

# Calibration with opticut

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# Chapter 1

## Calibration with single species

This is not necessarily great.

```
library(opticut)
```

```
## Loading required package: pbapply
```

```
## opticut 0.0-9      2016-10-30
```

```
library(dclone)
```

```
## Warning: package 'dclone' was built under R version 3.2.5
```

```
## Loading required package: coda
```

```
## Loading required package: parallel
```

```
## Loading required package: Matrix
```

```
## Warning: package 'Matrix' was built under R version 3.2.5
```

```
## dclone 2.1-2      2016-03-11
```

```
library(rjags)
```

```
## Linked to JAGS 4.2.0
```

```
## Loaded modules: basemod,bugs
```

```
source("~/repos/opticut/extras/calibrate.R")
```

```
set.seed(234)
```

```
n <- 50*4
```

```
K <- 4
```

```
g <- sample(1:K, n, replace=TRUE)
```

```
x <- rnorm(n, 0, 1)
```

```
z <- ifelse(g == 1, 1, 0)
```

```

b0 <- 1
b1 <- 1.5
a <- 0.2
mu <- b0 + z*b1 + x*a
#p <- plogis(mu)
#y <- rbinom(n, 1, p)
lam <- exp(mu)
y <- rpois(n, lam)
table(y, g)

```

```

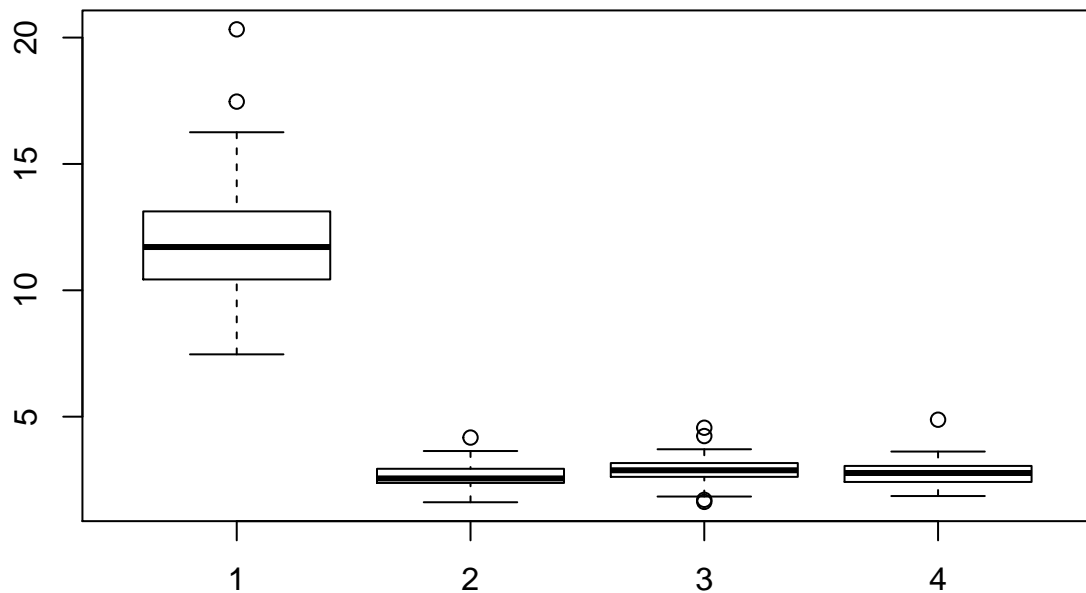
##      g
## y      1  2  3  4
## 0      0  4  5  2
## 1      0 10 12 12
## 2      0  9  9 10
## 3      1  8 13  7
## 4      1  6 11  6
## 5      0  7  8  5
## 6      2  0  3  4
## 7      0  1  0  1
## 8      4  0  2  0
## 9      7  0  0  0
## 10     4  0  0  0
## 11     4  0  0  0
## 12     3  0  0  0
## 13     4  0  0  0
## 14     2  0  0  0
## 15     1  0  0  0
## 16     4  0  0  0
## 17     4  0  0  0
## 18     2  0  0  0
## 20     1  0  0  0
## 21     1  0  0  0

```

```

boxplot(lam ~ g)

