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Traits influence responses to land-use and climate change in terrestrial vertebrates

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September 23, 2022

Submitted in part fulfilment of the requirements
for the degree of Doctor of Philosophy in Ecology at University College London

Word count: ~40,000

Declaration

I, Adrienne Etard, confirm that the work presented in this thesis is my own. Some parts have been conducted in collaboration with other researchers, and the contributions of co-authors to specific Chapters are described in the *Thesis outline of contents, authorship and collaborations* on page 6. All else is appropriately referenced, and where information has been derived from other sources, I confirm that this has been indicated in the thesis.

Acknowledgements

Firstly, I would like to thank my supervisors, Tim Newbold and Alex Pigot, whose guidance and continual support have been pivotal to the completion of this PhD thesis. A huge thanks to my primary supervisor, Tim Newbold, for making this PhD a fantastic experience. I am grateful for all the opportunities he has offered me, for his availability, his optimism, and for the kind, attentive, and constructive supervision. I am equally grateful to Alex Pigot for welcoming me into his lab, for his insights and feedback on my work, and for his advice. Thanks for the weekly lab meetings and chats that were once an essential part of the lockdown routine and made for stimulating conversations. I would also like to extend my thanks to Richard Pearson for sitting on my Upgrade committee.

Thanks to the wonderful people of CBER; both to those with whom I have directly collaborated and to all those who have contributed to make the workplace a friendly, constructive and open-minded environment. In particular, a big thanks to all present and past members of the Global Biodiversity Change group and of the Pigot's group.

To my friends and family, in London, the mountains, and elsewhere, for their support: thank you! A special mention goes to all the citizens of La Montaz, Mousse included. Another special mention goes to Tom K, for having been the loveliest housemate and friend. A huge thank you also to Yumi and Tom M.

And last but not least, a very special thanks to Tea Trains, Coffee Breaks, and Journal Club Cookies.

Abstract

Human activities have profoundly impacted global biodiversity. Currently, anthropogenic land-use and climate change figure among the major threats to the world's fauna. However, not all species respond similarly to these pressures. Interspecific variability in responses to human threats is notably underpinned by the fact that different species possess different ecological characteristics, some of them allowing species to cope with environmental changes, while others confer a disadvantage to species in modified environments. Understanding what renders species sensitive to anthropogenic pressures is vital to inform and prioritise conservation efforts. Yet, in terrestrial vertebrates, a group for which ecological data are the most abundant, it remains unclear which traits are associated with higher sensitivity to human pressures. The aims of my thesis are to investigate whether and which traits are associated with land-use responses and climate-change sensitivity in terrestrial vertebrates, and to highlight some of the consequences for ecosystem functioning. I first assess the global availability of ecological trait data for terrestrial vertebrates, identifying understudied groups and regions (e.g., Central-African reptiles). I then show that, at global scales, disturbed land uses negatively impact the functional diversity of vertebrate assemblages. Further, I find that in all classes, higher sensitivity to land-use and climate change is associated with narrower ranges, smaller habitat breadth and inability to use human-modified habitats. Both land-use responses and climate-change sensitivity are unevenly distributed among dietary groups, highlighting potential food-web disruptions in assemblages under pressure. Finally, I show that land-use responses are influenced by species' energetic requirements, so that energetic fluxes within vertebrate assemblages are likely modified under human-driven land-use change. Although the large-scale consequences of biodiversity changes for ecosystem functioning remain to be fully understood, my thesis highlights a compositional reshaping of vertebrate assemblages under human pressure and furthers our understanding of anthropogenic impacts on biodiversity.

Impact statement

As anthropogenic pressures on the world's biota keep increasing, it is vital to put into place conservation measures to prevent and reverse further species loss. Beyond ethical and moral considerations, there is an urgent need to protect biodiversity because it sustains a range of ecological processes essential to human well-being and planetary health. Effectively managing biodiversity and related ecosystem processes in a changing world requires an understanding of how different species respond to anthropogenic disturbances. My thesis integrates various data sources to investigate the influence of traits on species' land-use responses and on species' climate-change sensitivity – two of the most pressing threats to biodiversity – at global scales and comparatively across the four terrestrial vertebrate classes. By asking whether interspecific trait variation is associated with species' land-use responses and with climate-change sensitivity, my work consolidates our understanding of what renders species sensitive to environmental change, which can help prioritise conservation efforts.

Chapter 2 presents a trait data collection for terrestrial vertebrates, targeting seven commonly-used traits. I highlight the global taxonomic, geographical, and phylogenetic biases in the trait data, revealing knowledge gaps which could guide future data collection efforts. Chapter 2 was published in *Global Ecology and Biogeography* (Etard et al., 2020). The compiled data were made available and have since been used by other ecologists (e.g., Capdevila et al. (2022a)) and downloaded 272 times as of May 2022. Chapter 3 uses the collected trait data and reveals profound effects of land-use change on vertebrate functional diversity, which contributes to documenting global human impacts on vertebrates and also underlines the possible threats posed by land-use change to ecosystem processes sustained by vertebrates. Chapter 3 was published in *Ecology Letters* (Etard et al., 2022). In Chapter 4, I ask whether traits are associated with species' land-use responses and with species' climate-change sensitivity, comparatively across the four vertebrate classes. Chapter 4 thus puts into perspective the usefulness of trait data for understanding how species respond to these anthropogenic changes, which is valuable for conservation planning and prioritisation. In Chapter 5, I ask whether species' energetic requirements, estimated from metabolic rates, influence species persistence

in disturbed land uses. Chapter 5 thus integrates physiological data to further our fundamental understanding of how vertebrate species respond to land-use change and of the potential consequences for ecosystem functioning.

Beyond publishing two of my PhD Chapters, I have been able to disseminate my work at various international conferences (BES annual meetings in 2019, 2020 and 2021; BES Macroecology conference in 2019; IBS early-career conference in 2021). I will also present my PhD work at the IBS conference (June 2022, 10th Biennial meeting), and at the BES Macroecology conference (July 2022). I have contributed to the Living Planet Report 2020 (WWF, 2020) and to other published papers (Newbold et al., 2019, 2020b). Overall, my PhD work consolidates our knowledge of the role of vertebrate traits for understanding species responses to human pressures and highlights the value of trait data, and more widely, of ecological knowledge, for preserving vertebrate species in a changing world.

Thesis outline of contents, authorship and collaborations

Chapter 1: General introduction

Chapter 1 presents the background for this thesis, exposes the fundamental concepts, and highlights the research questions I investigated in the different Chapters.

Chapter 2: Global gaps and biases in trait data for terrestrial vertebrates

In Chapter 2, I present an analysis of the global gaps and biases in terrestrial vertebrate trait data. To this end, I collate data on seven traits commonly measured in terrestrial vertebrates. I then evaluate the availability of these trait data across the vertebrate classes, assessing whether there are taxonomic, phylogenetic and spatial biases. This Chapter was published in *Global Ecology and Biogeography* in 2020 (DOI: 10.1111/geb.13184; Etard et al. (2020)). The paper was co-authored by Sophie Morrill who collated some of the data on reptile traits as part of an MRes project at UCL, and by Tim Newbold, who participated in the development of the research questions, provided detailed feedback on the analyses, and contributed to the writing of the paper.

Chapter 3: Intensive human land uses negatively affect vertebrate functional diversity

In this Chapter, I investigate how land-use change affects the functional composition and functional diversity of local vertebrate assemblages. This Chapter was published in *Ecology Letters* in 2022 (DOI: 10.1111/ele.13926; Etard et al. (2022)) and co-authored by Alex Pigot and Tim Newbold, who helped construct the hypotheses, provided detailed feedback on the work, and took part in the writing of the paper.

Chapter 4: Narrow-ranging natural habitat specialists most sensitive to both land-use and climate change in terrestrial vertebrates

In this Chapter, I assess whether ecological traits as well as geographical range area are associated with species' land-use responses and species' estimated climate-change sensitivity, comparatively among terrestrial vertebrate classes. Rhiannon Osborne-Tonner contributed to this Chapter by collecting data on amphibian and reptile diet during her MSci project at UCL, which I used to complement my datasets. This Chapter was conducted in collaboration with Tim Newbold who helped develop the research questions and provided detailed feedback on the work and on the writing.

Chapter 5: Energetic constraints and trophic group explain species persistence in disturbed land uses

In Chapter 5, I evaluate the impacts of land-use change on community-level energetic requirements, and I assess whether species' energetic requirements influence species persistence in disturbed land uses. To this end, I use physiological data, compiling species resting metabolic rates (used as a proxy for energetic requirements) from the literature. Meghan Hayden and Laura Dee of the University of Colorado, Boulder, as well as Tim Newbold, contributed to the elaboration of the research questions for this Chapter. Meghan Hayden further contributed to this Chapter by retrieving information on net primary productivity for PREDICTS sites, using data from MODIS satellite imagery. All collaborators also provided feedback on the work and participated in writing the manuscript. This Chapter was submitted to a scientific journal and underwent a round of peer-review. I am preparing this Chapter for resubmission.

Chapter 6: General discussion

This final Chapter summarises the main findings of my thesis and assesses their contributions to the field.

Data and code access

Code access

The main pieces of R code for Chapters 2, 3, 4 and 5 are available at: https://github.com/AdrienneEtard/PhD_thesis_code.

This document was compiled with L^AT_EX; the source code and files are available at: <https://github.com/AdrienneEtard/Thesis>

Data

The data used in this thesis are freely accessible and sources are referenced throughout. In particular:

- the PREDICTS database (Hudson et al., 2014, 2017) can be downloaded from: <https://data.nhm.ac.uk/dataset/the-2016-release-of-the-predicts-database>;
- the trait datasets I compiled in Chapter 2 are available at: https://figshare.com/articles/dataset/Global_gaps_in_terrestrial_vertebrate_trait_data/10075421.

Contents

List of Tables	14
List of Figures	16
1 General introduction	22
1.1 Major drivers of global biodiversity change	23
1.1.1 Land-use change	23
1.1.2 Climate change	24
1.1.3 The future of biodiversity in the Anthropocene	26
1.2 Ecological importance of terrestrial vertebrates and current threats	26
1.3 Using trait-based approaches to understand global biodiversity change	27
1.3.1 Traits as common currencies across species	27
1.3.2 Using ecological traits to assess which species are at most risk from human-driven changes	28
1.3.3 Using physiological traits to understand land-use impacts on biodiversity and ecosystem functioning	30
1.3.4 Thesis aims	31
1.4 Detailed aims, hypotheses and outline of the following Chapters	32
2 Global gaps and biases in trait data for terrestrial vertebrates	35
2.1 Introduction	36
2.2 Methods	39
2.2.1 Trait data collection	39
2.2.2 Investigating gaps and biases in trait data	45
2.3 Results	47
2.3.1 Taxonomic biases in trait information	47

2.3.2	Phylogenetic biases in trait completeness	48
2.3.3	Spatial biases in trait completeness	51
2.4	Discussion	53
3	Intensive human land uses negatively affect vertebrate functional diversity	58
3.1	Introduction	58
3.2	Methods	60
3.2.1	Vertebrate assemblages	60
3.2.2	Functional traits and diversity metrics	62
3.2.3	Effects of land use and land-use intensity on FRic and FDis (Hypothesis 1)	63
3.2.4	Investigating functional under-dispersion (Hypothesis 2)	65
3.2.5	Functional loss and functional gain (Hypothesis 3)	65
3.3	Results	66
3.3.1	Effects of land use on FRic and FDis	66
3.3.2	Changes in the probability of occurrence of functional under-dispersion	69
3.3.3	Functional loss and gain	70
3.4	Discussion	72
4	Narrow-ranging natural habitat specialists most sensitive to both land-use and climate change in terrestrial vertebrates	76
4.1	Introduction	77
4.2	Methods	81
4.2.1	Ecological characteristics (Figure 4.1a)	81
4.2.2	Imputations of missing trait values	82
4.2.3	Characterizing the influence of traits on species' land-use responses (Figure 4.1b) . .	83
4.2.4	Characterizing the influence of traits on species' sensitivity to climate change (Figure 4.1c)	88
4.3	Results	90
4.3.1	Land-use responses	90
4.3.2	Climate-change sensitivity	96
4.3.3	Validations on complete trait data	101
4.4	Discussion	101

5 Energetic constraints and trophic group explain species persistence in disturbed land uses	105
5.1 Introduction	106
5.2 Methods	111
5.2.1 Vertebrate assemblage composition	111
5.2.2 Energy availability by land-use type and land-use intensity	111
5.2.3 Resting Metabolic Rates (RMR)	112
5.2.4 Trophic group and body mass information	112
5.2.5 Effects of land use, land-use intensity and trophic group on assemblage-level total RMR (prediction 1; Figure 5.2a)	113
5.2.6 Effects of land use, land-use intensity, trophic group and residual RMR on species occurrence probability (prediction 2; Figure 5.2b)	113
5.3 Results	114
5.3.1 Effects of land use, land-use intensity and trophic group on assemblage-level total RMR	114
5.3.2 Effects of land use, land-use intensity, trophic group and residual RMR on species' occurrence probability	116
5.4 Discussion	119
6 General discussion	123
6.1 Gaps and biases in the knowledge of terrestrial vertebrates	125
6.2 Functional reshaping of local vertebrate assemblages under land-use change	126
6.2.1 Functional diversity and functional composition (Chapter 3)	127
6.2.2 Energetic requirements (Chapter 5)	128
6.3 Uneven sensitivity of vertebrate species to land-use and climate change	128
6.3.1 Ecological characteristics (Chapter 4)	129
6.3.2 Mass-independent energetic requirements (Chapter 5)	130
6.4 Relevance of the findings to conservation	131
6.5 Conclusion	131
Bibliography	133
Appendix 1: Supporting information for Chapter 2	161
S2.6 Taxonomic corrections	161

S2.7	Additional information for trait compilation	164
S2.8	Cutting distribution maps by altitudinal limits	165
S2.9	Impact of taxonomic corrections on trait coverage	166
S2.10	Assemblage-level median, mean and standard deviation of trait completeness (maps)	167
S2.11	Phylogenetic patterns in trait completeness	172
S2.12	Poisson model summaries	177
S2.13	Spatial model summaries	178
S2.14	Trait coverage and taxonomic matching	179
Appendix 2: Supporting information for Chapter 3		180
S3.1	Land-use categories in PREDICTS and sample sizes	180
S3.2	Trait data & imputation of missing trait values	183
S3.2.1	Choice of imputation technique	185
S3.2.2	Phylogenetic signal in traits	185
S3.2.3	Implementation of ‘missforest’ imputations	186
S3.3	Degree of multicollinearity among traits	187
S3.4	Imputation performance	188
S3.5	Functional loss and functional gain	191
S3.6	Diagnostic plots	194
S3.7	Model robustness	203
S3.8	Model robustness – time since land-use conversion	210
Appendix 3: Supporting information for Chapter 4		213
S4.1	Compiling diet information	213
S4.2	Imputing missing trait values	217
S4.2.1	Trait data coverage	217
S4.2.2	Phylogenetic signal in traits	217
S4.2.3	Implementation of missing value imputations	218
S4.2.4	Imputation error	218
S4.3	Land-use types in PREDICTS and sample sizes (number of sampled sites across classes) . .	220
S4.4	Land-use responses: multicollinearity checks among the models’ explanatory variables . .	222
S4.5	Implementing Climate-niche Factor Analysis across terrestrial vertebrates	225
S4.5.1	Historical climate data: groups of intercorrelated variables	225

S4.5.2 CENFA estimation and resolution	225
S4.6 Climate-change sensitivity models: multicollinearity checks among models' explanatory variables	228
S4.7 Land-use responses: occurrence probability predictions from the partial models for artificial habitat use and diel activity	230
S4.8 Land-use responses: diagnostic plots for the full models	232
S4.9 Climate-change sensitivity: model summaries and diagnostic plots	235
S4.9.1 Summaries & diagnostic plots for models fitted on species with range area >100 km ²)	235
S4.9.2 Robustness checks of the PGLS models	239
S4.10 Validations on complete trait data subsets	246
Appendix 4: Supporting information for Chapter 5	249
Bibliography for the Appendices	257

List of Tables

Chapter 2	35
2.1 Data sources for each trait	41
2.2 Number of species for each analysis	45
Chapter 4	76
4.1 Summary of the effects of the ecological characteristics (except for primary diet) on species' responses to disturbed land uses ('within land-use type' effects) and on species' climate-change sensitivity, for each Class of terrestrial vertebrates.	92
4.2 ANOVA summaries for the PGLS models investigating the associations between the species-level ecological characteristics and species' estimated climate-change sensitivity.	100
Appendix 1	161
S2.1 Coefficients of the model investigating whether species range size explained the number of sampled traits	177
S2.2 Coefficients of the model investigating whether species range size explained the number of sampled traits, using range maps not cut by altitudinal limits	177
S2.3 Spatial model summary for amphibians	178
S2.4 Spatial model summary for reptiles	178
Appendix 2	180
S3.1 Land-use categories in the PREDICTS database	180
S3.2 Phylogenetic signal in continuous and categorical traits	186
S3.3 Variance inflation factors across considered (imputed) traits	187
S3.4 Sample sizes (number of pairs of sites) for the calculation of functional loss and functional gain across all species	192

S3.5	Sample sizes (number of pairs of sites) for the calculation of functional loss and functional gain within each class	193
S3.6	Summary of the model explaining FRic by land use and time since land-use conversion, fitted on the subset of data for which there are information on time since land-use conversion	210
S3.7	Summary of the model explaining FDis by land use and time since land-use conversion, fitted on the subset of data for which I have information on time since land-use conversion	212
Appendix 3		213
S4.1	Phylogenetic signal in continuous and categorical traits	217
S4.2	Land-use categories in the PREDICTS database	220
S4.3	Land-use responses: Generalised Variance Inflation Factors (amphibians, with diet)	222
S4.4	Land-use responses: Generalised Variance Inflation Factors (amphibians, without diet) . .	222
S4.5	Land-use responses: Generalised Variance Inflation Factors (birds)	223
S4.6	Land-use responses: Generalised Variance Inflation Factors (mammals)	223
S4.7	Land-use responses: Generalised Variance Inflation Factors (reptiles, with diet)	223
S4.8	Land-use responses: Generalised Variance Inflation Factors (reptiles, without diet)	224
S4.9	PGLS models: Generalised Variance Inflation Factors (amphibians, with diet breadth) . .	228
S4.10	PGLS models: Generalised Variance Inflation Factors (amphibians, without diet breadth) .	228
S4.11	PGLS models: Generalised Variance Inflation Factors (birds)	228
S4.12	PGLS models: Generalised Variance Inflation Factors (mammals)	229
S4.13	PGLS models: Generalised Variance Inflation Factors (reptiles)	229
S4.14	Summary for the PGLS model fitted on amphibians	235
S4.15	Summary for the PGLS model fitted on birds	236
S4.16	Summary for the PGLS model fitted on mammals	237
S4.17	Summary for the PGLS model fitted on reptiles	238
S4.18	Summary of the effects of the ecological characteristics (except for diet) on (a) species' responses to disturbed land uses ('within land-use type' effects) and (b) species climate-change sensitivity, for each class of terrestrial vertebrates, from validation models using empirical, non imputed trait values	247
Appendix 4		249
S5.1	Number of species for which RMR data was collected in each class & data sources, initial RMR coverage for PREDICTS species and phylogenetic signal in RMR	249

List of Figures

General introduction	22
1.1 Characteristics of the ‘Great Acceleration’	25
1.2 Typology of the traits used in this thesis	30
 Chapter 2	 35
2.1 Procedure used to identify the accepted names of species	40
2.2 Number of natural and artificial habitats used by species against number of strictly natural habitats used by species	44
2.3 Trait coverage and completeness across species	48
2.4 Within-family median trait completeness in herptiles	50
2.5 Relationship between number of sampled traits and geographical range size	51
2.6 Spatial distribution of assemblage-level median trait completeness in herptiles	52
2.7 Spatial model trends for herptiles	53
 Chapter 3	 58
3.1 Overview of the study design and functional diversity metrics	61
3.2 Effects of land use, land-use intensity and region on FRic (a) and on FDis (b) across all PREDICTS vertebrates.	67
3.3 Effects of land use, land-use intensity, region and taxonomic class on FRic (a) and FDis (b). .	68
3.4 Effects of land use, land-use intensity and region on the probability of occurrence of functional under-dispersion.	69
3.5 Effects of land use and land-use intensity on (a) functional loss and (b) functional gain across pairs of sites in temperate areas and tropical areas.	70

3.6	Effects of land use, land-use intensity, region and taxonomic class on functional loss and functional gain across pairs of sites.	71
Chapter 4		76
4.1	Framework of the study	80
4.2	Assessing the effects of ecological characteristics on species' land-use responses: methodology for (a) categorical characteristics and (b) continuous characteristics.	86
4.3	Predicted occurrence probability as a function of land use, land-use intensity, diet and their interactions, for each class of terrestrial vertebrates.	94
4.4	Proportion of the explained variance attributable to each of the main effects in the land-use and climate-change sensitivity models	95
4.5	Estimated effects of the categorical traits on climate-change sensitivity, from the PGLS models fitted in each class.	97
4.6	Effects of the continuous ecological characteristics on climate-change sensitivity, estimated from the PGLS models in each class.	98
Chapter 5		105
5.1	Map of PREDICTS sites, sample sizes and NPP by land use and land-use intensity	109
5.2	Framework for the predictions and models	110
5.3	Effects of land use, land-use intensity and trophic group on assemblage-level total RMR.	115
5.4	Slope estimates and predictions for the relationship between occurrence probability and residual RMR	117
5.5	Predicted occurrence probabilities in primary vegetation and for the most disturbed land uses where residual RMR was found to have a significant effect on occurrence probability	118
Appendix 1		161
S2.1	Difference in species number before and after taxonomic correction (A) and distribution of number of synonyms across datasets (B)	163
S2.2	(a) Body mass versus body length and (b) longevity versus age at sexual maturity in amphibians	164
S2.3	Generation length versus longevity data in mammals and birds	164
S2.4	Availability of altitudinal limits across species	165

S2.5	Range sizes before versus after cutting by altitudinal limits	165
S2.6	Trait coverage when no taxonomic correction is applied before matching sources by binomial names, versus when I applied the described procedure	166
S2.7	Spatial distribution of assemblage-level median trait completeness in each class	168
S2.8	Spatial distribution of assemblage-level mean trait completeness in each class	169
S2.9	Spatial distribution of assemblage-level standard deviation of trait completeness in each class	170
S2.10	Assemblage-level species richness against standard deviation in trait completeness in each class	171
S2.11	Within-family median trait completeness in mammals	173
S2.12	Within-family median trait completeness in birds	174
S2.13	Within-family standard deviation in trait completeness (herptiles)	175
S2.14	Within-family species richness against the within-family standard deviation of trait completeness	176
S2.15	Relationships between number of sampled traits and geographical range size using distribution maps not cut by altitudinal limits	177
S2.16	Comparison of trait coverage among datasets corrected for taxonomy in different ways . .	179

Appendix 2 180

S3.1	Number of sites in each land use and land-use intensity for which FRic and FDis were calculated, across all species	181
S3.2	Number of sites in each land use, land-use intensity and for which FRic and FDis were calculated, within each class	182
S3.3	Relationship between habitat breadth and geographical range size across species in each class	183
S3.4	Trait coverage for the vertebrate species sampled in the PREDICTS database	184
S3.5	Distribution of trait completeness across the vertebrate species sampled in the PREDICTS database	184
S3.6	Distribution of continuous traits considered in the calculation of the functional diversity metrics	188
S3.7	Frequency distribution for categorical traits considered in the calculation of the functional diversity metrics	189
S3.8	Out-of-bag imputation errors for the continuous traits (a) and categorical traits (b)	190
S3.9	Diagnostic plots for Model 1a	194

S3.10	Diagnostic plots for Model 1b	195
S3.11	Diagnostic plots for Model 2a	196
S3.12	Diagnostic plots for Model 2b	197
S3.13	Diagnostic plots for Model 3	198
S3.14	Diagnostic plots for Model 4a	199
S3.15	Diagnostic plots for Model 4b	200
S3.16	Diagnostic plots for Model 5a	201
S3.17	Diagnostic plots for Model 5b	202
S3.18	Effects of land use, land-use intensity and region on FRic (a) and FDis (b) across vertebrates, for the subset of species with complete trait data	203
S3.19	Effects of land use, land-use intensity and region on FRic (a) and FDis (b), for the subset of species with complete trait data, with geographical range size as an additional trait . . .	204
S3.20	Effects of land use and land-use intensity on FRic (a) and FDis (b), obtained when calculating FRic and FDis with each set of imputed traits	205
S3.21	Effects of land use and land-use intensity on FRic (a) and FDis (b), obtained when resampling primary vegetation sites twenty independent times	206
S3.22	Effects of land use, land-use intensity and taxonomic class on FRic (a) and FDis (c), for the subset of species with complete trait data	207
S3.23	Effects of land use, region, land-use intensity and taxonomic class on FRic and FDis obtained with the imputed trait data (black points) or with the complete data subsets (red points)	208
S3.24	Effects of land use, region, land-use intensity and taxonomic class on FRic and FDis, obtained when calculating FRic and FDis with each set of imputed traits	209
S3.25	Effects of land use on FRic for the model that includes time since land-use conversion (blue points) versus the model that does not take time since land-use conversion into account (red points)	211
A	Appendix 3	213
S4.1	Trait coverage across vertebrate classes, including coverage for diet information	217
S4.2	Out-of-bag estimation of imputation errors for the traits included in the analyses	219
S4.3	Sample sizes (number of PREDICTS sites) for the different land-use types, in each class .	221

S4.4	Groups of intercorrelated climatic variables using a cutoff of 0.65 for Pearson's correlation coefficient	225
S4.5	Possible impact of resolution on estimated geographical range area	226
S4.6	Estimated climate-change sensitivity estimations at three different resolutions	227
S4.7	Predicted occurrence probability as a function of land use, land-use intensity, artificial habitat use and their interactions in each class	230
S4.8	Predicted occurrence probability as a function of land use, land-use intensity, diel activity and their interactions in each class	231
S4.9	Land-use responses: diagnostic plots for the mixed-effects model fitted on amphibians	232
S4.10	Land-use responses: diagnostic plots for the mixed-effects model fitted on birds	232
S4.11	Land-use responses: diagnostic plots for the mixed-effects model fitted on mammals	233
S4.12	Land-use responses: diagnostic plots for the mixed-effects model fitted on reptiles	234
S4.13	Diagnostic plots for the PGLS model fitted on amphibians	235
S4.14	Diagnostic plots for the PGLS model fitted on birds	236
S4.15	Diagnostic plots for the PGLS model fitted on mammals	237
S4.16	Diagnostic plots for the PGLS model fitted on reptiles	238
S4.17	Effects for the PGLS models investigating associations between species-level ecological characteristics and climate-change sensitivity, either fitted on all species (y-axis), or fitted on the species whose range area was $>100 \text{ km}^2$ (x-axis)	239
S4.18	Effects for the PGLS models investigating associations between species-level ecological characteristics and climate-change sensitivity, either fitted on all birds and mammals (y-axis), or on the subset of bird and mammal species that were found not to be migratory (x-axis)	240
S4.19	Estimated effects of the categorical traits on climate-change sensitivity, from the PGLS models fitted on birds and mammals, excluding species identified as migratory	241
S4.20	Effects of the continuous ecological characteristics on climate-change sensitivity, estimated from the PGLS models fitted on birds and mammals, excluding species identified as migratory	242
S4.21	Effects for the PGLS models investigating associations between species-level ecological characteristics and climate-change sensitivity, either using climate-change sensitivity estimated at $5 \times 5 \text{ km}$ resolution (y-axis), or using climate-change sensitivity estimated at different resolutions (x-axis; see Methods in main text)	243

S4.22	Estimated effects of the categorical traits on climate-change sensitivity, using climate-change sensitivity estimated at different resolutions (see Methods in main text)	244
S4.23	Effects of the continuous ecological characteristics on climate-change sensitivity, using climate-change sensitivity estimated at different resolutions (see Methods in main text) . . .	245
S4.24	Predicted occurrence probability as a function of land use, land-use intensity, diet and their interactions in each class: from validation models using empirical, non imputed trait values	248

Appendix 4 **249**

S5.1	Formula for the abundance model, used to understand the role of shifts in the body mass of species on observed changes in tRMR	249
S5.2	Effects of land use, land-use intensity, trophic group, body mass and their interactions on assemblage-level total abundance	250
S5.3	Diagnostic plots for the linear mixed-effects model looking at the effects of land use, land-use intensity, trophic group and their interactions on assemblage-level total RMR	251
S5.4	Estimates for the models looking at the effects of land use, land-use intensity, trophic group and their interactions on assemblage-level total RMR	252
S5.5	Effects of land use, land-use intensity, trophic level and their interactions on assemblage-level total RMR: empirical versus imputed	253
S5.6	Diagnostic plots for the generalised mixed-effects model looking at the effects of land use, land-use intensity, trophic group, residual RMR and their interactions on species' probability of occurrence	254
S5.7	Model's coefficients from the occurrence model fitted using the 'lme4' package against coefficients from the model fitted using the 'MCMCglmm' package	255
S5.8	Slope estimates and predictions for the relationship between occurrence probability and residual RMR for the model fitted on empirical RMR values only	256

¹ 1 | General introduction

² Humans have been modifying Earth's ecosystems for thousands of years. Archaeological and palaeontological evidence suggest that human activities may have played a major role in the extinction of Australia's megafauna as early as fifty thousand years ago (Johnson et al., 2016; Miller et al., 2016; van der Kaars et al., 2017). The subsequent arrival of modern humans in other parts of the world has also been associated with extinctions, of the megafauna in particular (Broughton and Weitzel, 2018; Sandom et al., 2014). However, the global signature of human presence on Earth has never been as prominent as in recent decades. The past two hundred years have been characterised by a sharp increase in the rates of human-driven changes at the planetary scale, a phenomenon that has been termed 'the Great Acceleration' (Steffen et al., 2015).

¹⁰ To emphasize the recent impacts of human activities on the Earth's systems, Crutzen and Stoermer (2000) proposed that we have entered a new geological epoch, which they called 'the Anthropocene'. Although the formal acceptance of this epoch and the timing of its start are still debated within the stratigraphic community (Lewis and Maslin, 2015; Monastersky, 2015), the coined term reflects the profound effects of humans on planetary processes and on the biosphere, such that its use has largely surpassed the geological field (Malhi, 2017).

¹⁶ The Anthropocene can be characterised by Earth-system and socio-economic indicators (Biermann et al., 2016; Steffen et al., 2011). Two of the major signatures are the human-driven transformations of the land surface, and changes to atmospheric composition, which have led to the onset of anthropogenic climate change (Lewis and Maslin, 2015). Altogether, the development of human activities at unprecedented scales and magnitude has led to the alteration of many ecosystems. As a result of combined anthropogenic pressures, the world's biodiversity has been changing (Daru et al., 2021; Dirzo et al., 2014; Johnson et al., 2017; McGill et al., 2015). Decreases in a range of biodiversity indicators have been reported for many taxonomic groups (Butchart et al., 2010). Human-mediated invasions and translocations of species, coupled with local declines in native species, have promoted biotic homogenisation (Daru et al., 2021; Finderup Nielsen et al., 2019; Newbold et al., 2018). In addition, species have gone extinct at rates higher than expected from natural

26 background variability, with current extinction rates estimated to exceed those inferred from fossil records by
27 a hundred to a thousand times (Barnosky et al., 2011; De Vos et al., 2015). Biodiversity loss and ecosystem
28 change have become such major issues in the 21st century that the prevention of biodiversity erosion and
29 the protection of ecosystems have become priority goals on international agendas (Convention on Biological
30 Diversity, 2022; Hoban et al., 2020). Indeed, it is now well established that biodiversity is tightly linked
31 with ecosystem functioning and ecosystem services delivery (Hooper et al., 2005; Millennium Ecosystem
32 Assessment, 2005; Oliver et al., 2015), and thus ultimately with human well-being (Millennium Ecosystem
33 Assessment, 2005). However, the difficulty in achieving global conservation goals – such as the failure to
34 reach the Aichi targets (Buchanan et al., 2020) – highlights the need to strengthen global conservation efforts
35 if we are to protect biodiversity and related ecosystem services from global threats (Butchart et al., 2016).

36 **1.1 Major drivers of global biodiversity change**

37 Currently, land-use change is the primary driver of global biodiversity loss, and is responsible for causing
38 global declines in species richness and abundance through habitat modification (Chaudhary et al., 2018;
39 Maxwell et al., 2016; Newbold et al., 2015; Nowakowski et al., 2018; Powers and Jetz, 2019). Although
40 climate change is not currently the main driver of biodiversity change, the negative effects of climate change
41 on biodiversity could equate those of land-use change in their magnitude by 2070 (Newbold, 2018). Other
42 major drivers of biodiversity loss include overexploitation, pollution and the spread of non-native species. In
43 this thesis, my focus is on land-use and climate change as global drivers of biodiversity change.

44 **1.1.1 Land-use change**

45 Land cover describes the physical aspect and composition of the land surface from dominant biotic and
46 abiotic features, typically classifying the Earth's surface into determined sets of natural and artificial ensem-
47 bles (e.g., grasslands, forests, man-made structures, waterbodies, etc; Grekousis et al. (2015)). Land cover
48 can notably be characterised with satellite imagery (Wulder et al., 2018). Land use, however, describes the
49 human intent behind a particular land cover (Lambin et al., 2001). Land-use change thus refers the pro-
50 cess by which humans transform the landscape to achieve socio-economic needs. Land-use change includes
51 transitions from natural to anthropogenic landscapes, as exemplified by agricultural-driven deforestation in
52 tropical areas (Jayathilake et al., 2021). It also describes transitions between different forms of human-
53 dominated land uses, such as the expansion of urban areas over agricultural lands (Ustaoglu and Williams,
54 2017). Land-use change can also include transitions from anthropogenic land uses to natural habitats, for

example with the restoration of human-degraded landscapes (Banks-Leite et al., 2020). Land-use and land-cover classifications may overlap, and these two terms have often been used interchangeably (Comber et al., 2008). Although humans have been modifying terrestrial ecosystems for millennia – between 75% and 95% of the total land surface could have been altered by human activities at some point in history (Ellis et al., 2013, 2021) – it is only during the past three centuries that the terrestrial surface made the transition from mostly wild to mostly human-dominated (Ellis et al., 2010). The most important driver behind this transition has been agricultural expansion, with major increases in cropland and grazing areas from the mid-18th century onward (Figure 1.1a). In recent decades, the expansion of grazing areas and animal feed crops, fuelled by the rising demand for animal products, has been identified as the most important driver of land-use change (Alexander et al., 2015).

The effects of land-use change on biodiversity have been characterised at local, regional and global scales – although global-scale studies represent a small proportion of the published research (Davison et al., 2021). Overall, land-use change has a negative impact on species richness and abundance (Foley et al., 2005; Newbold et al., 2015). For example, urban areas, which have been expanding at faster rates than urban populations themselves (Seto et al., 2010), can have a considerable negative impact on biodiversity and ecosystem services, despite currently representing a small proportion of the terrestrial surface (about 1%; Goldewijk et al. (2017)). In particular, the expansion of impervious surfaces, which characterizes urban development, has been linked to a reduction in species richness (Souza et al., 2019; Yan et al., 2019) and to increases in environmental risks (e.g., due to flooding, Hou et al. (2022); or to heat-island effects). Another important aspect of land-use change for biodiversity outcomes and ecosystem service delivery is the level of intensity at which the land is used to fulfil its purpose. For instance, management practices in agricultural areas are a major determinant of local biodiversity and related ecosystem services such as pollination and pest control (Foley et al., 2005; Kehoe et al., 2015; Millard et al., 2021). In urban areas, introducing and managing green spaces can lead to positive biodiversity outcomes (Aronson et al., 2017; Ives et al., 2016), and can also help mitigate flooding risks and heat islands (Livesley et al., 2016). Yet, land-use intensity has not been explicitly considered by a majority of past studies investigating impacts of land-use change on biodiversity (Davison et al., 2021; Dullinger et al., 2021).

1.1.2 Climate change

According to the World Meteorological Organization, climate change is defined as long-term changes (i.e., over at least several decades) to the mean state or to the variability of the climate, attributable to human activity or to natural causes. There is a strong scientific consensus that current climate change (from ap-

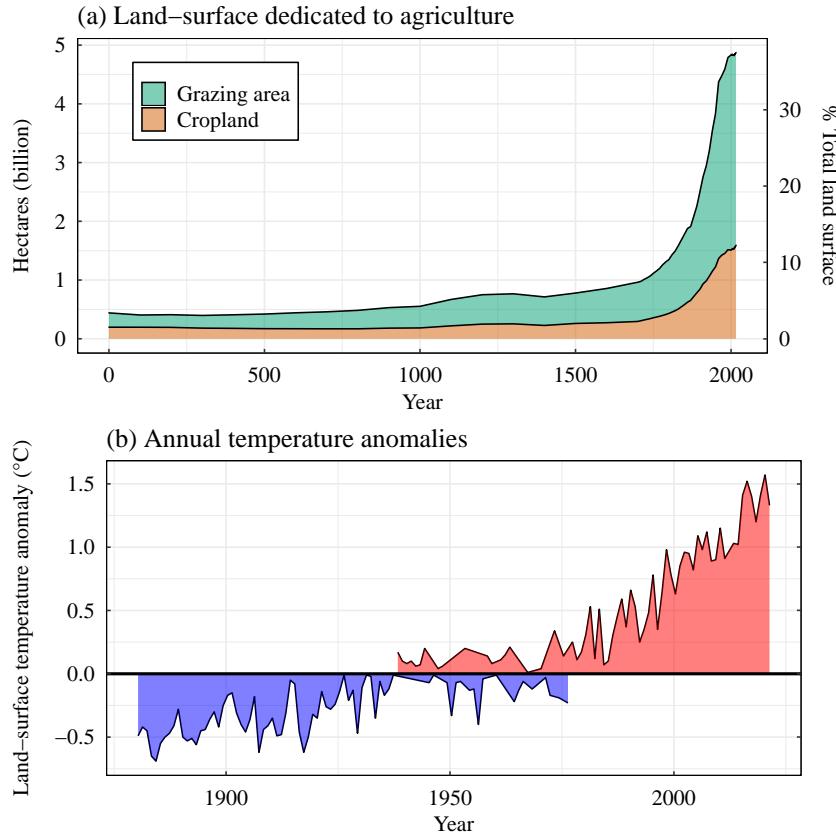


Figure 1.1: Characteristics of the ‘Great Acceleration’. (a) Land surface (and % land-surface) used for agricultural purposes between year 0 and 2016. Data from the HYDE database (Goldewijk et al., 2017), downloaded from <https://ourworldindata.org/land-use> (24/01/2022). (b) Annual land-surface temperature anomaly between 1880 and 2021. Data retrieved from the National Oceanic and Atmospheric Administration – National Centers for Environmental Information, Climate at a Glance: Global Time Series, published April 2022, retrieved 06/05/2022 from <https://www.ncdc.noaa.gov/cag/>. The anomalies are calculated with reference to the global temperature average for the 20th century.

proximately 1850) is the result of human-driven changes to atmospheric composition (Crowley, 2000; Intergovernmental Panel on Climate Change, 2013; Maibach et al., 2014). Current manifestations of climate change include rising average temperatures (Valipour et al. (2021); Figure 1.1b), increases in the frequency of extreme weather events (Seneviratne et al., 2012), and changes in global rainfall patterns (Dore, 2005; Trenberth, 2011).

There is accumulating empirical evidence that climate change affects biodiversity globally, with documented changes in phenology (Inouye, 2022), in the geographical distributions of species (Chen et al., 2011; Lenoir and Svenning, 2015; Soroye et al., 2020), and in species physiology (Chown et al., 2010; Pörtner and Farrell, 2008). Climate-change impacts on individual species have consequences for whole communities, through disruptions of species interactions, which can in turn exacerbate impacts on individual species (Cahill et al., 2013; Kharouba et al., 2018).

97 **1.1.3 The future of biodiversity in the Anthropocene**

98 Projecting future land-use and climate-change impacts on biodiversity highlights the key role of human-
99 development scenarios for global biodiversity outcomes (Newbold, 2018; Powell and Lenton, 2013), for the
100 long-term viability of animal populations (Spooner et al., 2018), and for ecosystem processes and services
101 (Lawler et al., 2014). As the world's population continues to grow and as the demand for food, energy and
102 other commodities keeps rising, rates of global land-use and climate change are unlikely to slow without
103 the implementation of strong international regulations and consumption changes (Intergovernmental Panel
104 on Climate Change, 2022; Stehfest et al., 2019). Under current development scenarios ('business as usual'),
105 future projections show that biodiversity will likely be negatively impacted overall, with decreases in species
106 abundance, increases in extinction rates, and shifts in the distribution of species (Newbold, 2018; Pereira
107 et al., 2010; Schipper et al., 2020). In this context, evaluating the effects of land-use and climate change
108 on biodiversity and associated ecosystem services has become vital in order to put into place mitigation
109 measures. Understanding how species have responded to past and current pressures can help assess how they
110 are likely to respond to future pressures. In particular, understanding what makes species more sensitive to
111 land-use and climate change can help conservation efforts and mitigate global human impacts on biodiversity.

112 **1.2 Ecological importance of terrestrial vertebrates and current threats**

113 In this thesis, I focus on terrestrial vertebrates, a group of more than 30,000 species that has been particu-
114 larly well sampled and studied (Titley et al., 2017), and for which there is available ecological information
115 for many species (such as geographical distributions, traits, occurrence, etc.), allowing for large-scale bio-
116 diversity assessments (e.g., Jenkins et al. (2013)). Terrestrial vertebrates play significant roles in ecosystem
117 functioning and support a wide range of processes, most notably as pollinators (Ratto et al., 2018), seed
118 dispersers (Tiffney, 2004), regulators of lower trophic levels (Lin et al., 2018; Luck et al., 2012; Mooney
119 et al., 2010; Salo et al., 2010; Zhang et al., 2018a), nutrient cyclers (Cunningham et al., 2018; Inger et al.,
120 2016; Wilson and Wolkovich, 2011) and ecosystem engineers (Severtsov, 2012). Vertebrates are also impor-
121 tant for human societies, both culturally and as sources of proteins (Albert et al., 2018; Alves et al., 2018;
122 H irons et al., 2016), and feature among the most charismatic species in the public's eye (Albert et al., 2018;
123 Courchamp et al., 2018).

124 Despite their cultural and ecological importance, terrestrial vertebrates are highly threatened by hu-
125 man activities. The latest Living Planet Report revealed that vertebrate populations have decreased by
126 70% on average since 1970 (WWF, 2020). According to the IUCN Red List of Threatened Species,

about 41% of assessed amphibian species, 26% of mammals, 21% of reptiles and 13% of birds are classified as threatened with extinction (IUCN 2022, <https://www.iucnredlist.org/resources/summary-statistics>). A recent assessment of vertebrates listed in the IUCN Red List of Threatened Species highlights habitat destruction as the predominant human threat (Cox et al., 2022), but direct exploitation also features among the major factors of decline (Monastersky, 2014). Although climate change is not the principal driver of current population declines (Caro et al., 2022), the first extinction of a mammal (the Bramble Cay melomys, *Melomys rubicola*) attributed to anthropogenic climate change was reported in 2016 (Watson, 2016). Future projections highlight that between 10% and 30% of vertebrate species could be locally lost by 2070 depending on climate-change scenarios (Newbold, 2018), and that up to one in six species could face extinction under current climate change (Urban, 2015). Further, despite having been well sampled and studied compared to other groups, there still remain important gaps and biases in our ecological knowledge of terrestrial vertebrates and of their responses to human threats (Hevia et al., 2017; Meiri and Chapple, 2016; Meyer et al., 2015; Oliver et al., 2021).

1.3 Using trait-based approaches to understand global biodiversity change

1.3.1 Traits as common currencies across species

Despite the global average declines reported for vertebrate populations, not all species respond similarly to environmental changes (Dornelas et al., 2019; Leung et al., 2020): while some species are impacted negatively, others benefit from global environmental changes (Newbold et al., 2018; Thomas, 2013). One of the reasons why species differ in their ability to cope with disturbances is that species present different characteristics, or traits. In my thesis, I use the term ‘traits’ to refer to *intrinsic* characteristics, i.e. measurable at the level of an individual. Thus, although the formal definition of a trait can vary depending on studies, I adopt the definition of McGill et al. (2006), considering traits to be characteristics that are measurable at an organismal level, comparable across different species, and that likely influence organismal fitness and performance. I further make a distinction between ‘traits’ and ‘ecological characteristics’, notably in Chapter 4, where I refer to ‘ecological characteristics’ as encompassing traits and geographical range area (as geographical range area does not meet the strict definition of a trait).

The idea that species traits mediate species’ responses to environmental change was formalised in the ‘response-effect’ framework, developed in the field of plant ecology (Lavorel and Garnier, 2002), where traits that influence species’ responses to environmental change were termed ‘response traits’ (and those that underpin ecological processes were termed ‘effect traits’). One of the appeals of trait-based approaches is

157 that individual species are no longer the fundamental unit of biodiversity investigations. Rather, traits become
158 the focus and act as ‘common currencies’ across species, which is of particular interest for conservation when
159 long-term population data are lacking. If species’ responses to human threats consistently relate to certain
160 traits, it may be possible to generalise patterns, and estimate the responses of species for which population
161 data are not available (Verberk et al., 2013).

162 **1.3.2 Using ecological traits to assess which species are at most risk from human-driven
163 changes**

164 Vertebrate ecological traits (which I distinguish from physiological traits, and which I define here as traits
165 relating to the life-history, diet, morphology, and habitat use of species; Figure 1.2) have been used to explain
166 species’ responses to global changes. Past studies have notably investigated whether species extinction risk
167 is associated with species traits (Chichorro et al., 2019; Lebreton, 2011; Ripple et al., 2017), which is of high
168 interest for conservation, but often lacks a consideration of specific threats (González-Suárez et al., 2013).

169 Other studies have focused on the influence of traits on species’ responses to particular human pressures.
170 For example, a range of correlative trait-based approaches have been used to understand whether traits are as-
171 sociated with species’ responses to climate change. On the one hand, past work has made use of spatial data
172 to investigate species’ responses to climate change. For instance, some studies have focused on explaining
173 interspecific variation in past or projected range shifts with traits (Di Marco et al., 2021; McCain and King,
174 2014; Pacifici et al., 2017; Schloss et al., 2012). However, studying observed responses to climate change
175 may be complicated by the fact that this requires disentangling the effects of climate change from effects of
176 other drivers of change on observed responses (MacLean and Beissinger, 2017). Thus, other studies have
177 used proxies instead of observed responses, which may provide complementary insights into the relation-
178 ships between traits and species’ ability to track climate change. For instance, Estrada et al. (2018) used a
179 ‘range-filling’ approach as a proxy for species’ ability to shift their distributions under climate change. The
180 ‘range-filling’ approach consists of investigating interspecific differences between the realised and poten-
181 tially suitable climatic niche of species, differences which are interpreted as being driven by intrinsic (i.e.,
182 traits) and/or extrinsic factors (e.g., non-climatic environmental factors, biotic interactions, etc.) limiting the
183 realised distribution of the species (Estrada et al., 2018; Svenning and Skov, 2004). Intrinsic factors found
184 to constrain species’ realised distribution could also limit species’ ability to track climate change, and as
185 such could confer higher climate-change sensitivity. On the other hand, demographic approaches relying on
186 long-term population data have also been employed to explain population- and species’ responses to recent
187 climate change (Spooner et al., 2018). These approaches may suffer from similar complications regarding

188 the disentangling of climatic and non-climatic effects on observed responses (Williams et al., 2022). Finally,
189 ecological traits have been used in a predictive fashion, notably with frameworks aiming to assess species'
190 vulnerability to climate change (Foden et al., 2013; Pacifici et al., 2015), assuming that given traits confer
191 higher sensitivity to climate change.

192 Past studies have also investigated whether traits are associated with species' responses to habitat dis-
193 turbance. Most of these studies have used spatial data, inferring effects of land-use change on biodiversity
194 from 'space-for-time' substitutions (Davison et al., 2021; De Palma et al., 2018), such as in Newbold et al.
195 (2013), Quesnelle et al. (2014) or Nowakowski et al. (2017).

196 Further, trait-based approaches have also been employed to understand the signature of human impacts
197 on the diversity and variability of traits within ecological communities. To this end, a range of functional
198 diversity indices have been developed (Legras et al., 2018; Schleuter et al., 2010). Functional diversity
199 indices have been employed to evaluate the effects of land-use disturbance on the trait diversity of vertebrate
200 assemblages (most often at local and regional scales; Flynn et al. (2009); La Sorte et al. (2018); Tinoco
201 et al. (2018)), or to assess the projected effects of climate change on the trait diversity of vertebrate species
202 (Stewart et al., 2022).

203 Although trait-based approaches using ecological traits have been widely employed to understand the
204 effects of human-driven changes on vertebrate species, past studies have mostly been conducted at local to
205 regional scales (Davison et al., 2021), and have tended to focus on given taxa among the vertebrate classes
206 (Hevia et al., 2017). Thus, global comparative assessments of the relationships between ecological traits and
207 species' responses to human pressures across terrestrial vertebrates are lacking. Because of the diversity of
208 approaches used to investigate associations between species traits and human pressures in past work, and
209 because of the different spatial scales and taxonomic focus of past studies, identifying a set of traits that
210 emerge as important determinants of sensitivity to both land-use and climate change across terrestrial verte-
211 brates remains challenging. For instance, Quesnelle et al. (2014) found that reproductive rates, as estimated
212 from litter or clutch sizes, were significantly associated with sensitivity to habitat loss in wetland vertebrates;
213 however, Nowakowski et al. (2017) did not find any effect of clutch size on amphibian sensitivity to habitat
214 modification. The meta-analysis by MacLean and Beissinger (2017) highlights the lack of consensus in pub-
215 lished studies on the associations between traits and species' range shifts under recent climate change. Thus,
216 there remains a need to assess whether and which traits are associated with species' responses to human
217 pressures in terrestrial vertebrates.

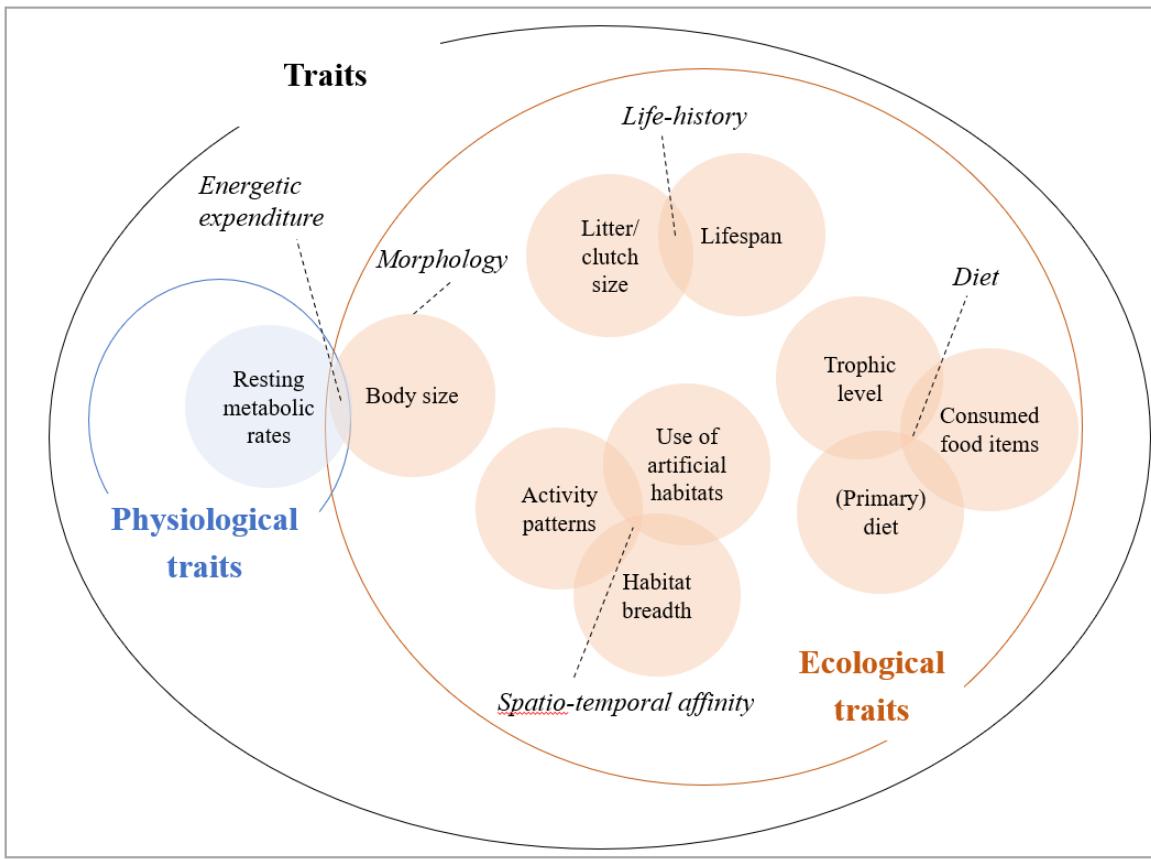


Figure 1.2: Typology of the traits used in this thesis. I distinguish *physiological traits*, which are typically measured to capture aspects of species' metabolism, performance or biochemistry, from *ecological traits*, in which I include traits relating to life-history, morphology, diet and spatio-temporal affinity of the species. Ecological traits may include traits for which the measurement needs to consider the relationship of a species to the surrounding environment (e.g., habitat breadth). I use physiological traits in Chapter 5 and ecological traits in Chapters 2, 3, 4, and 5.

218 1.3.3 Using physiological traits to understand land-use impacts on biodiversity and ecosys- 219 tem functioning

220 Physiological traits are typically measured to capture aspects of species' metabolism, performance or bio-
221 chemistry (Figure 1.2). Recent decades have seen advances in large-scale studies linking physiological traits
222 to macroecological patterns of species existence across levels of organisation (Burger et al., 2021; Chown
223 et al., 2004), as exemplified by the development of the ‘metabolic theory of ecology’ (Brown et al., 2004;
224 Gillooly et al., 2001). In particular, metabolic rates reflect the amount of energy used at the organismal level
225 (I define metabolic rates here as the rates at which an organism processes available energy, which is often
226 measured in the lab by estimating the amount of consumed O₂ by a whole organism over a period of time
227 (Auer et al., 2017; Sadowska et al., 2015)). As energy is a fundamental currency across all living organisms,
228 metabolic rates can be employed comparatively across species to investigate interspecific variation in ener-

229 genetic expenditure. Thus, past studies have focused on understanding intraspecific variation in metabolic rates
230 (Auer et al., 2017; Burton et al., 2011) as well as interspecific variation (such as variation in metabolic rates
231 with temperature; Clarke and Fraser (2004); or variation in metabolic rates with longevity – e.g., testing the
232 ‘pace of life’ theory; Stark et al. (2020)).

233 How species allocate their energy impacts almost all aspects of their persistence. Energetic expenditure
234 relates to food intake, which itself is constrained by the amount of available energy in the environment. Thus,
235 species’ energetic requirements are ultimately constrained and influenced by trade-offs between energetic-
236 expenditure allocation and resource intake (Auer et al., 2020). As land-use change profoundly modifies the
237 amount and the types of resources available, it follows that land-use change should impact the total amount
238 of energy processed by vertebrate assemblages. Further, the amount of energy required by species could
239 also be an important predictor of species’ ability to cope with a disturbed resource landscape. However,
240 to my knowledge, no study has yet investigated how land-use change affects the energetic requirements of
241 vertebrate assemblages, or whether species’ energetic requirements influence how species respond to land-
242 use change.

243 1.3.4 Thesis aims

244 As discussed above, studies investigating relationships between ecological traits and environmental change
245 have mostly been conducted at local to regional scale (Davison et al., 2021; Hevia et al., 2017), and have
246 mostly focused on single vertebrate classes or sub-taxa within particular classes. Thus, although response
247 traits to land-use and climate change have been identified in various vertebrate taxa, whether the effects of
248 such traits can be generalised geographically and taxonomically remains largely uncertain, emphasising the
249 need for global comparative assessments of the relationships between traits and species’ responses to human
250 threats.

251 In this thesis, I set out to fill in this gap by asking whether interspecific variation in ecological traits is as-
252 sociated with species’ land-use responses and with estimated climate-change sensitivity, at global scales, and
253 comparatively across the four terrestrial vertebrate classes. Such an assessment helps to understand which
254 species are at most risk from global changes, and may be useful for the prioritisation of conservation efforts.
255 My thesis also aims to highlight some of the consequences of land-use change for ecosystem functioning,
256 by investigating relationships between species’ energetic requirements (estimated from metabolic rates) and
257 land-use differences.

258 To investigate species’ land-use responses and species’ climate-change sensitivity, I use two indepen-
259 dent approaches. A fundamental difference between these approaches is the distinction between species’

260 *observed responses* and species' *sensitivity*. Throughout my thesis, assemblage-level and species-level *re-*
261 *sponses* to land use and land-use intensity are assessed using a ‘space-for-time’ approach (De Palma et al.,
262 2018). To this end, I use one of the most comprehensive databases recording species occurrence and abun-
263 dance in different land uses (the PREDICTS database, Hudson et al. (2014, 2017)). However, I estimate
264 *sensitivity* to climate change from properties of species' climatic niche space, and thus it is important to
265 emphasize that this does not allow a consideration of species' *responses* to climate change. Indeed, it is dif-
266 ficult to capture the responses of many species to climate change, given that this requires disentangling the
267 effects of climate change from that of other drivers of change over the considered time period (which can be
268 complex; MacLean and Beissinger (2017)), and also requires gathering data on the occurrence or abundance
269 of species over several decades (which may be particularly challenging when working at large taxonomic
270 scales). Conversely, I estimate species' *responses* to land-use change, as it is not possible to directly measure
271 species' sensitivity to land-use change from empirical data. However, other pressures may be interacting
272 with land use, such that quantifying land-use responses may require disentangling such interacting effects.
273 In my work, I did not take into account such possible interactive effects on responses, focusing on land use
274 only; this constitutes one of the limitations of my thesis.

275 **1.4 Detailed aims, hypotheses and outline of the following Chapters**

276 The overarching aims of my thesis are to investigate whether species traits are associated with species' land-
277 use responses and species' estimated climate-change sensitivity in terrestrial vertebrates, and to highlight
278 some of the consequences of global changes for ecosystem processes sustained by terrestrial vertebrates.
279 One of the obstacles that has hindered the application of trait-based approaches at large scales in animal
280 taxa is the lack of a centralised repository for readily available trait data, as emphasized by the recent calls
281 to compile and release trait data for animals (Junker et al., 2022; Kissling et al., 2018). Thus, collecting
282 trait data and investigating the current availability of the data for terrestrial vertebrates is an important and
283 necessary prerequisite to any analysis. In Chapter 2, I present a data collection of ecological traits for
284 terrestrial vertebrates. Because using similar traits in the different vertebrate classes is necessary to be able
285 to make comparisons among vertebrate classes, I target seven classes of traits that are commonly used across
286 taxonomic groups: body mass/size, lifespan, litter/clutch size, trophic level, diel activity, habitat breadth,
287 and habitat specialisation (characterising whether a species is able to use artificial habitats). I am not able
288 to consider intraspecific variation in the data compilation, since multiple measurements of trait values do
289 not exist for many vertebrate species. Chapter 2 also assesses the availability of the trait data across the

290 terrestrial vertebrate classes, and investigates whether the trait data present global taxonomic, phylogenetic
291 and spatial biases. On the basis of past work (Titley et al., 2017), I predict that amphibians and reptiles are
292 under-sampled compared to mammals and birds. Further, I hypothesize that trait data are less abundant for
293 the narrower-ranging species and in species-richer regions.

294 At the assemblage level, the diversity of species traits can be summarised with functional diversity indices
295 (Legras et al., 2018; Schleuter et al., 2010; Villéger et al., 2008). Past research has shown that land-use
296 disturbances affect the functional composition of vertebrate assemblages (Flynn et al., 2009; Tinoco et al.,
297 2018). However, to the best of my knowledge, a global assessment of how the functional diversity of local
298 terrestrial vertebrate assemblages respond to land use and land-use intensity, within and across taxonomic
299 classes, has not yet been undertaken. In Chapter 3, I aim to fill in this gap. To this end, I combine the trait
300 data collected in Chapter 2 with the PREDICTS database, after imputing missing trait values (as described
301 in Chapter 2). I hypothesize that the functional diversity of vertebrate assemblages in disturbed land uses is
302 lower than in undisturbed areas. I further predict that decreases in functional diversity in disturbed land uses
303 are driven by high levels of functional loss and that observed declines in functional diversity exceed those
304 expected from random species loss.

305 Chapter 3 highlights the effects of land-use change on the functional composition of vertebrate assem-
306 blages, but does not allow an assessment of the effects of particular traits on species' land-use responses,
307 as multidimensional interspecific trait variation is summarised into single indices of functional diversity.
308 Chapter 4 aims at assessing such effects, by investigating whether species traits are associated with species'
309 land-use responses and species' climate-change sensitivity. In addition to the traits considered in Chapter 3,
310 Chapter 4 includes dietary traits and species geographical range area. Although geographical range area is
311 not a trait *per se*, it has been shown to influence species' responses to land-use and climate change (Newbold
312 et al., 2018; Thuiller et al., 2005), and it is likely an important determinant of climate-change sensitivity.
313 Thus, in Chapter 4, I define 'ecological characteristics' as encompassing both traits and geographical range
314 areas. I investigate whether these ecological characteristics are associated with species' land-use responses
315 on the one hand and with species' estimated climate-change sensitivity on the other hand, comparatively
316 among the terrestrial vertebrate classes. To the best of my knowledge, Chapter 4 constitutes the first global
317 comparative assessment, across terrestrial vertebrate classes, of associations between species' ecological
318 characteristics and both land-use responses and estimated climate-change sensitivity.

319 Chapter 5 develops our understanding of the impacts of land-use change on ecosystem functioning by
320 focusing on species' energetic requirements, which is interesting for at least two reasons: first, because en-
321 ergetic requirements relate to resource intake, and as such reflect the amount of energy that is processed by

322 different trophic groups, which can inform about ecosystem functioning; second, because species persistence
323 is constrained by trade-offs in energy allocation among diverse processes (e.g., maintenance, growth, repro-
324 duction), such that energetic requirements are likely important determinants of species' ability to cope with
325 disturbances. Yet, there has been no study so far investigating relationships between energetic requirements
326 and land-use differences in terrestrial vertebrates. In Chapter 5, I collect resting metabolic rate estimates for
327 vertebrates, that is, the estimated minimum amount of energy necessary for organismal maintenance (esti-
328 mated at the species level), and I combine these estimates with the PREDICTS database. I then assess the
329 effects of land use on the total energetic requirements of vertebrate assemblages (conceptually comparable
330 to 'community metabolism'; Migné et al. (2015)). Second, I assess whether species' energetic requirements
331 influence species persistence in disturbed land uses (after controlling energetic requirements for the effects of
332 body mass and taxonomy, which explain most of the interspecific variation in metabolic rates; White (2011)).
333 Assuming that there is less energy available in disturbed land uses, I hypothesize that the assemblage-level
334 energetic requirements of vertebrates are lower in disturbed land uses compared to natural habitats, and that
335 species with lower mass-independent energetic requirements are favoured over species with higher mass-
336 independent energetic requirements in disturbed land uses. Chapter 5 highlights the impacts of land-use
337 change on vertebrate community metabolism and develops our understanding of the factors that shape how
338 species respond to changes in land use.

339 Finally, in Chapter 6, I summarise the findings of my thesis, I highlight some of the limitations, and I ex-
340 amine the relevance of my findings for the field. By investigating whether traits are associated with species'
341 land-use responses and climate-change sensitivity across the terrestrial vertebrates, my thesis furthers our
342 understanding of what could render species more sensitive to human threats, underlines possible modifica-
343 tions to ecosystem functioning, and stresses the role and usefulness of vertebrate trait data and ecological
344 knowledge for understanding species- and community-level responses to human pressures.

³⁴⁵ **2 | Global gaps and biases in trait data for**
³⁴⁶ **terrestrial vertebrates**

³⁴⁷ **Keywords**

³⁴⁸ Terrestrial vertebrates; traits; coverage; completeness; taxonomic biases; spatial biases; phylogenetic biases.

³⁴⁹ **Abstract**

³⁵⁰ Trait data are increasingly used in studies investigating the impacts of global changes on the structure and
³⁵¹ functioning of ecological communities. Despite a growing number of trait data collations for terrestrial
³⁵² vertebrates, there is to date no global assessment of the gaps and biases the data present. Here, I assess
³⁵³ whether terrestrial vertebrate trait data are taxonomically, spatially and phylogenetically biased. I compile
³⁵⁴ seven ecological traits and quantify coverage as the proportion of species for which an estimate is available.
³⁵⁵ For a species, I define completeness as the proportion of non-missing values across traits. I assess whether
³⁵⁶ coverage and completeness differ across classes and examine phylogenetic biases in trait data. To investi-
³⁵⁷ giate spatial biases, I test whether wider-ranging species have more complete trait data than narrow-ranging
³⁵⁸ species. Additionally, I test whether species-rich regions, which are of most concern for conservation, are
³⁵⁹ less well-sampled than species-poor regions. My results show that mammals and birds are well-sampled
³⁶⁰ even in species-rich regions. For reptiles and amphibians (herptiles), only body size presents a high cov-
³⁶¹ erage (>80%), as well as habitat related variables (for amphibians). Herptiles are poorly sampled for other
³⁶² traits. The shortfalls are particularly acute in some species-rich regions and for certain clades. Across all
³⁶³ classes, geographically rarer species have less complete trait information. Hence, trait information is less
³⁶⁴ available on average in some of the most diverse areas and in geographically rarer species, both critical for
³⁶⁵ biodiversity conservation. Gaps in trait data may impede our ability to conduct large scale analyses, while

366 biases can impact the validity of extrapolations. A short-term solution to the problem is to estimate missing
367 trait data using imputation techniques, while a longer-term and more robust filling of existing gaps requires
368 continued data collection efforts.

369 **2.1 Introduction**

370 Species traits are fundamental to ecological and evolutionary research. Comparative studies regularly use
371 trait data across organisms to understand evolutionary processes and species coexistence (Escudero and
372 Valladares, 2016; Zamudio et al., 2016), to investigate global patterns of life forms and functions (Díaz et
373 al., 2016), or to assess species' vulnerability to environmental changes (Böhm et al., 2016; Pacifici et al.,
374 2015; Pearson et al., 2014). Because traits influence species' ability to cope with environmental changes
375 (Newbold et al., 2013) and underpin species' contributions to ecosystem processes (Lavorel and Garnier,
376 2002; Violle et al., 2007; Wong et al., 2018), they play an increasingly important role in functional and
377 conservation ecology.

378 Past and recent efforts to collate and release trait data in the public domain have facilitated the develop-
379 ment of trait-based research. For instance, a global trait database has been published for plants (Kattge et al.,
380 2011). As of May 2020, data from this database had been used in 297 publications since its release (Ac-
381 tivity report, 18/06/2020, <https://www.try-db.org/TryWeb/Home.php>). Such databases hence
382 constitute invaluable research tools and have the potential to greatly advance the field.

383 Vertebrates are one of the most studied taxa (Titley et al., 2017). There are now diverse sources of eco-
384 logical traits for vertebrate groups (primates: Galán-Acedo et al., 2019; mammals: 'PanTHERIA', Jones et
385 al., 2009; amniotes: Myhrvold et al., 2015; amphibians: 'AmphiBIO', Oliveira et al., 2017). These datasets
386 stem from important efforts to collate published estimates of trait data and make them readily available. Trait
387 data have also been made available on online platforms (for instance, the Global Assessment of Reptile Dis-
388 tribution initiative: <http://www.gardinitiative.org/>; the IUCN Red List of Threatened Species:
389 <https://www.iucnredlist.org/>; BirdLife data zone: <http://datazone.birdlife.org/>
390 home).

391 Nevertheless, despite the importance of vertebrate species in global research outputs, there is no single
392 source for vertebrate ecological traits. Consequently, researchers wishing to conduct comparative studies
393 across vertebrate groups may have to collate trait data from a range of sources (such as in Cooke et al.
394 (2019a,b) or in González-Suárez et al. (2018)), a time-consuming prerequisite which may be a limiting
395 step of the research process. Indeed, collating data from heterogeneously-formatted sources presents many

396 challenges (Schneider et al., 2019), particularly when working across a large number of species. For instance,
397 traits may be measured differently across datasets; units may be inconsistent; and taxonomic resolution and
398 nomenclature may vary.

399 The lack of a curated, readily available global database for vertebrate ecological traits impedes our ability
400 to conduct cross-taxon comparative studies at global scales. However, efforts to collate data into a single
401 database are limited by the availability of underlying data. Because there exist important gaps in biodiversity
402 knowledge (Hortal et al., 2015), trait datasets are often incomplete, with many species lacking estimates for
403 many traits. The incompleteness of ecological trait data at the species level has been termed the ‘Raunkiær
404 shortfall’ by Hortal et al. (2015). Furthermore, incomplete trait data are likely to be biased. Biases in trait data
405 may be the consequence of uneven taxonomic and spatial collection effort, with a set of charismatic or easily
406 detectable species being more completely sampled. For instance, González-Suárez et al. (2012) investigated
407 biases in global trait information in mammals. They notably found that the availability of mammalian trait
408 data was geographically and phylogenetically biased, with larger and more widely distributed species being
409 better sampled. In addition, data availability also differed across IUCN Red List extinction risk categories,
410 with threatened species (Critically Endangered, Endangered or Vulnerable) being less well sampled for traits
411 than non-threatened species (Least Concern or Near Threatened).

412 A major issue with incomplete, biased data is the introduction of bias in subsequent analyses. Assessing
413 the amount of missing data as well as the so-called ‘missingness mechanism’ – whether missing data are
414 missing at random, as opposed to there being systematic biases in the way missing values are distributed, see
415 Baraldi and Enders (2010) – is an important prerequisite. Indeed, there exist diverse techniques to deal with
416 data missingness. The simplest one consists of retaining complete cases only by filtering out missing values
417 (case deletion, see Nakagawa and Freckleton (2008)). Nevertheless, case deletion may lead to biased param-
418 eter estimates and erroneous conclusions when values are not missing at random (González-Suárez et al.,
419 2012). Therefore, it is critical to determine the most appropriate way to deal with data incompleteness. For
420 instance, previous studies using terrestrial vertebrate trait data have employed multiple imputation techniques
421 to fill in the gaps (Cooke et al., 2019a; González-Suárez et al., 2012). Yet, imputation techniques could be
422 sensitive to non-randomness in trait data. Phylogenetic biases (where some clades are under-sampled com-
423 pared to other clades) could notably impact the performance of several imputation approaches. It is thus
424 vital to characterise the gaps in trait data prior to any analysis. However, there has been no study to date
425 investigating global patterns in the availability of trait data across terrestrial vertebrates.

426 Here, I aim to assess the global state of trait data in terrestrial vertebrates. I focus on a set of traits that are
427 available across the four classes and that are commonly used by ecologists: body size; litter or clutch size;

428 longevity; trophic level; activity time; habitat breath; and a measure of habitat specialisation. I quantify and
429 compare the gaps in trait data across classes by calculating the coverage of each trait across species, and the
430 completeness of trait estimates for each species (Box 1). I investigate taxonomic, spatial and phylogenetic
431 biases in trait coverage and completeness.

432 Given that biodiversity research is globally biased towards birds and mammals (Titley et al., 2017),
433 I hypothesise that herptiles are less well sampled for traits than mammals and birds, having both lower
434 coverage and completeness.

435 Furthermore, building upon previous studies conducted on mammals (González-Suárez et al., 2012), I
436 hypothesize that species rarity influences completeness, focusing on species geographical range size as one
437 aspect of rarity. Widely distributed species could be better sampled than narrowly distributed species because
438 their ranges overlap with more study sites, regardless of their abundance. As such, I test whether species
439 geographical range size explains trait completeness.

440 It is well established that global research effort is distributed unequally (United Nations Educational
441 Scientific and Cultural and Organization, 2015), with patterns underpinned by various geographical and so-
442 cieconomic factors. For instance, countries with higher gross domestic product tend to host a larger number
443 of research institutions (Martin et al., 2012). The proximity of research infrastructures and the accessibility
444 of survey sites play an important part in explaining the global distribution of knowledge (Hortal et al., 2015).
445 As a result of these factors, biodiversity data gaps tend to be greater in tropical areas (Collen et al., 2008).
446 Tropical areas have the greatest species richness, and so these data biases are of great concern for biodiversity
447 conservation. Whether species-rich regions are systematically under-sampled for traits compared to species-
448 poor regions is thus important to assess, given the significance of species-rich areas for global conservation.
449 Here, I investigate spatial biases in trait completeness, hypothesizing that species-rich areas are on average
450 less well sampled than species-poor areas.

451 Finally, I investigate phylogenetic biases in the trait data. I assess whether particular clades have received
452 more attention than others by looking for patterns in the distribution of trait completeness across the terminal
453 branches of phylogenetic trees in each class.

Box 1. Definitions

Functional trait: Sensu stricto, a characteristic measurable at the level of an individual and that influences organismal fitness or performance (Violle et al., 2007). In this thesis, I broaden this definition to include ‘ecological’ traits (e.g., the number of habitats used by a species), where the relationship of a species to the surrounding environment needs to be considered. Ecological traits may be estimated by aggregating data across multiple individuals.

Trait completeness: For a given species, the proportion of traits for which an estimate is available.

Trait coverage: For a given trait, the proportion of species for which an estimate is available.

454 2.2 Methods

455 I produced class-specific trait datasets that were made available on figshare (https://figshare.com/articles/dataset/Global_gaps_in_terrestrial_vertebrate_trait_data/10075421/)
456 2). Data compilation and all analyses were conducted with R version 3.5.1 (R Core Team, 2018). Distribu-
457 tion maps were processed using both R and the ‘ArcPy’ package available in ArcGIS v.10.6 (ESRI, 2011)
458 (implemented in Python 2.7; van Rossum, G. (1995)).

460 2.2.1 Trait data collection

461 Sources and taxonomic matching

462 I used freely accessible secondary sources in my compilation (Table 2.1), selected for their broad taxonomic
463 coverage and/or for their frequent use in macroecological studies. Across sources, similar species could
464 appear under synonymous names. This was a potential problem for matching sources by binomial names.
465 Indeed, synonymy can artefactually decrease trait coverage, when trait information is not available across
466 all synonyms. Notably, difficulties arise when species have been divided into several subspecies or when
467 different subspecies are clumped together. Systematic manual checks could not be applied considering the
468 scale of the data collection (there were >39,000 unique binomial names across sources). I developed a
469 procedure aiming at identifying one accepted name for each of the binomial names found across sources.
470 When I could not find an accepted name, I used the original name. Figure 2.1 summarizes the main steps;
471 similar solutions have been used in other large-scale studies (Cooke et al., 2019a).

472 Briefly, the procedure consisted of extracting synonyms from the IUCN (IUCN, 2020) or from the In-
473 tegrated Taxonomic Information System (ITIS; <https://www.itis.gov/>), using the ‘rredlist’ (Cham-
474 berlain, 2018) and ‘taxize’ (Chamberlain and Szöcs, 2013) R packages. One accepted name was assigned to

475 each synonym. I produced a "Synonym" dataset that I have also made available. I then normalized taxonomy
476 across sources by replacing binomial names with their identified accepted name where applicable.

477 Given that different taxonomic backbones could be used to correct for taxonomy, I make two versions
478 of the trait compilations available (corrected and not corrected for taxonomy), meaning that users are free to
479 apply their own corrections; for example, taxonomy could be aligned to that of class-specific sources, such
480 as The Reptile Database, the American Museum of Natural History's Amphibian Species of the World, the
481 Mammal Diversity Database or the International Ornithological Congress World Bird List. Datasets cor-
482 rected for taxonomy contain 11,634 species of birds, 5,381 mammals, 10,612 reptiles and 6,990 amphibians.
483 Where no taxonomic correction was applied when matching sources, the compiled datasets contain 13,501
484 birds, 5,791 mammals, 11,012 reptiles and 8,583 amphibians. For more information, see Appendix 1 (S2.1:
485 'Taxonomic corrections'; Figure S2.1).

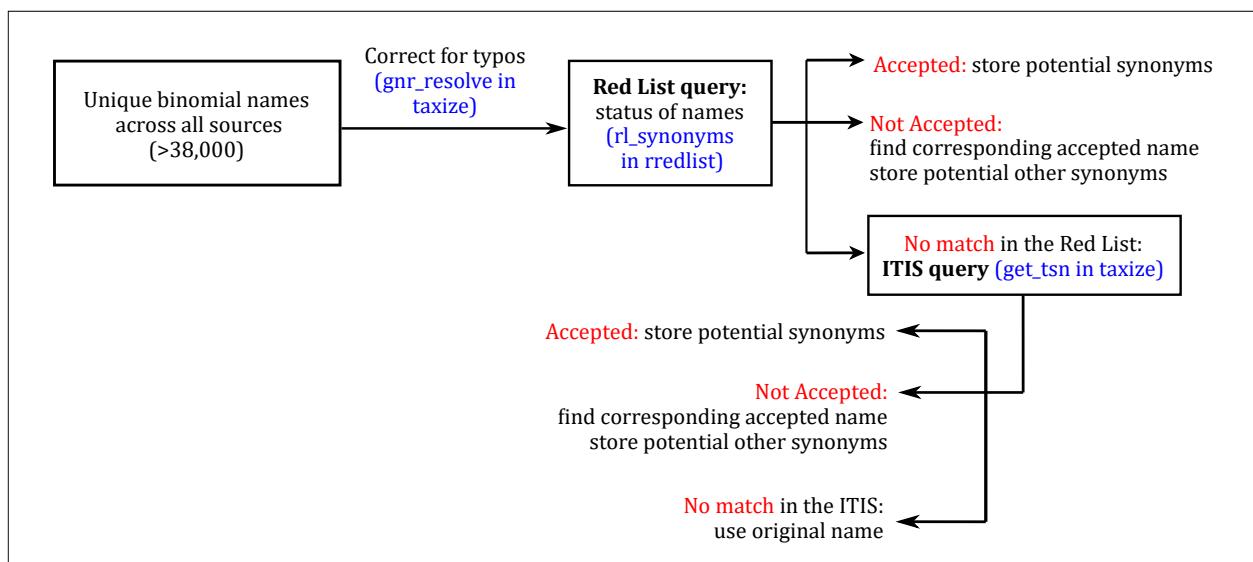


Figure 2.1: Procedure used to identify the accepted names of species. I extracted, where possible, the accepted names of species from either the IUCN Red List or the Integrated Taxonomic Information System (ITIS). Figure reproduced from Etard et al. (2020).

486 Compilation methods

487 For continuous traits, I took the median value within species when multiple estimates were available from
488 different sources, after removal of any repeated values, which were assumed to represent estimates duplicated
489 across secondary compilations and derived from the same underlying primary sources. Although intraspe-
490 cific variation is increasingly being recognized to have important effects on ecological systems (Bolnick
491 et al., 2011; Des Roches et al., 2018; González-Suárez and Revilla, 2012; Siefert et al., 2015), it was not

Table 2.1: Data sources for each trait. Abbreviations: A = amphibians; B = birds; BL = body length; BM = body mass; DA = diel activity time; GL = generation length; H = habitat data; LCS = litter or clutch size; L/ML = longevity or maximum longevity; M = mammals; MA = age at sexual maturity; R = reptiles; RS = range size; TL = trophic level. Note. Data sources may contain more traits than shown here. Tick marks in parentheses indicate that the trait was present in the data source but that another closely related trait with a better coverage was used instead. The tilde character (~) before a tick mark indicates that I derived trophic levels from species diet. ¹ <http://datazone.birdlife.org/home>; ² <https://www.iucnredlist.org/resources/spatial-data-download>; ³ <http://apiv3.iucnredlist.org/api/v3/docs##general>. Table reproduced from Etard et al. (2020).

Sources	Taxa	Traits									
		BM	BL	L/ML	MA	GL	LCS	TL	DA	RS	H
Oliveira et al. (2017)	Amphibians	(✓)	✓	(✓)	✓		✓	~✓	✓		
Cooper et al. (2008)							✓				
Sodhi et al. (2008)			✓								
Wilman et al. (2014)	Birds	✓						~✓	✓		
BirdLife ¹		✓				✓					✓
Jones et al. (2009)	Mammals	✓	(✓)	(✓)	(✓)		✓		✓		
Kissling et al. (2014)								✓			
Gainsbury et al. (2018)								✓			
Wilman et al. (2014)		✓							✓		
Pacifici et al. (2015)		✓				✓					
Scharf et al. (2015)	Reptiles	✓		✓	(✓)		✓	✓	✓		
Vidan et al. (2017)										✓	
Stark et al. (2018)		✓		✓			✓		✓		
Schwarz and Meiri (2017)								✓			
Novosolov et al. (2017)		✓						✓			
Novosolov et al. (2013)							✓				
Slavenko et al. (2016)		✓									
Feldman et al. (2016)		✓									
Meiri (2018)					✓		✓	✓	✓		
Meiri et al. (2015)								✓	✓		
Roll et al. (2017)										✓	
Myhrvold et al. (2015)	B, M, R	✓	✓	✓	(✓)		✓				
IUCN (2020) ²	A, B, M										✓
IUCN (2020) ³	All										✓

492 feasible to obtain measures of intraspecific variability from all sources; therefore, estimates were provided
493 as a single measure for each species. For some species and some traits, measures were provided separately
494 for females and males. In such cases, I first obtained the mean of these two measures.

495 Across sources, there were multiple traits related to each of body size and lifespan. For instance, body
496 mass and/or body length information could be provided. Different proxies were also available for lifespan,

497 such as the age at sexual maturity or generation length. In such cases, I focused on the trait presenting the
498 highest coverage.

499

- **Body size**

500 Adult body mass estimates were compiled for mammals, birds and reptiles. Body length information
501 was compiled for amphibians, because the coverage for body length was higher than that for body
502 mass. Body mass and body length are known to scale allometrically, although the allometric rela-
503 tionship differs across amphibian clades (Santini et al., 2018). In the amphibian dataset, Pearson's
504 correlation coefficient between log(Body mass) and log(Body length) was 0.71 (data points shown in
505 Appendix 1, S2.2:‘Additional information for trait compilation’, Figure S2.2).

506

- **Longevity**

507 I defined longevity as the lifespan of an individual and maximum longevity as the longest lifespan
508 reported. I used closely related traits when longevity/maximum longevity was not available or when
509 longevity/maximum longevity had a poorer coverage than a related trait. I selected the age at sexual
510 maturity for amphibians; Pearson's correlation coefficient between log(Age at sexual maturity) and
511 log(Maximum longevity) was 0.55 (Appendix 1, Figure S2.2). I compiled the generation length for
512 mammals and birds. The correlation between log(Generation length) and log(Longevity) was 0.74 for
513 mammals and 0.70 for birds (data points shown in Appendix 1, S2.2, Figure S2.3). Finally, I used
514 maximum longevity directly for reptiles.

515

- **Litter or clutch size**

516 The number of offspring (litter size) or eggs (clutch size) was compiled directly from the sources and
517 treated as equivalent across classes. I reported measures of central tendencies provided by the sources
518 where applicable; otherwise, I calculated range midpoints (mean of smallest and largest reported lit-
519 ter/clutch sizes).

520

- **Trophic level**

521 In all classes, species were described as omnivores, carnivores or herbivores. For reptiles and mam-
522 mals, this information was compiled directly from the sources. For amphibians and birds, trophic
523 levels were not provided. For these two classes, I inferred trophic levels from dietary information
524 (Table 2.1). For birds, I used the primary diet (based on food items recorded as composing $\geq 50\%$
525 of the diet of a species). Diet for amphibians was described without respect to the percentage use of
526 food items; simply as a binary record of whether or not food items were used. In both cases, species

527 recorded to only consume plant-based resources (seeds, nectar, fruit or other plant material) were
528 classified as herbivores. Species consuming only animal resources (invertebrates or vertebrates) were
529 classified as carnivores. Species consuming a mixture of plant and animal resources were classified as
530 omnivores.

531 **• Activity time**

532 Species were described as being either nocturnal or non-nocturnal. Despite a higher resolution of ac-
533 tivity time information in some of the sources (e.g., species being described as cathemeral, crepuscular
534 or diurnal), I adopted the classification of the source with the lowest resolution (EltonTraits: Wilman
535 et al. (2014), for birds), in order to have consistent information across classes. As such, all species
536 defined as diurnal, cathemeral or crepuscular were classified as non-nocturnal, as opposed to species
537 classified as strictly nocturnal.

538 **• Habitat breadth**

539 I used IUCN habitat data (IUCN, 2020), which describe species habitat preferences and the suitabil-
540 ity and importance of each habitat. I defined habitat breadth as the number of habitats a species was
541 known to use, using level 2 of the IUCN Habitat Classification Scheme for description of habitat
542 types (divided into: Forest, Savanna, Shrubland, Grassland, Wetland, Rocky areas, Caves and subter-
543 ranean, Desert, Marine, Marine intertidal or coastal/supratidal, Artificial, Introduced vegetation, and
544 Other/Unknown.) Note that the total number of habitats, determined by including those that qualify as
545 artificial, correlates positively with the number of natural habitats used (Figure 2.2).

546 **• Use of artificial habitats**

547 For a species, I recorded whether any artificial habitat was reported to be suitable in the IUCN habitat
548 data.

549 Finally, the compiled datasets contain an additional column, ‘Note’, where I reported species found to
550 be extinct or extinct in the wild (EW). I used species Red List status and information from Meiri (2018) to
551 flag such species. I reported 75 extinct/EW species for mammals, 160 for birds, 34 for amphibians and 53
552 for reptiles. It is likely that the datasets contain extinct species that I could not flag, because they were not
553 recorded as extinct in the sources I used.

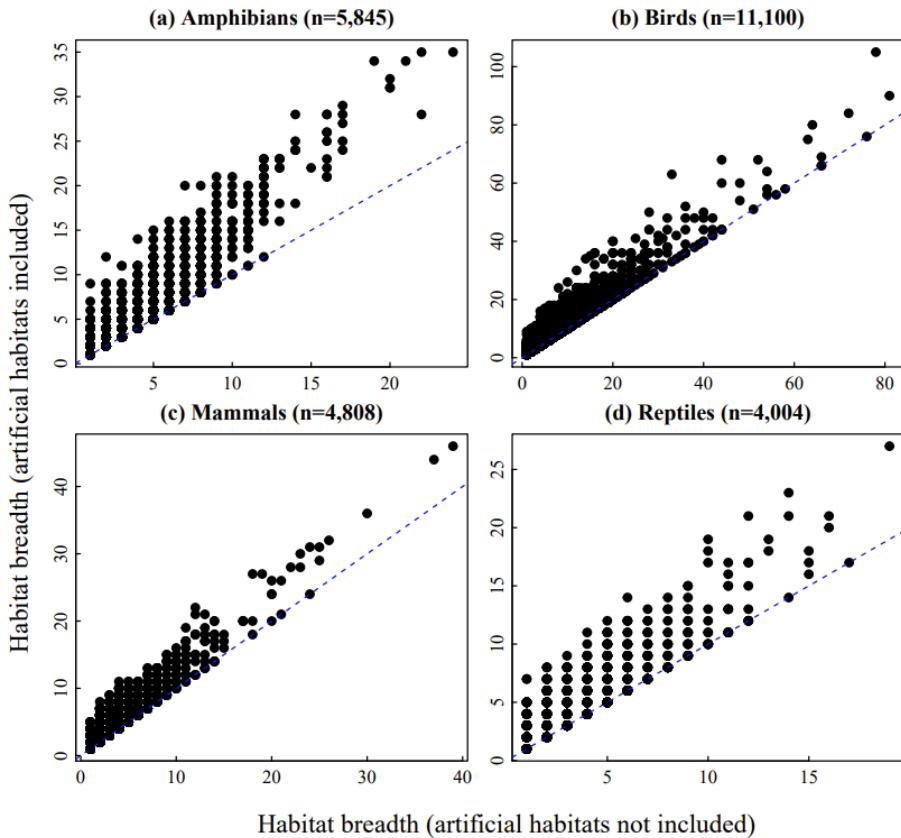


Figure 2.2: Number of natural and artificial habitats used by species against number of strictly natural habitats used by species. Pearson's correlation coefficients show a high positive correlation between these two metrics of habitat breadth in all terrestrial vertebrate classes: 0.92 for amphibians (a), 0.95 for birds (b), 0.94 for mammals (c), and 0.90 for reptiles (d).

554 Phylogenies

555 I used class-specific phylogenetic trees downloaded on 13 April 2020. For mammals, I used ‘complete’
 556 trees from Faurby et al. (2018, 2020), downloaded from <https://zenodo.org/record/3690867#.Xyc5wyhKhPZ>. For amphibians, birds and squamates, I obtained trees from <https://data.vertlife.org/>. The original sources were as follows: Jetz et al. (2012) for birds; Jetz and Pyron (2018) for amphibians; and Tonini et al. (2016) for squamates. For each class, a distribution of 1,000 trees was available.
 557
 558 For plotting purposes, I obtained consensus trees using the TreeAnnotator program of the BEAST software
 559 (Bouckaert et al., 2019).

562 Species distributions

563 I obtained extent-of-occurrence distribution maps for reptiles from Roll et al. (2017), available at: <https://datadryad.org/stash/dataset/doi:10.5061/dryad.83s7k> (downloaded 13 April 2020).
 564
 565 For mammals and amphibians, species distribution maps were obtained from the IUCN Red List (IUCN

566 (2020), downloaded 13 April 2020); for birds, they were obtained from BirdLife International (<http://datazone.birdlife.org/species/requestdis>, downloaded 17 April 2020).

568 For amphibians, mammals and birds, I selected areas of extant or probably extant presence only. Addi-
569 tionally, I selected areas where species were resident or present during the breeding season, and I excluded
570 areas occupied during the non-breeding season or where species were considered vagrant.

571 In addition, for all classes, I excluded occupied areas that fell outside the known elevational limits of
572 species, where such data were available. Lower and upper elevational limits were retrieved from the IUCN
573 Red List (queried using the ‘rredlist’ package) and were available for approximately half of the species
574 (Appendix 1, S2.3: ‘Cutting distribution maps by altitudinal limits’, Figure S2.4). Decreases in range sizes
575 were observed after cutting distribution maps by the known elevational limits (Appendix 1, Figure S2.5).

576 **2.2.2 Investigating gaps and biases in trait data**

577 I used trait coverage and completeness to investigate taxonomic, phylogenetic and spatial biases in the trait
578 data. Table 2.2 summarizes the sample sizes (number of species) in each of the following analyses. Note
579 that species for which completeness was 0% were included in all analyses (for more details, see Figure 2.3).
580 Also note that I did not filter out species identified as extinct or extinct in the wild, because they represented
581 a small proportion of the datasets (0.48% for amphibians, 1.4% for both birds and mammals, and 0.50% for
582 reptiles) and also because I could not exclude such species systematically, because it is likely that I did not
583 flag them all.

Table 2.2: Number of species for each analysis. All species represented in the trait datasets were included in (1). All species from the class-specific phylogenetic trees or from the distribution maps that matched with species in the trait datasets were included in (2) and (3). *Table reproduced from Etard et al. (2020).*

	(1) Taxonomic biases	(2) Phylogenetic biases	(3) Spatial biases
Amphibians	6,990	6,170	5,650
Birds	11,634	8,315	10,802
Mammals	5,381	5,171	5,046
Reptiles	10,612	9,404	9,382

584 **Taxonomic biases**

585 I tested whether completeness varied across taxonomic class using pairwise Wilcoxon rank sum tests. I
586 tested for the extent and performance of the taxonomic corrections by looking at trait coverage when taxo-
587 nomic corrections are applied and when no correction is applied (Appendix 1, S2.4: ‘Impact of taxonomic
588 corrections on trait coverage’, Figure S2.6).

589 **Phylogenetic biases**

590 Initially, to assess whether more closely related species were more likely to be similar in trait completeness,
591 I estimated the phylogenetic signal in completeness with Pagel’s λ (Pagel, 1999) in each class. I used a boot-
592 strapping approach, calculating λ for each of 50 trees randomly sampled in each class (using the ‘phylosig’
593 function of the ‘phytools’ R package; Revell, 2012). I then estimated the mean and 95% confidence intervals
594 (95% CIs) of λ . Sample sizes for computing λ (number of species represented in both the phylogenies and
595 trait datasets) are shown in Table 2.2.

596 I then plotted within-family median completeness in phylogenetic trees built at the family level, using
597 the consensus trees. Within-family median completeness was calculated using taxonomic information in the
598 trait datasets (sample sizes shown in Table 2.2).

599 **Spatial biases**

600 I first investigated whether wider-ranging species were more likely to be better sampled than narrow-ranging
601 species. I tested for a relationship between species range size and trait completeness. I fitted a generalized
602 linear model with a Poisson error distribution (directly using the number of sampled traits, ‘N_{traits}’, rather
603 than the proportion (completeness)). Class was added as a predictor interacting with range size; thus the
604 model was:

605 $N_{traits} \sim \log(\text{Range size}) * \text{Class.}$

606 Second, I mapped assemblage-level median completeness. Assemblages were characterized at the pixel level
607 at 50 km² resolution. I determined pixel-level composition and richness by stacking species geographical
608 distributions. I then calculated median completeness across species in each pixel. I show the resulting maps
609 for herptiles in the main text, and for mammals and birds in Appendix 1 (Figure S2.7; median completeness
610 was very high across most pixels for mammals and birds). In addition, I provide maps of assemblage-
611 level mean completeness and standard deviation for all classes in Appendix 1 (Figures S2.8 and S2.9 show
612 corresponding maps; Figure S2.10 shows standard deviation against species richness).

