

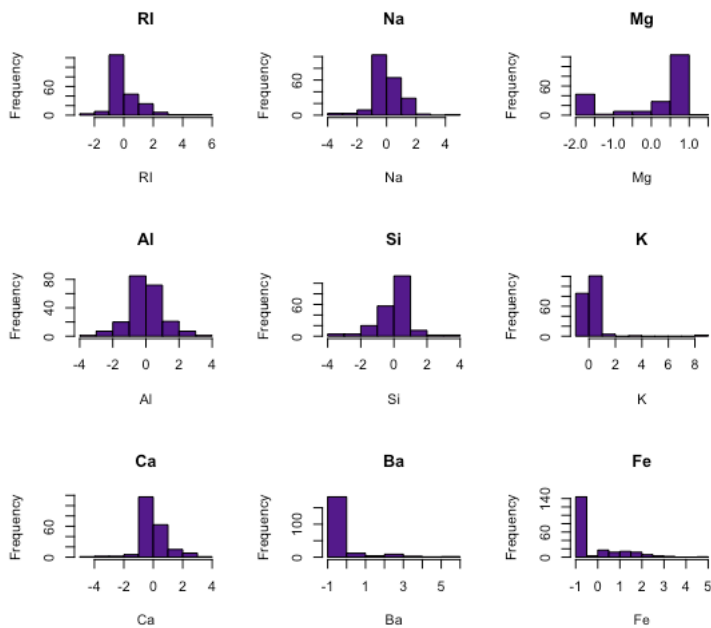
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SYST 568
Homework 1 – Preprocessing
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Question 3.1

- A. Using visualizations, explore the predictor variables to understand their distributions as well as the relationships between predictors.

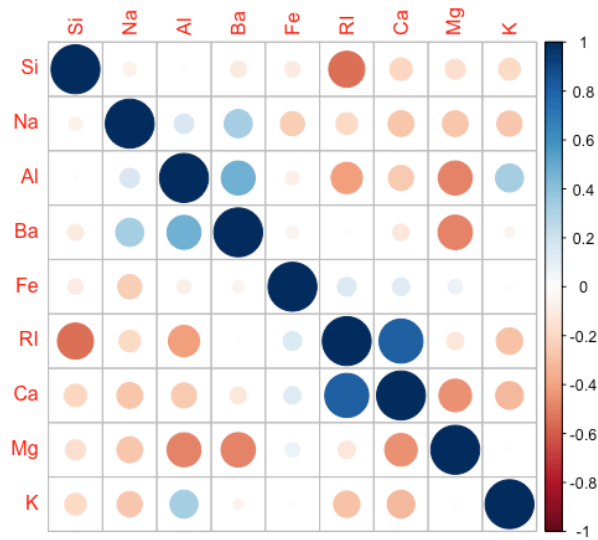
```
#install.packages("mlbench")
library(mlbench)
data(Glass)
colnames(Glass)
|
#subsetting dataset to remove 'type' column
elmns <- Glass[, 1:9]
par(mfrow = c(3,3))
for (i in 1:ncol(elmns)) {
  hist(elmns[,i], xlab = paste(names(elmns)[i]),
       main = paste(names(elmns)[i]), col = "purple4")
}
```