```



```
df <- data.frame(y=y, x=x, g=g)
df <- df[-(1:5),]
#o <- opticut(y ~ x, data=df, strata=g, dist="binomial")
o <- opticut(y ~ x, data=df, strata=g, dist="poisson")
summary(o)

## Multivariate opticut results, comb = rank, dist = poisson
##
## Call:
## opticut.formula(formula = y ~ x, data = df, strata = g, dist = "poisson")
##
## Best supported model with logLR >= 2:
##      split assoc      I    mu0    mu1 logLR w
## Sp 1      1    +++ 0.6293 2.744 12.06 245.8 1
## 3 binary splits

co <- calibrate(o, cbind("Sp 1"=(y[1:5])), model.matrix(~x, data.frame(x=x[1:5])))

## Compiling model graph
##   Resolving undeclared variables
##   Allocating nodes
## Graph information:
```

```
## Observed stochastic nodes: 5
## Unobserved stochastic nodes: 6
## Total graph size: 70
##
## Initializing model
```

```
y[1:5]
```

```
## [1] 4 2 10 4 10
```

```
g[1:5]
```

```
## [1] 3 4 1 4 1
```

```
co$gnew
```

```
## [1] "4" "3" "1" "2" "1"
```

```
round(co$pi, 2)
```

```
##      1      2      3      4
## [1,] 0.01 0.33 0.33 0.33
## [2,] 0.00 0.33 0.34 0.34
## [3,] 1.00 0.00 0.00 0.00
## [4,] 0.00 0.34 0.33 0.32
## [5,] 1.00 0.00 0.00 0.00
```

We get  $g=1$  cases correctly classified (this is where  $Sp1$  is to be found), But the rest is just all equally probable. All what we see is that it is not  $g=1$ . All makes sense, but not very useful. Luckily for us, we usually have  $>1$  species.

# Chapter 2

## Calibration with multiple species

This should work much better in principle.

```
set.seed(234)
n <- 50*4
K <- 4
g <- sample(1:K, n, replace=TRUE)
g[1:K] <- 1:K
x <- rnorm(n, 0, 1)
z <- ifelse(g %in% 1:2, 1, 0)
b0 <- 1
b1 <- 1.5
a <- 0.2
mu <- b0 + z*b1 + x*a
lam <- exp(mu)
y1 <- rpois(n, lam)

z <- ifelse(g %in% 2:3, 1, 0)
b0 <- 1
b1 <- 1.5
a <- 0.2
mu <- b0 + z*b1 + x*a
lam <- exp(mu)
y2 <- rpois(n, lam)
y <- cbind(y1=y1, y2=y2)

df <- data.frame(x=x, g=g)
df <- df[-(1:5),]
yy <- y[-(1:5),]
o <- opticut(yy ~ x, data=df, strata=g, dist="poisson")
summary(o)
```

```

## Multivariate opticut results, comb = rank, dist = poisson
##
## Call:
## opticut.formula(formula = yy ~ x, data = df, strata = g, dist = "poisson")
##
## Best supported models with logLR >= 2:
##   split assoc      I   mu0   mu1 logLR w
## y1   1+2   +++ 0.6344 2.823 12.62 331.6 1
## y2   2+3   +++ 0.6502 2.499 11.79 314.5 1
## 3 binary splits
co <- calibrate(o, y[1:5,], df[1:5,])

## Compiling model graph
##   Resolving undeclared variables
##   Allocating nodes
## Graph information:
##   Observed stochastic nodes: 10
##   Unobserved stochastic nodes: 7
##   Total graph size: 122
##
## Initializing model
y[1:5,]

##      y1 y2
## [1,] 14  3
## [2,] 14 17
## [3,]  3  6
## [4,]  4  2
## [5,]  5  1
g[1:5]

## [1] 1 2 3 4 1
co$gnew

## [1] "1" "2" "3" "4" "4"
round(co$pi, 2)

##      1      2      3      4
## [1,] 0.98 0.02 0.00 0.00
## [2,] 0.00 1.00 0.00 0.00
## [3,] 0.00 0.05 0.88 0.07
## [4,] 0.08 0.00 0.01 0.91
## [5,] 0.07 0.00 0.00 0.93

