



## Review

## Calcium signalling and calcium channels: Evolution and general principles

Alexei Verkhratsky <sup>a,b,c,\*</sup>, Vladimir Parpura <sup>d,e,\*\*</sup><sup>a</sup> Faculty of Life Sciences, The University of Manchester, Oxford Road, Manchester, M13 9PT, UK<sup>b</sup> Achucarro Center for Neuroscience, IKERBASQUE, Basque Foundation for Science, 48011 Bilbao, Spain<sup>c</sup> Department of Neurosciences, University of the Basque Country UPV/EHU, 48940 Leioa, Spain<sup>d</sup> Department of Neurobiology, Center for Glial Biology in Medicine, Atomic Force Microscopy & Nanotechnology Laboratories, Clevitan International Research Center, Evelyn F. McKnight Brain Institute, University of Alabama, Birmingham, AL 35294, USA<sup>e</sup> Department of Biotechnology, University of Rijeka, 51000 Rijeka, Croatia

## ARTICLE INFO

## Article history:

Received 12 November 2013

Accepted 21 November 2013

Available online 28 November 2013

## Keywords:

Calcium

Evolution

Prokaryotes

Eukaryotes

Channels

Transporters

## ABSTRACT

Calcium as a divalent cation was selected early in evolution as a signaling molecule to be used by both prokaryotes and eukaryotes. Its low cytosolic concentration likely reflects the initial concentration of this ion in the primordial soup/ocean as unicellular organisms were formed. As the concentration of calcium in the ocean subsequently increased, so did the diversity of homeostatic molecules handling calcium. This includes the plasma membrane channels that allowed the calcium entry, as well as extrusion mechanisms, i.e., exchangers and pumps. Further diversification occurred with the evolution of intracellular organelles, in particular the endoplasmic reticulum and mitochondria, which also contain channels, exchanger(s) and pumps to handle the homeostasis of calcium ions. Calcium signalling system, based around coordinated interactions of the above molecular entities, can be activated by the opening of voltage-gated channels, neurotransmitters, second messengers and/or mechanical stimulation, and as such is all-pervading pathway in physiology and pathophysiology of organisms.

© 2013 Elsevier B.V. All rights reserved.

## Contents

1. Introduction . . . . .	1
2. Early evolution of $\text{Ca}^{2+}$ signalling . . . . .	1
3. $\text{Ca}^{2+}$ homeostasis and signalling in prokaryotes . . . . .	2
4. Diversification of $\text{Ca}^{2+}$ channels in prokaryotes . . . . .	2
5. Conclusion . . . . .	3
Acknowledgements . . . . .	3
References . . . . .	3

## 1. Introduction

Calcium ion represents an important cytosolic signalling molecule as it can affect almost all cellular processes. The calcium signalling evolved around variations in the concentration of calcium within the cytosol, with calcium being sourced from the extracellular space and/or the intracellular calcium-storing

organelles. The flux of calcium across the plasma membrane and endomembranes, i.e. membranes demarcating internal organelles, critically relies on the operation of various calcium channels within the membranes. Here, we briefly outlined the evolution and general principles of calcium signalling as an introduction to the papers that follow discussing calcium channels, in the name-sake special issue of *European Journal of Pharmacology*.

2. Early evolution of  $\text{Ca}^{2+}$  signalling

Controlled environment is the essence of life. The very first cells appeared only after they were able to fence their entrails against the world by the means of a cellular membrane.

\* Corresponding author at: Faculty of Life Sciences, The University of Manchester, Manchester, M13 9PT, UK. Tel.: +44 161 2757324.

\*\* Correspondence to: Department of Neurobiology, 1719 6th Avenue South, CIRC 429, University of Alabama, Birmingham, AL 35294, USA. Tel.: +205 996 7369.

