How do climate change experiments actually change climate?

A.K. Ettinger^{1,2,a}, I. Chuine^{3,b}, B.I. Cook^{4,5,c}, J.S. Dukes^{6,d}, A.M. Ellison^{7,e}, M.R. Johnston^{8,f}, A.M. Panetta^{9,g}, C.R. Rollinson^{10,h}, Y. Vitasse^{11,12,i}, and E.M. Wolkovich^{1,8,j}

¹Arnold Arboretum of Harvard University, Boston, Massachusetts 02131, USA ²Tufts University, Medford, Massachusetts 02155, USA

³CEFE UMR 5175, CNRS, Université de Montpellier, Université Paul-Valéry Montpellier, EPHE, Montpellier, France

 $^4\mathrm{Lamont\text{-}Doherty}$ Earth Observatory, Columbia University, Palisades, New York 10964, USA

⁵NASA Goddard Institute for Space Studies, New York, New York 10025, USA
⁶Department of Forestry and Natural Resources and Department of Biological Sciences,
Purdue University, West Lafayette, Indiana 47907, USA

⁷Harvard Forest, Harvard University, Petersham, Massachusetts 01366, USA
⁸Department of Organismic and Evolutionary Biology, Harvard University, Cambridge,
Massachusetts 02138, USA

 $^9\mathrm{Department}$ of Ecology and Evolutionary Biology, University of Colorado, Boulder, Colorado 80309, USA

¹⁰The Morton Arboretum, Lisle, Illinois 60532, USA
¹¹Institute of Geography, University of Neuchâtel, Neuchâtel, Switzerland
¹²Swiss Federal Institute for Forest, Snow and Landscape Research WSL, Neuchâtel, Switzerland

^aCorresponding author; email: aettinger@fas.harvard.edu; phone: 781-296-4821; mailing address: 2204 Flagler Place NW, Washington, Distric of Columbia, 20001,USA

^bIsabelle.CHUINE@cefe.cnrs.fr

cbc9z@ldeo.columbia.edu

^djsdukes@purdue.edu

^eaellison@fas.harvard.edu

fjohnston.miriam@gmail.com

ganpa7214@colorado.edu

^hcrollinson@mortonarb.org

ⁱyann.vitasse@unine.ch

^jwolkovich@fas.harvard.edu

October 18, 2017

Statement of authorship All authors conceived of this manuscript, which was inspired by our discussions

at a Radcliffe Exploratory Seminar in 2016, and all authors contributed to manuscript revisions. A.E. and

E.W. conceived of the idea for the literature review, database compilation, and related Radcliffe Exploratory

Seminar. A.E. compiled the datasets; A.E. and C.R. analyzed the data and created the figures; A.E. wrote

the manuscript.

Data Accessibility The C3E database will be available at KNB (Ettinger and Wolkovich, 2017), along with

all R code from the analyses included in this paper. (Currently, metadata is published there; the full database

and R code are available to reviewers upon request.)

Running title Experimental climate change

Key words

Type of article Review and Synthesis

Number of words in abstract 182

Number of words in main text

Number of words in Box 1

2

Number of references

Number of figures

Number of tables

Number of text boxes

1 Abstract

To understand and forecast biological effects of climate change, scientists frequently use field experiments that alter temperature and precipitation (e.g., with infrared heaters, rain shields, and supplemental watering). These experimental results may be analyzed and interpreted in misleading ways, however. We reviewed publications on active warming experiments to compile a new database of daily climate data from 12 active warming experiments. We find that the common practice of summarizing and analyzing only the mean changes across treatments hides potentially important variation in treatment effects over space and time. Furthermore, treatments produce unintended secondary effects, such as soil drying in conjunction with warming. The implications of these complexities are rarely explored, but have important biological consequences. We show one example of such consequences with a case study of spring plant phenology, in which such secondary effects lead to inaccurate quantification of species' sensitivities to changes in temperature. Based on our synthesis, we present recommendations for future analyses, as well as experimental design and data sharing, that we believe will improve the ability of climate change experiments to accurately identify and forecast species' responses.

15 Introduction

Climate change is dramatically altering earth's biota, shifting the physiology, distribution, and abundance of organisms, with cascading community, ecosystem, and climate effects (Shukla and Mintz, 1982; Cox et al., 2000; Thomas et al., 2004; Parmesan, 2006; Field et al., 2007; Sheldon et al., 2011; Urban et al., 2012). Much uncertainty exists about how particular individuals, populations, species, communities, and ecosystems will respond as shifts in temperature and precipitation regimes become more extreme. Predicting biological responses to current and future climate change—and how they will feedback to affect earth's climate and ecosystem services—are among the most significant challenges facing scientists today.

Two common approaches for understanding biological effects of climate change are observational studies and process-based modeling; yet these approaches are insufficient for several reasons. Observational studies, which correlate recorded biological patterns with measured trends in climate, cannot disentangle the causal effects of warming from other factors that have also changed over time, such as successional stage or land use. Process-based models can overcome some of these challenges because they rely on explicit empirical

relationships between observed phenomena and climate. They, however, are limited by their underlying assumptions, which may be poorly constrained (e.g., Pearson and Dawson, 2004; Ibanez et al., 2006; Swab et al., 2012; Chuine et al., 2016). Specify what is meant by the previous sentence. In addition, neither approach is well-vetted for predicting future conditions that fall outside the range of historical variability; climate change will yield warmer temperatures than the previous 150 years, and possibly warmer than at any time in the last 2000 years (Ohlemüller et al., 2006; Williams and Jackson, 2007; Williams et al., 2007; Stocker et al., 2013).

Field-based experiments that alter temperature address these shortcomings, and are therefore critical for
determining mechanistic links between climate change and biological responses (e.g., Box et al., 1978; Williams
and Jackson, 2007; Gelman, 2014). Experiments can quantify biological responses to different levels of climate
change, and can create the "no-analog" climate scenarios forecasted for the future, particularly when they
employ active warming methods, such as forced air heaters, soil warming cables, or infrared heaters (Shaver
et al., 2000; Williams et al., 2007; Aronson and McNulty, 2009). In addition, active warming can be combined
with precipitation manipulations (e.g., snow removal, water additions, water reductions), offering the ability
to isolate effects of temperature and precipitation from other environmental changes (e.g., Price and Waser,
1998; Cleland et al., 2006; Sherry et al., 2007; Rollinson and Kaye, 2012). Compared with indoor growthchamber experiments, field-based experiments offer the possibility of preserving important, but unknown or
unquantified feedbacks among biotic and abiotic components of the studied systems.

