

BISC-577: Project # 2

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Question # 1

Uniquely mappable reads: Reads that map to a unique position in the reference genome. These reads cannot come from repeated regions of DNA.

Ambiguously mapping reads: A read may often map to more than one positions in the reference genome. This is often true for reads coming from the region of say short tandem repeats. It is also possible to get more than one mapping positions for a read if mismatches are allowed. With increasing number of allowed mismatches, the number of positions that read gets mapped to also increases.

PCR duplicate reads: Upon ligation with adapters, the fragments are PCR amplified so that they are enough to be detected by the flow channel. Multiple PCR copies of the same fragment if sequenced in two different wells.

Concordantly mapped paired-end reads: In a paired end/ mate pair experiment, a fragment is sequenced from both the ends. Thus while mapping, there is an 'expectation' that these 'mates' 1 and 2 will have certain orientation. These mates are expected to be separated by an 'insert size'. However it is possible that the sequenced read comes from say a structural variation, in which the sequence is likely to map in a flipped manner, resulting in discordant mapping.

Sequenced fragment length: The sequenced fragment length refers to the 'piece' of chunked sequence that is sequenced at single or both ends post ligation and PCR amplification

Uniquely mappable part of a genome: Genome sans the repeats(Tandem repeats, interspersed repeats)

Question # 2

Single End: <http://www.ncbi.nlm.nih.gov/sra/SRX175791>[accn]

SRA size: 2.7G

FastQ size : 20G

Paired End: <http://www.ncbi.nlm.nih.gov/sra/SRX109558>[accn]

SRA size: 9.7G

FastQ size : 25G + 25G = 50G

Question # 3

Organism: Homo Sapiens

The reference was downloaded as a 2 *bit* file from UCSC

Build: hg19

Reference came as a single 2 *bit* file and was converted to FASTA using 'twoBitToFa' utility available on UCSC.

Besides the 22 autosomes and the two sex chromosomes, the FASTA contains few scaffolds for some chromosomes and the mitochondrial sequence. In total there are 93 chromosomes with total base size 3,137,161,264.

Given that these datasets come from a WGS study, it would make sense to include all sequences(including scaffolds, mitochondrial) for mapping. The overhead of having extra sequences in the reference is going to result in increased time required for searching.

Question # 4**Building index:****bowtie2:** 96m38.714s**bwa** 60m55.489s

bwa's suffix-array and bwt files are 1.5G and 3G respectively. bowtie2 createx indexes[for hg19 4 forward, 2 reverse] in mutiple files totaling 3.5G as well.

Question # 5

Mapping results are presented in SAM format. SAM stands for Sequence Alignment/Map and is a generic format for storing alignments.

SAM contains reference sequence name, the leftmost positions where the alignment starts, the query sequence(read sequence), it's quality sequence. Match, mismatch information is encoded in CIGAR format. CIGAR is a space efficient way to store matches, mismatches.

bwa does not print out the number of concordant/discordant reads that were mapped explicitly, *bowtie2* does. *bwa* does not explicitly print out number of reads that are ambigulosuly mapped. Both print the total number of reads.

Paired End:**bwa:** 107m48s for 79367217 reads**bowtie2:** 69m19s for 79367217 reads**Single End****bwa:** 37m54s for 86574968 reads**bowtie2:** 16m37s for 86574968 reads

Memory requirement was bounded by 16GB for both bwa and bowtie2, not measured explicitly.

Question # 6**SAM size Single End:****bwa:** 17354752**bowtie2:** 18234575912**Paired End****bwa:** 53215198684**bowtie2:** 52379271708

Question # 7

BAM size:

Paired End

bwa: 16107276016

bowtie2: 16250076788

samtools recognizes multiple reads that map to the same coordinates as potential PCR duplicates. However if the mate pairs are mapped discordantly(for paired end experiments) and say mate pair 1 shares the same coordinate with some other read, they are NOT regarded as duplicates. single-end duplication is treated as defined earlier.