I then tested for a spatial correlation between species richness and median completeness. Given that median completeness was very high across most pixels for mammals and birds, I fitted such models for herptiles only. I fitted spatial autoregressive lag models to explain assemblage-level median completeness as a function of species richness (using the function ‘lagsarlm’ of the ‘spatialreg’ package (Bivand and Piras, 2015; Bivand et al., 2013a; Bivand et al., 2013b)). Given that responses could vary geographically, I included the biogeographical realm as an interacting factor (using the World Wide Fund for Nature (WWF) ecoregion shapefile to characterise realms, obtained from <https://www.worldwildlife.org/publications/terrestrial-ecoregions-of-the-world>); the considered realms were Afrotropics, Australasia, Indo-Malayan, Nearctic, Neotropics and Palaearctic. To improve normality, I arc-sin square-root transformed completeness values and log-transformed species richness. The ‘lagsarlm’ function allows for a consideration of spatial autocorrelation in the dependent variable by estimating the autoregressive lag coefficient, ρ , associated with an n-by-n matrix of spatial weights, W . The final model was:

$$\text{arcsin}(\sqrt{\text{Completeness}}) \sim \log(\text{Species richness}) * \text{realm} + \rho \cdot W \cdot \text{arcsin}(\sqrt{\text{Completeness}}).$$

The value of W was estimated using the functions ‘tri2nb’ and ‘nb2listw’ of the ‘spdep’ package (Bivand and Wong, 2018; Bivand et al., 2013b). Fitting the model using all grid cells was computationally intractable; therefore, I randomly sampled cells for this analysis (using 30% of the grid cells in each realm). I selected grid cells where species richness was higher than three to avoid sampling issues. I fitted separate models for amphibians and reptiles, because when adding class as an interacting predictor, the same cells (with the same coordinates) might be sampled for multiple classes, whereas the ‘tri2nb’ function does not tolerate duplicated coordinates.

2.3 Results

2.3.1 Taxonomic biases in trait information

Trait coverage for mammals and birds was overall high (Figure 2.3a; mean and median coverage across traits: 89% and 95% for mammals; 84% and 85% for birds). In both cases, litter/clutch size was the trait with the poorest coverage (61% for mammals and 59% for birds). Coverage exceeded 80% for all other traits (except trophic level for birds, at 75% coverage).

Conversely, trait coverage was more variable for herptiles, and poorer overall (Figure 2.3a; mean and median trait coverage: 47% and 32% for amphibians, 46% and 38% for reptiles). Coverage exceeded 80% only for body size in both reptiles and amphibians and for habitat related traits in amphibians only. In all

other cases coverage was <55%, with very little information available for longevity-related traits.

Trait completeness (proportion of non-missing trait values for a species) reflected similar biases (Figure 2.3b). The distribution of trait completeness varied significantly among classes (pairwise Wilcoxon rank sum test: p-value<0.0001 in all cases). Distributions were highly left skewed in mammals and birds (skewness: -2 and -1.6). 84% of all mammalian species and 80% of avian species fell in the 80–100% completeness range. Moreover, the completeness distribution was moderately right skewed for reptiles (skewness: 0.4), and slightly right skewed for amphibians (skewness: 0.02). 56% of all reptiles and 57% of amphibians fell in the 0-50% completeness range.

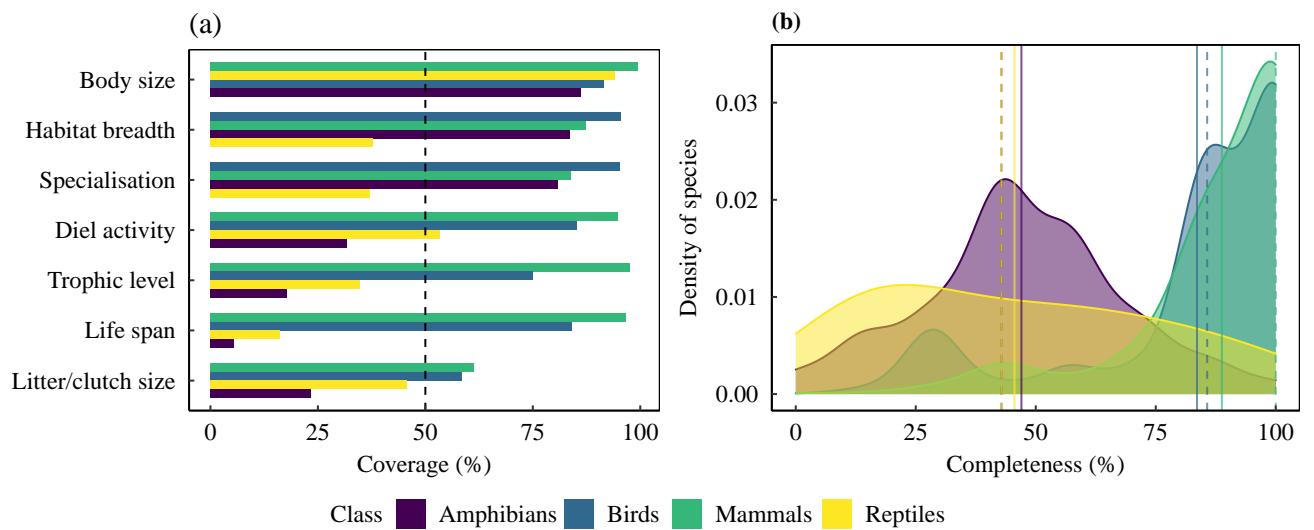


Figure 2.3: Trait coverage and completeness across species. (a) I defined coverage as the proportion of species for which an estimate is available for a given trait. The dashed line represents 50% coverage. (b) Trait completeness is the proportion of estimated traits for a species. Here, I show the distribution of completeness. Continuous lines represent the mean trait completeness for each class, whereas dashed lines represent the median trait completeness. Note that there were species with 0% completeness (230 species for amphibians – 3.3% of amphibian species in the trait dataset; 9 for birds – 0.077% of species; 7 for mammals – 0.13% of species; and 161 for reptiles – 1.5% of species). Species with 0% completeness were retained in the datasets when there was information for traits I did not select in the analyses, but no known value for the traits I did select. For instance, the body mass of the amphibian species *Rhinella centralis* was known, but other trait values (including body length) were missing, meaning that *Rhinella centralis* had 0% completeness for the set of traits I considered. *Figure reproduced from Etard et al. (2020)*.

2.3.2 Phylogenetic biases in trait completeness

As expected from the distribution of trait completeness in mammals and birds (Figure 2.3), within-family median trait completeness was high across most tips of the phylogenetic trees (Appendix 1, Figures S2.11 and S2.12; I present the avian and mammalian phylogenies in Appendix 1 because there was little variation in completeness across tips). For birds, λ was 0.71 (± 0.0053). For mammals, λ was 0.78 (± 0.0035). This indicated that, despite completeness generally being high across tips, the sampling was not evenly distributed across the phylogeny.

657 In herptiles, clusters of families with similar median trait completeness appeared (Figure 2.4). In am-
658 phibians, groups of families belonging to the order *Anura* (frogs) showed both the best and worst median
659 completeness (Figure 2.4a). The best-sampled families included the tailed frogs of the family *Ascaphidae*
660 (two species) and species of the family *Leiopelmatidae* (four species endemic to New Zealand). The family
661 *Ceratobatrachidae* (containing c. 90 species occurring in Southeast Asia and in some Pacific islands), the
662 family *Ranidae* (true frogs, 450 species considered here) and the family *Rhacophoridae* (shrub frogs, 382
663 species considered here) figured among the worst-sampled families. For amphibians, λ was $0.63 (\pm 0.0039)$.
664 In reptiles, most snakes were poorly sampled, whereas families in other suborders appeared to be sampled
665 better overall (Figure 2.4b). Within snakes, the pythons, boas, the three species of the family *Acrochordidae*
666 and the python-like species of the family *Loxocemidae* were better sampled than other snake families. In
667 reptiles, λ was $0.69 (\pm 0.0032)$. The sampling in herptiles was thus also uneven with regard to the phylogeny.

668 It is important to underline that Figure 2.4 shows within-family median completeness, masking the con-
669 siderable variation in species richness across families, hence masking potential important variation in com-
670 pleteness across species within families. For example, in the amphibian family *Allophrynididae* (three recog-
671 nized species), the within-family median completeness was 50%; but the dataset comprised two species of
672 completeness 14% and 86%, respectively. I present similar plots to those in Figure 2.4 showing the within-
673 family standard deviation in completeness in Appendix 1 (Figure S2.13). Within-family standard deviation
674 tended to increase with within-family species richness (Appendix 1, Figure S2.14).

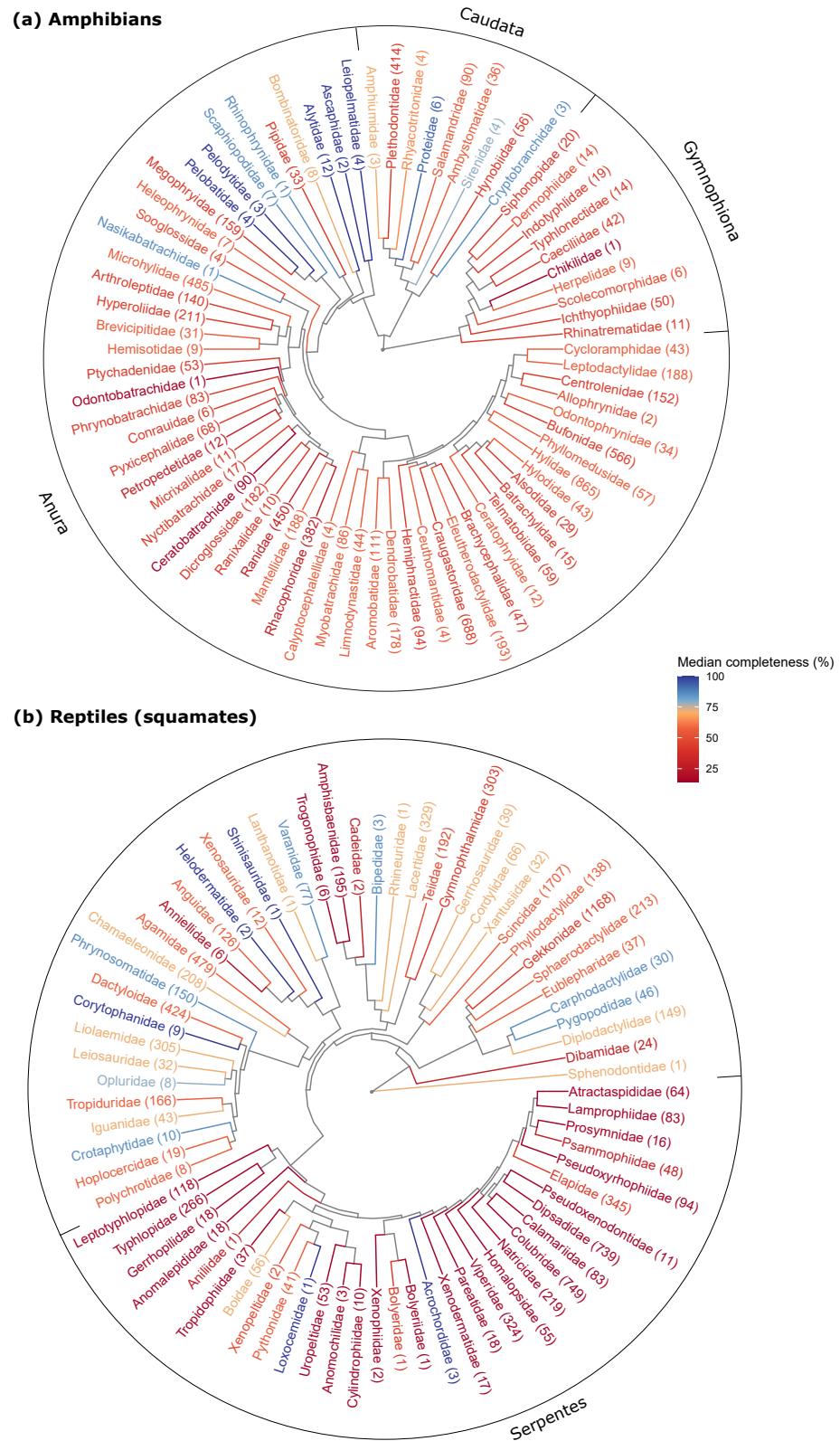


Figure 2.4: Within-family median trait completeness in herptiles. The number next to each family name represents the number of species included in the calculation of the median. *Figure reproduced from Etard et al. (2020).*

675 2.3.3 Spatial biases in trait completeness

676 Range size was significantly correlated with the number of sampled traits. Larger range sizes were associated
677 with a higher number of sampled traits (i.e., with higher completeness; Figure 2.5; Appendix 1, Table S2.1).
678 Similar results were obtained when using distribution maps not cut by elevational limits (Appendix 1, Table
679 S2.2; Figure S2.15). The rate of increase was steepest for reptiles, then for amphibians, then for birds and
680 mammals (slope estimates for birds and mammals were not significantly different from each other; Appendix
681 1, Table S2.1).

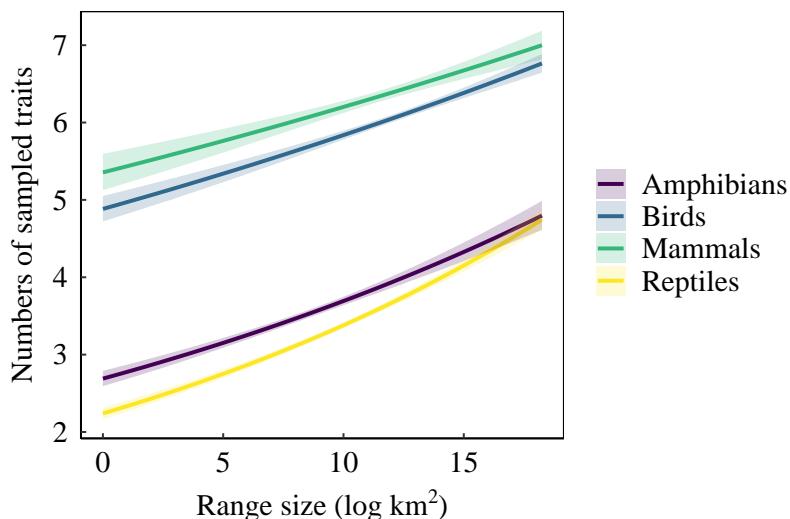
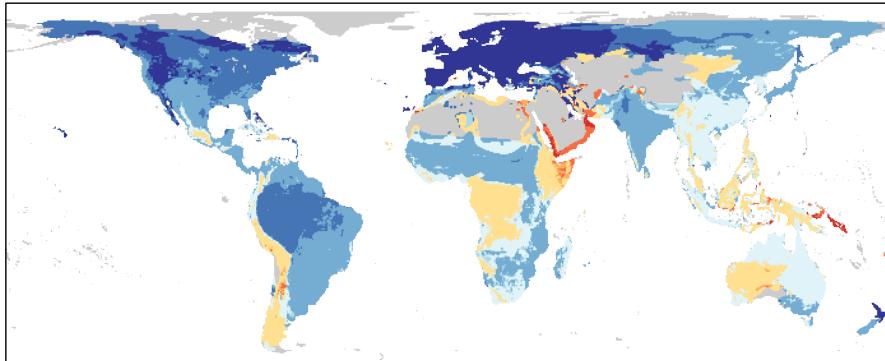


Figure 2.5: Relationship between number of sampled traits and geographical range size. The model was fitted using a Poisson error distribution. Class was added as a predictor interacting with range size. Rates of increase in number of sampled traits with range size were not significantly different for mammals and birds but differed for reptiles and amphibians, with the steepest rates of increase for reptiles. *Figure reproduced from Etard et al. (2020).*

682 There were marked spatial variations in median trait completeness in herptiles (Figure 2.6). North Amer-
683 ica and Europe were well sampled for both amphibians and reptiles. However, Southeast Asia and the Congo
684 basin were on average less well sampled. In other regions, contrasting patterns emerged between amphibians
685 and reptiles. For instance, median completeness was poorer for amphibians than for reptiles in Australia,
686 but opposite patterns were observed in South America. As in the phylogenetic analyses, assemblage-level
687 median completeness could mask potential important variation in completeness within species of a given
688 assemblage. Assemblage-level mean and standard deviation maps are shown in Appendix 1 (Figures S2.8
689 and S2.9). There was a trend for increasing standard deviation with increasing species richness, with a larger
690 spread in standard deviation at lower species richness (Appendix 1, Figure S2.10).

(a) Amphibians



Median completeness:

NA
0 – 10%
10 – 20%
20 – 30%
30 – 40%
40 – 50%
50 – 60%
60 – 70%
70 – 80%
80 – 90%
90 – 100%

(b) Reptiles

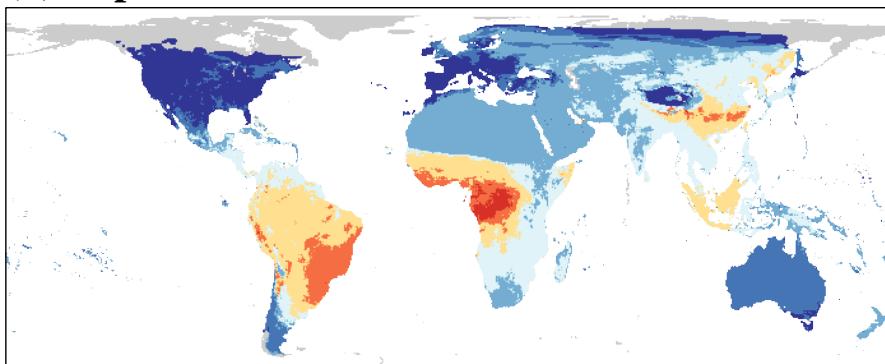


Figure 2.6: Spatial distribution of assemblage-level median trait completeness in herptiles. Similar maps for birds and mammals are shown in Appendix 1 (Figure S2.7). *Figure reproduced from Etard et al. (2020).*

691 Spatial models showed that species richness explained median trait completeness in herptiles in most
692 realms (Figure 2.7; Appendix 1, Tables S2.3 and S2.4); including spatial lags improved the models (am-
693 phibians: $\rho = 0.92$, p-value < 0.0001; reptiles: $\rho = 0.91$, p-value < 0.0001). For reptiles, completeness was
694 negatively correlated with species richness in the most species-rich realms (Afrotropics, Indo-Malayan and
695 Neotropics) and in the Palaearctic; the relationship was steepest in the Afrotropics and shallowest in the
696 Palaearctic. In the Australasian and Nearctic realms, completeness tended to increase with species richness.
697 For amphibians, negative relationships were observed in the Indo-Malay and Nearctic realms, whereas pos-
698 itive trends were observed in the Neotropics and the Palaearctic. The opposite trends between reptiles and
699 amphibians observed in the Australasian and Neotropical realms reflected patterns observed on the maps.
700 The Indo-Malayan was the only realm where median completeness tended to decrease with species richness
701 for both reptiles and amphibians.

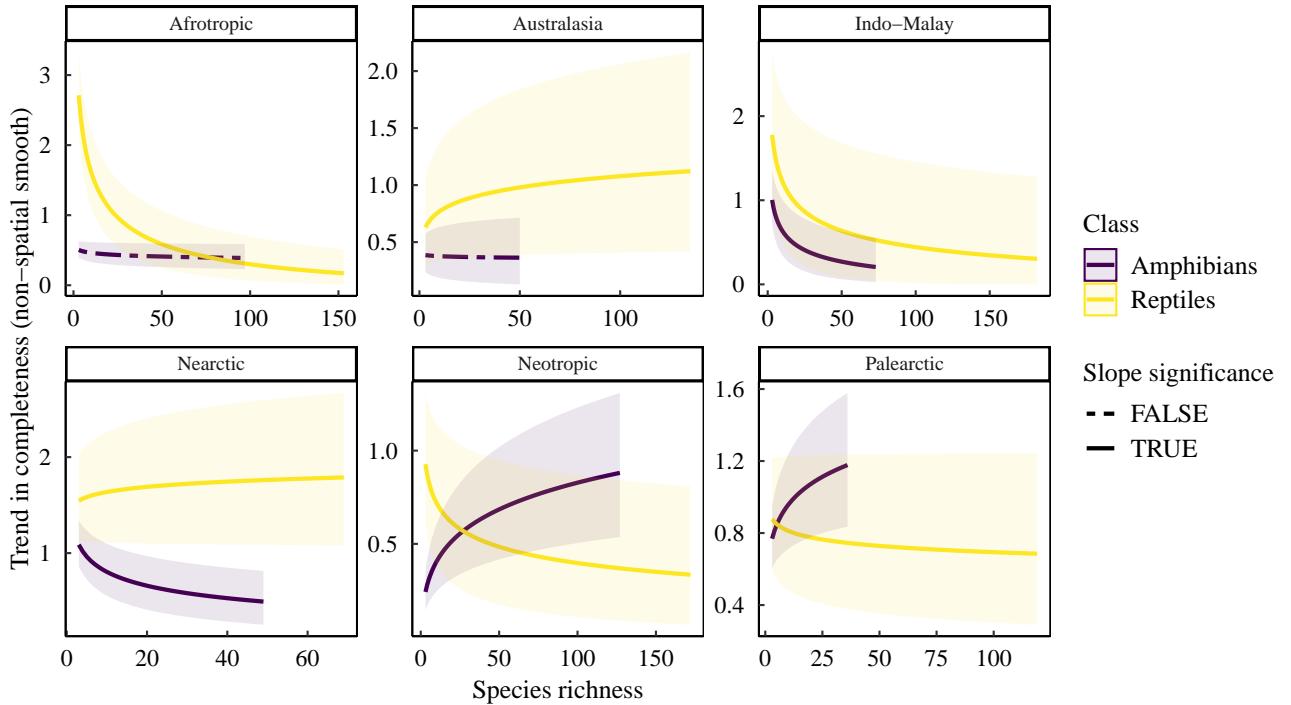


Figure 2.7: Spatial model trends for herptiles. The lines represent in-sample predictions (\pm standard error) for the trend components of the spatial models (trends after accounting for spatial autocorrelation). *Figure reproduced from Etard et al. (2020).*

702 2.4 Discussion

703 The results of this Chapter illustrate the taxonomic, spatial and phylogenetic dimensions of the knowledge
 704 gaps in trait data, termed the Raunkiær shortfall by Hortal et al. (2015). To the best of my knowledge,
 705 this work constitutes the first comparative assessment of global gaps for terrestrial vertebrate trait data,
 706 despite their use in numerous studies. I showed that the trait data present important taxonomic, spatial and
 707 phylogenetic biases, with contrasts in the availability of trait information between, on the one hand, herptiles
 708 and, on the other hand, birds and mammals.

709 Birds and mammals are globally well sampled for the set of traits I considered, even in the most species-
 710 rich assemblages. However, the availability of trait information for herptiles is lower overall and phyloge-
 711 netically and geographically biased. Several factors could interplay to shape these patterns. For instance,
 712 species that are more easily detectable (for example, wider ranging) and more charismatic are likely to be
 713 better sampled. Diverse socio-economic predictors could also contribute to geographical biases in trait data
 714 sampling; global biases in primary data collection are likely to be one of the most important contributors to
 715 the patterns I highlighted. Nevertheless, biases in the data could have been introduced at later stages, notably
 716 with the selection of sources and traits. The global compilation I obtained in this Chapter reflects, in part, the

717 interest and focus of the secondary data sources I used. It is possible that the addition of new sources from
718 regional journals or other authorities could diminish spatial biases in the data by increasing coverage for
719 certain areas. Nevertheless, I argue that by focusing on widely used traits, these results are likely to reflect
720 the “true” availability of the data in primary sources and that the shortfalls for other, less used traits would
721 be more pronounced.

722 I believe that the results presented here are robust to taxonomic uncertainty, although taxonomic match-
723 ing might potentially be improved further using class-specific sources, such as the Reptile Database or Am-
724 phibiaWeb, for identification of synonyms (but see Appendix 1, Figure S2.16). I have made two versions of
725 the data compilations available, one in which my own corrections were applied and one using the original
726 binomial names of the sources, meaning that users are free to use their own taxonomic backbones and iden-
727 tify synonyms within the compilations. I believe that taxonomic matching is a recurring issue when working
728 across thousands of species. Taxonomic synonymy artefactually inflates the numbers of identified species,
729 potentially lowering trait coverage (whereas clumping subspecies together can have the opposite effect).
730 Tackling this problem is difficult (Isaac et al., 2004; Jones et al., 2012), notably because there is no global
731 curated database recording the status of species names, and also because of the nature of taxonomy and the
732 debates around the species concept (May, 2011). Nevertheless, taxonomic uncertainty can have important
733 consequences. For instance, Cardoso et al. (2017) showed that inaccuracies and errors in species checklists
734 contributed to the overestimation of plant diversity in the Amazon (but see Freeman and Pennell (2021): the
735 relative underdescription of species in tropical areas compared to temperate areas –‘taxonomic debt’, also
736 referred to as ‘latitudinal taxonomic gradient’ by the authors– may lead to the underestimation of species
737 richness at low latitudes).

738 Biases in trait data have important implications for conservation planning. Past studies have shown that
739 narrow-ranging species, for which fewer trait data are available on average, have higher extinction risks
740 (Collen et al., 2016; Purvis et al., 2000; Ripple et al., 2017) and are more negatively impacted by anthro-
741 pogenic pressures than wider-ranging species (Newbold et al., 2018). Trait information is also less available
742 for herptiles in tropical regions such as the Congo basin, Southeast Asia and South America, which are some
743 of the most diverse areas of crucial importance for worldwide conservation (Barlow et al., 2018). Conse-
744 quently, trait information is on average less available where potentially more crucial to conservation plan-
745 ning. Indeed, trait information can be incorporated into vulnerability assessments and, as such, can help to
746 prioritize conservation efforts. Species traits have been found to mediate species responses to environmental
747 changes across diverse taxonomic groups, and thus can inform on the sensitivity of species to anthropogenic
748 pressures (Flynn et al., 2009; Newbold et al., 2013; Nowakowski et al., 2017). Traits are now commonly

749 used to estimate species vulnerability or extinction risks (Pacifici et al., 2015; Ramírez-Bautista et al., 2020).
750 As opposed to trend-based approaches, which rely on historical population trends (changes in abundance
751 or shifts in distributions) to predict species' vulnerability and extinction risks, trait-based approaches rely
752 on species' intrinsic sensitivity to particular threats. The appeal of trait-based approaches to extinction risk
753 estimation is that, by providing mechanistic insights, they diminish the amount of population information
754 needed. If the responses of species to a threat consistently relate to certain traits, it is possible to generalize
755 patterns across species for which population data are less available (Verberk et al., 2013). Integrating traits
756 into vulnerability assessments is hence of particular interest when field monitoring of species population
757 sizes or distributions is difficult to achieve, but biases in the data could mean that such information is lacking
758 for some of the most vulnerable species.

759 Traits that influence species responses to environmental changes have been termed 'response traits' (or
760 'response-mediating traits'; Luck et al. (2012)), as opposed to 'effect traits' that underpin ecosystem func-
761 tioning (Lavorel and Garnier, 2002). For instance, relative brain size and longevity have been characterized
762 as response traits in birds (Newbold et al., 2013; Sayol et al., 2020), whereas dietary characteristics (e.g.,
763 trophic levels or guilds) are both response and effect traits. Hortal et al. (2015) highlighted that, for plants,
764 both response and effect traits have been investigated, whereas for vertebrates the research has been more
765 focused on understanding species responses. This could be because the way vertebrate traits interact to shape
766 some ecosystem processes has not yet been characterized well.

767 Ecosystem processes sustained by animals might be harder to quantify and might be influenced by a com-
768 bination of traits. The traits compiled in this work are likely to have a role in diverse processes. Nevertheless,
769 there was one important omission, in that I did not compile species diet in this Chapter, potentially the most
770 straightforward trait to link with diverse processes, such as grazing, pollination, scavenging and seed dis-
771 persal. From a practical perspective, I chose traits that had been estimated at least for some of the species
772 in each class, and that were readily available. Diet was excluded because although estimates were available
773 for amphibians, birds and mammals, there was no readily available database for reptilian diet. Movement or
774 dispersal abilities were also excluded because information was not readily available for any class. Although
775 I expect that species diet and dispersal abilities would present similar sampling biases to the ones presented
776 in this work, the addition of such traits to the compilation would represent a valuable contribution and would
777 notably facilitate studies looking at the functional roles of reptiles.

778 For practical reasons, I did not consider intraspecific trait variation. Intraspecific variation has been
779 shown to have important effects on ecological systems, and a growing body of literature encourages trait-
780 based research to include intraspecific variability (Guralnick et al., 2016). There have been several calls to

781 produce open-access, global trait datasets (Weiss and Ray, 2019), including a representation of intraspecific
782 trait variation (Kissling et al., 2018). Notably, Schneider et al. (2019) designed a framework to store and
783 share inter- and intraspecific trait data, accompanied by an R package to standardize the data in a proposed
784 format. Such a proposition could constitute an important step towards the unification of individual datasets
785 into a single, comprehensive database for ecological trait data.

786 The current spatial and taxonomic gaps in trait data might limit our ability to scale studies up, whereas
787 biases in the data can affect the validity of extrapolations to groups or areas that are undersampled. More gen-
788 erally, biases and gaps in biodiversity data can have important implications for ecological studies. Data gaps
789 can hinder our ability to draw conclusions on observed macroecological patterns. For example, Chaudhary
790 et al. (2016) proposed that marine species richness follows a bimodal distribution, peaking at mid-latitudinal
791 locations, and argued that these patterns were not underpinned by knowledge gaps in species distributions.
792 However, Menegotto and Rangel (2018) attributed the tropical dip in marine species richness to a lack of
793 species distribution data, explained by lower sampling efforts in tropical areas ('Wallacean' shortfall; Hor-
794 tal et al. (2015)). Biases and gaps in trait data could also affect studies in closely related fields, such as
795 functional ecology – for instance, past studies have shown that functional diversity indices are sensitive to
796 missing data (Májeková et al., 2016; Pakeman, 2014) – or community assembly (Perronne et al., 2017).

797 Ecologists should, therefore, take particular care when designing trait-based studies, because both data
798 quality and data gaps are likely to influence the results and the generality of the conclusions. There exist
799 diverse methods to deal with missing trait values, should data missingness be problematic. Complete removal
800 of missing values ('case deletion') is commonly used but presents several issues, because it reduces sample
801 size and statistical power and introduces potential bias in data subsamples (Nakagawa and Freckleton, 2008).
802 For example, retaining complete cases only from the trait datasets would generate trait data disproportionately
803 representative of mammals and birds, which would be problematic for conducting cross-taxon analyses in
804 terrestrial vertebrates. As such, it is recommended that case deletion be applied only when data are missing
805 completely at random, which is rarely the case (Peugh and Enders, 2004).

806 Alternatives to case deletion consist of filling in the gaps. In recent years, the development of imputation
807 techniques has provided robust methods to handle missing data. Such imputation techniques have been used
808 to complete trait datasets in recent studies (Cooke et al., 2019a). Penone et al. (2014) used a simulation ap-
809 proach to evaluate the performance of four of these techniques, namely PhyloPars (Bruggeman et al., 2009),
810 random forest algorithms as implemented in R with missForest (Stekhoven, 2016; Stekhoven and Bühlmann,
811 2012), multivariate imputation by chained equations (MICE; van Buuren and Groothuis-Oudshoorn (2007))
812 and k-nearest neighbour (kNN; Troyanskaya et al. (2001)). Penone et al. (2014) introduced missing values

813 (10%–80%) in a complete trait dataset of carnivorans and measured imputation performance in different
814 scenarios. Given that phylogenetic non-randomness in missing trait values can impact imputation accuracy,
815 Penone et al. (2014) removed values in three different ways (completely at random; with a phylogenetic
816 bias; and with a body mass bias). Out of the four techniques, missForest and PhyloPars performed best when
817 species phylogenetic position was included as a predictor of missing trait values. Such imputations appeared
818 to be robust even when trait coverage was as low as 40%, which might be relevant for many reptilian and
819 amphibian traits. The performance was not significantly affected by phylogenetic non-randomness of the
820 data. Hence, missForest and PhyloPars appear to be well suited when traits are phylogenetically conserved,
821 because they allow species phylogenetic position to be included as a predictor of missing trait values. The
822 study by Penone et al. (2014) highlights that there are robust imputation techniques allowing to deal with
823 incomplete trait data where biases might otherwise be problematic. Nevertheless, it is important to highlight
824 that some imputation techniques, such as single or mean imputation, can be problematic because they do
825 not allow an estimation of uncertainty and suffer from a lack of accuracy (Nakagawa and Freckleton, 2008);
826 indeed, imputation techniques sometimes perform no better than case deletion. More work should be con-
827 ducted to assess imputation performance in various contexts (e.g., Johnson et al. (2021)), and the datasets
828 compiled in this Chapter might provide an opportunity for such studies.

829 Although robust imputation techniques can be useful for filling gaps in trait datasets, they are no substi-
830 tute for continued data collection efforts. The results of this Chapter show that data are particularly lacking
831 in herptiles, notably in the Afrotropics, the Neotropics and the Indo-Malayan realms. For these areas, incor-
832 porating regional databases into existing datasets could contribute to the reduction of global gaps. I believe
833 that both primary research and subsequent efforts to integrate new data and existing databases are required
834 if we are to collectively strive towards the unification of trait databases.

835 To conclude, this work constitutes, to my knowledge, the first assessment of the global gaps and biases
836 in terrestrial vertebrate trait information. I show that herptiles are undersampled compared with mammals
837 and birds, with important spatial and phylogenetic variability in the availability of trait information. Impu-
838 tation techniques are one possible solution to these problems. Nevertheless, I believe that primary research,
839 combined with efforts to complete existing datasets, is the only way to fill the current data gaps genuinely
840 and robustly. I hope that the compiled trait dataset and these findings can prove useful for guiding further
841 data collection efforts and for conducting macroecological analyses.

842 **3 | Intensive human land uses negatively af-**

843 **fect vertebrate functional diversity**

844 **Keywords**

845 Land use; land-use intensity; terrestrial vertebrates; functional diversity; traits.

846 **Abstract**

847 Land-use change is the leading driver of global biodiversity loss, thus characterising its impacts on the func-
848 tional structure of ecological communities is an urgent challenge. Using a database describing vertebrate
849 assemblages in different land uses, I assess how the type and intensity of land use affect the functional di-
850 versity of vertebrates globally. I find that human land uses alter local functional structure by driving declines
851 in functional diversity, with the strongest effects in the most disturbed land uses (intensely used urban sites,
852 cropland and pastures), and among amphibians and birds. Both tropical and temperate areas experience im-
853 portant functional losses, which are only partially offset by functional gains. Tropical assemblages are more
854 likely to show decreases in functional diversity that exceed those expected from species loss alone. These re-
855 sults indicate that land-use change non-randomly reshapes the functional structure of vertebrate assemblages,
856 raising concerns about the continuation of ecological processes sustained by vertebrates.

857 **3.1 Introduction**

858 Anthropogenic activities are profoundly transforming global biodiversity. Although multiple pressures act
859 in combination, land-use change currently poses the greatest threat to biodiversity (Maxwell et al., 2016;
860 Newbold et al., 2015). However, not all species respond similarly to land-use change. Traits have been found
861 to explain species' sensitivity to land-use change in diverse groups (Newbold et al., 2013; Nowakowski et

al., 2017; Quesnelle et al., 2014; Todd et al., 2017). Previous work has also shown that land-use change leads to non-random modification of assemblage trait composition (or functional diversity) (Chapman et al., 2018; Colin et al., 2018; Flynn et al., 2009; La Sorte et al., 2018; Newbold et al., 2013; Tinoco et al., 2018). Since it is widely acknowledged that biodiversity, and in particular trait diversity, may promote ecosystem functioning and stability, modification to the trait composition of assemblages could have far-reaching and adverse impacts on ecological processes (Hooper et al., 2012; Magioli et al., 2021; Oliver et al., 2015; Tilman and Downing, 1994).

Terrestrial vertebrates support many processes, ranging from pollination (Ratto et al., 2018), to seed dispersal to the regulation of lower trophic levels (Letnic et al., 2012; Mooney et al., 2010; Salo et al., 2010; Zhang et al., 2018a). However, we lack a global understanding of how the functional diversity of entire vertebrate assemblages responds to changes in land use. Most previous studies have been conducted at regional or local scales (Davison et al., 2021), but these may not be representative of global patterns. Indeed, recent global syntheses have highlighted how biodiversity responses can differ substantially between regions and across latitudes, with higher sensitivity reported for the tropics (Matuoka et al., 2020; Millard et al., 2021; Newbold et al., 2020b). Another key issue is the taxonomic coverage of past work. Few studies investigating effects of land use on functional diversity have considered several vertebrate classes together, and comparative studies remain rare. Thus, how land-use change affects the functional diversity of local vertebrate assemblages at global scales, and the potential geographical and taxonomic variation in the effects, still largely remains to be explored.

Here, I aim to assess how human land use and land-use intensity affect the functional diversity of vertebrate assemblages, across and within taxonomic classes. Building on recent work (Matuoka et al., 2020; Millard et al., 2021; Newbold et al., 2020b), I investigate differences in response between tropical and temperate regions. I use multiple response metrics to quantify functional diversity. First, functional richness measures the breadth and variety of trait combinations represented in an assemblage (Legras et al., 2018). Second, functional dispersion quantifies how similar species in a given assemblage are in terms of their traits (Laliberté and Legendre, 2010). These metrics can mask important alterations of assemblage composition if functional losses are compensated for by functional gains. To address this, I consider pairwise measures between assemblages, to explore levels of functional loss and functional gain across land uses (Figure 3.1).

To this end, I combine (1) the trait data across terrestrial vertebrates collected in Chapter 2 with (2) global records of species occurrence in eight land-use types of differing intensity of use (the PREDICTS database: Hudson et al. (2014, 2017); Figure 3.1; Appendix 2, Table S3.1). The PREDICTS database is currently the most comprehensive database of sampled species occurrence, and for most records also

894 abundance, across multiple land uses of different land-use intensity. Using the PREDICTS database allows
895 me to contrast biodiversity metrics among intact land uses (primary-vegetation sites, considered to be the
896 undisturbed reference condition), and all other human land-use types. Specifically, I test the following
897 hypotheses, both across and within taxonomic classes:

- 898 1. I expect decreases in functional diversity in human land uses compared to primary vegetation, caused
899 by contractions of occupied trait space. I expect such effects to be more pronounced where land is
900 used more intensively by humans. This hypothesis builds upon evidence that species with certain
901 traits are more sensitive to land-use disturbance (Newbold et al., 2013), meaning that disturbed land
902 uses will retain only disturbance-tolerant species, more functionally similar to one another. Given the
903 reported higher sensitivity of tropical assemblages to land-use disturbance, I predict that such effects
904 are stronger in the tropics.
- 905 2. I hypothesise that decreases in functional diversity in disturbed land uses exceed decreases expected
906 by chance, given local species loss. Thus, I expect disturbed land uses to promote functional under-
907 dispersion. Functional under-dispersion occurs when species within an assemblage are more similar,
908 in term of their traits, than expected by chance (Cadotte and Tucker, 2017; Wong et al., 2018) – or,
909 in other words, when functional dispersion is lower than expected given local species richness. I
910 predict that under-dispersion is more likely to occur in the highly disturbed sites, in both temperate
911 and tropical areas. This hypothesis is based on the idea that species are being removed non-randomly
912 from sensitive areas of the trait space, and increasingly so with higher disturbance level.
- 913 3. Finally, I expect decreases in functional diversity in human land uses to be driven by high functional
914 loss, whereby species are being removed from previously occupied areas of the trait space; I expect
915 no functional gain. This hypothesis is based on the idea that the functional trait space in undisturbed
916 land uses represents all of the possible regional trait combinations and that species with functional
917 attributes rendering them unable to persist in altered conditions will be filtered out (Cornwell et al.,
918 2006).

919 3.2 Methods

920 3.2.1 Vertebrate assemblages

921 I used vertebrate occurrence data from the PREDICTS database (Hudson et al., 2014, 2017), a collection of
922 studies that recorded species occurrence across multiple land uses and land-use intensities. In PREDICTS,

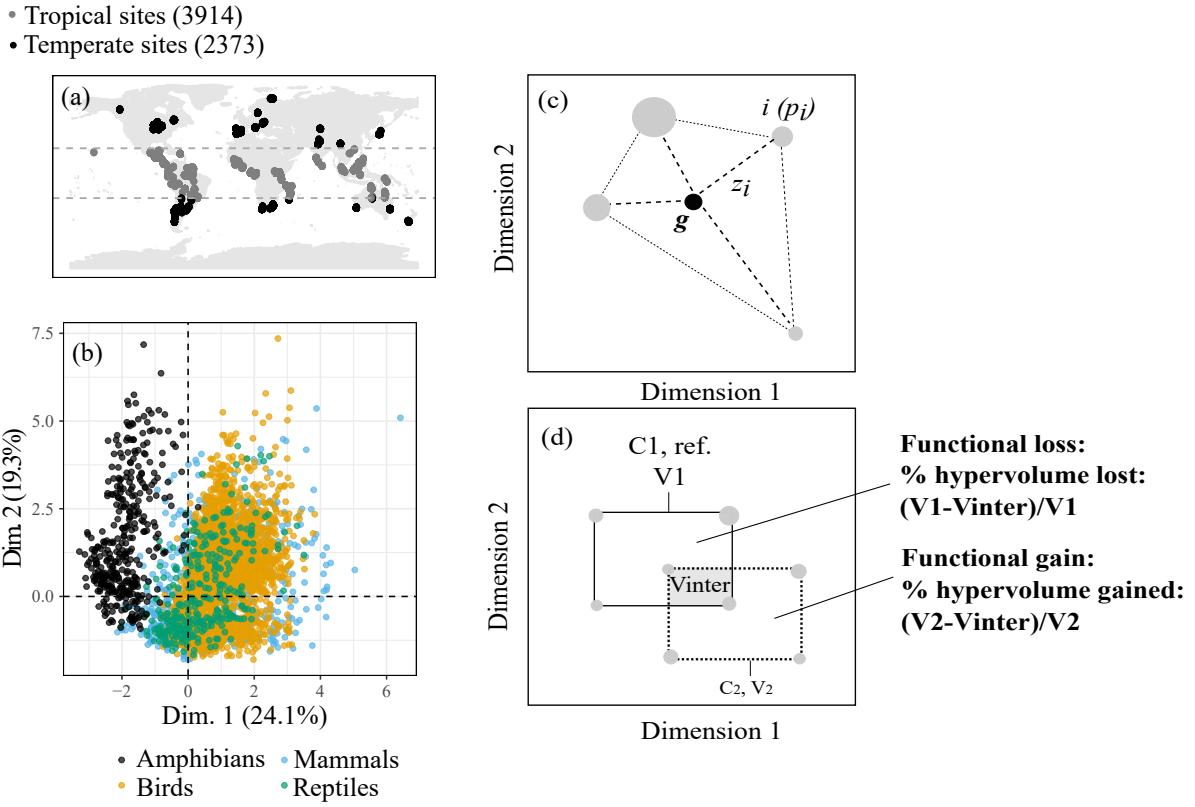


Figure 3.1: Overview of the study design and functional diversity metrics. I used occurrence data for vertebrate species from the PREDICTS database ((Hudson et al., 2014, 2017); 180 studies; 431,170 records; 4,339 species; 6,758 sampled sites). (a) shows the spatial distribution of sites I consider. I combine occurrence data with trait data compiled in Chapter 2 to calculate functional diversity metrics. (b) is a representation of the trait data in two dimensions, plotted across PREDICTS vertebrates. Traits that contributed most to dimension 1 were lifespan (29%) and litter/clutch size (22%), while traits that contributed most to dimension 2 were habitat breadth (47%) and use of artificial habitats (35%). (c) and (d) present the conceptual framework for the calculation of the functional diversity metrics: local measures (c) and pairwise metrics (d). (c) Given a trait space, functional richness is calculated as the hypervolume occupied by the minimum convex hull encompassing all species (Villéger et al., 2008). Functional dispersion is calculated as the mean distance of the species to the centroid, g (Laliberté and Legendre, 2010). (d) I compute functional loss as the proportion of hypervolume lost from the reference assemblage, and I define functional gain as the proportion of hypervolume of the disturbed assemblage that was gained (proportion of novel trait space in the disturbed assemblage). *Figure reproduced from Etard et al. (2022).*

each study contains several sites, which may be clustered into spatial blocks. Assemblage and land-use data are available at the site level: one site is characterised by a unique land use of given land-use intensity and provides occurrence data for a set of sampled taxa (and the same set of taxa is sought at all other sites within a study). Sites located between 23.5°N and 23.5°S of latitude were considered tropical, and otherwise temperate (Figure 3.1).

Land uses in PREDICTS were assigned to the following categories, based on the descriptions of the habitat given by the original collectors of the data: primary vegetation (considered to be the undisturbed reference); secondary vegetation; plantation forest; pasture; cropland; urban (considered human, or disturbed; Appendix 2, Table S3.1; Hudson et al., 2014, 2017). Secondary vegetation is further divided into three cat-

932 egories: mature, intermediate and young, depending on the stage of recovery of the vegetation. Land-use
933 intensity is reported as minimal, light or intense, according to criteria that depended on the land-use type in
934 question (e.g., crop diversity, degree of mechanisation and chemical inputs in cropland, or bushmeat harvest-
935 ing and selective logging in primary vegetation; Hudson et al., 2014). I excluded sites for which the land use
936 could not be characterised or for which the stage of recovery of secondary vegetation was unclear. As the
937 PREDICTS database is a collection of independent studies, the design of this study was not balanced: the
938 sample size varied across land uses (Appendix 2, Figures S3.1 & S3.2), and across taxonomic groups (3,103
939 species of birds; 531 mammals; 379 amphibians; 326 reptiles).

940 **3.2.2 Functional traits and diversity metrics**

941 Trait choice is a critical step when calculating functional diversity metrics, which are highly sensitive to
942 trait selection (Mouillot et al., 2021). However, trait selection trades off with data availability. Here, a
943 constraint was to use similar traits across the different classes. Thus, I used the seven traits compiled in
944 Chapter 2 across terrestrial vertebrates. Most of these traits were available for at least 50% of the species
945 in each class (except trophic level in amphibians and lifespan in herptiles; Appendix 2, Figure S3.4). In
946 addition, I chose traits that were ecologically relevant, broadening the biological definition of traits (i.e.,
947 a characteristic measurable at the level of an individual) to include measures of habitat breadth and habitat
948 specialisation (still theoretically measurable at the level of an individual). The final set constituted seven traits
949 that influence species' responses to environmental change: body mass, trophic level, lifespan, litter/clutch
950 size, diel activity, habitat breadth and use of artificial habitats. These traits related to life-history, habitat
951 specialisation and use of geographical space (e.g., habitat breadth significantly explains geographical range
952 size in all classes; Appendix 2, Figure S3.3). Here, I did not consider estimations of dispersal abilities or
953 home range size as these were available for a small fraction of the species (<3%, Paradis et al., 1998; Smith
954 and Green, 2005; Sutherland et al., 2000; Whitmee and Orme, 2013), neither did I include geographical
955 range size which is measured across many individuals, and hence cannot be considered a trait. As in Chapter
956 2, I did not consider intraspecific trait variation, thus assuming no effect of the environment on trait values.

957 Trait coverage was variable among classes and traits, with important gaps for reptiles and amphibians
958 (Appendix 2, Figures S3.4 & S3.5; Chapter 2). I imputed missing trait values using random forest algorithms
959 ('missForest' package: Stekhoven and Bühlmann (2012), Stekhoven (2016)), including traits, taxonomic
960 order and phylogenetic eigenvectors as predictors (Debastiani et al., 2021; Penone et al., 2014). To further
961 assess the sensitivity of the results to imputation (see next section), I imputed missing trait values eight times,
962 thereby obtaining eight sets of imputed traits. I randomly selected one imputed trait set for the calculation of

963 functional metrics. Imputations of missing trait values & imputation performance are detailed in Appendix
964 2, S3.2: ‘Trait data & imputation of missing trait values’ and S3.4: ‘Imputation performance’ (and Figures
965 S3.6-S3.8). Post-imputation, continuous traits were \log_{10} -transformed (except habitat breadth which was
966 square-rooted) and z-scored (standardised to unit variance and zero mean). In addition, I assessed whether
967 the results were robust to imputation error using a subset of the PREDICTS data considering only species
968 for which I had complete trait information (see next section).

969 Correlation among traits can be a safeguard against high sensitivity of functional metrics to trait omis-
970 sion, notably where omitted traits correlate strongly with traits that are already included in the calculation
971 (Mouillot et al., 2021). Nevertheless, high multicollinearity among traits has been reported as potentially
972 problematic for the calculation of functional diversity (Cadotte et al., 2011). Thus, I verified that the degree
973 of multicollinearity among traits was not problematically high (with a threshold of 5 for variance inflation
974 factors, Appendix 2, Table S3.3). Furthermore, I tested the sensitivity of the results to trait omission, by
975 investigating whether adding geographical range size in the calculation of functional metrics was likely to
976 affect the results.

977 3.2.3 Effects of land use and land-use intensity on FRic and FDis (Hypothesis 1)

978 For each assemblage, I measured functional richness using ‘FRic’ (Villéger et al., 2008), and functional
979 dispersion using ‘FDis’ (Laliberté and Legendre (2010); Figure 3.1), from the ‘FD’ package (Laliberté and
980 Legendre (2010); Laliberté et al. (2014)). I assessed the effects of land use, land-use intensity, and region
981 (temperate versus tropical) on FRic and FDis across and within taxonomic classes using linear mixed-effects
982 models ('lme4' package, Bates et al. (2015)). Land use and land-use intensity were not ranked in the models.
983 A random intercept of study identity accounted for variation in experimental design across studies, while a
984 random intercept representing spatial blocks of sampled sites, nested within study, accounted for spatial
985 structuring within studies. To improve normality and bound predictions between 0 and 1, I transformed FRic
986 and FDis using an arcsin-square-root transformation. The best-fitting model was sought using backwards
987 stepwise model selection, starting with the most complex model that included all two-way interactions among
988 the specified main effects. Model fits were compared using likelihood-ratio tests at each iteration of the
989 selection procedure.

990 Across vertebrates, the starting models included the effects of land use, land-use intensity and region
991 (temperate versus tropical). The best-fitting model for FRic was:

992 $\text{arcsin}(\sqrt{\text{FRic}}) \sim \text{Land use} + \text{Land-use intensity} + \text{Region} + \text{Land use : Land-use intensity} + \text{Land use : Region}$.

994 (Model 1a)

995 For FDis, the best-fitting model did not include interactions between land use and region, but the main
996 effect of region was retained:

997 $\text{arcsin}(\sqrt{\text{FDis}}) \sim \text{Land use} + \text{Land-use intensity} + \text{Region} + \text{Land use : Land-use intensity}$.

998 (Model 1b)

999 To investigate differences in responses across classes, I pooled some of the land uses together, because
1000 otherwise, sample sizes would have been too low. Mature, intermediate and young secondary vegetation were
1001 grouped together as ‘Secondary vegetation’, and cropland and pasture were grouped together as ‘Agricultural
1002 land uses’. The starting models included the effects of land use, land-use intensity, region and taxonomic
1003 class. For FRic, the best model was:

1004 $\text{arcsin}(\sqrt{\text{FRic}}) \sim \text{Land use} + \text{Land-use intensity} + \text{Region} + \text{Class} + \text{Land use : Land-use intensity} + \text{Land use : Class} + \text{Land-use intensity : Region} + \text{Class : Region}$.

1006 (Model 2a)

1007 For FDis, regional effects were dropped:

1008 $\text{arcsin}(\sqrt{\text{FDis}}) \sim \text{Land use} + \text{Land-use intensity} + \text{Class} + \text{Land use : Land-use intensity} + \text{Land use : Class} + \text{Land-use intensity : Class}$.

1010 (Model 2b)

1011 To assess whether the results were robust to imputation error, I used a subset of the PREDICTS data
1012 considering only species for which there were complete trait information (6,212 sites; 442 mammals; 1,975
1013 birds; 78 reptiles; 9 amphibians), and I fitted models again to this data subset. I did not have enough complete
1014 trait data among amphibians to be able to consider this class separately, so I first considered amphibians and
1015 reptiles together (herptiles), and reptiles only. In addition, I complemented this validation with a sensitivity
1016 analysis to variation in imputed values. I calculated FDis and FRic using each of the eight imputed trait
1017 datasets and fitted the previous models to each set. I then qualitatively evaluated the congruence of the
1018 estimates from the different models. Finally, because there tended to be more sites sampled in primary
1019 vegetation than in other land uses (Appendix 2, Figures S3.1 & S3.1), I ran additional sensitivity tests to
1020 assess whether the results were robust to resampling primary vegetation sites to a number equal to 50 (a
1021 sample size close to the median number of sites sampled in land uses other than primary vegetation in both
1022 regions (median = 37 for the temperate subset and 57 for the tropical subset, Appendix 2, Figure S3.1)).

1023 **3.2.4 Investigating functional under-dispersion (Hypothesis 2)**

1024 To assess whether effects of land use and land-use intensity on FDis differed from what would be expected
1025 by chance given changes in local species richness, I generated null expectations of FDis at each site. I
1026 randomised assemblage composition 500 times, drawing species from the corresponding study's species
1027 pool while maintaining local species richness. For each site, I thus obtained a null distribution for FDis.
1028 Then, I tested whether FDis differed from null expectations using Wilcoxon signed-rank tests. I created
1029 a binary variable which was assigned 1 if FDis was significantly lower than null expectations at a given
1030 site (significant under-dispersion), and 0 otherwise. I investigated how land use, land-use intensity, region
1031 and taxonomic class affected the probability of occurrence of under-dispersion using a generalised linear
1032 mixed-effects model with a binomial distribution of errors. The best-fitting model did not retain any effect
1033 of taxonomic class:

1034 $P_{\text{under-dispersion}} \sim \text{Land use} + \text{Land-use intensity} + \text{Region} + \text{Land use : Land-use intensity} + \text{Land use :}$
1035 Region.

1036 (Model 3)

1037 **3.2.5 Functional loss and functional gain (Hypothesis 3)**

1038 I calculated the proportion of trait space that was lost in disturbed land uses compared to reference land uses
1039 (functional loss) and the proportion of trait space that was gained in disturbed land uses (functional gain)
1040 (Figure 3.1c), across and within taxonomic classes. I selected studies where at least one site was sampled in
1041 primary vegetation. I then made within study pairwise comparisons between reference assemblages, sampled
1042 in primary vegetation, and disturbed assemblages. In addition, I considered all comparisons between pairs
1043 of primary-vegetation sites, to create reference pairs. I then investigated how land use, land-use intensity
1044 and region affected functional loss and gain across and within taxonomic classes using linear mixed-effects
1045 models, controlling for study identity in the random effects. Across vertebrates, the best-fitting model for
1046 functional loss was:

1047 $\text{arcsin}(\sqrt{\text{loss}}) \sim \text{Land use} + \text{Land-use intensity} + \text{Region} + \text{Land use : Land-use intensity} + \text{Land use :}$
1048 Region.

1049 (Model 4a)

1050 For functional gain, one interaction term (land use with region) was dropped:

1051 $\arcsin(\sqrt{\text{gain}}) \sim \text{Land use} + \text{Land-use intensity} + \text{Region} + \text{Land use : Land-use intensity}$.

1052 (Model 4b)

When considering the effects of taxonomic class, the best-fitting model for functional loss was:

1054 $\text{arcsin}(\sqrt{\text{loss}}) \sim \text{Land use} + \text{Land-use intensity} + \text{Class} + \text{Region} + \text{Land use :}$

1055 Land-use intensity + Land use : Class + Land use : Region + Land-use intensity : Class.

1056 (Model 5a)

1057 For functional gain (Model 5b), the fitted effects were the same as those of Model 2b. More details about
1058 the calculation of functional loss and gain can be found in Appendix S3.5.

1059 All data analyses were conducted using R version 3.5.1 (R Core Team, 2018). I made the code available
1060 on figshare (DOIs: <https://doi.org/10.6084/m9.figshare.14161883> and <https://doi.org/10.6084/m9.figshare.15163926>), as well as the main result datasets (<https://doi.org/10.6084/m9.figshare.15163971>).

1063 3.3 Results

3.3.1 Effects of land use on FRic and FDis

1065 Across all vertebrates, land use and land-use intensity significantly affected FRic and FDis (Figure 3.2).
1066 FRic tended to decrease with increasing disturbance level and higher intensity of land use. For FRic, relative
1067 effects differed between regions (Figure 3.2a). Although declines were overall more important for disturbed
1068 tropical assemblages, significant declines were observed for the temperate assemblages (e.g., a 37% average
1069 decline in intensely used urban areas; a 49% decline in pastoral areas of high land-use intensity). Never-
1070 theless, tropical assemblages typically showed more important reductions in FRic. For instance, declines
1071 averaged 63% for intensely used tropical cropland and 76% for urban areas. For FDis, relative effects were
1072 similar in both regions (Figure 3.2b). The most important average declines were observed for urban assem-
1073 blages of intense use (20% decline), and for lightly- and intensely used cropland (by 15% and 26%). Note
1074 that confidence intervals around the estimated average declines were large in some cases, highlighting some
1075 heterogeneity in the responses.

1076 Fitting the same models to the subset of species with complete trait data, I detected important declines in
1077 functional diversity in a number of land uses, showing that the conclusions are robust to trait imputation un-
1078 certainty (for example, FRic declined on average by 75% in intensely used temperate pastoral assemblages;
1079 by 48% for intensely used tropical cropland; and FDis declined by an average 37% in intensely used tropical

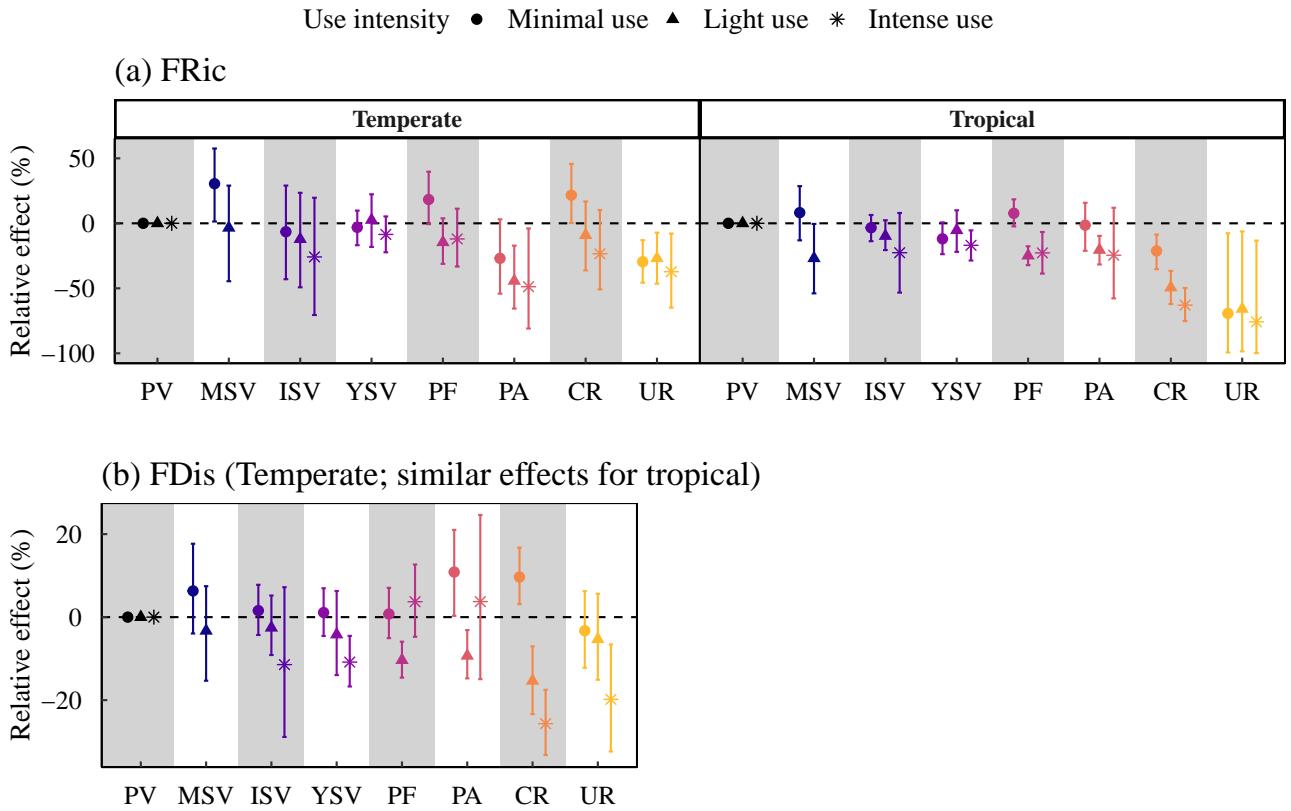


Figure 3.2: Effects of land use, land-use intensity and region on FRic (a) and on FDis (b) across all PREDICTS vertebrates. Effects were rescaled with reference to primary vegetation (PV) and are plotted as a % difference relative to PV within each land-use intensity. For FRic, the best-fitting model included interactions between land use and region, while these interactions were dropped for FDis, explaining the similar relative effects in both regions. Error bars represent 95% confidence intervals. MSV: mature secondary vegetation; ISV, intermediate secondary vegetation; YSV, young secondary vegetation; PF, plantation forest; PA, pasture; CR, cropland; UR, urban. Effects for intense use in MSV could not be estimated as there were not enough sampled sites. *Figure reproduced from Etard et al. (2022).*

1080 urban assemblages; Appendix 2, Figure S3.18). Furthermore, using the subset of species with complete trait
 1081 data, I found that the results were not sensitive to the inclusion of geographical range size as an additional
 1082 trait (Appendix 2, Figure S3.19). Finally, the results were not sensitive to variation across imputed trait val-
 1083 ues (Appendix 2, Figure S3.20) and were also robust to resampling in primary-vegetation sites (Appendix 2,
 1084 Figure S3.21).

1085 Responses of FRic and FDis to land use and land-use intensity differed among taxonomic classes (Figure
 1086 3.3). Within-class effects for FDis were similar between regions. The most notable decreases were observed
 1087 in lightly- and intensely used agricultural land uses in amphibians, birds and reptiles; and in intensely used
 1088 urban land uses for birds and mammals. For FRic, the effects in tropical and temperate regions were qual-
 1089 itatively similar in three out of four classes (birds, mammals and reptiles), although effect sizes tended to be
 1090 bigger for tropical assemblages. Birds and reptiles showed reductions in disturbed land uses in both tropical

and temperate regions, whereas I detected few significant effects for mammals. For birds, the most important average decline, of 50%, was observed in intensely used tropical urban land uses, while for reptiles I detected significant decreases in lightly- and intensely used agricultural sites (but I could not estimate effects for urban land uses due to the small sample size). Finally, the effects differed between tropical and temperate regions for amphibians, with no significant effects detected across temperate assemblages, but important reductions across tropical agricultural and urban assemblages.

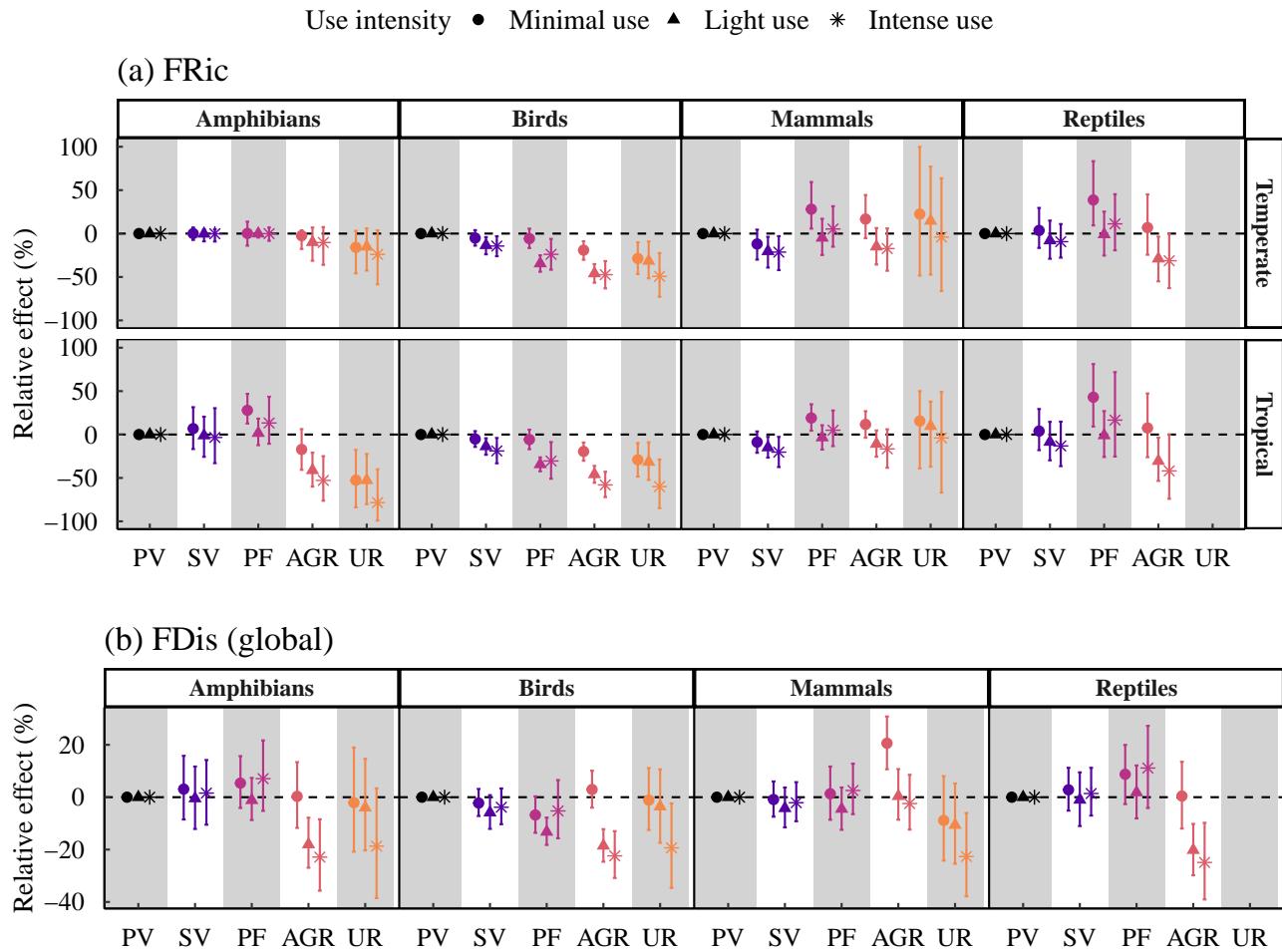


Figure 3.3: Effects of land use, land-use intensity, region and taxonomic class on FRic (a) and FDis (b). Effects were rescaled with reference to primary vegetation (PV) and are plotted as a % difference relative to PV within each land-use intensity. Error bars represent 95% confidence intervals. Effects for FRic were estimated from Model 2a, and from Model 2b for FDis. SV, secondary vegetation; PF, plantation forest; AGR, agricultural land uses (pasture and cropland); UR, urban. Effects for reptiles in urban land uses could not be estimated as there were not enough sampled sites. *Figure reproduced from Etard et al. (2022).*

Fitting similar models only for species with complete trait data showed that these patterns are unlikely to be affected by imputation uncertainty for birds; for mammals and reptiles, the main results could even be conservative (Appendix 2, Figures S3.22, S3.23). Indeed, although confidence intervals around the estimates

were large, I typically observed larger decreases in functional diversity when using the complete data subset, including an 86% decline in FRic for mammals in intensely used tropical agricultural areas. The results were also unaffected by variation across replicate sets of imputed trait values (Appendix 2, Figure S3.24).

3.3.2 Changes in the probability of occurrence of functional under-dispersion

Land use, land-use intensity and region significantly affected the probability of occurrence of functional under-dispersion across vertebrates. Functional under-dispersion was more likely to occur in tropical crop-land of all land-use intensities (Figure 3.4b), as well as in some of the lightly-used land uses (notably urban and plantation forest). Contrary to my expectations, and with the exception of tropical cropland, functional under-dispersion was not more likely to occur in intensely-used land uses. For minimally-used sites, changes in FDis were mostly consistent with changes expected given local species richness.

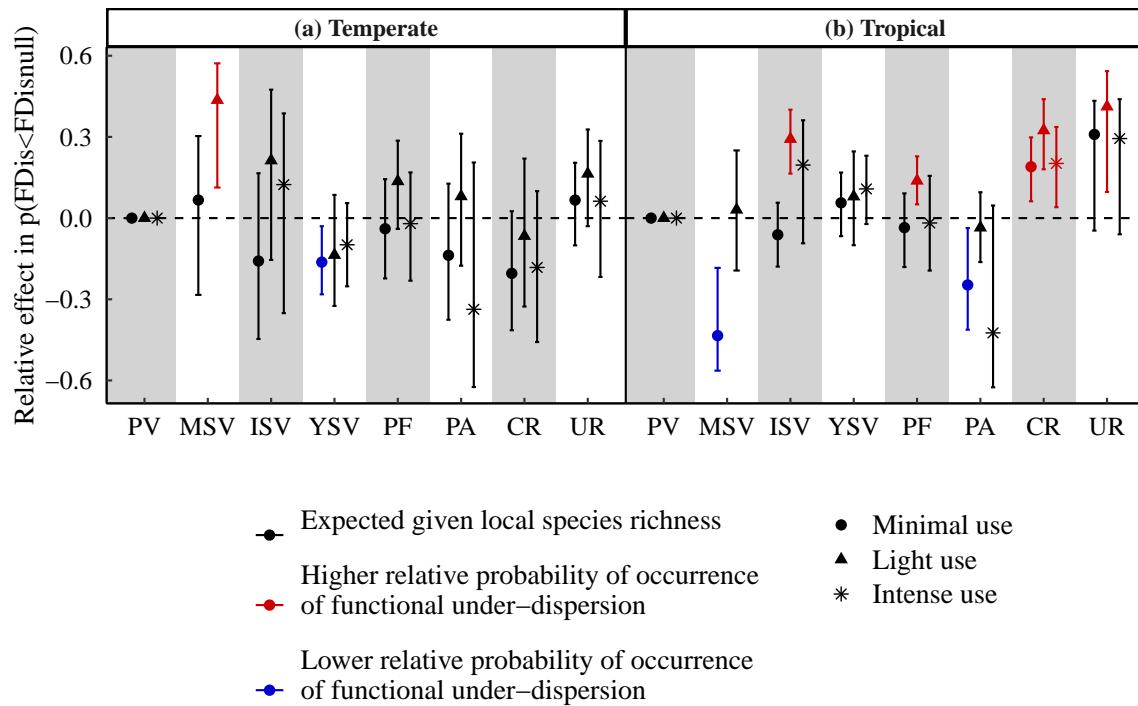


Figure 3.4: Effects of land use, land-use intensity and region on the probability of occurrence of functional under-dispersion. Error bars represent 95% confidence intervals. PV: primary vegetation; MSV, mature secondary vegetation; ISV, intermediate secondary vegetation; YSV, young secondary vegetation; PF, plantation forest; PA, pasture; CR, cropland; UR, urban. Effects are rescaled and represent the average difference in the probability of occurrence of functional under-dispersion between the reference (PV, probability of functional under-dispersion set at 0 within each land-use intensity) and the disturbed land uses. *Figure reproduced from Etard et al. (2022).*

3.3.3 Functional loss and gain

Across and within vertebrate classes, I detected high levels of functional loss, exceeding the natural turnover between primary-vegetation sites, both in temperate and tropical regions. Across vertebrates (Figure 3.5a), functional loss was notably high in temperate pastures (+27% above reference for minimal use; +73% for intense use), temperate urban sites (+27% for light use; +50% for intense use; effects for tropical urban sites could not be estimated), temperate and tropical cropland (+44% and +56% respectively for light use; effects for intense use could not be estimated). Important levels of functional loss were also observed in tropical plantation forest of light use intensity (+51%; effects for the intense use could not be estimated). High levels of functional loss were also observed within each class (Figure 3.6a) (although not all effects could be estimated because of limited sample sizes, Appendix 2, Table S3.5). The highest losses were observed in agricultural areas for amphibians and reptiles, with important losses also observed in temperate urban areas for both birds and amphibians (+35% for minimal use; effects for tropical urban areas could not be estimated).

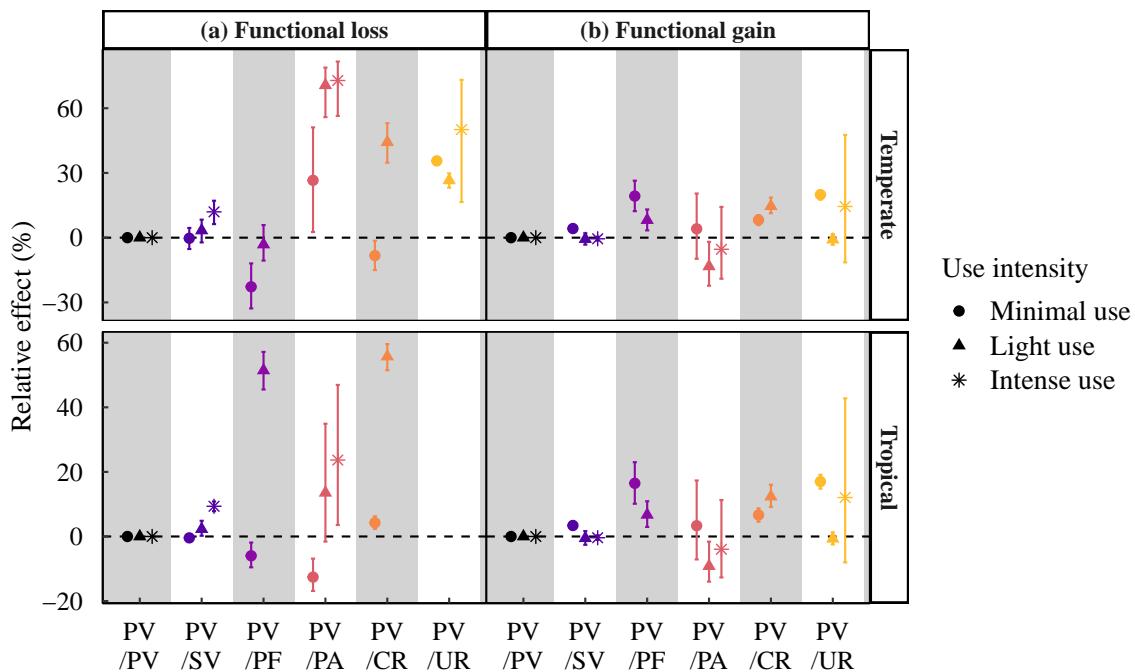


Figure 3.5: Effects of land use and land-use intensity on (a) functional loss and (b) functional gain across pairs of sites in temperate areas and tropical areas. PV: primary vegetation; SV, secondary vegetation; PF, plantation forest; PA, pasture; CR, cropland; UR, urban. Error bars represent 95% confidence intervals. Effects are rescaled and represent the average difference in functional loss and gain between the reference pairs (PV/PV, with loss and gain set at 0% within each land-use intensity), and pairs of primary vegetation with disturbed land uses. Negative effects mean that, on average, levels of functional loss (or gain) were lower than functional loss (or gain) observed between pairs of primary-vegetation sites, and not that there were absolute ‘negative losses’ or ‘negative gains’. *Figure reproduced from Etard et al. (2022).*

1123 Across vertebrates, average functional gain (average proportion of novel trait space in the disturbed
 1124 assemblage) was moderate and on average did not exceed 20% in any disturbed land uses (Figure 3.5b).
 1125 Patterns of functional gain were similar in both regions. The highest functional gains were observed for
 1126 minimally-used urban sites and plantation forest (range: +16% to +20%). On the other hand, important
 1127 levels of functional gain were observed in some classes (Figure 3.6b), with the highest functional gain for
 1128 mammals (+80% in intensely used urban sites).

1129 Diagnostic plots (qq-plots and residual distributions) for the models are shown in Appendix 2, Figures
 1130 S3.9–S3.17. Overall, the model residuals were appropriately distributed (but with some leptokurtic residual
 1131 distributions, to which mixed-effect models are generally robust (Schielzeth et al., 2020)).

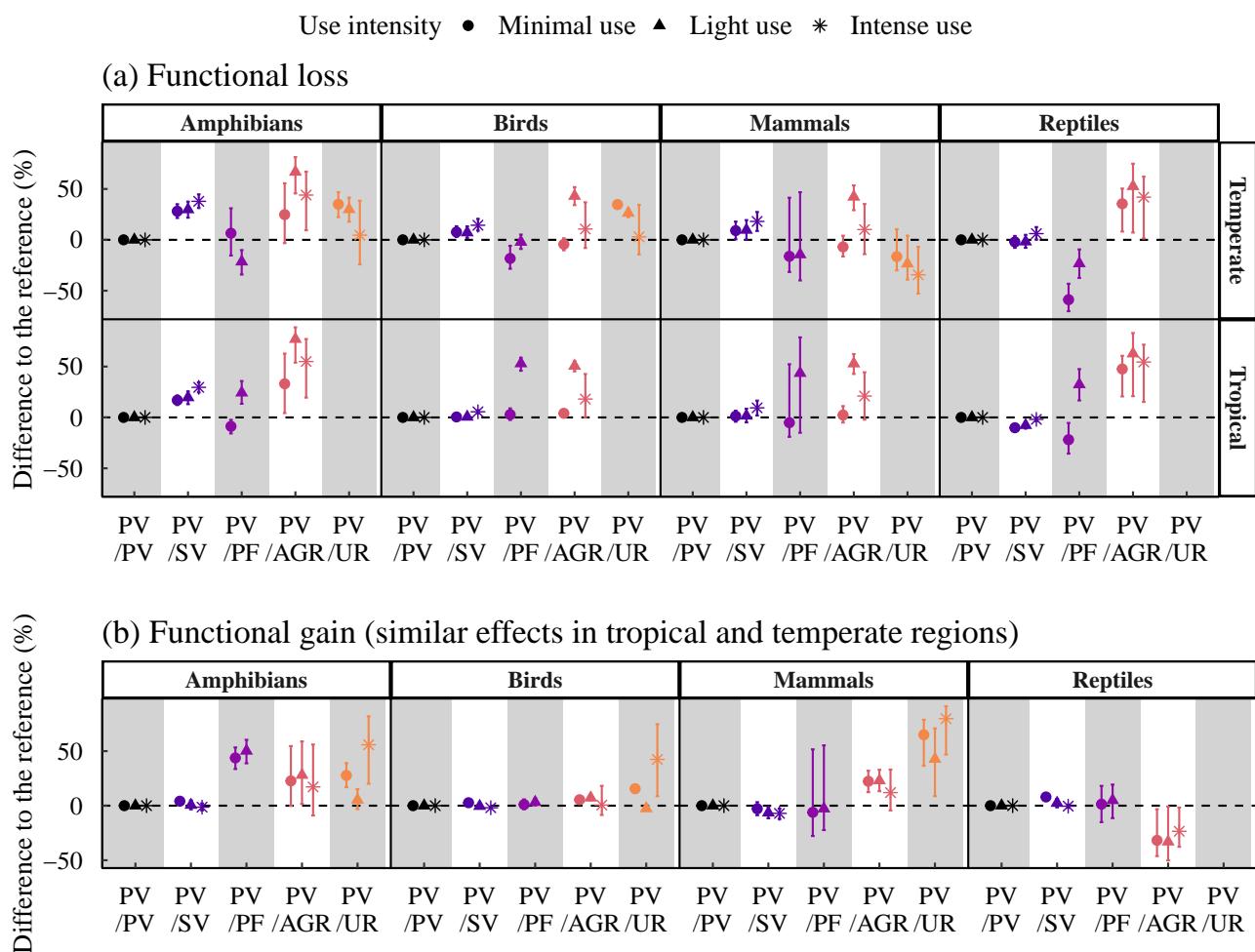


Figure 3.6: Effects of land use, land-use intensity, region and taxonomic class on functional loss (a) and functional gain (b) across pairs of sites. PV: primary vegetation; SV, secondary vegetation; PF, plantation forest; AGR, agricultural land uses (pasture and cropland); UR, urban. Error bars represent 95% confidence intervals. Effects are rescaled and represent the average difference in functional loss and gain between the reference pairs (PV/PV, with loss and gain set at 0% within each land-use intensity), and pairs of primary vegetation with disturbed land uses. Negative effects mean that, on average, levels of functional loss (or gain) were lower than functional loss (or gain) observed between pairs of primary-vegetation sites, and not that there were absolute ‘negative losses’ or ‘negative gains’. Figure reproduced from Etard et al. (2022).

1132 **3.4 Discussion**

1133 Here, I showed that the functional diversity of vertebrate assemblages is negatively impacted in human land
1134 uses, particularly in the most intensely used land types. The results of this Chapter extend previous studies
1135 that have been more taxonomically or geographically restricted (Flynn et al., 2009; Matuoka et al., 2020).
1136 Matuoka et al. (2020) found that the functional diversity of tropical bird assemblages was negatively affected
1137 by human disturbance, a pattern that did not appear in temperate assemblages. Yet, I found that functional
1138 diversity was negatively affected in both tropical and temperate areas, with important functional losses in all
1139 four vertebrate classes.

1140 Using multiple metrics allowed me to explore different facets of functional diversity. For instance, func-
1141 tional gain could locally offset functional loss in some disturbed land uses. This could indicate that despite
1142 no apparent negative effect on FRic, some disturbed land uses (e.g. lightly-used temperate cropland) could
1143 experience important functional loss, and highlights the importance of using a variety of indicators. This
1144 mechanism could be at play in mammalian assemblages, for which important levels of functional gain were
1145 observed in agricultural and urban sites. Further, functional gain in disturbed land uses could indicate that
1146 disturbances facilitate the introduction of functionally novel species, falling into previously unoccupied parts
1147 of the trait space. This may be because non-native species are more likely to become established in disturbed
1148 assemblages. Previous work has shown that land-use disturbance facilitates biological invasions in island
1149 ecosystems (Jesse et al., 2018; Sánchez-Ortiz et al., 2019), but to my knowledge, this has not been tested
1150 specifically across continental areas for invasive vertebrates (but see Pyšek et al. (2010)). It is also pos-
1151 sible that disturbed areas harbour synanthropic species that do not occur in primary vegetation, leading to
1152 substantial functional gain.

1153 Overall, the negative effects of land use on functional richness tended to be more pronounced in the
1154 tropics. This is congruent with past studies that have found tropical biodiversity to be disproportionately
1155 sensitive to human pressures (Martins and Pereira, 2017; Newbold et al., 2020b). There are a number of
1156 potential explanations for this. First, it could be that a long history of intense land-use disturbance at large
1157 scales in many temperate regions (e.g. Western Europe; Stephens et al. (2019)) means that biodiversity is
1158 now less sensitive to new disturbances, because the most sensitive species have been filtered out (Balmford,
1159 1996; Krauss et al., 2010; Le Provost et al., 2020; Munteanu et al., 2020). Species unable to cope with such
1160 disturbances may have gone extinct in the past, while the remaining species would be more disturbance-
1161 tolerant (Betts et al., 2019). Tropical regions, historically less disturbed at large scales, would then contain
1162 a higher proportion of disturbance-sensitive species than temperate regions. Consequently, the functional

richness in undisturbed tropical sites could be less resilient to new disturbances. This also highlights that time since land-use conversion could have important impacts on local functional diversity. Although I did not consider the effects of time since land-use conversion in this work (notably because PREDICTS contained data on time since land-use conversion only for about 22% of the sites), I expect that time since land-use conversion may affect assemblage composition, and thus, functional diversity, with potentially land-use-specific relationships between time since conversion and functional diversity (e.g., a positive relationship for recovering secondary vegetation or a negative relationship for urban areas; but I did not detect such effects when using the data subset for which there were information on time since land-use conversion [see Appendix 2, S3.8: ‘Model robustness – time since land-use conversion’]).

Second, it could be that tropical species are intrinsically more sensitive to disturbances than temperate species because of their evolutionary history. Natural climatic variability experienced by species as well as species history of exposure to disturbances have been proposed to influence sensitivity to disturbance. For instance, tropical species are, on average, nearer to their climatic limits than temperate species (Deutsch et al., 2008; Sunday et al., 2014). Tropical species could therefore experience more deleterious effects from interacting drivers of change, with land-use change bringing about novel climatic conditions pushing them beyond their tolerance limits (Frishkoff et al., 2016; Williams and Newbold, 2020).

In addition to filtering out sensitive species, land-use change is also expected to modify interactions among species, thereby influencing species persistence (Tylianakis et al., 2008; Valiente-Banuet et al., 2015). Although I detected a signal of functional under-dispersion (particularly in tropical cropland), which indicates that assemblages may be locally structured by environment filtering (Bregman et al., 2015), it is likely that several assembly rules underpin assemblage composition (Fournier et al., 2016). For instance, land-use changes could enhance competition among species, promoting over-dispersion by removing species that share similar resources. Such opposite signatures of environmental filtering and enhanced competition on functional dispersion could explain why I did not detect stronger effects of land use on functional under-dispersion occurrence.

Studies looking at impacts of global land use on functional diversity computed with species from all four terrestrial vertebrate classes remain rare. Lack of availability of standardised trait data across terrestrial vertebrates may have hindered such studies from being conducted in the past. To overcome this problem, I based the analyses on a large-scale collation of trait data (Chapter 2; Etard et al. (2020)), and I imputed missing trait values to obtain complete trait datasets in each class. I used random forest algorithms, currently thought to be one of the most robust technique for missing value imputations in trait datasets (Debastiani et al., 2021; Johnson et al., 2021; Penone et al., 2014). Replicating the analyses on complete trait data

1195 subsets showed that imputation uncertainty did not affect the main conclusions of this work and that the
1196 negative effects of human land uses were in some cases even stronger when using the complete data subsets.
1197 Furthermore, the results were highly consistent across imputed datasets and so insensitive to variation across
1198 imputed values. Although missing value imputation can offer a robust filling of missing entries, this study
1199 highlights the existing taxonomic biases both in trait data availability and in PREDICTS studies, and thus
1200 stresses the need to pursue data compilation efforts, particularly for the least-sampled classes (reptiles and
1201 amphibians).

1202 Another implication of trait data availability for vertebrates is that the choice of traits was constrained.
1203 Mouillot et al. (2021) showed that functional diversity metrics are sensitive to trait omission and that the
1204 sensitivity to trait omission decreases with increasing levels of correlation among traits. Here, I chose seven
1205 traits that were available across all classes at least for a subset of the species and that have been implicated in
1206 shaping species responses to environmental change. A notable omission was any metric of dispersal ability,
1207 which is likely to influence species' ability to respond to land-use change but is difficult to obtain for most
1208 species. In fact, past studies have shown that dispersal abilities can be predicted from ecological correlates,
1209 such as body mass, diet or geographical range size (Schloss et al., 2012; Sutherland et al., 2000). Since the
1210 results were robust to the omission of geographical range size, I am confident that the omission of dispersal
1211 abilities also does not affect the conclusions of this work.

1212 Functional diversity metrics are often used as a proxy for ecosystem functioning because of the concep-
1213 tual and mechanistic link between functional 'effect' traits and ecosystem processes (Lavorel and Garnier,
1214 2002; Violle et al., 2007). In many studies focused on vertebrates, however, functional diversity metrics do
1215 not correlate with a given ecosystem function (Hatfield et al., 2018). Here, I did not explicitly target given
1216 ecosystem functions, but I argue that evidence of functional loss of vertebrate assemblages indicates that
1217 processes sustained by vertebrates are put at risk by land-use change. My results further show that some
1218 disturbed land uses are more likely to experience functional under-dispersion, particularly tropical cropland
1219 and tropical urban areas, which again indicates a potential imperilment of ecological processes. Indeed, in
1220 such cases, decreases in functional dispersion exceed changes expected from the chance removal of species;
1221 such non-random modifications indicate that certain areas of the functional trait space are more sensitive to
1222 land-use disturbance. Future work could investigate the impacts of land-use change on particular ecosystem
1223 functions. The integration of trophic information (beyond the trophic levels I used here) to the species-trait
1224 dataset could be an interesting step in that direction, as dietary traits relate to resource use and are, as such,
1225 probably the most straightforward traits to link with ecosystem functions. Furthermore, my results suggest
1226 that the functional loss experienced within a class is unlikely to be compensated for by the persistence of

1227 functionally similar species in other classes. Indeed, I detected negative effects of human land use on func-
1228 tional richness in at least three out of four vertebrate classes (amphibians, birds, and reptiles), in accordance
1229 with past studies focusing on each of these groups (Gallmetzer and Schulze, 2015; Marcacci et al., 2021;
1230 Riemann et al., 2017; Sol et al., 2020). Although overall mammalian functional richness was less affected,
1231 high levels of functional gain suggest that the functional composition of mammalian assemblages is heavily
1232 modified in disturbed land uses.

1233 To conclude, the results of this Chapter highlight the negative impacts of human land uses on multiple
1234 dimensions of functional diversity, within and across terrestrial vertebrate classes, at a global scale. In
1235 many disturbed sites, decreases in functional diversity exceed changes expected from species loss alone,
1236 showing that human activities non-randomly reshape ecological assemblages. By intensifying functional
1237 loss and promoting functional under-dispersion, land-use change could have deleterious effects on ecosystem
1238 functioning, highlighting the necessity of putting into place effective conservation measures in the face of
1239 anthropogenic change.

1240 **4 | Narrow-ranging natural habitat specialists most sensitive to both land-use and**

1241 **climate change in terrestrial vertebrates**

1242

1243 **Keywords**

1244 Land use; land-use intensity; climate change; sensitivity; CENFA; dietary traits ; life-history traits; speciali-
1245 sation; geographical range area; terrestrial vertebrates.

1246 **Abstract**

1247 Land-use and climate change are two of the most important pressures on terrestrial biodiversity, however
1248 the factors that explain interspecific variation in responses to these pressures remain unclear. Although it is
1249 well established that extinction risk and some species' responses to human pressures relate to species traits,
1250 we lack large-scale comparative assessments across multiple clades linking traits to multiple human pres-
1251 sures. Here, I investigated whether a set of ecological characteristics that are commonly measured across
1252 terrestrial vertebrates (that is, ecological traits and geographical range area) are associated with (1) species'
1253 responses to different land-use types and (2) species' sensitivity to climate change. My aim was to test
1254 whether generalisable patterns in species' responses to these pressures arise with regards to species' ecolog-
1255 ical characteristics, which helps assess the global signature of human pressures on vertebrate biodiversity
1256 and is also of interest for the prioritisation of conservation efforts. Among the set of characteristics I consid-
1257 ered, I found that only three were consistently associated with both land-use responses and climate-change
1258 sensitivity across terrestrial vertebrate classes: geographical range area, habitat breadth and specialisation
1259 on natural habitats. The association of other traits with species' land-use responses and with climate-change

1260 sensitivity often depended on class and land-use type. My work highlights that narrow-ranged species with
1261 narrow habitat breadth and natural habitat specialism are typically more sensitive to human pressures. Fur-
1262 ther, I found that invertebrate eaters and fruit/nectar eaters tended to be negatively affected in disturbed land
1263 uses in all classes, and that invertebrate- and plant/seed- eating birds had higher climate-change sensitiv-
1264 ity, raising concerns about the continuation of ecological processes sustained by these species under global
1265 changes. My work stresses the need for putting into place conservation and mitigation measures to protect
1266 biodiversity and related services from human impacts.

1267 **4.1 Introduction**

1268 Land-use change is currently the most important driver of global biodiversity loss (Newbold et al., 2015)
1269 and is likely to continue to cause species loss in the coming decades (Li et al., 2022; Powers and Jetz, 2019;
1270 Stehfest et al., 2019). However, biodiversity faces multiple pressures acting in combination (Maxwell et al.,
1271 2016). In particular, the impacts of climate change on biodiversity are projected to equate or even surpass
1272 those of land-use change in their magnitude by 2070 (Newbold, 2018). Thus, it has become more vital than
1273 ever to put into place mitigation and conservation measures to protect biodiversity from human pressures.

1274 It is now well established that species differ in their ability to cope with environmental changes (Ferreira
1275 et al., 2022; Matich and Schalk, 2019; Newbold et al., 2013). As such, global average declines in biodiver-
1276 sity indices mask substantial interspecific variation in responses to disturbances (Leung et al., 2020). Such
1277 interspecific variation has important consequences for the prioritisation of conservation efforts and the defi-
1278 nition of protected areas (Morelli et al., 2021). Mitigating the impacts of land-use and climate-change on the
1279 world's biota requires to understand which species are put at most risk by these pressures, in other words to
1280 understand the factors that are associated with species' sensitivity to land-use and climate change.