Climate experiments allow ecologists to draw conclusions about how climate change may affect species' growth, survival, and future distributions (Dukes and Mooney, 1999; Hobbie et al., 1999; Morin et al., 2010; Chuine et al., 2012; Reich et al., 2015; Gruner et al., 2016). But is it reasonable to extrapolate findings from these experiments to the real world? Do they actually alter climate in the ways that we think they do? Recent research suggests that climate manipulations do not alter climate in ways that are consistent with observed changes over time (Wolkovich et al., 2012; Menke et al., 2014). However, we lack a robust assessment of how active warming experiments alter the climate conditions experienced by organisms, and the extent to which these conditions are similar to current field conditions or anticipated climate change.

Here, we investigate if and how climate change experiments actually change climate. Using plot-level daily microclimate data from 12 active warming experiments (yielding 41 experiment years) we show the direct and indirect ways that experimental manipulations alter climate. We highlight the challenges associated with quantifying and interpreting experimental shifts in climate and the resulting biological responses. Finally, we use findings from our synthesis to make recommendations for future climate change experiments (Box 1). We focus on *in situ* active warming manipulations, because recent analyses indicate that active warming methods are the most controlled and consistent (Kimball, 2005; Kimball et al., 2008; Aronson and McNulty, 2009; Wolkovich et al., 2012). The data we use were collected between 1991 and 2014 from North American and European climate change experiments (Figure 1, Tables S1, S2) and have been merged into a new, publicly available Climate from Climate Change Experiments (C3E) database (Ettinger and Wolkovich, 2017).

4 Complexities in interpreting experimental climate change

Climate change experiments often include detailed monitoring of climate variables at the plot level, yielding large amounts of data, such as daily or hourly temperature and other climate variables, over the course of the experiment. Biologists, however, are generally interested in the biological responses (e.g., community dynamics, species' growth, abundance, or phenology), which are collected on much coarser timescales (e.g., weekly or annually). Not surprisingly, then, authors typically provide detailed information on the observed biological responses, but report only the mean change in climate over the course of the experiment and whether it matched their target level of change (e.g., Price and Waser, 1998; Rollinson and Kaye, 2012; Clark et al., 2014a,b).

Though the published focus is often on shifts in mean climate variables, imposed climate manipulations actually result in much more complex shifts. The magnitude of change in these manipulations may vary in time and space, and the presence of experimental equipment often unintentionally alters environmental conditions. These factors, discussed below, challenge our interpretation of how experimental warming studies can be used to forecast effects of climate change.

⁷⁸ Effects on local climate vary over time and space

Reporting only the mean temperature difference across the duration of the study hides potentially important variations in daily, seasonal, and annual temperatures among treatments. Using the C3E database, we found that active warming reduces above-ground daily temperature range (DTR) (Table S3, see also Table S2, which details the different methods used to measure temperature). Active warming decreased above-ground

- DTR by differentially affecting maximum and minimum temperatures: warming increased daily minima by 0.84°C per °C of warming target, but only increased daily maxima by 0.51°C per °C of target warming (Tables S3).
- We observed strong seasonal and annual variations in experimental warming effects (Figures 2, 3, Table S4).
- 87 These may be driven by interactions between warming treatments and daily, seasonal, and annual weather
- patterns, since the magnitude of warming may vary as weather conditions change. Both infrared heaters and
- soil cables fail to achieve the target temperatures during rainstorms (Peterjohn et al., 1993; Hoeppner and
- Dukes, 2012) and with windy conditions (Kimball, 2005; Kimball et al., 2008). In addition, treatments are
- often applied inconsistently within or across years. Heat applications are frequently shut off during winter
- 92 months, and some heating methods, even if left on throughout the year, are not capable of applying constant
- warming year-round (e.g. Clark et al., 2014a,b; Hagedorn et al., 2010).
- Treatment effects also vary spatially, adding further complication to interpreting effects of climate change
- 95 experiments. The C3E database contains four studies that used blocked designs, allowing us to examine
- 96 spatial variation in the amount of warming (i.e. the difference between treatment and control plots within
- ⁹⁷ a block). We found that the amount of observed warming varied significantly by more than 1°C among
- 98 blocks (Figure 3, Table S5); block-to-block variation in warming treatment varied by 60-100% of target
- 99 temperatures. These differences in warming levels among blocks may be caused by fine-scale variation in
- vegetation, slope, aspect, soil type, or other factors that can alter wind or soil moisture, which in turn affect
- warming (Peterjohn et al., 1993; Kimball, 2005; Kimball et al., 2008; Hoeppner and Dukes, 2012; Rollinson
- 102 and Kaye, 2015).
- Of course, identical experimental treatments across space and time are not necessary for robust analysis of
- experimental results or for forecasting. Indeed, the spatial and temporal variation we report could improve
- and refine models, and—at least in some regions—may be consistent with contemporary patterns of climate
- change (Stocker et al., 2013). Taking advantage of this variation, however, requires understanding and
- 107 reporting it (e.g., Milcu et al., 2016). In contrast, fine-scale spatial and temporal variations in warming
- treatments are rarely analyzed explicitly, so the implications for interpretation of experimental findings are
- unclear.

Experimental infrastructure alters local climate

Experimental structures themselves can alter temperature and other important biotic and abiotic variables in
ways that are not generally examined nor reported in experimental climate change studies. The importance of
controls that mimic a treatment procedure without actually applying the treatment is widely acknowledged
in biology (e.g., Spector, 2001; Johnson and Besselsen, 2002; Quinn and Keough, 2002). Though some
researchers install treatments with non-functional warming equipment in experimental climate change studies,
the magnitude and implications of structural effects on climate are rarely discussed or interpreted.

To investigate the magnitude of infrastructure effects, we compared temperature and soil moisture data from five active warming studies at two sites: Duke Forest and Harvard Forest (Farnsworth et al., 1995; Clark et al., 2014b; Marchin et al., 2015; Pelini et al., 2011). These were the only studies in the C3E database that monitored climate in two types of control plots: structural controls (i.e., 'shams' or 'disturbance controls,' which contained all the warming infrastructure, such as soil cables or infrared heating units but with no heat applied) and ambient controls with no infrastructure added. Other studies monitored environmental conditions in only structural controls (n=3) or only ambient controls (n=4).

We found that experimental structures altered above-ground and soil temperatures in opposing ways: aboveground temperatures were higher in the structural controls than in ambient controls, whereas soil temperatures were lower in structural controls compared with ambient controls (Figure 4a-d). This general pattern
was consistent across different temperature models (mean, minimum, and maximum temperatures), although
the magnitude varied among seasons, studies, and years (Figure 4a-d, Tables S6-S11). We also found that
experimental infrastructure decreased soil moisture relative to ambient conditions (Figure 4e, Tables S8, S11).

