

# KGD: Software for GBS-based relationship calculations

## v0.9.2

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## Background

R code is available for the analysis of genotyping-by-sequencing (GBS) data, primarily to construct a genomic relationship matrix ('G matrix') for the genotyped individuals, with a focus on the KGD (Kinship using GBS with Depth adjustment) method of Dodds *et al.* (2015). The code can be used

on its own or incorporated into other R programs. There are QC tools (primarily graphical output), relationship estimation tools, pedigree verification tools and pedigree 'mix and match' tools. The latter two operations require additional input information about the samples genotyped. There are also tools implementing some methods for population genetics that are currently under development.

In this document, 'Individual' or 'sample' generally refers to the genotyping unit (possibly combined, if the same individual or sample is genotyped multiple times). Familial relationships are given the labels 'Father', 'Mother' and 'Offspring' (as appropriate).

Many of the methods used are as described in Dodds *et al.* (2015), Bilton *et al.* (2019) and Dodds *et al.* (2019). Unless specified otherwise, relatedness estimates in this documentation refer to those using the 'G5' method of Dodds *et al.* (2015).

## Program structure

There are separate analysis program files, the first (GBS-Chip-Gmatrix.R) is for genotype QC and relationship matrix construction while there are also programs for pedigree verification and/or assignment, using the related estimates (GBSPedAssign.R) and for population genetics analyses (GBS-PopGen.R). These programs can be invoked from another program file (using the *source* command), or users can insert all or parts of these programs into their own code. For the purposes of this documentation, it is assumed the first method is used, with calling program named GBSRun.R. If any of these programs are being called multiple times in an R session, care should be taken to make sure any default values still apply for the subsequent call(s), otherwise they should be removed (rm) or set specifically.

## Calling program (GBSRun.R)

Variables that can be set or commands that can be run using the main program (i.e., QC and relatedness estimation) are shown in the following table. See other sections (pedigree analysis, population genetics) for details on variables and commands relevant to those analysis.

Variable command	Type <sup>1</sup>	Description
<b>genofile</b>	V	Name (including path) of the genotype file. Default value is "HapMap.hmc.txt".
<b>gform</b>	V	Type of genotype file. Default is "uneak"; other options are "Tassel", "TagDigger", "ANGSDcounts" or "Chip".
<b>sampdepth.thresh</b>	V	Minimum mean sample depth for retaining sample results. Default is 0.01.
<b>snpdepth.thresh</b>	V	Minimum mean SNP depth for retaining SNPs. Default is 0.01.
<b>hirel.thresh</b>	V	Lower threshold for reporting highly related individuals, and upper threshold for displaying positive control pairs which don't seem sufficiently related. Default is 0.9.
<b>triallelic.thresh</b>	V	Upper threshold for the proportion of ignored reads for the third allele – SNPs with a higher proportion are removed (as a triallelic variant). Relevant only to the ANGSDcounts input format. Default is 0.005 (0.5%).
<b>cex.pointsize</b>	V	Relative value of pointsize used in output graphics. This has a default value of 1.
<b>functions.only</b>	V	Set to TRUE to source GBS-Chip-Gmatrix.R for setting up functions (not reading data etc). Default is FALSE.
<b>alleles.keep</b>	V	Set to TRUE to retain an updated version of alleles. This object is needed for some downstream uses, e.g. for writing VCF files

		or for the calculation of linkage disequilibrium (using GUS-LD). Default is FALSE.
<b>outlevel</b>	V	Integer (1-9) determining the level of output created – higher numbers give more output. At present only two levels are active; 5 to 9 give the full output while 1 to 4 gives less output. A value less than 8 will suppress the sampled alleles setup and analysis (reducing time). Default is 9 (all available output)
<b>use.Rcpp</b>	V	Set to FALSE to prevent the C++ versions of functions being used. Default is TRUE.
<b>nThreads</b>	V	The number of OpenMP threads to be used by C++. The default is 4. Using 0 means all available threads would be used.
<b>iemm.thresh</b>	V	Identity excess mismatch rate threshold for displaying non-matching results putatively from the same individual. Currently only used in the posC-EMM plot.
<b>negC</b>	V	character string containing a regular expression to be matched to <i>seqID</i> to identify negative controls. The default is an empty string, in which case no checking is done. See below for more details.
<b>negCsettings</b>	V	a list of qualifiers to be passed to R's <i>grep</i> function when pattern matching for <i>negC</i> in <i>seqID</i> . The default is an empty list. See below for more details.
<b>QQprobpts</b>	V	numeric vector of probability points for plotting reference lines on QQ plots. The default is <code>c(0.5,0.8,0.9,0.95,0.99)</code> . Set to <code>numeric(0)</code> to remove the lines. A new setting will be used in any subsequent <code>GBSsummary()</code> call.
<b>source</b>	C	Invoke GBS-Chip-Gmatrix.R code, to run QC procedures and define functions, e.g. the genomic relationship matrix function ( <i>calcG</i> )
<b>calcG</b>	C	Calculate genomic relationship matrices. May be invoked several times with different options.

<sup>1</sup> Type is V for a variable to be set, or C for a command to be invoked or function to be run.

## Relatedness estimation program (GBS-Chip-Gmatrix.R)

This program performs some QC diagnostics, rudimentary data cleaning and defining a function (*calcG*) for relatedness estimation and reporting. A number of other functions are defined, such as those for checking and report on positive controls. Any procedures or output relating to depth are not implemented for chip data. The use of depth information to construct the GRM can be modified (see *depth2K* section).

If there are negative controls specified and found, they are summarised (normal output), reported (in *negCStats.csv*) and removed from the data before further analysis. The negative controls are defined by matching the text *negC* in *seqID*. For example, if *negC* is "NEG" then any *seqID* containing the string "NEG" is treated as a negative control. The *grep* function with usual default is used for pattern matching so that *negC* is used as a regular expression. This can be modified by setting *grep* arguments in *negCsettings*. For example,

```
negCsettings <- list(fixed=TRUE)
```

will match *negC* as is to *seqID*. i.e. does not treat it as a regular expression. Negative control checks are not invoked for chip data.

Samples with very low depth are dropped from the analyses. The threshold is a mean depth of *sampdepth.thresh* (default of 0.01, but can be set in the calling program) or with a maximum depth of one (including those with no genotype calls). Samples that are dropped are reported in the program output and are available in *seqID.removed*. The remaining number of samples is also reported in the output.

SNPs with no data or with a MAF (minor allele frequency) of zero are dropped. The remaining number of SNPs is reported.

Some basic statistics are reported: Proportion of missing genotypes is the number of SNP x individual combinations with no allele calls; Mean sample depth is the average depth (number of reads of either allele) for a sample.

The default action when sourcing GBS-Chip-Gmatrix.R is to read the data file and run some QC procedures, as well as define various functions. If `functions.only` is set to TRUE, then only the function definition occurs. The default action can then be mimicked using the pair of commands:

```
readGBS()  
GBSSummary()
```

These functions are not yet described in the documentation. Additional processing can be inserted between these statements, for example to manually remove samples or SNPs. The following objects need to be maintained correctly, before *GBSSummary* is run: *nsnps*, *SNP\_Names*, *seqID*, *nind*, *alleles*. If *GBSSummary* has been run once, it could be re-run, e.g. after merging results from the same individual. In that case (detected by the presence of *depth*), processing that uses *alleles* (which is not recalculated in *mergeSamples* unless *keep.alleles* is set to TRUE) is omitted. This means that *depth* needs to correspond to actual depth (may need to replace with values in *depth.orig*). Alternatively, if *alleles* is present and corresponds to the current data, then *depth* could be removed so that it gets recalculated. This is mainly to obtain *genon* and *depth* which will be assumed to be present and correct, but it should be noted that *p* is not recalculated. *p* should remain unchanged when samples are merged, but could change, for example if the *sampdepth.thresh* is changed between calls to *GBSSummary*.

Some functions have been coded in C++ to improve speed. These will be used instead of the corresponding R functions if the libraries Rcpp and RcppArmadillo are installed, and `use.Rcpp` is TRUE (the default value).

### Output - files

*negCStats.csv* contains call rates, along with mean sample depths for each sample identified as a negative control.

*SampleStats.csv* contains call rates for each sample, along with mean sample depths (for GBS data).

*AlleleFreq.png* is a plot of allele frequencies calculated using different methods (and as given, if the uneak format is used).

*CallRate.png* shows a histogram of sample call rates (proportion of SNPs with a result for a sample).

*SampDepth.png* plots mean sample depth against median sample depth.

*SampDepth-scored.png* plots mean sample depth, over SNPs that are scored for the individual, against mean sample depth over all SNPs for the individual.

*SampDepthHist.png* is a histogram of mean sample depths

*SampDepthCR.png* plots mean sample depth against call rate.

*SNPDepthHist.png* is a histogram of SNP depths (number of reads of either allele averaged over samples)

*SNPCallRate.png* is a histogram of SNP call rates (proportion of samples with a result for a SNP)

*SNPDepth.png* plots SNP depth against mean SNP depth (on a log scale). This may reveal SNPs that are called infrequently, but when they are called have good depth (these SNPs may be near the boundary of a size selection step in the laboratory).

*finplot.png* plots Hardy-Weinberg disequilibrium (HWD) against MAF, shaded by the SNP depth. HWD is the proportion of (reference allele) homozygotes minus the expected proportion (under Hardy-Weinberg equilibrium). HWD is the same whichever allele is used in the calculation. The 'fin plot' may reveal sets of SNPs that do not follow Mendelian inheritance, for example apparent SNPs in duplicated regions.

*HWdisMAFsig.png* is similar to the fin plot, but with shading by *l10pstar*, the  $\log_{10}$  p-value corresponding to the depth-adjusted chi-squared test statistic of Hardy Weinberg equilibrium (versions prior to v0.702 used the likelihood ratio test statistic for HWD).

*LRT-QQ.png* is a QQ plot for the likelihood ratio test statistic for HWD. Grey lines connect the x and y axis values corresponding to the cumulative proportions given in *QQprobpts*.

*LRT-hist.png* is a histogram of the likelihood ratio test statistic for HWD.

*X2star-QQ.png* is a QQ plot for the depth-adjusted chi-square test statistic for HWD. To allow comparison with values shown in *HWdisMAFsig.png*,  $l10pstar \approx 0.77 + 0.218 * x2star$ . Grey lines connect the x and y axis values corresponding to the cumulative proportions given in *QQprobpts*. The right-hand side vertical axis shows the  $-\log_{10}$  probabilities for the depth-adjusted chi-square test statistic (*x2star*) (the variable used for colour shading in *HWdisMAFsig.png*).

*MAF.png* is a histogram of the MAFs for each SNP (based on observed genotypes).

## Variables defined

These include:

Variable	Description
<b><i>nind</i></b>	Number of samples analysed (after initial QC)
<b><i>nsnps</i></b>	Number of SNPs analysed (after initial QC)
<b><i>seqID</i></b>	Identifiers for each sample
<b><i>seqID.removed</i></b>	Identifiers for samples removed during initial QC.
<b><i>SNP_Name</i></b>	Identifiers for each SNP
<b><i>chrom</i></b>	chromosome label (character), if <i>gform</i> is Tassel
<b><i>pos</i></b>	chromosome position (numeric), if <i>gform</i> is Tassel
<b><i>alleles</i></b>	matrix ( <i>nind</i> x $2 * nsnps$ ) of read counts. The results for each SNP are in consecutive columns.
<b><i>genon</i></b>	matrix ( <i>nind</i> x <i>nsnps</i> ) of numeric genotype calls 0 (homozygous alternate allele), 1 (heterozygous), 2 (homozygous reference allele), NA for missing
<b><i>depth.orig</i></b>	matrix ( <i>nind</i> x <i>nsnps</i> ) of counts for each sample and SNP
<b><i>sampdepth</i></b>	mean depth for each sample
<b><i>snpdepth</i></b>	mean depth for each SNP
<b><i>callrate</i></b>	mean call rate for each sample
<b><i>SNPcallrate</i></b>	mean call rate for each SNP
<b><i>p</i></b>	allele frequencies on the basis of allele counts
<b><i>pg</i></b>	allele frequencies on the basis of genotype calls
<b><i>HWdis</i></b>	Hardy-Weinberg disequilibrium (raw)
<b><i>x2star</i></b>	Depth-adjusted chi-squared test statistic of Hardy Weinberg equilibrium
<b><i>l10pstar</i></b>	$\log_{10}$ p-value corresponding to <i>x2star</i>



### Function to read TagDigger format files (*readTD*)

This function is for reading TagDigger files. It can be used by the main program (if `functions.only` is FALSE, the default), but can also be used to read additional files (e.g. to compare results in two different files). The variables *nsnps*, *seqID*, *nind*, and *alleles* are defined. See the section on the TagDigger format for more information.

Usage: *readTD*(*genofilefn0* = *genofile*, *skipcols*=0)

Arguments:

<i>genofilefn0</i>	the name of the file to read. Defaults to <i>genofile</i> .
<i>skipcols</i>	the number of columns of input to ignore. Defaults to 0.

Value: NULL

### Function to remove samples from objects (*samp.remove*)

This function removes samples from the relevant objects (*alleles*, *depth*, *sampdepth*, *seqID* *nind*). It would normally be used between calls to *readGBS* and *GBSsummary*.

