

# Propensity Score Weighting and Generalized Boosted Regression in R

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# 1 Load Packages

Stata DTA files can be imported directly into R using the **haven** and **sjlabelled** packages.

Propensity score weighting can be accomplished with base R. However, we need the **lmtest** and **sandwich** packages to estimate clustered covariance matrices in this example. Using these packages, we can obtain estimates and standard errors that are identical to Stata's **regress** program.

Generalized boosted regression requires the **gbm** package.

Finally, the **tidyverse** package can be optionally loaded for its convenient data manipulation and plotting functions.

```
library(tidyverse)
library(haven)
library(sjlabelled)
library(lmtest)
library(sandwich)
library(gbm)
```

## 2 Propensity Score Weighting

### 2.1 Load Data with Propensity Scores

```
d <- read_dta("data/chpt5_2_original.dta") %>%
  haven::zap_formats() %>%
  sjlabelled::remove_all_labels() %>%
  as_tibble()
head(d$ps)
```

```
## [1] 0.3096053 0.3064355 0.1935465 0.3243693 0.3178224 0.2095492
```

### 2.2 Estimate ATE and ATT Weights

Here we calculate separate weights for estimating the average treatment effect (ATE) and the average treatment effect for the treated (ATT).

For ATE, the weight estimates are calculated as follows for the treatment group:

$$\omega = \frac{1}{\hat{e}(x)}$$

And for the control group:

$$\omega = \frac{1}{1 - \hat{e}(x)}$$

For ATT, the weight is 1 for a treated case. The weight for a comparison case is:

$$\omega = \frac{\hat{e}(x)}{1 - \hat{e}(x)}$$

```
d.weights <- d %>%
  mutate(ate_w = ifelse(kuse == 0, 1/(1-ps), 1/ps),
         att_w = ifelse(kuse == 0, ps/(1-ps), 1))
```

## 2.3 Outcome Analysis

### 2.3.1 Weighted Regression with ATE Weights

After creating the weights, use the `weights` argument in `lm()` to run a weighted outcome analysis and `lmtest::coeftest()` to control for clustering effects.

This analysis showed that children who used Aid to Families With Dependent Children (AFDC) had an average letter-word identification score that was 5.16 points lower than children who never used AFDC,  $p < .01$ .

```
m3 <- lm(lwss97 ~ kuse + male + black + age97 + pcged97 + mratio96,
        data = d.weights, weights = ate_w)
lmtest::coeftest(m3, vcov. = vcovCL(m3, cluster = d.weights$pcg_id))
```

```
##
## t test of coefficients:
##
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  84.19992    4.83032  17.4315 < 2.2e-16 ***
## kuse         -5.16399    1.42438  -3.6254 0.0003031 ***
## male        -1.62201    1.09186  -1.4855 0.1377180
## black       -2.49898    1.34670  -1.8556 0.0638009 .
## age97         0.73868    0.18075   4.0867 4.727e-05 ***
## pcged97       0.99264    0.35596   2.7886 0.0053938 **
## mratio96      1.13856    0.32220   3.5337 0.0004286 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

### 2.3.2 Weighted Regression with ATT Weights

When considering only individuals assigned to the treatment condition, children who used AFDC had an average letter-word identification score that was 4.62 points lower than children who never used AFDC,  $p < .01$ .

```
m4 <- lm(lwss97 ~ kuse + male + black + age97 + pcged97 + mratio96,
        data = d.weights, weights = att_w)
lmtest::coeftest(m4, vcov. = vcovCL(m4, cluster = d.weights$pcg_id))
```

```
##
## t test of coefficients:
##
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)  85.29467    5.05331  16.8790 < 2.2e-16 ***
## kuse         -4.62058    1.41182  -3.2728 0.001102 **
## male        -1.58995    1.14705  -1.3861 0.166020
## black       -2.74605    1.41605  -1.9392 0.052756 .
## age97         0.61577    0.20001   3.0787 0.002136 **
## pcged97       0.92698    0.36718   2.5246 0.011738 *
## mratio96      1.26018    0.33556   3.7555 0.000183 ***
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

## 2.4 Check Imbalance

To assess balance before and after propensity score weighting, use logistic regression for dummy covariates and OLS regression for continuous covariates. Some examples are included below, and the full code can be

found in Section 7.3.1 of the PSA-R code.

