## **R** Basics

Laurent Gatto 1g390@cam.ac.uk

Computational Statistics for Genome Biology  $24^{th}$  June 2013

### Plan

#### Introduction

### Data types and structures

Basic data types Higher order objects

## Manipulating data

Subsetting

### **Useful functions**

**Plotting** 

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### Hello world

## [1] 2

## [1] 1

У

```
x <- 1  ## a variable
x
## [1] 1
x = 2  ## overwrite the value x
x</pre>
```

y <- length(x) ## calling a function

### **Getting help**

- Just ask!
- ► help.start() and the HTML help button in the Windows GUI.
- ▶ help and ?: help("data.frame") or ?help.
- help.search, apropos
- Online manuals and mailing lists
- ► Local R user groups

```
ls()
## [1] "x" "y"
rm(y)
ls()
## [1] "x"
```

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```
c(1, 3, 9, -1)
## [1] 1 3 9 -1
```

A vector contains an indexed set of values

- index starts at 1;
- all items are of the same storage mode;
- one of logical, numeric, complex or character,

numeric can futher be broken into integer, single and double types (only important when passing these to C or Fortran code, though).

```
mode(1)
## [1] "numeric"
typeof(1)
## [1] "double"
mode(1L)
## [1] "numeric"
typeof(1L)
## [1] "integer"
```

```
mode("1")
## [1] "character"
typeof("1")
## [1] "character"
mode (TRUE)
## [1] "logical"
typeof (FALSE)
## [1] "logical"
## as we are talking about booleans...
TRUE & TRUE
## [1] TRUE
```

The different modes an types can be retrieved and coerced with the is.\* and as.\* functions.

```
x <- 1
typeof(x)
```

```
## [1] "double"
```

y <- as.integer(x)

## [1] "integer"

is.integer(y)

## [1] TRUE

typeof(y)

## **Special values**

```
NULL
NA
NaN
Inf
-Inf
is.null()
is.na()
is.infinite()
```

What are the mode and types of these?

All these are objects with a certain class.

```
class(x)
```

## [1] "numeric"

class("a character")

## [1] "character"

## Creating vectors with functions

numeric(4)

## [1] 0 0 0 0

```
vector(mode = "character", length = 3)
## [1] "" ""
vector(mode = "numeric", length = 4)
## [1] 0 0 0 0
```

Creating vectors with functions (2)

```
x \leftarrow c(1, 4, 7, 10) ## concatenate
X
## [1] 1 4 7 10
y <- 1:5 ## integer sequence
У
```

 $z \leftarrow seq(from = 1, to = 10, by = 2)$ 

## [1] 1 2 3 4 5

## [1] 1 3 5 7 9

Z

## Arguments by position or name

```
z1 <- seq(from = 1, to = 10, by = 2)
z2 <- seq(1, 10, 2)
```

## [1] TRUE TRUE TRUE TRUE TRUE

z1 == z2

all(z1 == z2)

## [1] TRUE

## [1] TRUE

identical(z1, z2)

# **Vectorised** arithmetic

```
x <- 1:5
y <- 5:1
X
## [1] 1 2 3 4 5
## [1] 5 4 3 2 1
x + y
## [1] 6 6 6 6 6
x^2
```

## [1] 1 4 9 16 25

### **Matrices**

are 2-dimensional vectors

```
m \leftarrow matrix(1:12, nrow = 4, ncol = 3)
m
## [,1] [,2] [,3]
## [1,] 1 5 9
## [2,] 2 6 10
## [3,] 3 7 11
## [4,] 4 8 12
dim(m)
## [1] 4 3
```

What if I don't get the data or dimensions right?