Colder soil temperatures; warmer air temperatures in structural controls, some potential explanations: - Drier because infrastructure reduces rainfall that can hit the ground - Soil temperature colder because blocking sunlight, so less radiative heating - Air temperatures are warmer because metal panels radiatively warm the air around them (and this could continue into the night) and could also block wind, which could stabilize the air and allow it to warm more.

There are several possible reasons for the observed climatic differences between ambient and structural controls. Infrastructure materials may shade the plots, reduce airflow, reduce albedo relative to surroundings,

or otherwise change the energy balance. Structures also interfere with snow accumulation, thereby reducing

particularly if we wish to apply results to forecasting.

155

snowpack and its insulation. This likely plays a bigger role in soil temperature differences at the Harvard Forest sites (exp04, exp07, exp08), where average annual snowfall is over one meter, than at Duke Forest 139 (exp03,exp10), where average snow accumulation each winter is 20 cm or less. Although there is little discussion of measured temperature (or other) differences between ambient and structural control plots in published 141 work (e.g., Farnsworth et al., 1995; Pelini et al., 2011; Clark et al., 2014a,b), Clark et al. (2014b) mention 142 that "control of the air temperature was less precise, in part due to air scooping on windy days." Marchin 143 et al. (2015) note that structural controls had mean spring air temperatures about 0.5°C or more above ambient temperatures and Peterjohn et al. (1994) reported cooler soil temperatures in structural controls than in ambient controls at shallow soil depths. Similarly, we found the greatest difference in soil temperature 146 between structural and ambient controls in shallow soils (e.g. exp10, soil depth = 2cm). Further, while the 147 focus to date has been largely on these abiotic impacts of experimental structures, such structures may also 148 alter herbivory and other biotic conditions (Kennedy, 1995; Moise and Henry, 2010; Wolkovich et al., 2012; 149 Hoeppner and Dukes, 2012). 150 Most warming experiments calculate focal response variables relative to ambient controls (e.g., Marchin 151 et al., 2015), which our analyses suggest will not properly account for infrastructure effects. Because the 152 design of these experiments may influence abiotic and biotic responses in warming experiments, improved documentation and analysis of infrastructure effects is an important next step in climate change experiments, 154

56 Secondary and feedback effects of climate change manipulations

Climate change experiments often seek to manipulate one or two climate variables, usually temperature 157 and precipitation, but manipulating either of these variables also alters the other. Wet soils: more NRG 158 into latent heating (evapotransition) and thus less NRG sensible heating (heating the air). Precipitation 159 treatments typically reduce temperatures in climate change manipulations (Sherry et al., 2007; Rollinson and Kaye, 2012; McDaniel et al., 2014b): McDaniel et al. (2014) observed that a twenty percent increase in precipitation reduced mean hourly temperatures by 0.3°C over the course of their two-year experiment. 162 Experimental warming typically increases vapor pressure deficit and reduces soil water content (e.g., Sherry 163 et al., 2007; Morin et al., 2010; Pelini et al., 2014; Templer et al., 2016). Of the twelve experiments in the 164 C3E database, we examined the ten that measured and reported soil moisture and found that experimental 165

warming reduced soil moisture by 3.0%, on average (Figure 5, Table S13), and that this reduction occurred
at a rate of 0.43% per degree of target warming (Table S12). Thus, although active warming experiments
may not be explicitly designed to manipulate soil moisture, soil moisture is unavoidably affected by changing
temperatures. Warming increases evaporative demand (note: VPD is a quantity of the air so perhaps do not
want to use this term for places just warming the soils) and thus soils dry. No need to mention turbulence
(which typically goes up with drier air but not always and this seemed confusing to me anyway).

Warming and precipitation treatments, and their secondary effects on soil moisture and other abiotic factors, can also alter the biotic environment, which may produce cascading effects. Many studies have found
shifts from herbaceous to woody plant communities with experimental warming (e.g., Rollinson and Kaye,
2012; McDaniel et al., 2014b,a; Harte et al., 2015); this, in turn, can alter microbial and herbaceous plant
communities. These community shifts may change competitive dynamics and affect resource levels, such as
moisture, carbon, and nutrients in the soil (McDaniel et al., 2014b,a; Harte et al., 2015), and cause positive
feedbacks to local climate change (Harte et al., 2015).

The widespread presence of unintended secondary effects of climate change manipulations highlights the importance of measuring environmental conditions at the plot level, and using these measurements in analysis and interpretation of results. Many climate change experiments—including seven of the 12 in the C3E database—analyze warming and/or precipitation treatments as simple categorical predictors (e.g., as in a two-way ANOVA). Our findings, however, demonstrate a clear need for alternative modelling approaches to fully understand the experimental results and to make mechanistic links between changes in climate and ecological responses. One straightforward alternative is to include the continuous climate data (e.g., plot-level mean temperatures) as predictors of the focal response variable, such as phenological state or species density (e.g., Marchin et al., 2015; Pelini et al., 2014).

Biological implications

We have highlighted a suite of factors that complicate interpretation of warming experiments. These largely unintended alterations are likely to have biological implications for many of the major responses studied in warming experiments (e.g., Figure 7). Interpretation of experimental climate change effects on biological responses may be misleading, because the intended climate treatments (i.e., categorical comparisons or target

warming levels) are generally used as explanatory variables in analyses. The interpretation is likely to be
altered by using fine-scale, measured climate as explanatory variables. Detailed examination of multiple microclimate variables (e.g., plot-level temperature and soil moisture) will allow a more complete understanding
of the indirect, as well as direct, effects of treatments on abiotic and biotic drivers of focal responses.

Plant phenology provides one example of a biological response that is muted in experiments versus observational studies (Figure 7b). This is because phenology has a complex dependence on temperature and water availability (as well as other factors). Although phenology is generally advanced by higher spring temperatures, it can also be delayed by increased winter temperature (which delays endodormancy break). In addition, reduced water availability during the spring can slow cell elongation and delay budburst (Peñuelas et al., 2004; Ourcival and Rambal, 2011; Craine et al., 2012; Matthews and Mazer, 2016).

Effects of these different drivers may be responsible for the observed discrepancy between observational and experimental phenological responses to warming (Wolkovich et al., 2012). Other biological responses may be exaggerated in experiments when direct and indirect effects of climate manipulations work in concert (Figure 7c). Accounting for both direct and indirect effects of warming is critical for accurate interpretation of the consequences of climate change (Kharouba et al., 2015). Since climate change experiments have indirect effects on the biotic as well as abiotic environment (Hoeppner and Dukes, 2012; Pelini et al., 2014; Diamond et al., 2016), a critical question is the extent to which these indirect effects are accurate forecasts of future shifts that are likely to occur with climate change, or due to side-effects that are unlikely to occur outside of experimental systems (Moise and Henry, 2010; Diamond et al., 2013).