1281 By capturing key aspects of species' morphology, life-history, ecological strategies or demography, traits
1282 can inform on species' use of resources and space, as well as on some community and population-level
1283 processes (Capdevila et al., 2022b). As such, traits can help understand what drives species' responses
1284 to environmental change. Thus, to explain interspecific differences in responses to human disturbance, a
1285 number of studies have investigated whether species traits influence species' responses to human pressures,
1286 in particular to land-use change (Newbold et al., 2013; Nowakowski et al., 2017; Quesnelle et al., 2014;
1287 Tinoco et al., 2018) and climate change (Angert et al., 2011; Di Marco et al., 2021; Estrada et al., 2018;
1288 McCain and King, 2014; Pacifici et al., 2017; Pearson et al., 2014; Schloss et al., 2012).

1289 From these past studies, several traits have been identified as important correlates of species' responses

to land-use and climate change within vertebrate taxa (for example, body mass and generation length were found to influence bird responses to land-use change in Newbold et al. (2013); and body mass and activity time were found to be associated with mammal responses to climate change in McCain and King (2014)). However, past work has mostly been conducted at local to regional scales (Davison et al., 2021; Hevia et al., 2017), such that it remains unclear whether the effects of traits on species' responses to environmental change can be generalised across vertebrate taxa and regions. Yet, at least two meta-analyses have investigated whether traits explained responses to human pressures across diverse taxa, one focused on climate-change responses (MacLean and Beissinger, 2017), and one on species extinction risk (Chichorro et al., 2019). MacLean and Beissinger (2017) found that habitat breadth and historic range limit were consistently associated with interspecific variation in range shifts under contemporary climate change across a range of taxa (including plants, birds and butterflies), but they did not detect any effect of life-history traits, such as body size or fecundity. Similarly, Chichorro et al. (2019) highlighted the effects of geographical range area and habitat breadth on species extinction risks in different taxa (including terrestrial vertebrates), with other traits having inconsistent effects. However, as underlined by Chichorro et al. (2019), the studies included in the meta-analysis often considered extinction risk without an explicit consideration of the pressures to which the species were exposed. Yet, a given trait could be associated with opposite responses depending on the pressure under consideration (González-Suárez et al., 2013).

Further, previous studies have often been restricted in their taxonomic coverage, with very few studies considering several vertebrate classes together, so that comparative investigations among vertebrate classes remain rare. In addition, there has not yet been a global assessment of the association between vertebrate traits and both land-use responses and climate-change sensitivity. Here, I test whether general patterns in species' land-use responses and climate-change sensitivity arise with regards to species traits. I include species geographical range area in the analysis, as it is one aspect of rarity that has been shown to influence species' responses to land use and to climate change (Newbold, 2018; Thuiller et al., 2005). Since geographical range area does not meet the strict definition of a trait, I henceforth refer to all traits and range area as 'ecological characteristics'. Thus, I examine associations with a set of ecological characteristics that are commonly measured across terrestrial vertebrates, at global scales (Figure 4.1a). Considering ecological characteristics that are available at least for a subset of the species in each class allows for a cross-taxon comparative assessment. Further, it also allows me to ask whether such commonly measured ecological characteristics show consistent associations with species' land-use responses and climate-change sensitivity. I ask two questions: (1) are any ecological characteristics associated with interspecific variation in responses to land use and with climate-change sensitivity? (2) If so, are these ecological characteristics similar across

1322 classes; are they similar between land-use responses and climate-change sensitivity; and are associations in
1323 the same direction, such that I can identify a set of characteristics that are associated with a high sensitivity
1324 of species to human pressures? Conversely, are such associations both taxon- and pressure- dependent?

1325 Given the different nature of the threats I consider, I use two independent approaches, one for land-use
1326 change and one for climate-change sensitivity. Thus, I do not consider interactive effects between these
1327 pressures. To infer species' responses to land-use change, I use a space-for-time substitution approach,
1328 modelling occurrence probability across different land-use types (Figure 1b). I estimate species' expected
1329 sensitivity to future climate change from properties of species' climatic niches (Figure 1c); species niche
1330 properties have been shown to be strong indicators of species' climate-change sensitivity (Thuiller et al.
1331 2005), and are also straightforward to use at large scales given the availability of species distribution data,
1332 from which climatic niche spaces can be constructed. I then bring these two approaches together to look for
1333 any emerging pattern in species' responses to land use or in their climate-change sensitivity, with regards to
1334 species' ecological characteristics.

1335 Among the characteristics I consider (Figure 4.1a), some may directly influence species survival by me-
1336 diating resource acquisition and use. These characteristics are body mass, diet, and diet breadth. Other
1337 characteristics (e.g., lifespan and litter/clutch size) may indirectly affect species persistence over time by
1338 influencing species reproductive output and demographic processes (Capdevila et al., 2022b). Finally, re-
1339 sponds to human pressures are known to be dependent on species' degree of specialisation, which I capture
1340 with characteristics reflecting specialisation in time (i.e., diel activity) and reflecting use of space (e.g., habi-
1341 tat breadth and geographical range area).

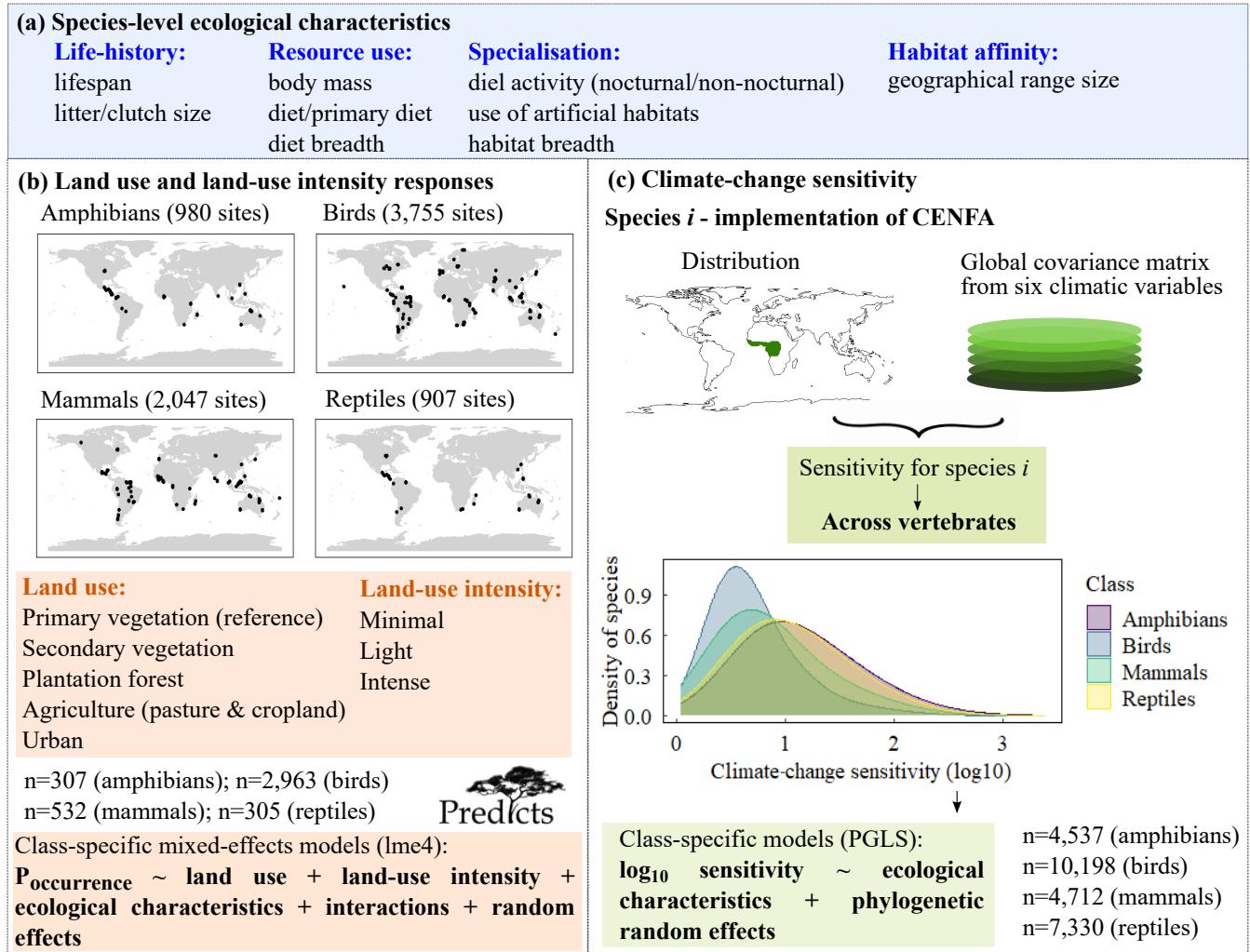


Figure 4.1: Framework of the study. (a) I collected ecological trait data and geographical range areas across terrestrial vertebrates (termed ‘ecological characteristics’). I then used two independent approaches to assess the influence of these characteristics on species’ responses to land use and on species’ climate-change sensitivity. (b) To assess the influence of traits on responses to land use and land-use intensity in each vertebrate class, I combined the ecological characteristics with the PREDICTS database. (c) To estimate species’ sensitivity to climate change, I used the CENFA framework (Rinnan and Lawler, 2019), which relies on the combination of species’ distributions with climatic variables to estimate sensitivity from properties of the species’ climatic niche space. I then built class-specific models to assess whether the ecological characteristics were associated with species’ sensitivity to climate change.

1342 **4.2 Methods**

1343 **4.2.1 Ecological characteristics (Figure 4.1a)**

1344 **Traits**

1345 I obtained the six following traits from Chapter 2 (in which I presented a trait data compilation across
1346 terrestrial vertebrates): body size (body mass and/or length, depending on the class); a proxy for species
1347 lifespan (generation length for mammals and birds; age at sexual maturity for amphibians; and maximum
1348 longevity for reptiles); litter or clutch size; diel activity; habitat breadth; and use of artificial habitats. I
1349 chose these traits because 1) they were available across all vertebrate classes, at least for a subset of species,
1350 allowing for a comparative assessment; and 2) they relate to species life-history, ecology, and resource use,
1351 such that they might influence species' land-use responses and climatic niche properties (and thus expected
1352 climate-change sensitivity). I couldn't capture intraspecific variation in trait values, and instead I used single
1353 mean values for all traits.

1354 I enhanced the trait data from Chapter 2 with species-level estimates of diet, lacking in the published
1355 database but likely important for understanding species' sensitivity to human pressures. For birds and mam-
1356 mals, I collected estimates of species primary diet (i.e., the diet inferred from the combination of food items
1357 totalling more than 50% of species' consumption), from the EltonTraits database (Wilman et al., 2014). For
1358 amphibians and reptiles, obtaining species *primary* diet was not possible, as there were no data available
1359 on the relative consumption of different food items. For amphibians, the AmphiBIO database (Oliveira
1360 et al., 2017) provided information on species consumption of different food items (just in terms of pres-
1361 ence/absence in the diet, but without estimation of their percent use), so I inferred diet on the basis of these
1362 reported food items (however the coverage was low, with more than 75% of the species missing diet infor-
1363 mation; Appendix 3, Figure S4.1). For reptiles, there was no available data collection describing diet. For
1364 both reptiles and amphibians, I supplemented the existing datasets with data on species consumption from
1365 published sources (recording the presence/absence of different food items in species consumption; data com-
1366 piled by Rhiannon Osborne-Tonner), for an additional 108 amphibians and for 239 reptiles (see Appendix 3,
1367 S4.1: 'Compiling diet information').

1368 I standardised the diet data across the vertebrate classes, by grouping species in five different diet cate-
1369 gories: vertebrate eaters; invertebrate eaters; plant/seed eaters; fruit/nectar eaters; and omnivores (for mam-
1370 mals and birds, species were classified as omnivores when all food items had a percent use $\leq 50\%$; and for
1371 amphibians and reptiles, when species were known to consume both plant and animal matter, or both in-

1372 vertebrates and vertebrates). I also calculated species diet breadth – the total number of recorded food items
1373 (in terms of presence/absence) known to be consumed by a species. More information on the compilation of
1374 dietary information can be found in Appendix 3, S4.1: ‘Compiling diet information’.

1375 **Geographical range area**

1376 I used extent-of-occurrence maps from BirdLife International for birds (<http://datazone.birdlife.org/species/requestdis>), from the IUCN Red List for mammals and amphibians (IUCN, 2020),
1377 and from Roll et al. (2017) for reptiles (all downloaded in April 2020). I excluded areas occupied during
1378 non-breeding seasons and areas falling outside species known elevational limits (following Chapter 2). The
1379 range maps were then converted to the raster format (‘raster’ package, version 3.5.15 Hijmans (2022)), and
1380 I estimated species geographical range areas using a resolution of 1 km × 1 km with Behrmann’s equal-area
1381 projection. Although range area cannot be considered a trait (which is a property measurable at the level of
1382 individual organisms), I included range area in the analyses because past work has shown that range area is
1383 an important correlate of species’ responses to land-use (Newbold et al., 2018) and climate change (Thuiller
1384 et al., 2005). In addition, range area may correlate with other aspects of species’ ecology that I could not
1385 include directly in the analysis because of limited data availability, such as dispersal ability (Capurucho et al.,
1386 2020).
1387

1388 **Phylogenies**

1389 I used information on species’ phylogenetic position in the imputations of missing trait values (see next
1390 section), and also to control for phylogenetic relationships in the models investigating the association be-
1391 tween species’ ecological characteristics and species’ estimated climate-change sensitivity. Class-specific
1392 phylogenetic trees were downloaded April 2020 from <https://zenodo.org/record/3690867#.Xyc5wyhKhPZ> for mammals (Phylacine 1.2; Faurby et al. (2018, 2020)); and from <https://data.vertlife.org/> for amphibians (Jetz and Pyron, 2018), birds (Jetz et al., 2012) and squamates (Tonini
1394 et al., 2016). For each class, I used a consensus tree obtained with the TreeAnnotator programme of the
1395 BEAST software (Bouckaert et al. 2014), from an available distribution of 1000 trees.
1396

1397 **4.2.2 Imputations of missing trait values**

1398 For some of the traits and classes, there was a substantial proportion of missing trait values (Figure S4.1).
1399 To fill these gaps, I imputed missing trait values using random forests, as implemented with the ‘missforest’
1400 function of the ‘missForest’ package in R (version 1.4, Stekhoven (2016) and Stekhoven and Bühlmann

1401 (2012)). ‘missforest’ is one of the best methods for missing-value imputations when working with continuous
1402 and categorical variables, and when including species phylogenetic position as a predictor (Debastiani et al.,
1403 2021; Penone et al., 2014). After showing that several traits were strongly phylogenetically conserved
1404 (Table S4.1), I included ten phylogenetic eigenvectors in the imputations (Penone et al., 2014), as well as
1405 taxonomic orders as a categorical variable (included to account for the taxonomic positions of species that
1406 were not represented in the phylogenies). Full details are given in Appendix 3 (S4.2: ‘Imputing missing
1407 trait values’). After imputation, continuous traits were \log_{10} -transformed to improve normality (except for
1408 habitat and diet breadth, which I square-root transformed; this transformation was more appropriate here
1409 because the distributions of habitat breadth and diet breadth tended to be less right-skewed than that of the
1410 other traits, and the range of values was smaller).

1411 4.2.3 Characterizing the influence of traits on species’ land-use responses (Figure 4.1b)

1412 Vertebrate assemblage composition

1413 To compare vertebrate assemblages in different land-use types, I used the PREDICTS database (Hudson
1414 et al., 2014, 2017). PREDICTS is a collection of independent studies that have sampled biodiversity in sites
1415 of varying land use and land-use intensity. Samples are mostly of species abundance, sometimes species
1416 occurrence, and rarely just overall species richness. It is one of the most comprehensive such databases
1417 to date, with 4,107 vertebrate species sampled across 7,689 sites considered in this work (Figure 4.1b). In
1418 PREDICTS, sites are assigned to one of the following land-use categories: primary vegetation (native vege-
1419 tation); secondary vegetation, plantation forest, pasture, cropland, and urban (disturbed land uses; see Table
1420 S4.2 and Hudson et al. (2014, 2017) for more details). Each site is also characterised in terms of land-use
1421 intensity based on land-use-specific criteria (such as mechanisation degree, crop diversity and agricultural
1422 inputs for cropland; Hudson et al. (2014)). Land-use intensity is divided into three categories to reflect the
1423 degree of human transformation and impacts on the land: minimal, light or intense. Here, I considered
1424 minimally-used primary vegetation to be the least-disturbed reference land use against which I compare all
1425 other land-use types. I grouped pasture and cropland together into a category I termed ‘agricultural’. As the
1426 design of the PREDICTS database is not balanced, sample sizes varied among classes and land-use types
1427 (Figure S4.3).

1428 **Full models (all-predictor models)**

1429 Within each vertebrate class, I investigated whether interactions among the ecological characteristics, land
1430 use and land-use intensity explained species occurrence probability. I fitted four binomial mixed-effects
1431 models (one for each class), using the ‘lme4’ package (version 1.1-23; Bates et al. (2015)), with random
1432 effects accounting for study, site and species identity to account for the nested design of the database, taxo-
1433 nomic non-independence, and repeated observations among species. I did not consider interactions among
1434 the ecological characteristics, but I included interactions between land use and ecological characteristics, and
1435 between land-use intensity and ecological characteristics. Before fitting the models, I checked the degree of
1436 multicollinearity among explanatory variables using generalised variance inflation factors (GVIF; Fox and
1437 Monette (1992)), with a threshold of 5 for the detection of multicollinearity (Tables S4.3-S4.8). For amphibi-
1438 ans and reptiles, including both diet and diet breadth was problematic, so I excluded diet from the set of
1439 predictors for these classes on the basis of the GVIF scores. Models investigating the effects of diet were
1440 built separately (see next section, ‘Partial models’).

1441 I did not use phylogenetic random effects directly in the models because of the computational load re-
1442 quired by such models when working with several hundred or thousands of species. However, I checked the
1443 phylogenetic signal in the models’ residuals using Pagel’s λ (Pagel, 1999). Thus, in each class, the model
1444 fitted was:

1445

1446 $P_{\text{occurrence}} \sim \text{land use} + \text{land-use intensity} + \text{species-level ecological characteristics} +$
1447 $\text{land use : species-level ecological characteristics} +$
1448 $\text{land-use intensity : species-level ecological characteristics} +$
1449 $(1|\text{study identity}) + (1|\text{site identity}) + (1|\text{species identity}).$

1450

1451 To verify that the models’ estimates were robust to any violation of distributional assumptions, I fitted
1452 the models again using a Bayesian framework (using the ‘MCMCglmm’ package version 2.32, Hadfield
1453 (2010)).

1454 **Partial models (single-predictor models)**

1455 In addition to the full models, I fitted partial models for each class. These were fitted to visualise occurrence
1456 patterns for each trait independently of other traits. The structure of the models was similar to that of the full
1457 models, except that I included a single species-level characteristic at a time in each model.

1458 **Effects of categorical ecological characteristics on species' occurrence probability (Figure 4.2a)**

1459 The influence of categorical traits on species' responses to land use and land-use intensity can be visualised
1460 in two ways: either by comparing occurrence probability in different land-use types relative to species with
1461 similar traits (I term such effects 'among land-use type effects', Figure 4.2a); or by comparing occurrence
1462 probability in a given land-use type relative to species with different traits (I term such effects 'within land-
1463 use type effects', Figure 4.2a).

- 1464 • Within land-use type effects (Figure 4.2a): from the full, all-predictor models fitted for each class, I
1465 focused on the interactive effects between land use and ecological characteristics (and between land-
1466 use intensity and ecological characteristics). These interactions indicated whether, in a given land-use
1467 type, there were any significant differences in occurrence probability between species with different
1468 traits. In other words, I looked at whether any trait level lowered or increased occurrence probability
1469 in each land-use type, compared to a reference trait level. I used this approach for all the categorical
1470 predictors, except diet (interpreting within land-use type effects for primary diet being complicated by
1471 the fact that there were more than two levels for this trait).
- 1472 • Among land-use type effects (Figure 4.2a): from a partial model, I predicted occurrence probability
1473 in the different land uses for all different levels of the trait. The partial models allowed to visualise
1474 occurrence patterns across land-use types for single explanatory variables, without having to account
1475 for the values of other variables. I used this approach to evaluate the influence of diet on species'
1476 land-use responses.

1477 **Effects of continuous ecological characteristics on species' occurrence probability (Figure 4.2b)**

1478 For a given continuous ecological characteristic, any effect of land use or land-use intensity can be as-
1479 sessed through changes in the slope of the relationship between the ecological characteristic and occurrence
1480 probability (Figure 4.2b). When an ecological characteristic negatively impacts occurrence probability in a
1481 disturbed land use, I expect the slope of the relationship to be more negative than the slope for the reference
1482 land use (minimally-used primary vegetation). Focussing on slopes does not allow to infer absolute changes
1483 in occurrence probability across land-use types (e.g., a positive slope in a disturbed land use does not mean
1484 that there are absolute increases in occurrence probability in that land use, but only that higher values of the
1485 ecological characteristic are associated with relatively higher occurrence probability in that land-use type).
1486 This is because I do not assess changes in the mean occurrence probability here (which would require to

1487 consider the intercept of the relationship between the ecological characteristic and occurrence probability in
 1488 different land-use types). Thus, I only capture ‘within land-use type’ effects for continuous predictors.

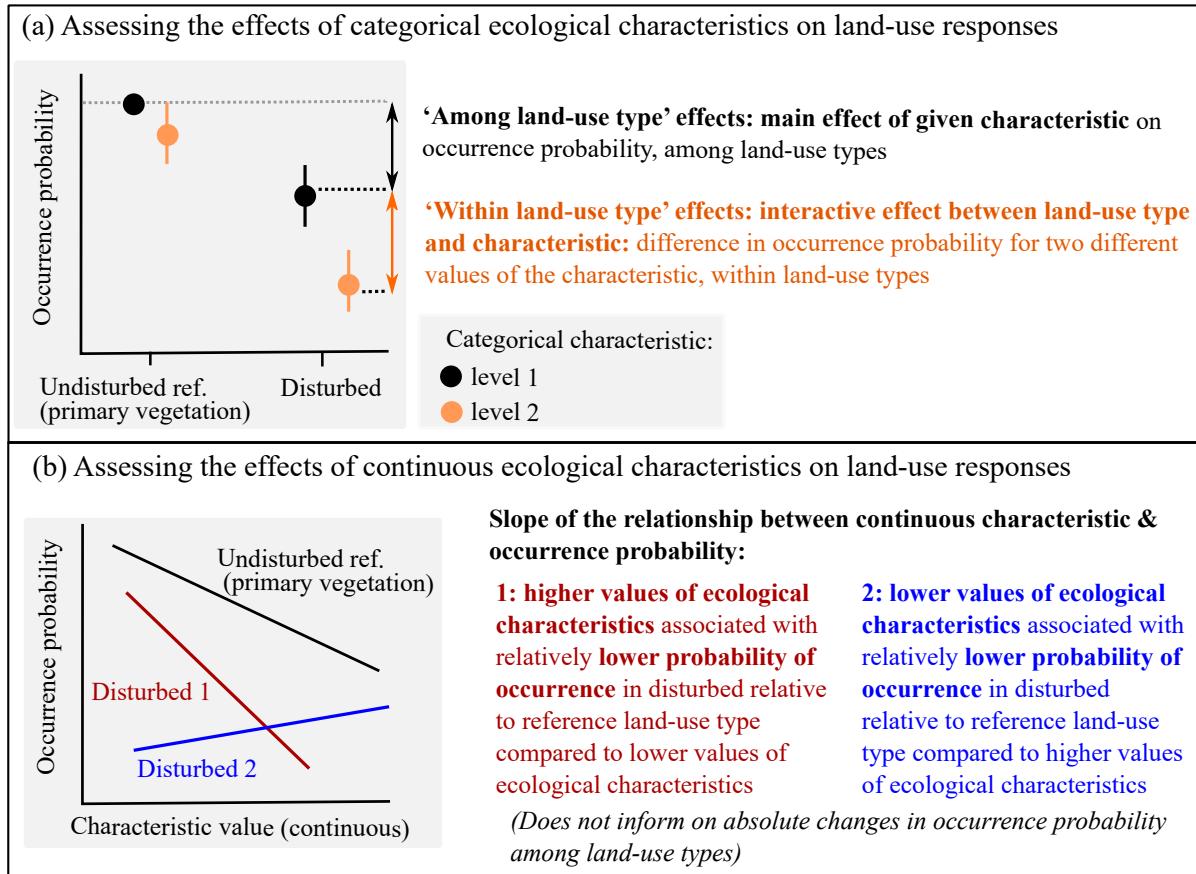


Figure 4.2: Assessing the effects of ecological characteristics on species’ land-use responses: methodology for (a) categorical characteristics and (b) continuous characteristics. (a) For all categorical characteristics, except diet, I look at ‘within land-use type’ effects, asking whether there are significant differences in occurrence probability among species with different ecological characteristics in a given land-use type. For diet, I look at ‘among land-use type’ effects, comparing species occurrence probability in disturbed land uses versus that in primary vegetation (I chose this approach here because visualising ‘within land-use type’ effects for diet is complicated by the fact that there were more than two levels for this categorical trait). (b) For continuous characteristics, I focus on the relationship with occurrence probability, and I investigate how the slope of this relationship is affected by land-use type, i.e. a ‘within land-use type’ effect.

1489 **Validation on complete trait data subset (no imputed trait values)**

1490 To assess whether the results were robust to trait imputation uncertainty, I fitted the models again for the
 1491 subset of species for which I had complete, non-imputed data for all ecological characteristics. The models’
 1492 structure was unchanged for birds and mammals. For amphibians, I excluded both diet and litter/clutch size
 1493 because of multicollinearity issues, and I also excluded lifespan proxy and body mass (as there were too
 1494 many missing values in the dataset, 85% and 59% respectively). For reptiles, I excluded both diet and body

¹⁴⁹⁵ mass because of multicollinearity issues.

1496 **4.2.4 Characterizing the influence of traits on species' sensitivity to climate change (Figure**
1497 **4.1c)**

1498 I estimated climate-change sensitivity across vertebrate species using the ‘Climate-niche Factor Analysis’
1499 (CENFA) approach developed by Rinnan and Lawler (2019), implemented with the ‘CENFA’ R package
1500 version 1.1.1 (Rinnan, 2021). CENFA is a spatial approach for estimating species’ climate-change sensitivity,
1501 exposure, and vulnerability. CENFA combines distribution data with climatic variables to estimate sensitivity
1502 and vulnerability from properties of species’ climatic niches (see Rinnan and Lawler (2019) for details).
1503 CENFA has been used in previous studies focused on a small number of species or on a few taxonomic
1504 groups, but to my knowledge has not yet been applied across all terrestrial vertebrates.

1505 **Historical climate data**

1506 I used global climate data from WorldClim version 2.1 (Fick and Hijmans, 2017). I downloaded 19 climatic
1507 variables at a resolution of 2.5 arcminutes (~ 4.6 km at the Equator). I removed variables that were strongly
1508 collinear with any other climatic variables (using a threshold of 0.65 for Spearman correlation coefficients).
1509 I obtained six groups of intercorrelated variables (using the ‘removeCollinearity’ function from the ‘virtu-
1510 alspecies’ R package version 1.5.1 (Leroy et al., 2015); Figure S4.4), and randomly selected one climatic
1511 variable in each group. The final set comprised six climatic variables: annual mean temperature (bio1), mean
1512 diurnal temperature range (bio2), maximum temperature of the warmest month (bio5), annual precipitation
1513 (bio12), precipitation seasonality (bio15), and precipitation of the coldest quarter (bio19).

1514 **Estimating climate-change sensitivity from CENFA**

1515 All climatic variables and distribution files were re-projected to a resolution of $5\text{ km} \times 5\text{ km}$ in the Behrmann
1516 equal-area projection. I picked this resolution because the coarser the resolution, the more climate-change
1517 sensitivity tended to be underestimated for narrowly distributed species (Figures S4.5 & S4.6). However,
1518 finer resolutions demand a large amount of memory space when working at global scales across all terrestrial
1519 vertebrates. I found the $5\text{-km} \times 5\text{-km}$ resolution to be an acceptable trade-off between computational load
1520 and accuracy of the sensitivity estimations. However, when working at $5\text{ km} \times 5\text{ km}$ resolution, there were
1521 still some narrowly distributed species for which sensitivity was likely underestimated (Figure S4.6). Thus,
1522 I chose to exclude species with a range area $\leq 100\text{ km}^2$ from further analyses (i.e., excluding narrow-ranging
1523 species whose distributions could intersect up to 4 grid cells). In doing so, the sample size was reduced by
1524 660 species for amphibians, by 142 species for birds, by 129 species for mammals, and by 615 species for

1525 reptiles (the final sample sizes were: n=4,537 for amphibians; n=10,198 for birds; n=4,721 for mammals;
1526 n=7,330 for reptiles). My results were overall robust to the exclusion of these species (see Results section).

1527 I then combined the climate data with the species' distributions to estimate sensitivity to climate change,
1528 applying the CENFA framework across terrestrial vertebrates (Figure 4.1c). Further details of the imple-
1529 mentation of the CENFA framework are given in Appendix 3 (S4.5: 'Implementing Climate-niche Factor
1530 Analysis across terrestrial vertebrates').

1531 **Climate-change sensitivity models**

1532 I used phylogenetic least-square (PGLS) regressions, implemented in the 'caper' R package version 1.0.1
1533 (Orme et al., 2018), to assess the effects of ecological characteristics on species' estimated sensitivity to
1534 climate change, while controlling for phylogenetic relationships among species. I combined the ecological
1535 characteristics and the phylogenies using the 'comparative.data' function from the 'caper' package, and then
1536 built class-specific models to explain climate-change sensitivity with the ecological characteristics (Figure
1537 4.1c). Before fitting the models, I checked for multicollinearity among the predictors using GVIF scores.
1538 Across all classes, the models included all the main effects of the ecological characteristics, except for am-
1539 phibians, for which I dropped diet breadth (which was strongly collinear with diet; Tables S4.9-S4.13). For
1540 the continuous predictors, I fitted third-order polynomials to allow for non-linearity of the responses (I in-
1541 cluded third order polynomials for the climate-change sensitivity models but not for the land-use models
1542 because the PGLS model had a simpler structure than the land-use models, were less computationally inten-
1543 sive, and also because the number of estimated parameters was already high for the land-use models without
1544 allowing for third-order polynomials). As such, the general form of the PGLS models was:
1545 $\log_{10}(\text{climate-change sensitivity}) \sim \text{poly}(\log_{10}(\text{continuous ecological characteristics}), 3) +$
1546 categorical ecological characteristics +
1547 phylogenetic random effects.

1548 **Models' robustness checks**

- 1549 • To check whether the results were robust to the exclusion of species whose range area was ≤ 100
1550 km^2 , I repeated the models on all species (including those with range area $\leq 100 \text{ km}^2$: n=5,208 for
1551 amphibians; n=10,340 for birds; n=4,844 for mammals; n=7,951 for reptiles).
- 1552 • For birds and mammals, I further checked whether the results were robust to the exclusion of migratory
1553 species, as the breeding range and the non-breeding range of migratory species can differ, possibly

1554 impacting the definition of the species climatic niche spaces and thus the estimation of climate-change
1555 sensitivity. I identified migratory mammals using the dataset from Gnanadesikan et al. (2017), and
1556 migratory birds from BirdLife international (check how to cite / acknowledge). I ran the models again
1557 after excluding 1,882 bird species and 155 mammal species identified as migratory.

- 1558 • I further checked whether the results were sensitive to the use of different resolutions. As I showed
1559 in Figures S4.5 & S4.6, resolution can have an impact on the estimation of climate-change sensitiv-
1560 ity (e.g., coarser resolutions tend to underestimate the climate-change sensitivity of narrow-ranging
1561 species, Figures S4.5 & S4.6). Here, I verified whether the results were robust to the use of different
1562 resolutions. I thus fitted the models again using estimates of climate-change sensitivity quantified from
1563 different resolutions depending on species' geographical range area: for species whose range area fell
1564 in the $]100-400 \text{ km}^2]$ interval, I used a $5 \text{ km} \times 5 \text{ km}$ resolution (thus excluding those narrow-ranging
1565 species whose range area could intersect up to 4 grid cells, as for the main models); for species whose
1566 range area fell in the $]400-10,000 \text{ km}^2]$ interval, I used a $10 \text{ km} \times 10 \text{ km}$ resolution; and finally, for
1567 species whose range area $<10,000 \text{ km}^2$, I used a $50 \text{ km} \times 50 \text{ km}$ resolution. Thus, in each case, I used
1568 a cut-off value for the left bound that corresponds to the surface area covered by a species that would
1569 intersect up to four grid cells. This cut-off value is a conservative choice that aims to avoid overesti-
1570 mating the climate-change sensitivity of the more narrowly-distributed species given any resolution.

1571 **Validations on complete trait data**

1572 Finally, to assess the degree to which the results were robust to trait imputation uncertainty, I fitted
1573 the models again for the subset of species for which I had empirical (i.e., non-imputed) trait estimates.
1574 Diet was excluded for amphibians and reptiles on the basis of high collinearity ($\text{GVIF} > 5$). I fitted
1575 first-order polynomials here because of the substantially reduced sample size compared to the main
1576 models.

1577 **4.3 Results**

1578 **4.3.1 Land-use responses**

1579 **'Within land-use type' effects (Table 4.1a)**

1580 Land-use, land-use intensity, species' ecological characteristics and their interactions had significant effects
1581 on species occurrence probability. Significant interactive effects between land use and ecological charac-

1582 teristics (and between land-use intensity and ecological characteristics) reflected differences in the ability
1583 of species with different ecological characteristics to cope within the disturbed land-use types (Table 4.1a).
1584 Across all classes, species with narrower geographical range areas, smaller habitat breadth and inability to
1585 exploit artificial habitats showed greater decreases in occurrence probability within disturbed land uses, than
1586 species with larger range areas, larger habitat breadth and ability to exploit artificial habitats (the only ex-
1587 ceptions were opposite effects found for mammals and reptiles for habitat breadth in two of the land-use
1588 types). The effects of the other ecological characteristics differed in direction depending on class and land
1589 use, impeding any generalisation (Table 4.1a). For instance, I found that being smaller and longer-lived was
1590 associated with decreases in occurrence probability for birds found in agricultural areas, but with increases
1591 in occurrence probability for urban birds; and that longer-lived species tended to be more negatively affected
1592 for mammals and reptiles, whereas I found evidence of opposite trends for amphibians.

1593 I would like to highlight that the ‘within land-use type’ effects summarised in Table 4.1a do not nec-
1594 essarily reflect occurrence patterns among land-use types. For example, in all classes, ‘among land-use
1595 type’ effects derived from partial models showed that occurrence probability in disturbed land uses was
1596 strongly negatively affected for natural habitat specialists, compared with primary vegetation levels (Figure
1597 S4.7). On the other hand, in most classes and disturbed land uses, artificial habitat users either increased or
1598 showed no significant difference in occurrence probability. One exception was for reptiles, where the effect
1599 of habitat specialisation was mostly non-significant within land-use types (Table 4.1a), with both natural
1600 habitat specialists and artificial habitat users showing important declines in some disturbed land uses (e.g.,
1601 intensely-used agricultural areas, Figure S4.7d). Similarly, the occurrence probability of both nocturnal and
1602 non-nocturnal species was negatively impacted in disturbed land uses compared with primary vegetation
1603 (Figure S4.8), such that land-use responses were not distinguishable between nocturnal and non-nocturnal
1604 species for all classes and land-use types.

Table 4.1: Summary of the effects of the ecological characteristics (except for primary diet) on (a) species' responses to disturbed land uses ('within land-use type' effects) and (b) species' climate-change sensitivity, for each class of terrestrial vertebrates. The symbol - indicates where a characteristic has a significant negative effect on occurrence probability within a disturbed land-use type (within any of the land-use intensities), or where the characteristic renders species significantly more sensitive to climate change. A + indicates a significantly positive effect of a characteristic on occurrence probability in a land-use type (within any of the land-use intensities), or significantly lower sensitivity to climate change. For the land-use effects, I report 'within land-use type' effects here, that is, within a disturbed land use whether there were significant differences in occurrence probability among species with different trait values (see Figure 4.2). These effects were derived from the interactive terms of the full, all-predictor models.

(a) "Within land-use type" effects of the species-level characteristics on occurrence probability - <=> more sensitive; + <=> less sensitive															Predictors			
-	-	-	-	(-)	-	0	-	0	-	-	-	-	0	-	0	NA	More narrowly-distributed	
0	-	-	0	+	-	-	-	0	-	-	0	-	0	+	NA		Smaller habitat breadth	
-	-	(-)	-	-	-	0	0	-	0	0	-	-	NA	Natural habitat specialist				
0	+	+	-	-	+	0	+	-	+	+	-	0	0	0	NA		Non-nocturnal	
-	0	+	(-)	-	0	+	+	-	0	0	+	-	0	NA			Narrower diet breadth	
0	0	+	0	(+)	+	+	0	0	-	0	-	0	+	0	NA			Smaller body mass
+	0	0	0	0	-	0	0	+	-	0	0	0	-	NA				Smaller litter/clutch size
(-)	0	(+)	+	0	0	0	-	+	(+)	+	0	-	0	NA				Shorter-lived

1605 **Effects of diet on species' occurrence probability (Figure 4.3)**

1606 In all classes, diet had significant effects on occurrence probability in disturbed land uses (Figure 4.3).
1607 Changes in occurrence probability in disturbed land uses differed among classes and dietary groups. Overall,
1608 invertebrate eaters tended to be negatively affected in disturbed land uses (e.g., -66% average declines in
1609 occurrence probability for amphibians in intensely used agricultural areas, compared with minimally-used
1610 primary vegetation). Omnivores were both negatively and positively impacted, showing both important
1611 decreases (e.g., -81% for reptiles in intensely used plantation forest) as well as strong increases (e.g., +43%
1612 for lightly used urban areas in birds). Overall, fruit/nectar eaters showed important declines in occurrence
1613 probability for mammals and birds, as opposed to plants/seeds eaters, whose occurrence probability tended
1614 to be strongly positively affected for birds, and dependent on land-use intensity for mammals (with increases
1615 in minimally-used land-types, but not in more intensely-used land-types). Finally, I also detected significant
1616 changes in occurrence probability for vertebrate eaters, with some declines for mammals in agricultural areas
1617 (-75% on average in intense uses), but also some increases (e.g., +43% on average for birds in lightly used
1618 agricultural areas).

1619 **Explanatory power for the full models & variance explained by each characteristic (Figure 4.4)**

1620 Overall, land use, land-use intensity and the ecological characteristics explained a small amount of the total
1621 variation in species' occurrence probability (marginal R²: 0.15 for amphibians; 0.054 for birds; 0.15 for
1622 mammals; 0.13 for reptiles), in part because the random effects explained a substantial proportion (condi-
1623 tional R²: 0.59 for amphibians; 0.61 for birds; 0.72 for mammals; 0.57 for reptiles). The relative importance
1624 of traits explaining the most variation differed among classes, with interactions between land use and habitat
1625 breadth explaining the most variation in amphibians and birds, but interactions between land use and body
1626 mass explaining the most variation for mammals, and interactions between land use and lifespan explaining
1627 the most variation for reptiles (Figure 4.4a).

1628 Finally, the models' diagnostics showed evidence of deviations from distributional assumptions (diag-
1629 nistic plots for the full models are shown in Figures S4.9-S4.12). However, when estimated from a Bayesian
1630 framework, the models' estimates were mostly congruent (results not shown), so the frequentist approach I
1631 used with 'lme4' was robust despite the deviations from distributional assumptions. The phylogenetic sig-
1632 nals in the models' residuals were low and not significant (Pagel's $\lambda < 0.01$ for amphibians and reptiles, $p \approx$
1633 1; $\lambda = 0.13$ for mammals, $p = 0.09$; $\lambda = 0.01$ for birds, $p = 0.56$), despite not having included phylogenetic
1634 random effects.

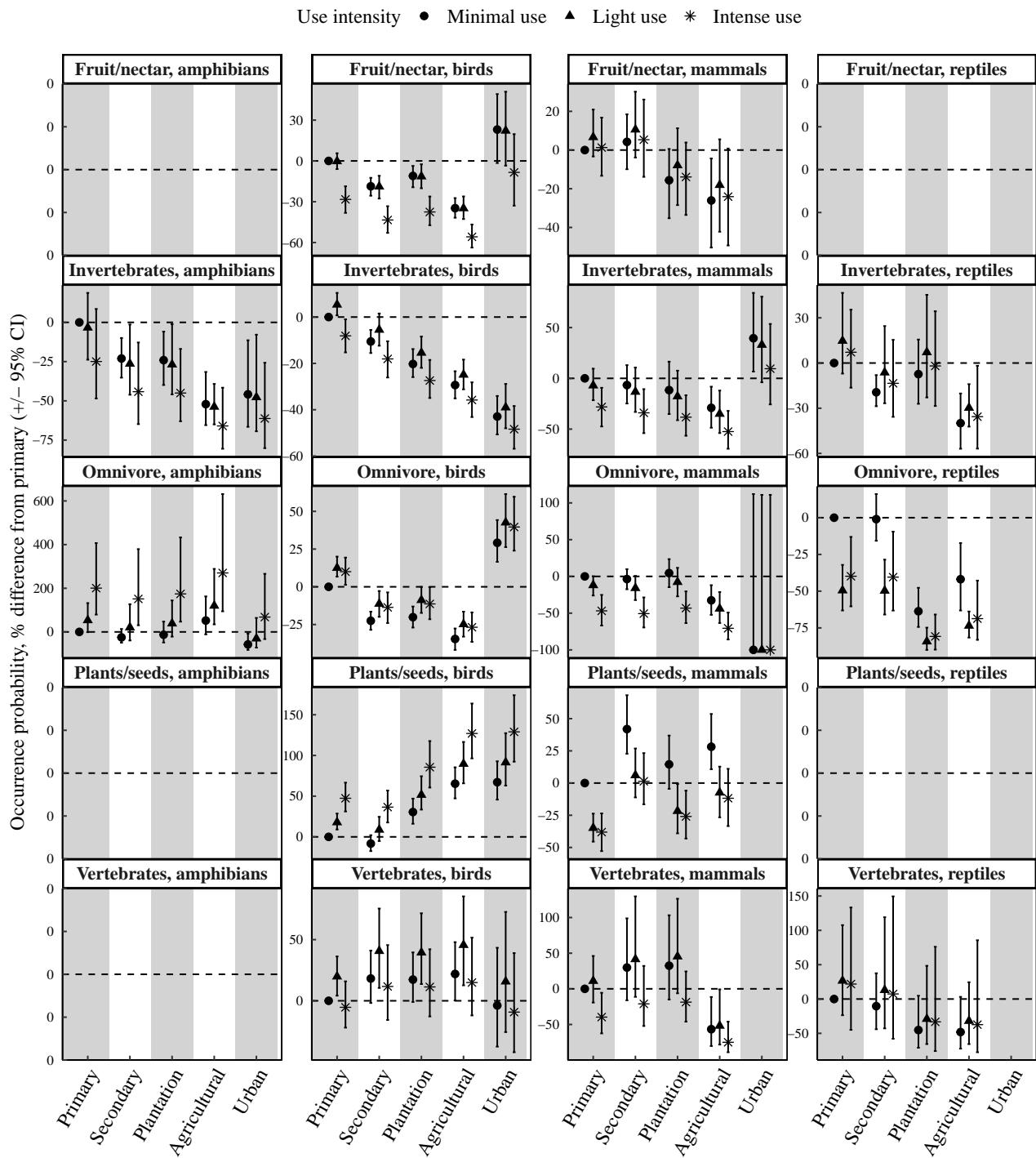


Figure 4.3: Predicted occurrence probability as a function of land use, land-use intensity, diet and their interactions, for each class of terrestrial vertebrates (mean \pm 95% confidence interval; the predictions are rescaled with reference to minimally-used primary vegetation). The predictions were obtained from the partial models fitted for each class, considering only diet among the ecological characteristics. Empty plots are drawn where there were no data for a diet category for a given class. Effects could not be estimated for urban reptiles, as well as for urban vertebrate, fruit/nectar and plant/seed eaters for mammals because there were no sampled species. Primary: primary vegetation; Secondary: secondary vegetation; Plantation: plantation forest; Agricultural: cropland and pasture.

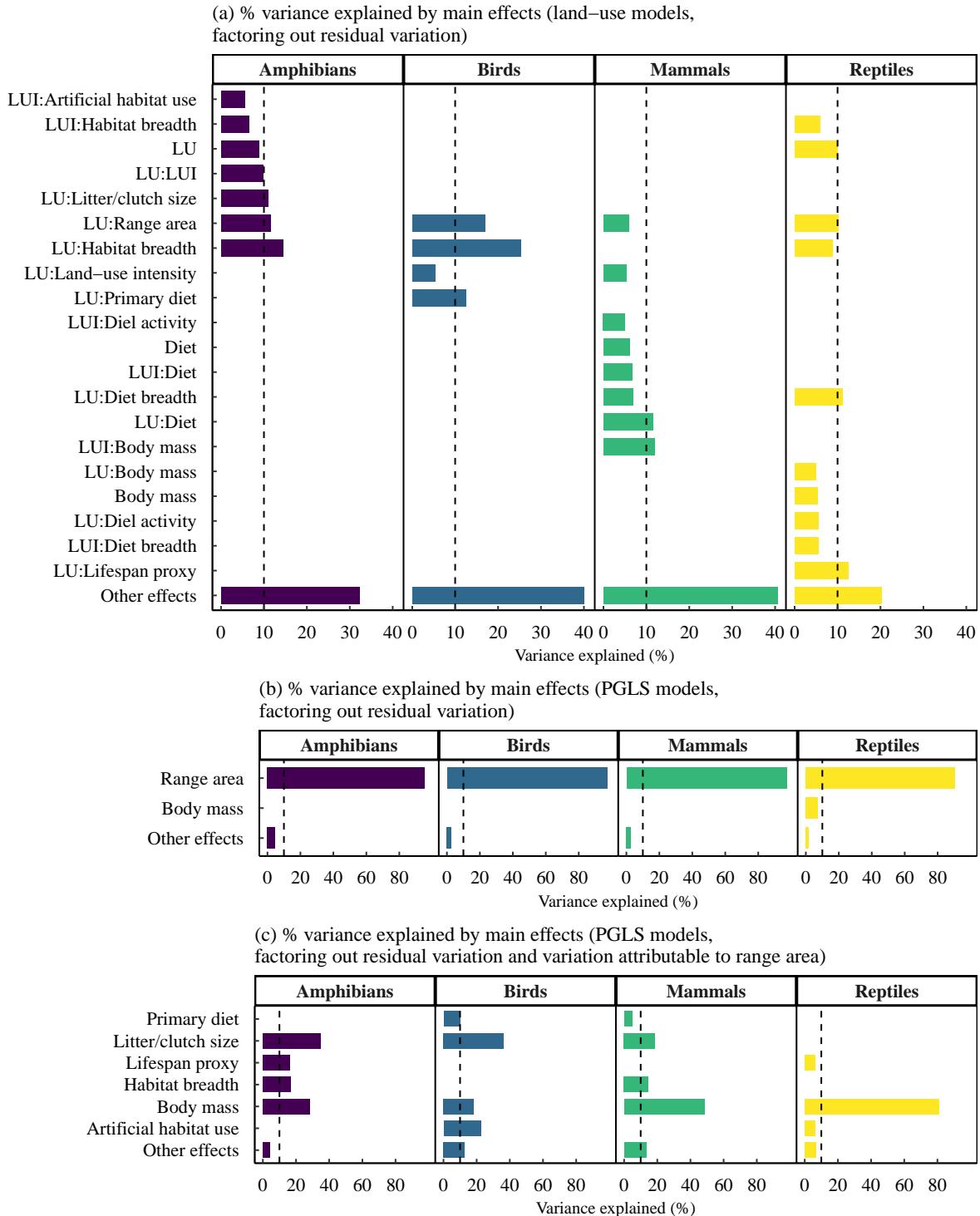


Figure 4.4: Proportion of the explained variance attributable to each of the main effects for (a) the mixed-effects models fitting the effects of land use, land-use intensity, and ecological characteristics on species occurrence probability (after factoring out residual variation); (b) for the phylogenetic least-squares regressions investigating whether the ecological characteristics explained climate-change sensitivity (after factoring out residual variation); and (c) for the phylogenetic least-squares regressions investigating whether the ecological characteristics explained climate-change sensitivity (after factoring out the variance explained by geographical range area and the residual variation). The dashed vertical lines mark 10% explained variance (for visualisation purposes). I individually show all the effects that explain more than 5% of the overall variation. Effects that individually explain less than 5% of the overall variation are grouped together as ‘Other effects’. LU: land use; LUI: land-use intensity.

1635 **4.3.2 Climate-change sensitivity**

1636 The ecological characteristics were significantly associated with climate-change sensitivity in all classes
1637 (Tables 4.1b & 4.2, Figures 4.5 & 4.6); models' coefficients shown in Tables S4.14-S4.17). Overall, climate-
1638 change sensitivity was highest for amphibians (median \log_{10} -sensitivity: 1.1; 95% interpercentile range:
1639 0.40-2.2), then reptiles (median \log_{10} -sensitivity: 1.0; 95% interpercentile range: 0.32-2.1), then mammals
1640 (median \log_{10} -sensitivity: 0.76; 95% interpercentile range: 0.22-2.0) and birds (median \log_{10} -sensitivity:
1641 0.62; 95% interpercentile range: 0.21-1.77). In all classes, narrower geographical range area, smaller habitat
1642 breadth and being specialised on natural habitats were consistently associated with higher climate-change
1643 sensitivity (Table 4.1b). However, other characteristics did not have consistent associations with climate-
1644 change sensitivity across classes, in different cases varying in both significance and direction. For instance,
1645 I found opposite associations between body mass and climate-change sensitivity for mammals, amphibians
1646 and reptiles on the one hand, and birds on the other hand.

1647 The PGLS models explained an important proportion of the overall variation in estimated climate-change
1648 sensitivity (adjusted $R^2 = 0.64$ for amphibians; 0.62 for birds; 0.63 for mammals and reptiles). Geographical
1649 range area explained the majority of this variation in climate-change sensitivity (about 60% in all classes;
1650 Figure 4.4b), which largely reflects the design of the CENFA approach. When factoring out residual variation
1651 and variation explained by geographical range area, the relative importance of the traits as correlates of
1652 climate-change sensitivity varied among classes (Figure 4.4c), with body mass explaining the most variation
1653 for mammals and reptiles, and litter/clutch size explaining the most variation for amphibians and birds.

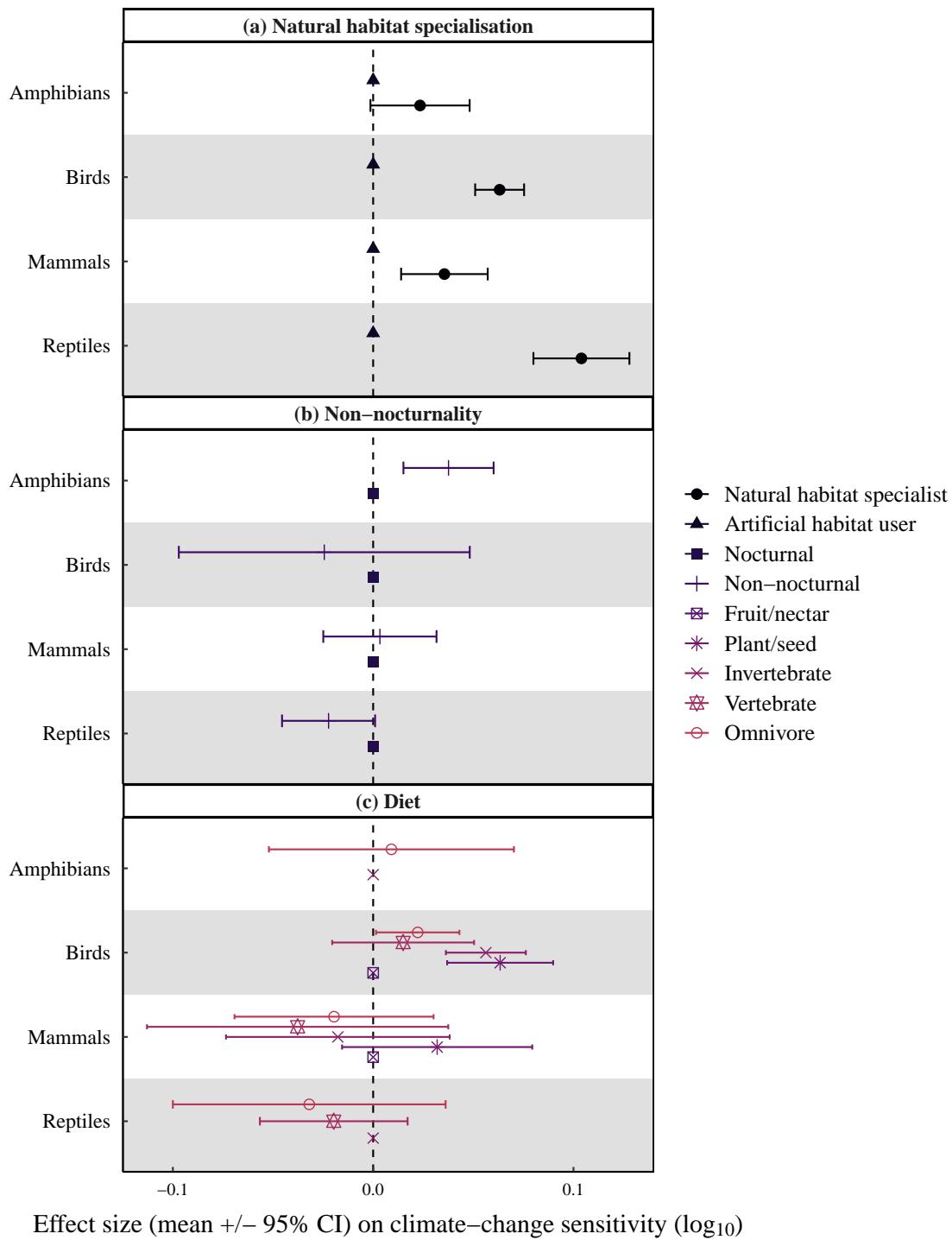


Figure 4.5: Estimated effects of the categorical traits on climate-change sensitivity, from the PGLS models fitted in each class (mean effect \pm 95% confidence interval). For each categorical trait, I show the effect size for all levels referring to the reference level (vertical dashed line). (a) For artificial habitat use, the reference level is ‘Artificial habitat user’; (b) for diel activity, the reference level is ‘Nocturnal’; (c) for diet, the reference level for mammals and birds is ‘Fruit/nectar’ eaters, but it is ‘Invertebrate’ eaters for amphibians and reptiles.

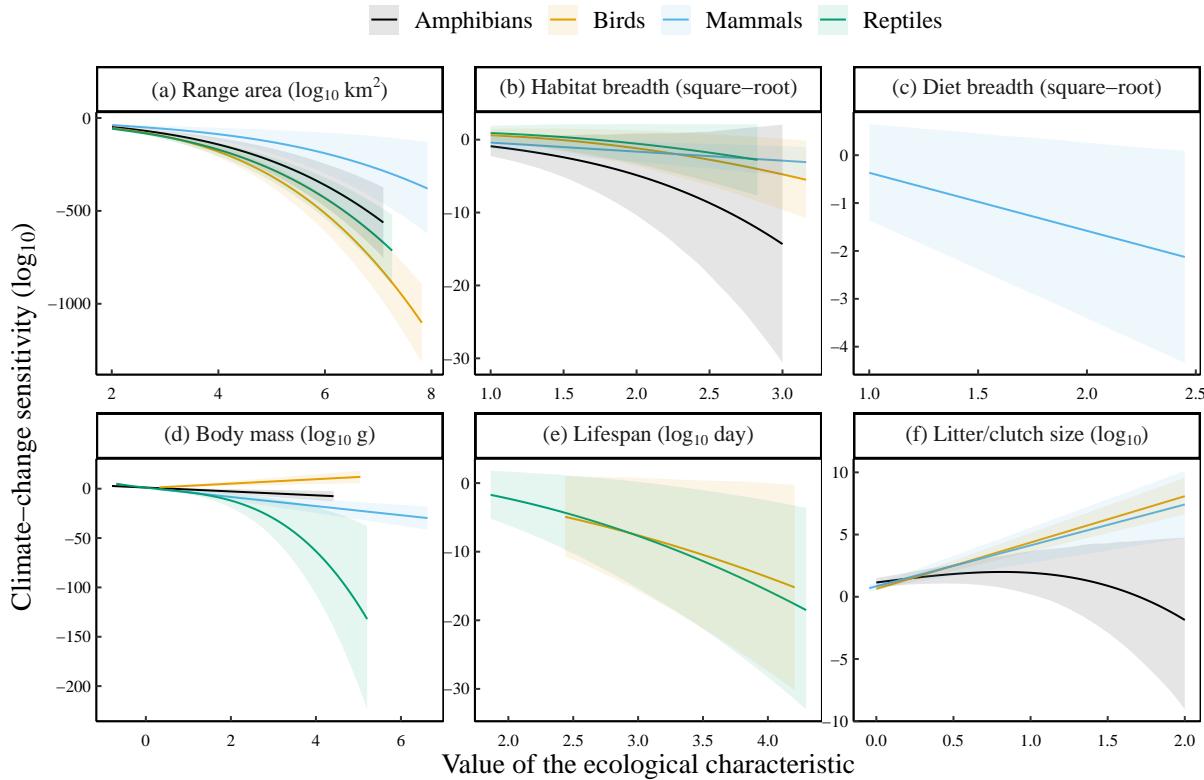


Figure 4.6: Effects of the continuous ecological characteristics on climate-change sensitivity, estimated from the PGLS models in each class. The lines represent the estimated relationships between climate-change sensitivity and ecological characteristics; the shaded areas are 95% confidence intervals. I plotted the estimated relationships only when they were found to be significant.

1654 Robustness of the PGLS models

1655 The PGLS models were robust to distributional assumptions (Figures S4.13-S4.16). Further, I found that the
 1656 results were robust to the different robustness checks, that is, to:

- 1657 • the exclusion of species whose range area was $\leq 100 \text{ km}^2$. When fitting the models on all species
 1658 (including those with range area $\leq 100 \text{ km}^2$), I found that the relationship between climate-change sen-
 1659 sitivity and geographical range area was reversed in all classes (with smaller-ranging species estimated
 1660 to be less sensitive). This result is likely an artefact caused by the underestimation of climate-change
 1661 sensitivity for the most narrow-ranging species, which would support the exclusion of such species
 1662 from the analysis. Other results were generally not sensitive to the exclusion of species whose range
 1663 area was $\leq 100 \text{ km}^2$ (Figure S4.17).
- 1664 • the exclusion of migratory birds and mammals. The models' coefficients and effects were congruent
 1665 with those estimated from the main models which included the migratory species (Figures S4.18,

1666 S4.19, S4.20).

- 1667 • the use of different resolutions for the estimation of climate-change sensitivity. The models' coeffi-
1668 cients and effects were congruent with those estimated from the main models (Figures S4.21, S4.22,
1669 S4.23).

Table 4.2: ANOVA summaries for the PGLS models investigating the associations between the species-level ecological characteristics and species' estimated climate-change sensitivity.

Class	Model terms	Df	Sum Square	Mean Square	F-value	Pr(>F)
Amphibians	poly(log ₁₀ (Body mass, 3))	3	9.85E-02	3.28E-02	3.41E+01	9.03E-22 ***
	poly(log ₁₀ (Lifespan proxy), 3)	3	5.62E-02	1.87E-02	1.94E+01	1.60E-12 ***
	poly(log ₁₀ (Litter/clutch size), 3)	3	1.22E-01	4.07E-02	4.22E+01	6.68E-27 ***
	poly(log ₁₀ (Range area), 3)	3	7.30E+00	2.43E+00	2.52E+03	0.00E+00 ***
	poly(sqrt(Habitat breadth), 3)	3	5.86E-02	1.95E-02	2.03E+01	4.69E-13 ***
	Specialisation	1	3.17E-03	3.17E-03	3.29E+00	6.96E-02 .
	Diel activity	1	1.03E-02	1.03E-02	1.07E+01	1.07E-03 **
	Diet	3	8.61E-04	2.87E-04	2.98E-01	8.27E-01
Birds	poly(log ₁₀ (Body mass, 3))	3	5.78E-02	1.93E-02	2.87E+01	1.87E-18 ***
	poly(log ₁₀ (Lifespan proxy), 3)	3	1.55E-02	5.18E-03	7.70E+00	3.87E-05 ***
	poly(log ₁₀ (Litter/clutch size), 3)	3	1.15E-01	3.83E-02	5.70E+01	1.63E-36 ***
	poly(log ₁₀ (Range area), 3)	3	1.10E+01	3.66E+00	5.45E+03	0.00E+00 ***
	poly(sqrt(Habitat breadth), 3)	3	1.29E-02	4.30E-03	6.39E+00	2.53E-04 ***
	poly(sqrt(Diet breadth), 3)	3	1.01E-02	3.38E-03	5.02E+00	1.77E-03 **
	Specialisation	1	7.10E-02	7.10E-02	1.06E+02	1.15E-24 ***
	Diel activity	1	1.58E-03	1.58E-03	2.34E+00	1.26E-01
Mammals	Primary diet	4	3.15E-02	7.88E-03	1.17E+01	1.70E-09 ***
	poly(log ₁₀ (Body mass, 3))	3	9.00E-02	3.00E-02	3.44E+01	5.41E-22 ***
	poly(log ₁₀ (Lifespan proxy), 3)	3	7.32E-03	2.44E-03	2.80E+00	3.85E-02 *
	poly(log ₁₀ (Litter/clutch size), 3)	3	3.46E-02	1.15E-02	1.32E+01	1.33E-08 ***
	poly(log ₁₀ (Range area), 3)	3	6.83E+00	2.28E+00	2.61E+03	0.00E+00 ***
	poly(sqrt(Habitat breadth), 3)	3	2.73E-02	9.11E-03	1.05E+01	7.50E-07 ***
	poly(sqrt(Diet breadth), 3)	3	8.64E-03	2.88E-03	3.31E+00	1.94E-02 *
	Specialisation	1	8.99E-03	8.99E-03	1.03E+01	1.33E-03 **
Reptiles	Diel activity	1	6.08E-06	6.08E-06	6.98E-03	9.33E-01
	Primary diet	4	9.32E-03	2.33E-03	2.67E+00	3.04E-02 *
	poly(log ₁₀ (Body mass), 3)	3	8.53E-01	2.84E-01	3.18E+02	3.24E-194 ***
	poly(log ₁₀ (Lifespan proxy), 3)	3	6.40E-02	2.13E-02	2.39E+01	2.25E-15 ***
	poly(log ₁₀ (Litter/clutch size), 3)	3	4.55E-02	1.52E-02	1.70E+01	5.50E-11 ***
	poly(log ₁₀ (Range area), 3)	3	1.03E+01	3.42E+00	3.83E+03	0.00E+00 ***
	poly(sqrt(Habitat breadth), 3)	3	1.34E-02	4.48E-03	5.01E+00	1.81E-03 **
	poly(sqrt(Diet breadth), 3)	3	6.37E-03	2.12E-03	2.38E+00	6.79E-02 .
	Specialisation	1	6.58E-02	6.58E-02	7.36E+01	1.13E-17 ***
	Diel activity	1	2.76E-03	2.76E-03	3.09E+00	7.89E-02 .
	Diet	2	1.34E-03	6.69E-04	7.48E-01	4.73E-01

1670 **4.3.3 Validations on complete trait data**

1671 Running the land-use and PGLS models again using data subsets for which I had empirical, non-imputed
1672 values only for the ecological characteristics showed that the conclusions of this work are likely robust
1673 to imputation uncertainty. Overall, across all classes, the associations of geographical range area, habitat
1674 breadth and use of artificial habitats with sensitivity to climate change and land-use responses were consistent
1675 with the main models (Table S4.10, Figure S4.24).

1676 **4.4 Discussion**

1677 Here, I investigated whether species' ecological characteristics were associated with species' sensitivity to
1678 two human pressures (climate change and land-use change), across terrestrial vertebrate classes. Overall, I
1679 found that geographical range area, habitat breadth and specialisation on natural habitats were consistently
1680 associated with sensitivity to both pressures across terrestrial vertebrate classes: narrower ranges, narrower
1681 habitat breadth and inability to exploit artificial habitats were associated with more negative land-use re-
1682 sponds and with higher climate-change sensitivity. My results are in line with previous meta-analyses that
1683 have found species' extinction risks to be associated with habitat breadth and range area (Chichorro et al.,
1684 2019), range shifts under contemporary climate change to be associated with species' historical range limits
1685 and habitat breadth (MacLean and Beissinger, 2017), and with many other studies on land-use responses or
1686 extinction risks (Newbold et al., 2018; Nowakowski et al., 2017; Purvis et al., 2000; Ripple et al., 2017).
1687 However, to the best of my knowledge, this work constitutes the first global study to compare patterns among
1688 vertebrate classes and between the two major human pressures of climate change and land use. The results
1689 of this Chapter have important implications for conservation, as they mean that land-use and climate change
1690 are non-randomly affecting all terrestrial vertebrates, with a consistently higher risk for geographically rarer
1691 species and habitat specialists. Geographical rarity has been employed by the IUCN for many years in vul-
1692 nerability assessments (Rodrigues et al., 2006), and this work provides further support for its integration in
1693 large-scale assessments. My results also lend support to the idea that habitat specialisation is a likely indica-
1694 tor of species' sensitivity to both land-use and climate change across all vertebrates, thus indices reflecting
1695 habitat specialisation should be highly relevant to consider in large-scale vulnerability assessments, as done
1696 by Foden et al. (2013).

1697 This work further highlights the class-specific associations between most traits and likely responses to
1698 human-driven environmental changes, but again highlighting a non-random reshaping of vertebrate biodi-
1699 versity under global changes. In the case of land use, I find that the directionality of the responses not only

1700 often depends on taxonomic class but also on land-use type, further complicating the patterns. In line with
1701 past work highlighting the low explanatory power of traits when used to explain species' responses to human
1702 pressures (Angert et al., 2011; Cannistra and Buckley, 2021; Verberk et al., 2013), I found that most traits
1703 explained a small proportion of the interspecific variation in land-use responses and in climate-change sensi-
1704 tivity. The only exception was range area, which explained a large proportion of the interspecific variation in
1705 climate-change sensitivity. Given that the estimates of climate-change sensitivity were based on properties of
1706 species' climatic niches (Rinnan and Lawler, 2019), it was not surprising that range area explained such an
1707 important part of the interspecific variation in sensitivity, as it is likely that broader range areas are associated
1708 with broader climatic tolerances, and thus with lower estimated climate-change sensitivity on average.

1709 Despite their generally low explanatory power, traits have been used to assess species vulnerability to
1710 human threats, in particular to climate change (Böhm et al., 2016; Foden et al., 2013). One of the con-
1711 ceptual appeals behind the use of traits is that if clear patterns in responses to environmental change can
1712 be identified across taxa, then it could be possible to generalize their effects in space and time (Hamilton
1713 et al., 2020; Verberk et al., 2013), which is of interest for conservation -- notably for data-deficient species
1714 and/or those lacking estimates of abundance or population sizes. This work does not provide support for the
1715 generalisation at large scales of the effects of life-history and dietary traits on sensitivity to either land-use or
1716 climate change. The class-specific influence of traits on climate-change sensitivity, coupled with their low
1717 explanatory power, could be one of the reasons why trait-based approaches have been shown to perform less
1718 well and show less congruence than trend-based approaches (which rely on the use of long-term population
1719 data) for climate-change vulnerability assessments (Wheatley et al., 2017). Indeed, low explanatory power
1720 is likely to be particularly problematic for drawing inferences and predictions on species' responses to envi-
1721 ronmental change from traits. I would like to emphasize that this does not mean that life-history and dietary
1722 traits are unimportant for understanding species' responses; it means, however, that the effects of traits and
1723 their relative importance and predictive power could depend on interactions between the considered taxa and
1724 the pressures affecting them.

1725 Further, it is possible that general underlying patterns remain undetected or are being masked by inter-
1726 actions and trade-offs among traits, which I did not consider here. For instance, larger species tend to have
1727 larger dispersal distances and movement abilities (Jenkins et al., 2007), which could be beneficial to resource
1728 acquisition in disturbed areas (Hillaert et al., 2018); however, such species also tend to have higher energetic
1729 requirements (White, 2011) and lower reproductive output, which could be detrimental to their persistence in
1730 the face of environmental change. Interactions and trade-offs among traits have been shown to be important
1731 for understanding which species are likely to persist in disturbed environments (Sayol et al., 2020), but little

1732 is known about their potential effects at large scales and for different types of pressure. Thus, developments
1733 of my work could focus on the effects of trait interactions on species' sensitivity to climate change and
1734 land-use responses. I was also unable to consider intraspecific variation in this work. Intraspecific variation
1735 and the potential for acclimation and evolutionary adaptation are likely important determinants of species'
1736 ability to cope with human disturbance (Carlson et al., 2014; Rohr et al., 2018), but considering such effects
1737 at large scales is challenging because of the lack of available data.

1738 Moreover, I investigated climate-change sensitivity and land-use responses separately, meaning that I
1739 did not consider the combined effects of these pressures. However, human pressures act in combination
1740 (Capdevila et al., 2022a; Harfoot et al., 2021; Segan et al., 2016), and a number of confounding factors and
1741 threats that I could not account for could possibly influence species' sensitivity. For example, larger species
1742 might be more sensitive to warming than smaller species (Hantak et al., 2021; Merckx et al., 2018), but larger
1743 species could also be better able to persist in fragmented landscapes, such that habitat fragmentation and cli-
1744 mate warming may have opposite effects on the responses of such species. Further, larger species might also
1745 be disproportionately exposed to other threats such as overexploitation and human-wildlife conflicts (Rip-
1746 ple et al., 2014, 2017). Interactions among traits, among types of pressure, and among traits and pressures
1747 should ideally be considered together to understand species' responses to human disturbances (Hantak et al.,
1748 2021). However, considering all these effects simultaneously may be challenging because of data-limitation
1749 issues, model complexity, and difficulty in assessing and disentangling individual and interactive effects. In
1750 addition, I would like to emphasize that my results, based on correlative assessments of the associations be-
1751 tween ecological characteristics and species' sensitivity to climate change and species' land-use responses,
1752 do not allow to infer any causal links between traits and species' responses to global changes. Reinforcing
1753 our mechanistic understanding of how traits influence species' ability to cope with different disturbances
1754 may help understand opposite signatures of human impacts on vertebrate trait diversity (e.g., on body mass
1755 (Hantak et al., 2021; Merckx et al., 2018; Rapacciulo et al., 2017; Ripple et al., 2014, 2017)). Further work
1756 could help elucidate the mechanistic links between species traits and responses to environmental change,
1757 perhaps supported by long-term population data and demographic models (Hernández-Yáñez et al., 2022).

1758 To conclude, the results of this Chapter indicate that land-use and climate change are likely to impact
1759 terrestrial vertebrates non-randomly with respect to their ecological characteristics, which could have impor-
1760 tant consequences for ecosystem functioning (Duffy, 2003; Luck et al., 2012). I found that narrow-ranging
1761 species, species with smaller habitat breadth and species with natural habitat specialisation are disproportio-
1762 nately sensitive to both land-use and climate change. Further, I detected substantial declines in occur-
1763 rence probability of certain dietary groups in disturbed land-use types, most notably invertebrate eaters

1764 and fruit/nectars eaters in all classes. I also found higher climate-change sensitivity for invertebrate- and
1765 plant/seed-eating birds. My findings thus highlight the potential risks from global changes for ecosystem
1766 processes and services sustained by those species, such as pest control, seed dispersal or pollination (Civan-
1767 tos et al., 2012; Fricke et al., 2022; González-Varo et al., 2013), highlighting the need to put into place
1768 mitigation and conservation measures in the face of global changes.

¹⁷⁶⁹ 5 | Energetic constraints and trophic group ¹⁷⁷⁰ explain species persistence in disturbed ¹⁷⁷¹ land uses

¹⁷⁷² Keywords

¹⁷⁷³ Land use; land-use intensity; metabolic rates; energetic constraints; energetic requirements; terrestrial vertebrates; trophic group; occurrence.

¹⁷⁷⁵ Abstract

¹⁷⁷⁶ Land-use change is the primary driver of global biodiversity loss. In terrestrial vertebrates, previous work
¹⁷⁷⁷ has shown that sensitivity to land-use change depends on species traits, but the extent to which energetic
¹⁷⁷⁸ constraints explain species' responses to disturbed land uses remains largely unexplored. Here, I investigate
¹⁷⁷⁹ relationships between the energetic requirements of terrestrial vertebrates (estimated from resting metabolic
¹⁷⁸⁰ rates) and land-use change, at two levels of organisation. First, at the assemblage level I hypothesize that
¹⁷⁸¹ total energetic requirements in disturbed land uses are lower than in undisturbed land uses, assuming that
¹⁷⁸² there is less energy available in these areas overall. Second, after controlling for the effects of body mass and
¹⁷⁸³ taxonomy on metabolic rates, I predict that species with relatively lower energetic expenditure are favoured
¹⁷⁸⁴ over species with relatively higher energetic expenditure in disturbed land uses, as resource efficiency will
¹⁷⁸⁵ be beneficial in these resource-poor environments. Because trophic group influences species' ability to
¹⁷⁸⁶ assimilate various types of food, I investigate whether my predictions are consistent among trophic groups
¹⁷⁸⁷ (here, omnivores, carnivores or herbivores). The results challenged both hypotheses. I found that total
¹⁷⁸⁸ assemblage-level energetic requirements did not systematically decrease in disturbed land uses. For instance,

1789 I detected significant increases for urban areas in all trophic groups, highlighting that disturbed areas may not
1790 be as energy-poor as I initially assumed. Second, I found a positive effect of metabolic rates (after controlling
1791 for body mass and taxonomy) on species probability of occurrence across all trophic groups for at least one
1792 of the most disturbed land uses I considered (pasture, cropland and urban). Species for which there are
1793 exploitable resources in disturbed environments may benefit from having larger mass-independent energetic
1794 expenditure: they may display a set of characteristics rendering them more able to cope with disturbances
1795 and more able to acquire available resources, such as higher activity levels or bigger brain sizes. The findings
1796 of this Chapter highlight that land-use change has substantial impacts on vertebrate community metabolism.

1797 5.1 Introduction

1798 Land-use change is currently the strongest driver of global biodiversity declines (Maxwell et al., 2016;
1799 Newbold et al., 2015), with major and long-lasting impacts on the structure and functioning of ecological
1800 communities (Bregman et al., 2016; Fukasawa and Akasaka, 2019; Magioli et al., 2021; Marcacci et al.,
1801 2021). With land-use change likely to continue to intensify (Stehfest et al., 2019), it is vital to put into
1802 place conservation and mitigation measures to minimise future losses of biodiversity and negative impacts
1803 on ecosystem functioning. To this end, pressing questions remain as to what renders species able or unable
1804 to cope with human disturbance, and how losses of sensitive species might influence ecosystem functioning
1805 (Dirzo et al., 2014; Young et al., 2016).

1806 Land-use change acts as an environmental filter affecting species persistence (Edwards et al., 2021;
1807 Evans et al., 2018). Past studies have shown that sensitivity to land-use change is distributed unevenly across
1808 the tree of life (Nowakowski et al., 2018), and across behavioural (Lowry et al., 2013; Samia et al., 2015)
1809 and ecological strategies (De Palma et al., 2015; Møller, 2009; Newbold et al., 2013). For instance, long-
1810 lived and large forest specialist birds respond more negatively to land-use change than generalist species
1811 (Newbold et al., 2013). In addition, human land uses impose energetic constraints on species, by modifying
1812 the amount and type of available resources (Inger et al., 2016; Zuñiga-Palacios et al., 2021). Consequently,
1813 land-use change brings about modifications in dietary and foraging strategies (Møller, 2009; Ramesh et al.,
1814 2017; Sévèque et al., 2020), which can have cascading effects on local trophic structure (e.g., mesopredator
1815 release; Crooks and Soulé (2010)). The impacts of land-use change on biodiversity also depend on land-
1816 use intensity (Davison et al., 2021; Newbold et al., 2015), that is, the magnitude of human disturbance in
1817 a given land-use type, which can vary importantly according to local practices and management (Dullinger
1818 et al., 2021). In particular, land-use intensity affects the types and availability of resources (by homogenising

1819 resources for example), which in turn may affect biodiversity patterns (Weiner et al., 2011).

1820 Energetic requirements are likely to be important predictors of species' ability to cope with a disturbed
1821 environment where resource availability and resource types are strongly impacted (Auer et al., 2020). Past
1822 research has shown that metabolic rates (the amount of energy required for organismal maintenance and for
1823 other processes such as growth and reproduction) are heritable and subject to selection, for instance with
1824 species in energy-rich environments having evolved faster metabolic rates than species in less energy-rich
1825 environments (Mueller and Diamond, 2001). However, metabolic rates have received less attention than
1826 other life-history traits in studies aiming to understand different species' responses to land-use change. Body
1827 mass, known to explain much of the variation in metabolic rates (Bushuev et al., 2018; Hudson et al., 2013;
1828 White and Seymour, 2003), has been considered in past studies examining effects of land-use disturbance on
1829 assemblage composition (Hevia et al., 2017). Since metabolic rates correlate positively with body mass, the
1830 effects of land-use change on local energetic requirements have been indirectly assessed through investiga-
1831 tions of changes in body-mass composition along land-use gradients (Newbold et al., 2020a; Tinoco et al.,
1832 2018). However, to the best of my knowledge, no study has yet directly reported the effects of land-use
1833 change on assemblage-level energetic requirements.

1834 Thus, the first objective of this Chapter is to investigate how land-use change impacts assemblage-level
1835 energetic requirements in terrestrial vertebrates. To this end, I use metabolic rates as a proxy for maintenance
1836 energetic requirements at the species level. I predict that, at the assemblage level, total energetic requirements
1837 in disturbed land uses are lower than in undisturbed land uses. This prediction relies on the assumption that
1838 there is less energy available overall in disturbed land uses than in undisturbed land uses (Figure 5.1c), with
1839 disturbed land uses being characterised by human appropriation of net primary productivity (Krausmann et
1840 al., 2013). Because species' ability to exploit particular resources and meet energetic demands also depends
1841 on food types and diet (McNab, 1986; Mendoza and Araújo, 2019), I test whether this prediction holds
1842 true within trophic groups (i.e., within herbivores, omnivores, and carnivores). Here, I consider energetic
1843 requirements at the assemblage level, not controlling for species' body mass, to obtain estimates of total
1844 energy expenditure in different land-use types (Figure 5.2a). Thus, any changes in assemblage-level energetic
1845 requirements could be ultimately driven by shifts in the size-spectrum of local assemblages.

1846 Body mass explains much of the variation in metabolic rates, but species with similar body masses still
1847 display important variation in metabolic rates (Mueller and Diamond, 2001). Some of this variation can
1848 be explained by taxonomic position (which also accounts for differences in thermoregulatory strategy be-
1849 tween endotherms and ectotherms). However, there remains substantial interspecific variation in metabolic
1850 rates even after accounting for body mass and taxonomic affiliation (Mueller and Diamond, 2001; White

1851 and Kearney, 2013). Food availability and food type might explain this residual variation in metabolic rates
1852 among species (Mueller and Diamond, 2001). Species that evolved in areas with abundant resources assim-
1853 ilate and burn calories at high rates, regardless of their body mass – while species that evolved with limiting
1854 resources could process resources at lower rates, irrespectively of their body mass (Mueller and Diamond,
1855 2001). Building upon this idea and the ‘allocation’ principle (i.e., the trade-offs in resource allocation; Auer
1856 et al. (2020)), species with lower energetic expenditure (than expected from body mass and taxonomy) may
1857 be able to cope with food scarcity better than those with larger energy expenditure (than expected from
1858 body mass and taxonomy). Indeed, regardless of their body mass, such species should need fewer resources
1859 to meet energetic demands (Clarke and Fraser, 2004). Thus, as land-use disturbance modifies the types and
1860 abundance of resources, I expect species energetic requirements –after removing the effects of body mass and
1861 taxonomic position– to influence responses to land-use change. Yet, whether energetic constraints explain
1862 species’ responses to land-use change in terrestrial vertebrates has not been investigated before (Hevia et al.,
1863 2017). Thus, the second objective of this Chapter is to characterise the influence of energetic constraints
1864 on species occurrence probability in disturbed land uses, compared to undisturbed land uses. I approximate
1865 energetic constraints at the species level from residual variation in metabolic rates, that is, the variation not
1866 explained by body mass or taxonomy. I predict that species with negative deviations (lower rates than ex-
1867 pected from body mass and taxonomy) are more likely to persist in disturbed land uses than species with
1868 positive deviations (higher rates than expected from body mass and taxonomy; Figure 5.2b). Given that
1869 trophic group can influence resource acquisition and species’ ability to assimilate various types of food, I
1870 investigate whether this prediction is consistent among trophic groups.

1871 To test both predictions, I use a space-for-time substitution approach, obtaining vertebrate assemblage
1872 composition data from the PREDICTS database (Hudson et al., 2014, 2017). I combine this database with
1873 trophic-group information (characterising species as either omnivores, herbivores, or carnivores), and with
1874 species-level estimates of resting metabolic rates and body mass.

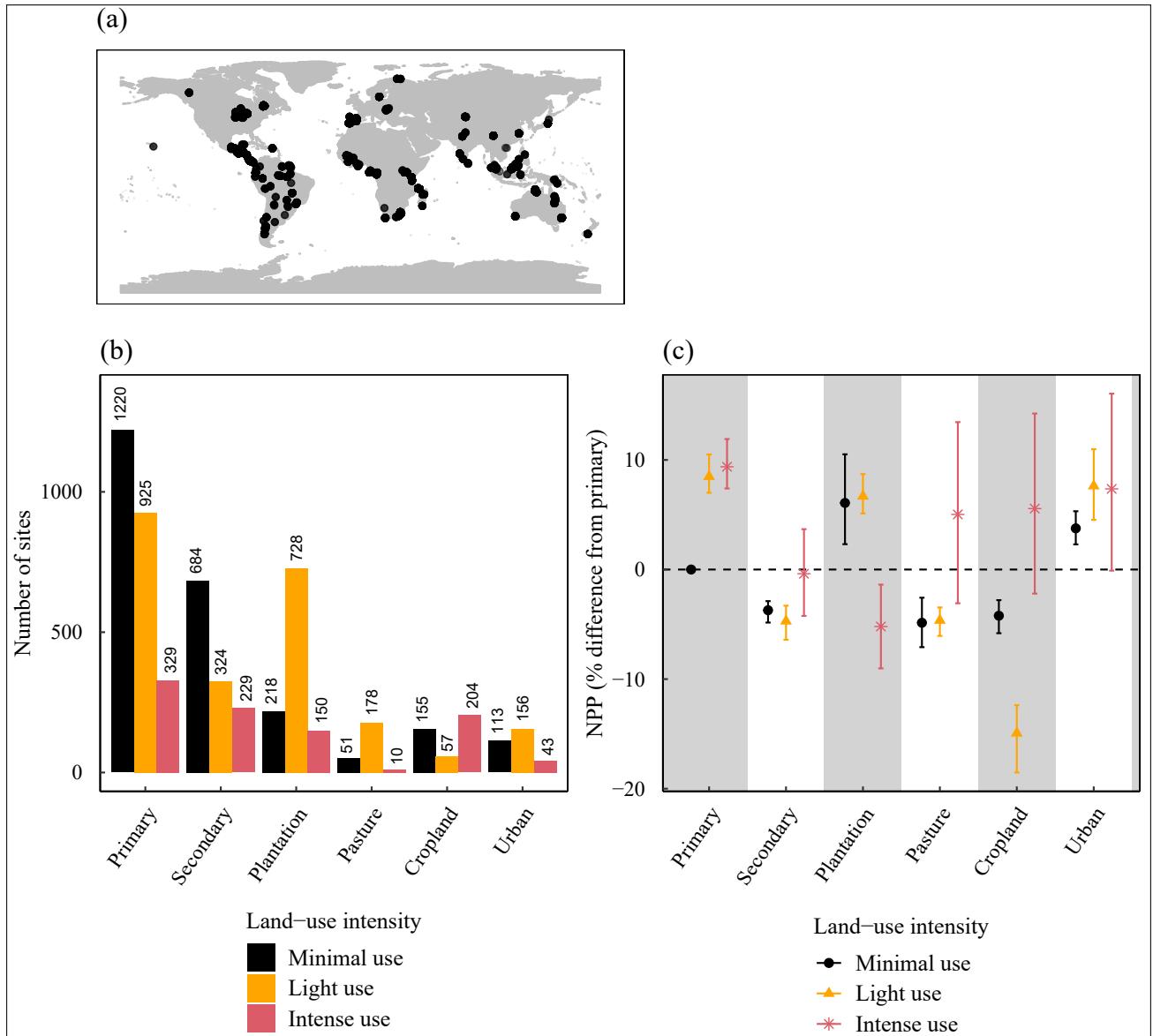
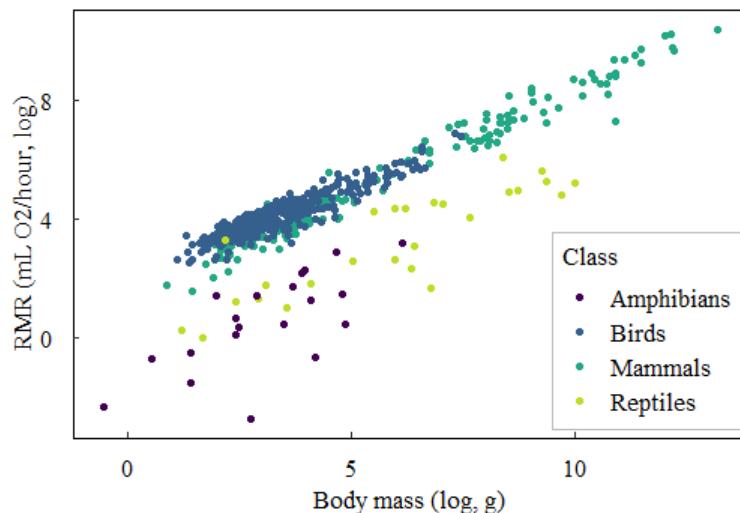


Figure 5.1: (a) Spatial distribution of the sampled sites from the PREDICTS database for terrestrial vertebrates (5,774 sites); (b) Number of sites in each land-use and land-use-intensity category; (c) Net primary productivity by land use and land-use intensity across PREDICTS sites (derived from MODIS satellite imagery), with model predictions plotted relative to minimally used primary vegetation (and rescaled with reference to that land-use type). Primary: primary vegetation; secondary: secondary vegetation; plantation: plantation forest.

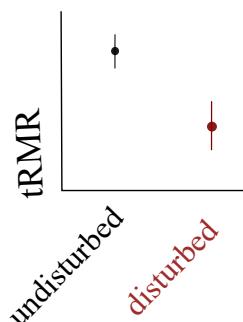
(a) Assemblage-level: there is less energy available to utilize by vertebrate assemblages in disturbed land uses than in undisturbed land uses.



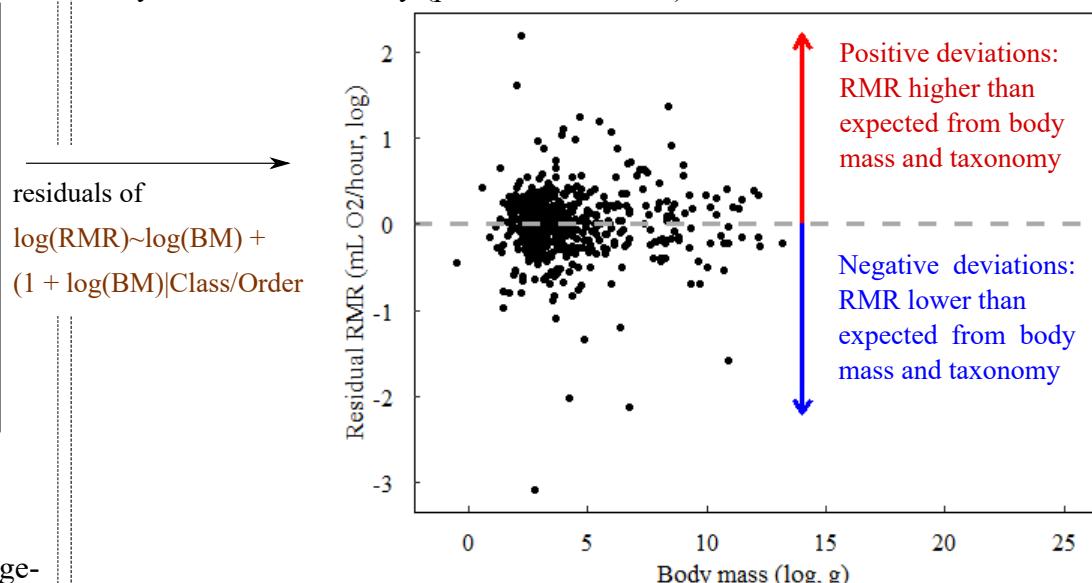
Prediction 1: within all trophic groups, total assemblage-level **mass-dependent** metabolic rates decrease in disturbed land uses compared to undisturbed land uses.

$$\text{Assemblage level:} \quad t\text{RMR} = \sum_i (\text{RMR}_i * a_i)$$

$$\text{Across assemblages - model 1:} \quad \log(t\text{RMR}) \sim \text{LU} + \text{LUI} + \text{TG} + \text{LU:LUI} + \text{LU:TG} + \text{LUI:TG} + \text{LU:LUI:TG} + RE$$



(b) Species-level: species with lower energetic expenditure than expected from body mass and taxonomy (negative deviations) do better in disturbed land uses than species with higher energetic expenditure than expected from body mass and taxonomy (positive deviations).



Prediction 2: within all trophic groups, species occurrence probability in disturbed land uses is negatively affected by **residual RMR**. The slope of the relationship between occurrence probability and residual RMR is significantly lower than the slope estimate for the undisturbed land uses.

Across species - model 2:

$$\begin{aligned} P_{\text{occurrence}} \sim & \text{LU} + \text{LUI} + \text{resRMR} + \text{TG} + \\ & \text{LU:LUI} + \text{LU:TG} + \text{LU:resRMR} + \\ & \text{LUI:TG} + \text{LUI:resRMR} + \text{TG:resRMR} + \\ & \text{LU:TG:resRMR} + \text{LUI:TG:resRMR} + RE \end{aligned}$$

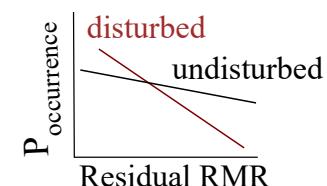


Figure 5.2: Framework for the predictions and models. I use resting metabolic rates (RMR) as a proxy for the amount of energy processed by vertebrate species. **(a)** I hypothesize that there is less energy overall that can be utilized by vertebrate species in disturbed land uses compared to undisturbed land uses. As such, I expect decreases in assemblage-level total metabolic rates ($t\text{RMR}$). $t\text{RMR}$ is calculated as the sum, over all species occurring at a site, of species RMR (RMR_i) weighted by species abundance (a_i). **(b)** After removing the effects of body mass and taxonomy, I expect species with lower energy expenditure than expected to do better than species with higher energy expenditure than expected. I predict that the slope of the relationship between residual RMR and occurrence probability is more negative in disturbed land uses than in undisturbed land uses. For both (a) and (b), I test whether the predictions are consistent among trophic groups. LU: Land use; LUI: land-use intensity; TG: trophic group; RE: random effects. In *model 1*, random effects include the identity of the PREDICTS studies from which the assemblages are derived. In *model 2*, random effects include study, site, and species identity.

1875 **5.2 Methods**

1876 **5.2.1 Vertebrate assemblage composition**

1877 I obtained vertebrate assemblage composition in different land uses from the PREDICTS database (Hudson
1878 et al., 2014, 2017). The PREDICTS database is a large collection of published studies that measure bio-
1879 diversity across different land uses and is one of the most comprehensive global databases of its type. In
1880 each PREDICTS study, species occurrence and often abundance were recorded across different sites. Each
1881 site was assigned to one of the following land-use types: primary vegetation (natural, undisturbed vegeta-
1882 tion), secondary vegetation (recovering after complete destruction of primary vegetation), plantation forest
1883 (woody crops), pasture (areas grazed by livestock), cropland (herbaceous crops) and urban (built-up areas).
1884 The land-use categories were assigned based on habitat descriptions from the original studies (Hudson et al.,
1885 2014), sometimes in consultation with the original study authors. Each site was also classified in terms of
1886 land-use intensity as either minimal, light or intense. The land-use-intensity assignment was also made on
1887 the basis of the habitat description in the original studies, and depended on criteria specific to each land use
1888 (such as degree of mechanisation, yield or chemical inputs for cropland; or the amount of green space in
1889 urban areas; Hudson et al. (2014)).

1890 I subset the PREDICTS database for studies that sampled terrestrial vertebrates, and for which both land
1891 use and land-use intensity had been characterised. I further subset the database for species for which I could
1892 obtain an estimate of resting metabolic rates (see below). I thus obtained 154 studies for 529 species sampled
1893 across 5,774 sites (Figure 5.1a). Sample sizes varied across land uses and land-use intensities (Figure 5.1b).

