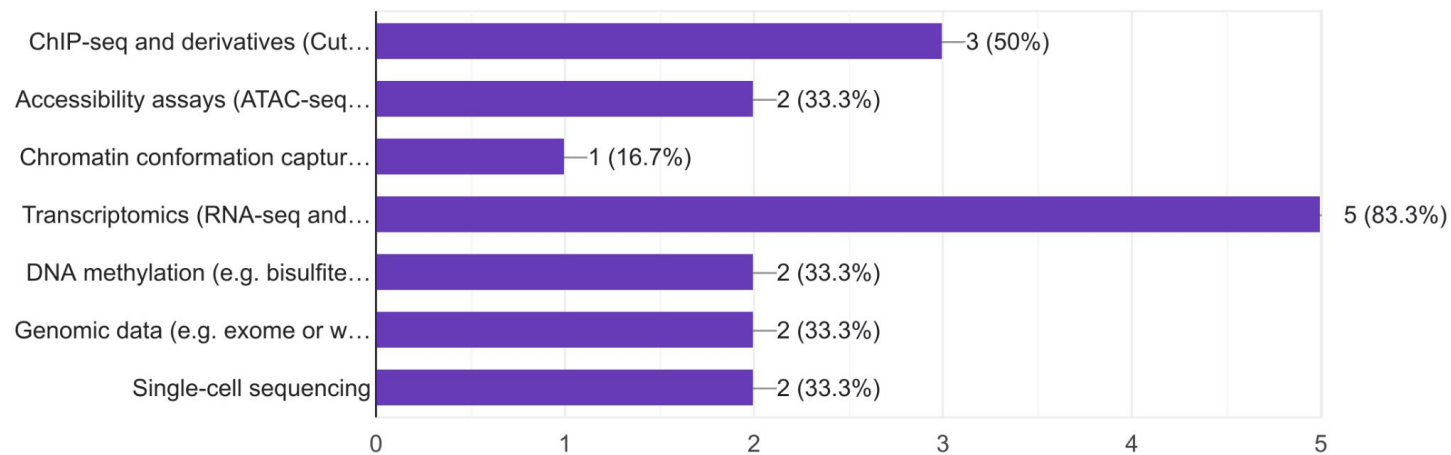
Prior knowledge about epigenetics & regulatory genetics

20 responses



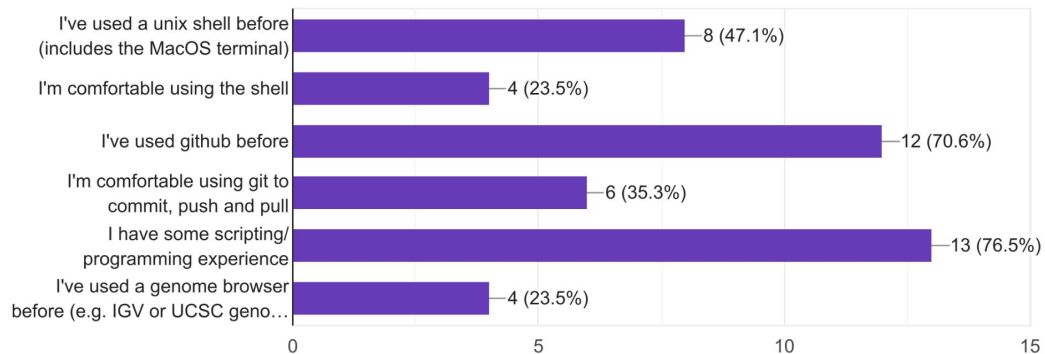
If you have worked with Next Generation Sequencing data before, what kind?

6 responses



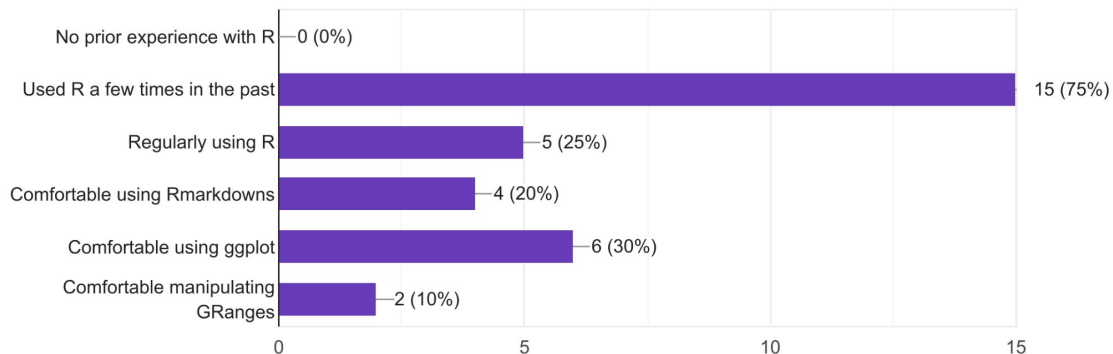
General computer skills; please check all that are relevant

