SS4850 Final Project

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Explanatory analysis

```
HCVdata <- read.csv(file = 'hcvdat0.csv')</pre>
suppressMessages(library(dplyr))
drop <- c("X")
HCVdata = HCVdata[,!(names(HCVdata) %in% drop)]
colnames(HCVdata) = c("Category", "Age", "Sex", "ALB", "ALP", "ALT", "AST",
                      "BIL", "CHE", "CHOL", "CREA", "GGT", "PROT")
# 1 and 2 for blood donor and suspect blood donor, 2,3,4 for hepatitis,
fibrosis, cirrhosis respectively
HCVdata$Category <- factor(HCVdata$Category,labels=c(1,2,3,4,5),</pre>
                           levels=c('0=Blood Donor',
                                     '0s=suspect Blood Donor',
                                     '1=Hepatitis',
                                     '2=Fibrosis',
                                     '3=Cirrhosis'))
# 0 for male, 1 for female
HCVdata$Sex <- factor(HCVdata$Sex,levels=c("m","f"),labels=c("0","1"))</pre>
HCVdata=na.omit(HCVdata[c("Category", "Age", "Sex", "ALB", "ALP", "ALT", "AST",
                          "BIL", "CHE", "CHOL", "CREA", "GGT", "PROT")])
for(var in 1:13)
  HCVdata[,var]=as.numeric(HCVdata[,var])
}
tail(HCVdata)
##
       Category Age Sex ALB
                             ALP ALT
                                         AST BIL CHE CHOL CREA
                                                                    GGT PROT
## 608
              5 52
                      2
                         39
                            37.0 1.3 30.4 21 6.33 3.78 158.2 142.5 82.7
              5
## 609
                 58
                      2 34 46.4 15.0 150.0 8 6.26 3.98 56.0 49.7 80.6
              5 59
                      2 39 51.3 19.6 285.8 40 5.77 4.51 136.1 101.1 70.5
## 610
              5 62
                      2 32 416.6 5.9 110.3 50 5.57 6.30 55.7 650.9 68.5
## 611
              5 64
                      2 24 102.8 2.9 44.4 20 1.54 3.02 63.0 35.9 71.3
## 612
## 613
              5 64
                      2 29 87.3 3.5 99.0 48 1.66 3.63 66.7 64.2 82.0
```

EDA!!!

set.seed(4850)

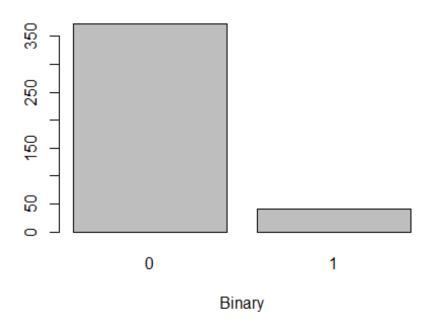
#barplot on binary

```
#class information, distribution of "category"
library(dplyr)
HCVdata %>%
  group_by(Category) %>%
  summarise(no rows = length(Category))
## # A tibble: 5 x 2
##
     Category no_rows
##
        <dbl>
                <int>
## 1
            1
                   526
## 2
            2
                    7
## 3
            3
                    20
## 4
            4
                    12
## 5
            5
                    24
#mean of each variable
colMeans(HCVdata)
                                        ALB
                                                              ALT
                                                                        AST
## Category
                   Age
                              Sex
                                                   ALP
BIL
## 1.303905 47.417657 1.383701 41.624278 68.123090 26.575382 33.772835
11.018166
##
         CHE
                  CHOL
                             CREA
                                         GGT
                                                  PROT
    8.203633 5.391341 81.669100 38.198472 71.890153
#median of each variable
apply(HCVdata,2,median)
## Category
                           Sex
                                    ALB
                                              ALP
                                                       ALT
                                                                 AST
                                                                          BIL
                 Age
##
       1.00
               47.00
                          1.00
                                  41.90
                                            66.20
                                                     22.70
                                                               25.70
                                                                         7.10
##
        CHE
                CHOL
                          CREA
                                    GGT
                                             PROT
       8.26
                5.31
                         77.00
                                  22.80
                                            72.10
train and test split 7:3
## train and test set 7:3
HCVdata<-HCVdata[complete.cases(HCVdata),]</pre>
set.seed(4850)
sample <- sample.int(n = nrow(HCVdata), size = floor(.70*nrow(HCVdata)),</pre>
replace = F)
train <- HCVdata[sample, ]</pre>
test <- HCVdata[-sample, ]
roc with random forest
#binary category
#1 for HCV, 0 for donor(regular/suspect)
```

train\$binary <- ifelse(as.numeric(train\$Category)>2,1,0)
test\$binary <- ifelse(as.numeric(test\$Category)>2,1,0)

```
counts <- table(train$binary)
barplot(counts, main="HCV distribution",
    xlab="Binary")</pre>
```

