**The importance of light and mycorrhizal fungi in determining the distribution of epiphytic orchids in mega-diverse tropical forests**

# Rationale and positioning with regard to the state-of-the-art

Understanding the various **factors that limit distribution** of plant and animal species is a longstanding question in ecology (Krebs, 1972). In land plants, the distribution of a species is simultaneously limited by climate, dispersal, and biotic interactions (Krebs, 1972). The most relevant climatic factors restricting plant species distribution globally are temperature and humidity. At local scales, microsite conditions such as light and substrate physicochemical quality play an important role as well (Krebs, 1972). Dispersal determines how far seeds are dispersed from mother plants and is limited by the presence of barriers that impede their dissemination (Nathan & Muller-landau, 2000). Biotic interactions can limit or increase species distributions. Antagonistic interactions such as interspecific competition will exclude some species from a given site, while mutualisms such as mycorrhizal symbioses can contribute to expand plant species distribution areas (Krebs, 1972; Dunn *et al.*, 2009; Afkhami *et al.*, 2014).

Whereas the effects of environmental conditions and dispersal on species distributions are relatively well understood, the ecological effects of **obligate inter-specific interactions**, such as symbioses, are far less understood (Dunn *et al.*, 2009; Afkhami *et al.*, 2014). Recent evidence suggests that the distribution ofspecies that rely on other species to complete their life cycle depends on the presence of suitable partners (partner availability)(Slatyer *et al.*, 2013) and on the level of specialization (partner breadth) (Batstone *et al.*, 2018). Species that either associate with widespread partners or are generalist towards them are expected to have a broader spatial distribution, whereas species that associate with a limited number of partners or with partners that only sporadically occur in nature, are expected to have a narrow distribution. Furthermore, a species can interact with more than one type of partner at the same time, such as mycorrhiza, pollinators, or dispersal agents, which adds complexity to the determinants of species distribution.

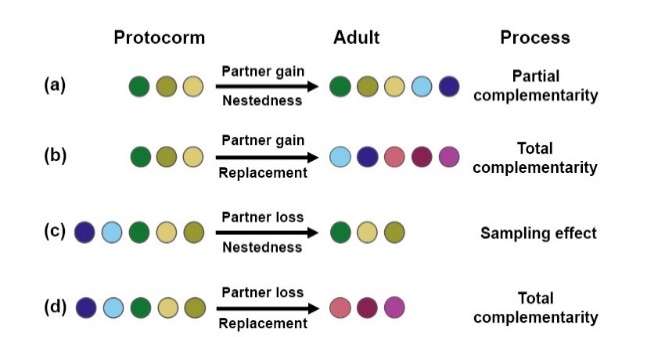
Current understanding of the determinants of species distribution is based mostly on the biology of the adult individuals. However, in sessile organisms such as plants, the distribution of adults can largely depend on the species' requirements during the earliest developmental life stages (the **regeneration niche** (Grubb, 2018)). The regeneration niche includes preferences for different substrates (Lusk, 1995), differential responses of seedlings to ecological gradients (Denslow, 1980), and the availability of microscopic mutualistic organisms (McCormick & Jacquemyn, 2014). Local availability of mutualists, particularly of symbiotic fungi, is one of the least studied aspect of plants' regeneration niche.

**Epiphytes** are plants that live non-parasitically on tree trunks and branches, and therefore need to find a **suitable host tree** for physical support. Because of their life style, epiphytes face strong ecological gradients at relatively short distances. Probably the most important environmental factor in tropical forest is the **availability of light**, which changes considerably through the canopy of the forest. To cope with these variable light conditions, epiphytes have specific adaptations to regulate their photosynthetic capacity and growth rate (Zotz, 2016). Nevertheless, whether light affects seed germination in epiphytes is an understudied issue, and the scarce information available is far from conclusive (Zotz, 2016). By contrast with epiphytes belonging to other families, the regeneration niche of **epiphytic orchids** depends on two additional factors. First, seeds fully rely on **mycorrhizal fungi for germination**. Since these fungi are not inherited from maternal plants (Benzing, 1990; Rasmussen, 2002), the availability and spatial distribution of suitable mycorrhizal fungi are a key component determining the probability of germination and therefore the distribution of epiphytes in the forest canopy. Second, while the early life stages lack chlorophyll and fully depend on mycorrhiza for nutrition, adults of most species are capable of photosynthesis (**partial mycoheterotrophy**), potentially reducing their dependency on mycorrhiza as compared to seedlings (Leake, 1994). Epiphytic orchids thus need to find suitable host trees and mycorrhizal fungi for germination, but also might experience changes in their environmental and nutritional requirements throughout development, meaning that the **regeneration and adult niches might differ substantially**.