17 responses



Prior experience with R/Bioconductor; please check all that are relevant

20 responses



Some expectations given by your fellow students

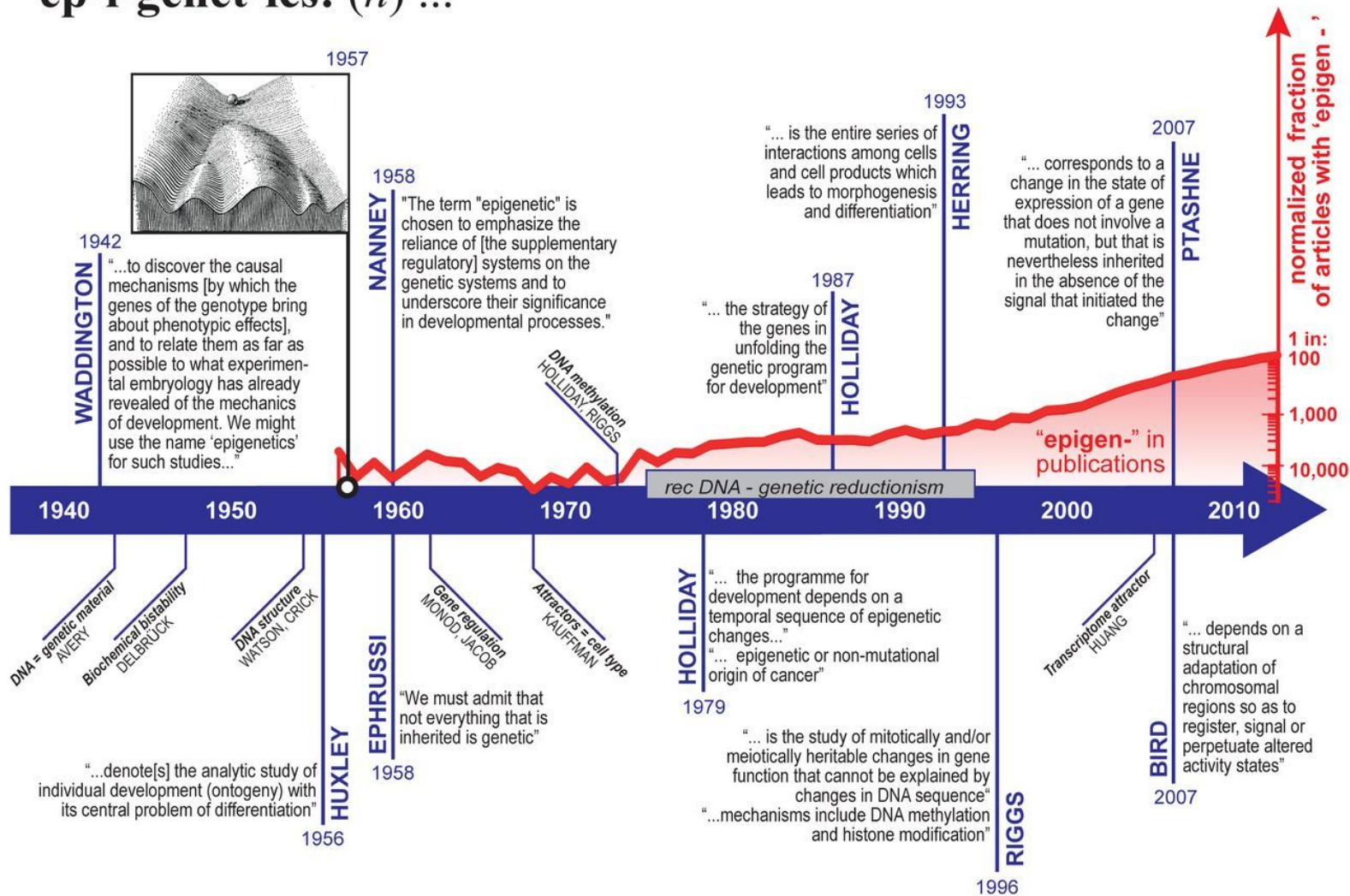
- “I hope to get more comfortable with the standard tools and procedures of epigenetics analysis. Also getting some useful tips and tricks regarding programming.”
- “get some general knowledge about how to make use of genome browsers”
- “Getting acquainted with analysis of regulatory data”
- “Become more comfortable with using R, learning how to extract information from the genomic data”
- “multi-omics knowledge, how to integrate those, and gene regulation dynamics”

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ep•i•genet•ics: (n) ...