HCV distribution



```
#Load libraries
library(randomForest)
## randomForest 4.6-14
## Type rfNews() to see new features/changes/bug fixes.
##
## Attaching package: 'randomForest'
## The following object is masked from 'package:dplyr':
##
       combine
##
rfmod <- randomForest(formula = binary ~ .-Category, data = train, ntree =</pre>
10, maxnodes= 100, norm.votes = F)
## Warning in randomForest.default(m, y, ...): The response has five or fewer
## unique values. Are you sure you want to do regression?
suppressMessages(library(dplyr))
testm <- mutate(test,</pre>
                rf_pred = predict(rfmod, newdata = test,
                                    type="response"))
```

```
suppressMessages(library(pROC))
rocrf <- roc(testm$binary,testm$rf_pred)
## Setting levels: control = 0, case = 1
## Setting direction: controls < cases
AUCrf <- auc(rocrf)
rs <- rocrf[['rocs']]</pre>
```

Mixed model GLMM

```
binary random: AST
suppressMessages(library(ggpubr))
library(ggplot2)
agePlot=ggplot(train, aes(x=Age,y=binary))+geom_point()
ALPPlot=ggplot(train, aes(x=ALP,y=binary))+geom_point()
ALBPlot=ggplot(train, aes(x=ALB,y=binary))+geom_point()
# regular logistic model
bmod <- glm(binary ~.-Category, binomial, data=, train)</pre>
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
#GLMM model random effect:Age
library(MASS)
##
## Attaching package: 'MASS'
## The following object is masked from 'package:dplyr':
##
##
       select
set.seed(4850)
suppressMessages(modpql1 <- glmmPQL(binary</pre>
~Sex+ALB+Age+ALT+ALT+BIL+CHE+CHOL+CREA+GGT+PROT,
                  data=train, random= ~ 1 AST, family=binomial))
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
suppressMessages(drop1(modpql1, test="Chi"))
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
```

```
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Single term deletions
##
## Model:
## binary ~ Sex + ALB + Age + ALT + ALT + BIL + CHE + CHOL + CREA +
       GGT + PROT
##
          Df AIC LRT Pr(>Chi)
## <none>
## Sex
           1
## ALB
           1
## Age
           1
## ALT
           1
## BIL
           1
## CHE
## CHOL
           1
## CREA
## GGT
           1
## PROT
           1
testm1 <- mutate(test,</pre>
                glmm_pred = predict(modpql1, newdata = test,
                                    type="response"))
suppressMessages(library(pROC))
roc1 <- roc(testm1$binary,testm1$glmm pred)</pre>
## Setting levels: control = 0, case = 1
## Setting direction: controls < cases
AUC <- auc(roc1)
rs <- roc1[['rocs']]
coords(roc1, "best", ret=c("threshold",
"specificity", "sensitivity", "accuracy", "precision"))
                threshold specificity sensitivity accuracy precision
## threshold 8.117251e-06 0.952381 0.6666667 0.9444444 0.2857143
```

Random effect:CHE

```
#GLMM model random effect:CHE
library(MASS)
set.seed(4850)
suppressMessages(modpql2 <- glmmPQL(binary</pre>
~Sex+ALB+Age+ALT+ALT+BIL+AST+CHOL+CREA+GGT+PROT,
                  data=train, random= ~ 1 CHE, family=binomial))
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
suppressMessages(drop1(modpql2, test="Chi"))
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Single term deletions
##
## Model:
## binary ~ Sex + ALB + Age + ALT + ALT + BIL + AST + CHOL + CREA +
##
       GGT + PROT
          Df AIC LRT Pr(>Chi)
##
## <none>
## Sex
## ALB
## Age
           1
## ALT
## BIL
           1
## AST
           1
## CHOL
           1
## CREA
           1
## GGT
           1
## PROT
testm2 <- mutate(test,</pre>
                glmm pred = predict(modpq12, newdata = test,
```