E-mail addresses: [Alexej.Verkhratsky@manchester.ac.uk](mailto:Alexej.Verkhratsky@manchester.ac.uk) (A. Verkhratsky), [vlad@uab.edu](mailto:vlad@uab.edu) (V. Parpura).

This membrane in the animal kingdom is made of lipids, so that it is poorly, if at all, permeable to the majority of biologically relevant hydrophilic molecules and ions; the exceptions are hydrophobic compounds, which can be dissolved in lipids. This cellular separation from the surround was the first step in the long lasting story of biological evolution, which pretty much builds around a simple and effective principle of *divide et impera*, i.e., divide the world into external environment and internal space and govern everything which goes into or out of the living cell/organism.

Some of the first cells appeared in the primordial ocean in which the main elements were ions derived from the salts enriching the Earth's crust, the most abundant ions being  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Cl}^-$ ,  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$ . Out of the two divalent cations which can bind to the same sites in the cell,  $\text{Ca}^{2+}$  emerged with binding reactions that are ~100 times faster than  $\text{Mg}^{2+}$  (Williams, 2007). The concentrations of these ions in the primeval ocean are not precisely known. However, some paleontologists suggest that  $\text{Ca}^{2+}$  concentration was very low, somewhere in the range of 100 nM (Kazmierczak et al., 2013). Hence, the very first cells had acquired a very low  $\text{Ca}^{2+}$  content in their cytoplasms and lived in a low  $\text{Ca}^{2+}$  environment. Indeed, even today, some organisms like the cyanobacteria (which are probably the most ancient organisms that still live today) have a low  $\text{Ca}^{2+}$  requirement and are alkalophilic (Brock, 1973; Gerloff and Fishbeck, 1969; Kazmierczak et al., 2013). Low  $\text{Ca}^{2+}$  in the cytosol of primeval cells is also compatible with energetics based around ATP and the usage of DNA/RNA for genetic encoding, because both cannot tolerate high  $\text{Ca}^{2+}$  concentrations; at the levels above 10  $\mu\text{M}$  of  $\text{Ca}^{2+}$ , this ion induces the precipitation of phosphates, causes aggregation of proteins and nucleic acids and disrupts lipid membranes (Case et al., 2007; Jaiswal, 2001; Williams, 2007).

Washout of  $\text{Ca}^{2+}$  ions from the Earth's crust, in combination with a decreased alkalinisation of the ancient ocean, led to a continuous increase in  $\text{Ca}^{2+}$  concentration in the sea water, which in turn initiated the evolution of a  $\text{Ca}^{2+}$  homeostatic system that kept cytosolic  $\text{Ca}^{2+}$  at a low level. The molecules governing such homeostasis seem to evolve rather early in the genealogical tree as the most primitive bacteria were already in possession of  $\text{Ca}^{2+}$  pumps and  $\text{Ca}^{2+}$  exchangers. An increase in environmental  $\text{Ca}^{2+}$  concentration in combination with an evolving  $\text{Ca}^{2+}$  homeostatic system assured the build-up of a transmembrane  $\text{Ca}^{2+}$  gradient, which lies at the very base of  $\text{Ca}^{2+}$  signalling. This gradient soon was utilised by prokaryotes to develop  $\text{Ca}^{2+}$  permeable channels, which formed a pathway for a transmembrane  $\text{Ca}^{2+}$  influx and, thus, made  $\text{Ca}^{2+}$  signalling possible. In this respect, an increase in the ocean  $\text{Ca}^{2+}$  concentration could be regarded as a trigger of evolution of complex homeostatic and signalling systems.