Usage: *samp.remove*(*samppos* = NULL, *keep*=FALSE)

Arguments:

<i>samppos</i>	the positions of the samples to remove. Defaults to NULL.
<i>keep</i>	If TRUE, the samples with positions <i>samppos</i> will be kept and other samples removed. Default value is FALSE

### Function to remove SNPs from objects (*snp.remove*)

This function removes samples from the relevant objects (*p*, *nsnps*, *SNP\_Names*, *alleles*, *depth*, and some others). It would normally be used between calls to *readGBS* and *GBSsummary*.

Usage: *snp.remove*(*samppos* = NULL, *keep*=FALSE)

Arguments:

<i>snppos</i>	the positions of the SNPs to remove. Defaults to NULL.
<i>keep</i>	If TRUE, the SNPs with positions <i>snppos</i> will be kept and other SNPs removed. Default value is FALSE

### Depth functions (*depth2K*, *depth2Kbb*, *depth2Kmodp*, *depth2Kchoose*)

The GBS-Chip-Gmatrix.R program defines a default function for calculating “*K* values”, as well as alternate functions (using alternate allele sampling models) and a function to reset the default to one of the alternatives. These functions are relevant for used both self-relatedness estimation and pedigree assignment diagnostics. If a different depth model is required for calculating the self-relatedness, this *depth2K* function should be re-defined before using the *calcG* function (defined below). *K* is the probability of observing an AA genotype, given that the true genotype is AB and the read depth is *k*. These models will be discussed in more detail elsewhere. The function is used within *calcG* for calculating the self-relatedness for G5, and in the pedigree assignment program, for calculating expected mismatch rates.

A function *depth2K* is defined. This function takes a vector of read depths and returns the corresponding set of *K* values. Initially the function is defined using a binomial sampling model (the number of A alleles is binomial with probability parameter 0.5 and sample size the read depth).

*depth2Kbb* is an alternate depth function which uses a beta-binomial model. This model has two parameters,  $\alpha$  and  $\beta$ , but here these are set to be equal, so that  $P(AA|AB, k=1) = 0.5$ .

Usage: *depth2Kbb* (*depthvals*, *alph*=Inf)

Arguments:

<i>depthvals</i>	a vector of read depths
<i>alph</i>	the value of $\alpha$ (and also $\beta$ ) – the default is to use Inf, in which case the binomial model is used.

*depth2Kmodp* is an alternate depth function which uses a modified *p* value for 2<sup>nd</sup> and subsequent reads. The modified *p* can be thought of as the probability of seeing the same allele as in the previous read (for that SNP) for a true AB genotype, although because we are only interesting in the probability of all reads being the same allele, it is also the probability of seeing the same allele as *all* previous reads (for a true AB genotype).

Usage: *depth2Kmodp* (depthvals, modp=0.5)

Arguments:

depthvals	a vector of read depths
modp	the modified probability – the default is 0.5, which gives the binomial model. Normally a value $\geq 0.5$ would be used to reflect an increased chance of seeing the same allele as in the previous read.

*depth2Kchoose* is function to re-define *depth2K* to one of the alternative models.

Usage: *depth2K* <- *depth2Kchoose* (dmodel="bb", param)

Arguments:

dmodel	the model to use, either "modp" (to use <i>depth2Kmodp</i> ), or "bb" to use <i>depth2Kbb</i> – the default is "bb" (also used if any other string is used)
param	the parameter to use for the alternative function, used for alph for the bb model, and modp for the modp model.

### Function for calculating identity mismatch rates (*mismatch.ident*)

Mismatch rates for comparing two results from putatively the same individual. Currently under development. Used by *posCreport*.

Usage *mismatch.ident*(seqID1, seqID2, snpsubset=1:nsnps, puse=p, mindepth.mm=1)

Arguments:

seqID1	seqID for first result
seqID2	seqID for second result
snpsubset	a vector of integers (between 1 and <i>nsnps</i> , inclusive) of SNPs to be compared. The default is to use all SNPs.
puse	a length <i>nsnps</i> vector of allele frequencies to use in the calculations. The default is to use allele frequencies calculated on the basis of allele counts.
mindepth.mm	the minimum depth for a genotype to be used in the comparison

Value: a list containing mmrate (raw mismatch rate), ncompare (number of SNPs compared) and exp.mmrate, the expected mismatch rate.

### Function for reporting on positive controls (*posCreport*)

A function, *posCreport*, for reporting on samples which are supposedly from the same individual. These will normally be one or more positive controls, but may also be repeat runs.

Usage *posCreport*(mergeIDs, Guse, indsubset, Gindsubset, snpsubset=1:nsnps, puse=p)

Arguments:

mergeIDs	a vector of identifiers, ordered as in <i>Guse</i> , where samples from the same individuals are given the same identifier
Guse	the G matrix for comparing samples
sfx	text to be included in output file names to allow output from multiple calls or runs to be identified
indsubset	a vector of integers (between 1 and <i>length(mergeIDs)</i> , inclusive) of individuals in <i>mergeIDs</i> (and <i>Guse</i> ) to be compared. The default is to use all individuals.
Gindsubset	a vector of integers (between 1 and <i>nind</i> , inclusive) of the individuals from the full data in <i>Guse</i> (normally the same as used for <i>indsubset</i> when calling <i>calcG</i> to obtain <i>Guse</i> )
snpsubset	a vector of integers (between 1 and <i>nsnps</i> , inclusive) of SNPs to be compared for mismatch rates. The default is to use all SNPs. If an empty set ( <i>integer(0)</i> ) is given, comparisons using relatedness estimates only will be undertaken (this is useful if comparisons are to be made from saved relatedness estimates from different analyses).
puse	a length <i>nsnps</i> vector of allele frequencies to use in the mismatch calculations. The default is to use allele frequencies calculated on the basis of allele counts.

Value: a data frame containing columns mergeID (the ID given in *mergeIDs*), nresults (the number of runs with this ID), selfrel (the average self-relatedness), meanrel (the mean relatedness between all pairs with the given value of mergeID), minrel (the minimum relatedness between all pairs with



the given value of *mergeID*), *meandepth* (mean of *sampdepth*), *mindepth* (minimum *sampdepth*), *meanCR* (mean call rate). Only values of *mergeID* with *nresults* >1 are included.

**Details:** The function displays pairs of results (relatedness estimate, mean depth for each sample and IEMM) where the estimated relatedness is less than 1 and below the *selfrel* by at least  $1 - \text{hirel.thresh}$ , and outputs the files:

*posCchecks<sfx>.txt* a copy of the results displayed on the default output (i.e. low relatedness pairs)

*posCreport<sfx>.csv* contains the data frame that was returned by the function

*SelfRel<sfx>.png* a plot of *meanrel* against *selfrel*. The line of equality is shown in red. A grey line gives the boundary where relatedness is lower than 1 and lower than *selfrel* by more than  $1 - \text{hirel.thresh}$  (as a guide for results to check).

*posC-MM<sfx>.png* a plot of mean (over pairs of the same individual) raw mismatch rate against mean expected mismatch rate. The line of equality is shown in red, while a grey line denotes when the difference (identity EMM, IEMM) is greater than the threshold *iemm.thresh*.

*posC-EMM<sfx>.png* a scatterplot of matrix mean of *selfrel*, *meanrel* and IEMM.

### Functions for merging results for the same individual (*mergeSamples*, *mergeSamples2*)

A function, *mergeSamples*, for merging samples from the same individual. The function *mergeSamples2* is similar, see below.

**Usage** *mergeSamples* (*mergeIDs*, *indsubset*)

**Arguments:**

<i>mergeIDs</i>	a vector of identifiers for all <i>nind</i> samples, such that samples that have the same identifier are to be merged
<i>indsubset</i>	a vector of integers (between 1 and <i>nind</i> , inclusive) of samples to be retained. The default is to use all all samples.

**Value:** a list of the following objects:

<i>mergeIDs</i>	a vector of identifiers, as per the input, but ordered as in the other output objects (and with unique values)
<i>nind</i>	the length of <i>mergeIDs</i>
<i>seqID</i>	normally one of the <i>seqIDs</i> that correspond to the <i>mergeIDs</i> . If the <i>seqIDs</i> can be broken into five parts, using an underscore ( <code>_</code> ) as a separator, then the second part will be replaced by "merged", the third part by the number of results merged and the fourth part by "0"
<i>genon</i>	genotype (0/1/2) matrix after merging, if <i>genon</i> exists before merging
<i>depth.orig</i>	depth matrix after merging, if <i>genon</i> exists before merging
<i>alleles</i>	alleles matrix after merging, if <i>alleles.keep</i> is TRUE or , if <i>genon</i> does not exist before merging
<i>sampdepth</i>	sample mean read depths after merging, if <i>genon</i> exists before merging
<i>snpdepth</i>	SNP mean read depths after merging, if <i>genon</i> exists before merging
<i>pg</i>	allele frequencies based on genotype calls, after merging
<i>nmerged</i>	number of results merged (1, if not merged) for each individual.

Normally these objects would be used to replace their corresponding values before the merge, but this is not done automatically (it is up to the user). Note that some objects are not merged (e.g. the allele depth matrix, *alleles*, if *alleles.keep* is FALSE) and that the diagnostics produced when sourcing GBS-Chip-Gmatrix.R are not re-done by this function. If *mergeSamples* is run before *GBSSummary* (when *functions.only* is TRUE, and after the data is read), then it produces a reduced set of outputs, corresponding to those existing before the merge. In particular it will have an *alleles* object, but neither *genon* nor *depth.orig*.

*mergeSamples* will fail when the number of elements for a merged object (*genon*, and *alleles* if *alleles.keep* is TRUE) exceeds the maximum integer allowed (currently  $2^{31}-1$ ). *mergeSamples2*

provides a strategy to allow larger merges when some of the records do not require merging, with the limit being  $2^{31}-1$  elements in the subset of *genon* containing *mergeIDs* with at least two observations in the input data. *mergeSamples2* does not contain some of the other functionality of *mergeSamples* (yet), for example the objects created in *GBSSummary*, such as *genon*, must be present, and the output object will not contain *alleles* (even if *alleles.keep* is TRUE).

### Plate plot function (*plateplot*)

The *plateplot* function can be used for quality control based on laboratory plate layouts. The user must supply this layout in *plateinfo*. If there are more than one observation for a given plate position, then the sum for those observations is used (but currently the function is intended to be used with a single value for each position). Positions without data will be displayed as white.

**Usage:** `plateplot(plateinfo, plotvar=sampdepth, vardesc="", sfx="", neginfo, negcol="grey", colpal = hcl.colors(80,palette="YlGnBu", rev=TRUE))`

#### Arguments:

<i>plateinfo</i>	a data frame with elements row, column and optionally subplate (anything else ignored). The rows in <i>plateinfo</i> should correspond to the elements in <i>seqID</i> (at time of use). row and column contain plate row and column labels for an observation. These can be text or numeric, but plate physical order and sort order of these label should match. If <i>plateinfo</i> has a variable <i>subplate</i> present, this should represent plate identifiers for an earlier step in the lab process than for the plate represented by the <i>row</i> and <i>column</i> positions. In this case an additional plot is produced distinguishing the <i>subplate</i> levels.
<i>plotvar</i>	a variable with values to be plotted (using a colour scale). The default is <i>sampdepth</i> .
<i>vardesc</i>	A descriptive name for <i>plotvar</i> , used to label the colour scale on the plot(s). A length zero string (the default) will result in the name of the variable given in <i>plotvar</i> to be used.
<i>sfx</i>	A suffix to use in output file names to identify which function call has produced that output.
<i>neginfo</i>	A data frame with elements row, column identifying the positions of any negative controls. These will be identified on the plots with two diagonal lines (X) through the position. If not given negative control plotting is ignored.
<i>negcol</i>	the colour of the lines identifying negative control positions. The default is "grey".
<i>colpal</i>	a colour palette to be used for displaying <i>plotvar</i> . The entire palette is used to cover the data range (the value corresponding to a given colour will change depending on the data). It is advised that the palette does not include white as this will be the colour for any positions without data (e.g., failed), which are useful to have identified. The default palette uses the "YlGnBu" (yellow – green – blue) palette (or similar, if not available). Other possibilities include viridis colours (yellow low, blue high, e.g., <code>rev(hcl.colors(80),rev=TRUE)</code> , requires R.version ≥ 3.6) or heatmap colours (e.g. yellow to red: <code>rev(heat.colors(80))[25:80]</code> )

**Details:** Colour plots of the variable in plate layout are produced as given below. The function does not return an object to R.

*Plate<sfx>.png* Colour plot (using *colpal*) of the variable in plate layout. A colour legend is included with the minimum, mean and maximum of the variable indicated.

*Subplate<sfx>.png* Colour plot of the variable in plate layout with darker colours indicating higher values and different base colours used for each level of *subplate*. A colour legend is included with the minimum, mean and maximum of the variable indicated. This plot is produced only when *plateinfo* contains a *subplate* column.

### Hardy-Weinberg statistics function (*HWpops*)

A function, *HWpops*, for calculating Hardy-Weinberg statistics, by population if provided, is defined.

Usage: *HWpops*(snpsubset, indsubset, populations=NULL, depthmat = depth.orig)

Arguments:

snpsubset	a vector of integers (between 1 and <i>nsnps</i> , inclusive) of SNPs calculate the statistics for. The default is to use all SNPs.
indsubset	a vector of integers (between 1 and <i>nind</i> , inclusive) of individuals to use for the calculations. The default is to use all individuals.
populations	a vector of length <i>nind</i> containing population labels. The default is NULL in which case a single population of all individuals is used.
depthmat	matrix ( <i>nind</i> x <i>nsnps</i> ) of depth values. The default is depth.orig which should normally be used.

