

转录组流程

step1: sra2fastq

下载SRA数据

新建一个名为 `SRR_Acc_List.txt` 的文档，将SRR号码保存在文档内，一个号码占据一行。文件可以在我的GitHub下载获取：

https://raw.githubusercontent.com/jmzeng1314/GEO/master/airway_RNAseq/SRR_Acc_List.txt 当然了，你自己去GEO里面找到SRA再找到文件才是正路。

- prefetch下载数据

```
wkd=/home/jmzeng/project/airway/ #设置工作目录
source activate rna
cat SRR_Acc_List.txt | while read id; do (prefetch ${id}
);done
ps -ef | grep prefetch | awk '{print $2}' | while read id;
do kill ${id}; done #在内地下载速度很慢，所以我杀掉这些下载进程
```

- 或者直接使用我已经下载好的sra数据

```
mkdir $wkd/raw
cd $wkd/raw
ls /public/project/RNA/airway/sra/* | while read id; do (
fastq-dump --gzip --split-3 -O ./ ${id} ); done ## 批量转换
sra到fq格式。
source deactivate
```

- 得到的SRA数据如下

```
/public/project/RNA/airway/sra/
```

```
|— [1.6G] SRR1039508.sra  
|— [1.4G] SRR1039509.sra  
|— [1.6G] SRR1039510.sra  
|— [1.5G] SRR1039511.sra  
|— [2.0G] SRR1039512.sra  
|— [2.2G] SRR1039513.sra  
|— [3.0G] SRR1039514.sra  
|— [1.9G] SRR1039515.sra  
|— [2.1G] SRR1039516.sra  
|— [2.6G] SRR1039517.sra  
|— [2.3G] SRR1039518.sra  
|— [2.0G] SRR1039519.sra  
|— [2.1G] SRR1039520.sra  
|— [2.4G] SRR1039521.sra  
|— [2.0G] SRR1039522.sra  
|— [2.2G] SRR1039523.sra
```

- sra格式转fastq格式

格式转还用到的软件是fastq-dump

```
for i in $(ls /public/project/RNA/airway/sra/*sra)
do
    echo $i
    fastq-dump --split-3 --skip-technical --clip --gzip $i ## 批量转换
done
```

- 得到fastq数据如下

原始数据是双端测序结果，fastq-dump配合--split-3参数，一个样本被拆分成两个fastq文件

```
|— [1.3G]  SRR1039508_1.fastq.gz
|— [1.3G]  SRR1039508_2.fastq.gz
|— [1.2G]  SRR1039509_1.fastq.gz
|— [1.2G]  SRR1039509_2.fastq.gz
|— [1.3G]  SRR1039510_1.fastq.gz
|— [1.3G]  SRR1039510_2.fastq.gz
|— [1.2G]  SRR1039511_1.fastq.gz
|— [1.2G]  SRR1039511_2.fastq.gz
|— [1.6G]  SRR1039512_1.fastq.gz
|— [1.6G]  SRR1039512_2.fastq.gz
|— [950M]  SRR1039513_1.fastq.gz
|— [952M]  SRR1039513_2.fastq.gz
|— [2.4G]  SRR1039514_1.fastq.gz
.....
|— [1.5G]  SRR1039522_1.fastq.gz
|— [1.5G]  SRR1039522_2.fastq.gz
|— [1.8G]  SRR1039523_1.fastq.gz
|— [1.8G]  SRR1039523_2.fastq.gz
```

step2: check quality of sequence reads

fastqc生成质控报告，multiqc将各个样本的质控报告整合为一个。

```
ls *.gz | xargs fastqc -t 2
multiqc ./
```

- 得到结果如下

```
|— [4.0K]  multiqc_data
|   |— [2.1M]  multiqc_data.json
|   |— [6.8K]  multiqc_fastqc.txt
|   |— [2.2K]  multiqc_general_stats.txt
```

```
|   |— [ 16K]  multiqc.log
|   |— [3.4K]  multiqc_sources.txt
|— [1.5M]  multiqc_report.html
|— [236K]  SRR1039508_1_fastqc.html
|— [279K]  SRR1039508_1_fastqc.zip
|— [238K]  SRR1039508_2_fastqc.html
|— [286K]  SRR1039508_2_fastqc.zip
|— [236K]  SRR1039510_1_fastqc.html
|— [278K]  SRR1039510_1_fastqc.zip
|— [241K]  SRR1039510_2_fastqc.html
|— [292K]  SRR1039510_2_fastqc.zip
.....
|— [220K]  SRR1039522_fastqc.zip
|— [234K]  SRR1039523_1_fastqc.html
|— [273K]  SRR1039523_1_fastqc.zip
|— [232K]  SRR1039523_2_fastqc.html
|— [274K]  SRR1039523_2_fastqc.zip
```

每个id_fastqc.html都是一个质量报告，multiqc_report.html是所有样本的整合报告

step3: filter the bad quality reads and remove adaptors.