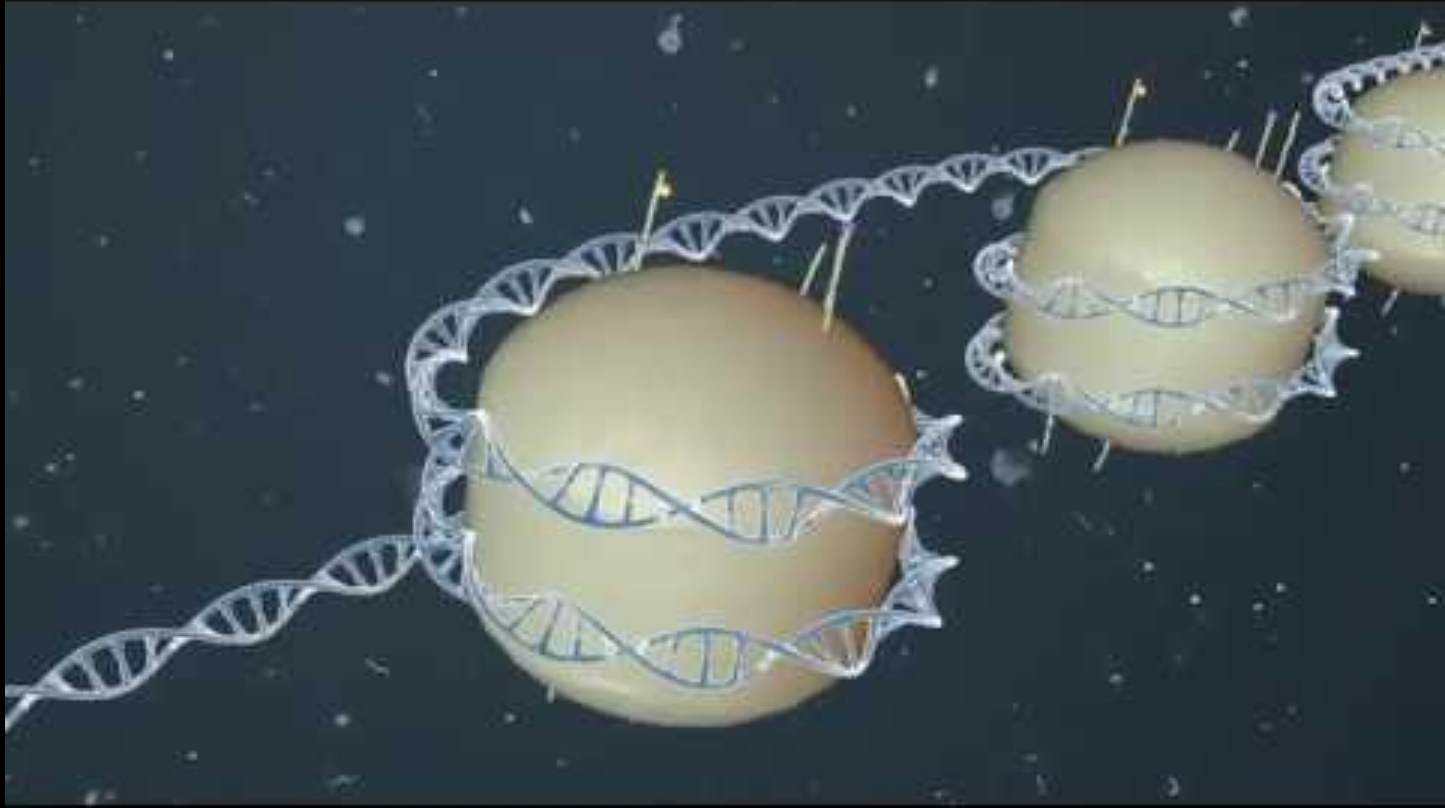


(Oliveira Pisco, Fouquier d'Hérouël and Huang, 2016)

Epigenetic(s)

- Waddington (1942) :
“the causal interactions between genes and their products which bring the phenotype into being”
- Ptashne (2007) (also Nanney 1958):
"A change in the state of expression of a gene that does not involve a mutation, but that is nevertheless **inherited in the absence of the signal** (or event) that initiated the change."
- Bird (2007) and common contemporary usage in molecular biology:
"molecular **modifications of DNA and chromatin** that do not alter the sequence"

Basic primer on epigenetics



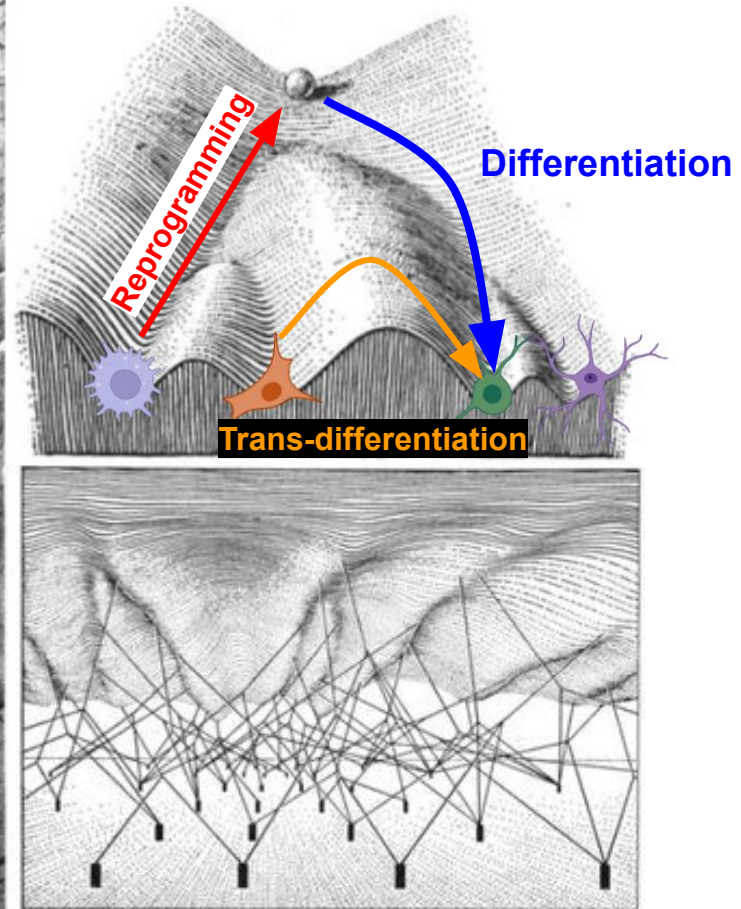
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Waddington's epigenetic landscape