1894 **5.2.2 Energy availability by land-use type and land-use intensity**

1895 The predictions of this Chapter rely on the assumption that resource types and abundance are modified in
1896 disturbed environments, with less energy available in disturbed compared to undisturbed land uses over-
1897 all. To test this assumption, I used terrestrial net primary productivity (NPP) across land uses as a proxy
1898 for available energy. NPP quantifies the amount of atmospheric carbon fixed by plants and accumulated
1899 as biomass. NPP estimates were derived by Meghan Hayden, using imagery from the Moderate Resolu-
1900 tion Imaging Spectroradiometer (MODIS) on board NASA's Terra satellite. NPP estimates were based on
1901 a yearly composite of measures made at 8-day intervals, captured at 500-m spatial resolution (Running and
1902 Zhao, 2015). Meghan Hayden obtained NPP for 4,062 of the PREDICTS sites (matching the sites to the NPP
1903 data using the sampling year available in PREDICTS). I fit a linear mixed-effects model ('lme4' package,

1904 version 1.1-23, Bates et al. (2015)) explaining site-level NPP by land use and land-use intensity, with a ran-
1905 dom intercept accounting for study identity, to control for differences in experimental design across studies.
1906 Model predictions showed that NPP decreased significantly in several land uses (e.g., pasture and cropland)
1907 compared with the primary vegetation reference level, although the strength and in some cases direction of
1908 the difference varied among land-use and land-use intensity combinations (e.g., increases in urban land uses;
1909 Figure 5.1c).

1910 **5.2.3 Resting Metabolic Rates (RMR)**

1911 As a proxy for species-level energetic expenditure, I used estimates of the minimum amount of energy re-
1912 quired for organismal maintenance, i.e., basal metabolic rates (BMR) for endotherms, and resting metabolic
1913 rates (RMR) for ectotherms. From the literature, I obtained estimates of BMR for 719 species of birds and
1914 685 mammals, and estimates of RMR for 126 amphibians and 173 reptiles (Appendix 4, Table S5.1). For
1915 endotherms, BMR are measured when species are in their thermoneutral zone, that is, when there is little to
1916 no energy expenditure allocated to thermoregulation. Thus, BMR estimates were derived from lab studies
1917 that mostly measured oxygen consumption of the organisms at rest under controlled conditions and in the
1918 thermoneutral zone of the species. For an ectotherm, there is no ‘basal’ metabolic rate, as body tempera-
1919 ture mainly depends on environmental temperature. Their metabolic rates follow a hump-shaped relationship
1920 with environmental temperature, highest at an optimal temperature which corresponds to a performance peak.
1921 To be able to compare endotherms’ BMR with ectotherms’ RMR, Stark et al. (2020) used the metabolic rates
1922 that correspond to a performance peak for both groups (i.e., BMR in the thermoneutral zone for endotherms,
1923 and metabolic rates at optimal temperature for ectotherms). Thus, I used the data compiled in Stark et al.
1924 (2020) for ectotherms, and from the sources specified in Table S5.1 for endotherms. The units for BMR
1925 and RMR were standardized to mL of dioxygen consumed per hour (mLO₂/h). As in Stark et al. (2020), I
1926 henceforth refer to both basal and resting metabolic rates as RMR. I then matched the RMR data with the
1927 PREDICTS data, obtaining occurrence records for 529 species with known RMR values.

1928 **5.2.4 Trophic group and body mass information**

1929 I used body mass and trophic group information for terrestrial vertebrates compiled in Chapter 2. Body
1930 mass was compiled as a single measure at the species level, meaning I was unable to consider intraspecific
1931 variation. Trophic group described species as either carnivores, omnivores, or herbivores. Because there
1932 were gaps in the availability of the data, more so for trophic group than for body mass (Chapter 2), I imputed
1933 the missing trait values. To impute missing body mass and trophic groups, I used random forests algorithms

1934 (using the ‘missForest’ R package, *()*), including as additional predictors phylogenetic information, added
1935 in the form of 10 phylogenetic eigenvectors (Diniz-Filho et al., 2012) following Penone et al. (2014), and
1936 also taxonomic order. I considered a wider set of life-history traits in the missing values imputations: lifes-
1937 pan, litter/clutch size, habitat breadth and use of artificial habitats (compiled in Chapter 2). Phylogenetic
1938 eigenvectors were extracted from the class-specific phylogenies using the ‘PVR’ package (Santos, 2018).

1939 **5.2.5 Effects of land use, land-use intensity and trophic group on assemblage-level total
1940 RMR (prediction 1; Figure 5.2a)**

1941 Here, I subset the PREDICTS database for species for which local abundance at the sites was recorded. The
1942 subset contained 459 species sampled across 108 studies in 2,592 sites. Assemblage-level total RMR (tRMR)
1943 was then calculated by summing abundance-weighted RMR for the species occurring in each site. I fitted
1944 a linear mixed-effects model to explain \log_e -tRMR as a function of land use, land-use intensity and trophic
1945 group, with a random intercept accounting for study identity to control for differences in experimental design
1946 across studies. I started with a model allowing all two-way interactions among the predictors. I then tested
1947 whether adding the three-way interaction among land use, land-use intensity and trophic group improved the
1948 fit of the model, using a likelihood-ratio test. The model that included the three-way interaction was retained
1949 ($P \ll 0.01$; *model 1*, Figure 5.2). In addition, because it is well established that resting metabolic rates are
1950 influenced by temperature (Clarke and Fraser, 2004), I checked whether including annual mean temperature
1951 in the model affected the conclusions. Annual mean temperature at each PREDICTS site was estimated from
1952 WorldClim version 2.1 (Fick and Hijmans, 2017), using a 2.5 arc-minute resolution. Adding annual mean
1953 temperature did not improve model fit (likelihood-ratio test: $P=0.08$), thus I did not consider its effects any
1954 further.

1955 **Disentangling the effects of body mass and abundance on tRMR.**

1956 Since RMR correlates strongly with body mass, changes in tRMR are likely to be driven in part by changes
1957 in the size-spectrum of ecological assemblages. I fitted an additional model to explain changes in species’
1958 abundance (given presence) by land use, land-use intensity, trophic group, body mass and their interactions,
1959 to understand the role of shifts in the body mass of species on observed changes in tRMR (Appendix 4,
1960 Figure S5.1).

1961 **5.2.6 Effects of land use, land-use intensity, trophic group and residual RMR on species**
1962 **occurrence probability (prediction 2; Figure 5.2b)**

1963 To control for the effects of body mass and taxonomy on RMR, I used the residual variation in RMR after
1964 accounting for these variables, from a linear mixed-effects model fitting \log_e -RMR as a function of \log_e -body
1965 mass with nested random taxonomic effects (random intercepts and slopes: $1 + \log(\text{body mass})|\text{Class}/\text{Order}$;
1966 Figure 5.2). Hence, I used a metric that describes how much more energy (positive deviations) or less energy
1967 (negative deviations) than expected from body mass and taxonomic position a species spends for organismal
1968 maintenance. Similar approaches have been used in previous papers (Furness and Speakman, 2008; Naya
1969 et al., 2013). As detailed earlier, I expect species with lower residual RMR to do better in disturbed land uses
1970 than species with higher residual RMR (prediction 2; Figure 5.2b) because, given any body mass, investing
1971 less energy in maintenance could contribute to persistence in a context of resource scarcity.

1972 To test the second prediction, I fitted a binomial mixed-effects model explaining species occurrence
1973 with land use, land-use intensity, trophic group and residual RMR. I started with a complete model that
1974 included all two-way interactions among the main effects. Because I wanted to test whether the second
1975 prediction was valid for each trophic group, I needed to account for potential differences in the slope of the
1976 relationships between occurrence probability and residual RMR among trophic groups. Thus, I performed
1977 a forward stepwise selection procedure to test whether adding three-way interactions among (1) land use,
1978 trophic group and residual RMR and (2) among land-use intensity, trophic group and residual RMR improved
1979 model fit, using likelihood-ratio tests. The final model included both three-way interactions (Figure 5.2b;
1980 *model 2*). I fitted random effects that accounted for species identity, as well as for study and site identity
1981 within PREDICTS.

1982 **Model validation.**

1983 I checked the phylogenetic signal in the model residuals using Pagel's λ (Pagel, 1999). Non-significant
1984 phylogenetic signal in the residuals would indicate that fitting species identity in the model's random effects
1985 was sufficient to account for residual phylogenetic variation in RMR.

1986 5.3 Results

1987 5.3.1 Effects of land use, land-use intensity and trophic group on assemblage-level total 1988 RMR

1989 Land use, land-use intensity, trophic group and their interactions had significant effects on assemblage-level
1990 total RMR (Figure 5.3). Overall, and contrary to my expectations, assemblage-level total RMR did not
1991 show systematic decreases in disturbed land uses. In fact, urban land uses were associated with strong
1992 significant increases in tRMR in all trophic groups (e.g., a 200% average increase in tRMR in lightly-used
1993 urban areas for carnivores, compared with primary vegetation levels; +207% on average in lightly-used urban
1994 areas for herbivores; +107% for minimally-used urban areas for omnivores). In other land uses, responses
1995 depended on trophic group and land-use intensity. Whilst for herbivores, disturbed land uses were typically
1996 associated with increases in tRMR, I detected decreases in tRMR for omnivores and carnivores in several
1997 land uses, most notably in intensely-used pasture for carnivores (-84%). Such effects could reflect changes
1998 in the size-spectrum of local assemblages (Appendix 4, Figure S5.2). For instance, in minimally-used urban
1999 areas, larger herbivores tended to occur at higher abundances compared to primary vegetation level; and in
2000 intensely-used pastures, carnivores tended to occur at lower abundances overall (Figure S5.2).

2001 The model residuals were appropriately distributed (see diagnostic plots in Appendix 4, Figure S5.3).
2002 Investigating the sensitivity of the results to imputation uncertainty showed that the conclusions were robust
2003 to the removal of all imputed estimates of RMR (the correlation coefficient was 0.72 between the two sets of
2004 model coefficients; Figure S5.4). Comparing model predictions showed that effects were mostly congruent,
2005 although there were some differences (Figure S5.5). In particular, for herbivores, effect sizes tended to be
2006 bigger for the model fitted on empirical data compared with the model that included imputed data. Thus, the
2007 main results appear to be conservative if anything. The model fitted on empirical data had larger standard
2008 errors, likely due to the reduction in sample size.

2009 5.3.2 Effects of land use, land-use intensity, trophic group and residual RMR on species' 2010 occurrence probability

2011 Species' occurrence probability was significantly affected by land use, land-use intensity, trophic group,
2012 residual RMR and their interactions (Figures 5.4, 5.5). Contrary to my expectations, species with higher
2013 residual RMR (relative to their body mass and taxonomic position) tended to do better than species with lower
2014 residual RMR in a number of disturbed land uses. Overall, land-use type was more important for determining

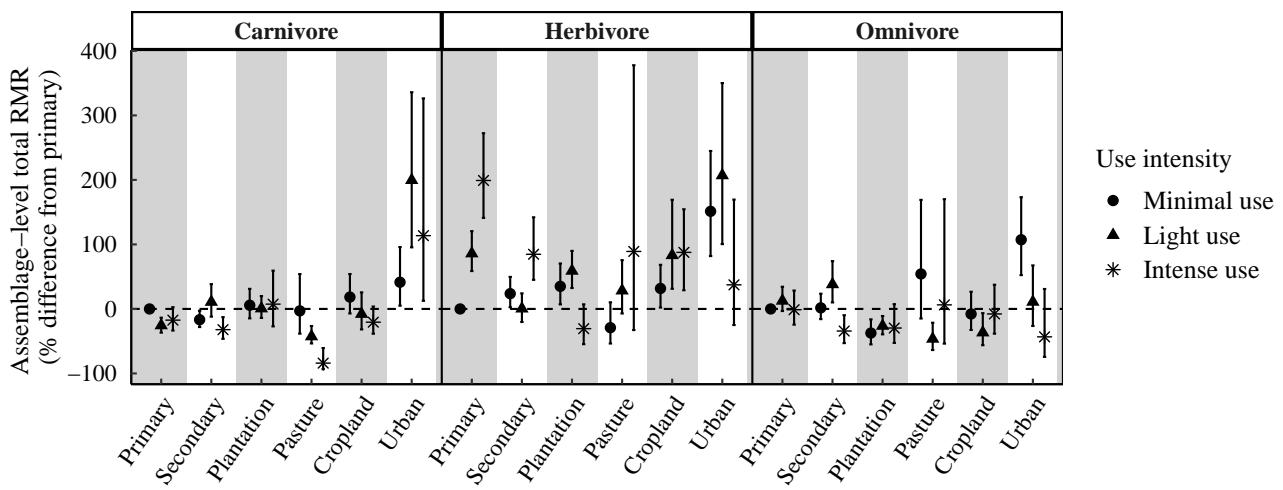
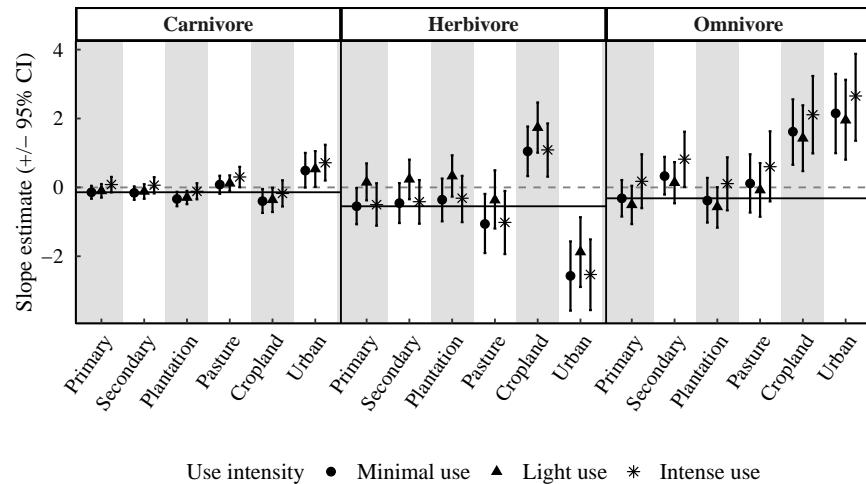


Figure 5.3: Effects of land use, land-use intensity and trophic group on assemblage-level total RMR (mean estimate $\pm 95\%$ confidence interval). Model predictions are rescaled with reference to minimally-used primary vegetation, considered to be the undisturbed baseline. Primary: primary vegetation; secondary: secondary vegetation; plantation: plantation forest.

2015 the relationship between occurrence probability and residual RMR than land-use intensity (Figure 5.4a).

(a) Slope of the relationships between occurrence probability (log-odds) and residual RMR



(b) Predicted effects of residual RMR on occurrence probability (minimal use intensity)

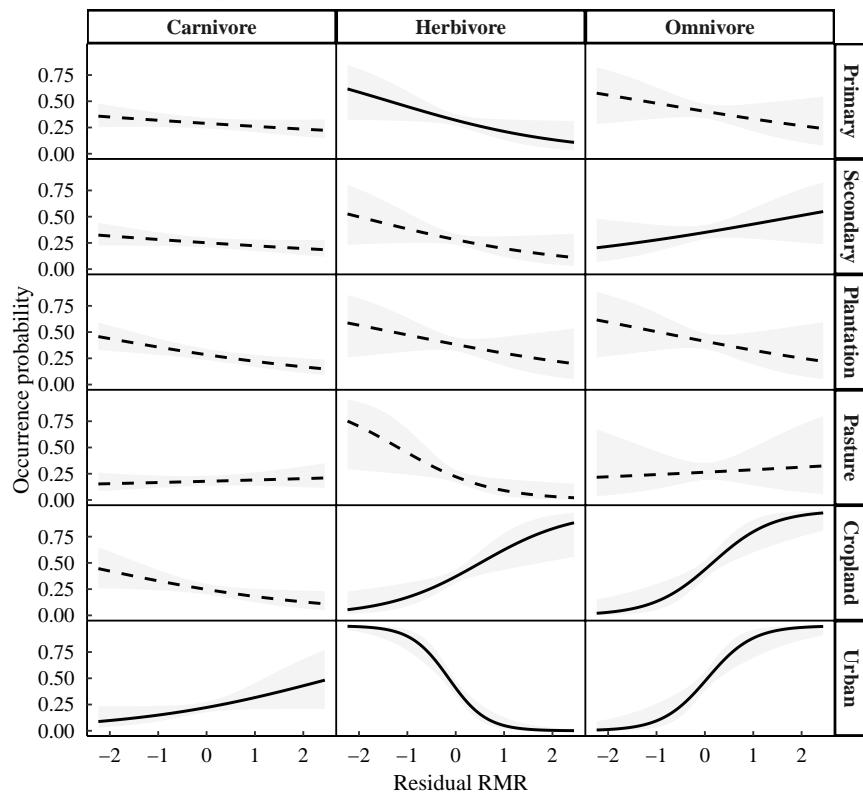


Figure 5.4: (a) Slope estimates for the relationship between residual RMR and occurrence probability in each land-use type and for the three levels of land-use intensity (mean estimate $\pm 95\%$ confidence interval). The black horizontal line indicates the slope for the reference level (primary vegetation) for minimal land-use intensity. The grey dashed line marks 0. (b) Effect of residual RMR on species probability of occurrence within each trophic group and for each land-use type. I plotted the predictions for minimal land-use intensity only. Solid lines represent significant relationships (with slopes significantly different from 0), and dashed lines are plotted where the slope was not significantly different from 0. Shaded areas represent 95% confidence intervals. Primary: primary vegetation; secondary: secondary vegetation; plantation: plantation forest.

2016 For minimally-used primary vegetation (reference), residual RMR tended to be negatively associated
 2017 with species occurrence probability in all trophic groups (but with a significant slope for herbivores only;
 2018 Figure 5.4a). However, the directionality of this relationship was reversed in some disturbed land uses in
 2019 all trophic groups (secondary vegetation, cropland and urban for omnivores; cropland for herbivores; urban
 2020 for carnivores), with significant positive slopes, also significantly higher than those observed for primary
 2021 vegetation (Figure 5.4a); in these land uses, residual RMR was thus positively associated with occurrence
 2022 probability. One notable exception was the opposite pattern for urban herbivores (Figure 5.4b), where resid-
 2023 ual RMR had a more negative effect on occurrence probability than in minimally-used primary vegetation.

2024 I would like to emphasize that positive effects of residual RMR on occurrence probability in some of the
 2025 most disturbed land uses (e.g., urban for carnivores) do not mean that there were absolute increases in species
 2026 occurrence probability in disturbed land uses compared to primary vegetation (and vice-versa). I illustrate
 2027 this point in Figure 5.5. For carnivores with a median value for residual RMR, occurrence probability was
 2028 reduced by an average 24% in urban land uses compared to primary vegetation (but there was a significant
 2029 positive association between residual RMR and occurrence probability for carnivores in urban areas; Figure
 2030 5.4).

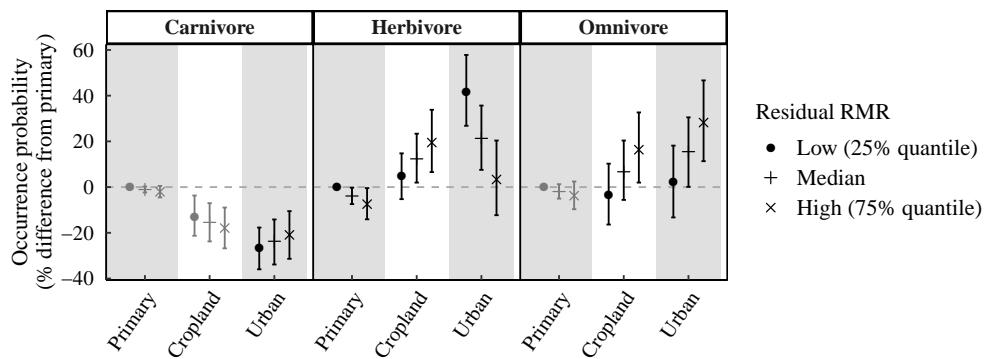


Figure 5.5: Predicted occurrence probabilities (mean effect \pm 95% confidence interval) in primary vegetation (primary) and for the most disturbed land uses where residual RMR was found to have a significant effect on occurrence probability. For visualisation purposes, I discretised residual RMR in three levels. The predicted probabilities of occurrence were rescaled with reference to primary vegetation for the lowest value of residual RMR (25% quantile). Here, the predictions are plotted for minimal land-use intensity (effects would be similar for light and intense land-use intensities). Black points and error bars are plotted where the relationship between occurrence probability and residual RMR was significant (and dark grey points and error bars represent non-significant trends).

2031 Finally, the model showed some degree of deviation from distributional assumptions (diagnostic plots,
 2032 Appendix 4, Figure S5.6). Nevertheless, the model's coefficients were similar when estimated with a
 2033 Bayesian framework, suggesting that the estimates were robust (Figure S5.7). The phylogenetic signal in
 2034 the model residuals was weak and non-significant ($\lambda=0.004$, $P=0.24$). Re-fitting the model using the com-

2035 plete data subset (i.e., excluding imputed RMR estimates) showed that the conclusions are likely robust to
2036 imputation uncertainty (Figure S5.8), with congruent results overall, although there were a few differences
2037 in the predictions between the two models – notably, for herbivores in urban land uses (Figure S5.8).

2038 **5.4 Discussion**

2039 The results of this Chapter provide insights into the relationship between land-use change and energetic
2040 requirements at two levels of organisation (at the assemblage level and the species level), and contradict
2041 both my initial predictions. I found that total energetic requirements did not show systematic decreases in
2042 disturbed land uses. On the contrary, there were strong increases in all trophic groups, most notably in urban
2043 land uses. Further, in all trophic groups, species with higher energetic expenditure (than expected from body
2044 mass and taxonomy) tended to do better in some of the most disturbed land uses (that is, cropland and urban)
2045 than species with lower energetic expenditure (than expected from body mass and taxonomy).

2046 At the assemblage level, the results first highlight the effects of land-use change on vertebrate com-
2047 munity metabolism. Contrary to my expectations, I found differing effects of land use on total energetic
2048 requirements among trophic groups, reflecting changes in the size-spectrum of ecological assemblages (i.e.,
2049 changes in the distribution of abundance along the body-mass gradient). On the one hand, decreases in total
2050 energetic requirements, such as observed for carnivores in intensely used pastures, are likely due to overall
2051 reductions in local abundance (with carnivorous species in pastures perceived as a threat to livestock (Eeden
2052 et al., 2018); or because of increased competition for fewer resources). On the other hand, increases in total
2053 energetic requirements could reflect higher levels of resource intake in some disturbed land-use types. On the
2054 basis of net primary productivity, I hypothesized that there would be fewer resources available in disturbed
2055 areas. However, it could be that the carrying capacity of some disturbed land-use types actually increases
2056 for some trophic groups compared to that of primary vegetation, because of the presence of novel or more
2057 abundant food sources. For instance, in urban areas, both wildlife feeding and the presence of anthropogenic
2058 food sources, such as human refuse or pet food, could lead to an increase in resource availability all year-
2059 round for species with a non-specialised diet that are able to exploit such food sources (Fischer et al., 2012).
2060 Past research on urban carnivores has shown that some species have adapted to urban environments by ex-
2061 ploiting anthropogenic food sources, and also benefit from physical protection (with some human structures
2062 providing shelter) and from reduced occurrence of natural enemies (Bateman and Fleming, 2012). Herbi-
2063 vores and omnivores could also benefit from increased resources in urban areas, notably where the urban
2064 matrix includes semi-natural habitats, which is congruent with observed increases in occurrence probability

2065 in minimally used urban areas for these trophic groups. Further, anthropogenic ‘bottom-up food forcing’ has
2066 been shown to affect food-web dynamics, with decreases in predation rates, loss of anti-predator abilities
2067 (Geffroy et al., 2020) and mesopredator release (Fischer et al., 2012). Thus, in urban areas, increases in
2068 anthropogenic food sources could provide support for a higher abundance of medium-sized species released
2069 from predation, increasing total energetic requirements.

2070 I hypothesized that resource availability would be the main driver behind changes in energetic require-
2071 ments. However, resource availability likely interacts with other factors to explain the patterns. In particular,
2072 past work has shown that brain size is an important determinant of species’ ability to cope with disturbance,
2073 notably in urban settings (Sayol et al., 2020). Since brain size scales allometrically with body mass, it is
2074 possible that I observe increases in total energetic requirements where having larger brains is advantageous
2075 for resource acquisition, and in turn where resources are both exploitable and abundant enough to sustain
2076 the larger energetic expenditures associated with bigger brain sizes. In addition, body size, which explains
2077 an important proportion of the interspecific variation in mobility, likely interacts with characteristics of the
2078 landscape (such as fragmentation) with consequences for species persistence in disturbed land uses (Merckx
2079 et al., 2018). Hence, the patterns I observe in this Chapter could emerge from interactions and trade-offs
2080 among resource availability, diet, body size and human management of the landscape.

2081 At the species level, the results underline the role of energetic constraints on species’ responses to land-
2082 use change. After controlling for the effects of body mass and taxonomy, I found that residual energetic
2083 expenditure was significantly associated with species occurrence probability in disturbed land uses. Con-
2084 trary to my expectations, in several disturbed land uses and in all trophic groups, species with higher residual
2085 energetic expenditure tended to have a higher occurrence probability than species with lower residual ener-
2086 getic expenditure. It could be that species with lower residual energetic requirements are less well equipped
2087 than species with higher residual energetic requirements at making use of the available food sources in dis-
2088 turbed land uses. Species in disturbed land uses may need to display higher levels of feeding innovation
2089 (Coogan et al., 2018), have larger brains (Sayol et al., 2020), or be bolder and more active to make use of
2090 the available resources. In mammals, past research has shown that larger brains are associated with larger
2091 residual energetic expenditure (Isler and Van Schaik, 2006). At the individual level, past research suggests
2092 that metabolic rates are linked to differences in behaviour, with bolder and more active individuals exhibiting
2093 higher metabolic rates than less active individuals, with consequences for food acquisition (Biro and Stamps,
2094 2010). Although I am not aware of similar evidence at the species level, I propose that residual metabolic
2095 rates interact with species’ ecological traits, behavioural traits and foraging strategy in influencing species’
2096 responses to land use. Among species with an adaptable diet, able to make use of the resources available

2097 in a disturbed landscape, those with higher residual metabolic rates could present a set of behavioural char-
2098 acteristics that render them better at acquiring the available resources, hence be more able to cope with
2099 land-use disturbance. This could be the case for urban carnivores and omnivores, as well as herbivores in
2100 cropland. Thus, it is possible that disturbed land uses favour species with higher residual resting metabolic
2101 rates because such species are overall better competitors when faced with disturbance. In addition, release
2102 from predation in some disturbed land uses – most notably in urban areas – could favour bolder species, as
2103 they become less exposed to predation risks. Conversely, and in accordance with my initial hypothesis, the
2104 results suggest that when there are no or few exploitable resources in disturbed areas, spending less energy
2105 than expected from body mass and taxonomy might be beneficial for persistence (this could be the case for
2106 urban herbivores, which include species specialised on fruit, nectar, and other plant materials; some of these
2107 resources likely become less abundant in disturbed areas).

2108 To summarise, I propose that, for species with adaptable diets, increases in exploitable resources cou-
2109 pled with decreases in predation rates could lead to increases in overall energetic expenditure, associated
2110 with shifts towards assemblages composed of a higher abundance of medium-sized, bolder, more active and
2111 larger-brained species. On the other hand, species with specialised diets might be better able to persist and
2112 cope with resource scarcity when their energetic expenditure is lower than expected from body mass. These
2113 ideas could be further tested by considering more refined dietary groups (as those compiled in Chapter 4)
2114 rather than trophic groups, as diet should be easier to link with resource availability in a given land-use type.
2115 This could constitute a development for this work.

2116 Finally, I would like to emphasize that energetic budget allocation is a fundamental aspect of organismal
2117 fitness (Burger et al., 2021). Past studies suggest that individuals exposed to food scarcity can lower their
2118 energetic expenditure to increase their chances of survival. For instance, Zhang et al. (2018b) showed that,
2119 subjected to food restrictions, the Chinese bulbul (*Pycnonotus sinensis*) was able to lower its basal metabolic
2120 rate not only passively through body and organ mass reductions, but also by modifying enzymatic activity
2121 and metabolism in the muscles and liver. Other studies have also found that bird nestlings downregulate
2122 energetic costs allocated to maintenance when exposed to food shortage (Brzék and Konarzewski, 2001;
2123 Moe et al., 2004), or to disturbances such as urban noise (Brischoux et al., 2017). Intraspecific variation
2124 in energetic expenditure could affect individual responses to land-use change, which in turn could affect
2125 species' ability to adapt to human pressures. However, the lack of spatially explicit estimates of metabolic
2126 rates precluded its consideration in this Chapter.

2127 To conclude, the findings of this Chapter suggest important effects of land-use change on energetic
2128 fluxes, and thus potentially on ecosystem functioning. Further research is warranted as to the effects of these

2129 energetic patterns for ecosystem processes. Indeed, assemblage-level energy fluxes may serve as important
2130 indicators of change for ecosystem processes, such as decomposition (Barnes et al., 2014). Understanding
2131 these impacts is particularly important given increasing land-use change and biodiversity loss across the
2132 globe.

2133 6 | General discussion

2134 In recent centuries, large-scale anthropogenic modifications to the Earth's systems have accelerated and
2135 reached unprecedented levels (Steffen et al., 2015). Two of the major signatures of human impacts on the
2136 Earth's systems are the transformation of the land surface (Ellis et al., 2010), notably fuelled by the rising de-
2137 mand for agricultural commodities, and anthropogenic climate change onset by human-driven modifications
2138 to atmospheric composition (Lewis and Maslin, 2015). Such transformations have had important impacts
2139 on the world's biota. By modifying species' habitats at large scales and altering local climatic conditions
2140 at rates exceeding natural variability, human changes have put biodiversity under pressure (Maxwell et al.,
2141 2016). Empirical evidence showing the extent and magnitude of human impacts on biodiversity has been ac-
2142 cumulating (Daru et al., 2021; Newbold et al., 2015; Young et al., 2016). In particular, land-use and climate
2143 change have already driven declines in species richness and abundance, and altered species distributions,
2144 phenology and physiology (Butchart et al., 2010; Chen et al., 2011; Chown et al., 2010; Dirzo et al., 2014;
2145 Inouye, 2022; Lenoir and Svenning, 2015; Newbold et al., 2015; Pörtner and Farrell, 2008; Soroye et al.,
2146 2020), and increased extinctions rates to unprecedented levels (Barnosky et al., 2011; Ceballos et al., 2015;
2147 De Vos et al., 2015) – in some cases putting hundreds of millions of years of evolution at risk (IUCN, 2020;
2148 Nowakowski et al., 2018). As anthropogenic pressures on biodiversity are unlikely to reduce given current
2149 scenarios of human development (Stehfest et al., 2019), and as international targets aiming to protect biodi-
2150 versity and related ecosystem services have failed to be met (Buchanan et al., 2020), it is vital that we keep
2151 pursuing conservation and mitigating efforts to minimise or even reverse human impacts on biodiversity.

2152 Human pressures impact species unevenly; for instance, past work has highlighted phylogenetic and
2153 spatial biases in species vulnerability to human pressures (Ducatez and Shine, 2017; Fritz et al., 2009;
2154 Weeks et al., 2022; Yessoufou et al., 2012). Some species (termed 'winners' in past work) may benefit
2155 from global changes, while other species (termed 'losers') are likely to decline. Understanding the factors
2156 that underpin interspecific variation in responses to human pressures is valuable to conservation planning,
2157 as it can help target and prioritise species at most risk from different threats. One of the reasons why

species differ in their responses to environmental change is that species possess different characteristics, or traits. In my thesis, following McGill et al. (2006), I defined traits as characteristics measurable at the level of an individual (i.e., intrinsic), comparable across different species, with likely impacts on organismal fitness or performance. Asking whether species traits relate to species' responses to land-use and climate change can thus help understand interspecific differences in responses to human threats (Munstermann et al., 2022), and may help assess which species are more likely to be 'winners' or 'losers' under particular threatening processes. However, past studies that have tackled this question in terrestrial vertebrates have often been limited in both taxonomic and spatial coverage, so that it remains unclear whether there are general patterns in trait-sensitivity associations with human pressures across vertebrate species. Further, comparative studies of the sensitivity to both land-use and climate change among terrestrial vertebrate classes have been lacking. Understanding whether different human pressures are likely to favour similar species across diverse taxonomic groups could help conservation efforts. In my thesis, I aimed to fill in these gaps, investigating whether and which traits are associated with land-use responses and climate-change sensitivity comparatively across terrestrial vertebrates and at global scales. My thesis also aimed to highlight some of the possible consequences for ecosystem functioning.

To this end, I started by compiling trait data across terrestrial vertebrates, and by assessing the gaps and biases in the availability of trait data (Chapter 2). Bringing together these compiled data and a database containing species records in different land-use types (PREDICTS), I assessed the effects of land use and land-use intensity on the functional diversity of local terrestrial vertebrate assemblages (Chapter 3). I then asked whether species ecological characteristics (in which I included species ecological traits and species geographical range area) were associated with species' land-use responses and with species' estimated climate-change sensitivity, comparatively across the four vertebrate classes (Chapter 4). Finally, I investigated the effects of land use and land-use intensity on the total energetic requirements of vertebrate assemblages, and I further assessed whether species' energetic requirements influenced species' land-use responses (Chapter 5). In this final Chapter, I synthesise the key findings of my work, and I assess their contributions to the broader knowledge and their relevance for the field. I highlight some of the limitations of my work and further challenges, notably reflecting on the current challenges to the application of trait-based approaches at large scales in animal taxa.

2186 6.1 Gaps and biases in the knowledge of terrestrial vertebrates

2187 Although terrestrial vertebrates have been extensively studied in the past, there remain important gaps in our
2188 knowledge. Some of these gaps are illustrated in Chapter 2, which demonstrates the biases in the availability
2189 of ecological trait data (i.e., the ‘Raunkiær’ shortfall (Hortal et al., 2015)). After collating data for seven
2190 commonly-used ecological traits, I showed that the sampling of these traits presented taxonomic, phyloge-
2191 netic and spatial biases. Mammals and birds were overall well sampled for most traits (with a median trait
2192 coverage of 85% for birds, and of 95% for mammals). However, amphibians and reptiles presented acute
2193 gaps, with a 32% median coverage for amphibians, and 38% for reptiles. Chapter 2 further showed that such
2194 gaps were non-randomly distributed with regards to species phylogenetic position, with certain clades being
2195 under-sampled compared to others (for example, the family *Ranidae*, or true frogs; or the blind snakes of
2196 the family *Typhlopidae*, and the worm snakes of the family *Amphisbaenidae*). Hence, Chapter 2 showed that
2197 amphibians and reptiles are understudied compared to mammals and birds. Knowledge gaps are thus acute
2198 for the most diverse vertebrate class (reptiles) as well as for the most threatened vertebrate class (amphibians;
2199 IUCN (2020)).

2200 Chapter 2 also highlights that knowledge gaps for amphibians and reptiles were non-randomly distributed
2201 across geographical space. For instance, the availability of trait data was significantly positively associated
2202 with species richness in several biogeographic realms (e.g., in the Australasian realm for reptiles, and in
2203 the Neotropics for amphibians), and significantly negatively associated with species richness in other realms
2204 (e.g., in the Indo-Malayan realm for both amphibians and reptiles). Chapter 2 thus highlighted some critically
2205 under-sampled regions for traits in amphibians and reptiles (e.g., the Congo-Basin). As discussed in Chapter
2206 2, such geographical biases may be driven by uneven primary data collection efforts, themselves possibly
2207 explained by interacting socioeconomic factors (Collen et al., 2008; Hortal et al., 2015; Martin et al., 2012;
2208 United Nations Educational Scientific and Cultural Organization, 2015). Importantly, Chapter 2 shows
2209 that trait information may be less available in some of the most species-rich regions, critically important
2210 for global biodiversity conservation (Barlow et al., 2018). Chapter 2 further showed that, in all classes, the
2211 availability of trait information depended on species’ rarity (captured by geographical range area), with more-
2212 widely distributed species being on average better sampled for traits than less widely distributed species.
2213 Such trends could be explained by a sampling bias towards more easily detectable species, and is concerning
2214 as narrow-ranging species have been shown to be at higher risks of extinction (Chichorro et al., 2019).

2215 It is worth highlighting that I targeted traits that are commonly used by ecologists in Chapter 2; knowl-
2216 edge gaps are likely to be even more important when looking at other traits, that are maybe less frequently

used (such as dispersal abilities, which are available for a small fraction of vertebrate species only, despite their likely importance for understanding species' responses to anthropogenic pressures; Schloss et al. (2012); Lenoir and Svenning (2015)). Chapter 4 and Chapter 5 provide with an illustration of this point: in Chapter 4, no information about reptile diet could be obtained from an existing published database; while in Chapter 5, the availability of physiological data on resting metabolic rates was limited to a small number of species, even in mammals and birds. Further, lack of available data prevented me from considering intraspecific variation in my thesis, despite its likely importance for understanding species- and assemblage-level responses to human-driven change (Carlson et al., 2014; Guralnick et al., 2016; Rohr et al., 2018).

The 'Raunkiær' shortfall, coupled with the lack of a centralised repository for animal trait data, may have been a major hindrance to the application of trait-based approaches at large scales in vertebrate species. In my thesis, I employed imputation techniques throughout to estimate missing trait values. However, complementing existing datasets with empirical data is the only way to fill the current data gaps robustly. It may be that integrating regional databases, potentially in different languages, could help fill some of the gaps.

Finally, many studies have called for the integration and standardisation of trait databases (Junker et al., 2022; Kissling et al., 2018; Schneider et al., 2019; Weiss and Ray, 2019), to produce open-access, interoperable, and unified products. Developing an appropriate framework for data standardisation and integration would require overcoming implementation and conceptual difficulties, notably pertaining to the definition of comparable traits across organisms and to the integration of taxonomic nomenclatures (Salguero-Gómez et al., 2021). However, integration and standardisation of trait databases would represent a major step for animal ecology, preventing the duplication of research efforts, promoting collaboration and opening new research avenues. By highlighting the current gaps in terrestrial vertebrate trait data, Chapter 2 could help guide future collection efforts.

6.2 Functional reshaping of local vertebrate assemblages under land-use change

In Chapter 3 and Chapter 5, my thesis highlights two different dimensions of the functional reshaping of vertebrate assemblages under land-use change. While Chapter 3 focuses on the functional diversity of local vertebrate assemblages by using various indices to summarise the diversity of ecological traits across vertebrate assemblages of the PREDICTS database, Chapter 5 makes use of physiological data to quantify energetic requirements at the assemblage level. To my knowledge, my work constitutes the first global assessment of the responses of functional diversity to land use and land-use intensity across vertebrate classes,

2247 and of the changes in total vertebrate energetic requirements with land use and land-use intensity.

2248 **6.2.1 Functional diversity and functional composition (Chapter 3)**

2249 In Chapter 3, after combining the trait data compiled in Chapter 2 with the PREDICTS database, I investi-
2250 gated how land use and land-use intensity affect the functional diversity (i.e., functional richness and func-
2251 tional dispersion) as well as the functional composition (i.e., functional loss and functional gain) of terrestrial
2252 vertebrate assemblages, across and within vertebrate classes, at global scales. I investigated potential spatial
2253 variation in responses by looking for differences between temperate and tropical areas. The findings of this
2254 Chapter are threefold. First, across all vertebrate species, the functional diversity of vertebrate assemblages
2255 is negatively impacted by human land uses. Both functional richness and functional dispersion showed im-
2256 portant average decreases in some disturbed land uses (e.g., a 63% average decline in functional richness in
2257 intensely-used tropical cropland; a 20% average decline in intensely-used urban areas). Further, decreases in
2258 functional dispersion exceeded the decreases expected from species loss in a number of disturbed land uses
2259 (most notably in tropical cropland), providing evidence of functional clustering of vertebrate assemblages
2260 in these land uses. Second, Chapter 3 highlights the spatial and taxonomic variation in responses: overall,
2261 functional richness tended to be more negatively affected in the tropics than in temperate areas (but responses
2262 were similar between the two areas for functional dispersion). The functional diversity of amphibians and
2263 birds was overall more negatively impacted than that of mammals and reptiles. Finally, Chapter 3 showed
2264 that vertebrate assemblages in disturbed land uses were subject to high levels of functional loss, and in some
2265 cases functional gain, highlighting important functional turnover in vertebrate assemblages.

2266 Chapter 3 thus highlights that the functional composition of vertebrate assemblages is reshaped in dis-
2267 turbed land uses. My results are mostly in line with previous studies that have been more taxonomically or
2268 geographically restricted (Flynn et al., 2009; Marcacci et al., 2021; Matuoka et al., 2020). Although the func-
2269 tional diversity of tropical areas emerges as being more sensitive than that of temperate areas, I also found
2270 important decreases in functional diversity in temperate areas, a pattern that had not been detected in past
2271 work (Matuoka et al., 2020). My results indicate that land-use change is a potential threat to ecosystem pro-
2272 cesses, in particular those sustained by species falling in sensitive areas of the trait space. Further, losses of
2273 functional diversity in tropical areas are particularly problematic given that those areas are more species-rich
2274 than temperate areas, harbour many biodiversity hotspots and protected areas, and are currently subjected to
2275 some of the highest rates of land-use change (Hansen et al., 2013; Laurance et al., 2012; Spracklen et al.,
2276 2015).

2277 **6.2.2 Energetic requirements (Chapter 5)**

2278 In Chapter 5, I combined the PREDICTS database with species-level estimates of resting metabolic rates.
2279 I investigated whether the total energetic requirements of vertebrate assemblages differed among different
2280 land-use types and trophic groups (classified as omnivores, herbivores, and carnivores). Contrary to my
2281 expectations, I found that the minimum amount of energy required by vertebrate assemblages did not show
2282 systematic decreases in disturbed land-use types. Across all three trophic groups, total energetic require-
2283 ments (estimated from total abundance-weighted resting metabolic rates) even showed strong increases in
2284 some disturbed land uses (e.g., an average increase of 200% in lightly-used urban areas for carnivores).
2285 These findings highlight that disturbed land uses have significant impacts on ecosystem functioning and em-
2286 phasize that land-use change may promote significant changes to the local energetic balance of vertebrate
2287 assemblages. To my knowledge, this work constitutes the first direct global quantification of the impacts of
2288 land-use and land-use intensity on vertebrate energetic requirements. My findings are tightly linked to stud-
2289 ies that have investigated changes in the body-mass composition of vertebrate assemblages under land-use
2290 change (e.g., Newbold et al. (2020b)), and also to studies drawing from food-web theory (e.g, impacts of
2291 habitat loss and fragmentation on the body size distribution in a trophic chain; Hillaert et al. (2020)).

2292 Altogether, the findings of Chapter 3 and 5 show that land-use change reshapes the functional compo-
2293 sition of vertebrate assemblages. Land-use change may reduce native ecological trait diversity, constricting
2294 used areas of the trait space, thus potentially disrupting ecosystem processes sustained by the native species
2295 that are located in sensitive areas of the trait space. Further work could investigate the contributions of non-
2296 native species to the functional reshaping of local vertebrate assemblages. Changes in species composition
2297 in disturbed land uses have consequences for ecosystem functioning, notably significantly impacting the
2298 amount of energy locally processed by vertebrate species.

2299 **6.3 Uneven sensitivity of vertebrate species to land-use and climate change**

2300 In Chapter 4 and Chapter 5, I investigated associations between species-level characteristics and species'
2301 sensitivity to land-use and climate change. Chapter 4 focuses on ecological characteristics (ecological traits
2302 and geographical range area), and to my knowledge constitutes the first work to investigate associations
2303 between species' ecological characteristics and two human pressures, at global scales and comparatively
2304 across vertebrate classes. Chapter 5 uses physiological data and focuses on species' energetic requirements.
2305 To my knowledge, there has yet not been a study investigating whether species' energetic requirements

2306 influence species' responses to land use and land-use intensity in vertebrate species.

2307 **6.3.1 Ecological characteristics (Chapter 4)**

2308 In Chapter 4, I complemented the trait data from Chapter 2 with species-level dietary information and geo-
2309 graphical range area. On the one hand, I combined these species-level data with the PREDICTS database,
2310 and I investigated how species' ecological characteristics influenced their responses to land use and land-use
2311 intensity. On the other hand, after estimating climate-change sensitivity from properties of species' climatic
2312 niche space, I investigated the associations between species' ecological characteristics and climate-change
2313 sensitivity. First, I found that narrower ranges, smaller habitat breadth, and inability to exploit artificial habi-
2314 tats were consistently associated with more negative land-use responses and higher climate-change sensitiv-
2315 ity across vertebrate classes. Second, the associations of other traits were both class- and pressure-dependent.
2316 Overall, invertebrate eaters and fruit/nectar eaters tended to be negatively affected in disturbed land uses in
2317 all classes; in addition, invertebrate- and plant/seed-eating birds had higher climate-change sensitivity.

2318 Chapter 4 thus highlights that both land-use and climate change are likely to favour similar species,
2319 that is, wider-ranging species, those with larger habitat breadth and those able to use artificial habitats. My
2320 work aligns with previous analyses on extinction risks (Chichorro et al., 2019) and climate-change responses
2321 (MacLean and Beissinger, 2017), and shows that similar species might be favoured by both these human
2322 pressures. Further, the narrow-ranging species, which are on average less well known for trait data (as
2323 shown in Chapter 2) and potentially for many other aspects of their biology, are those that are more likely
2324 to be more sensitive to human pressures. Such species may however make distinct and unique contributions
2325 to ecosystem functioning (Dee et al., 2019; Mouillot et al., 2013). Further investigations of the ecosystem
2326 processes sustained by these geographically rarer species and their contributions to ecosystem functioning
2327 may be helpful to the mitigation of human impacts.

2328 I would like to underline some of the limitations of this work: first, as discussed in Chapter 4, I consid-
2329 ered land-use change and climate change separately, thus not accounting for potentially interactive effects
2330 between these pressures. However, human pressures are likely to interact, and combined effects of land-
2331 use and climate change could have more deleterious (synergistic) effects than individual pressures acting
2332 independently (Williams and Newbold, 2020; Williams et al., 2022). Second, my findings rely on the use
2333 of the PREDICTS database (in this Chapter and in Chapters 3 and 5) and on the use of geographical dis-
2334 tributions, and as such I used a 'space-for-time' approach to infer land-use responses and climate-change
2335 sensitivity from spatial data (Blois et al., 2013). Thus, I assumed that vertebrate assemblages and current
2336 species distributions were at equilibrium (De Palma et al., 2018), and I did not consider potential recovery

2337 effects, long-term population declines or any other temporal dynamics. However, it is likely that populations
2338 are not at equilibrium (Damgaard, 2019; De Palma et al., 2018). For instance, Sales et al. (2022) showed
2339 that range contractions of megafaunal mammals over the Late Pleistocene could lead to an underestimation
2340 of the realised climate niches of these species when using current distribution data, so that the estimated
2341 climate-change sensitivity of these species may be overestimated (Sales et al., 2022). Further, biotic lags and
2342 delays in biodiversity responses emphasise the need to account for land-use history (De Palma et al., 2018;
2343 Le Provost et al., 2020). Using long-term species records and population data may provide insights into the
2344 long-term effects of land-use and climate change on vertebrate species.

2345 Finally, in Chapter 4, I conducted a correlative assessment of the associations between species ecological
2346 characteristics and species' responses to land use and species' climate-change sensitivity. I found a number
2347 of significant associations, but traits had overall low explanatory power in the models, thus putting into
2348 question the degree to which traits can be used to infer species' responses to environmental change. Further,
2349 my results do not allow for mechanistic interpretations of how traits influence species' responses to land-use
2350 and climate change. Using long-term population data, mechanistic models and *in silico* and field experiments
2351 may help uncover some of these mechanisms (Boult and Evans, 2021; Ries et al., 2004).

2352 **6.3.2 Mass-independent energetic requirements (Chapter 5)**

2353 Chapter 5 highlights the effects of the interactions between species' energetic requirements and trophic
2354 group on responses to land use and land-use intensity (after controlling metabolic rates for the effects of
2355 body mass and taxonomy, which explain a large proportion of the interspecific variation in metabolic rates).
2356 The key finding from this analysis contradicted my initial prediction: in all trophic groups, species with
2357 larger energetic expenditure (relative to body mass and taxonomy) tended to do better in some of the most
2358 disturbed land-use types, compared to species with lower energetic expenditure. Thus, my work highlights
2359 that land-use change may favour species that have higher rates of mass-independent energy consumption,
2360 which could be linked with interspecific differences in behaviour, as discussed in Chapter 5 in more details.
2361 Species with larger energetic expenditure may display a set of characteristics that render them better able to
2362 cope with anthropogenic disturbances, such as higher activity levels (Biro and Stamps, 2010; Coogan et al.,
2363 2018). Such species may also have a larger brain (a metabolically more consuming organ per unit mass; Isler
2364 and Van Schaik (2006)), which could play a significant role on species' ability to persist in disturbed areas
2365 (Sayol et al., 2020).

2366 Finally, I would like to emphasise that my work gives important indications of how human pressures
2367 may affect ecosystem functioning. For example, the higher sensitivity of invertebrate eaters to both land-use

and climate change may indicate that the processes that such species underpin, such as pest control, might be put at risk (Civantos et al., 2012). However, we lack large-scale quantifications and empirical measurements of ecosystem processes sustained by vertebrates (Luck et al., 2012; Wenny et al., 2011). Linking vertebrate traits to particular ecosystem functions at large scales remains challenging, maybe because of a lack of empirical data, but also maybe because ecosystem processes sustained by vertebrate species are difficult to quantify (potentially because of the mobility of these species). Vertebrates nevertheless contribute significantly to ecosystem functioning (Breviglieri and Romero, 2017; Ratto et al., 2018; Wandrag et al., 2015; Zhang et al., 2018a), emphasising the need to mitigate human impacts on them.

6.4 Relevance of the findings to conservation

Overall, my findings show that land-use and climate change may favour similar vertebrate species: ‘winners’ are likely to be wider-ranging species that are able to occupy diverse habitats, including human-disturbed habitats; ‘winners’ might also be able to allocate more energy to sustain higher activity levels. On the other hand, ‘losers’ are likely narrow-ranging species, with natural habitat specialism and unlikely to be able to persist in human-disturbed areas. Thus, considerations of geographical rarity and specialisation indices appear to be highly relevant for characterising species at risk from both land-use and climate change when working at large scales and across all terrestrial vertebrates. In fact, such criteria have been employed by the IUCN Red List for assessing species’ extinction risk (Rodrigues et al., 2006), as well as in predictive vulnerability assessments (e.g., Foden et al. (2013)). Further, considerations of species’ geographical rarity and specialisation can be useful to species-based as well as area-based conservation prioritization, for instance for identifying key areas for biodiversity conservation and for targeting species of interest (Asaad et al., 2017; Mace et al., 2006). My results thus lend support to the idea that vulnerability assessments and hierarchisation frameworks should take geographical rarity and specialisation into account (Le Berre et al., 2019).

6.5 Conclusion

My thesis constitutes, to my knowledge, the first attempt to use trait-based approaches at global scales to investigate associations between sensitivity to human pressures and species traits, comparatively across the terrestrial vertebrate classes and human pressures (land-use and climate change). I demonstrated that there exist major gaps and biases in our global ecological knowledge of terrestrial vertebrate species, particularly affecting amphibians and reptiles and some of the most species-rich regions, which is problematic given the

2397 higher sensitivity of these areas to human pressures. My work indicates that land-use and climate change
2398 are reshaping vertebrate biodiversity. I highlighted two dimensions of such functional reshaping, using
2399 ecological characteristics on the one hand, and physiological data on the other hand. Land-use and climate
2400 change may tend to favour species that are wider-ranging, have larger habitat breadth and are able to use
2401 artificial habitats; and land-use change alone tends to favour species that are able to allocate more energy to
2402 organismal maintenance than expected from body mass and taxonomy. Thus, my work lends further support
2403 to the idea that human activities promote the homogenisation of the biota, with a set of ‘winning’ species
2404 likely to benefit from global changes at the expense of sensitive ‘losers’, putting at risk ecosystem processes
2405 sustained by those sensitive species. Overall, my work highlights the necessity of strengthening conservation
2406 and mitigation efforts in the face of global human-driven changes.

2407

Bibliography

- 2408 Albert, C., Luque, G. M., and Courchamp, F. (2018). The twenty most charismatic species. *PLoS ONE*,
2409 13(7), 1–12. DOI: 10.1371/journal.pone.0199149.
- 2410 Alexander, P., Rounsevell, M. D., Dislich, C., Dodson, J. R., Engström, K., and Moran, D. (2015). Drivers
2411 for global agricultural land use change: The nexus of diet, population, yield and bioenergy. *Global Envi-*
2412 *ronmental Change*, 35, 138–147. DOI: 10.1016/j.gloenvcha.2015.08.011.
- 2413 Alves, R. R. N., Souto, W. M. S., Fernandes-Ferreira, H., Bezerra, D. M. M., Barboza, R. R. D., and Vieira,
2414 W. L. S. (2018). Chapter 7 - The Importance of Hunting in Human Societies. *Ethnozoology - Animal*
2415 *in our lives*. Ed. by R. R. N. Alves and U. P. Albuquerque. Academic Press, 95–118. DOI: 10.1016/
2416 B978-0-12-809913-1.00007-7.
- 2417 Angert, A. L., Crozier, L. G., Rissler, L. J., Gilman, S. E., Tewksbury, J. J., and Chunco, A. J. (2011). Do
2418 species' traits predict recent shifts at expanding range edges? *Ecology Letters*, 14(7), 677–689. DOI:
2419 10.1111/j.1461-0248.2011.01620.x.
- 2420 Aronson, M. F., Lepczyk, C. A., Evans, K. L., Goddard, M. A., Lerman, S. B., MacIvor, J. S., Nilon, C. H.,
2421 and Vargo, T. (2017). Biodiversity in the city: key challenges for urban green space management. *Fron-*
2422 *tiers in Ecology and the Environment*, 15(4), 189–196. DOI: 10.1002/fee.1480.
- 2423 Asaad, I., Lundquist, C. J., Erdmann, M. V., and Costello, M. J. (2017). Ecological criteria to identify areas
2424 for biodiversity conservation. *Biological Conservation*, 213(Part B). SI:Measures of biodiversity, 309–
2425 316. DOI: <https://doi.org/10.1016/j.biocon.2016.10.007>.
- 2426 Auer, S. K., Killen, S. S., and Rezende, E. L. (2017). Resting vs. active: a meta-analysis of the intra- and
2427 inter-specific associations between minimum, sustained, and maximum metabolic rates in vertebrates.
2428 *Functional Ecology*, 31(9), 1728–1738. DOI: 10.1111/1365-2435.12879.
- 2429 Auer, S. K., Solowey, J. R., Rajesh, S., and Rezende, E. L. (2020). Energetic mechanisms for coping with
2430 changes in resource availability. *Biology Letters*, 16(11), 1–8. DOI: 10.1098/rsbl.2020.0580.
- 2431 Balmford, A. (1996). Extinction filters and current resilience: The significance of past selection pressures
2432 for conservation biology. *Trends in Ecology and Evolution*, 11(5), 193–196. DOI: 10.1016/0169–
2433 5347(96)10026-4.
- 2434 Banks-Leite, C., Ewers, R. M., Folkard-Tapp, H., and Fraser, A. (2020). Countering the effects of habitat
2435 loss, fragmentation, and degradation through habitat restoration. *One Earth*, 3(6), 672–676. DOI: 10.
2436 1016/j.oneear.2020.11.016.
- 2437 Baraldi, A. N. and Enders, C. K. (2010). An introduction to modern missing data analyses. *Journal of School*
2438 *Psychology*, 48(1), 5–37. DOI: 10.1016/j.jsp.2009.10.001.

- 2439 Barlow, J., França, F., Gardner, T. A., Hicks, C. C., Lennox, G. D., Berenguer, E., Castello, L., Economo,
2440 E. P., Ferreira, J., Guénard, B., Gontijo Leal, C., Isaac, V., Lees, A. C., Parr, C. L., Wilson, S. K., Young,
2441 P. J., and Graham, N. A. (2018). The future of hyperdiverse tropical ecosystems. *Nature*, 559, 517–526.
2442 DOI: 10.1038/s41586-018-0301-1.
- 2443 Barnes, A. D., Jochum, M., Mumme, S., Haneda, N. F., Farajallah, A., Widarto, T. H., and Brose, U. (2014).
2444 Consequences of tropical land use for multitrophic biodiversity and ecosystem functioning. *Nature Communications*, 5, 1–7. DOI: 10.1038/ncomms6351.
- 2445 Barnosky, A. D., Matzke, N., Tomaia, S., Wogan, G. O., Swartz, B., Quental, T. B., Marshall, C., McGuire,
2446 J. L., Lindsey, E. L., Maguire, K. C., Mersey, B., and Ferrer, E. A. (2011). Has the Earth's sixth mass
2447 extinction already arrived? *Nature*, 471, 51–57. DOI: 10.1038/nature09678.
- 2448 Bateman, P. W. and Fleming, P. A. (2012). Big city life: Carnivores in urban environments. *Journal of Zoology*, 287(1), 1–23. DOI: 10.1111/j.1469-7998.2011.00887.x.
- 2449 Bates, D., Mächler, M., Bolker, B. M., and Walker, S. C. (2015). Fitting linear mixed-effects models using
2450 lme4. *Journal of Statistical Software*, 67, 1–48. DOI: 10.18637/jss.v067.i01.
- 2451 Betts, M. G. et al. (2019). Extinction filters mediate the global effects of habitat fragmentation on animals.
2452 *Science*, 366(6470), 1236–1239. DOI: 10.1126/science.aax9387.
- 2453 Biermann, F., Bai, X., Bondre, N., Broadgate, W., Arthur Chen, C. T., Dube, O. P., Erisman, J. W., Glaser, M.,
2454 Hel, S. van der, Lemos, M. C., Seitzinger, S., and Seto, K. C. (2016). Down to Earth: Contextualizing the
2455 Anthropocene. *Global Environmental Change*, 39, 341–350. DOI: 10.1016/j.gloenvcha.2015.
2456 11.004.
- 2457 Biro, P. A. and Stamps, J. A. (2010). Do consistent individual differences in metabolic rate promote con-
2458 sistent individual differences in behavior? *Trends in Ecology and Evolution*, 25(11), 653–659. DOI:
2459 10.1016/j.tree.2010.08.003.
- 2460 Bivand, R. and Piras, G. (2015). Comparing Implementations of Estimation Methods for Spatial Economet-
2461 rics. *Journal of Statistical Software*, 63(18), 1–36. DOI: 10.18637/jss.v063.i18.
- 2462 Bivand, R. and Wong, D. W. S. (2018). Comparing implementations of global and local indicators of spatial
2463 association. *TEST*, 27(3), 716–748. DOI: 10.1007/s11749-018-0599-x.
- 2464 Bivand, R., Hauke, J., and Kossowski, T. (2013a). Computing the Jacobian in Gaussian spatial autoregressive
2465 models: An illustrated comparison of available methods. *Geographical Analysis*, 45(2), 150–179. DOI:
2466 10.1111/gean.12008.
- 2467 Bivand, R. S., Pebesma, E., and Gomez-Rubio, V. (2013b). Applied spatial data analysis with R, Second
2468 edition. DOI: 10.1007/978-1-4614-7618-4.
- 2469 Blois, J. L., Williams, J. W., Fitzpatrick, M. C., Jackson, S. T., and Ferrier, S. (2013). Space can substitute
2470 for time in predicting climate-change effects on biodiversity. *Proceedings of the National Academy of
2471 Sciences of the United States of America*, 110(23), 9374–9379. DOI: 10.1073/pnas.1220228110.
- 2472 Böhm, M., Cook, D., Ma, H., Davidson, A. D., García, A., Tapley, B., Pearce-Kelly, P., and Carr, J. (2016).
2473 Hot and bothered: Using trait-based approaches to assess climate change vulnerability in reptiles. *Bio-
2474 logical Conservation*, 204, 32–41. DOI: 10.1016/j.biocon.2016.06.002.
- 2475 Bolnick, D. I., Amarasekare, P., Araújo, M. S., Bürger, R., Levine, J. M., Novak, M., Rudolf, V. H., Schreiber,
2476 S. J., Urban, M. C., and Vasseur, D. A. (2011). Why intraspecific trait variation matters in community
2477 ecology. *Trends in Ecology and Evolution*, 26(4), 183–192. DOI: 10.1016/j.tree.2011.01.009.

- 2480 Bouckaert, R. et al. (2019). BEAST 2.5: An advanced software platform for Bayesian evolutionary analysis.
2481 *PLoS Computational Biology*, 15(4). DOI: 10.1371/journal.pcbi.1006650.
- 2482 Boult, V. L. and Evans, L. C. (2021). Mechanisms matter: Predicting the ecological impacts of global change.
2483 *Global Change Biology*, 27(9), 1689–1691. DOI: 10.1111/gcb.15527.
- 2484 Bregman, T. P., Lees, A. C., Seddon, N., Macgregor, H. E., Darski, B., Aleixo, A., Bonsall, M. B., and
2485 Tobias, J. A. (2015). Species interactions regulate the collapse of biodiversity and ecosystem function in
2486 tropical forest fragments. *Ecology*, 96(10), 2692–2704. DOI: 10.1890/14-1731.1.
- 2487 Bregman, T. P., Lees, A. C., MacGregor, H. E., Darski, B., Moura, N. G. de, Aleixo, A., Barlow, J., and
2488 Tobias, J. A. (2016). Using avian functional traits to assess the impact of land-cover change on ecosystem
2489 processes linked to resilience in tropical forests. *Proceedings of the Royal Society B: Biological Sciences*,
2490 283(1844), 20161289. DOI: 10.1098/rspb.2016.1289.
- 2491 Breviglieri, C. P. B. and Romero, G. Q. (2017). Terrestrial vertebrate predators drive the structure and func-
2492 tioning of aquatic food webs. *Ecology*, 98(8), 2069–2080. DOI: 10.1002/ecy.1881.
- 2493 Brischoux, F., Meillère, A., Dupoué, A., Lourdais, O., and Angelier, F. (2017). Traffic noise decreases
2494 nestlings' metabolic rates in an urban exploiter. *Journal of Avian Biology*, 48(7), 905–909. DOI: 10 .
2495 1111/jav.01139.
- 2496 Broughton, J. M. and Weitzel, E. M. (2018). Population reconstructions for humans and megafauna suggest
2497 mixed causes for North American Pleistocene extinctions. *Nature Communications*, 9(1), 1–12. DOI:
2498 10.1038/s41467-018-07897-1.
- 2499 Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M., and West, G. B. (2004). Toward a metabolic theory
2500 of ecology. *Ecology*, 85(7), 1771–1789. DOI: 10.2106/JBJS.J.01311.
- 2501 Bruggeman, J., Heringa, J., and Brandt, B. W. (2009). PhyloPars: Estimation of missing parameter values
2502 using phylogeny. *Nucleic Acids Research*, 37(suppl2), W179–W184. DOI: 10.1093/nar/gkp370.
- 2503 Brzęk, P. and Konarzewski, M. (2001). Effect of food shortage on the physiology and competitive abilities
2504 of sand martin (*Riparia riparia*) nestlings. *Journal of Experimental Biology*, 204(17), 3065–3074. DOI:
2505 10.1242/jeb.204.17.3065.
- 2506 Buchanan, G. M., Butchart, S. H., Chandler, G., and Gregory, R. D. (2020). Assessment of national-level
2507 progress towards elements of the Aichi Biodiversity Targets. *Ecological Indicators*, 116, 106497. DOI:
2508 10.1016/j.ecolind.2020.106497.
- 2509 Burger, J. R., Hou, C., Hall, C. A. S., and Brown, J. H. (2021). Universal rules of life: metabolic rates,
2510 biological times and the equal fitness paradigm. *Ecology Letters*, 24(6), 1262–1281. DOI: 10.1111/
2511 ele.13715.
- 2512 Burton, T., Killen, S. S., Armstrong, J. D., and Metcalfe, N. B. (2011). What causes intraspecific variation
2513 in resting metabolic rate and what are its ecological consequences? *Proceedings of the Royal Society B:*
2514 *Biological Sciences*, 278(1724), 3465–3473. DOI: 10.1098/rspb.2011.1778.
- 2515 Bushuev, A., Tolstenkov, O., Zubkova, E., Solovyeva, E., and Kerimov, A. (2018). Basal metabolic rate
2516 in free-living tropical birds: The influence of phylogenetic, behavioral, and ecological factors. *Current*
2517 *Zoology*, 64(1), 33–43. DOI: 10.1093/cz/zox018.
- 2518 Butchart, S. H. M. et al. (2010). Global Biodiversity: Indicators of Recent Declines. *Science*, 328(5982),
2519 1164–1168. DOI: 10.1126/science.1187512.

- 2520 Butchart, S. H., Di Marco, M., and Watson, J. E. (2016). Formulating Smart Commitments on Biodiversity:
2521 Lessons from the Aichi Targets. *Conservation Letters*, 9(6), 457–468. DOI: 10.1111/conl.12278.
- 2522 Cadotte, M. W. and Tucker, C. M. (2017). Should Environmental Filtering be Abandoned? *Trends in Ecology
2523 and Evolution*, 32(6), 429–437. DOI: 10.1016/j.tree.2017.03.004.
- 2524 Cadotte, M. W., Carscadden, K., and Mirochnick, N. (2011). Beyond species: Functional diversity and the
2525 maintenance of ecological processes and services. *Journal of Applied Ecology*, 48(5), 1079–1087. DOI:
2526 10.1111/j.1365-2664.2011.02048.x.
- 2527 Cahill, A. E., Aiello-Lammens, M. E., Caitlin Fisher-Reid, M., Hua, X., Karanewsky, C. J., Ryu, H. Y.,
2528 Sbeglia, G. C., Spagnolo, F., Waldron, J. B., Warsi, O., and Wiens, J. J. (2013). How does climate change
2529 cause extinction? *Proceedings of the Royal Society B: Biological Sciences*, 280(1750), 20121890. DOI:
2530 10.1098/rspb.2012.1890.
- 2531 Cannistra, A. F. and Buckley, L. B. (2021). Improving range shift predictions: enhancing the power of traits.
2532 *bioRxiv*, 2021.02.15.431292. DOI: 10.1101/2021.02.15.431292.
- 2533 Capdevila, P., Noviello, N., McRae, L., Freeman, R., and Clements, C. (2022a). Body mass and latitude
2534 as global predictors of vertebrate populations exposure to multiple threats. *bioRxiv*, 2022.02.04.479091.
2535 DOI: 10.1101/2022.02.04.479091.
- 2536 Capdevila, P., Stott, I., Cant, J., Beger, M., Rowlands, G., Grace, M., and Salguero-Gómez, R. (2022b). Life
2537 history mediates the trade-offs among different components of demographic resilience. *Ecology Letters*,
2538 1–14. DOI: 10.1111/ele.14004.
- 2539 Capuracho, J. M., Ashley, M. V., Tsuru, B. R., Cooper, J. C., and Bates, J. M. (2020). Dispersal ability
2540 correlates with range size in Amazonian habitat-restricted birds. *Proceedings of the Royal Society B:
2541 Biological Sciences*, 287(1939). DOI: 10.1098/rspb.2020.1450rspb20201450.
- 2542 Cardoso, D. et al. (2017). Amazon plant diversity revealed by a taxonomically verified species list. *Proceedings
2543 of the National Academy of Sciences of the United States of America*, 114(40), 10695–10700. DOI:
2544 10.1073/pnas.1706756114.
- 2545 Carlson, S. M., Cunningham, C. J., and Westley, P. A. (2014). Evolutionary rescue in a changing world.
2546 *Trends in Ecology and Evolution*, 29(9), 521–530. DOI: 10.1016/j.tree.2014.06.005.
- 2547 Caro, T., Rowe, Z., Berger, J., Wholey, P., and Dobson, A. (2022). An inconvenient misconception: Climate
2548 change is not the principal driver of biodiversity loss. *Conservation Letters*, 1–6. DOI: 10.1111/
2549 conl.12868.
- 2550 Ceballos, G., Ehrlich, P. R., Barnosky, A. D., García, A., Pringle, R. M., and Palmer, T. M. (2015). Ac-
2551 celerated modern human–induced species losses: Entering the sixth mass extinction. *Science
2552 Advances*, 1(5), e1400253. DOI: 10.1126/sciadv.1400253.
- 2553 Chamberlain, S. (2018). *rredlist: 'IUCN' Red List Client*. R package version 0.5.0.
- 2554 Chamberlain, S. A. and Szöcs, E. (2013). taxize : taxonomic search and retrieval in R [version 2; referees: 3
2555 approved]. *F1000Research*, 2(191). DOI: 10.12688/f1000research.2-191.v2.
- 2556 Chapman, P. M., Tobias, J. A., Edwards, D. P., and Davies, R. G. (2018). Contrasting impacts of land-use
2557 change on phylogenetic and functional diversity of tropical forest birds. *Journal of Applied Ecology*,
2558 55(4), 1604–1614. DOI: 10.1111/1365-2664.13073.

- 2559 Chaudhary, A., Pourfaraj, V., and Mooers, A. O. (2018). Projecting global land use-driven evolutionary
2560 history loss. *Diversity and Distributions*, 24(2), 158–167. DOI: 10.1111/ddi.12677.
- 2561 Chaudhary, C., Saeedi, H., and Costello, M. J. (2016). Bimodality of Latitudinal Gradients in Marine Species
2562 Richness. *Trends in Ecology & Evolution*, 31(9), 670–676. DOI: <https://doi.org/10.1016/j.tree.2016.06.001>.
- 2564 Chen, I. C., Hill, J. K., Ohlemüller, R., Roy, D. B., and Thomas, C. D. (2011). Rapid range shifts of species
2565 associated with high levels of climate warming. *Science*, 333(6045), 1024–1026. DOI: 10.1126/science.1206432.
- 2567 Chichorro, F., Juslén, A., and Cardoso, P. (2019). A review of the relation between species traits and extinction
2568 risk. *Biological Conservation*, 237, 220–229. DOI: 10.1016/j.biocon.2019.07.001.
- 2569 Chown, S. L., Gaston, K. J., and Robinson, D. (2004). Macrophysiology: large-scale patterns in physiological
2570 traits and their ecological implications. *Functional Ecology*, 18(2), 159–167. DOI: 10.1111/j.0269-
2571 8463.2004.00825.x.
- 2572 Chown, S. L., Hoffmann, A. A., Kristensen, T. N., Angilletta, M. J., Stenseth, N. C., and Pertoldi, C. (2010).
2573 Adapting to climate change: A perspective from evolutionary physiology. *Climate Research*, 43(1-2),
2574 3–15. DOI: 10.3354/cr00879.
- 2575 Civantos, E., Thuiller, W., Maiorano, L., Guisan, A., and Arajo, M. B. (2012). Potential impacts of climate
2576 change on ecosystem services in Europe: The case of pest control by vertebrates. *BioScience*, 62(7),
2577 658–666. DOI: 10.1525/bio.2012.62.7.8.
- 2578 Clarke, A. and Fraser, K. P. (2004). Why does metabolism scale with temperature? *Functional Ecology*,
2579 18(2), 243–251. DOI: 10.1111/j.0269-8463.2004.00841.x.
- 2580 Colin, N., Villéger, S., Wilkes, M., de Sostoa, A., and Maceda-Veiga, A. (2018). Functional diversity measures
2581 revealed impacts of non-native species and habitat degradation on species-poor freshwater fish
2582 assemblages. *Science of the Total Environment*. DOI: 10.1016/j.scitotenv.2017.12.316.
- 2583 Collen, B., Ram, M., Zamin, T., and McRae, L. (2008). The Tropical Biodiversity Data Gap: Addressing
2584 Disparity in Global Monitoring. *Tropical Conservation Science*, 1(2), 75–88. DOI: 10.1177/
2585 194008290800100202.
- 2586 Collen, B., Dulvy, N. K., Gaston, K. J., Gärdenfors, U., Keith, D. A., Punt, A. E., Regan, H. M., Böhm,
2587 M., Hedges, S., Seddon, M., Butchart, S. H., Hilton-Taylor, C., Hoffmann, M., Bachman, S. P., and
2588 Akçakaya, H. R. (2016). Clarifying misconceptions of extinction risk assessment with the IUCN Red
2589 List. *Biology Letters*, 12(4), 20150843. DOI: 10.1098/rsbl.2015.0843.
- 2590 Comber, A. J., Wadsworth, R. A., and Fisher, P. F. (2008). Using semantics to clarify the conceptual confusion
2591 between land cover and land use: The example of ‘forest’. *Journal of Land Use Science*, 3(2-3),
2592 185–198. DOI: 10.1080/17474230802434187.
- 2593 Convention on Biological Diversity (2022). *Report of the open-ended working group on the Post-2020 Global
2594 Biodiversity Framework on its third meeting (part II)*. Tech. rep. United Nations Convention on Biological
2595 Diversity.
- 2596 Coogan, S. C., Raubenheimer, D., Zantis, S. P., and Machovsky-Capuska, G. E. (2018). Multidimensional
2597 nutritional ecology and urban birds. *Ecosphere*, 9(4), e02177. DOI: 10.1002/ecs2.2177.

- 2598 Cooke, R. S., Bates, A. E., and Eigenbrod, F. (2019a). Global trade-offs of functional redundancy and
2599 functional dispersion for birds and mammals. *Global Ecology and Biogeography*, 28(4), 484–495. DOI:
2600 10.1111/geb.12869.
- 2601 Cooke, R. S., Eigenbrod, F., and Bates, A. E. (2019b). Projected losses of global mammal and bird ecological
2602 strategies. *Nature Communications*, 10(2279). DOI: 10.1038/s41467-019-10284-z.
- 2603 Cooper, N., Bielby, J., Thomas, G. H., and Purvis, A. (2008). Macroecology and extinction risk correlates of
2604 frogs. *Global Ecology and Biogeography*, 17(2), 211–221. DOI: 10.1111/j.1466-8238.2007.
2605 00355.x.
- 2606 Cornwell, W. K., Schwilk, D. W., and Ackerly, D. D. (2006). A trait-based test for habitat filtering: Convex
2607 hull volume. *Ecology*, 87(6), 1465–1471. DOI: 10.1890/0012-9658 (2006)87[1465:ATTFHF]
2608 2.0.CO;2.
- 2609 Courchamp, F., Jaric, I., Albert, C., Meinard, Y., Ripple, W. J., and Chapron, G. (Apr. 2018). The paradoxical
2610 extinction of the most charismatic animals. *PLOS Biology*, 16(4), 1–13. DOI: 10.1371/journal.
2611 pbio.2003997.
- 2612 Cox, N. et al. (2022). A global reptile assessment highlights shared conservation needs of tetrapods. *Nature*,
2613 605, 285–290. DOI: 10.1038/s41586-022-04664-7.
- 2614 Crooks, K. and Soulé, M. (2010). Mesopredator release and avifaunal extinctions in a fragmented system.
2615 *Nature*, 400, 563–566. DOI: 10.1038/23028.
- 2616 Crowley, T. J. (2000). Causes of climate change over the past 1000 years. *Science*, 289, 270–277. DOI:
2617 10.1126/science.289.5477.270.
- 2618 Crutzen, P. and Stoermer, E. (2000). Sustaining Earth's Life Support Systems – the Challenge for the Next
2619 Decade and Beyond. *IGBP newsletter*, 41, 17–18.
- 2620 Cunningham, C. X., Johnson, C. N., Barmuta, L. A., Hollings, T., Woehler, E. J., and Jones, M. E. (2018).
2621 Top carnivore decline has cascading effects on scavengers and carrion persistence. *Proceedings of the
2622 Royal Society B: Biological Sciences*, 285(1892), 20181582. DOI: 10.1098/rspb.2018.1582.
- 2623 Damgaard, C. (2019). A Critique of the Space-for-Time Substitution Practice in Community Ecology. *Trends
2624 in Ecology and Evolution*, 34(5), 416–421. DOI: 10.1016/j.tree.2019.01.013.
- 2625 Daru, B. H., Davies, T. J., Willis, C. G., Meineke, E. K., Ronk, A., Zobel, M., Pärtel, M., Antonelli, A.,
2626 and Davis, C. C. (2021). Widespread homogenization of plant communities in the Anthropocene. *Nature
2627 Communications*, 12(1), 1–10. DOI: 10.1038/s41467-021-27186-8.
- 2628 Davison, C. W., Rahbek, C., and Morueta-Holme, N. (2021). Land-use change and biodiversity: Challenges
2629 for assembling evidence on the greatest threat to nature. *Global Change Biology*, 27(21), 5414–5429.
2630 DOI: 10.1111/gcb.15846.
- 2631 De Palma, A., Kuhlmann, M., Roberts, S. P., Potts, S. G., Börger, L., Hudson, L. N., Lysenko, I., Newbold,
2632 T., and Purvis, A. (2015). Ecological traits affect the sensitivity of bees to land-use pressures in European
2633 agricultural landscapes. *Journal of Applied Ecology*, 52(6), 1567–1577. DOI: 10.1111/1365-2664.
2634 12524.
- 2635 De Palma, A., Sánchez-Ortiz, K., Martin, P. A., Chadwick, A., Gilbert, G., Bates, A. E., Börger, L., Contu,
2636 S., Hill, S. L., and Purvis, A. (2018). Challenges With Inferring How Land-Use Affects Terrestrial Bio-
2637 diversity: Study Design, Time, Space and Synthesis. 58, 163–199. DOI: 10.1016/bs.aecr.2017.
2638 12.004.

- 2639 De Vos, J. M., Joppa, L. N., Gittleman, J. L., Stephens, P. R., and Pimm, S. L. (2015). Estimating the normal
2640 background rate of species extinction. *Conservation Biology*, 29(2), 452–462. DOI: 10.1111/cobi.12380.
- 2641
- 2642 Debastiani, V. J., Bastazini, V. A. G., and Pillar, D (2021). Using phylogenetic information to impute missing
2643 functional trait values in ecological databases. *Ecological Informatics*, 63, 101315. DOI: 10.1016/j.ecoinf.2021.101315.
- 2644
- 2645 Dee, L. E., Cowles, J., Isbell, F., Pau, S., Gaines, S. D., and Reich, P. B. (2019). When Do Ecosystem Services
2646 Depend on Rare Species? *Trends in Ecology and Evolution*, 34(8), 746–758. DOI: 10.1016/j.tree.2019.03.010.
- 2647
- 2648 Des Roches, S., Post, D. M., Turley, N. E., Bailey, J. K., Hendry, A. P., Kinnison, M. T., Schweitzer, J. A.,
2649 and Palkovacs, E. P. (2018). The ecological importance of intraspecific variation. *Nature Ecology and
2650 Evolution*, 2, 57–64. DOI: 10.1038/s41559-017-0402-5.
- 2651
- 2652 Deutsch, C. A., Tewksbury, J. J., Huey, R. B., Sheldon, K. S., Ghalambor, C. K., Haak, D. C., and Martin,
2653 P. R. (2008). Impacts of climate warming on terrestrial ectotherms across latitude. *Proceedings of the
2654 National Academy of Sciences of the United States of America*, 105(18), 6668–6672. DOI: 10.1073/pnas.0709472105.
- 2655
- 2656 Di Marco, M., Pacifici, M., Maiorano, L., and Rondinini, C. (2021). Drivers of change in the realised climatic
niche of terrestrial mammals. *Ecography*, 44(8), 1180–1190. DOI: 10.1111/ecog.05414.
- 2657
- 2658 Díaz, S. et al. (2016). The global spectrum of plant form and function. *Nature*, 529, 167–171. DOI: 10.1038/nature16489.
- 2659
- 2660 Diniz-Filho, J. A. F., Bini, L. M., Rangel, T. F., Morales-Castilla, I., Olalla-Tárraga, M. Á., Rodríguez,
2661 M. Á., and Hawkins, B. A. (2012). On the selection of phylogenetic eigenvectors for ecological analyses.
Ecography, 35, 239–249. DOI: 10.1111/j.1600-0587.2011.06949.x.
- 2662
- 2663 Dirzo, R., Young, H. S., Galetti, M., Ceballos, G., Isaac, N. J., and Collen, B. (2014). Defaunation in the
Anthropocene. *Science*, 345(6195), 401–406. DOI: 10.1126/science.1251817.
- 2664
- 2665 Dore, M. H. (2005). Climate change and changes in global precipitation patterns: What do we know? *Environment
International*, 31(8), 1167–1181. DOI: 10.1016/j.envint.2005.03.004.
- 2666
- 2667 Dornelas, M., Gotelli, N. J., Shimadzu, H., Moyes, F., Magurran, A. E., and McGill, B. J. (2019). A balance
2668 of winners and losers in the Anthropocene. *Ecology Letters*, 22(5), 847–854. DOI: 10.1111/ele.13242.
- 2669
- 2670 Duceatz, S. and Shine, R. (2017). Drivers of Extinction Risk in Terrestrial Vertebrates. *Conservation Letters*,
10(2), 186–194. DOI: 10.1111/conl.12258.
- 2671
- 2672 Duffy, J. E. (2003). Biodiversity loss, trophic skew and ecosystem functioning. *Ecology Letters*, 6(8), 680–
687. DOI: 10.1046/j.1461-0248.2003.00494.x.
- 2673
- 2674 Dullinger, I., Essl, F., Moser, D., Erb, K., Haberl, H., and Dullinger, S. (2021). Biodiversity models need to
2675 represent land-use intensity more comprehensively. *Global Ecology and Biogeography*, 30(5), 924–932.
DOI: 10.1111/geb.13289.
- 2676
- 2677 Edwards, F. A., Edwards, D. P., Hamer, K. C., and Fayle, T. M. (2021). Tropical land-use change alters
2678 trait-based community assembly rules for dung beetles and birds. *Oecologia*, 195(3), 705–717. DOI:
10.1007/s00442-020-04829-z.

- 2679 Eeden, L. M. van et al. (2018). Carnivore conservation needs evidence-based livestock protection. *PLoS Biology*, 16(9), 1–8. DOI: 10.1371/journal.pbio.2005577.
- 2680
- 2681 Ellis, E. C., Goldewijk, K. K., Siebert, S., Lightman, D., and Ramankutty, N. (2010). Anthropogenic trans-
2682 formation of the biomes, 1700 to 2000. *Global Ecology and Biogeography*, 19(5), 589–606. DOI: 10 .
2683 1111/j.1466-8238.2010.00540.x.
- 2684 Ellis, E. C., Kaplan, J. O., Fuller, D. Q., Vavrus, S., Goldewijk, K. K., and Verburg, P. H. (2013). Used planet:
2685 A global history. *Proceedings of the National Academy of Sciences of the United States of America*,
2686 110(20), 7978–7985. DOI: 10.1073/pnas.1217241110.
- 2687 Ellis, E. C., Gauthier, N., Goldewijk, K. K., Bird, R. B., Boivin, N., Díaz, S., Fuller, D. Q., Gill, J. L., Kaplan,
2688 J. O., Kingston, N., Locke, H., McMichael, C. N., Ranco, D., Rick, T. C., Rebecca Shaw, M., Stephens,
2689 L., Svenning, J. C., and Watson, J. E. (2021). People have shaped most of terrestrial nature for at least
2690 12,000 years. *Proceedings of the National Academy of Sciences of the United States of America*, 118(17),
2691 1–8. DOI: 10.1073/pnas.2023483118.
- 2692 Escudero, A. and Valladares, F. (2016). Trait-based plant ecology: moving towards a unifying species coex-
2693 istence theory: Features of the Special Section. *Oecologia*, 180, 919–922. DOI: 10 .1007/s00442–
2694 016–3578–5.
- 2695 ESRI (2011). ArcGIS desktop: Release 10. *Environmental Systems Research Institute*.
- 2696 Estrada, A., Morales-Castilla, I., Meireles, C., Caplat, P., and Early, R. (2018). Equipped to cope with climate
2697 change: traits associated with range filling across European taxa. *Ecography*, 41, 770–781. DOI: 10 .
2698 1111/ecog.02968.
- 2699 Etard, A., Morrill, S., and Newbold, T. (2020). Global gaps in trait data for terrestrial vertebrates. *Global
2700 Ecology and Biogeography*, 29(12), 2143–2158. DOI: 10 .1111/geb.13184.
- 2701 Etard, A., Pigot, A. L., and Newbold, T. (2022). Intensive human land uses negatively affect vertebrate
2702 functional diversity. *Ecology Letters*, 25(2), 330–343. DOI: <https://doi.org/10.1111/ele.13926>.
- 2703
- 2704 Evans, B. S., Reitsma, R., Hurlbert, A. H., and Marra, P. P. (2018). Environmental filtering of avian com-
2705 munities along a rural-to-urban gradient in Greater Washington, D.C., USA. *Ecosphere*, 9(11), e02402.
2706 DOI: 10 .1002/ecs2.2402.
- 2707 Faurby, S., Davis, M., Pedersen, R., Schowanek, S. D., Antonelli1, A., and Svenning, J.-C. (2018). PHY-
2708 LACINE 1.2: The Phylogenetic Atlas of Mammal Macroecology. *Ecology*, 99(11), 2626–2626. DOI:
2709 10 .1002/ecy.2443.
- 2710 Faurby, S., Pedersen, R., Davis, M., Schowanek, S. D., Jarvie, S., Antonelli, A., and Svenning, J.-C. (2020).
2711 PHYLACINE 1.2.1: An update to the Phylogenetic Atlas of Mammal Macroecology. DOI: 10 .5281/
2712 zenodo .3690867.
- 2713 Feldman, A., Sabath, N., Pyron, R. A., Mayrose, I., and Meiri, S. (2016). Body sizes and diversification rates
2714 of lizards, snakes, amphisbaenians and the tuatara. *Global Ecology and Biogeography*, 25(2), 187–197.
2715 DOI: 10 .1111/geb.12398.
- 2716 Ferreira, D. F., Gibb, R., López-Baucells, A., Nunes, N. J., Jones, K. E., and Rocha, R. (2022). Species-
2717 specific responses to land-use change in island insectivorous bats. *Journal for Nature Conservation*, 67,
2718 126177. DOI: 10 .1016/j.jnc.2022.126177.

- 2719 Fick, S. E. and Hijmans, R. J. (2017). WorldClim 2: new 1-km spatial resolution climate surfaces for global
2720 land areas. *International Journal of Climatology*, 37(12), 4302–4315. DOI: 10.1002/joc.5086.
- 2721 Finderup Nielsen, T., Sand-Jensen, K., Dornelas, M., and Bruun, H. H. (2019). More is less: net gain in
2722 species richness, but biotic homogenization over 140 years. *Ecology Letters*, 22(10), 1650–1657. DOI:
2723 10.1111/ele.13361.
- 2724 Fischer, J. D., Cleton, S. H., Lyons, T. P., and Miller, J. R. (2012). Urbanization and the predation paradox:
2725 The role of trophic dynamics in structuring vertebrate communities. *BioScience*, 62(9), 809–818. DOI:
2726 10.1525/bio.2012.62.9.6.
- 2727 Flynn, D. F., Gogol-Prokurat, M., Nogeire, T., Molinari, N., Richers, B. T., Lin, B. B., Simpson, N., May-
2728 field, M. M., and DeClerck, F. (2009). Loss of functional diversity under land use intensification across
2729 multiple taxa. *Ecology Letters*, 12(1), 22–33. DOI: 10.1111/j.1461-0248.2008.01255.x.
- 2730 Foden, W. B., Butchart, S. H. M., Stuart, S. N., Vié, J. C., Akçakaya, H. R., Angulo, A., DeVantier, L. M.,
2731 Gutsche, A., Turak, E., Cao, L., Donner, S. D., Katariya, V., Bernard, R., Holland, R. A., Hughes, A. F.,
2732 O'Hanlon, S. E., Garnett, S. T., Şekercioğlu, Ç. H., and Mace, G. M. (2013). Identifying the World's Most
2733 Climate Change Vulnerable Species: A Systematic Trait-Based Assessment of all Birds, Amphibians and
2734 Corals. *PLoS ONE*, 8(6), 1–13. DOI: 10.1371/journal.pone.0065427.
- 2735 Foley, J. A., DeFries, R., Asner, G. P., Barford, C., Bonan, G., Carpenter, S. R., Chapin, F. S., Coe, M. T.,
2736 Daily, G. C., Gibbs, H. K., Helkowski, J. H., Holloway, T., Howard, E. A., Kucharik, C. J., Monfreda, C.,
2737 Patz, J. A., Prentice, I. C., Ramankutty, N., and Snyder, P. K. (2005). Global consequences of land use.
2738 *Science*, 309(5734), 570–574. DOI: 10.1126/science.1111772.
- 2739 Fournier, B., Mouly, A., and Gillet, F. (2016). Multiple assembly rules drive the co-occurrence of orthopteran
2740 and plant species in grasslands: Combining network, functional and phylogenetic approaches. *Frontiers
2741 in Plant Science*, 7(1224), 1–12. DOI: 10.3389/fpls.2016.01224.
- 2742 Fox, J. and Monette, G. (1992). Generalized Collinearity Diagnostics. *Journal of the American Statistical
2743 Association*, 87(417), 178–183. DOI: 10.1080/01621459.1992.10475190.
- 2744 Freeman, B. G. and Pennell, M. W. (2021). The latitudinal taxonomy gradient. *Trends in Ecology and Evolution*,
2745 36(9), 778–786. DOI: 10.1016/j.tree.2021.05.003.
- 2746 Fricke, E. C., Ordonez, A., Rogers, H. S., and Svenning, J.-C. (2022). The effects of defaunation on plants'
2747 capacity to track climate change. *Science*, 375(6577), 210–214. DOI: 10.1126/science.abk3510.
- 2748 Frishkoff, L. O., Karp, D. S., Flanders, J. R., Zook, J., Hadly, E. A., Daily, G. C., and M'Gonigle, L. K.
2749 (2016). Climate change and habitat conversion favour the same species. *Ecology letters*, 19(9), 1081–
2750 1090. DOI: 10.1111/ele.12645.
- 2751 Fritz, S. A., Bininda-Emonds, O. R. P., and Purvis, A. (2009). Geographical variation in predictors of
2752 mammalian extinction risk: big is bad, but only in the tropics. *Ecology Letters*, 12(6), 538–549. DOI:
2753 10.1111/j.1461-0248.2009.01307.x.
- 2754 Fukasawa, K. and Akasaka, T. (2019). Long-lasting effects of historical land use on the current distribution
2755 of mammals revealed by ecological and archaeological patterns. *Scientific Reports*, 9(1), 1–11. DOI:
2756 10.1038/s41598-019-46809-1.
- 2757 Furness, L. J. and Speakman, J. R. (2008). Energetics and longevity in birds. *AGE*, 30, 75–87. DOI: 10.
2758 1007/s11357-008-9054-3.

- 2759 Gainsbury, A. M., Tallowin, O. J., and Meiri, S. (2018). An updated global data set for diet preferences in
2760 terrestrial mammals: testing the validity of extrapolation. *Mammal Review*, 48(3), 160–167. DOI: 10 .
2761 1111/mam.12119.
- 2762 Galán-Acedo, C., Arroyo-Rodríguez, V., Andresen, E., and Arasa-Gisbert, R. (2019). Ecological traits of the
2763 world's primates. *Scientific data*, 6(55). DOI: 10 .1038/s41597-019-0059-9.
- 2764 Gallmetzer, N. and Schulze, C. H. (2015). Impact of oil palm agriculture on understory amphibians and
2765 reptiles: A Mesoamerican perspective. *Global Ecology and Conservation*, 4, 95–109. DOI: 10 .1016/
2766 j.gecco.2015.05.008.
- 2767 Geffroy, B., Sadoul, B., Putman, B. J., Berger-Tal, O., Garamszegi, L. Z., Møller, A. P., and Blumstein, D. T.
2768 (2020). Evolutionary dynamics in the anthropocene: Life history and intensity of human contact shape
2769 antipredator responses. *PLoS Biology*, 18(9), 1–17. DOI: 10 .1371/journal.pbio.3000818.
- 2770 Gillooly, J. F., Brown, J. H., West, G. B., Savage, V. M., and Charnov, E. L. (2001). Effects of Size and Tem-
2771 perature on Metabolic Rate. *Science*, 293(5538), 2248–2251. DOI: 10 .1126/science.1061967.
- 2772 Gnanadesikan, G. E., Pearse, W. D., and Shaw, A. K. (2017). Evolution of mammalian migrations for refuge,
2773 breeding, and food. *Ecology and Evolution*, 7(15), 5891–5900. DOI: 10 .1002/ece3.3120.
- 2774 Goldewijk, K. K., Beusen, A., Doelman, J., and Stehfest, E. (2017). Anthropogenic land use estimates for the
2775 Holocene - HYDE 3.2. *Earth System Science Data*, 9(2), 927–953. DOI: 10 .5194/essd-9-927-
2776 2017.
- 2777 González-Suárez, M. and Revilla, E. (2012). Variability in life-history and ecological traits is a buffer against
2778 extinction in mammals. *Ecology Letters*, 16(2), 242–251. DOI: 10 .1111/ele.12035.
- 2779 González-Suárez, M., Lucas, P. M., and Revilla, E. (2012). Biases in comparative analyses of extinction risk:
2780 Mind the gap. *Journal of Animal Ecology*, 81(6), 1211–1222. DOI: 10 .1111/j.1365-2656.2012.
2781 01999.x.
- 2782 González-Suárez, M., Gómez, A., and Revilla, E. (2013). Which intrinsic traits predict vulnerability to ex-
2783 tinction depends on the actual threatening processes. *Ecosphere*, 4(6), 76. DOI: 10 .1890 / ES12-
2784 00380 .1.
- 2785 González-Suárez, M., Zanchetta Ferreira, F., and Grilo, C. (2018). Spatial and species-level predictions of
2786 road mortality risk using trait data. *Global Ecology and Biogeography*, 27(9), 1093–1105. DOI: 10 .
2787 1111/geb.12769.
- 2788 González-Varo, J. P., Biesmeijer, J. C., Bommarco, R., Potts, S. G., Schweiger, O., Smith, H. G., Steffan-
2789 Dewenter, I., Szentgyörgyi, H., Woyciechowski, M., and Vilà, M. (2013). Combined effects of global
2790 change pressures on animal-mediated pollination. *Trends in Ecology and Evolution*, 28(9), 524–530.
2791 DOI: 10 .1016/j.tree.2013.05.008.
- 2792 Grekousis, G., Mountrakis, G., and Kavouras, M. (2015). An overview of 21 global and 43 regional land-
2793 cover mapping products. *International Journal of Remote Sensing*, 36(21), 5309–5335. DOI: 10 .1080/
2794 01431161.2015.1093195.
- 2795 Guralnick, R. P., Zermoglio, P. F., Wieczorek, J., LaFrance, R., Bloom, D., and Russell, L. (2016). The
2796 importance of digitized biocollections as a source of trait data and a new VertNet resource. *Database*,
2797 2016(baw158). DOI: 10 .1093/database/baw158.
- 2798 Hadfield, J. D. (2010). MCMC Methods for Multi-Response Generalized Linear Mixed Models: The MCM-
2799 Cglmm R Package. *Journal of Statistical Software*, 33(2), 1–22. DOI: 10 .18637/jss.v033.i02.

