# Package 'ymse'

# November 17, 2019

Title Y	mse (	various,

**Version** 0.6.7

**Description** Supplies a number of more or less useful functions and methods suitable for, eg. estimating dice roll probabilities, calculate latin squares, perform binary search, adjust colours in HSV space, produce prime numbers, find maximum acf/pacf/ccf, convert floats to simple ratio, produce averaged shifted histogram drop variables from formulae using regex, flatten a nested list, compute the similarity between two character vectors, plot a simple loess smooth, and other assorted tasks.

**Depends** R (>= 3.5.0)

Imports stats, utils, graphics, grDevices

**License** GPL (>= 2)

**Encoding** UTF-8

LazyData true

RoxygenNote 7.0.0

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acf\_max

Maximum ACF, PACF and CCF

### **Description**

Find lag that maximizes correlation

```
acf_max(
  х,
  . . . ,
  plot = FALSE,
  show = plot,
  ci = 0.95,
  ma.ci = TRUE,
  max.type = c("pos", "neg", "abs"),
  most.signif = FALSE
)
pacf_max(
  х,
  . . . .
  plot = FALSE,
  show = plot,
  ci = 0.95,
  max.type = c("pos", "neg", "abs")
)
ccf_max(
  х,
  у,
  plot = FALSE,
  show = plot,
  ci = 0.95,
  max.type = c("pos", "neg", "abs")
```

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### **Arguments**

x, y	univariate numeric vector or time series
	further arguments passed to acf, pacf, ccf
plot	logical; return a plot
show	indicate on the plot the maximum correlation
ci	confidence interval used, by default 95%
	should the confidence limits assume an MA input (TRUE, the default), or white noise as is default for plot.acf?
• •	what maximum should be returned, the positive (default), negative, or absolute maximum?
most.signif	should the most significant correlation be returned. Only applicable if ma.ci=TRUE

### **Examples**

```
x \leftarrow c(5, 5, 3, 6, 3, 6, 9, 6, 3, 1, 3, 2, 8, 9, 4, 3, 6, 6,
       6, 7, 5, 2, 5, 1, 5, 5, 0, 3, 7, 3, 6, 6, 2, 2, 6, 5)
y \leftarrow c(8, 9, 7, 5, 3, 5, 6, 9, 6, 3, 4, 5, 9, 7, 8, 5, 5, 7,
       4, 7, 7, 2, 5, 6, 5, 7, 5, 3, 5, 6, 7, 0, 5, 3, 8, 4)
acf_max(x, plot=TRUE, max.type="abs")
acf_max(x, max.type="neg")
acf_max(x, max.type="neg", most.signif=TRUE)
pacf_max(x, plot=TRUE)
pacf_max(x, max.type="abs")
ccf_max(x, y, plot=TRUE)
ccf_max(x, y, max.type="neg")
# Same plot
plot(acf(x, plot=FALSE), ci.type="ma")
acf_max(x, plot=TRUE)
acf_max(x, ci=0.99, plot=TRUE)
ccf_max(x, y, ci=0, max.type="pos", plot=TRUE)
```

addrows

Add rows to a data.frame

# Description

An "rbind for data.frames", sort of.

```
addrows(dtf, nrw, top = FALSE)
```

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#### **Arguments**

dtf	data.frame; original data.frame
nrw	data.frame; the new row(s) to be added
top	logical; should the new rows be added to the top or the bottom (default)?

#### **Details**

Can only bind two objects at a time, but will bind data.frames with non-matching column names and -classes. In such cases the original data.frame will serve as template.

#### **Examples**

```
dtf <- data.frame(A=letters[1:5],</pre>
                   B=1:5,
                   C=as.factor(5:1),
                   D=as.Date(0:4, origin="2000-01-01"),
                   stringsAsFactors=FALSE)
nrw <- data.frame(A=letters[1:5],</pre>
                   B=4:8,
                   C=5:1,
                   D=as.Date(5:1, origin="1990-01-01"),
                   stringsAsFactors=FALSE)
str(dtf)
dtf.a <- addrows(dtf, nrw, top=FALSE)</pre>
str(dtf.a)
# adding a single row with little concern for data types and column names
b <- type.convert(beaver1[80:90,])</pre>
b$activ <- as.logical(b$activ)</pre>
addrows(b, data.frame(350, 1200, 37.02, 1))
```

adjustcolorHSV

Adjust Colors in One or More Directions Conveniently.

### **Description**

Adjust or modify a vector of colors by "turning knobs" on one or more coordinates in  $(h,s,v,\alpha)$  space, typically by up or down scaling them.

```
adjustcolorHSV(
   col,
   alpha.f = 1,
   h.f = 1,
   s.f = 1,
   v.f = 1,
   offset = c(0, 0, 0, 0),
   transform = diag(c(h.f, s.f, v.f, alpha.f)),
```

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```
h = NULL,
s = NULL,
v = NULL,
alpha = NULL)
```

#### **Arguments**

vector of colors, in any format that col2rgb() accepts

alpha.f, h.f, s.f, v.f

factors scaling the opacity, hue, saturation and value of col

offset

a length 4 numeric vector specifying the linear offset applied to the hue, saturation, value and alpha values

transform

a 4x4 diagonal matrix specifying the scaling applied to the hue, saturation, value and alpha values

h, s, v, alpha fixed vlues for hue, saturation, value and alpha. Overrides any corresponding scaling factor or offset

#### **Details**

Essentially an HSV version of the RGB-based adjustcolor. One important distinction is that the h.f value wraps around to fit the [0, 1] range, rather than simply "clamping" it between 0 and 1.

#### Value

A character vector the same length as col containing color data in standard hexadeximal RGBA format.

#### **Examples**

```
# Halve the saturation and value of the default palette colours
plot(2:8, cex=5, lwd=4, pch=21, bg=2:8,
    col=adjustcolorHSV(2:8, s.f=0.5, v.f=0.6))

# Offset the hue of the default palette colours by 0.5, inverting the colours
plot(2:8, cex=5, lwd=4, pch=21, bg=2:8,
    col=adjustcolorHSV(2:8, offset=c(0.5, 0, 0, 0)))
```

ahist

Average shifted histogram

### Description

Create a smoothed histogram by averaging several histograms shifted by fractions of a bin-width

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#### Usage

```
ahist(
    x,
    n.breaks = nclass.FD(x),
    n.shifts = 3,
    type = c("histogram", "polygon", "line", "table"),
    freq = FALSE,
    plot = TRUE,
    add = FALSE,
    ...
)
```

### **Arguments**

```
a vector of values for which the histogram is desired

n.breaks an integer giving the number of bins to be used

n.shifts an integer giving the number of shifts to be performed

type if plot=TRUE, the type of plot to be used

freq should frequency counts be used, or density (default)

plot logical; if TRUE (default), a graphical output will be returned

add logical; if TRUE the plot will be added to the current plot

further graphical parameters to ymse::plot.histogram, polygon, or lines
```

#### Value

an object of class "histogram"

# **Examples**

```
set.seed(1)
n <- 6

x <- sample(sample(0:20, 8), 6*n, replace=TRUE) + rnorm(6*n, -8, 0.5)
x <- c(x, rgamma(5*n, 3, 0.5), rnorm(4*n, 15, 2))
x <- round(x*5)/5

hist(x, freq=FALSE, breaks="FD", col="lightblue")
ahist(x, type="hist", border=2, col=7, freq=FALSE, lwd=2)
ahist(x, type="poly", border=2, col=7, freq=FALSE, lwd=2)
ahist(x, type="line", col=2, freq=FALSE, lwd=2)
ahist(x, type="table", col=2, freq=FALSE, lwd=2)
ahist(x, plot=FALSE)</pre>
```

align\_char

Align character strings

### **Description**

Align character strings

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#### Usage

```
align_char(x, pattern = ".", ..., lpad = " ", rpad = " ")
```

#### **Arguments**

x numeric or character vector

pattern pattern, passed to regexpr, whose match the character strings will be aligned by

... further arguments passed to regexpr

lpad, rpad character strings used for padding. Repeated to length

#### See Also

align\_num, for alignment more aimed at numeric strings

#### **Examples**

```
x <- c("Tom und Jerry", "Abbott og Costello", "Milo och Stich", "et alii")
cat(align_char(x, pat="[[:alpha:]]"), sep="\n")
cat(align_char(x, pat=" "), sep="\n")
cat(align_char(x, pat=" [A-Z]"), sep="\n")
cat(align_char(x, pat=" [a-z]"), sep="\n")
cat(align_char(x, pat="t", ignore.case=TRUE), sep="\n")
cat(align_char(x, pat="x"), sep="\n")</pre>
```

align\_num

Align numbers

### **Description**

Align numbers for neat vertical printing

#### Usage

```
align_num(x, lpad = " ", rpad = "0", dec = ".", min.dec = 0, rm.dec = TRUE)
```

#### **Arguments**

x numeric or character vector

1pad, rpad character strings used for padding. Repeated to length
dec decimal seperator, or any other character to align by
min.dec pad with zeros to reach a minimum number of decimal points
rm.dec remove zeros at the end of whole numbers

### See Also

align\_char, for alignment more aimed at character strings

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#### **Examples**

```
x <- c(22100, 100, 1015, 13.018, 0.1, 0.01234)
cat(align_num(x), sep="\n") # Default
cat(format(x, scientific=FALSE), sep="\n")
cat(align_num(x, rpad=" "), sep="\n")
cat(align_num(x, rpad=" ", rm.dec=FALSE), sep="\n")
cat(align_num(x, rpad=" ", min.dec=1), sep="\n")
cat(align_num(x, rpad=" ", min.dec=2), sep="\n")
cat(align_num(x, rpad="\U00b7", min.dec=0), sep="\n")
cat(align_num(x, lpad="'", rpad="'", min.dec=0), sep="\n")
cat(align_num(c("1.000.000", "10.000.000", "1.000,85"), dec=","), sep="\n")
# corner cases
x <- c("100.", "1.2", ".1111")
cat(align_num(x, rpad=" ", rm.dec=TRUE), sep="\n")
cat(align_num(round(as.numeric(x)), rpad=" ", rm.dec=TRUE), sep="\n")
# matching on more than one character
# so far not much more advanced than this
# working on align_num2 more suited for character strings
s <- c("cataract", "hematology", "pancreatic")
cat(align_num(s, dec="a", rpad=" "), sep="\n")
cat(align_num(s, dec="at", rpad=" "), sep="\n")</pre>
a <- c("Tom and Jerry", "Milo and Stich", "Abbott and Costello")
cat(align_num(a, dec="and", rpad=" "), sep="\n")
```

arfilter

AR filter

### **Description**

Filter a time series using AR coefficients

# Usage

```
arfilter(x, mod, x.mean = mod$x.mean, init = "focb")
```

#### **Arguments**

x a time series mod an AR model

x.mean the mean used. By default the mean of the original model. Set to zero for no

demeaning

init how the initial values should be chosen. First observation carried backwards

(default), mean of the first values, or the first values in reverse.

#### See Also

armodel

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### **Examples**

```
set.seed(1)
arap <- ar(AirPassengers)
spec.ar(arap)
spec.pgram(arfilter(rnorm(10000), arap), span=21, na.action=na.omit)
arm <- armodel(c(1.3, -0.4))
spec.ar(arm)
plot(x <- rnorm(200), type="1")
lines(scale(arfilter(x, arm), center=FALSE), col="red", lwd=2)</pre>
```

arfit

AR model fit

### **Description**

Fit a specified AR model to a univariate time series

# Usage

```
arfit(x, mod, x.mean = mod$x.mean)
```

### **Arguments**

x a time seriesmod an AR model

x.mean the mean used. By default the mean of the original model. Set to zero for no

demeaning

#### See Also