Random effect:Sex

```
#GLMM model random effect:Sex
library(MASS)
set.seed(4850)
suppressMessages(modpql3 <- glmmPQL(binary</pre>
~CHE+ALB+Age+ALT+ALT+BIL+AST+CHOL+CREA+GGT+PROT,
                  data=train, random= ~ 1 Sex, family=binomial))
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
suppressMessages(drop1(modpql3, test="Chi"))
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Warning: glm.fit: fitted probabilities numerically 0 or 1 occurred
## Single term deletions
##
## Model:
```

```
## binary ~ CHE + ALB + Age + ALT + ALT + BIL + AST + CHOL + CREA +
       GGT + PROT
##
          Df AIC LRT Pr(>Chi)
##
## <none>
## CHE
## ALB
## Age
## ALT
## BIL
           1
## AST
## CHOL
## CREA
## GGT
           1
## PROT
           1
testm3 <- mutate(test,</pre>
                glmm pred = predict(modpql3, newdata = test,
                                    type="response"))
suppressMessages(library(pROC))
roc3 <- roc(testm3$binary,testm3$glmm pred)</pre>
## Setting levels: control = 0, case = 1
## Setting direction: controls < cases
coords(roc3,"best",ret=c("threshold",
"specificity", "sensitivity", "accuracy", "precision"))
##
             threshold specificity sensitivity accuracy precision
## threshold 0.4924259 0.9937888 0.9375 0.9887006 0.9375
```

Deep learning

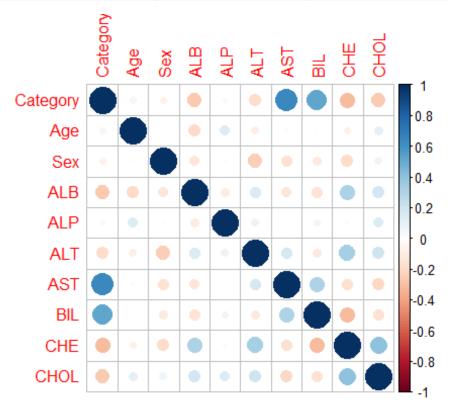
```
binary
```

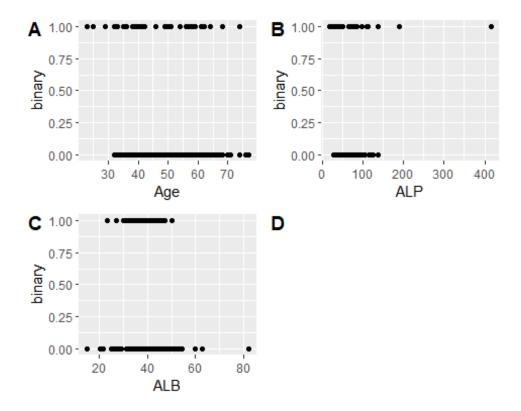
```
library(dplyr)
library(keras)
library(tensorflow)
library(tfdatasets)

set.seed(4850)
# Store the overall correlation in `M`
M <- cor(train[,1:10])

# Plot the correlation plot with `M`
library(corrplot)</pre>
```