- 2800 Hamilton, A. T., Schäfer, R. B., Pyne, M. I., Chessman, B., Kakouei, K., Boersma, K. S., Verdonschot, P. F., Verdonschot, R. C., Mims, M., Khamis, K., Bierwagen, B., and Stamp, J. (2020). Limitations of trait-based approaches for stressor assessment: The case of freshwater invertebrates and climate drivers. *Global Change Biology*, 26(2), 364–379. DOI: 10.1111/gcb.14846.
- 2801
2802
2803
- 2804 Hansen, M. C., Potapov, P. V., Moore, R., Hancher, M., Turubanova, S. A., Tyukavina, A., Thau, D., Stehman, S. V., Goetz, S. J., Loveland, T. R., Kommareddy, A., Egorov, A., Chini, L., Justice, C. O., and Townshend, J. R. G. (2013). High-Resolution Global Maps of 21st-Century Forest Cover Change. *Science*, 342(6160), 850–853. DOI: 10.1126/science.1244693.
- 2805
2806
2807
- 2808 Hantak, M. M., McLean, B. S., Li, D., and Guralnick, R. P. (2021). Mammalian body size is determined by interactions between climate, urbanization, and ecological traits. *Communications Biology*, 4(1), 1–10. DOI: 10.1038/s42003-021-02505-3.
- 2809
2810
- 2811 Harfoot, M. B., Johnston, A., Balmford, A., Burgess, N. D., Butchart, S. H., Dias, M. P., Hazin, C., Hilton-Taylor, C., Hoffmann, M., Isaac, N. J., Iversen, L. L., Outhwaite, C. L., Visconti, P., and Geldmann, J. (2021). Using the IUCN Red List to map threats to terrestrial vertebrates at global scale. *Nature Ecology and Evolution*, 5(11), 1510–1519. DOI: 10.1038/s41559-021-01542-9.
- 2812
2813
2814
- 2815 Hatfield, J. H., Harrison, M. L. K., and Banks-Leite, C. (2018). Functional Diversity Metrics: How They Are Affected by Landscape Change and How They Represent Ecosystem Functioning in the Tropics. *Current Landscape Ecology Reports*, 3, 35–42. DOI: 10.1007/s40823-018-0032-x.
- 2816
2817
- 2818 Hernández-Yáñez, H., Kim, S. Y., and Che-Castaldo, J. P. (2022). Demographic and life history traits explain patterns in species vulnerability to extinction. *PLoS ONE*, 17(2), e0263504. DOI: 10.1371/journal.pone.0263504.
- 2819
2820
- 2821 Hevia, V., Martín-López, B., Palomo, S., García-Llorente, M., de Bello, F., and González, J. A. (2017). Trait-based approaches to analyze links between the drivers of change and ecosystem services: Synthesizing existing evidence and future challenges. *Ecology and Evolution*, 7(3), 831–844. DOI: <https://doi.org/10.1002/ece3.2692>.
- 2822
2823
2824
- 2825 Hijmans, R. J. (2022). *raster: Geographic Data Analysis and Modeling*. R package version 3.5-15.
- 2826 Hillaert, J., Hovestadt, T., Vandegehuchte, M. L., and Bonte, D. (2018). Size-dependent movement explains why bigger is better in fragmented landscapes. *Ecology and Evolution*, 8(22), 10754–10767. DOI: 10.1002/ece3.4524.
- 2827
2828
- 2829 Hillaert, J., Vandegehuchte, M. L., Hovestadt, T., and Bonte, D. (2020). Habitat loss and fragmentation increase realized predator-prey body size ratios. *Functional Ecology*, 34(2), 534–544. DOI: 10.1111/1365-2435.13472.
- 2830
2831
- 2832 Hiron, M., Comberti, C., and Dunford, R. (2016). Valuing Cultural Ecosystem Services. *Annual Review of Environment and Resources*, 41, 545–574. DOI: 10.1146/annurev-environ-110615-085831.
- 2833
- 2834 Hoban, S. et al. (2020). Genetic diversity targets and indicators in the CBD post-2020 Global Biodiversity Framework must be improved. *Biological Conservation*, 248, 108654. DOI: 10.1016/j.biocon.2020.108654.
- 2835
2836
- 2837 Hooper, D. U., Chapin, F. S., Ewel, J. J., Hector, A., Inchausti, P., Lavorel, S., Lawton, J. H., Lodge, D. M., Loreau, M., Naeem, S., Schmid, B., Setälä, H., Symstad, A. J., Vandermeer, J., and Wardle, D. A. (2005). Effects of biodiversity on ecosystem functioning: A consensus of current knowledge. *Ecological Monographs*, 75, 3–35. DOI: 10.1890/04-0922.
- 2838
2839
2840

- 2841 Hooper, D. U., Adair, E. C., Cardinale, B. J., Byrnes, J. E., Hungate, B. A., Matulich, K. L., Gonzalez, A.,
2842 Duffy, J. E., Gamfeldt, L., and Connor, M. I. (2012). A global synthesis reveals biodiversity loss as a
2843 major driver of ecosystem change. *Nature*, 486, 105–108. DOI: 10.1038/nature11118.
- 2844 Hortal, J., Bello, F. de, Diniz-Filho, J. A. F., Lewinsohn, T. M., Lobo, J. M., and Ladle, R. J. (2015). Seven
2845 Shortfalls that Beset Large-Scale Knowledge of Biodiversity. *Annual Review of Ecology, Evolution, and*
2846 *Systematics*, 46(1), 523–549. DOI: 10.1146/annurev-ecolsys-112414-054400.
- 2847 Hou, Y., Ding, W., Liu, C., Li, K., Cui, H., Liu, B., and Chen, W. (2022). Influences of impervious sur-
2848 faces on ecological risks and controlling strategies in rapidly urbanizing regions. *Science of The Total*
2849 *Environment*, 825, 153823. DOI: 10.1016/j.scitotenv.2022.153823.
- 2850 Hudson, L. N., Isaac, N. J., and Reuman, D. C. (2013). The relationship between body mass and field
2851 metabolic rate among individual birds and mammals. *Journal of Animal Ecology*, 82(5), 1009–1020.
2852 DOI: 10.1111/1365-2656.12086.
- 2853 Hudson, L. N. et al. (2014). The PREDICTS database: A global database of how local terrestrial biodiversity
2854 responds to human impacts. *Ecology and Evolution*, 4(24), 4701–4735. DOI: 10.1002/ece3.1303.
- 2855 — (2017). The database of the PREDICTS (Projecting Responses of Ecological Diversity In Changing
2856 Terrestrial Systems) project. *Ecology and Evolution*, 7(1), 145–188. DOI: 10.1002/ece3.2579.
- 2857 Inger, R., Cox, D. T., Per, E., Norton, B. A., and Gaston, K. J. (2016). Ecological role of vertebrate scavengers
2858 in urban ecosystems in the UK. *Ecology and Evolution*, 6(19), 7015–7023. DOI: 10.1002/ece3 .
2859 2414.
- 2860 Inouye, D. W. (2022). Climate change and phenology. *Wiley Interdisciplinary Reviews: Climate Change*,
2861 13(3), e764. DOI: 10.1002/wcc.764.
- 2862 Intergovernmental Panel on Climate Change (2013). *Climate Change 2013 – The Physical Science Basis*.
2863 Tech. rep. Available at https://www.ipcc.ch/site/assets/uploads/2018/03/WG1AR5_SummaryVolume_FINAL.pdf. WMO, UNEP.
- 2864 — (2022). *Climate Change 2022 – Mitigation of Climate Change – Summary for Policymakers*. Tech. rep.
2865 Available at https://report.ipcc.ch/ar6wg3/pdf/IPCC_AR6_WGIII_SummaryForPolicymakers.pdf. WMO, UNEP.
- 2866 Isaac, N. J., Mallet, J., and Mace, G. M. (2004). Taxonomic inflation: Its influence on macroecology and
2867 conservation. *Trends in Ecology and Evolution*, 19(9), 464–469. DOI: 10.1016/j.tree.2004.06.
2870 004.
- 2871 Isler, K. and Van Schaik, C. P. (2006). Metabolic costs of brain size evolution. *Biology Letters*, 2(4), 557–
2872 560. DOI: 10.1098/rsbl.2006.0538.
- 2873 IUCN (2020). The IUCN Red List of Threatened Species. Version 2020.1.
- 2874 Ives, C. D., Lentini, P. E., Threlfall, C. G., Ikin, K., Shanahan, D. F., Garrard, G. E., Bekessy, S. A., Fuller,
2875 R. A., Mumaw, L., Rayner, L., Rowe, R., Valentine, L. E., and Kendal, D. (2016). Cities are hotspots for
2876 threatened species. *Global Ecology and Biogeography*, 25(1), 117–126. DOI: 10.1111/geb.12404.
- 2877 Jayathilake, H. M., Prescott, G. W., Carrasco, L. R., Rao, M., and Symes, W. S. (2021). Drivers of deforesta-
2878 tion and degradation for 28 tropical conservation landscapes. *Ambio*, 50(1), 215–228. DOI: 10.1007/
2879 s13280-020-01325-9.

- 2880 Jenkins, C. N., Pimm, S. L., and Joppa, L. N. (2013). Global patterns of terrestrial vertebrate diversity and
2881 conservation. *Proceedings of the National Academy of Sciences of the United States of America*, 110(28),
2882 E2603–E2610. DOI: 10.1073/pnas.1302251110.
- 2883 Jenkins, D. G., Brescacin, C. R., Duxbury, C. V., Elliott, J. A., Evans, J. A., Grablom, K. R., Hillegass, M.,
2884 Lyon, B. N., Metzger, G. A., Olandese, M. L., Pepe, D., Silvers, G. A., Suresch, H. N., Thompson,
2885 T. N., Trexler, C. M., Williams, G. E., Williams, N. C., and Williams, S. E. (2007). Does size matter
2886 for dispersal distance? *Global Ecology and Biogeography*, 16(4), 415–425. DOI: 10.1111/j.1466–
2887 8238.2007.00312.x.
- 2888 Jesse, W. A., Behm, J. E., Helmus, M. R., and Ellers, J. (2018). Human land use promotes the abundance
2889 and diversity of exotic species on Caribbean islands. *Global Change Biology*, 24(10), 4784–4796. DOI:
2890 10.1111/gcb.14334.
- 2891 Jetz, W., Thomas, G. H., Joy, J. B., Hartmann, K., and Mooers, A. O. (2012). The global diversity of birds in
2892 space and time. *Nature*, 491, 444–448. DOI: 10.1038/nature11631.
- 2893 Jetz, W. and Pyron, R. A. (2018). The interplay of past diversification and evolutionary isolation with present
2894 imperilment across the amphibian tree of life. *Nature Ecology and Evolution*, 2, 850–858. DOI: 10.
2895 1038/s41559-018-0515-5.
- 2896 Johnson, C. N., Alroy, J., Beeton, N. J., Bird, M. I., Brook, B. W., Cooper, A., Gillespie, R., Herrando-Pérez,
2897 S., Jacobs, Z., Miller, G. H., Prideaux, G. J., Roberts, R. G., Rodríguez-Rey, M., Saltré, F., Turney, C. S.,
2898 and Bradshaw, C. J. (2016). What caused extinction of the pleistocene megafauna of Sahul? *Proceedings
2899 of the Royal Society B: Biological Sciences*, 283(1824), 20152399. DOI: 10.1098/rspb.2015.
2900 2399.
- 2901 Johnson, C. N., Balmford, A., Brook, B. W., Buettel, J. C., Galetti, M., Guangchun, L., and Wilmshurst,
2902 J. M. (2017). Biodiversity losses and conservation responses in the Anthropocene. *Science*, 356(6335),
2903 270–275. DOI: 10.1126/science.aam9317.
- 2904 Johnson, T. F., Isaac, N. J., Paviolo, A., and González-Suárez, M. (2021). Handling missing values in trait
2905 data. *Global Ecology and Biogeography*, 30(1), 51–62. DOI: 10.1111/geb.13185.
- 2906 Jones, K. E. et al. (2009). PanTHERIA: a species-level database of life history, ecology, and geography of
2907 extant and recently extinct mammals. *Ecology*, 90(9), 2648–2648. DOI: 10.1890/08-1494.1.
- 2908 Jones, O. R., Purvis, A., and Quicke, D. L. (2012). Latitudinal gradients in taxonomic overdescription rate
2909 affect macroecological inferences using species list data. *Ecography*, 35(4), 333–340. DOI: 10.1111/
2910 j.1600-0587.2011.06956.x.
- 2911 Junker, R. R., Albrecht, J., Becker, M., Keuth, R., Farwig, N., and Schleuning, M. (2022). Towards an animal
2912 economics spectrum for ecosystem research. *Functional Ecology*, 00, 1–16. DOI: 10.1111/1365–
2913 2435.14051.
- 2914 Kattge, J. et al. (2011). TRY - a global database of plant traits. *Global Change Biology*, 17(9), 2905–2935.
2915 DOI: 10.1111/j.1365-2486.2011.02451.x.
- 2916 Kehoe, L., Kuemmerle, T., Meyer, C., Levers, C., Václavík, T., and Kreft, H. (2015). Global patterns of
2917 agricultural land-use intensity and vertebrate diversity. *Diversity and Distributions*, 21(11), 1308–1318.
2918 DOI: 10.1111/ddi.12359.
- 2919 Kharouba, H. M., Ehrlén, J., Gelman, A., Bolmgren, K., Allen, J. M., Travers, S. E., and Wolkovich, E. M.
2920 (2018). Global shifts in the phenological synchrony of species interactions over recent decades. *Pro-*

- 2921 *ceedings of the National Academy of Sciences of the United States of America*, 115(20), 5211–5216.
2922 DOI: 10.1073/pnas.1714511115.
- 2923 Kissling, W. D. et al. (2018). Towards global data products of Essential Biodiversity Variables on species
2924 traits. *Nature Ecology and Evolution*, 2, 1531–1540. DOI: 10.1038/s41559-018-0667-3.
- 2925 Kissling, W. D., Dalby, L., Fløjgaard, C., Lenoir, J., Sandel, B., Sandom, C., Trøjelsgaard, K., and Svenning,
2926 J. C. (2014). Establishing macroecological trait datasets: Digitalization, extrapolation, and validation
2927 of diet preferences in terrestrial mammals worldwide. *Ecology and Evolution*, 4(14), 2913–2930. DOI:
2928 10.1002/ece3.1136.
- 2929 Krausmann, F., Erb, K. H., Gingrich, S., Haberl, H., Bondeau, A., Gaube, V., Lauk, C., Plutzar, C., and
2930 Searchinger, T. D. (2013). Global human appropriation of net primary production doubled in the 20th
2931 century. *Proceedings of the National Academy of Sciences of the United States of America*, 110(25),
2932 10324–10329. DOI: 10.1073/pnas.1211349110.
- 2933 Krauss, J., Bommarco, R., Guardiola, M., Heikkinen, R. K., Helm, A., Kuussaari, M., Lindborg, R., Öckinger,
2934 E., Pärtel, M., Pino, J., Pöyry, J., Raatikainen, K. M., Sang, A., Stefanescu, C., Teder, T., Zobel, M., and
2935 Steffan-Dewenter, I. (2010). Habitat fragmentation causes immediate and time-delayed biodiversity loss
2936 at different trophic levels. *Ecology Letters*, 13(5), 597–605. DOI: <https://doi.org/10.1111/j.1461-0248.2010.01457.x>.
- 2938 La Sorte, F. A., Lepczyk, C. A., Aronson, M. F., Goddard, M. A., Hedblom, M., Katti, M., MacGregor-Fors,
2939 I., Mörtberg, U., Nilon, C. H., Warren, P. S., Williams, N. S., and Yang, J. (2018). The phylogenetic and
2940 functional diversity of regional breeding bird assemblages is reduced and constricted through urbaniza-
2941 tion. *Diversity and Distributions*, 24(7), 928–938. DOI: 10.1111/ddi.12738.
- 2942 Laliberté, E. and Legendre, P. (2010). A distance-based framework for measuring functional diversity from
2943 multiple traits. *Ecology*, 91(1), 299–305. DOI: 10.1890/08-2244.1.
- 2944 Laliberté, E., Legendre, P., and Shipley, B. (2014). FD: measuring functional diversity from multiple traits,
2945 and other tools for functional ecology. *R Package version 1.0-12*.
- 2946 Lambin, E. F. et al. (2001). The causes of land-use and land-cover change: moving beyond the myths. *Global
2947 Environmental Change*, 11(4), 261–269. DOI: 10.1016/S0959-3780(01)00007-3.
- 2948 Laurance, W. F. et al. (2012). Averting biodiversity collapse in tropical forest protected areas. *Nature*,
2949 489(7415), 290–294. DOI: 10.1038/nature11318.
- 2950 Lavorel, S. and Garnier, E. (2002). Predicting changes in community composition and ecosystem functioning
2951 from plant traits: Revisiting the Holy Grail. *Functional Ecology*, 16(5), 545–556. DOI: 10.1046/j.
2952 1365-2435.2002.00664.x.
- 2953 Lawler, J. J., Lewis, D. J., Nelson, E., Plantinga, A. J., Polasky, S., Withey, J. C., Helmers, D. P., Martin-
2954 uzzi, S., Pennington, D., and Radeloff, V. C. (2014). Projected land-use change impacts on ecosystem
2955 services in the United States. *Proceedings of the National Academy of Sciences of the United States of
2956 America*, 111(20), 7492–7497. DOI: 10.1073/pnas.1405557111.
- 2957 Le Berre, M., Noble, V., Pires, M., Médail, F., and Diadema, K. (2019). How to hierarchise species to
2958 determine priorities for conservation action? A critical analysis. *Biodiversity and Conservation*, 28(12),
2959 3051–3071. DOI: 10.1007/s10531-019-01820-w.
- 2960 Le Provost, G., Badenhausen, I., Le Bagousse-Pinguet, Y., Clough, Y., Henckel, L., Violle, C., Bretagnolle,
2961 V., Roncoroni, M., Manning, P., and Gross, N. (2020). Land-use history impacts functional diversity

- 2962 across multiple trophic groups. *Proceedings of the National Academy of Sciences of the United States of*
2963 *America*, 117(3), 1573–1579. DOI: 10.1073/pnas.1910023117.
- 2964 Lebreton, J. D. (2011). The impact of global change on terrestrial vertebrates. *Comptes Rendus - Biologies*,
2965 334(5-6), 360–369. DOI: 10.1016/j.crvi.2011.01.005.
- 2966 Legras, G., Loiseau, N., and Gaertner, J. C. (2018). Functional richness: Overview of indices and underlying
2967 concepts. *Acta Oecologica*, 87, 34–44. DOI: 10.1016/j.actao.2018.02.007.
- 2968 Lenoir, J. and Svenning, J. C. (2015). Climate-related range shifts - a global multidimensional synthesis and
2969 new research directions. *Ecography*, 38(1), 15–28. DOI: 10.1111/ecog.00967.
- 2970 Leroy, B., Meynard, C. N., Bellard, C., and Courchamp, F. (2015). virtualspecies, an R package to generate
2971 virtual species distributions. *Ecography*, 39(6), 599–607. DOI: 10.1111/ecog.01388.
- 2972 Letnic, M., Ritchie, E. G., and Dickman, C. R. (2012). Top predators as biodiversity regulators: The dingo
2973 *Canis lupus dingo* as a case study. *Biological Reviews*, 87(2), 390–413. DOI: 10.1111/j.1469–
2974 185X.2011.00203.x.
- 2975 Leung, B., Hargreaves, A. L., Greenberg, D. A., McGill, B., Dornelas, M., and Freeman, R. (2020). Clustered
2976 versus catastrophic global vertebrate declines. *Nature*, 588(7837), 267–271. DOI: 10.1038/s41586–
2977 020–2920–6.
- 2978 Lewis, S. L. and Maslin, M. A. (2015). Defining the Anthropocene. *Nature*, 519(7542), 171–180. DOI: 10.
2979 1038/nature14258.
- 2980 Li, G., Fang, C., Li, Y., Wang, Z., Sun, S., He, S., Qi, W., Bao, C., Ma, H., Fan, Y., Feng, Y., and Liu, X.
2981 (2022). Global impacts of future urban expansion on terrestrial vertebrate diversity. *Nature Communications*,
2982 13(1), 1–12. DOI: 10.1038/s41467–022–29324–2.
- 2983 Lin, F., Jia, S., Luskin, M. S., Ye, J., Hao, Z., Wang, X., and Yuan, Z. (2018). Global signal of top-down
2984 control of terrestrial plant communities by herbivores. *Proceedings of the National Academy of Sciences*,
2985 115(24), 6237–6242. DOI: 10.1073/pnas.1707984115.
- 2986 Livesley, S. J., McPherson, E. G., and Calfapietra, C. (2016). The Urban Forest and Ecosystem Services:
2987 Impacts on Urban Water, Heat, and Pollution Cycles at the Tree, Street, and City Scale. *Journal of
2988 Environmental Quality*, 45(1), 119–124. DOI: 10.2134/jeq2015.11.0567.
- 2989 Lowry, H., Lill, A., and Wong, B. B. (2013). Behavioural responses of wildlife to urban environments.
2990 *Biological Reviews*, 88(3), 537–549. DOI: 10.1111/brv.12012.
- 2991 Luck, G. W., Lavorel, S., McIntyre, S., and Lumb, K. (2012). Improving the application of vertebrate trait-
2992 based frameworks to the study of ecosystem services. *Journal of Animal Ecology*, 81(5), 1065–1076.
2993 DOI: 10.1111/j.1365–2656.2012.01974.x.
- 2994 Mace, G. M., Possingham, H. P., and Leader-Williams, N (2006). Prioritizing choices in conservation. *Key
2995 Topics in Conservation Biology*, 17–34.
- 2996 MacLean, S. A. and Beissinger, S. R. (2017). Species' traits as predictors of range shifts under contemporary
2997 climate change: A review and meta-analysis. *Global Change Biology*, 23(10), 4094–4105. DOI: 10.
2998 1111/gcb.13736.
- 2999 Magioli, M., Ferraz, K. M. P. M. d. B., Chiarello, A. G., Galetti, M., Setz, E. Z. F., Paglia, A. P., Abrego,
3000 N., Ribeiro, M. C., and Ovaskainen, O. (2021). Land-use changes lead to functional loss of terrestrial

- 3001 mammals in a Neotropical rainforest. *Perspectives in Ecology and Conservation*, 19(2), 161–170. DOI:
3002 10.1016/j.pecon.2021.02.006.
- 3003 Maibach, E., Myers, T., and Leiserowitz, A. (2014). Climate scientists need to set the record straight: There
3004 is a scientific consensus that human-caused climate change is happening. *Earth's Future*, 2(5), 295–298.
3005 DOI: 10.1002/2013ef000226.
- 3006 Májeková, M., Paal, T., Plowman, N. S., Bryndová, M., Kasari, L., Norberg, A., Weiss, M., Bishop, T. R.,
3007 Luke, S. H., Sam, K., Le Bagousse-Pinguet, Y., Lepš, J., Götzenberger, L., and De Bello, F. (2016).
3008 Evaluating Functional diversity: Missing trait data and the importance of species abundance structure
3009 and data transformation. *PLoS ONE*, 11(2), e0149270. DOI: 10.1371/journal.pone.0149270.
- 3010 Malhi, Y. (2017). The Concept of the Anthropocene. *Annual Review of Environment and Resources*, 42, 77–
3011 104. DOI: 10.1146/annurev-environ-102016-060854.
- 3012 Marcacci, G., Westphal, C., Wenzel, A., Raj, V., Nölke, N., Tscharntke, T., and Grass, I. (2021). Taxonomic
3013 and functional homogenization of farmland birds along an urbanization gradient in a tropical megacity.
3014 *Global Change Biology*, 27(20), 4980–4994. DOI: 10.1111/gcb.15755.
- 3015 Martin, L. J., Blossey, B., and Ellis, E. (2012). Mapping where ecologists work: biases in the global distri-
3016 bution of terrestrial ecological observations. *Frontiers in Ecology and the Environment*, 10(4), 195–201.
3017 DOI: 10.1890/110154.
- 3018 Martins, I. S. and Pereira, H. M. (2017). Improving extinction projections across scales and habitats using the
3019 countryside species-area relationship. *Scientific Reports*, 7(12899). DOI: 10.1038/s41598-017-
3020 13059-y.
- 3021 Matich, P. and Schalk, C. M. (2019). Move it or lose it: Interspecific variation in risk response of pond-
3022 breeding anurans. *PeerJ*, 7, 1–16. DOI: 10.7717/PEERJ.6956.
- 3023 Matuoka, M. A., Benchimol, M., Almeida-Rocha, J. M. de, and Morante-Filho, J. C. (2020). Effects of
3024 anthropogenic disturbances on bird functional diversity: A global meta-analysis. *Ecological Indicators*,
3025 116, 106471. DOI: 10.1016/j.ecolind.2020.106471.
- 3026 Maxwell, S. L., Fuller, R. A., Brooks, T. M., and Watson, J. E. M. (2016). Biodiversity: The ravages of guns,
3027 nets and bulldozers. *Nature*, 536, 143–145. DOI: 10.1038/536143a.
- 3028 May, R. M. (2011). Why worry about how many species and their loss? *PLoS Biology*, 9(8), e1001130. DOI:
3029 10.1371/journal.pbio.1001130.
- 3030 McCain, C. M. and King, S. R. B. (2014). Body size and activity times mediate mammalian responses to
3031 climate change. *Global Change Biology*, 20(6), 1760–1769. DOI: 10.1111/gcb.12499.
- 3032 McGill, B. J., Enquist, B. J., Weiher, E., and Westoby, M. (2006). Rebuilding community ecology from
3033 functional traits. *Trends in Ecology and Evolution*, 21(4), 178–185. DOI: 10.1016/j.tree.2006.
3034 02.002.
- 3035 McGill, B. J., Dornelas, M., Gotelli, N. J., and Magurran, A. E. (2015). Fifteen forms of biodiversity trend in
3036 the anthropocene. *Trends in Ecology and Evolution*, 30(2), 104–113. DOI: 10.1016/j.tree.2014.
3037 11.006.
- 3038 McNab, B. K. (1986). The Influence of Food Habits on the Energetics of Eutherian Mammals. *Ecological
3039 Monographs*, 56(1), 1–19. DOI: 10.2307/2937268.

- 3040 Meiri, S., Feldman, A., and Kratochvíl, L. (2015). Squamate hatchling size and the evolutionary causes of
3041 negative offspring size allometry. *Journal of Evolutionary Biology*, 28(2), 438–446. DOI: 10.1111/jeb.12580.
- 3042
- 3043 Meiri, S. (2018). Traits of lizards of the world: Variation around a successful evolutionary design. *Global
3044 Ecology and Biogeography*, 27(10), 1168–1172. DOI: 10.1111/geb.12773.
- 3045 Meiri, S. and Chapple, D. G. (2016). Biases in the current knowledge of threat status in lizards, and bridging
3046 the ‘assessment gap’. *Biological Conservation*, 204, 6–15. DOI: 10.1016/j.biocon.2016.03.
3047 009.
- 3048 Mendoza, M. and Araújo, M. B. (2019). Climate shapes mammal community trophic structures and humans
3049 simplify them. *Nature Communications*, 10(5197). DOI: 10.1038/s41467-019-12995-9.
- 3050 Menegotto, A. and Rangel, T. F. (2018). Mapping knowledge gaps in marine diversity reveals a latitudinal
3051 gradient of missing species richness. *Nature Communications*, 9(4713). DOI: 10.1038/s41467-
3052 018-07217-7.
- 3053 Merckx, T. et al. (2018). Body-size shifts in aquatic and terrestrial urban communities. *Nature*, 558, 113–
3054 116. DOI: 10.1038/s41586-018-0140-0.
- 3055 Meyer, C., Kreft, H., Guralnick, R., and Jetz, W. (2015). Global priorities for an effective information basis
3056 of biodiversity distributions. *Nature Communications*, 6(8221). DOI: 10.1038/ncomms9221.
- 3057 Migné, A., Golléty, C., and Davoult, D. (2015). Effect of canopy removal on a rocky shore community
3058 metabolism and structure. *Marine Biology*, 162(2), 449–457. DOI: 10.1007/s00227-014-2592-
3059 6.
- 3060 Millard, J., Outhwaite, C. L., Kinnersley, R., Freeman, R., Gregory, R. D., Adedoja, O., Gavini, S., Kioko,
3061 E., Kuhlmann, M., Ollerton, J., Ren, Z.-X., and Newbold, T. (2021). Global effects of land-use intensity
3062 on local pollinator biodiversity. *Nature Communications*, 12(2902). DOI: 10.1038/s41467-021-
3063 23228-3.
- 3064 Millennium Ecosystem Assessment (2005). *Millennium Ecosystem Assessment: Ecosystems and Human
3065 Well-being: Synthesis*. Tech. rep. Available at: <https://www.millenniumassessment.org/documents/document.356.aspx.pdf>.
- 3066
- 3067 Miller, G., Magee, J., Smith, M., Spooner, N., Baynes, A., Lehman, S., Fogel, M., Johnston, H., Williams, D.,
3068 Clark, P., Florian, C., Holst, R., and DeVogel, S. (2016). Human predation contributed to the extinction
3069 of the Australian megafaunal bird *Genyornis newtoni* ~ 47 ka. *Nature Communications*, 7(10496), 1–7.
3070 DOI: 10.1038/ncomms10496.
- 3071 Moe, B., Brunvoll, S., Mork, D., Brobak, T. E., and Bech, C. (2004). Developmental plasticity of physiology
3072 and morphology in diet-restricted European shag nestlings (*Phalacrocorax aristotelis*). *Journal of
3073 Experimental Biology*, 207(23), 4067–4076. DOI: 10.1242/jeb.01226.
- 3074 Möller, A. P. (2009). Successful city dwellers: A comparative study of the ecological characteristics of urban
3075 birds in the Western Palearctic. *Oecologia*, 159(4), 849–858. DOI: 10.1007/s00442-008-1259-8.
- 3076 Monastersky, R. (2014). Life - a status report. *Nature*, 516(7530), 161. DOI: 10.1038/516158a.
- 3077 — (2015). The human age. *Nature*, 519, 144–147. DOI: 10.1038/519144a.
- 3078 Mooney, K. A., Gruner, D. S., Barber, N. A., Bael, S. A. V., Philpott, S. M., and Greenberg, R. (2010). Inter-
3079 actions among predators and the cascading effects of vertebrate insectivores on arthropod communities

- 3080 and plants. *Proceedings of the National Academy of Sciences*, 107(16), 7335–7340. DOI: 10.1073/pnas.1001934107.
- 3082 Morelli, F., Benedetti, Y., Hanson, J. O., and Fuller, R. A. (2021). Global distribution and conservation of
3083 avian diet specialization. *Conservation Letters*, 14(4), e12795. DOI: 10.1111/conl.12795.
- 3084 Mouillot, D., Bellwood, D. R., Baraloto, C., Chave, J., Galzin, R., Harmelin-Vivien, M., Kulbicki, M.,
3085 Lavergne, S., Lavorel, S., Mouquet, N., Paine, C. E. T., Renaud, J., and Thuiller, W. (May 2013). Rare
3086 Species Support Vulnerable Functions in High-Diversity Ecosystems. *PLOS Biology*, 11(5), e1001569.
3087 DOI: 10.1371/journal.pbio.1001569.
- 3088 Mouillot, D., Loiseau, N., Grenié, M., Algar, A. C., Allegra, M., Cadotte, M. W., Casajus, N., Denelle, P.,
3089 Guégan, M., Maire, A., Maitner, B., McGill, B. J., McLean, M., Mouquet, N., Munoz, F., Thuiller, W.,
3090 Villéger, S., Violle, C., and Auber, A. (2021). The dimensionality and structure of species trait spaces.
3091 *Ecology Letters*, 24, 1988–2009. DOI: 10.1111/ele.13778.
- 3092 Mueller, P. and Diamond, J. (2001). Metabolic rate and environmental productivity: Well-provisioned ani-
3093 mals evolved to run and idle fast. *Proceedings of the National Academy of Sciences of the United States
3094 of America*, 98(22), 12550–12554. DOI: 10.1073/pnas.221456698.
- 3095 Munstermann, M. J., Heim, N. A., McCauley, D. J., Payne, J. L., Upham, N. S., Wang, S. C., and Knope,
3096 M. L. (2022). A global ecological signal of extinction risk in terrestrial vertebrates. *Conservation Biol-
3097 ogy*, 36(3), e13852. DOI: 10.1111/cobi.13852.
- 3098 Munteanu, C., Kamp, J., Nita, M. D., Klein, N., Kraemer, B. M., Müller, D., Koskina, A., Prishchepov,
3099 A. V., and Kuemmerle, T. (2020). Cold War spy satellite images reveal long-term declines of a philopatric
3100 keystone species in response to cropland expansion. *Proceedings of the Royal Society B: Biological
3101 Sciences*, 287(20192897). DOI: 10.1098/rspb.2019.2897.
- 3102 Myhrvold, N. P., Baldridge, E., Chan, B., Sivam, D., Freeman, D. L., and Ernest, S. K. M. (2015). An am-
3103 niote life-history database to perform comparative analyses with birds, mammals, and reptiles. *Ecology*,
3104 96(11), 3109–3109. DOI: 10.1890/15-0846R.1.
- 3105 Nakagawa, S. and Freckleton, R. P. (2008). Missing inaction: the dangers of ignoring missing data. *Trends
3106 in Ecology and Evolution*, 23(11), 592–596. DOI: 10.1016/j.tree.2008.06.014.
- 3107 Naya, D. E., Spangenberg, L., Naya, H., and Bozinovic, F. (2013). How does evolutionary variation in basal
3108 metabolic rates arise? A statistical assessment and a mechanistic model. *Evolution*, 67(5), 1463–1476.
3109 DOI: 10.1111/evo.12042.
- 3110 Newbold, T. (2018). Future effects of climate and land-use change on terrestrial vertebrate community diver-
3111 sity under different scenarios. *Proceedings of the Royal Society of London Series B, Biological Sciences*,
3112 285(1881), 20180792. DOI: 10.1098/rspb.2018.0792.
- 3113 Newbold, T., Scharlemann, J. P. W., Butchart, S. H. M., Sekercioğlu, C. H., Alkemade, R., Booth, H., and
3114 Purves, D. W. (2013). Ecological traits affect the response of tropical forest bird species to land-use in-
3115 tensity. *Proceedings of the Royal Society of London Series B, Biological Sciences*, 280(1750), 20122131.
3116 DOI: 10.1098/rspb.2012.2131.
- 3117 Newbold, T. et al. (2015). Global effects of land use on local terrestrial biodiversity. *Nature*, 520, 45–50.
3118 DOI: 10.1038/nature14324.
- 3119 Newbold, T., Hudson, L. N., Contu, S., Hill, S. L., Beck, J., Liu, Y., Meyer, C., Phillips, H. R., Scharlemann,
3120 J. P., and Purvis, A. (2018). Widespread winners and narrow-ranged losers: Land use homogenizes biodi-

- 3121 diversity in local assemblages worldwide. *PLoS Biology*, 16(12), e2006841. DOI: 10.1371/journal.
3122 pbio.2006841.
- 3123 Newbold, T., Adams, G. L., Robles, G. A., Boakes, E. H., Ferreira, G. B., Chapman, A. S., Etard, A., Gibb,
3124 R., Millard, J., Outhwaite, C. L., and Williams, J. J. (2019). Climate and land-use change homogenise
3125 terrestrial biodiversity, with consequences for ecosystem functioning and human well-being. *Emerging
3126 Topics in Life Sciences*, 3(2), 207–219. DOI: 10.1042/ETLS20180135.
- 3127 Newbold, T., Bentley, L. F., Hill, S. L., Edgar, M. J., Horton, M., Su, G., Şekercioğlu, Ç. H., Collen, B., and
3128 Purvis, A. (2020a). Global effects of land use on biodiversity differ among functional groups. *Functional
3129 Ecology*, 34(3), 684–693. DOI: 10.1111/1365-2435.13500.
- 3130 Newbold, T., Oppenheimer, P., Etard, A., and Williams, J. J. (2020b). Tropical and Mediterranean biodi-
3131 versity is disproportionately sensitive to land-use and climate change. *Nature Ecology and Evolution*, 4,
3132 1630–1638. DOI: 10.1038/s41559-020-01303-0.
- 3133 Novosolov, M., Raia, P., and Meiri, S. (2013). The island syndrome in lizards. *Global Ecology and Biogeog-
3134 raphy*, 22(2), 184–191. DOI: 10.1111/j.1466-8238.2012.00791.x.
- 3135 Novosolov, M., Rodda, G. H., North, A. C., Butchart, S. H., Tallowin, O. J., Gainsbury, A. M., and Meiri, S.
3136 (2017). Population density–range size relationship revisited. *Global Ecology and Biogeography*, 26(10),
3137 1088–1097. DOI: 10.1111/geb.12617.
- 3138 Nowakowski, A. J., Thompson, M. E., Donnelly, M. A., and Todd, B. D. (2017). Amphibian sensitivity to
3139 habitat modification is associated with population trends and species traits. *Global Ecology and Biogeog-
3140 raphy*, 26(6), 700–712. DOI: 10.1111/geb.12571.
- 3141 Nowakowski, A. J., Frishkoff, L. O., Thompson, M. E., Smith, T. M., and Todd, B. D. (2018). Phylogenetic
3142 homogenization of amphibian assemblages in human-altered habitats across the globe. *Proceedings of
3143 the National Academy of Sciences of the United States of America*, 115(15), E3454–E3462. DOI: 10.
3144 1073/pnas.1714891115.
- 3145 Oliveira, B. F., São-Pedro, V. A., Santos-Barrera, G., Penone, C., and Costa, G. C. (2017). AmphiBIO, a
3146 global database for amphibian ecological traits. *Scientific Data*, 4, 170123. DOI: 10.1038/sdata.
3147 2017.123.
- 3148 Oliver, R. Y., Meyer, C., Ranipeta, A., Winner, K., and Jetz, W. (2021). Global and national trends, gaps,
3149 and opportunities in documenting and monitoring species distributions. *PLoS Biology*, 19(8), e3001336.
3150 DOI: 10.1371/journal.pbio.3001336.
- 3151 Oliver, T. H., Heard, M. S., Isaac, N. J., Roy, D. B., Procter, D., Eigenbrod, F., Freckleton, R., Hector, A.,
3152 Orme, C. D. L., Petchey, O. L., Proença, V., Raffaelli, D., Suttle, K. B., Mace, G. M., Martín-López,
3153 B., Woodcock, B. A., and Bullock, J. M. (2015). Biodiversity and Resilience of Ecosystem Functions.
3154 *Trends in Ecology and Evolution*, 30, 673–684. DOI: 10.1016/j.tree.2015.08.009.
- 3155 Orme, D., Freckleton, R., Thomas, G., Petzoldt, T., Fritz, S., Isaac, N., and Pearse, W. (2018). *caper: Com-
3156 parative Analyses of Phylogenetics and Evolution in R*. R package version 1.0.1.
- 3157 Pacifici, M. et al. (2015). Assessing species vulnerability to climate change. *Nature Climate Change*, 5, 215–
3158 224. DOI: 10.1038/nclimate2448.
- 3159 Pacifici, M., Visconti, P., Butchart, S. H. M., Watson, J. E. M., Cassola, F. M., and Rondinini, C. (2017).
3160 Species' traits influenced their response to recent climate change. *Nature Climate Change*, 7, 205–208.
3161 DOI: 10.1038/nclimate3223.

- 3162 Pagel, M. (1999). Inferring the historical patterns of biological evolution. *Nature*, 401, 877–884. DOI: 10 .
3163 1038/44766.
- 3164 Pakeman, R. J. (2014). Functional trait metrics are sensitive to the completeness of the species' trait data? *Methods in Ecology and Evolution*, 5(1), 9–15. DOI: 10 .1111/2041-210X.12136.
- 3166 Paradis, E., Baillie, S. R., Sutherland, W. J., and Gregory, R. D. (1998). Patterns of natal and breeding
3167 dispersal in birds. *Journal of Animal Ecology*, 67(4), 518–536. DOI: 10 .1046/j.1365-2656 .
3168 1998.00215.x.
- 3169 Pearson, R. G., Stanton, J. C., Shoemaker, K. T., Aiello-Lammens, M. E., Ersts, P. J., Horning, N., Fordham,
3170 D. A., Raxworthy, C. J., Ryu, H. Y., Mcnees, J., and Akçakaya, H. R. (2014). Life history and spatial
3171 traits predict extinction risk due to climate change. *Nature Climate Change*, 4, 217–221. DOI: 10 .
3172 1038/nclimate2113.
- 3173 Penone, C., Davidson, A. D., Shoemaker, K. T., Di Marco, M., Rondinini, C., Brooks, T. M., Young, B. E.,
3174 Graham, C. H., and Costa, G. C. (2014). Imputation of missing data in life-history trait datasets: Which
3175 approach performs the best? *Methods in Ecology and Evolution*, 5(9), 961–970. DOI: 10 .1111/2041-
3176 210X.12232.
- 3177 Pereira, H. M. et al. (2010). Scenarios for global biodiversity in the 21st century. *Science*, 330(6010), 1496–
3178 1501. DOI: 10 .1126/science.1196624.
- 3179 Perronne, R., Munoz, F., Borgy, B., Reboud, X., and Gaba, S. (2017). How to design trait-based analyses
3180 of community assembly mechanisms: Insights and guidelines from a literature review. *Perspectives in
3181 Plant Ecology, Evolution and Systematics*, 25, 29–44. DOI: 10 .1016/j.ppees.2017.01.004.
- 3182 Peugh, J. L. and Enders, C. K. (2004). Missing data in educational research: A review of reporting practices
3183 and suggestions for improvement. *Review of Educational Research*, 74(4), 525–556. DOI: 10 .3102 /
3184 00346543074004525.
- 3185 Pörtner, H. O. and Farrell, A. P. (2008). Physiology and Climate Change. *Science*, 322(5902), 690–692. DOI:
3186 10 .1126/science.1163156.
- 3187 Powell, T. W. R. and Lenton, T. M. (2013). Scenarios for future biodiversity loss due to multiple drivers
3188 reveal conflict between mitigating climate change and preserving biodiversity. *Environmental Research
3189 Letters*, 8(2), 025024. DOI: 10 .1088/1748-9326/8/2/025024.
- 3190 Powers, R. P. and Jetz, W. (2019). Global habitat loss and extinction risk of terrestrial vertebrates under
3191 future land-use-change scenarios. *Nature Climate Change*, 9(4), 323–329. DOI: 10 .1038/s41558-
3192 019-0406-z.
- 3193 Purvis, A., Gittleman, J. L., Cowlishaw, G., and Mace, G. M. (2000). Predicting extinction risk in declining
3194 species. *Proceedings of the Royal Society B: Biological Sciences*, 267(1456), 1947–1952. DOI: 10 .
3195 1098/rspb.2000.1234.
- 3196 Pyšek, P., Bacher, S., Chytrý, M., Jarošík, V., Wild, J., Celesti-Grapow, L., Gassó, N., Kenis, M., Lambdon,
3197 P. W., Nentwig, W., Pergl, J., Roques, A., Sádlo, J., Solarz, W., Vilà, M., and Hulme, P. E. (2010).
3198 Contrasting patterns in the invasions of European terrestrial and freshwater habitats by alien plants,
3199 insects and vertebrates. *Global Ecology and Biogeography*, 19(3), 317–331. DOI: 10 .1111/j.1466-
3200 8238.2009.00514.x.

- 3201 Quesnelle, P. E., Lindsay, K. E., and Fahrig, L. (2014). Low reproductive rate predicts species sensitivity
3202 to habitat loss: A meta-analysis of wetland vertebrates. *PLoS ONE*, 9(3), e90926. DOI: 10.1371/journal.pone.0090926.
- 3204 R Core Team (2018). *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical
3205 Computing. Vienna, Austria.
- 3206 Ramesh, T., Kalle, R., and Downs, C. T. (2017). Space use in a South African agriculture landscape by the
3207 caracal (*Caracal caracal*). *European Journal of Wildlife Research*, 63(1). DOI: 10.1007/s10344-
3208 016-1072-3.
- 3209 Ramírez-Bautista, A., Thorne, J. H., Schwartz, M. W., and Williams, J. N. (2020). Trait-based climate vul-
3210 nerability of native rodents in southwestern Mexico. *Ecology and Evolution*, 10(12), 5864–5876. DOI:
3211 10.1002/ece3.6323.
- 3212 Rapacciulo, G., Marin, J., Costa, G. C., Helmus, M. R., Behm, J. E., Brooks, T. M., Hedges, S. B., Rade-
3213 loff, V. C., Young, B. E., and Graham, C. H. (2017). The signature of human pressure history on the
3214 biogeography of body mass in tetrapods. *Global Ecology and Biogeography*, 26(9), 1022–1034. DOI:
3215 <https://doi.org/10.1111/geb.12612>.
- 3216 Ratto, F., Simmons, B. I., Spake, R., Zamora-Gutierrez, V., MacDonald, M. A., Merriman, J. C., Tremlett,
3217 C. J., Poppy, G. M., Peh, K. S., and Dicks, L. V. (2018). Global importance of vertebrate pollinators for
3218 plant reproductive success: a meta-analysis. *Frontiers in Ecology and the Environment*, 16(2), 82–90.
3219 DOI: 10.1002/fee.1763.
- 3220 Revell, L. J. (2012). phytools: An R package for phylogenetic comparative biology (and other things). *Meth-
3221 ods in Ecology and Evolution*, 3(2), 217–223. DOI: 10.1111/j.2041-210X.2011.00169.x.
- 3222 Riemann, J. C., Ndriantsoa, S. H., Rödel, M. O., and Glos, J. (2017). Functional diversity in a fragmented
3223 landscape — Habitat alterations affect functional trait composition of frog assemblages in Madagascar.
3224 *Global Ecology and Conservation*, 10, 173–183. DOI: 10.1016/j.gecco.2017.03.005.
- 3225 Ries, L., Fletcher, R. J., Battin, J., and Sisk, T. D. (2004). Ecological responses to habitat edges: Mechanisms,
3226 models, and variability explained. *Annual Review of Ecology, Evolution, and Systematics*, 35, 491–522.
3227 DOI: 10.1146/annurev.ecolsys.35.112202.130148.
- 3228 Rinnan, D. S. (2021). *CENFA: Climate and Ecological Niche Factor Analysis*. R package version 1.1.1.
- 3229 Rinnan, D. S. and Lawler, J. (2019). Climate-niche factor analysis: a spatial approach to quantifying species
3230 vulnerability to climate change. *Ecography*, 42(9), 1494–1503. DOI: <https://doi.org/10.1111/ecog.03937>.
- 3232 Ripple, W. J., Estes, J. A., Beschta, R. L., Wilmers, C. C., Ritchie, E. G., Hebblewhite, M., Berger, J.,
3233 Elmhausen, B., Letnic, M., Nelson, M. P., Schmitz, O. J., Smith, D. W., Wallach, A. D., and Wirsing,
3234 A. J. (2014). Status and ecological effects of the world's largest carnivores. *Science*, 343(6167). DOI:
3235 10.1126/science.1241484.
- 3236 Ripple, W. J., Wolf, C., Newsome, T. M., Hoffmann, M., Wirsing, A. J., and McCauley, D. J. (2017). Ex-
3237 tinction risk is most acute for the world's largest and smallest vertebrates. *Proceedings of the National
3238 Academy of Sciences*, 114(40), 10678–10683. DOI: 10.1073/pnas.1702078114.
- 3239 Rodrigues, A. S., Pilgrim, J. D., Lamoreux, J. F., Hoffmann, M., and Brooks, T. M. (2006). The value of the
3240 IUCN Red List for conservation. *Trends in Ecology and Evolution*, 21(2), 71–76. DOI: 10.1016/j.tree.2005.10.010.

- 3242 Rohr, J. R., Civitello, D. J., Cohen, J. M., Roznik, E. A., Sinervo, B., and Dell, A. I. (2018). The complex
3243 drivers of thermal acclimation and breadth in ectotherms. *Ecology Letters*, 21(9), 1425–1439. DOI: 10 .
3244 1111/ele.13107.
- 3245 Roll, U. et al. (2017). The global distribution of tetrapods reveals a need for targeted reptile conservation.
3246 *Nature Ecology and Evolution*, 1, 1677–1682. DOI: 10 .1038/s41559-017-0332-2.
- 3247 Running, S. W. and Zhao, M. (2015). Daily GPP and Annual NPP (MOD17A2/A3) products NASA Earth
3248 Observing System MODIS Land Algorithm (User's guide V3). Available at: https://lpdaac.usgs.gov/documents/495/MOD17_User_Guide_V6.pdf.
- 3250 Sadowska, E. T., Stawski, C., Rudolf, A., Dheyongera, G., Chrząćik, K. M., Baliga-Klimczyk, K., and
3251 Koteja, P. (2015). Evolution of basal metabolic rate in bank voles from a multidirectional selection ex-
3252 periment. *Proceedings of the Royal Society B: Biological Sciences*, 282(1806), 20150025. DOI: 10 .
3253 1098/rspb.2015.0025.
- 3254 Sales, L. P., Galetti, M., Carnaval, A., Monsarrat, S., Svenning, J., and Pires, M. M. (2022). The effect of
3255 past defaunation on ranges, niches, and future biodiversity forecasts. *Global Change Biology*, 28(11),
3256 3683–3693. DOI: 10 .1111/gcb.16145.
- 3257 Salguero-Gómez, R., Jackson, J., and Gascoigne, S. J. L. (2021). Four key challenges in the open-data revo-
3258 lution. *Journal of Animal Ecology*, 90(9), 2000–2004. DOI: 10 .1111/1365-2656.13567.
- 3259 Salo, P., Banks, P. B., Dickman, C. R., and Korpimäki, E. (2010). Predator manipulation experiments: Im-
3260 pacts on populations of terrestrial vertebrate prey. *Ecological Monographs*, 80(4), 531–546. DOI: 10 .
3261 1890/09-1260.1.
- 3262 Samia, D. S., Nakagawa, S., Nomura, F., Rangel, T. F., and Blumstein, D. T. (2015). Increased tolerance to
3263 humans among disturbed wildlife. *Nature Communications*, 6(8877). DOI: 10 .1038/ncomms9877.
- 3264 Sánchez-Ortiz, K., Taylor, K. J., Palma, A. de, König, C., Pyšek, P., Weigelt, P., Essl, F., and Purvis, A.
3265 (2019). Effects of land-use change and related pressures on alien and native subsets of island communi-
3266 ties. *PLoS ONE*, 15(12), e0227169. DOI: 10 .1371/journal.pone.0227169.
- 3267 Sandom, C., Faurby, S., Sandel, B., and Svenning, J. C. (2014). Global late Quaternary megafauna extinc-
3268 tions linked to humans, not climate change. *Proceedings of the Royal Society B: Biological Sciences*,
3269 281(1787), 20133254. DOI: 10 .1098/rspb.2013 .3254.
- 3270 Santini, L., Benítez-López, A., Ficetola, G. F., and Huijbregts, M. A. J. (2018). Length–mass allometries in
3271 amphibians. *Integrative Zoology*, 13(1), 36–45. DOI: 10 .1111/1749-4877.12268.
- 3272 Santos, T. (2018). Package ‘PVR’. *Phylogenetic Eigenvectors Regression and Phylogenetic Signal-Representation*
3273 *Curve*. R package version 0.3.0.
- 3274 Sayol, F., Sol, D., and Pigot, A. L. (2020). Brain Size and Life History Interact to Predict Urban Tolerance
3275 in Birds. *Frontiers in Ecology and Evolution*, 8, 58. DOI: 10 .3389/fevo .2020 .00058.
- 3276 Scharf, I., Feldman, A., Novosolov, M., Pincheira-Donoso, D., Das, I., Böhm, M., Uetz, P., Torres-Carvajal,
3277 O., Bauer, A., Roll, U., and Meiri, S. (2015). Late bloomers and baby boomers: Ecological drivers of
3278 longevity in squamates and the tuatara. *Global Ecology and Biogeography*, 24(4), 396–405. DOI: 10 .
3279 1111/geb.12244.
- 3280 Schielzeth, H., Dingemanse, N. J., Nakagawa, S., Westneat, D. F., Allegue, H., Teplitsky, C., Réale, D.,
3281 Dochtermann, N. A., Garamszegi, L. Z., and Araya-Ajoy, Y. G. (2020). Robustness of linear mixed-

- 3282 effects models to violations of distributional assumptions. *Methods in Ecology and Evolution*, 11(9),
3283 1141–1152. DOI: 10.1111/2041-210X.13434.
- 3284 Schipper, A. M., Hilbers, J. P., Meijer, J. R., Antão, L. H., Benítez-López, A., Jonge, M. M. de, Leemans,
3285 L. H., Scheper, E., Alkemade, R., Doelman, J. C., Mylius, S., Stehfest, E., Vuuren, D. P. van, Zeist, W. J.
3286 van, and Huijbregts, M. A. (2020). Projecting terrestrial biodiversity intactness with GLOBIO 4. *Global
3287 Change Biology*, 26(2), 760–771. DOI: 10.1111/gcb.14848.
- 3288 Schleuter, D., Daufresne, M., Massol, F., and Argillier, C. (2010). A user’s guide to functional diversity
3289 indices. *Ecological Monographs*, 80(3), 469–484. DOI: 10.1890/08-2225.1.
- 3290 Schloss, C. A., Nunez, T. A., and Lawler, J. J. (2012). Dispersal will limit ability of mammals to track climate
3291 change in the Western Hemisphere. *Proceedings of the National Academy of Sciences*, 109(22), 8606–
3292 8611. DOI: 10.1073/pnas.1116791109.
- 3293 Schneider, F. D., Fichtmueller, D., Gossner, M. M., Güntsch, A., Jochum, M., König-Ries, B., Le Provost,
3294 G., Manning, P., Ostrowski, A., Penone, C., and Simons, N. K. (2019). Towards an ecological trait-data
3295 standard. *Methods in Ecology and Evolution*, 10(12), 2006–2019. DOI: 10.1111/2041-210X.13288.
- 3296 Schwarz, R. and Meiri, S. (2017). The fast-slow life-history continuum in insular lizards: a comparison
3297 between species with invariant and variable clutch sizes. *Journal of Biogeography*, 44(12), 2808–2815.
3298 DOI: 10.1111/jbi.13067.
- 3300 Segan, D. B., Murray, K. A., and Watson, J. E. (2016). A global assessment of current and future biodiversity
3301 vulnerability to habitat loss-climate change interactions. *Global Ecology and Conservation*, 5, 12–21.
3302 DOI: 10.1016/j.gecco.2015.11.002.
- 3303 Seneviratne, S. I. et al. (2012). Changes in climate extremes and their impacts on the natural physical envi-
3304 ronment. *Managing the Risks of Extreme Events and Disasters to Advance Climate Change Adaptation:
3305 Special Report of the Intergovernmental Panel on Climate Change*. Ed. by C. B. Field, V. Barros, T. F.
3306 Stocker, and Q. Dahe. Vol. 9781107025, 109–230. DOI: 10.1017/CBO9781139177245.006.
- 3307 Seto, K. C., Sánchez-Rodríguez, R., and Fragkias, M. (2010). The new geography of contemporary ur-
3308 banization and the environment. *Annual Review of Environment and Resources*, 35(1), 167–194. DOI:
3309 10.1146/annurev-environ-100809-125336.
- 3310 Sévèque, A., Gentle, L. K., López-Bao, J. V., Yarnell, R. W., and Uzal, A. (2020). Human disturbance has
3311 contrasting effects on niche partitioning within carnivore communities. *Biological Reviews*, 95(6), 1689–
3312 1705. DOI: 10.1111/brv.12635.
- 3313 Severtsov (2012). The Significance of Vertebrates in the Structure and Functioning of Ecosystems. *Biology
3314 Bulletin*, 40(7), 571–579. DOI: 10.1134/S1062359013070054.
- 3315 Siefert, A. et al. (2015). A global meta-analysis of the relative extent of intraspecific trait variation in plant
3316 communities. *Ecology Letters*, 18(12), 1406–1419. DOI: 10.1111/ele.12508.
- 3317 Slavenko, A., Tallowin, O. J., Itescu, Y., Raia, P., and Meiri, S. (2016). Late Quaternary reptile extinctions:
3318 size matters, insularity dominates. *Global Ecology and Biogeography*, 25(11), 1308–1320. DOI: 10.
3319 1111/geb.12491.
- 3320 Smith, A. M. and Green, D. M. (2005). Dispersal and the metapopulation paradigm in amphibian ecology
3321 and conservation: are all amphibian populations metapopulations? *Ecography*, 28(1), 110–128.

- 3322 Sodhi, N. S., Bickford, D., Diesmos, A. C., Lee, T. M., Koh, L. P., Brook, B. W., Sekercioglu, C. H., and
3323 Bradshaw, C. J. (2008). Measuring the meltdown: Drivers of global amphibian extinction and decline.
3324 *PLoS ONE*, 3(2), e1636. DOI: 10.1371/journal.pone.0001636.
- 3325 Sol, D., Trisos, C., Múrria, C., Jeliazkov, A., González-Lagos, C., Pigot, A. L., Ricotta, C., Swan, C. M.,
3326 Tobias, J. A., and Pavoine, S. (2020). The worldwide impact of urbanisation on avian functional diversity.
3327 *Ecology Letters*, 23(6), 962–972. DOI: 10.1111/ele.13495.
- 3328 Soroye, P., Newbold, T., and Kerr, J. (2020). Climate change contributes to widespread declines among
3329 bumble bees across continents. *Science*, 367(6478), 685–688. DOI: 10.1126/science.aax8591.
- 3330 Souza, F. L., Valente-Neto, F., Severo-Neto, F., Bueno, B., Ochoa-Quintero, J. M., Laps, R. R., Bolzan,
3331 F., and Roque, F. d. O. (2019). Impervious surface and heterogeneity are opposite drivers to maintain
3332 bird richness in a Cerrado city. *Landscape and Urban Planning*, 192, 103643. DOI: 10.1016/j.landurbplan.2019.103643.
- 3334 Spooner, F. E., Pearson, R. G., and Freeman, R. (2018). Rapid warming is associated with population decline
3335 among terrestrial birds and mammals globally. *Global Change Biology*, 24(10), 4521–4531. DOI: 10.1111/gcb.14361.
- 3337 Spracklen, B. D., Kalamandeen, M., Galbraith, D., Gloor, E., and Spracklen, D. V. (Dec. 2015). A Global
3338 Analysis of Deforestation in Moist Tropical Forest Protected Areas. *PLoS ONE*, 10(12), e0143886. DOI:
3339 10.1371/journal.pone.0143886.
- 3340 Stark, G., Tamar, K., Itescu, Y., Feldman, A., and Meiri, S. (2018). Cold and isolated ectotherms: drivers
3341 of reptilian longevity. *Biological Journal of the Linnean Society*, 125(4), 730–740. DOI: 10.1093/biolinnean/bly153.
- 3343 Stark, G., Pincheira-Donoso, D., and Meiri, S. (2020). No evidence for the ‘rate-of-living’ theory across the
3344 tetrapod tree of life. *Global Ecology and Biogeography*, 29(5), 857–884. DOI: 10.1111/geb.13069.
- 3345 Steffen, W., Grinevald, J., Crutzen, P., and McNeill, J. (2011). The anthropocene: Conceptual and historical
3346 perspectives. *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering
3347 Sciences*, 369(1938), 842–867. DOI: 10.1098/rsta.2010.0327.
- 3348 Steffen, W., Broadgate, W., Deutsch, L., Gaffney, O., and Ludwig, C. (2015). *The trajectory of the Anthro-
3349 pocene: The Great Acceleration*. DOI: 10.1177/2053019614564785.
- 3350 Stehfest, E., Zeist, W. J. van, Valin, H., Havlik, P., Popp, A., Kyle, P., Tabeau, A., Mason-D'Croz, D.,
3351 Hasegawa, T., Bodirsky, B. L., Calvin, K., Doelman, J. C., Fujimori, S., Humpenöder, F., Lotze-Campen,
3352 H., Meijl, H. van, and Wiebe, K. (2019). Key determinants of global land-use projections. *Nature Communications*, 10(2166). DOI: 10.1038/s41467-019-09945-w.
- 3354 Stekhoven, D. J. (2016). *Nonparametric Missing Value Imputation using Random Forest*. R package version
3355 1.5. DOI: 10.1093/bioinformatics/btr597.
- 3356 Stekhoven, D. J. and Bühlmann, P. (2012). Missforest-Non-parametric missing value imputation for mixed-
3357 type data. *Bioinformatics*, 28(1), 112–118. DOI: 10.1093/bioinformatics/btr597.
- 3358 Stephens, L. et al. (2019). Archaeological assessment reveals Earth’s early transformation through land use.
3359 *Science*, 365(6456), 897–902. DOI: 10.1126/science.aax1192.
- 3360 Stewart, P. S., Voskamp, A., Santini, L., Biber, M. F., Devenish, A. J., Hof, C., Willis, S. G., and Tobias, J. A.
3361 (2022). Global impacts of climate change on avian functional diversity. *Ecology Letters*, 25(3), 673–685.
3362 DOI: 10.1111/ele.13830.

- 3363 Sunday, J. M., Bates, A. E., Kearney, M. R., Colwell, R. K., Dulvy, N. K., Longino, J. T., and Huey, R. B.
3364 (2014). Thermal-safety margins and the necessity of thermoregulatory behavior across latitude and el-
3365 evation. *Proceedings of the National Academy of Sciences of the United States of America*, 111(15),
3366 5610–5615. DOI: 10.1073/pnas.1316145111.
- 3367 Sutherland, G. D., Harestad, A. S., Price, K., and Lertzman, K. P. (2000). Scaling of natal dispersal distances
3368 in terrestrial birds and mammals. *Ecology and Society*, 4(1), 16. DOI: 10.5751/es-00184-040116.
- 3369 Svenning, J. C. and Skov, F. (2004). Limited filling of the potential range in European tree species. *Ecology
Letters*, 7(7), 565–573. DOI: 10.1111/j.1461-0248.2004.00614.x.
- 3370 Thomas, C. D. (2013). The Anthropocene could raise biological diversity. *Nature*, 502(7469), 7. DOI: 10.
3371 1038/502007a.
- 3372 Thuiller, W., Lavorel, S., and Araújo, M. B. (2005). Niche properties and geographical extent as predictors
3373 of species sensitivity to climate change. *Global Ecology and Biogeography*, 14(4), 347–357. DOI: 10.
3374 1111/j.1466-822X.2005.00162.x.
- 3375 Tiffney, B. H. (2004). Vertebrate dispersal of seed plants through time. *Annual Review of Ecology, Evolution,
and Systematics*, 35, 1–29. DOI: 10.1146/annurev.ecolsys.34.011802.132535.
- 3376 Tilman, D. and Downing, J. A. (1994). Biodiversity and stability in grasslands. *Nature*, 367, 363–365. DOI:
3377 10.1038/367363a0.
- 3378 Tinoco, B. A., Santillán, V. E., and Graham, C. H. (2018). Land use change has stronger effects on functional
3379 diversity than taxonomic diversity in tropical Andean hummingbirds. *Ecology and Evolution*, 8(6), 3478–
3380 3490. DOI: 10.1002/ece3.3813.
- 3381 Titley, M. A., Snaddon, J. L., and Turner, E. C. (2017). Scientific research on animal biodiversity is sys-
3382 tematically biased towards vertebrates and temperate regions. *PLoS ONE*, 12(12), e0189577. DOI: 10.
3383 1371/journal.pone.0189577.
- 3384 Todd, B. D., Nowakowski, A. J., Rose, J. P., and Price, S. J. (2017). Species traits explaining sensitivity
3385 of snakes to human land use estimated from citizen science data. *Biological Conservation*, 206, 31–36.
DOI: 10.1016/j.biocon.2016.12.013.
- 3386 Tonini, J. F. R., Beard, K. H., Ferreira, R. B., Jetz, W., and Pyron, R. A. (2016). Fully-sampled phylogenies
3387 of squamates reveal evolutionary patterns in threat status. *Biological Conservation*, 204, Part A, 23–31.
DOI: 10.1016/j.biocon.2016.03.039.
- 3388 Trenberth, K. E. (2011). Changes in precipitation with climate change. *Climate Research*, 47(1-2), 123–138.
DOI: 10.3354/cr00953.
- 3389 Troyanskaya, O., Cantor, M., Sherlock, G., Brown, P., Hastie, T., Tibshirani, R., Botstein, D., and Altman,
3390 R. B. (2001). Missing value estimation methods for DNA microarrays. *Bioinformatics*, 17(6), 520–525.
DOI: 10.1093/bioinformatics/17.6.520.
- 3391 Tylianakis, J. M., Didham, R. K., Bascompte, J., and Wardle, D. A. (2008). Global change and species
3392 interactions in terrestrial ecosystems. *Ecology Letters*, 11(12), 1351–1363. DOI: 10.1111/j.1461–
3393 0248.2008.01250.x.
- 3394 United Nations Educational Scientific and Cultural and Organization (2015). *UNESCO Global Science Re-
3395 port: Towards 2030*. Tech. rep. Available at: [http://uis.unesco.org/sites/default/
3396 files/documents/unesco-science-report-towards-2030-part1.pdf](http://uis.unesco.org/sites/default/files/documents/unesco-science-report-towards-2030-part1.pdf).

- 3403 Urban, M. C. (2015). Accelerating extinction risk from climate change. *Science*, 348(6234), 571–573. DOI: 10.1126/science.aaa4984.
- 3405 Ustaoglu, E. and Williams, B. (2017). Determinants of Urban Expansion and Agricultural Land Conversion in 25 EU Countries. *Environmental Management*, 60(4), 717–746. DOI: 10.1007/s00267-017-0908-2.
- 3408 Valiente-Banuet, A., Aizen, M. A., Alcántara, J. M., Arroyo, J., Cocucci, A., Galetti, M., García, M. B., García, D., Gómez, J. M., Jordano, P., Medel, R., Navarro, L., Obeso, J. R., Oviedo, R., Ramírez, N., Rey, P. J., Traveset, A., Verdú, M., and Zamora, R. (2015). Beyond species loss: The extinction of ecological interactions in a changing world. *Functional Ecology*, 29(3), 299–307. DOI: 10.1111/1365-2435.12356.
- 3413 Valipour, M., Bateni, S. M., and Jun, C. (2021). Global surface temperature: A new insight. *Climate*, 9(5), 1–4. DOI: 10.3390/cli9050081.
- 3415 van Buuren, S. and Groothuis-Oudshoorn, K. (2007). MICE: Multivariate Imputation by Chained Equations in R. *Journal of Statistical Software*, 45(3), 1–67. DOI: 10.18637/jss.v045.i03.
- 3417 van der Kaars, S., Miller, G. H., Turney, C. S., Cook, E. J., Nürnberg, D., Schönfeld, J., Kershaw, A. P., and Lehman, S. J. (2017). Humans rather than climate the primary cause of Pleistocene megafaunal extinction in Australia. *Nature Communications*, 8(14142). DOI: 10.1038/ncomms14142.
- 3420 van Rossum, G. (1995). *Python tutorial*. Centrum voor Wiskunde en Informatica (CWI).
- 3421 Verberk, W. C. E. P., Noordwijk, C. G. E. van, and Hildrew, A. G. (2013). Delivering on a promise: integrating species traits to transform descriptive community ecology into a predictive science. *Freshwater Science*, 32(2), 531–547. DOI: 10.1899/12-092.1.
- 3424 Vidan, E., Roll, U., Bauer, A., Grismer, L., Guo, P., Maza, E., Novosolov, M., Sindaco, R., Wagner, P., Belmaker, J., and Meiri, S. (2017). The Eurasian hot nightlife: Environmental forces associated with nocturnality in lizards. *Global Ecology and Biogeography*, 26(11), 1316–1325. DOI: 10.1111/geb.12643.
- 3428 Villéger, S., Mason, N. W. H., and Mouillot, D. (2008). New multidimensional functional diversity indices for a multifaceted framework in functional ecology. *Ecology*, 89(8), 2290–2301. DOI: 10.1890/07-1206.1.
- 3431 Violle, C., Navas, M. L., Vile, D., Kazakou, E., Fortunel, C., Hummel, I., and Garnier, E. (2007). Let the concept of trait be functional! *Oikos*, 116(5), 882–892. DOI: 10.1111/j.0030-1299.2007.15559.x.
- 3434 Wandrag, E. M., Dunham, A. E., Miller, R. H., and Rogers, H. S. (2015). Vertebrate seed dispersers maintain the composition of tropical forest seedbanks. *AoB PLANTS*, 7, plv130. DOI: 10.1093/aobpla/plv130.
- 3437 Watson, J. (2016). Bring climate change back from the future. *Nature*, 534(7608), 437. DOI: 10.1038/534437a.
- 3439 Weeks, B. C., Naeem, S., Lasky, J. R., and Tobias, J. A. (2022). Diversity and extinction risk are inversely related at a global scale. *Ecology Letters*, 25(3), 697–707. DOI: 10.1111/ele.13860.
- 3441 Weiner, C. N., Werner, M., Linsenmair, K. E., and Blüthgen, N. (2011). Land use intensity in grasslands: Changes in biodiversity, species composition and specialisation in flower visitor networks. *Basic and Applied Ecology*, 12(4), 292–299. DOI: 10.1016/j.baae.2010.08.006.

- 3444 Weiss, K. C. and Ray, C. A. (2019). Unifying functional trait approaches to understand the assemblage
3445 of ecological communities: synthesizing taxonomic divides. *Ecography*, 42(12), 2012–2020. DOI: 10 .
3446 1111/ecog.04387.
- 3447 Wenny, D. G., DeVault, T. L., Johnson, M. D., Kelly, D., Sekercioglu, C. H., Tomback, D. F., and Whelan,
3448 C. J. (2011). Perspectives in ornithology the need to quantify ecosystem services provided by birds. *The
3449 Auk*, 128(1), 1–14. DOI: 10 .1525/auk.2011.10248.
- 3450 Wheatley, C. J., Beale, C. M., Bradbury, R. B., Pearce-Higgins, J. W., Critchlow, R., and Thomas, C. D.
3451 (2017). Climate change vulnerability for species—Assessing the assessments. *Global Change Biology*,
3452 23(9), 3704–3715. DOI: 10 .1111/gcb.13759.
- 3453 White, C. R. (2011). Allometric estimation of metabolic rates in animals. *Comparative Biochemistry and
3454 Physiology - A Molecular and Integrative Physiology*, 158(3), 346–357. DOI: 10 .1016/j.cbpa .
3455 2010.10.004.
- 3456 White, C. R. and Kearney, M. R. (2013). Determinants of inter-specific variation in basal metabolic rate.
3457 *Journal of Comparative Physiology B: Biochemical, Systemic, and Environmental Physiology*, 183(1),
3458 1–26. DOI: 10 .1007/s00360-012-0676-5.
- 3459 White, C. R. and Seymour, R. S. (2003). Mammalian basal metabolic rate is proportional to body mass^{2/3}.
3460 *Proceedings of the National Academy of Sciences of the United States of America*, 100(7), 4046–4049.
3461 DOI: 10 .1073/pnas .0436428100.
- 3462 Whitmee, S. and Orme, C. D. L. (2013). Predicting dispersal distance in mammals: A trait-based approach.
3463 *Journal of Animal Ecology*, 82(1), 211–221. DOI: 10 .1111/j.1365-2656.2012.02030.x.
- 3464 Williams, J. J. and Newbold, T. (2020). Local climatic changes affect biodiversity responses to land use: A
3465 review. *Diversity and Distributions*, 26(1), 76–92. DOI: 10 .1111/ddi .12999.
- 3466 Williams, J. J., Freeman, R., Spooner, F., and Newbold, T. (2022). Vertebrate population trends are influenced
3467 by interactions between land use, climatic position, habitat loss and climate change. *Global Change
3468 Biology*, 28(3), 797–815. DOI: 10 .1111/gcb .15978.
- 3469 Wilman, H., Belmaker, J., Simpson, J., de la Rosa, C., Rivadeneira, M. M., and Jetz, W. (2014). EltonTraits
3470 1.0: Species-level foraging attributes of the world's birds and mammals. *Ecology*, 95(7), 2027–2027.
3471 DOI: 10 .1890/13-1917 .1.
- 3472 Wilson, E. E. and Wolkovich, E. M. (2011). Scavenging: How carnivores and carrion structure communities.
3473 *Trends in Ecology and Evolution*, 26(3), 129–135. DOI: 10 .1016/j.tree .2010 .12 .011.
- 3474 Wong, M. K., Guénard, B., and Lewis, O. T. (2018). Trait-based ecology of terrestrial arthropods. *Biological
3475 Reviews*, 94(3), 999–1022. DOI: 10 .1111/brv .12488.
- 3476 Wulder, M. A., Coops, N. C., Roy, D. P., White, J. C., and Hermosilla, T. (2018). Land cover 2.0. *International
3477 Journal of Remote Sensing*, 39(12), 4254–4284. DOI: 10 .1080/01431161 .2018 .1452075.
- 3478 WWF (2020). *Living Planet Report 2020 - Bending the curve of biodiversity loss*. Tech. rep. Available at:
3479 <https://www.zsl.org/sites/default/files/LPR%202020%20Full%20report.pdf>. Almond, R.E.A., Grooten M. and Petersen, T. (Eds). WWF, Gland, Switzerland.
- 3481 Yan, Z., Teng, M., He, W., Liu, A., Li, Y., and Wang, P. (2019). Impervious surface area is a key predictor
3482 for urban plant diversity in a city undergone rapid urbanization. *Science of the Total Environment*, 650,
3483 Part 1, 335–342. DOI: 10 .1016/j.scitotenv .2018 .09 .025.

- 3484 Yessoufou, K., Daru, B. H., and Davies, T. J. (2012). Phylogenetic Patterns of Extinction Risk in the Eastern
3485 Arc Ecosystems, an African Biodiversity Hotspot. *PLoS ONE*, 7(10), 1–8. DOI: 10.1371/journal.
3486 pone.0047082.
- 3487 Young, H. S., McCauley, D. J., Galetti, M., and Dirzo, R. (2016). Patterns, Causes, and Consequences of
3488 Anthropocene Defaunation. *Annual Review of Ecology, Evolution, and Systematics*, 47(1), 333–358.
3489 DOI: 10.1146/annurev-ecolsys-112414-054142.
- 3490 Zamudio, K. R., Bell, R. C., and Mason, N. A. (2016). Phenotypes in phylogeography: Species' traits, en-
3491 vironmental variation, and vertebrate diversification. *Proceedings of the National Academy of Sciences*,
3492 113(29), 8041–8048. DOI: 10.1073/pnas.1602237113.
- 3493 Zhang, J., Qian, H., Girardello, M., Pellissier, V., Nielsen, S. E., and Svenning, J.-C. (2018a). Trophic inter-
3494 actions among vertebrate guilds and plants shape global patterns in species diversity. *Proceedings of the
3495 Royal Society B: Biological Sciences*, 285(1883), 20180949. DOI: 10.1098/rspb.2018.0949.
- 3496 Zhang, Y., Yang, K., Yang, P., Su, Y., Zheng, W., and Liu, J. (2018b). Food restriction decreases BMR, body
3497 and organ mass, and cellular energetics, in the Chinese Bulbul (*Pycnonotus sinensis*). *Avian Research*,
3498 9(39). DOI: 10.1186/s40657-018-0131-8.
- 3499 Zuñiga-Palacios, J., Zuria, I., Castellanos, I., Lara, C., and Sánchez-Rojas, G. (2021). What do we know (and
3500 need to know) about the role of urban habitats as ecological traps? Systematic review and meta-analysis.
3501 *Science of the Total Environment*, 780, 146559. DOI: 10.1016/j.scitotenv.2021.146559.

Appendix 1: Supporting information for Chapter 2

S2.6 Taxonomic corrections

Across the different sources of trait data, similar species could appear under different binomial names. This was a problem when matching datasets by species. Moreover, it is possible that within a source, a given species was appearing under two or more different, synonymous names. As such, taxonomic synonymy created duplicated rows for the same species, overall falsely increasing the total number of species and potentially inflating the number of missing trait values. Taxonomic synonymy was hence a major issue. Due to the large number of species across datasets, extensive manual checks could not be applied. The presence of typos in species names had the same effect as synonymy, erroneously duplicating species. I attempted to correct for taxonomy first by correcting for typos, and second by identifying species which were entered under non-accepted names and replacing these with the accepted name. To this end, I developed an automated procedure, complemented with a few manual entries where errors were opportunistically spotted. Such errors in taxonomy were notably spotted when attempting to retrieve trait data for subsets of species, for analyses not directly related to the work conducted here. Taxonomic synonymy was as such checked manually for 91 species (56 birds, 7 mammals and 28 reptiles); in that case, information was extracted from other diverse sources (such as the Reptile Database (<http://www.reptile-database.org/>); Avibase (<https://avibase.bsc-eoc.org/avibase.jsp?lang=EN&pg=home>); AmphibiaWeb (<https://amphibiaweb.org/>); and additional manual checks using the IUCN Red List for mammals). A column in the Synonym dataset mentions where manual checks were applied (in which case the Synonym dataset was manually corrected).

Automated procedure and outputs

Extracting names from the IUCN Red List and the Integrated Taxonomic Information System (ITIS)

The objectives of the automated procedure were to (1) extract species synonymous binomial names from the

IUCN Red List or from the ITIS, using the ‘rredlist’ (Chamberlain, 2018) and ‘taxize’ (Chamberlain and Szöcs, 2013) R packages; and (2) identify the status of each name (accepted or not accepted). I started by generating a list of all names featuring in any of the sources. These ‘original’ names were corrected for typos (using the ‘gnr_resolve’ function in the ‘taxize’ package). Then, the IUCN Red List was queried and any listed synonyms were stored, as well as the status of each synonym (accepted or not accepted). When species were not found in the IUCN Red List, synonyms were extracted from the ITIS. When species were not found in the ITIS either, corrected names (original names corrected for typos) were used. Family and order designations were extracted using the same procedure and some entries were retrieved from the Global Biodiversity Information Facility taxonomic backbone when not available in the IUCN Red List or in the ITIS (GBIF, <https://www.gbif.org/tools/species-lookup>).

NB: for species entered with the forms *Genus cf.*, *Genus aff.* or *Genus spp.*, the accepted binomial name was left empty.

Output. I generated a ‘Synonym’ dataset containing records of binomial names (14,124 recorded names for birds; 8,743 for amphibians; 6,090 for mammals; and 11,678 for reptiles), and for each I recorded their status and their potential synonyms.

Harmonising taxonomy in trait datasets. Taxonomy across datasets was finally homogenised by replacing synonyms with a uniquely identified accepted name. As a consequence, the total number of identified unique species decreased (Figure S2.1a). The species presenting the highest number of synonyms was the East African mole rat (*Tachyoryctes splendens*), for which I found 12 synonymous names (Figure S2.1b).

The automated procedure was not perfect, and taxonomic errors are likely to have persisted in the trait datasets. The IUCN Red List and the ITIS were not comprehensive taxonomic sources, and for clades with high degrees of synonymy in names, such as reptiles or amphibians, neither the IUCN Red List or the ITIS contained enough information. Taxonomy may be further improved by using class-specific sources in an automated procedure.

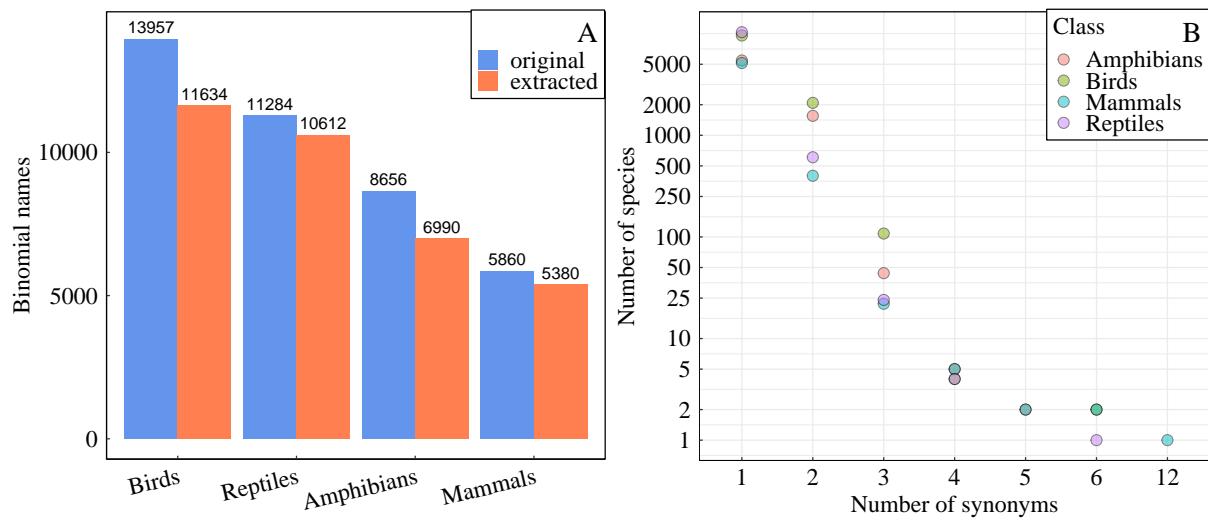


Figure S2.1: Difference in species number before and after taxonomic correction (A) and distribution of number of synonyms across datasets (B). (A) shows the number of species (binomial names) extracted from all sources (blue bars), and the number of uniquely identified accepted names (in red). Replacing non-accepted synonyms by one identified accepted name reduced the number of species in all classes, with the largest reduction for birds. (B) shows the distribution of the number of synonymous names. In all four classes, more than 5,000 species were known under one name only. Nevertheless, a large number of species had two identified synonyms (range: 400 species for mammals - 2086 for birds). The most potentially replicated species was the East African mole rat *Tachyoryctes splendens*, for which 12 synonyms were identified.

S2.7 Additional information for trait compilation

Correlations among closely related traits

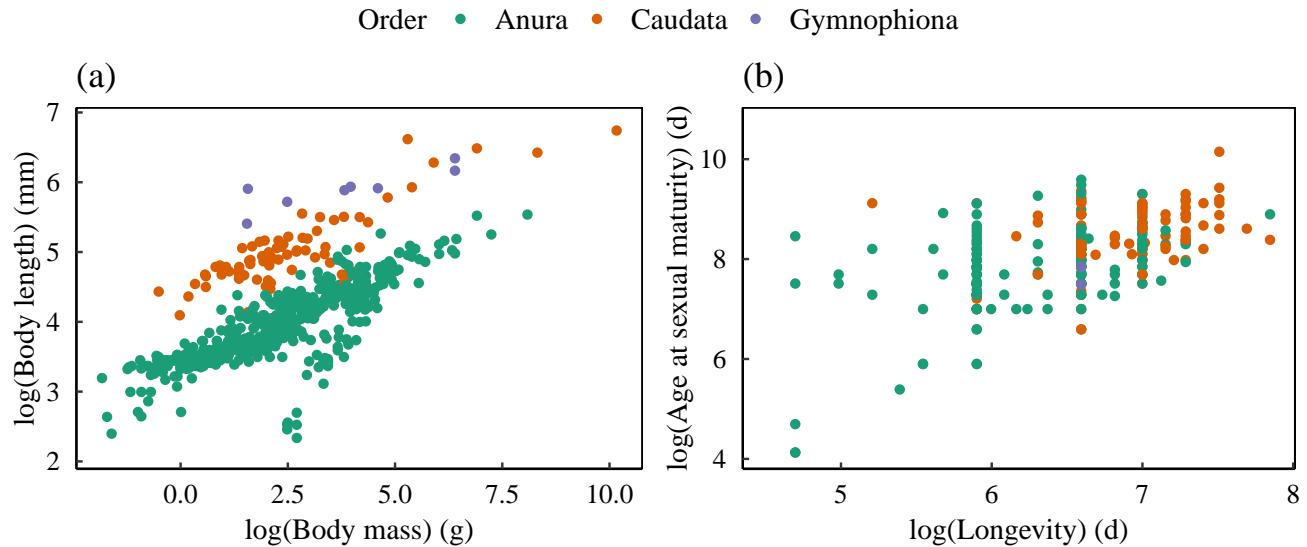


Figure S2.2: (a) Body mass versus body length and (b) longevity versus age at sexual maturity in amphibians. Pearson's correlation coefficient was 0.71 in (a) and 0.55 in (b) (taxonomic order was considered in these coefficients).

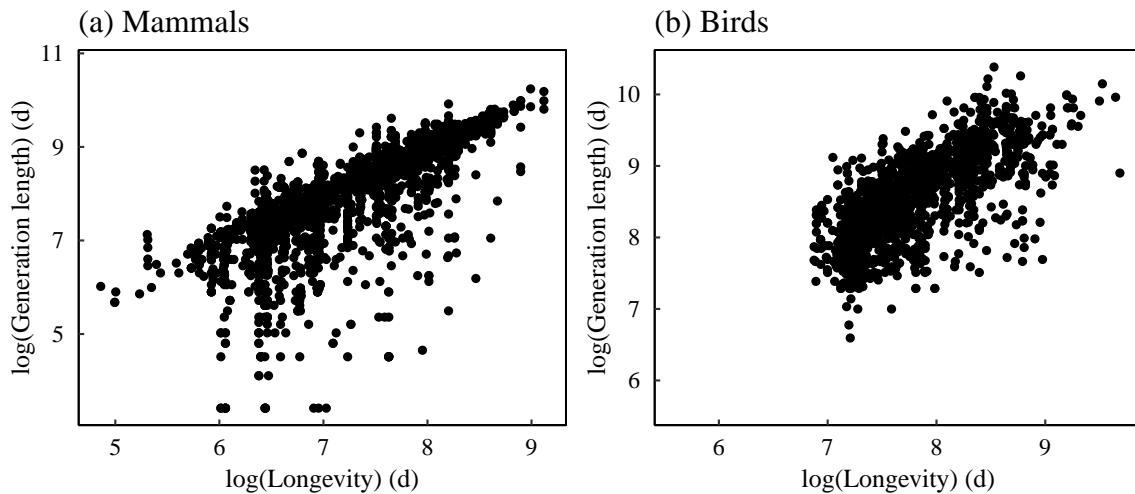


Figure S2.3: Generation length versus longevity data in (a) mammals and (b) birds. Pearson's correlation coefficient was 0.74 in (a) and 0.70 in (b).

S2.8 Cutting distribution maps by altitudinal limits

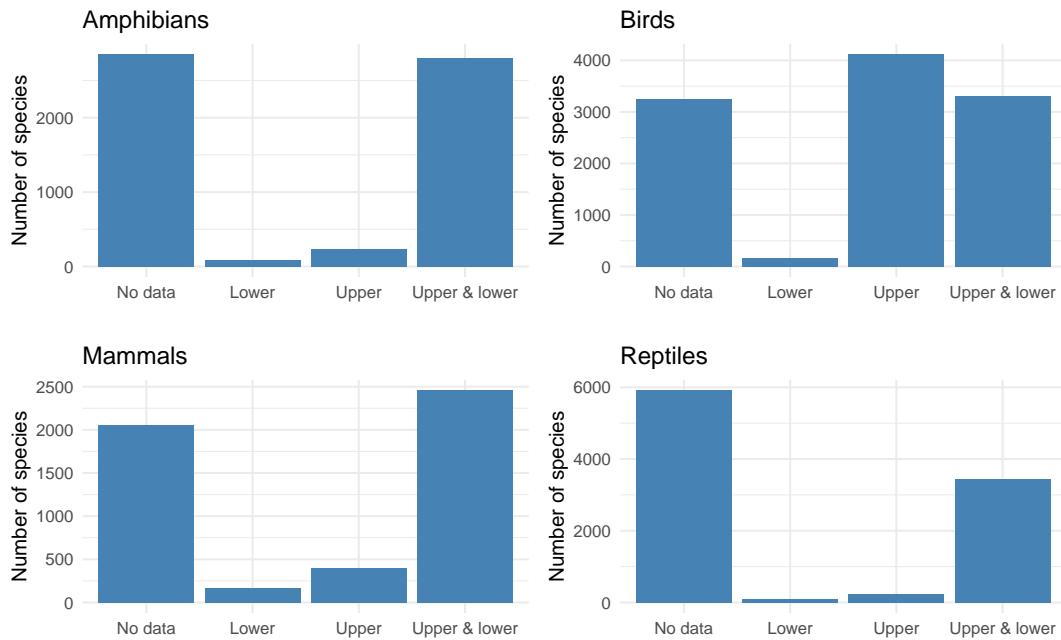


Figure S2.4: Availability of altitudinal limits across species. Upper and lower altitudinal limits were extracted from the IUCN Red List (IUCN, 2020).

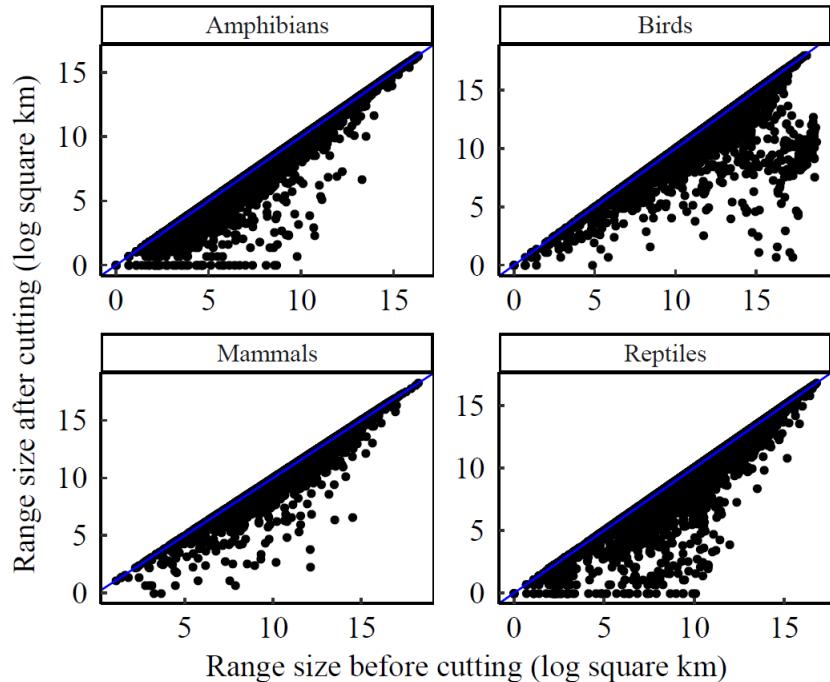


Figure S2.5: Range sizes before versus after cutting the range maps by altitudinal limits.