Histograms showing distributions of each predictor variable



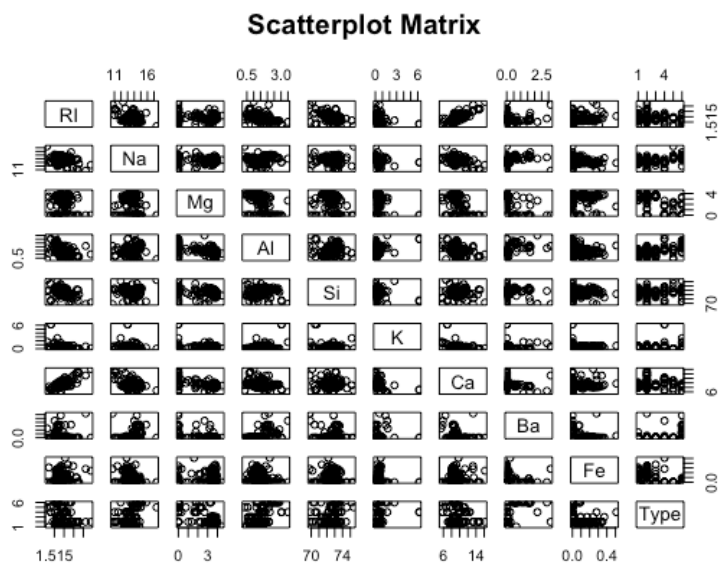
Correlation Plot of the predictors

```
par(mfrow=c(1,1))
library(corrplot)
corrplot( cor( Glass[, -10] ), order="hclust")
```



Pairwise scatterplot

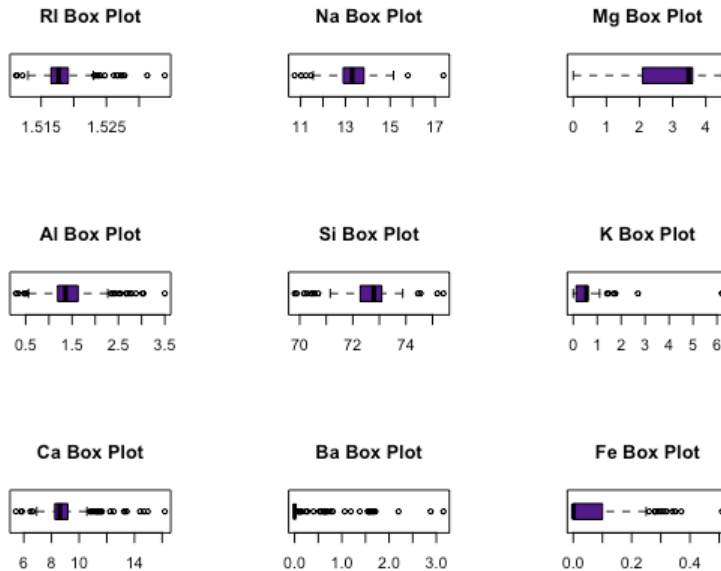
```
##relationship (Correlation) between predictors
par(mfrow=c(1,1))
pairs(Glass, main = "Scatterplot Matrix")
```



The histograms above indicate that RI, Na, Al, and Si are the only predictors that resemble a normal distribution. The other predictors are asymmetrical. According to the correlation plot, we see that RI possesses a negative correlation with Si and a positive correlation with Ca. This can also be clearly seen by the scatterplot matrix. The correlation plot would suggest that Mg – Al and Mg – Ba are both negatively correlated, but a clear correlation is not as apparent in the scatterplot matrix.

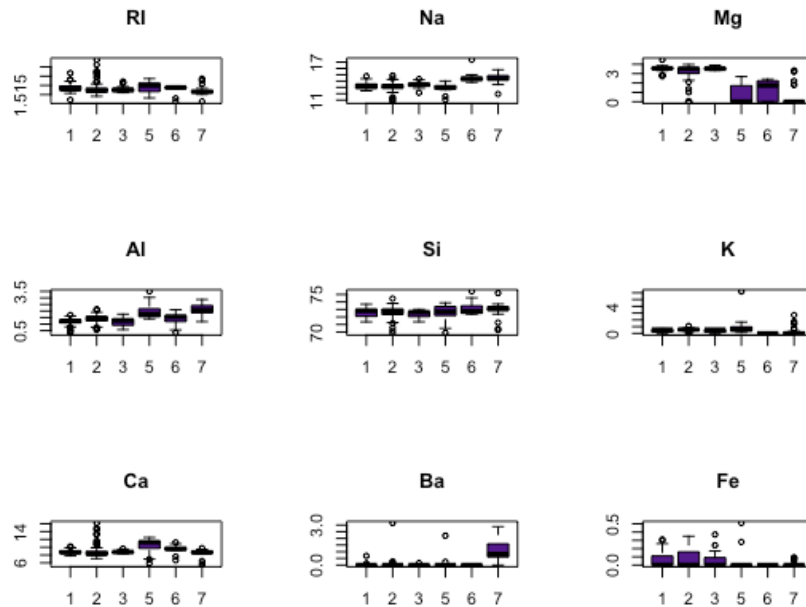
- B. Do there appear to be any outliers in the data? Are any predictors skewed?
Box Plot Visualization to display outliers

```
#3.1.b
#boxplots representing each predictor variable.
#for loop works very nicely here to iterate the whole dataset
full_elmns <- Glass[,1:10]
par(mfrow = c(3,3))
for (i in 1:ncol(full_elmns)) {
  boxplot(full_elmns[,i], main = paste(names(full_elmns[i]), "Box Plot"),
          horizontal = T, col = "purple4")
}
```