In model c5 below, the treatment dummy variable is significant, meaning that there is no sufficient balance after the propensity score weighting.

To assess balance before propensity score weighting, remove the `weights` argument.

```
c1 <- glm(male ~ kuse, family = quasibinomial, data = d.weights, weights = ate_w)
lmtest::coeftest(c1, vcov. = vcovCL(c1, cluster = d.weights$pcg_id))
```

```
##
## z test of coefficients:
##
##           Estimate Std. Error z value Pr(>|z|)
## (Intercept)  0.142911   0.075538  1.8919   0.0585 .
## kuse        -0.060271   0.150143 -0.4014   0.6881
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
c2 <- glm(male ~ kuse, family = quasibinomial, data = d.weights, weights = att_w)
lmtest::coeftest(c2, vcov. = vcovCL(c2, cluster = d.weights$pcg_id))
```

```
##
## z test of coefficients:
##
##           Estimate Std. Error z value Pr(>|z|)
## (Intercept)  0.152522   0.077410  1.9703   0.0488 *
## kuse        -0.079497   0.147480 -0.5390   0.5899
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
c5 <- lm(age97 ~ kuse, weights = ate_w, data = d.weights)
lmtest::coeftest(c5, vcov. = vcovCL(c5, cluster = d.weights$pcg_id))
```

```
##
## t test of coefficients:
##
##           Estimate Std. Error t value Pr(>|t|)
## (Intercept)  6.51389    0.10900 59.7615 < 2.2e-16 ***
## kuse         0.61064    0.21883  2.7905  0.005362 **
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
c6 <- lm(age97 ~ kuse, weights = att_w, data = d.weights)
lmtest::coeftest(c6, vcov. = vcovCL(c6, cluster = d.weights$pcg_id))
```

```
##
## t test of coefficients:
##
##           Estimate Std. Error t value Pr(>|t|)
## (Intercept)  6.55501    0.11499 57.0057 < 2e-16 ***
## kuse         0.56178    0.21939  2.5606  0.01059 *
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

## 3 Estimate Propensity Scores Using Generalized Boosted Regression

### 3.1 Load Data and Sort

Generalized boosted regression (GBR) is an iterative method for creating propensity scores. Therefore, to create reproducible results, we need to use the `set.seed()` function in R.

After importing the data, missing data is deleted listwise, and the data is sorted in a random order. Note the use of the `set.seed()` function to create the same set of random numbers from the uniform distribution.

According to the `gbm` package vignette, if the data is sorted in a systematic way, then the data should be shuffled before running `gbm`.