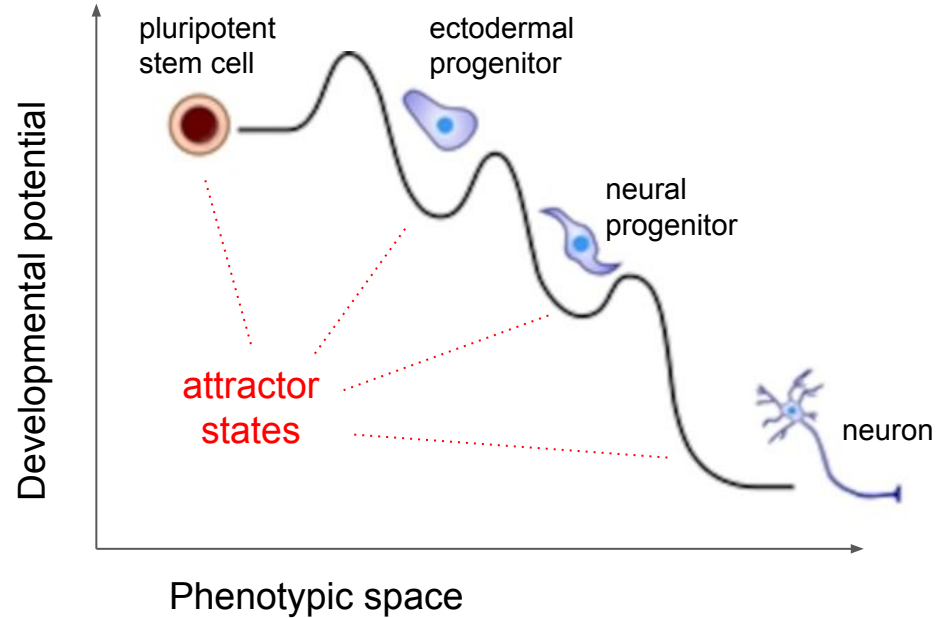
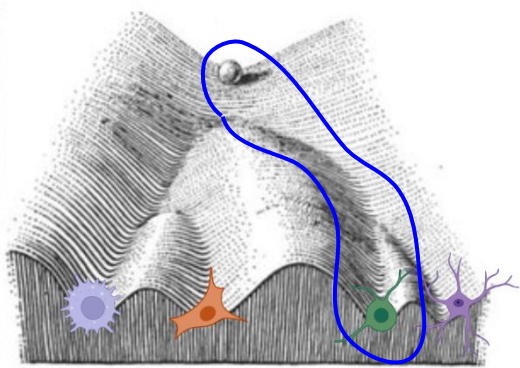


Left from Waddington (1940) "Organisers & Genes"



Right from Waddington (1957) "The strategy of the genes"

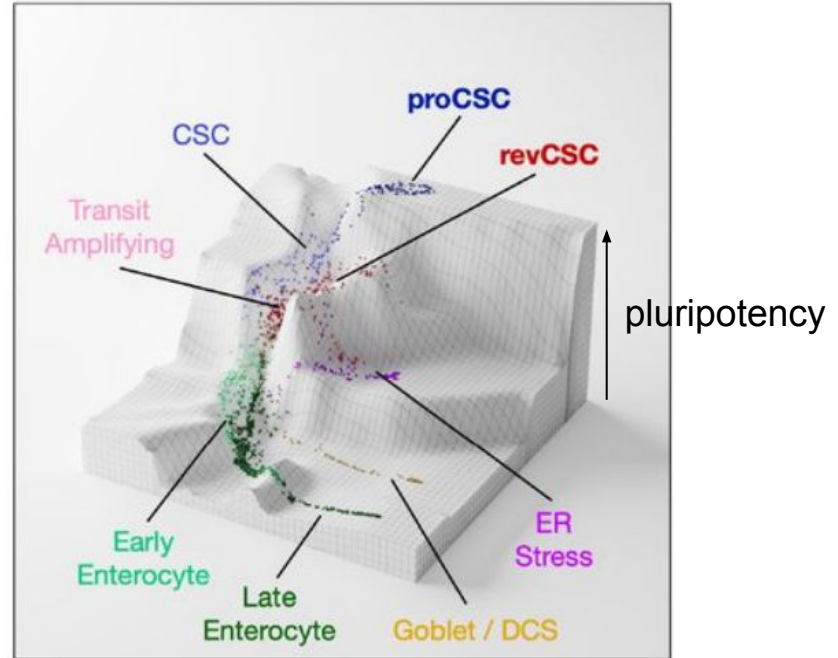
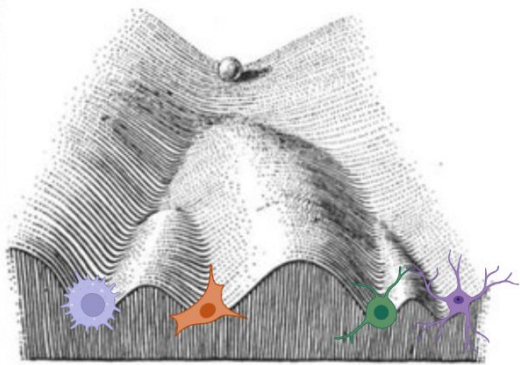
Maintaining and changing cell identity



(Adapted from an original figure by KT Rodolfa 2008)