Although specialized partnerships with host trees have been reported, most evidence so far indicates that most epiphytic orchids can grow on a broad range of tree species (Burns & Zotz, 2010; Wagner *et al.*, 2015; Rasmussen & Rasmussen, 2018), suggesting that tree availability may not be a major constraint to the distribution of epiphytic orchids. Less, however, is known about how mycorrhizal fungi determine the distribution of epiphytic orchids. In terrestrial orchids, there is a continuum from specialists(McKendrick *et al.*, 2002; Bidartondo & Read, 2008) to generalists (Jacquemyn *et al.*, 2016; Ogura-Tsujita *et al.*, 2018). For epiphytic orchids, we do not really know whether they are specialized or generalized (Otero *et al.*, 2007). The little evidence available suggests that epiphytic orchids may have a higher diversity of mycorrhizal interactions than terrestrial orchids (Martos *et al.*, 2012) and that they tend to be generalists (Suárez & Kottke, 2016; Herrera *et al.*, 2018), although highly specialized taxa have also been documented (Otero *et al.*, 2007; Riofrío *et al.*, 2013). Studies over small spatial scales have shown that coexisting terrestrial orchids tend to have distinctive mycorrhizal communities and show strong spatial segregation, suggesting that mycorrhizal partners play a role in determining their distribution (Jacquemyn *et al.*, 2014). No such data are available for epiphytic orchids.

Little is also known about **how mycorrhizal communities vary** among host trees and over the host tree surface, and how this affects the distribution of epiphytic orchids. It has been proposed that epiphyte turnover among host trees may be mediated by mycorrhizal fungi (Clements, 1987) and that orchids with many mycorrhizal partners have more host tree species (Gowland *et al.*, 2013). However, exceptions to this pattern have been reported as well(Gowland *et al.*, 2013), suggesting that interactions of epiphytic orchids with their both trees and mycorrhizal fungi can be diverse and complex. Similarly, the vertical turnover in epiphytic orchids is known to relate to changes in bark characteristics as well as the epiphytes' requirements for light (Rasmussen & Rasmussen, 2018). Yet, whether those factors influence the distribution of mycorrhizal partners remains to be assessed (Gowland *et al.*, 2013). In general, solid evidence of how host trees and mycorrhiza affect epiphytic orchid distribution is still lacking, because **studies were conducted on a limited spatial scale and rarely included ecological gradients**.

Because seedlings often experience high mortality in later stages of their development (Harper, 1977; Batty *et al.*, 2001), several factors may affect the distribution of epiphytic orchids. These relate to seedling predation and pronounced differences between the *regeneration* and *adult niches* of the species (Eriksson & Ehrlén, 2009). Plant physiological needs often change over ontogeny, and the successful transition from seedling (protocorm) to adult might depend on acquiring new mycorrhizal partners that help fullfill those new needs (Batstone *et al.*, 2018). Such ontogenetic partner turnover can result from complementarity or sampling effects in time (Batstone *et al.*, 2018) (Fig. 1). **Complementarity** consists of a replacement of partners from the seedling to the adult stage, under the assumption that new partners potentially play complementary roles. On the other hand, **sampling effects** consist of adults retaining a subset of their partners from the seedling stage. Ontogenetic partner turnover through total complementarity is risky because a lack of suitable new partners can compromise survival to adulthood. Evidence from terrestrial orchids suggests that partner gains are common (Rasmussen *et al.*, 2015), while total complementarity (Rasmussen *et al.*, 2015) and partner losses (Bidartondo & Read, 2008) are less well documented. In epiphytic orchids, the role of mycorrhiza turnover on the successful transition to adulthood remains to be assessed (Jersáková & Malinová, 2007; McCormick & Jacquemyn, 2014).



**Fig. 1** Hypothetical scenarios of ontogenetic partner turnover and their putative driving processes. Colours denote different partners. Text above arrows indicates changes in partner number; text below arrows indicates the prevailing component of composition turnover (nestedness or replacement)

# Scientific research objectives

The major aim of this research is to understand **how multiple partners interacting with abiotic conditions influence the distribution of epiphytic orchids in hyper-diverse tropical forests**. Specifically, the project aims at understanding how the availability of suitable mycorrhizal fungi affects germination and recruitment of epiphytic orchids along natural light gradients, and how this is affected by specific characteristics of the host tree. To this end, I will address three key aspects of the interaction: i) the availability of free-living fungi on the substrate as a key component of microsite quality; ii) changes in the interaction over the vertical gradient of light of the forest; and iii) ontogenetic turnover of mycorrhizal partners as a putative barrier to post-germination establishment. In particular, I will test the hypotheses that:

H1: the influence of partner availability on epiphytic orchid distribution depends on partner breadth and abiotic conditions.

H2: the composition and the availability of free-living fungi changes over the vertical gradient of light and over the trunk of the host tree.

H3: seedlings associate with a larger diversity of mycorrhizal fungi than adults allowing them to easily find a suitable partner in the complex canopy of the rainforest.

H4: ontogenetic changes in mycorrhizal partners occur and result from sampling effects rather than total complementarity.

This study will provide **three innovative aspects to the field**. Specifically, it will:

**(1) address an unresolved question in plant ecology**: how more than one partner affects plant species' distribution. This is not trivial because a considerable proportion of tropical plant diversity relies on more than one partner for successful establishment. In epiphytic orchids in particular, most studies have focused on bipartite interactions, *i.e.*, epiphyte-host tree or epiphyte-fungi interactions, while a tripartite network approach (epiphyte-mycorrhiza-host tree) better reflects the actual situation.