S2.9 Impact of taxonomic corrections on trait coverage

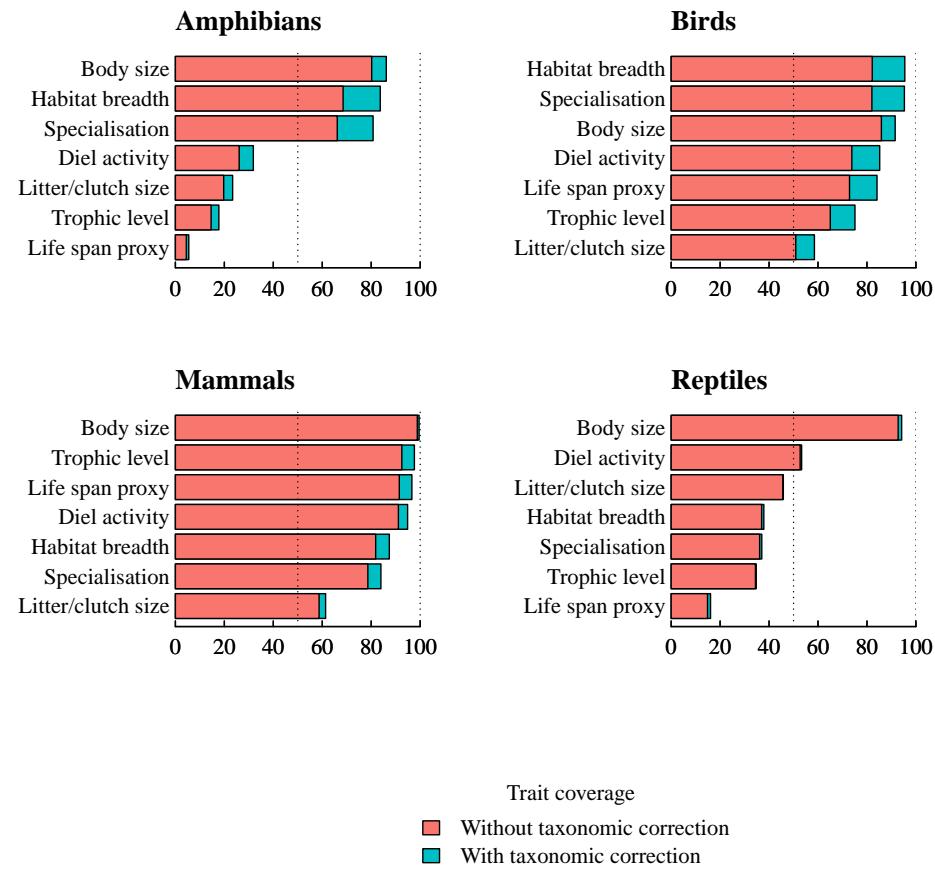
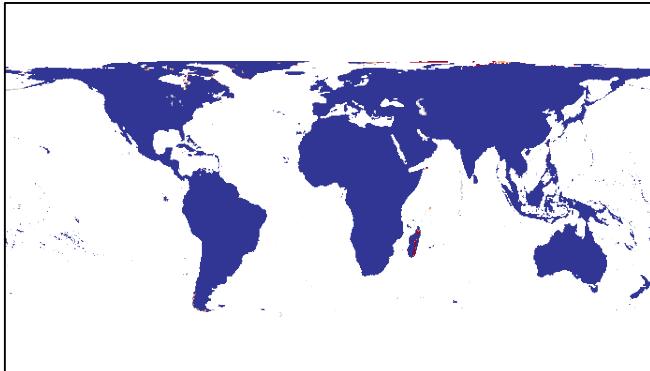


Figure S2.6: Trait coverage when no taxonomic correction is applied before matching sources by binomial names, versus when I applied the described procedure. The identification of synonymous names resulted in an increase in trait coverage in most cases.

S2.10 Assemblage-level median, mean and standard deviation of trait completeness (maps)

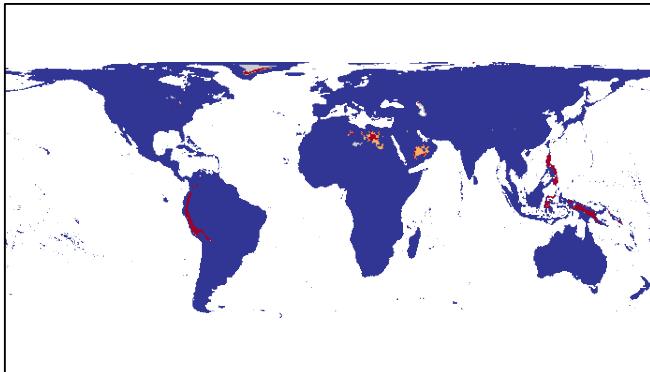
(a) Mammals



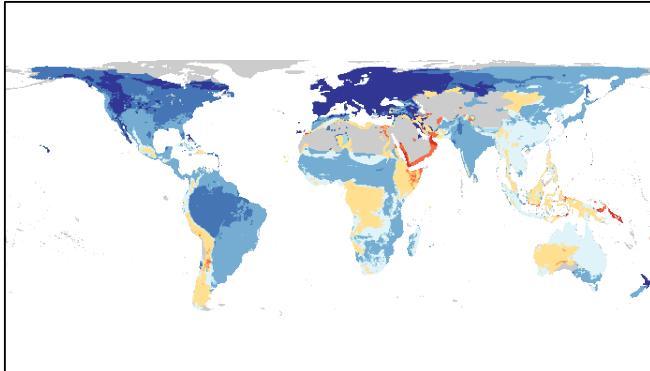
Median completeness
(birds and mammals):

- NA
- 80 – 90%
- 90 – 91%
- 91 – 92%
- 92 – 93%
- 93 – 94%
- 94 – 95%
- 95 – 96%
- 96 – 97%
- 97 – 98%
- 98 – 99%
- 99 – 100%

(b) Birds



(c) Amphibians



Median completeness
(herptiles):

- NA
- 0 – 10%
- 10 – 20%
- 20 – 30%
- 30 – 40%
- 40 – 50%
- 50 – 60%
- 60 – 70%
- 70 – 80%
- 80 – 90%
- 90 – 100%

(d) Reptiles

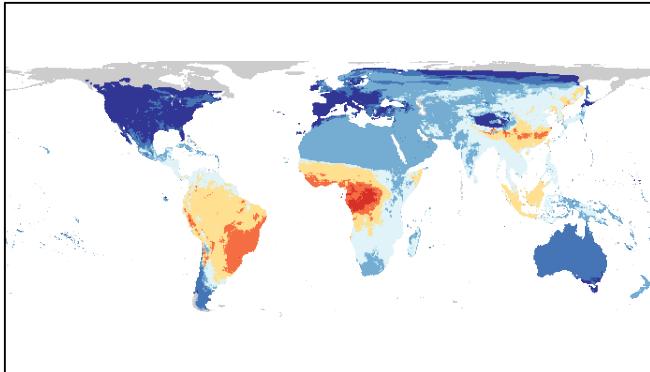
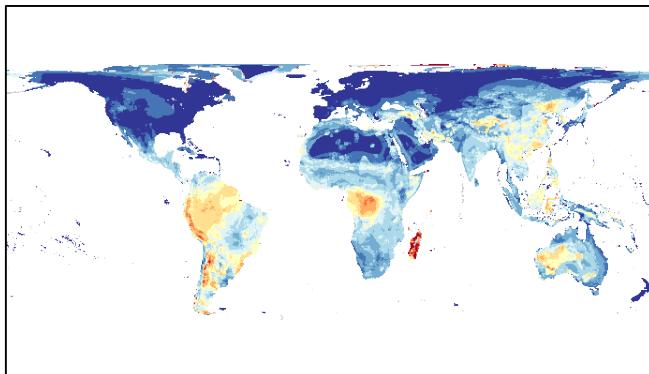


Figure S2.7: Spatial distribution of assemblage-level median trait completeness in each class. Note that the color breaks differ for mammals and birds and for herptiles.

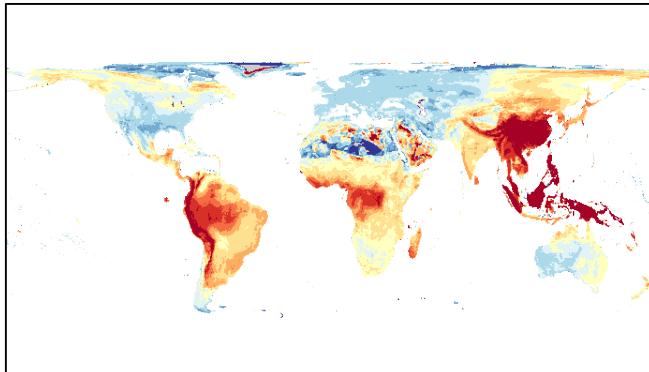
(a) Mammals



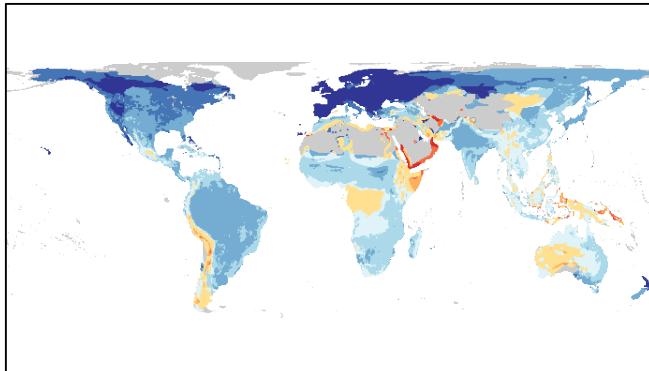
Mean completeness
(birds and mammals):

- NA
- 80 – 90%
- 90 – 91%
- 91 – 92%
- 92 – 93%
- 93 – 94%
- 94 – 95%
- 95 – 96%
- 96 – 97%
- 97 – 98%
- 98 – 99%
- 99 – 100%

(b) Birds



(c) Amphibians



Mean completeness
(herptiles):

- NA
- 0 – 10%
- 10 – 20%
- 20 – 30%
- 30 – 40%
- 40 – 50%
- 50 – 60%
- 60 – 70%
- 70 – 80%
- 80 – 90%
- 90 – 100%

(d) Reptiles

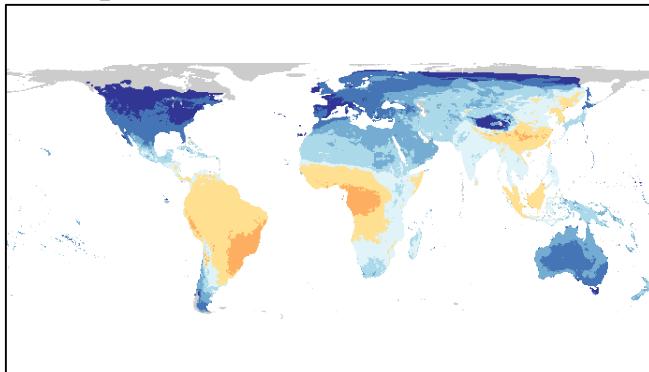
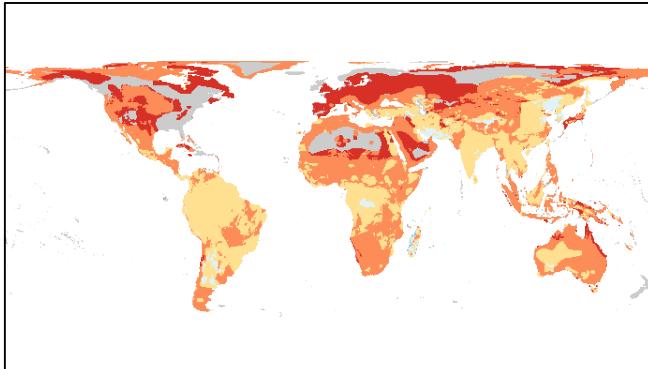


Figure S2.8: Spatial distribution of assemblage-level mean trait completeness in each class. Note that the color breaks differ for mammals and birds and for herptiles.

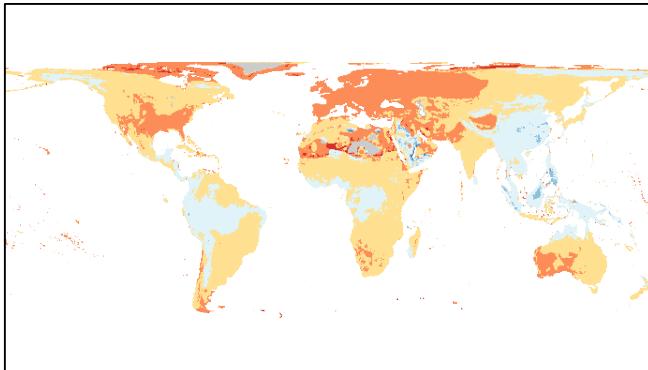
(a) Mammals



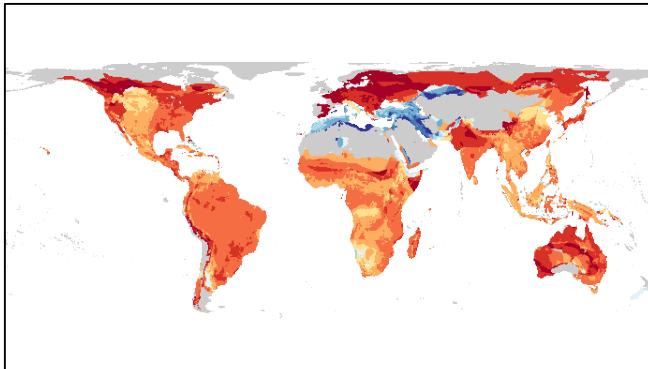
Standard deviation
of completeness
(birds and mammals):

- NA
- 0 – 5%
- 5 – 10%
- 10 – 15%
- 15 – 20%
- 20 – 25%
- 25 – 30%

(b) Birds



(c) Amphibians



Standard deviation
of completeness
(herptiles):

- NA
- 0– 10%
- 10 – 15%
- 15 – 20%
- 20 – 25%
- 25 – 30%
- 30 – 35%
- 35 – 40%
- 40 – 50%
- 50 – 60%
- 60 – 75%

(d) Reptiles

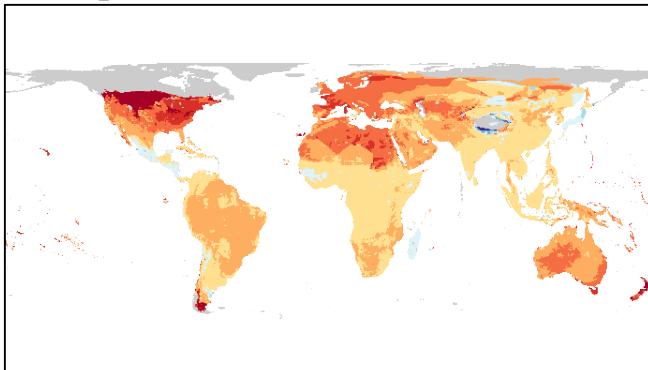


Figure S2.9: Spatial distribution of assemblage-level standard deviation of trait completeness in each class. Note that the color breaks differ for mammals and birds and for herptiles.

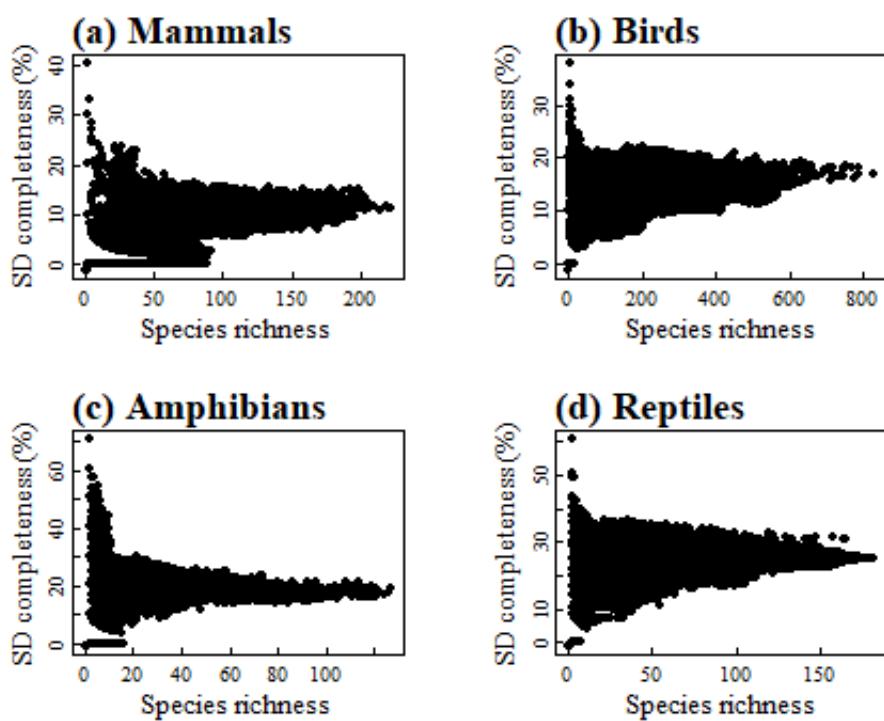


Figure S2.10: Assemblage-level species richness against standard deviation in trait completeness in each class.

S2.11 Phylogenetic patterns in trait completeness

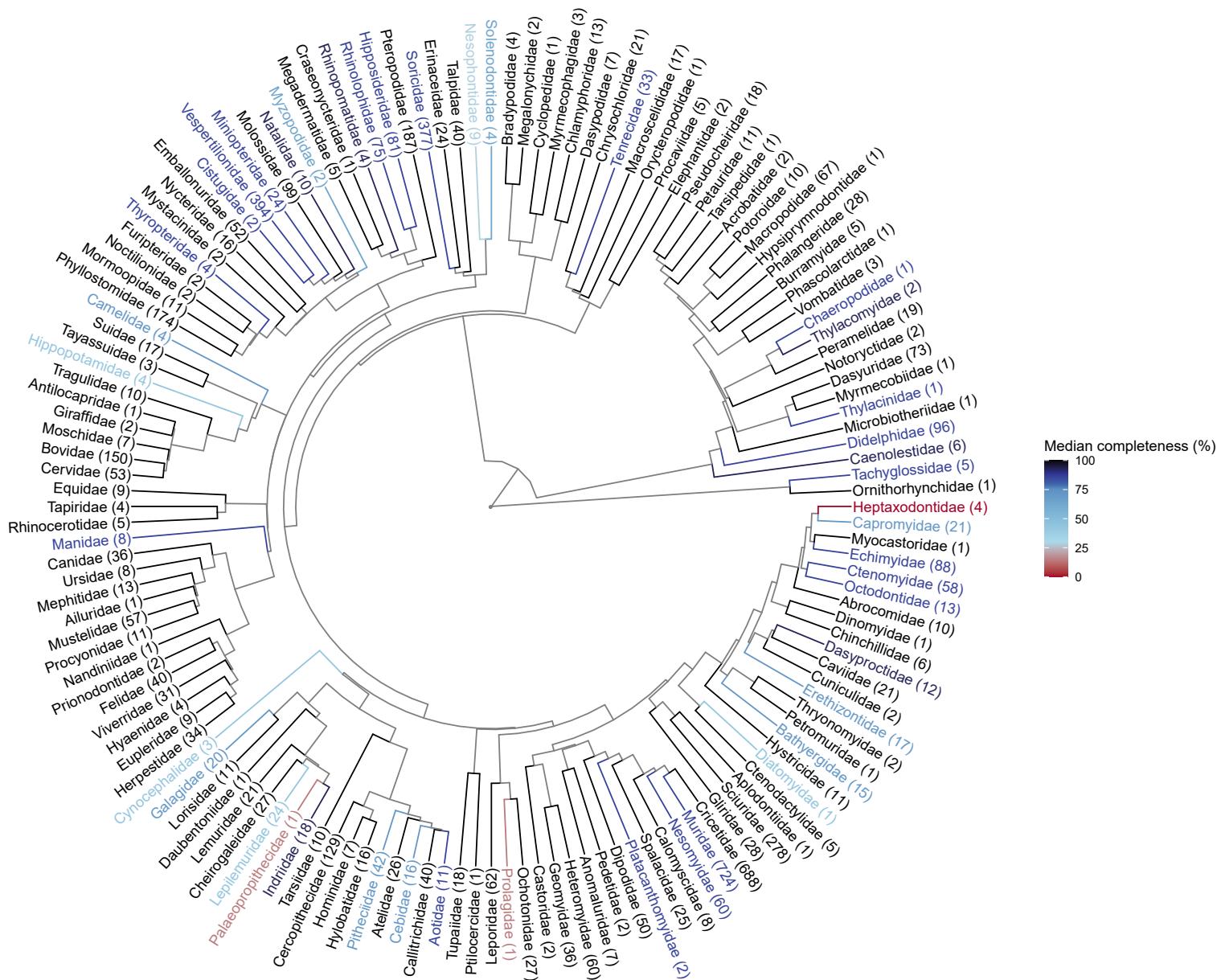


Figure S2.11: Within-family median trait completeness in mammals.

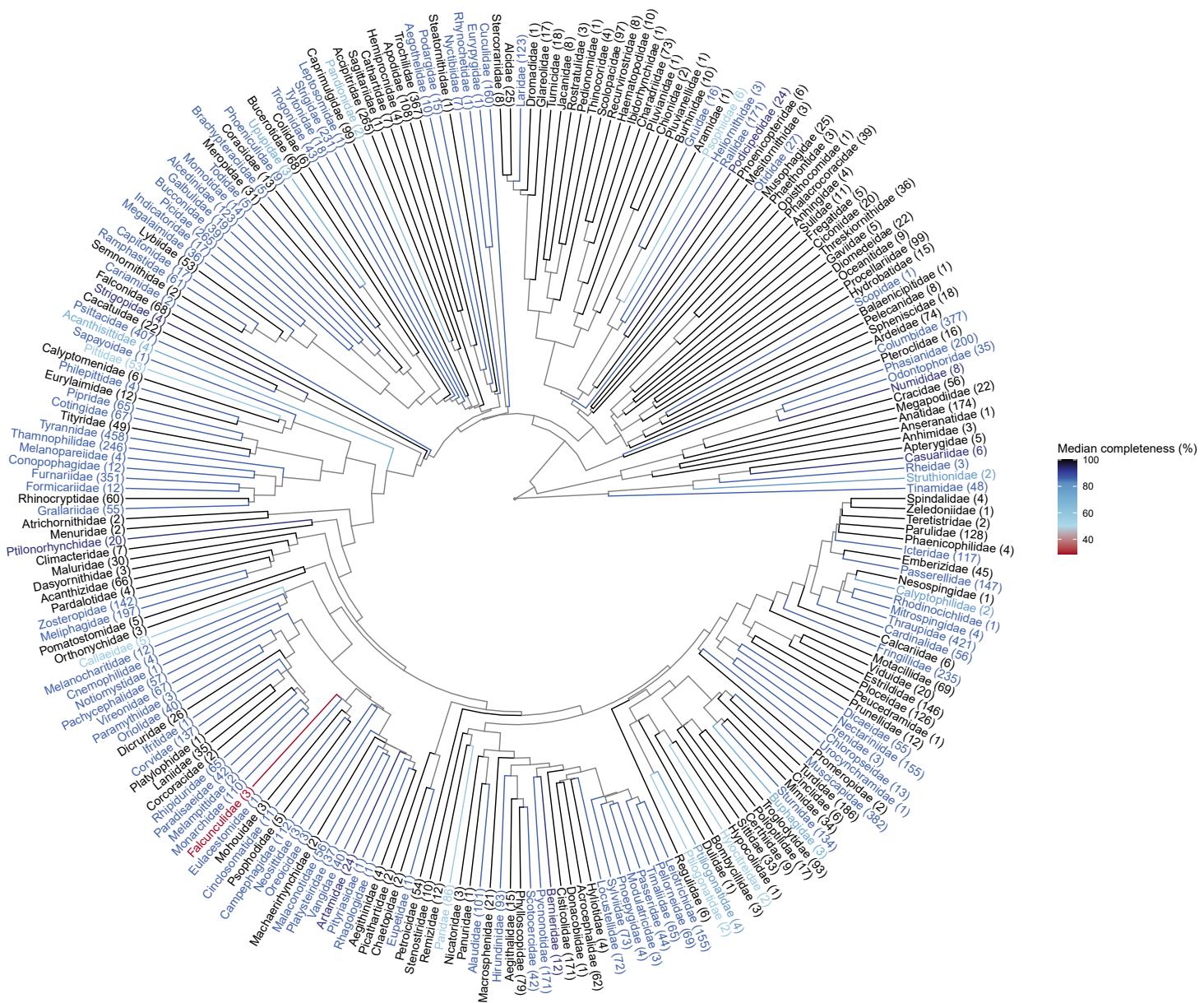


Figure S2.12: Within-family median trait completeness in birds.

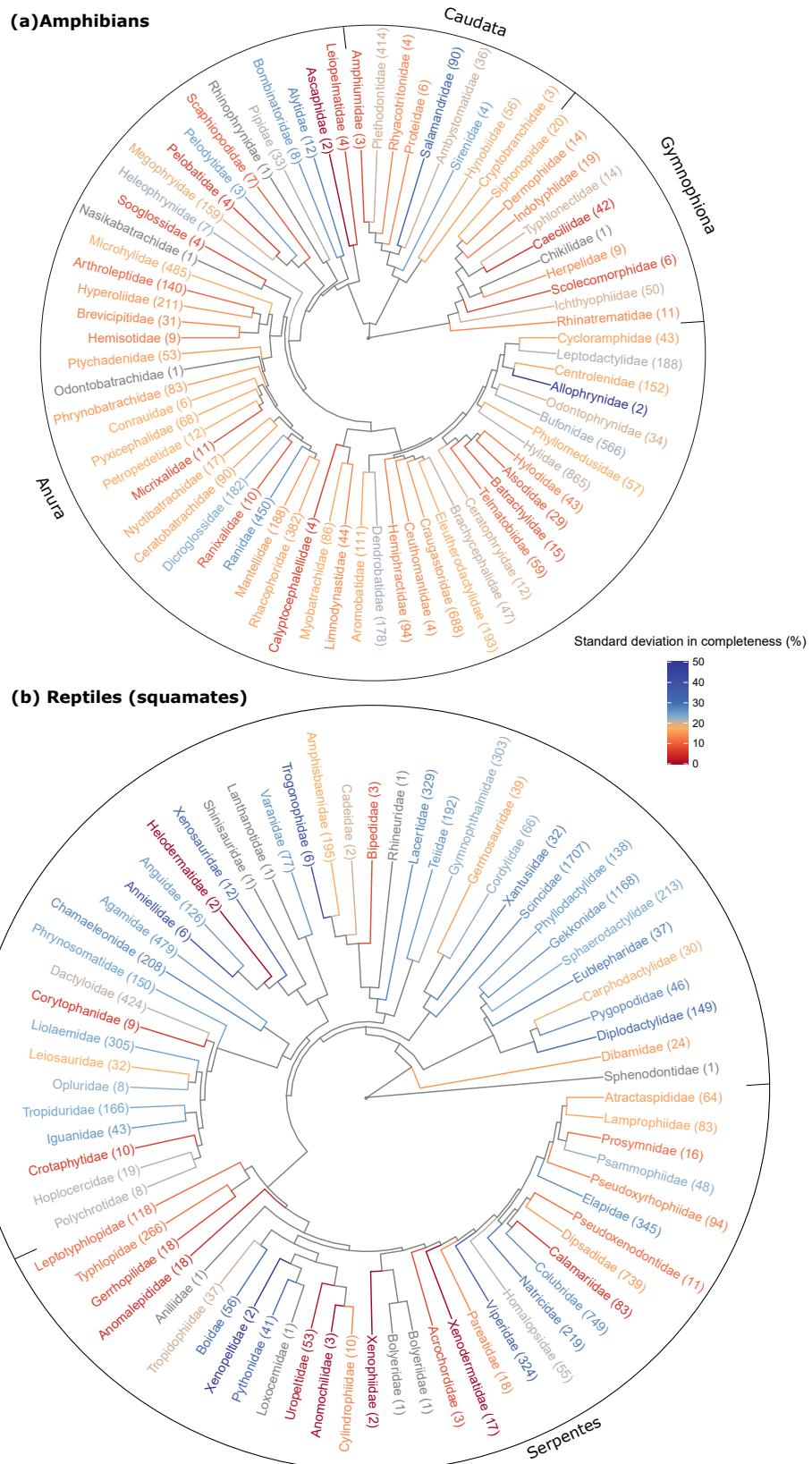


Figure S2.13: Within-family standard deviation in trait completeness (herptiles).

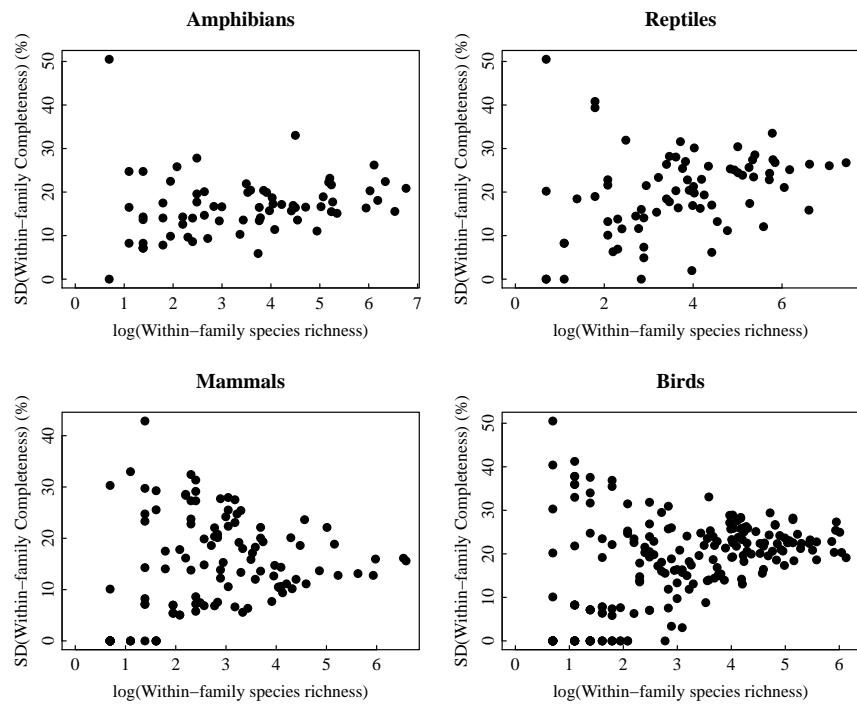


Figure S2.14: Within-family species richness against the within-family standard deviation of trait completeness.

S2.12 Poisson model summaries

Table S2.1: Coefficients of the model investigating whether species range size explained the number of sampled traits. Class was added as an interacting predictor. The reference level for class is mammals. The model was fitted using a Poisson error distribution.

	Estimate	Std. Error	z value	Pr(> z)
Intercept	1.678	0.022	76.809	< 2e - 16
log Range Size	0.015	0.002	8.086	6.16e - 16
Class Birds	-0.092	0.028	-3.350	0.000809
Class Amphibians	-0.689	0.029	-24.099	< 2e - 16
Class Reptiles	-0.872	0.027	-31.856	< 2e - 16
log Range Size:Class Birds	0.003	0.002	1.415	0.157
log Range Size:Class Amphibians	0.017	0.003	6.427	1.30e - 10
log Range Size:Class Reptiles	0.026	0.002	11.159	< 2e - 16

Table S2.2: Coefficients of the model investigating whether species range size explained the number of sampled traits, using range maps not cut by altitudinal limits. Class was added as an interacting predictor. The reference level for class is mammals. The model was fitted using a Poisson error distribution.

	Estimate	Std. Error	z value	Pr(> z)
Intercept	1.665	0.023	72.070	< 2e - 16
log Range Size	0.015	0.002	8.167	3.16e - 16
Class Birds	-0.110	0.029	-3.763	0.0002
Class Amphibians	-0.700	0.030	-23.721	< 2e - 16
Class Reptiles	-0.928	0.029	-32.403	< 2e - 16
log Range Size:Class Birds	0.004	0.002	1.840	0.066
log Range Size:Class Amphibians	0.018	0.003	6.564	5.24e - 11
log Range Size:Class Reptiles	0.031	0.002	12.630	< 2e - 16

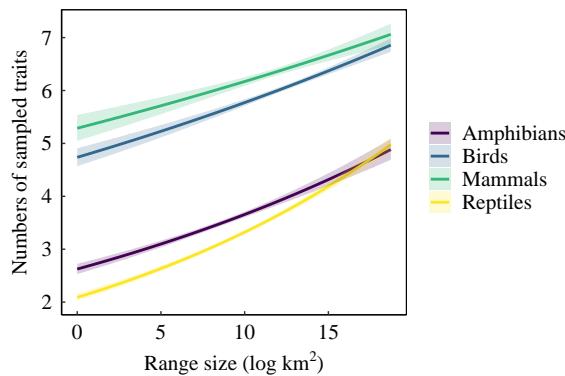


Figure S2.15: Relationship between number of sampled traits and geographical range size using distribution maps not cut by altitudinal limits. The model was fitted using a Poisson error distribution. Class was added as a predictor interacting with range size. Rates of increase were not significantly different for mammals and birds, but differed for reptiles and amphibians, with steeper rates of increase for reptiles overall.

S2.13 Spatial model summaries

Table S2.3: Spatial model summary for amphibians. The spatial model was fitted to explain assemblage-level median completeness with species richness. Biogeographic realm was added as an interacting explanatory variable.

	Estimate	Std. Error	z value	Pr(> z)
Intercept - Realm: Afrotropic	0.0738	0.0064	11.4908	0
log(Species richness)	-0.0025	0.0017	-1.4261	0.1538
Realm: Australasia	-0.0109	0.0095	-1.1453	0.2521
Realm: Indo-Malay	0.0455	0.0119	3.8294	0.0001
Realm: Nearctic	0.0441	0.0082	5.3905	0.000000
Realm: Neotropic	-0.0377	0.0083	-4.5538	0.00001
Realm: Palearctic	0.0047	0.0067	0.6992	0.4844
log(Species richness):Australasia	0.0018	0.0038	0.4789	0.6320
log(Species richness):Indo-Malay	-0.0147	0.0039	-3.7294	0.0002
log(Species richness):Nearctic	-0.0097	0.0030	-3.2003	0.0014
log(Species richness):Neotropic	0.0144	0.0026	5.6454	0.000000
log(Species richness):Palearctic	0.0109	0.0029	3.7358	0.0002

Table S2.4: Spatial model summary for reptiles. The spatial model was fitted to explain assemblage-level median completeness with species richness. Biogeographic realm was added as an interacting explanatory variable.

	Estimate	Std. Error	z value	Pr(> z)
Intercept - Realm: Afrotropic	0.2001	0.0144	13.9349	0
log(Species richness)	-0.0316	0.0031	-10.0547	0
Realm: Australasia	-0.1284	0.0189	-6.7851	0
Realm: Indo-Malay	-0.0453	0.0263	-1.7215	0.0852
Realm: Nearctic	-0.0788	0.0140	-5.6366	0.000000
Realm: Neotropic	-0.0932	0.0145	-6.4425	0
Realm: Palearctic	-0.1030	0.0131	-7.8787	0
log(Species richness):Australasia	0.0386	0.0046	8.4019	0
log(Species richness):Indo-Malay	0.0124	0.0061	2.0397	0.0414
log(Species richness):Nearctic	0.0346	0.0038	9.1601	0
log(Species richness):Neotropic	0.0220	0.0034	6.4231	0
log(Species richness):Palearctic	0.0286	0.0033	8.6153	0

S2.14 Trait coverage and taxonomic matching

Here, I briefly explore the robustness of my work to taxonomic uncertainty by comparing trait coverage obtained with the procedure I developed for taxonomic matching against trait coverage obtained when extracting synonyms from class-specific sources, which could potentially contain more taxonomic information than the ITIS or the IUCN Red List, notably for herptiles. I corrected the datasets for taxonomy again using the ‘rangeBuilder’ R package (Rabosky et al., 2016), which allows to extract accepted names from class-specific sources. Overall, my results are robust to the use of a different taxonomic backbone (Figure S2.16); the main conclusions of this work are likely unaffected by taxonomic uncertainty.

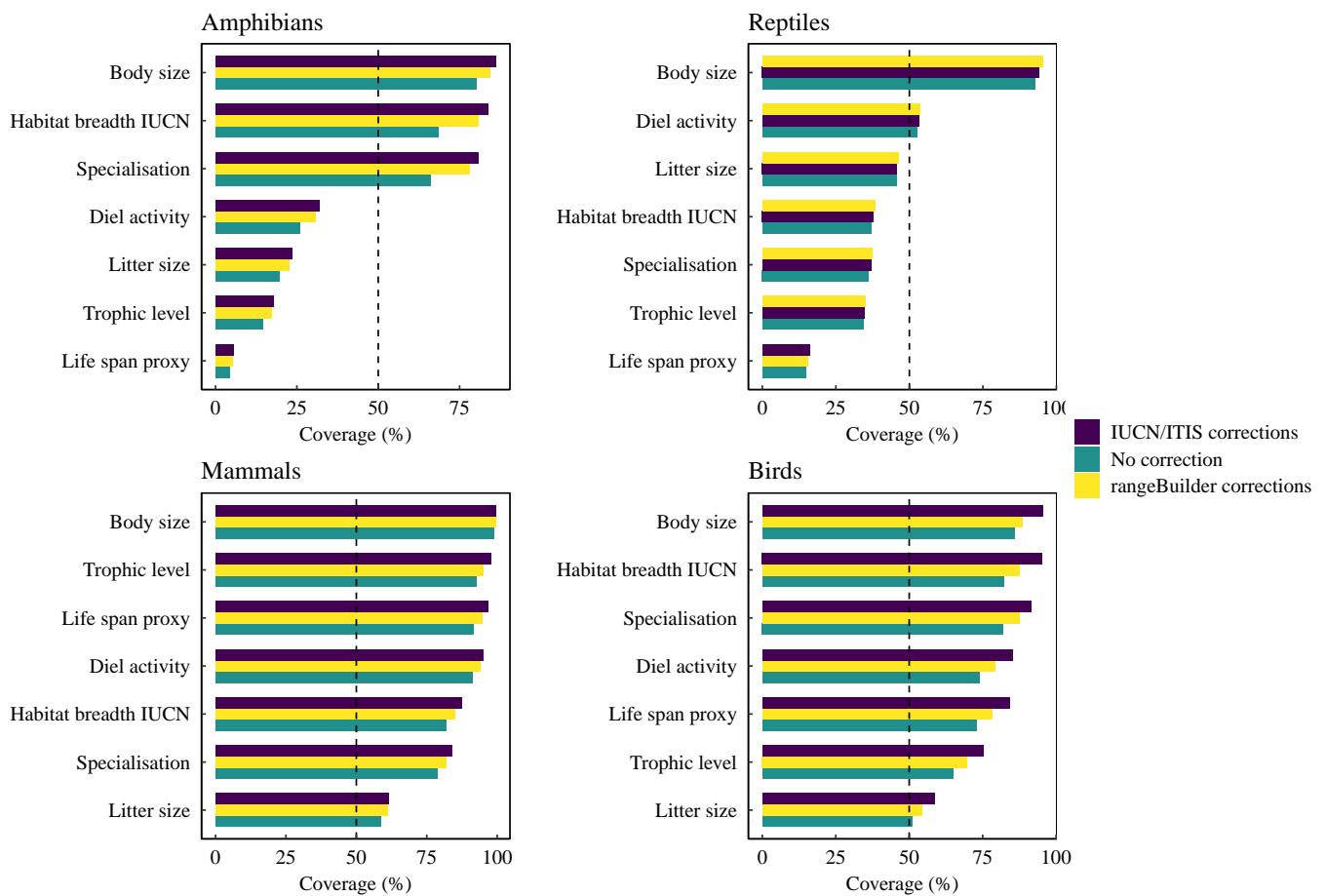


Figure S2.16: Comparison of trait coverage among datasets corrected for taxonomy in different ways: using the described procedure (purple bars); using datasets corrected for taxonomy with the ‘rangeBuilder’ package (yellow bars; Rabosky et al. (2016)); and using datasets uncorrected for taxonomy (green bars).

Appendix 2: Supporting information for Chapter 3

S3.1 Land-use categories in PREDICTS and sample sizes

Table S3.1: Land-use categories in the PREDICTS database. See Hudson et al. (2014, 2017) for more details.

Land-use category	Definition
Primary vegetation	Native vegetation, undisturbed since its establishment under current climatic conditions. No known alterations due to human activities or to extreme natural events.
Mature secondary vegetation	Vegetation recovering after complete destruction of primary vegetation & where succession is near complete – the structure approaches that of primary vegetation.
Intermediate secondary vegetation	Vegetation recovering after complete destruction of primary vegetation at a mid-successional stage.
Young secondary vegetation	Vegetation recovering after complete destruction of primary vegetation at an early successional stage.
Plantation forest	Previously cleared areas planted with crop trees or shrubs grown and harvested for human consumption or for commercial purposes (includes wood, fruit, oil, biofuel, rubber, etc).
Pasture	Areas grazed by livestock, permanently or regularly. Can be improved through cultivation techniques.
Cropland	Previously cleared areas planted with herbaceous crops and harvested for human or animal consumption (including animal feed and crops used in the food industry), or for commercial purposes (e.g., crops grown for the textile industry).
Urban	Previously cleared areas built up by humans. Vegetation is managed for civic or personal purposes.

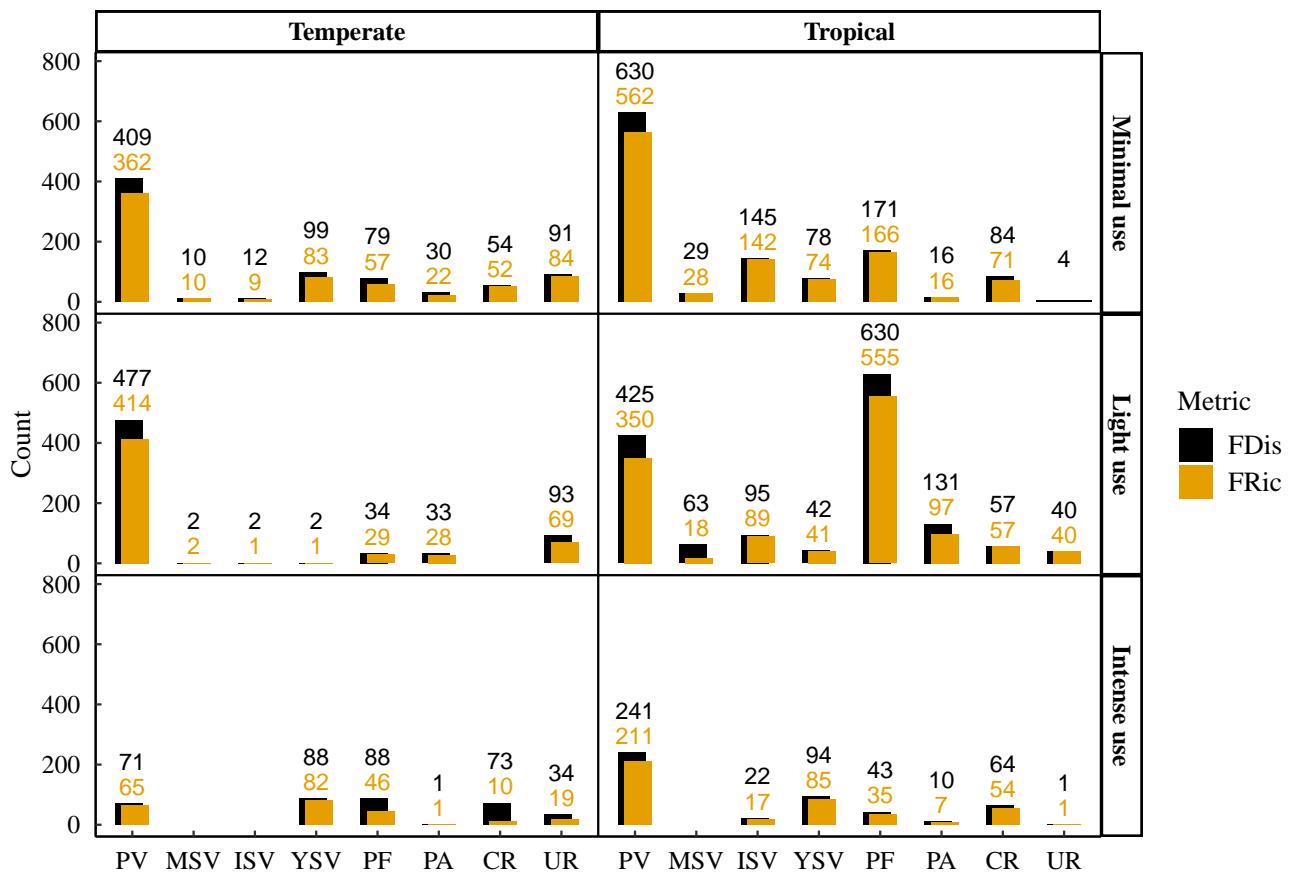


Figure S3.1: Number of sites in each land use and land-use intensity for which FRic and FDis were calculated, across all species. The number of sites for FRic can be smaller than the number of sites for FDis because FRic could not necessarily be computed for all assemblages in which FDis was estimated (in assemblages where species richness was 2 or 3, FRic could not be computed).

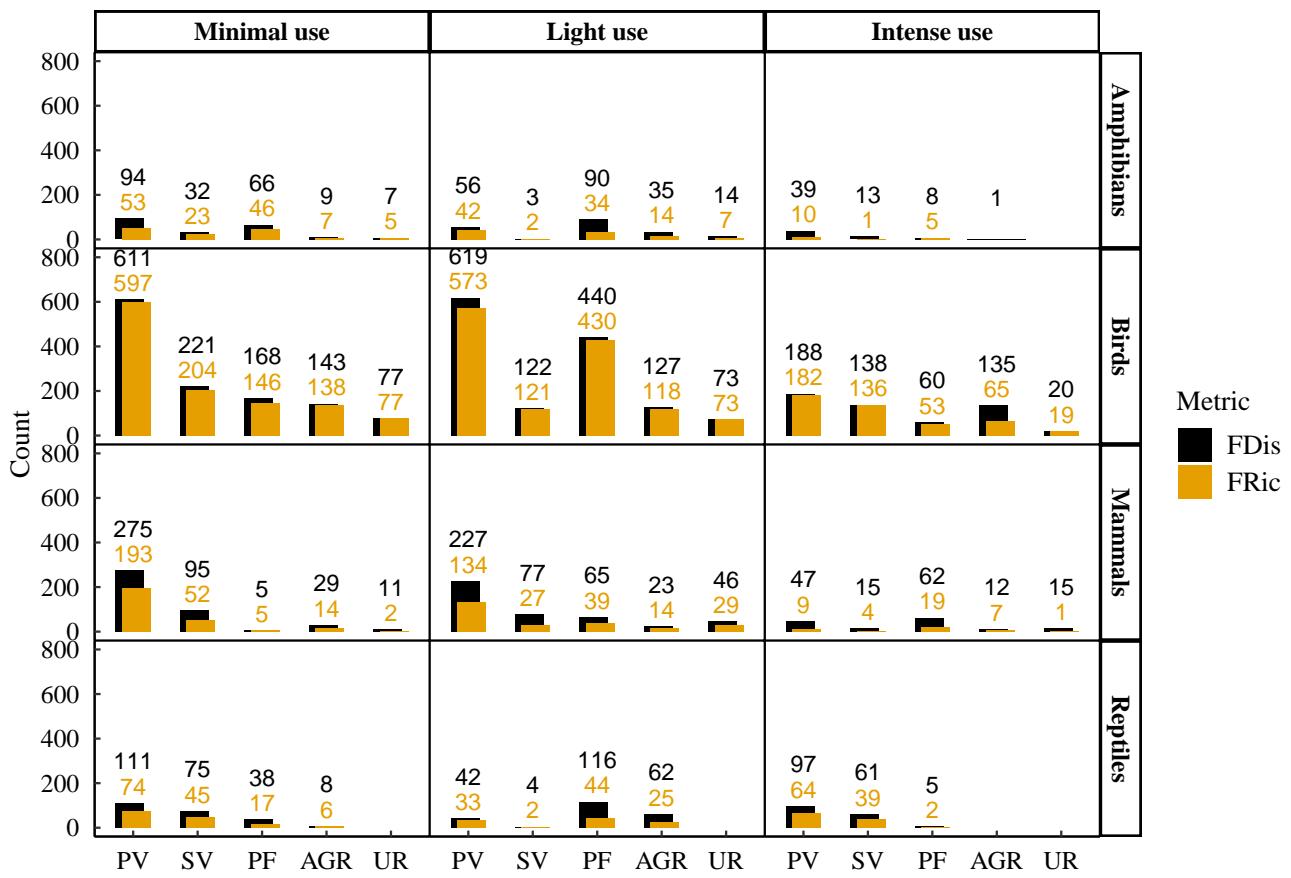


Figure S3.2: Number of sites in each land use, land-use intensity and for which FRic and FDis were calculated, within each class. The number of sites for FRic can be smaller than the number of sites for FDis because FRic could not necessarily be computed for all assemblages in which FDis was estimated (in assemblages where species richness was 2 or 3, FRic could not be computed).

S3.2 Trait data & imputation of missing trait values

I used the compilation of trait data across terrestrial vertebrates from Chapter 2. I selected seven traits that were available at least for a subset of the species in each class: body mass, trophic level, lifespan, litter/clutch size, diel activity, habitat breadth and use of artificial habitats. These traits related to life-history, habitat specialisation and use of geographical space (for instance, habitat breadth is a significant predictor of geographical range size in all classes, Figure S3.3).

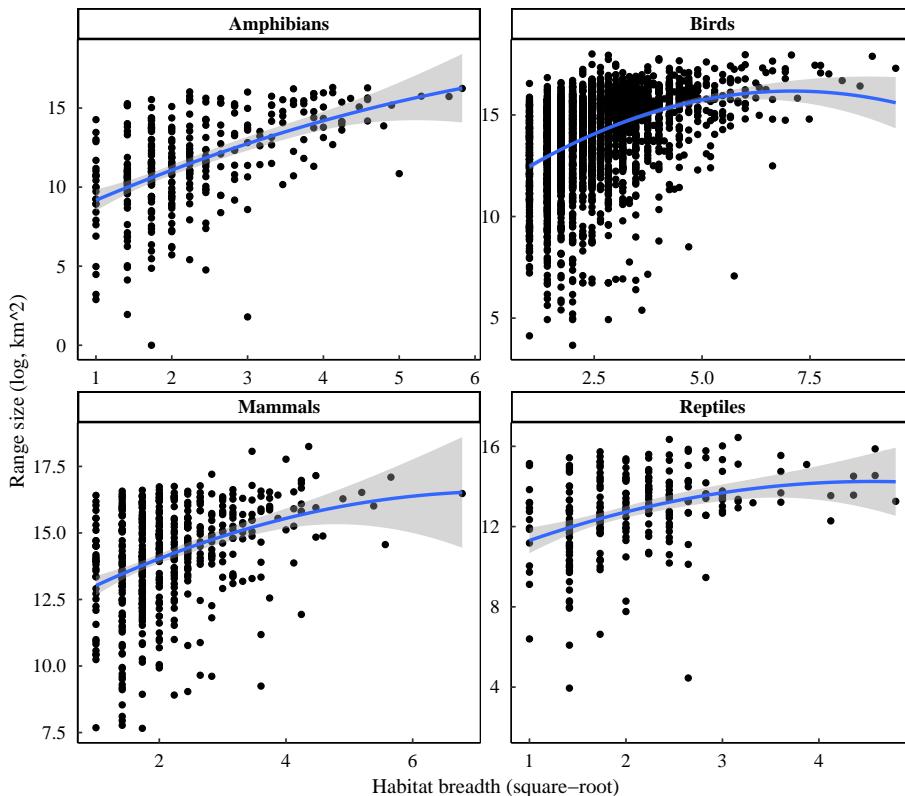


Figure S3.3: Relationship between habitat breadth and geographical range size across species in each class. The derivation of geographical range sizes is described in Chapter 2. The blue lines are the lines of best fit for the regression of geographical range size against habitat breadth in each class, allowing for a quadratic term in the models ($\text{range size} \sim \text{habitat breadth} + \text{habitat breadth}^2$); the shaded areas are the 95% confidence interval of the lines of best fit.

Trait-data coverage was highly variable among classes and traits, with important geographical and phylogenetic biases in trait data for reptiles and amphibians (Chapter 2; Figure S3.4, Figure S3.5). To obtain complete species-trait datasets, I imputed missing trait values. Further, in order to assess the sensitivity of the models to variation in imputed values, I imputed the missing trait values eight independent times. This allowed me to assess the congruence of the model predictions when using the different imputed trait datasets in the analyses.

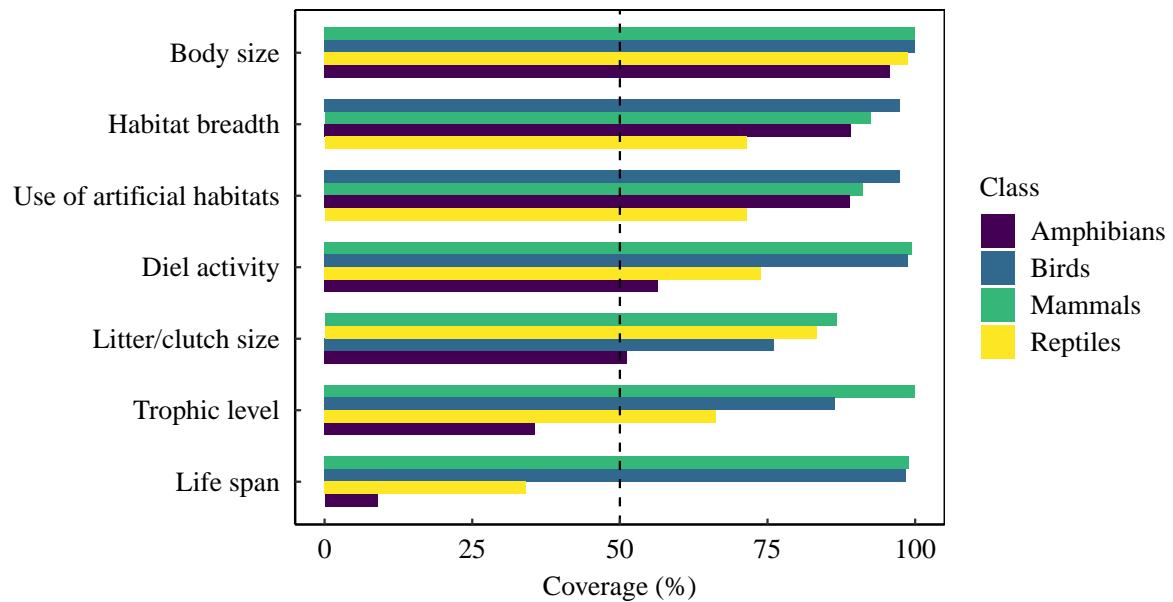


Figure S3.4: Trait coverage for the vertebrate species sampled in the PREDICTS database. For a given trait, coverage is calculated as the percentage of species for which an estimate was available.

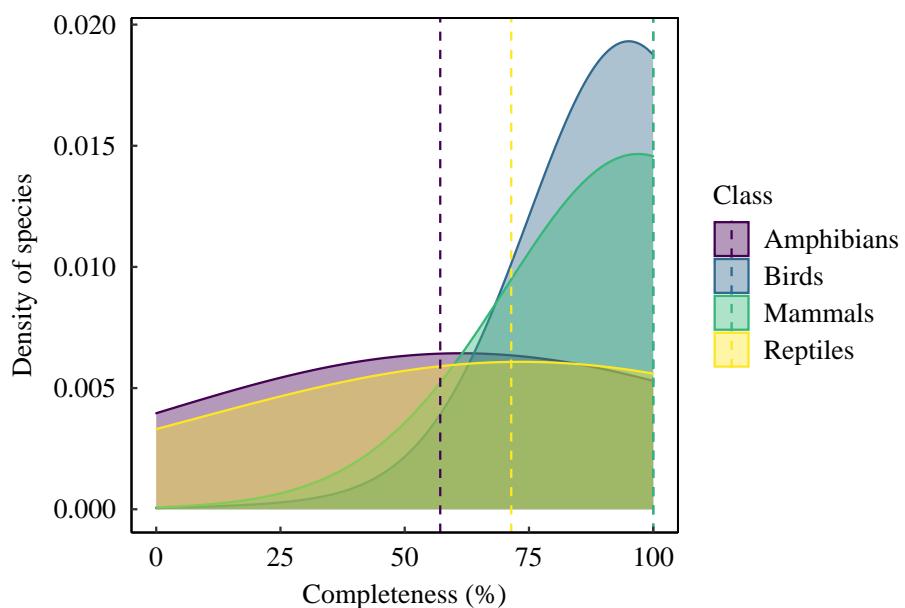


Figure S3.5: Distribution of trait completeness across the vertebrate species sampled in the PREDICTS database. For a given species, trait completeness is calculated as the proportion of traits for which an estimate was available. Dashed lines represent the median trait completeness.

S3.2.1 Choice of imputation technique

There exist several imputation techniques (Debastiani et al., 2021; Johnson et al., 2021; Penone et al., 2014), such as K-nearest neighbour (Troyanskaya et al., 2001), multivariate imputation by chained equations (van Buuren and Groothuis-Oudshoorn, 2007), random forest algorithms (implementable in R with the ‘missForest’ function from the ‘missForest’ package; Stekhoven, 2016; Stekhoven and Bühlmann, 2012), and phylogenetic imputations (implementable in R with PhyloPars, Bruggeman et al., 2009). Penone et al. (2014) assessed the performance of these four techniques and showed that ‘missforest’ and ‘PhyloPars’ performed better when traits were phylogenetically conserved, and when the species phylogenetic position was included as a predictor of missing trait values. ‘PhyloPars’ can only handle continuous data, while ‘missForest’ is compatible with mixed-type (including categorical) data. When no phylogenetic information was included, ‘mice’ was found to be the best method, with fast imputations of mixed-type data (Penone et al., 2014). Therefore, to assess whether ‘missForest’ or ‘mice’ was more appropriate here, I measured the phylogenetic signal in the traits. For continuous traits, I used Pagel’s λ (Pagel, 1999), and for categorical traits I used Borges’ δ (Borges et al., 2018). Strong phylogenetic signals would indicate that traits are phylogenetically conserved, and hence ‘missForest’ would be the most suited approach for imputing missing trait values, with the inclusion of species’ phylogenetic positions as a predictor.

S3.2.2 Phylogenetic signal in traits

Across all classes, similar traits were used for calculating functional diversity metrics: body mass, litter/clutch size, lifespan (using different proxies in different vertebrate classes: generation length for birds and mammals, longevity for reptiles, and age at sexual maturity for amphibians), trophic level, diel activity, habitat breadth and use of artificial habitats. In addition, I included some class-specific traits for the imputations, as certain class-specific traits could be useful predictors of other traits (such as body length for instance in amphibians (Santini et al., 2018)). Table S3.2 details the traits that were included for the imputations in each class and the phylogenetic signal for each of these traits. Continuous traits were log-10 transformed before assessing Pagel’s λ to improve normality. Pagel’s λ was estimated using the ‘phylosig’ function of the ‘phytools’ package (Revell, 2012), and Borges’ δ was assessed using code provided by Borges et al. (2018), available at : https://github.com/mrborges23/delta_statistic. To test for the significance of δ , I generated null distributions of δ for each categorical trait by randomising trait vectors 50 times, and calculating δ for each randomised vector – following the guidelines proposed by Borges et al. (2018). I then tested whether the observed medians were greater than the null distributions using one-sided Wilcoxon rank sum tests.

I used class-specific phylogenies to estimate phylogenetic signal, all downloaded on 13th April 2020. Trees from Faurby et al. (2018, 2020) were used for mammals (downloaded from <https://zenodo.org/record/3690867#.Xyc5wyhKhPZ>). For amphibians, birds and reptiles (squamates only), I downloaded trees from <https://data.vertlife.org/>. Trees were from Jetz et al. (2012) for birds, from Jetz and Pyron (2018) for amphibians and from Tonini et al. (2016) for squamates. For each class, I downloaded a distribution of 1,000 trees, from which I obtained consensus trees to estimate phylogenetic signal (to that end, I used the TreeAnnotator programme of the BEAST software (Bouckaert et al., 2014)).

Table S3.2: Phylogenetic signal in continuous and categorical traits. BM: body mass; BL: body length; GL: generation length; MA: age at sexual maturity; ML: maximum longevity; L: longevity; LCS: litter/clutch size; HB: habitat breadth; TL: trophic level; DA: diel activity; UA: use of artificial habitats. Continuous traits were log-10 transformed to improve normality before estimating Pagel's λ – except for habitat breadth which was square-rooted. A star indicates a significant signal (p-value<0.05 for the log-likelihood ratio test in the case of λ ; and a significant difference from the simulated null distribution of δ for categorical traits). 'NA' indicates traits that were not considered for a given class. All traits showed significant phylogenetic signal, with signals for BM, BL, L, GL, MA and LCS being particularly strong (above 0.8) across the four classes.

Class	Pagel's λ								Borges' δ		
	BM	BL	GL	MA	ML	L	LCS	HB	TL	DA	UA
Amphibians	0.98*	0.94*	NA	0.85*	0.82*	NA	0.93*	0.99*	18*	3.4*	4.5*
Birds	0.99*	NA	0.97*	NA	NA	NA	0.95*	0.60*	13*	32e3*	1.8*
Mammals	0.99*	NA	0.97	NA	NA	NA	0.99	0.71	26*	17*	1.3*
Reptiles	1.0*	NA	NA	NA	0.94*	0.98*	1.0*	0.52*	6.3*	6.4*	1.4*

S3.2.3 Implementation of ‘missforest’ imputations

As phylogenetic signals were strong in many categorical and continuous traits (Table S3.2), I imputed missing trait values using random forest algorithms, as implemented in R with ‘missforest’ from the ‘missForest’ package (Stekhoven, 2016; Stekhoven and Bühlmann, 2012). Another advantage of ‘missforest’ was that, being a non-parametric approach, no prior assumption about data distribution was required. The data were therefore not transformed prior to imputations. In addition, Penone et al. (2014) showed that including phylogenetic information did not decrease the accuracy of imputations for traits that were less phylogenetically conserved, such as habitat breadth in this work.

Phylogenetic relationships were included as additional predictors in the form of phylogenetic eigenvectors (Diniz-Filho et al., 2012), extracted from the phylogenies using the ‘PVR’ package (Santos, 2018). Following Penone et al. (2014), I included the first ten phylogenetic eigenvectors as additional predictors of missing trait values in each class, enough to minimise imputation error. As not all species were represented in the phylogenies, I also added taxonomic order as a predictor for all species. All traits in Table S3.2 were included in the imputations. Tuning parameters of ‘missforest’ were set to ten maximum iterations and to

one hundred trees grown in each forest.

S3.3 Degree of multicollinearity among traits

Multicollinearity among traits can be problematic when calculating functional diversity indices (Cadotte et al., 2011). After imputing missing trait values and before estimating functional metrics, I assessed whether the degree of multicollinearity among categorical and continuous traits was problematically high. To this end, I used generalised variance inflation factors (Fox and Monette, 1992). Given a regression model, variance inflation factors quantify the overestimation in the variance of estimated regression coefficients due to multicollinearity among the predictors. A GVIF value of 5 or 10 is commonly used as a threshold to select out collinear predictors (Dormann et al., 2013). I used the ‘stepwise.vif’ function of the ‘Rnalytica’ package (<https://github.com/awsm-research/Rnalytica>), with a threshold of 5, to determine the GVIF of each trait. I used the imputed traits from the 8th imputation iteration to assess whether multicollinearity was problematically high. Continuous traits were log-10 transformed (except for habitat breadth which was square-rooted). The degree of multicollinearity among traits was not detected to be problematically high, as all traits had a GVIF value below 2 (Table S3.3). As such, all seven traits were included in the calculation of the functional diversity indices.

Table S3.3: Variance inflation factors across considered (imputed) traits.

Trait	GVIF
Diel activity	1.1
Trophic level	1.3
Use of artificial habitats	1.4
Body mass (log10)	1.5
Habitat breadth (square-root)	1.5
Litter/clutch size (log10)	1.6
Lifespan proxy (log10)	1.7

S3.4 Imputation performance

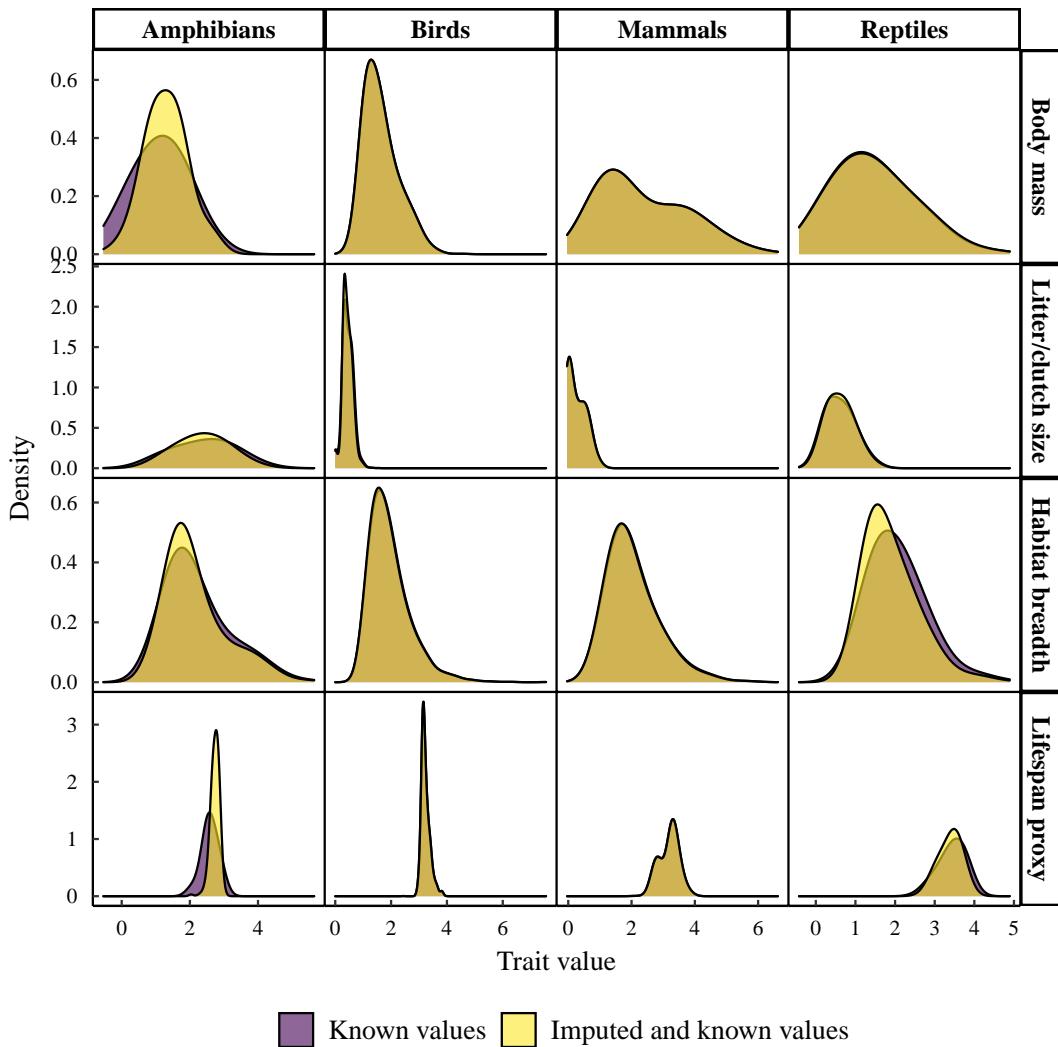


Figure S3.6: Distribution of continuous traits considered in the calculation of the functional diversity metrics (shown as density plots), before and after missing value imputations, in each class and for the species occurring in the PREDICTS database. All traits were log10-transformed, except habitat breadth, which was square-rooted.

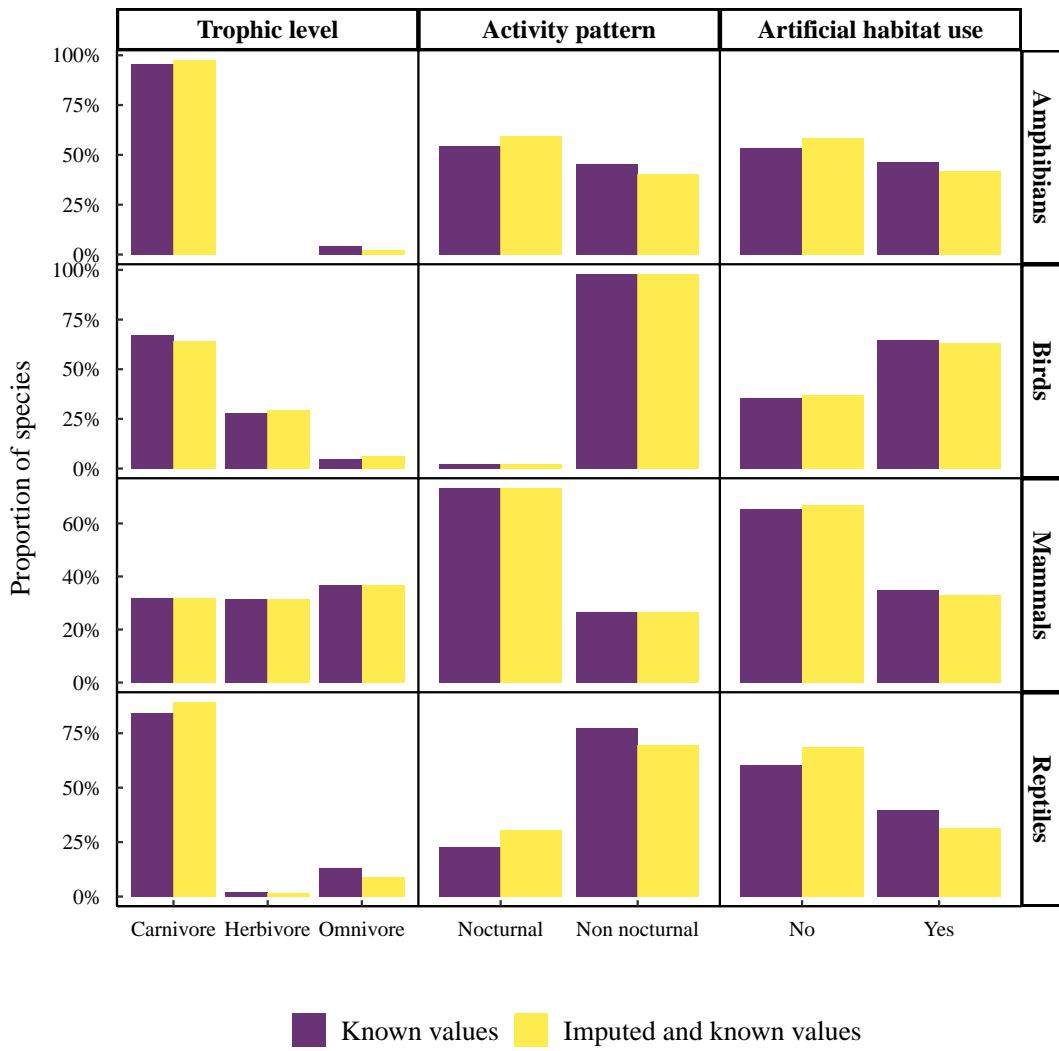


Figure S3.7: Frequency distribution for categorical traits considered in the calculation of the functional diversity metrics (shown as % of total species in each category) before and after missing value imputations, in each class, for the species occurring in the PREDICTS database.

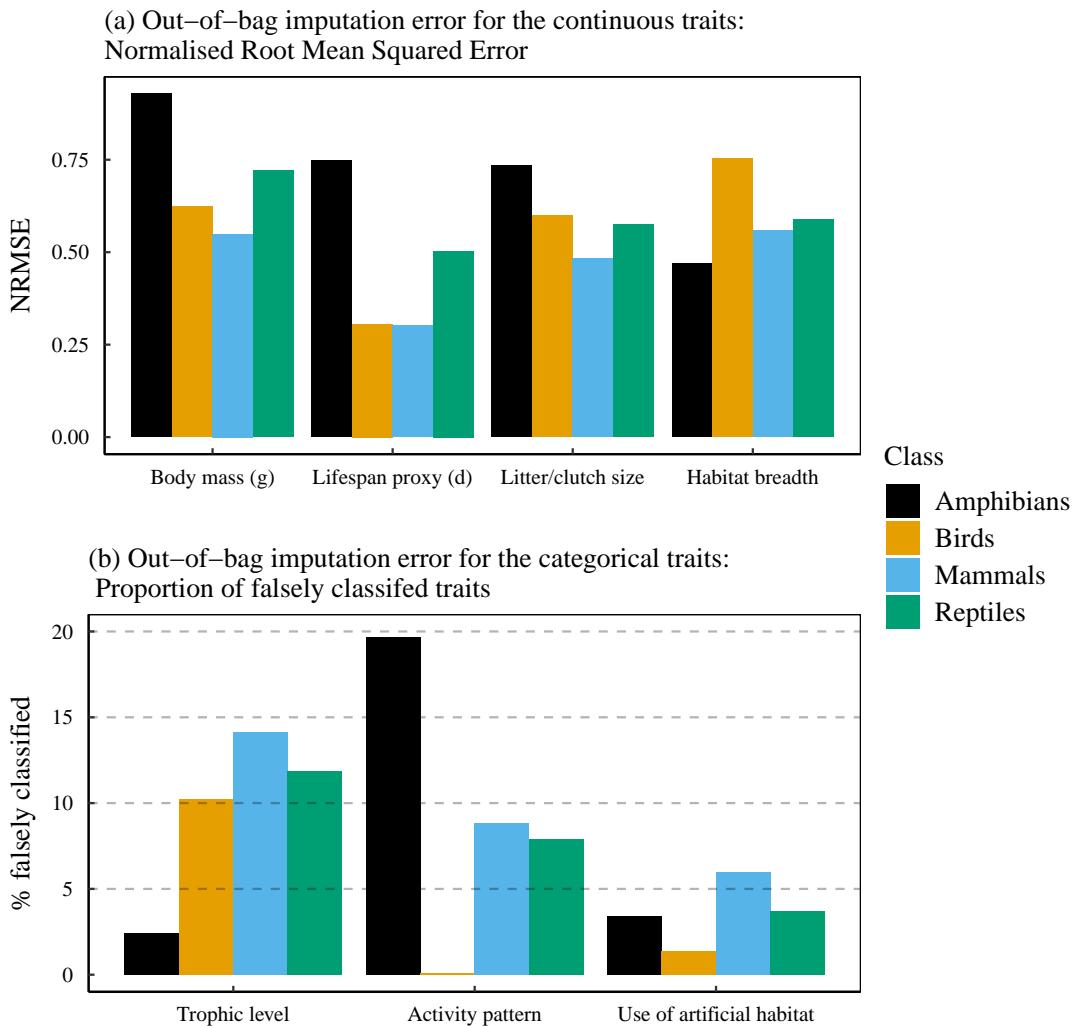


Figure S3.8: Out-of-bag imputation errors for the continuous traits (a) and categorical traits (b). For continuous traits, the Normalised Root Mean Squared Error (NRMSE) was obtained by dividing the Mean Squared Error (MSE) by the variance of the known trait distribution, then square-rooting the result. The MSE was returned for each trait by the ‘missForest’ function (‘missForest’ package, Stekhoven, 2016; Stekhoven and Bühlmann, 2012) and corresponds to an out-of-bag error. For categorical traits, the error was estimated as the out-of-bag proportion of falsely classified traits.

S3.5 Functional loss and functional gain

Across all vertebrates, I estimated functional loss and functional gain using 84 studies for the tropical subset and 39 studies for the temperate subset (51,514 and 30,470 pairwise comparisons between sites respectively, Table S3.4). Because of this large number of pairwise comparisons, I did not develop a null modelling approach (if I used 100 randomisations per pair of sites, I would need to compute functional loss and functional gain for more than 8 million pairs, which would be very computationally demanding). I grouped mature, intermediate and young secondary vegetation together in this analysis. I could not estimate the effects in all land uses (for instance, sample sizes for tropical urban sites were too small; Figure S3.1).

Within classes, I used 18 tropical studies and 1 temperate study for amphibians; 38 tropical studies and 21 temperate studies for birds; 28 tropical studies and 9 temperate studies for mammals; and 11 tropical studies and 7 temperate studies for reptiles. As sample sizes differed among pairs of land uses and land-use intensities (Table S3.5), I was not able to estimate all effects, notably for the intensely-used land uses.

To calculate functional loss and functional gain, the Gower distance matrix (obtained from the global species by trait dataset for the species occurring in PREDICTS) was first subsetted to the species occurring in a given pair of sites. Cailliez corrections were applied when the distance matrix was not Euclidian (Cailliez corrections consist of applying the smallest positive constant to the distances so as to make them Euclidian (Cailliez, 1983); ‘ade4’ R package (Dray et al., 2007)). I then performed a principal coordinates analysis on the (corrected) Gower distance matrix, retaining the first two axes to reduce the computational load in the calculation of convex hulls. Sites that contained fewer than three functionally different species were excluded (the computation of a convex hull requiring more species in the assemblage than PCoA axes). Then I estimated the hypervolume of trait space occupied by each assemblage of a given pair, as well as the volume of the shared trait space (intersection), from which I derived functional loss and functional gain.

Table S3.4: Sample sizes (number of pairs of sites) for the calculation of functional loss and functional gain across all species.

Region	Pairs	Minimal use	Light use	Intense use
Temperate	PV-PV	7626	22546	492
	PV-SV	511	72	–
	PV-PF	9	166	–
	PV-PA	8	40	–
	PV-CR	150	–	–
	PV-UR	6306	1197	7
Tropical	PV-PV	8547	4016	16722
	PV-SV	6584	1124	9713
	PV-PF	580	1378	–
	PV-PA	36	20	22
	PV-CR	1700	1088	–
	PV-UR	–	–	–

Table S3.5: Sample sizes (number of pairs of sites) for the calculation of functional loss and functional gain within each class.

Class	Region	Pair of land uses	Minimal use	Light use	Intense use
Amphibians	Temperate	PV/PV	–	45	–
Amphibians	Temperate	PV/SV	8	70	–
Amphibians	Temperate	PV/AGR	3	–	–
Amphibians	Temperate	PV/UR	6	100	–
Amphibians	Tropical	PV/PV	501	241	307
Amphibians	Tropical	PV/SV	838	–	90
Amphibians	Tropical	PV/PF	422	91	–
Amphibians	Tropical	PV/AGR	1	3	1
Birds	Temperate	PV/PV	7,382	19,300	491
Birds	Temperate	PV/SV	150	1	–
Birds	Temperate	PV/PF	9	166	–
Birds	Temperate	PV/AGR	145	40	–
Birds	Temperate	PV/UR	6,300	992	–
Birds	Tropical	PV/PV	5,059	3,117	9,014
Birds	Tropical	PV/SV	3,491	1,058	5,225
Birds	Tropical	PV/PF	156	994	–
Birds	Tropical	PV/AGR	1,626	1,085	–
Mammals	Temperate	PV/PV	110	3,030	–
Mammals	Temperate	PV/SV	25	–	–
Mammals	Temperate	PV/AGR	5	–	–
Mammals	Temperate	PV/UR	–	105	7
Mammals	Tropical	PV/PV	1,989	637	64
Mammals	Tropical	PV/SV	230	65	8
Mammals	Tropical	PV/PF	2	–	–
Mammals	Tropical	PV/AGR	109	20	21
Reptiles	Temperate	PV/PV	132	2	1
Reptiles	Temperate	PV/SV	250	1	–
Reptiles	Temperate	PV/AGR	5	–	–
Reptiles	Tropical	PV/PV	989	137	5,140
Reptiles	Tropical	PV/SV	1,760	1	3,456
Reptiles	Tropical	PV/PF	–	190	–

S3.6 Diagnostic plots

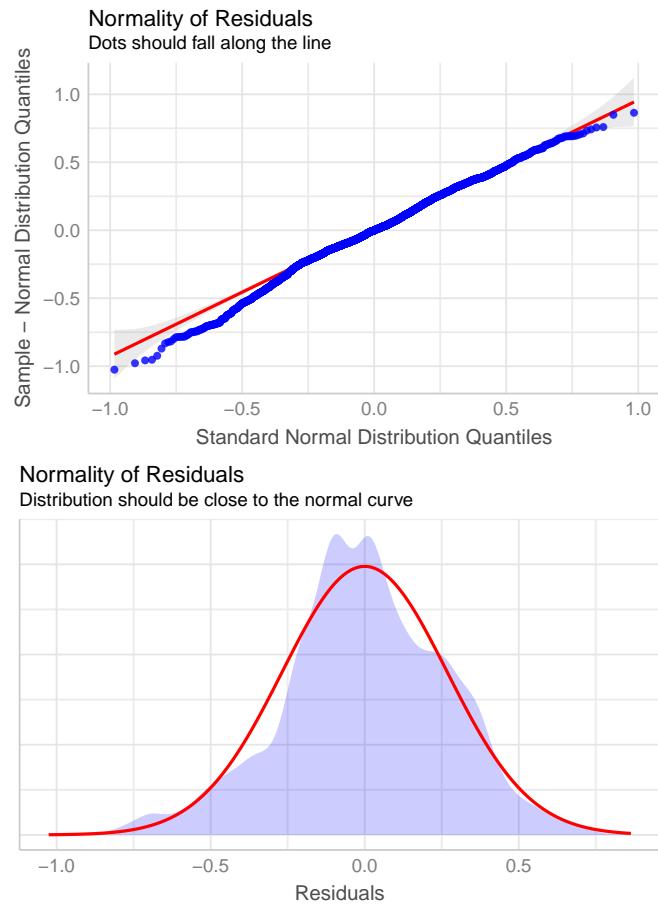


Figure S3.9: Diagnostic plots for Model 1a, obtained using the 'performance' R package (Lüdecke et al., 2021).

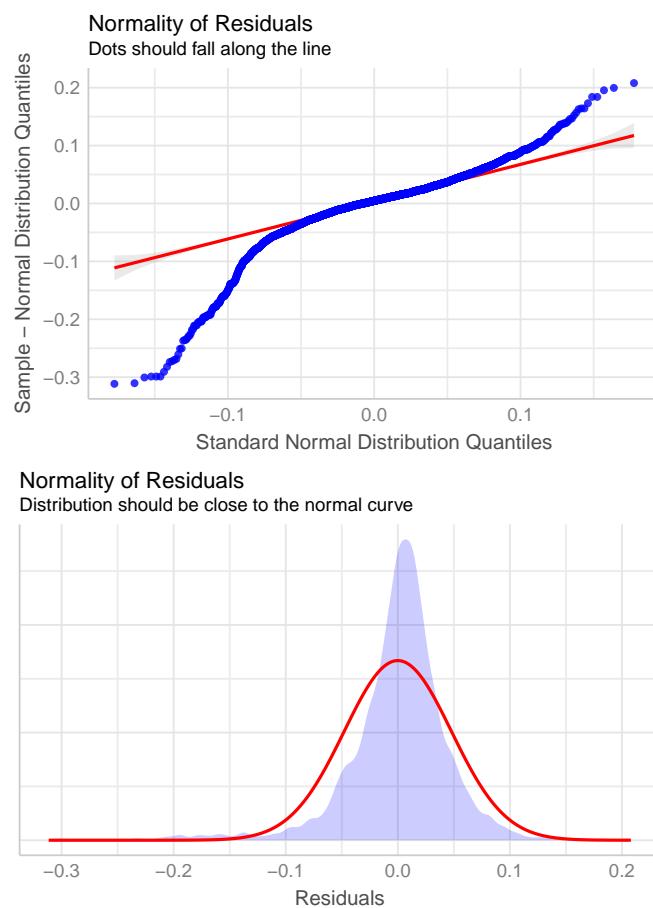


Figure S3.10: Diagnostic plots for **Model 1b**, obtained using the ‘performance’ R package (Lüdecke et al., 2021).

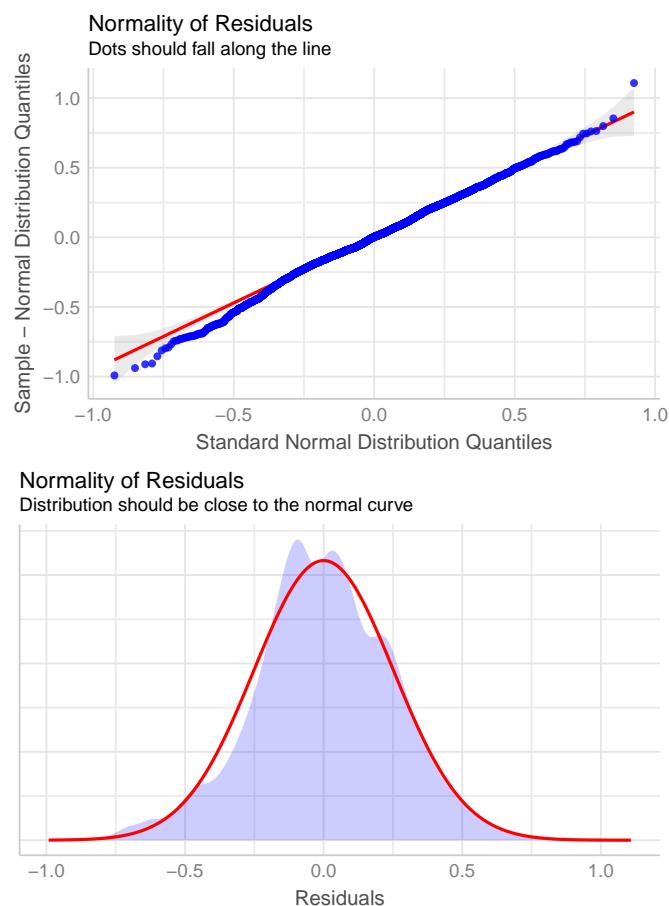


Figure S3.11: Diagnostic plots for Model 2a, obtained using the ‘performance’ R package (Lüdecke et al., 2021).

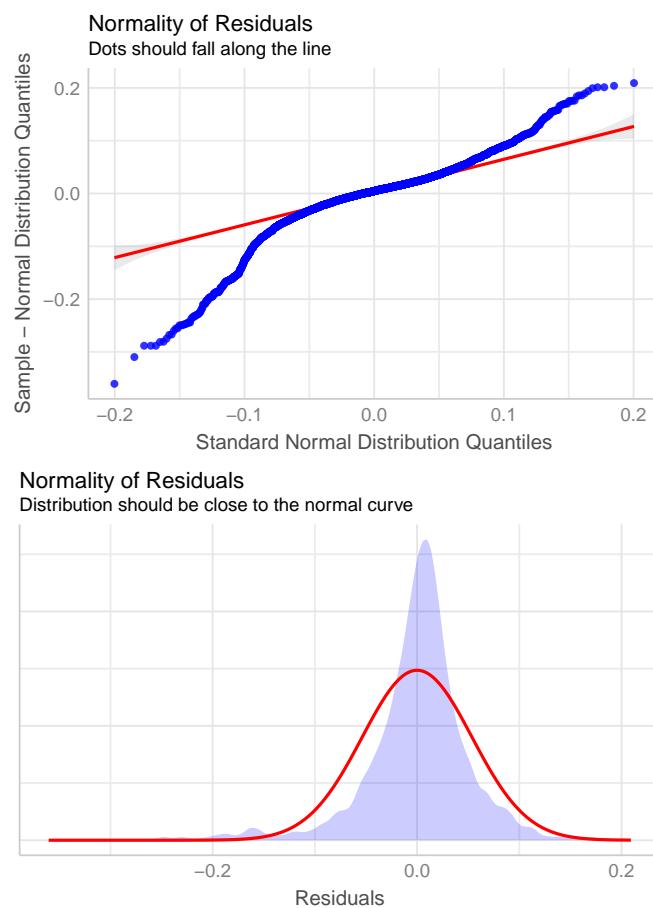


Figure S3.12: Diagnostic plots for Model 2b, obtained using the ‘performance’ R package (Lüdecke et al., 2021).

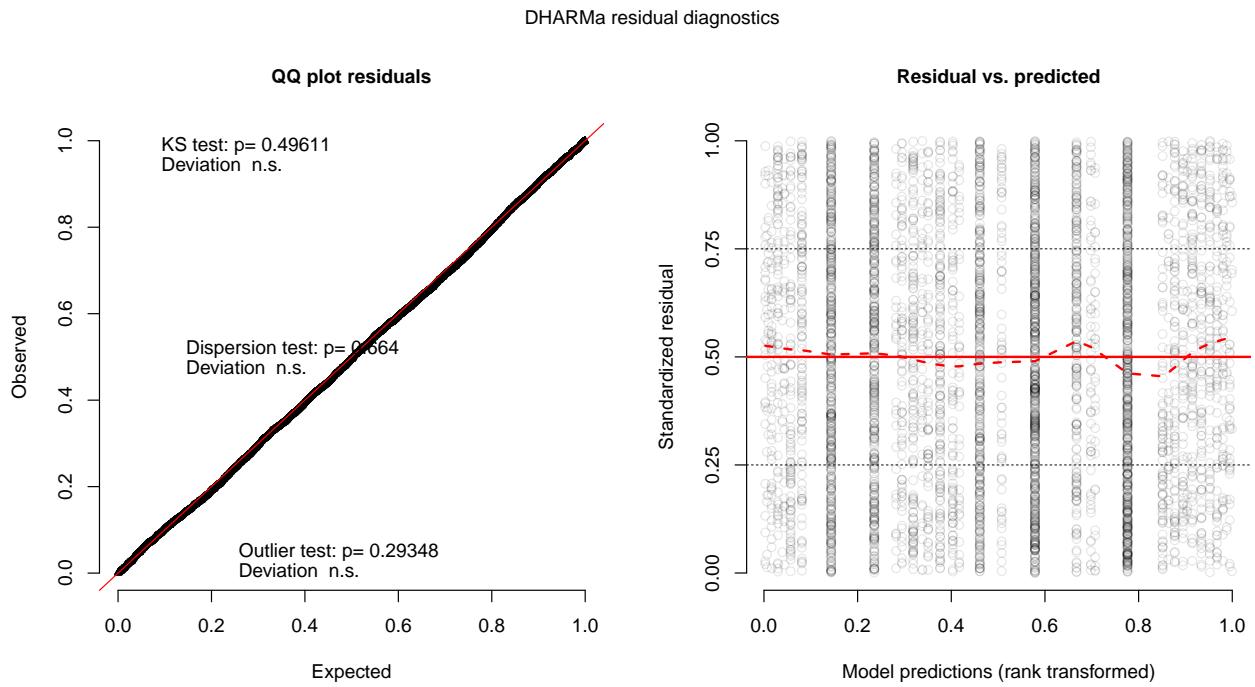


Figure S3.13: Diagnostic plots for Model 3, obtained using the ‘DHARMA’ R package (Hartig, 2021).

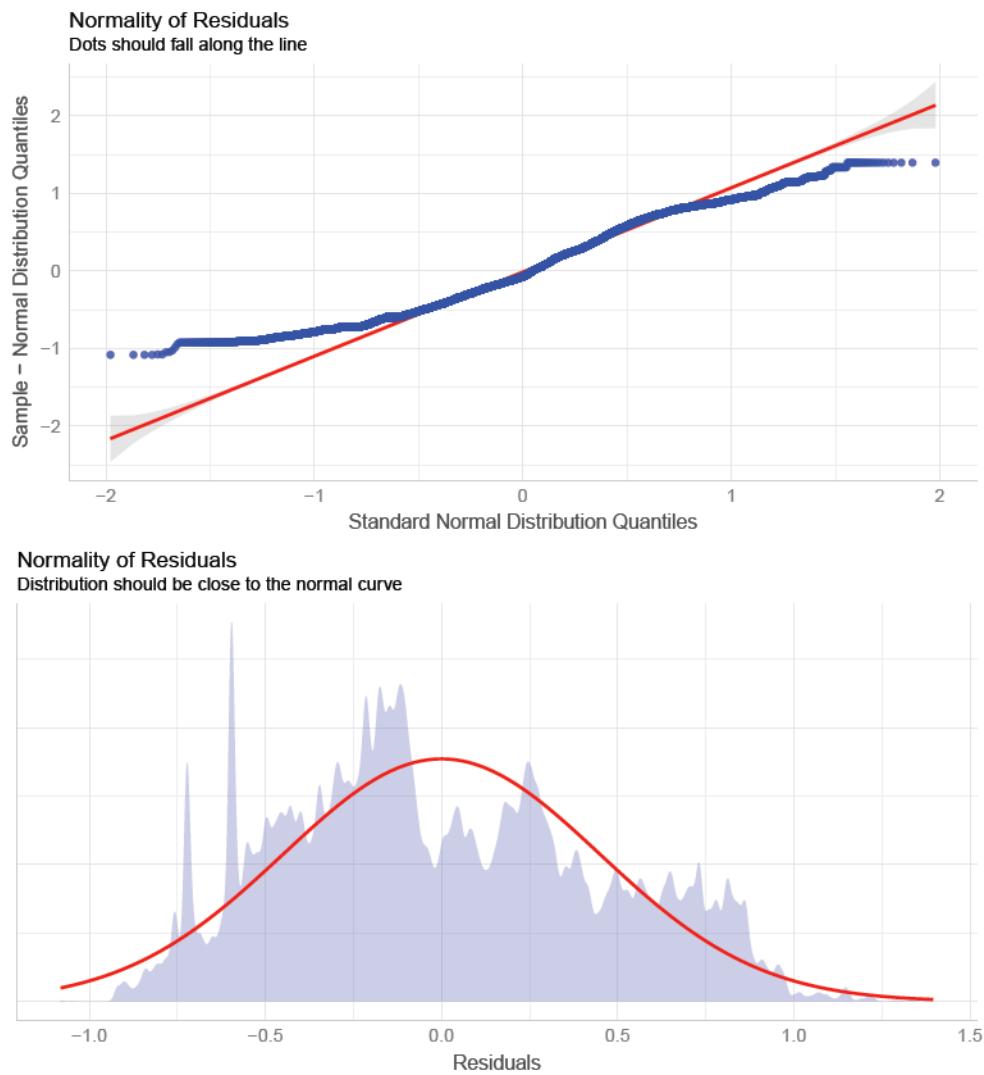


Figure S3.14: Diagnostic plots for Model 4a, obtained using the ‘performance’ R package (Lüdecke et al., 2021).