```
armodel for examples
```

### **Examples**

```
 set.seed(1) \\ x \leftarrow runif(50) + sin(1:50/10) \\ plot(x); lines(arfilter(x, armodel(c(1.5, -0.5, 0.5)), x.mean=mean(x)))
```

arimpulse

Impulse response of an AR model

# Description

Get and plot the impulse response of an AR model

```
arimpulse(mod, pulse = 1, n.ahead = 20, plot = TRUE, ...)
```

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### **Arguments**

mod an AR model

pulse numeric vector; the initial pulse. Magnitude is added to the model mean

n. ahead the length of the computed response

plot logical; sgould the result be plotted?

... further arguments to plot

#### See Also

armodel for examples

armodel

Create an AR model object

### **Description**

Specify the characteristics of an AR model

# Usage

```
armodel(
  coefs,
  mean = 0,
  intercept = 0,
  var.pred = 1,
  frequency = 1,
  x.name = "Synthetic AR model"
)
```

# Arguments

coefs a vector of model coefficients

mean the mean of the process

intercept the intercept in the model

var.pred the portion of the variance not explained by this model

frequency the sampling frequency of the process

x.name name of the series

```
arimpulse
```

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#### **Examples**

```
# short decay
ar.mod <- armodel(c(0.5))
arimpulse(ar.mod, pulse=1)
# long decay
ar.mod <- armodel(c(0.8))
arimpulse(ar.mod, pulse=1)
# negative second coefficient reduce damping, signal returns to normal
# more quickly
ar.mod <- armodel(c(0.8, -0.1))
arimpulse(ar.mod, pulse=1)
# second coefficient reduce damping too much, overdamping, oscillations
ar.mod <- armodel(c(0.8, -0.5))
arimp <- arimpulse(ar.mod, pulse=1, n.ahead=40)$pred</pre>
polyroot(c(1, -ar.mod$ar)) # complex conjugate roots
acf(arimp) # period ~= 6?
phi1 <- ar.mod$ar[1]</pre>
phi2 <- ar.mod$ar[2]</pre>
f \leftarrow (1/(2*pi)) * acos((phi1*(phi2-1))/(4*phi2))
1/f # period = 6.78
sp <- spec.ar(ar.mod, plot=FALSE)</pre>
1/sp\$freq[which.max(sp\$spec)] # period = 6.79
# decaying oscillations
ar.mod1 <- armodel(c(0.8, -0.6, -0.5, 0.2, -0.2))
arimpulse(ar.mod1, n.ahead=100)
Mod(1/polyroot(c(1, -ar.mod1\$ar))) # barely inside the unit circle
# growing oscillations
ar.mod2 \leftarrow armodel(c(0.8, -0.7, -0.5, 0.2, -0.2))
arimpulse(ar.mod2, n.ahead=100)
Mod(1/polyroot(c(1, -ar.mod2\$ar))) # barely outside the unit circle
ar.mod3 <- armodel(c(1.8, -1.1, 0.2, -0.2, 0.2))
arimpulse(ar.mod3, n.ahead=100)
spec.ar(ar.mod3)
resid(arfit(rnorm(10), armodel(c(0.5, -0.1), frequency=2)))
```

as.array.list

Coerce a list to an array

#### **Description**

Coerce a list consisting of data.frames or matrices of equal size to a 3d array

```
## S3 method for class 'list'
as.array(x, ...)
```

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### **Arguments**

```
x a list of equal sized data.frames or matrices
... (not used)
```

### Value

A list of length l with elements of m rows and n columns wix result in an  $m \times n \times l$  array.

### **Examples**

aug\_median

Centre weighted mean

### **Description**

Offering a continuous link between median and arithmetic mean

### Usage

```
aug_median(x, p = 1, rank = FALSE, na.rm = FALSE)
```

### **Arguments**

X	numeric vector
р	positive numeric narrowness of the weight. 1 gives triangular weighting. Higher values gives narrower weights, approaching meadian, lower values gives broader weights, approaching arithmetic mean
rank	logical. Should should ranks or numeric values determine relative weights?
na.rm	logical. Should missing values be removed?

### **Details**

A weighted arithmetic mean is calculated over the input vector, where most weight is given to the median value(s), and monotonically less towards either extreme. Faloff depends on p, with small values resulting in a gentler falloff and less difference between minimum and maximum weights.

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#### **Examples**

```
x <- c(0, 8, 8, 8, 9)
aug_median(x)

# 0 and 9 are considered equidistant from 8
aug_median(x, rank=TRUE)

# Nearly a point weight placed at the median
aug_median(x, 100)
median(x)

# Nearly uniform weights
aug_median(x, 0.001)
mean(x)</pre>
```

bartlett

Maurice Stevenson Bartlett's car data

#### **Description**

This is an example data set Bartlett used for a lecture course on stochastic processes, Statistics Department, University College, London. The data represents the times, in seconds, when cars passed an observation point by a road.

Bartlett attributes the data to a Dr A. J. Miller who supplied them as a class example. According to Adery C. A. Hope the data was recorded on a rural Swedish road.

# Usage

bartlett

#### **Format**

A numeric vector representing time points in seconds

#### M. S. Bartlett's notes

Analyse the above data with a view to examining:

- i whether the times of passing constitute a Poisson process;
- ii if not, whether some form of "bunching" or "clustering" seems to be present.

Possible analyses include:

- **a** testing the homogeneity of the consecutive random time-intervals, by means of a partitioning of the degrees of freedom for the total (approximate)  $\chi^2$ ;
- **b** testing the homogeneity of counts in consecutive fixed time-intervals, choosing an appropriate interval, and partitioning the degrees of freedom corresponding to the total dispersion by means of an analysis of variance;
- c testing the correlation between the consecutive random time-intervals;
- **d** examining the overall distribution of counts in fixed time-intervals;

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e examining the overall distribution of the consecutive random time-intervals

You should undertake at least sufficient of these to answer the questions asked.

#### **Source**

```
The Spectral Analysis of Point Processes (p. 280), M. S. Bartlett, 1963
Also mentioned in:
Statistical Estimation of Density Functions (p. 252), M. S. Bartlett, 1963
A Simplified Monte Carlo Significance Test Procedure (p. 583), Adery C. A. Hope, 1968
```

### **Examples**

```
cpgram(diff(bartlett))
bartlett2 <- bartlett - bartlett[1]

x <- rep(0, tail(bartlett2, 1)*10)
x[bartlett2*10] <- 1

par(mfrow=c(2, 1), mar=c(2, 3, 1, 1))
plot(x, type="1", ann=FALSE)
lines(cumsum(x)/sum(x), col="red", lwd=2)

sp <- spectrum(x, main="", xlim=c(0, 0.1), ylim=c(1e-3, 0.04))
spec <- predict(loess(sp$spec[1:3000] ~ sp$freq[1:3000], span=0.15), se=TRUE)
lines(sp$freq[1:3000], spec$fit, col="red", lwd=2)
lines(sp$freq[1:3000], spec$fit - qt((0.99 + 1)/2, spec$df)*spec$se,
    lty=1, col="lightblue")
lines(sp$freq[1:3000], spec$fit + qt((0.99 + 1)/2, spec$df)*spec$se,
    lty=1, col="lightblue")</pre>
```

binsearch

Binary search

# Description

Find the position of a given value in a sorted array

# Usage

```
binsearch(val, arr, L = 1L, H = length(arr))
binclosest(val, arr, L = 1L, H = length(arr))
```

### **Arguments**

val	the value to search for
arr	a sorted array to make the search in
L	a lower bound
Н	an upper bound

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#### **Details**

While both val and arr can be either integer or double, the algorithm is limited by integer storage in how long the array can be. L and H can be used to limit the range of indices to be search within. binsearch will return either the index of the exact match, or the index just below if no exact match is found. This means that if val is less than the lowest value in arr (and L=1), a 0 will be returned, which can lead to issues as such an index does not exist in R. An array indexed by 0 will return a zero length object. binclosest will return the index of the closest match, and therefore a 1 in the situation where binsearch returns a 0. If there is a tie the lower index will be returned.

In either case, if there are duplicate matches, the lower index will be returned.

#### Value

A single integer representing an index on the input array.

#### **Examples**

```
binsearch(15, (1:9)*3.333)
binsearch(2, (1:9)*3.333)
binclosest(2, (1:9)*3.333)
binsearch(18, seq_len(2e9))
## Not run:
binsearch(18, seq_len(3e9))
## End(Not run)
binsearch(18, seq_len(3e9), H=2e9)
binsearch(2000, seq_len(3e7)*100 + 0.1)
set.seed(1)
x <- sort(sample(1:300, 30))</pre>
r <- sort(sample(1:300, 30))
plot(sapply(r, binsearch, x), type="l")
lines(sapply(r, binclosest, x), col="red")
x < -c(1, 2, 3, 5, 8, 9)
binclosest(6, x)
binclosest(7, x)
binclosest(5, x)
```

bix

Bix attributes

### **Description**

bix provides access to the bix attribute of a variable. The first form returns the value of the levels of its argument and the second sets the attribute.

```
bix(d)
bix(d) <- value
```

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### **Arguments**

```
d a "dice" object
value value to begin index at
```

### **Examples**

```
d <- dice(6)
d
bix(d)
bix(d) <- 3
d
expand(d)</pre>
```

caleidoscope

Caleidoscopic effect on a matrix

# Description

Flip a matrix vertically and horizontally before recombining into a new large matrix

### Usage

```
caleidoscope(m, odd = TRUE)
```

# Arguments

m a matrix

odd logical; should the resulting matrix have odd dimensions?

#### **Details**

Three copies of m will be made. One flipped horizontally, one flipped vertically, and one flipped both horizontally and vertically. Then they are recombined with the original matrix in the upper right corner, and the flipped copies in the upper left, lower righ and lower left corners, respectively.

#### Value

A matrix of either  $2 \times$  or  $2 \times -1$  the number of rows and columns of the input matrix.

```
caleidoscope(matrix(1:4, 2), odd=FALSE)
image(caleidoscope(1:9 %0% 1:9))
image(caleidoscope(matrix(runif(180*200)^2, 180)), col=rainbow(256, start=0.58))
```

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cbapply

Apply function to contents of clipboard

# Description

Read in clipboard contents as lines, apply a function on them, and write results back to the clipboard

### Usage

```
cbapply(FUN, ..., collapse = FALSE, write = TRUE, eval = FALSE)
```

# **Arguments**

FUN	function to be applied
	optional arguments to FUN
collapse	collapse the lines into a single string separated by newlines
write	write the results back to the clipboard
eval	parse and evaluate the results

```
## Not run:
# Copy to clipboard
a <- 10
b <- 20
s <- a + b
s
# end
# Run
ev <- cbapply(FUN=function(x) paste(x, "+ 2"), eval=TRUE)</pre>
ev; a; b; s
# Clipboard contents changed to
a <- 10 + 2
b <- 20 + 2
s \leftarrow a + b + 2
s + 2
# end
# Copy to clipboard
One Two
Three
# end
cbapply(FUN=toupper, write=FALSE)
# Clipboard contents unchanged
One Two
Three
```

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```
# end
## End(Not run)
```

central.tendency

Central tendency measures

### **Description**

Central tendency measures

#### Usage

```
pseudomedian(x, na.rm = TRUE)

cmode(x, single = TRUE, ...)

dmode(x, single = TRUE, na.rm = FALSE)

midrange(x, na.rm = FALSE)

srmean(x, na.rm = FALSE)
```

# Arguments

```
    x numeric vector
    na.rm remove NAs before starting calculations
    single return a single value (for cmode and dmode)
    send further arguments to underlying function, e.g. density for cmode
```

#### See Also

means

```
xx <- c(1, 3, 4, 5, 7, 8, 9, 9, 7, 5, 4, 5, 3, 8)
median(xx)
pseudomedian(xx)

# Discrete mode
dmode(c(2, 3, 3, 4, 5))
dmode(c(2, 3, 3, 2, 5))
dmode(c(2, 1, 3, NA, 1))
dmode(c(2, 1, 3, NA, NA))

# Continuous mode
cmode(c(2, 1, 3, NA, NA))

# Continuous mode
cmode(c(2, 3, 3, 4, 5))
cmode(c(2, 3, 3, 4, 5))
cmode(c(2, 3, 3, 4, 4, 5), n=512)
cmode(c(2, 2, 3, 3, 6, 6, 6, 7), single=FALSE, adjust=0.5)</pre>
```

20 combodice

```
# Slightly robust mean
set.seed(1)
r <- round(rexp(12)*c(-100, 100))
mean(r)
srmean(r)
weighted.mean(sort(r), c(0.5, rep(1, length(r)-2), 0.5))</pre>
```

combodice

Combine dice

#### **Description**

Generate probability density functions for combinations of dice.