**(2)** provide the **first experimental test** of how tripartite interactions affect orchid distribution within a vertical light gradient. Combining **careful field observations and experiments with cutting-edge analyses and molecular techniques**, I will be able to decipher:

* how mycorrhizal fungi are distributed over the host tree trunk.
* how the vertical gradient of light within a host-tree affects epiphytic orchid germination.
* whether orchid mycorrhizal partners are replaced or retained over an individual's lifetime, and the underlying mechanisms.

**(3)** provide the **first comprehensive overview of the patterns and potential drivers of tropical epiphytic orchid distribution**. This research **will push the state-of-the-art forward**, moving from local studies of focal orchid species towards an integrative approach over larger scales, and provide novel insights into:

* how climatic factors influence the interaction network and community structure of epiphytic orchids.
* how orchid partner breadth and partner availability influence the large-scale distribution of epiphytic orchids.

To test the proposed research hypotheses, I will build and analyse tripartite interaction networks to infer changes in the orchid-mycorrhiza-host tree interactions over a geographical gradient of temperature and moisture. I expect to find a continuum of orchid-partner interactions, from strict generalists to strict specialists. Forests with more stressful abiotic conditions (e.g., warm and dry) will harbour orchid species that depend more strongly on their partners, while moist forests will host a wider variety of strategies. Solid evidence of how light gradients affect fungi availability and mycorrhizal symbioses in epiphytic orchids is still lacking. This project will address for the first time the effect of two partners on the distribution of epiphytic orchids in megadiverse communities across geographical and local ecological gradients.

Our knowledge on how seedling and adult niche requirements ultimately affect epiphytic orchid distribution is only fragmentary since the few studies available deal mainly with adult plants, but neglect germination or transitions between ontogenetic stages. Such knowledge, however, is key to design effective, evidence-based orchid conservation actions. In addition, the technology for massive DNA sequencing of microscopic fungi has been developed only in the last 15-10 years, which now allows to quantify the molecular diversity of fungal partners with an unprecedented resolution. A major current need in the field is an integrated approach that addresses (i) how host trees and mycorrhizal fungi limit epiphytic orchid distribution across geographical ecological gradients, and (ii) how the regeneration niche as defined by mycorrhizal fungi and the abiotic environment, and the transition to the adult stage determine individual establishment in epiphytic orchid populations.

# Research methodology and work plan

The methodology combines extensive field sampling and *in-situ* experiments with cutting-edge molecular and statistical analyses. I will conduct fieldwork in Colombia, where I will collaborate with Dr. Nicola Flanagan, professor at Pontificia Universidad Javeriana, Cali. I will undertake laboratory and computing tasks in Belgium, in the research group of Dr. Hans Jacquemyn, professor at KU Leuven.

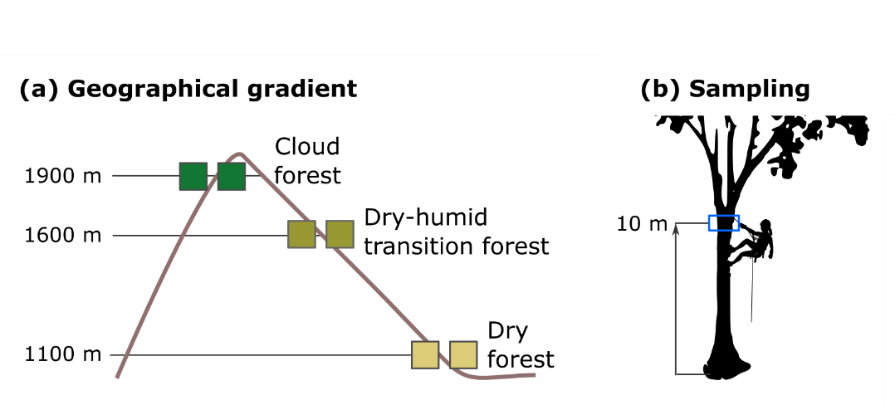
The work will be structured in three work packages (WP). In WP 1, I will study how the tripartite interaction between orchid, host tree, and mycorrhizal fungi varies across a geographical gradient of temperature and humidity (H1). To this end, I will work in different forest types across the Colombian Andes (Fig. 2). In WP 2, I will investigate the distribution of free-living mycorrhizal fungi over a local gradient of light (H2) represented by three trunk heights. Field sampling for this WP will be conducted in dry-humid transition forests. In WP 3, I will assess whether mycorrhizal fungi communities change between the seedling and the adult stages of epiphytic orchids (H3) and infer potential underlying ecological mechanisms (H4). To this end, I will perform seed germination experiments in dry-humid transition forest plots. Each WP will produce a research paper (*Deliverables 1, 2, and 3, respectively*) that I will submit to high-impact scientific journals.

**WP1 Assessing the influence of partner availability on epiphytic orchid distribution.**

It is reasonable to assume that the frequency of seed germination will be higher in orchid species that associate selectively with common instead of rare fungal partners, implying that **partner availability in the substrate** is probably an important determinant of orchid occurrence. Yet, orchid species differ in their degree of specialization in the interaction (**partner breadth**), meaning that they can associate selectively with a few fungal partners (*specialist species*), or indistinctively with many of them (*generalist species*). Theory predicts that natural selection favors highly specialized interactions as a way to avoid cheaters. This view has been recently challenged on the basis that generalist interactions can be advantageous under environmental heterogeneity, where species should not be very choosy in order to associate with the partners at hand(Frederickson, 2013; Batstone *et al.*, 2018). In this way, generalist species can broaden their distribution by shifting partners (**partner turnover**) when the preferred partner is not locally available. Species exhibiting such ability have more chances of germinating in a variety of microsites than highly-specialized species.

