

NEURON
PROJECT

Questions (general)

- ~> How do neurons deal with morphological and biophysical variation?
- ~> What is the connection between the two?

"average"
classes vs. **VARIATION**

notes

The diagram illustrates a process flow from left to right:

- SNP_c** (with a circled **DANS**) has a curved arrow pointing to **CP_n**.
- PD** (with a curved arrow) leads to **degru**.
- CP_n** has a downward-pointing arrow labeled **! 2-4% coverage**.
- A bracket below **CP_n** and **degru** points to the text **↳ high energy degru**.

w/o attenuation
↳ also in mitral cells OR (G)

* pacemaking

1-3 Hz in rat, cell-to-cell variations
 hz 2-4 ms

The diagram shows a cross-section of a rat brain. Two groups of small circles representing neurons are labeled: SCN (Suprachiasmatic Nucleus) on the left and VTA (Ventral Tegmental Area) on the right. Below the brain, a vertical arrow points down to the text "DD n". To the right of the brain, a graph displays four sharp, narrow spikes of electrical activity, representing the rhythmic pacemaking of SCN neurons.

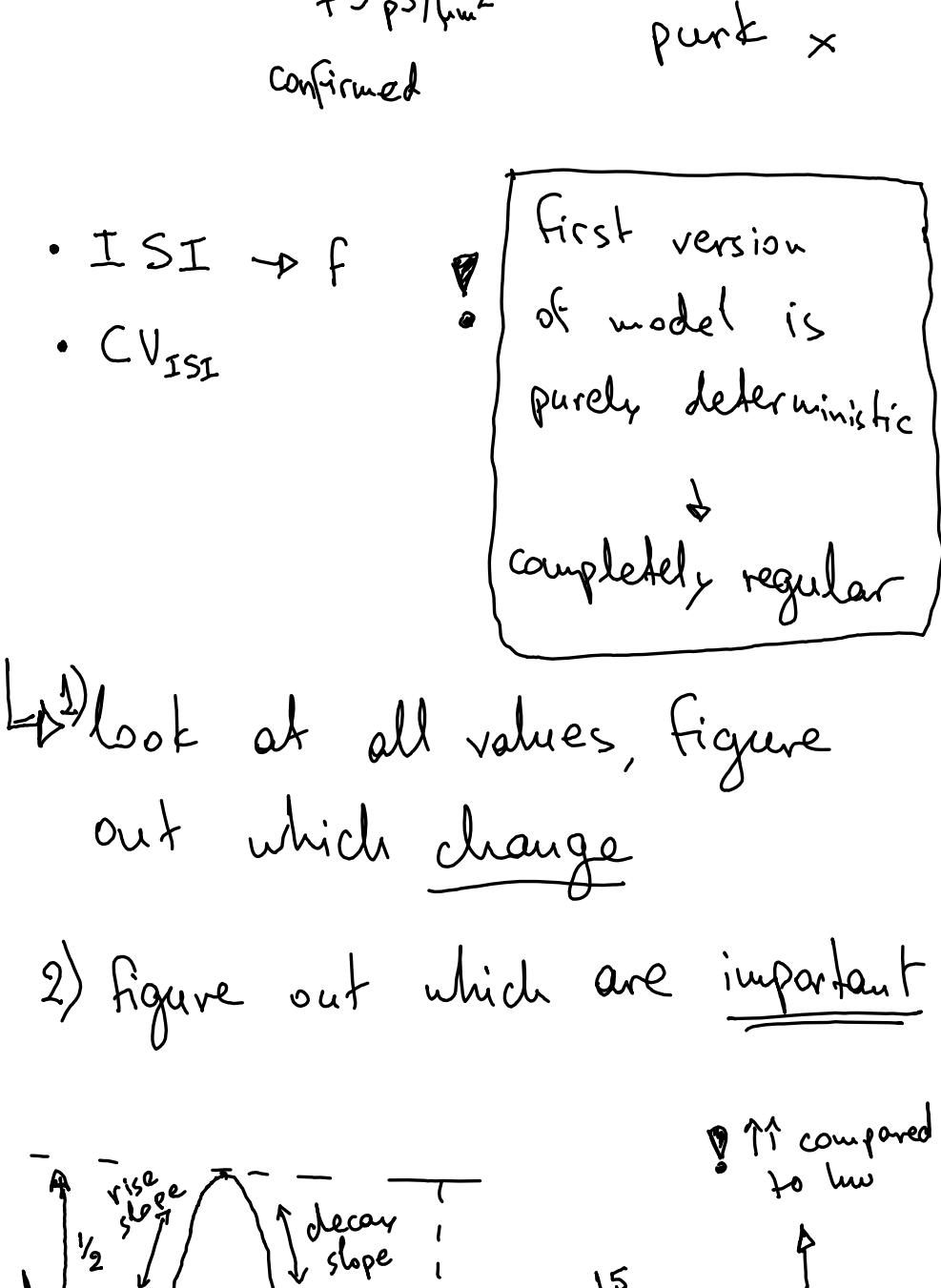
→ small

I_H HCN channels

to understand firing / excitability profile

Cable Theory

→ $d_m^{\frac{3}{2}} = d_{D_1}^{\frac{3}{2}} + d_{D_2}^{\frac{3}{2}}$



The diagram illustrates the generation of an action potential and its subsequent after-hyperpolarization (AHP). It features a solid line representing membrane potential and dashed lines representing resting potential levels. An arrow labeled "induced by" points to the initial depolarization phase. The peak of the action potential is labeled "AP". A vertical dashed line marks the time of repolarization. The period following repolarization is labeled "AHP ampl". An arrow labeled "AP recovery" points to the final phase where the membrane potential returns to baseline.

A hand-drawn diagram consisting of a large, roughly triangular or V-shaped outline. Inside this shape, near the top center, is a small, curved arrow pointing upwards and to the left. Below the arrow, the letters "thr" are handwritten in black ink.

Simple criterion:

$$\frac{dV}{dt} \geq 10 \text{ mV/ms}$$

→ usually ampl-freq. → NEG
bw as well?
(because of slope
changes)