### Usage

```
combodice(x, FUN, ..., method = c("outer", "expand.grid", "convolve"), name)
```

a list of dice objects, or objects that can be interpreted as such

#### **Arguments**

Х

	3
FUN	function passed on to outer or apply, depending on method
	further arguments passed to FUN
method	method for computation. One of outer, expand.grid or convolve
name	name used for the resulting PDF. Will use x object if none is given

#### **Details**

Each of the methods have their advantages and disadvantages. Outer and expand.grid work with roughly the same speed and memory, and can take the same kind of input, but FUN is interpreted differently, reflecting their use of outer and apply respectively. Convolve is much quicker than the other two, but is restricted to only summing distributions. While the first two can handle non-integer values, but only integer probabilities, the third can handle non-integer probabilities, but only integer values.

### Value

A table giving the relative probability of each value

### See Also

dusd

combodice 21

```
# Fudge dice
dF.2 <- as.table(c("-1"=2, "0"=2, "1"=2))
dF.1 <- as.table(c("-1"=1, "0"=4, "1"=1))
fudgedice2221 <- list(dF.2, dF.2, dF.2, dF.1)</pre>
combodice(fudgedice2221)
# Heterogeneous-class list and non-integer values
die1 <- as.table(c("2.6"=2, "3"=1, "5"=1))</pre>
die2 <- c(0, 1.4)
die3 <- as.dice(as.table(c("1"=2, "2"=2, "3"=2)))</pre>
diel <- list(die1, die2, die3)</pre>
combodice(diel)
# Regular d6 pair
re <- combodice(list(1:6, 1:6))</pre>
# Sichermann pair
si \leftarrow combodice(list(c(1, 2, 2, 3, 3, 4), c(1, 3, 4, 5, 6, 8)))
re; si # Identical
# One regular and one "average" d6
combodice(list(1:6, c(2, 3, 3, 4, 4, 5)))
# One 1/2 coin, one D4 and one d6, multiplied together
combodice(list(1:2, 1:4, 1:6), "*")
# Probability of getting n 1s throwing 1d4, 1d6 and 2d8
f \leftarrow function(x) sum(x == 1)
combodice(list(1:4, 1:6, 1:8, 1:8), FUN=f, method="exp")
# 3d6, discarding the lowest
discard_lowest <- function(x) sum(x[-which.min(x)])</pre>
combodice(list(1:6, 1:6, 1:6), discard_lowest, method="exp")
# 1d4, 2d6 and 1d20, discarding lowest and highest
olympic <- function(x) sum(x[-c(which.min(x), which.max(x))])
combodice(list(1:4, 1:6, 1:6, 1:20), olympic, method="exp")
# Dice pool. 3 d10 with target value 7
f \leftarrow function(x) sum(x >= 7)
combodice(lapply(rep(1, 3), seq, 10), f, method="ex")/10^3
# Equivalent using binomial PDF
dbinom(0:3, 3, 0.4)
# I have a d20 with a slight bump at the 4 and 10 facets,
\# which makes 16 and 11 less likely, but the nearby 3, 18, 19 and 20
# correspondingly more likely. How does this affect the PDF?
d201 < - dice(20)
d201[c(16, 11)] \leftarrow 0.6
d201[c(3, 20, 18, 19)] <- 1.2
mean(d201)
```

22 comparison\_with\_ties

```
c0 <- combodice(list(dice(6), dice(10), dice(20)), method="conv", name="fair")
cl <- combodice(list(dice(6), dice(10), d20l), method="conv", name="uneven")
set_mar()
plot(c0, type="o", pch=16, col="grey")
points(cl, col=2, type="o", lwd=1, pch=16, cex=0.6)
legend("topright", c("fair", "bumpy"), bty="n", col=c("grey", "red"), lwd=2:1)</pre>
```

# Description

Compare numeric values, returning an inbetween value for ties

### Usage

```
x %tgt% y

tgt(x, y, bias = 0.5)
x %tlt% y

tlt(x, y, bias = 0.5)
```

#### **Arguments**

x, y numeric values to be compared
bias what bias should be given to ties? 0.5, the default, is considered neutral as it's halfway between 1 and 0 (true and false).

# See Also

```
Comparison, tied_triple_test
```

```
1:5 %tlt% 3
1:5 %tgt% 3

c(1, 4, 3, 1) %tlt% c(1, 3, 3, 2)
c(1, 4, 3, 1) %tgt% c(1, 3, 3, 2)

# Calculate MannWhitney U statistic set.seed(1)
x <- sort(round(runif(20)*13, 1))
y <- sort(round(runif(15)*10, 1))
o <- outer(x, y, "%tgt%")

sum(o)
wilcox.test(x, y, exact=FALSE)$statistic
```

default\_par 23

default\_par

Default par

### **Description**

Sets par settings to their default values

#### Usage

```
default_par()
```

#### **Details**

Default par settings can be retreived by data(.def.par). A new default can be specified by editing def.par or making a def.par <- par(no.readonly=TRUE) type call.

### See Also

```
Other par_and_plot_margins_functions: revert_par(), set_mar()
```

dice

Create, modify or convert from/to dice objects

### **Description**

Create, modify or convert from/to dice objects

### Usage

```
dice(dval)
is.dice(x, ...)
as.dice(x, ...)
## S3 method for class 'dice'
print(x, ...)
## S3 method for class 'dice'
as.table(x, ...)
```

### **Arguments**

```
dval an integer vectorx an arbitrary R object... further arguments passed to methods
```

```
expand, table
```

24 dput2

#### **Examples**

```
# Regular d6 dice
dice(6)

# d4 dice with sides 0, 1, 2, 4
dice(c(0:3))

# d4 dice with two 2s and two 5s
dice(c(2, 2, 5, 5))
```

dput2

Write an Object to console

# Description

Writes an ASCII text representation of an R object to the console for easy copy/paste sharing

### Usage

```
dput2(
    x,
    width = 65,
    assign = c("front", "end", "none"),
    breakAtParen = FALSE,
    compact = TRUE,
    exdent = NULL
)
```

### **Arguments**

x an object

width integer; column width

assign character; should assignment be included?
breakAtParen logical; should lines break at parenthesis begins

compact remove spaces around ' = ' assignments

exdent a non-negative integer specifying the exdentation of lines after the first. default

2 if assign="front", else 0.

### **Details**

This is similar to the way dput is used to print ASCII representations of objects to the console. The differences are that dput2 lets you specify the width of the resulting column, and assignment of the object to the name used in the call will by default be included. Line breaks are by default only done on whitespace, but can be set to happen at parenthesis begins as well. This should not break code and can make for a more compact representation, but it can also make the code harder to read.

```
dput, deparse, explode
```

drop\_pattern 25

#### **Examples**

```
xmpl <- faithful[sort(sample(1:nrow(faithful), 50)), ]</pre>
dput(xmpl)
cat(deparse(xmpl, width.cutoff=65), sep='\n')
dput2(xmpl, compact=FALSE)
dput2(xmp1)
dput2(xmpl, assign="end")
dput2(xmpl, assign="none")
dput2(xmpl, 80)
# no line breaks on whitespaces or parens within character strings
xmpl <- mtcars[1:5, ]</pre>
" rrrrrrr ( bbbbbbb )",
                  "v v v v v v v v v v",
                  "( g-god, d-god, _-__)",
                  "100*(part)/(total)")
dput2(xmpl, 15)
dput2(xmpl, 15, breakAtParen=TRUE)
```

drop\_pattern

Drop predictors

### **Description**

Drop predictor variables according to a (regex) pattern

#### Usage

```
drop_pattern(form, pattern, ...)
```

### Arguments

form a formula object

pattern predictors matching this pattern will be dropped

... further arguments passed on to grep1

#### **Details**

form is divided into its individual terms, any term matching pattern is removed, before form is updated and returned. In case no match is made, form is returned unmodified. In case all predictors match, only the intercept is retained. In any case the response variable(s) are kept as is.

### Value

A formula object

```
drop_randfx
```

26 drop\_randfx

#### **Examples**

```
f6 <- y ~ aa*bb + aa + ac + cc + acab

drop_pattern(f6, "a") # Drop all containing a
drop_pattern(f6, "a{2}") # Drop all containing exactly 2 consecutive as
drop_pattern(f6, "^[^a]*a[^a]*$") # All containing exactly 1 a
drop_pattern(f6, ":") # Drop interaction
drop_pattern(f6, "^[:]*a[^:]*$") # Drop all containing a, but not interaction
drop_pattern(f6, "^((?!a).)*$", perl=TRUE) # Drop all not containing a

# Degenerate cases
drop_pattern(f6, "[abc]") # Drop all
drop_pattern(f6, "q") # Drop none</pre>
```

drop\_randfx

Drop random effects

### **Description**

Drop random effects from a mixed effects model formula

#### Usage

```
drop_randfx(form)
```

### **Arguments**

form

a formula object

#### **Details**

form is divided into its individual terms, any term containg a vertical bar (|) is removed, before form is updated and returned. In case form has no random effect terms, form is returned unmodified. In case all effects are random, only the intercept is retained. In any case the response variable(s) are kept as is.