*Task 1.1 Sampling natural communities of epiphytic orchids*

The study will be conducted in natural, mostly undisturbed tropical forest ecosystems. I will sample communities of epiphytic orchids in three forest types along a humidity-altitude gradient in the western mountain range of the Colombian Andes (Fig. 2a). The forest types encompass dry forests in the Cauca River Valley, dry-humid transition forests (DHTF) in the valley slope, and cloud forests in the Pacific slope. In each forest type I will choose two plots 5 km apart and 10 host trees per plot. I will sample plants growing at a height of 10 m on tree trunks using the single-rope climbing method (Fig. 2b), collecting three 2-cm root fragments in up to five individuals per orchid species of all the species found. Sampling will not destroy the plants.



**Fig.** **2** Sampling design. **(a)** Geographical gradient including three forest types in the Andes Mountain Range, with two plots per altitude. **(b)** Sampling method.

*Identifying species -* Based on diversity inventories in the study locations, I expect to find approximately 120 orchid species in total (García-Revelo, S. unpubl.; Ospina, NH, unpubl.). Some orchid species are expected to be unique to a particular forest type, while others are expected to occur in at least two of them. I will register the **host tree species** where the different epiphytic orchid species occur (host-tree breadth), and the **mycorrhizal partners** in orchid roots (mycorrhizal fungi breadth). I will **identify orchids and host trees** with the help of a local taxonomist (Nhora Ospina, Universidad del Valle, Cali) and confirm ambiguous tree and orchid records using genetic barcoding.

*Characterizing abiotic conditions -* To characterize **regional climate** I will use climatic data from meteorological stations near the plots. To describe the **light environment of host trees** I will measure photosynthetically active radiation (PAR, mol·m-2·day-1) by taking three hemispheric photographs in each tree at 10 m, where orchid roots will be sampled. To improve PAR estimation accuracy, I will measure the diffuse to direct PAR ratio with BF5 diffuse PAR sensors (Delta-T Devices, UK) in three randomly chosen host trees. The completion of this task is a milestone (M1). Potential delays (*e.g.*, due to bad weather) have been considered in the task schedule (Table 1).

*Task 1.2 Obtaining export licenses and sample shipping*

I will perform DNA extraction and sequencing analyses in the laboratory of Dr. Hans Jacquemyn at KU Leuven in Belgium. Therefore, root samples have to be exported. I will apply for export licenses, for which the Colombian government has legal mechanisms in place. Licenses for research purposes are often granted within 1-3 months (N Ospina, Pers. comm.). The completion of this task represents a milestone (M2). Therefore, I have scheduled a four-month span between license application and sample shipping, to prevent unplanned delays in the schedule. In the low-likelihood scenario that this process delays further, I have two alternative contingency plans: (1) Use Dr. Flanagan's licenses backed by her institution (PUJ); (2) Extract mycorrhizal DNA in her molecular biology lab and export PCR products.

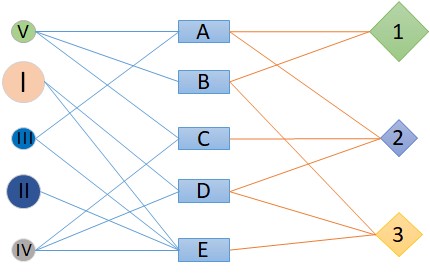
*Task 1.3 Identifying mycorrhiza with molecular analyses*

Since mycorrhizal fungi are microscopic and may form complex interactions with both host trees and orchids, I will combine **meta-barcoding of mycorrhizal DNA** with **network analysis tools** to assess mycorrhizal partner breadth (Jacquemyn *et al.*, 2011, 2015). An individual orchid can host 1-30 different fungal partners (Herrera, 2018). To accurately describe **mycorrhizal diversity** associated with each individual plant I will extract DNA from 0.5 g mycorrhizal root fragments using UltraClean Plant DNA Isolation Kit (Mo Bio Laboratories Inc., CA, USA). I will use two complementary primer pairs (ITS3/ITS4OF and ITS86F/ITS4) for detailed characterization of diverse orchid mycorrhizal communities (Waud *et al.*, 2014). I will use Illumina sequencing to obtain mycorrhizal operational taxonomic units (OTUs), which are the commonly used units of microbial diversity. This task is a milestone (M4), but I do not foresee major risks that endanger its completion.