```
What if I don't get the data or dimensions right?
matrix(1:11, 4, 3) ## recycling
## Warning: data length [11] is not a sub-multiple
or multiple of the number of rows [4]
## [,1] [,2] [,3]
## [1,] 1 5 9
## [2,] 2 6 10
```

```
## [3,] 3 7 11
## [4,] 4 8 1
```

matrix(1:12, 3, 3) ## [,1] [,2] [,3]

## [1,] 1 4 7 **##** [2,] 2 5 8

## [3,] 3 6 9

```
x <- 1:12
class(x)
## [1] "integer"
dim(x)
## NULL
dim(x) < -c(4, 3)
X
## [,1] [,2] [,3]
## [1,] 1 5 9
## [2,] 2 6 10
## [3,] 3 7 11
## [4,] 4 8 12
class(x)
## [1] "matrix"
```

### **Arrays**

are n-dimensional vectors

## [2,] 10 12 14 16

```
array(1:16, dim = c(2, 4, 2))
## , , 1
##
## [,1] [,2] [,3] [,4]
## [1,] 1 3 5 7
## [2,] 2 4 6 8
##
## , , 2
##
## [,1] [,2] [,3] [,4]
## [1,] 9 11 13 15
```

### Lists

are ordered collections of elements that can be arbitrary R objects.

```
(11 \leftarrow list(a = 1:3, f = length))
## $a
## [1] 1 2 3
##
## $f
## function (x) .Primitive("length")
ll[1] ## a list of length 1
## $a
## [1] 1 2 3
ll[[1]] ## or ll$a - first element
## [1] 1 2 3
```

```
11
## $a
## [1] 1 2 3
##
## $f
## function (x) .Primitive("length")
11$f(11)
## [1] 2
```

### **Data.frames**

## 4 ctrl 1.9970

are 2-dimensional lists (with elements of same length!).

```
dfr <- data.frame(type = c(</pre>
                    rep("case", 2),
                    rep("ctrl", 2)),
                  time = rnorm(4)
dfr
## type time
## 1 case -0.9144
## 2 case -0.4354
## 3 ctrl -0.3966
```

```
dfr[1, ]
## type time
## 1 case -0.9144
dfr[1, "time"]
## [1] -0.9144
dfr$time
## [1] -0.9144 -0.4354 -0.3966 1.9970
```

## **Environments**

are unordered sets of objects.

```
e <- new.env()
e[["a"]] <- 1:3
assign("b", "CSAMA", envir = e)
ls(e)
## [1] "a" "b"
```

e\$a

## [1] 1 2 3

get("b", e)

## [1] "CSAMA"

#### Names

We have seen that function arguments have names, and named our data.frame columns. We can also name matrix/data.frame columns and rows, dimensions, and vector items.

```
x <- c(a = 1, b = 2)
x

## a b
## 1 2
names(x)
## [1] "a" "b"</pre>
```

2005 4 6

## 2006 8 4 5 ## 2007 5 2 7

##

#### **Factors**

for categorical data

```
## [1] Control Case Control Case Case
## [6] Control Case Case Case Control
## [11] Case Control Case Case Case
## [16] Control Case Control Case Case
## [21] Control Control Control Control Case
## [26] Case
## Levels: Case Control
```

## Data types summary

	1 dim	2 dims	n dims
same type		matrix	array
diff types	list	$\mathtt{data.frame}^*$	list

(\*elements of same length)

### Higher order objects

When the data to be stored is more complex, special objects are created to store and handle it in a specialised manner. These higher order objects are constructed using the data types we have seen so far as building blocks.

Let's look at how microarray data is handled in Bioconductor.

The eSet model has been re-used for other technologies.

```
library(Biobase)
data(sample.ExpressionSet)
sample.ExpressionSet
## ExpressionSet (storageMode: lockedEnvironment)
## assayData: 500 features, 26 samples
##
     element names: exprs, se.exprs
## protocolData: none
## phenoData
     sampleNames: A B ... Z (26 total)
##
## varLabels: sex type score
## varMetadata: labelDescription
## featureData: none
## experimentData: use 'experimentData(object)'
```

## Annotation: hgu95av2

```
class(sample.ExpressionSet)
## [1] "ExpressionSet"
## attr(,"package")
## [1] "Biobase"
slotNames(sample.ExpressionSet)
## [1] "experimentData"
                            "assayData"
                            "featureData"
## [3] "phenoData"
## [5] "annotation"
                            "protocolData"
```

## [7] ".\_\_classVersion\_\_"

?ExpressionSet

assayData expression values in identical sized matrices.
phenoData sample annotation in AnnotatedDataFrame.

featureData feature annotation in AnnotatedDataFrame.

annotation type of chip as a character.

protocolData scan dates as a character.



sample meta-data

### The assayData slot

## Features Samples 500

##

26

Stored the expression data of the assay.