- 运行如下代码，得到名为config的文件，包含两列数据

```
mkdir $wkd/clean
cd $wkd/clean
ls /home/jmzeng/project/airway/raw/*_1.fastq.gz >1
ls /home/jmzeng/project/airway/raw/*_2.fastq.gz >2
paste 1 2 > config
```

- 打开文件 qc.sh ，并且写入如下内容

trim_galore, 用于去除低质量和接头数据

```

source activate rna
bin_trim_galore=trim_galore
dir='/home/jmzeng/project/airway/clean'
cat $1 |while read id
do
    arr=(${id})
    fq1=${arr[0]}
    fq2=${arr[1]}
    $bin_trim_galore -q 25 --phred33 --length 36 --stringency
3 --paired -o $dir $fq1 $fq2
done
source deactivate

```

- 运行qc.sh

```

bash qc.sh config #config是传递进去的参数

```

- 结果显示如下

```

├─ [2.9K]  SRR1039508_1.fastq.gz_trimming_report.txt
├─ [1.2G]  SRR1039508_1_val_1.fq.gz
├─ [3.1K]  SRR1039508_2.fastq.gz_trimming_report.txt
├─ [1.2G]  SRR1039508_2_val_2.fq.gz
├─ [2.9K]  SRR1039509_1.fastq.gz_trimming_report.txt
.....
├─ [2.9K]  SRR1039522_1.fastq.gz_trimming_report.txt
├─ [1.4G]  SRR1039522_1_val_1.fq.gz
├─ [3.1K]  SRR1039522_2.fastq.gz_trimming_report.txt
├─ [1.4G]  SRR1039522_2_val_2.fq.gz
├─ [2.9K]  SRR1039523_1.fastq.gz_trimming_report.txt
├─ [1.7G]  SRR1039523_1_val_1.fq.gz
├─ [3.1K]  SRR1039523_2.fastq.gz_trimming_report.txt
├─ [1.7G]  SRR1039523_2_val_2.fq.gz

```

step4: alignment

star, hisat2, bowtie2, tophat, bwa, subread都是可以用于比到的软件

- 先运行一个样本，测试一下

```
mkdir $wkd/test
cd $wkd/test
source activate rna
ls $wkd/clean/*gz |while read id;do (zcat ${id}|head -1000>
$(basename ${id} ".gz"));done
id=SRR1039508
hisat2 -p 10 -x /public/reference/index/hisat/hg38/genome
-1 ${id}_1_val_1.fq -2 ${id}_2_val_2.fq -S
${id}.hisat.sam
subjunc -T 5 -i /public/reference/index/subread/hg38 -r
${id}_1_val_1.fq -R ${id}_2_val_2.fq -o ${id}.subjunc.sam
bowtie2 -p 10 -x /public/reference/index/bowtie/hg38 -1
${id}_1_val_1.fq -2 ${id}_2_val_2.fq -S ${id}.bowtie.sam
bwa mem -t 5 -M /public/reference/index/bwa/hg38
${id}_1_val_1.fq ${id}_2_val_2.fq > ${id}.bwa.sam
```

- 批量比对代码

```
cd $wkd/clean
ls *gz|cut -d"_" -f 1 |sort -u |while read id;do
ls -lh ${id}_1_val_1.fq.gz ${id}_2_val_2.fq.gz
hisat2 -p 10 -x /public/reference/index/hisat/hg38/genome
-1 ${id}_1_val_1.fq.gz -2 ${id}_2_val_2.fq.gz -S
${id}.hisat.sam
subjunc -T 5 -i /public/reference/index/subread/hg38 -r
${id}_1_val_1.fq.gz -R ${id}_2_val_2.fq.gz -o
${id}.subjunc.sam
bowtie2 -p 10 -x /public/reference/index/bowtie/hg38 -1
${id}_1_val_1.fq.gz -2 ${id}_2_val_2.fq.gz -S
${id}.bowtie.sam
bwa mem -t 5 -M /public/reference/index/bwa/hg38
${id}_1_val_1.fq.gz ${id}_2_val_2.fq.gz > ${id}.bwa.sam
done
```