*Task 1.4 Assessing inter-specific interactions and network analyses*

Network analysis captures patterns of interactions between species (Newman, 2018). I will build **tripartite networks** comprised of three types of nodes: i) epiphytic orchids, ii) mycorrhizal fungi, and iii) host trees (Fig. 3). Tripartite network analysis provides information on the *number of links per species* (which species are more connected) and *layer interdependence* (a measure of how much information about one layer predicts information in another layer) (De Bacco *et al.*, 2017), and allows to *detect communities within the network* ('blocks', a metric similar to modularity of bipartite networks) (Kivelä *et al.*, 2014; Kéfi *et al.*, 2016; De Bacco *et al.*, 2017). Tripartite network analysis is only recently being used to address multiple interactions and, despite the field evolves rapidly (Kivelä *et al.*, 2014), networks involving more than two partners have to be decomposed into bipartite networks to obtain other relevant metrics (Olesen *et al.*, 2007), such as *modularity* (Olesen *et al.*, 2007), *nestedness*  (Bascompte *et al.*, 2003), and *specialization* (Blüthgen *et al.*, 2006). In the context of network analysis, a high modularity indicates there are subsets of strongly connected species interlinked through a few interactions (Olesen *et al.*, 2007). A high nestedness indicates that there is a core of the most generalist species interacting among them (Bascompte *et al.*, 2003). Partner breadth of a species can be quantified with the degree of interaction specialization at the species (d') and network (H') level (Blüthgen *et al.*, 2006).

With this information, I will be able to test H1, which states that the influence of partner availability on epiphytic orchid distribution depends on partner breadth and abiotic conditions. In particular, I expect network nestedness and specialization to vary between forests. In cloud forests, where abiotic stress is lowest, epiphyte abundance is high and competition for space is strong (Benzing, 1990), there will be a wide array of orchid-mycorrhiza-host tree interactions, from generalists to specialists. This is because generalization often allows plants to use a wider variety of microsites (Batstone *et al.*, 2018) and specialization potentially reduces competition among coexisting orchids (Jacquemyn *et al.*, 2014). In dry forests, where abiotic conditions are harsher, physiological adaptation will be a strong determinant of orchid distribution, and interaction networks will probably be more generalized to increase the probability of successful establishment.



**Fig. 3** Schematic representation of a tripartite interaction network between orchids (circles), mycorrhiza (squares) and host trees (rhombi). Characters depict different species; lines depict inter-specific interactions.

**WP2 Assessing how mycorrhizal fungi are distributed over the host tree trunk.**

The interaction between orchids and mycorrhizal fungi is asymmetric. Orchid seeds have to find suitable mycorrhizal fungi for germination, while fungi do not necessarily depend on the orchid for their survival. However, it is likely to assume that the spatial distribution of fungi themselves depends on characteristics of the environment. Spatial variation in the composition of mycorrhizal fungi communities is likely to affect seed germination and consequently the distribution of epiphytic orchids. If the composition of fungal communities is heterogeneous over the substrate, orchid species that specialize on a limited number of fungi, may suffer from establishment limitation. In contrast, generalist orchid species that associate with a variety of mycorrhizal fungi are expected to be less limited by the presence of mycorrhizal and therefore will occupy a broader range of microhabitats.

*Task 2.1 Assessing variation in mycorrhizal communities*

To investigate variation in mycorrhizal communities, I will sample 20 trees from the dry-humid transition forest plots from WP 1. I will establish sampling points at heights of 10, 6 and 2 m in the trunk, representing a vertical light gradient (Fig. 4). At each height, I will measure light using hemispheric photography as described in WP 1. I will collect three bark samples per height to obtain bark fungi and perform fungal DNA extraction and sequencing analyses as described in WP 1. This will provide a picture of the pool of mycorrhizal fungi locally available in different regions of the trunk. The proportion of mycorrhizal fungi actually interacting with orchids will be identified by combining three pieces of information: 1) OTUs lists derived from the regional scanning in WP 1, 2) the mycorrhizal fungi associated to the adult orchids present nearby, and 3) the mycorrhizal fungi identified from seedlings derived from *in-situ* germination assays.

*Task 2.2 Collecting root fragments from adult orchids*

To identify mycorrhizal fungi in the adult stage, I will sample the 20 trees at the three heights for the 4 species selected, and collect 3 root fragments in 3-5 adult individuals (Fig 4a). These samples will be shipped to Belgium along with those of WP1. I will extract fungal DNA and analyze the sequences as described in WP 1. During this sampling, I will also collect ripe capsules of these species in order to obtain the seeds for setting the germination assays (Task 2.3).

*Task 2.3 Setting* in-situ *germination assays*

I will extract the seeds from the capsules to prepare seed packets (Shao *et al.*, 2017). I will use plastic wraps (Shao *et al.*, 2017) to attach three seed packets per orchid species at the three heights in 10 trees per plot (720 seed packets in total) (Fig. 4b). The completion of this task is a milestone (M3).