```
exprs(sample.ExpressionSet)[1:4, 1:3]
##
                       Α
                              В
## AFFX-MurIL2_at 192.74 85.753 176.76
## AFFX-MurIL10_at 97.14 126.196 77.92
## AFFX-MurIL4_at 45.82 8.831 33.06
## AFFX-MurFAS_at 22.54 3.601 14.69
dim(sample.ExpressionSet)
```

### The phenoData slot

stores the meta data about the samples.

## varMetadata: labelDescription

```
phenoData(sample.ExpressionSet)

## An object of class 'AnnotatedDataFrame'

## sampleNames: A B ... Z (26 total)

## varLabels: sex type score
```

### The featureData slot

## NULL

stores the meta data about the feautres.

```
featureData(sample.ExpressionSet)
```

- -

## An object of class 'AnnotatedDataFrame': none

#### AnnotatedDataFrame

consists of a collection of samples and the values of variables measured on those samples. There is also a description of each variable measured. AnnotatedDataFrame associates a data.frame with its metadata.

```
head(pData(sample.ExpressionSet))

## sex type score
## A Female Control 0.75

## B Male Case 0.40

## C Male Control 0.73

## D Male Case 0.42

## E Female Case 0.93

## F Male Control 0.22
```

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# Manipulating data Subsetting

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- ► One of the most powerful features of R is its ability to manipulate subsets of vectors and arrays.
- ► As seen, subsetting is done with, [], [, ], ...

# Subsetting with positive indices

```
x < -1:10
x[3:7]
## [1] 3 4 5 6 7
x[9:11]
## [1] 9 10 NA
x[0:1]
```

x[c(1, 7, 2, NA)]
## [1] 1 7 2 NA

## [1] 1

### Assignments with positive indices

```
x[2] <- 20
x[4:5] <- x[4:5] * 100
## x[1:6] ?
```

# Assignments with positive indices

**##** [1] 1 20 3 400 500 6

```
x[2] <- 20
x[4:5] <- x[4:5] * 100
## x[1:6] ?
```

```
## x[1:6] ?

x[1:6]
```

### Subsetting with negative indices

```
x <- 1:10
## x[-c(3:7)] ?
```

# Subsetting with negative indices

```
x <- 1:10
## x[-c(3:7)] ?
```

```
## [1] 1 2 8 9 10
```

x[-c(3:7)]

# Subsetting with logical predicates

x[x > 5]

## [1] 6 7 8 9 10

```
x[c(TRUE, TRUE, rep(FALSE, 8))]
## [1] 1 2
x > 5
## [1] FALSE FALSE FALSE FALSE TRUE TRUE
## [8] TRUE TRUE TRUE
```

# Subsetting with logical predicates

```
## x[c(TRUE, FALSE)] ?
```

# Subsetting with logical predicates

```
## x[c(TRUE, FALSE)] ?
```

```
x[c(TRUE, FALSE)] ## recycled
```

## [1] 1 3 5 7 9

# Subsetting by names

x[c("a", "d")]

## a <NA> ## 1 NA

```
x \leftarrow c(a = 1, b = 2, c = 2)
x[c("a", "c")]
## a c
## 1 2
```

# Subsetting matrices

**##** [3,] 3 6 9 12

```
M <- matrix(1:12, 3)
M[1, ] ## row -> vector (or drop = FALSE)
## [1] 1 4 7 10
M[, 1] ## column -> vector (or drop = FALSE)
## [1] 1 2 3
M[2, 3] < -0
M
## [,1] [,2] [,3] [,4]
## [1,] 1 4 7 10
## [2,] 2 5 0 11
```

# Subsetting matrices (2)

## [2,] -1 -1 -1 11 ## [3,] -1 -1 9 12