### Value

A formula object

#### See Also

drop\_pattern

```
f1 <- Reaction \sim (1 + Days | Subject)
f2 <- Reaction \sim (1 | mygrp/mysubgrp) + (1 | Subject)
f3 <- Reaction \sim x1 + x2 + (1 + Days | Subject)
f4 <- Reaction \sim x1 * x2 + (1 | mygrp/mysubgrp) + (1 | Subject)
f5 <- Reaction \sim x1 + x2
```

dtf\_clean 27

```
sapply(list(f1, f2, f3, f4, f5), drop_randfx)
```

dtf\_clean

Data cleanup

# Description

Create a data.frame from a messy table

#### Usage

```
dtf_clean(
    x,
    header = TRUE,
    na.strings = c("NA", "N/A"),
    stringsAsFactors = FALSE,
    ...
)
```

### **Arguments**

x a messy table the form of a character string
header does the table include headers? (default TRUE)
na.strings a vector of character strings which will be interpreted as missing values
stringsAsFactors
should strings be read as factors? (default FALSE)
... further arguments passed to read.table

# **Examples**

x2 <- '

Date	1	Emp1	   	Cas	e	   	Priority		PriorityCountinLast7days
2018-06-01   2018-06-03   2018-06-02   2018-06-03	İ	A B	i	"A "A "B "B	2" 2"	i I	0 0 0 0	i	0   1   2   3

28 dusd

```
x3 <- "
   Date
          | Emp1 | Case | Priority | PriorityCountinLast7days
 2018-06-01 | A
                | A|1 |
                               0 |
 2018-06-03 | A
                | A|2 |
                               0 |
                                                       1
 2018-06-02 | B
                               0 |
                                                       2
               | B|2 |
 2018-06-03 | B
                | B|3 |
                                                       3
                               0 |
x4 <- "
Maths | English | Science | History | Class
 0.1 | 0.2
             0.3
                      0.2
                                 | Y2
 0.9 | 0.5
              0.7
                        0.4
                                 | Y1
 0.2 | 0.4
               0.6
                        0.2
                                 | Y2
 0.9 | 0.5
               0.2
                        0.7
                                 | Y1
x5 <- "
      Season | Team | W | AHWO
1 | 2017/2018 | TeamA | 2 | 1.75
2 | 2017/2018 | TeamB | 1 | 1.85
3 | 2017/2018 | TeamC | 1 | 1.70
 | 2017/2018 | TeamD | 0 | 3.10
 | 2016/2017 | TeamA | 1 | 1.49
 | 2016/2017 | TeamB | 3 | 1.51
7 | 2016/2017 | TeamC | 2 | 1.90
8
 | 2016/2017 | TeamD | 0 | N/A
lapply(c(x1, x2, x3, x4), dtf_clean)
## End(Not run)
```

dusd

Discrete (Uniform) Sum Distributions

# Description

Generate distributions of the sum of discrete (uniform) random variables. Two different approaches.

```
dusd1(xr = 1:6, n = 2, FUN = "+")

dusd2(xi = rep(1, 6), n = 2, bix = 1, round, limit = 1e-13)
```

dusd 29

#### **Arguments**

xr	numeric vector; a vector of equiprobable values
n	integer; the number of distributions to be summed
FUN	function passed on to outer
xi	numeric vector; a vector of probabilities, with indices representing values
bix	logical; where does the index of xi start?
round	integer; number of digits to round to after each convolution
limit	numeric; values (frequencies or counts) less than this will be omitted.

#### **Details**

dusd1 works by recursively taking the outer sum of xr, while dusd2 recursively convolves xi. Although convolution is more efficient, it can introduce small errors, and with repeated convolutions those errors can compound. By rounding to a slightly lower precision after each convolution the generation of spurious singletons and general imprecicions can be mitigated.

#### Value

dusd1 returns an array of size  $length(xr)^n$  representing every possible outcome. dusd2 returns a probability mass function in the form of a table.

#### See Also

combodice for a more flexible implementation of the same ideas

```
# five coin flips
plot(table(dusd1(0:1, 5)))
plot(dusd2(c(1, 1), 5, bix=0))
plot(as.table(dbinom(0:5, 5, 0.5)))
# ten flips with a loaded coin
plot(table(dusd1(c(1, 1, 2), 10)))
plot(dusd2(c(2, 1), 10))
plot(dbinom(0:10, 10, 1/3), type="h", lwd=2)
# sample from a multi-roll d4 distribution
sample(dusd1(1:4, 5), 20, replace=TRUE)
plot(ecdf(dusd1(1:4, 5)))
tt <- dusd2(xi=rep(1, 4), n=3)
plot(tt)
tt <- tt/sum(tt)
rr <- replicate(50000, sample(names(tt), prob=tt))</pre>
barplot(apply(rr, 1, table), beside=TRUE)
# distribution of the sum of three d6 rolls
plot(table(dusd1(xr=1:6, 3)))
plot(dusd2(xi=rep(1, 6), n=3))
# d6 die with faces 2, 3, 5, 7, 11, 13 (prime numbers)
plot(table(dusd1(xr=c(2, 3, 5, 7, 11, 13), 3)))
```

30 dusd

```
# Probalility of getting 7 or 8 with an 8-sided die in n out of 5 throws
1 <- 6/8
h <- 1-1
d \leftarrow as.dice(c(1, h), bix=0)
dusd2(d, 5)
# need integer "probabilities" for dusd1
table(dusd1(d*4, 5))/(4^{5})
# or an equivalent die
table(dusd1(c(0, 0, 0, 1), 5))/(4^5)
# Loaded die
p \leftarrow c(0.5, 1, 1, 1, 1, 1.5); sum(p)
plot(dusd2(xi=p, n=2))
# A loaded die with prime number faces
s <- vector(length=13)</pre>
s[c(2, 3, 5, 7, 11, 13)] \leftarrow c(0.5, 1, 1, 1, 1, 1.5)
plot(dusd2(xi=s, n=3))
# tricky to do with dusd2
plot(table(dusd1(xr=c(0.1105, 2, exp(1)), 10)))
# Demonstrating CLT
# dusd1 struggles with many iterations
\# remember it returns an array of size length(xr)^n
plot(table(dusd1(xr=c(1, 2, 9), 12)))
s <- vector(length=9)</pre>
s[c(1, 2, 9)] <- 1
plot(dusd2(xi=s, 12, round=9)) # much quicker
plot(dusd2(xi=s/sum(s), 12)) # for frequencies instead of counts
# Impossible with dusd1
clt \leftarrow dusd2(xi=s, 15, round=9)
plot(clt, lwd=0.5, col="#00000088")
# small floating-point errors from convolution.
tail(dusd2(xi=s, 15))
# dusd2 isn't always quicker
## Not run:
plot(table(dusd1(xr=c(1, 220, 3779), 12)), lwd=1)
s2 <- vector(length=3779)
s2[c(1, 220, 3779)] <- 1
plot(dusd2(xi=s2, 12, round=8), lwd=1)
# making sure the length of xi is highly composite (or more precicely 'smooth')
# improves speed
# 3779 is prime, 3780 == 2*2*3*3*3*5*7
s3 <- vector(length=3780)
s3[c(1, 220, 3779)] <- 1
plot(dusd2(xi=s3, 12, round=9), lwd=1)
## End(Not run)
```

Elo\_rating 31

|--|

### **Description**

Calculate updated Elo ratings based on existing rating and results from matches

### Usage

```
elo_upd(ra, rb, score, k = 16, sum = length(rb) > 1)
elo_upd_pw(ra, rb, score, k = 16)
```

### **Arguments**

ra	rating of player A
rb	rating of player B
score	score respective to player A. Numeric or character; 1, 0.5, 0; win, tie, loss
k	a measure of how big the correction should be
sum	calculate the new rating based on sum of scores and opponents ratings

```
# as in example from
# https://en.wikipedia.org/wiki/Elo_rating_system#Mathematical_details
# per 2019-10-04
ra <- 1613
rb <- c(1609, 1477, 1388, 1586, 1720)
score <- c(0, 0.5, 1, 1, 0)
elo_upd(ra, rb, score, k=32, sum=FALSE)
elo_upd(ra, rb, score, k=32, sum=TRUE)
elo_upd_pw(c(1400, 1500), c(1300, 1400), c("w", "t"))
results <- read.table(text="
Player1 Player2 Result
Alice
         Bob
                   Win
Charlie Dennis
                   Loss
Elena
         Frank
                   Loss
June
         Rashida Tie", header=TRUE, stringsAsFactors=FALSE)
scores <- read.table(text="</pre>
Player Score
         1150
Alice
Charlie 1150
Frank
         1150
Bob
         800
Dennis
         800
Elena
         800
June
Rashida 1100", header=TRUE, stringsAsFactors=FALSE)
```

32 entropy

```
rownames(scores) <- scores$Player

r2 <- results
r2[,1:2] <- scores[as.matrix(r2[,1:2]), 2]

r2u <- elo_upd_pw(r2)

scores.new <- data.frame(Score=c(r2u))
rownames(scores.new) <- as.matrix(results[,1:2])

scores.new <- round(scores.new[rownames(scores),, drop=FALSE])
scores.new$diff <- scores.new$Score - scores$Score
scores.new</pre>
```

entropy

Information entropy

### **Description**

Computes the information entropy (also called Shannon entropy) of a set of discrete values, or a tabulated such set.

### Usage

```
entropy(x, ...)
## S3 method for class 'table'
entropy(x, base = 2, ...)
## S3 method for class 'data.frame'
entropy(x, base = 2, ...)
## S3 method for class 'matrix'
entropy(x, base = 2, ...)
## Default S3 method:
entropy(x, base = 2, ...)
```

### **Arguments**

a vector, table, data.frame or matrix. In the case of table, data.frame and matrix each row is treated as a separate set of counts or proportions, with columns representing species, types, categories etc.
 further arguments passed to methods
 the log base to be used.

```
entropy(c(5, 5, 4, 4, 2, 3, 5)) # default is unit bits entropy(c(5, 5, 4, 4, 2, 3, 5), base=exp(1)) # unit nats
```

every\_nth 33

every\_nth

Select every n'th element

### **Description**

Select every second, third, fourth etc. element (or slice/hyperplane) of an object

### Usage

```
every_nth(...)
## Default S3 method:
every_nth(x, n = 2, start = 1, ...)
## S3 method for class 'matrix'
every_nth(x, n = 2, start = 1, margin = 1, ...)
## S3 method for class 'array'
every_nth(x, n = 2, start = 1, margin = 1, ...)
## S3 method for class 'data.frame'
every_nth(x, n = 2, start = 1, margin = 1, ...)
## S3 method for class 'list'
every_nth(x, n = 2, start = 1, ...)
```

### **Arguments**

```
    further arguments passed to methods
    an object to be selected from
    selection "step size"
    integer in [1:n] specifying the start of selection
    what margin to select along
```

34 expand

### **Examples**

```
m <- matrix(1:64, 8)
every_nth(m, n=3, start=3, margin=2)

d <- data.frame(A=1:8, B=2:9, Q=letters[rep(1:3, length.out=8)])
every_nth(d, start=2)

a <- array(1:6^4, rep(6, 4))
every_nth(a)

l <- list(a=1:3, b=2:6, c=8:5, d=9:7, e=list(ea=1:2, eb=1), f=2:6)
every_nth(l, n=2, start=2)</pre>
```

expand

Expand

# Description

Expand a "table", a "table"-like object, or a list of "table"-like objects

# Usage

```
expand(x, ...)
```

# **Arguments**

x an object to be expanded

... further arguments passed to or from methods

### Value

A vector with values and their repetitions specified by x

### See Also

```
dice, table
```

```
x <- c(4, 2, 2, 2, 3, 3, 2, 4, 6, 6)
(xt <- table(x))
(xd <- dice(x))

expand(xt)
expand(xd)

expand(list(xt, xd, x))

xn <- as.table(1:4)
names(xn) <- LETTERS[1:length(xn)]
expand(xn)</pre>
```

explode 35

explode

Explode object

#### **Description**

Presents an R object in an exploded, or expanded, form

#### Usage

```
explode(x, indent = 2)
```

# Arguments

x an R oject, or a character string describing an R object indent how many spaces for indention (and exdention) at each level

#### **Details**

If x is an R oject it is first deparsed and converted into a character string describing the object. This string is then unwrapped, or exploded, according to these rules: newline and exdention after each open parenthesis, newline and indention after each close parenthesis, and newline after each comma. Parentheses and commas forming part of character strings are ignored.

#### Value

An exploded representation of the object is printed to console, and returned invisibly. The output is in most cases a complete and reproducible representation of the object, similarly to dput, but less compact and more reaviling of its inner structure.