↳ refractory period → INACTIVATED
 Na^+ channels

Ion channels

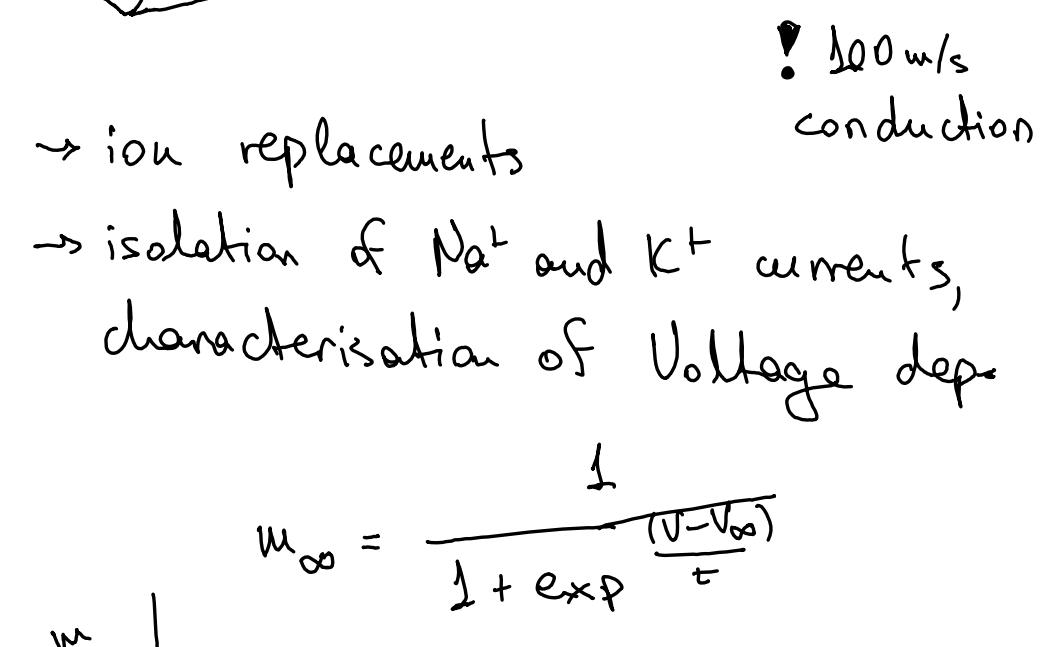
Jean-Marc

Gaillard

notes

Hodgkin-Huxley

Isopotential recording

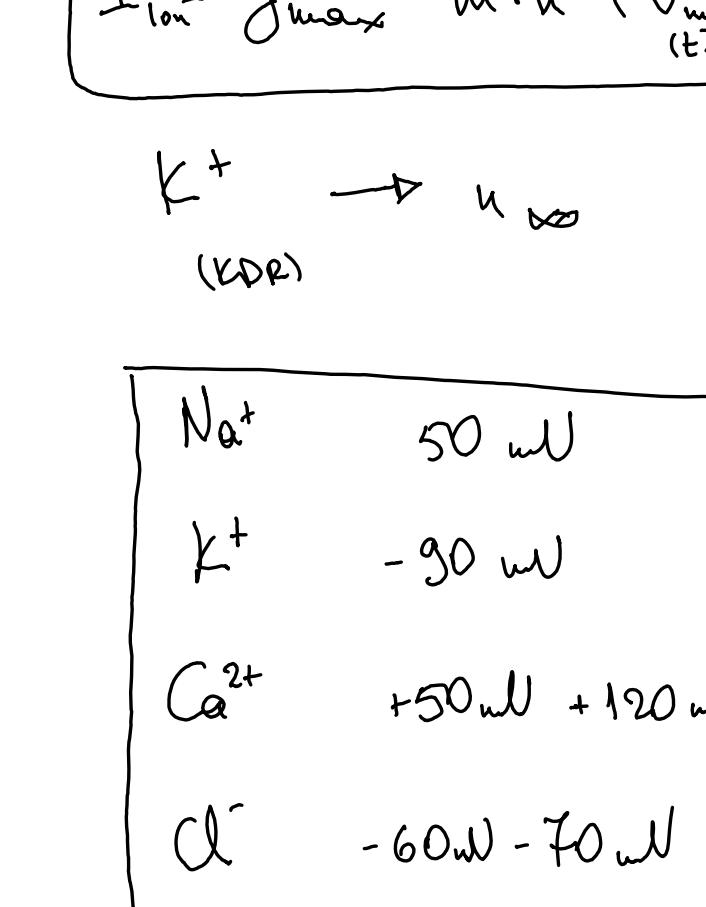


! 100 m/s

conduction

→ ion replacements

→ isolation of Na^+ and K^+ currents,
characterisation of Voltage dep.



$$I_{\text{ion}} = g_{\text{max}} \cdot m \cdot h \cdot (V_m - E_{\text{ion}})$$

$$g_{\text{max}} = g_{\text{unit}} \cdot N$$

$$g_{\text{Na}} \quad g_L \quad \rightarrow E_{\text{rev}} = -50 \text{ mV}$$

g_{KDR} 1 gating var.

g_A → 2 gating var!

$$g_H$$

$$g_{\text{CaL}}$$

$$g_{\text{SK}}$$

! dif. g_{max} per compartment

→ we can monitor V_m in any part of the neuron

axon fixed & 800 μm length

→ Educated, simplified guesses

NEURON

- 1) build compartments geom.
 - 2) define biophysics of compartment.
-

- 1) ↳ play first w/ avg wt model
- 2) ↳ explore conductance changes
(to match to phys)
↳ (grid) (?) search ranges
in an efficient way
- 3) plug conductances in to morphs