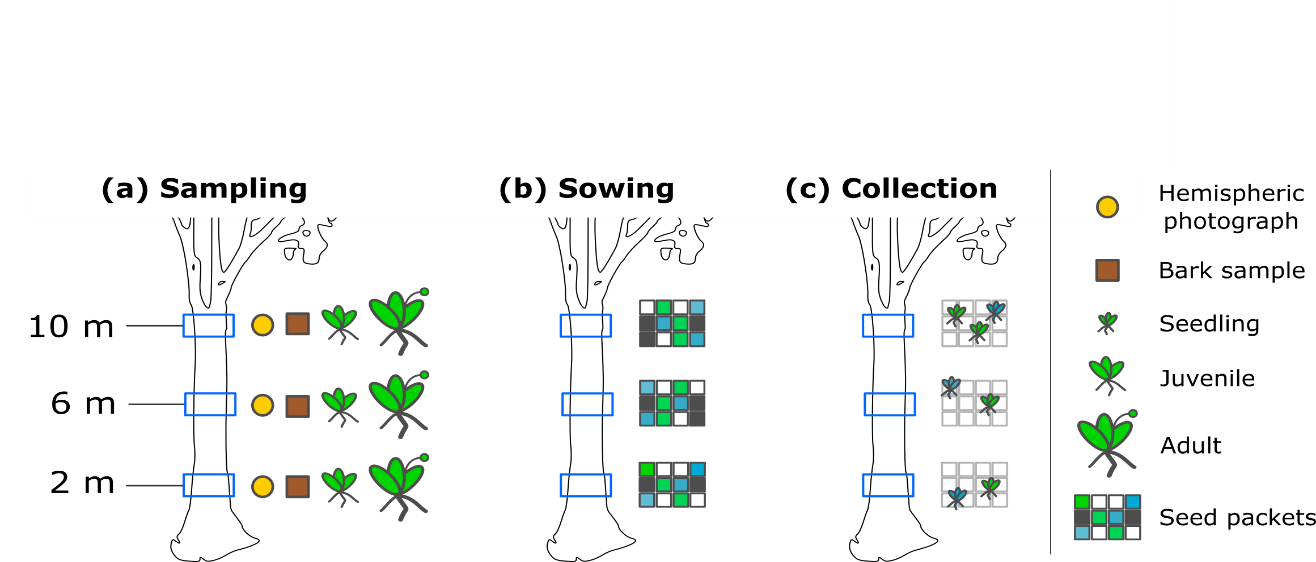
*Task 2.4.* *Identifying mycorrhiza with molecular analyses*

I will extract fungal DNA from bark and mycorrhiza of adult orchids as described in *Task 1.4* (WP1).

This task is a milestone (M5), but I do not foresee major risks that endanger its completion.

*Task 2.5* *Collecting protocorm fragments*

Eight to ten months after sowing I will harvest the seedlings (Fig. 4c) to collect protocorm fragments. Mycorrhizal DNA extraction and sequencing will be performed as described in Task 1.3. This task is a milestone (M6). A potential risk is obtaining a low germination success (medium likelihood). I will implement two preventive measures to increase the probability of germination: (i) the use of an effective sowing method (Shao *et al.*, 2017), and (ii) a high number of replicates. In addition, this experiment will be monitored every two months by a member of Dr. Flanagan's lab. This will ensure that I do not miss the germination period for not being physically there.



**Fig.** **4** Study design. **(a)** Bark and adult sampling at 3 trunk heights, **(b)** *in-situ* germination assays (each colour denotes a different species), **(c)** collection of seedlings.

*Task 2.6.* *Identifying mycorrhiza with molecular analyses*

I will extract fungal DNA from protocorms as described in *Task 1.4* (WP1). This task is a milestone (M7), but I do not foresee major risks that endanger its completion.

*Task 2.7 Data analysis*

I will determine which bark fungi correspond to mycorrhizal taxa. With this information, I will be able to test H2, which states that the availability of free-living fungi changes over the vertical gradient of light. To this end, I will first describe mycorrhiza community composition, richness and turnover across tree heights. I will perform non-metric multidimensional scaling (NMDS) analysis over a mycorrhizal OTU presence-absence matrix and assess differences in species composition with PERMANOVA, estimate richness with rarefaction curves (Magurran, 2004), and turnover using Jaccard index (Cardoso *et al.*, 2015). Second, I will test whether mycorrhizal fungi communities vary over the tree trunk by building linear models controlled by the host tree species.

**WP3 Assessing turnover in mycorrhizal fungi over epiphytic orchid ontogeny.**

To assess turnover in mycorrhizal communities across different life stages, I will use the same study species and experimental design as in WP2. I will compare the **richness** and **composition** of mycorrhizal fungi communities extracted from **seedling** and **adult** roots. To test the hypothesis that seedlings will have a greater richness of mycorrhizal fungi than adults (**H3**), I will calculate **mycorrhizal OTU richness in seedlings and adults** for each species.

*Sampling -* I will use the data of the four species collected in WP2: OTUs obtained from adult orchid roots in *Task 2.2*, and OTUs obtained from seedling roots from germination assays in *Task 2.3*.

*Task 3.1 Data analysis*

I will estimate **mycorrhizal OTU richness in seedlings and adults** for each species, and I will test whether seedlings have a greater richness of mycorrhizal fungi than adults by building linear models. I expect seedlings to associate with a greater number of fungi OTUs. A confirmation of this hypothesis would indicate that seedlings are more generalist than adults, suggesting that the establishment of mycorrhizal associations is opportunistic during early life stages.

To test the hypothesis that ontogenetic partner turnover occurs through sampling effects rather than complementarity (**H4**), I will calculate OTU composition turnover from seedlings to adults.

*Sampling -* I will use the data of the four species collected in WP2: OTUs obtained from adult orchid roots in Task 2.2, and OTUs obtained from seedling roots from germination assays in Task 2.3.