Value: a list of the following objects:

HWdis	a matrix of Hardy-Weinberg disequilibrium values, populations in rows, SNPs in columns, population names used for row names.
l10LRT	a matrix of $-\log_{10}$ p-values from the likelihood ratio test assuming observed genotypes are true, populations in rows, SNPs in columns, population names used for row names.
x2star	a matrix of x2star values (depth-adjusted chi-squared test statistic of Hardy Weinberg equilibrium), populations in rows, SNPs in columns, population names used for row names.
l10pstar	a matrix of $-\log_{10}$ p-values corresponding to <i>x2star</i> , populations in rows, SNPs in columns, population names used for row names.
maf	a matrix of minor allele frequencies (based on observed genotypes), populations in rows, SNPs in columns, population names used for row names.
l10pstar.pop	(if there is more than one population) a vector of $-\log_{10}$ p-values for each SNP corresponding to a combined within populations test of <i>x2star</i> . Non-missing <i>x2star</i> values are summed over populations and the p-value calculated from the chi-squared distribution with degrees of freedom equal to the number of <i>x2star</i> values summed.

### Allele frequency function (*calcp*)

A function, *calcp*, for calculating allele frequencies (for all SNPs), is defined.

Usage: *calcp*(indsubset, pmethod="A")

Arguments:

indsubset	a vector of integers (between 1 and <i>nind</i> , inclusive) of the individuals for which are to be used for allele frequency estimation. The default is to use all individuals.
pmethod	a method for calculating the frequencies, being one of "A" (calculate on the basis of allele counts – the default method) or "G" (calculate on the basis of genotype calls)

Value: a vector of allele frequencies

Warning when using this after *mergeSamples*: pmethod A uses the object *alleles*, which is not recreated during the merge, so indsubset refers to sample positions prior to the merge. pmethod G uses *genon* whose positions are those following the merge.

### Genomic relatedness function (*calcG*)

A function, *calcG*, for calculating the genomic relatedness, is defined.

Usage: *calcG*(snpsubset, sfx="", puse, indsubset, depth.min=0, depth.max=Inf, npc=0, calclevel=9, cocall.thresh=0, mdsplot=FALSE, mindepth.idr = 0.1, withPlotly=FALSE, withHeatmaply=withPlotly, plotly.group=NULL, plotly.group2=NULL, samp.info=NULL, samptype="diploid")

Arguments:

snpsubset	a vector of integers (between 1 and <i>nsnps</i> , inclusive) of the SNPs to use in the calculation. The default is to use all SNPs.
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sfx	A suffix to use in output file names to identify which function call has produced that output.
puse	a vector or matrix of (reference) allele frequencies to use in the calculations. The default is to use allele frequencies calculated on the basis of allele counts. The values (for the snps in <i>snpsubset</i> ) should be greater than 0 and less than 1. For the vector form, either a vector of the same length as <i>snpsubset</i> or one of length <i>nsnps</i> (in which case the function subsets these using <i>snpsubset</i> ) can be supplied. If a matrix is supplied, it is taken as the matrix of allele frequencies for individuals x snps. The number of rows can be the length of <i>indsubset</i> or <i>nind</i> , while the number of columns can be the length of <i>snpsubset</i> or <i>nsnps</i> , and if needed the function subsets to the required set.
indsubset	a vector of integers (between 1 and <i>nind</i> , inclusive) of the individuals for which relatedness matrices will be calculated. The default is to calculate for all individuals.
depth.min	The minimum depth for a SNP result for an individual to be used.
depth.max	The maximum depth for a SNP result for an individual to be used.
npc	The number of principal components of the 'G5' relatedness matrix (Gpool for <i>samptype</i> = "pooled") to display. If <i>npc</i> ≤ 0, then the heatmap plot is omitted, but otherwise <i> npc </i> is used for <i>npc</i> . If <i>npc</i> = 0 (the default) the principal component analysis is omitted.
calclevel	specifies the amount of calculation and output produced: 1 gives G5 (see below) and intermediate results only, 2 gives G5 and reports using G5, 3 gives all types of G available and 9 gives these and all reporting available.
cocall.thresh	Samples may be removed so that co-call rates (the proportion of SNPs with a call in both of a pair of samples) for heatmap and PCA analyses are above this value. Firstly, if <i>cocall.thresh</i> ≥ 0, samples with a maximum SNP depth of 1 are removed. The further samples are removed successively, with the sample appearing the most often in pairs not meeting the criterion removed at each step, until all pairs meet the criterion. The removal of these samples under the default threshold allows the heatmap and PCA analyse to be performed (no NAs in the relationship matrix used).
mdsplot	if TRUE and the conditions for plotting the principal components is met, a two-dimensional multidimensional scaling plot (principal coordinates plot) is also produced. The default is FALSE.
mindepth.idr	minimum depth for including samples in the self-relatedness (or inbreeding) regression on log(sample depth), applied after any filtering specified in the other call parameters.
withPlotly	If TRUE, then plotly graphs are produced, else if FALSE the standard plots are produced. The default is FALSE.
withHeatmaply	If TRUE, then plotly graphs are produced for the heatmap, else if FALSE the standard plots are produced. The default is the <i>withPlotly</i> setting.
plotly.group	A character vector of length equal to the number of individuals (in <i>indsubset</i> ). Gives grouping on the plotly graphs in terms of different coloured points. The default is NULL (no colouring).
plotly.group2	A character vector of length equal to the number of individuals (in <i>indsubset</i> ).. Gives grouping on the plotly in terms of different points. The default is NULL (no grouping with symbols).
samp.info	A list where each element is a character vector of length equal to the number of individuals (in <i>indsubset</i> ). Used to provide "hover" information for plotly graphs. The default is NULL, in which case the seqID is used.
samptype	the type of samples being analysed. Currently only two values are allowed: "diploid" (the default) and "pooled" for the case where pools of individuals are sequenced. Any other value is treated as "diploid".

Value: a list of relatedness structures: G1, G4d (diagonal elements of G4), G5, Gpool, samp.removed (positions of samples removed to ensure the *cocall.thresh* criterion) and PC, the output of the principal components analysis (if *|npc|* > 0). The *Gn* relatedness matrices are

described in Dodds *et al.* (2015), except that a range of allele sampling models can be incorporated for the diagonal of G5 – see the depth2K section below). Gpool is NULL except when *samptype* is “pooled”.

For *samptype* = “pooled”, Gpool is another relatedness matrix calculated in the same way as G5 (including the diagonal adjustment for depth), except that 2 x the proportion of reference alleles is used as the numeric genotype (rather than the number of reference alleles in the observed genotype). This is similar to the approach of Reverter *et al.* (2016) for pooled SNP-chip data, and Cericola *et al.* (2018) for pooled GBS data, except that no adjustment is made for pool size as in those studies – this is left to the user (e.g. Gpool could be multiplied by the pool size as in Cericola *et al.* (2018) or scaled so that diagonal elements are 1 as in Reverter *et al.* (2016). Note that the diagonal adjustment for depth has not been theoretically verified, and therefore Gpool should be considered as under development. When *samptype* = “pooled” the outputs are based on Gpool in preference to G5.

When *puse* is a matrix, individual-specific allele frequencies are used (normally these would be population-specific, such as the breed or strain, including crosses). The calculations follow the method of Auvray *et al.* (2014) which is based on Harris and Johnson (2010).

Some summary information is output. Gpool is used for the self-relatedness regression when *samptype* is “pooled”.

**Details:** The function also produces a set of output files, as detailed below. If *withPlotly* is TRUE and both the plotly and heatmaply packages are available, interactive plotly plots are produced for some of the plots.

*Co-call-<sfx>.png* is a histogram of co-call rates (the proportion of SNPs with a call in both of a pair of samples) for all sample pairs.

*MAF<sfx>.png* is a histogram of the MAFs for the subset of SNPs used (if not all SNPs).

*HighRelatedness<sfx>.csv* contains pairs of samples, their G5 relatedness (G5rel) and self-relatednesses (SelfRel1 and SelfRel2), where the relatedness is > *hirel.thresh* (default value of *hirel.thresh* is 0.9).

*Heatmap-G5<sfx>.png* is a heatmap plot using G5 relatedness (Gpool for *samptype* = “pooled”). This is not produced if *npc* ≤ 0. If *fcolo* for the relevant individuals has more than one colour, colour bars are added to the plot.

*HeatmapOrder<sfx>.csv* contains a list of the samples in the order they are plotted on the heatmap. *rowInd* is the index values (written on the heatmap plot), *seqIDInd* is the position of the individual in *seqID.csv*; *seqID* is also included. For “standard” cases, where all *seqID* samples are included, the values of *rowInd* and *seqIDInd* will be the same.

*Heatmap-G5<sfx>.html* is a plotly version of *Heatmap-G5<sfx>.png*. *plotly.group* and *plotly.group2* are not used.

*Gcompare<sfx>.png* is a plot comparing relatedness estimates for G1, G3 and G5.

*G<sfx>-diag.png* is a plot of diagonal elements (self-relatedness estimates) of G4 against those of G5 (illustrating the effect of correcting for depth). If *samptype* is “pooled” then a comparison of diagonal elements of G4, G5 and Gpool is produced.

*G<sfx>-diag.html* is a plotly plot of diagonal elements of G4 against those of G5. Produced if *withPlotly* is TRUE. If *samptype* is “pooled” then Gpool is used instead of G5.

*G<sfx>diagdepth.png* is a plot of diagonal elements of G5 (or Gpool if *samptype* is “pooled”) against the logged sample depth. We do not expect there to be a relationship between these variables (unless planned) so this serves as a diagnostic for e.g. non-Mendelian SNPs and/or the assumption of random sampling of alleles during sequencing.

*G<sfx>diagdepth.html* is a plotly version of *G<sfx>diagdepth.png*.

*PC1v2G5<sfx>.png* (if  $|npc| > 0$ ) is a plot of 2<sup>nd</sup> versus the 1<sup>st</sup> principal components. Points are plotted with open (if 100 or more samples) or closed circles. If only one component was requested, a histogram of the 1<sup>st</sup> component is produced.

*PC1v2G5<sfx>.html* is the plotly version of *PC1v2G5<sfx>.png*.

*PCG5<sfx>.pdf* (if  $|npc| > 2$ ) is a scatterplot matrix of the first  $|npc|$  principal components.

*PC1vInb<sfx>.png* (if  $|npc| > 0$ ) is a plot of estimated inbreeding (using G5 or Gpool) against the 1<sup>st</sup> principal component. An unexpected trend may indicate allele sampling does not follow the analysis model.

*PC1vDepth<sfx>.png* (if  $|npc| > 0$ ) is a plot of sample depth against the 1<sup>st</sup> principal component. An unexpected trend may indicate that the data contain artefacts.

*MDS1v2G5<sfx>.png* (if  $|npc| > 0$ ) is a plot of 2<sup>nd</sup> versus the 1<sup>st</sup> principal coordinates (multidimensional scaling axes).

*MDS1v2G5<sfx>.html* is the plotly version of *MDS1v2G5<sfx>.png*.

There is a vector *fcolo* (length *nind*) of colours to be used for the individuals in these plots. It defaults to all black, but can be set after sourcing the program (and/or running GBSsummary) and before calling *calcG*.

### Output genomic relationship matrix (*writeG*)

A function, *writeG*, for saving genomic relationship matrices, is defined.

Usage: *writeG* (*Guse*, *outname*, *outtype*=0, *indsubset*, *IDuse*, *metadf*=NULL)

#### Arguments:

<i>Guse</i>	the G matrix of relationships to output, should be a square matrix, or a list containing an element G5 (for <i>outtypes</i> 1 to 5) and/or PC (for <i>outtype</i> 6)
<i>outname</i>	text used in the naming of the output file(s)
<i>outtype</i>	constant or vector containing the type(s) of output required. If <i>outtype</i> contains any of the following values, the corresponding output is produced: 1 an R datasets file containing the G matrix and corresponding <i>seqID</i> 2 a .csv file containing the G matrix with row and column headings 3 a .csv file containing the G matrix in "long" format, i.e. one row for every (unique) relationship pair including selfs; columns are IDs of first and second individual, followed by the relatedness value 4 a .csv file containing inbreeding for each individual; first column contains IDs, second column contains inbreeding estimates 5 two tab delimited files (.tsv) for input into the t-SNE interactive browser at <a href="http://projector.tensorflow.org/">http://projector.tensorflow.org/</a> (allows exploration of dimension-reduced data from the PCA or t-SNE methods). 6 a .csv file containing the principal components (requires <i>Guse</i> to be a list with element PC)
<i>indsubset</i>	a vector of integers (between 1 and <i>nind</i> , inclusive) of the individuals in the G matrix. The default assumes all individuals.
<i>IDuse</i>	a vector of IDs to use in the output, corresponding to the order in <i>Guse</i> , the default is to use values of <i>seqID</i> as the identifiers (in which case <i>seqID</i> must exist)
<i>metadf</i>	a data frame with the same number of rows as the G matrix, containing sample information to pass to the t-SNE browser.