```



# Chapter 3

## Dolina and calibration

```
data(dolina)
dolina$samp$stratum <- as.integer(dolina$samp$stratum)
Y <- dolina$xtab#[dolina$samp$method=="Q",]
X <- dolina$samp#[dolina$samp$method=="Q",]
inew <- 1:nrow(Y) %in% sample(nrow(Y), 10)
Y <- Y[,colSums(Y[!inew,] > 0) >= 20]
#Y <- ifelse(Y > 0, 1, 0)
x <- X[!inew,]
xnew <- dolina$samp[inew,]
y <- Y[!inew,]
ynew <- Y[inew,]
o <- opticut(y ~ stratum + lmoist + method, data=x, strata=mhab, dist="poisson")
#o <- opticut(y ~ 1, data=x, strata=mhab, dist="poisson")

## binary calibration
cal <- calibrate(o, ynew, xnew)

## Compiling model graph
##   Resolving undeclared variables
##   Allocating nodes
## Graph information:
##   Observed stochastic nodes: 190
##   Unobserved stochastic nodes: 29
##   Total graph size: 1901
##
## Initializing model

#cal <- calibrate(o, ynew)
round(cal$pi,2)

##           LI    DW    TL    RO
```

```
## 11R3T 0.05 0.83 0.04 0.08
## 12A4Q 0.00 0.12 0.87 0.00
## 13A2T 0.46 0.05 0.47 0.03
## 1A1T 0.44 0.07 0.45 0.04
## 1A6T 0.43 0.05 0.44 0.08
## 3H2T 0.00 0.09 0.00 0.90
## 4A7T 0.38 0.07 0.38 0.18
## 4R3Q 0.00 0.00 0.00 1.00
## 7R1Q 0.00 0.00 0.00 1.00
## 9L3T 0.81 0.00 0.18 0.00
```

```
round(cor(cal$pi),2)
```

```
##      LI    DW    TL    RO
## LI  1.00 -0.29  0.23 -0.63
## DW -0.29  1.00 -0.18 -0.25
## TL  0.23 -0.18  1.00 -0.70
## RO -0.63 -0.25 -0.70  1.00
```

```
data.frame(predicted=cal$gnew,
            truth=dolina$samp[inew,"mhab"],
            OK=cal$gnew == dolina$samp[inew,"mhab"])
```

```
##      predicted truth    OK
## 1           DW     RO FALSE
## 2           TL     LI FALSE
## 3           TL     LI FALSE
## 4           TL     LI FALSE
## 5           TL     LI FALSE
## 6           RO     DW FALSE
## 7           LI     LI  TRUE
## 8           RO     RO  TRUE
## 9           RO     RO  TRUE
## 10          LI     TL FALSE
```

```
cm <- table(pred=cal$gnew, true=X[inew,"mhab"])
cm <- cm[rownames(cm),rownames(cm)]
cm
```

```
##      true
## pred DW LI RO TL
## DW  0  0  1  0
## LI  0  1  0  1
## RO  1  0  2  0
## TL  0  4  0  0
```

```
sum(diag(cm)) / sum(cm)
```

```
## [1] 0.3
```

```
## multinomial calibration using calibrate.default
```

```
oo <- lapply(1:ncol(y), function(i) {  
  glm(y[,i] ~ mhab + stratum + lmoist + method, x, family=poisson)  
})
```

```
xnew <- model.matrix(~ stratum + lmoist + method, xnew)
```

```
cal <- calibrate(oo, ynew, xnew, K=4)
```

```
## Compiling model graph
```

```
##   Resolving undeclared variables
```

```
##   Allocating nodes
```

```
## Graph information:
```

```
##   Observed stochastic nodes: 190
```

```
##   Unobserved stochastic nodes: 29
```

```
##   Total graph size: 2187
```

```
##
```

```
## Initializing model
```

```
round(cal$pi,2)
```

```
##      [,1] [,2] [,3] [,4]
```

```
## 11R3T 0.07 0.66 0.11 0.16
```

```
## 12A4Q 0.00 0.02 0.98 0.00
```

```
## 13A2T 0.60 0.04 0.35 0.01
```