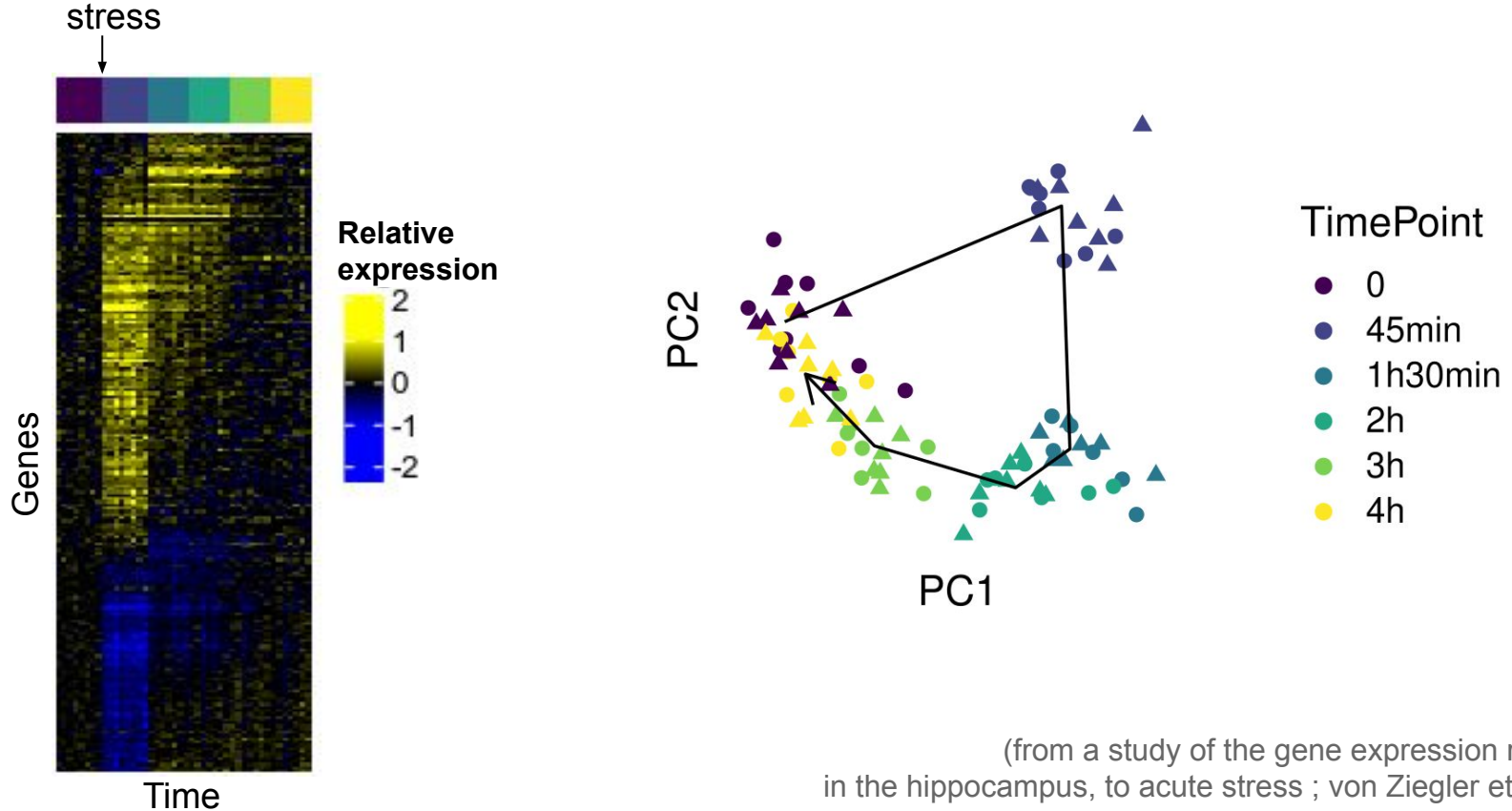
Maintaining and changing cell identity

From concept to quantitative measurement



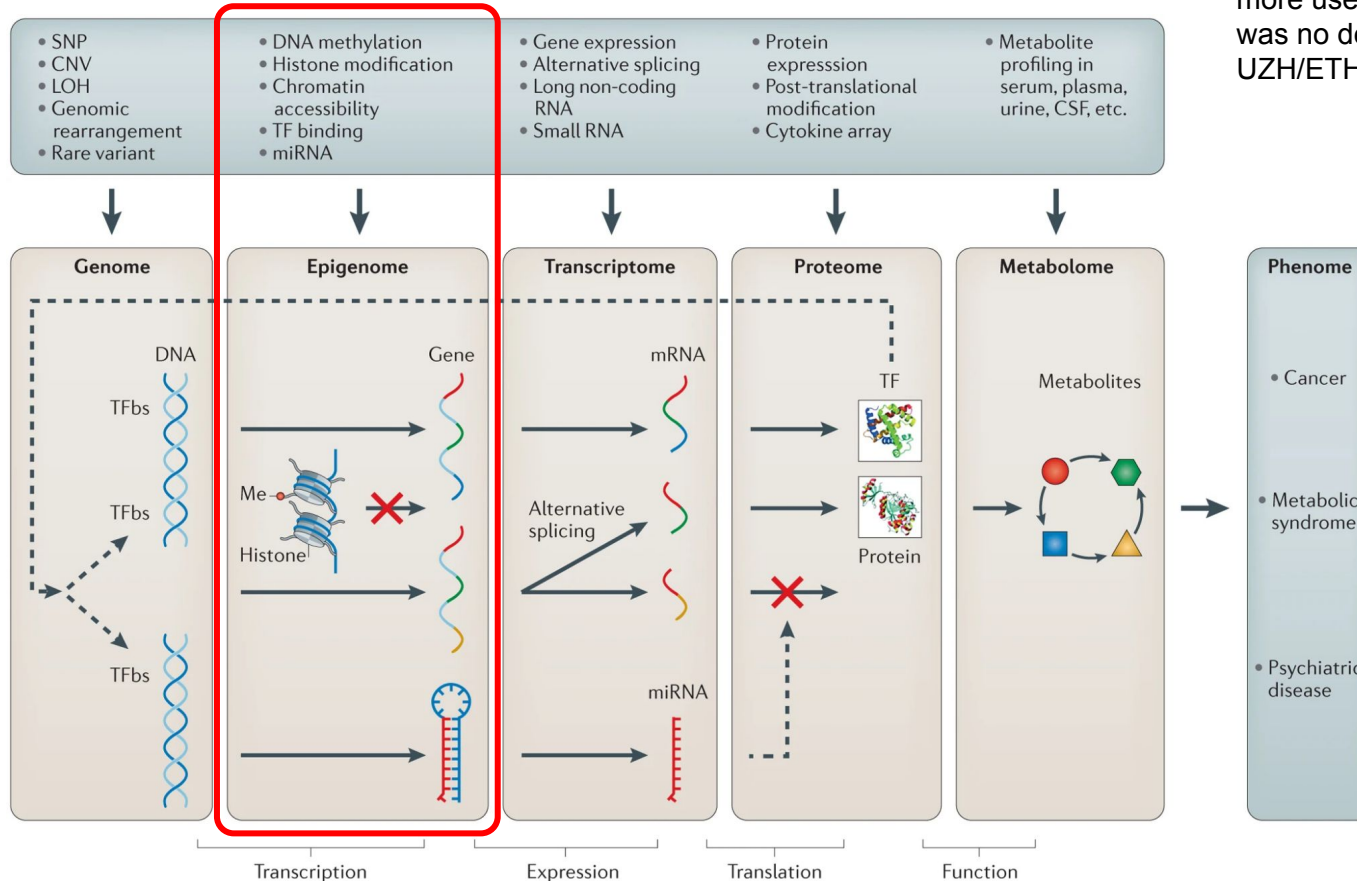
(adapted from
Qin et al., [bioRxiv 2023](#))