这里是演示多个比对工具，但事实上，对RNA-seq数据来说，不要使用bwa和bowtie这样的软件，它需要的是能进行跨越内含子比对的工具。

- sam文件转bam

```
ls *.sam | while read id ;do (samtools sort -O bam -@ 5 -o  
$(basename ${id} ".sam").bam ${id});done  
rm *.sam
```

- 为bam文件建立索引

```
ls *.bam | xargs -i samtools index {}
```

- reads的比对情况统计

```
ls *.bam | xargs -i samtools flagstat -@ 2 {} >  
ls *.bam | while read id ;do ( samtools flagstat -@ 1 $id >  
$(basename ${id} ".bam").flagstat );done  
source deactivate
```

- 最终结果显示如下

```
|— [1.8G]  SRR1039508.bowite2.bam  
|— [2.9M]  SRR1039508.bowite2.bam.bai  
|— [ 444]  SRR1039508.bowite2.flagstat  
|— [ 10G]  SRR1039508.bowite2.sam  
|— [1.7G]  SRR1039509.bowite2.bam  
.....  
|— [2.0G]  SRR1039521.bowite2.bam  
|— [2.9M]  SRR1039521.bowite2.bam.bai  
|— [ 444]  SRR1039521.bowite2.flagstat  
|— [ 10G]  SRR1039521.bowite2.sam  
|— [2.3G]  SRR1039522.bowite2.bam  
|— [3.0M]  SRR1039522.bowite2.bam.bai  
|— [ 444]  SRR1039522.bowite2.flagstat  
|— [ 12G]  SRR1039522.bowite2.sam  
|— [2.5G]  SRR1039523.bowite2.bam  
|— [3.0M]  SRR1039523.bowite2.bam.bai
```

```
| [ 444] SRR1039523.bowtie2.flagstat  
| [ 14G] SRR1039523.bowtie2.sam
```

step5: counts

```
mkdir $wkd/align  
cd $wkd/align  
source activate rna  
# 如果一个一个样本单独计数，输出多个文件使用代码是：  
for fn in {508..523}  
do  
featureCounts -T 5 -p -t exon -g gene_id -a  
/public/reference/gtf/genencode/genencode.v25.annotation.gtf.gz  
-o $fn.counts.txt SRR1039$fn.bam  
done  
# 如果是批量样本的bam进行计数，使用代码是：  
mkdir $wkd/align  
cd $wkd/align  
source activate rna  
gtf="/public/reference/gtf/genencode/genencode.v25.annotation.g  
tf.gz"  
featureCounts -T 5 -p -t exon -g gene_id -a $gtf -o  
all.id.txt *.bam 1>counts.id.log 2>&1 &  
# 这样得到的 all.id.txt 文件就是表达矩阵啦，但是，这个  
featureCounts有非常多的参数可以调整。  
source deactivate
```

- 得到的文件如下


```

1 # Program:featureCounts v1.6.1;
Command:"featureCounts" "-T" "5" "-p" "-t" "exon" "-g"
"gene_id" "-a" "/public/reference/gtf/gencode/ge
2 Geneid Chr Start End Strand Length
/home/llwu/RNA/airway/2.align/bowite2/SRR1039523.bowite2.ba
m
3 ENSG00000223972.5
chr1;chr1;chr1;chr1;chr1;chr1;chr1;chr1;chr1
11869;12010;12179;12613;12613;12975;13221;13221;13453
12227;1
4 ENSG00000227232.5
chr1;chr1;chr1;chr1;chr1;chr1;chr1;chr1;chr1;chr1;chr1
14404;15005;15796;16607;16858;17233;17606;17915;18268;2
5 ENSG00000278267.1 chr1 17369 17436 -
68 9
6 ENSG00000243485.4
chr1;chr1;chr1;chr1;chr1;chr1
29554;30267;30366;30564;30976;30976
30039;30667;30503;30667;31097;31109
7 ENSG00000237613.2 chr1;chr1;chr1;chr1;chr1
34554;35245;35277;35721;35721
35174;35481;35481;36073;36081 -;-;-;-;-
8 ENSG00000268020.3 chr1 52473 53312 +
840 0
9 ENSG00000240361.1 chr1 62948 63887 +
940 0
10 ENSG00000186092.4 chr1 69091 70008 +
918 0

```

上面的文件，请务必仔细了解。

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