### 3. $\text{Ca}^{2+}$ homeostasis and signalling in prokaryotes

All prokaryotic organisms living today have a low (80–100 nM) cytosolic free  $\text{Ca}^{2+}$  concentration ( $[\text{Ca}^{2+}]_c$ )—(Gandola and Rosen, 1987; Watkins et al., 1995) and several systems for  $\text{Ca}^{2+}$  extrusion that include plasmalemmal  $\text{Ca}^{2+}$  pumps (which are structurally similar to eukaryotic P-type  $\text{Ca}^{2+}$  pumps), as well as  $\text{Ca}^{2+}/\text{H}^+$  and  $\text{Na}^+/\text{Ca}^{2+}$  exchangers (Berkelman et al., 1994; Case et al., 2007; Ivey et al., 1993; Kanamaru et al., 1993; Shemarova and Nesterov, 2005). The prokaryotic cells also have intracellular  $\text{Ca}^{2+}$  signals, reflecting the activation of transmembrane  $\text{Ca}^{2+}$  fluxes through  $\text{Ca}^{2+}$  selective channels. These channels are, indeed, widespread in prokaryotic organisms, being arguably the most ancient ion channels (Shemarova and Nesterov, 2005).

There is evidence about a non-proteinaceous nature of ancient proto- $\text{Ca}^{2+}$  channels. These  $\text{Ca}^{2+}$  channels could have been constructed from large (molecular weight of 60 to 1000 kDa) polymers of

poly-3-hydroxybutyrate and smaller (12 kDa) polymers of  $\text{Ca}^{2+}$  polyphosphate (Reusch, 1999; Reusch et al., 1995). These two polymers were reported to form a transmembrane complex that behaves very much like a  $\text{Ca}^{2+}$  channel, displaying characteristic selectivity for divalent cations and being inhibited by transition metal cations like  $\text{La}^{3+}$ ,  $\text{Co}^{2+}$  and  $\text{Cd}^{2+}$ . Furthermore, these channels show elementary voltage-dependent openings when studied under patch-clamp (Reusch, 1999; Reusch et al., 1995).

Prokaryotic organisms are also in possession of  $\text{Ca}^{2+}$  channels constructed from protein helices (Durell and Guy, 2001; Matsushita et al., 1989; Tisa et al., 2000). Bacterial voltage-dependent  $\text{Ca}^{2+}$  channels contain a single domain assembled from six transmembrane  $\alpha$ -helix segments S1–S6 (Durell and Guy, 2001), being therefore different from eukaryotes where  $\text{Ca}^{2+}$  channels have a four-domain structure. Bacterial  $\text{Ca}^{2+}$  channels are, however, functionally similar to eukaryotic analogues, having similar voltage-dependence and pharmacological properties, i.e. sensitivity to phenylalkylamines, dihydropyridines and  $\text{La}^{3+}$  (Matsushita et al., 1989). For instance, the voltage-dependence of  $\text{Ca}^{2+}$  channels in *Escherichia coli* resembles that of low-voltage-activated (T)  $\text{Ca}^{2+}$  channels in eukaryotes (Tisa et al., 2000).

### 4. Diversification of $\text{Ca}^{2+}$ channels in prokaryotes

In eukaryotes,  $\text{Ca}^{2+}$  signalling systems became more complex; this is primarily associated with the development of intracellular organelles with their specific  $\text{Ca}^{2+}$  signalling mechanisms. Complexity of  $\text{Ca}^{2+}$  signalling in eukaryotes is also linked to the appearance of several types of  $\text{Ca}^{2+}$  permeable channels with distinct gating characteristics and differential  $\text{Ca}^{2+}$  permeability. In eukaryotes,  $\text{Ca}^{2+}$  fluxes through the plasma membrane are controlled by two highly  $\text{Ca}^{2+}$  selective channels, the voltage-gated  $\text{Ca}^{2+}$  channels and the store-operated Orai channels. In addition, the plasma membrane contains numerous cationic channels that include ligand-gated channels, numerous channels of the transient receptor potential (TRP) family, cyclic-nucleotide-sensitive cationic channels, mechanically-sensitive cationic channels and sperm-associated cation channels. All this remarkable diversity of  $\text{Ca}^{2+}$  permeable channels occurred very early in the evolution of unicellular organisms (Cai and Clapham, 2012), although some of their precursors have appeared even earlier in bacteria and fungi.