```
set.seed(1000)
d2 <- read_dta("data/g3aca1.dta") %>%
  haven::zap_formats() %>%
  sjlabelled::remove_all_labels() %>%
  as_tibble() %>%
  select(intbl, ageyc, fmale, blk, whit, hisp, pcedu, ipovl, pceft, fthr,
         dicsagg2, dicsint2, dccereg2, dccscom2, dccpros2, draggr2) %>%
  drop_na() %>%
  add_column(runif = runif(nrow(.))) %>%
  arrange(runif)
```

### 3.2 Generate Propensity Scores

### 3.3 Fit Generalized Boosted Regression Model

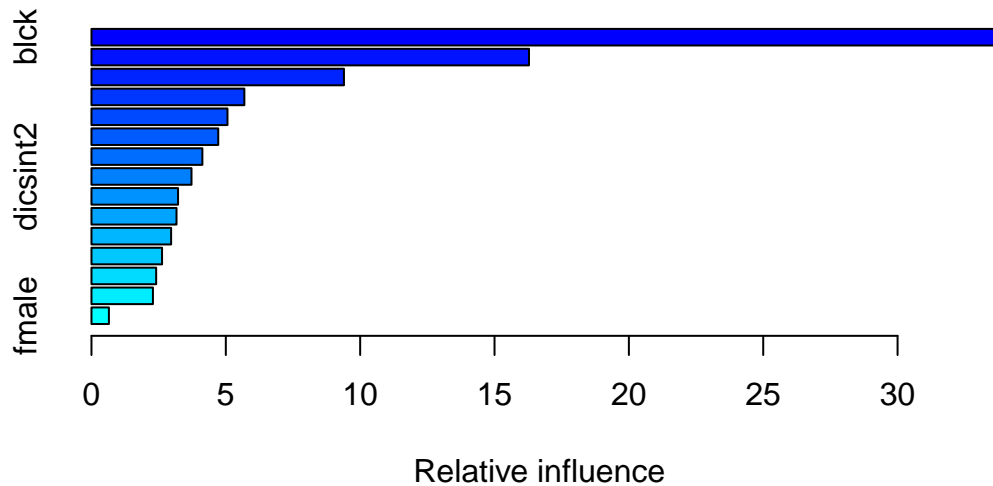
The `gbm::gbm()` function has many arguments that can be fine-tuned. See `?gbm` for a detailed description of each argument.

A summary of the fitted model provides us with *relative influence*, which is the percentage of log likelihood explained by each input variable.

```
# Correction: Remove the column of random numbers from the formula
f5 <- as.formula(paste("intbl ~ ", paste(names(select(d2, -intbl, -runif)),
                                         collapse = " + ")))

set.seed(1000)
m5 <- gbm::gbm(formula = f5,
               data = d2,
               distribution = "bernoulli",
               n.trees = 1000, # number of trees to fit
               train.fraction = 0.8, # a random 80% subsample for estimation
               interaction.depth = 4, # allow all four-way interactions
               shrinkage = 0.0005) # small shrinkage to ensure smooth fit

summary(m5)
```



```
##          var      rel.inf
## blk      blk 33.6768868
## ageyc    ageyc 16.2834103
## draggr2  draggr2 9.3962151
## whit     whit  5.6895119
## ipovl    ipovl  5.0614691
## pemft    pemft  4.7179839
## pcedu     pcedu  4.1280063
## dicsint2 dicsint2 3.7223185
## dicsagg2 dicsagg2 3.2228567
## dccscom2 dccscom2 3.1648839
## dccereg2 dccereg2 2.9647178
## dccpros2 dccpros2 2.6269129
## hisp     hisp   2.4079157
## fthr     fthr   2.2874508
## fmale    fmale  0.6494603
```

### 3.4 Estimate Propensity Scores

After fitting the model, propensity scores can be generated by using the `predict.gbm()` function.

