# Day6 exercise solutions

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```
# Set global code chunk options
knitr::opts_chunk$set(warning = FALSE)
# load required libraries
library(skimr)
library(ggplot2)
library(ggpubr)
library(magrittr)
library(tidyr)
##
## Attaching package: 'tidyr'
## The following object is masked from 'package:magrittr':
##
##
       extract
library(dplyr)
## Attaching package: 'dplyr'
## The following objects are masked from 'package:stats':
##
##
       filter, lag
## The following objects are masked from 'package:base':
##
       intersect, setdiff, setequal, union
##
library(tibble)
library(lme4)
## Loading required package: Matrix
## Attaching package: 'Matrix'
```

```
## The following objects are masked from 'package:tidyr':
##
## expand, pack, unpack

library(lattice)
library(reshape2)

##
## Attaching package: 'reshape2'

## The following object is masked from 'package:tidyr':
##
## smiths

# define functions
`%notin%` <- Negate(`%in%`)</pre>
```

## Problem 1

## 1.A)

```
# Load data
hearing <- read.table(file = "data/hearing.txt", sep = "\t", header = TRUE)
hearing <- within(hearing, {
   ListID <- factor(ListID, levels = c("List1", "List2", "List3", "List4"))
})
skim(hearing)</pre>
```

Table 1: Data summary

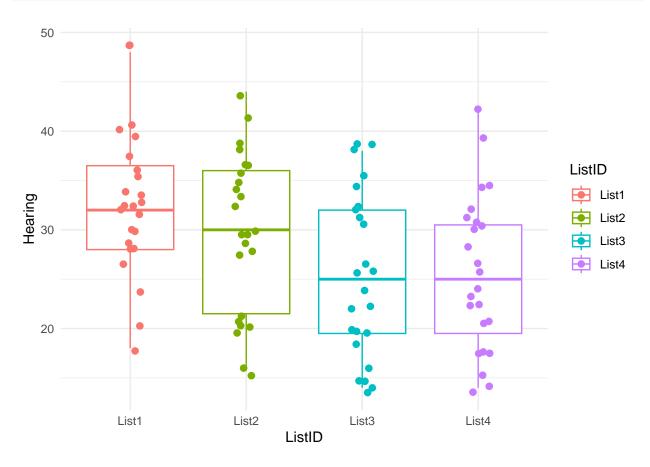
Name	hearing
Number of rows	96
Number of columns	3
Column type frequency:	
factor	1
numeric	2
Group variables	None

#### Variable type: factor

skim_variable	n_missing	complete_rate	ordered	n_unique	top_counts
ListID	0	1	FALSE	4	Lis: 24, Lis: 24, Lis: 24

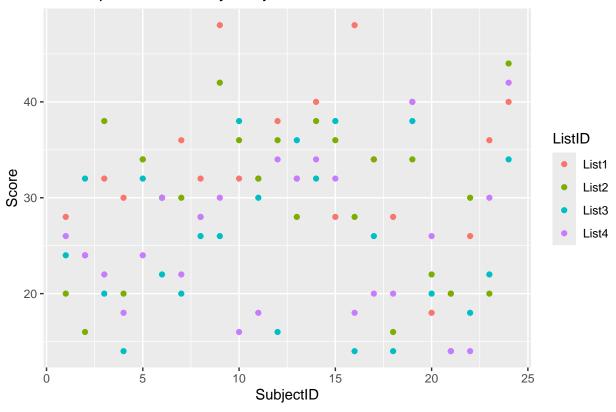
#### Variable type: numeric

skim_variable	n_missing	complete_rate	mean	$\operatorname{sd}$	p0	p25	p50	p75	p100	hist
SubjectID	0	1	12.50	6.96	1	6.75	12.5	18.25	24	
Hearing	0	1	28.31	8.37	14	20.00	30.0	34.00	48	



```
# Scatterplot of subjectids on hearing scores
ggplot(hearing, aes(x = SubjectID, y = Hearing, color = ListID)) +
  geom_point() +
  labs(title = "Scatterplot of Scores by SubjectID", x = "SubjectID", y = "Score")
```

## Scatterplot of Scores by SubjectID



## 1.B)