The ligand-gated cationic channels have very early evolutionary roots. The pentameric receptors (which in vertebrates mediate acetylcholinergic, GABAergic, glycinergic and serotonergic transmissions) are present in cyanobacteria and proteobacteria as orthologous proton-activated channels (Corringer et al., 2012). Similarly, an early analogue of ionotropic glutamate receptors, the glutamatergic receptor GluR0, is also present in bacteria (Traynelis et al., 2010). Functional ancestral ionotropic purinoreceptors of P2X class are found in protozoa, such as social amoeba *Dictyostelium discoideum* and in algae *Ostreococcus tauri*, whereas P2X protein homologues were identified in three basal fungi *Allomyces macrogyrus*, *Spizellomyces punctatus*, and *Batrachochytrium dendrobatidis* (Burnstock and Verkhratsky, 2009; Cai, 2012).

The first true homologue of voltage-gated  $\text{Ca}^{2+}$  channels appeared in fungi, represented by Cch1. This fungal protein is similar to the vertebrate channels in its overall structure, being constructed from four repeats of six-transmembrane domains with P-loop selectivity filters (Cai and Clapham, 2012; Zelter et al., 2004). Similarly, proteins homologous to sperm-associated cation channels, generally believed to be associated with animal reproduction, were identified in the basal fungus *Allomyces macrogyrus* (Cai and Clapham, 2012).

The  $\text{Ca}^{2+}$  permeable channels of the TRP family appeared in yeasts, which are in possession of the specific TRPY1 channel that

is localised in the vacuolar membrane and arguably is involved in  $\text{Ca}^{2+}$  release from this organelle (Palmer et al., 2001). More closer relatives to animals, the choanoflagellates, already have several TRP proteins homologous to mammalian TRPC, TRPV, TRPM, TRPML and TRPA channels (Cai and Clapham, 2012). Similarly, choanoflagellates *Monosiga brevicollis*, *Salpingoeca rosetta* and amoeboid animal *Capsaspora owczarzaki* already have proteins for Orai-stromal interaction molecule (STIM) store-operated  $\text{Ca}^{2+}$  influx complex; these proteins, however, are absent in fungi, indicating that they appeared in ancestral animals (Cai, 2008; Cai and Clapham, 2012).

The origin and development of intracellular  $\text{Ca}^{2+}$  channels is also associated with early animals and is rather complex. The intracellular  $\text{Ca}^{2+}$  channels are represented by two types of endoplasmic reticulum channels, the ryanodine and inositol 1,4,5 trisphosphate receptors, as well as by the mitochondrial  $\text{Ca}^{2+}$  channels, also known as mitochondrial  $\text{Ca}^{2+}$  uniports (Baughman et al., 2011; De Stefani et al., 2011; Kirichok et al., 2004; Verkhratsky, 2005). The evolution of endoplasmic reticulum channels begun in protists, which have developed quite an extended family of these molecules represented by 36 members of 6 families that share certain properties with mammalian ryanodine and inositol 1,4,5 trisphosphate receptors (Plattner and Verkhratsky, 2013). Subsequent animal evolution led to a tuning down of this extended number of ancestral forms.

## 5. Conclusion

Calcium signalling system is based around coordinated interactions of  $\text{Ca}^{2+}$  channels (that provide for the diffusional  $\text{Ca}^{2+}$  transport along electro-chemical gradients) and  $\text{Ca}^{2+}$  transporters (that move  $\text{Ca}^{2+}$  across membranes against electro-chemical gradients consuming energy). Evolution of  $\text{Ca}^{2+}$  channels resulted in the appearance of remarkably diversified classes of  $\text{Ca}^{2+}$  permeable channels, regulated by various physiological stimuli. These  $\text{Ca}^{2+}$  channels include highly selective voltage-gated and store-operated (Orai) channels and much less selective cationic channels that can be activated by neurotransmitters, second messengers or mechanical stimulation. Properties of these channels and their roles in physiology and pathophysiology form the subject of the special collection of papers that appear in this issue of *European Journal of Pharmacology*.