212 Drying, light removal, change in diurnal temperature ranges are the main artifacts (says Lizzie).

Ben says must couch in terms of how to interpret and compare results: Some very wet places will probably not see significant warming to affect Tmax more than Tmin, but thing would be different in California (aka dried places).

Diurnal temp range (minima changing more than maxima changes) in general (CITE), but not true in all places; in some places where soil is drying because that increased maxima temperatures more. Soil drying is not an expected outcome of

Soil drying with warming is very regional: places that are dry (US Southwest, US west,)âĂę other places like Eastern US where things will be warmer and precip will increase (and eastern US is trending wetter).

- Tmin has been increasing more rapidly than Tmax, reducing diurnal temperature range (DTR), but these trends are strongest and most significant from 1950-1980. Largely insignificant trends from 1980 and on. (Thorne et al., 2016; Vose et al., 2005) Soils are expected to dry out regionally with warming because of reductions in precipitation and increased evaporative demand in a warmer atmosphere. (Dai, 2013)
- The southwest is one region where models show robust drying (Seager et al., 2013)
- The northeast US, however, has been trending wetter over time and is expected to be wetter in the future (Seager et al., 2014; Shuman and Burrell, 2017)
- Another artefact to discuss, that we do not mention anywhere yet: the fact that in warming studies, the
 temperature increase is instantaneous, while climate change obviously happens gradually (see for example
 the modelling study by Luo and Reynolds, Ecology, 1999). This also relates to possible differences between
 short and longer term studies, cf. the different conclusions drawn from FACE experiments in the first years
 vs. later on.

• Conclusions

As climate change continues across the globe, ecologists are challenged to not only document impacts but make quantitative, robust predictions. Our ability to meet this challenge requires a nuanced mechanistic 235 understanding of how climate directly and indirectly alters biological processes. Climate change experiments, 236 which have been underway for nearly four decades (e.g., Tamaki et al., 1981; Carlson and Bazzaz, 1982), 237 provide invaluable information about biological responses to climate change. Yet the full range of changes 238 in environmental conditions imposed by these experiments is rarely presented. We have compiled the first 239 database of fine-scale climate data from multiple warming experiments and shown how time, space, and experimental artifacts may hinder simple interpretations of these climate change experiments. We hope this provides a foundation for gaining the most knowledge and utility from existing experiments, for designing better experiments and models in the future (see Box 1), and for improved understanding of biological 243 responses and feedbacks in a changing world

Acknowledgements

- We are grateful to those who shared their experimental climate data with us and others in the C3E database.

 We thank the Radcliffe Institute for Advanced Study at Harvard University, which provided funding for an

 Exploratory Seminar at which the ideas in this paper were conceived. This research was also supported by

 the National Science Foundation (NSF DBI 14-01854 to A.E.). Any opinion, findings, and conclusions or

 recommendations expressed in this material are those of the authors and do not necessarily reflect the views
- of the National Science Foundation.

261

262

263

264

266

267

268

269

271

Box 1: Recommendations for future climate change experiments

- 1. Collect and analyze fine-scale climate data. This includes analyzing and interpreting minimum and
 maximum values, as well as variance and critical thresholds (e.g., the number and duration of freeze-thaw
 events and accumulated chilling hours, McDaniel et al., 2014b; Vasseur et al., 2014). We suggest saving
 the raw data from data loggers (often collected at hourly or higher resolution) to allow quantification
 of variance (and other summaries) at different temporal resolutions. In assessing which frequency of
 measurements is most appropriate for analyses (e.g., hourly, twice daily), it is critical to consider the
 chronobiology of the event and organisms of interest. For ants, this might mean that temperatures be
 monitored every minute (Helm and Shavit, 2017); for bacteria, even more frequently.
 - 2. Analyze measured climate variables rather than targets. There can be substantial variation in the effects of warming and precipitation treatments among plots and across time (Figure 3). Analyzing measured climate will allow much more in-depth understanding of the drivers and biological effects of variation in temperature and moisture.
 - 3. Publish high quality, usable data and metadata. Given that in situ active climate manipulations are logistically challenging and expensive (Aronson and McNulty, 2009), and that they often produce a large volume of fine-scale climate data, good curation and data sharing will ensure wider use and deeper understanding of these valuable data. When studying biological implications of a global challenge as large as climate change, progress will come from designing and reporting experiments in ways that facilitate an eventual global data set.
 - 4. Include both structural and ambient controls and collect, use, and report data collected within them.

- Fewer than half of the studies in our C3E database reported data from these two control types (5 out of 12 studies); however, all experiments that did include both control types showed significant effects of infrastructure (Figure 4).
- 5. Design relevant manipulations by consulting observational records and forecasts, including seasonal and annual variation in projected warming. When it is not possible or desirable to match anticipated changes in climate, studies should report how imposed treatments compare to projected changes and past observations (e.g., Hoover et al., 2014). In addition, if continuous treatments are not applied throughout the study, the seasonality and timing of treatments should be explicitly reported and the climate should be monitored throughout.
- 6. Maximize the duration of climate change experiments by running some experiments for as long as possible. Long-term responses of individuals and populations can differ from transient responses (Saleska et al., 2002; Franklin, 1989; Giasson et al., 2013; Harte et al., 2015). Well-designed and well-supported longer warming experiments will allow study of how inter-annual variations interact with climate change treatments, particularly when combined with observational studies and modeling (Luo et al., 2011).

References

- Aronson, E. L., and S. G. McNulty. 2009. Appropriate experimental ecosystem warming methods by ecosystem, objective, and practicality. Agricultural and Forest Meteorology 149:1791–1799.
- Box, G. E., W. G. Hunter, J. S. Hunter, et al. 1978. Statistics for Experimenters: An Introduction to Design,

 Data Analysis, and Model Building. John Wiley and Sons New York.
- Carlson, R. W., and F. A. Bazzaz. 1982. Photosynthetic and growth response to fumigation with so2 at elevated co2 for c3 and c4 plants. Oecologia 54:50–54.
- Chuine, I., M. Bonhomme, J.-M. Legave, I. García de Cortázar-Atauri, G. Charrier, A. Lacointe, and
 T. Améglio. 2016. Can phenological models predict tree phenology accurately in the future? the unrevealed hurdle of endodormancy break. Global change biology 22:3444–3460.
- Chuine, I., X. Morin, L. Sonié, C. Collin, J. Fabreguettes, D. Degueldre, J.-L. Salager, and J. Roy. 2012.

 Climate change might increase the invasion potential of the alien c4 grass setaria parviflora (poaceae) in

 the mediterranean basin. Diversity and Distributions 18:661–672.