```
M < 9
## [,1] [,2] [,3] [,4]
## [1,] TRUE TRUE TRUE FALSE
## [2,] TRUE TRUE TRUE FALSE
## [3,] TRUE TRUE FALSE FALSE
M[M < 9] < -1
Μ
## [,1] [,2] [,3] [,4]
## [1,] -1 -1 10
```

# **Subsetting lists**

## [1] 1 2 3

```
11 <- list(a = 1:3, b = "CSAMA", c = length)
11[1] ## still a list, but of length 1
## $a
## [1] 1 2 3</pre>
```

ll[[1]] ## first element of the list

### Subsetting ExpressionSet instances

It is reasonable to expect that subsetting operations work also for higher order objects.

```
sample.ExpressionSet[1:10, 1:2]
## ExpressionSet (storageMode: lockedEnvironment)
## assayData: 10 features, 2 samples
##
     element names: exprs, se.exprs
## protocolData: none
## phenoData
##
     sampleNames: A B
## varLabels: sex type score
## varMetadata: labelDescription
## featureData: none
## experimentData: use 'experimentData(object)'
## Annotation: hgu95av2
```

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#### Data 10

read.table creates a data.frame from a spreadsheet file.
write.table writes a data.frame/matrix to a spreadsheet (tsv,

csv).

**save** writes an binary representation of R objects to a file (cross-platform).

load load a binary R file from disk.

Specialised data formats often have specific i/o functionality (microarray CEL files, XML, . . . )

```
read.table("./Data/data.csv", sep = ",",
          header = TRUE, row.names = 1)
##
           A B C D
## 1 -0.15330 10 x 10
## 2 -0.13868 3 n 9
## 3 -0.43323 2 f 8
## 4 1.64569 4 o 7
## 5 0.23381 6 b 6
## 6 0.98770 9 m 5
## 7 -0.25565 7 c 4
## 8 -0.74719 1 1 3
## 9 -0.02001 5 e 2
## 10 -0.95000 8 v 1
```

```
read.csv("./Data/data.csv", row.names = 1)
##
           A B C D
## 1 -0.15330 10 x 10
## 2 -0.13868 3 n 9
## 3 -0.43323 2 f 8
## 4 1.64569 4 o 7
## 5 0.23381 6 b 6
## 6 0.98770 9 m 5
## 7 -0.25565 7 c 4
## 8 -0.74719 1 1 3
```

## 9 -0.02001 5 e 2 ## 10 -0.95000 8 v 1

```
x <- read.csv("./Data/data.csv", row.names = 1)
save(x, file = "./Data/data.rda")
rm(x)
load("./Data/data.rda")
x[1:3, ]
## A B C D
## 1 -0.1533 10 x 10</pre>
```

## 2 -0.1387 3 n 9 ## 3 -0.4332 2 f 8

# String manipulation (1)

## [1] "abcdef"

```
paste("abc", "def", sep = "-")
## [1] "abc-def"
paste0("abc", "def")
```

# String manipulation (2)

```
month.name[1:4]
## [1] "January" "February" "March" "April"
grep("Feb", month.name)
## [1] 2
grep("Feb", month.name, value = TRUE)
## [1] "February"
grepl("Feb", month.name)
## [1] FALSE TRUE FALSE FALSE FALSE FALSE
## [8] FALSE FALSE FALSE FALSE
```

# String manipulation (3)

```
month.name[1]
```

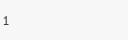
## [1] "January"

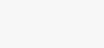
length(month.name[1])

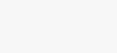
nchar(month.name[1])

## [1] 7

## [1] 1













# String manipulation (4)

```
strsplit("abc-def", "-")
## [[1]]
## [1] "abc" "def"
```

# String manipulation (4)

## [1] "ghi" "jkl"

```
strsplit("abc-def", "-")
## [[1]]
## [1] "abc" "def"
strsplit(c("abc-def", "ghi-jkl"), "-")
## [[1]]
## [1] "abc" "def"
##
## [[2]]
```

# Comparing and matching (1)

## [1] "j" "f" "i" "a" "b" "g"