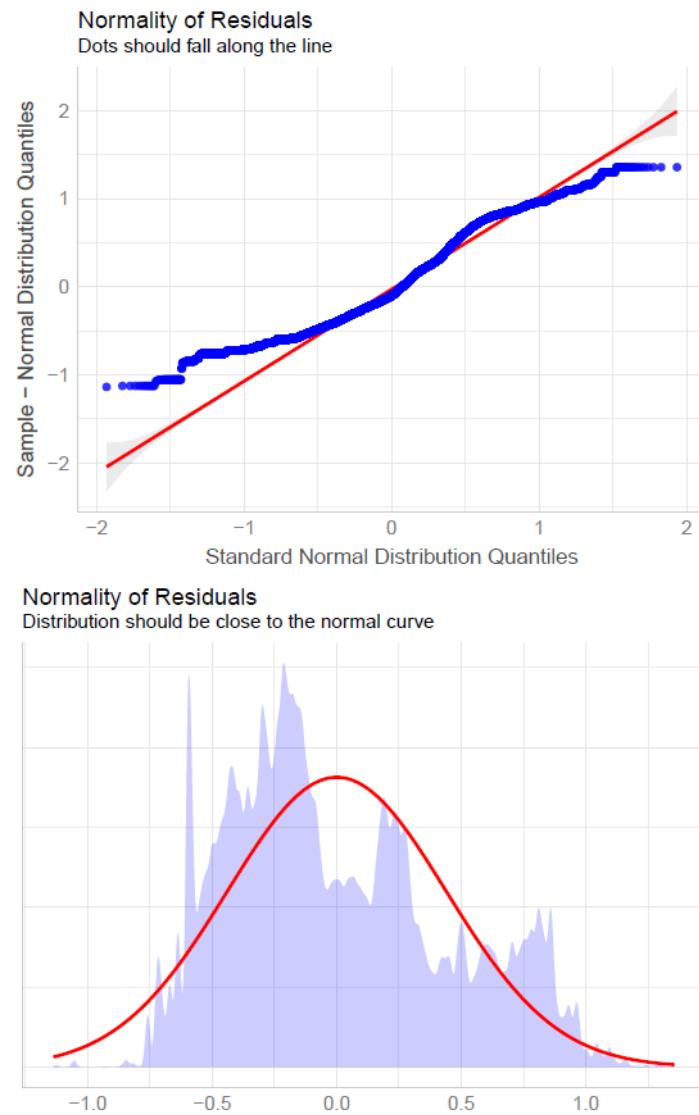


Figure S3.15: Diagnostic plots for Model 4b, obtained using the ‘performance’ R package (Lüdecke et al., 2021).

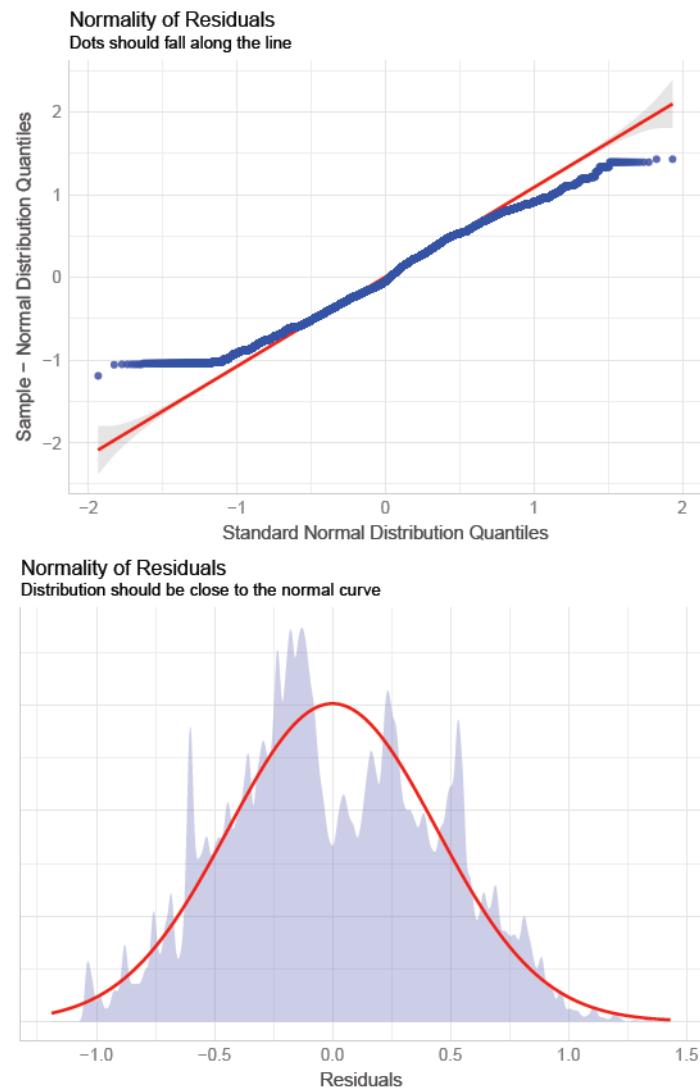


Figure S3.16: Diagnostic plots for Model 5a, obtained using the ‘performance’ R package (Lüdecke et al., 2021).

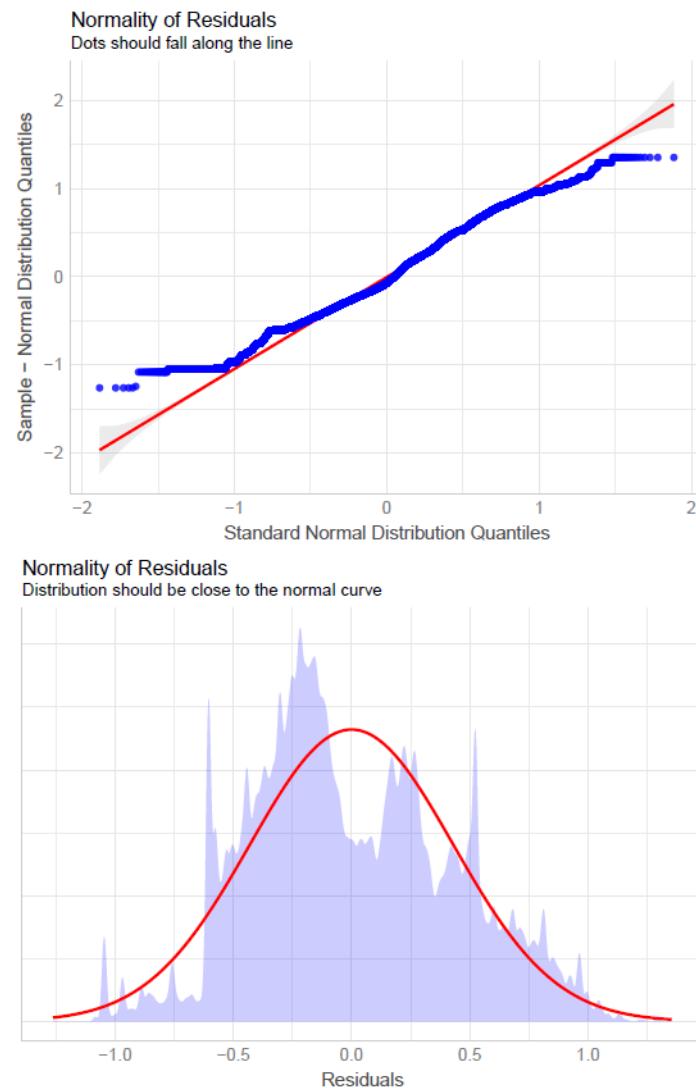


Figure S3.17: Diagnostic plots for Model 5b, obtained using the ‘performance’ R package (Lüdecke et al., 2021).

S3.7 Model robustness

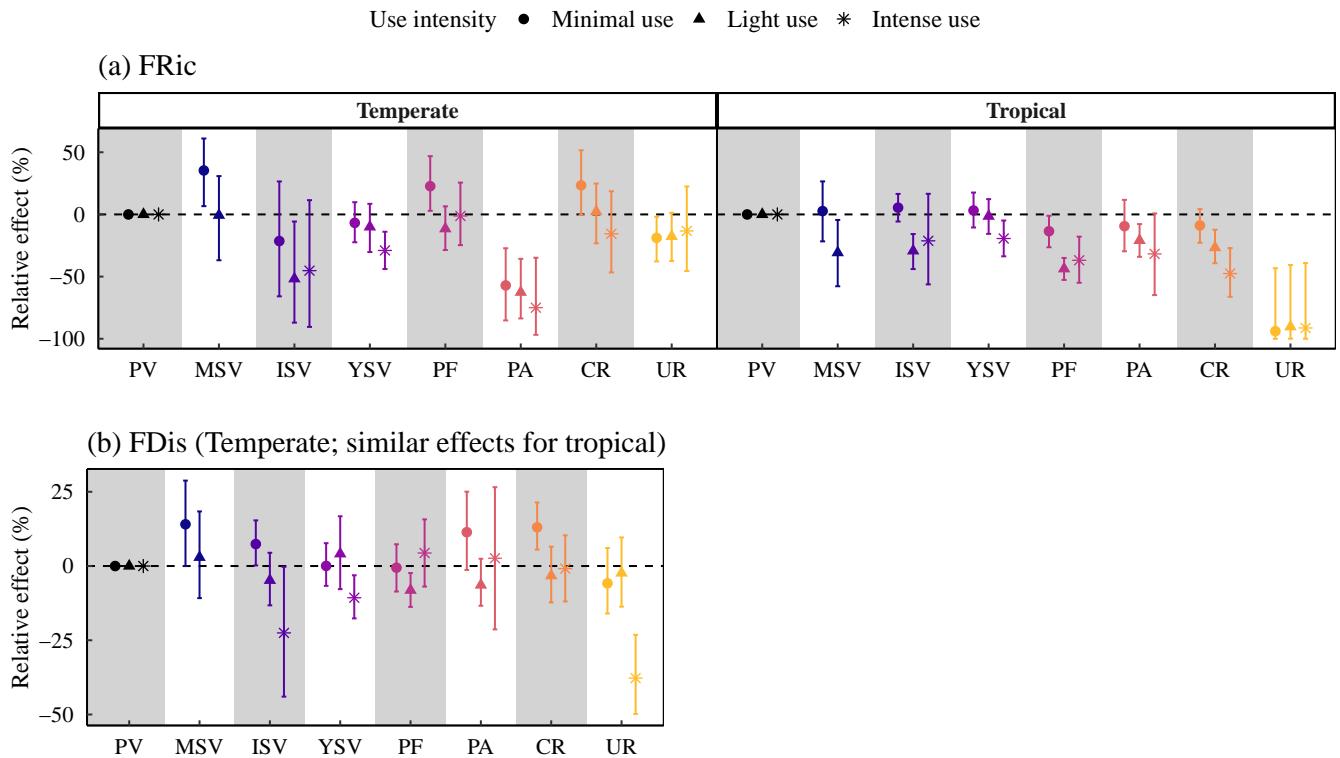


Figure S3.18: Effects of land use, land-use intensity and region on FRic (a) and FDis (b) across vertebrates, for the subset of species with complete trait data (i.e., excluding species with less than 100% trait completeness). Effects are plotted as a % difference relative to the reference land use (primary vegetation, PV). For FRic, I fitted **Model 1a**, which included the effects of land use, land-use intensity and region, as well as interactions between land use and land-use intensity and between land use and region. For FDis, I fitted **Model 1b**, which included effects of land use, land-use intensity and region, and interactions between land use and land-use intensity. Error bars represent 95% confidence intervals. MSV: mature secondary vegetation; ISV: intermediate secondary vegetation; YSV: young secondary vegetation; PF: plantation forest; PA: pasture; CR: cropland; UR: urban. Effects for Intense use in MSV could not be estimated as there weren't enough sampled sites.

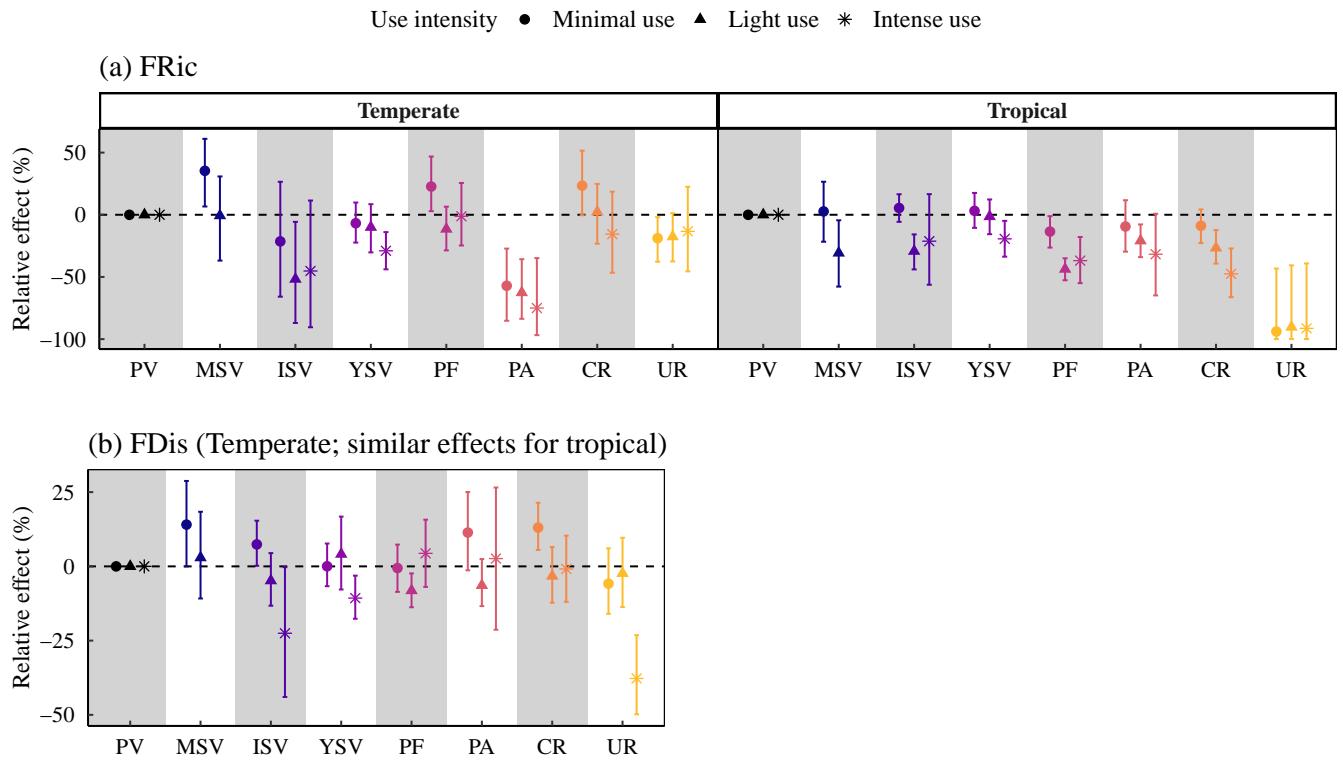


Figure S3.19: Effects of land use, land-use intensity and region on FRic (a) and FDis (b), for the subset of species with complete trait data (i.e., excluding species with less than 100% trait completeness), with geographical range size as an additional trait considered in the calculation of functional diversity metrics. Effects are plotted as a % difference relative to the reference land use (primary vegetation, PV). For FRic, I fitted **Model 1a**, which included the effects of land use, land-use intensity and region, as well as interactions between land use and land-use intensity and between land use and region. For FDis, I fitted **Model 1b**, which included effects of land use, land-use intensity and region, and interactions between land use and land-use intensity. Error bars represent 95% confidence intervals. MSV: mature secondary vegetation; ISV: intermediate secondary vegetation; YSV: young secondary vegetation; PF: plantation forest; PA: pasture; CR: cropland; UR: urban. Effects for Intense use in MSV could not be estimated as there weren't enough sampled sites.

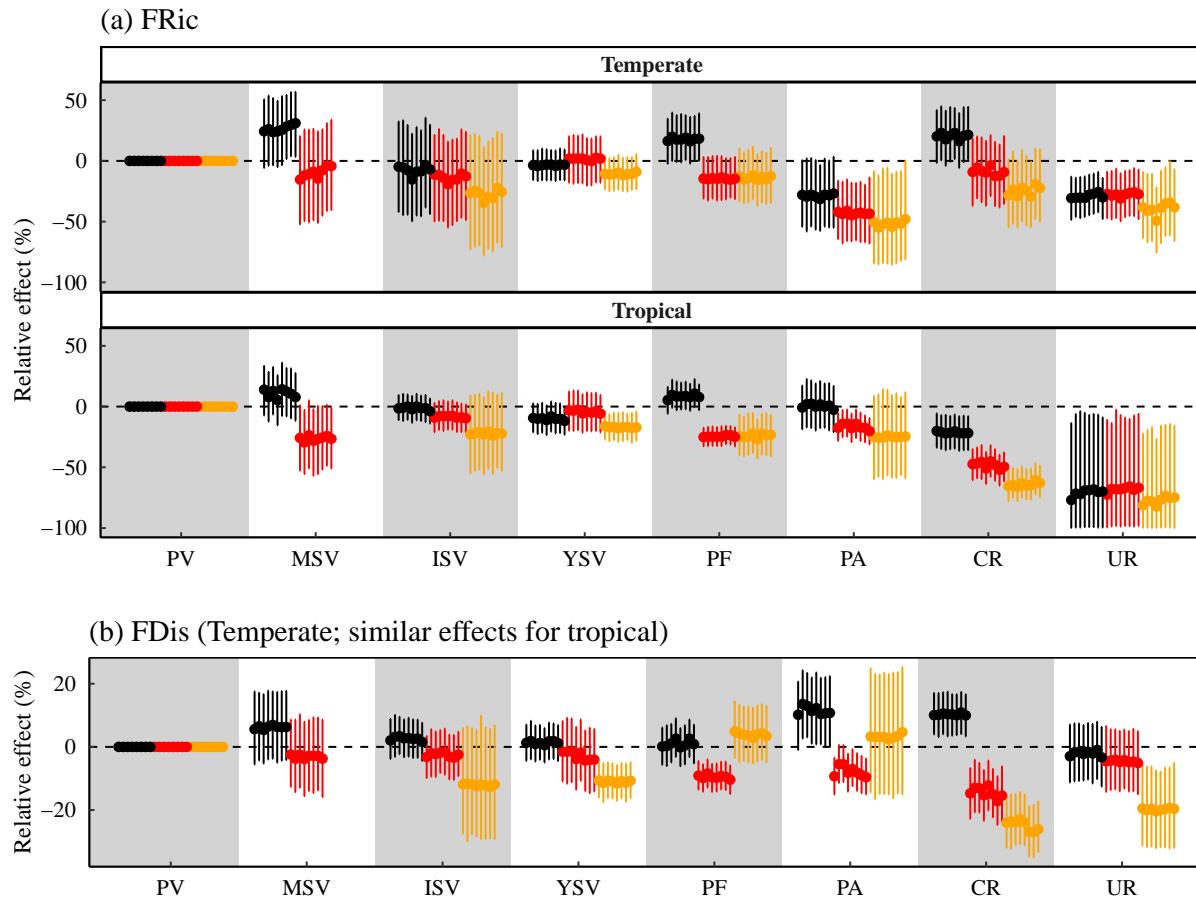


Figure S3.20: Effects of land use and land-use intensity on FRic (a) and FDis (b), obtained when calculating FRic and FDis with each set of imputed traits (eight in total). Effects are plotted as a % difference relative to the reference land use (primary vegetation, PV). For FRic, I fitted **Model 1a, which included the effects of land use, land-use intensity and region, as well as interactions between land use and land-use intensity and between land use and region. For FDis, I fitted **Model 1b**, which included effects of land use, land-use intensity and region, and interactions between land use and land-use intensity. Error bars represent 95% confidence intervals. MSV: mature secondary vegetation; ISV: intermediate secondary vegetation; YSV: young secondary vegetation; PF: plantation forest; PA: pasture; CR: cropland; UR: urban. Effects for Intense use in MSV could not be estimated as there weren't enough sampled sites.**

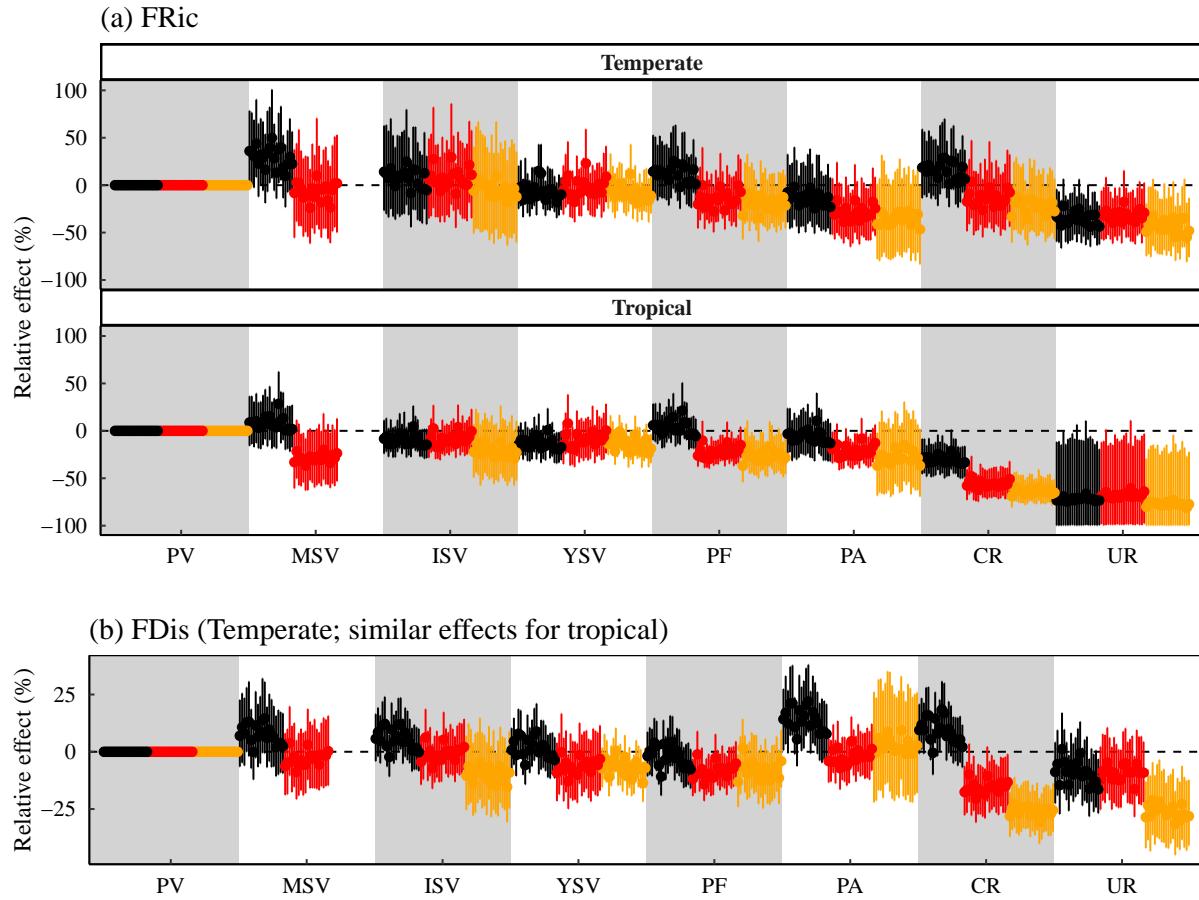


Figure S3.21: Effects of land use and land-use intensity on FRic (a) and FDis (b), obtained when re-sampling primary vegetation sites twenty independent times. I fixed the sample size for primary vegetation sites at 50. For FRic, we fitted **Model 1a**, which included the effects of land use, land-use intensity and region, as well as interactions between land use and land-use intensity and between land use and region. For FDis, I fitted **Model 1b**, which included effects of land use, land-use intensity and region, and interactions between land use and land-use intensity. Error bars represent 95% confidence intervals. MSV: mature secondary vegetation; ISV: intermediate secondary vegetation; YSV: young secondary vegetation; PF: plantation forest; PA: pasture; CR: cropland; UR: urban. Effects for Intense use in MSV could not be estimated as there weren't enough sampled sites.

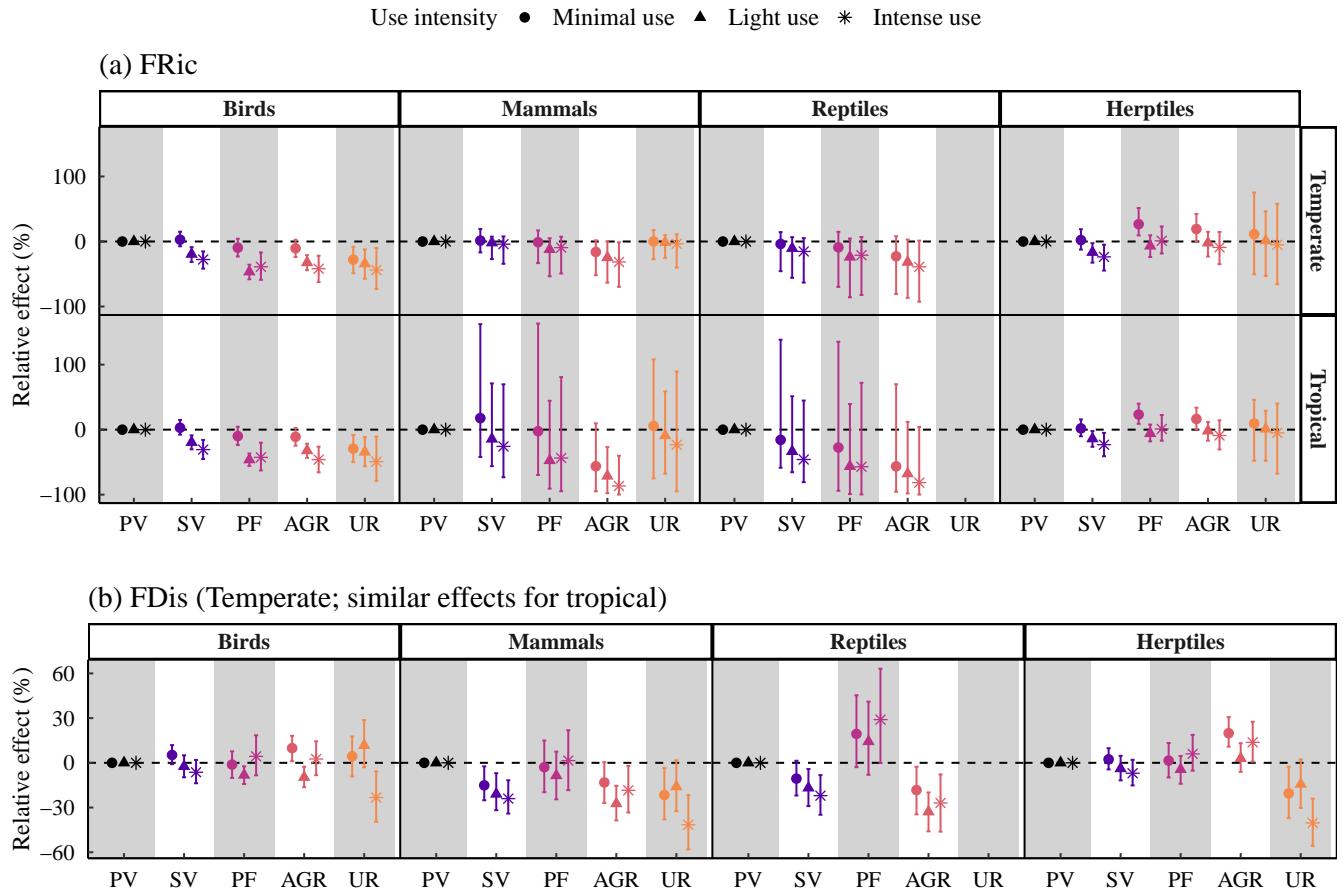
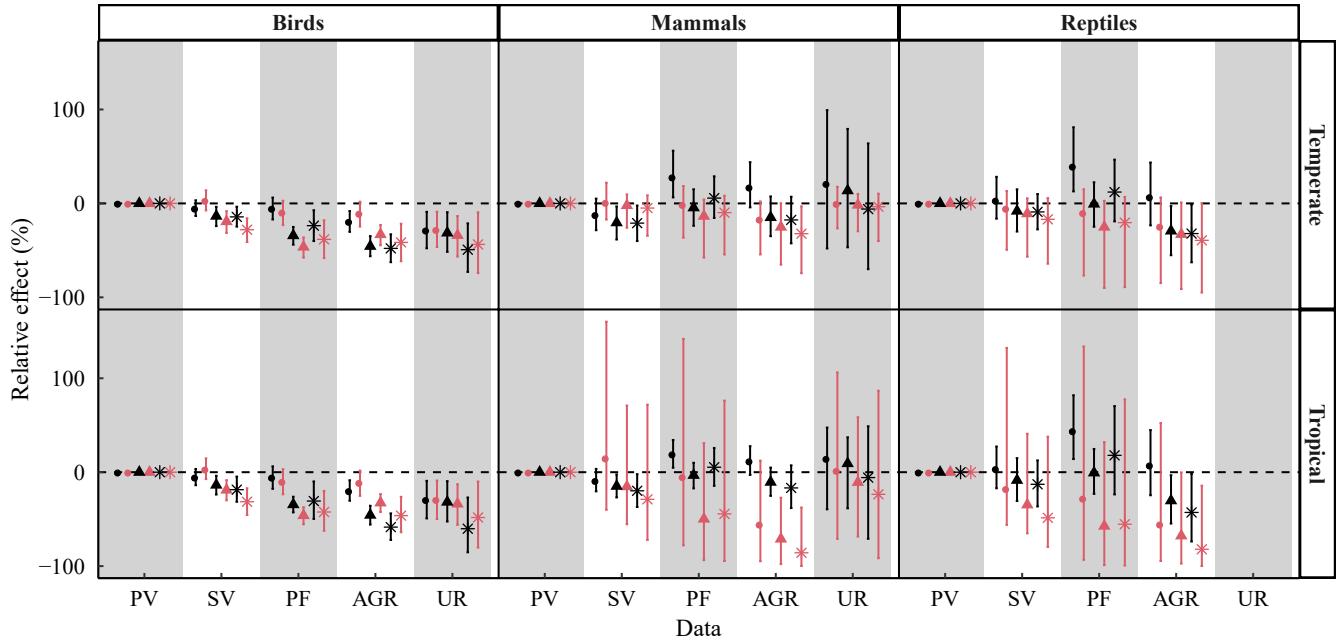


Figure S3.22: Effects of land use, land-use intensity and taxonomic class on FRic (a) and FDis (c), for the subset of species with complete trait data (i.e., excluding species with less than 100% trait completeness). Effects are plotted as a % difference relative to the reference land use (primary vegetation, PV). I did not include the effects of region here as sample sizes were not large enough for some classes. For FRic, the model included the effects of land use, land-use intensity and class, and interactions between land use and land-use intensity as well as land use and class. For FDis, the model included an additional interaction between land-use intensity and class. Error bars represent 95% confidence intervals. SV: secondary vegetation; PF: plantation forest; PA: pasture; CR: cropland; UR: urban. Effects for reptiles in urban land uses could not be estimated as there weren't enough sampled sites.

— complete and imputed trait data — complete trait data subset Use intensity • Minimal use ▲ Light use * Intense use

(a) FRic



(b) FDis

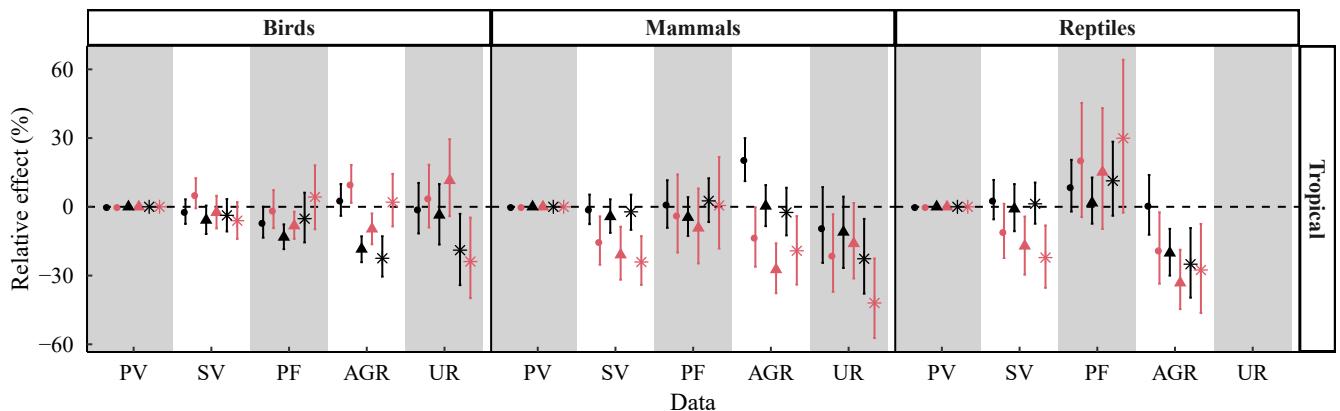


Figure S3.23: Effects of land use, region, land-use intensity and taxonomic class on FRic and FDis obtained with the imputed trait data (black points) or with the complete data subsets (red points). Effects are plotted as a % difference relative to the reference land use (primary vegetation, PV). For FRic, I fitted Model 2a, and I fitted Model 2b for FDis. Error bars represent 95% confidence intervals. SV: secondary vegetation; PF: plantation forest; AGR: agricultural (cropland and pasture); UR: urban. Effects for reptiles in urban land uses could not be estimated as there weren't enough sampled sites.

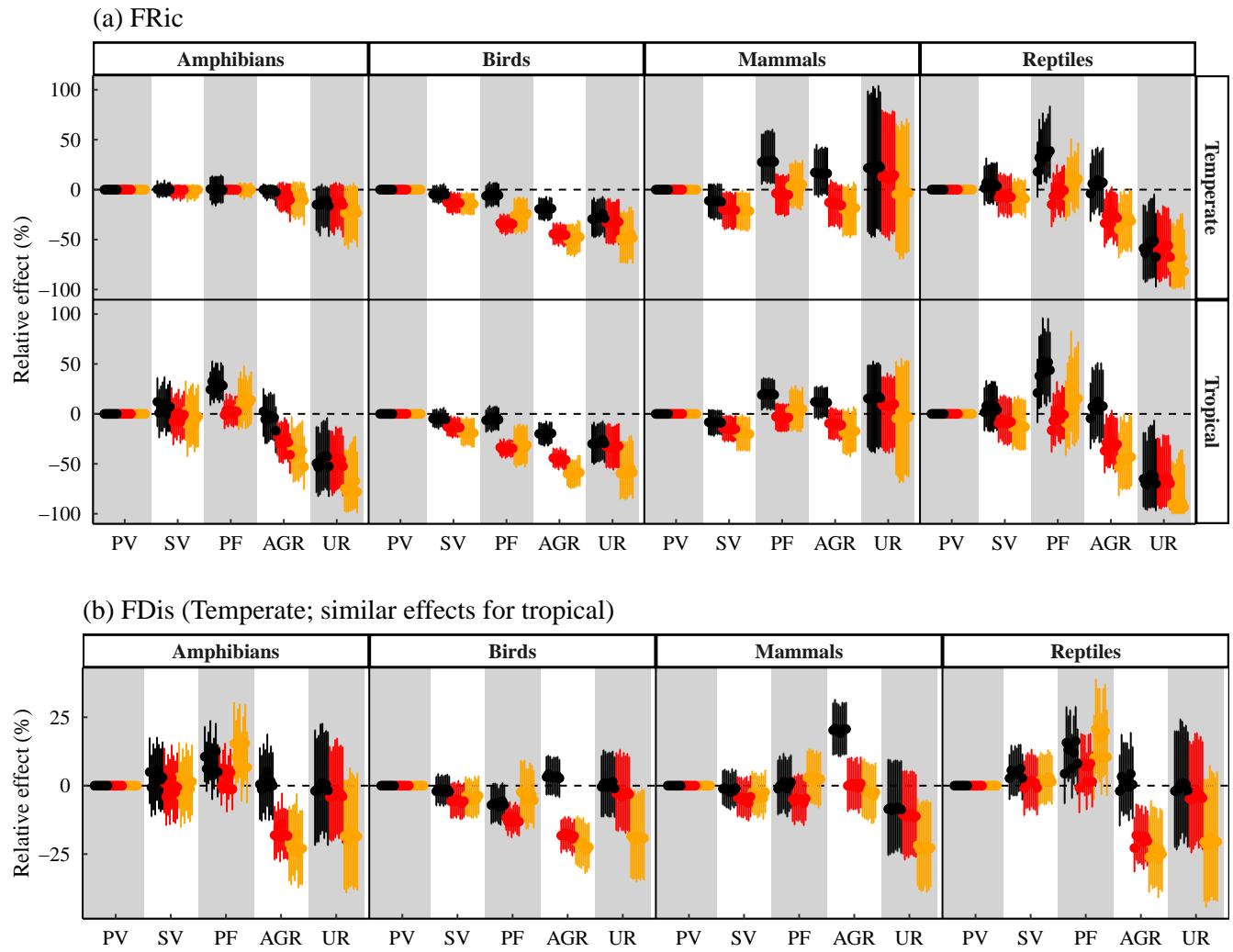


Figure S3.24: Effects of land use, region, land-use intensity and taxonomic class on FRic and FDis, obtained when calculating FRic and FDis with each set of imputed traits (eight in total). Effects are plotted as a % difference relative to the reference land use (primary vegetation, PV). For FRic, I fitted Model 2a, and Model 2b for FDis. Error bars represent 95% confidence intervals. SV: secondary vegetation; PF: plantation forest; AGR: agricultural (cropland and pasture); UR: urban. Effects for reptiles in urban land uses could not be estimated as there weren't enough sampled sites.

S3.8 Model robustness – time since land-use conversion

Time since land-use conversion could have important impacts on assemblage composition and thus, on local functional diversity. I did not investigate these effects because PREDICTS contained data on time since land-use conversion only for about 22% of the sites, considerably reducing samples sizes. Here, I investigated whether the results are likely robust to the inclusion of time since land-use conversion using the subset of sites for which time since land-use conversion was provided. To this end, I found the best-fitting models explaining FRic and FDis, using backwards stepwise selection, starting with complete models that included the effects of land use, time since land-use conversion, region, land-use intensity (for FRic only) and all two-way interactions among these predictors.

- For FRic, the best-fitting model included the main effects of land use and time since land-use conversion, but no interaction between these predictors. The model's summary (Table S3.6) showed that time since conversion had a significant negative effect on FRic, but the relationship between FRic and time since land-use conversion was similar in different land uses (as there were no interactions between land use and time since conversion retained in the best-fitting model, such that the slopes were similar in different land uses, and so the rate at which FRic decreased with time was similar in different land uses). The intercept was only different for urban land uses (significantly lower). Thus, based on this data subset, I expect time since land-use conversion to have a similar effect in different land-use types.

Table S3.6: Summary of the model explaining FRic by land use and time since land-use conversion, fitted on the subset of data for which there are information on time since land-use conversion.

	Estimate	Std. Error	t value
Intercept: Primary vegetation	1.156	0.073	15.921
Mature secondary vegetation	0.178	0.093	1.907
Intermediate secondary vegetation	0.018	0.072	0.249
Young secondary vegetation	-0.078	0.051	-1.532
Plantation forest	-0.018	0.082	-0.224
Pasture	-0.005	0.093	-0.054
Cropland	0.133	0.152	0.875
Urban	-0.316	0.133	-2.368
Time since conversion (log years)	-0.094	0.021	-4.566

I then compared this model's predictions with a simpler model that did not account for time since land-use conversion ($\text{FRic} \sim \text{Land use}$). The predictions (Figure S3.25) showed that including time since land-use

conversion did not bias the results, as I found a similar significant effect with both models in urban land uses, and elsewhere the effects were congruent. Thus, given this data subset, I argue that the results are robust to the inclusion of time since land-use conversion.

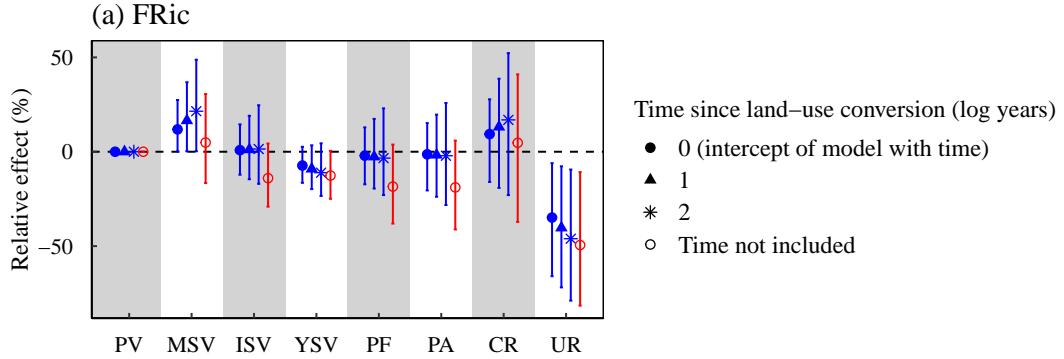


Figure S3.25: Effects of land use on FRic for the model that includes time since land-use conversion (blue points) versus the model that does not take time since land-use conversion into account (red points).

- For FDis, the best-fitting model included the main effects of land use, time since land-use conversion as well as interactions between land use and time since land-use conversion (I did not consider land-use intensity in the starting model because of sample size issues). Nevertheless, the main effect of time since land-use conversion was not significant (Table S3.7), and the relationship between time since land-use conversion and FDis was also not significant in most disturbed land uses (except for plantation forest). Thus, I argue the available data do not allow us to properly investigate the relationship between time since land-use conversion and FDis.

Table S3.7: Summary of the model explaining FDis by land use and time since land-use conversion, fitted on the subset of data for which I have information on time since land-use conversion.

	Estimate	Std. Error	t value
Intercept: Primary vegetation	0.366	0.011	32.219
Mature secondary vegetation	0.032	0.055	0.577
Intermediate secondary vegetation	-0.015	0.050	-0.298
Young secondary vegetation	0.020	0.015	1.386
Plantation forest	0.074	0.023	3.213
Pasture	-0.017	0.048	-0.346
Cropland	-0.013	0.042	-0.317
Urban	0.031	0.054	0.573
Time since conversion (log years)	-0.004	0.004	-1.186
Mature secondary vegetation:Time since conversion (log years)	-0.005	0.015	-0.335
Intermediate secondary vegetation:Time since conversion (log years)	0.011	0.016	0.650
Young secondary vegetation:Time since conversion (log years)	-0.008	0.007	-1.170
Plantation forest:Time since conversion (log years)	-0.023	0.007	-3.077
Pasture:Time since conversion (log years)	0.010	0.015	0.688
Cropland:Time since conversion (log years)	0.007	0.012	0.620
Urban:Time since conversion (log years)	-0.016	0.022	-0.714

Appendix 3: Supporting information for Chapter 4

S4.1 Compiling diet information

For mammals and birds, diet information was obtained from the EltonTraits database (Wilman et al., 2014). Before processing the data, the taxonomy was aligned to that of the trait datasets compiled in Chapter 2. Primary diet – that is, the diet inferred from the combination of food items that represent more than 50% of species consumption – was directly available for birds, but not for mammals. For both classes, diet was described as the percent use of different food items (namely: invertebrates, vertebrates – either ectotherms, endotherms, fish or unknown –, carrion, fruit, nectar, seed or other plant material). In order to have a consistent classification scheme across mammals and birds, I chose not to use the provided primary diet for birds, and instead I applied my own procedure to infer primary diet from recorded food items across birds and mammals. I first grouped the different vertebrate food items together with carrion to create a single ‘vertebrate’ food item category. I then used the percent uses of the food items to infer primary diet, classifying species’ primary diet into the following categories: vertebrate eaters; invertebrate eaters; plant/seed eaters; fruit/nectar eaters; and omnivores [these categories are similar to those employed for birds’ primary diet in EltonTraits]. When all food items had a percent use below (or equal to) 50% percent, species were classified as omnivores. For each species, I calculated diet breadth as the number of consumed food items (regardless of the percent use of those items; I kept vertebrate food items grouped together in the calculation of diet breadth, such that I did not count carrion as a separate food item).

For amphibians, diet information was partly extracted from the AmphiBIO database (Oliveira et al., 2017), and partly compiled from the literature (see next paragraph). In AmphiBIO, diet information was recorded as the consumption of six food items (leaves, flowers, seeds, fruit, arthropods and vertebrates), but the percent use of these items was not recorded (only whether they were consumed or not). From AmphiBIO, I classified amphibians into the different diet categories, depending on the combinations of consumed food

items (with species consuming both plant and animal matter classified as omnivores; species consuming both vertebrates and invertebrates also classified as omnivores; and species consuming plant or seed only, fruit or nectar only, vertebrates only or invertebrates only classified into the corresponding groups).

To increase diet data coverage for amphibians, I used data compiled by Rhiannon Osborne-Tonner, collected during her MSci project at UCL. Rhiannon Osborne-Tonner collected data from published papers and from the grey literature, targeting species occurring in the PREDICTS database (which I used for inferring land-use responses). Using these data, I was able to supplement my datasets with diet information for an additional 108 amphibians from 26 published sources (all found to be invertebrate eaters; see below for the list of sources). For reptiles, there was no readily available diet information (except for trophic level information, see Chapter 2). Thus, I used diet data compiled by Rhiannon Osborne-Tonner, collected from the literature, again specifically targeting reptiles occurring in the PREDICTS database. I was thus able to add diet information for 239 reptiles. Finally, I calculated diet breadth across amphibians and reptiles from the recorded food items. The diet data compiled by Rhiannon Osborne-Tonner are available at: https://figshare.com/articles/Reptile_Diet_csv/12024309 (DOI: 10.6084/m9.figshare.12024309.v1) and https://figshare.com/articles/Untitled_Item/12024312 (DOI: 10.6084/m9.figshare.12024312.v4).

Complementary data sources for amphibians

- Aguilar-López & Pineda (2013). An exotic species of earthworm preyed by *Craugastor rhodopis*. Herpetology Notes, 6, 335-336.
- Arroyo, S. B., Serrano-Cardozo, V. H., & Ramírez-Pinilla, M. P. (2008). Diet, microhabitat and time of activity in a Pristimantis (Anura, Strabomantidae) assemblage. Phylomedusa: Journal of Herpetology, 7(2), 109-119.
- Loc Barragán, J. A. & Woolrich, G. (2016). *Smilisca baudinii*. Diet. Mesoamerican Herpetology, 3(3).
- Berry, P. Y., & Bullock, J. A. (1962). The Food of the Common Malayan Toad, *Bufo melanostictus* Schneider. Copeia, 1962(4), 736–741.
- Blommers-Schlösser, R. M. A. (1975). A unique case of mating behaviour in a Malagasy tree frog, *Gephyromantis liber* (Peracca, 1893), with observations on the larval development (Amphibia, Ranidae). Beaufortia, 23(296), 15–25.

- Cappo, M. (1986). Frogs as predators of organisms of aquatic origin in the Magela Creek System, Northern Territory. MSc thesis. Dept of Zoology, University of Adelaide. <https://hdl.handle.net/2440/110846>.
- Costello, J.M. (2013). Differences in Morphology and Behavior in Green Frogs (*Lithobates clamitans*) from Urban and Rural Sites in New York and New Jersey. PhD thesis. City University of New York.
- Brito de Carvalho, C. & Freitas, E. & Faria, R. & Batista, R. & Batista, C. & Coelho, W. & Bocchiglieri, A. (2008). Natural history of *Leptodactylus mystacinus* and *Leptodactylus fuscus* (Anura: Leptodactylidae) in the Cerrado of Central Brazil. *Biota Neotropica*, 8.
- Batista, R. & Brito de Carvalho, C. & Freitas, E.B. & Franco, S.C. & Batista, C.C. & Coelho, Wellington & Faria, Renato. (2011). Diet of *Rhinella schneideri* (Werner, 1894) (Anura: Bufonidae) in the Cerrado, Central Brazil. *Herpetology Notes*, 4:17-21.
- Fonseca-Pérez, K. A., Molina, C., & Tárano, Z. (2017). Diet of *Dendropsophus microcephalus* and *Scarthyla vigilans* (Anura: Hylidae) at a locality in north-western Venezuela with notes on microhabitat occupation. *Papéis Avulsos De Zoologia*, 57(7), 93-104.
- García-R, J.C. & Posso-Gómez, C. E. & Cárdenas-Henao, H. (2015). Diet of direct-developing frogs (Anura: Craugastoridae: Pristimantis) from the Andes of western Colombia. *Acta Biológica Colombiana*, 20:79-87.
- Zumbado-Ulato, H. & Acosta-Chaves, V. (2012). *Cochranella granulosa* (Granular glass frog) feeding behaviour. Herpetological review.
- Yap, C.H. & Jafaar, I. (2012). Feeding analysis of *Hylarana cf. labialis*, *Leptobrachium hendricksoni*, and *Occidozyga laevis* (Amphibia: Anura) from a lowland dipterocarp forest in Kedah, Malaysia. *Herpetological Review*, 43(1).
- Schwenk, K. (2000). Feeding: Form, function and evolution in tetrapod vertebrates. Academic Press.
- Sabagh, L.T., Ferreira V.L., & Rocha C.F. (2010). Living together, sometimes feeding in a similar way: the case of the syntopic hylid frogs *Hypsiboas raniceps* and *Scinax acuminatus* (Anura: Hylidae) in the Pantanal of Miranda, Mato Grosso do Sul State, Brazil. *Brazilian Journal of Biology*, 70(4):955-9.
- Luría-Manzano R. & Ramírez-Bautista A. (2017). Diet comparison between rainforest and cave populations of *Craugastor alfredi* (Anura: Craugastoridae): does diet vary in contrasting habitats? *Journal of Natural History*, 51:39-40, 2345-2354.

- Martínez-Coronel, M. & Pérez-Gutiérrez, M. (2011). Diet composition of *Craugastor lineatus* (Anura: Craugastoridae) of Chiapas, Mexico. *Acta Zoológica Mexicana* 27(2):215-230.
- McAlister, W. H. (1963). Evidence of mild toxicity in the saliva of the hognose snake (Heterodon). *Herpetologica*, 19:132-137.
- Moreno-Barbosa, S. & Hoyos, J. (2014). Ontogeny of the diet in Anurans (Amphibia) collected at La Vieja river basin in the departamento of Quindío (Colombia). *Caldasia*, 36 (365-372).
- Piatti, L. & Souza, F. (2011). Diet and resource partitioning among anurans in irrigated rice fields in Pantanal, Brazil. *Brazilian journal of biology*, 71 (653-61).
- Savini, C. O. K., Chuang, M. F., & Ishida, C. (2004). Diet Selection in the Green Paddy Frog (*Rana erythraea*). International field biology course 2004.
- Simon, M. P., & Toft, C. A. (1991). Diet Specialization in Small Vertebrates: Mite-Eating in Frogs. *Oikos*, 61(2):263–278.
- Sugai, J.L.M.M., Terra, J.S., & Ferreira, V.L. (2012): Diet of *Leptodactylus fuscus* (Amphibia: Anura: Leptodactylidae) in the Pantanal of Miranda river, Brazil. *Biota Neotropica*, 12: 99-104.
- Toft, C. A. (1981). Feeding Ecology of Panamanian Litter Anurans: Patterns in Diet and Foraging Mode. *Journal of Herpetology*, 15(2):139–144.
- Vences, M., Glaw, F. & Zapp, C.(1999). Stomach content analysis in Malagasy frogs of the genera *Tomopterna*, *Aglyptodactylus*, *Boophis* and *Mantidactylus*. *Herpetozoa*, 11(3/4):109-116.
- Yap, C.H. & Jaafar, I. (2011). Stomach Content Analysis of Tropical Forest Toads *Ingerophrynus parvus* and *Phrynoïdes aspera* (ANURA : BUFONIDAE) from Kedah, Malaysia. Taxonomist and Ecologist Conference 2011.

Complementary data sources for reptiles

The 148 sources are listed in the dataset available from https://figshare.com/articles/Reptile_Diet_csv/12024309 (DOI: 10.6084/m9.figshare.12024309.v1).

S4.2 Imputing missing trait values

S4.2.1 Trait data coverage

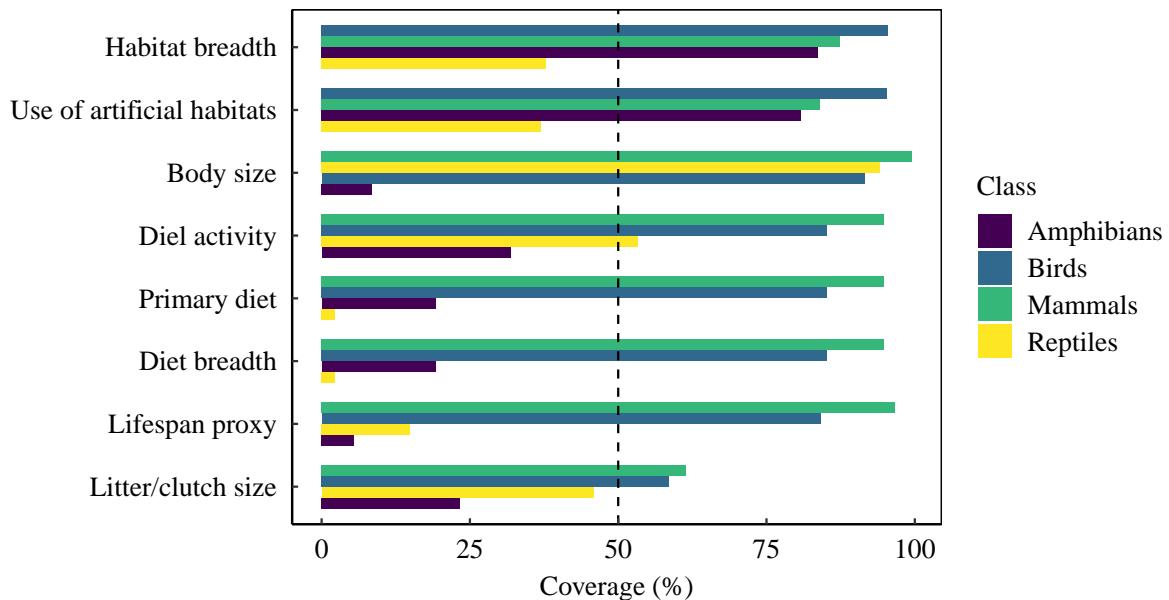


Figure S4.1: Trait coverage, including coverage for diet information, calculated as the proportion of species for which trait values are not missing. The dashed line represents 50% coverage.

S4.2.2 Phylogenetic signal in traits

I measured the phylogenetic signal in traits using Pagel's λ (for continuous traits) and Borges' δ (for categorical traits). I found evidence of phylogenetic conservatism in all traits.

Table S4.1: Phylogenetic signal in continuous and categorical traits. BM: body mass; BL: body length; GL: generation length; MA: age at sexual maturity; ML: maximum longevity; L: longevity; LCS: litter/clutch size; HB: habitat breadth; DA: diel activity; UA: use of artificial habitats. Continuous traits were log-10 transformed to improve normality before estimating Pagel's λ – except for habitat breadth and diet breadth which were square-rooted. A star indicates a significant signal ($P<0.05$ for the log-likelihood ratio test in the case of λ ; and a significant difference from the simulated null distribution of δ for categorical traits). 'NA' indicates traits that were not considered for a given class.

Class	Pagel's λ										Borges' δ		
	BM	BL	GL	MA	ML	L	LCS	HB	Diet breadth	Diet	DA	UA	
Amphibians	0.98*	0.94*	NA	0.85*	0.82*	NA	0.93*	0.99*	0.61*	3.4*	3.4*	4.5*	
Birds	0.99*	NA	0.97*	NA	NA	NA	0.95*	0.60*	0.72*	6.4*	32e3*	1.8*	
Mammals	0.99*	NA	0.97	NA	NA	NA	0.99	0.71	0.99*	26*	52*	1.3*	
Reptiles	1.0*	NA	NA	NA	0.94*	0.98*	1.0*	0.52*	0.84*	2.2*	6.4*	1.4*	

S4.2.3 Implementation of missing value imputations

I imputed missing trait values using random forest algorithms, as implemented in R with ‘missForest’ (Stekhoven, 2016; Stekhoven and Bühlmann, 2012). Phylogenetic relationships were included as additional predictors in the form of phylogenetic eigenvectors (Diniz-Filho et al., 2012), extracted from the phylogenies using the PVR package (Santos, 2018). I used class-specific phylogenetic trees from <https://zenodo.org/record/3690867#.Xyc5wyhKhPZ> for mammals (Phylacine 1.2; Faurby et al. (2018, 2020)); and from <https://data.vertlife.org/> for amphibians (Jetz & Pyron 2018), birds (Jetz et al. 2012) and squamates (Tonini et al. 2016). For each class, I used a consensus tree obtained with the TreeAnnotator programme of the BEAST software (Bouckaert et al. 2014), from an available distribution of 1000 trees.

Following Penone et al. (2014), I included the first ten phylogenetic eigenvectors as additional predictors of missing trait values in each class. As not all species were represented in the phylogenies, I also added taxonomic order as a predictor for all species. All traits in Table S4.1 were included in the imputations. Habitat & diet breadth were considered as categorical variables for the imputations (and so, discretised), in order to ensure that only integer estimates were obtained from the imputations for these traits. Tuning parameters of ‘missforest’ were set to ten maximum iterations and to one hundred trees grown in each forest.

S4.2.4 Imputation error

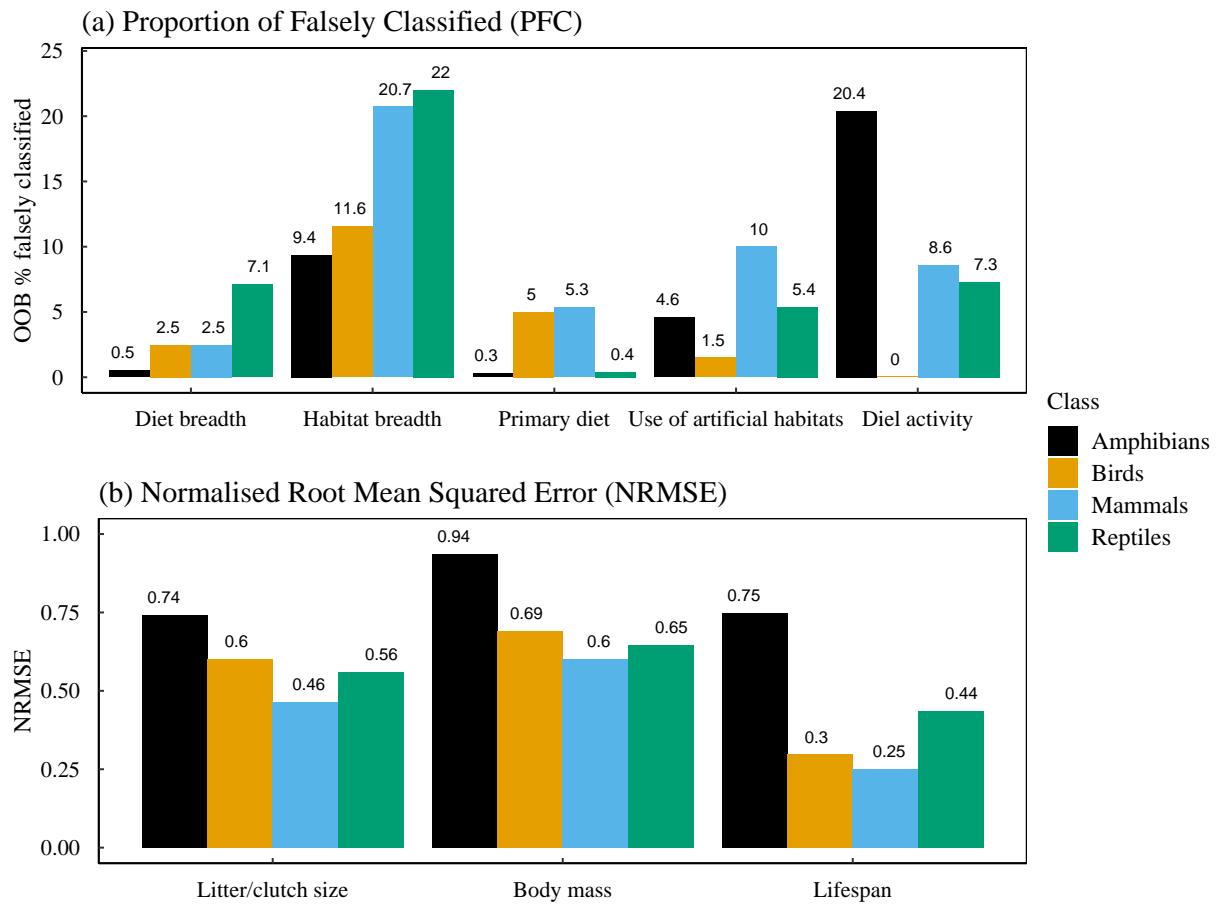


Figure S4.2: Out-of-bag estimation of imputation errors for the traits included in the analyses. (a) For the categorical traits, I show the proportion of falsely classified traits ('PFC', out-of-bag estimates); (b) For the continuous traits, I calculate the normalised root-mean-squared error (NRMSE), from the out-of-bag mean square error that I divide by the standard deviation of the known trait distribution. The lower the NRMSE, the lower the imputation error, with values close to 0 indicating lower imputation error and values close to 1 tending to indicate higher imputation error.

S4.3 Land-use types in PREDICTS and sample sizes (number of sampled sites across classes)

Table S4.2: Land-use categories in the PREDICTS database. See Hudson et al. (2014, 2017) for more details.

Land-use category	Definition
Primary vegetation	Native vegetation, undisturbed since its establishment under current climatic conditions. No known alterations due to human activities or to extreme natural events.
Mature secondary vegetation	Vegetation recovering after complete destruction of primary vegetation & where succession is near complete – the structure approaches that of primary vegetation.
Intermediate secondary vegetation	Vegetation recovering after complete destruction of primary vegetation at a mid-successional stage.
Young secondary vegetation	Vegetation recovering after complete destruction of primary vegetation at an early successional stage.
Plantation forest	Previously cleared areas planted with crop trees or shrubs grown and harvested for human consumption or for commercial purposes (includes wood, fruit, oil, biofuel, rubber, etc).
Pasture	Areas grazed by livestock, permanently or regularly. Can be improved through cultivation techniques.
Cropland	Previously cleared areas planted with herbaceous crops and harvested for human or animal consumption (including animal feed and crops used in the food industry), or for commercial purposes (e.g., crops grown for the textile industry).
Urban	Previously cleared areas built up by humans. Vegetation is managed for civic or personal purposes.

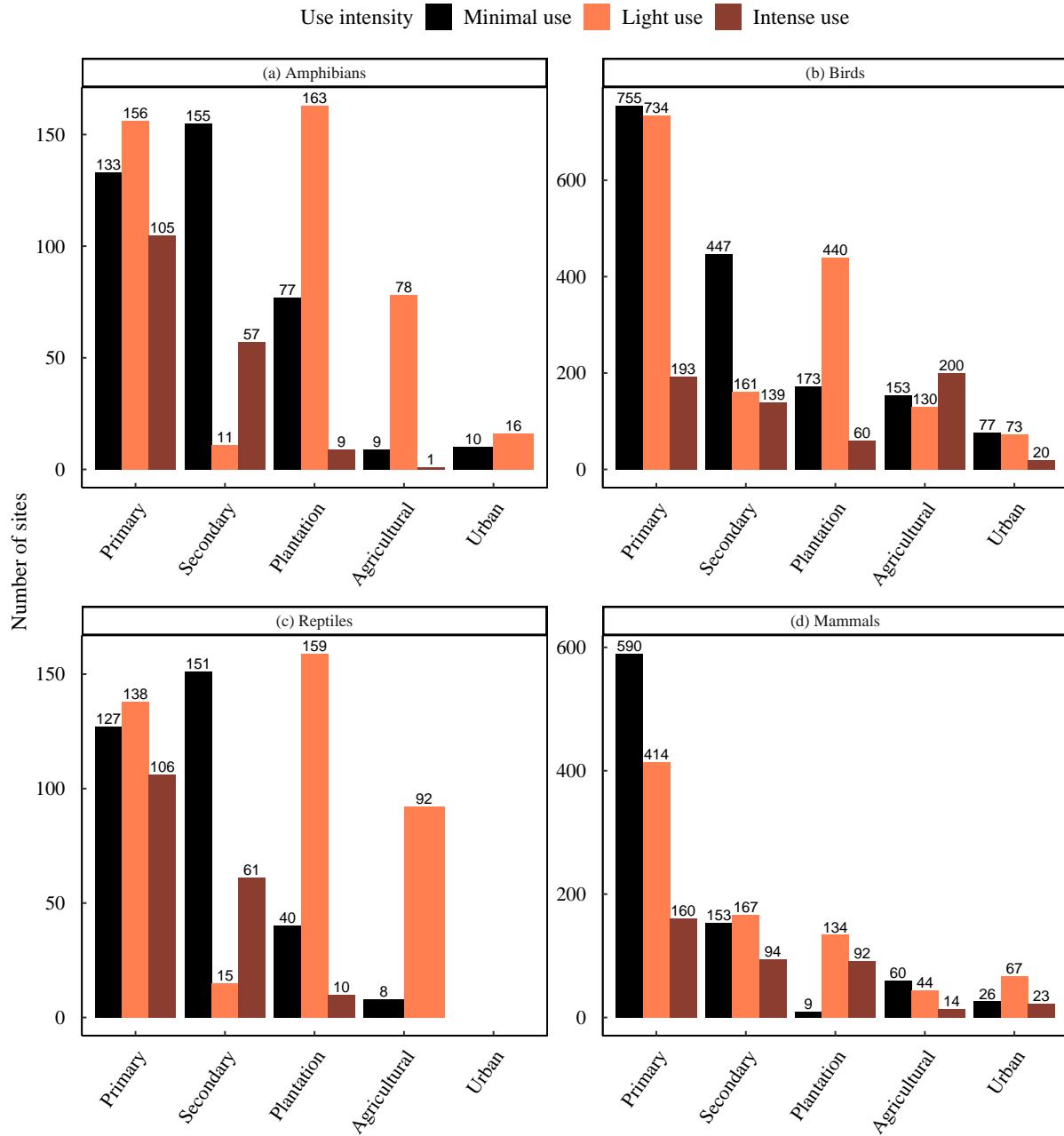


Figure S4.3: Sample sizes (number of PREDICTS sites) for the different land-use types, in each class.

S4.4 Land-use responses: multicollinearity checks among the models' explanatory variables

Table S4.3: Generalised Variance Inflation Factors among the candidate explanatory variables for the mixed-effects model fitted across amphibians, prior to the exclusion of diet. The model aimed at investigating the effects of land use, land-use intensity and the species-level ecological characteristics on species occurrence probability.

Predictor	GVIF
Diel activity	1.7
Lifespan proxy (\log_{10})	1.8
Specialisation	1.8
Range area (\log_{10})	2.0
Body mass (\log_{10})	2.0
Land use	2.0
Litter/clutch size (\log_{10})	2.5
Land-use intensity	2.6
Habitat breadth (square-root)	3.2
Diet breadth (square-root)	22.8
Primary diet	23.6

Table S4.4: Generalised Variance Inflation Factors among the explanatory variables for the mixed-effects model fitted across amphibians (after the exclusion of diet), looking at the effects of land use, land-use intensity and the species-level ecological characteristics on species occurrence probability.

Predictor	GVIF
Diel activity	1.6
Diet breadth (square-root)	1.7
Lifespan proxy (\log_{10})	1.8
Specialisation	1.8
Range area (\log_{10})	1.9
Land use	2.0
Body mass (\log_{10})	2.0
Litter/clutch size (\log_{10})	2.4
Land-use intensity	2.6
Habitat breadth (square-root)	3.1

Table S4.5: Generalised Variance Inflation Factors among the explanatory variables for the mixed-effects model fitted across birds, looking at the effects of land use, land-use intensity and the species-level ecological characteristics on species occurrence probability.

Predictor	GVIF
Diel activity	1.1
Land use	1.2
Land-use intensity	1.2
Litter/clutch size (\log_{10})	1.3
Range area (\log_{10})	1.4
Diet breadth (square-root)	1.5
Specialisation	1.6
Lifespan proxy (\log_{10})	1.7
Habitat breadth (square-root)	1.8
Body mass (\log_{10})	1.9
Primary diet	2.3

Table S4.6: Generalised Variance Inflation Factors among the explanatory variables for the mixed-effects model fitted across mammals, looking at the effects of land use, land-use intensity and the species-level ecological characteristics on species occurrence probability.

Predictor	GVIF
Diel activity	1.2
Range area (\log_{10})	1.2
Specialisation	1.4
Land-use intensity	1.4
Diet breadth (square-root)	1.7
Land use	1.8
Habitat breadth (square-root)	1.8
Litter/clutch size (\log_{10})	2.7
Body mass (\log_{10})	3.0
Lifespan proxy (\log_{10})	3.4
Primary diet	4.4

Table S4.7: Generalised Variance Inflation Factors among the candidate explanatory variables for the mixed-effects model fitted across reptiles, prior to the exclusion of diet. The model aimed at investigating the effects of land use, land-use intensity and the species-level ecological characteristics on species occurrence probability.

Predictor	GVIF
Diel activity	1.1
Specialisation	1.3
Range area (\log_{10})	1.3
Habitat breadth (square-root)	1.6
Lifespan proxy (\log_{10})	1.9
Litter/clutch size (\log_{10})	2.8
Land use	3.2
Land-use intensity	3.5
Body mass (\log_{10})	3.9
Diet breadth (square-root)	5.8
Primary diet	9.9

Table S4.8: Generalised Variance Inflation Factors among the explanatory variables for the mixed-effects model fitted across reptiles (*after the exclusion of diet*), looking at the effects of land use, land-use intensity and the species-level ecological characteristics on species occurrence probability.

Predictor	GVIF
Diel activity	1.1
Diet breadth (square-root)	1.2
Specialisation	1.2
Range area (\log_{10})	1.3
Habitat breadth (square-root)	1.6
Lifespan proxy (\log_{10})	1.9
Litter/clutch size (\log_{10})	2.7
Land use	3.2
Body mass (\log_{10})	3.2
Land-use intensity	3.4

S4.5 Implementing Climate-niche Factor Analysis across terrestrial vertebrates

S4.5.1 Historical climate data: groups of intercorrelated variables

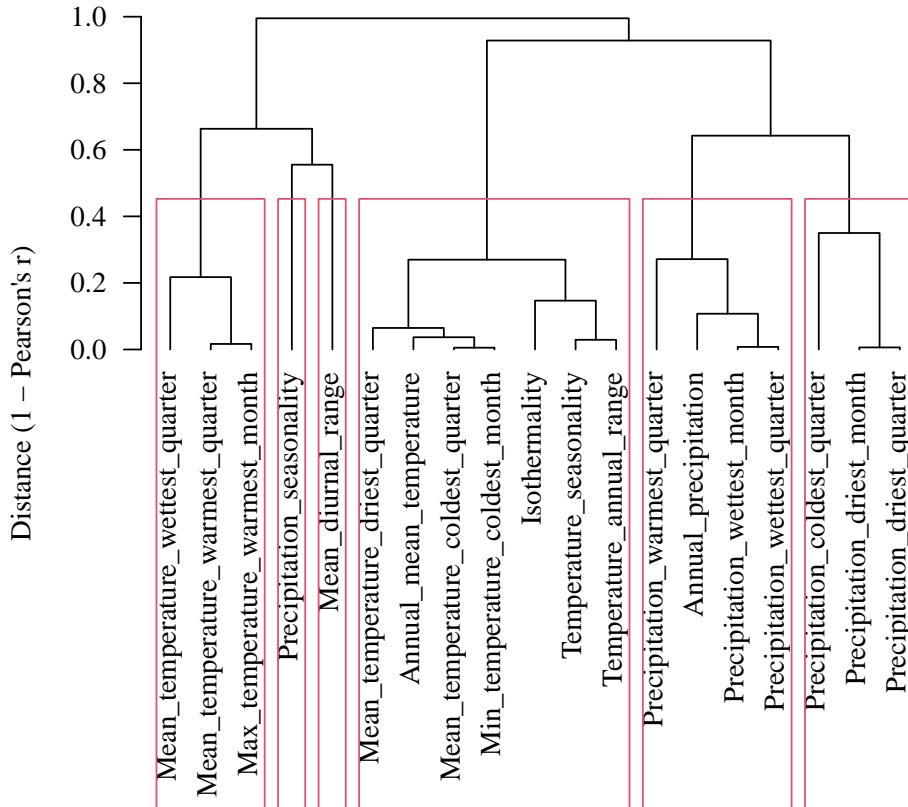


Figure S4.4: Groups of intercorrelated climatic variables using a cutoff of 0.65 for Pearson's correlation coefficient, obtained using the ‘removeCollinearity’ R function (‘virtualspecies’ package, Leroy et al. (2015)). Variables figuring in the same red boxes were correlated with Pearson's correlation coefficient > 0.65.

S4.5.2 CENFA estimation and resolution

I estimated climate-change sensitivity across terrestrial vertebrates with the CENFA framework using three different resolutions for the species distribution files and the climatic variables: 50 km², 10 km² and 5 km². The finer the resolution, the better the species' distribution is likely to be captured, particularly for narrow-ranging species (Figure S4.5). When working with coarser resolutions, the actual geographical distribution of a narrow-ranging species might be overestimated (Figure S4.5), such that the climatic niche breadth of the species might also be overestimated; consequently, its climate-change sensitivity may be underestimated. However, finer resolutions are more computationally demanding, which can be limiting when working across

several thousand species.

Thus, I looked for a resolution that provided the best compromise between accuracy of sensitivity estimation and computational load (Figure S4.6). Hence, I chose to work with a resolution of 5 km^2 . At this resolution, there were still some narrow-ranging species for which sensitivity was likely overestimated (Figure S4.6). To prevent any impact of these species on the analyses, I removed species with the smallest geographical range areas, using a conservative threshold of 100 km^2 for geographical range area (Figure S4.6).

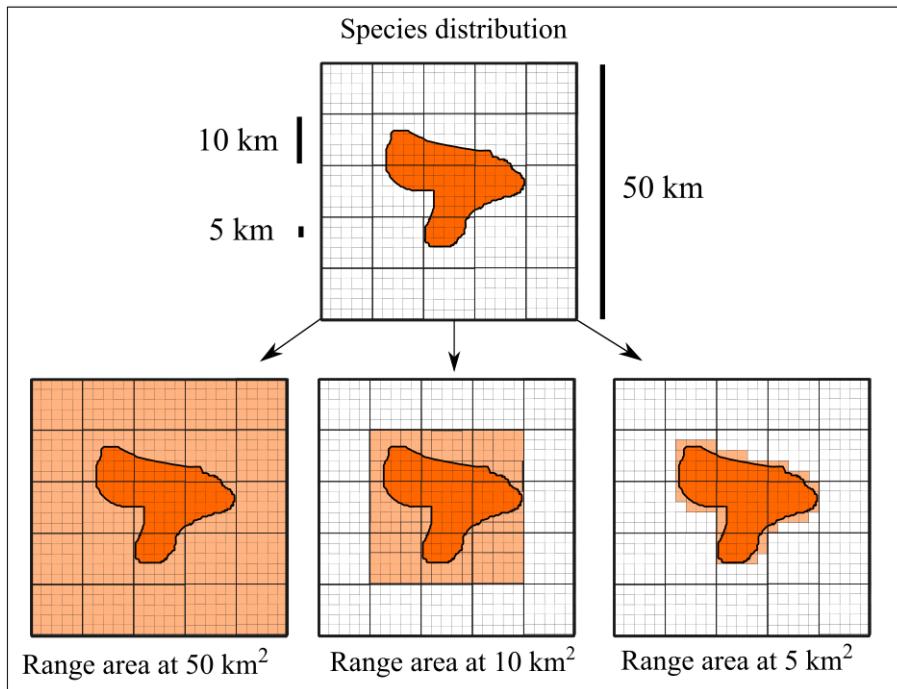


Figure S4.5: Possible impact of resolution on estimated geographical range area. I represent a virtual distribution for a species (orange shape). The species distribution is more accurately captured at finer resolutions (e.g., 5 km^2) than at coarser resolutions (e.g., 10 km^2 or 50 km^2). A possible consequence is that coarser resolutions can tend to disproportionately overestimate the geographical range area of narrow-ranging species, because the aggregation of grid cells where the species is found to be present can artificially augment the amount of occupied area at coarser resolutions, and relatively more so if the species is narrow-ranging.

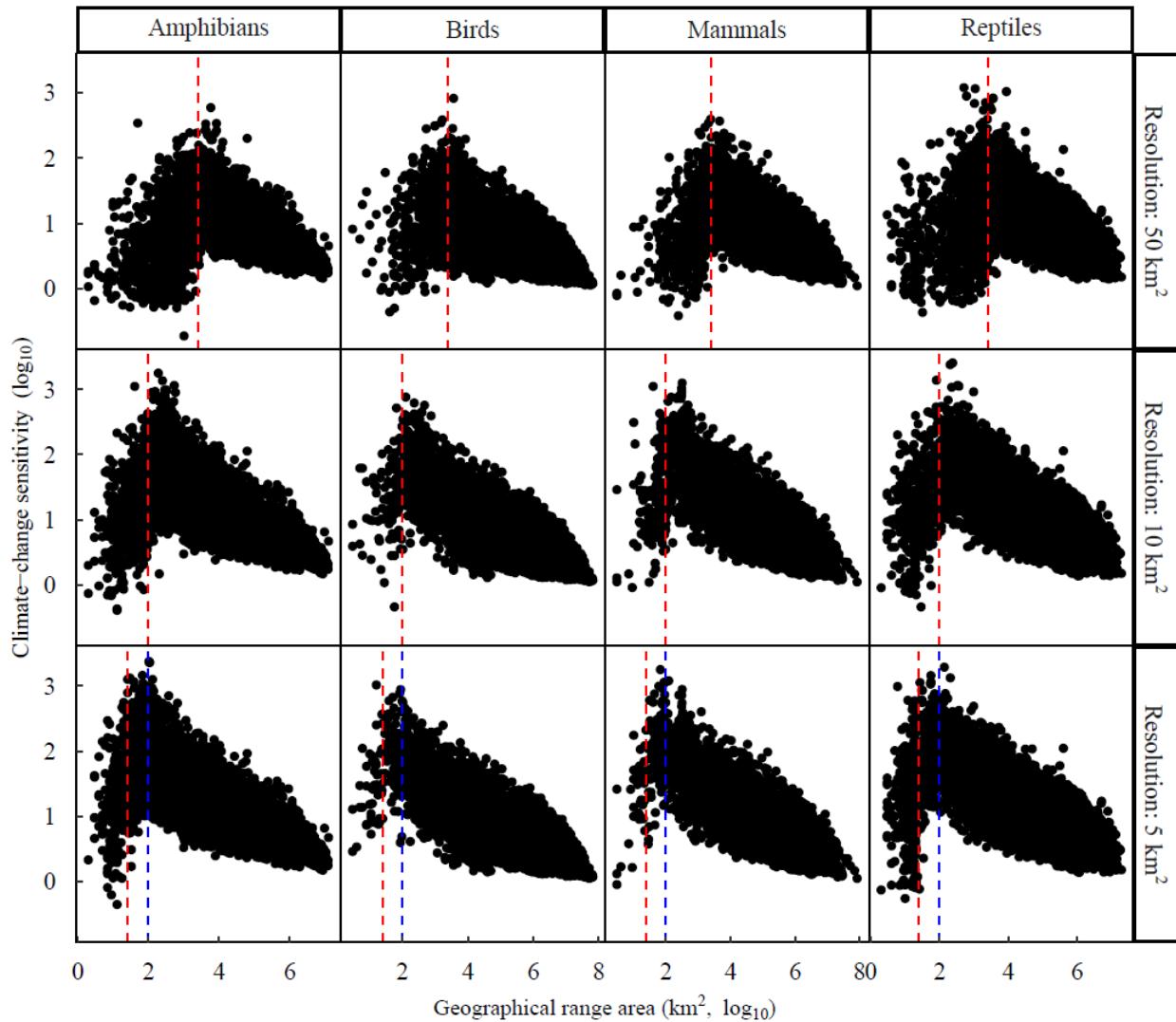


Figure S4.6: Estimated climate-change sensitivity estimations at three different resolutions (50 km^2 , 10 km^2 and 5 km^2) against geographical range area (estimated at 1 km^2). With the red dashed lines, I highlight the range areas that correspond to the surface area of one grid cell (i.e., $2,500 \text{ km}^2$, 100 km^2 and 25 km^2 respectively). Climate-change sensitivity was estimated using the CENFA framework (Rinnan and Lawler, 2019). I chose to work at a resolution of 5 km^2 and I excluded species whose range area was $\leq 100 \text{ km}^2$ (blue dashed line), that is, species whose distribution could intersect up to four grid cells at a resolution of 5 km^2 .

S4.6 Climate-change sensitivity models: multicollinearity checks among models' explanatory variables

Table S4.9: Generalised Variance Inflation Factors among the candidate explanatory variables for the phylogenetic least-square regression fitted across amphibians, prior to the exclusion of diet breadth. The model aimed at investigating the association between the species-level ecological characteristics and species climate-change sensitivity.

Predictor	GVIF
Diel activity	1.1
Lifespan proxy (\log_{10})	1.2
Range area (\log_{10})	1.3
Body mass (\log_{10})	1.4
Litter/clutch size (\log_{10})	1.5
Specialisation	1.6
Habitat breadth (square-root)	1.9
Primary diet	17.1
Diet breadth (square-root)	17.1

Table S4.10: Generalised Variance Inflation Factors among the explanatory variables for the phylogenetic least-square regression fitted across amphibians, after excluding diet breadth. The model aimed at investigating the association between the species-level ecological characteristics and species climate-change sensitivity.

Predictor	GVIF
Diel activity	1.1
Primary diet	1.1
Lifespan proxy (\log_{10})	1.2
Range area (\log_{10})	1.3
Body mass (\log_{10})	1.4
Litter/clutch size (\log_{10})	1.5
Specialisation	1.6
Habitat breadth (square-root)	1.9

Table S4.11: Generalised Variance Inflation Factors among the explanatory variables for the phylogenetic least-square regression fitted across birds. The model aimed at investigating the association between the species-level ecological characteristics and species climate-change sensitivity.

Predictor	GVIF
Diel activity	1.1
Range area (\log_{10})	1.2
Litter/clutch size (\log_{10})	1.3
Diet breadth (square-root)	1.5
Specialisation	1.6
Habitat breadth (square-root)	1.8
Lifespan proxy (\log_{10})	1.9
Body mass (\log_{10})	2.0
Primary diet	2.1

Table S4.12: Generalised Variance Inflation Factors among the explanatory variables for the phylogenetic least-square regression fitted across mammals. The model aimed at investigating the association between the species-level ecological characteristics and species climate-change sensitivity.

Predictor	GVIF
Range area (\log_{10})	1.2
Diel activity	1.3
Specialisation	1.3
Habitat breadth (square-root)	1.5
Diet breadth (square-root)	1.6
Body mass(\log_{10})	2.3
Litter/clutch size (\log_{10})	2.4
Primary diet	2.7
Lifespan proxy (\log_{10})	3.0

Table S4.13: Generalised Variance Inflation Factors among the explanatory variables for the phylogenetic least-square regression fitted across reptiles. The model aimed at investigating the association between the species-level ecological characteristics and species climate-change sensitivity.

Predictor	GVIF
Diel activity	1.1
Range area(\log_{10})	1.2
Specialisation	1.4
Habitat breadth (square-root)	1.5
Lifespan proxy (\log_{10})	1.6
Litter/clutch size (\log_{10})	2.0
Body mass (\log_{10})	2.9
Diet breadth (square-root)	2.9
Primary diet	3.6

S4.7 Land-use responses: occurrence probability predictions from the partial models for artificial habitat use and diel activity

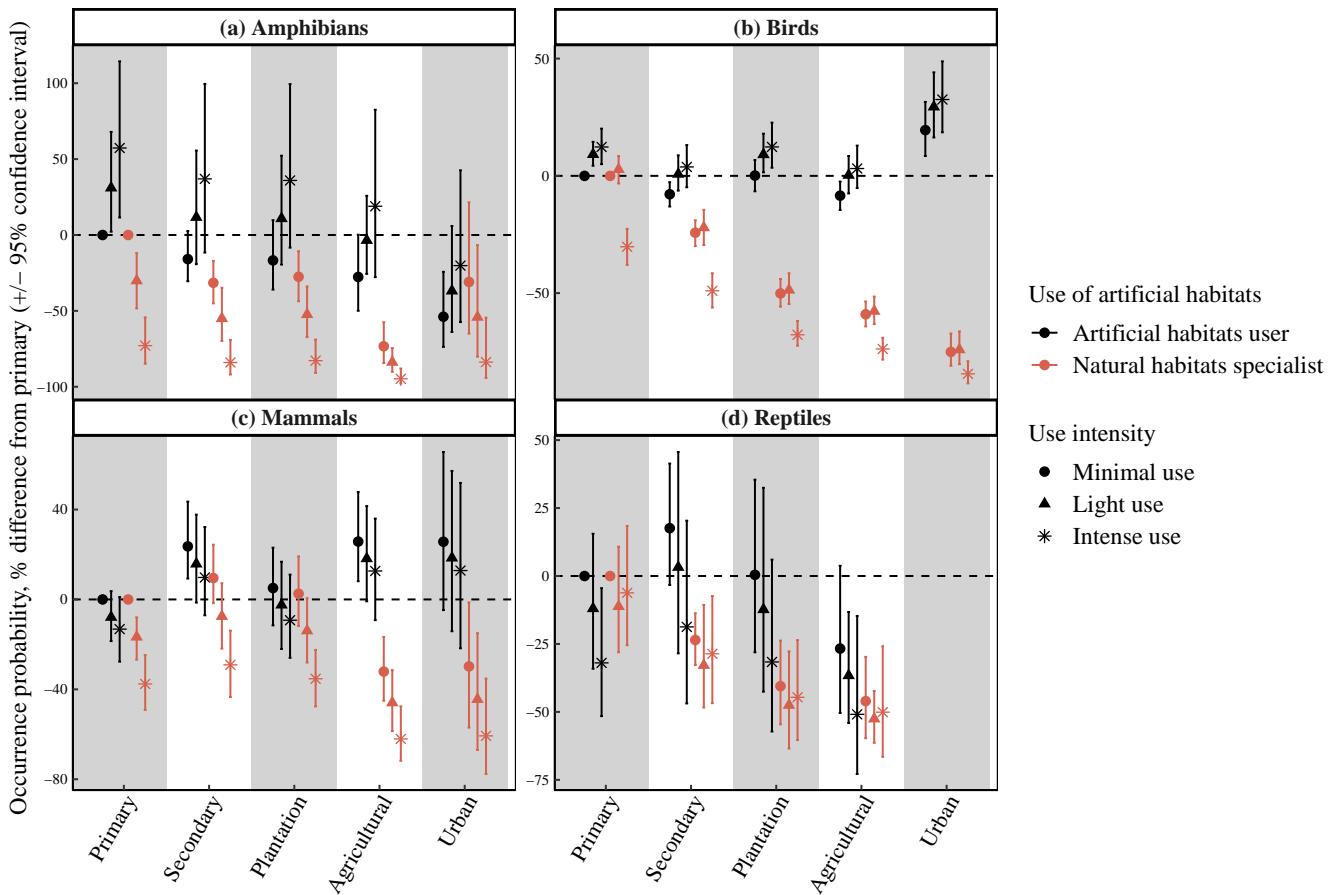


Figure S4.7: Predicted occurrence probability as a function of land use, land-use intensity, artificial habitat use and their interactions, for each class of terrestrial vertebrates (mean $\pm 95\%$ confidence interval; the predictions are rescaled with reference to minimally-used primary vegetation). The predictions were obtained from the partial models fitted in each class for artificial habitat use. Effects could not be estimated for urban reptiles, as there weren't any sampled sites. Primary: primary vegetation; Secondary: secondary vegetation; plantation: plantation forest; agricultural: cropland and pasture.

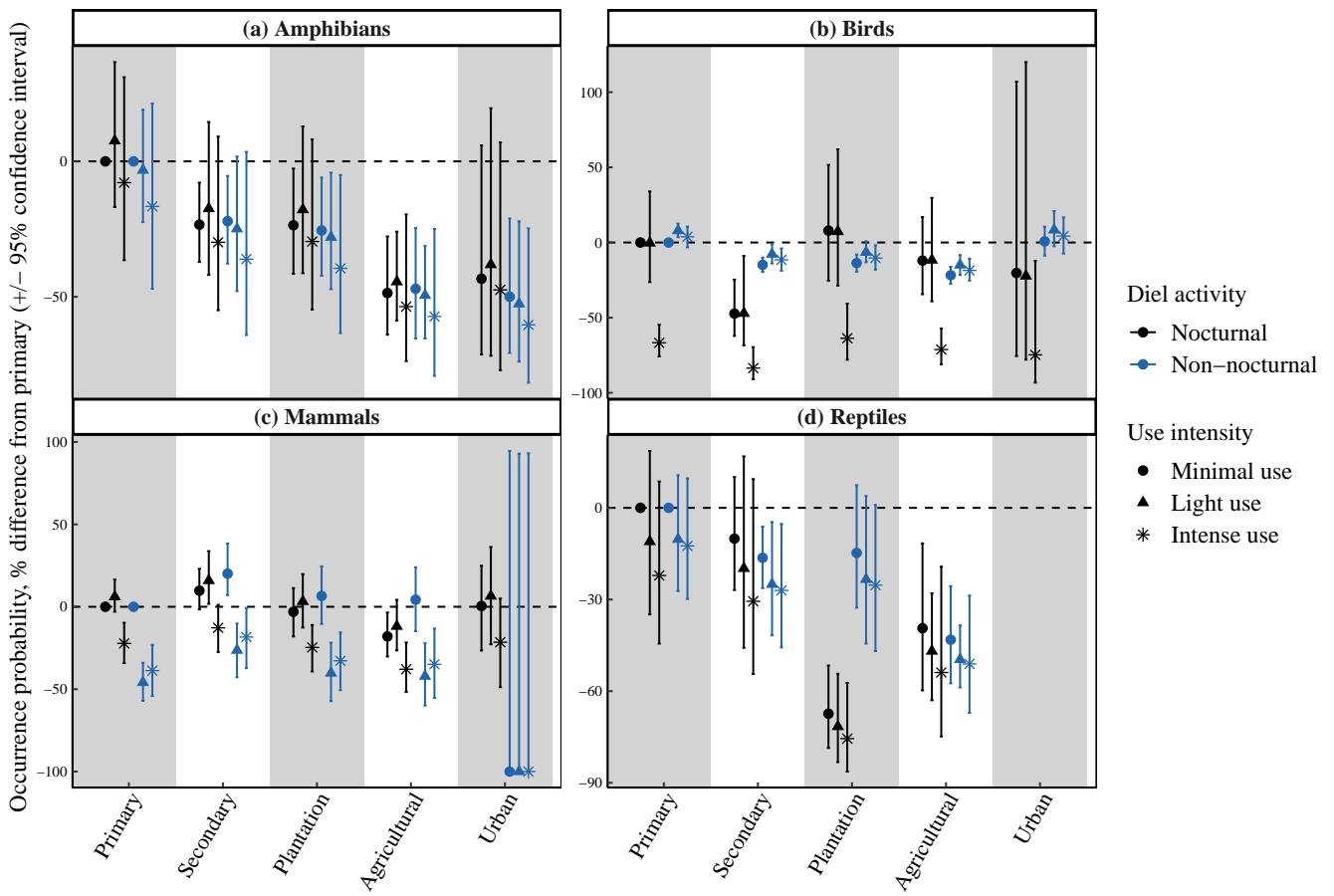


Figure S4.8: Predicted occurrence probability as a function of land use, land-use intensity, diel activity and their interactions, for each class of terrestrial vertebrates (mean $\pm 95\%$ confidence interval; the predictions are rescaled with reference to minimally-used primary vegetation). The predictions were obtained from the partial models fitted in each class for diel activity. Effects could not be estimated for urban reptiles, as there weren't any sampled sites. Error bars are large for non-nocturnal urban mammals because there were very few sampled species (only five). Primary: primary vegetation; Secondary: secondary vegetation; plantation: plantation forest; agricultural: cropland and pasture.