*Task 3.2 Data analysis*

I will obtain **mycorrhizal OTU composition in seedlings and adults** for each species. I will assess OTU turnover between both developmental stages, and partition total turnover into its nestedness and replacement components (Cardoso *et al.*, 2015). High nestedness values indicate that adults associate with a subset of seedlings' OTUs, whereas high replacement values indicate that adults associate with a set of OTUs that are absent in seedlings. I expect nestedness to represent a greater proportion of OTU turnover than replacement (Fig. 1c). This result would indicate that mycorrhyzal turnover over the ontogeny is due to sampling effects instead of total complementarity, thereby supporting the general hypothesis that switching partners over ontogeny is a risky strategy.

**WORK PLAN**

The proposal consists of three work packages (**WP**), seven milestones (**M**) and three deliverables (**D**) (*Table 1*). Ideal start date is October 1st, 2019.

**Table 1** Gantt chart showing work packages (WP), milestones (M) and deliverables (D) of the project. Blue: research tasks; green: administrative process.



# References

**Afkhami M, McIntyre P, Strauss S**. **2014**. Mutualist-mediated effects on species ’ range limits across large geographic scales. *Ecology Letters*: 1265–1273.

**De Bacco C, Power EA, Larremore DB, Moore C**. **2017**. Community detection , link prediction , and layer interdependence in multilayer networks. *Physical Review E* **95**: 042317.

**Bascompte J, Jordano P, Melia CJ**. **2003**. The nested assembly of plant – animal mutualistic networks. **100**: 9383–9387.

**Batstone RT, Carscadden KA, Afkhami ME, Frederickson ME**. **2018**. Using niche breadth theory to explain generalization in mutualisms. *Ecology* **99**: 1039–1050.

**Batty AL, Dixon KW, Brundrett M, Sivasithamparam K**. **2001**. Constraints to symbiotic germination of terrestrial orchid seed in a mediterranean bushland. : 511–520.

**Benzing DH**. **1990**. *Vascular epiphytes: General Biology and Related Biota*. Cambridge: Cambridge University Press.

**Bidartondo M, Read D**. **2008**. Fungal specificity bottlenecks during orchid germination and development. *Molecular Ecology* **17**: 3707–3716.

**Blüthgen N, Menzel F, Blüthgen N**. **2006**. Measuring specialization in species interaction networks. *BMC Ecology* **6**.

**Burns KC, Zotz G**. **2010**. A hierarchical framework for investigating epiphyte assemblages: networks, meta-communities, and scale. *Ecology* **91**: 377–385.

**Cardoso P, Rigal F, Carvalho JC**. **2015**. BAT - Biodiversity Assessment Tools, an R package for the measurement and estimation of alpha and beta taxon, phylogenetic and functional diversity. *Methods in Ecology and Evolution* **6**: 232–236.

**Clements M**. **1987**. Orchid-fungus-host associations of epiphytic orchids. In: Saito K, Tanaka R, eds. Proceedings of the 12th World Orchid Conference. Tokyo.

**Denslow JS**. **1980**. Gap partitioning among tropical rainforest trees. *Biotropica* **12**: 47–55.

**Dunn RR, Harris NC, Colwell RK, Koh LP, Sodhi NS**. **2009**. The sixth mass coextinction : are most endangered species parasites and mutualists ? : 3037–3045.

**Eriksson O, Ehrlén J**. **2009**. Seedling recruitment and population ecology. In: Leck M, Parker V, Simpson R, eds. Seedling Ecology and Evolution. Cambridge University Press.

**Frederickson ME**. **2013**. Rethinking Mutualism Stability: Cheaters and the Evolution of Sanctions. *The Quarterly Review of Biology* **88**: 269–295.

**Gowland K, van der Merwe M, Linde C, Clements M, Nicotra A**. **2013**. The host bias of three epiphytic Aeridinae orchid species is reflected, but not explained, by mycorrhizal fungal associations. *American Journal of Botany* **100**: 764–777.

**Grubb PJ**. **2018**. The maintenance of species richness in plant communities: the importance of the regeneration niche. *Biological Reviews* **52**: 107–145.

**Harper JL**. **1977**. *Population Biology of Plants*. New York: Academic Press.

**Herrera P**. **2018**. Interacción entre orquídeas epífitas y hongos micorrízicos en bosques tropicales del Sur de Ecuador.

**Herrera P, Kottke I, Molina MC, Méndez M, Suárez JP**. **2018**. Generalism in the interaction of Tulasnellaceae mycobionts with orchids characterizes a biodiversity hotspot in the tropical Andes of Southern Ecuador. *Mycoscience* **59**: 38–48.

**Jacquemyn H, Brys R, Merckx VSFT, Waud M, Lievens B, Wiegand T**. **2014**. Coexisting orchid species have distinct mycorrhizal communities and display strong spatial segregation. *New Phytologist* **202**: 616–627.

**Jacquemyn H, Brys R, Waud M, Busschaert P, Lievens B**. **2015**. Mycorrhizal networks and coexistence in species-rich orchid communities. *New Phytologist* **206**: 1127–1134.

**Jacquemyn H, Merckx V, Brys R, Tyteca D, Cammue BPA, Honnay O, Lievens B**. **2011**. Analysis of network architecture reveals phylogenetic constraints on mycorrhizal specificity in the genus Orchis (Orchidaceae). *New Phytologist* **192**: 518–528.