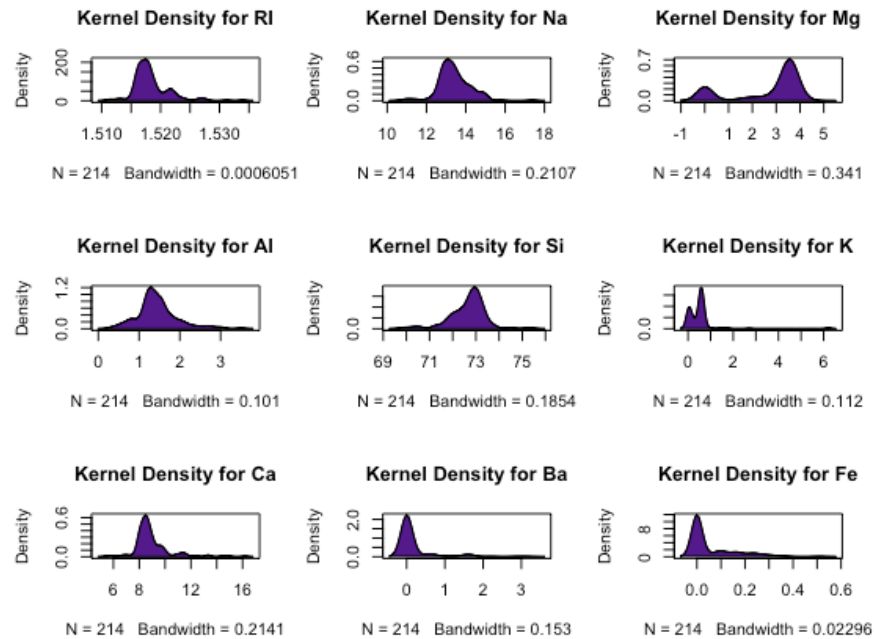
Box Plot Visualization that shows predictor dependency on Glass type

```
#boxplots to visualize distribution
#this shows how each predictor does per Type
par(mfrow = c(3,3))
for (i in 1:ncol(elmns)) {
  boxplot(full_elmns[,i] ~ full_elmns[,10], main = paste(names(full_elmns[i]), "Box Plot"),
    horizontal = F, col = "purple4")
}
```



Kernel Density Visualization to determine Skewedness

```
#boxplots to visualize distribution
#this shows how each predictor does per Type
par(mfrow = c(3,3))
for (i in 1:ncol(elmns)) {
  boxplot(full_elmns[,i] ~ full_elmns[,10], main = paste(names(full_elmns[i]), "Box Plot"),
    horizontal = F, col = "purple4")
}
```