```
## 1A1T  0.64 0.03 0.31 0.02
```

```
## 1A6T  0.44 0.04 0.48 0.04
```

```
## 3H2T  0.00 0.60 0.00 0.40
```

```
## 4A7T  0.49 0.04 0.39 0.09
```

```
## 4R3Q  0.00 0.00 0.00 1.00
```

```
## 7R1Q  0.00 0.00 0.00 1.00
```

```
## 9L3T  0.82 0.01 0.18 0.00
```

```
round(cor(cal$pi),2)
```

```
##      [,1] [,2] [,3] [,4]
```

```
## [1,]  1.00 -0.41  0.12 -0.63
```

```
## [2,] -0.41  1.00 -0.36 -0.04
```

```
## [3,]  0.12 -0.36  1.00 -0.62
```

```
## [4,] -0.63 -0.04 -0.62  1.00
```

```
data.frame(predicted=levels(x$mhab)[cal$gnew],
```

```
  truth=dolina$samp[inew,"mhab"],
```

```
  OK=levels(x$mhab)[cal$gnew] == dolina$samp[inew,"mhab"])
```

```
##      predicted truth    OK
## 1      DW      RO FALSE
## 2      TL      LI FALSE
## 3      LI      LI  TRUE
## 4      LI      LI  TRUE
## 5      TL      LI FALSE
## 6      DW      DW  TRUE
## 7      LI      LI  TRUE
## 8      RO      RO  TRUE
## 9      RO      RO  TRUE
## 10     LI      TL FALSE
```

```
cm <- table(pred=levels(x$mhab)[cal$gnew], true=X[inew,"mhab"])
cm <- cm[rownames(cm),rownames(cm)]
cm
```

```
##      true
## pred DW LI RO TL
## DW  1  0  1  0
## LI  0  3  0  1
## RO  0  0  2  0
## TL  0  2  0  0
```

```
sum(diag(cm)) / sum(cm)
```

```
## [1] 0.6
```

```
library(optpart)
```

```
## Warning: package 'optpart' was built under R version 3.2.5
```

```
## Loading required package: cluster
```

```
## Warning: package 'cluster' was built under R version 3.2.5
```

```
## Loading required package: labdsv
```

```
## Loading required package: mgcv
```

```
## Warning: package 'mgcv' was built under R version 3.2.5
```

```
## Loading required package: nlme
```

```
## Warning: package 'nlme' was built under R version 3.2.5
```

```
## This is mgcv 1.8-15. For overview type 'help("mgcv-package")'.
```

```
## Loading required package: MASS
```

```
##
```

```
## Attaching package: 'labdsv'
```

```

## The following object is masked from 'package:stats':
##
##      density
## Loading required package: plotrix
## Warning: package 'plotrix' was built under R version 3.2.5
##
## Attaching package: 'optpart'
## The following object is masked from 'package:labdsv':
##
##      clustify
data(shoshsite)
data(shoshveg)

#elev <- cut(shoshsite$elevation, breaks=c(0, 7200, 8000, 9000, 20000))
#levels(elev) <- c("low", "mid1", "mid2", "high")
elev <- cut(shoshsite$elevation, breaks=c(0, 7800, 8900, 20000))
levels(elev) <- c("low", "mid", "high")

sveg <- as.matrix(shoshveg)
sveg[sveg > 0] <- 1

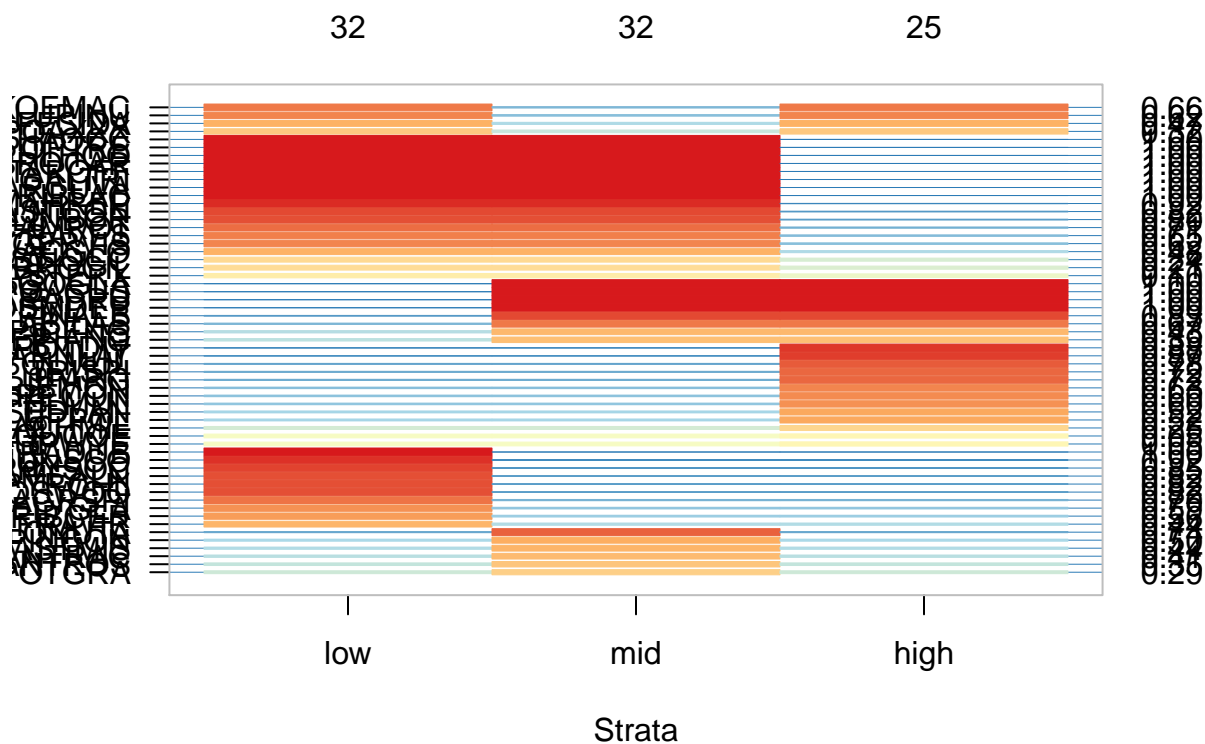
set.seed(1)
ss <- seq_len(nrow(sveg)) %in% sample.int(nrow(sveg), floor(nrow(sveg) * 0.9))
sveg1 <- sveg[ss,]
sveg1 <- sveg1[,colSums(sveg1) > 5]
elev1 <- elev[ss]
dim(sveg)

## [1] 150 368
dim(sveg1)

## [1] 135 85
sveg2 <- sveg[!ss, colnames(sveg1)]
elev2 <- elev[!ss]

o <- opticut(sveg1 ~ 1, strata=elev1, dist="binomial")
plot(o, sort=1)