Regulating cell-type-specific responses to stimuli



Situating the course

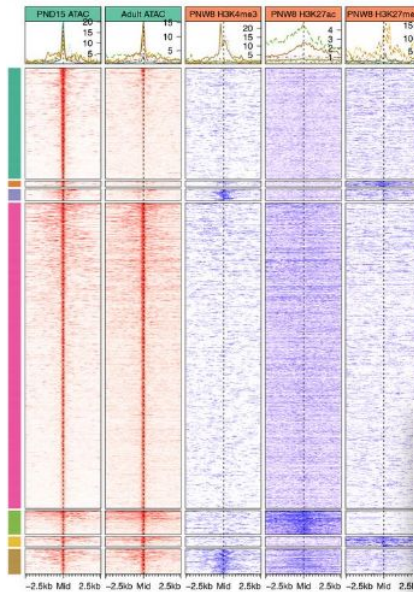
While genomics and transcriptomics are much more used in the clinics, until this course there was no dedicated teaching on epigenomics at UZH/ETH.



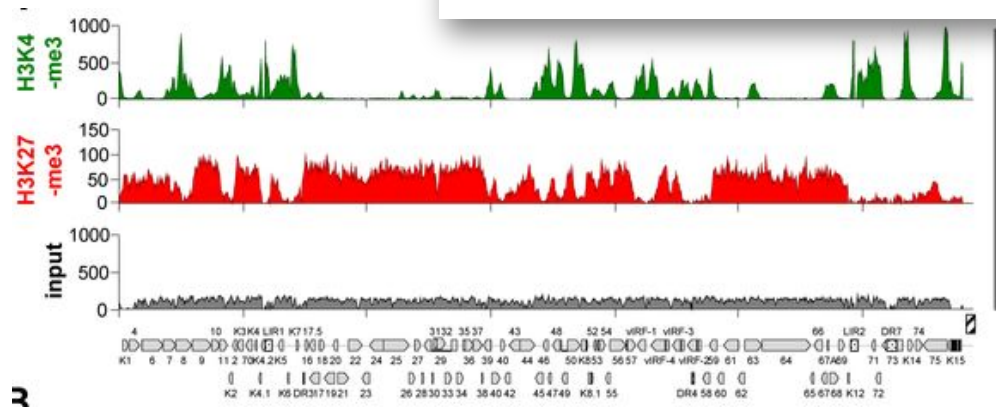
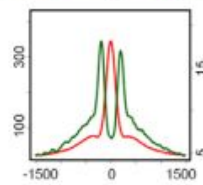
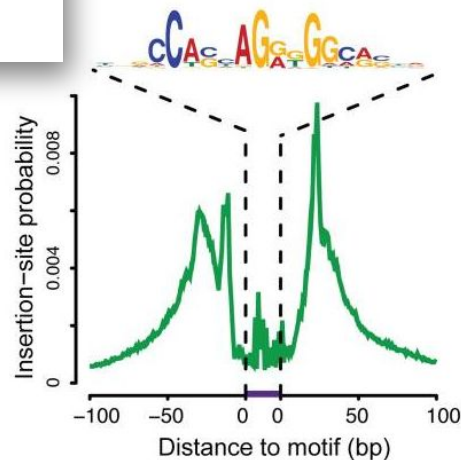
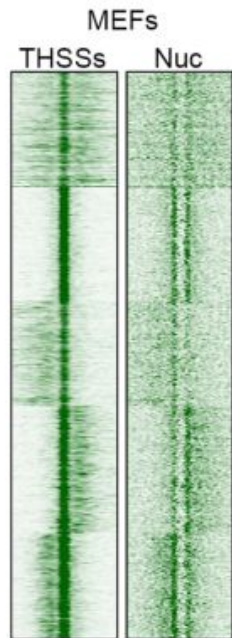
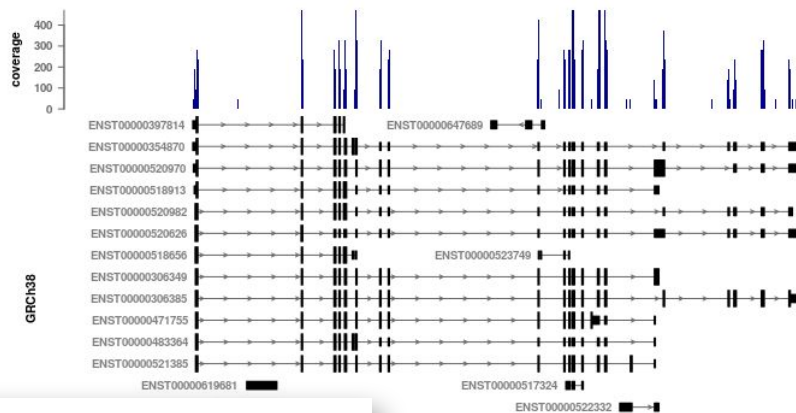
Students interested in transcriptomic analysis may want to follow the excellent 401-6282-00L (UZH Sta426) course (emphasis on stats)

Aims of the course

1. to enable students to be, if not wholly independent with respect to epigenomics data analysis, at least able to **autonomously explore, visualize and interpret** such data;
2. to understand and critically appraise, from a genomics perspective and through **hands-on data exploration**, the key **concepts underlying chromatin regulation** of transcription and its impact on various biological phenomena.