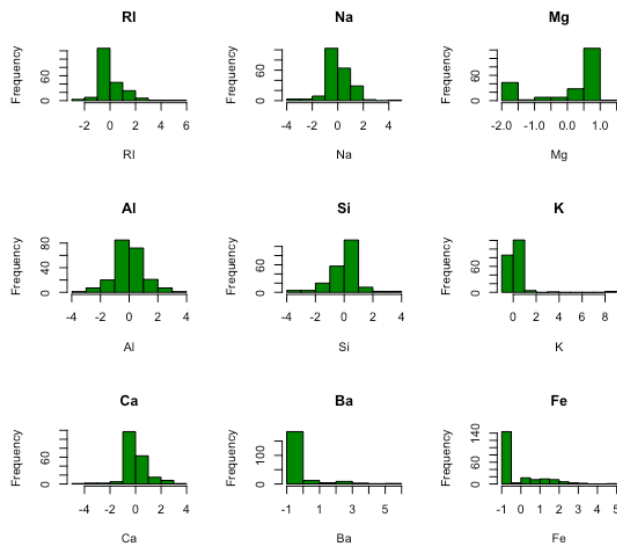


We see outliers in each predictor except for Mg, which doesn't have any values outside of either whisker. All the rest have some to many outliers, especially predictors RI, K, CA, and Ba. According to the density graphs, we see that Mg is left skewed, while Fe, Ba are hugely right skewed.

- C. Are there any relevant transformations of one or more predictors that might improve the classification model?

```
#install.packages("caret")
#install.packages("car")
library(car)
library(caret)

gls <- Glass[,1:9]
par(mfrow = c(3,3))
for (i in 1:ncol(glassTrans)) {
  hist(glassTrans[,i], xlab = paste(names(glassTrans[i])),
       main = paste(names(glassTrans[i])), col = "green4")
}
```



We use BoxCox to transform the distributions of each variable in order to remove skewness. In addition to this visual, I ran the skewness values of the original dataset as well as the transformed dataset to compare and see which skewness levels were smaller.

```
skew.g1<-sapply(Glass[,1:9], function(x){round(skewness(x),4)})
skew.g2<-sapply(glassTrans[,1:9],function(x){round(skewness(x),4)})
skew.g1
skew.g2
```

```
> skew.g1
      RI      Na      Mg      Al      Si      K      Ca      Ba      Fe
1.6027 0.4478 -1.1365 0.8946 -0.7202 6.4601 2.0184 3.3687 1.7298
> skew.g2
      RI      Na      Mg      Al      Si      K      Ca      Ba      Fe
1.5657 0.0338 -1.1365 0.0911 -0.6509 6.4601 -0.1940 3.3687 1.7298
> |
```

From the results, there are a few predictors that would benefit from a transformation. RI, NA, Al, Si, and Ca show less skewness in the transformed dataset than in the original Dataset. Of those predictors, transforming Ca, Al, and Na present the most beneficial change as the difference in their skewness values is 1.824, 0.8035, and 0.414 respectively.

Question 3.2

- A. Investigate the frequency distributions for the categorical predictors. Are any of the distributions degenerate in the ways discussed earlier in this chapter?

In order to determine degenerate distributions, we have to find predictors that have zero variance, meaning that predictor only has one single value. There is a function called 'nearZeroVar,' which calculates this with ease.