```
model_A <- lm(Hearing ~ ListID, data = hearing)
sum_model_A <- summary(model_A)</pre>
```

#### « comments »

Only 11.01% of variability in hearing measures are explained by different lists

We have enough evidence to state that the mean hearing score for List 3 and 4 are different than list 1, while for list 2 we cannot state that!

### 1.C)

```
# fit the mixed model
lm_mixed <- lmer(Hearing ~ 1 + ListID + (1|SubjectID), data = hearing)
sum_model_mixed <- summary(lm_mixed)
print(sum_model_mixed)</pre>
```

```
## Linear mixed model fit by REML ['lmerMod']
## Formula: Hearing ~ 1 + ListID + (1 | SubjectID)
      Data: hearing
##
## REML criterion at convergence: 635.4
##
## Scaled residuals:
##
       Min
                  1Q
                      Median
                                    3Q
                                            Max
## -1.86533 -0.56158 -0.01092 0.63222 2.69167
##
## Random effects:
## Groups
              Name
                          Variance Std.Dev.
## SubjectID (Intercept) 26.04
                                   5.103
## Residual
                                   6.027
                          36.33
## Number of obs: 96, groups: SubjectID, 24
##
## Fixed effects:
##
              Estimate Std. Error t value
                             1.612 20.315
## (Intercept)
                32.750
## ListIDList2
                -3.083
                             1.740
                                   -1.772
## ListIDList3
                -7.500
                             1.740 -4.311
## ListIDList4
                -7.167
                             1.740 -4.119
##
## Correlation of Fixed Effects:
##
               (Intr) LsIDL2 LsIDL3
## ListIDList2 -0.540
## ListIDList3 -0.540 0.500
## ListIDList4 -0.540 0.500 0.500
# fit the model without ListID (null model)
lm_mixed_null <- lmer(Hearing ~ (1 | SubjectID), data = hearing)</pre>
sum_model_mixed_null <- summary(lm_mixed_null)</pre>
# Likelihood ratio test
anova(lm_mixed_null, lm_mixed)
## refitting model(s) with ML (instead of REML)
## Data: hearing
## Models:
## lm_mixed_null: Hearing ~ (1 | SubjectID)
## lm_mixed: Hearing ~ 1 + ListID + (1 | SubjectID)
                                BIC logLik deviance Chisq Df Pr(>Chisq)
                 npar
                         AIC
## lm_mixed_null
                    3 674.22 681.91 -334.11
                                              668.22
## lm_mixed
                    6 657.70 673.09 -322.85
                                              645.70 22.52 3 5.083e-05 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
« comments »
```

The more complex model with ListID as a fixed effect is significantly more accurate at representing the data than the simpler model.

## 1.D)

Both models from  $1.\mathrm{B}$  and  $1.\mathrm{C}$  have the same estimates.

# Exercise 2

## (a)