```
set.seed(1)
x <- sample(letters[1:10], 6)
y <- sample(letters[1:10], 6)
x
## [1] "c" "d" "e" "g" "b" "h"</pre>
y
```

# Comparing and matching (2)

union(x, y)

```
intersect(x, y)
## [1] "g" "b"
setdiff(x, y)
## [1] "c" "d" "e" "h"
```

## [1] "c" "d" "e" "g" "b" "h" "j" "f" "i" "a"

# Comparing and matching (3)

## [1] NA NA NA 6 5 NA

match(x, y)

```
x %in% y
## [1] FALSE FALSE FALSE TRUE TRUE FALSE
```

```
x == y
## [1] FALSE FALSE FALSE FALSE TRUE FALSE
```

# **Generating data (1)**

```
seq(1, 7, 3)
```

## [1] 1 4 7

rep(1:2, 2)

## [1] 1 1 2 2



rep(1:2, each = 2)

















# **Generating data (2)**

```
runif(5)
## [1] 0.6870 0.3841 0.7698 0.4977 0.7176
```

rnorm(5)

## [1] 2.4047 0.7636 -0.7990 -1.1477 -0.2895

#### About the data

```
table(sample(letters, 100, replace = TRUE))
##
## a b c d e f g h i j k l m n o p q r s t u v w x y
## 2 2 4 4 2 2 4 2 6 4 5 7 9 3 1 3 5 3 5 5 6 4 5 2 2
## z
## 3
summary(rnorm(100))
## Min. 1st Qu. Median Mean 3rd Qu. Max.
## -1.6800 -0.8280 -0.0081 -0.0089 0.6090 2.6600
head(x)
## [1] "c" "d" "e" "g" "b" "h"
tail(x)
## [1] "c" "d" "e" "g" "b" "h"
```

```
M \leftarrow matrix(rnorm(1000), ncol = 4)
head(M)
## [,1] [,2] [,3] [,4]
## [1,] 0.7796 -0.3399 -1.44689 -0.1658
## [2,] 0.7132 0.6063 1.01951 0.5571
## [3,] -0.5429 1.3411 1.17855 1.4443
## [4.] 0.8858 0.7673 -0.01026 0.9014
## [5.] -0.3486 0.1937 0.26862 -0.2220
## [6,] -1.0081 1.1406 1.34203 0.1062
```

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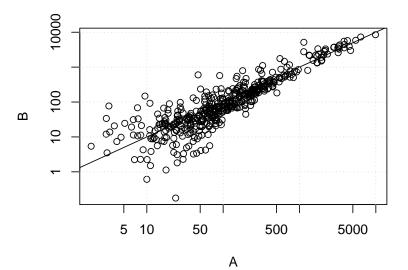
# **Plotting**

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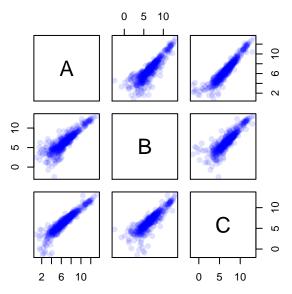
- ► Scatterplots with plot
- ► Boxplots with boxplot
- ► Barplots with barplot
- ► Histograms with hist
  - smoothScatter

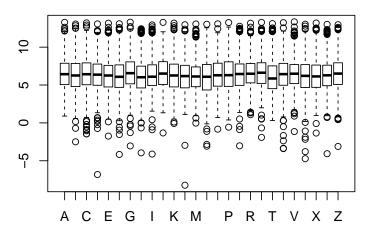
```
plot(exprs(sample.ExpressionSet[, 1]),
        exprs(sample.ExpressionSet[, 2]),
        log = "xy",
        xlab = sampleNames(sample.ExpressionSet)[1],
        ylab = sampleNames(sample.ExpressionSet)[2])
abline(0, 1)
grid()
```

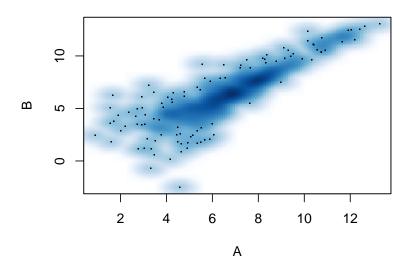


To create subplots, one can use par(mfrow = c(2,2)), layout, or (for scatterplots)