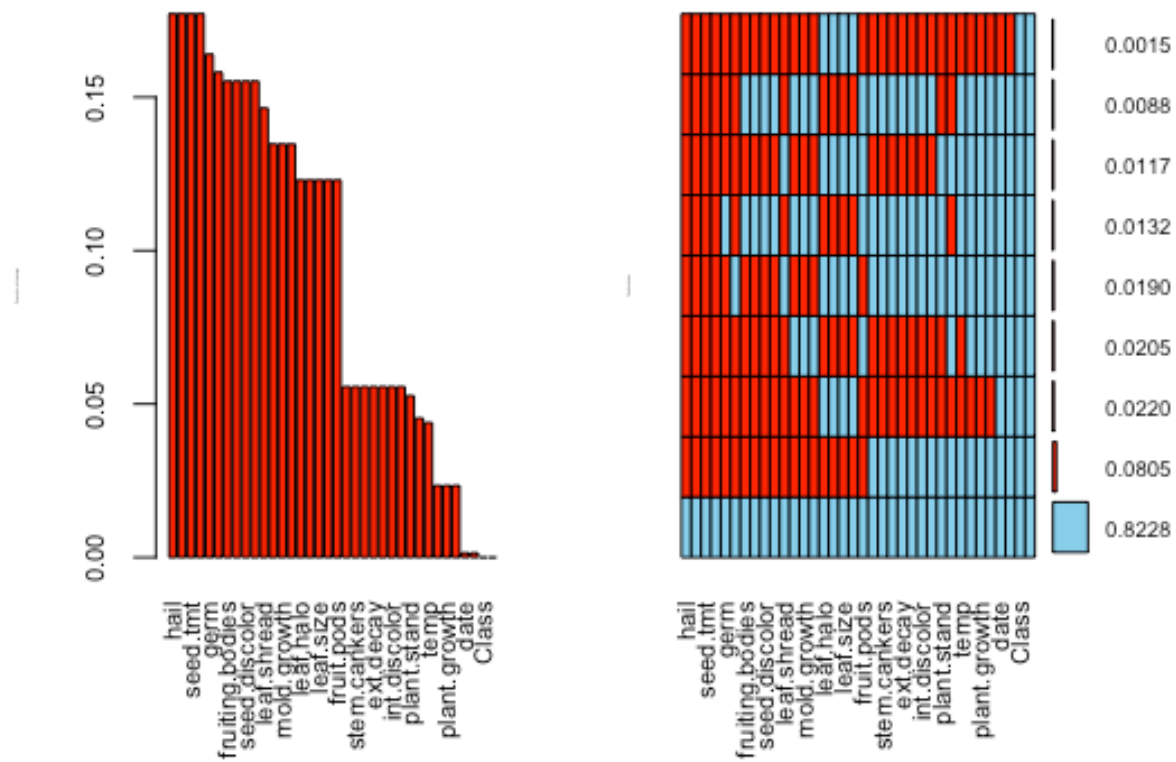
```
> #3.2.a
> nzv <- nearZeroVar(Soybean, saveMetrics= TRUE)
> nzv
```

	freqRatio	percentUnique	zeroVar	nzv
Class	1.010989	2.7818448	FALSE	FALSE
date	1.137405	1.0248902	FALSE	FALSE
plant.stand	1.208191	0.2928258	FALSE	FALSE
precip	4.098214	0.4392387	FALSE	FALSE
temp	1.879397	0.4392387	FALSE	FALSE
hail	3.425197	0.2928258	FALSE	FALSE
crop.hist	1.004587	0.5856515	FALSE	FALSE
area.dam	1.213904	0.5856515	FALSE	FALSE
sever	1.651282	0.4392387	FALSE	FALSE
seed.tmt	1.373874	0.4392387	FALSE	FALSE
germ	1.103627	0.4392387	FALSE	FALSE
plant.growth	1.951327	0.2928258	FALSE	FALSE
leaves	7.870130	0.2928258	FALSE	FALSE
leaf.halo	1.547511	0.4392387	FALSE	FALSE
leaf.marg	1.615385	0.4392387	FALSE	FALSE
leaf.size	1.479638	0.4392387	FALSE	FALSE
leaf.shread	5.072917	0.2928258	FALSE	FALSE
leaf.malf	12.311111	0.2928258	FALSE	FALSE
leaf.mild	26.750000	0.4392387	FALSE	TRUE
stem	1.253378	0.2928258	FALSE	FALSE
lodging	12.380952	0.2928258	FALSE	FALSE
stem.cankers	1.984293	0.5856515	FALSE	FALSE
canker.lesion	1.807910	0.5856515	FALSE	FALSE
fruiting.bodies	4.548077	0.2928258	FALSE	FALSE
ext.decay	3.681481	0.4392387	FALSE	FALSE
mycelium	106.500000	0.2928258	FALSE	TRUE
int.discolor	13.204545	0.4392387	FALSE	FALSE
sclerotia	31.250000	0.2928258	FALSE	TRUE
fruit.pods	3.130769	0.5856515	FALSE	FALSE
fruit.spots	3.450000	0.5856515	FALSE	FALSE
seed	4.139130	0.2928258	FALSE	FALSE
mold.growth	7.820896	0.2928258	FALSE	FALSE
seed.discolor	8.015625	0.2928258	FALSE	FALSE
seed.size	9.016949	0.2928258	FALSE	FALSE
shriveling	14.184211	0.2928258	FALSE	FALSE
roots	6.406977	0.4392387	FALSE	FALSE

The findings results in No zero variance predictors, but there are three predictors that have very low variance (or near-zero variance): **leaf.mild**, **mycelium**, and **sclerotia**.