Details: One or more files are written to the default directory, according to *outtype*:



**<outname>.RData** an R data file containing the G matrix and corresponding *IDuse* values, produced when *outtype* contains a 1. The G matrix is named based on the object specified in *Guse*, removing text up to \$ and from [, if either of these are present. As an example using `writeG(Gfull$G5[1:100,1:100],outtype=1)` will result in the G matrix being named G5.

**<outname>.csv** a csv file containing the G matrix, produced when *outtype* contains a 2. The first column is labelled with the name of the object passed to *IDuse* and contains the values of *IDuse*. The other columns are labelled with the values of *IDuse*.

**<outname>-long.csv** a csv file containing the unique relatedness values, one row for every pair of individuals (including selfs), produced when *outtype* contains a 3. The columns are labelled id1, id2 (lower case to avoid warning messages when opening with Excel) and rel. *IDuse* is used for the ID values.

**<outname>-Inbreeding.csv** a csv file containing inbreeding values (self-relatedness minus 1), produced when *outtype* contains a 4. The first column is labelled with the name of the object passed to *IDuse* and the second column as Inbreeding. *IDuse* is used for the ID values.

**<outname>-pca\_vectors.tsv** a tsv file containing the G matrix in a format suitable for the t-SNE browser, produced when *outtype* contains a 5.

**<outname>-pca\_metadata.tsv** a tsv file containing sample information (from *metadf*, or *IDuse* if *metadf* is NULL) in a format suitable for the t-SNE browser, produced when *outtype* contains a 5.

**<outname>-PC.csv** a csv file containing principal components, produced when *outtype* contains a 6 and *Guse* is a list containing PC (*Guse* is assumed to be the output from *calcG*). The first columns are from *metadf*, if given, or the object passed to *IDuse*. Subsequent columns are the principal components, labelled PC1, PC2 etc.

### Relatedness comparison function (*GCompare*)

This is a function to help make comparisons between different estimates of relatedness on the same set (or overlapping subsets) of individuals. These different estimates may come from different genotyping technologies (e.g. SNP chip vs GBS), different protocols (e.g. GBS with different restriction enzymes, different levels of multiplexing samples, different SNP callers) or using different SNP filters.

The program inputs a set of (genomic) relationship matrices (GRMs) and a corresponding set of individual IDs. The output is a set of scatterplots (possibly as scatterplot matrices) and corresponding regression output. The relatedness estimates between each pair of (different) individuals for each pair of GRMs are compared, as are those for the self-relatedness estimates for each individual. For any pair of GRM, all individuals common to the GRM are used. If there are duplicated IDs within any set a warning is printed and only the first observation for the individual is used.

The information in the upper and lower panels is determined by the functions *regpanel* (for upper panel which normally shows regression results) and *plotpanel* (for lower panel which normally shows the scatterplot). These can be redefined before running *GCompare* (e.g. to change the colour of the line of equality, to draw other reference lines etc). Setting one of them to NULL will suppress that half of the scatterplot matrix. Setting *regpanel* to NULL will also suppress the statistics being added to the output plots when there are only 2 G matrices.

Additionally, if the *MethComp* (Carstensen, 2015) R package is installed, there can be corresponding sets of plots using this package, with scatterplots of relatedness estimates below the diagonal, and 'Bland-Altman' (BA) plots (Altman and Bland, 1983) above the diagonal. The Bland-Altman plots have the differences on the vertical and the means on the horizontal axis, for the two relatedness estimates. These plots take a lot more CPU time than the regression plots.

**Usage:** GCompare (Glist, IDlist, Gnames = paste0("G.",1:length(Glist)), plotname = "", whichplot="both", doBA=FALSE, ...)

**Arguments:**

Glist	a list of G matrices
IDlist	a list of ID variables, paired to the G matrices and in the same order as the data in the corresponding G matrix
Gnames	a set of labels to use for the G matrices (defaults to G1, G2, ...)
plotname	text to use in the naming of output files
whichplot	variable to choose which plot types are produced, can be one of: "diag": compare diagonals (self-relatedness) "off": compare off-diagonals (relatedness between individuals) "both": compare both diagonals and off-diagonals. This is the default.
doBA	Additionally produce Bland-Altman plots. The default is FALSE.
...	Arguments to be passed to the plotting functions (e.g. col= for coloring). These need to be relevant to the plot types being produced (e.g. if a vector of colours, then it should not be used with <i>whichplot</i> ="both").

**Details:** One or more plots are produced, depending on the options used. Regression statistics relating to each comparison are displayed. A set of ignorable warnings is issued.

*Gcompare- <plotname>-diag.png* a plot of the diagonal comparison(s). If more than 2 G matrices, this will be a scatterplot matrix with regression results in the upper matrix panels. A red line is drawn where values are equal.

*Gcompare- <plotname>-offdiag.png* a plot of the diagonal comparison(s). If more than 2 G matrices, this will be a scatterplot matrix with regression results in the upper matrix panels. A red line is drawn where values are equal.

*GcompareBA- <plotname>-diag.png* a scatterplot matrix BA plot of the diagonal comparison(s). The regression plots are in the lower diagonal and the BA plots in the upper diagonal. A grey line indicates equality ( $y=x$  for lower plots,  $y=0$  for upper plots). The BA plots have 3 additional horizontal lines being the mean & mean  $\pm 1.96sd$  ('95% limits of agreement').

*GcompareBA- <plotname>-offdiag.png* a scatterplot matrix BA plot of the off-diagonal comparison(s). See description of the BA plot for the diagonals for more details.

### Bend a genomic relationship matrix (*Gbend*)

The function *Gbend* will 'bend' a square matrix to make it positive definite. The bending method used was proposed by Schaeffer (2013) and involves an eigen-decomposition of the input matrix, modification of the eigenvalues followed by a reconstruction. The method given here simply increases all eigenvalues below a threshold to that threshold. There are no guarantees to the performance of this method (use at your own risk!). The output matrix should be compatible with GBLUP methods.

**Usage:** Gbend (GRM, mineval=0.001, doplot=TRUE, sfx="", evalsum="free")

**Arguments:**

GRM	the square matrix to bend (usually a genomic relatedness matrix)
mineval	the lower threshold for eigenvalues. Values less than <i>mineval</i> are set at <i>mineval</i> . The default is 0.001.
doplot	If TRUE (the default), produce output plots (see below).
sfx	A suffix/label for the plot names and titles
evalsum	If "fixed", modified eigenvalues are rescaled to the original sum. The default is "free" (do not rescale).

**Details:** A matrix with the same dimensions as *GRM* is returned. If *doplot* is TRUE, three plots are produced:

*Eigenvalues<sfx>.png* a plot of the ordered original eigenvalues diagonal comparison(s). Negative eigenvalues are plotted in blue (positive values in black). A line is drawn at the *mineval* threshold.

*Self-Bending<sfx>.png* a plot of the diagonal values (self-relatedness values for a GRM) of GRM after vs before bending. The red lines shows where these are equal.

*Rel-Bending<sfx>.png* a plot of the off-diagonal values (between sample relatedness values for a GRM) of GRM after vs before bending. The red lines shows where these are equal.

### Output data in variant call format (*writeVCF*)

A function, *writeVCF*, for saving data in VCF format is defined.

Usage: *writeVCF*(indsubset, snpsubset, outname=NULL, ep=0.001, puse = p, IDuse, usePL = FALSE, contig.meta = FALSE)

#### Arguments:

indsubset	a vector of integers (between 1 and <i>nind</i> , inclusive) of the individuals in to be output. The default assumes all individuals.
snpsubset	a vector of integers (between 1 and <i>nsnps</i> , inclusive) of the SNPs to output. The default is to use all SNPs.
outname	base name of the output file which will have the extension “.vcf” appended. The default is “GBSdata”.
ep	the probability of a sequencing error. Default to 0.001 (changed in v0.8.2).
puse	a vector of length <i>nsnps</i> of (reference) allele frequencies to use in the calculation of posterior genotype probabilities. The default is to use <i>p</i> (normally the allele frequencies calculated based on allele counts).
IDuse	a vector of IDs of length <i>nind</i> to use in the output. The default is to use values of <i>seqID</i> as the identifiers (in which case <i>seqID</i> must exist)
usePL	indicator which, if set to TRUE, will result in the output containing phred-scaled likelihoods instead of genotype likelihoods. The default is FALSE.
contig.meta	indicator which, if set to TRUE, will add contig meta info to the file (ID's only). Required for input into ANGSD. The default is FALSE.

Details: A VCF format (<https://samtools.github.io/hts-specs/VCFv4.3.pdf>) file of the requested data is written. The file contains four fields of information relating to a genotype:

GT: the inferred genotype (0/0, 0/1, 1/1, and ./ for homozygous for reference allele, heterozygous, homozygous for alternate allele and missing, respectively)

GP: the three posterior genotype probabilities with priors calculated from the allele frequencies and assuming Hardy-Weinberg equilibrium, ordered corresponding to genotypes 0/0, 0/1, 1/1 (in GT format),

GL: (if usePL=FALSE) three log<sub>10</sub>-scaled likelihoods, calculated as in Li (2011), in the same order as GP,

PL: (IF usePL=TRUE) phred-scaled likelihoods (-10 \* (GL – max(GL)) rounded to the nearest integer,

AD: allelic depth (read depth for reference and alternate alleles).

The “chromosome” is specified as the SNP\_Name and the position is numbered sequentially from 1. The variants are all denoted as C (REF allele) and G (ALT allele). Currently there is no facility for incorporating other genomic information passed either in the input file (e.g. if Tassel format) or as additional information.

*<outname>.vcf* a vcf formatted file of the data specified.

### Output GBS data (*writeGBS*)

A function, *writeGBS*, for saving data is defined. Currently the only supported format is the UNEAK format (see the section “GBS via UNEAK”).

Usage: *writeGBS*(indsubset, snpsubset, outname= "HapMap.hmc.txt", outformat=gform, seqIDuse=seqID)

#### Arguments:

<code>indsubset</code>	a vector of integers (between 1 and <i>nind</i> , inclusive) of the individuals in to be output. The default assumes all individuals.
<code>snpsubset</code>	a vector of integers (between 1 and <i>nsnps</i> , inclusive) of the SNPs to output. The default is to use all SNPs.
<code>outname</code>	name of the output file. The default is "HapMap.hmc.txt".
<code>outformat</code>	the format of the output file. The default value is <i>gform</i> (the format of the input file). Currently only the "uneak" format will produce an output file. Any other value of <i>outformat</i> will produce a warning message.
<code>seqIDuse</code>	a vector of IDs of length <i>nind</i> to use in the output (and which will be read as <i>seqID</i> if the file is read back in). The default is to use values of <i>seqID</i> .

Details: A data file with the specified format is written. The function requires that the object *alleles* exists and that it corresponds to the genotype matrix (*genon*). It may be necessary to set *alleles.keep* to TRUE before data manipulation to ensure this is the case.

*<outname>* a file of the data specified.

#### **Gender prediction (*genderassign*)**

The function *genderassign* can be used to predict gender using the methods described in Bilton *et al.* (2019). The assignment boundaries are specified by two functions *upperbounday(x)* and *lowerboundary(x)* where x represents the proportion of heterozygotes on the homogametic sex chromosome. These functions can be modified before using *genderassign*, but only *upperboundary* can be non-linear (not checked). Their initial values are:

```
upperboundary <- function(x){ 20*pmax(rep(0,length(x)),x)^2+0.2}
lowerboundary <- function(x){ 0.1 + x}
```

Usage: *genderassign* (*ped.df*, *index\_Y\_SNPs*, *index\_X\_SNPs*, *sfx=""*, *hetgamsex* = "M", *homgamsex* = "F", *hetchrom* = "Y", *homchrom* = "X", *dojitter* = FALSE)

#### Arguments:

<code>ped.df</code>	a dataframe of individuals for gender prediction, as if read from a pedigree file (see Input formats section). This optionally contains variables Sex (with values M, F or U for male, female, unknown) and Relationship (character, e.g. "progeny", "sire" or "dam")
<code>index_Y_SNPs</code>	a vector of positions of SNPs on the homogametic sex chromosome (Y chromosome for X/Y systems) to use.
<code>index_X_SNPs</code>	a vector of positions of SNPs on the heterogametic sex chromosome (X chromosome for X/Y systems) to use.
<code>sfx</code>	text to be included in output file names
<code>plotname</code>	text to use in the naming of output files
<code>hetgamsex</code>	gender label for the heterogametic sex. The default is "M" (for males, assumes X/Y system). Use "F" for the Z/W system.
<code>homgamsex</code>	gender label for the homogametic sex. The default is "F" (for females, assumes X/Y system). Use "M" for the Z/W system.
<code>hetchrom</code>	chromosome label for the heterogametic sex chromosome. The default is "Y" (assumes X/Y system). Use "W" for the Z/W system.
<code>homchrom</code>	chromosome label for the homogametic sex chromosome. The default is "X" (assumes X/Y system). Use "Z" for the Z/W system.
<code>dojitter</code>	if set to TRUE, values on the y axis are jittered. The default is FALSE.

Details: Outputs a dataframe containing the input data frame (*ped.df*), the predicted gender, *new\_prop\_X* (the ratio of heterozygosity proportion of SNPs in *index\_X\_SNPs* compared to their expected proportions given the depth), *proportion\_SNPs\_Y* (proportion of SNPs in *index\_Y\_SNPs* with a result) and *sampdepth* (the mean sample depth for the individual). An output file and plot are also produced.

*gender\_prediction<sfx>.csv* a .csv file containing the same information as the output dataframe.