S4.8 Land-use responses: diagnostic plots for the full models

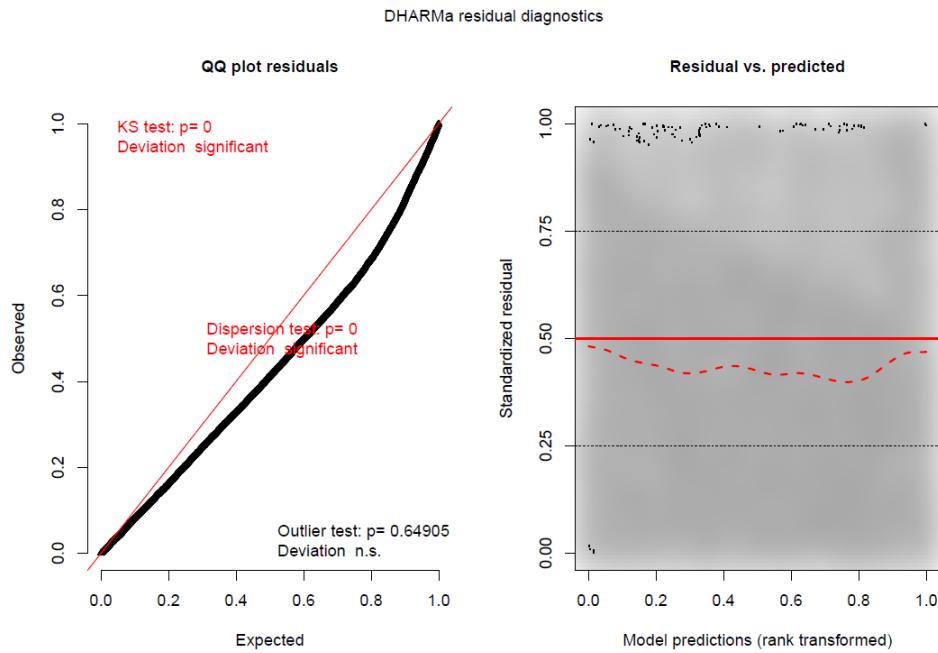


Figure S4.9: Diagnostic plots for the mixed-effects model fitted on amphibians, looking at the effects of land use, land-use intensity and the species-level ecological characteristics on species occurrence probability (full model). The diagnostic plots were obtained from the ‘DHARMA’ R package (Hartig, 2021).

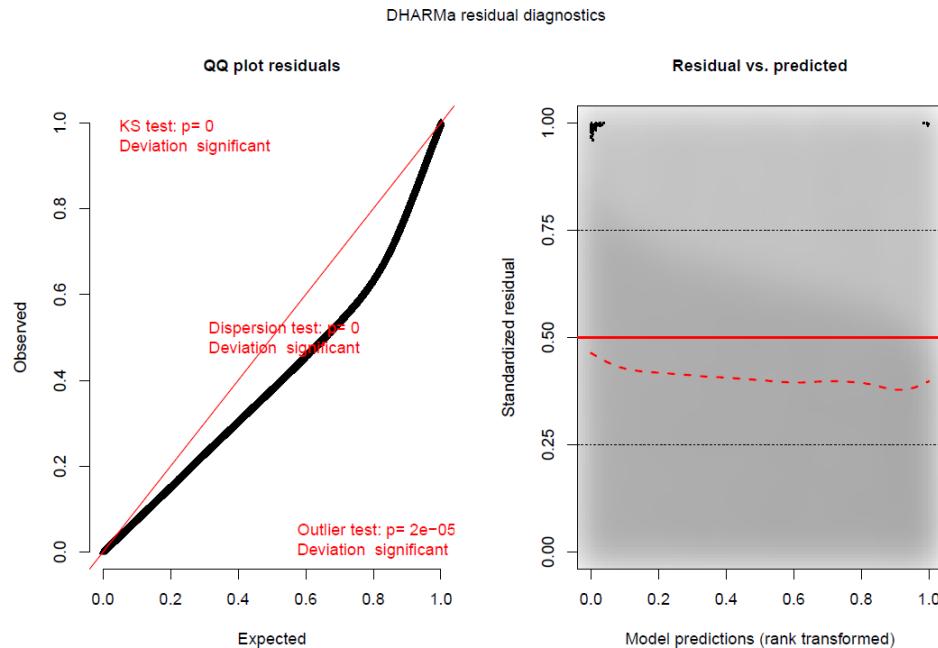


Figure S4.10: Diagnostic plots for the mixed-effects model fitted on birds, looking at the effects of land use, land-use intensity and the species-level ecological characteristics on species occurrence probability (full model). The diagnostic plots were obtained from the ‘DHARMA’ R package (Hartig, 2021).

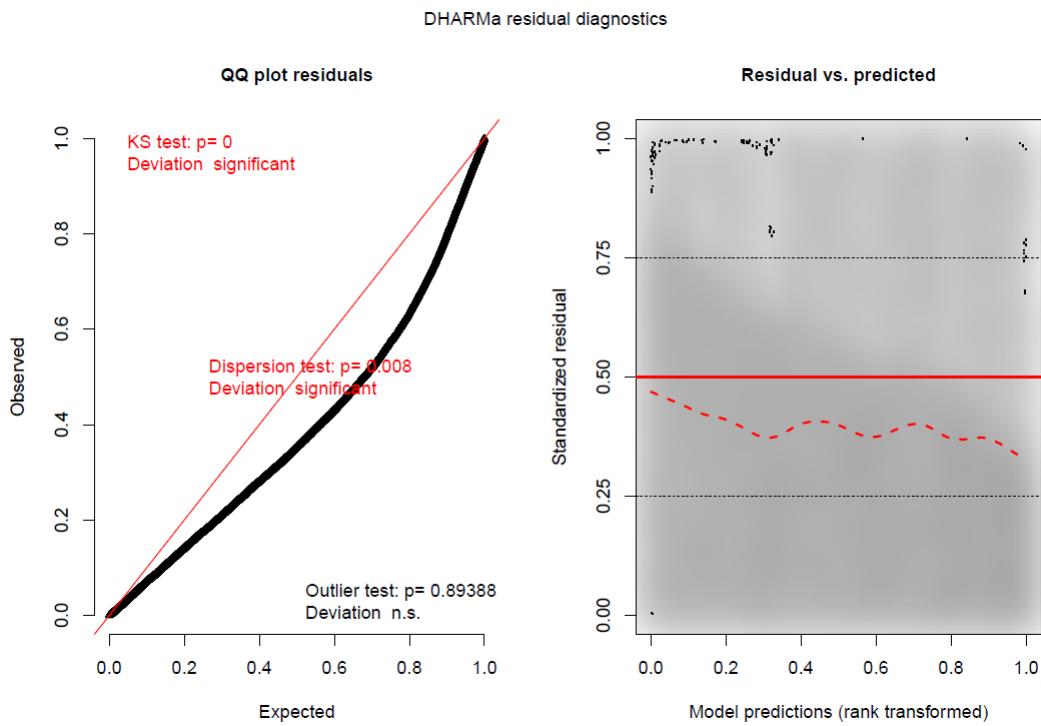


Figure S4.11: Diagnostic plots for the mixed-effects model fitted on mammals, looking at the effects of land use, land-use intensity and the species-level ecological characteristics on species occurrence probability (full model). The diagnostic plots were obtained from the ‘DHARMA’ R package (Hartig, 2021).

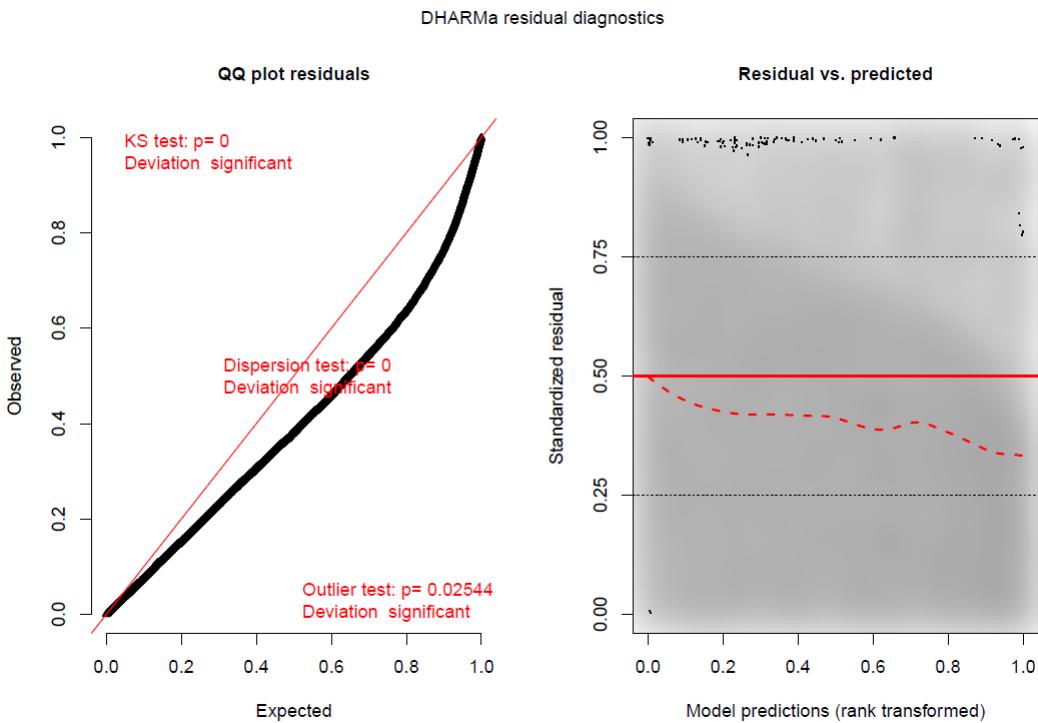


Figure S4.12: Diagnostic plots for the mixed-effects model fitted on reptiles, looking at the effects of land use, land-use intensity and the species-level ecological characteristics on species occurrence probability (full model). The diagnostic plots were obtained from the 'DHARMA' R package (Hartig, 2021).

S4.9 Climate-change sensitivity: model summaries and diagnostic plots

S4.9.1 Summaries & diagnostic plots for models fitted on species with range area $>100 \text{ km}^2$

Table S4.14: Summary for the PGLS model fitted on amphibians, investigating associations between species-level ecological characteristics and climate-change sensitivity, excluding species whose range area was $\leq 100 \text{ km}^2$ (sample size: n=4,537).

	Estimate	Std. Error	t value	Pr(> t)
Intercept	1.15	0.21	5.49	< 0.001
$\log_{10}(\text{Body mass})$	-1.97	0.61	-3.22	0.001
$\log_{10}(\text{Body mass})^2$	-0.26	0.42	-0.60	0.55
$\log_{10}(\text{Body mass})^3$	0.46	0.37	1.24	0.22
$\log_{10}(\text{Lifespan proxy})$	-0.21	0.59	-0.36	0.72
$\log_{10}(\text{Lifespan proxy})^2$	-0.14	0.43	-0.32	0.75
$\log_{10}(\text{Lifespan proxy})^3$	0.58	0.35	1.66	0.10
$\log_{10}(\text{Litter/clutch size})$	1.59	0.54	2.96	0.003
$\log_{10}(\text{Litter/clutch size})^2$	-0.06	0.38	-0.16	0.87
$\log_{10}(\text{Litter/clutch size})^3$	-0.74	0.31	-2.37	0.02
$\log_{10}(\text{Range area})$	-26.60	0.34	-77.15	< 0.001
$\log_{10}(\text{Range area})^2$	4.27	0.29	14.57	< 0.001
$\log_{10}(\text{Range area})^3$	-1.65	0.28	-5.96	< 0.001
square-root(Habitat breadth)	-2.26	0.43	-5.32	< 0.001
square-root(Habitat breadth) ²	0.81	0.30	2.67	0.01
square-root(Habitat breadth) ³	-0.59	0.28	-2.10	0.04
Specialisation: Natural habitat specialist	0.02	0.01	1.85	0.06
Diel activity: Non-nocturnal	0.04	0.01	3.28	0.001
Primary diet: Omnivore	0.01	0.03	0.29	0.77
Primary diet: Plants/seeds	0.04	0.13	0.31	0.76
Primary diet: Vertebrates	0.13	0.15	0.87	0.39

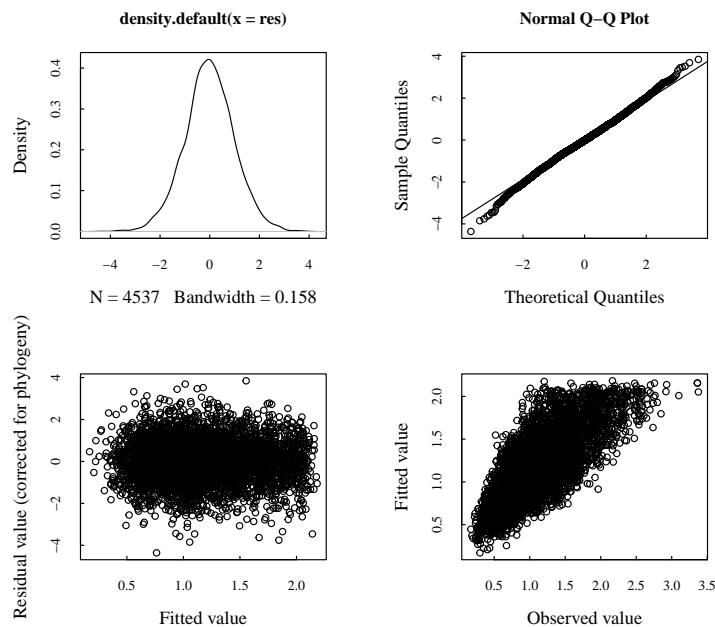


Figure S4.13: Diagnostic plots for the PGLS model fitted on amphibians, investigating associations between species-level ecological characteristics and climate-change sensitivity, excluding species whose range area was $\leq 100 \text{ km}^2$ (sample size: n=4,537).

Table S4.15: Summary for the PGLS model fitted on birds, investigating associations between species-level ecological characteristics and climate-change sensitivity, excluding species whose range area was $\leq 100 \text{ km}^2$ (sample size: n=10,198).

	Estimate	Std. Error	t value	Pr(> t)
Intercept	0.64	0.08	7.91	< 0.001
$\log_{10}(\text{Body mass})$	2.24	0.67	3.36	0.001
$\log_{10}(\text{Body mass})^2$	0.16	0.42	0.37	0.71
$\log_{10}(\text{Body mass})^3$	-0.22	0.37	-0.60	0.55
$\log_{10}(\text{Lifespan proxy})$	-0.23	0.59	-0.38	0.70
$\log_{10}(\text{Lifespan proxy})^2$	-0.84	0.39	-2.16	0.03
$\log_{10}(\text{Lifespan proxy})^3$	-0.10	0.28	-0.36	0.72
$\log_{10}(\text{Litter/clutch size})$	3.72	0.39	9.46	< 0.001
$\log_{10}(\text{Litter/clutch size})^2$	-0.42	0.33	-1.26	0.21
$\log_{10}(\text{Litter/clutch size})^3$	-0.40	0.27	-1.47	0.14
$\log_{10}(\text{Range area})$	-30.69	0.27	-113.09	< 0.001
$\log_{10}(\text{Range area})^2$	7.22	0.24	29.92	< 0.001
$\log_{10}(\text{Range area})^3$	-2.73	0.23	-11.74	< 0.001
square-root(Habitat breadth)	0.86	0.33	2.59	0.01
square-root(Habitat breadth) ²	-0.89	0.24	-3.63	< 0.001
square-root(Habitat breadth) ³	-0.22	0.23	-0.95	0.34
square-root(Diet breadth)	-0.50	0.31	-1.64	0.10
square-root(Diet breadth) ²	-0.11	0.25	-0.44	0.66
square-root(Diet breadth) ³	0.32	0.24	1.36	0.18
Specialisation: Natural habitat specialist	0.06	0.01	10.13	< 0.001
Diel activity: Non-nocturnal	-0.02	0.04	-0.66	0.51
Primary diet: Invertebrates	0.06	0.01	5.53	< 0.001
Primary diet: Omnivores	0.02	0.01	2.09	0.04
Primary diet: Plants/seeds	0.06	0.01	4.69	< 0.001
Primary diet: Vertebrates	0.01	0.02	0.83	0.41

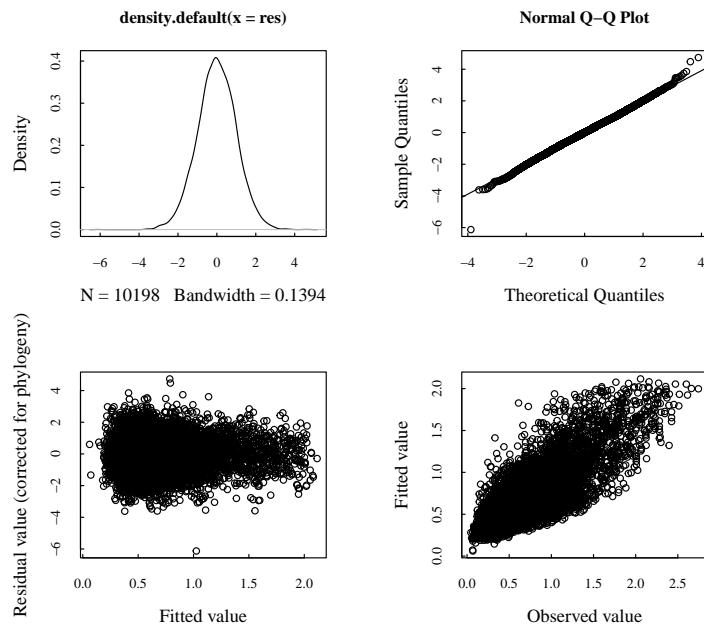


Figure S4.14: Diagnostic plots for the PGLS model fitted on birds, investigating associations between species-level ecological characteristics and climate-change sensitivity, excluding species whose range area was $\leq 100 \text{ km}^2$ (sample size: n=10,198).

Table S4.16: Summary for the PGLS model fitted on mammals, investigating associations between species-level ecological characteristics and climate-change sensitivity, excluding species whose range area was $\leq 100 \text{ km}^2$ (sample size: n=4,712).

	Estimate	Std. Error	t value	Pr(> t)
Intercept	0.84	0.16	5.37	< 0.001
$\log_{10}(\text{Body mass})$	-4.62	0.94	-4.93	< 0.001
$\log_{10}(\text{Body mass})^2$	0.40	0.56	0.72	0.47
$\log_{10}(\text{Body mass})^3$	0.59	0.44	1.33	0.18
$\log_{10}(\text{Lifespan proxy})$	1.60	1.03	1.55	0.12
$\log_{10}(\text{Lifespan proxy})^2$	-0.79	0.49	-1.60	0.11
$\log_{10}(\text{Lifespan proxy})^3$	-0.15	0.43	-0.35	0.73
$\log_{10}(\text{Litter/clutch size})$	3.29	0.71	4.63	< 0.001
$\log_{10}(\text{Litter/clutch size})^2$	0.06	0.42	0.14	0.89
$\log_{10}(\text{Litter/clutch size})^3$	-0.16	0.33	-0.47	0.64
$\log_{10}(\text{Range area})$	-24.17	0.31	-78.21	< 0.001
$\log_{10}(\text{Range area})^2$	4.15	0.28	15.09	< 0.001
$\log_{10}(\text{Range area})^3$	-0.90	0.26	-3.45	0.001
square-root(Habitat breadth)	-1.24	0.34	-3.60	< 0.001
square-root(Habitat breadth) ²	0.22	0.27	0.82	0.41
square-root(Habitat breadth) ³	-0.03	0.26	-0.10	0.92
square-root(Diet breadth)	-1.21	0.47	-2.55	0.01
square-root(Diet breadth) ²	0.33	0.36	0.91	0.36
square-root(Diet breadth) ³	0.11	0.34	0.33	0.74
Specialisation: Natural habitat specialist	0.04	0.01	3.22	0.001
Diel activity: Non-nocturnal	0.003	0.01	0.23	0.82
Primary diet: Invertebrates	-0.02	0.03	-0.62	0.54
Primary diet: Omnivores	-0.02	0.03	-0.77	0.44
Primary diet: Plants/seeds	0.03	0.02	1.32	0.19
Primary diet: Vertebrates	-0.04	0.04	-0.98	0.33

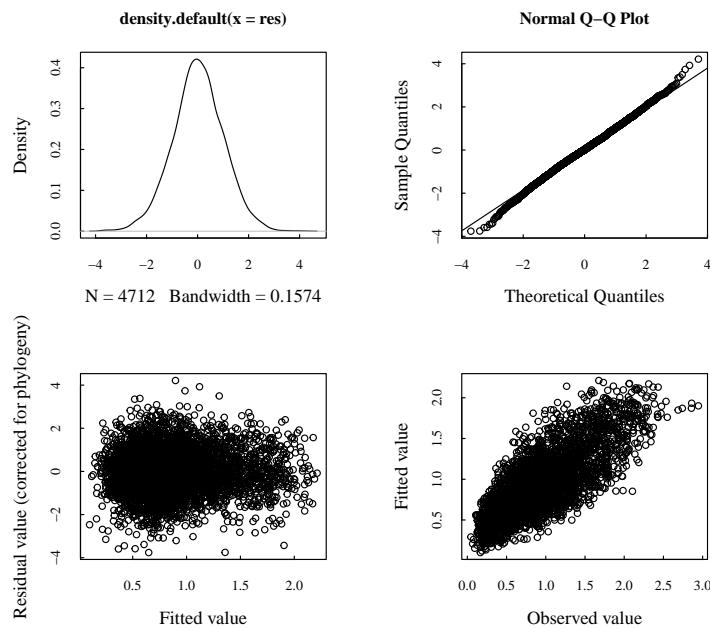


Figure S4.15: Diagnostic plots for the PGLS model fitted on mammals, investigating associations between species-level ecological characteristics and climate-change sensitivity, excluding species whose range area was $\leq 100 \text{ km}^2$ (sample size: n=4,712).

Table S4.17: Summary for the PGLS model fitted on reptiles, looking at the effects of species-level ecological characteristics on climate-change sensitivity, excluding species whose range area was $\leq 100 \text{ km}^2$ (sample size: n=7,330).

	Estimate	Std. Error	t value	Pr(> t)
Intercept	1.02	0.13	7.70	< 0.001
$\log_{10}(\text{Body mass})$	-4.62	0.62	-7.47	< 0.001
$\log_{10}(\text{Body mass})^2$	0.97	0.41	2.35	0.02
$\log_{10}(\text{Body mass})^3$	-0.96	0.33	-2.90	0.004
$\log_{10}(\text{Lifespan proxy})$	0.89	0.51	1.74	0.08
$\log_{10}(\text{Lifespan proxy})^2$	-1.27	0.39	-3.27	0.001
$\log_{10}(\text{Lifespan proxy})^3$	-0.29	0.33	-0.88	0.38
$\log_{10}(\text{Litter/clutch size})$	0.84	0.68	1.24	0.22
$\log_{10}(\text{Litter/clutch size})^2$	-0.34	0.45	-0.76	0.45
$\log_{10}(\text{Litter/clutch size})^3$	0.26	0.35	0.74	0.46
$\log_{10}(\text{Range area})$	-31.88	0.31	-101.48	< 0.001
$\log_{10}(\text{Range area})^2$	5.15	0.27	19.03	< 0.001
$\log_{10}(\text{Range area})^3$	-1.97	0.26	-7.53	< 0.001
square-root(Habitat breadth)	0.56	0.35	1.59	0.11
square-root(Habitat breadth) ²	-0.67	0.27	-2.49	0.01
square-root(Habitat breadth) ³	0.05	0.26	0.21	0.83
square-root(Diet breadth)	-0.33	0.37	-0.90	0.37
square-root(Diet breadth) ²	-0.02	0.27	-0.08	0.94
square-root(Diet breadth) ³	0.31	0.26	1.22	0.22
Specialisation: Natural habitat specialist	0.10	0.01	8.50	< 0.001
Diel activity: Non-nocturnal	-0.02	0.01	-1.88	0.06
Primary diet: Omnivore	-0.03	0.03	-0.92	0.36
Primary diet: Vertebrates	-0.02	0.02	-1.05	0.30

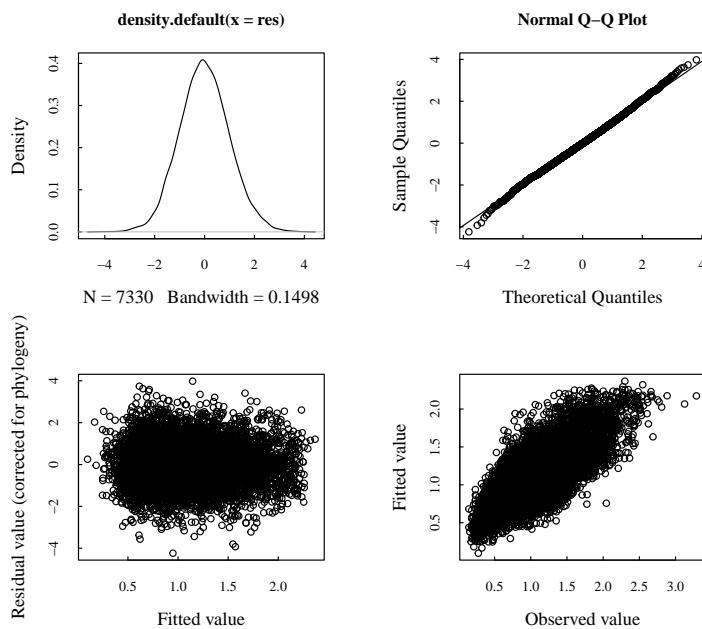


Figure S4.16: Diagnostic plots for the PGLS model fitted on reptiles, investigating associations between species-level ecological characteristics and climate-change sensitivity, excluding species whose range area was $\leq 100 \text{ km}^2$ (sample size: n=7,330).

S4.9.2 Robustness checks of the PGLS models

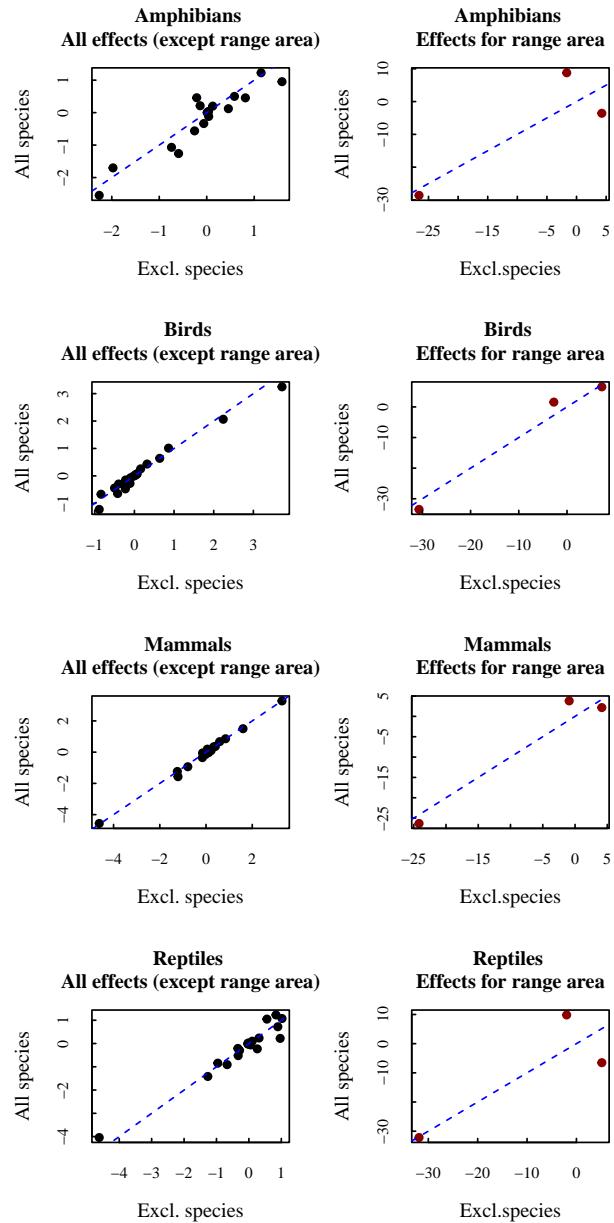


Figure S4.17: Effects for the PGLS models investigating associations between species-level ecological characteristics and climate-change sensitivity, either fitted on all species (y-axis), or fitted on the species whose range area was $>100 \text{ km}^2$ (x-axis). Overall, the estimates from both sets of models were congruent, except for those estimated for geographical range area (I show the effects for range area separately from the effects of other characteristics). Across all classes, the relationship between sensitivity and geographical range area was reversed between the two sets of models. I found that sensitivity was positively associated with geographical range area when including all species, likely because of the underestimation of climate-change sensitivity for the most narrow-ranging species when working with a resolution of 5 km^2 (see Figure S4.6). The dashed line is the identity line ($y=x$).

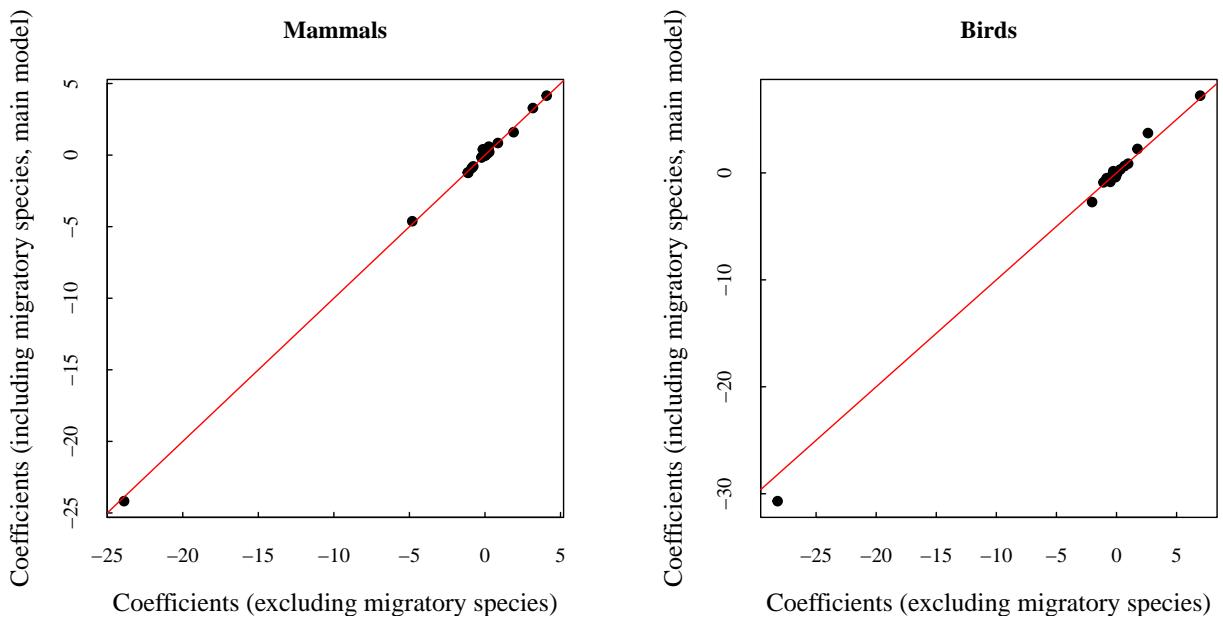


Figure S4.18: Effects for the PGLS models investigating associations between species-level ecological characteristics and climate-change sensitivity, either fitted on all birds and mammals (y-axis), or on the subset of bird and mammal species that were found not to be migratory (x-axis). Overall, the estimates from both sets of models were congruent. The dashed line is the identity line ($y=x$).

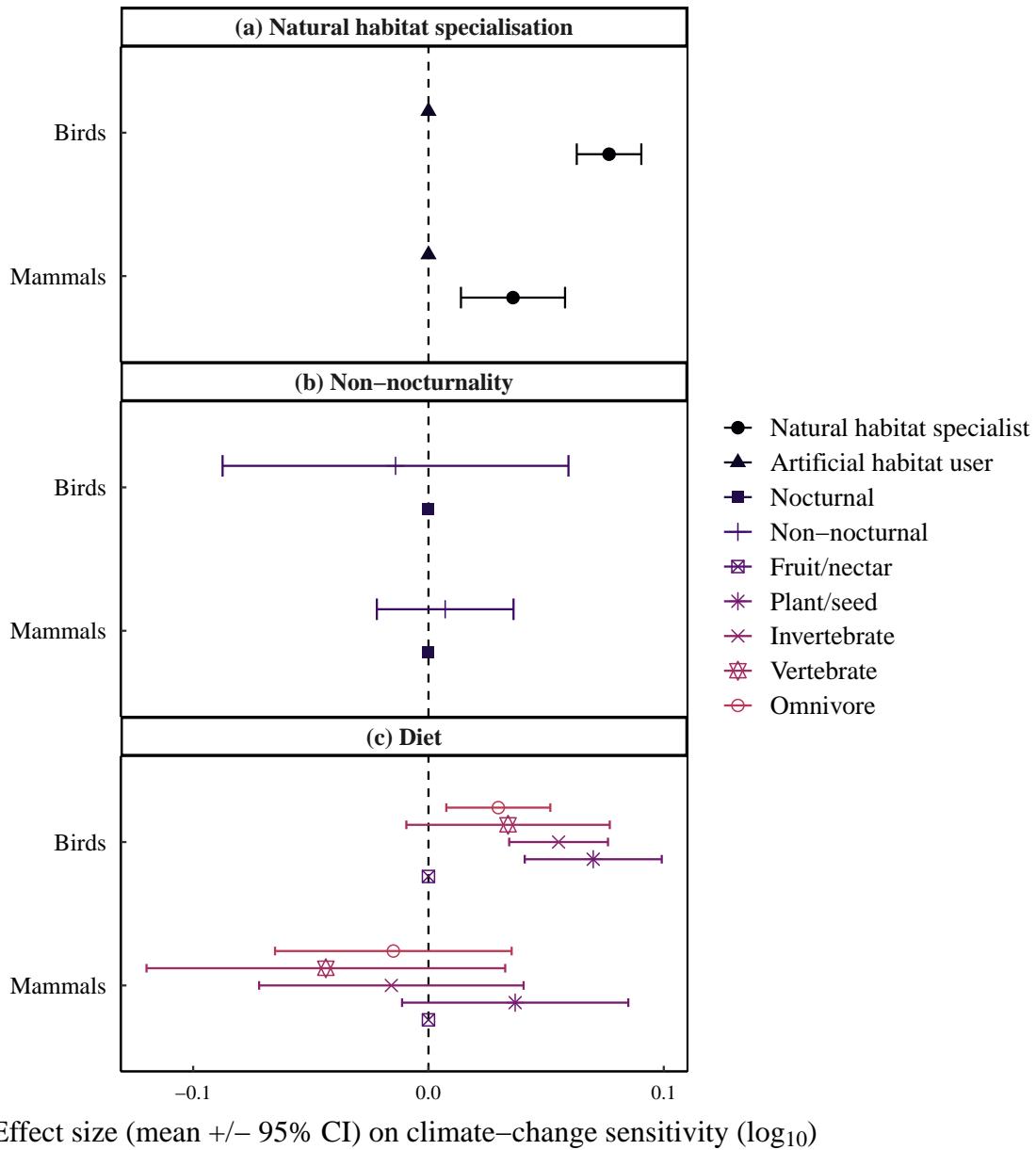


Figure S4.19: Estimated effects of the categorical traits on climate-change sensitivity, from the PGLS models fitted on birds and mammals, excluding species identified as migratory (mean effect \pm 95% confidence interval). For each categorical trait, I show the effect size for all levels referring to the reference level (vertical dashed line). **(a)** For artificial habitat use, the reference level is ‘Artificial habitat user’; **(b)** for diel activity, the reference level is ‘Nocturnal’; **(c)** for diet, the reference level (for mammals and birds) is ‘Fruit/nectar’ eaters.

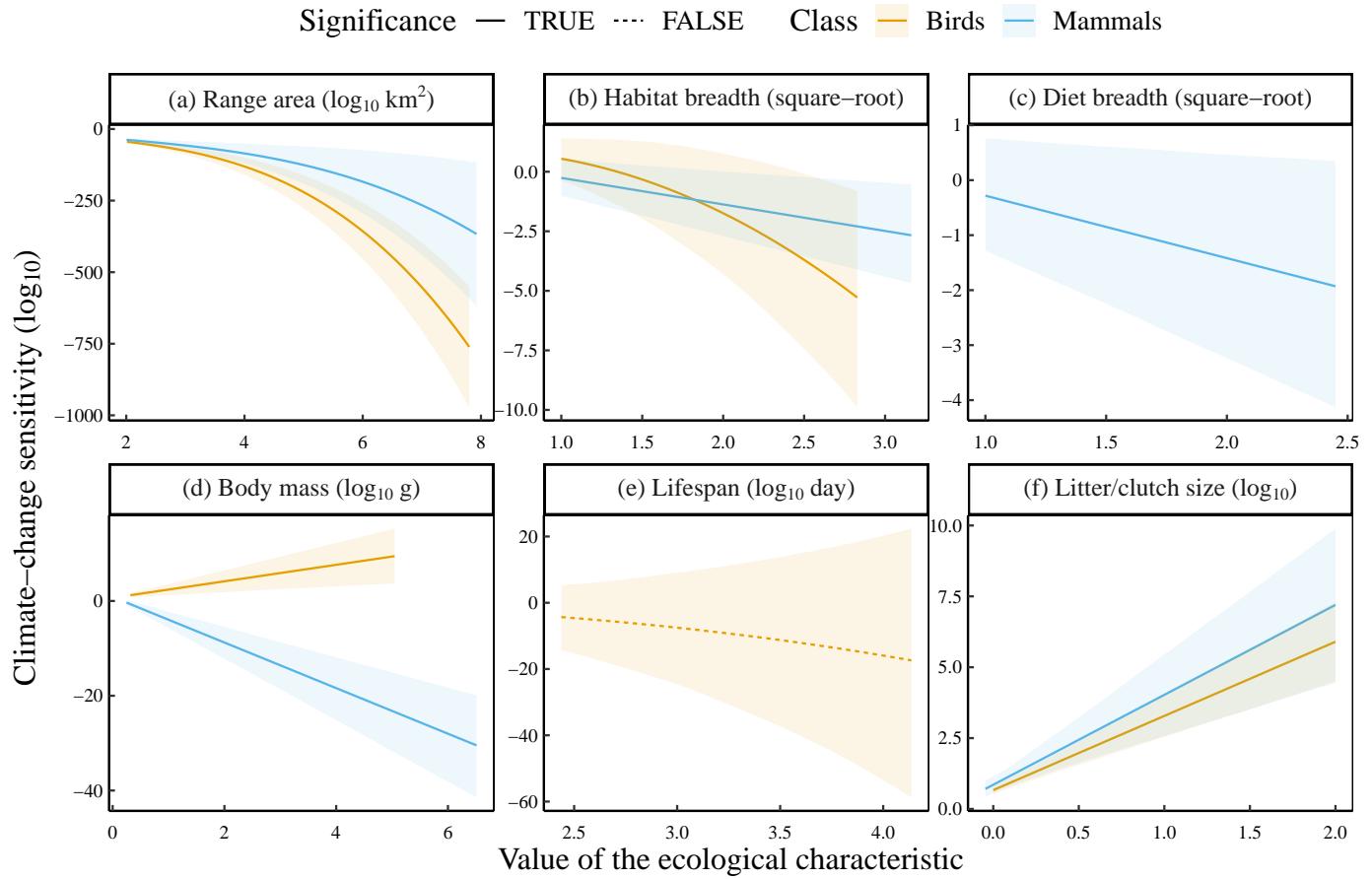


Figure S4.20: Effects of the continuous ecological characteristics on climate-change sensitivity, estimated from the PGLS models fitted on birds and mammals, excluding species identified as migratory. The lines represent the estimated relationships between climate-change sensitivity and ecological characteristics; the shaded areas are 95% confidence intervals.

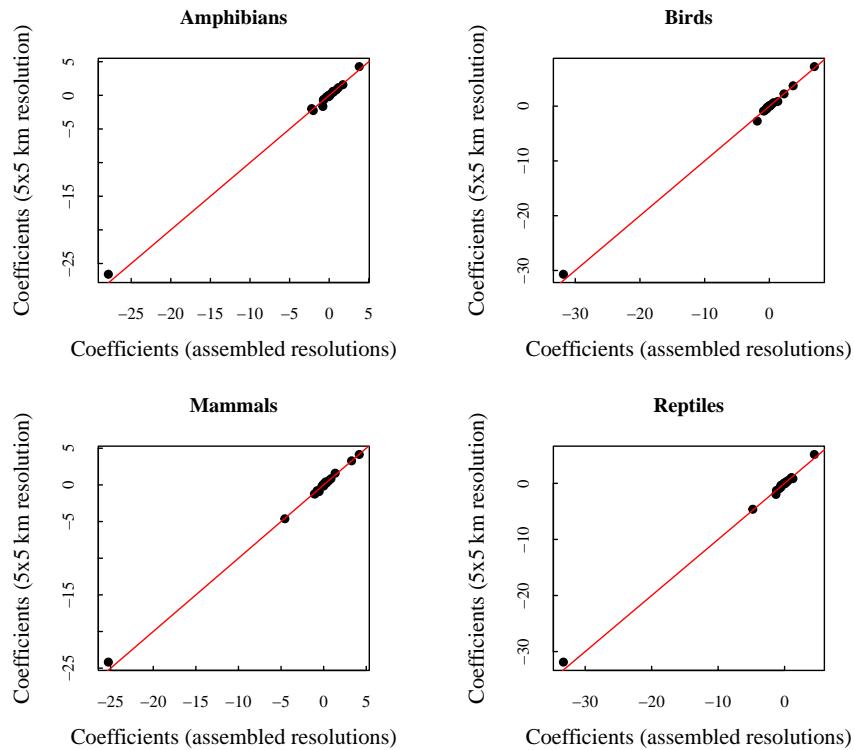


Figure S4.21: Effects for the PGLS models investigating associations between species-level ecological characteristics and climate-change sensitivity, either using climate-change sensitivity estimated at 5×5 km resolution (y-axis), or using climate-change sensitivity estimated at different resolutions (x-axis; see Methods in main text). Overall, the estimates from both sets of models were congruent. The dashed line is the identity line ($y=x$).

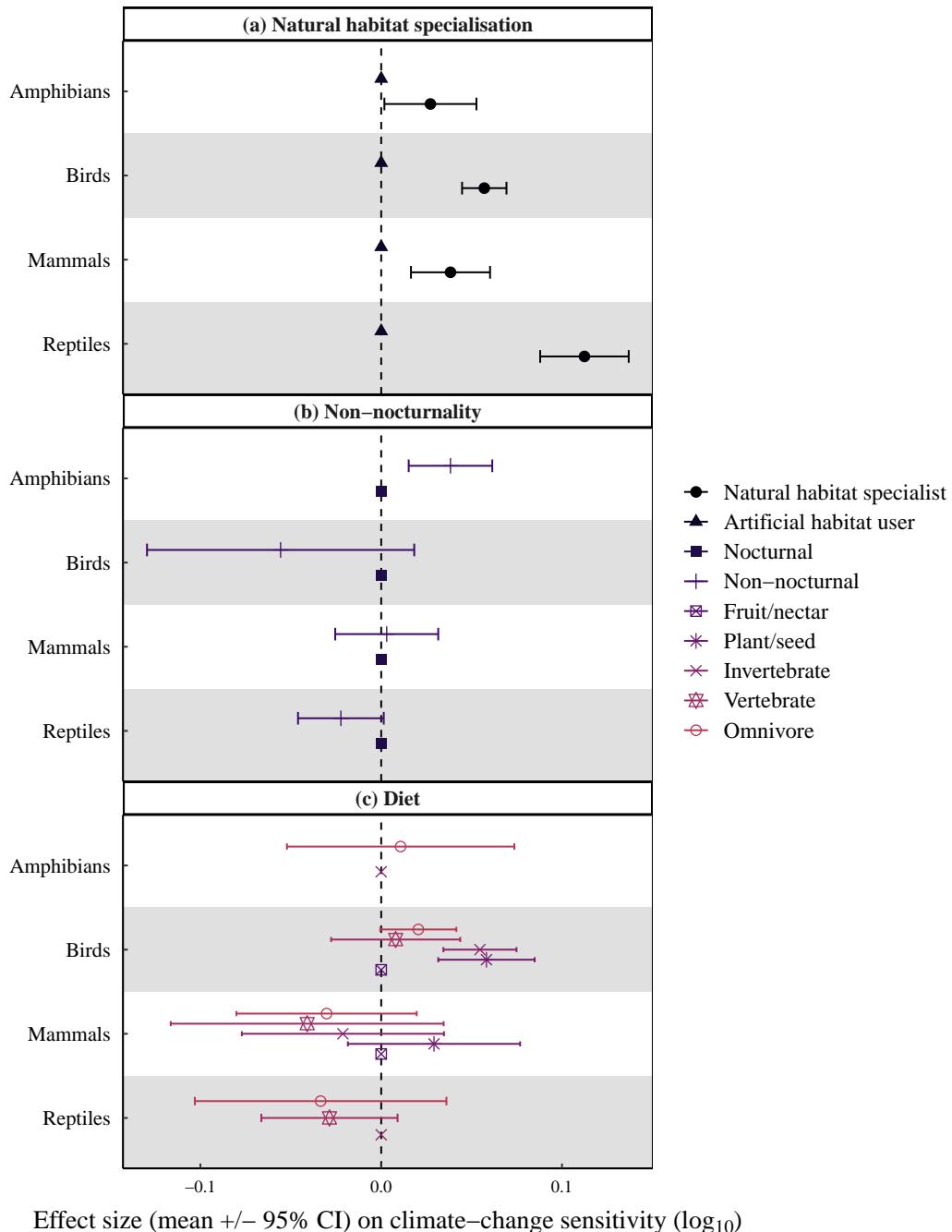


Figure S4.22: Estimated effects of the categorical traits on climate-change sensitivity, using climate-change sensitivity estimated at different resolutions (see Methods in main text; mean effect \pm 95% confidence interval). For each categorical trait, I show the effect size for all levels referring to the reference level (vertical dashed line). (a) For artificial habitat use, the reference level is ‘Artificial habitat user’; (b) for diel activity, the reference level is ‘Nocturnal’; (c) for diet, the reference level (for mammals and birds) is ‘Fruit/nectar’ eaters.

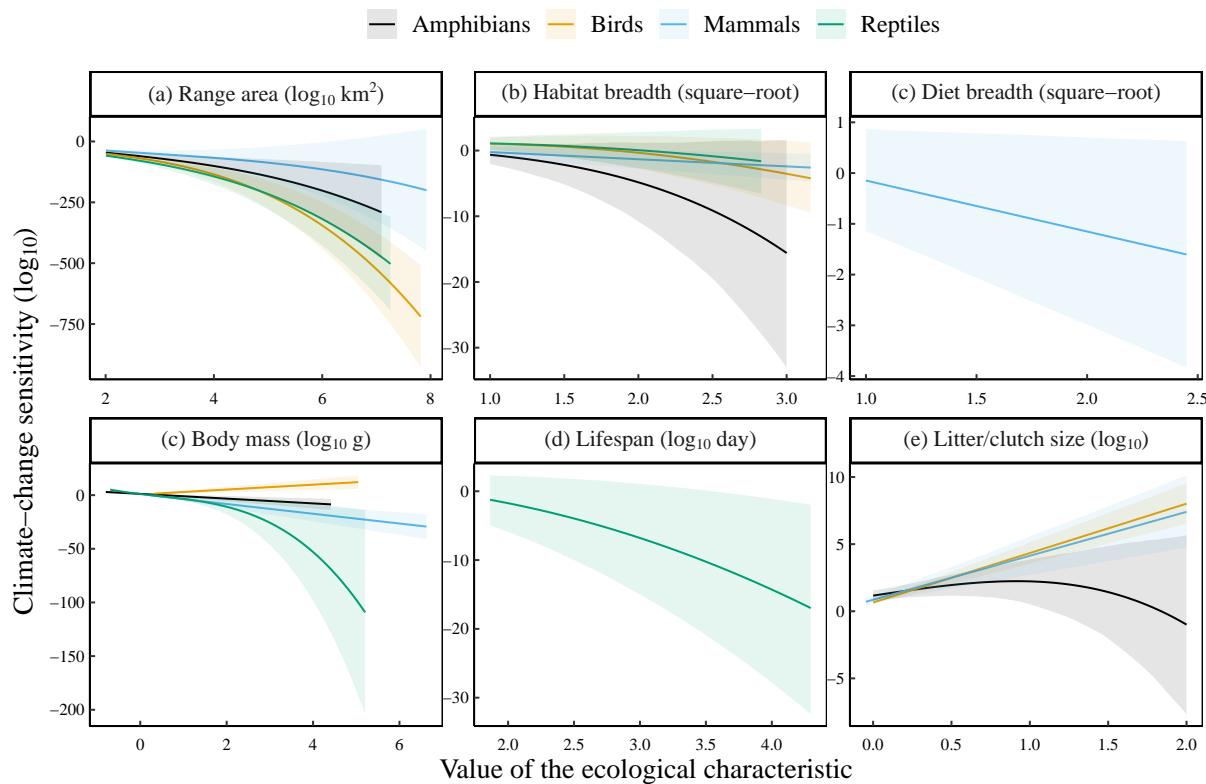


Figure S4.23: Effects of the continuous ecological characteristics on climate-change sensitivity, using climate-change sensitivity estimated at different resolutions (see Methods in main text). The lines represent the estimated relationships between climate-change sensitivity and ecological characteristics; the shaded areas are 95% confidence intervals.

S4.10 Validations on complete trait data subsets

Table S4.18: Summary of the effects of the ecological characteristics (except for diet) on (a) species' responses to disturbed land uses ('within land-use type' effects) and (b) species climate-change sensitivity, for each class of terrestrial vertebrates, from the models fitted on empirical trait values (excluding all imputed values). The symbol  indicates where a characteristic has a significant negative effect on occurrence probability within a disturbed land-use type (within any of the land-use intensities), or where the characteristic renders species significantly more sensitive to climate change. A  indicates a significantly positive effect of a characteristic on occurrence probability in a land-use type (within any of the land-use intensities), or significantly lower sensitivity to climate change. For the land-use effects, I report 'within land-use type effects' here, that is, within a disturbed land use whether there were significant differences in occurrence probability among species with different trait values. These effects were derived from the interactive terms of the full models.

(a) "Within land-use type" effects of the species-level characteristics on occurrence probability - <=> more sensitive; + <=> less sensitive																Predictors				(b) Climate-change sensitivity: - <=> more sensitive + <=> less sensitive				
Secondary vegetation				Plantation forest				Agricultural				Urban												
																								
0	-	0	0	0	-	0	0	0	-	-	+	+	-	0	NA	More narrowly-distributed		Habitat affinity	-	-	-	(-)		
0	-	0	0	-	-	0	0	-	-	0	-	-	-	0	NA	Smaller habitat breadth		0	-	-	-	-		
-	-	0	0	0	-	0	0	-	-	-	+	0	-	0	NA	Natural habitat specialist		Specialisa -tion	0	-	0	-		
0	+	+	0	+	+	(+)	+	0	+	+	0	0	0	0	NA	Non-nocturnal		Resource use	0	0	0	0		
-	0	(+)	0	(-)	0	+	0	-	0	(+)	+	-	-	0	NA	Narrower diet breadth		0	0	0	0			
NA	(+)	+	NA	NA	+	+	NA	NA	-	0	NA	NA	+	0	NA	Smaller body mass		Life-history	0	(+)	-	0		
NA	0	0	-	NA	-	(-)	0	NA	-	(-)	0	NA	0	-	NA	Smaller litter/clutch size		(+)	+	+	0			
NA	0	0	-	NA	(-)	0	0	NA	+	0	+	NA	-	0	NA	Shorter-lived		(+)	0	0	0			

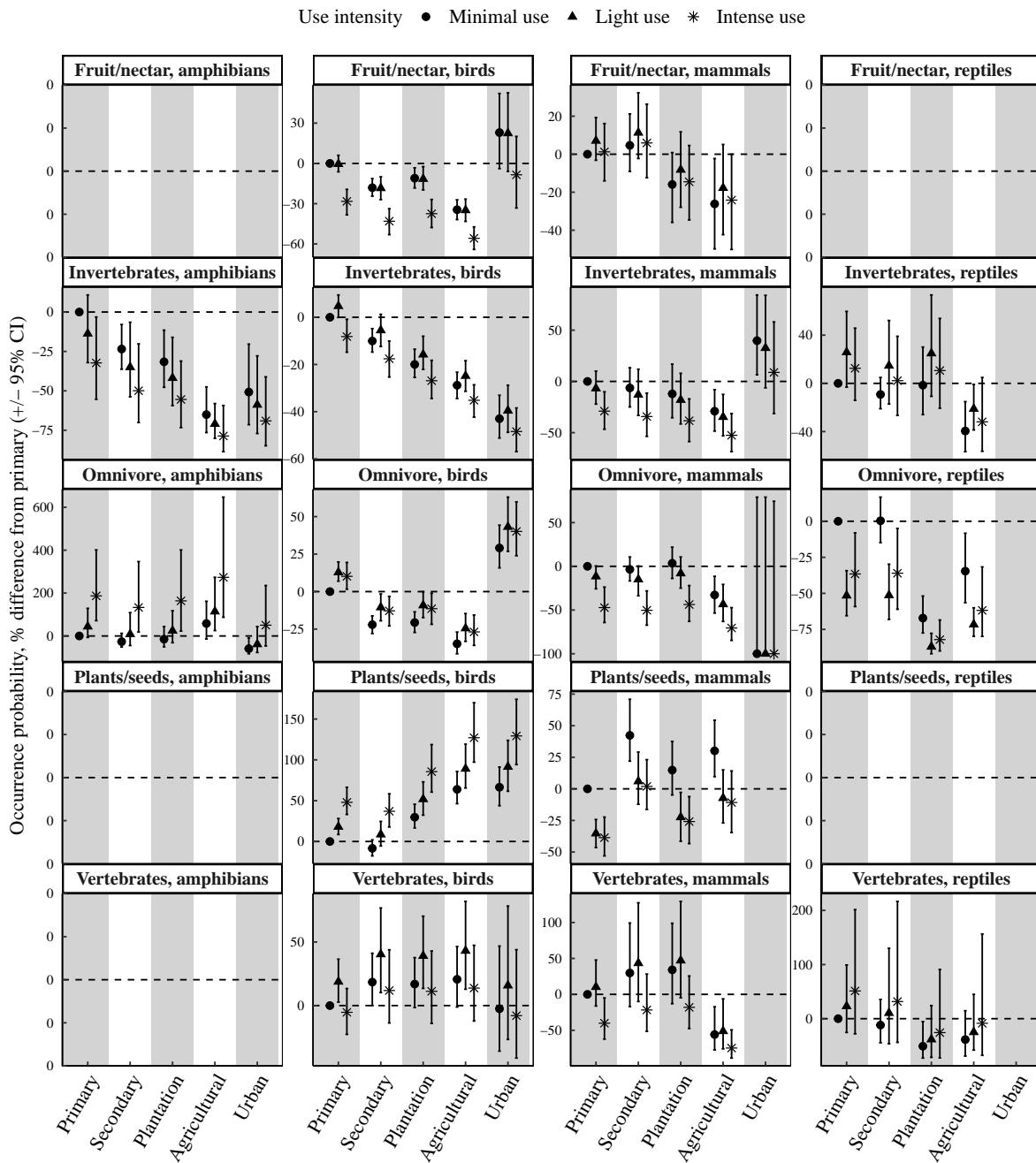


Figure S4.24: Predicted occurrence probability as a function of land use, land-use intensity, diet and their interactions in each class. The predictions were obtained from the partial models fitted in each class for diet, estimated using empirical trait data subsets (i.e., excluding imputed trait values). Empty plots are drawn where there were no data for a diet category in a given class (e.g., amphibian fruit/nectar eaters). Effects could not be estimated for urban reptiles, as well as for urban vertebrate, fruit/nectar and plant/seed eaters for mammals because there were no sampled species. The predictions are rescaled with reference to minimally-used primary vegetation. Primary: primary vegetation; Secondary: secondary vegetation; plantation: plantation forest; agricultural: cropland and pasture.

Appendix 4: Supporting information for Chapter 5

Table S5.1: Number of species for which RMR data was collected in each class & data sources, initial RMR coverage for PREDICTS species and phylogenetic signal in RMR.

Class	RMR data	Coverage for PREDICTS species	Phylogenetic signal (Pagel's λ , $\pm 95\%$ CI)
Amphibians	126 species from Stark et al. (2020)	16/379 species (4%)	0.89 (0.86-0.91)
Birds	719 species from McNab (2009) Fristoe et al. (2015) Londoño et al. (2015) Stark et al. (2020)	317/3129 species (10%)	0.97 (0.95-0.98)
Mammals	685 species from PanTHERIA (Jones et al., 2009) Fristoe et al. (2015) Stark et al. (2020)	148/556 species (27%)	0.99 (0.98-0.99)
Reptiles	173 species from Stark et al. (2020)	24/329 species (7.3%)	0.90 (0.86-0.92)

$$\begin{aligned} \log(\text{Abundance}) = & \text{LU} + \text{LUI} + \text{TG} + \log(\text{BM}) + \\ & \text{LU:LUI} + \text{LU:TG} + \text{LU:}\log(\text{BM}) + \text{LUI:TG} + \text{LUI:}\log(\text{BM}) + \text{TG:}\log(\text{BM}) + \\ & \text{LU:TG:}\log(\text{BM}) + \text{LUI:TG:}\log(\text{BM}) \end{aligned}$$

Figure S5.1: Formula for the abundance model, used to understand the role of shifts in the body mass of species on observed changes in tRMR (see Chapter 5, section 5.2.5, ‘Disentangling the effects of body mass and abundance on tRMR’). I fitted a model to explain changes in species abundance (given presence) by land use, land-use intensity, trophic group, body mass and their interactions. The model included all two-way interactions among these predictors. To account for potential differences in the slope of the relationship between abundance and body mass among the different trophic groups, I also included two three-way interactions in the model (among land use, trophic group and body mass; and among land-use intensity, trophic group and body mass). Random effects included study, site and species identity. LU: land use; LUI: land-use intensity; TG: trophic group; BM: body mass.

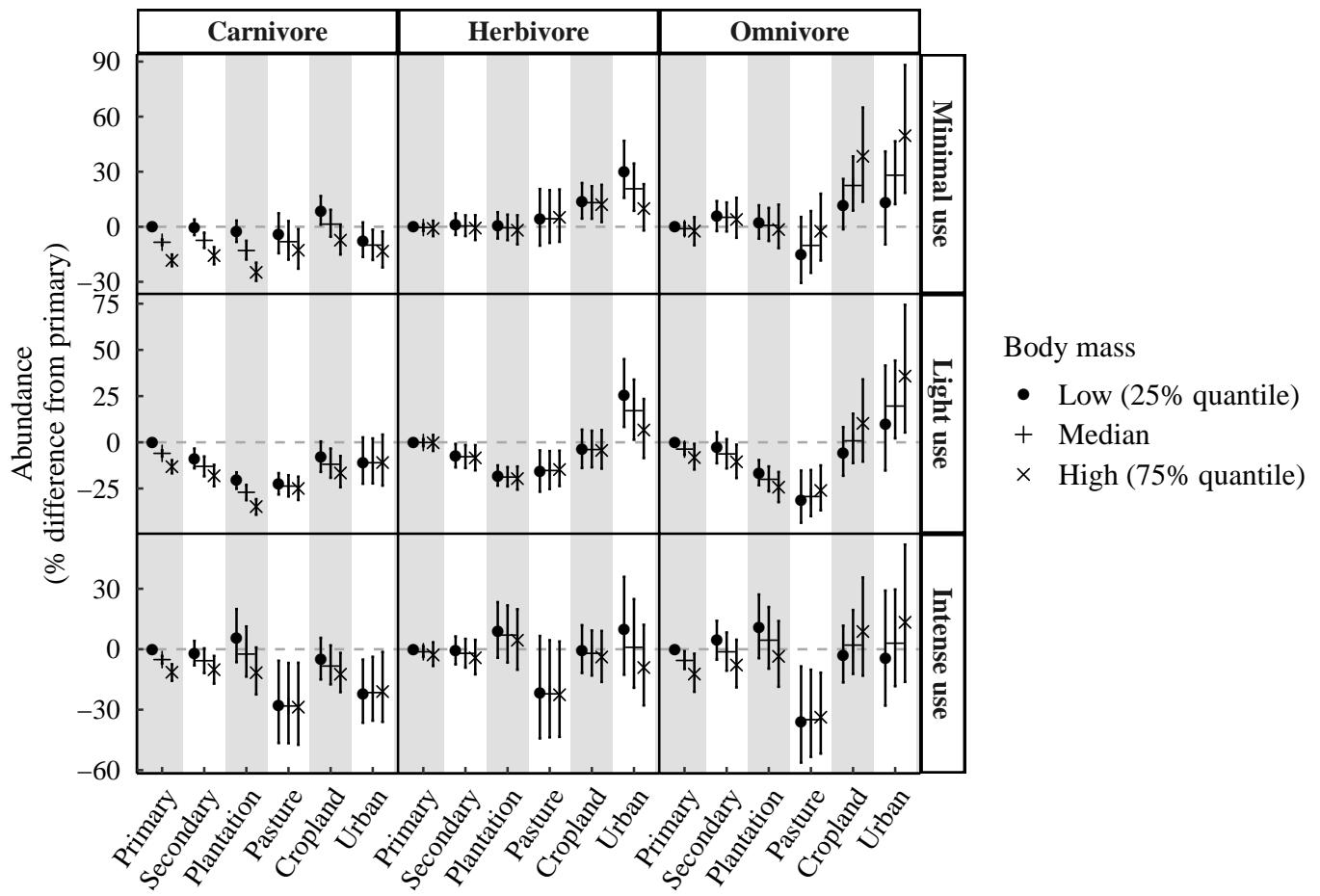


Figure S5.2: Effects of land use, land-use intensity, trophic group, body mass and their interactions on assemblage-level total abundance, estimated from the model specified in Figure S5.1. The predictions are rescaled with reference to minimally used primary vegetation, considered to be the undisturbed baseline. Primary: primary vegetation; secondary: secondary vegetation; plantation: plantation forest. For visualisation purposes, I plotted the predictions for three body mass levels (but body mass was considered as a continuous variable in the model).

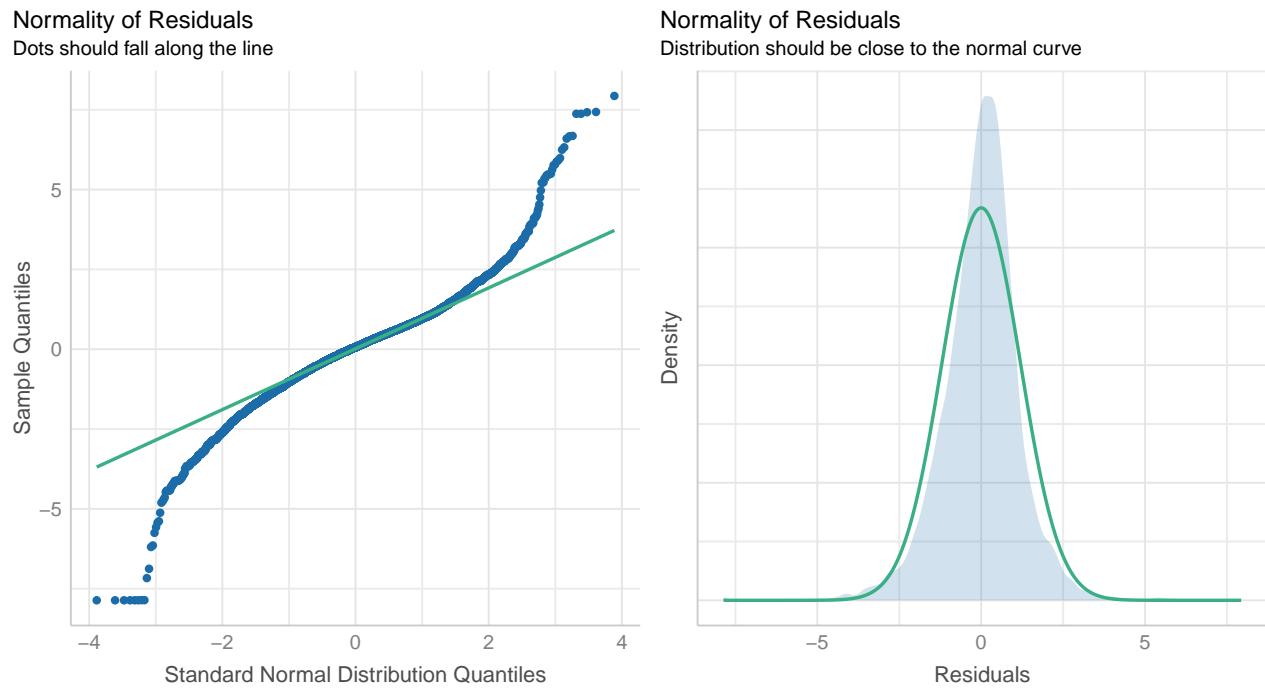
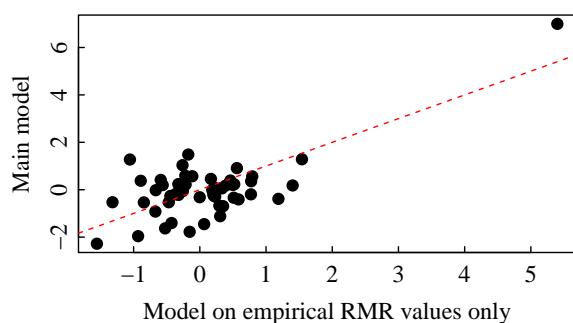


Figure S5.3: Diagnostic plots (qq-plot and residual distribution) for the linear mixed-effects model looking at the effects of land use, land-use intensity, trophic group and their interactions on assemblage-level total RMR (*model I*). The diagnostic plots were obtained with the ‘performance’ R package (Lüdecke et al., 2021).

(a) Coefficient estimates



(b) Standard error estimates

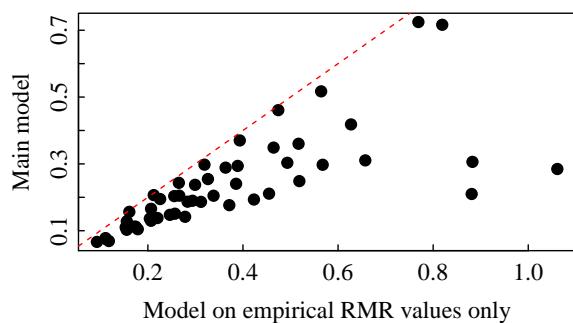


Figure S5.4: Estimates for the models looking at the effects of land use, land-use intensity, trophic group and their interactions on assemblage-level total RMR. I plotted the estimates from the model fitted on the empirical and imputed RMR values (*model 1*) on the y-axis, and the estimates from the model fitted on the empirical RMR values only on the x-axis.

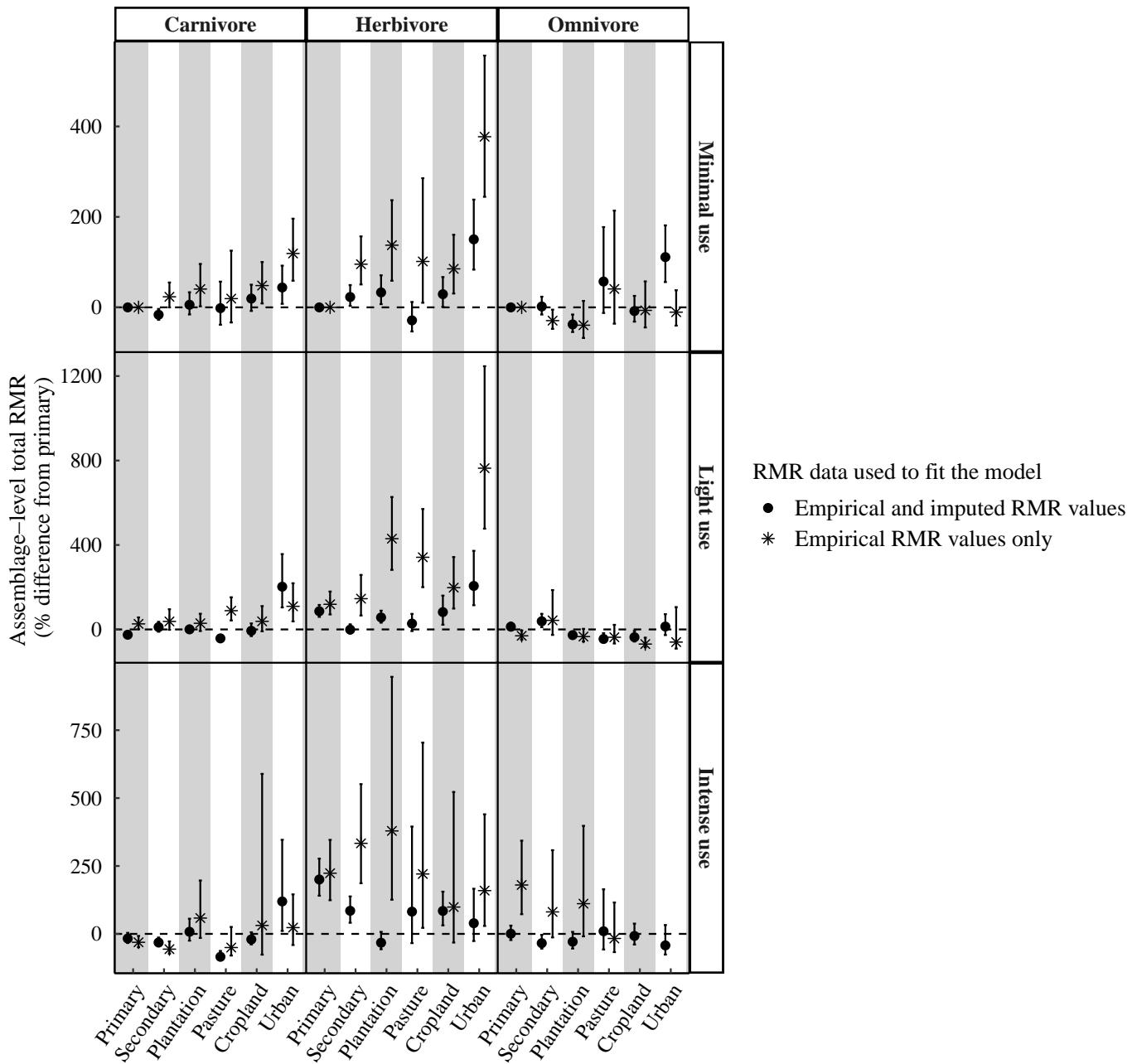


Figure S5.5: Effects of land use, land-use intensity, trophic level and their interactions on assemblage-level total RMR, estimated from the model fitted on the empirical and imputed RMR values (*model 1*) and from the model fitted on the empirical values only. The predictions are rescaled with reference to minimally used primary vegetation, considered to be the undisturbed baseline. Primary: primary vegetation; secondary: secondary vegetation; plantation: plantation forest.

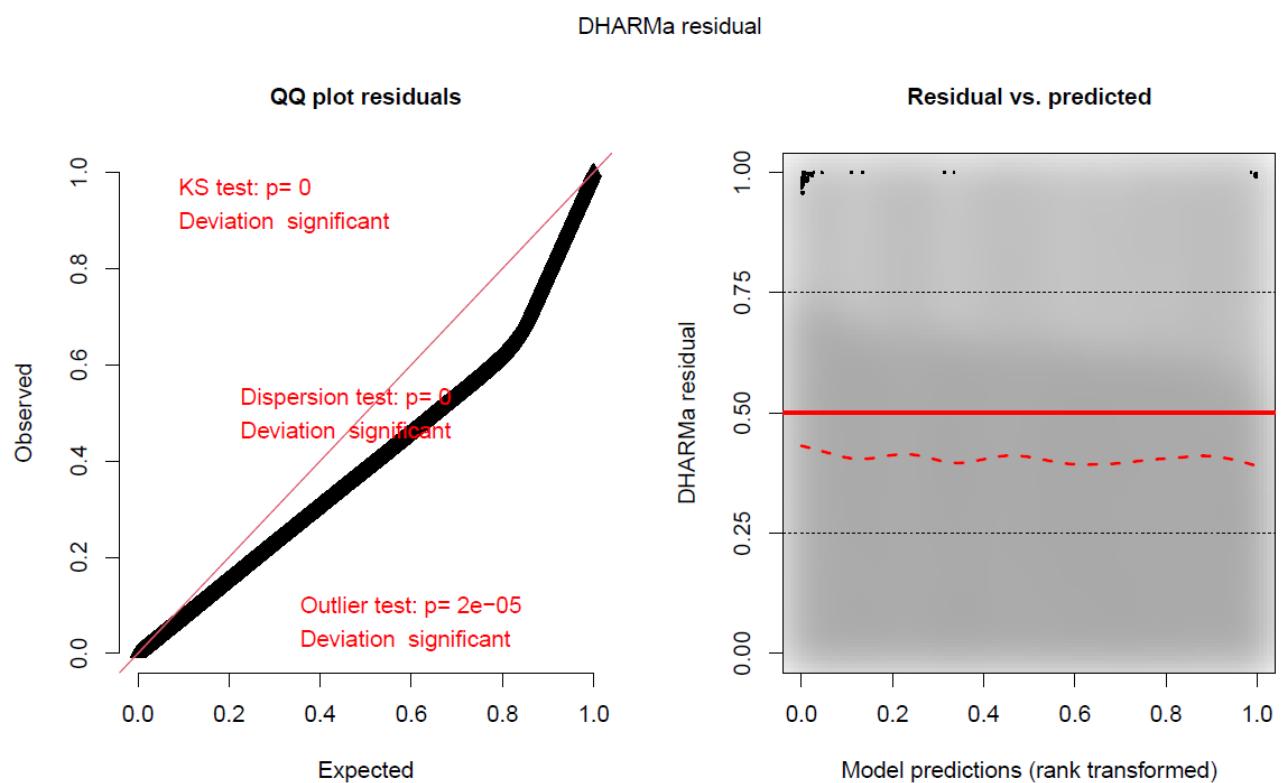


Figure S5.6: Diagnostic plots for the generalised mixed-effects model looking at the effects of land use, land-use intensity, trophic group, residual RMR and their interactions on species' probability of occurrence (*model 2*). The diagnostic plots were obtained with the 'DHARMA' R package (Hartig, 2021).

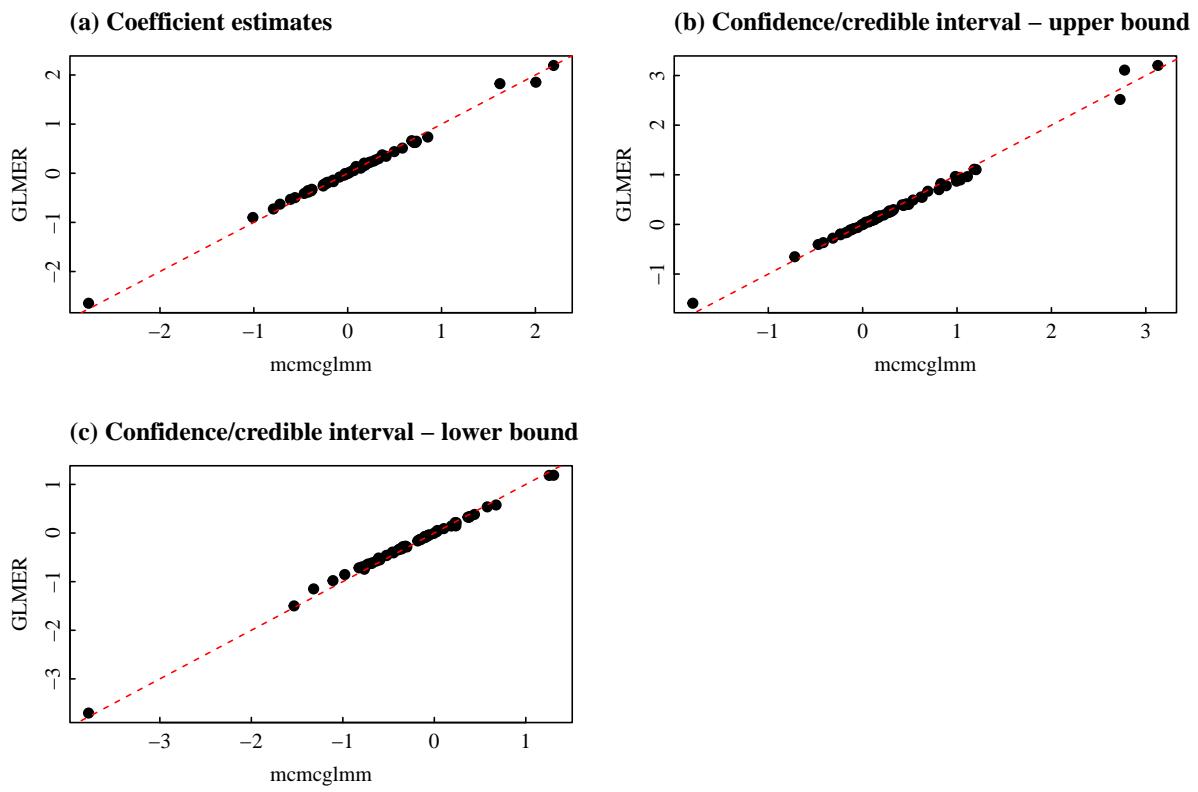
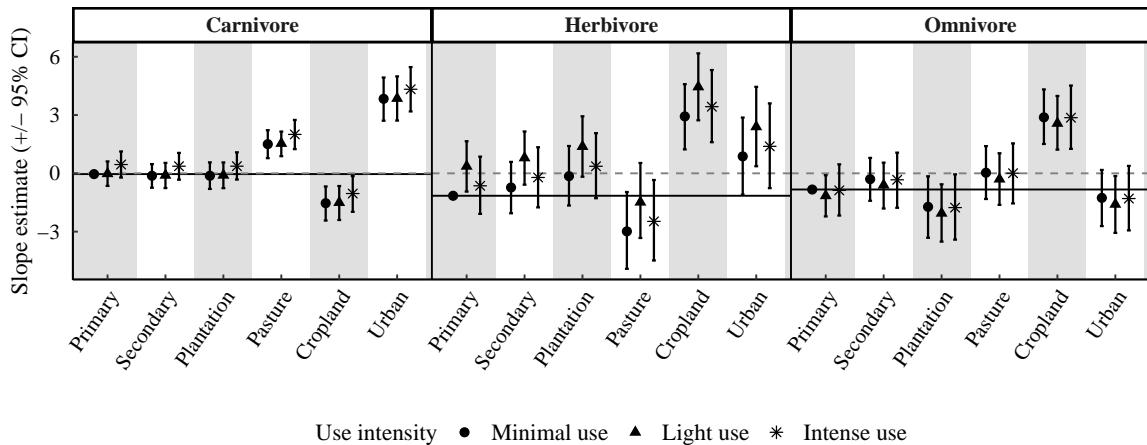


Figure S5.7: Model's coefficients from the occurrence model fitted using the 'lme4' package (Bates et al., 2015) against coefficients from the model fitted using a Bayesian framework with the 'MCMCglmm' package (Hadfield, 2010). The models were fitted to investigate the effects of land use, land-use intensity, trophic group and residual RMR on species occurrence probability (model 2).

(a) Slope of the relationships between occurrence probability (log-odds) and residual RMR



(b) Predicted effects of residual RMR on occurrence probability (minimal use intensity)

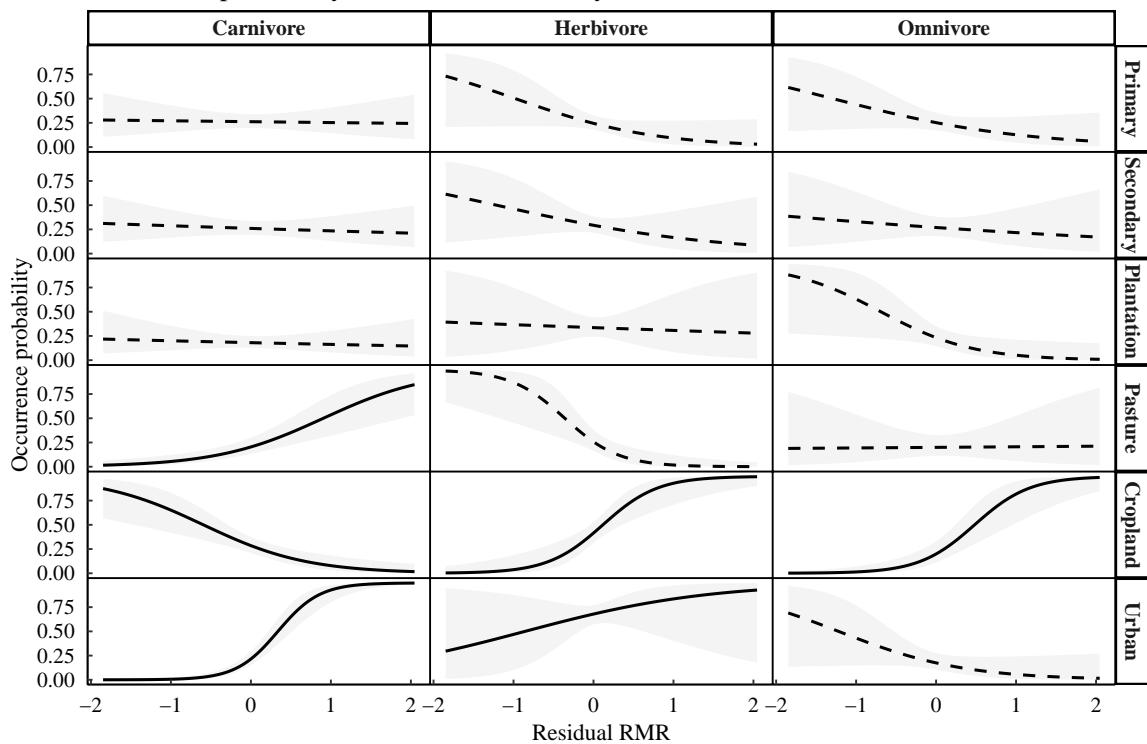


Figure S5.8: (a) Slope estimates for the relationship between residual RMR and occurrence probability in each land-use type and for the three levels of land-use intensity, from the model fitted using the empirical RMR values (i.e., *model 2*, excluding imputed RMR values). The black horizontal line indicates the slope for the reference level (primary vegetation) for minimal land-use intensity. The grey dashed line marks 0. Error bars are 95% confidence intervals. **(b) Effect of residual RMR on species probability of occurrence within each trophic level and for each land use-type from the model fitted using the empirical RMR values (i.e., *model 2*, excluding imputed RMR values).** I plotted the predictions for minimal land-use intensity only. Solid lines represent significant relationships. Primary: primary vegetation; secondary: secondary vegetation; plantation: plantation forest.

Bibliography for the Appendices

- Bates, D., Mächler, M., Bolker, B. M., and Walker, S. C. (2015). Fitting linear mixed-effects models using lme4. *Journal of Statistical Software*, 67, 1–48. DOI: [10.18637/jss.v067.i01](https://doi.org/10.18637/jss.v067.i01).
- Borges, R., Machado, J. P., Gomes, C., Rocha, A. P., and Antunes, A. (2018). Measuring phylogenetic signal between categorical traits and phylogenies. *Bioinformatics*, 35(11), 1862–1869. DOI: [10.1093/bioinformatics/bty800](https://doi.org/10.1093/bioinformatics/bty800).
- Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C. H., Xie, D., Suchard, M. A., Rambaut, A., and Drummond, A. J. (2014). BEAST 2: A Software Platform for Bayesian Evolutionary Analysis. *PLoS Computational Biology*, 10(4). DOI: [10.1371/journal.pcbi.1003537](https://doi.org/10.1371/journal.pcbi.1003537).
- Bruggeman, J., Heringa, J., and Brandt, B. W. (2009). PhyloPars: Estimation of missing parameter values using phylogeny. *Nucleic Acids Research*, 37(suppl2), W179–W184. DOI: [10.1093/nar/gkp370](https://doi.org/10.1093/nar/gkp370).
- Cadotte, M. W., Carscadden, K., and Mirochnick, N. (2011). Beyond species: Functional diversity and the maintenance of ecological processes and services. *Journal of Applied Ecology*, 48(5), 1079–1087. DOI: [10.1111/j.1365-2664.2011.02048.x](https://doi.org/10.1111/j.1365-2664.2011.02048.x).
- Cailliez, F. (1983). The analytical solution of the additive constant problem. *Psychometrika*, 48, 305–308. DOI: [10.1007/BF02294026](https://doi.org/10.1007/BF02294026).
- Chamberlain, S. (2018). *rredlist: ‘IUCN’ Red List Client*. R package version 0.5.0.
- Chamberlain, S. A. and Szöcs, E. (2013). taxize : taxonomic search and retrieval in R [version 2; referees: 3 approved]. *F1000Research*, 2(191). DOI: [10.12688/f1000research.2-191.v2](https://doi.org/10.12688/f1000research.2-191.v2).
- Debastiani, V. J., Bastazini, V. A. G., and Pillar, D (2021). Using phylogenetic information to impute missing functional trait values in ecological databases. *Ecological Informatics*, 63, 101315. DOI: [10.1016/j.ecoinf.2021.101315](https://doi.org/10.1016/j.ecoinf.2021.101315).
- Diniz-Filho, J. A. F., Bini, L. M., Rangel, T. F., Morales-Castilla, I., Olalla-Tárraga, M. Á., Rodríguez, M. Á., and Hawkins, B. A. (2012). On the selection of phylogenetic eigenvectors for ecological analyses. *Ecography*, 35, 239–249. DOI: [10.1111/j.1600-0587.2011.06949.x](https://doi.org/10.1111/j.1600-0587.2011.06949.x).
- Dormann, C. F., Elith, J., Bacher, S., Buchmann, C., Carl, G., Carré, G., Marquéz, J. R., Gruber, B., Lafourcade, B., Leitão, P. J., Münkemüller, T., McClean, C., Osborne, P. E., Reineking, B., Schröder, B., Skidmore, A. K., Zurell, D., and Lautenbach, S. (2013). Collinearity: A review of methods to deal with it and a simulation study evaluating their performance. *Ecography*, 36(1), 27–46. DOI: [10.1111/j.1600-0587.2012.07348.x](https://doi.org/10.1111/j.1600-0587.2012.07348.x).
- Dray, S., Dufour, A.-B., and Chessel, D. (2007). The ade4 Package – II: Two-Table and K-Table Methods. *R News*, 7(2), 47–52.

- Faurby, S., Davis, M., Pedersen, R., Schowanek, S. D., Antonelli, A., and Svenning, J.-C. (2018). PHYLACINE 1.2: The Phylogenetic Atlas of Mammal Macroecology. *Ecology*, 99(11), 2626–2626. DOI: 10.1002/ecy.2443.
- Faurby, S., Pedersen, R., Davis, M., Schowanek, S. D., Jarvie, S., Antonelli, A., and Svenning, J.-C. (2020). PHYLACINE 1.2.1: An update to the Phylogenetic Atlas of Mammal Macroecology. DOI: 10.5281/zenodo.3690867.
- Fox, J. and Monette, G. (1992). Generalized Collinearity Diagnostics. *Journal of the American Statistical Association*, 87(417), 178–183. DOI: 10.1080/01621459.1992.10475190.
- Fristoe, T. S., Burger, J. R., Balk, M. A., Khaliq, I., Hof, C., and Brown, J. H. (2015). Metabolic heat production and thermal conductance are mass-independent adaptations to thermal environment in birds and mammals. *Proceedings of the National Academy of Sciences of the United States of America*, 112(52), 15934–15939. DOI: 10.1073/pnas.152162112.
- Hadfield, J. D. (2010). MCMC Methods for Multi-Response Generalized Linear Mixed Models: The MCMCglmm R Package. *Journal of Statistical Software*, 33(2), 1–22. DOI: 10.18637/jss.v033.i02.
- Hartig, F. (2021). *DHARMA: Residual Diagnostics for Hierarchical (Multi-Level / Mixed) Regression Models*. R package version 0.4.1.
- Hudson, L. N. et al. (2014). The PREDICTS database: A global database of how local terrestrial biodiversity responds to human impacts. *Ecology and Evolution*, 4(24), 4701–4735. DOI: 10.1002/ece3.1303.
- (2017). The database of the PREDICTS (Projecting Responses of Ecological Diversity In Changing Terrestrial Systems) project. *Ecology and Evolution*, 7(1), 145–188. DOI: 10.1002/ece3.2579.
- IUCN (2020). The IUCN Red List of Threatened Species. Version 2020.1.
- Jetz, W., Thomas, G. H., Joy, J. B., Hartmann, K., and Mooers, A. O. (2012). The global diversity of birds in space and time. *Nature*, 491, 444–448. DOI: 10.1038/nature11631.
- Jetz, W. and Pyron, R. A. (2018). The interplay of past diversification and evolutionary isolation with present imperilment across the amphibian tree of life. *Nature Ecology and Evolution*, 2, 850–858. DOI: 10.1038/s41559-018-0515-5.
- Johnson, T. F., Isaac, N. J., Paviolo, A., and González-Suárez, M. (2021). Handling missing values in trait data. *Global Ecology and Biogeography*, 30(1), 51–62. DOI: 10.1111/geb.13185.
- Jones, K. E. et al. (2009). PanTHERIA: a species-level database of life history, ecology, and geography of extant and recently extinct mammals. *Ecology*, 90(9), 2648–2648. DOI: 10.1890/08-1494.1.
- Lüdecke, D., Ben-Shachar, M. S., Patil, I., Waggoner, P., and Makowski, D. (2021). performance: An R Package for Assessment, Comparison and Testing of Statistical Models. *Journal of Open Source Software*, 6(60), 3139. DOI: 10.21105/joss.03139.
- Leroy, B., Meynard, C. N., Bellard, C., and Courchamp, F. (2015). virtualspecies, an R package to generate virtual species distributions. *Ecography*, 39(6), 599–607. DOI: 10.1111/ecog.01388.
- Londoño, G. A., Chappell, M. A., Castañeda, M. d. R., Jankowski, J. E., and Robinson, S. K. (2015). Basal metabolism in tropical birds: Latitude, altitude, and the ‘pace of life’. *Functional Ecology*, 29(3), 338–346. DOI: 10.1111/1365-2435.12348.

- McNab, B. K. (2009). Ecological factors affect the level and scaling of avian BMR. *Comparative Biochemistry and Physiology - A Molecular and Integrative Physiology*, 152(1), 22–45. DOI: 10.1016/j.cbpa.2008.08.021.
- Oliveira, B. F., São-Pedro, V. A., Santos-Barrera, G., Penone, C., and Costa, G. C. (2017). AmphiBIO, a global database for amphibian ecological traits. *Scientific Data*, 4, 170123. DOI: 10.1038/sdata.2017.123.
- Pagel, M. (1999). Inferring the historical patterns of biological evolution. *Nature*, 401, 877–884. DOI: 10.1038/44766.
- Penone, C., Davidson, A. D., Shoemaker, K. T., Di Marco, M., Rondinini, C., Brooks, T. M., Young, B. E., Graham, C. H., and Costa, G. C. (2014). Imputation of missing data in life-history trait datasets: Which approach performs the best? *Methods in Ecology and Evolution*, 5(9), 961–970. DOI: 10.1111/2041-210X.12232.
- Rabosky, A. D., Cox, C., Rabosky, D., Title, P., Holmes, I., Feldman, A., and McGuire, J. (2016). Coral snakes predict the evolution of mimicry across New World snakes. *Nature Communications*, 7, 11484.
- Revell, L. J. (2012). phytools: An R package for phylogenetic comparative biology (and other things). *Methods in Ecology and Evolution*, 3(2), 217–223. DOI: 10.1111/j.2041-210X.2011.00169.x.
- Rinnan, D. S. and Lawler, J. (2019). Climate-niche factor analysis: a spatial approach to quantifying species vulnerability to climate change. *Ecography*, 42(9), 1494–1503. DOI: <https://doi.org/10.1111/ecog.03937>.
- Santini, L., Benítez-López, A., Ficetola, G. F., and Huijbregts, M. A. J. (2018). Length–mass allometries in amphibians. *Integrative Zoology*, 13(1), 36–45. DOI: 10.1111/1749-4877.12268.
- Santos, T. (2018). Package ‘PVR’. *Phylogenetic Eigenvectors Regression and Phylogenetic Signal-Representation Curve*. R package version 0.3.0.
- Stark, G., Pincheira-Donoso, D., and Meiri, S. (2020). No evidence for the ‘rate-of-living’ theory across the tetrapod tree of life. *Global Ecology and Biogeography*, 29(5), 857–884. DOI: 10.1111/geb.13069.
- Stekhoven, D. J. (2016). *Nonparametric Missing Value Imputation using Random Forest*. R package version 1.5. DOI: 10.1093/bioinformatics/btr597.
- Stekhoven, D. J. and Bühlmann, P. (2012). Missforest-Non-parametric missing value imputation for mixed-type data. *Bioinformatics*, 28(1), 112–118. DOI: 10.1093/bioinformatics/btr597.
- Tonini, J. F. R., Beard, K. H., Ferreira, R. B., Jetz, W., and Pyron, R. A. (2016). Fully-sampled phylogenies of squamates reveal evolutionary patterns in threat status. *Biological Conservation*, 204, Part A, 23–31. DOI: 10.1016/j.biocon.2016.03.039.
- Troyanskaya, O., Cantor, M., Sherlock, G., Brown, P., Hastie, T., Tibshirani, R., Botstein, D., and Altman, R. B. (2001). Missing value estimation methods for DNA microarrays. *Bioinformatics*, 17(6), 520–525. DOI: 10.1093/bioinformatics/17.6.520.
- van Buuren, S. and Groothuis-Oudshoorn, K. (2007). MICE: Multivariate Imputation by Chained Equations in R. *Journal of Statistical Software*, 45(3), 1–67. DOI: 10.18637/jss.v045.i03.
- Wilman, H., Belmaker, J., Simpson, J., de la Rosa, C., Rivadeneira, M. M., and Jetz, W. (2014). EltonTraits 1.0: Species-level foraging attributes of the world’s birds and mammals. *Ecology*, 95(7), 2027–2027. DOI: 10.1890/13-1917.1.