**Jacquemyn H, Waud M, Merckx VSFT, Brys R, Tyteca D, Hedrén M, Lievens B**. **2016**. Habitat-driven variation in mycorrhizal communities in the terrestrial orchid genus Dactylorhiza. *Nature Publishing Group*: 1–9.

**Jersáková J, Malinová T**. **2007**. Spatial aspects of seed dispersal and seedling recruitment in orchids. *New Phytologist* **176**: 237–241.

**Kéfi S, Miele V, Wieters EA, Navarrete SA, Berlow EL**. **2016**. How Structured Is the Entangled Bank? The Surprisingly Simple Organization of Multiplex Ecological Networks Leads to Increased Persistence and Resilience. *PLoS Biology* **14**: e1002527.

**Kivelä M, Arenas A, Barthelemy M, Gleeson JP, Moreno Y, Porter MA**. **2014**. Multilayer networks. *Journal of Complex Networks* **2**: 203–271.

**Krebs C**. **1972**. *Ecology: the experimental analysis of distribution and abundance.* New York: Harper & Row New York.

**Leake JR**. **1994**. The biology of myco-heterotrophic ('saprophytic’) plants. *New Phytologist* **69**: 171–216.

**Lusk CH**. **1995**. Seed Size, Establishment Sites and Species Coexistence in a Chilean Rain Forest. *Journal of Vegetation Science* **6**: 249–256.

**Magurran A**. **2004**. *Measuring biological diversity*. Oxford: Blackwell.

**Martos F, Munoz F, Pailler T, Kottke I, Gonneau C, Selosse MA**. **2012**. The role of epiphytism in architecture and evolutionary constraint within mycorrhizal networks of tropical orchids. *Molecular Ecology* **21**: 5098–5109.

**McCormick MK, Jacquemyn H**. **2014**. What constrains the distribution of orchid populations? *New Phytologist* **202**: 392–400.

**McKendrick SL, Leake JR, Lee Taylor D, Read DJ**. **2002**. Symbiotic germination and development of the myco-heterotrophic orchid Neottia nidus-avis in nature and its requirement for locally distributed Sebacina spp. *New Phytologist*: 233–247.

**Nathan R, Muller-landau HC**. **2000**. Spatial patterns of seed dispersal , their determinants and consequences for recruitment. **15**: 278–285.

**Newman M**. **2018**. *Networks*. Oxford: Oxford University Press.

**Ogura-Tsujita Y, Gebauer G, Xu H, Fukasawa Y, Umata H, Tetsuka K, Kubota M, Schweiger JMI, Yamashita S, Maekawa N, *et al.*** **2018**. The giant mycoheterotrophic orchid Erythrorchis altissima is associated mainly with a divergent set of wood-decaying fungi. *Molecular Ecology* **27**: 1324–1337.

**Olesen JM, Bascompte J, Dupont YL, Jordano P**. **2007**. The modularity of pollination networks.

**Otero JT, Aragón S, Ackerman JD**. **2007**. Site variation in spatial aggregation and phorophyte preference in Psychilis monensis (Orchidaceae). *Biotropica* **39**: 227–231.

**Rasmussen HN**. **2002**. Recent developments in the study of orchid mycorrhiza. *Plant and Soil* **244**: 149–163.

**Rasmussen HN, Dixon KW, Jersáková J, Tesitelová T**. **2015**. Germination and seedling establishment in orchids : a complex of requirements. *Annals of Botany* **116**: 391–402.

**Rasmussen HN, Rasmussen FN**. **2018**. The epiphytic habitat on a living host : reflections on the orchid – tree relationship. : 456–472.

**Riofrío ML, Cruz D, Torres E, de La Cruz M, Iriondo JM, Suárez JP**. **2013**. Mycorrhizal preferences and fine spatial structure of the epiphytic orchid Epidendrum rhopalostele. *American Journal of Botany* **100**: 2339–2348.

**Shao S-C, Burgess KS, Cruse-Sanders JM, Liu Q, Fan X-L, Huang H, Gao J-Y**. **2017**. Using In Situ Symbiotic Seed Germination to Restore Over-collected Medicinal Orchids in Southwest China. *Frontiers in Plant Science* **8**: 1–10.

**Slatyer RA, Hirst M, Sexton JP**. **2013**. Niche breadth predicts geographical range size: a general ecological pattern. *Ecology Letters* **16**: 1104–1114.

**Suárez JP, Kottke I**. **2016**. Main fungal partners and different levels of specificity of orchid mycorrhizae in the tropical mountain forests of Ecuador. *Lankesteriana* **16**: 299–305.

**Wagner K, Mendieta-Leiva G, Zotz G**. **2015**. Host specificity in vascular epiphytes: a review of methodology, empirical evidence and potential mechanisms. *AoB PLANTS* **7**: plu092-plu092.

**Waud M, Busschaert P, Ruyters S, Jacquemyn H, Lievens B**. **2014**. Impact of primer choice on characterization of orchid mycorrhizal communities using 454 pyrosequencing. *Molecular Ecology Resources* **14**: 679–699.

**Zotz G**. **2016**. *Plants on Plants – The Biology of Vascular Epiphytes*. Springer International Publishing.