## Acknowledgements

We would like to thank Manoj K. Gottipati for comments on a previous version of the manuscript. Authors' research was supported by Alzheimer's Research Trust (UK) Programme Grant (ART/PG2004A/1) to A.V. and by the National Institutes of Health (The Eunice Kennedy Shriver National Institute of Child Health and Human Development award HS078678).

## References

- Baughman, J.M., Perocchi, F., Grgis, H.S., Plovanich, M., Belcher-Timme, C.A., Sancak, Y., Bao, X.R., Strittmatter, L., Goldberger, O., Bogorad, R.L., Koteliansky, V., Mootha, V.K., 2011. Integrative genomics identifies MCU as an essential component of the mitochondrial calcium uniporter. *Nature* 476, 341–345.
- Berkelman, T., Garret-Engele, P., Hoffman, N.E., 1994. The pacL gene of *Synechococcus* sp. strain PCC 7942 encodes a  $\text{Ca}^{2+}$ -transporting ATPase. *J. Bacteriol.* 176, 4430–4436.
- Brock, T.D., 1973. Lower pH limit for the existence of blue-green algae: evolutionary and ecological implications. *Science* 179, 480–483.
- Burnstock, G., Verkhratsky, A., 2009. Evolutionary origins of the purinergic signalling system. *Acta Physiol.* 195, 415–447.
- Cai, X., 2008. Unicellular  $\text{Ca}^{2+}$  signaling 'toolkit' at the origin of metazoa. *Mol. Biol. Evol.* 25, 1357–1361.
- Cai, X., 2012. P2X receptor homologs in basal fungi. *Purinergic Signal.* 8, 11–13.
- Cai, X., Clapham, D.E., 2012. Ancestral  $\text{Ca}^{2+}$  signaling machinery in early animal and fungal evolution. *Mol. Biol. Evol.* 29, 91–100.
- Case, R.M., Eisner, D., Gurney, A., Jones, O., Muallem, S., Verkhratsky, A., 2007. Evolution of calcium homeostasis: from birth of the first cell to an omnipresent signalling system. *Cell Calcium* 42, 345–350.
- Corringer, P.J., Poitevin, F., Prevost, M.S., Sauguet, L., Delarue, M., Changeux, J.P., 2012. Structure and pharmacology of pentameric receptor channels: from bacteria to brain. *Structure* 20, 941–956.
- De Stefani, D., Raffaello, A., Teardo, E., Szabo, I., Rizzuto, R., 2011. A forty-kilodalton protein of the inner membrane is the mitochondrial calcium uniporter. *Nature* 476, 336–340.
- Durell, S.R., Guy, H.R., 2001. A putative prokaryote voltage-gated  $\text{Ca}^{2+}$  channel with only one 6TM motif per subunit. *Biochem. Biophys. Res. Commun.* 281, 741–746.
- Gandola, P., Rosen, B.P., 1987. Maintenance of intracellular calcium in *Escherichia coli*. *J. Biol. Chem.* 262, 12570–12574.
- Gerloff, G.C., Fishbeck, K.A., 1969. Quantitative cation requirements of several green and blue-green algae. *J. Phycol.* 5, 109–114.
- Ivey, D.M., Guffanti, A.A., Zemske, J., Pinner, E., Karpel, R., Padan, E., Schuldiner, S., Krulwich, T.A., 1993. Cloning and characterization of a putative  $\text{Ca}^{2+}/\text{H}^+$  antiporter gene in *Escherichia coli* upon functional complementation of  $\text{Na}^+/\text{H}^+$  antiporter-deficient strains by the overexpressed gene. *J. Biol. Chem.* 268, 11296–11303.