*GenderPlot<sfx>.png* a plot of the results similar to Figure 1 of Bilton *et al.* (2019), where females are plotted in red, males in blue, unknowns in grey. The light blue shaded region indicates individuals predicted to be male, while the light red shaded region indicates individuals predicted to be female.

### Finplot functions (*finplot*, *HWsigplot*, *finclass*)

These functions allow different colours to be applied to the normal 'finplot'. A finplot refers to a plot of *HWdis* against *maf* for each SNP. The functions *finplot* (coloured by depth) and *HWsigplot* (coloured by the significance of a Hardy-Weinberg test) are used in the normal running of the GBS-Chip-Gmatrix.R code (perhaps via *GBSSummary*) – see Output - files, but they can also be used outside of this to give plots with other options specified. *finclass* allows colouring based on a factor or character variable.

**Usage:** *finplot* (*HWdiseq*=*HWdis*, *MAF*=*maf*, *plotname*="finplot", *finpalette*=*palette.aquatic*, *finxlim*=*c*(0,0.5), *finylim*=*c*(-0.25, 0.25))

#### Arguments:

<i>HWdiseq</i>	a vector of <i>nsnp</i> y values for the plot (normally a set of HW disequilibrium values). The default is to use <i>HWdis</i>
<i>MAF</i>	a vector of <i>nsnp</i> x values for the plot (normally a set of minor allele frequencies). The default is to use <i>maf</i>
<i>plotname</i>	The name of the .png output file. The default is "finplot"
<i>finpalette</i>	A set of 50 colours to use for portraying <i>snpdepth</i> . The default is to use <i>palette.aquatic</i> (grey to blue). Other inbuilt palettes are <i>palette.terrain</i> (from <i>terrain.colors</i> ) and <i>palette.temperature</i> (blue through white to red).
<i>finxlim</i>	a numeric vector of length 2 with the x-coordinate limits. The default is (0,0.5) (the bounds for minor allele frequencies).
<i>finylim</i>	a numeric vector of length 2 with the y-coordinate limits. The default is (-0.25,0.25) (the bounds for Hardy Weinberg disequilibrium).

**Details:** A finplot is produced, coloured by *snpdepth* (mean depth per SNP), truncated at 256. *snpdepth* is mapped to *finpalette* in a non-linear way, such that there is more separation at lower depths. If *finpalette* contains colours close to white, the background of the plot is set to grey.

*<plotname>.png* a plot of SNPs coloured by *snpdepth*. Normally a finplot.

**Usage:** *HWsigplot* (*HWdiseq*=*HWdis*, *MAF*=*maf*, *ll*=*l10LRT*, *plotname*="HWdisMAFsig", *finpalette*=*palette.aquatic*, *finxlim*=*c*(0,0.5), *finylim*=*c*(-0.25, 0.25), *llname*="-log10 LRT", *sortord*=*ll*)

#### Arguments:

<i>HWdiseq</i>	as in the <i>finplot</i> function
<i>MAF</i>	as in the <i>finplot</i> function
<i>ll</i>	a vector of <i>nsnps</i> significance statistics to use in colouring the plot. The default is to use $\log_{10}$ likelihood values from the likelihood ratio test (without adjustment for depth) of Hardy-Weinberg equilibrium.
<i>plotname</i>	The name of the .png output file. The default is "HWdisMAFsig"
<i>finpalette</i>	A set of 50 colours to use for portraying <i>ll</i> . The default is to use <i>palette.aquatic</i> (grey to blue). See the <i>finplot</i> function for other inbuilt palettes. The palette used by the normal running of GBS-Chip-Gmatrix.R is <i>colorRampPalette</i> ( <i>c</i> ("deepskyblue2","red"))(50).
<i>finxlim</i>	as in the <i>finplot</i> function
<i>finylim</i>	as in the <i>finplot</i> function
<i>llname</i>	a label to use to describe the colours used. The default is "-log10 LRT".
<i>sortord</i>	a variable of length <i>nsnps</i> for plotting order. The default is variable specified by <i>ll</i> . The effect of sorting is that higher values are plotted last, and therefore makes these less likely to be overplotted.



Details: A finplot, but coloured by *ll*, is produced. The main code uses this function with *ll=ll10pstar*, *llname="-log10p X2\*"* and *finpalette=colorRampPalette(c("deepskyblue2","red"))(50)* (light blue to red).

*<plotname>.png* a plot of SNPs coloured by *ll*.

Usage: `finclass (HWdiseq=HWdis, MAF=maf, colobj, classname=NULL, plotname="finclass", finxlim=c(0,0.5), finylim=c(-0.25, 0.25))`

Arguments:

HWdiseq	as in the <i>finplot</i> function
MAF	as in the <i>finplot</i> function
colobj	a list as produced by <i>colourby</i> (see: Colouring functions ( <i>colourby</i> , <i>changecol</i> , <i>colkey</i> ) specifying the colour for each SNP
classname	a label to use to describe the colours used. The default is to not use a label.
plotname	The name of the .png output file. The default is "finclass"
finxlim	as in the <i>finplot</i> function
finylim	as in the <i>finplot</i> function

Details: A finplot, but with SNPs coloured by the *sampcol* item of *colobj*, is produced. This can be a useful graphic for displaying filtered and unfiltered SNPs, for example.

*<plotname>.png* a plot of SNPs coloured by *colobj\$sampcol*.

### HDplot function (*HDplot*)

This function produces an HDplot, as described by McKinney *et al.* (2017) with the proportion of (observed) heterozygous individuals on the x axis, and the read-ratio deviation (from 1:1) is plotted on the y axis. The function allows different colouring options (see details below). The function saves some intermediate variables to the workspace to save re-calculation when requesting another colouring scheme (remove the variable *HD.saved.KGD* if the function is to be applied to different data).

Usage: `HDplot (plotname="HDplot", colourtype = "depth", finpalette=palette.aquatic, HDxlim=c(0,1), HDylim=c(-Inf, Inf), HDcol=NULL, sortcol = "asc")`

Arguments:

plotname	The name of the .png output file. The default is "HDplot"
colourtype	specifies the colouring scheme, when HDcol is NULL. Can be one of "depth" (colour by mean read depth as in the <i>finplot</i> function), "HW" (colour by the raw Hardy-Weinberg significance) or "HW*" (colour by the depth-adjusted Hardy-Weinberg significance). The latter two schemes colour in the same way as in the <i>HWsigplot</i> function. A legend is also added to the plot.
finpalette	A set of 50 colours to use for portraying the information specified by <i>colourtype</i> . The default is to use <i>palette.aquatic</i> (grey to blue). See the <i>finplot</i> function for other inbuilt palettes. Use <i>colorRampPalette(c("deepskyblue2","red"))(50)</i> when <i>colourtype</i> is "HW" or "HW*" to match the <i>HWsigplot(s)</i> from the normal running of GBS-Chip-Gmatrix.R.
HDxlim	a numeric vector of length 2 with the x-coordinate limits. The default is (0,1) (the bounds for heterozygosity).
HDylim	a numeric vector of length 2 with the y-coordinate limits. The default is (-Inf,Inf) in which case the data bounds are used. If the limits are set inside the data bounds, points outside the limits are plotted as "^" and "v" on the closest limit.
HDcol	a character vector of length <i>nsnps</i> specifying the colour for each SNP. If given, these colours override the action of <i>colourtype</i> . The default is NULL.



`sortcol` an character option to plot the points in sorted colouring variable order (when *HDcol* is NULL), either ascending ("asc") or descending ("desc"). Any other value will plot in data order. Points plotted last will be more noticeable.

Details: An "HDplot", with SNPs coloured as specified is produced. This can be a useful graphic for detecting the presence of paralogous loci and for determining whether a SNP filter to be used will likely remove those SNPs.

*<plotname>.png* an HDplot of SNPs.

### Miscellaneous functions

These are functions to help make manipulations of the data easier.

Usage: `upper.vec (sqMatrix)`

Argument:

`sqMatrix` a square matrix

Details: Outputs numeric vector of upper triangular (off-diagonal) values, ordered by row then column.

#### Extract off-diagonal values from a square matrix (upper.vec)

This function extracts the upper triangular values from a square matrix and places them in a vector. This function is used by other functions (usually for comparing matrix values).

Usage: `upper.vec (sqMatrix, diag=FALSE)`

Argument:

`sqMatrix` a square matrix

`diag` if FALSE only include off-diagonal values, if TRUE include diagonal values as well

Details: Outputs numeric vector of upper triangular values, ordered by row then column.

#### Extracting first 'field' from seqID (seq2samp)

This function extracts the text, that precedes a specified delimiter, from a character variable. The normal use of this is to extract the first 'field' from *seqID*, when *seqID* contains information about the sequencing process separated by a delimiter (e.g. an underscore), and the first piece of information is a sample identifier. The sample identifier should not contain the delimiter.

Usage: `seq2samp (seqIDvec=seqID, splitby="_",...)`

Arguments:

`seqIDvec` a character vector, usually of *seqIDs*. The default is *seqID*

`splitby` the character to delimit the end of the text to be extracted. The default is an underscore (" \_ ")

... further arguments to be passed to `strsplit` (e.g., using `fixed=TRUE` will treat *splitby* as plain text rather than a regular expression)

Details: Outputs a character vector.

#### Colouring functions (colourby, changecol, colkey)

Some functions are provided to help specify plotting colours based on a factor or character variable. *colourby* creates a set of colours, *changecol* allows these to be modified and *colkey* plots a key to the colours.

Usage: `colourby (colgroup, groupsort=FALSE, maxlight=1, hclpals=character(0))`

Arguments:

`colgroup` a character vector or factor, whereby each level (unique value) will be given a different colour.

`groupsort` specifies whether the *colgroup* levels are sorted before assigning colours. The default is FALSE which uses the order encountered in *colgroup*.

`maxlight` A value in (0,1] limiting the 'lightness' of the colours assigned

**hclpals** a character vector containing colour palettes to use from `hcl.pals()` (instead of rainbow colours). Ignored if `hcl.colors` is not available. If more than one palette is specified, they are used sequentially and evenly, but there is no check for duplicated colours across the palettes.

**Details:** Outputs list with three elements:

**collabels** The discrete values in *colgroup*  
**collist** The set of corresponding colours to each element of *collabels*  
**sampcol** The set of colours in *collist*, corresponding to *colgroup*.

The function chooses a set of colours that span the rgb range. When there are more than 8 levels and if *hclpals* is empty, every 2<sup>nd</sup> colour is made greyer, to help distinguish the colours. Normally the function is applied to samples, with *colgroup* of length *nind* corresponding to each *seqID*, although it can be applied to other items, e.g. to the SNPs.

**Usage:** `changecol (colobject,colposition,newcolour)`

**Arguments:**

**colobject** a list that was created with *colourby*  
**colposition** the (set of) position(s) (integer(s)) in the *collabels* (and *collist*) element of *colobject* to be changed  
**newcolour** the (set of) new colours (character (vector)) to use in the *colposition* position(s) of the *collist* element of *colobject*

**Details:** Outputs a list with the same structure as *colobject*. The number of elements in *colposition* and *newcolour* should match.

**Usage:** `colkey (colobj, sfx="", srt.lab=0, plotch=16, horiz=TRUE)`

**Arguments:**

**colobj** a list that was created with *colourby* (or *changecol*)  
**sfx** text to be used in naming the output file  
**srt.lab** string rotation setting for the labels. Common values are 0 (the default, horizontal text) or 90 (vertical text).  
**plotch** an integer or vector of integers of length equal to the length of *colgroup* in *colobj*. Denotes the plotting symbol(s) to use. Defaults to 16 (solid circle).  
**horiz** a logical value. If TRUE (default), the symbols are drawn horizontally with the text below. If FALSE, the symbols are drawn vertically with the text to the right.

**Details:** A plot is produced.

*ColourKey<sfx>.png* A plot showing a set of dots coloured with *colobj\$collist* colours and labelled with *colobj\$collevels*.

## Pedigree analysis (GBSPedAssign.R)

The following table shows variables and commands that are specific to the pedigree program.

Variable command	/ Type <sup>1</sup>	Description
<b>pedfile</b>	V	Name of file containing pedigree and/or parent group information
<b>groupsfile</b>	V	Name of file containing which individuals are in which parent groups. This file is ignored if <i>matesfile</i> is defined.
<b>matesfile</b>	V	Name of file containing which mate pairs are in which parent groups
<b>GCheck</b>	V	The name (as a string) of the G matrix to use for parent verification or assignment. This must be set before calling GBSPedAssign.R.
<b>indsubset</b>	V	The subset of individuals used to calculate the matrix specified in <i>GCheck</i> .