- Clark, J. S., J. Melillo, J. Mohan, and C. Salk. 2014a. The seasonal timing of warming that controls onset of the growing season. Global Change Biology 20:1136–1145.
- Clark, J. S., C. Salk, J. Melillo, and J. Mohan. 2014b. Tree phenology responses to winter chilling, spring warming, at north and south range limits. Functional Ecology 28:1344–1355.
- ³⁰³ Cleland, E. E., N. R. Chiariello, S. R. Loarie, H. A. Mooney, and C. B. Field. 2006. Diverse responses of
- phenology to global changes in a grassland ecosystem. Proceedings of the National Academy of Sciences
- of the United States of America 103:13740–13744.
- Cox, P. M., R. A. Betts, C. D. Jones, S. A. Spall, and I. J. Totterdell. 2000. Acceleration of global warming
 due to carbon-cycle feedbacks in a coupled climate model. Nature 408:184–187.
- Craine, J. M., E. M. Wolkovich, E. G. Towne, and S. W. Kembel. 2012. Flowering phenology as a functional trait in a tallgrass prairie. New Phytologist 193:673–682.
- Dai, A. 2013. Increasing drought under global warming in observations and models. Nature Climate Change
 3:52–58.
- Diamond, S. E., L. M. Nichols, S. L. Pelini, C. A. Penick, G. W. Barber, S. H. Cahan, R. R. Dunn, A. M.
- Ellison, N. J. Sanders, and N. J. Gotelli. 2016. Climate warming destabilizes forest ant communities.
- Science Advances 2:e1600842.
- Diamond, S. E., C. A. Penick, S. L. Pelini, A. M. Ellison, N. J. Gotelli, N. J. Sanders, and R. R. Dunn.
- 2013. Using physiology to predict the responses of ants to climatic warming. Integrative and comparative
- biology 53:965–974.
- Dukes, J. S., and H. A. Mooney. 1999. Does global change increase the success of biological invaders? Trends in Ecology and Evolution 14:135–139.
- Ettinger, A., and E. Wolkovich. 2017. Climate from climate change experiments (c3e).
- Farnsworth, E., J. Nunez-Farfan, S. Careaga, and F. Bazzaz. 1995. Phenology and growth of three temperate
- forest life forms in response to artificial soil warming. Journal of Ecology pages 967–977.
- Field, C. B., D. B. Lobell, H. A. Peters, and N. R. Chiariello. 2007. Feedbacks of terrestrial ecosystems to
 climate change*. Annu. Rev. Environ. Resour. 32:1–29.

- Franklin, J. F. 1989. Importance and justification of long-term studies in ecology. Pages 3–19 in Long-term
- studies in ecology. Springer.
- Gelman, A. 2014. Experimental reasoning in social science experiments, chap. 7, pages 185–195. New Haven,
- 328 CT: Yale University Press.
- Giasson, M.-A., A. M. Ellison, R. Bowden, P. M. Crill, E. Davidson, J. Drake, S. Frey, J. Hadley, M. Lavine,
- J. Melillo, et al. 2013. Soil respiration in a northeastern us temperate forest: a 22-year synthesis. Ecosphere
- 331 4:1–28.
- Gruner, D. S., M. E. Bracken, S. A. Berger, B. K. Eriksson, L. Gamfeldt, B. Matthiessen, S. Moorthi,
- U. Sommer, and H. Hillebrand. 2016. Effects of experimental warming on biodiversity depend on ecosystem
- type and local species composition. Oikos .
- Hagedorn, F., M. Martin, C. Rixen, S. Rusch, P. Bebi, A. Zürcher, R. T. Siegwolf, S. Wipf, C. Escape, J. Roy,
- et al. 2010. Short-term responses of ecosystem carbon fluxes to experimental soil warming at the swiss
- alpine treeline. Biogeochemistry 97:7–19.
- Harte, J., S. R. Saleska, and C. Levy. 2015. Convergent ecosystem responses to 23-year ambient and manip-
- ulated warming link advancing snowmelt and shrub encroachment to transient and long-term climate-soil
- carbon feedback. Global change biology 21:2349–2356.
- Helm, B., and A. Shavit. 2017. Dissecting and reconstructing time and space for replicable biological research,
- pages 233–249. New Haven, CT: Yale University Press.
- Hobbie, S. E., A. Shevtsova, and F. S. Chapin III. 1999. Plant responses to species removal and experimental
- warming in alaskan tussock tundra. Oikos pages 417–434.
- Hoeppner, S. S., and J. S. Dukes. 2012. Interactive responses of old-field plant growth and composition to
- warming and precipitation. Global Change Biology 18:1754–1768.
- Hoover, D. L., A. K. Knapp, and M. D. Smith. 2014. Resistance and resilience of a grassland ecosystem to
- climate extremes. Ecology 95:2646–2656.
- January, I., J. S. Clark, M. C. Dietze, K. Feeley, M. Hersh, S. LaDeau, A. McBride, N. E. Welch, and M. S.
- Wolosin. 2006. Predicting biodiversity change: Outside the climate envelope, beyond the species-area
- 351 curve. Ecology 87:1896–1906.

- Johnson, P. D., and D. G. Besselsen. 2002. Practical aspects of experimental design in animal research. ILAR journal 43:202–206.
- Kennedy, A. 1995. Temperature effects of passive greenhouse apparatus in high-latitude climate change experiments. Functional Ecology pages 340–350.
- Kharouba, H. M., M. Vellend, R. M. Sarfraz, and J. H. Myers. 2015. The effects of experimental warming on the timing of a plant–insect herbivore interaction. Journal of Animal Ecology 84:785–796.
- Kimball, B. 2005. Theory and performance of an infrared heater for ecosystem warming. Global Change Biology 11:2041–2056.
- Kimball, B. A., M. M. Conley, S. Wang, X. Lin, C. Luo, J. Morgan, and D. Smith. 2008. Infrared heater arrays for warming ecosystem field plots. Global Change Biology 14:309–320.
- Luo, Y., J. Melillo, S. Niu, C. Beier, J. S. Clark, A. T. Classen, E. Davidson, J. S. Dukes, R. Evans, C. B.
- Field, et al. 2011. Coordinated approaches to quantify long-term ecosystem dynamics in response to global
- change. Global Change Biology 17:843–854.
- Marchin, R. M., C. F. Salk, W. A. Hoffmann, and R. R. Dunn. 2015. Temperature alone does not explain
- phenological variation of diverse temperate plants under experimental warming. Global change biology
- 367 21:3138–3151.
- Matthews, E. R., and S. J. Mazer. 2016. Historical changes in flowering phenology are governed by
- temperature× precipitation interactions in a widespread perennial herb in western north america. New
- ³⁷⁰ Phytologist 210:157–167.
- 371 McDaniel, M., J. Kaye, and M. Kaye. 2014a. Do "hot moments" become hotter under climate change?
- soil nitrogen dynamics from a climate manipulation experiment in a post-harvest forest. Biogeochemistry
- 373 121:339-354.
- McDaniel, M., R. Wagner, C. Rollinson, B. Kimball, M. Kaye, and J. Kaye. 2014b. Microclimate and ecological
- threshold responses in a warming and wetting experiment following whole tree harvest. Theoretical and
- applied climatology 116:287–299.
- Menke, S. B., J. Harte, and R. R. Dunn. 2014. Changes in ant community composition caused by 20 years
- of experimental warming vs. 13 years of natural climate shift. Ecosphere 5:1–17.