- Jaiswal, J.K., 2001. Calcium—how and why? *J. Biosci.* 26, 357–363.
- Kanamaru, K., Kashiwagi, S., Mizuno, T., 1993. The cyanobacterium *Synechococcus* sp. PCC 7942 possesses 2 distinct genes encoding cation-transporting P-Type ATPases. *FEBS Lett.* 330, 99–104.
- Kazmierczak, J., Kempe, S., Kremer, B., 2013. Calcium in the early evolution of living systems: a biohistorical approach. *Curr. Org. Chem.* 17, 1738–1750.
- Kirichok, Y., Kapivinsky, G., Clapham, D.E., 2004. The mitochondrial calcium uniporter is a highly selective ion channel. *Nature* 427, 360–364.
- Matsushita, T., Hirata, H., Kusaka, I., 1989. Calcium channels in bacteria. Purification and characterization. *Ann. N.Y. Acad. Sci.* 560, 426–429.
- Palmer, C.P., Zhou, X.L., Lin, J., Loukin, S.H., Kung, C., Saimi, Y., 2001. A TRP homolog in *Saccharomyces cerevisiae* forms an intracellular  $\text{Ca}(2+)$ -permeable channel in the yeast vacuolar membrane. *Proc. Nat. Acad. Sci. U.S.A.* 98, 7801–7805.
- Plattner, H., Verkhratsky, A., 2013.  $\text{Ca}^{2+}$  signalling early in evolution—all but primitive. *J. Cell Sci.* 126, 2141–2150.
- Reusch, R.N., 1999. Polyphosphate/poly-(R)-3-hydroxybutyrate) ion channels in cell membranes. *Prog. Mol. Subcell. Biol.* 23, 151–182.
- Reusch, R.N., Huang, R., Bramble, L.L., 1995. Poly-3-hydroxybutyrate/polyphosphate complexes form voltage-activated  $\text{Ca}^{2+}$  channels in the plasma membranes of *Escherichia coli*. *Biophys. J.* 69, 754–766.
- Shemarova, I.V., Nesterov, V.P., 2005. Evolution of mechanisms of calcium signaling: the role of calcium ions in signal transduction in prokaryotes. *Zh. Evol. Biokhim. Fiziol.* 41, 12–17.
- Tisa, L.S., Sekelsky, J.J., Adler, J., 2000. Effects of organic antagonists of  $\text{Ca}^{2+}$ ,  $\text{Na}^+$ , and  $\text{K}^+$  on chemotaxis and motility of *Escherichia coli*. *J. Bacteriol.* 182, 4856–4861.
- Traynelis, S.F., Wollmuth, L.P., McBain, C.J., Menniti, F.S., Vance, K.M., Ogden, K.K., Hansen, K.B., Yuan, H., Myers, S.J., Dingledine, R., 2010. Glutamate receptor ion channels: structure, regulation, and function. *Pharmacol. Rev.* 62, 405–496.
- Verkhratsky, A., 2005. Physiology and pathophysiology of the calcium store in the endoplasmic reticulum of neurons. *Physiol. Rev.* 85, 201–279.
- Watkins, N.J., Knight, M.R., Trewalas, A.J., Campbell, A.K., 1995. Free calcium transients in chemotactic and non-chemotactic strains of *Escherichia coli* determined by using recombinant aequorin. *Biochem. J.* 306, 865–869.
- Williams, R.J.P., 2007. The evolution of the biochemistry of calcium. In: Krebs, J., Michalak, M. (Eds.), *Calcium: A Matter of Life and Death*. Elsevier, Amsterdam, pp. 23–48.
- Zelter, A., Bencina, M., Bowman, B.J., Yarden, O., Read, N.D., 2004. A comparative genomic analysis of the calcium signaling machinery in *Neurospora crassa*, *Magnaporthe grisea*, and *Saccharomyces cerevisiae*. *Fungal Genet. Biol.: FG & B* 41, 827–841.