corrplot 0.84 loaded predictorsCorr=corrplot(M, method="circle")





```
#normalize
spec <- feature spec(train, binary ~ .-Category ) %>%
  step_numeric_column(all_numeric(), normalizer_fn = scaler_standard()) %>%
  fit()
## Warning in normalizePath(path.expand(path), winslash, mustWork):
path[1]="C:
## \Users\Admin\.conda\envs\test-env/python.exe": The system cannot find the
file
## specified
## Warning in normalizePath(path.expand(path), winslash, mustWork):
path[1]="C:
## \Users\Admin\.conda\envs\test-env/python.exe": The system cannot find the
file
## specified
spec
## -- Feature Spec ------
## A feature_spec with 12 steps.
## Fitted: TRUE
## -- Steps -----
## The feature_spec has 1 dense features.
## StepNumericColumn: Age, Sex, ALB, ALP, ALT, AST, BIL, CHE, CHOL, CREA,
```

```
GGT, PROT
## -- Dense features -----
layer <- layer dense features(</pre>
  feature_columns = dense_features(spec),
  dtype = tf$float32
)
suppressMessages(layer(train))
## tf.Tensor(
## [[ 0.653347
               -0.6670947 -0.568868 ... -0.4312038
                                                          0.7929118
##
   -0.76765865]
## [-0.44780117 -0.22397378 -0.13247223 ... -0.32766175 -0.35917082
##
    -0.76765865]
## [-0.05207561 2.561358 0.41447717 ... 1.2864846
                                                          0.079717
      1.2995006 ]
##
## ...
## [-0.9811694 -0.68571323 -1.2147336 ... 0.36570016 0.55518115
##
   -0.76765865]
## [-1.1188128
                  0.05530416 -0.1441094 ... -0.14276527 -2.059866
##
   1.2995006 ]
## [ 0.08556774 -0.71922666 0.8683287 ... 0.6338001 -0.03000497
    -0.76765865]], shape=(412, 12), dtype=float32)
input <- layer input from dataset(train[,2:13])</pre>
output <- input %>%
  layer_dense_features(dense_features(spec)) %>%
  layer_dense(units = 64, activation = "relu") %>%
  layer dense(units = 64, activation = "relu") %>%
  layer dense(units = 1)
dpmod <- keras_model(input, output)</pre>
dpmod %>%
  compile(
    loss = "mse",
    optimizer = optimizer rmsprop(),
    metrics = list("mean absolute error")
  )
build_model <- function() {</pre>
  input <- layer input from dataset(train[,2:13])</pre>
```