# See Also

```
dput, dput2
```

36 factors

factorise

**Factorise** 

# Description

Find the prime factors of a given integer

# Usage

```
factorise(x)
```

# Arguments

Χ

integer

### Value

An integer vector

# See Also

factors for unique prime factors or all integer factors

# **Examples**

```
factorise(320)
factorise(2 * 2 * 2 * 3 * 3 * 5)
prod(factorise(5641324))
## Not run:
factorise(nextn(60000000, c(2, 3)))
factorise(72*999983)
## End(Not run)
```

factors

Factors

# Description

Find the integers a given number is divisible by

# Usage

```
factors(x, prime = FALSE)
```

# Arguments

x an integer

prime should only prime factors be returned?

file\_ext 37

### Value

An integer vector

### Note

The trivial factors 1 and x itself are not included.

### See Also

factorise for prime factorisation

# **Examples**

```
factors(210)
factors(210, prime=TRUE)
```

file\_ext

File extension

# Description

Separate file name and extension from a file path

# Usage

```
file_ext(x)
file_name(x)
file_name_ext(x)
```

# **Arguments**

Х

a character vector

### **Details**

If the supplied file name has several extensions, f.ex. like foobar.tar.bz, only the last extension will be considered.

### Value

file\_ext returns the file extension of each file path. file\_name returns the file name, no extension, of each file path. file\_name\_ext returns both name and extension, but arranged in separate columns of a matrix.

### See Also

basename

38 fitrange

#### **Examples**

```
x <- c("/hg/.gi.tar.gz", "ff/hg/hh.pdf", "git", ".History", ".History.log")
file_ext(x)
file_name(x)
file_name_ext(x)</pre>
```

fingerknit

Render R example

### **Description**

Render example from R code stored on the clipboard

### Usage

```
fingerknit()
```

#### **Details**

Similar to reprex::reprex() or knitr::spin(text=, envir=new.env(),report=FALSE), but stripped down to the very basics. Input is plain valid R code taken from the clipboard. It is run in a fresh environment and both commands and results are catured. Commands are kept as is, but results are commented out. Instead of using three backticks to indicate code for markdown, each line has four whitespaces prepended. fingerknit output is also valid R code.

### Value

The clipboard is used for both input and output, but the output is also returned invisibly as a character string. Warnings and errors are not captured, but printed to console as normal. If an error is encountered nothing is returned and the clipboard data remains unchanged.

fitrange

Fit to a range

### **Description**

Linearly shift and scale a numeric vector so that it fits to a given range.

### Usage

```
fitrange(x, lower = -1, upper = 1)
```

### **Arguments**

x a numeric vector

lower the lower bound of the new vector upper the upper bound of the new vector

flatten 39

### See Also

norma

### **Examples**

```
range(fitrange(runif(10, -2, 1.5), 0, 1))
fitrange(c(2, 3, 5, 7, 4), 1, 0)
# same, but without warning
1 - fitrange(c(2, 3, 5, 7, 4), 0, 1)
```

flatten

Flatten list

### **Description**

Flatten a (nested) list to a list of its leaves

### Usage

```
flatten(x, flatten.df = FALSE, keep.order = TRUE)
```

# Arguments

x a list object

flatten.df should data.frames also be flattened?

keep.order keep the order of the original list, same as seen when using str

### **Details**

The nodes of the supplied list is traversed from root to leaf and successively unlisted until no lists are left (except possibly for data.frames).

### Value

A single level list of x's leaves.

```
xl <- list(
    O=NA,
    R=list(
        j=1:3,
        h="(a)",
    q=data.frame(
        a=1:2,
        b=c("A, K", "B, L"),
        stringsAsFactors=FALSE
    )
    ),
    N=1,
    L=FALSE
}</pre>
```

40 incdiff

```
flatten(x1, flatten.df=TRUE, keep.order=FALSE)
flatten(x1, flatten.df=TRUE, keep.order=TRUE)
str(x1)
```

gcd

Greatest common divisor

# Description

Find the largest integer, that when two numbers are divided by it, returns an integer in both cases

### Usage

```
gcd(x, y)
```

### **Arguments**

х, у

integers whose greates common divisor is to be found

### **Examples**

```
gcd(sequence(10:16), rep(10:16, 10:16))
```

incdiff

Increase difference

### **Description**

Rearrange a sorted numeric sequence so that the difference between subsequent elements is increased

# Usage

```
incdiff(x, step = 2)
```

# Arguments

x a numeric sequence

step how long a step the difference is considered for.

### **Details**

With step=2 (default) only the difference between immediate neighbours are considered; the difference between every second element will remain small, or rather reduced, compared to the original sequence. With step=3 say, differences of both lag 1 and 2 is increased, but the difference of lag 1 will be less than if a step of 2 was used.

indexvalue 41

#### **Examples**

```
x <- 1:100
diff(x)

diff(incdiff(x, 2))
diff(incdiff(x, 3))

diff(incdiff(x, 2), 2)
diff(incdiff(x, 3), 2)

# incdiff will introduce a periodicity equal to the step length acf(incdiff(x, 10))

# useful for making a sequence of colours more distinct y <- seq(0.4, 1, 1=18)
cols1 <- hsv(y, 1, y)
cols2 <- hsv(y, 1, incdiff(y, 3))

plot(y, col=cols1, pch=16, cex=5, ylim=c(0.4, 1.5))
points(y+0.5, col=cols2, pch=16, cex=5)</pre>
```

indexvalue

Index-value representation of arrays

### **Description**

Represent an array as columns of dimensional indices and value

### Usage

```
indexvalue(x, reverse = FALSE)
```

### **Arguments**

x an array or something that can be coerced into an array

reverse logical; convert from Index–value representation to regular array representation?

#### **Details**

An n-dimensional array will be unfolded to a n+1-column data frame where the first n columns represent the indices of the n dimensions, and the last column gives the value found at each index tuple. The reverse process can also be performed.

### See Also

```
latin_sq
```

42 intsect

#### **Examples**

```
arr <- array(1:(2*3*4), dim=c(2, 3, 4))
arr.is <- indexvalue(arr)</pre>
# can be used to permutate an array
indexvalue(arr.is[,c(2, 1, 3, 4)], rev=TRUE)
aperm(arr, c(2, 1, 3))
\# can interpret values (symbols) as dimensional indices and permute them as well
arr2 <- array(rep(1:6, 4), dim=c(2, 3, 4))
arr2.is <- indexvalue(arr2)</pre>
indexvalue(arr2.is[,c(1, 2, 4, 3)], rev=TRUE)
# a latin square will produce an "orthogonal array"
set.seed(1)
lsq <- latin_sq(5)</pre>
iv <- indexvalue(lsq)</pre>
i٧
\mbox{\tt\#} any permutation of a latin square is also a latin square
indexvalue(iv[, c(1, 3, 2)], reverse=TRUE)
```

intsect

Intersect

### **Description**

Performs set intersection on a list of vectors

# Usage

```
intsect(x)
```

### **Arguments**

Х

list of sets (vectors of same mode or factors)

### **Details**

The intersection between the sets in the list is found. This means no duplicate values are returned, whether or not there were any in the input.

#### Value

A vector of same mode as input, or a single factor object if input was factor.

```
intsect(list(0:6, c(2, 4, 6, 8), 3:8))
fc <- factor(LETTERS[sample(1:5, 20, rep=TRUE)])
fcl <- split(fc, sample(1:3, 20, rep=TRUE))
intsect(fcl)</pre>
```

in\_range 43

in\_range

Test if values is in a given range

# Description

Checks either whether both extrema are in the given range, or if each individual value is.

# Usage

```
in\_range(x, lower, upper, inc = c(TRUE, TRUE), na = NA)
```

### **Arguments**

x any atomic or vector-like object

lower lower range

upper upper range

inc logical vector of length one or two; should the lower and upper ranges, respectively, be considered inclusive?

na should NAs in x return TRUE, FALSE, NA, or something else?

### **Examples**

```
in_range(c(1:3, NA), 1, 4, na=NA)
in_range(1:4, 1, 4)
in_range(matrix(1:4, 2), 1, 4)
in_range(1:4, 1, 4, inc=1:0)
in_range(1:4, 1, 4, inc=0:1)
in_range(1:4, 1, 4, inc=0)

x <- as.Date(0:3, origin="2000-01-01")
in_range(x, "2000-01-01", "2000-01-04")
in_range(letters[1:4], "a", "d", inc=1:0)
in_range(letters[1:4], "a", "da", inc=1:0)
# no upper range
in_range(c(10^rnorm(9), NA), 0, NA)
in_range(c(10^rnorm(9), NA), 0, NA, na=TRUE)</pre>
```

is\_coprime

Coprimality check

### **Description**

Test whether to integers are coprime, that is, have no factors in common

### Usage

```
is_coprime(x, y)
```

44 keep\_finite

### **Arguments**

x, y integers to be tested for coprimality

### Value

A logical vector

### **Examples**

```
is_coprime(sequence(10:16), rep(10:16, 10:16)) is_coprime(2*3*5*7, 11*13)
```

is\_prime

Primality check

# Description

Test integers for whether they are prime or not

# Usage

```
is_prime(x)
```

# Arguments

Χ

vector of integers

# See Also

primes

 $keep\_finite$ 

Keep finite values

# Description

Remove NAs codeNaNs and codeInfs from data

### Usage

```
keep_finite(x, ...)
## Default S3 method:
keep_finite(x, ...)
## S3 method for class 'matrix'
keep_finite(x, margin = 1, keep = c("any", "complete"), ...)
## S3 method for class 'data.frame'
keep_finite(x, margin = 1, keep = c("any", "complete"), ...)
```

lag\_vector 45

### **Arguments**

a vector or matrix
 further arguments passed to methods
 if x is matrix, which margin to keep finites by
 keep if x is matrix, keep rows/columns with any finite values, or keep only complete rows/columns.

------

#### Value

If x is a matrix and margin is 1 or 2, a matrix is returned. Else a vector.

### **Examples**

lag\_vector

Lag an arbitrary vector

### **Description**

Lag an arbitrary vector

### Usage

```
lag_vector(x, k, type = c("cycle", "na.fill", "trim"), ...)
```

# **Arguments**

x vector to be lagged
 k integer vector specifying the number of lags
 type how to deal with non-overlapping sections
 further arguments passed to methods

46 latin\_sq

### **Examples**

```
x <- 1:9
lag_vector(x, c(0, 1, -2, 3))
lag_vector(x, c(0, 1, -2, 3), "na")
lag_vector(x, c(0, 1, -2, 3), "trim")</pre>
```

latin\_sq

Latin square

### **Description**

Generate latin squares, either randomly or ordered

### Usage

```
latin_sq(n, random = TRUE, reduce = TRUE)
```

### **Arguments**

n integer; number of unique values (aka. symbols)
random logical; should the square be generated randomly?
reduce logical; should the square be in reduced form?

# **Details**

Computation time increses rapidly with n. On my computer generating a random square with n=12 takes about ten minutes, marking the upper limit of practicability, or even stretching it a little. A latin square in reduced form will have elements in the first row and the first column in a sorted order. By setting reduced=TRUE the first row and the first column will always be 1:n.

### Value

A square integer matrix of size n^2

### See Also

indexvalue

```
set.seed(1)
ls <- latin_sq(9, reduce=TRUE)
image(ls, col=randcolours(ncol(ls)))

# The more "classic" representation with latin capital letters
ls[] <- LETTERS[ls]
ls</pre>
```

markov\_seq 47

markov\_seq

Discrete markov sequence

### **Description**

Generate a random discrete markov sequence

### Usage

```
markov_seq(n = 100, tmat = rbind(1:3, 3:1, 2:0), init = 1)
```

### **Arguments**

```
n length of the sequence
tmat a transition matrix
init the initial state
```

### **Examples**

math\_constants

Mathematical constants

### **Description**

Various mathemathical constants available as global variables

#### **Format**

An object of class numeric of length 1.