```
psb <- gbm::predict.gbm(m5, data = d2, type = "response") # Create ps
```

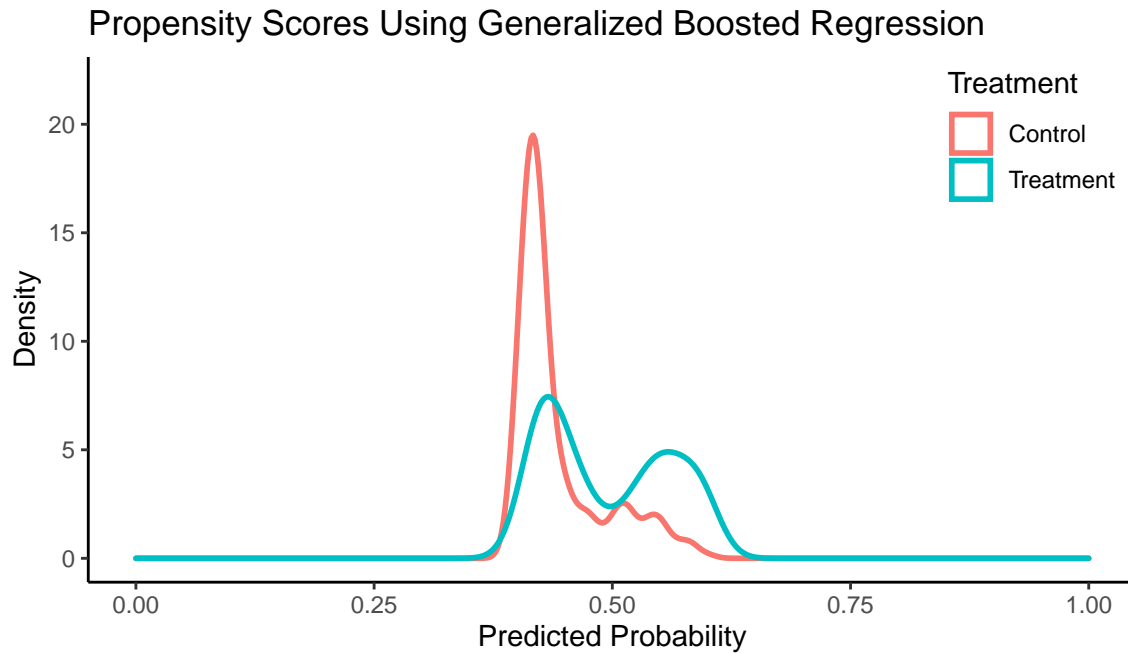
```
## Using 1000 trees...
```

```
head(psb)
```

```
## [1] 0.4441531 0.4405950 0.5288983 0.5958331 0.3985049 0.5562076
```

### 3.5 Plot Propensity Score Distributions

```
d2 %>%
  mutate(psb = psb,
         intbl = factor(intbl, labels = c("Control", "Treatment"))) %>%
  ggplot(aes(x = psb, color = intbl)) + theme_classic() +
  geom_density(size = 1) + xlim(0, 1) + ylim(0, 22) +
  labs(x = "Predicted Probability", y = "Density",
       title = "Propensity Scores Using Generalized Boosted Regression",
       color = "Treatment") +
  theme(legend.position = c(0.9, 0.85))
```



### 3.6 Summary Statistics of Propensity Scores

```
summary(psb)
```

##	Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
##	0.3913	0.4189	0.4383	0.4674	0.5202	0.6082

## 4 Bonus: Using the WeightIt Package

### 4.1 Estimate ATE and ATT Weights

```
# Load Packages
library(WeightIt)
library(cobalt)

# Estimate ATE and ATT weights and Check with Previous Results
ate_w2 <- WeightIt::get_w_from_ps(ps = d$ps,
                                  treat = d$kuse,
                                  estimand = "ATE")

table(ate_w2 == d.weights$ate_w)

##
## TRUE
## 1003

att_w2 <- WeightIt::get_w_from_ps(ps = d$ps,
                                  treat = d$kuse,
                                  estimand = "ATT")

table(att_w2 == (d.weights$ate_w * d.weights$ps))

##
## TRUE
## 1003
```

### 4.2 Estimate Propensity Scores using Generalized Boosted Regression

```
# Estimate PS with GBM
set.seed(1000)
m5.alt <- WeightIt::weightit(
  formula = f5,
  data = d2,
  method = "gbm",
  estimand = "ATE",
  distribution = "bernoulli",
  stop.method = "es.mean",
  n.trees = 10000, # different
  nTrain = 0.8 * nrow(d2), # different
  interaction.depth = 4,
  shrinkage = 0.0005
)
```



### 4.3 Check Imbalance with the Cobalt Package

```
cobalt::love.plot(m5.alt, thresholds = c(m = .1), binary = "std", abs = T) +  
  labs(title = "Covariate Balance (ATE)")
```