```



```
co <- calibrate(o, sveg2)
```

```
## Compiling model graph
##   Resolving undeclared variables
##   Allocating nodes
## Graph information:
##   Observed stochastic nodes: 1275
##   Unobserved stochastic nodes: 100
##   Total graph size: 7586
##
## Initializing model
```

```
data.frame(predicted=co$gnew,
            truth=elev2,
            OK=co$gnew == elev2,
            round(co$pi,2))
```

	predicted	truth	OK	low	mid	high
## X1V90L019	high	mid	FALSE	0.00	0.01	0.99
## X1V90P002	low	low	TRUE	0.89	0.11	0.00
## X1V90L031	low	low	TRUE	0.95	0.05	0.00
## X1V891031	low	low	TRUE	0.99	0.01	0.00

```
## X5V96I296      high  high  TRUE 0.00 0.00 1.00
## X5V96M122      high  high  TRUE 0.00 0.00 1.00
## X4V93R081      low   mid  FALSE 1.00 0.00 0.00
## X1V90I051      high  high  TRUE 0.00 0.00 1.00
## X1V89I022      mid   mid  TRUE 0.01 0.99 0.00
## X4V89P048      mid   high FALSE 0.02 0.93 0.04
## X4V95S079      mid   mid  TRUE 0.01 0.99 0.00
## X2V95S211      high  high  TRUE 0.00 0.00 1.00
## X1V90L024      high  high  TRUE 0.00 0.00 1.00
## X5V96K201      high  high  TRUE 0.00 0.00 1.00
## X5V96I014      mid   mid  TRUE 0.04 0.96 0.00
```

```
sum(co$gnew == elev2) / nrow(sveg2)
```

```
## [1] 0.8
```

## 3.1 Conclusions

- More species, merrier results (see simple simulation)
- Binary partitions might not be the best, especially for count models
- Multinomial models (K levels in g) are more efficient (see dolina example)
- Many species in binomial context work quite well, if K is not unreasonably high.

## 3.2 Todo

- OK use notation:  $j=1 \dots p$  &  $r=1 \dots S$
- OK implement test cases for lm/glm
- OK write internal function (.calibrate)
- OK implement user interface: calibrate, calibrate.opticut(object, ynew, xnew, ...)
- add NB, beta, ZI cases (ZI: inits!) [drop all the 1-species models!]
- gaussian needs  $\text{sig}^2$