### **Details**

```
e Euler's number pi Archimedes' number, the circle constant phi Golden ratio feig1 Feigenbaum's first constant, \delta; bifurcation velocity feig2 Feigenbaum's second constant, \alpha; reduction parameter eu.ma Euler–Mascheroni constant khin Khintchine's constant glai.kin Glaisher-Kinkelin constant
```

48 means

math\_constants\_char

High precision mathematical constants

### **Description**

Character strings representing various mathemathical constants to ~100 decimal points

#### **Format**

An object of class character of length 1.

### **Details**

```
e.char Euler's number pi.char Archimedes' number, the circle constant phi.char Golden ratio feig1.char Feigenbaum's first constant, \delta; bifurcation velocity feig2.char Feigenbaum's second constant, \alpha; reduction parameter eu.ma.char Euler–Mascheroni constant khin.char Khintchine's constant glai.kin.char Glaisher-Kinkelin constant
```

means

Generalized means

# Description

Harmonic, geometric, quadratic, cubic, power and Lehmer means.

### Usage

```
harm(x, na.rm = TRUE)
geom(x, zero.rule = c("1p", "rm", "1"), na.rm = TRUE)
quad(x, na.rm = TRUE)
cubi(x, na.rm = TRUE)
powr(x, p = 1.5, na.rm = TRUE)
lehm(x, p = 2, na.rm = TRUE)
```

merge\_multiple 49

# Arguments

Х	numeric vector of values whose *mean is to be computed
na.rm	logical; should NA values be removed? (default TRUE)
zero.rule	for the geometric mean, how should zeros be dealt with? Add one before, and subtract one after the calculation (see lop1p), remove all zeros, or replace all zeros with 1.
р	exponential power. For the power mean p=-1, p=2 and p=3 gives the harmonic, quadratic and cubic means, respectively. For the Lehmer mean p=0, p=1 and p=2 gives the harmonic, arithmetic and contraharmonic means, respectively.

### Notice

For some of these means zeros and/or negative values are undefined, or make otherwise little sense in context. Workarounds are given for the geometric mean, but if you end up using it on data  $\leq 0$ , the wise call would be to reconsider whether using a geometric mean really makes sense in that case.

### See Also

```
central.tendency
```

# **Examples**

merge\_multiple

 ${\it Merge\ multiple}\ {\it data.frame} s$ 

### **Description**

Merge multiple data. frames

50 merge\_multiple

#### Usage

```
merge_multiple(
    x,
    by,
    all = FALSE,
    sort = TRUE,
    incomparables = NULL,
    include = NULL,
    exclude = NULL
)
```

# **Arguments**

```
x a list of data. frames with at least one column in common

by name of the column to be merged by, by default the full intersect of column
names between data. frames

all include all rows, including those with no match

sort sort the output on the by column(s)

incomparables values which cannot be matched.

include, exclude

numric, logical or character vector specifying which columns to include in or
```

#### **Details**

If there are duplicate columns that aren't being used to merge by, one of two things will happen. If the parent data.frames of the duplicate columns are named, then that name will be appended to the relevant column names. If the data.frames aren't named, then the data.frames idices in the parent list are appended to the relevant column names.

Inclusion and exclusion are performed in sequence, so that if both include and exclude are specified, exclude acts on the result from include.

### **Examples**

```
dtf1 <- data.frame(ast=1:4, bar=1:4, kat=c("A", "B", "C", "D"))
dtf2 <- data.frame(ast=1:6, bar=1:6, jun=9:4)
dtf3 <- data.frame(ast=2:6, bar=2:6, kat=c("A", "B", "C", "D", "E"))
dtf4 <- data.frame(ast=3:4, bar=3:4)
dtf5 <- data.frame(ast=1:-3, bar=0:4, git=0:4)

ll <- list(d1=dtf1, d2=dtf2, dtf3, A=dtf4, dtf5)

merge_multiple(ll, by="bar")
merge_multiple(ll, by="bar", all=TRUE, include=1:2)
merge_multiple(ll, by="bar", all=TRUE, exclude="kat")
merge_multiple(x=ll, by=c("bar", "ast"), all=TRUE)</pre>
```

exclude from the merge

multidensity 51

multidensity

Plot multiple kernel density estimates

# Description

Plot multiple kernel density estimates in the same window, together with a legend

# Usage

```
multidensity(
  Х,
  main,
  xlab = "",
  ylab = "Density",
  xlim,
  ylim,
  col = 1:9,
  lty = 1:2,
  lwd = 1,
  add = FALSE,
  frame.plot = TRUE,
  legend = TRUE,
  x.legend = "topleft",
  y.legend = NULL,
  bty = "o",
  box.col = "#FFFFFF00",
  bg.legend = "#FFFFFAA",
  cex.legend = 0.7,
  x.intersp = 1,
  y.intersp = 1.5,
  inset = 0,
  xpd.legend = NA,
  horiz = FALSE,
)
```

# Arguments

X	a list or data.frame of numeric values
main	a main title for the plot. Defaults to the call made to density
xlab, ylab	labels for the x and y axes
xlim, ylim	the x and y limits of the plot
col, lty, lwd	the line colours, types and widths for lines appearing in plot and legend
add	if TRUE, add to the current plot
frame.plot	an integer indicating whether a box should be drawn around the plot before the legend (1), after the legend (2), or not at all (0). Logical values are coerced to integer, so TRUE implies 1, and FALSE implies 0
legend	logical; if TRUE (the default) a legend is included with the plot

52 narm

x.legend, y.legend

the x and y co-ordinates to be used to position the legend. They can be specified

by keyword or in any way which is accepted by xy.coords

bty legend box type

box.col line colour for the legend box

bg.legend background colour for the legend box cex.legend character expanson faftor for legend

x.intersp, y.intersp

horizontal and vertical character interspacing for legend

inset the legends inset distance from the margins as a fraction of the plot region

xpd.legend the value of xpd to be used while drawing the legend

horiz logical; if TRUE, set the legend horizontally rather than vertically

... further arguments passed to density

#### Value

An invisible list of the "density" objects the plot is based on.

#### See Also

```
density, ahist
```

### **Examples**

narm Remove NAs

# Description

Remove NAs from vector or matrix

norma 53

### Usage

```
narm(x, ...)
## Default S3 method:
narm(x, ...)
## S3 method for class 'matrix'
narm(x, margin = 1, keep = c("any", "complete"), ...)
## S3 method for class 'data.frame'
narm(x, margin = 1, keep = c("any", "complete"), ...)
```

### **Arguments**

x a vector or matrix
 ... further arguments passed to methods
 margin if x is matrix, which margin to remove NAs by
 keep if x is matrix, keep rows/columns with any non-NA values, or keep only complete rows/columns.

### Value

If x is a matrix and margin is 1 or 2, a matrix is returned. Else a vector.

# **Examples**

norma

Normalize

### Description

Linearly shift and scale a numeric vector so that it has a given range, about a given centre.

54 pairwise

### Usage

```
norma(x, c = 0, r = 2)
```

# **Arguments**

x a numeric vector

c the centre (as in the midrange) for the new vector

r the range of the new vector

### See Also

```
fitrange
```

# **Examples**

```
range(norma(runif(9, -2, 0.1), 0, 2))
```

pairwise

Apply function to columns/elements pairwise

# Description

Pairwise application of a function to the columns of a matrix/data.frame or elements of a list

# Usage

```
pairwise(x, FUN, ..., comm = FALSE)
```

### **Arguments**

x a matrix or data.frame

FUN any function that takes two vectors as input and returs a single value

... further arguments passed to FUN

comm logical; is FUN commutative? If true, only the lower triangle, including the

diagonal, is computed

### Value

An  $n \times n$  square matrix with n the number of columns of x.

### See Also

```
similarity for a few more examples
```

pcamean 55

#### **Examples**

pcamean

PCA mean

### **Description**

Takes the average of several PCA objects

### Usage

```
pcamean(...)
```

### **Arguments**

prcomp, princomp or factanal objects, or a single list of such objects

# **Details**

I don't know if this kind of calculation has any sort of merit. It was written more as an impromptu challenge than as a solution to any problem

### See Also

```
prcomp, princomp, factanal
```

56 primes

```
xxs <- replicate(r, {
   xx$random <- sample(c(0:1, 0:4), 8, r=TRUE)
   xx
   }, simplify=FALSE)

xxm <- Reduce("+", xxs) / r
   xxl <- lapply(xxs, princomp)

biplot(pcamean(xxl))
biplot(princomp(xxm))</pre>
```

plot.histogram

 ${\it Plot}$  histogram  ${\it object}$ 

# Description

A a very minor modification of graphics::plot.histogram.

Only difference is that lwd now specifies the width of the histogram bars' outline.

#### See Also

```
plot.histogram, plot.stl, ahist
```

plot.stl

Plot stl object

# Description

A a very minor modification of stats::stl.

Only difference is that the distance between the plotting window and the x and y labels is set by par("mgp")[1], as it is for regular plots.

### See Also

```
plot.stl, plot.histogram
```

primes

Prime number generator

# Description

Prime generator based on the sieve of Eratosthenes

### Usage

```
primes(n)
```

quartz.png 57

#### **Arguments**

n

integer; all prime numbers up to this will be returned

### **Details**

```
Effective for primes up to ~100,000,000.
On my lightweight laptop: 1e7 -> 0.32s, 5e7 -> 1.7s, 1e8 -> 3.7s, 2e8 -> 7.6s, 3e8 -> 15s
```

#### Source

https://stackoverflow.com/questions/3789968/generate-a-list-of-primes-up-to-a-certain-number/3791284#3791284

#### See Also

is\_prime

quartz.png

Save as PNG

### **Description**

Save the contents of the current Quartz window as PNG file

# Usage

```
quartz.png(file = "%Y%m%d_%H", width = 550, dir, force = FALSE)
```

# Arguments

file file name. If it contains any "%" it is passed on as a format string to format(Sys.time(), file).

A .png file extension is added automatically.

width pixel width of the PNG file

dir directory to save to. Defaults to current working directory

force force overwriting of existing file with same name. By default duplicate path

names are resolved by appending \_N, using successive integers, to the end of the

file name.

### Value

A PNG file is written to disk and a message is written to the console, giving the new file's path and pixel dimensions. The file path is also returned invisibly.

58 quick\_table

#### **Examples**

```
## Not run:
set.seed(1)
i_h100 <- round(runif(100, 2, 30), 2)
i_cd <- rexp(100, 1/i_h100)
mydata <- data.frame(i_cd, i_h100)

mydata$i_h100_2m <- cut(mydata$i_h100, seq(2, 30, by=2))
i_cd_2m <- aggregate(i_cd ~ i_h100_2m, mydata, mean)

set_mar(x=2.5)
plot.default(i_cd_2m, xaxt="n", main="Groupwise means", xlab="", cex.main=1)
axis(1, i_cd_2m[,1], as.character(i_cd_2m[,1]), cex.axis=0.6, las=2)

quartz.png()
p <- quartz.png("test", 550)
file.info(p)

## End(Not run)</pre>
```

quick\_table

Tabulate data

### **Description**

Quick and simple function for creating contingency tables

### Usage

```
quick_table(x, na.rm = FALSE, order = c("frequency", "value", "none"))
```

### **Arguments**

x a vector or factor object
na.rm should NAs be included

order how should the results be ordered, if any?

