MPI-parallel Molecular Dynamics Trajectory Analysis with the H5MD Format in the MDAnalysis Python Package

Edis Jakupovic[‡], Oliver Beckstein^{‡*}

Abstract—Fill here

Index Terms—Molecular Dynamics Simulations, High Performance Computing, Python, MDAnalysis, HDF5, H5MD, MPI I/O

Introduction

As HPC resources continue to increase, the size of molecular dynamics (MD) simulation files are now commonly terabytes in size, making serial analysis of these trajectory files impractical. Parallel analysis is a necessity for the efficient use of both HPC resources and a scientist's time. MDAnalysis is a widely used Python library that can read and write over 20 popular MD file formats while providing the same user-friendly interface [MADWB11], [GLB+16]. Previous work that focused on developing a task-based approach to parallel analysis found that an IO bound task only scaled to 12 cores due to a file IO bottleneck [SFMLIP+19]. Our previous feasibility study suggested that parallel reading via MPI-IO and HDF5 can lead to good scaling although it only used a reduced size custom HDF5 trajectory and did not provide a usable implementation of a true MD trajectory reader [KPF+20].

H5MD, or "HDF5 for molecular data", is an HDF5-based file format that is used to store MD simulation data, such as particle coordinates, box dimensions, and thermodynamic observables [dBCH14]. HDF5 is a structured, binary file format that organizes data into 2 objects: groups and datasets, which follows a hierarchical, tree-like structure, where groups represent nodes of the tree, and datasets represent the leaves [Col14]. The HDF5 library can be built on top of a message passing interface (MPI) implementation so that a file can be accessed in parallel on a parallel filesystem such as Lustre or BeeGFS. We implemented a parallel MPI-IO capable HDF5-based file format trajectory reader into MDAnalysis, H5MDReader, that adheres to H5MD specifications. H5MDReader interfaces with h5py, a high level Python package that provides a Pythonic interface to the HDF5 format such that accessing a file in parallel is as easy as passing

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a keyword argument into h5py.File, and all of parallel disk access occurs under the hood.

We benchmarked H5MDReader's parallel reading capabilities with MDAnalysis on three HPC clusters: ASU Agave, SDSC Comet, and PSC Bridges. The benchmark consisted of a simple split-apply-combine scheme of an IO-bound task that split a 90k frame (113GB) trajectory into n chunks for n processes, where each process a task on their chunk of data, and then gathered the results back to the root process. For the computational task, we computed the time series root mean squared distance (RMSD) of the positions of the alpha carbons in the protein to their initial coordinates at the first frame of the trajectory. The RMSD calculation is not only a very common task performed to analyze the dynamics of the structure of a protein, but more importantly is a very fast computation that is heavily bounded by how quickly data can be read from the file. Therefore it provided an excellent analysis candidate to test the I/O capabilities of H5MDReader.

Across the three HPC clusters tested, the benchmarks were done on both a BeeGFS and Lustre parallel filesystem which is highly suited for multi-node MPI parallelization. We tested various algorithmic optimizations for our benchmark, including altering the stripe count, loading only necessary coordinate information with numpy.Masked_arrays, and front loading all IO by loading the entire trajectory into memory prior to the RMSD calculation.

BRIEFLY DISCUSS RESULTS AND CHUNKING

Methods

We implemented a simple split-apply-combine parallelization algorithm that divides the number of frames in the trajectory evenly among all available processes. Each process receives a unique start and stop for which to iterate through their section of the trajectory and compute the RMSD at each frame. The data files used in our benchmark included a topology file YiiP_system.pdb and a trajectory file YiiP_system_9ns_center100x.h5md with 90100 frames. The trajectory file was converted on the fly with MDAnalysis to several different file formats. Table 1 gives all of these formats with how they are identified in this paper as well as their corresponding file size.