```
termites <- read.table(file = "data/termites.txt", sep = " ", header = TRUE)
# Remove NA entries
termites <- termites %>% select_if(~ !any(is.na(.)))
# EDA
skim(termites)
```

Table 4: Data summary

Name	termites
Number of rows	16
Number of columns	15
Column type frequency:	
numeric	15
Group variables	None

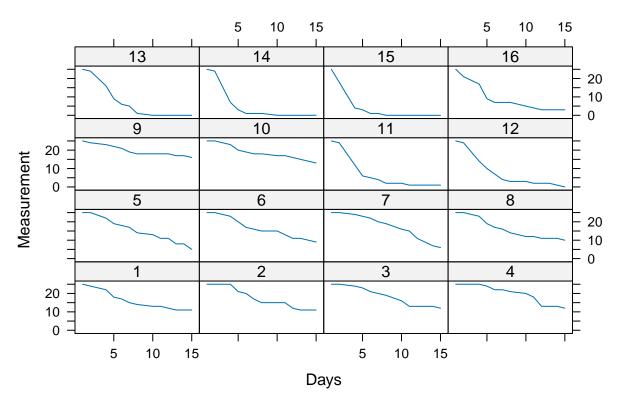
### Variable type: numeric

skim_variable	n_missing	complete_rate	mean	sd	p0	p25	p50	p75	p100	hist
dish	0	1	8.50	4.76	1	4.75	8.5	12.25	16	
dose	0	1	7.50	2.58	5	5.00	7.5	10.00	10	
day1	0	1	25.00	0.00	25	25.00	25.0	25.00	25	
day2	0	1	23.94	1.88	18	24.00	24.5	25.00	25	
day4	0	1	19.00	6.63	4	15.50	22.5	23.25	25	
day5	0	1	15.56	7.51	3	9.00	19.0	21.25	24	
day6	0	1	13.81	7.79	1	6.75	17.0	20.25	22	
day7	0	1	12.62	7.49	1	4.75	16.0	18.25	22	
day8	0	1	11.31	7.60	0	2.75	14.0	18.00	21	
day10	0	1	10.31	7.27	0	2.75	13.0	16.00	20	
day11	0	1	9.50	7.00	0	1.75	12.5	15.00	18	
day12	0	1	8.38	6.23	0	1.75	11.0	12.25	18	
day13	0	1	7.81	5.88	0	1.75	10.0	11.50	17	
day14	0	1	7.50	5.83	0	1.00	9.0	11.50	17	
day15	0	1	6.81	5.56	0	0.75	7.5	11.25	16	

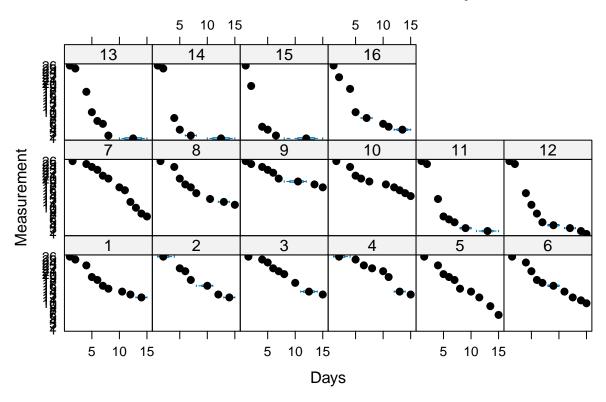
(b)

```
# Reshape data
termites <- melt(termites, id.vars = c("dish", "dose"),
                  variable.name = "day",
                  value.name = "measurement")
str(termites)
## 'data.frame':
                   208 obs. of 4 variables:
                 : int 1 2 3 4 5 6 7 8 9 10 ...
  $ dose
                 : int 555555551010\dots
                 : Factor w/ 13 levels "day1", "day2", ...: 1 1 1 1 1 1 1 1 1 1 1 ...
  $ day
   $ measurement: int 25 25 25 25 25 25 25 25 25 25 ...
# Convert day to numeric
termites$day <- as.numeric(gsub("day", "", as.character(termites$day)))</pre>
# Graphical EDA
lattice::xyplot(measurement ~ as.numeric(gsub("day", "", day)) | as.factor(dish),
      data = termites, type = "1",
      layout = c(4, 4),
      main = "Measurements Over Time for Each Dish",
      xlab = "Days", ylab = "Measurement")
```

### **Measurements Over Time for Each Dish**



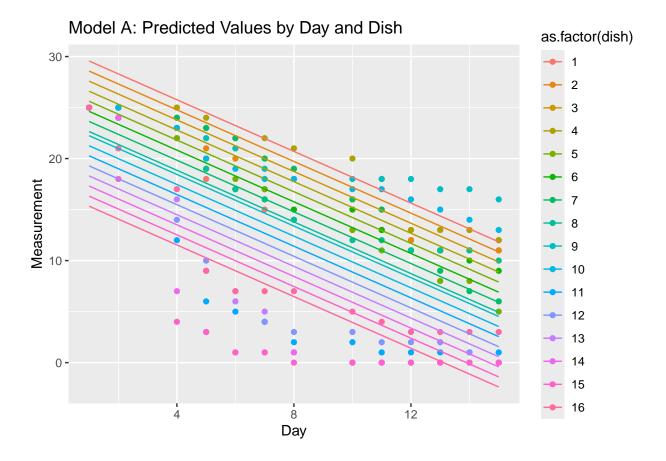
# **Measurement Distribution Across Days**



(c)