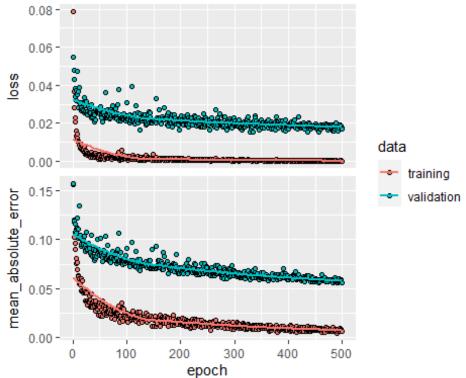
```
output <- input %>%
    layer dense features(dense features(spec)) %>%
    layer_dense(units = 64, activation = "relu") %>%
    layer_dense(units = 64, activation = "relu") %>%
    layer_dense(units = 1)
  dpmod <- keras_model(input, output)</pre>
  dpmod %>%
    compile(
      loss = "mse",
      optimizer = optimizer rmsprop(),
      metrics = list("mean_absolute_error")
    )
  dpmod
}
# Display training progress by printing a single dot for each completed
epoch.
print_dot_callback <- callback_lambda(</pre>
  on_epoch_end = function(epoch, logs) {
    if (epoch %% 80 == 0) cat("\n")
    cat(".")
  }
)
dpmod <- build_model()</pre>
history <- dpmod %>% fit(
  x = train[2:13],
  y = train$binary,
  epochs = 500,
  validation split = 0.2,
  verbose = 0,
  callbacks = list(print_dot_callback)
)
##
##
##
##
```

```
##
...
##
...
##
...

library(ggplot2)

plot(history)

## `geom_smooth()` using formula 'y ~ x'
```



```
# it will stop when no more improvement
early_stop <- callback_early_stopping(monitor = "val_loss", patience = 20)

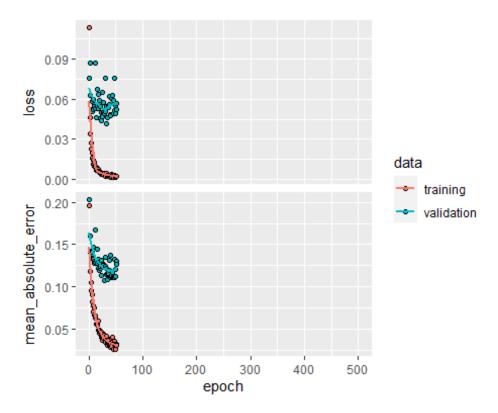
dpmod <- build_model()

history <- dpmod %>% fit(
    x = train[2:13],
    y = train$binary,
    epochs = 500,
    validation_split = 0.2,
```

```
verbose = 0,
  callbacks = list(early_stop)
)

plot(history)

## `geom_smooth()` using formula 'y ~ x'
```



```
AUCnn <- auc(rocnn)
rs <- rocnn[['rocs']]</pre>
suppressMessages(summary(dpmod))
## Model: "model_2"
##
                             Output Shape Param # Connected to
## Layer (type)
## ALB (InputLayer)
                             [(None,)]
                                               0
##
                             [(None,)]
## ALP (InputLayer)
                                               0
##
## ALT (InputLayer)
                             [(None,)]
                                               0
## AST (InputLayer)
                             [(None,)]
                                               0
##
## Age (InputLayer)
                             [(None,)]
                                               0
##
## BIL (InputLayer)
                             [(None,)]
                                               0
##
## CHE (InputLayer)
                             [(None,)]
                                               0
##
## CHOL (InputLayer)
                             [(None,)]
                                               0
##
## CREA (InputLayer)
                             [(None,)]
                                               0
##
## GGT (InputLayer)
                             [(None,)]
##
```

```
## PROT (InputLayer)
                              [(None,)]
                                                 0
##
## Sex (InputLayer)
                              [(None,)]
                                                 0
##
## dense_features_3 (DenseFe (None, 12)
                                                 0
                                                           ALB[0][0]
                                                           ALP[0][0]
##
##
                                                           ALT[0][0]
##
                                                           AST[0][0]
##
                                                           Age[0][0]
##
                                                           BIL[0][0]
##
                                                           CHE[0][0]
##
                                                           CHOL[0][0]
                                                           CREA[0][0]
##
##
                                                           GGT[0][0]
                                                           PROT[0][0]
##
##
                                                           Sex[0][0]
##
## dense_8 (Dense)
                              (None, 64)
                                                 832
dense_features_3[0][0]
##
## dense_7 (Dense)
                              (None, 64)
                                                 4160
                                                           dense_8[0][0]
##
## dense_6 (Dense)
                              (None, 1)
                                                 65
                                                           dense_7[0][0]
##
## Total params: 5,057
## Trainable params: 5,057
## Non-trainable params: 0
##
#RM, (AST ALT ALP) glmm, nn
coords(rocrf, "best", ret=c("threshold",
"specificity", "sensitivity", "accuracy", "precision")) ##the random forest
model
```

```
threshold specificity sensitivity accuracy precision
## threshold 0.2833333
                         0.9689441
                                             1 0.9717514 0.7619048
coords(roc1, "best", ret=c("threshold",
"specificity", "sensitivity", "accuracy", "precision"))# the AST GLMM model
                threshold specificity sensitivity accuracy precision
##
## threshold 8.117251e-06
                                        0.6666667 0.9444444 0.2857143
                             0.952381
coords(roc2, "best", ret=c("threshold",
"specificity", "sensitivity", "accuracy", "precision"))#the CHE GLMM model
##
             threshold specificity sensitivity accuracy precision
## threshold 0.3358878
                                     0.8571429 0.9863014
coords(roc3, "best", ret=c("threshold",
"specificity", "sensitivity", "accuracy", "precision"))# the Sex GLMM model
             threshold specificity sensitivity accuracy precision
## threshold 0.4924259
                         0.9937888
                                        0.9375 0.9887006
coords(rocnn, "best", ret=c("threshold",
"specificity", "sensitivity", "accuracy", "precision"))# the nn GLMM model
##
             threshold specificity sensitivity accuracy precision
## threshold 0.2531132
                         0.9813665
                                             1 0.9830508 0.8421053
#importance: impurity gini importance
c(importance(rfmod),importance(rfmod))
## [1] 3.610843e-01 2.775558e-18 4.248468e-01 3.345247e+00 7.598513e+00
## [6] 1.973172e+01 1.716077e+00 1.254098e+00 3.677537e-01 4.640634e-01
## [11] 2.672794e+00 6.762646e-15 3.610843e-01 2.775558e-18 4.248468e-01
## [16] 3.345247e+00 7.598513e+00 1.973172e+01 1.716077e+00 1.254098e+00
## [21] 3.677537e-01 4.640634e-01 2.672794e+00 6.762646e-15
log(importance(rfmod))
##
        IncNodePurity
## Age
          -1.0186438
## Sex
          -40.4256800
## ALB
          -0.8560265
## ALP
            1.2075404
## ALT
            2.0279526
## AST
            2.9822275
## BIL
           0.5400411
## CHE
           0.2264169
## CHOL
           -1.0003418
## CREA
          -0.7677341
           0.9831243
## GGT
## PROT
          -32.6273622
```