How does one make plots like these, and what do they tell us?

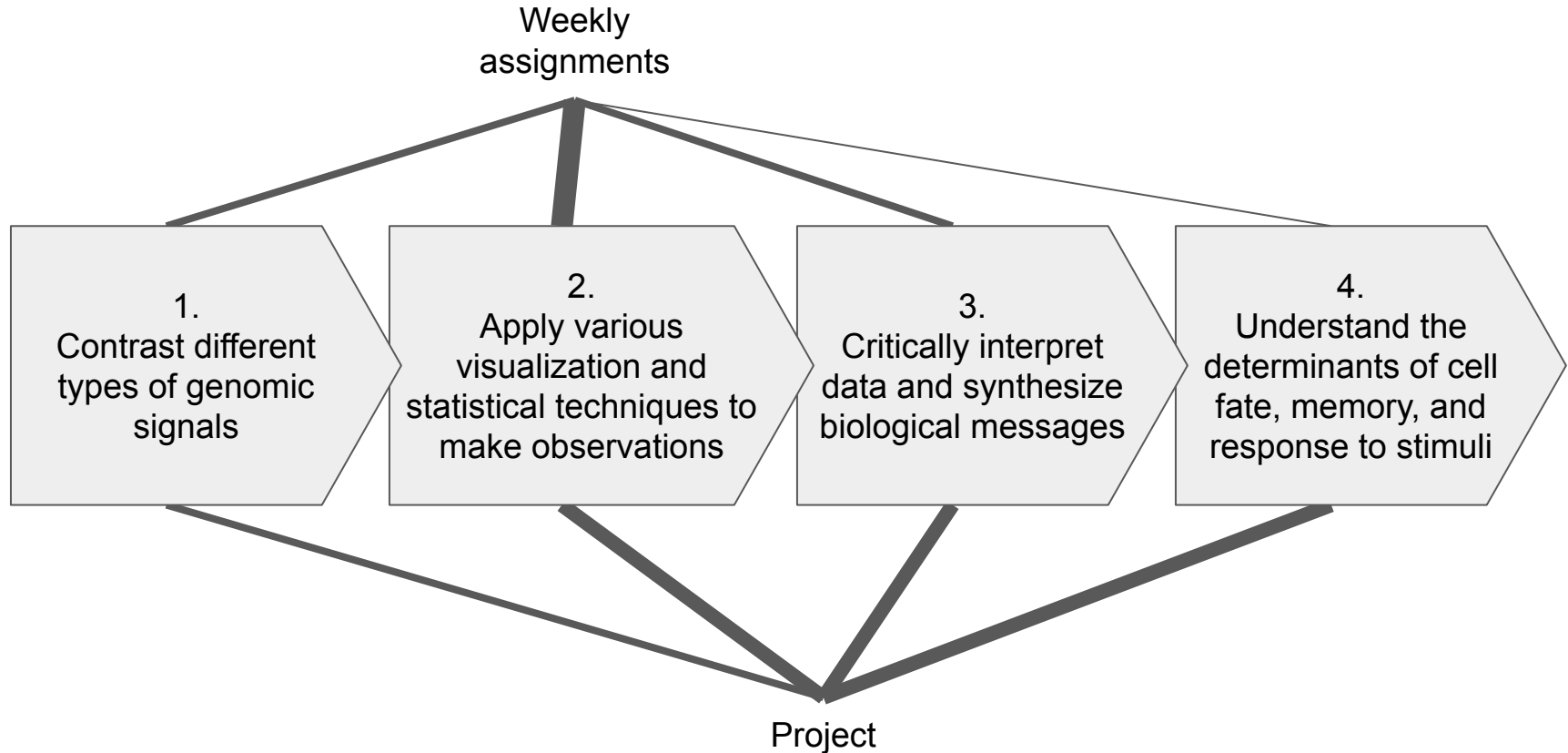


BCBL1

Grading and expectations

- 50% of the grade is based on **weekly exercises**
 - Exercises should be **submitted via github**, by thursday noon the following week
 - The best half of the exercises will make up the grade
- 50% of the grade is based on the **project** (alone or in groups of 2-3)
 - The project can be either:
 - Re-producing the analyses from a publication (in a critical fashion)
 - Analyzing new data (e.g. yours or in collaboration with a group)
 - The project *must be discussed and approved in advance*
 - The expected outputs of the project are:
 - a report (e.g. ~10-15 pages) with embedded full code and figures, and including an introduction and discussion of the results (40% of the grade)
 - a short presentation (10%)

Learning objectives



Tentative schedule of the course

Each week:

- **~15min of debriefing** on the exercises from the previous week
- **~30min of “lecture”** to provide background and theory
- **~60min of hands-on** demonstration (live coding)