```
pairs(log2(exprs(sample.ExpressionSet)[, 1:4]),
    pch = 19,
    col = "#0000FF20")
```







We have not covered lattice and ggplot2.

#### References

- http://gallery.r-enthusiasts.com/allgraph.php
- R Graphics manual: http://rgm3.lab.nig.ac.jp/RGM/r\_image\_list
- ▶ http://www.cookbook-r.com/Graphs/ (ggplot2)
- ggplot2: Elegant Graphics for Data Analysis, Hadley Wickham (2009)
- ► Lattice: Multivariate Data Visualization with R, Deepayan Sarkar (2008)

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#### Flow control

for (var in seq) expr
while (cond) expr
repeat expr break

```
for (i in 1:4) {
    ## bad
   print(i^2)
## [1] 1
## [1] 4
## [1] 9
## [1] 16
(1:4)^2 ## good
## [1] 1 4 9 16
```

### The apply family and friends

- ▶ Applies a function to each element of an input, being a list or a vector (sapply, lapply), a matrix or a data frame (apply) or an environment (eapply).
- Same functionality than an explicit for loop, but often more elegant, function-centric, not faster.

```
sapply(month.name[1:2], paste0, "_2012")
##
           January February
   "January_2012" "February_2012"
##
lapply(month.name[1:2], paste0, "_2012")
## [[1]]
## [1] "January_2012"
##
## [[2]]
## [1] "February_2012"
```

```
M \leftarrow matrix(1:9, ncol = 3)
М
## [,1] [,2] [,3]
## [1,] 1 4 7
## [2,] 2 5 8
## [3,] 3 6 9
apply(M, 1, sum) ## better rowSums
## [1] 12 15 18
apply(M, 2, sum) ## better colSums
## [1] 6 15 24
## [1] 2
```

```
mean(rnorm(100))
## [1] 0.05783
replicate(3, mean(rnorm(100)))
## [1] -0.01148 -0.19392 0.07020
replicate(2, rnorm(3))
## [,1] [,2]
## [1,] 1.1553 0.9439
## [2,] 1.5614 -1.8573
## [3,] 0.3231 -0.5995
```

### **Conditionals**

```
if (cond) expr1 else expr2
ifelse(cond, expr1, expr2)
switch
```

```
x < -2
if (x > 0) {
   ## bad
   log2(x)
} else {
   log2(-x)
## [1] 1
log2(abs(x)) ## better
## [1] 1
```

## **Exception handling**

try(exprs) will either return the value of the expression expr, or an object of class try-error.

tryCatch provides a more configurable mechanism for condition handling and error recovery.

## **Writing functions**

```
myFun <- function(param1, param2, ...) {
    ## function body
    ## acting on copies of the params
    ans <- param1 + param2
    return(ans)</pre>
```

#### **Function facts**

- ► Single return value.
- ► To return multiple items, use a list or a proper object (see OO programming).
- ► The return value is either the last statement, or explicit return using return (can be called from any where in a function)

# **Function facts (cont.)**

## [1] 1

```
► Functions act on a pass-by-copy semantic.
 x <- 1
  f <- function(x) {</pre>
    x < -x + 10
     X
  f(x)
  ## [1] 11
  X
```

# **Function facts (cont.)**

► Functions live/act in their own environment and have access to *global* variables.