- ³⁷⁹ Milcu, A., R. Puga-Freitas, A. M. Ellison, M. Blouin, S. Scheu, T. Girin, G. Frechet, L. Rose, M. Scherer-
- Lorenzen, S. Barot, et al. 2016. Systematic variability enhances the reproducibility of an ecological study.
- ³⁸¹ bioRxiv page 080119.
- Moise, E. R., and H. A. Henry. 2010. Like moths to a street lamp: exaggerated animal densities in plot-level
- global change field experiments. Oikos 119:791–795.
- Morin, X., J. Roy, L. Sonié, and I. Chuine. 2010. Changes in leaf phenology of three european oak species in
- response to experimental climate change. New Phytologist 186:900–910.
- Ohlemüller, R., E. S. Gritti, M. T. Sykes, and C. D. Thomas. 2006. Towards european climate risk surfaces:
- the extent and distribution of analogous and non-analogous climates 1931–2100. Global ecology and
- biogeography 15:395–405.
- ourcival, J., and S. Rambal. 2011. Phenological responses to extreme droughts in a mediterranean forest.
- 390 Glob Change Biol 17:1036–1048.
- Parmesan, C. 2006. Ecological and evolutionary responses to recent climate change. Annual Review of
- Ecology Evolution and Systematics 37:637–669.
- Pearson, R. G., and T. P. Dawson. 2004. Bioclimate envelope models: what they detect and what they hide
- response to hampe (2004). Global Ecology and Biogeography 13:471–473.
- Pelini, S., S. Diamond, L. Nichols, K. Stuble, A. M. Ellison, N. Sanders, R. Dunn, and N. Gotelli. 2014. Geo-
- graphic differences in effects of experimental warming on ant species diversity and community composition.
- Ecosphere 5:1-12.
- Pelini, S. L., F. P. Bowles, A. M. Ellison, N. J. Gotelli, N. J. Sanders, and R. R. Dunn. 2011. Heating up the
- forest: open-top chamber warming manipulation of arthropod communities at harvard and duke forests.
- Methods in Ecology and Evolution 2:534–540.
- Peñuelas, J., I. Filella, X. Zhang, L. Llorens, R. Ogaya, F. Lloret, P. Comas, M. Estiarte, and J. Terradas.
- 2004. Complex spatiotemporal phenological shifts as a response to rainfall changes. New Phytologist
- 403 161:837–846.
- ⁴⁰⁴ Peterjohn, W. T., J. M. Melillo, F. P. Bowles, and P. A. Steudler. 1993. Soil warming and trace gas fluxes:
- experimental design and preliminary flux results. Oecologia 93:18–24.

- Price, M. V., and N. M. Waser. 1998. Effects of experimental warming on plant reproductive phenology in a subalpine meadow. Ecology 79:1261–1271.
- Quinn, G. P., and M. J. Keough. 2002. Experimental design and data analysis for biologists. Cambridge
 University Press.
- Reich, P. B., K. M. Sendall, K. Rice, R. L. Rich, A. Stefanski, S. E. Hobbie, and R. A. Montgomery. 2015.
- 411 Geographic range predicts photosynthetic and growth response to warming in co-occurring tree species.
- Nature Clim.Change 5:148–152.
- Rollinson, C. R., and M. W. Kaye. 2012. Experimental warming alters spring phenology of certain plant functional groups in an early successional forest community. Global Change Biology 18:1108–1116.
- 2015. Modeling monthly temperature in mountainous ecoregions: importance of spatial scale for ecological research. Climate Research 64:99–110.
- 417 Saleska, S. R., M. R. Shaw, M. L. Fischer, J. A. Dunne, C. J. Still, M. L. Holman, and J. Harte. 2002.
- Plant community composition mediates both large transient decline and predicted long-term recovery of soil carbon under climate warming. Global Biogeochemical Cycles 16.
- Seager, R., D. Neelin, I. Simpson, H. Liu, N. Henderson, T. Shaw, Y. Kushnir, M. Ting, and B. Cook. 2014.
- Dynamical and thermodynamical causes of large-scale changes in the hydrological cycle over north america
- in response to global warming. Journal of Climate 27:7921–7948.
- Seager, R., M. Ting, C. Li, N. Naik, B. Cook, J. Nakamura, and H. Liu. 2013. Projections of declining
 surface-water availability for the southwestern united states. Nature Climate Change 3:482.
- Shaver, G. R., J. Canadell, F. S. Chapin, J. Gurevitch, J. Harte, G. Henry, P. Ineson, S. Jonasson, J. Melillo,
- 426 L. Pitelka, et al. 2000. Global warming and terrestrial ecosystems: A conceptual framework for analysis
- ecosystem responses to global warming will be complex and varied. ecosystem warming experiments hold
- great potential for providing insights on ways terrestrial ecosystems will respond to upcoming decades of
- climate change. documentation of initial conditions provides the context for understanding and predicting
- ecosystem responses. BioScience 50:871–882.
- Sheldon, K. S., S. Yang, and J. J. Tewksbury. 2011. Climate change and community disassembly: impacts
- of warming on tropical and temperate montane community structure. Ecology Letters 14:1191–1200.