### Value

A data.frame with columns val (the original values and class of x) and freq (the count, or frequency, of each value in x, integer). The rows are sorted by frequency in descending order.

```
set.seed(1)
m <- sample(c(rep(NA, 5), rpois(45, 3)))
quick_table(m)

x <- LETTERS[c(2, 2, 2, 2, 3, 1, 1)]
quick_table(x, order="freq")
quick_table(x, order="value")
quick_table(x, order="none")</pre>
```

rainbowHCL 59

rainbowHCL

HCL rainbow palette

### **Description**

HCL version of rainbow. Create a vector of n contiguous colours by specifying a range of Hues, and fixed Chroma and Luminance

# Usage

```
rainbowHCL(
    n,
    c = 100,
    l = 75,
    start = 0,
    end = max(1, n - 1)/n,
    alpha = 1,
    s = NULL,
    v = NULL
)
```

# **Arguments**

```
n number of colours

c, 1 the 'chroma' and 'luminance' to be used to complete the HSV color descriptions

start, end the hue in [0,1] at which the rainbow begins/ends

alpha the alpha transparency, a number in [0,1], see argument alpha in hsv

s, v 'saturation' and 'value' passed to adjustcolorHSV. Overrides 'chroma' and 'luminance' if specified.
```

```
mat2grid <- function(x) {
    eg <- expand.grid(1:NCOL(x), NROW(x):1)
    gd <- data.frame(eg, c(t(x)), stringsAsFactors=FALSE)
    colnames(gd) <- c("x", "y", "z")
    gd
    }

n <- 25
hcl0 <- rainbowHCL(n)
hcl1 <- rainbowHCL(n, c=150, l=85)
hcl2 <- rainbowHCL(n, s=1, v=1)
hsv0 <- rainbow(n)

cols <- rbind(hcl0, hcl1, hcl2, hsv0)

pos <- mat2grid(cols)
plot(pos[,1:2], pch=17, cex=3.5, col=pos[,3], ylim=c(0.5, 4.5))</pre>
```

60 randcolours

randcolours

Random colours

# Description

Generate a randomly selected colour palette

# Usage

```
randcolours(
    n,
    l = c(0.2, 0.9),
    c1 = c(0, 1),
    c2 = c(0, 1),
    alpha = 1,
    space = c("Luv", "Lab")
)
```

### **Arguments**

n	number of colours
1	lightness range
c1	colour channel one range
c2	colour channel two range
alpha	alpha channel range
space	should the parameters be interpreted as Luv or Lab components?

### **Details**

The range of 1, c1, c2 and alpha, will be interpreted as the wanted range of each colour component, whether their length is 1, 2, or more. Although they all should nominally lie within [0, 1], only alpha must do so to achieve a valid output. The others can exceed this range, at an icreased risk of clipping.

```
set.seed(3)
n <- 20
plot(1:n, col=randcolours(n), pch=16, cex=5)</pre>
```

resolve\_dup 61

resolve\_dup

Resolve duplicate

### Description

Resolve duplicate names by appending successive integers

### Usage

```
resolve_dup(x, candidates, ignore.extension = FALSE)
```

### **Arguments**

```
x character string; name to be resolved candidates character vector; possible duplicate names ignore.extension
```

logical; append to the end of x, even if it has something that can be interpreted as an extension

# **Examples**

```
x <- c("my.var", "aaa.png", "aaa.jpg", "aaa_1.png", "doc-folder")
resolve_dup("aaa.jpg", x)
resolve_dup("aaa.png", x)
resolve_dup("aaa_1.png", x)
resolve_dup("doc-folder", x)
resolve_dup("New Document", x)
resolve_dup("my.var", x, ignore.ext=TRUE)
x <- c(x, resolve_dup("aaa.png", x))
resolve_dup("aaa.png", x)</pre>
```

revert\_par

Revert par

### **Description**

Reverts par settings back to old.par

# Usage

```
revert_par()
```

### See Also

```
Other par_and_plot_margins_functions: default_par(), set_mar()
```

62 rle2

rle2

Run Length Encoding

### **Description**

Compute the lengths and values of runs of equal values in a vector

### Usage

```
rle2(
    x,
    na.unique = FALSE,
    output = c("data.frame", "rle", "named vector", "lengths", "values")
)
```

### **Arguments**

x a numeric or character vector
na.unique should every NA be conidered unique?
output what form of output

### Value

Return value depends on output.

data.frame A data.frame with lengths and values columns rle An object of class "rle"

named vector A vector of lengths with values as names lengths. The lengths as a single vector

values The values as a single vector

```
x <- c(NA, NA, 1, 2, 3, 3, NA, NA, NA, 2, 2, 2, NA, 1, 1, NA, NA)
rle2(x)

m <- matrix(c(
    0.7, 0.2, 0.1,
    0.2, 0.6, 0.2,
    0.1, 0.2, 0.7
), 3, byrow=TRUE)

set.seed(1)
y <- LETTERS[markov_seq(n=100, m)]
rle2(y, out="named")

# Same result as rle
rle2(x, na.unique=TRUE, output="rle")
rle(x)</pre>
```

seq\_range 63

```
# inverse.rle works as long as output is "rle"
inverse.rle(rle2(x, output="rle"))
```

seq\_range

Generate a sequence spanning a given range

### **Description**

Generate a sequence spanning a given range

### Usage

```
seq_range(x, ..., spread)
```

# **Arguments**

a single numeric, a range, or a sequence
 further arguments passed to seq
 use spread\_seq to spread out the range by a given factor

### **Details**

If x is a single number, the range is interpreted to be [0, x]. If x is length two, the numbers are interpreted as the left and right extrema of the sequence interval. If x is longer than two, the sequence is based upon its range.

# **Examples**

```
seq_range(c(1, 4), by=0.5)
seq_range(c(1, 4), by=0.5, spread=2)
seq_range(4)

x <- sample(1:10, 3)
seq_range(x)</pre>
```

set\_mar

Set plot margins

# Description

Moves axis titles and labels closer to the plotting window and shrinks the margins

# Usage

```
set_mar(x = 1.8, y = 1.8, main = 1, right = 1, cex.main = 1, ...)
```

64 similarity

### **Arguments**

```
x margin width for the x axis, default 2
y margin width for the x axis, default 2
main margin width for the main title, default 1
right margin width for the right edge, default 1
cex.main The magnification to be used for main titles relative to the current setting of cex, default 1
... further arguments passed to par
```

### **Details**

Old par settings are stored in .old.par before a call to par of the form par(mar=c(x, y, main, right), mgp=c(1.9, is made.)

#### See Also

```
par
Other par_and_plot_margins_functions: default_par(), revert_par()
```

### **Examples**

```
ymse:::old.par
get("old.par", envir=ymse:::ymseEnv)
ls(envir=ymse:::ymseEnv)

par(col.axis=2)
plot(1:4)

set_mar()
plot(1:4)

default_par()
plot(1:4)

revert_par()
plot(1:4)

ymse:::old.par
head(get("old.par", envir=ymse:::ymseEnv))
```

similarity

Similarity measure

# Description

Calculate the similarity between two character vectors based on a similarity matrix

### Usage

```
similarity(x, y, sm = smat(x, y), sfun = sum, ...)
```

similarity 65

#### **Arguments**

x a character vecor or two-column data.frame/matrix
y a character vector. Ignored if x is data.frame/matrix
sm a similarity matrix. By default a unit matrix
sfun function used to summarise the elementwise similarities
... further arguments passed to sfun

### See Also

smat

```
# In its most basic form similarity() gives the Hamming distance
similarity(c(1, 0, 1, 0), c(1, 1, 0, 0))
# Symmetry not required.
bef <- c(1, 2, 3, 1, 2, 3, 1, 2, 3)
aft <- c(0, 2, 2, 1, 2, 2, 1, 1, 2)
# Here a decrease in value of 1 is considered
# more similar than an increase in value of 1.
sm1 <- t(structure(c(</pre>
3, 0, 0, 0,
2, 3, 0, 0,
0, 2, 3, 0,
0, 0, 2, 3),
.Dim=c(4L, 4L),
.Dimnames=list(c("0", "1", "2", "3"), c("0", "1", "2", "3"))))
# Symmetric version
sm2 <- t(structure(c(</pre>
3, 1, 0, 0,
1, 3, 1, 0,
0, 1, 3, 1,
0, 0, 1, 3),
.Dim=c(4L, 4L),
.Dimnames=list(c("0", "1", "2","3"), c("0", "1", "2", "3"))))
similarity(bef, aft, sm1)
similarity(bef, aft, sm2)
# Pre-aligned fragments of insulin genes
data(insulin)
# Transition-transversion matrix
data(smt)
# Using pairwise() to run similarity() over all column pairs
pairwise(insulin, similarity, smt, sfun=mean)
# Imagined result from questionnaire
qu <- data.frame(
  Alice=c("happy", "sad", "angry", "unsure", "happy", "sad", "happy", "angry"),
```

66 simple\_loess

```
Bob=c("happy", "sad", "angry", "angry", "happy", "angry", "angry", "sad"),
  Charlie=c("sad", "sad", "unsure", "unsure", "happy", "sad", "angry", "sad"),
  stringsAsFactors=FALSE
)
\ensuremath{\mathtt{\#}} Similarity matrix describing the relative similitudes of the moods
emsm <- as.matrix(read.table(text="</pre>
       happy sad angry unsure
happy 5
              0
                     1
                            1
               5
                     2
                            1
  sad 0
angry
                            2
                            3",
unsure
                     2
header=TRUE))
pairwise(qu, similarity, sm=emsm/5, sfun=mean)
```

simple\_loess

Simplified Local Polynomial Regression Fitting

# Description

A simplified interface to the loess and predict.loess combo.

### Usage

```
simple_loess(...)
## Default S3 method:
simple_loess(
   y,
   x = seq_along(y),
   xout = sort(x),
   span = 0.75,
   periodic = FALSE,
   ...
)
## S3 method for class 'data.frame'
simple_loess(df, xout = sort(df[, 1]), ...)
```

### **Arguments**

	further arguments passed to loess
у	the response values to be regressed
X	the regressor, by default an integer sequence along y
xout	values used for prediction, unless it is an integer of length 1. In that case xout specifies the number of equally spaced values on the interval of $x$ to be used. By default the same as $x$
span	parameter controlling the degree of smoothing
periodic	should the input be treated as periodic?
df	a data.frame with x-values in the first column and y-vlues in the second

simple\_table 67

### Value

A data.frame with columns xout and y.predicted

### **Examples**

```
# Simple equally spaced vector
h \leftarrow c(-0.63, 0.2, -0.44, 1.6, 0.33, -0.74, -0.82, 0.29, 0.74, 0.58, -0.3)
plot(h)
lines(simple_loess(h))
# More complicated unequally space x-values
x \leftarrow c(4, 3, 2, 5, 6, 7, 9, 10, 12, 13, 14, 15, 16, 17, 18, 19)
y \leftarrow c(3, 2, 4, 5, 6, 5, 5, 3, 4, 7, 10, 10, 8, 9, 7, 8)
plot(x, y)
lines(simple_loess(y, x), col="gray40")
points(simple_loess(y=y, x=x, xout=5L), col=2, cex=2)
points(simple_loess(y=y, x=x, xout=17), col=3, cex=2)
points(simple_loess(y=y, x=x, xout=seq(8, 12, 0.3)), col=3, pch=16)
lines(simple_loess(y=y, x=x, xout=50L), col=4, lty=2)
# data.frame input
dtf <- data.frame(x, y)</pre>
simple_loess(dtf)
```

simple\_table

Read a simple table

### **Description**

Read tables given in more or less elaborate human-readable formats

# Usage

```
simple_table(
    x,
    header = TRUE,
    rem.dup.header = header,
    na.strings = c("NA", "N/A"),
    stringsAsFactors = FALSE,
    ...
)
```