<b>rel.thresh</b>	V	The relatedness threshold to use for parent verification or assignment, if the corresponding parent sex-specific threshold ( <i>rel.threshF</i> or <i>rel.threshM</i> ) has not been set. This has a default value of 0.4.
<b>rel.threshF</b>	V	The relatedness threshold to use for father verification or assignment. This has a default value of <i>rel.thresh</i> .
<b>rel.threshM</b>	V	The relatedness threshold to use for mother verification or assignment. This has a default value of <i>rel.thresh</i> .
<b>mindepth.mm</b>	V	Minimum depth to be used for calculating mismatch proportions in parent matching. Default is 1 (use all results).
<b>snpsubset</b>	V	The subset of SNPs to be used for calculating mismatch rates or for bootstrapping (usually the same set as used for calculating the matrix specified in <i>GCheck</i> ). Default is all SNPs.
<b>emm.thresh</b>	V	The excess mismatch rate threshold to use for parent assignment. This has a default value of 0.01.
<b>emm.thresh2</b>	V	The excess mismatch rate threshold to use for parent-pair assignment. This has a default value 2 x <i>emm.thresh</i> .
<b>emmdiff.thresh2</b>	V	The excess mismatch rate difference (from that for the most related father and most related mother) threshold to use for suggesting an alternate parent-pair assignment. This has a default value of 0.
<b>inb.thresh</b>	V	The lower threshold for the difference between parent relatedness and twice the estimated inbreeding to exclude a parent-pair match with the inbreeding check. This has a default value of 0.2.
<b>minr4inb</b>	V	The lower threshold on parent relatedness to exclude a parent-pair match with the inbreeding check. This has a default value of NULL (no minimum).
<b>boot.thresh</b>	V	If the relatedness with the 2 <sup>nd</sup> best parent is within <i>boot.thresh</i> of that for the best parent, a bootstrapping procedure will be invoked to further compare these possible matches. Default value of 0.05.
<b>depth.min</b>	V	Minimum mean depth of SNPs to be used for bootstrapping. Default value is 0.
<b>depth.max</b>	V	Maximum mean depth of SNPs to be used for bootstrapping. Default value is 0.
<b>puse</b>	V	Allele frequencies to be used bootstrapping. Default is to use <i>p</i> .
<b>nboot</b>	V	Number of bootstrap replicates. Default value is 1000.
<b>boota.thresh</b>	V	The upper threshold on bootstrap reliability for excluding a parent match with the bootstrapping check. This has a default value of 99.
<b>matchmethod</b>	V	The method used to find the best 2 matching parents (fathers and/or mothers). The default value is “rel” where the maximum relatedness is used. The alternative is “EMM” where minimum EMM is used. At this stage bootstrapping and alternate assignments were based on using “rel” so may not give sensible results with “EMM”.
<b>source</b>	C	Invoke GBS-PedAssign.R code to verify parents (if given) or assign parents (if parent groups are given)

<sup>1</sup> Type is V for a variable to be set, or C for a command to be invoked or function to be run.

This program uses a relatedness matrix and excess mismatch rate (EMM) results to verify given pedigrees and/or to find the best matching parents from groups of potential parents using the methods described in Dodds *et al.* (2019). Both these tasks require a pedigree file (with name given in *pedfile*). For parent matching a groups file (with name given in *groupsfile*) or a mate pairs file (with name given in *matesfile*) is also required. If there is a *matesfile*, these combinations are

used for checking and *groupsfile* (if given) is ignored. See below for the formats for these files. Father (Mother) verification is undertaken if the pedigree file contains a FatherID (MotherID) variable. Father (Mother) matching is undertaken if a groups file is given and the pedigree file contains a FatherGroup (MotherGroup) variable. The Group fields are read as text fields. Similarly, parent-pair matching is undertaken if a mates file is given and the pedigree file contains a MatesGroup field.

For parent matching, mismatch statistics are calculated for reporting and using, in addition to relatedness values, for assigning parentage. The 'raw' mismatch rate is the proportion of apparent (i.e. using observed genotypes) mismatches (i.e., genotypes inconsistent with parentage). 'Excess' rates are the differences between raw rates and rates that are expected given the genotype uncertainty due to the GBS process (manuscript in prep). A number of variables (see below) control how the mismatch rates are calculated and used. Mismatch rates are calculated for offspring-parent pairs and for offspring-parent trios (if matching to both parents). If both parents are being matched, the apparent parent-pair mismatch rates (offspring and parent genotypes incompatible) are given for each combination of the best two matching parents.

Before calling the program, the variable *GCheck* must be set to the name (as a string) of the G matrix to use. If this is for a subset of individuals, *indsubset* must be set to the indices of those individuals (as used in *calcG*). In addition, *rel.thresh* (and/or *rel.threshF* and/or *rel.threshM* for fathers and mothers, respectively) may be set to override the default relatedness value of 0.4 for declaring a parentage verification (or to allow parent assignment). A number of other variables control calculated results and reporting for parent matching. *mindepth.mm* may be set to override the default minimum depth (1) for a SNP for the individuals being compared when calculating (excess) mismatch rates for parentage matching. The default value is recommended for calculating excess rates, but raw rates are likely to be more useful when using a higher threshold. *snpsubset* may be set to indices of SNPs to be considered for use in calculating mismatch rates and for bootstrapping (see below, this will usually be the same subset as used for calculating the G matrix being used). The excess mismatch rate thresholds for declaring parentage are set by *emm.thresh* (parent-offspring pair; default value of 0.01) and *emm.thresh2* (parent-offspring trio; default value of twice *emm.thresh*). An alternative parentage is suggested when a possible pair (mother and father) have an excess mismatch rate that is lower than that for the best (i.e., most highly related) father and best mother by more than *emmdiff.thresh2* (default value of 0).

For parent pair matching, the estimated relatedness between the parent pairs (all four combinations of best and 2<sup>nd</sup> best matching fathers and mothers, or the best and 2<sup>nd</sup> best pairs, if *matesfile* is used) are calculated. The relatedness for the best matching pair of parents is compared with the estimated inbreeding for the individual. High values of parent relatedness (compared with the inbreeding of the individual) may indicate that one of the parents has been incorrectly assigned to a relative of the other parent. A parent-pair match will be excluded as a match if the parent relatedness exceeds offspring inbreeding by at least *inb.thresh* (default value 0.2).

A bootstrapping procedure is available to provide a metric on the closeness of parent-offspring match compared to that with the 2<sup>nd</sup> best parent. The procedure resamples SNPs (with replacement), recalculates the relatedness values (for the offspring and each of the two best parents) and reports the percentage of times that the best parent is still the better of the two among the bootstrap replicates. This should not be taken as a significance level test, as the resampled SNPs are not independent. As bootstrapping is quite time-consuming, it is invoked only when there are 2 possible parents with similar (within *boot.thresh*) parent-offspring relatedness values, and if the best parent exceeds the relatedness and excess mismatch thresholds. The number of bootstrap replicates is set by *nboot* (default value 1000). Three other variables (*depth.min*, *depth.max*, *puse*) mirror those used in *calcG* to allow the bootstrapping to calculate relatedness in the same way as was used for the G matrix being used in parentage assignment. These variables should be set to the same values as those used for calculating the G matrix. An assignment is flagged (see below) if the best parent is the better one in the bootstrap samples in less than *boota.thresh* percent (default value 99) of the replicates. Bootstrapping is not conducted if *matesfile* is used.

The output files contain variables to indicate whether the parentage should be accepted. These variables are called *FatherAssign* and *MotherAssign* for single parent matching of fathers and mothers, respectively. The codes used as values for these variables are:

Assign code	Description
<b>N</b>	Relatedness estimate for best matching parent is below <i>rel.threshF</i> or <i>rel.threshM</i> (for fathers and mothers, respectively).
<b>E</b>	Excess mismatch rate for best matching parent exceeds <i>emm.thresh</i> .
<b>A</b>	Alternate assignment: the 2 <sup>nd</sup> best parent appears acceptable. This parent has relatedness exceeding <i>rel.threshF</i> or <i>rel.threshM</i> (for fathers and mothers, respectively) and excess mismatch rate that is lower than <i>emm.thresh</i> when the best parent had excess mismatch rate exceeding this threshold.
<b>B</b>	Best matching parent is the better one in less than <i>boota.thresh</i> % of the bootstrap replicates.
<b>Y</b>	Best matching parent passes all assignment criteria

The variable for indicating whether a parent-pair match should be accepted is *BothAssign* and takes values as shown:

Assign code	Description
<b>N</b>	Relatedness estimate for best matching parent is below <i>rel.threshF</i> or <i>rel.threshM</i> (for fathers and mothers, respectively).
<b>M</b>	Mother assigned, father not assigned.
<b>F</b>	Father assigned, mother not assigned.
<b>E</b>	Excess mismatch rate for best matching parent-pair exceeds <i>emm.thresh2</i> , except when one parent assigned and the other has an E code, then the parent assignment is made.
<b>A</b>	An alternate parent-pair appears acceptable. This pair has excess mismatch rate less than <i>emm.thresh2</i> and lower than that for the best parent-pair by more than <i>emmdiff.thresh2</i> . If the alternate pair also passes the other checks, the pair is indicated by the value of <i>Alternate</i> , e.g. a value of F1M2 indicates that the alternate pair is the best father and 2 <sup>nd</sup> best mother.
<b>B</b>	At least one of the parents has a B code. (It may still be possible to assign the other parent).
<b>I</b>	The best parent-pair relatedness exceeds twice the offspring inbreeding by at least <i>inb.thresh</i> , and is above <i>minr4inb</i> (if that threshold has been set). An alternate pair may be indicated by the value of <i>Alternate</i> , similarly to the A code offspring.
<b>Y</b>	Best matching parent passes all assignment criteria

Where more than one of the assign codes is possible, the one that ranks the highest (in the order given in the above tables) is used.

This program outputs summary statistics and a number of files. The %s of verified fathers and mothers are given, as well as the mean relatedness estimates for matching and non-matching fathers and mothers. The files, where relevant, are as follows:

*PedVerify.csv* returns the pedigree file with additional columns, as shown below:

Variable name	Description
<b>FatherRel</b>	Relatedness estimate between individual and it's specified father
<b>FatherEMM</b>	The specified father-offspring EMM

<b>FatherMatch</b>	TRUE if <i>FatherRel</i> > <i>rel.threshF</i> and <i>FatherEMM</i> < <i>emm.thresh</i>
<b>MotherRel</b>	Relatedness estimate between individual and it's specified mother
<b>MotherEMM</b>	The specified mother-offspring EMM
<b>MotherMatch</b>	TRUE if <i>MotherRel</i> > <i>rel.threshM</i> and <i>MotherEMM</i> < <i>emm.thresh</i>
<b>FandMEMM</b>	The specified parent pair – offspring EMM
<b>FandMmatch</b>	TRUE if <i>FatherMatch</i> and <i>MotherMatch</i> are both TRUE and <i>FandMEMM</i> < <i>emm.thresh2</i>

*FatherVerify.png* is a scatterplot matrix showing *FatherRel*, *FatherEMM* (see above), the position of the individual in the pedigree file and the position of the recorded father in the pedigree file. This is useful for seeing the distribution of relatedness values, and possibly for detecting sample tracking issues (if the order in the pedigree file relates to the order samples are processed at a particular stage).

*MotherVerify.png* is a scatterplot matrix like *FatherVerify.png* but for mother verification.

*FatherMatches.csv* shows the results of the father matching. It returns the first two columns of the pedigree file with additional columns, as shown below:

Variable name	Description
<b>BestFatherMatch</b>	IndivID of the father from the <i>FatherGroup</i> having the highest estimated relatedness to the individual (or lowest EMM, if <i>matchmethod</i> is "EMM").
<b>FatherMatch2nd</b>	IndivID of the father from the <i>FatherGroup</i> having the 2 <sup>nd</sup> highest estimated relatedness to the individual (or 2 <sup>nd</sup> lowest EMM, if <i>matchmethod</i> is "EMM")
<b>Fatherrel</b>	The estimated relatedness for <i>BestFatherMatch</i>
<b>Fatherrel2nd</b>	The estimated relatedness for <i>FatherMatch2nd</i>
<b>Father12rel</b>	The estimated relatedness between <i>BestFatherMatch</i> and <i>FatherMatch2nd</i> .
<b>mmrateFather</b>	The (raw) mismatch rate for <i>BestFatherMatch</i>
<b>mmnumFather</b>	The number of snps used to calculate <i>mmrateFather</i>
<b>exp.mmrateFather</b>	The expected mismatch rate for <i>BestFatherMatch</i>
<b>mmrateFather2</b>	The (raw) mismatch rate for <i>FatherMatch2nd</i>
<b>exp.mmrateFather2</b>	The expected mismatch rate for <i>FatherMatch2nd</i>
<b>Fathersd</b>	The bootstrap sd of <i>Fatherrel</i> values (for bootstrapped cases, the variable is present only if there are bootstrapped caess)
<b>FatherReliability</b>	The % of bootstrap results where <i>Fatherrel</i> > <i>Fatherrel2nds</i> (for bootstrapped cases, the variable is present only if there are bootstrapped caess)
<b>FatherAssign</b>	The code for father assignment.

*MotherMatches.csv* shows the results of the mother matching (with columns as for *FatherMatches.csv* but for mothers instead of fathers).

*BothMatches.csv* shows the results of both father and mother matching (for individuals with both *FatherGroup* and *MotherGroup*). It contains the columns of *FatherMatches.csv* and *MotherMatches.csv* with additional columns, as shown below:

Variable name	Description
<b>mmrateF&lt;fatherrank&gt;M&lt;motherrank&gt;</b>	The (raw) mismatch rate for possible parent matches, where <fatherrank> is 1 to indicate <i>BestFatherMatch</i> and 2 to indicate <i>FatherMatch2nd</i> , and similarly for <motherrank>.