- Sherry, R. A., X. Zhou, S. Gu, J. A. A. 3rd, D. S. Schimel, P. S. Verburg, L. L. Wallace, and Y. Luo. 2007.
- Divergence of reproductive phenology under climate warming. Proceedings of the National Academy of
- Sciences of the United States of America 104:198–202.
- 436 Shukla, J., and Y. Mintz. 1982. Influence of land-surface evapotranspiration on the earth's climate. Science
- 215:1498-1501.
- Shuman, B. N., and S. A. Burrell. 2017. Centennial to millennial hydroclimatic fluctuations in the humid
- northeast united states during the holocene. Quaternary Research pages 1–11.
- Spector, R. 2001. Progress in the search for ideal drugs. Pharmacology 64:1–7.
- 441 Stocker, T., D. Qin, G. Plattner, M. Tignor, S. Allen, J. Boschung, A. Nauels, Y. Xia, B. Bex, and B. Midgley.
- 2013. Ipcc, 2013: climate change 2013: the physical science basis. contribution of working group i to the
- fifth assessment report of the intergovernmental panel on climate change.
- Swab, R. M., H. M. Regan, D. A. Keith, T. J. Regan, and M. K. J. Ooi. 2012. Niche models tell half the story:
- spatial context and life-history traits influence species responses to global change. Journal of Biogeography
- 39:1266–1277.
- 447 Tamaki, G., M. A. Weiss, and G. E. Long. 1981. Evaluation of plant density and temperature in predator-prey
- interactions in field cages. Environmental Entomology 10:716–720.
- Templer, P. H., N. G. Phillips, A. M. Ellison, and S. L. Pelini. 2016. Ecosystem warming increases sap flow
- rates of northern red oak trees. Ecosphere 7.
- Thomas, C. D., A. Cameron, R. E. Green, M. Bakkenes, L. J. Beaumont, Y. C. Collingham, B. F. N. Erasmus,
- M. F. de Siqueira, A. Grainger, L. Hannah, L. Hughes, B. Huntley, A. S. van Jaarsveld, G. F. Midgley,
- 453 L. Miles, M. A. Ortega-Huerta, A. T. Peterson, O. L. Phillips, and S. E. Williams. 2004. Extinction risk
- from climate change. Nature 427:145–148.
- Thorne, P., M. Donat, R. Dunn, C. Williams, L. Alexander, J. Caesar, I. Durre, I. Harris, Z. Hausfather,
- P. Jones, et al. 2016. Reassessing changes in diurnal temperature range: Intercomparison and evaluation
- of existing global data set estimates. Journal of Geophysical Research: Atmospheres 121:5138–5158.
- 458 Urban, M. C., J. J. Tewksbury, and K. S. Sheldon. 2012. On a collision course: competition and dispersal

- differences create no-analogue communities and cause extinctions during climate change. Proceedings of
- the Royal Society B-Biological Sciences 279:2072–2080.
- Vasseur, D. A., J. P. DeLong, B. Gilbert, H. S. Greig, C. D. Harley, K. S. McCann, V. Savage, T. D. Tunney,
- and M. I. O'Connor. 2014. Increased temperature variation poses a greater risk to species than climate
- warming. Proceedings of the Royal Society of London B: Biological Sciences 281:20132612.
- 464 Vose, R. S., D. R. Easterling, and B. Gleason. 2005. Maximum and minimum temperature trends for the
- globe: An update through 2004. Geophysical Research Letters 32.
- Williams, J. W., and S. T. Jackson. 2007. Novel climates, no-analog communities, and ecological surprises.
- Frontiers in Ecology and the Environment 5:475–482.
- Williams, J. W., S. T. Jackson, and J. E. Kutzbacht. 2007. Projected distributions of novel and disappearing
- climates by 2100 ad. Proceedings of the National Academy of Sciences of the United States of America
- 104:5738-5742.
- Wolkovich, E. M., B. I. Cook, J. M. Allen, T. M. Crimmins, J. L. Betancourt, S. E. Travers, S. Pau,
- J. Regetz, T. J. Davies, N. J. B. Kraft, T. R. Ault, K. Bolmgren, S. J. Mazer, G. J. McCabe, B. J. McGill,
- 473 C. Parmesan, N. Salamin, M. D. Schwartz, and E. E. Cleland. 2012. Warming experiments underpredict
- plant phenological responses to climate change. Nature 485:494–497.

Figures Figures

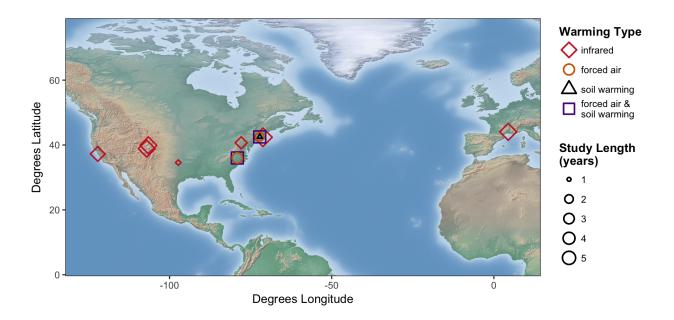


Figure 1: Climate data from 12 climate change experiments in North America and Europe are included in the C3E database and analyzed here. See Supplemental Materials, Tables S1 and S2 for details.

Daily Mean Soil Temperature Difference

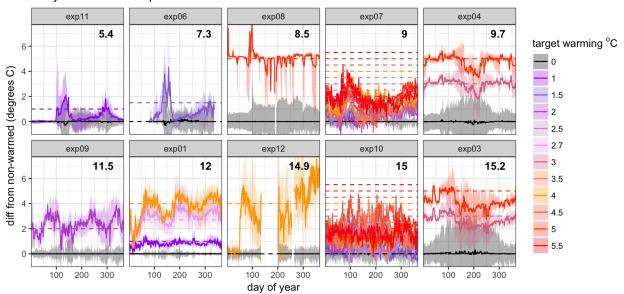


Figure 2: Deviations in daily observed warming from mean soil temperature for 10 study sites. Solid lines show observed difference between warming treatment (colors) and control (black) plots, averaged across replicates and years; shading shows 95% confidence intervals. Dashed lines represent target warming levels. Two sites not shown here did not monitor soil temperature; we also excluded data from plots that manipulated precipitation. Mean annual temperature for experimental sites are shown in the upper right corner of each panel; panels are arranged by increasing annual temperature.

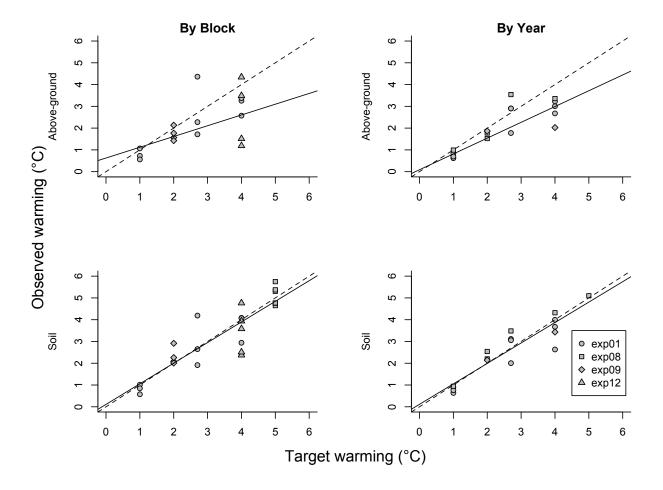


Figure 3: Observed warming (i.e., the difference between treatment and control plots) over space and time, for above-ground and below-ground temperatures, excluding data from plots that manipulated precipitation. The solid line is the fitted relationship between observed and target warming and the dashed line shows when observed warming is exactly equal to target warming (1:1). See Supplemental Materials (especially Tables S4 and S5) for details.