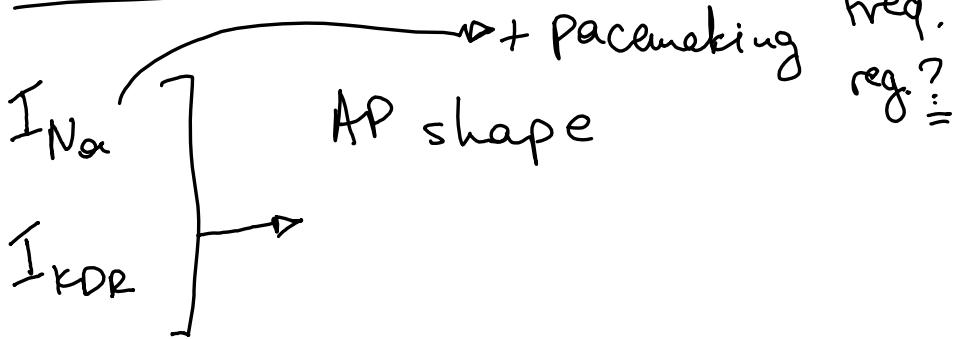
infer
parameters
from recordings

→ clone github repository
locally

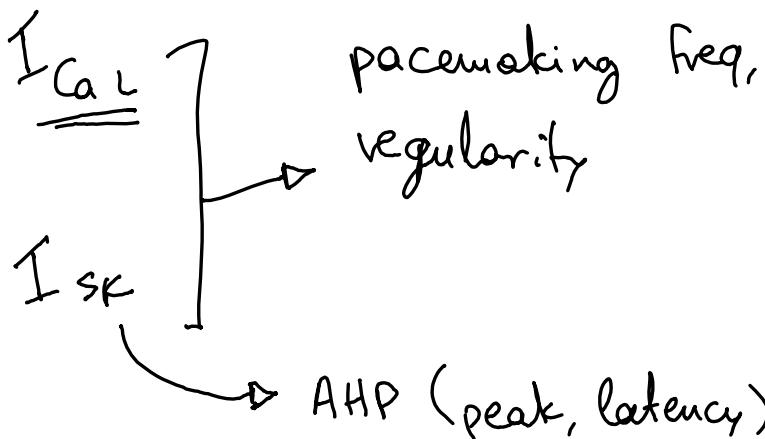
21.11.2025

- ✓ 1) Literature
- ✓ 2) Data → sets
- 3) model → running → play w/ pattern.
- 4) model → implement KO morphs

Currents



I_{I+} \longrightarrow sag, \downarrow rebound delay



Features altered in KO

g_{Na}, g_{CaL} (g_{SK})

- frequency $\downarrow 65\%$.
- regularity $\rightarrow CV_{ISI} \uparrow 172\%$

- AP ampl $\uparrow 10\% \text{ (Na}^+?)$
- rise \searrow
decay \nwarrow AP h_w $\uparrow 11\%$.
in our case:
decay contr. more than rise $\rightarrow K_w 3$

g_{KDR}

- AHP latency $\uparrow 74\%$.

~~g_{KDR}, g_{SK}~~
 $K_w 3$ SK

✓ check thr, slope \uparrow , slope \downarrow

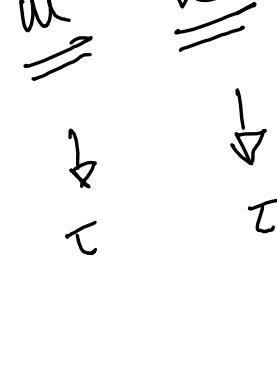
dV/dt peaks, d^2V/dt^2 peaks

Goals

- ✓ 1) features to target
 - ✓ 2) parameters to target
 - 3) models to work on
- search algorithms?

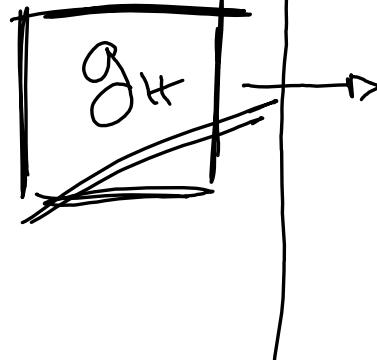
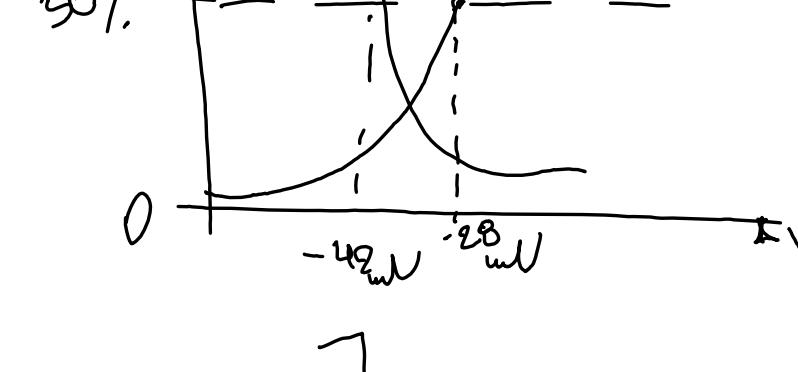
24.11.2025

$g_{Ca^{2+}}$ g_{Sk} g_{Na}



ACTIVATION AND INACTIVATION

VARIABLES OF g_{Na}



contributes to
sub-threshold
DEPOLARISATION



contributes to
REPOLARISATION

! HOWEVER, overall minor
contribution + major contr.
in features that we do
not see any change in
(sag ampl., rebound delay)