<b><i>mmnumF&lt;fatherrank&gt;M&lt;motherrank&gt;</i></b>	The number of SNPs used to calculate <i>mmrateF&lt;fatherrank&gt;M&lt;motherrank&gt;</i>
<b><i>exp.mmrateF&lt;fatherrank&gt;M&lt;motherrank&gt;</i></b>	The expected mismatch rate corresponding to <i>mmrateF&lt;fatherrank&gt;M&lt;motherrank&gt;</i>
<b><i>relF&lt;fatherrank&gt;M&lt;motherrank&gt;</i></b>	The estimated relatedness between the pair of possible parents
<b><i>Inb</i></b>	The estimated inbreeding of the offspring
<b><i>BothAssign</i></b>	The code for the parent-pair assignment
<b><i>Alternate</i></b>	An alternative (to F1M1) parent pair

*GroupsParentCounts.csv* returns the groups file with additional columns, as shown below:

Variable name	Description
<b><i>FatherFreq</i></b>	Number of offspring where this father is the <i>BestFatherMatch</i> in this group
<b><i>MotherFreq</i></b>	Number of offspring where this mother is the <i>BestMotherMatch</i> in this group

*MatePairMatches.csv* shows the results of father and mother pair matching (for individuals with *MatesGroup*). It contains the a subset of the columns of *BothMatches.csv*, given above (*BothMatches.csv* is not written with a mate pairs analysis). There are no *FatherAssign* or *MotherAssign* fields, no bootstrap results and no information parent pairs across the best and 2<sup>nd</sup> best matches (e.g. no F1M2 results). The fields occur in a different order to those in *BothMatches.csv*.

*BestFatherMatches.png* is a plot of the raw mismatch rate for *BestFatherMatch* against the estimated relatedness (*Fatherrel*). Points are coloured using *fcolo* and a grey vertical line indicates the value of *rel.thresh* used.

*BestFatherMatchesE.png* is the same *BestFatherMatches.png* except that the excess mismatch rate is plotted. A grey horizontal line indicates the value of *emm.thresh* used.

*Best2FatherMatches.png* is a plot of the estimated relatedness for *FatherMatch2nd* (*Fatherrel2nd*) against that for *BestFatherMatch* (*Fatherrel*). Points are coloured using a scale based on the excess mismatch rate (*mmrateFather - exp.mmrateFather*) for father-offspring and the line of equality is drawn (by definition all points fall below the line). Vertical and horizontal grey lines indicate the value of *rel.threshF* or *rel.threshM* (for fathers and mothers, respectively) used.

*ExpMM-Father.png* is a plot of the raw mismatch rate against the expected mismatch rate for *BestFatherMatch*. A red line shows where these are equal and a grey line shows the boundary for an E assign code. Points are coloured using *fcolo* and the symbols indicate *FatherAssign*.

*BestMotherMatches.png*, *BestMotherMatchesE.png*, *Best2MotherMatches.png* and *ExpMM-Mother.png* are the corresponding plots to *BestFatherMatches.png*, *BestFatherMatchesE.png* and *Best2FatherMatches.png* and *ExpMM-Father.png*, respectively, for mothers.

*ParRel-Inb.png* is a plot of estimated parent-pair relatedness against offspring estimated inbreeding. Points are coloured according to the mean depth in the offspring (as depth is more critical for inbreeding than relatedness estimation), and with a symbol corresponding to *BothAssign* (see *ExpMM-Both.png* for a key).

*MMrateBoth.png* is a scatterplot matrix plot of the four combinations of parent-pair raw mismatch rates that were saved in *BothMatches.csv*. Points are coloured using *fcolo* and the lines of equality are drawn (in red).

*MMrateBothE.png* is a scatterplot matrix plot of the four combinations of parent-pair excess mismatch rates (or a single scatter plot of 2<sup>nd</sup> versus best EMM, when *matesfile* is used). Points

are coloured using *fcolo*, the lines of equality are drawn (in red) and the symbols for the points denote *BothAssign*. The key for the symbols can be found in ExpMM-Both.png.

*ExpMM-BothE.png* is a plot of raw versus expected parent-pair mismatch rates. Points are coloured using *fcolo* and the symbols for the points denote *BothAssign*.

### EMM sum of squared for beta-binomial model (*ssbbmm*)

A function to use for determining the fit of a beta-binomial model in terms of trio matches. Should only be run after a trio parentage assignment. *Under development and likely to change in future updates.*

Usage: `ssbbmm(bbpar, ,uuse=uY)`

Arguments:

<code>bbpar</code>	the parameter for the beta-binomial model
<code>uuse</code>	the offspring set to use (positions in <i>BothMatches</i> ) as true trios

Details: Returns the sum of squared trio EMM values for the `uuse` offspring and their assigned parents. The function can be used to estimate the beta-binomial parameter, e.g. with `optimize(ssbbmm,lower=0,upper=20, tol=0.001)`

### EMM sum of squared for modified p model (*ssmpmm*)

A function to use for determining the fit of a modified p model in terms of trio matches. Should only be run after a trio parentage assignment. *Under development and likely to change in future updates.*

Usage: `ssmpmm(mppar, ,uuse=uY)`

Arguments:

<code>mppar</code>	the parameter for the modified p model
<code>uuse</code>	the offspring set to use (positions in <i>BothMatches</i> ) as true trios

Details: Returns the sum of squared trio EMM values for the `uuse` offspring and their assigned parents. The function can be used to estimate the modified p parameter, e.g. with `optimize(ssmpmm, lower=0.5,upper=0.8, tol=0.001)`

### Add tagID function (*addtagIDs*)

This is a function will add alternative IDs for offspring, father and mother.

Usage: `addtagIDs(sampinfo, indvar, matchtype ="both")`

Arguments:

<code>sampinfo</code>	a dataframe containing the relevant IDs for all individuals
<code>indvar</code>	quoted text giving the name of the variable in <code>sampinfo</code> that corresponds to <code>IndivID</code> in the pedigree file
<code>tagvar</code>	quoted text giving the name of the variable in <code>sampinfo</code> that contains the tag (identifier) to be returned
<code>matchtype</code>	one of "both", "father" or "mother" (not case-sensitive) specifying which data frame to update

Details: Returns a data frame with the additional IDs added. It is not output to a file.

### Parentage PC plot function (*bestparPCA*)

This is a function generates a PC2 vs PC1 plot with lines joining each progeny with its best father and mother.

Usage: `bestparPCA(Gobj, sfx="", keypos=NULL)`

Arguments:

<code>Gobj</code>	an object produced from <code>calcG</code> (usually the same one used to obtain the <code>GCheck</code> matrix for parentage), with $npc \geq 2$
-------------------	--

sfx                    text to be included in output file name to allow output from multiple calls or runs to be identified

keypos                the location (if given) on the plot of the legend of assign codes, using a value as accepted by the legend command (e.g. "topleft")

Details: Generates a PC plot.

*PC-BestParents<sfx>.png*      a plot of PC2 vcs PC1 relating to the parentage analysis. Points are coloured according to fcolo. Parents are shown as dots and offspring have symbols representing their assignment codes. Blue and pink lines are drawn from best father and best mother, respectively, to offspring.

## Population genetics analysis (GBS-PopGen.R)

This R code makes available some functions for population genetics analyses. These are currently under development and only a brief description is provided here. The methods and syntax of this code is likely to change in the future.

### Heterozygosity measures (*heterozygosity*)

This function gives various measures of observed and expected heterozygosity. keep.alleles should be set to TRUE to use this function.

Usage: heterozygosity(indsubsetgf=1:nind,snpsubsetgf=1:nsnps,maxiter=100,convtol=0.001)

Arguments:

indsubsetgf          a vector of integers (between 1 and *nind*, inclusive) of individuals to use for the heterozygosity measures. The default is to use all individuals.

snpsubsetgf          a vector of integers (between 1 and *nsnps*, inclusive) of SNPs to use for the heterozygosity measures. The default is to use all SNPs.

maxiter                maximum number of iterations to use in the estimation process. The default is 100.

convtol                convergence tolerance – the difference between genotype frequency estimates in successive iterations that is sufficiently small to assume convergence. The default value is 0.001.

Details: A data frame is returned with a (unlabelled) row for each SNP and the columns:

Variable name	Description
<b>ohetstar</b>	Observed heterozygosity on the raw scale
<b>ehetstar</b>	Observed heterozygosity on the raw scale (the proportion of genotype results expected to contain reads from both alleles)
<b>ohet</b>	Observed heterozygosity (estimated) on the true genotype scale
<b>ohet2</b>	An alternative measure of <i>ohet</i>
<b>ehet</b>	Expected heterozygosity (estimated) on the true genotype scale

### F<sub>ST</sub> calculations (*Fst.GBS* and *Fst.GBS.pairwise*)

These functions calculate approximate F<sub>ST</sub> (estimates) accounting for read depth.

Usage: Fst.GBS(snpsubset, indsubset, populations, varadj=0) and/or Fst.GBS.pairwise(snpsubset, indsubset, populations,sortlevels=TRUE, ...)

Arguments:

snpsubset            a vector of integers (between 1 and *nsnps*, inclusive) of SNPs to calculate F<sub>ST</sub> for. The default is to use all SNPs.

indsubset            a vector of integers (between 1 and *nind*, inclusive) of individuals to use for the calculations. The default is to use all individuals.

populations          a vector of length *nind* containing population labels

varadj                use varadj=1 to get Fst as Weir p166, varadj=0 for usual Fst.

sortlevels            determines whether populations are listed as encountered in *populations* (sortlevels=FALSE) or sorted (sortlevels=TRUE)

... arguments passed to Fst.GBS (currently only *varadj*)

**Details:**  $F_{ST}$  with depth adjustment. The adjustment is “approximate” and may result in estimates outside [0,1]. The *.pairwise* version calculates the statistics for each pair of populations. The (pairwise) means and medians are displayed. The values for each SNP are returned in a vector (Fst.GBS) or three-dimensional array (Fst.GBS.pairwise) with first two dimensions being the population and the third dimension being the SNP.

### MAF plots by population (*popmaf*)

Plot minor allele frequency distributions by population.

**Usage:** *popmaf*(*snpsubset*, *indsubset*, *populations*=NULL, *subpopulations*=NULL, *indcol*, *colobj*, *minsamps*=10, *mafmin*=0, *sortlevels*=TRUE, *unif*=FALSE)

**Arguments:**

<i>snpsubset</i>	a vector of integers (between 1 and <i>nsnps</i> , inclusive) of SNPs calculate $F_{ST}$ for. The default is to use all SNPs.
<i>indsubset</i>	a vector of integers (between 1 and <i>nind</i> , inclusive) of individuals to use for the calculations. The default is to use all individuals.
<i>populations</i>	a vector of length <i>nind</i> containing population labels. The default is NULL in which case a MAF plot for all <i>indsubset</i> individuals is given.
<i>subpopulations</i>	a vector of length <i>nind</i> containing subpopulation labels. These are treated as being nested within <i>populations</i> .
<i>indcol</i>	A colour assignment for each individual. The plot is coloured by <i>indcol</i> if all individuals in that population have that colour (otherwise black).
<i>colobj</i>	an object created by <i>colorby</i> (undocumented). If present it will be used for populations and their colours.
<i>minsamps</i>	Minimum number of samples in a (sub)population to invoke plotting. The default is 10.
<i>mafmin</i>	Minimum (sub)population MAF to include a SNP in the plot. The default is 0.
<i>sortlevels</i>	determines whether data are processed by populations as encountered ( <i>sortlevels</i> =FALSE) or sorted ( <i>sortlevels</i> =TRUE)
<i>unif</i>	determines whether the plots are drawn with the same vertical axis range. The default is FALSE (population-specific range).

**Details:** A MAF distribution is plotted, possibly with a different colour for each population, and different shading for each subpopulation. Some summary statistics are also displayed.

### Manhattan plots (*manhatplot*)

Simple plotting of results as a Manhattan plot.

**Usage:** *manhatplot*(*value*, *chrom*, *pos*, *plotname*, *qdistn*=*qunif*, ...)

<i>value</i>	vector of statistic values to be plotted (for each SNP).
<i>chrom</i>	chromosome name (numeric or character) for each SNP.
<i>pos</i>	position (numeric) on chromosome for each SNP.
<i>plotname</i>	text used as prefix for names of output plots
<i>keyrot</i>	rotation angle for chromosome key. The default is 0, but 90 is a better choice for longer chromosome labels.
<i>qdistn</i>	function name of null distribution. The default is the uniform distribution ( <i>qunif</i> ) which could be used e.g., with p-values. A more common example would be the chi-squared distribution ( <i>qchisq</i> ).
...	further arguments passed to <i>qdistn</i> , for example the <i>df</i> (degrees of freedom) parameter (if using <i>qdistn</i> = <i>qchisq</i> ).

**Details:** Two plots are produced.

*<plotname>-Manhat.png* a Manhattan plot of *value*. Points are sorted by *chrom* and *pos* with a different colour for each value of *chrom*.

`<plotname>-QQ.png` a QQ plot *value* using the distribution specified in *qdistn*.

### Pairs of SNPs from different chromosomes (*snpselection*, *snpselectionUR*)

These functions allow pairs of SNPs from different chromosomes to be selected, normally for use in calculating linkage disequilibrium between unlinked SNPs to allow effective population size estimates.