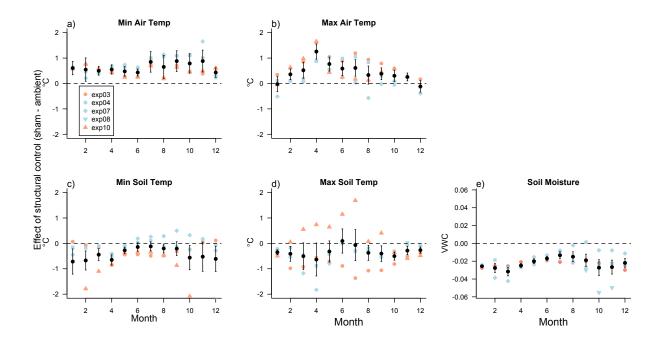


Figure 4: Deviations in measured abiotic variables by month in structural controls compared to ambient controls (i.e., with no control chambers or warming infrastructure in place). Above-ground temperatures were higher, whereas below-ground temperature and soil moisture were lower in structural controls compared with ambient controls. We show overall (fixed) effects in black from monthly mixed-effects models; site-level random effects are shown by symbols in blue (for the three studies conducted at Harvard Forest in Massachusetts, USA) and pink (the two studies conducted at Duke Forest in North Carolina, USA).

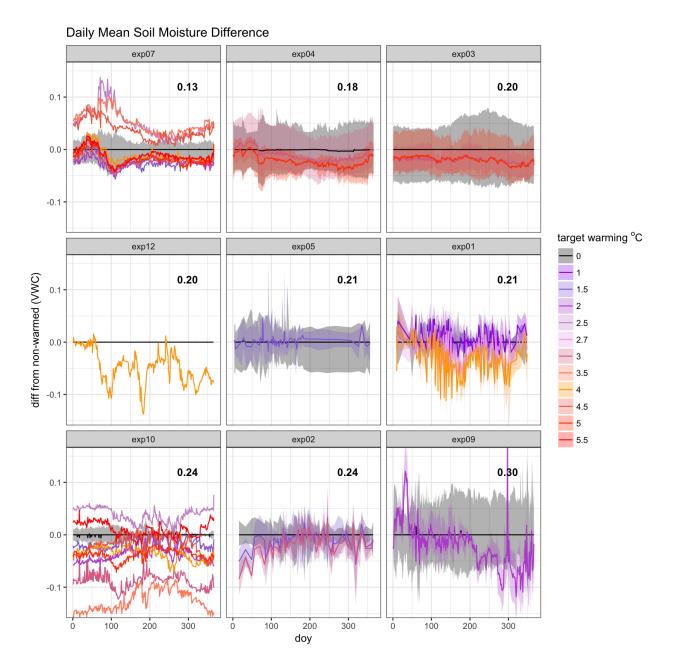


Figure 5: **Deviations in daily observed soil moisture**, shown for the nine study sites that continuously monitored soil moisture, excluding data from plots that manipulated precipitation. Black lines represent control plots, and colored lines represent warming treatments with various target warming levels. The number of temperature treatment levels vary from one (e.g. exp08, exp11) to nine (exp07 and exp10, which used an unreplicated regression design). Mean annual soil moisture for the experimental site is shown in the upper right corner of each plot, and plots are arranged by increasing mean soil moisture. All experiments measured soil moisture in volumetric water content (VWC, as a proportion of the soil volume in the sample, scaled from 0 to 1).

٠

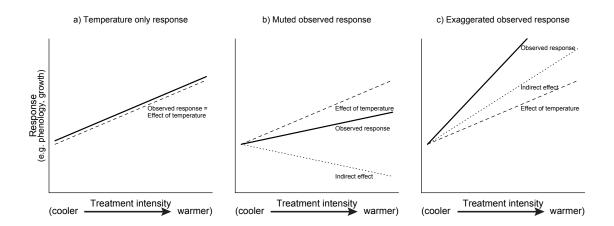


Figure 6: Possible biological responses to experimental climate change and their interpretation. Direct responses to temperature alone (a) can be easily understood. Complications arise when biological responses are a mix of the direct and indirect effects of experimental warming. Then experimental warming may cause biological responses to be muted (b) or exaggerated (c). Slopes of these example lines assume a linear response with additive direct and indirect effects. The relationship between these effects could be more complex (e.g., nonlinear; antagonistic, multiplicative, or otherwise interactive).

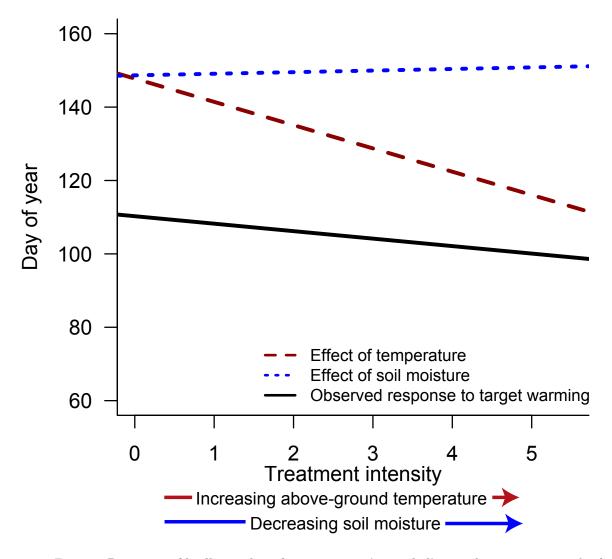


Figure 7: Response of budburst day of year to experimental climate change is an example of a muted response (a): the observed response to target warming temperature (i.e., the coefficient of a model fit with only target temperature as the explanatory variable, black line) is weaker than the effect of temperature in a more biologically meaningful model that includes both measured above-ground temperature (dashed red line) and soil moisture (dotted blue line). This is because both temperature and soil moisture affect the timing of plant budburst, and experimental warming dries out the soil (b) in addition to increasing temperatures. Or could say "This is because experimental warming dries out the soil (b) in addition to increasing temperatures, and both climate variables affect the timing of budburst. Whereas increasing temperatures advance budburst decreasing soil moisture has a delaying effect." See Tables S14 & S15 for statistical details.