```
x <- 1
f <- function() {</pre>
    x < -x + 10
    X
f()
## [1] 11
## [1] 1
```

### Plan

### Introduction

# **Data types and structures**

Basic data types
Higher order objects

# Manipulating data

Subsetting

**Useful functions** 

**Plotting** 

**Programming** 

**Packages** 

- ▶ Primary mechanism to distribute R software is via packages.
- Packages are installed in <u>libraries</u> (directories) on your had disk, and they are loaded with the library function.
- There are software, data and annotation packages.
- ► The Comprehensive R Archive Network (CRAN) is the main package repository. It provides an automatic build framework for package authors.
- ► The Bioconductor project manages its own CRAN-style repository.
- R-forge https://r-forge.r-project.org/

### Package installation

- ► From within R, using install.packages takes care of dependencies.
- ▶ Update all installed packages with update.packages.
- ► For Bioconductor packages, use biocLite:

```
source("http://www.bioconductor.org/biocLite.R")
## or, if you have already done so in the past
library("BiocInstaller")
biocLite("packageName")
```

### Getting information about packages

- ► CRAN/Bioconductor/R-forge web pages
- Documentation

```
help(package = "Biobase")
```

Vignettes (mandatory for Bioconductor packages)

```
vignette(package = "Biobase")
```

```
vignette("Bioconductor", package = "Biobase")
```

Demos

```
demo("lattice", package = "lattice")
```

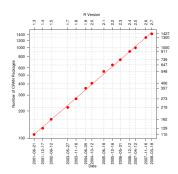
```
packageDescription("Biobase")
## Package: Biobase
## Title: Biobase: Base functions for Bioconductor
## Version: 2.21.4
## Author: R. Gentleman, V. Carey, M. Morgan, S. Falcon
## Description: Functions that are needed by many other packages or
          which replace R functions.
##
## Suggests: tools, tkWidgets, ALL
## Depends: R (>= 2.10), BiocGenerics (>= 0.3.2), utils
## Imports: methods, BiocGenerics
## Maintainer: Bioconductor Package Maintainer
          <maintainer@bioconductor.org>
##
## License: Artistic-2.0
## Collate: tools.R strings.R environment.R vignettes.R packages.R
##
## LazyLoad: yes
## biocViews: Infrastructure, Bioinformatics
## Packaged: 2013-06-11 02:32:03 UTC; biocbuild
## Built: R 3.1.0; x86_64-unknown-linux-gnu; 2013-06-18 00:33:27 UTC;
##
          unix
##
## -- File: /home/lgatto/R/x86_64-unknown-linux-gnu-library/3.1/Biobase/Meta/pa
```

## **Package versions**

- New Bioconductor releases appear twice a year. Bioconductor versions are tied to ℝ versions.
- $\blacktriangleright$  Stable packages versions are x.y.z where x >= 1 and y is even
- Devel packages have y odd.

Bioconductor 636 reviewed packages
CRAN 3889 packages
R-forge 1313 projects

(19<sup>th</sup> June 2012)



## Finding packages

- ▶ BiocViews http://bioconductor.org/packages/ release/BiocViews.html.
- CRAN Task Views http://cran.r-project.org/web/views/.
- ▶ sos to search inside contributed R packages.

### toLatex(sessionInfo())

- ► R Under development (unstable) (2013-06-16 r62969), x86\_64-unknown-linux-gnu
- ► Locale: LC\_CTYPE=en\_GB.UTF-8, LC\_NUMERIC=C, LC\_TIME=en\_GB.UTF-8, LC\_COLLATE=en\_GB.UTF-8, LC\_MONETARY=en\_GB.UTF-8, LC\_MESSAGES=en\_GB.UTF-8, LC\_PAPER=C, LC\_NAME=C, LC\_ADDRESS=C, LC\_TELEPHONE=C, LC\_MEASUREMENT=en\_GB.UTF-8, LC\_IDENTIFICATION=C
- ► Base packages: base, datasets, graphics, grDevices, methods, parallel, stats, utils
- ▶ Other packages: Biobase 2.21.4, BiocGenerics 0.7.2, knitr 1.2
- ► Loaded via a namespace (and not attached): digest 0.6.3, evaluate 0.4.3, formatR 0.7, stringr 0.6.2, tools 3.1.0

#### References

- ► W. N. Venables, D. M. Smith and the R Development Core Team, An Introduction to R (get it with help.start())
- ▶ R. Gentleman, R Programming for Bioinformatics, CRC Press, 2008

- ▶ This work is licensed under a CC BY-SA 3.0 License.
- Course web page and more material: https://github.com/lgatto/TeachingMaterial

Thank you for your attention