```
## dose
               0.11676
                          0.26441
                                    0.442
                                             0.659
              -1.26588
                          0.07451 -16.989
## day
                                          < 2e-16 ***
## dish
              -0.98764
                          0.14340
                                   -6.887
                                          6.9e-11 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 4.739 on 204 degrees of freedom
## Multiple R-squared: 0.6928, Adjusted R-squared: 0.6883
## F-statistic: 153.4 on 3 and 204 DF, p-value: < 2.2e-16
```

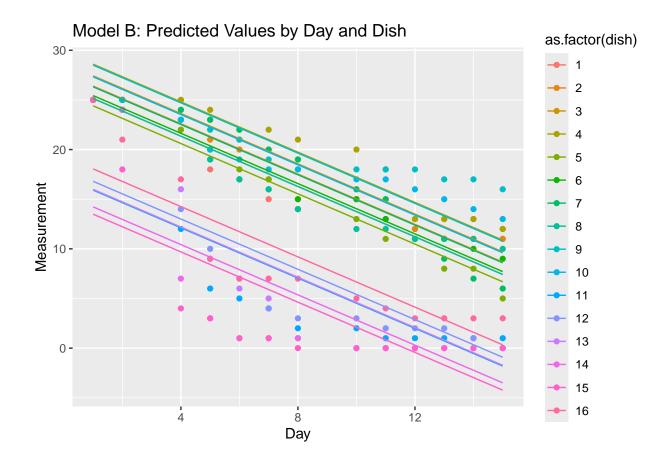
```
# Plot model
ggplot(termites, aes(x = as.numeric(gsub("day", "", day)), y = measurement, color = as.factor(dish))) +
   geom_point() +
   geom_line(aes(y = predicted_A)) +
   labs(title = "Model A: Predicted Values by Day and Dish", x = "Day", y = "Measurement")
```



It is problematic to use dish as a dependent variable, as repeated measurements are taken on the same dish. This violates the assumption of independence of observations.

(d)

```
# Fit a linear mixed model
model_B <- lmer(measurement ~ dose + day + (1 | dish), data = termites)</pre>
summary(model_B)
## Linear mixed model fit by REML ['lmerMod']
## Formula: measurement ~ dose + day + (1 | dish)
##
     Data: termites
##
## REML criterion at convergence: 1137.6
## Scaled residuals:
              1Q Median
##
      Min
                                3Q
                                       Max
## -2.0822 -0.5606 -0.0373 0.4065 3.4749
##
## Random effects:
## Groups Name
                         Variance Std.Dev.
## dish
            (Intercept) 18.66
                                  4.320
                         10.97
                                  3.312
## Residual
## Number of obs: 208, groups: dish, 16
## Fixed effects:
              Estimate Std. Error t value
##
## (Intercept) 34.68963
                           3.51844
                                   9.859
## dose
              -1.46346
                           0.44167 -3.313
## day
              -1.26588
                           0.05208 -24.305
## Correlation of Fixed Effects:
##
        (Intr) dose
## dose -0.941
## day -0.123 0.000
confint(model_B, method = "boot", nsim = 100, oldNames = FALSE)
## Computing bootstrap confidence intervals ...
##
                           2.5 %
                                     97.5 %
## sd_(Intercept)|dish 2.191328 5.8291817
## sigma
                       2.887144 3.6774336
## (Intercept)
                       27.335859 42.0978916
## dose
                       -2.353942 -0.5765336
## day
                       -1.369631 -1.1562214
termites$predicted_B <- predict(model_B)</pre>
# Plot model
ggplot(termites, aes(x = as.numeric(gsub("day", "", day)), y = measurement, color = as.factor(dish))) +
  geom_point() +
 geom line(aes(y = predicted B)) +
 labs(title = "Model B: Predicted Values by Day and Dish", x = "Day", y = "Measurement")
```



The linear mixed model is more appropriate for this data as it accounts for the repeated measurements taken on the same dish, this can also be seen visually in the plots, where each prediction in model\_B seem to be closer to the actual measurements compared to model\_A. Dose now seems highly correlated with the measurements whereas in model\_A it was not significant.

(e)

```
# Fit a linear mixed model
model_C <- lmer(measurement ~ dose + day + (day | dish) + day, data = termites)