In order to obtain detailed timing information we instrumented code as follows:

```
class timeit(object):
def __enter__(self):
self._start_time = time.time()
```

[‡] Arizona State University

^{*} Corresponding author: obeckste@asu.edu

name	format	file size (GB)
H5MD_default	H5MD	113
H5MD_chunked	H5MD	113
H5MD_contiguous	H5MD	113
H5MD_gzipx1	H5MD	77
H5MD_gzipx9	H5MD	75
XTC	XTC	35
DCD	DCD	113
TRR	TRR	113

TABLE 1

```
return self

def __exit__(self, exc_type, exc_val, exc_tb):
end_time = time.time()
self.elapsed = end_time - self._start_time
    # always propagate exceptions forward
return False
```

The timeit class was used as a context manager to record how long our benchmark spent on particular lines of code. Below, we give example code of how each benchmark was performed:

```
import MDAnalysis as mda
2 from MDAnalysis.analysis.rms import rmsd
3 from mpi4py import MPI
4 import numpy as np
6 def benchmark(topology, trajectory):
      with timeit() as init_top:
         u = mda.Universe(topology)
      with timeit() as init_traj:
          u.load_new(trajectory,
10
                      driver="mpio",
11
                      comm=MPI.COMM_WORLD)
12
      t_init_top = init_top.elapsed
13
      t_init_traj = init_traj.elapsed
14
      CA = u.select_atoms("protein and name CA")
15
      x_ref = CA.positions.copy()
16
17
      total_io = 0
18
      total_rmsd = 0
19
      rmsd_array = np.empty(bsize, dtype=float)
20
      for i, frame in enumerate(range(start, stop)):
21
          with timeit() as io:
22
23
              ts = u.trajectory[frame]
24
          total_io += io.elapsed
          with timeit() as rms:
2.5
              rmsd_array[i] = rmsd(CA.positions,
                                     x ref,
27
                                     superposition=True)
28
29
          total_rmsd += rms.elapsed
30
      with timeit() as wait_time:
31
         comm.Barrier()
32
      t_wait = wait_time.elapsed
33
34
      with timeit() as comm_gather:
35
          rmsd_buffer = None
36
37
          if rank == 0:
              rmsd_buffer = np.empty(n_frames, dtype=fl
38
          comm.Gatherv(sendbuf=rmsd_array,
39
40
                        recvbuf=(rmsd buffer,
41
                                  sendcounts),
                        root=0)
42.
```

The time $t^{\rm initialize_top}$ records the time it takes to load a universe from the topology file. $t^{\rm initialize_traj}$ records the time it takes to open the trajectory file. The HDF5 file is opened with the mpio driver and the MPI.COMM_WORLD communicator to ensure the file is accessed in parallel via MPI I/O. It's important to separate the topology and trajectory initialization times, as the topology file is not opened in parallel and represents a fixed cost each

t_comm_gather = comm_gather.elapsed

process must pay to open the file. $t^{I/O}$ represents the time it takes to read the data for each frame into the corresponding MDAnalysis.Universe.trajectory.ts attribute. MD-Analysis reads data from MD trajectory files one frame, or "snapshot" at a time. Each time the u.trajectory[frame] is iterated through, MDAnalysis reads the file and fills in numpy arrays corresponding to that timestep. Each MPI process runs an identical copy of the script, but receives a unique start and stop variable such that the entire file is read in parallel. $t^{compute}$ gives the total RMSD computation time. t^{wait} records how long each process waits before the results are gathered with comm.Gather(). Gathering the results is done collectively by MPI, which means all processes must finish their iteration blocks before the results can be returned. Therefore, it's important to measure t^{wait} as it represents the existence of "straggling" processes. If one process takes substantially longer than the others to finish its iteration block, all processes are slowed down. tcomm_gather measures the time MPI spends communicating the results from each process back to the root process.

We applied this benchmark scheme to H5MD test files on Agave, Bridges, and Comet. We also tested 3 algorithmic optimizations: Lustre file striping, loading the entire trajectory into memory, and using Masked Arrays to only load the alpha carbon coordinates required for the RMSD calculation. For striping, we ran the benchmark on Bridges and Comet with a file stripe count of 48 and 96. For the into memory optimization, we used MDAnalysis.Universe.transfer_to_memory() to read the entire file in one go and pass all file I/O to the HDF5 library. For the masked array optimization, we allowed u.load_new() to take a list or array of atom indices as an argument, sub, so that the MDAnalysis.Universe.trajectory.ts arrays are instead initialized as ma.masked_array's and only the indices corresponding to sub are read from the file.

Performance was quantified by measuring the I/O timing returned from the benchmarks, and strong scaling was assessed by calculating the speedup $S(N) = t_1/t_N$ and the efficiency E(N) = S(N)/N.