- B. Roughly 18% of the data are missing. Are there particular predictors that are more likely to be missing? Is the pattern of missing data related to the classes?

```
library(VIM)
##the aggr function allows me to find and sort the frequency of missing values per
##categorical predictor. I am then able to display the proportion of missing values
##in a separate visual
aggr(Soybean, delimiter = NULL, prop = c(T,T), bars = TRUE, numbers = TRUE, plot = TRUE,
      sortVars = T, sortCombs = T, labels = names(Soybean), cex.lab = 0.1, cex.axis = 0.7, cex.numbers = 0.6)
```



Variables sorted by number of missings:

Variable	Count
hail	0.177159590
sever	0.177159590
seed.tmt	0.177159590
lodging	0.177159590
germ	0.163982430
leaf.mild	0.158125915
fruiting.bodies	0.155197657
fruit.spots	0.155197657
seed.discolor	0.155197657
shriveling	0.155197657
leaf.shread	0.146412884
seed	0.134699854
mold.growth	0.134699854
seed.size	0.134699854
leaf.halo	0.122986823
leaf.marg	0.122986823
leaf.size	0.122986823
leaf.malf	0.122986823
fruit.pods	0.122986823
precip	0.055636896
stem.cankers	0.055636896
canker.lesion	0.055636896
ext.decay	0.055636896
mycelium	0.055636896
int.discolor	0.055636896
sclerotia	0.055636896
plant.stand	0.052708638
roots	0.045387994
temp	0.043923865
crop.hist	0.023426061
plant.growth	0.023426061
stem	0.023426061
date	0.001464129
area.dam	0.001464129
Class	0.000000000
leaves	0.000000000

Using the VIM package in R for handling missing data points, I am able to use the `aggr` function to bin the frequencies of missing data points per categorical variable. The first visual presents a sorted bar chart of the proportion of missing data per categorical variable; it gives the percentage of missing data points per categorical variable. The second visual includes the

combined present and missing percentages for each categorical variable, and it shows the total amount of available data (82%, which is correct since there are 18% of missing data). We can see that hail, server, and seed.tmt are the top three categorical variables that present the most amount of missing data.

In order to find potential patterns of missing data by class type, we have to use a data manipulation package like dplyr (pronounced dee-plyer). This package allows us to customize and manipulate the return information from a dataset to meet our requirements. It is basically SQL for R.

```
Soybean %>%  
  mutate(Total = n()) %>% ##finding the total number to be used later  
  filter(!complete.cases()) %>% ##here, I'm finding all na's or missing vals  
  group_by(Class) %>% #grouping my results by class  
  mutate(Missing = n(), Proportion=Missing/Total) %>% ##new columns to create  
  select(Class, Missing, Proportion)%>% #columns to select  
  unique() ##distinct values  
  
# A tibble: 5 x 3  
# Groups:   Class [5]  
  Class                Missing Proportion  
  <fct>                <int>      <dbl>  
1 phytophthora-rot      68      0.0996  
2 diaporthe-pod-&-stem-blight 15      0.0220  
3 cyst-nematode         14      0.0205  
4 2-4-d-injury          16      0.0234  
5 herbicide-injury       8      0.0117  
> |
```

From the data above, we see that the phytophthora-rot class shows up missing 68 times, which is almost 10% of the missing data. The remaining ~18% of missing data shows up in classes 2-5 above. We conclude that out of the 19 classes, the 5 classes above comprise 100% of the missing data in this dataset.