$(g_{Na}) \rightsquigarrow$ indirectly
affecting
 $g_{IPR} \text{ (?)}$ (Moubarak
et. al
2022)



$g_{Na} \text{ (1)}$

$g_{Ca} \text{ (2)}$

$g_{IPR} \times$

$E_h = -40 \text{ mV}$

$g_{Sk} \text{ (4)}$

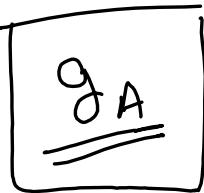
$g_A \times$

g_L

ADDITIONAL INFO

24.11.2025||

2007 paper



→ higher density @
the same, uniform
in dendrites

? Also dif. parameters

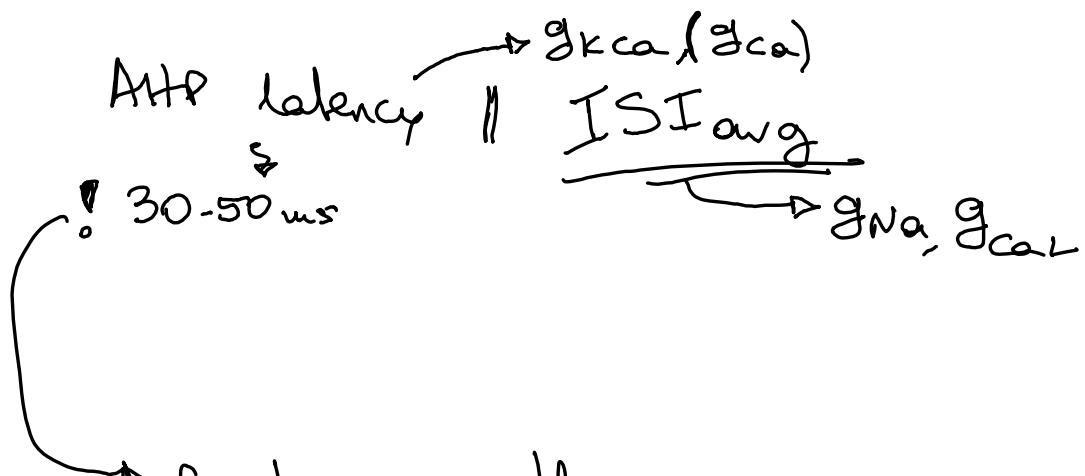
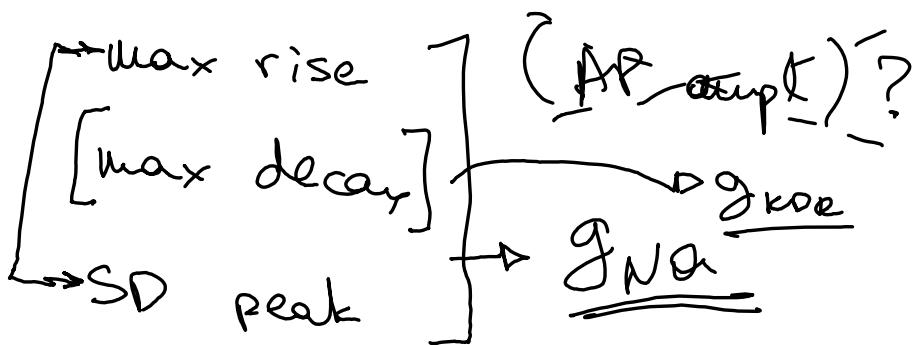
Moubarak

2022

↳ indirect correlation

of decay w/ g_{Na}

! \uparrow freq. \rightarrow \uparrow regularity



If it was the main determinant of fir. freq., we would have a way higher rate (10-20 Hz)

Conductances ranges to test

→ 5-10 fold change

→ $25-165 \text{ } \mu\text{S}/\text{cm}^2$

g_{Na}

range used in Moubarek
papers

↳ ! to look up:

CONDUCTANCE RANGES

in HT models

↳ specifically in DA neurons?

24.11.2025

Features

- (AP ampl?)
- AP rise
 - AP decay
 - SD peak
 - AHP latency
 - ISI avg

Parameters

- g_{Na}
- g_{Ca}
- g_{KDR}
- g_{KCa}

↓

possible addition
(later, no priority): thr

As a feature that
does not change
among the 2 genotypes.

for starters

- explore SD only
- keep ABD and nABD homogeneous

Presentation

- 30 - 40 mins
- all students present
- single project line