Results and Discussion

TODO

Conclusions

TODO

Acknowledgments

Funding was provided by the National Science Foundation for a REU supplement to award ACI1443054. The SDSC Comet computer at the San Diego Supercomputer Center was used under allocation TG-MCB130177. The authors acknowledge Research Computing at Arizona State University for providing HPC resources that have contributed to the research results reported within this paper. We would like to acknowledge Gil Speyer and Jason Yalim from the Research Computing Core Facilities at Arizona State University for advice and consultation.

REFERENCES

[Col14] Andrew Collette. Python and hdf5. In Meghan Blanchette and Rachel Roumeliotis, editors, Python and HDF5. O'Reilly Media, Inc., 1005 Gravenstein Highway North, Sebastopol, CA 95472., 2014.

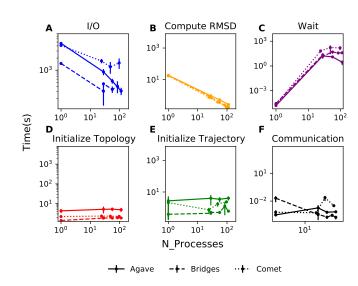


Fig. 1: caption

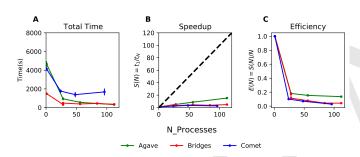


Fig. 2: caption

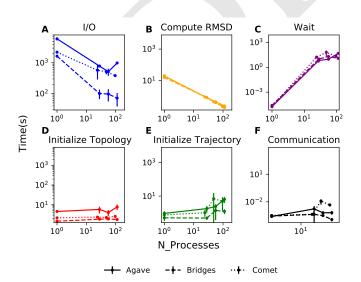


Fig. 3: caption

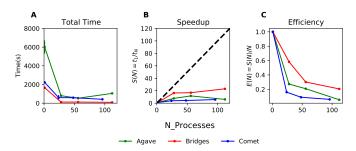


Fig. 4: caption

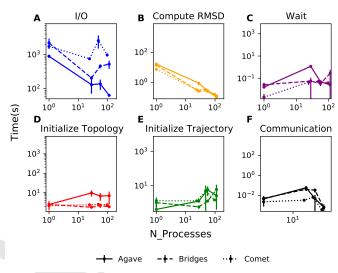


Fig. 5: caption

[dBCH14] Pierre de Buyl, Peter H. Colberg, and Felix Höfling. H5MD: A structured, efficient, and portable file format for molecular data. *Computer Physics Communications*, 185(6):1546 – 1553, 2014. doi:10.1016/j.cpc.2014.01.018.

[GLB+16]

Richard J. Gowers, Max Linke, Jonathan Barnoud, Tyler J. E. Reddy, Manuel N. Melo, Sean L. Seyler, David L Dotson, Jan Domański, Sébastien Buchoux, Ian M. Kenney, and Oliver Beckstein. MDAnalysis: A Python package for the rapid analysis of molecular dynamics simulations. In Sebastian Benthall and Scott Rostrup, editors, *Proceedings of the 15th Python in Science Conference*, pages 98–105, Austin, TX, 2016. SciPy. URL: https://www.mdanalysis.org, doi:10.25080/Majora-629e541a-00e.

[KPF+20] Mahzad Khoshlessan, Ioannis Paraskevakos, Geoffrey C. Fox, Shantenu Jha, and Oliver Beckstein. Parallel performance of molecular dynamics trajectory analysis. *Concurrency* and Computation: Practice and Experience, 32:e5789, 2020. doi:10.1002/cpe.5789.

[MADWB11] Naveen Michaud-Agrawal, Elizabeth Jane Denning,

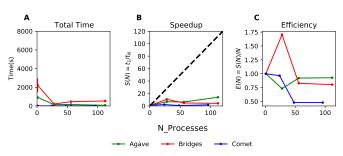


Fig. 6: caption

Thomas B. Woolf, and Oliver Beckstein. MDAnalysis: A toolkit for the analysis of molecular dynamics simulations. *J Comp Chem*, 32:2319–2327, 2011. doi:10.1002/jcc.21787.

[SFMLIP+19] Shujie Fan, Max Linke, Ioannis Paraskevakos, Richard J. Gowers, Michael Gecht, and Oliver Beckstein. PMDA - Parallel Molecular Dynamics Analysis. In Chris Calloway, David Lippa, Dillon Niederhut, and David Shupe, editors, *Proceedings of the 18th Python in Science Conference*, pages 134 – 142, Austin, TX, 2019. SciPy. doi:10.25080/Majora-7ddc1dd1-013.