### **Arguments**

```
    x a teble represented as a character string
    header are the table columns named? By default TRUE
    rem.dup.header remove duplicated headers.
    na.strings a character vector of strings which are to be interpreted as NA values
```

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stringsAsFactors

should character vectors be converted to factors? By default FALSE

... further arguments passed to read.table

# Value

 $\boldsymbol{A}$  data. frame containing a representation of the data.

impics				
x1 <- "				
			Priority	PriorityCountinLast7days
2018-06-01     2018-06-03		A1	   0   0	0
2018-06-02	В	B2	0	2
2018-06-03		B3 ⊦	0 +	3 -+
n				
x2 <- "				
Date	Emp1	Case	Priority	PriorityCountinLast7days
2018-06-01   2018-06-03   2018-06-02   2018-06-03	A   B	A 1   A 2   B 2   B 3	0   0   0	1 1 2
n				
x3 <- " Maths   Engli	ish   So	cience	History	Class
0.1   0.2	(	0.3	0.2	Y2
0.9   0.5	6	0.7	0.4	Y1
0.2   0.4	(	0.6	0.2	Y2
0.9   0.5	6	0.2	0.7	Y1
x4 <- "				
Seaso	on   1	Γeam	W   AHWO	
1   2017/201 2   2017/201 3   2017/201 4   2017/201 5   2016/201 6   2016/201 7   2016/201 8   2016/201	8   Te 8   Te 8   Te 7   Te 7   Te	eamB   eamC   eamD   eamA   eamB   eamC	2   1.75 1   1.85 1   1.70 0   3.10 1   1.49 3   1.51 2   1.90 0   N/A	

smat 69

```
X5 <- "
A T G C
------
A | 6 | 0 | 4 | 0 |
|---:-----
T | 0 | 6 | 0 | 4 |
|---:-----
G | 4 | 0 | 6 | 0 |
|-------
C | 0 | 4 | 0 | 6 |
"
```

x6 <- "

date 	Material	Description	    -
10/04/2013	WM.5597394	PNEUMATIC	i
11/07/2013	GB.D040790	RING	I

lapply(c(x1, x2, x3, x4, x5, x6), simple\_table)

smat

Similarity matrix

# Description

Create a similarity matrix

# Usage

```
smat(x, y, s, byrow = FALSE)
```

# Arguments

X	an object containing the values the similarity matrix should be computed for
у	same as $x$ . If given the union of values in $x$ and $y$ are used, if not the unique values of $x$ are used
S	a vector for filling the matrix. By default producing an identity matrix
byrow	should s fill the matrix by row?

### Value

A square matrix with the values of s and row-/colnames of the unique values in  $\{x, y\}$ .

70 speedskate

### See Also

```
similarity
```

### **Examples**

```
smat(1:3)

smat(c("f", "e", "d"), s=c(
4, 1, 1,
1, 3, 2,
1, 2, 3
))
```

speedskate

2018 MarbleLympics speed skating times

# Description

Intermediate and total times for all 16 runs, arranged by lane and heat number.

# Usage

speedskate

### **Format**

A list containing two data.frames, one for each lane. Columns are heat and rows are time checks in seconds.

# Source

https://www.youtube.com/watch?v=fA-O6f\_jArk

```
tt <- t(do.call(cbind, speedskate))
pairs(tt)
cor(tt)
outer(
  colnames(tt),
  colnames(tt),
  Vectorize(function(i,j) cor.test(tt[,i],tt[,j])$p.value)
)</pre>
```

spread\_seq 71

spread\_seq

Spread sequence

### **Description**

Spread out the values of a numeric vector

### Usage

```
spread_seq(
    x,
    f = 1.1,
    ...,
    node = c("midrange", "first", "last", "mean", "median", "min", "max")
)
```

# Arguments

x a numeric vector

f numeric or item matching a function. If numeric, a multiplicative factor applied to the distance between points, otherwise a function to be applied to differences

... further arguments passed to f if matching a function

node the location of x that will remain unchanged

```
x \leftarrow c(-1, 0, 2, NA, 4, 5, 6, 8, 9)
spread_seq(c(-1, 1))
spread_seq(x, 1.5, "midrange")
spread_seq(x, 1.5, "first")
spread_seq(x, 1.5, "last")
spread_seq(x, 1.5, "mean")
spread_seq(x, 1.5, "median")
spread_seq(c(3, 4, 1, 9, 6))
spread_seq(c(3, 4, 1, 9, 6), 2, "min")
spread_seq(c(3, 4, 1, 9, 6), 2, "max")
spread_seq(c(3, 4, 1, 9, 6), "/", 2)
spread_seq(c(3, 4, 1, 9, 6), 0.5)
y \leftarrow sort(c(3, 4, 1, 9, 6))
plot(y)
lines(spread_seq(y, "log1p"))
f <- function(x) sqrt(abs(x)*2)*sign(x)</pre>
y \leftarrow c(3, 4, 1, 9, 6)
plot(y)
lines(spread_seq(y, f=f))
```

72 tied\_triple\_test

summary.stl

Summarizing seasonal decomposition

# Description

Summary method for class "stl".

### Usage

```
## S3 method for class 'stl'
summary(object, digits = getOption("digits"), ...)
```

# Arguments

```
object an object of class "stl"

digits the number of significant digits to use when printing

further arguments passed to or from other methods
```

### **Details**

This function is a slight modification to stats:::summary.stl, the main change being the addition of the variance statistic, which can be considered a parametric (normal) compliment to the existing IQR statistic.

### **Examples**

```
set.seed(1)
x <- ts(rnorm(1e4, sd=1), frequency=12)
a <- stl(x, s.window="periodic")
stats:::summary.stl(a)
summary(a)
b <- stl(co2, s.window="periodic")
summary(b)</pre>
```

tied\_triple\_test

Tied triple test

# Description

Compare numeric values, returning an inbetween value for ties

tied\_triple\_test 73

#### Usage

```
x %ttt% y

ttt(x, y)

is.ttt(x)

## S3 method for class 'ttt'
print(x, symbols = TRUE, ...)

## S3 method for class 'ttt'
table(...)
```

### **Arguments**

x, y numeric values to be comparedsymbols should symbols be used instead of numeric values?further arguments passed to methods

#### See Also

Comparison, comparison\_with\_ties

```
1:5 %ttt% 3
ttt(1:3, 2)
print(ttt(1:3, 2), FALSE)
c(1, 6, 3, 0) %ttt% c(1, 3, 3, 2)
# Equivalent
as.integer(c(1, 6, 3, 0) %ttt% c(1, 3, 3, 2))
sign(c(1, 6, 3, 0) - c(1, 3, 3, 2))
# Demonstrating table method
dtf \leftarrow data.frame(x=1:5, y=3)
dtf" <- ttt(dtfx, dtfy)
x \leftarrow c(8, 4, 6, 8, 9, 6, 5, 7, 0, 3, 2, 1, 5, 6, 4, 7, 6,
       3, 1, 9, 5, 6, 7, 7, 4, 5, 8, 6, 2, 5, 9, 5, 4, 8)
y \leftarrow c(1, 3, 2, 4, 6, 0, 5, 3, 7, 5, 7, 4, 5, 6, 0, 1, 4,
       2, 4, 3, 1, 5, 3, 9, 2, 2, 4, 7, 5, 6, 8)
ou <- outer(sort(x), sort(y), "%ttt%")</pre>
ta <- table(ou)
pa <- capture.output(ta)</pre>
par(mar=c(1, 2, 3, 2))
image(ou, col=topo.colors(length(ta)), axes=FALSE)
title(pa)
```

74 var\_th

box()

var\_th

Theoretical variance

### **Description**

Use Calculate the theoretical variance of base probability distributions

### Usage

```
var_th(
   p,
   distribution = c("uniform", "exponential", "gamma", "t", "students-t", "bates",
   "binomial", "nbinom", "negative binomial", "beta", "f", "geometric", "hypergeometric",
   "lognormal", "log-normal", "weibull", "signed-rank", "rank-sum", "logistic")
)
```

### **Arguments**

n

a named vector of parameter values, or a single unnamed numeric if only one parameter. Use a data frame with appropriately named columns to calculate caveral variances of the same distribution.

several variances of the same distribution.

 $\hbox{\tt distribution}$ 

the name of the distribution to calculate the variance of

### **Details**

The parameters and their names are the same as used in their respective density function. In some cases, like gamma, (negative) binomial etc. more than one convention is followed.

### See Also

Distributions

```
var_th(p=data.frame(min=1:2, max=5:6), dist="unif")
var(runif(1e5, 1, 5))

var_th(p=2:3, dist="exp")
var(rexp(1e5, 2))

var_th(p=data.frame(shape=3:2, scale=c(0.8, 1)), dist="gamma")
var(rgamma(1e5, shape=3, scale=0.8))

var_th(p=c(shape=3, rate=1.25), dist="gamma")
var(rgamma(1e5, shape=3, rate=1.25))

var_th(p=18:20, dist="t")
var(rt(1e5, 18))

var_th(p=c(a=1, b=2, n=3), dist="bates")
```

weekday 75

```
var(rbates(1e5, a=1, b=2, nr=3))
var_th(p=c(size=10, prob=0.8), dist="binom")
var(rbinom(1e5, 10, 0.8))
var_th(p=c(size=10, prob=0.8), dist="nbinom")
var(rnbinom(1e5, size=10, prob=0.8))
var_th(p=c(size=10, mu=2), dist="nbinom")
var(rnbinom(1e5, size=10, mu=2))
var_th(p=data.frame(shape1=c(1, 2), shape2=c(1.5, 1)), dist="beta")
var(rbeta(1e5, shape1=1, shape2=1.5))
var(rbeta(1e5, shape1=2, shape2=1))
var_th(p=c(df1=6, df2=11), dist="f")
var(rf(1e5, 6, 11))
var_th(p=c(m=3, n=3, k=2), dist="hypergeom")
var(rhyper(1e5, m=3, n=3, k=2))
var_th(p=c(meanlog=0, sdlog=1), dist="log-normal")
var(rlnorm(1e5, meanlog=0, sdlog=1))
var_th(p=c(shape=2, scale=1), dist="weibull")
var(rweibull(1e5, shape=2, scale=1))
var_th(p=20, dist="signed-rank")
var(rsignrank(1e5, n=20))
var_th(p=c(m=13, n=10), dist="rank-sum")
var(rwilcox(1e5, m=13, n=10))
```

weekday

Week-day names

### **Description**

Convert numeric, character, factor and date-time vectors to week-day names

#### Usage

```
weekday(x, ...)
## Default S3 method:
weekday(
    x,
    short = TRUE,
    language = c("english", "nn norwegian", "bm norwegian"),
    ...
)
## S3 method for class 'Date'
weekday(x, ...)
```

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```
## S3 method for class 'POSIXt'
weekday(x, ...)
```

### **Arguments**

x a vector

... further arguments passed to methods

short if TRUE the names will be returned in shortened form language what language the names should be returned in

### **Details**

This function follows the ISO 8601 standard, meaning that Monday is considered the first day of the week.

# **Examples**

```
weekday(c("c", "b", "a"))
weekday(c("3", "2", "1"))
weekday(3:1)

weekday(Sys.Date())
weekday(Sys.Date(), short=FALSE, lang="nn nor")
```

ymse

ymse: A collection of more or less useful functions

### **Description**

There is no grand "theme" to ymse, other than that none of the functions, and in some cases function groups and classes, seemed to fit too well in any other package or merit their own package entirely.

# ymse functions

addrows Add rown to a data.frame ahist Create an average shifted histogram

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