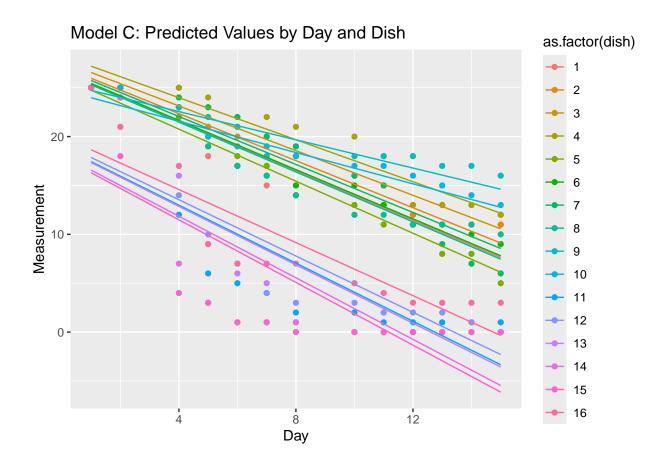
## boundary (singular) fit: see help('isSingular')

summary(model_C)

## Linear mixed model fit by REML ['lmerMod']
## Formula: measurement ~ dose + day + (day | dish) + day

## Data: termites
##
## REML criterion at convergence: 1115.1
##</pre>
```

```
## Scaled residuals:
            1Q Median
##
       Min
                                   30
                                            Max
## -2.47711 -0.51473 0.04884 0.42996 2.89506
##
## Random effects:
                        Variance Std.Dev. Corr
## Groups Name
            (Intercept) 4.77731 2.1857
## dish
                        0.06521 0.2554
##
                                           1.00
                        9.66642 3.1091
## Residual
## Number of obs: 208, groups: dish, 16
## Fixed effects:
              Estimate Std. Error t value
## (Intercept) 32.49049
                          2.18765 14.852
                           0.27571 -4.245
## dose
              -1.17024
## day
              -1.26588
                          0.08041 -15.743
##
## Correlation of Fixed Effects:
        (Intr) dose
##
## dose -0.945
## day 0.085 0.000
## optimizer (nloptwrap) convergence code: 0 (OK)
## boundary (singular) fit: see help('isSingular')
confint(model_C, method = "boot", nsim = 100, oldNames = FALSE)
## Computing bootstrap confidence intervals ...
##
## 56 message(s): boundary (singular) fit: see help('isSingular')
## 6 warning(s): Model failed to converge with max|grad| = 0.0021905 (tol = 0.002, component 1) (and ot
##
                                 2.5 %
                                           97.5 %
## sd_(Intercept)|dish
                            1.1820088 3.3756471
## cor_day.(Intercept)|dish 0.2938783 1.0000000
## sd_day|dish
                            0.1161900 0.3920406
## sigma
                            2.7200329 3.3872942
## (Intercept)
                           28.0584328 36.8895443
## dose
                           -1.6583837 -0.6167504
                            -1.4377197 -1.0974096
## day
termites$predicted_C <- predict(model_C)</pre>
# Plot model
ggplot(termites, aes(x = as.numeric(gsub("day", "", day)), y = measurement, color = as.factor(dish))) +
 geom_point() +
  geom_line(aes(y = predicted_C)) +
 labs(title = "Model C: Predicted Values by Day and Dish", x = "Day", y = "Measurement")
```



Again, this model seems to be more accurate than the previous models.

(f)

```
boostrap_ci <- function(data, formula, parameter, N = 1000, conf = 0.90) {
    estimates <- numeric(N)

for (i in 1:N) {
        resample <- data[sample(nrow(data), replace = TRUE), ]
        model <- lmer(formula, data = resample)
        estimates[i] <- fixef(model)[[parameter]]
    }

    return(quantile(estimates, c((1 - conf) / 2, 1 - (1 - conf) / 2)))
}

ci <- boostrap_ci(termites, measurement ~ dose + day + (1 | dish) + day, "dose")
ci</pre>
```

```
## 5% 95%
## -1.607911 -1.305789
```

The estimate of model\_B where we have only a random intercept for dish falls within the 90% confidence interval, therefore we can conclude that the estimate is significant.