**Usage:** *snpselection* (chromosome, position, nsnpupperchrom=100, seltype="centre", randseed=NULL, snpsubset, chromuse)

chromosome	a character vector of the chromosome name for each SNP
position	numeric vector of position (normally in bp) on chromosome for each SNP
nsnpupperchrom	the (maximum) number of SNPs to choose on each chromosome. The default is 100.
seltype	the method to select SNPs from those available on a chromosome. Available values are "centre" – choose the <i>nsnpupperchrom</i> SNPs closest to the centre (mean SNP position) of the chromosome; "even" – space the SNPs as evenly as possible (by SNP order on the chromosome, not using position values); or "random" – randomly selected
randseed	a seed value for the random number generator (to allow reproducible selection using "random" <i>seltype</i> )
snpsubset	a vector of integers (between 1 and <i>nsnps</i> , inclusive) of SNPs to choose from
chromuse	a character vector of the chromosomes (names, as in <i>chromosome</i> ) to use. The default is to use all chromosomes. For example, this option allows restricting the selection of SNPs to the mapped autosomal SNPs.

**Details:** the output is an array with two columns of SNP positions, containing all pairs of selected SNPs where the pair are on different chromosomes.

**Usage:** *snpselectionUR* (URobj, nsnpupperchrom=100, nchrom, ...)

URobj	a UR (unrelated samples) object from the <a href="#">GUSbase</a> package.
nsnpupperchrom	the (maximum) number of SNPs to choose on each chromosome. The default is 100.
nchrom	the number of chromosomes to use. If specified, the first <i>nchrom</i> chromosomes are used. The default is to use all chromosomes.
...	additional parameters to be passed to <i>snpselection</i> .

**Details:** provides convenient access to the *snpselection* function for a UR object in GUSbase.

### Effective population size (*Nefromr2*)

The *Nefromr2* function is experimental may be enhanced and/or modified at a later date. Calculates effective population size ( $N_e$ ) from a set of linkage disequilibrium  $r^2$  values. Assumes that these are from unlinked pairs of SNPs, and that the  $N_e$  is for the generation of the genotyped individuals (i.e. present day, rather than historic). The [GUS-LD](#) package (Bilton *et al.*, 2018) provides a method to estimate linkage disequilibrium while accounting for read depth and sequencing error.

**Usage:** *Nefromr2* (r2auto, nLD, alpha=1, weighted=FALSE, minN=1)

r2auto	a vector of linkage disequilibrium $r^2$ values UR (unlinked autosomal pairs of SNPs).
nLD	the number of individuals used to calculate the $r^2$ values. Either a vector of the same length as <i>r2auto</i> or a single value (assumed to be the same for all pairs of SNPs).
alpha	the $\alpha$ (mutation) parameter. The default value is 1.
weighted	a setting to invoke using a weighted mean of $r^2$ values (when TRUE) in the calculations. The default is FALSE.
minN	a minimum number of individuals used to retain a pair of SNPs. The default value is 1 (i.e., no minimum).

Details: a set of statistics is displayed:

n	the mean sample size for a pair of SNPs
Neauto	the estimated Ne based on mean $r^2$ , without bias correction
Neauto.adj.b1	the estimated Ne with bias correction using $\beta=1$ (for phase unknown data)
Neauto.adj.b2	the estimated Ne with bias correction using $\beta=2$ (for phase known data)
Neauto.med	the estimated Ne based on the median $r^2$ (without bias correction)
Neauto.med.adj.b1	the estimated Ne using medians, with 'bias correction' using $\beta=1$ . For illustration only. Do not use.
Neauto.med.adj.b2	the estimated Ne using medians, with 'bias correction' using $\beta=2$ . For illustration only. Do not use.

See Barbato *et al.* (2015) for a description and definition of  $\alpha$  and  $\beta$ .

## Input formats

The genotype input format is set with *gform*, one of “uneak” (the default), “Tassel”, “TagDigger” or “Chip”.

### GBS via UNEAK

The default input format ('uneak') is a 'hapmap count' formatted file as produced by the UNEAK pipeline (Lu *et al.* 2013). This is a tab-separated flat text file with the first column being the SNP identifier, then a column for each genotyped individual (or sample, or other genotyping unit), followed by 5 columns of summary information (HetCount\_allele1, HetCount\_allele2, Count\_allele1, Count\_allele2, Frequency). Only the last of these 5 is used. Each row is for a different SNP. The column for each individual contains the genotype information as the allele depth (number of reads of that allele) for the 'reference' and 'alternate' alleles, respectively. The designation of reference and alternate is arbitrary for this software. The numbers of reads are separated by a pipe symbol (“|”). There is a header line, which, for the genotype columns, is taken as the identifiers of the individuals.

### GBS via Tassel

An additional format ('Tassel') is available that may be easier to use for GBS data that has been manipulated in Tassel. It is similar to the uneak format, but allele depths in a genotype are separated by a comma (“,”), has two columns before genotype data (, and no columns following the genotype data. The first two columns are the chromosome and position (which together, separated by an underscore, serve as the SNP identifier), respectively. As with the “uneak” format, this is a tab-separated flat text file with a header row.

### GBS via TagDigger

TagDigger (<https://github.com/lvclark/tagdigger>, Clarke and Sacks, 2016) is a tool for SNP calling from a given set of tags (sequences). It is likely to be used in a production environment, where the set of SNPs being called is unlikely to change much with additional samples being added. The 'TagDigger' format requires a comma delimited file with sample results in rows and SNP results in pairs of columns (count of reference allele, count of alternate allele). The first column contains the sample identifier. The header row, apart from the first value, contains SNP/allele identifiers. It is assumed that these identifiers have a SNP identifier followed by an underscore, followed by the allele identifier. The text preceding the underscore is taken as the SNP name (the other text is ignored).

TagDigger files will be read with the fread function from the data.table package, if that package is installed. This is faster than the method used when the package is not available. Files compressed



with the gzip (.gz) format can be read by both methods on linux platforms, but not for fread on other platforms.

## GBS via ANGSD

ANGSD (<http://www.popgen.dk/angsd>, Korneliussen *et al.*, 2014) is a program for analysing sequencing data, and can output SNP information. The 'ANGSDcount' format reads files created by the `-dumpCounts 4` option of ANGSD. This file has a header row, followed by a row for each SNP. There is a column for each of the 4 possible alleles (A, C, G, T) for each SNP and sample. The columns for a sample are together. The header contains an identifier for each column consisting of the sample identifier followed by underscore and the allele (e.g. ind0\_A). After reading this file, SNPs are checked for which alleles are most common. The two most common alleles are taken as the variant of interest, and other alleles are ignored, except that a SNP is discarded if the proportion of reads for the third most common alleles exceeds the threshold `triallelic.thresh`. SNPs are named as 'SNP' followed by the zero-padded position. SNPs that have been dropped by the triallelic threshold can be identified by finding gaps in the SNP\_Name sequence.

## vcf files

A python helper script `vcf2ra.py` is available to convert .vcf files (named `<infile>.vcf`) to the 'Tassel' format (named `<infile>.vcf.ra.tab`). The .vcf file must have either the AD (allelic depth) field, or both the AO (alternate allele observation count) and RO (reference allele observation count) fields.

Some sites are filtered out and placed in ancillary output files:

- indels are removed and reported in `<infile>.vcf.indel`,
- SNPs that are more than biallelic are removed and reported in `<infile>.vcf.polyallele`
- redundant sites are removed and reported in `<infile>.vcf.posred`

## Chip

Fully recorded genotypes can be entered via the "Chip" format. This comma-separated format has results for each individual in the rows and SNP results in a column. There is a header row (SNP identifiers) and the first column contains individual identifiers. Subsequent columns contain the SNP results. Genotype data is given in 0/1/2 format, representing first homozygote, heterozygote and second homozygote, respectively. Designation of which allele is the 'first' is arbitrary.

## Pedigree file

An optional pedigree file (given by *pedfile*) can be given, and will be used to verify or find parent matches. This is a comma separated file (csv). All individuals to be considered as offspring or parents need to have a row in this file. The columns of this file are specified below. The names must be exactly as specified. Additional columns may be present in the file.

Variable name	Required?	Description
<b>IndivID</b>	Y	identifies individuals in the pedigree and groups files
<b>seqID</b>	Y	matches <i>IndivID</i> to the identifier in the genotype file
<b>FatherID</b>	N	Recorded <i>IndivID</i> of father
<b>MotherID</b>	N	Recorded <i>IndivID</i> of mother
<b>FatherGroup</b>	N	Group label for group of potential fathers for the given <i>IndivID</i>
<b>MotherGroup</b>	N	Group label for group of potential mothers for the given <i>IndivID</i>
<b>MatesGroup</b>	N	Group label for group of potential parent (mate) pairs for the given <i>IndivID</i> .

Father and mother group labels should be distinct. If required, they are entered for the progeny. The information linking these labels to the set of possible parents is placed in the groups file.

Similarly, if *matesfile* is given, the *MatesGroup* field needs to give the label to identify possible parent pairs in the mates file.

## Groups file

If parent matching is required, then a groups file (or a mates file, see below) (given by *groupsfile*) describing the group labels in the pedigree file is required. This is a comma separated file (csv). The columns (both required) of this file are specified below. The names must be exactly as specified. Additional columns may be present in the file.

Variable name	Description
<b><i>IndivID</i></b>	identifier for potential parent, matching <i>IndivID</i> in the pedigree file
<b><i>ParGroup</i></b>	Group label for the group that <i>IndivID</i> belongs to

There should be one row for each group a potential parent belongs to.

## Mates file

When parent matching is required and the mating pairs are known, then a mates file (given by *matesfile*) describing the mate group labels in the pedigree file is required. This is a comma separated file (csv). The columns (all required) of this file are specified below. The names must be exactly as specified. Additional columns may be present in the file.

Variable name	Description
<b><i>MaleID</i></b>	identifier for potential male parent, matching <i>IndivID</i> in the pedigree file
<b><i>FemaleID</i></b>	identifier for potential female parent, matching <i>IndivID</i> in the pedigree file
<b><i>MatesGroup</i></b>	Group label for the group of mate pairs that could be the parent of <i>IndivID</i>

There should be one row for each group a potential parent belongs to.

## Optional packages

The software has been designed to run without the need for any R packages to be installed, but can use such packages if available. Sometimes there will be messages relating to these packages, but these messages can be ignored. A list of optional packages and their use follows.

Package name	Usage
<b><i>Rcpp</i></b>	Various functions have C++ versions for improved efficiency, but require this package
<b><i>RcppArmadillo</i></b>	This package is required for some of the C++ functions to be used
<b><i>data.table</i></b>	Reading tagdigger files, writing VCF files
<b><i>plotly</i></b>	Interactive graphics output from calcG
<b><i>heatmaply</i></b>	Interactive heatmap from calcG
<b><i>parallelDist</i></b>	Parallelized calculation of distance for the heatmap in calcG
<b><i>MethComp</i></b>	Bland-Altman plots in GCompare

## Example

This folder contains an example run (possibly using an earlier version). Files in directory :  
GBSRun.R HapMap.hmc.txt.gz Ped-GBS.csv Ped-Groups.csv

GBSRun.R

```
genofile <- "HapMap.hmc.txt.gz"
```

```

source("<source directory>/GBS-Chip-Gmatrix.R")
Gfull <- calcG()
GHwdgm.05 <- calcG(which(HWdis > -0.05),"HWdgm.05",npc=4) # recalculate
using Hardy-Weinberg disequilibrium cut-off at -0.05

pedfile <- "Ped-GBS.csv"
groupsfile <- "Ped-Groups.csv"

rel.thresh <- 0.2
GCheck <- "GHwdgm.05$G5"
source("<source directory>/GBSPedAssign.R")

```

<source directory> should be replaced with the location of the relevant .R files before running.

linux command:

R CMD BATCH --no-save GBSRun.R &

Files in directory after running code:

AlleleFreq.png	GcompareHWdgm.05.png	MotherVerify.png
Best2FatherMatches.png	Gcompare.png	PC1v2G5HWdgm.05.png
Best2MotherMatches.png	Gdiagdepth.png	PCG5HWdgm.05.pdf
BestFatherMatchesE.png	G-diag.png	Ped-GBS.csv
BestFatherMatches.png	GHwdgm.05diagdepth.png	Ped-Groups.csv
BestMotherMatchesE.png	GHwdgm.05-diag.png	PedVerify.csv
BestMotherMatches.png	GroupsParentCounts.csv	SampDepthCR.png
CallRate.png	HapMap.hmc.txt.gz	SampDepthHist.png
Co-call-Hwdgm.05.png	Heatmap-G5HWdgm.05.png	SampDepth.png
Co-call-.png	Heatmap-G5.png	SampDepth-scored.png
ExpMM-Father.png	HighRelatedness.csv	SampleStats.csv
ExpMM-Mother.png	HWdisMAFsig.png	seqID.csv
FatherMatches.csv	LRT-hist.png	SNPCallRate.png
FatherVerify.png	LRT-QQ.png	SNPDepthHist.png
finplot.png	MAFHWdgm.05.png	SNPDepth.png
GBSRun.R	MAF.png	
GBSRun.Rout	MotherMatches.csv	

A workshop using this example was given at the 2015 MapNet meeting (Rotorua, New Zealand). Instructions (KGDCourseInstructions-Mapnet2015.pdf) and course notes (KGDCourse-Mapnet2015.pdf) are available in the Example folder.

## ParExample

The folder gives an example of the code for a parentage analysis, based on the example given in Dodds *et al.* (2019). Example code is given in GBSParDeer.R. The example code assumes all necessary files are in the working directory. The example data (allele counts, pedigree file, groups file) can be obtained from <https://gsajournals.figshare.com/s/7ca45accf6ae82047c86>. An annotated description of the commands in GBSParDeer.R is in GBSParentage-Annotated.pdf.

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