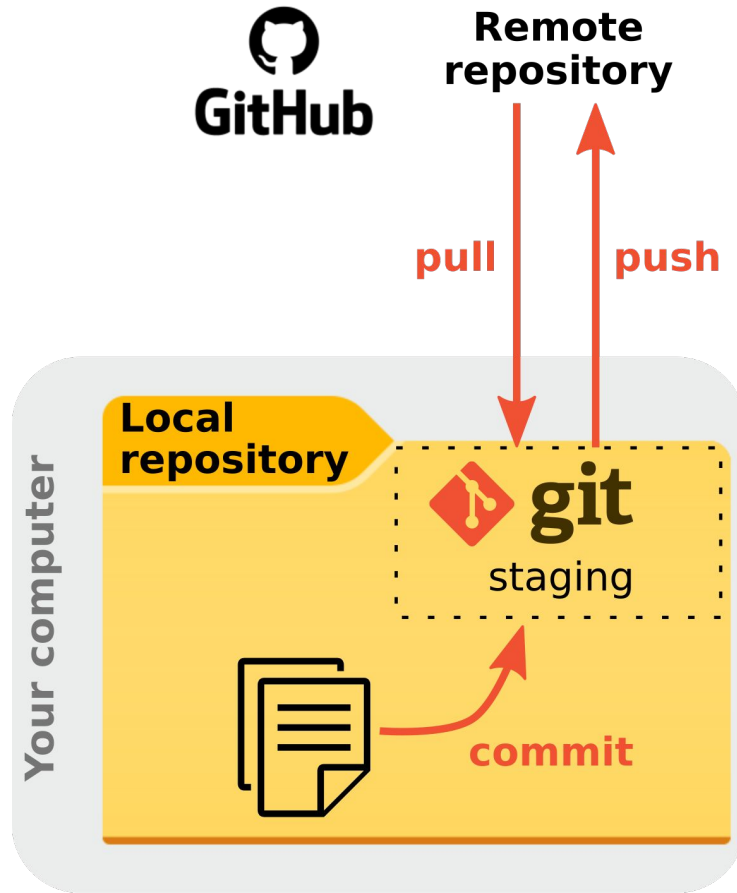
W	Date	Lecture (~30min)	Mixed/hands-on (~60min)
1	24/02	Introduction to the course	Introduction to the practical tools for the course
2	03/03	Genome builds, transcriptome assemblies and annotations	AnnotationHub, Genomic Ranges, Features and their manipulating in R
3	10/03	Overview of NGS chromatin assays and their analysis	Primary ChIPseq analysis pipeline
4	17/03	Specificities and QCs on primary analysis	Finding data from the literature
5	24/03	Functional elements & the histone code	Visualization and exploration of histone modifications
6	31/03	Types of transcriptional regulators, modes and dynamics of their binding	Motif analysis
7	07/04	DNA accessibility, nucleosome positioning, and TF footprints	Analysis and exploration of ATAC-seq data
8	14/04	Differential analysis	Differential analysis
9	21/04	Normalization and enrichment analysis	Normalization and enrichment analysis
10	28/04	From repression to expression and back	Clustering genomic signals and characterizing the clusters
11	05/05	DNA methylation and CpG islands	DNAme visualization
12	12/05	Chromatin conformation, domains and looping	Working with distal regulatory elements
13	19/05	Single-cell chromatin assays	Student presentations
14	26/05	Chromatin and disease; open questions	Student presentations

The tools we'll be using... and some documentation

- Unix shell environment : [short primer](#) / [long intro](#)
- R & Bioconductor : [primer](#)
 - (installation instructions for [R](#), [rstudio](#) and [bioconductor](#))
- R markdowns : [primer](#)
- git & github : [tutorials](#)

Particularly
important for the
course

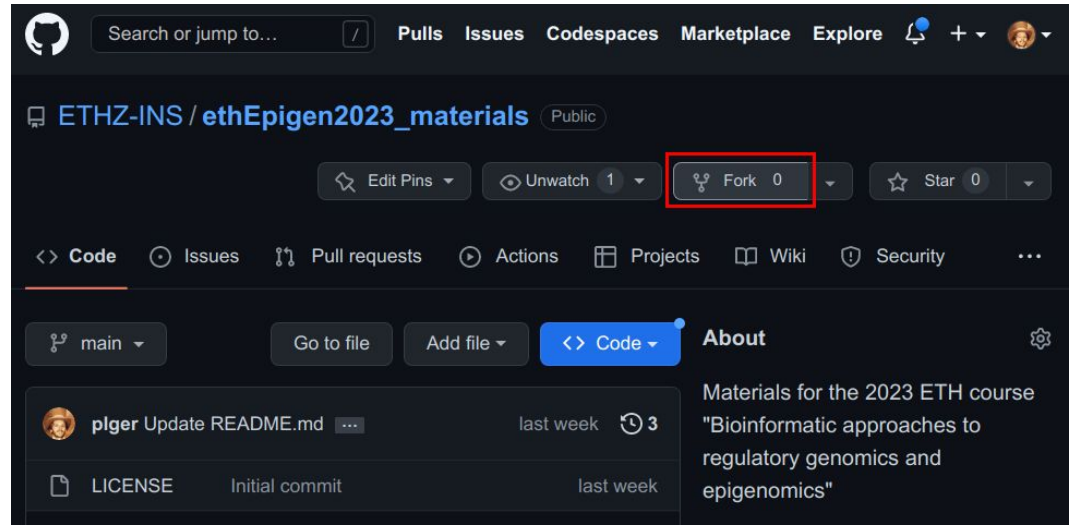
Introduction to git and github



Setting up your github repository for the course:

- If you don't already have one, create a github account
- Send us your github username on slack
- Fork the course's repository, and send me the link to your forked repo

To fork, go to
https://github.com/ETHZ-INS/ethEpiGen2023_materials, make sure you are logged into github, and click the "Fork" button



This week's assignment

1. If you haven't done it yet, install R, Rstudio, and Bioconductor
2. Install the packages we'll need (see the `install.R` file on the repo)
3. Create a R markdown named "`assignment.Rmd`" in which you:
 - a. load the epiwraps library (using `library(epiwraps)`)
 - b. print the session info (using `sessionInfo()`)
4. Render your markdown, and push **both the Rmd and html files** to your github repo, in the directory `week01/`